

The medfate reference book

true

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Preface

This is a reference book for the models implemented in **medfate** and **medfatedland**, two R packages that provide functions for the simulation of functioning and dynamics of Mediterranean forests and landscapes.

How to use this book

This *reference book* is meant to help you understand the static and dynamic models included in packages **medfate** (ver. **3.2.0**) and **medfatedland** (ver. **2.0.5**). Hands-on user guides to run simulation functions and static functions can be found as *package vignettes* within the packages. As any reference book, you are not expected to read the book linearly, but to jump to specific book sections whenever you have doubts about the design or implementation of certain processes or calculations.

The first part of the book presents the main functions of the package and describes the soil, vegetation and meteorology inputs that apply to all simulation models. After that, the book presents in detail each simulation model using a set of chapters. The first of those provides an overview of the model design and the remaining ones describe the details of formulations by processes. Process formulations that are common to more than one model are described only once to avoid repeating material already presented, but links are provided whenever needed. Static models and their corresponding R functions are presented grouped at the end of the book, before the appendices, which include parametrization information and a list of all symbols for reference.

In this book we use `objectname` or `variablename` to refer to an R objects or a variable within data frames, and `functionname()` to refer to a package function. Whenever relevant, we indicate the correspondence between mathematical symbols, their units and the names used within the R package.

Our aim is to continuously update this reference book along with package developments, so that users have detailed and up-to-date information about the models at the time functions are run. Since the book evolves with the package, after any model application we recommend users to store a **PDF version** of the reference book to be sure it matches the version the package reported in their application report or article.

An ongoing project

You should look at **medfate** and **medfatedland** R packages as an ongoing research project for the development of forest ecosystem modelling tools. The models included in the package may contain bugs, and provide wrong predictions (all models are *wrong*, remember?). Package functions may evolve in the future, so that you may have to modify your scripts after package updates (hopefully not too often). Nevertheless, we feel the tool is mature enough to become useful for applications and that is why we made an effort to document it thoroughly. The R packages are the result of collaborative work between modellers and experts in different disciplines. If your own expertise includes any of the processes that are modelled in the package and you feel that your expertise could be helpful to improve the package, you are more than welcome to contact us!

Funding

I have been intensively working on **medfate** since 2013, when I obtained a *Ramon y Cajal* research fellowship from the Spanish government (Reference: RYC-2012-11109; Amount: 308.600 €). An investment on a long-term modelling project could not have started with this initial financial help and that of the following competitive research projects:

- **Title:** Forecasting of ecosystem services on forests: impacts and adaptation to extreme climate events (FORESTCAST). **Financial Entity:** Ministerio de Economía y Competitividad (CGL2014-59742-C2-2-R). **Amount:** 152.460 €. **Duration from:** 01/01/2015 **to:** 31/12/2017. **PI:** Lluís Brotons/Miquel De Cáceres.
- **Title:** Modeling plant water use and drought resistance strategies across scales (DRESS). **Financial Entity:** Ministerio de Economía y Competitividad (CGL2017-89149-C2-2-R). **Amount:** 93.170 €. **Duration from:** 01/01/2018 **to:** 31/12/2021. **PI:** Miquel De Cáceres.
- **Title:** Boosting process-based models to project forest dynamics and associated ecosystem services at stand-to-regional scales (BOMFORES). **Financial Entity:** Ministerio de Ciencia e Innovación (PID2021-126679OB-I00). **Amount:** 75.020 €. **Duration from:** 01/09/2022 **to:** 31/08/2024. **PI:** Miquel De Cáceres.

Besides this specific funding, this model developments could not have occurred without the institutional support of the Forest Science and Technology Center of Catalonia (CTFC), where I worked until March 2021, and Center for Ecological Research and Forestry Applications (CREAF), where I currently coordinate its Ecosystem Modelling Facility.

Acknowledgements

The idea of an R package devoted to Mediterranean vegetation evolved after a previous attempt at modelling the dynamics of plant cohorts within a landscape model that tried to extend the MEDFIRE model. I made these initial efforts when I was still a post-doc at CTFC under the supervision of Lluís Brotons, to whom I am grateful for having greatly expanded my view of ecology and its applications. Undertaking this initial endeavor made us contact two excellent researchers in fire and drought ecology respectively, Juli G. Pausas (CIDE-CSIC, Spain) and Jordi Martínez-Vilalta (CREAF-UAB, Spain). While interacting with Juli was rewarding and fostered my interest in wildfires and fire ecology, Jordi had a profound influence on the project. Collaborating with him caused a slow but steady drift of my research interests towards the soil water balance, plant hydraulics and physiological responses to drought. His influence on my work was not so apparent when we published water balance model (and launched the R package) with the help of the forest hydrologist Pilar Llorens (CSIC, Spain) and other researchers (Rafa Poyatos, Lluís Coll and Pere Casals). However, it became undeniable when I started studying plant hydraulics, trying to understand and implement Sperry's model in the package. Jordi made me interact with people in his research group and introduced me to Maurizio Mencuccini, another great ecophysicist with whom he had been working for a long time. I have to thank Maurizio and Jordi for encouraging me to continue working on this development project and overcome the difficulty that entails learning scientific disciplines quite far from my previous experience.

The development of the package has benefited from many colleagues who have provided me with ideas or have struggled to use the models within it. Asaf Karavani was among the first package users besides me. Together, we used the package to predict soil moisture dynamics in forest plots where mushroom production was being monitored. Antoine Cabon (CTFC, Spain) was also an early user of the package. I have enjoyed endless discussions with him about several processes and how to implement them. While he was the Ph.D. student and I one of his advisors, I felt we both learned together. Moreover, Antoine has contributed with some functions to the package (and I hope he will contribute more!). Another key person for the development of **medfate** was working with Victor Granda (Ecosystem Modelling Facility at CREAF,

Spain), whose great programming skills and profound knowledge about R technologies has encouraged me to embrace new programming habits and skills. I owe the idea of separating packages **meteoland** and **medfate** to Roberto Molowny (Ecosystem Modelling Facility at CREAF, Spain) an excellent modeller who has also contributed to the design of dispersal in **medfate**. More recently, collaboration with another ecophysiologist, Nicolas Martin-StPaul (INRA, France), has stimulated improvements both in **meteoland** and **medfate**. In fact, Nicolas has became another key person in my research, with whom I share the interest for one of the interfaces between fire and drought, namely fuel moisture dynamics. Thanks to him and other colleagues (Julien Ruffault and François Pimont), since ver. 3.0 **medfate** includes an additional sub-model of plant hydraulics, based on SurEau. Collaboration with Shengli Huang (Forest Service, US) on adapting the package for the US Forest Inventory and Analysis (FIA) program has expanded the horizons of its application. Collaboration with Josep Maria Espelta (CREAF) and Francisco Lloret (UAB-CREAF) has been key to the design of regeneration processes. The package have also benefited from the feed back of other users, such as Francesco d'Adamo (CREAF, Spain), Paula Martín (CTFC, Spain) and María Gonzalez Sanchís (UPV, Spain).

I would not like to finish the acknowledgements without thanking my former colleagues at CTFC (located in the small town of Solsona, Catalonia, Spain), for their collaboration and support. Particularly, Lluís Coll, Aitor Ameztegui and Pere Casals have been continuously supporting my efforts and finding new model applications. Moreover, they have contributed to the knowledge within the package with their expertise in forest dynamics, forest management, soils and fire issues. I also thank Mario Beltrán and Assu Gil for teaching me forestry basics.

Finally, I would like to thank José Manuel, who takes care of me and has to endure my endless talking about the package and its models, to my family for being there at difficult times, including my three nephews (Aran, Jan and Nil) that constantly remind me the joy of life.



Part I

Preliminaries

Chapter 1

Introduction

This chapter provides an overview of the purpose of developing the **medfate** and **medfatedland** R packages, their main simulation functions and expected applications.

1.1 Purpose

Being able to anticipate the impact of global change on forest function and dynamics is one of the major environmental challenges in contemporary societies. However, uncertainties in the mechanisms underpinning forest function and practical constraints in how to integrate available information still hinder the availability of robust and reliable predictive models. Despite the amount of knowledge accumulated about the function and dynamics of forest ecosystems and the plethora of observations and modelling tools already available, further efforts are necessary to achieve the challenge of integrating different global change drivers into simulation frameworks useful for research and applications.

1.1.1 Package **medfate**

The R package **medfate** has been designed as a platform to simulate the function and dynamics of individual forest stands at temporal scales from days to years. Climate is the main environmental driver covered by the package and the ecological, hydrological and physiological processes surrounding these are fundamental for the simulation models included in the package. In particular, the package allows the simulation of daily water balance of soils and plants within forest stands. It also allows simulating the growth, mortality and regeneration processes of a set of woody plant cohorts competing for light and water within a forest stand. Finally, additional package functions allow relating the amount of plant biomass and the water status of plant live and dead tissues to fuel characteristics and, hence, fire hazard.

The version of the reference manual that you are reading is intended to reflect **version 3.2.0** of the package.

1.1.2 Package **medfatedland**

The R package **medfatedland** has been designed to extend the capabilities of **medfate** to a spatial context, most usually at the landscape or regional level. It allows running the stand-level models available in **medfate** on a set of spatial units (normally points or cells) within a target area. The package allows coordinating the dynamics of multiple stands via the evaluation of demand-based management scenarios. Additionally, the package allows considering spatial hydrological processes in forested watersheds. Hence, **medfatedland** can also be used as a tool for eco-hydrological applications.

The version of the reference manual that you are reading is intended to reflect **version 2.0.5** of the package.

1.1.3 Companion packages

Packages **medfate** and **medfateland** are more easily used in conjunction with two other packages, called **meteoland** and **medfateutils**. Package **meteoland** assists the estimation of weather variables over landscapes, whereas **medfateutils** provides multiple functions to assist the creation of inputs and the parameterization of simulation models. Together the four R packages conform a modelling framework designed to help simulating the function and dynamics of forest ecosystems, with a particular focus on drought impacts under Mediterranean conditions (Fig. 1.1).

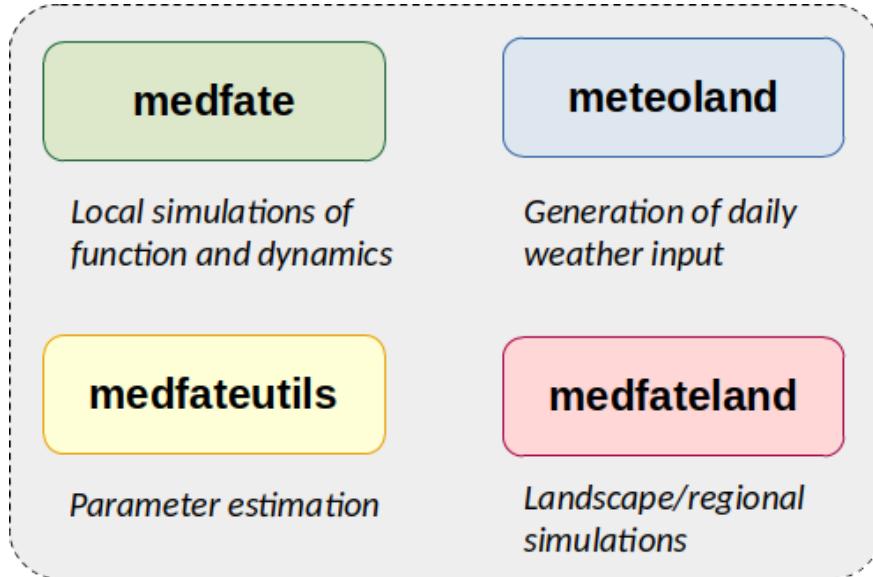


Figure 1.1: Set of packages conforming the medfate modelling framework

1.2 Installation

1.2.1 Requirements

Both **medfate** and **medfateland** have a user interface in R, but models have been coded in **C++ language**, and the resulting simulation functions are linked to R via the **Rcpp** package. This means that whenever installation is done from sources you will need a C++ compiler (i.e. Rtools in Windows). Note that compilers will always be a requirement when installing packages from GitHub.

1.2.2 Installing medfate

Package **medfate** can be found at CRAN, where it is updated every few months. Hence, it can be installed as any other R package using:

```
install.packages("medfate")
```

Users can also download and install the latest stable versions GitHub as follows (required package **remotes** should be installed/updated first):

```
remotes::install_github("emf-creaf/medfate")
```

Among other dependencies, installing **medfate** requires package **meteoland**, as it links to some of its C++ functions. Note that some package dependencies are only *suggested*, so that they are not automatically installed when installing **medfate**.

1.2.3 Installing medfate`land`

Since both packages evolve together, installing **medfate`land`** normally requires an up-to-date version of **medfate**. The package **medfate`land`** is currently only available at GitHub, and can be installed using:

```
remotes::install_github("emf-creaf/medfateland")
```

Among other dependencies, installing **medfate`land`** requires packages **medfate** and **meteoland**, as it links to some of their C++ functions.

1.3 Main functions in medfate

1.3.1 Dynamic simulation functions

Dynamic simulation functions in **medfate** include three nested simulation levels (Fig. 1.2):

1. Forest water and energy balance can be studied for a given forest stand using function **spwb()** (soil-plant-water-balance). The same function is used to run three different models (**Granier**, **Sperry** or **Cochard**), that correspond to two levels of complexity (*basic* or *advanced*; see Fig. 1.2). The user should choose one model or another depending on the intended application and data availability. Water balance simulations include hydrological processes (rainfall interception, soil infiltration, percolation and evapotranspiration) and plant physiological processes related to transpiration (hydraulics, photosynthesis and stomatal regulation). Most processes are implemented at the daily scale, although some operate at subdaily time steps for *advanced* models. Function **spwb()** has a closely related function **spwb_day()** that allows focusing on sub-daily processes. Finally, one can simulate plant physiological processes only, while having soil moisture as a prescribed dynamic input like weather, using function **pwb()** (plant-water-balance).
2. Changes in primary (leaf area) and secondary (sapwood) growth are key to evaluate the influence of climatic conditions on plant and forest structure and function. Function **growth()** extends the previous models because it allows simulating carbon balance, tissue growth and mortality of a set of plant cohorts competing for light and water in a given forest stand. Analogously to the simulation of water balance, **growth()** has a closely related function **growth_day()** that allows focusing on sub-daily processes (i.e. carbon balance).
3. Finally, function **fordyn()** complements growth and mortality processes with recruitment (from seeds) and resprouting, thus completing the minimum set of demographic processes needed to simulate Mediterranean forest dynamics. The function splits the period to be simulated by years and makes internal calls to **growth()** for the simulation of growth and mortality. **fordyn()** is suited to simulate the inter-annual variation in forest structure and composition, while accounting for the biophysical and physiological processes provided by the former models. In addition, it allows simulating human interventions (i.e. tree or shrub cuts) on forest stands.

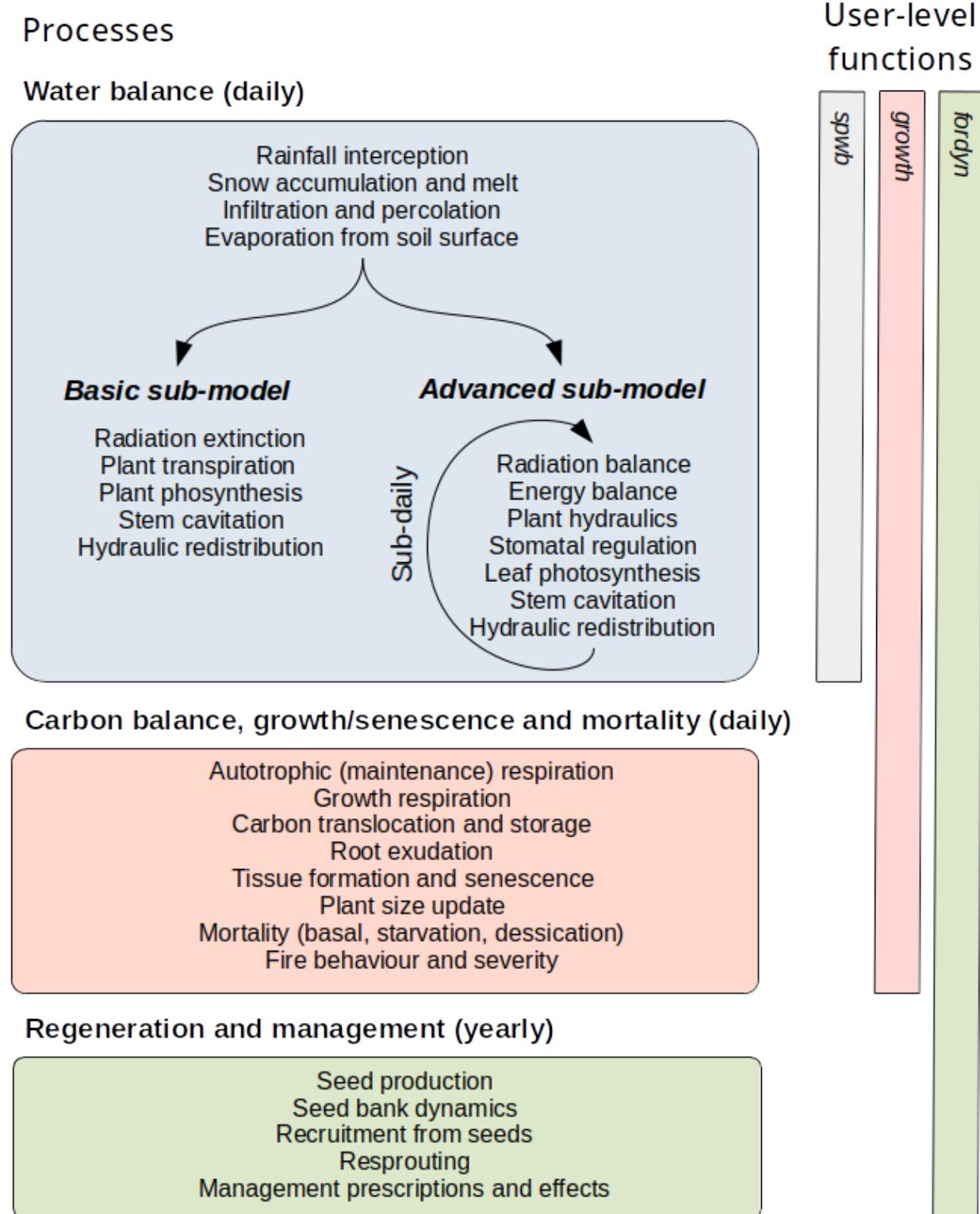


Figure 1.2: Nested relationships between medfate simulation functions and processes included in each model

1.3.2 Sub-model functions

Many of the sub-models included in **medfate** are implemented as C++ functions and internally called by the simulation functions implementing top-level models. Most sub-models have, however, their corresponding R function to make them directly available to the user. This facilitates understanding the different sub-models and a more creative use of the package. However, most users will not normally use them in their simulation workflows. Sub-model functions are grouped by *subject*, which is included in the name of the function. The different sub-model functions are (by subject):

Group	Description
<code>biophysics_*</code>	Physics and biophysics
<code>carbon_*</code>	Carbon balance
<code>fuel_*</code>	Fuel properties
<code>fire_*</code>	Fire behaviour and severity
<code>hydraulics_*</code>	Plant hydraulics
<code>hydrology_*</code>	Canopy and soil hydrology
<code>light_*</code>	Light extinction and absorption
<code>moisture_*</code>	Live tissue moisture
<code>pheno_*</code>	Leaf phenology
<code>photo_*</code>	Leaf photosynthesis
<code>root_*</code>	Root distribution and conductance calculations
<code>soil_*</code>	Soil hydraulics and thermodynamics
<code>transp_*</code>	Stomatal regulation, transpiration and photosynthesis
<code>wind_*</code>	Canopy turbulence

1.3.3 Static functions

Package **medfate** includes a number of functions to examine static properties of plants conforming forest stands, summary functions at the stand level or vertical profiles of several physical properties:

- `plant_*`: Cohort-level information (species name, id, leaf area, height...).
- `species_*`: Cohort-level attributes aggregated by species (e.g. basal area).
- `herb_*`: Attributes of the herbaceous layer (e.g. fuel density or leaf area index).
- `stand_*`: Stand-level attributes (e.g. basal area).
- `vprofile_*`: Vertical profiles (light, wind, fuel density, leaf area density).

Vegetation functioning and dynamics have strong, but complex, effects on fire hazard. On one hand, growth and death of organs and individuals changes the amount of standing live and dead fuels, as well as downed dead fuels. On the other, day-to-day changes in soil and plant water content changes the physical properties of fuel, notably moisture content of live and dead fuels. Package **medfate** provides functions to estimate fuel properties and potential fire behaviour in forest stands. Specifically, `fuel_FCCS()` calculates fuel characteristics for a set of fuel strata; and a fire behaviour model is implemented in function `fire_FCCS()` to calculate the intensity of surface fire reaction and the rate of fire spread of surface fires assuming a steady-state fire. Fuel and fire behavior functions allow obtaining the following:

1. Fuel characteristics by stratum.
2. Surface fire behavior (i.e. reaction intensity, rate of spread, fireline intensity and flame length).
3. Crown fire behavior.
4. Fire potential ratings of surface fire behavior and crown fire behavior.

1.4 Main functions in medfateland

Package **medfateland** allows simulating forest functioning and dynamics on sets forests stands distributed across space, with or without spatial processes (Fig. 1.3).

When spatial processes are omitted, the package offers a set of simulation functions that are analogous to those of package **medfate** but allow processing multiple forest stands sequentially or in parallel:

- Forest water and energy balance can be studied for a set of forest stands using functions `spwb_spatial()` or `spwb_spatial_day()`.
- Growth and mortality of a set of plant cohorts competing for light and water can be simulated for a set of forest stands using functions `growth_spatial()` or `growth_spatial_day()`.
- Forest dynamics arising from competition for light and water can be simulated for a set of forest stands using function `fordyn_spatial()`.

The package offers a simulation function where the dynamics of forest stands are related by management decisions:

- Function `fordyn_scenario()` allows simulating `fordyn()` on forest stands, while applying a demand-based forest management scenario. Simulations with this function also include seed dispersal among forest stands.

Finally, there are three functions that are meant to simulate watersheds, including spatial processes:

- Functions `spwb_land()` and `growth_land()` include distributed hydrological models that allow simulating daily water balance or growth processes on the cells of a watershed while accounting for overland runoff, subsurface flow and groundwater flow between cells.
- Forest dynamics arising from competition for light and water can be simulated in a watershed using function `fordyn_land()`. Simulations with this function also include seed dispersal among grid cells.

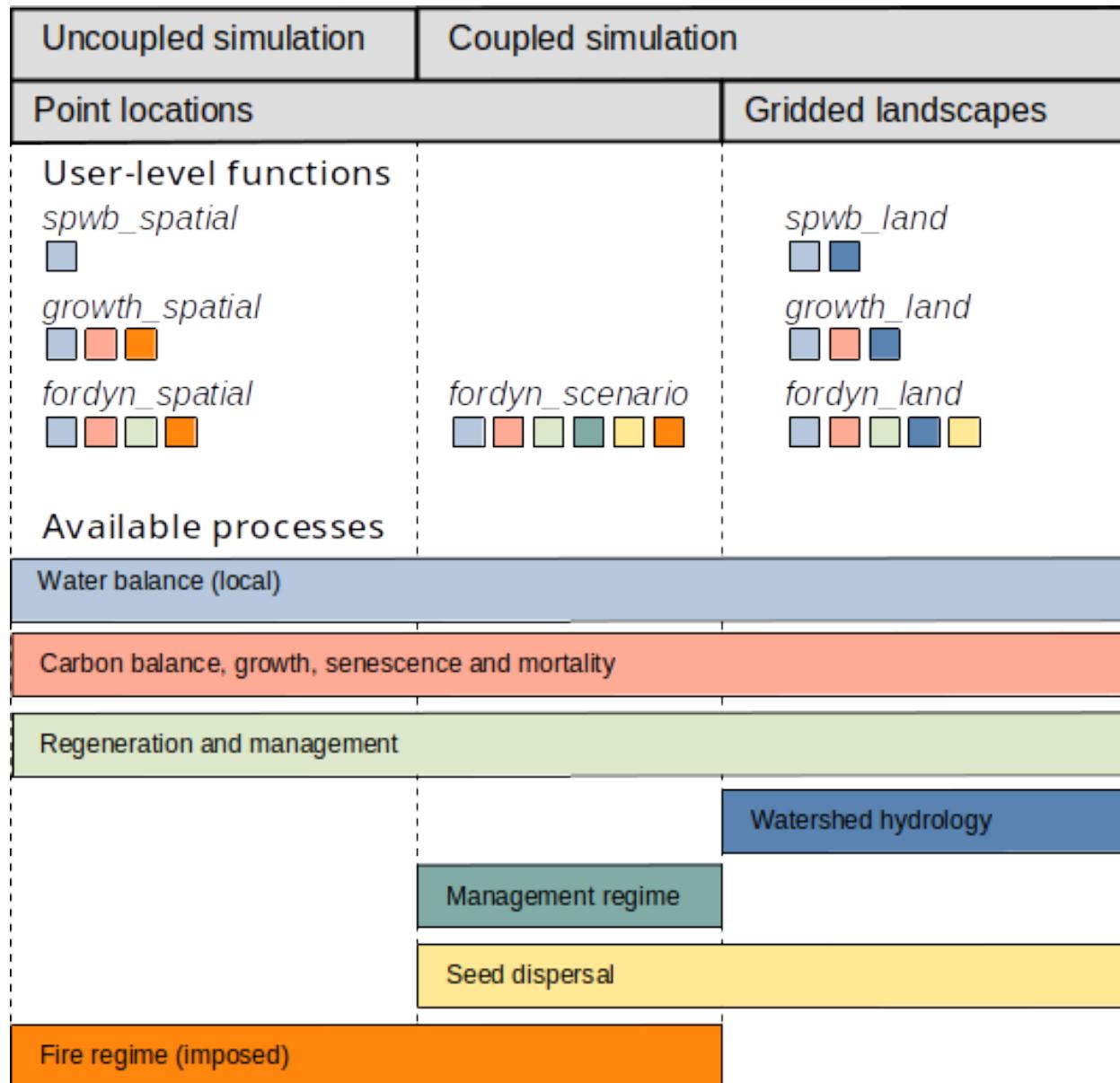


Figure 1.3: Overview of simulation functions in medfateiland and the processes available for each of them. Colored squares below each simulation function indicate the processes included

Chapter 2

Model inputs

This chapter describes the inputs required by simulation models of **medfate** and **medfate`land`** packages. First, we describe the R structures required for the two packages. Then, we provide the details for four different kinds of inputs:

- (1) *Vegetation existing in the forest stand.* This information should be available at a level of detail appropriate for the model design, which in our case includes not only species composition and/or structure, but also functional parameters of woody plants. Vegetation structure is considered static in `spwb()` calls, but becomes dynamic in calls to `growth()` or `fordyn()`. Hence, in these functions the input vegetation describes the starting point.
- (2) *Information on the physical structure of soil.* Soil information is also required but, unlike vegetation, soil's physical structure is considered static.
- (3) *Climatic forcing.* Weather is always a dynamic input that defines the temporal variation of environmental conditions in which physical and biological processes occur.
- (4) *Topographic and geographic information.* Topography and latitude are also required, because internal calculations of some physical variables require them.

The last section describes additional input parameters that are used to specify global options of simulation functions.

Since preparing inputs for simulations can be daunting for most users, package **medfateutils** provides a set of utility functions aimed at facilitating the preparation of vegetation and soil inputs, whereas package **meteoland** provides functions to assess daily weather over landscapes.

2.1 Input structures

2.1.1 Input structures in medfate

Package **medfate** deals with simulations at stand level and, hence, the data structures defined in the package are meant to encapsulate information about a target forest stands and the properties of the soil where it grows:

- S3 class `soil` is a list of soil physical properties, including both parameters and state variables. Soil objects are described in subsection 2.3.3.
- S3 class `forest` is a list of forest inventory data (i.e. mainly tree and shrub measurements, but can include herbaceous vegetation) corresponding to a single forest stand. Forest plot objects are described in subsection 2.4.4.

- S3 classes `spwbInput` and `growthInput` are lists containing all the vegetation parameters and state variables necessary to run water-balance simulations (function `spwb()` and alike) and forest growth simulations (function `growth()` and alike). Objects of classes `spwbInput` and `growthInput` are created using functions whose input includes a `forest` object and a `soil` object.

The figure below (Fig. 2.1) can be used to illustrate the workflow of soil/forest input creation for the main simulation functions in `medfate`. Note the different inputs in the case of `fordyn()`.

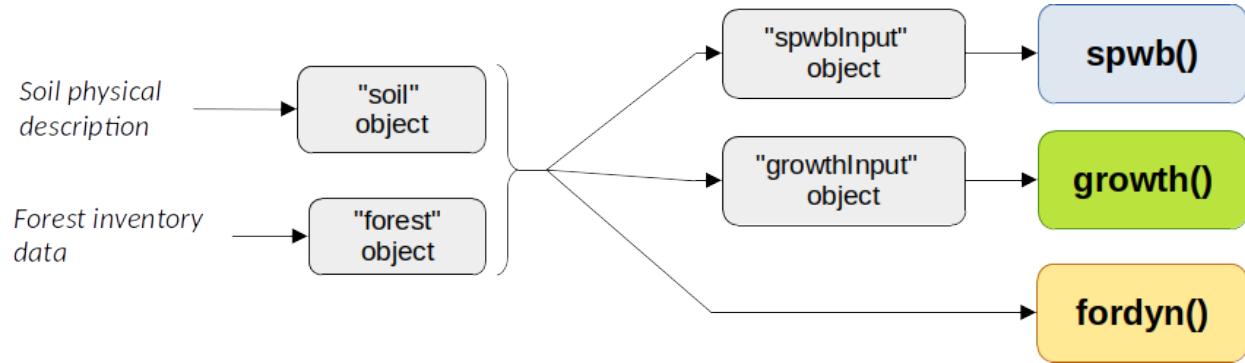


Figure 2.1: Schematic workflow of input creation for simulation functions in `medfate`

2.1.2 Input structures in `medfateLand`

Spatial information for `medfateLand` needs to be provided in `sf` objects (i.e. data frames with geometry, see package `sf`), whose actual columns required depend on the simulation function to be used, as described in the following table.

Column	Description	Use
<code>geom</code>	Spatial geometries (points, polygons, ...)	Always required
<code>id</code>	Stand identifiers	Required for <code>*_spatial()</code> , <code>*_spatial_day()</code> and <code>fordyn_scenario()</code>
<code>elevation</code>	Elevation values above sea level (in m)	Always required
<code>slope</code>	Slope values (in degrees)	Always required
<code>aspect</code>	Aspect values (in degrees)	Always required
<code>land_cover_type</code>	Land cover type: <i>wildland</i> , <i>agriculture</i> , <i>rock</i> , <i>artificial</i> or <i>water</i>	Required for <code>*_land()</code> , optional for <code>*_spatial()</code> and <code>*_spatial_day()</code>
<code>forest</code>	Objects of class <code>forest</code>	Always required
<code>soil</code>	Objects of class <code>soil</code>	Always required
<code>state</code>	Objects of class <code>spwbInput</code> or <code>growthInput</code>	Optional to recover the final state of a previous simulation
<code>meteo</code>	Weather data frames	Optional
<code>crop_factor</code>	Crop evapo-transpiration factor	Only required for <i>agriculture</i> land cover type
<code>management_arguments</code>	Management arguments for each stand	Optional for <code>fordyn_spatial()</code>
<code>management_units</code>	Management unit corresponding to each stand	Required for <code>fordyn_scenario()</code>
<code>represented_area</code>	Area represented by each stand or cell	Required for <code>fordyn_scenario()</code> and <code>*_land()</code>

Column	Description	Use
<code>waterOrder</code>	Integers indicating cell processing order.	Required for <code>*_land()</code>
<code>waterQ</code>	Lists of water discharge values to neighbors.	Required for <code>*_land()</code>
<code>queenNeigh</code>	Lists of integers identifying the (up to 8) queen neighbors.	Required for <code>*_land()</code>
<code>channel</code>	Logical values indicating whether cells belong to the channel network.	Required for <code>*_land()</code>
<code>depth_to_bedrock</code>	Depth to bedrock (mm).	Required for <code>*_land()</code>
<code>bedrock_conductivity</code>	(saturated) conductivity (in $\text{m} \cdot \text{day}^{-1}$).	Required for <code>*_land()</code>
<code>bedrock_porosity</code>	Bedrock porosity ([0-1]).	Required for <code>*_land()</code>
<code>snowpack</code>	Snow water equivalent content of the snowpack (mm).	Required for <code>*_land()</code>
<code>aquifer</code>	Water content of the aquifer in each cell.	Required for <code>*_land()</code>

2.2 Latitude and topography

Topography has a relatively small direct influence in **medfate** simulations, because most topographic effects are assumed to be taken into account in the estimation of weather inputs. Nonetheless, *elevation* in meters is needed to estimate atmospheric pressure and air density (see 2.5), whereas *aspect* (in degrees from North) and *slope* (also in degrees) are needed in addition to *latitude* (also in degrees North) to estimate sunrise/sunset hours and potential radiation. Topographic variables should be readily accessible for users having access to a digital elevation model of the target area.

2.3 Soil description

Soils are described in **medfate** using $S > 1$ different soil layers ($s \in \{1, 2, \dots, S\}$). Here the word *soil* refers to depths that plant rooting systems can reach (i.e., the critical zone), including cracks within the bedrock. The number and size of soil layers may correspond to changes in soil properties along depth, but also can be chosen to reflect differences in plant rooting depths. The physical properties of the soil are needed to estimate soil hydraulic properties and are considered **static soil parameters** in simulations. In contrast, soil moisture content, other variables that depend on moisture (e.g. soil water potential and soil conductivity) and soil temperature, if the soil energy balance is considered, are all **soil state variables**.

2.3.1 Soil physical properties

Soil physical properties, such as *texture* (i.e. volume percent of sand, silt and clay), *organic matter content*, *bulk density* or *rock fragment content*, can differ between soil layers, and this has important consequences for soil water retention capacity and soil hydraulics. Specifying layers with an elevated rock fragment content (e.g. layers of weathered rock where soil particles are scarce) may be important in seasonally-arid climates like the Mediterranean, because plants often extend their roots into cracks existing in the parent rock to access water during summer (Ruffault *et al.* 2013).

For each soil layer s , the following physical parameters are needed:

Symbol	Units	R	Description
d_s	mm	<code>widths</code>	Width of soil layer s
$P_{clay,s}$	%	<code>clay</code>	Percent volume of clay (within soil particles fraction) in layer s
$P_{sand,s}$	%	<code>sand</code>	Percent volume of sand (within soil particles fraction) in layer s
OM_s	%	<code>om</code>	Percentage of organic mater per dry weight (can be missing) in layer s
BD_s	$g \cdot cm^{-3}$	<code>bd</code>	Bulk density in layer s
$P_{rocks,s}$	%	<code>rfc</code>	Rock fragment content as percent volume (within whole soil) in layer s

The percent volume of silt is 100% minus the percent volume of clay and sand. Whenever possible, soil physical properties should be measured in soil profiles conducted at the target forest plot (although soil profiles rarely reach rooting depths of deeply-rooted plants). Soil input data should be arranged in a `data.frame` with soil layers in rows and physical variables in columns (see function `defaultSoilParams()`).

Package **medfateutils** includes a function `soilgridsParams()` to fetch soil information from SoilGrids.org, a global soil database currently providing soil data at 250m scale. This can be helpful to users lacking local soil measurements, but the uncertainty of SoilGrids estimates can be high for some areas and soil properties, especially soil depth and rock fragment content.

The depth of a given soil layer s (Z_s ; in mm) is defined as the sum of layer widths from the surface to the target layer:

$$Z_s = \sum_{i=1}^s d_s \quad (2.1)$$

And the overall *soil depth* (Z_{soil} ; in mm) is the sum of widths across all soil layers. As the soil may include deep rocky layers, the value of Z_{soil} may be larger than the soil depth reported in soil profiles.

2.3.2 Water retention curves

The **water retention curve** of a soil is the relationship between *volumetric soil moisture content* (θ , in $m^3 \cdot m^{-3}$ of soil, excluding rock fragments) and the corresponding *soil water potential* (Ψ , in MPa), i.e. the amount of work that must be done per unit quantity of pure water in order to transport reversibly and isothermally an infinitesimal quantity of water from a reference pool. The shape of the water retention curve (also called the *soil moisture characteristic curve*) depends on physical properties (mainly texture and bulk density, but also organic matter content). Since soil layers usually differ in their physical properties they also normally have different water retention curves.

Two water retention curve models are available in **medfate** (fig. 2.2):

1. *Saxton model*: In this model, volumetric soil moisture $\theta(\Psi)$ corresponding to a given water potential Ψ (in MPa) below field capacity (i.e., $\Psi < \Psi_{fc}$) is calculated using:

$$\theta(\Psi) = (\Psi/A)^{(1/B)} \quad (2.2)$$

where A and B depend on the texture and, if available, organic matter in the soil. If organic matter is available, A and B are calculated from P_{clay} , P_{sand} and OM following Saxton & Rawls (2006). Otherwise, they are calculated from P_{clay} and P_{sand} as indicated in Saxton *et al.* (1986). Volumetric changes between field capacity and saturation (i.e., $\Psi_{fc} \leq \Psi < 0$) are estimated using a linear function.

2. *Van Genuchten model*: The well known van Genuchten (1980) model is:

$$\theta(\Psi) = \theta_{res} + \frac{\theta_{sat} - \theta_{res}}{[1 + (\alpha \cdot \Psi)^n]^{1-1/n}} \quad (2.3)$$

where $\theta(\psi)$ is the water retention, θ_{sat} is the saturated water content, θ_{res} is the residual water content, α is related to the inverse of the air entry pressure, and n is a measure of the pore-size distribution.

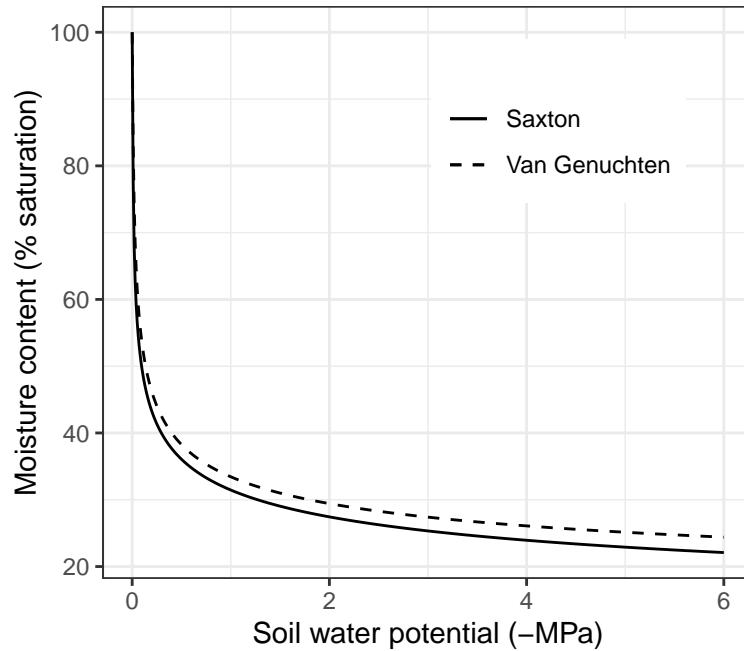


Figure 2.2: Water retention curves under the Saxton and Van Genuchten models, for the same physical attributes (25% sand, 25% clay, 50% silt and bulk density of $1.5 g \cdot cm^{-3}$). Parameters of the Van Genuchten curve have been estimated using pedotransfer functions given in Tóth *et al.* (2015).

2.3.3 Soil initialization

Soil initialization is needed to estimate hydrological parameters for each soil layer s from its physical attributes. Soil initialization is done using function **soil()**, which returns an object of a S3 class with the same name (see 2.1.1). Simulation functions in **medfate** need an object of class **soil** to be run.

Function **soil()** adds the following information to the physical soil description:

Symbol	Units	R	Description
$P_{macro,s}$	%	macro	Percentage of macroporosity corresponding to each soil layers
γ_{soil}	$mm \cdot day^{-1}$	Gsoil	The maximum daily evaporation rate from soil (see 5.5)
κ_{soil}		Ksoil	Extinction parameter to regulate the amount of water extracted from each soil layer when simulating evaporation from bare soil (see 5.5)
$\theta_{sat,s}$	$m^3 \cdot m^{-3}$	VG_thetaSat	Volumetric soil moisture at saturation
$\theta_{res,s}$	$m^3 \cdot m^{-3}$	VG_thetaRes	Residual volumetric soil moisture
n_s		VG_n	Parameter n of the Van Genuchten (1980) model
α_s	MPa^{-1}	VG_alpha	Parameter α of the Van Genuchten (1980) model
$K_{sat,s}$	$mmol \cdot s^{-1} \cdot m^{-1} \cdot MPa^{-1}$	Ksat	Saturated soil conductivity
K_{drain}	$mm \cdot d^{-1}$	Kdrain	Hydraulic conductivity for deep drainage.

Macroporosity values are calculated using the equations given in Stolf *et al.* (2011), while γ_{soil} and κ_{soil} are derived from texture. Parameters of the van Genuchten model for each layer are estimated from the physical description of the layer using one of two pedotransfer functions (see help for **soil()**):

1. Using the USDA texture classification and the average texture class parameters given by Carsel & Parrish (1988).
2. Directly from the soil texture, organic matter and bulk density, using the pedotransfer functions in Tóth *et al.* (2015).

Soil layer saturated conductivity ($K_{sat,s}$) is estimated using Saxton & Rawls (2006) or Saxton *et al.* (1986), depending on whether an estimate of organic mater is available. K_{drain} is currently initialized from soil saturated conductivity in the deepest soil layer, but should be defined depending on the permeability of the bedrock material.

Besides defining soil parameters, function `soil()` also initializes *soil state variables*: soil moisture content, soil temperature (although set to a missing value) and water table depth. Users can edit the `soil` object manually (it is actually a `list`), for example to provide specific parameters of the Van Genuchten retention curve calibrated from soil measurements.

2.3.4 Water content and water table depth

The *volume of water* (V_s in mm = $l \cdot m^{-2}$ of ground area) in a soil layer s is calculated from its water potential (Ψ_s) and water retention curve using:

$$V_s(\Psi_s) = d_s \cdot ((100 - P_{rocks,s})/100) \cdot \theta_s(\Psi_s) \quad (2.4)$$

where d_s is the depth of the soil layer (in mm) and $P_{rocks,s}$ is the percentage of rock fragments. The overall volume of water in the soil (accessible by roots) is simply the sum of water content over soil layers:

$$V_{soil} = \sum_{s=1}^S V_s(\Psi_s) \quad (2.5)$$

A number of fixed water volumes (and their corresponding moisture) are important to remember:

- *Water holding capacity* ($V_{fc,s}$, in mm) of the soil layer s is defined as the volume of water at *field capacity*, i.e. at $\Psi_{fc} = -0.033$ MPa, the amount of water held in the soil after gravitational water has drained away:

$$V_{fc,s} = V_s(-0.033) = d_s \cdot ((100 - P_{rocks,s})/100) \cdot \theta_{fc,s} \quad (2.6)$$

where $\theta_{fc,s} = \theta_s(-0.033)$ is the corresponding moisture at field capacity.

- *Water content at saturation* ($V_{sat,s}$, in mm) is calculated analogously, but replacing $\theta_{fc,s}$ by $\theta_{sat,s} = \theta_s(0)$, the moisture at *saturation*, the amount of soil moisture when all easily drained spaces between soil particles (i.e. macropores) are also filled with water.
- *Water content at wilting point* ($V_{wp,s}$, in mm) is calculated replacing $\theta_{fc,s}$ by $\theta_{wp,s} = \theta_s(-1.5)$, the moisture at *wilting point*, i.e. at $\Psi_{fc} = -1.5$ MPa, a conventional amount of soil moisture beyond which plants are assumed have problems extracting water and start to wilt. However, plants can often extract water from much drier soils.
- The *amount of extractable water* (in mm) is the difference between water content at field capacity and the water content at a conventional minimum water potential, which can be set at wilting point or lower values (by default at -5 MPa in `medfate`).

The depth of water table is the depth of saturated soil (in mm). In `medfate`, *water table depth* (Z_{wt} , in mm) equals soil depth (i.e. Z_{soil}) when all soil layers are below field capacity. When some layers are between field capacity and saturation, water table depth is calculated as:

$$Z_{wt} = \sum_s d_s \cdot \min [1, (\theta_{sat,s} - \theta(\Psi_s)) / (\theta_{sat,s} - \theta_{fc,s})] \quad (2.7)$$

2.4 Vegetation description

The representation of vegetation within a target forest stand is done in **medfate** primarily by means of **cohorts of woody plants** (i.e. trees and shrubs). A woody plant cohort represents a set of woody plants that belong to the same species (or functional group) and are more or less of the same *size*. For example, trees in a stand may be grouped by species and diameter class, whereas shrubs may be grouped by species and height. In practice however, plant cohorts are entities that are treated separately when modeling the forest stand. This representation in plant cohorts was chosen so that package functions can be easily applied to **forest inventory data**, for example from national forest surveys. The description of woody plant cohorts includes a measure of *abundance* of the cohort in the stand, which in the case of trees is *density* as stems per hectare and for shrubs is *percent cover* (shrubs are often multi-stemmed). The difference in measurement units for abundance is the main motivation for distinguishing between tree cohorts and shrub cohorts, rather than their size or growth form. Additionally, the description of vegetation in the forest stand may include the cover and mean height of a **herbaceous layer**. The presence or abundance of herbs is not always recorded in forest inventory surveys, but may be relevant vegetation component in open habitats like shrublands.

Note that one limitation of the representation of vegetation in **medfate** is that it is not spatially-explicit (i.e. plants cannot have explicit coordinates within forest stands) and hence spatial (horizontal) interactions between plant cannot be taken into account explicitly (but see 2.4.7).

2.4.1 Structural attributes of woody plant cohorts

A woody plant cohort i is either a *tree cohort* or a *shrub cohort*, and is defined using a set of structural attributes shown in the following table, where columns **spwb**, **growth** and **fordyn** indicate whether attributes are required by simulation functions **spwb()**/**pwb()**, **growth()** and **fordyn()**, respectively:

Symbol	Units	R	Description	spwb	growth	fordyn
SP_i		Species	Species identity	Y	Y	Y
H_i	cm	Height	Average tree or shrub height	Y	Y	Y
CR_i		CR	Crown ratio (i.e. ratio between crown length and plant height)	Y	Y	N
N_i	$\text{ind} \cdot \text{ha}^{-1}$	N	Density of tree individuals	N	Y	Y
DBH_i	cm	DBH	Tree diameter at breast height	N	Y	Y
$Cover_i$	%	Cover	Shrub percent cover	N	Y	Y
LAI_i^{live}	$m^2 \cdot m^{-2}$	LAI_live	(Maximum) leaf area index	Y	Y	N
LAI_i^{dead}	$m^2 \cdot m^{-2}$	LAI_dead	Dead leaf area index	Y	Y	N
LAI_i^ϕ	$m^2 \cdot m^{-2}$	LAI_expanded	expanded leaf area index	Y	Y	N
$Z_{50,i}$	mm	Z50	Depth above which 50% of the fine root mass is located	Y	Y	Y
$Z_{95,i}$	mm	Z95	Depth above which 95% of the fine root mass is located	Y	Y	Y

Height (H) values refer to average height of individuals included in the cohort, and the same for crown ratio (CR) and diameter at breast height (DBH). While plant size (i.e. height or diameter) is relatively easy to tally, other measurements are not usually made in the field. Package **medfate** includes utility functions that provide estimates CR and LAI from forest inventory data (e.g. heights, DBH and density for measured trees), using allometric relationships calibrated for Catalonia, Spain (see chapter 21).

Leaf area index

LAI variables refer to *one-sided leaf area* of plants per surface area of the stand. Leaves standing on branches of woody plants can be alive or dead. We will call *total leaf area index* of a woody plant cohort i (LAI_i^{all})

to the sum of its dead and live (and unfolded) leaf area:

$$LAI_i^{all} = LAI_i^\phi + LAI_i^{dead} \quad (2.8)$$

For simulations not involving growth, winter-deciduous phenology is a prognosed variable, and the current level of leaves unfolded may be lower than the maximum (live) amount of leaves. In simulations involving growth, both LAI^ϕ and LAI^{live} can change, the former due to leaf phenology (unfolding is simulated explicitly and has carbon costs) and the latter due to leaf allocation limits during plant development.

2.4.2 Structural attributes of herbaceous vegetation

The herbaceous layer of the stand is described collectively, without distinguishing among plant cohorts. Two attributes, cover and height, are required:

Symbol	Units	R	Description
H_{herb}	cm	herbHeight	Average height of herbaceous vegetation
$Cover_{herb}$	%	herbCover	Percent cover of herbaceous vegetation
LAI_{herb}	$m^2 \cdot m^{-2}$	LAI_herb	Leaf area index of herbaceous vegetation

Leaf area index of herbaceous layer (LAI_{herb}), is normally estimated from H_{herb} and $Cover_{herb}$, but could be directly prescribed by the user. Rooting depth parameters are not needed for herbaceous vegetation, which is assumed to extract water from the topmost soil layer. Note that neither phenology or growth/senescence processes are modelled in the case of herbaceous vegetation.

2.4.3 Leaf area index of the forest stand

If there are c woody plant cohorts, the leaf area index corresponding to living leaves, unfolded leaves and dead leaves can be aggregated across cohorts using:

$$LAI_{woody}^{live} = \sum_{i=1}^c LAI_i^{live} \quad (2.9)$$

$$LAI_{woody}^\phi = \sum_{i=1}^c LAI_i^\phi \quad (2.10)$$

$$LAI_{woody}^{dead} = \sum_{i=1}^c LAI_i^{dead} \quad (2.11)$$

We can also estimate the overall leaf area index of woody plant cohorts (LAI_{woody}^{all}):

$$LAI_{woody}^{all} = \sum_{i=1}^c LAI_i^{woody} = LAI_{woody}^\phi + LAI_{woody}^{dead} \quad (2.12)$$

Finally, we can also estimate the overall leaf area index of the whole stand (LAI_{stand}^{all}), which also takes into account the herbaceous layer (LAI_{herb}):

$$LAI_{stand}^{all} = LAI_{woody}^{all} + LAI_{herb} = LAI_{woody}^\phi + LAI_{woody}^{dead} + LAI_{herb} \quad (2.13)$$

2.4.4 Forest plot objects

While the previous section described the structural forest variables required to run simulations in **medfate**, forest plot data input in the package is easier if one uses a format that follows closely forest inventory plot descriptions. In this format, each forest plot is represented in an object of class **forest**, a list that contains several elements. Here we use an example forest stand provided with the package:

```
data(exampleforestMED)
exampleforestMED

## $treeData
##           Species   N   DBH Height Z50  Z95
## 1 Pinus halepensis 168 37.55     800 100  600
## 2      Quercus ilex 384 14.60     660 300 1000
##
## $shrubData
##           Species Cover Height Z50  Z95
## 1 Quercus coccifera  3.75     80 200 1000
##
## $herbCover
## [1] 10
##
## $herbHeight
## [1] 20
##
## attr(),"class")
## [1] "forest" "list"
```

The most important items of **forest** are the two data frames describing woody vegetation, **treeData** (for trees) and **shrubData** for shrubs, but the herbaceous layer can also be described using **herbCover** and **herbHeight**. With the aim to help users in the task of constructing **forest** objects, package **medfateutils** includes functions to map user data in into tables **treeData** and **shrubData**.

forest objects are a convenient format to start calculations with **medfate**, because there are many static functions that take forest objects as input. For example, a **summary.forest()** function provides the basal area, density, cover and leaf area index of the forest stand, and its different components:

```
summary(exampleforestMED, SpParamsMED)
```

```
## Tree BA (m2/ha): 25.0333016 adult trees: 25.0333016 saplings: 0
## Density (ind/ha) adult trees: 552 saplings: 0 shrubs (estimated): 749.4923076
## Cover (%) adult trees: 100 saplings: 0 shrubs: 3.75 herbs: 10
## LAI (m2/m2) total: 1.7585845 adult trees: 1.5543216 saplings: 0 shrubs: 0.030626 herbs: 0.1736363
## Fuel loading (kg/m2) total: 0.5588728 adult trees: 0.5255004 saplings: 0 shrubs: 0.0140795 herbs
## PAR ground (%): 40.0075402 SWR ground (%): 50.7329667
```

2.4.5 Vertical leaf distribution

The vegetation input structures allow the package to determine the vertical distribution of leaves in the stand. The leaf area of any woody plant cohort is assumed to be distributed vertically following a **truncated Gaussian function** whose standardized values -1.5 and 1.5 correspond to crown base height ($H_{crown,i}$; in cm) and total plant height (H_i), respectively. Crown base height is defined as the height corresponding to

the first living branch. It is calculated from the crown ratio of the cohort (CR_i ; a proportion between 0 and 1), which in turn can be estimated as explained in (21.11).

Simulation models in **medfate** divide the vertical dimension into vertical layers (by default are 100 cm width, but see control parameter `verticalLayerSize`). Let us define $LAI_{i,j}^{all} = LAI_{i,j}^{\phi} + LAI_{i,j}^{dead}$ as the leaf area index of cohort i in layer j including both functional leaves and dead leaves standing on branches. The truncated Gaussian distribution defines the $LAI_{i,j}^{\phi}$ and $LAI_{i,j}^{dead}$ for all plant cohorts and vertical layers.

Dividing the leaf area of a given layer by its width, one obtains the *leaf area density* (LAD in $m^2 \cdot m^{-3}$). Figure 2.3 illustrates the leaf area density profile (see function `vprofile_leafAreaDensity()`) corresponding to the forest stand described in the example `forest` object that we showed above.

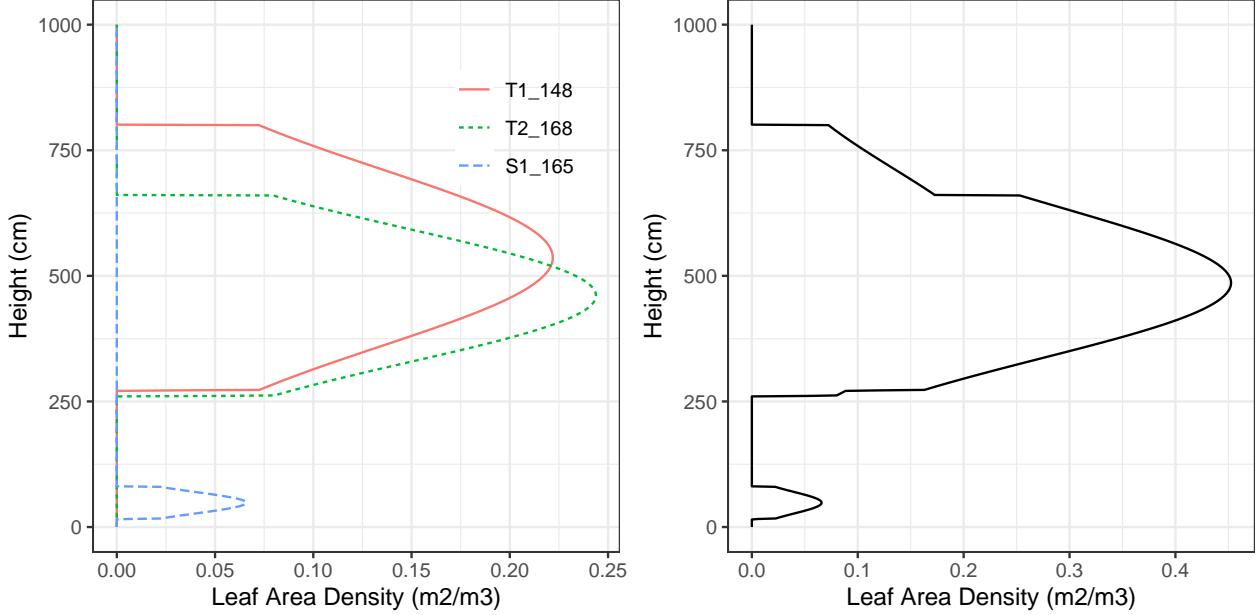


Figure 2.3: Leaf area density distribution of woody vegetation in a forest stand. The left panel shows the distribution for each plant cohort separately, whereas the right panel shows the overall density distribution.

The leaf area density profile determines the light extinction rates through the forest canopy. The same truncated Gaussian distribution is used to distribute leaf and small branch biomass along the vertical dimension.

When present, we can include herbaceous vegetation (i.e. LAI_{herb}) in the leaf area density profile. Its vertical distribution is also assumed to follow a truncated Gaussian function:

2.4.6 Vertical root distribution

Roots can be distinguished functionally into two different organs: **coarse roots** are responsible for mechanical support and transport of water and nutrients towards the stem, whereas **fine roots** are responsible for absorbing water and nutrients.

Analogously to the aboveground vegetation parameters, inputs describing depths corresponding to 50% and 95% of fine roots ($Z_{50,i}$ and $Z_{95,i}$) are used to completely specify the distribution of roots across soil layers. The root system of each plant cohort i is described using $FRP_{i,s}$, the proportion of fine roots (with respect to the whole plant) in each soil layer s . $FRP_{i,s}$ values are normally defined using the linear dose response model (Schenk & Jackson 2002; Collins & Bras 2007):

$$FRP_{i,s} = Y_i(Z_s) - Y_i(Z_{s-1}) \quad (2.14)$$

$$Y_i(z) = \frac{1}{1 + (z/Z_{50,i})^{c_i}} \quad (2.15)$$

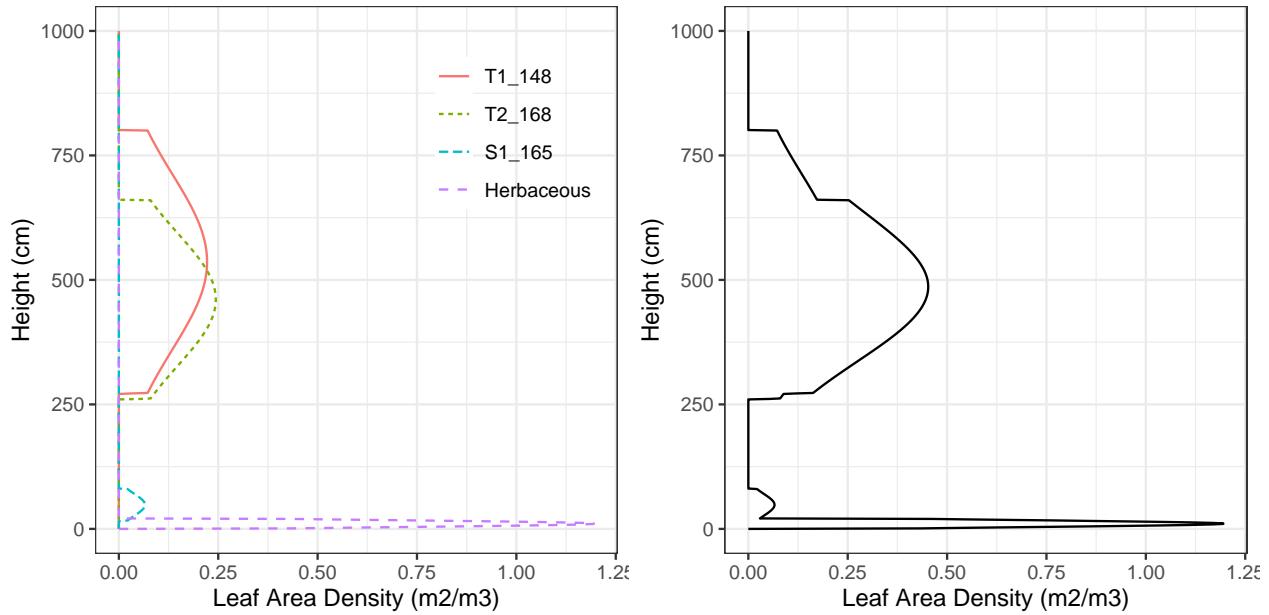


Figure 2.4: Leaf area density distribution in a forest stand, including the herbaceous layer. The left panel shows the distribution for each plant cohort separately, whereas the right panel shows the overall density distribution.

where $Y_i(z)$ is the cumulative fraction of fine root mass located between surface and depth z ; $Z_{50,i}$ is the depth (in mm) above which 50% of the root mass is located; and c_i is a shape parameter related to $Z_{50,i}$ and $Z_{95,i}$ as $c_i = 2.94/\ln(Z_{50,i}/Z_{95,i})$.

The linear dose response model is quite flexible with respect to fine root distribution (see function `root_ldrDistribution()`). The following figure shows the fine root distribution profile of the same cohorts as in fig. 2.3 (see `vprofile_rootDistribution()`):

The actual input to simulation functions `spwb()`, `growth()` and alike, describes fine root distribution using the matrix of $FRP_{i,s}$ values - not $Z_{50,i}$ and $Z_{95,i}$ -, which means that the proportions of fine roots can be modified manually or be defined by another model. In fact, `medfate` also allows calculating $FRP_{i,s}$ values assuming a conic distribution of fine roots (see `root_conicDistribution()`).

2.4.7 Water pools and root horizontal distribution

By default, models in `medfate` assume that soil moisture under all plant cohorts is the same (i.e. water sources corresponding to vertical soil layers are shared among cohorts). Therefore, it neglects spatial variation in soil moisture, against moisture variation patterns observed in mixed stands (Schume *et al.* 2004). However, variations in soil moisture beneath plant cohorts (and, implicitly, horizontal variation of soil moisture) can also be simulated if required by the user (see parameter `rhizosphereOverlap` in 2.6). This involves considering that a given plant cohort will perform water uptake from the **water pool** surrounding its roots, whereas it may not have access to the water beneath other plants. However, there can exist some degree of horizontal overlap between water pools exploited by different plants.

Considering water pools involves partitioning the stand area into fractions corresponding to the abundance of each plant cohort. More specifically, the model defines as many water pools as plant cohorts, with proportions defined by their LAI values:

$$f_{pool,i} = \frac{LAI_i^{live}}{LAI_{stand}^{live}} \quad (2.16)$$

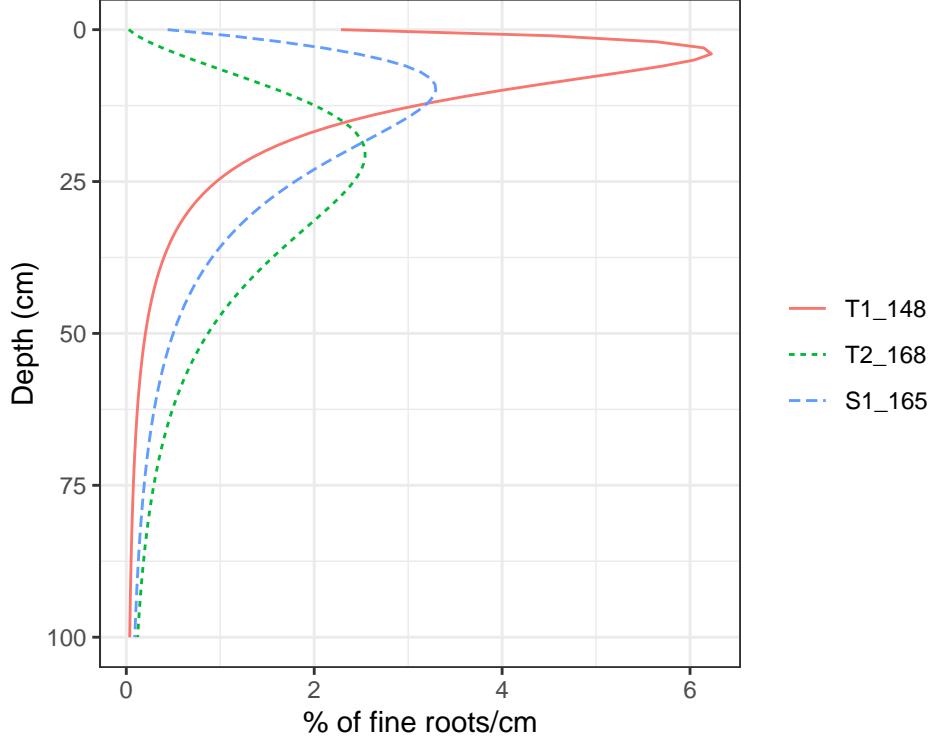


Figure 2.5: Examples of root density profile according to the linear dose response model.

Models assume that the rhizosphere of each plant cohort occupies its own water pool but may extend into the water pools under other plant cohorts. In other words, the root systems of different cohorts may overlap horizontally. Moreover, the horizontal overlap of root systems will vary across soil layers. A given plant cohort i will have its roots in layer s partitioned among different water pools. We thus need to define $fr_{i,s,j}$, the (horizontal) proportion of fine roots of cohort i in layer s of the water pool j , with the restriction that:

$$\sum_j fr_{i,s,j} = 1 \quad \forall i, s \quad (2.17)$$

A simplifying assumption is to force a complete independence of water pools, which is equal to say that $fr_{i,s,j} = 1$ for $i = j$ and $fr_{i,s,j} = 0$ when $i \neq j$ (this is the simplification adopted when using `rhizosphereOverlap = "none"`). In the general case of partial overlap (`rhizosphereOverlap = "partial"`), it is important to realize that proper estimation of $fr_{i,s,j}$ is challenging when we do not have explicit plant coordinates, root lateral widths, etc. At this point, let us assume we have a reasonable estimate of Vol_i , the **soil volume explored by coarse roots** of an individual of cohort i . Assuming that the proportion of fine roots in each layer is proportional to the proportion of total soil volume explored by coarse roots that corresponds to the same layer we have that $Vol_{i,s}$, the soil volume explored by coarse roots in layer s is:

$$Vol_{i,s} = FRP_{i,s} \cdot Vol_i \quad (2.18)$$

In `medfate`, coarse roots are represented by *axial* and *radial* components. The length of axial component of a given layer ($L_{axial,i,s}$) is simply equivalent to its bottom depth, whereas the radial component ($L_{radial,i,s}$) is unknown. However, we can assume that the soil volume explored by coarse roots corresponds to a cylinder:

$$Vol_{i,s} = \pi \cdot L_{radial,i,s}^2 \cdot L_{axial,i,s} \quad (2.19)$$

and substituting the soil volume $Vol_{i,s}$ in the equation above we can isolate $L_{radial,i,s}$ (see function `root_coarseRootLengthsFromVolume()`). Once we have the radial component we can estimate the area

(m^2) covered by coarse roots of an individual of the cohort in a given layer s :

$$Area_{i,s} = \pi \cdot L_{radial,i,s}^2 \quad (2.20)$$

We can then compare $Area_{i,s}$ to the area of the water pool for an individual of cohort i , which in m^2 is:

$$AreaPool_i = 10000 \cdot f_{pool,i}/N_i \quad (2.21)$$

If $Area_{i,s} \leq AreaPool_i$, then the roots of the individual do not exceed the area of the pool, in other words $f_{r,i,s,i} = 1$ and $f_{r,i,s,j} = 0$ for all $j \neq i$. If $Area_{i,s} > AreaPool_i$ then the excess should be counted as overlap with the other pools. Assuming that plants are randomly distributed (i.e. no clumping) the probability of finding the pool of a cohort is equal to the proportion of its water pool (see function `root_horizontalProportions()`). All these calculations were done assuming Vol_i were known. Estimation of Vol_i depends on the complexity of the water balance model and is explained in A.4.6.

2.4.8 Functional traits and vegetation initialization

Besides the physical representation of vegetation, forest ecosystem models require information regarding plant functional traits, because this influences the outcome of physical, physiological and hydrological processes. In the following chapters, we will indicate for each simulated process its required plant functional parameters. A description of all parameters is provided in data frame `SpParamsDefinition` included in the package.

Normally, functional traits are described at the species level because infra-specific parameters are hard to get. Even at the species level, many functional (e.g. physiological) traits are hard to obtain, so in `medfate` we provide a default species parameter table for woody species found in Spain, with many of them occurring elsewhere in the Mediterranean Basin (`SpParamsMED`). User can modify this data frame to account for intra-specific trait variation (see function `modifySpParams()`).

The package provides functions `forest2spwbInput()` and `forest2growthInput()` that prepare the `forest` vegetation input for simulation functions `spwb()` and `growth()`, respectively (see 2.1.1). This initialization mostly consists in compiling the necessary functional traits from the species parameter table, following the SP_i attribute of woody plant cohorts. Some new model parameters are estimated from the initial ones, while taking into account the structural attributes of cohorts (an example of these derived quantities is stem conductance, which estimated from stem xylem conductivity, Huber value and plant height). Users can take the output of these initialization functions and replace parameter values for specific plant cohorts (see function `modifyCohortParams()`).

2.4.9 Update of structural variables during simulations

Vegetation characteristics stay constant during simulations using functions `spwb()` or `pwb()`, although the actual (unfolded) leaf area and dead leaf area may vary depending on leaf phenology.

In contrast, growth simulation requires updating the structure of vegetation, i.e. plant heights, tree diameters, tree density and shrub cover. Function `growth()` can modify any of the vegetation attributes. Finally, function `fordyn()` modifies not only structural variables of the initial cohorts but also involves the removal of dead (or cut) woody cohorts and recruitment of new woody cohorts during simulations.

2.5 Metereological input

Weather input data must include variables calculated at the **daily** scale. Weather data should be arranged in a data frame with days in **rows** and variables in **columns**. Dates are needed in `medfate` to estimate parameters like solar declination or the day of the year (*DOY*). Importantly, *dates* should be included as row names (in format *year-month-day*) of the input meteorology data frame.

The following table indicates the symbols, units, definitions and the variable name in R of all weather variables. Some of them are optional, as indicated in the right-most column.

Symbol	Units	R param	Description	Optional
T_{min}	°C	MinTemperature	Minimum temperature	No
T_{max}	°C	MaxTemperature	Maximum temperature	No
RH_{min}	%	MinRelativeHumidity	Minimum relative humidity	No
RH_{max}	%	MaxRelativeHumidity	Maximum relative humidity	No
P	$L \cdot m^{-2} = mm$	Precipitation	Precipitation (either rainfall or snow).	No
Rad	$MJ \cdot m^{-2}$	Radiation	Solar radiation after accounting for clouds	No
u	$m \cdot s^{-1}$	WindSpeed	Wind speed	Yes
C_{atm}	ppm	CO2	Atmospheric (above-canopy) CO_2 concentration	Yes
P_{atm}	kPa	Patm	Surface atmospheric pressure	Yes
P_{fire}	[0-1]	FireProbability	Probability of wildfire occurrence	Yes

While the weather variables listed in the table above are input requirements, **medfate** derives other atmospheric variables from them, using also topographic information (see utility functions of the **meteoland** reference manual):

Symbol	Units	R param	Description
ρ_{air}	$kg \cdot m^{-3}$		Air density
e_{atm}	kPa		Atmospheric water vapor pressure
T_{mean}	°C	MeanTemperature	Mean daily temperature
PET	$L \cdot m^{-2} = mm$	PET	Potential evapotranspiration, calculated using Penman's (1948) equation

Missing wind speed (u) and CO_2 values will be replaced by defaults defined in control parameters. As an alternative to providing daily atmospheric CO_2 concentration values, simulation functions allow the user to provide CO_2 concentration values in **annual** time steps using a separate vector. When missing, atmospheric pressure (P_{atm}) is also derived from elevation using an utility function available in **meteoland**.

Since it allows producing data frames with appropriate variable units and row/column names, we recommend meteorological input to be generated using package **meteoland** (De Cáceres *et al.* 2018), but other sources are possible.

2.6 Simulation control

Simulation control parameters are a list of global parameter values, initialized using function **defaultControl()**, that the user can modify to change the general behavior of simulation functions. Here is the set of global control parameters currently accepted in **medfate**:

```
names(defaultControl())
## [1] "fillMissingRootParams"          "fillMissingSpParams"
## [3] "verbose"                      "subdailyResults"
## [5] "standResults"                  "soilResults"
## [7] "plantResults"                  "leafResults"
## [9] "temperatureResults"            "fireHazardResults"
```

```

## [11] "fireHazardStandardWind"
## [13] "transpirationMode"
## [15] "defaultWindSpeed"
## [17] "snowpack"
## [19] "rockyLayerDrainage"
## [21] "rhizosphereOverlap"
## [23] "verticalLayerSize"
## [25] "cavitationRefill"
## [27] "ndailysteps"
## [29] "capacitance"
## [31] "multiLayerBalance"
## [33] "klatstem"
## [35] "numericParams"
## [37] "fracRootResistance"
## [39] "thermalCapacityLAI"
## [41] "refillMaximumRate"
## [43] "allowDessication"
## [45] "sinkLimitation"
## [47] "herbDynamics"
## [49] "phloemConductanceFactor"
## [51] "equilibriumOsmoticConcentration"
## [53] "constructionCosts"
## [55] "maximumRelativeGrowthRates"
## [57] "mortalityBaselineRate"
## [59] "mortalityRWCThreshold"
## [61] "recrTreeDensity"
## [63] "ingrowthTreeDensity"
## [65] "allowRecruitment"
## [67] "recruitmentMode"
## [69] "minimumTreeCohortDensity"
## [71] "dynamicallyMergeCohorts"
## [73] "seedProductionTreeHeight"
## [75] "probRecr"
## [77] "minMoistureRecr"
## [79] "recrTreeHeight"
## [81] "recrShrubHeight"
## [83] "recrShrubZ50"
## [85] "recrShrubZ95"

## [11] "fireHazardStandardDFMC"
## [13] "soilFunctions"
## [15] "defaultCO2"
## [17] "leafPhenology"
## [19] "unlimitedSoilWater"
## [21] "unfoldingDD"
## [23] "windMeasurementHeight"
## [25] "hydraulicRedistributionFraction"
## [27] "nsubsteps"
## [29] "taper"
## [31] "maximumStemConductance"
## [33] "klatleaf"
## [35] "fracLeafResistance"
## [37] "averageFracRhizosphereResistance"
## [39] "boundaryLayerSize"
## [41] "subdailyCarbonBalance"
## [43] "allowStarvation"
## [45] "shrubDynamics"
## [47] "allocationStrategy"
## [49] "nonSugarConcentration"
## [51] "minimumRelativeStarchForGrowth"
## [53] "senescenceRates"
## [55] "mortalityMode"
## [57] "mortalityRelativeSugarThreshold"
## [59] "recrTreeDBH"
## [61] "ingrowthTreeDBH"
## [63] "allowSeedBankDynamics"
## [65] "allowResprouting"
## [67] "removeEmptyCohorts"
## [69] "minimumShrubCohortCover"
## [71] "seedRain"
## [73] "seedProductionShrubHeight"
## [75] "minTempRecr"
## [77] "minFPARRecr"
## [79] "recrShrubCover"
## [81] "recrTreeZ50"
## [83] "recrTreeZ95"

```

Control parameters are employed when initializing vegetation inputs and state variables using functions `forest2spwbInput()` and `forest2growthInput()`, since they are needed also to make decisions at the point of initialization. Control parameters are also stored in the result of these functions, so the user does not need to specify again control parameters when calling simulation functions such as `spwb()` or `growth()`. Function `fordyn()` is different than the other simulation functions, because the list control parameters is directly passed to the function (see 2.1.1).

Even if the list of control parameters is long, not all control parameters are relevant to all simulation functions (this will be indicated in the following chapters), and most of them should not be altered from their default values. However, there are a set of parameters that are worth learning:

- `verbose` [=TRUE]: Whether extra console output is desired during simulations.
- `fillMissingRootParams` [= TRUE]: Boolean flag to indicate that initialization functions should provide estimates for Z50 and Z95 if these are lacking in the forest data. Note that if `fillMissingRootParams` is set to FALSE then simulations may fail if the user does not provide values for Z50 and Z95 in tree or shrub data.

- `fillMissingSpParams [=TRUE]`: Boolean flag to indicate that initialization functions should provide estimates for functional parameters if these are lacking in the species parameter table `SpParams`. Note that if `fillMissingSpParams` is set to FALSE then simulations may fail if the user does not provide values for required parameters.
- `transpirationMode [=“Granier”]` is an important control parameter, because it allows the user to switch between the *basic* (i.e. “`Granier`”) and *advanced* (i.e., either “`Sperry`” or “`Cochard`”) water balance models, corresponding to chapters 3 and 7, respectively.
- `rhizosphereOverlap [=“total”]` is also a relevant parameter, since changing its default value allows considering that plants extract soil water from partially or totally unrelated water pools, as explained in 2.4.7.

Part II

Basic water balance modelling

Chapter 3

Basic water balance model

This chapter provides an overview of the basic soil and plant water balance model included in **medfate**. The model allows simulating vertical water fluxes in a forest stand, and performs soil and plant water balance on a daily step basis for the period corresponding to input weather data.

The model is run using function `spwb()`, for a set of days, or function `spwb_day()`, for a single day. Alternatively, one can use function `pwb()` to evaluate plant transpiration and drought stress resulting from given weather and soil moisture dynamics. The basic water balance model corresponds to the control variable `transpirationMode = "Granier"`.

The description of the model presented here conforms to a large extent to the description given in De Cáceres *et al.* (2015), but includes several modifications that have been implemented since the publication of this scientific article. The information provided in this chapter should be enough to understand what the model does and when it can be useful, but the reading the following ones should provide a more detailed understanding on how the different processes are formulated.

3.1 Design principles

The water balance model considers only the vertical spatial dimension of the stand and not the horizontal distribution of plants within it. In other words, the model is not spatially explicit (i.e., plants do not have horizontal coordinates nor interact for space explicitly). Woody vegetation in the stand is divided into groups, here referred to as *plant cohorts* of different species, height and leaf area index (*LAI*). Height and *LAI* values determine competition for light. *LAI* values also drive competition for soil water, along with fine root distribution. The model can also consider a layer herbaceous vegetation, which also has a height value and *LAI*, but where plant cohorts are not distinguished and with fine roots confined to the topmost layer.

The soil water balance follows the design principles of SIERRA (Mouillot *et al.* 2001; Ruffault *et al.* 2013, 2014) and BILJOU (Granier *et al.* 1999, 2007), although some soil hydrology features are taken from other models (Kergoat 1998). Hydrological processes include water interception loss (Gash *et al.* 1995), plant transpiration, evaporation from soil (Ritchie 1972) and the partition between infiltration and runoff (Boughton 1989). Infiltrated water is either retained by capillarity or percolates through soil layers, depending on macro-porosity, and water exceeding soil water holding capacity is lost beyond the rooting zone via deep drainage. Snow pack dynamics can also be simulated. Potential evapo-transpiration (*PET*) is a required input and the model determines maximum herbaceous and maximum canopy transpiration (Tr_{max}) using an empirical relationship between the *LAI* of the stand and the ratio Tr_{max}/PET (Granier *et al.* 1999). Actual plant transpiration is estimated using a function that depends not only on Tr_{max} but also on current soil moisture levels, cohort-specific drought resistance, fine root distribution and the degree of shading of the target plant cohort. When soil water deficit progresses transpiration rates are consequently reduced, but

minimum transpiration rates of woody vegetation are not zero, because of the role of cuticular transpiration and imperfectly closed stomata (Duursma *et al.* 2018). Woody plant water status (i.e. plant water potential and, hence, plant water content) follows variations of soil moisture in the soil layers where fine roots are located.

By default, the model assumes that soil moisture under all plant cohorts is the same (i.e. water sources corresponding to vertical soil layers are shared among cohorts). However, 3D variations in soil moisture beneath plant cohorts can be simulated in spatially-explicit forest ecosystem models (Manoli *et al.* 2017; Rötzer *et al.* 2017). The water balance model does not simulate horizontal spatial processes explicitly, but allows considering more or less independent water pools for plant cohorts. If partial or completely independent water pools are considered, hydrological processes are replicated for the fraction of soil corresponding to each woody cohort. Transpiration of each plant cohort depends on the soil moisture beneath itself and, depending on the degree of rhizosphere overlap, on the soil moisture beneath other plant cohorts.

3.2 State variables

The following are state variables in the model under all simulations:

- Cumulative degree days (a) to budburst, (b) to complete unfolding or (c) to senescence ($S_{eco,d}$, $S_{unf,d}$ or $S_{sen,d}$; all in $^{\circ}\text{C}$), are tracked by the model to determine leaf phenological status (see 4.1).
- Soil moisture content dynamics on each layer s are tracked daily using $W_s = \theta_s(\Psi)/\theta_{fc,s}$, the *proportion of soil moisture in relation to field capacity*, where moisture at field capacity, $\theta_{fc,s}$, is assumed to correspond to $\Psi_{fc} = -0.033$ MPa. Note that W_s values larger than one are possible if the soil is between field capacity and saturation (which can happen if deep drainage is not allowed).
- Plant water potential ($\Psi_{plant,i}$) of each plant cohort i are also tracked daily and follows the soil water potential of layers (or water pools) where fine roots are located.

Additional state variables depend on the activation of specific control flags:

- *If snow accumulation/melting is considered*, the model tracks S_{snow} , the snow water equivalent (in mm) of the snow pack storage over the surface.
- *If stem cavitation is not completely reversible*, the model tracks PLC_i , the proportion of stem conductivity lost by plants of cohort i .
- *If the degree of rhizosphere overlap between plant cohorts is not total* (i.e. if `rhizosphereOverlap` = "none" or `rhizosphereOverlap` = "partial"), the model also tracks $W_{i,s}$, the proportion of soil moisture in relation to field capacity for layer s within the areal fraction of the stand covered by the woody cohort i .

3.3 Water balance

Daily variations in soil water content (ΔV_{soil} in mm) can be summarized as:

$$\Delta V_{soil} = Pr + Sm - In - Ru - Dd - Es - Tr_{herb} - Tr_{woody} \quad (3.1)$$

where Pr is precipitation as rainfall, Sm is water from melting the snow pack (if considered), In is the interception loss (i.e., water evaporated after being intercepted by the canopy), Ru is surface runoff, Dd is deep drainage (i.e. water percolated to layers beyond soil depth), Es is evaporation from soil, Tr_{herb} is the transpiration made by herbaceous plants and Tr_{woody} is the transpiration made by woody plants.

While the above describes the default water balance equations, variations can occur depending on specific control flags:

- a. If snow dynamics are considered, the water balance of the snow pack (ΔS_{snow} in mm of snow water equivalents) is defined as:

$$\Delta S_{snow} = Ps - Sm \quad (3.2)$$

where Ps is precipitation as snowfall and Sm is snow melt. Evaporation from bare soil cannot occur if there is a snow pack over the soil (i.e., if $S_{snow} > 0$ then $Es = 0$).

- b. If plant water pools are considered, the water balance equation applies not only to the soil of the overall stand but also to the soil beneath each woody plant cohort. The fraction of stand area corresponding to the water pool of each cohort is used to keep the water balance at the two scales aligned. Moreover, the water balances of soils beneath the different plant cohort are more or less correlated depending on the degree of rhizosphere overlap.

3.4 Process scheduling

For every day to be simulated, the model performs the following steps:

1. Update leaf area values according to the phenology of species and recalculate radiation extinction (sections 4.1 and 4.2).
2. If snow dynamics are included, increase snow pack from snow precipitation (Ps) and decrease it following snow melt (Sm) (section 5.2).
3. Determine rainfall interception loss (In) and net rainfall (Pr_{net}) (section 5.3)
4. Increase soil water content due to net rainfall (Pr_{net}), surface runoff (Ro) and snow melt (Sm), after accounting for surface runoff (Ru) and deep drainage (Dd) (section 5.4).
5. Decrease soil water content of the topmost soil layer due to bare soil evaporation (Es), if snow is not present (section 5.5), and herbaceous transpiration (Tr_{herb} ; section 5.6).
6. Determine woody plant transpiration (Tr_{woody}) and photosynthesis (chapter 6).
7. Update plant water potential (Ψ_{plant}) and plant drought stress for each plant cohort and decrease soil water content due to woody plant transpiration.

Details of the formulation of each of these processes are provided within the referred chapters and sections. If plant water pools are considered, steps 4-5 are performed for each plant water pool separately, step 6 involves extracting water from different plant water pools depending on the proportion of fine roots of each cohort in each pool (see 6.1.5).

3.5 Inputs and outputs

3.5.1 Soil, vegetation and meteorology

Soil

Soil input requirements are fully described in section 2.3.

Vegetation

Vegetation input requirements are fully described in section 2.4. Trees and shrubs do not need to be characterized with different variables in soil water balance calculations, since both kinds of woody cohorts have a LAI value. In most cases, users only need to estimate the leaf area index corresponding to live leaves, i.e. LAI^{live} , because normally at the starting point all leaves are expanded (i.e. $LAI^\phi = LAI^{live}$) and one can assume no dead leaves in the canopy (i.e., $LAI^{dead} = 0$). Vegetation characteristics stay constant during simulations using function `spwb()`, although the actual expanded leaf area (LAI^ϕ) and dead leaf area (LAI^{dead}) may vary if the species is winter deciduous.

Meteorology

The minimum weather variables required to run the model are min/max temperatures (T_{min} and T_{max}), min/max relative humidity (RH_{min} and RH_{max}), precipitation (P) and solar radiation (Rad). Wind speed (u) is also needed, but the user may use missing values if not available (a default value will be used in this case). Wind speed is assumed to have been measured at a specific height above the canopy (by default at 2 m). Atmospheric CO_2 concentration (C_{atm}) may also be specified, but if missing a default constant value is assumed, which is taken from the control parameters. Definitions and units of these variables were given in section 2.5.

3.5.2 Vegetation functional parameters

The following sets of functional parameters should be supplied for each woody plant cohort. In practice, they are normally estimated from the species-specific parameter table and imputation of missing values (see A). Functional attributes are filled for each cohort i by function `forest2spwbInput()` from species identity (i.e. SP_i). However, different parameters can be specified for different cohorts of the same species if desired (see section 2.4.8).

A first set of functional parameters refers to leaf phenology (`paramsPhenology`):

Symbol	Units	R	Description
		<code>PhenoType</code>	Phenology type (oneflush-evergreen, progressive-evergreen, winter-deciduous, winter-semideciduous)
LD	years	<code>LeafDuration</code>	Leaf duration of leaves (in years).
$t_{0,eco}$	days	<code>t0gdd</code>	Date to start the accumulation of degree days.
S_{eco}^*	°C	<code>Sgdd</code>	Degree days corresponding to leaf budburst (see section 4.1.2).
T_{eco}	°C	<code>Tbgdd</code>	Base temperature for the calculation of degree days to leaf budburst (see section 4.1.2).
S_{sen}^*	°C	<code>Ssen</code>	Degree days corresponding to leaf senescence (see section 4.1.3).
Ph_{sen}	hours	<code>Phsen</code>	Photoperiod corresponding to start counting senescence degree-days (see section 4.1.3).
T_{sen}	°C	<code>Tbsen</code>	Base temperature for the calculation of degree days to leaf senescence (see section 4.1.3).
x_{sen}	{0,1,2}	<code>xsen</code>	Discrete values, to allow for any absent/proportional/more than proportional effects of temperature on senescence (see section 4.1.3).
y_{sen}	{0,1,2}	<code>ysen</code>	Discrete values, to allow for any absent/proportional/more than proportional effects of photoperiod on senescence (see section 4.1.3).

A second set of parameters relate to light extinction and water interception (`paramsInterception`):

Symbol	Units	R	Description
k_{PAR}		<code>kPAR</code>	PAR extinction coefficient (see section 4.2).
s_{water}	$mm\ H_2O \cdot LAI^{-1}$		Crown water storage capacity (i.e. depth of water that can be retained by leaves and branches) per LAI unit (see section 5.3).

A third set includes parameters related to plant anatomic and morphological attributes (`paramsAnatomy`):

Symbol	Units	R param	Description
$1/H_v$	$m^2 \cdot m^{-2}$	<code>A12As</code>	Ratio of leaf area to sapwood area
RLR	$m^2 \cdot m^{-2}$	<code>Ar2Al</code>	Fine root area to leaf area ratio
SLA	$m^2 \cdot kg^{-1}$	<code>SLA</code>	Specific leaf area
ρ_{leaf}	$g \cdot cm^{-3}$	<code>LeafDensity</code>	Leaf tissue density

Symbol	Units	R param	Description
ρ_{wood}	$g \cdot cm^{-3}$	WoodDensity	Wood tissue density
$\rho_{fineroot}$	$g \cdot cm^{-3}$	FineRootDensity	Fine root tissue density
SRL	$cm \cdot g^{-1}$	SRL	Specific root length
RLD	$cm \cdot cm^{-3}$	RLD	Fine root length density (i.e. density of root length per soil volume)
$r_{6.35}$		r635	Ratio between the weight of leaves plus branches and the weight of leaves alone for branches of 6.35 mm

A fourth set of parameters are related to transpiration and photosynthesis (`paramsTranspiration`):

Symbol	Units	R	Description
g_{swmin}	$mol H_2O \cdot s^{-1} \cdot m^{-2}$	Gwmin	Minimum stomatal conductance to water vapour
$T_{max,LAI}$		Tmax_LAI	Empirical coefficient relating LAI with the ratio of maximum transpiration over potential evapotranspiration (see section 6.1.1).
$T_{max,sqLAI}$		Tmax_sqLAI	Empirical coefficient relating squared LAI with the ratio of maximum transpiration over potential evapotranspiration (see section 6.1.1).
$\Psi_{extract}$	MPa	Psi_Extract	Water potential at which plant transpiration is 50% of its maximum (see section 6.1.2).
$c_{extract}$		Exp_Extract	Parameter of the Weibull function regulating transpiration reduction (see section 6.1.2).
c_{stem}, d_{stem}	(unitless), MPa	VCstem_d	Parameters of the vulnerability curve for stem xylem
WUE_{max}	$g C \cdot mm^{-1}$	WUE	Water use efficiency at VPD = 1kPa and without light or CO ₂ limitations (see section 6.1.6).
WUE_{PAR}		WUE_par	Coefficient describing the progressive decay of WUE with lower light levels (see section 6.1.6).
WUE_{CO2}		WUE_co2	Coefficient for WUE dependency on atmospheric CO ₂ concentration (see section 6.1.6).
WUE_{VPD}		WUE_vpd	Coefficient for WUE dependency on vapor pressure deficit (see section 6.1.6).

A fifth (final) set of parameters are related to water storage and water relations in plant tissues (`paramsWaterStorage`):

Symbol	Units	R	Description
$LFMC_{max}$	%	maxFMC	Maximum live fuel moisture content, corresponding to fine fuels (< 6.35 mm twigs and leaves).
ϵ_{leaf}	MPa	LeafEPS	Modulus of elasticity of leaves
ϵ_{stem}	MPa	StemEPS	Modulus of elasticity of symplastic xylem tissue
$\pi_{0,leaf}$	MPa	LeafPI0	Osmotic potential at full turgor of leaves
$\pi_{0,stem}$	MPa	StemPI0	Osmotic potential at full turgor of symplastic xylem tissue
$f_{apo,leaf}$	[0-1]	LeafAF	Apoplastic fraction in leaf tissues
$f_{apo,stem}$	[0-1]	StemAF	Apoplastic fraction in stem tissues
V_{leaf}	$l \cdot m^{-2}$	Vleaf	Leaf water capacity per leaf area unit
$V_{sapwood}$	$l \cdot m^{-2}$	Vsapwood	Sapwood water capacity per leaf area unit

3.5.3 Control parameters

Control parameters modulate the overall behaviour of water balance simulations (see section 2.6). Most importantly, `transpirationMode` defines the transpiration model, which in turn defines the complexity of the water balance model. If `transpirationMode = "Granier"` (the default value) then the basic water balance model is used.

The other relevant control parameters for the basic water balance model are:

- `soilFunctions [= "SX"]`: Soil water retention curve and conductivity functions, either 'SX' (for Saxton) or 'VG' (for Van Genuchten).
- `defaultWindSpeed [= 2.5]`: Default value for wind speed (in $m \cdot s^{-1}$) when this is missing (only used for leaf fall, see section 4.1).
- `defaultCO2 [= 386]`: Default atmospheric (abovecanopy) CO_2 concentration (in micromol $CO_2 \cdot mol^{-1} = ppm$). This value will be used whenever CO_2 concentration is not specified in the weather input.
- `snowpack [= TRUE]`: Whether dynamics of snow pack are included (see section 5.2).
- `leafPhenology [= TRUE]`: Whether leaf phenology is simulated (see section 4.1). If FALSE then all species are assumed to be evergreen.
- `rockyLayerDrainage [= TRUE]`: Whether water exceeding the field capacity of the deepest layer can be drained via macropore flow towards a non-reachable compartment, only for layers with > 95% rocks (see section 5.4).
- `unlimitedSoilWater [= FALSE]`: Boolean flag to indicate the simulation of plant transpiration assuming that soil water is always at field capacity.
- `unfoldingDD [= 300]`: Degree-days for complete leaf unfolding after budburst has occurred (S_{unf}^* ; see section 4.1.2).
- `rhizosphereOverlap [= "total"]`: A string indicating the degree of rhizosphere spatial overlap between plant cohorts:
 - "none" - no overlap (independent water pools).
 - "partial" - partial overlap determined by coarse root volume.
 - "total" - total overlap (plants extract from common soil pools).
- `verticalLayerSize [= 100]`: Size of vertical layers (in cm) for the calculation of light extinction (see section 2.4.5).
- `windMeasurementHeight [= 200]`: Height (in cm) over the canopy corresponding to wind measurements.
- `cavitationRefill [= "total"]`: A string indicating how refilling of embolized conduits is done (see section 6.2.2. Allowed values are:
 - "none" - no refilling.
 - "annual" - every first day of the year.
 - "total" - instantaneous complete refilling.

3.5.4 Model output

Functions `spwb()` and `pwb()` return a list of a class with the same name. The first four elements of this list (i.e., `latitude`, `topography`, `weather` and `spwbInput`) are simply copies of model inputs. The next element is `spwbOutput`, which contains the state of the input object at the end of the simulation (this can be used to perform further simulations starting with current values of state variables). The next five elements correspond to *water balance flows*, *soil state* variables, *stand-level* variables, *plant-level* and *fire hazard* results:

Element	Description
WaterBalance	Climatic input and water balance flows of eq.(3.1) (i.e. net precipitation, infiltration, runoff, transpiration...). All of them in $mm = l \cdot m^{-2}$.
Soil	Soil variables for each soil layer: Moisture relative to field capacity (W_s), water potential (Ψ_s) and volumetric water content (V_s).
Stand	Stand-level variables, such as LAI_{stand}^ϕ , LAI_{stand}^{dead} , the water retention capacity of the canopy (S_{canopy}) or the fraction of light reaching the ground (L_{ground}^{PAR} and L_{ground}^{SWR}).
Plants	Variables defined at the level of woody plant cohort, such as expanded leaf area (LAI_i^{ϕ}), transpiration, photosynthesis, water potential, etc.
FireHazard	Fire hazard variables (fuel moisture, rate of spread, fire potentials, etc). Only returned if <code>fireHazardResults = TRUE</code> .

Elements `WaterBalance`, `Soil`, `Stand` and `FireHazard` are data frames with dates in rows and variables in columns, whereas `Plants` is itself a list with several data frames, with dates in rows and woody plant cohorts in columns:

Element	Symbol	Units	Description
LAI	LAI_i^ϕ	$m^2 \cdot m^{-2}$	Leaf area index (expanded).
LAIlive	LAI_i^{live}	$m^2 \cdot m^{-2}$	Leaf area index (live).
AbsorbedSWRFraction		[0-1]	Fraction of shortwave radiation absorbed.
Transpiration Tr_i		mm	Transpiration per unit ground area.
GrossPhotosynthesis G_P		$g C \cdot m^{-2}$	Gross photosynthesis per unit ground area.
PlantPsi	$\Psi_{plant,i}$	MPa	Plant water potential.
StemPLC	PLC_i	[0-1]	Degree of stem embolisation.
PlantWaterBalance		mm	Internal daily plant water balance (balance of soil extraction and transpiration).
LeafRWC	$RWC_{leaf,i}$	%	Mean leaf relative water content.
StemRWC	$RWC_{stem,i}$	%	Mean stem relative water content.
LFMC	$LFMC_i$	%	Live fuel moisture content (as percent of dry weight), corresponding to fine fuels (< 6.35 mm twigs and leaves).
PlantStress	DDS_i	[0-1]	Drought stress level suffered by each plant cohort (relative whole-plant transpiration).

The output of simulations can be inspected using `plot`, `shinyplot` and `summary` functions specific to `spwb` objects (examples are given in the corresponding package *vignette*).

As it simulates water balance for only one day, function `spwb_day()` returns a much more reduced output. This function is most useful with advanced water balance modelling (chapter 7).

3.6 Applications

The basic water balance model is easier to parameterize and is faster the advanced water balance model. These features make it appropriate for applications that do not seek a high predictive capacity of water status at the plant level, but require robust estimates of water fluxes at the stand, landscape or regional levels. In our opinion the basic water balance model may be useful for the following applications:

1. To monitor or forecast temporal variation in transpiration fluxes and soil water content in particular stands or in multiple stands across a landscape or region.
2. Monitor or forecast temporal variation of plant drought stress in particular stands or in multiple stands across a landscape or region (for example to anticipate mortality events).

For example, Karavani *et al.* (2018) employed the water balance to provide sound estimates of soil moisture dynamics to predict mushroom production in a network forest stands. The model is particularly interesting to test the relationship between forest structure/composition and the water balance at the stand level or the drought stress suffered by plants, as done in Ameztegui *et al.* (2017). See also another application in Rolo & Moreno (2019).

Chapter 4

Leaf phenology and radiation extinction

Leaf phenology determines seasonal variations in the surface area of leaves, hence influencing the energy and water exchange between plants and the atmosphere. Radiation extinction is an important process to consider in a model where plants have different heights.

4.1 Leaf phenology

4.1.1 Phenological types and leaf expanded area

Plant species can have either *progressive evergreen*, *one-flush evergreen*, *winter-deciduous*, *winter semi-deciduous* or *drought-deciduous* leaf phenology (see Fig. 4.1). Progressive evergreen plants are assumed to shed old leaves at the same time as new leaves appear, so that the leaf area remains constant throughout the year. One-flush evergreen plants concentrate leaf unfolding and leaf senescence during similar periods, so that leaf area is also approximately constant year-round. In winter-deciduous plants, leaf shedding and leaf bud formation occurs in autumn. Buds remain under dormancy until spring, when budburst and unfolding occurs. The difference between semi-deciduous and deciduous plants is that the former retain dead leaves in the plant until next season unfolding period, so that leaf fall is retarded. Finally, drought-deciduous plants behave like progressive-evergreen plants except that they can shed their leaves in response to drought.

Evergreen plants thus maintain constant leaf area over the year, whereas in deciduous plants leaf-expanded status is updated daily, represented by ϕ_i , the fraction of maximum leaf area:

$$LAI_i^\phi = LAI_i^{live} \cdot \phi_i \quad (4.1)$$

whereas for evergreen species $LAI_i^\phi = LAI_i^{live}$ (or equivalently $\phi_i = 1$ at all times).

The general structure of process based phenological models explained in Chuine *et al.* (2013). These models aim determine, for a given phenological phase n (endodormancy, ecodormancy, maturation, etc), the day of its finalisation d_n , such that the following equation holds:

$$S_{n,d} = \sum_{d_{n-1}}^{d_n} R_{n,d} = S_n^* \quad (4.2)$$

where $S_{n,d}$ is the state of development on day d in phase n , and d_{n-1} is the end of the previous phase. $R_{n,d}$ is the rate of development during phase n on day d , which depends on environmental variables (temperature, photoperiod, ...) and S_n^* is the critical threshold to achieve the change of phase.

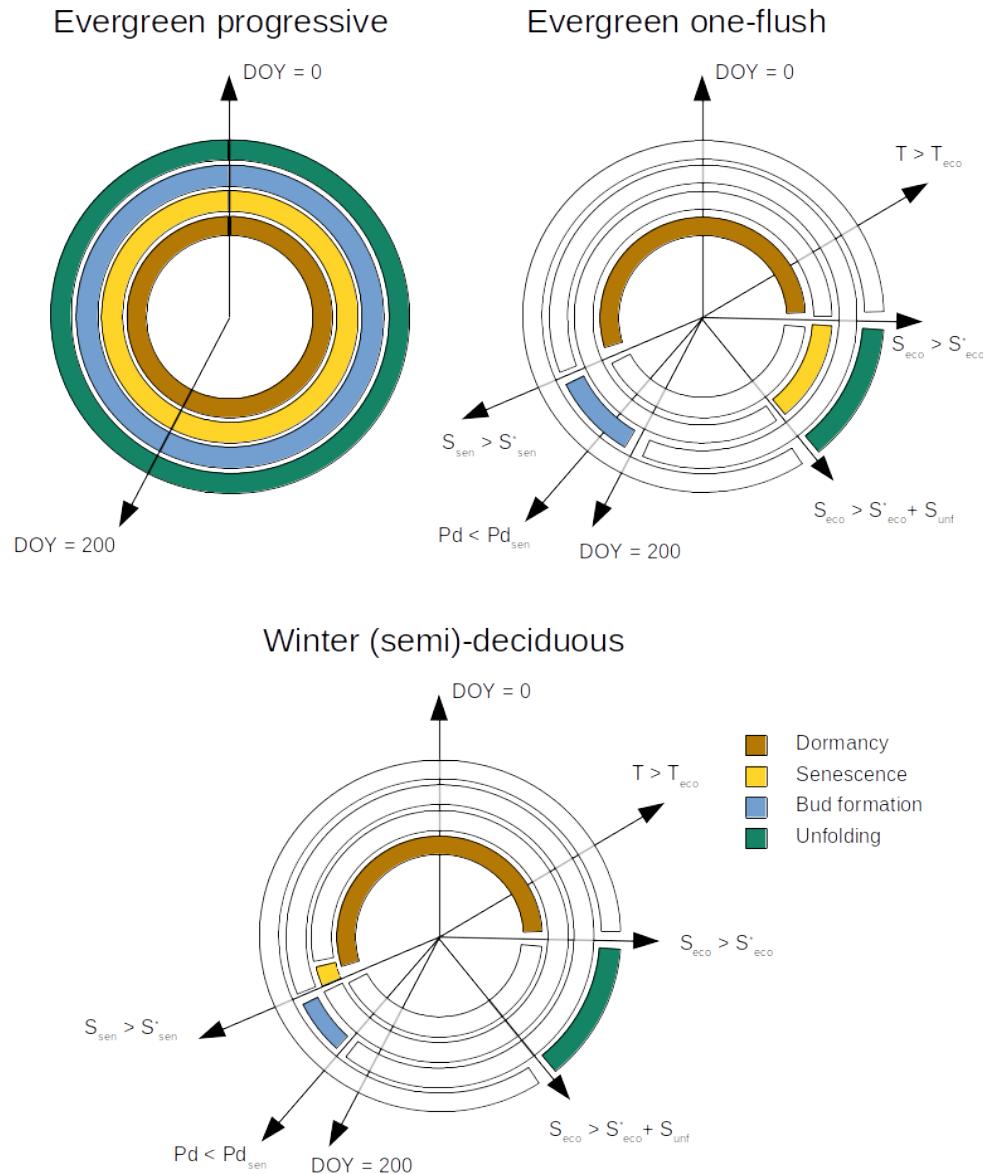


Figure 4.1: Schematic representation of leaf phenology types

In the water balance model leaf area index (*LAI*) values of winter (semi-)deciduous plants are adjusted for leaf phenology following a leaf development model in spring and a leaf senescence model in autumn. Function `pheno_updateLeaves()` updates the status of expanded leaves and dead leaves in a simulation object.

4.1.2 Bud burst and leaf unfolding

Transition from ecodormancy to bud burst is estimated using a very simple one-phase ecodormancy model (also called the spring warming model) implemented in function `pheno_leafDevelopmentStatus()`. Given a base temperature (T_{eco} , in °C), the rate of development during the ecodormancy phase (R_{eco} , in °C) is zero for those days where mean temperature T_{mean} is below T_{eco} and $T_{mean} - T_{eco}$ for those days where temperatures become warmer than this threshold:

$$R_{eco,d}(T_d) = \begin{cases} 0 & T_d \leq T_{eco} \\ T_d - T_{eco} & T_d > T_{eco} \end{cases} \quad (4.3)$$

Degree accumulation starts after the year date surpasses parameter $t_{0,eco}$. If $DOY > t_{0,eco}$, daily $R_{eco,d}$ values are added to the cumulative sum $S_{eco,d}$ and budburst occurs when $S_{eco,d} > S_{eco}^*$. Both T_{eco} and S_{eco}^* are plant-specific parameters in `spwbInput`.

After budburst, we model progressive leaf unfolding in a similar manner, by accumulating degree days using the same definition as for $R_{eco,d}$:

$$R_{unf,d} = R_{eco,d} \quad (4.4)$$

and then defining an unfolding development status $S_{unf,d} = \sum_{d,eco}^d R_{unf,d}$, which we use to determine $\phi_{i,d}$, the degree of leaf expansion of cohort i at day d :

$$\phi_{i,d} = S_{unf,d}/S_{unf}^* \quad (4.5)$$

until $S_{unf,d} \geq S_{unf}^*$ and the unfolding process ends, where S_{unf}^* is a simulation control parameter `unfoldDD`.

4.1.3 Leaf senescence

Leaf senescence follows the models developed by Delpierre *et al.* (2009). The daily rate of development of senescence $R_{sen,d}$ is defined on the basis of daily photoperiod (Ph_d , in hours) and temperature (T_d , in °C):

$$R_{sen,d}(Ph_d, T_d) = \begin{cases} 0 & T_d \geq T_{sen} \text{ or } Ph_d > Ph_{sen} \\ (T_d - T_{sen})^{x_{sen}} \cdot (Ph_d/Ph_{sen})^{y_{sen}} & T_d < T_{sen} \text{ and } Ph_d < Ph_{sen} \end{cases} \quad (4.6)$$

where Ph_{sen} is the maximum photoperiod to start counting the senescence; x_{sen} and y_{sen} are exponents regulating the importance of temperature and photoperiod on leaf senescence. The state of development of senescence $S_{sen,d}$ is defined

$$S_{sen,d} = \sum R_{sen,d} \quad (4.7)$$

and when $S_{sen,d} \geq S_{sen}^*$ then expanded leaves are assumed to suddenly die. This model is implemented in function `pheno_leafSenescenceStatus()` and returns $\phi_{i,d} = 1$ for all days until $S_{sen,d} \geq S_{sen}^*$ when it returns $\phi_{i,d} = 0$. Parameters T_{sen} , Ph_{sen} and S_{sen}^* are plant-specific parameters in `spwbInput`.

4.1.4 Leaf abscission

The drop of ϕ_i causes live expanded leaves to become dead leaves. To avoid a sudden decrease of leaf area, dead leaves are kept in the canopy and they are reduced daily using a negative exponential function of wind speed:

$$LAI_i^{dead} = LAI_i^{dead} \cdot e^{-u/10} \quad (4.8)$$

where u is wind speed in ($m \cdot s^{-1}$).

4.2 Radiation extinction

The proportion of photosynthetically active radiation (PAR) and short-wave radiation (SWR; 400-3000 nm) decreases through the canopy following the Beer-Lambert's light extinction equation. L_{herb}^{PAR} , the proportion of PAR that reaches the herbaceous layer, is calculated as:

$$L_{herb}^{PAR} = e^{-\sum_{i=1}^c k_{PAR,i} \cdot LAI_i^{all}} \quad (4.9)$$

where $k_{PAR,i}$ is the PAR extinction coefficient of woody plant cohort i . If we add the extinction caused by the herbaceous layer, we have that L_{ground}^{PAR} , the proportion of PAR that reaches the ground, is calculated as:

$$L_{ground}^{PAR} = L_{herb}^{PAR} e^{-0.5 \cdot LAI_{herb}} \quad (4.10)$$

Where herbaceous vegetation is assumed to have a fixed extinction coefficient.

We can also define L_i^{PAR} as the proportion of PAR available for a given plant cohort i , which we associate to the PAR available at a height corresponding to half of the cohort's crown. This height will include some self-shading of the target cohort.

The proportion of short-wave radiation (SWR) energy absorbed by each plant cohort needs to be calculated to divide the transpiration of the stand among cohorts (chapter 6), and the radiation absorbed by the soil is needed to calculate soil evaporation (section 5.5). Foliage absorbs a higher proportion of PAR than SWR; thus, the extinction coefficient is higher for PAR than for SWR. However, values for the ratio of extinction coefficients are rather constant. Following Friend et al. (1997) it is assumed that the extinction coefficient for PAR is 1.35 times larger than that for SWR (i.e. $k_{SWR,i} = k_{PAR,i}/1.35$). Figure 4.2 shows the PAR and SWR extinction profiles (see functions `vprofile_PARExtinction()` and `vprofile_SWRExtinction()`) corresponding to the leaf area density distribution of Fig. 2.4.

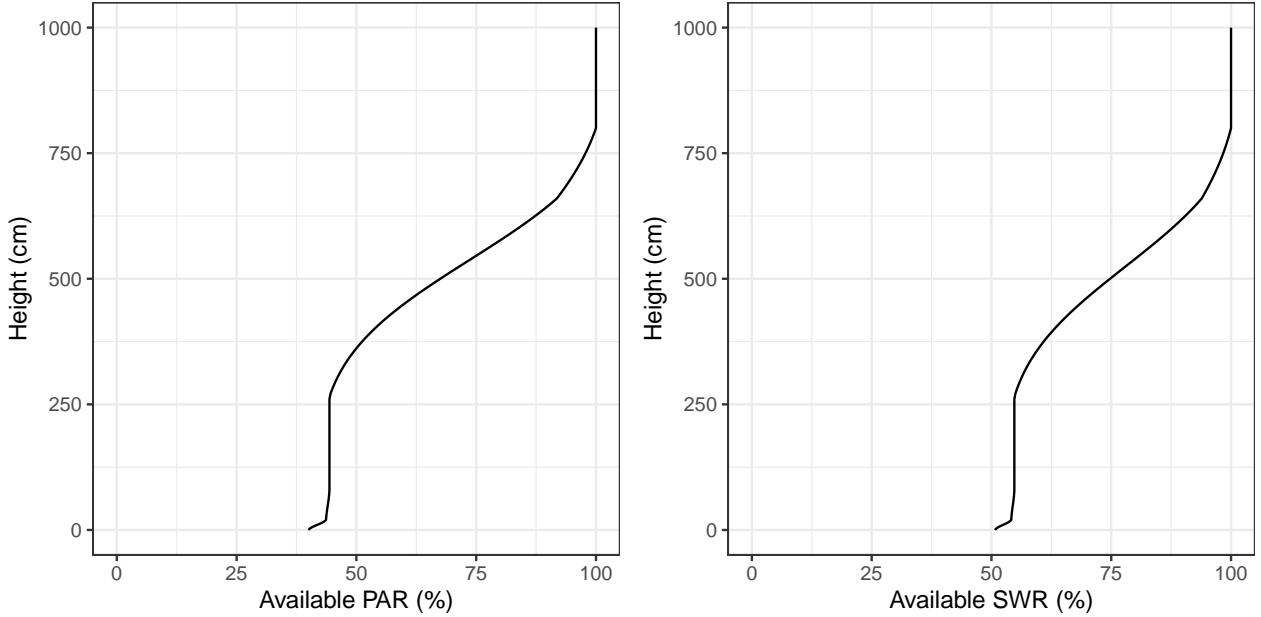


Figure 4.2: Light extinction in a forest stand. Note the sharper decrease of PAR (left panel) in comparison to SWR (right panel)

To calculate radiation absorption, where the vertical dimension of the plot is divided into layers (as explained in 2.4.5), and the SWR absorbed is calculated for each plant cohort in each layer. Let l be the number of vertical layers. The fraction of radiation incident on layer j that is absorbed in the same layer is:

$$f_j = 1 - e^{-\sum_{i=1}^c k_{SWR,i} \cdot LAI_{i,j}^{all}} \quad (4.11)$$

where $LAI_{i,j}^{all}$ is the leaf area index of cohort i in layer j . Hence, the fraction transmitted is $(1 - f_j)$. The fraction of radiation incident on layer j that is absorbed by expanded leaves of plant cohort i in that layer (f_{ij}) is calculated from the relative contribution of these leaves to the total absorption in the layer:

$$f_{ij} = f_j \cdot \frac{k_{SWR,i} \cdot LAI_{i,j}^\phi}{\sum_{h=1}^c k_{SWR,h} \cdot LAI_{h,j}^{all}} \quad (4.12)$$

The fraction of canopy radiation absorbed by a plant cohort i across all layers is found by adding the fraction absorbed in each layer:

$$f_i = \sum_{j=1}^l f_{ij} \cdot \prod_{h>j}^l (1 - f_h) \quad (4.13)$$

where for each layer the fraction of the radiation incident in the canopy that reaches the layer is found by multiplying the transmitted fractions across the layers above it. For example, the fraction of SWR absorbed by each of the three cohorts of the example in Fig. 2.3 would be (see function `light_cohortAbsorbedSWRFraction()`):

```
##      T1_148      T2_168      S1_165
## 0.247978703 0.204200474 0.006792845
```


Chapter 5

Forest hydrology

Forest hydrological processes are key for soil and plant water balances. This chapter details the design and implementation of most of the vertical hydrological processes included in package **medfate**. These processes determine the value of the water flows mentioned in eq. (3.1). The only process that is not described here is woody plant transpiration (Tr_w), as it will be covered in detail in chapter 6.

Although each process can be simulated in specific sub-model functions, function `hydrology_verticalInputs()` allows simulating soil water input processes altogether, including canopy water interception, snow accumulation/melt, soil infiltration and percolation.

5.1 Snow and rainfall

Precipitation (P) is considered be snow precipitation (Ps) when $T_{mean} < 0$, and is considered rainfall (Pr) otherwise. Thus, we have:

$$P = Pr + Ps \quad (5.1)$$

Interception of snow by the canopy is neglected, and all snow is assumed to accumulate in a single storage compartment S_{snow} over the soil (i.e. canopy snow storage capacity is neglected). Rainfall interception is described in section 5.3.

5.2 Snow pack dynamics

A very simple snow submodel is used for snow pack dynamics (accumulation and melt), taken from Kergoat (1998). When mean air temperature is above 0 Celsius ($T_{mean} > 0$), a simple energy budget relates snow melt, Sm (mm), to air temperature and soil-level radiation (see function `hydrology_snowMelt()`):

$$Sm = \frac{Rad \cdot L_{ground}^{SWR} \cdot (1 - \alpha_{ice}) + \tau_{day} \cdot T_{mean} \cdot \rho_{air} \cdot C_p / r_s}{\lambda_{ice}} \quad (5.2)$$

where Rad is solar radiation ($MJ \cdot m^{-2}$), L_{ground}^{SWR} is the fraction of (short-wave) radiation reaching the ground, $\alpha_{ice} = 0.9$ is the albedo of snow, $\tau_{day} = 86400$ is the day duration in seconds, ρ_{air} is the air density ($kg \cdot m^{-3}$), depending on temperature and elevation (see utility functions of the **meteoland** reference manual), $C_p = 1013.86 \cdot 10^{-6} MJ \cdot kg^{-1} \cdot C^{-1}$ is the specific heat capacity of the air and $r_s = 100 s \cdot m^{-1}$ is the snow aerodynamic resistance and $\lambda_{ice} = 0.33355 MJ \cdot kg^{-1}$ is the (latent) heat of fusion of snow.

5.3 Rainfall interception loss

As mentioned above, interception loss is only modelled for liquid precipitation (i.e. snow interception is not modelled). Rainfall interception loss, In , is estimated using the Gash *et al.* (1995) analytical interception model for sparse canopies, where rain is assumed to fall in a single event during the day. First, the amount of rainfall needed to saturate the canopy is calculated:

$$P_G = -\frac{S_{canopy}/C_{canopy}}{ER_{ratio}} \cdot \ln(1 - ER_{ratio}) \quad (5.3)$$

where S_{canopy} is the canopy water storage capacity (in mm) – i.e. the minimum amount of water needed to saturate the canopy –, C_{canopy} is the canopy cover and ER_{ratio} is the ratio of evaporation rate to rainfall rate during the rainfall event. Although interception models are normally applied to single-canopy stands, we apply the sparse Gash model to the whole stand (including shrubs and the herbaceous layer). Moreover, in our implementation stem interception is lumped with crown interception, so that S_{canopy} represents both. Following Watanabe & Mizutani (1996) we estimate S_{canopy} , the canopy water storage capacity, from adjusted LAI values:

$$S_{canopy} = LAI_{herb} + \sum_i s_{water,i} \cdot LAI_i^\phi \quad (5.4)$$

where $s_{water,i}$ is the depth of water that can be retained by leaves, branches and stems of cohort i per unit of leaf area index ($mm \cdot LAI^{-1}$). To estimate the stand cover, C_{canopy} , we use the complement of the percentage of PAR that reaches the ground, i.e. $C_{canopy} = 1 - L_{ground}^{PAR}$ (Deguchi *et al.* 2006).

Simplifying assumptions are made to determine ER_{ratio} . Values of the evaporation-to-rainfall ratio are calculated from daily potential evapotranspiration and rainfall, while accounting for seasonal variation in rainfall intensity ($mm \cdot h^{-1}$). Minimum values for rainfall intensity are assumed for convective storms ($5.6 mm \cdot h^{-1}$) and synoptic storms ($1.5 mm \cdot h^{-1}$) from Miralles *et al.* (2010). Synoptic storms are assumed between December and June, and convective storms are assumed for the remaining months, as typical in the Mediterranean Basin.

The amount of water evaporated from interception (In , in mm), is calculated as:

$$In = C_{canopy} \cdot P_G + C_{canopy} \cdot ER_{ratio} \cdot (Pr - P_G) \text{ if } Pr > P_G \quad (5.5)$$

$$In = C_{canopy} \cdot Pr \text{ if } Pr \leq P_G \quad (5.6)$$

where Pr is the daily gross rainfall (in mm). Net rainfall (Pr_{net} , also in mm) includes throughfall and stemflow, and is calculated as the difference between gross rainfall and interception loss. Fig. 5.1 below shows examples of relative throughfall (including stemflow), calculated according to the Gash *et al.* (1995) interception model, under different situations (see function `hydrology_rainInterception()`).

5.4 Runoff, infiltration and percolation

The amount of water that reaches the soil is the sum of net rainfall (Pr_{net}), runoff (Ro , in mm) and melted snow (Sm). The amount of water infiltrating into the soil is $Pr_{net} + Sm + Ro - Ru$, where Ru (in mm) is the water lost by surface runoff (see function `hydrology_infiltrationAmount()`).

Surface runoff, is calculated using the USDA SCS curve number method, as in Boughton (1989):

$$Ru = \frac{(Pr_{net} + Ro + Sm - 0.2 \cdot V_{fc,soil})^2}{(Pr_{net} + Ro + Sm - 0.8 \cdot V_{fc,soil})} \quad (5.7)$$

where $V_{fc,soil}$ (in mm) is the overall soil water retention capacity.

Following Granier *et al.* (1999), part of the water reaching one soil layer percolates quickly through the macropores. The amount of water reaching each layer through macropores is modelled using an extinction function that depends on macroporosity (see function `hydrology_infiltrationRepartition`). The

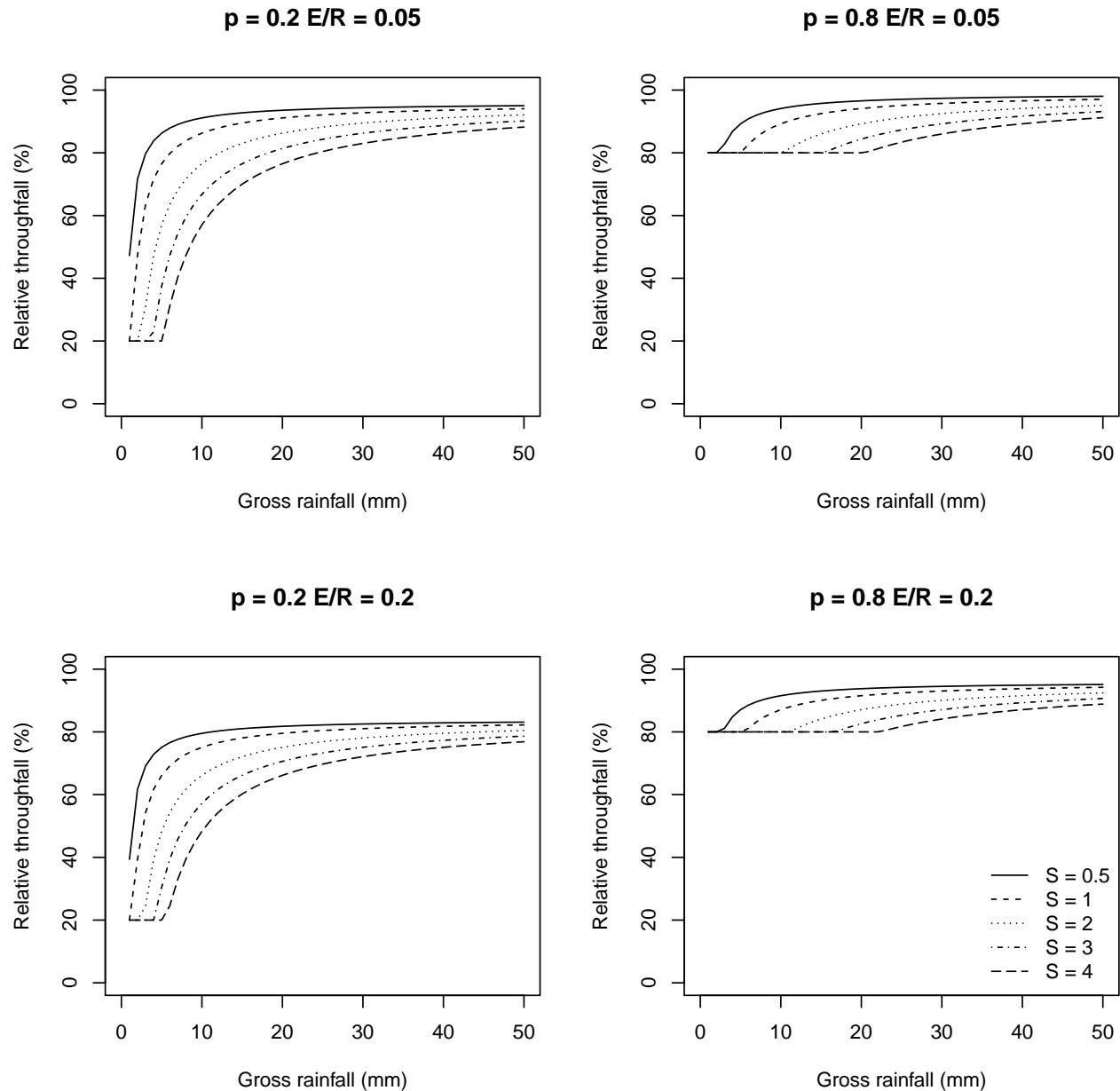


Figure 5.1: Examples of canopy interception with different S_{canopy} (canopy water storage capacity), ER_{ratio} (ratio between evaporation and rainfall rates) and p (throughfall coefficient; $p = 1 - C_{canopy}$).

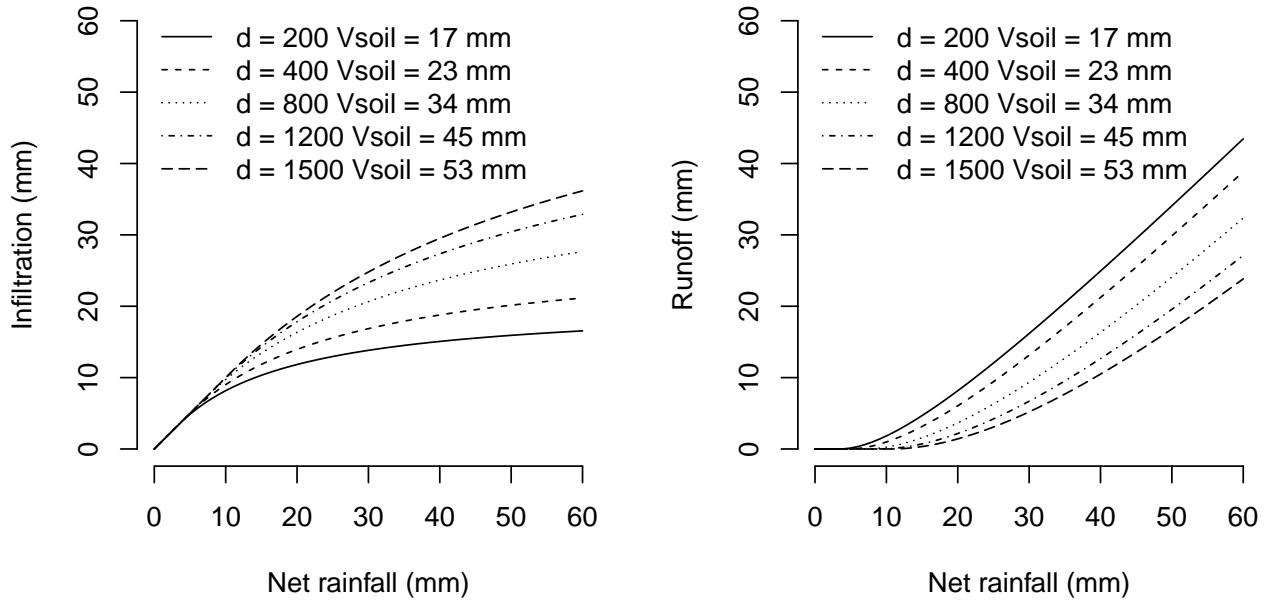


Figure 5.2: Examples of infiltration/runoff calculation for different values of net rainfall and overall retention capacity, V_{soil} , calculated from different soil depths (topsoil+subsoil) and assuming that soil texture is 15% clay and 25% sand. Rock fragment content was 25% and 40% for the topsoil and subsoil, respectively.

remaining water is retained by the micropores refilling the current soil layer. When this soil layer reaches its field capacity the excess of water also percolates to the soil layer below.

Percolation in deepest layer may be treated differently in the model. If it is a rocky layer (i.e if rock fragment content > 95%) and `rockyLayerDrainage = FALSE`, the model assumes that most cracks in the rock are not hydraulically connected to groundwater, so that they conform water pockets from which plants can extract water. Hence, in this situation the percolation excess occurs only if saturation is surpassed. Otherwise, percolation excess is estimated if field capacity is surpassed, as in the other soil layers. When a percolation excess exists, the remaining water fills soil layers over field capacity up to saturation, from bottom to top. If the soil becomes saturated and there is still an excess of water, this is assumed as infiltration excess and added to the surface runoff (R_u).

Deep drainage (D_d) is estimated after percolation of soil water inputs. First, hydraulic head is estimated summing the amount of water contained in macropores across soil layers (i.e. water over field capacity). Then, an estimation of the daily maximum drainage is obtained by multiplying hydraulic head by the saturated vertical hydraulic conductivity K_{drain} . Drainage of saturated water is then applied from top to bottom until the maximum drainage is achieved or macropores are empty in all soil layers. As before, the drainage from the deepest rocky layer may be prevented if `rockyLayerDrainage = FALSE`.

5.5 Bare soil evaporation

Evaporation from the soil surface is the last component of the soil water balance to be calculated before calculating plant transpiration. Bare soil evaporation cannot happen if there is snow over the soil surface (i.e., if $S_{snow} > 0$).

Potential evaporation from the soil (PE_{soil} ; in $\text{mm} \cdot \text{day}^{-1}$) is defined as the product between PET and L_{ground}^{SWR} , the proportion of SWR absorbed by the ground:

$$PE_{soil} = PET \cdot L_{ground}^{SWR} \quad (5.8)$$

Actual evaporation from the soil surface is modeled as in Mouillot *et al.* (2001), who in turn followed Ritchie (1972). First, the model determines t_{soil} , the time needed to evaporate the current water deficit (difference between field capacity and current moisture) in the surface soil layer:

$$t_{soil} = \left\{ \frac{V_{fc,1} \cdot (1 - W_1)}{\gamma_{soil}} \right\} \quad (5.9)$$

where $V_{fc,1}$ is the water retention capacity of layer 1, W_1 is the proportion of moisture in relation to field capacity of layer 1 and γ_{soil} is the maximum daily evaporation ($mm \cdot day^{-1}$). The calculated time is used to determine the ‘supplied’ evaporation, SE_{soil} :

$$SE_{soil} = \gamma_{soil} \cdot (\sqrt{t_{soil} + 1} - \sqrt{1}) \quad (5.10)$$

The amount of water actually evaporated from the soil, Es , is then calculated as the minimum between supply, SE_{soil} , and demand (Federer 1982), i.e. PE_{soil} (see function `hydrology_soilEvaporationAmount`):

$$Es = \min(PE_{soil}, SE_{soil}) \quad (5.11)$$

Finally, Es is distributed along the soil profile according to an exponential decay function with an extinction coefficient κ_{soil} (Mouillot *et al.* 2001). Figure 5.3 shows the cumulative evaporation from soils for different values of maximum evaporation rate and extinction coefficient. When the extinction coefficient is smaller a higher proportion of the evaporated water is removed from the subsoil and less from the topsoil. This causes more water being available to calculate t in the next step.

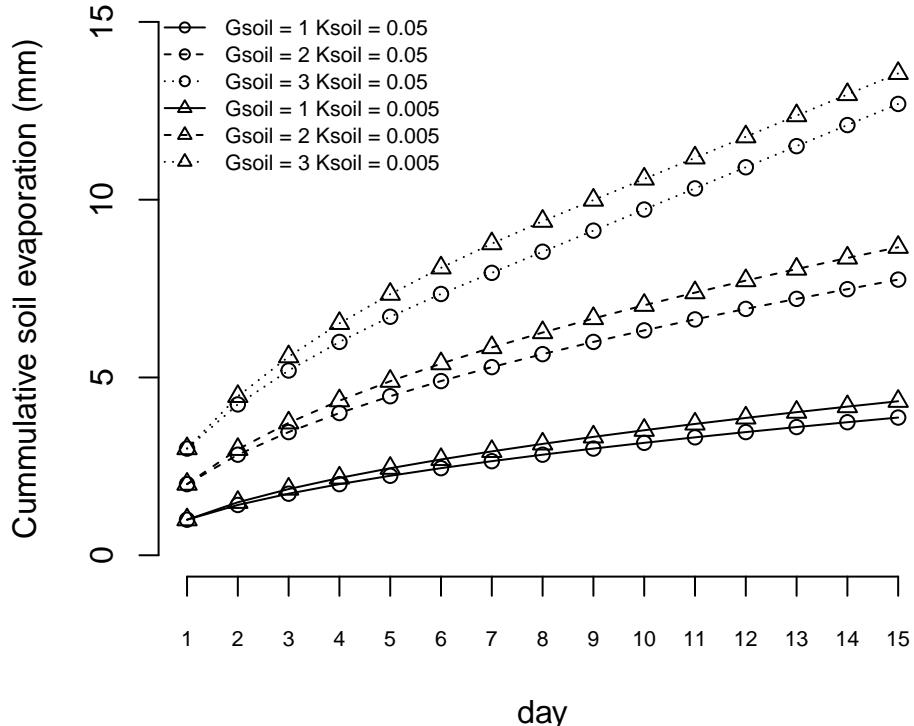


Figure 5.3: Cumulative bare soil evaporation for different values of maximum evaporation rate (γ_{soil}) and extinction coefficient (κ_{soil}). Three soil layers (0 – 30 cm; 30 – 150 cm; 150 – 400 cm) are initialized at field capacity ($V_1 = 50mm$; $V_2 = 201mm$; $V_3 = 35mm$). PE_{soil} was assumed not to be limiting.

5.6 Transpiration of the herbaceous layer

Transpiration of herbaceous layer is modelled analogously to the transpiration of woody plants. First, potential evapotranspiration from the herbaceous layer (PET_{herb} ; in $mm \cdot day^{-1}$) is defined as the product between PET and L_{herb}^{SWR} , the proportion of SWR reaching the herbaceous layer:

$$PET_{herb} = PET \cdot L_{herb}^{SWR} \quad (5.12)$$

Thus, the more dense the woody plant canopy, the lower herbaceous transpiration will be. Maximum herbaceous transpiration $Tr_{herb,max}$ depends on both PET_{herb} and the amount of transpiring surface, i.e. LAI_{herb} . To estimate Tr_{max} the model uses the empirical equation of Granier *et al.* (1999), as in section 6.1.1:

$$\frac{Tr_{herb,max}}{PET} = -0.006 \cdot (LAI_{herb})^2 + 0.134 \cdot LAI_{herb} \quad (5.13)$$

Finally, actual herbaceous transpiration (Tr_{herb}) is reduced according to the water potential in the topmost soil layer (Ψ_1), assuming that transpiration reduction follows Weibull function with fixed coefficients:

$$Tr_{herb} = Tr_{herb,max} \cdot \exp \left\{ \ln(0.5) \cdot \left[\frac{\Psi_1}{-1.5} \right]^2 \right\} \quad (5.14)$$

Chapter 6

Woody plant transpiration, photosynthesis and water status

This chapter describes how daily woody plant transpiration (Tr_{woody}) is modelled in the basic water balance model, completing the flows needed to evaluate eq. (3.1). The model uses the approach of BILJOU (Granier *et al.* 1999) to estimate woody maximum transpiration, and then divides this value among the contribution of plant cohorts and estimate actual transpiration depending on soil moisture and cohort functional parameters, which determine plant drought. The chapter also describes how is daily photosynthesis estimated, a process that in this model does not influence transpiration but is necessary for growth calculations (chapter 15).

6.1 Plant transpiration and photosynthesis

6.1.1 Maximum transpiration

Potential evapotranspiration (PET , in mm) is assumed to represent open water evaporation potential. Maximum canopy transpiration $Tr_{woody,max}$ depends on both PET and the amount of transpiring surface, i.e. the current cumulative LAI of the woody vegetation, LAI_{woody}^ϕ . To estimate $Tr_{woody,max}$ the model uses the empirical equation of Granier *et al.* (1999), where $Tr_{woody,max}/PET$ is a function of LAI_{woody}^ϕ :

$$\frac{Tr_{woody,max}}{PET} = -0.006 \cdot (LAI_{woody}^\phi)^2 + 0.134 \cdot LAI_{woody}^\phi + 0.036 \quad (6.1)$$

This equation has already been adopted for Mediterranean biomes (Fyllas & Troumbis 2009; Ruffault *et al.* 2013).

Since medfate's **ver. 2.1**, empirical coefficients can be species-specific and Granier *et al.* (1999) empirical equation has been generalized to:

$$\frac{Tr_{woody,max}}{PET} = T_{max,LAI} \cdot LAI_{woody}^\phi + T_{max,sqLAI} \cdot (LAI_{woody}^\phi)^2 \quad (6.2)$$

where $T_{max,LAI}$ and $T_{max,sqLAI}$ are species-specific parameters (if missing, they are given default values 0.134 and -0.006, respectively).

The maximum transpiration for a given plant cohort i is calculated using the portion of Tr_{max} defined by f_i the fraction of total absorbed SWR that is due to cohort i (see section 4.2):

$$Tr_{max,i} = Tr_{woody,max} \cdot \frac{f_i^{0.75}}{\sum_j f_j^{0.75}} \quad (6.3)$$

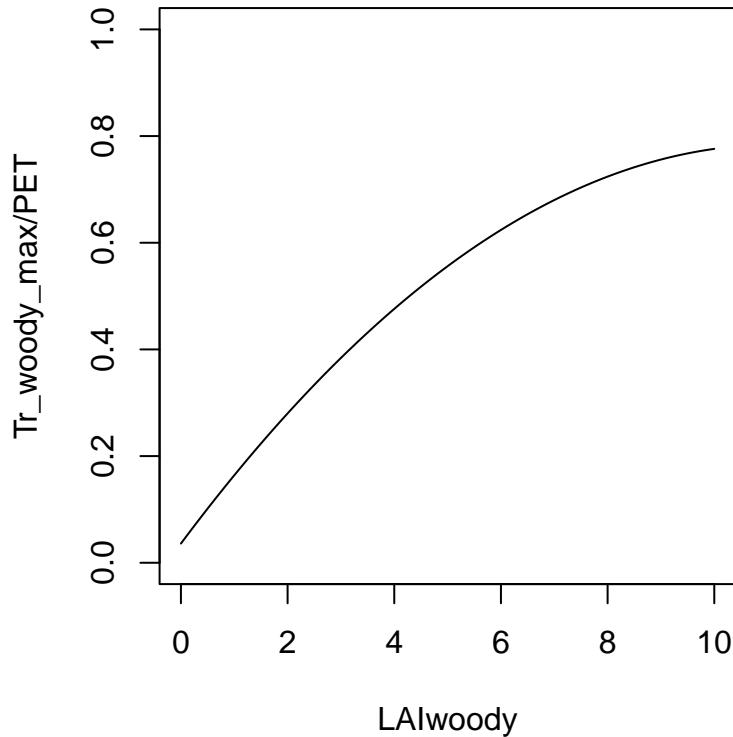


Figure 6.1: Empirical relationship between $Tr_{woody,max}/PET$ and LAI_{woody}^ϕ

Before **ver 2.5.0** $Tr_{max,i}$ was linearly related to f_i , but an exponent 0.75 is now used to decrease *ad hoc* the proportion of transpiration corresponding to cohorts absorbing high SWR fractions with respect to those absorbing low amounts of SWR, which has been found to better match the transpiration predicted using the advanced transpiration model. Note that the actual amount of radiation energy absorbed by plants is much more difficult to determine, involving diffuse and direct radiation, as well as net long-wave radiation (see chapter 9).

6.1.2 Actual plant transpiration and plant water potential

Actual plant transpiration depends on soil moisture and is calculated for each plant cohort and each soil layer separately. $Tr_{i,s}$ (in mm) represents the transpiration made by cohort i from layer s . Actual plant transpiration from a given layer is regulated by soil moisture and the resistance to water flow through the plant. For each plant cohort i and soil layer s , the model first estimates the a whole-plant relative water transpiration, $K_{i,s}$, which varies between 0 and 1 depending on $\Psi_{extract,i}$, the potential at which transpiration is 50% of maximum, and Ψ_s , the water potential in layer s . This relationship is modelled using a Weibull function (see function `hydraulics_psi2K()`):

$$K_{i,s} = K_i(\Psi_s) = \exp \left\{ \ln(0.5) \cdot \left[\frac{\Psi_s}{\Psi_{extract,i}} \right]^{c_{extract,i}} \right\} \quad (6.4)$$

where $c_{extract,i}$ is an exponent that modulates the steepness of the decrease in relative conductance when soil potential becomes negative and $\ln(0.5)$ is used to ensure that $K_i(\Psi_{extract,i}) = 0.5$ (Fig. 6.2).

If cavitation has occurred in previous steps then the capacity of the plant to transport water is impaired. Hence $K_{i,s}$ from eq. (6.4) cannot be larger than the one-complement of PLC_i , so that:

$$K_{i,s}^{PLC} = \min\{K_{i,s}, 1.0 - PLC_i\} \quad (6.5)$$

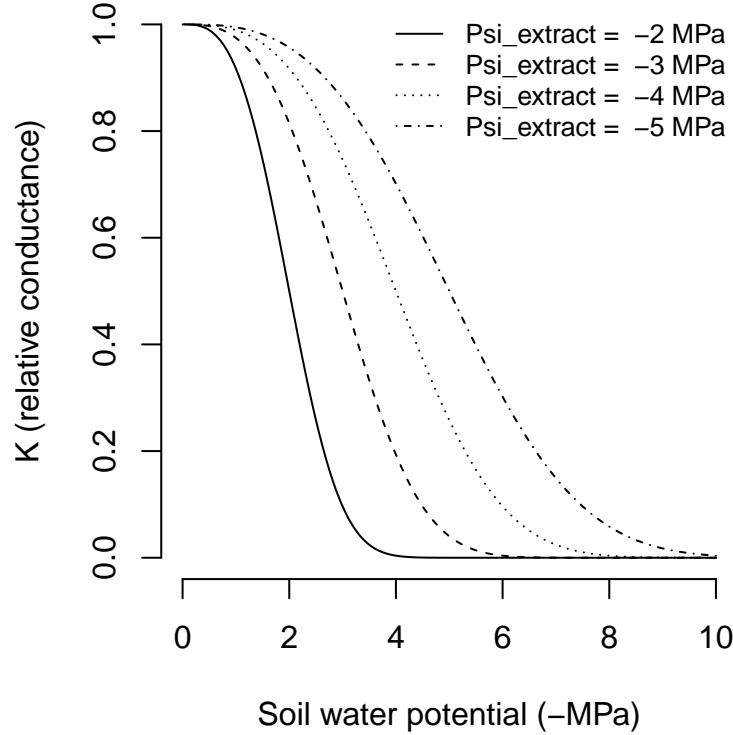


Figure 6.2: Whole-plant relative water conductance functions for different water potential values ($c_{extract} = 3$ in all cases)

where PLC_i is the proportion of conductance lost for cohort i (see 6.2.2).

Until **ver 2.5.0**, actual transpiration of plant cohort i from a given soil layer s , $Tr_{i,s}$, was defined as the product of (Mouillet *et al.* 2001): the maximum transpiration of the plant cohort ($Tr_{max,i}$), the relative whole-plant transpiration ($K_{i,s}^{PLC}$) corresponding to the species and water potential in layer s , and the proportion of plant fine roots in layer s , $FRP_{i,s}$:

$$Tr_{i,s} = Tr_{max,i} \cdot K_{i,s}^{PLC} \cdot FRP_{i,s} \quad (6.6)$$

Since **ver 2.5.0**, this equation has been slightly modified and is now:

$$Tr_{i,s} = Tr_{max,i} \cdot K_{i,s}^{PLC} \cdot \frac{K_{unsat,s}^{0.5} \cdot FRP_{i,s}}{\sum_r K_{unsat,r}^{0.5} \cdot FRP_{i,r}} \quad (6.7)$$

where $K_{unsat,s}$ is the unsaturated soil conductivity of layer s . This modification was introduced to allow the plant to increase the amount of water drawn from layers with higher levels of moisture in seasons where the usual water pools are depleted. This formulation decreases the differences in soil moisture between soil layers, which in the advanced model is achieved thanks to hydraulic redistribution process.

The total amount of water transpired by woody cohort i , Tr_i (in mm) is the sum of $Tr_{i,s}$ values over all soil layers:

$$Tr_i = \sum_s Tr_{i,s} \quad (6.8)$$

Analogously, the total amount of water transpired by woody plants, Tr_{woody} (in mm), is the sum of Tr_i values over all plant cohorts:

$$Tr_{woody} = \sum_i Tr_i = \sum_s \sum_i Tr_{i,s} \quad (6.9)$$

Assuming no water limitations (i.e. $K_{i,s} = 1$), we have that $Tr_{woody} = Tr_{woody,max}$ and $Tr_i = Tr_{max,i}$. Cohort transpiration and stand transpiration will be lower than maximum values if soil water potential in any layer is negative enough to cause a significant reduction in whole-plant conductance. At the plant level, the transpiration of a given plant cohort will be lower than that of others if:

- a. The cohort is under the shade (it reduces f_i and hence $Tr_{max,i}$)
- b. The cohort has a lower amount of leaf area (it reduces f_i and hence $Tr_{max,i}$)
- c. The soil layers exploited by the cohort have more negative water potentials (it reduces $K_{i,s}$ and $K_{i,s}^{PLC}$).

6.1.3 Plant water potential

The basic water balance model does not allow estimating a water potential drop from soil to the leaf. Moreover, in a multilayered soil it is difficult to know what would be the water potential of the plant. Despite these limitations, a gross surrogate of ‘plant’ water potential ($\Psi_{plant,i}$; in MPa) may be obtained averaging whole-plant relative conductance values across soil layers and inverting the whole-plant relative transpiration function:

$$\Psi_{plant,i} = K_i^{-1} \left(\sum_s K_{i,s} \cdot FRP_{i,s} \right) \quad (6.10)$$

where K_i^{-1} is the inverse of the relative whole-plant transpiration eq. (6.4). The plant water potential is used to determine plant stress levels and cavitation, as explained in 6.2.1 and 6.2.2.

Importantly, changes in $\Psi_{plant,i}$ induce changes in plant water content (see section 10.3), which may induce a release of water in the transpiration stream or an absorption of water from it, which leads to the cohort transpiration, Tr_i , being different from its water extraction from the soil, Ex_i .

6.1.4 Minimum transpiration

The description of actual transpiration in the previous section incorporates the assumption that plants can regulate water losses up to zero transpiration, neglecting water leaks through the leaf cuticle and/or incomplete stomatal closure (Duursma *et al.* 2018). To avoid this assumption we can estimate minimum transpiration for a given plant cohort, $Tr_{min,i}$ (in mm), and prevent $Tr_i < Tr_{min,i}$.

Minimum transpiration flow per leaf area E_{min} in $mol\ H_2O \cdot s^{-1} \cdot m^{-2}$ can be calculated if we know (1) the minimum leaf conductance to water vapor ($g_{sw,min}$; in $mol\ H_2O \cdot s^{-1} \cdot m^{-2}$); (2) the leaf vapor pressure deficit (VPD_{leaf} ; in kPa); and (3) atmospheric pressure (P_{atm} ; also in kPa):

$$E_{min} = g_{sw,min} \cdot \frac{VPD_{leaf}}{P_{atm}} \quad (6.11)$$

In turn, the vapor pressure deficit in the leaf (VPD_{leaf}) is calculated as:

$$VPD_{leaf} = e_{leaf} - e_{air} \quad (6.12)$$

Where e_{air} is the water vapor pressure of the air surrounding the leaf (but beyond the leaf boundary layer) and e_{leaf} is the vapor pressure inside the leaf. While e_{air} can be derived from the weather input, e_{leaf} should be calculated from leaf temperature (T_{leaf}) and leaf water potential (Ψ_{leaf}):

$$e_{leaf}(T_{leaf}, \Psi_{leaf}) = e_{sat}(T_{leaf}) \cdot \exp \left\{ \frac{2.17 \cdot \Psi_{leaf}}{T_{leaf} + 273.15} \right\} \quad (6.13)$$

where $e_{sat}(T)$ is a function giving the saturated vapor pressure for temperature T (see utility functions of the **meteoland** reference manual). Given the simplicity of the basic water balance model, here we make the assumption that $\Psi_{leaf} = \Psi_{plant}$ (from the previous time step). Furthermore, we assume that leaf

temperature is coupled with the temperature of the surrounding air. Given the non-linearity of e_{leaf} we average the vapor pressure values provided by $e_{leaf}(T_{\min}, \Psi_{plant})$ and $e_{leaf}(T_{\max}, \Psi_{plant})$.

Once the leaf level E_{\min} is known for a given cohort i , we can scale this instantaneous rate to daily transpiration (in mm) at the cohort level using:

$$Tr_{\min,i} = E_{\min,i} \cdot 24 \cdot 3600 \cdot LAI_i^\phi \cdot 0.018 \quad (6.14)$$

6.1.5 Transpiration with water pools

Considering water pools (i.e. setting `rhizosphereOverlap` = "partial" or `rhizosphereOverlap` = "total" in the control parameters) involves partitioning the stand area into fractions corresponding to the abundance of each plant cohort. More specifically, the model defines as many plant water pools as plant cohorts, with proportions defined by their LAI values. For example, the fraction corresponding to plant cohort j would be:

$$f_{pool,j} = \frac{LAI_j^{live}}{LAI_{stand}^{live}} \quad (6.15)$$

Overall soil moisture is then related to moisture of plant water pools by the following equation, that is fulfilled for each soil layer s :

$$W_s = \sum_j f_{pool,j} \cdot W_{j,s} \quad (6.16)$$

where $W_{j,s}$ is the soil moisture (relative to field capacity) of layer s in the water pool of cohort j , i.e. under the area of the stand attributed to cohort j .

When transpiration occurs and water pools are considered, the model assumes that the rhizosphere of each plant cohort occupies its own water pool, but may enter into the water pools under other plant cohorts if the overlap is partial. In other words, the root systems of different cohorts may overlap horizontally. A given plant cohort i will have its roots in layer s partitioned among different water pools. Let us assume that we know $fr_{i,s,j}$, the (horizontal) proportion of fine roots of cohort i in layer s of the water pool j (see section 2.4.7). These proportions sum one when summed across water pools:

$$\sum_j fr_{i,s,j} = 1 \quad \forall i, s \quad (6.17)$$

The proportion of fine roots in a given water pool, with respect to the whole root system, will then be equal to:

$$FRP_{i,s,j} = fr_{i,s,j} \cdot FRP_{i,s} \quad (6.18)$$

Estimating transpiration with water pools is analogous to estimating transpiration without them, but replacing the role of $FRP_{i,s}$ by $FRP_{i,s,j}$, and $K_{i,s}$ by $K_{i,s,j}$, in eq. (6.7), which leads to the estimation of $Tr_{i,s,j}$. Analogous replacements have to be made when estimating the plant water potential in eq. (6.10).

Finally, note that the amount of water extracted from the soil layer s of given water pool j (i.e. from $W_{j,s}$) will not be $Tr_{i,s,j}$ (whose units are referred to the overall stand) but $Tr_{i,s,j}/f_{pool,j}$.

6.1.6 Plant photosynthesis

Because it is useful for growth, and for compatibility with the 'Sperry' transpiration mode (chapter 7), the water balance model using 'Granier' transpiration mode also calculates gross assimilated carbon. Gross photosynthesis for a given plant cohort i (in $g C \cdot m^{-2}$) is estimated as a function of transpiration:

$$A_{g,i} = Tr_i \cdot WUE_{max,i} \cdot (L_i^{PAR})^{WUE_{PAR,i}} \cdot (1 - e^{WUE_{CO2,i} \cdot C_{air}}) \cdot VPD^{WUE_{VPD}} \quad (6.19)$$

where Tr_i is the transpiration of plant cohort i , $WUE_{max,i}$ is the maximum water use efficiency of the cohort (in $g C \cdot mm^{-1}$) - a species-specific parameter where WUE assumed to be estimated under conditions

of $VPD = 1\text{ kPa}$, no atmospheric CO₂ limitations and maximum light availability -; L_i^{PAR} is the proportion of photosynthetically-active radiation available at the mid-crown of cohort i , according to the Beer-Lambert extinction model; C_{air} is the atmospheric CO₂ concentration (in ppm); VPD is the vapor pressure deficit (in kPa); $WUE_{CO_2,i}$ and $WUE_{VPD,i}$ are species-specific coefficients regulating the influence of CO₂ concentration and VPD on WUE, respectively; and $WUE_{PAR,i}$ is a species-specific exponent indicating how relative WUE decreases with lower light levels.

Different effects are taken into account in the estimation of gross photosynthesis (eq. (6.19)). The dependency of photosynthesis on drought stress is represented in the Tr_i term, whereas the dependency on CO₂ concentration is given by the negative exponential term and the progressive reduction of photosynthesis under higher VPD (assuming a reduction of stomatal conductance but the same water flow) is modelled using negative $WUE_{VPD,i}$ exponents. Finally, the reduction of photosynthesis due to low light availability (for cohorts in the understory) is represented by the reduction of relative WUE given by low values of L_i^{PAR} .

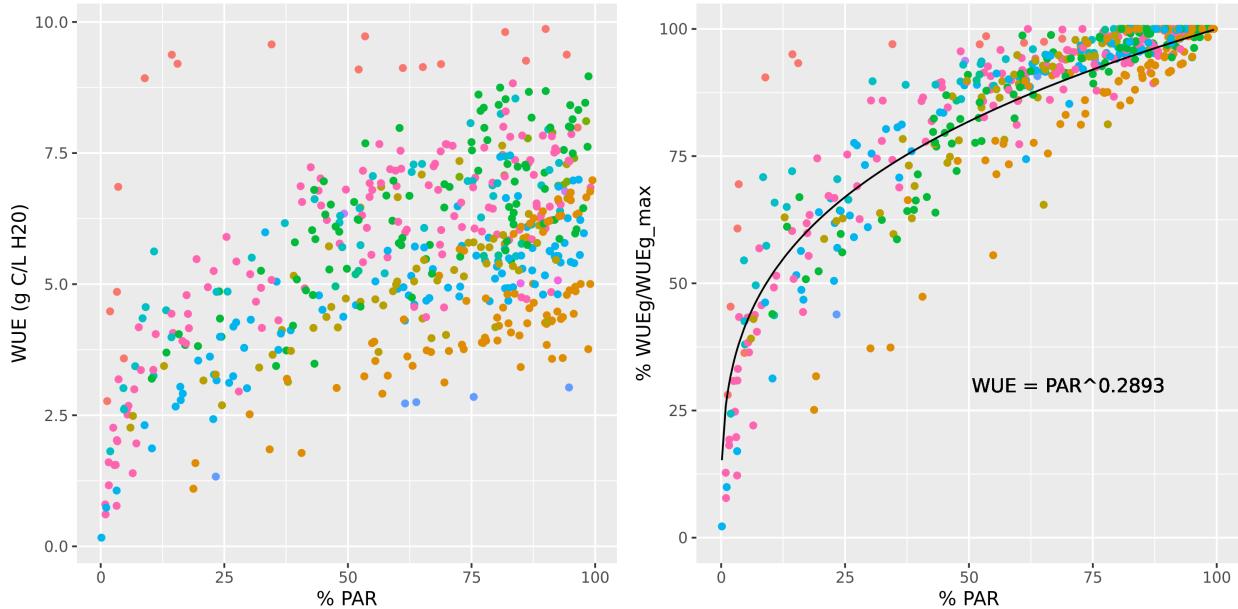


Figure 6.3: Relationship between the percentage of PAR available to a cohort and the water use efficiency (left) or the water use efficiency relative to the maximum value observed in the stand for the same species (right). WUE values were estimated using the advanced water balance model and a non-linear regression was fitted.

6.2 Plant drought stress and cavitation

6.2.1 Daily drought stress

Similarly to Mouillot *et al.* (2002), daily drought stress of a given plant cohort i , DDS_i , is defined as the one-complement of relative whole-plant transpiration (see eq. (6.4)) estimated from its plant water potential ($\Psi_{plant,i}$):

$$DDS_i = \phi_i \cdot (1 - K_i(\Psi_{plant,i})) \quad (6.20)$$

Leaf-phenological status is included to prevent winter deciduous plants from suffering drought stress during winter. Daily drought stress values can be later used to define other drought stress indices for larger temporal scales.

6.2.2 Cavitation and hydraulic recovery

The water balance model is normally run assuming that although soil drought may reduce transpiration, embolized xylem conduits are automatically refilled when soil moisture recovers (in other words, cavitation is reversible and `cavitationRefill = "total"`). It is possible to simulate irreversible cavitation by setting `cavitationRefill = "none"`, `cavitationRefill = "rate"` or `cavitation = "annual"` in the control parameters. Any of these options cause the model to keep track of the maximum value of cavitation so far experienced using the proportion of lost conductance for the plant cohort i (PLC_i), which is modelled from Ψ_{plant} using a Weibull function:

$$PLC_i = \max \left\{ PLC_i, 1 - \exp \left\{ \ln(0.5) \cdot \left[\frac{\Psi_{plant,i}}{\Psi_{critic,i}} \right]^r \right\} \right\} \quad (6.21)$$

For simulations of less than one year one can use `cavitationRefill = "none"` to keep track of the maximum cavitation. However, for simulations of several years, it is normally advisable to allow recovery. If `cavitation = "annual"`, PLC_i values are set to zero at the beginning of each year, assuming that embolized plants overcome the conductance loss by creating new xylem tissue. Finally, if `cavitationRefill = "rate"` the model simulates stem refilling at daily steps as a function of symplasmic water potential. First, a daily recovery rate (r_{refill} ; in $cm^2 \cdot m^{-2} \cdot day^{-1}$) is estimated as a function of $\Psi_{plant,i}$:

$$r_{refill}(\Psi_{plant,i}) = r_{max,refill} \cdot \max\{0.0, (\Psi_{plant,i} + 1.5)/1.5\} \quad (6.22)$$

Where $r_{max,refill}$ is the control parameter `refillMaximumRate` indicating a maximum refill rate. The right part of the equation normalizes the water potential, so that $r_{refill} = r_{refill,max}$ if $\Psi_{plant,i} = 0$ and $r_{refill} = 0$ if $\Psi_{plant,i} <= -1.5 MPa$. The proportion of conductance lost is then updated using:

$$PLC_i = \max\{0.0, PLC_i - (r_{refill}/H_v)\} \quad (6.23)$$

where H_v is the Huber value (sapwood area to leaf area ratio) in units of $cm^2 \cdot m^{-2}$.

Part III

Advanced water balance modelling

Chapter 7

Advanced water balance model

This chapter provides an overview of the advanced water and energy balance model available in `medfate`. It shares some characteristics with the basic water balance model described in previous chapters. Like the previous one, the model is run using function `spwb()`, for a set of days, or function `spwb_day()`, for a single day. Compared to the basic model, the advanced water balance model adds more detail in some processes (radiation transfer, plant hydraulics and photosynthesis) and incorporates other processes that were missing (canopy/soil energy balances and plant water balance). Importantly, two alternative approaches are possible regarding *plant hydraulics* and *stomatal conductance* sub-models:

- If `transpirationMode = "Sperry"`, the sub-models of plant hydraulics and stomatal conductance follow Sperry *et al.* (2017) and were described in detail in De Cáceres *et al.* (2021).
- If `transpirationMode = "Cochard"`, the sub-models of plant hydraulics and stomatal conductance follow SurEau-ECOS and were described in Ruffault *et al.* (2022).

The following model overview is structured as in chapter 3. Process formulations shared with the basic water balance model were presented in previous chapters and will not be repeated. The chapters following this one provide a detailed description of those sub-modules that are exclusive of the advanced model. Whenever necessary, we will detail the difference between the two sub-models of plant hydraulics and stomatal conductance corresponding to De Cáceres *et al.* (2021) and Ruffault *et al.* (2022). We recommend readers to familiarize themselves with basic water balance modelling processes before reading this and the subsequent chapters. In addition, users of this model should have a background on the processes implemented to fully understand and exploit model outputs. Since the scope of this book is focused on the model, interested readers should address relevant eco-physiological bibliography complement the material exposed here.

7.1 Design principles

The model performs soil/plant water balance and soil/canopy energy balance for a given forest stand and for the period corresponding to input weather data. Soil water balance is calculated on a daily step basis whereas energy balances and plant transpiration are determined at subdaily steps (typically 1h steps). The model considers only the vertical spatial dimension of the stand, and not the horizontal distribution of plants within it. Still, the forest stand is divided into: (a) canopy vertical layers; (b) woody plant cohorts of different species and a herbaceous layer.

Hydrological processes are the same as in chapter 3. Thus, the model includes water interception loss (Gash *et al.* 1995), plant transpiration, evaporation from soil (Ritchie 1972), transpiration of the herbaceous layer and partition between infiltration and runoff (Boughton 1989). Infiltration is partitioned among soil layers, depending on macroporosity, and water exceeding soil water holding capacity is lost via deep drainage. Snow pack dynamics can also be simulated.

A great deal of the complexity of the advanced model concerns the estimation of woody plant transpiration and photosynthesis. The model simulates stomatal regulation of gas exchange separately for sunlit and shade leaves at sub-daily steps, involving detailed calculations of plant hydraulics, leaf energy balance and photosynthesis. Compared to the basic water balance model of chapter 3, the higher level of complexity of the advanced model allows a more precise estimation of carbon and water flows, as well as a more nuanced evaluation of plant water status.

To calculate radiation balance and incident PAR at the leaf level, the leaf area of each plant cohort is divided into vertical layers according to its crown limits. Photosynthesis and transpiration of sunlit and shade leaves are treated separately (De Pury & Farquhar 1997). Given an input direct and diffuse short-wave irradiance at the top of the canopy, the amount of light absorbed by sunlit leaves, shade leaves and the soil follows the model of Anten & Bastiaans (2016). The scheme for long-wave radiation transfer between the atmosphere, the different canopy layers and the soil follows the SHAW model (Flerchinger *et al.* 2009).

Energy balances are conducted at two levels: the canopy (or canopy layers) level and the leaf level. The model keeps track of temperature variation within the canopy (i.e. canopy energy balance) and in the uppermost soil surface (i.e. soil energy balance) as the result of energy exchanges between them and with the atmosphere. Heat conduction also occurs among soil layers. Energy balance can be performed assuming a single canopy layer, with soil and canopy energy balance equations being similar to those of Best *et al.* (2011) for JULES. Alternatively, energy balance can be conducted for each vertical layer following Bonan *et al.* (2014). In this case, the model allows considering within-canopy gradients in temperature, water vapor pressure, and CO_2 as well as the turbulent transport of heat, water vapor and CO_2 between canopy layers (Ma & Liu 2019), determined using a canopy turbulence model by Katul *et al.* (2004).

The model also performs the energy balance at the leaf level to determine transpiration. At this scale, radiation inputs include shortwave radiation from the atmosphere absorbed by the leaf and the net balance of long-wave radiation. Leaf temperature is determined assuming that the temperature of the surrounding air is that of the canopy (or canopy layer). After determining stomatal regulation, the model up-scales the transpiration flux to the canopy (or canopy layer) scale and the corresponding latent heat is used to complete the calculation of the energy balance at the canopy (or canopy layer) level. Latent heat fluxes from evaporation from the soil and evaporation of intercepted rainfall are also included in the canopy/soil energy balances.

Plant hydraulics and stomatal regulation follow different designs depending on the sub-model employed:

- If `transpirationMode = "Sperry"`, plant hydraulics follows the steady-state approach of Sperry *et al.* (1998) model, and stomatal regulation follows the profit maximization approach of Sperry *et al.* (2017). A variant of this sub-model (i.e., if `capacitance = TRUE`) can also consider an explicit water balance within plant compartments De Cáceres *et al.* (2021). In this case, the amount of water within plant compartments is explicitly tracked and the rate of extraction of soil water can be decoupled from the rate of transpiration.
- If `transpirationMode = "Cochard"`, plant water compartments are always included, and water flows follow Darcy's law, as described in Ruffault *et al.* (2022). In this case, stomatal regulation follows a Jarvis-type model.

Like in the basic water model, the advanced model assumes by default that soil moisture under all plant cohorts is the same (i.e. water sources corresponding to vertical soil layers are shared among cohorts). However, variations in soil moisture beneath plant cohorts (and, implicitly, horizontal variation of soil moisture) can be simulated. In this case, hydrological processes are replicated for the fraction of soil corresponding to each cohort. Transpiration of each plant cohort depends on the soil moisture beneath itself and, depending on root overlap, on the soil moisture beneath other plant cohorts.

7.2 State variables

The following are state variables in the model under all simulations:

- Cumulative degree days (a) to budburst, (b) to complete unfolding or (c) to senescence ($S_{eco,d}$, $S_{unf,d}$ or $S_{sen,d}$; all in $^{\circ}\text{C}$), are tracked by the model to determine leaf phenological status (see 4.1).
- Daily soil moisture content dynamics on each layer s are tracked using $W_s = \theta_s(\Psi_s)/\theta_{fc,s}$, the proportion of soil moisture in relation to field capacity, where moisture at field capacity, $\theta_{fc,s}$, is assumed to correspond to $\Psi_{fc} = -0.033$ MPa. Note that W_s values larger than one are possible if the soil is between field capacity and saturation (which can happen if deep drainage is not allowed).
- The air temperature within the canopy (T_{can} ; $^{\circ}\text{C}$) and the temperature of each soil layer s ($T_{soil,s}$; $^{\circ}\text{C}$) are tracked for every subdaily step.

Additional state variables depend on the activation of specific control flags (see subsection 7.5.3):

- If plant capacitance effects are considered, the model also tracks $\Psi_{apo,stem,i}$, the water potential of the stem apoplastic compartment, as well as the water potential of stem and leaf symplastic compartments, $\Psi_{symp,stem,i}$ and $\Psi_{symp,leaf,i}$ respectively.
- If cavitation is not completely reversible, the proportion of stem hydraulic conductance loss for each plant cohort i , $PLC_{stem,i}$, becomes a state variable.
- If plant water pools are considered, the model also tracks $W_{i,s}$, the proportion of soil moisture in relation to field capacity for layer s within the areal fraction of stand covered by cohort i .
- If multi-layer canopy energy balance is activated, the model tracks for each vertical layer $j \in \{1, l\}$:
 - Air temperature, $T_{air,j}$
 - Water vapor pressure, $e_{air,j}$
 - CO_2 concentration, $C_{air,j}$.

7.3 Water and energy balances

7.3.1 Water balance

Daily variations in soil water content (ΔV_{soil} in mm) can be summarized as:

$$\Delta V_{soil} = Pr + Sm - In - Ru - Dd - Es - Tr_{herb} - Ex_{woody} \quad (7.1)$$

where Pr is precipitation as rainfall, Sm is water reaching the soil from snow melt (if snow-pack dynamics are considered), In is the interception loss (i.e., water evaporated after being intercepted by the canopy), Ru is surface runoff, Dd is deep drainage (i.e. water percolated to layers beyond soil depth), Es is evaporation from soil surface, Tr_{herb} is the transpiration of the herbaceous layer and Ex_{woody} is soil water extraction done by woody plants.

Although the advanced water balance model includes hydraulic redistribution, this does not change the total amount of water in the soil, so the water balance equation is very similar to that of the basic water balance model (eq.(3.1)). Soil water extraction (Ex_{woody}) and plant transpiration (Tr_{woody}) will be equal if capacitance effects are not considered. Otherwise the amount of water stored in the woody plant tissues changes according to:

$$\Delta V_{plant} = Ex_{woody} - Tr_{woody} \quad (7.2)$$

While the above describes the default water balance equation, variations can occur depending on specific control flags:

- If snow dynamics are considered, the water balance of the snow pack is defined as:

$$\Delta S_{snow} = Ps - Sm \quad (7.3)$$

where Ps is precipitation as snowfall and Sm is snow melt. Evaporation from bare soil cannot occur if there is a snow pack over the soil surface (i.e., if $S_{snow} > 0$ then $Es = 0$).

- b. If plant water pools are considered, the water balance equation applies not only to the soil of the overall stand but also to the soil beneath each plant cohort. The fraction of stand area covered by each cohort is used to keep the water balance at the two scales aligned. Moreover, the water balances of soils beneath the different plant cohort are more or less correlated depending on root overlap.

Unlike the basic water balance, in this model water balance components contributing to latent heat (E_s , Tr_{woody} , Tr_{herb} , In and Sm) are included in the energy balance equations of the canopy and soil, therefore coupling water and energy balances.

7.3.2 Single-layer canopy energy balance

For the sake of performing energy balance, the canopy can be considered a single layer that exchanges energy with the soil and the atmosphere (Best *et al.* 2011). First, it absorbs shortwave radiation from the atmosphere ($K_{abs,can}$) and long-wave radiation resulting from net radiation balance ($L_{net,can}$). Other energy fluxes considered are convective exchanges between the canopy and atmosphere ($H_{can,atm}$) and between the canopy and the soil ($H_{can,soil}$). Finally, energy is released from the canopy to the atmosphere through latent heat (LE_{can}) produced via transpiration (Tr) and evaporation of water intercepted by the canopy (In). Combining all energy flows (all of them in $W \cdot m^{-2}$), the instantaneous energy balance equation for the canopy is:

$$TC_{can} \cdot \frac{\delta T_{can}}{\delta t} = K_{abs,can} + L_{net,can} - LE_{can} - H_{can,atm} - H_{can,soil} \quad (7.4)$$

where TC_{can} is the canopy thermal capacitance (in $J \cdot m^{-2} \cdot K^{-1}$).

7.3.3 Multiple-layer canopy energy balance

Alternatively, the model allows performing an energy balance for each vertical layer in the canopy, including its heat storage capacity (Ma & Liu 2019). The energy balance equation for canopy layer j is:

$$TC_j \cdot \frac{\delta T_{air,j}}{\delta t} = K_{abs,j} + L_{net,j} - LE_j + H_j \quad (7.5)$$

where $K_{abs,j}$ is the short-wave radiation absorbed by leaves in the layer, $L_{net,j}$ is the net long-wave radiation balance of the layer, LE_j is the latent heat flux and H_j is the sensible heat flux, which includes heat coming from leaves or soil surfaces as well as turbulent heat transport from adjacent layers. Finally, TC_j is the thermal capacitance (in $J \cdot m^{-2} \cdot K^{-1}$) of canopy layer j .

7.3.4 Soil energy balance

Like the canopy, the uppermost soil layer absorbs short-wave radiation from the atmosphere ($K_{abs,soil}$) and has a net long-wave radiation balance ($L_{net,soil}$). The uppermost soil layer also exchanges heat with the canopy ($H_{can,soil}$) and releases energy in the form of latent heat (LE_{soil}), which includes snow melt (Sm) and evaporation from the soil surface (E_s). Finally, the soil layer exchanges thermal energy by conduction with the layer below ($G_{1,2}$) The energy balance equation for the uppermost soil layer is:

$$TC_{soil,1} \cdot \frac{\delta T_{soil,1}}{\delta t} = K_{abs,soil} + L_{net,soil} + H_{can,soil} - LE_{soil} - G_{1,2} \quad (7.6)$$

where $TC_{soil,1}$ is the thermal capacitance of soil layer 1 (the uppermost layer).

Instantaneous soil temperature changes on each soil layer depend on the balance between upward and downward energy flows. The downward energy flow for the uppermost soil layer depends on its energy exchanges with the canopy and the atmosphere, as shown in eq. (7.6), whereas the remaining energy flows

are generated by heat conduction and depend on the temperature gradients between soil layers. The energy balance for a soil layer $s > 1$ is:

$$TC_{soil,s} \cdot \frac{\delta T_{soil,s}}{\delta t} = G_{s-1,s} - G_{s,s+1} \quad (7.7)$$

where $G_{s-1,s}$ is the energy inflow from layers above, $G_{s,s+1}$ is the energy outflow towards layers below and $TC_{soil,s}$ is the thermal capacitance of soil layer s (in $J \cdot m^{-2} \cdot K^{-1}$).

Unlike Best *et al.* (2011), the model considers the snow pack and the uppermost soil layer as a single layer in terms of energy exchange, but treats this layer differently depending on whether snow is present or not. The snow pack absorbs shortwave radiation (section 5.2) and snow melt is included as latent heat in the energy balance, but the snow layer does not exchange long-wave radiation with the soil nor the atmosphere. $K_{abs,soil}$ is set to zero when snow pack is present (because shortwave radiation is absorbed by snow), but the soil layer still exchanges long-wave radiation and convective energy with the canopy and the atmosphere. In addition, remember that bare soil evaporation does not occur when the snow pack is present (section 5.5).

7.4 Process scheduling

For every day to be simulated, the model performs the following steps:

1. Update leaf area values according to the phenology of species (sections 4.1).
2. If snow dynamics are included, increase snow pack from snow precipitation (P_s) and decrease it following snow melt (Sm) (section 5.2).
3. Determine rainfall interception loss (In) and net rainfall (Pr_{net}) (section 5.3).
4. Increase soil water content due to net rainfall (Pr_{net}), surface runon (Ro) and snow melt (Sm), after accounting for surface runoff (Ru) and deep drainage (Dd) (section 5.4).
5. Decrease soil water content due to bare soil evaporation (Es), if snow is not present (section 5.5), and herbaceous transpiration (Tr_{herb} ; section 5.6).
6. Determine sub-daily air temperature and direct/diffuse irradiance variations (chapter 8).
7. Determine shortwave radiation absorbed by plant cohorts and the soil at sub-daily steps (sections 9.1.3 and 9.1.4).
8. If `transpirationMode = "Sperry"`, update the water supply function of each plant cohort, according to the hydraulic model and the current soil water potential (chapter 10). If plant capacitance effects are considered, the supply function involves only the soil-to-stem part of the hydraulic pathway. Otherwise, it describes the complete soil-to-leaf pathway.
9. Determine instantaneous plant cohort transpiration, photosynthesis rates and close soil/canopy energy balance at sub-daily steps (chapters 12 and 13). This involves the following actions for each sub-daily step:
 - a. Determine long-wave radiation exchange between soil, canopy layers and atmosphere (section 9.2).
 - b. If `transpirationMode = "Sperry"`, for each woody cohort:
 - i. If plant capacitance effects are considered update the stem-to-leaf hydraulic supply function.
 - ii. Calculate leaf energy balance and photosynthesis (for shade and sunlit leaves of each plant cohort) corresponding to each transpiration value of the supply function (section 11.1). If plant capacitance effects are considered this concerns the stem-to-leaf hydraulic supply function.
 - iii. Determine stomatal conductance, transpiration and photosynthesis on shade and sunlit leaves of each plant cohort according to Sperry's (2017) profit maximization strategy (section 12.1.2).
 - iv. If plant capacitance effects are considered perform water balance of plant compartments at high temporal resolution (1 second steps), while re-estimating the flow from the soil to the stem at each step according to changes in stem water potential (see 10.5).
 - c. If `transpirationMode = "Cochard"`, for each plant cohort repeat the following in small sub-steps (e.g. 10 min):
 - i. Determine long-wave radiation exchange between soil, canopy layers and atmosphere (section 9.2).
 - ii. Calculate leaf energy balance and photosynthesis (for shade and sunlit leaves of each plant cohort) corresponding to each transpiration value of the supply function (section 11.1). If plant capacitance effects are considered this concerns the stem-to-leaf hydraulic supply function.
 - iii. Determine stomatal conductance, transpiration and photosynthesis on shade and sunlit leaves of each plant cohort according to Sperry's (2017) profit maximization strategy (section 12.1.2).
 - iv. If plant capacitance effects are considered perform water balance of plant compartments at high temporal resolution (1 second steps), while re-estimating the flow from the soil to the stem at each step according to changes in stem water potential (see 10.5).

- i. Perform energy balance of sunlit and shade leaves (section 11.1), according to the current transpiration flow.
 - ii. Re-estimate leaf and stem cuticular transpiration.
 - iii. Re-estimate stomatal regulation, transpiration and photosynthesis for sunlit and shade leaves.
 - iv. Update water potentials, flows and conductances across the hydraulic network.
 - d. Scale transpiration (Tr_{woody}) and net photosynthesis (A_n) from sunlit/shade leaves to the plant cohort level (section 12.3).
 - e. Scale water extraction from soil layers (Ex_{woody}) to the woody plant cohort level.
 - f. Complete energy balance of the canopy and the soil, after translating plant transpiration, evaporation from bare soil, snow melt and interception loss to latent heat and calculating convective heat exchange for both the canopy and the soil. Alternatively, perform energy balance for individual canopy layers, while including turbulent heat exchange and turbulent transport of water vapor and CO_2 between layers (chapter 13).
10. Determine day-level photosynthesis and transpiration from instantaneous rates and update soil moisture (section 14.1), effectively closing the soil water balance at the daily scale.
11. Determine drought stress for each woody plant cohort (section 14.3).

Details of the formulation of each of these processes are provided within the referred chapters and sections (some processes have been already described in previous chapters/sections).

7.5 Inputs and outputs

7.5.1 Soil, vegetation and meteorology

Soil

Soil input requirements are the same as for the basic water balance model and were fully described in section 2.3. Genuchten (1980) water retention curves are forced for this model, because rhizosphere conductance is modelled using van Genuchten hydraulic equations (see subsection 10.2).

Vegetation

Vegetation input requirements were fully described in section 2.4. Trees and shrubs do not need to be characterized with different variables in soil water balance calculations, since both kinds of cohort have a LAI value. In most cases, users only need to estimate the leaf area index corresponding to live leaves, i.e. LAI^{live} , because normally at the starting point all leaves are expanded (i.e. $LAI^\phi = LAI^{live}$) and one can assume no dead leaves in the canopy (i.e., $LAI^{dead} = 0$). Vegetation characteristics stay constant during simulations using function `spwb()`, although the actual expanded leaf area (LAI^ϕ) and dead leaf area may vary if the species is winter deciduous.

Meteorology

The minimum weather variables required to run the model are min/max temperatures (T_{min} and T_{max}), min/max relative humidity (RH_{min} and RH_{max}), precipitation (P) and solar radiation (Rad). Wind speed (u) is also needed, but the user may use missing values if not available (a default value will be used in this case). Wind speed is assumed to have been measured at a specific height above the canopy (by default at 2 m). Atmospheric CO_2 concentration (C_{atm}) may also be specified, but if missing a default constant value is assumed, which is taken from the control parameters. Definitions and units of these variables were given in section 2.5.

7.5.2 Vegetation functional parameters

The advanced water balance model requires many functional parameters to be specified for plant cohorts. A first set of parameters refers to leaf phenology (`paramsPhenology`):

Symbol	Units	R	Description
		PhenologyType	phenology type (oneflush-evergreen, progressive-evergreen, winter-deciduous, winter-semideciduous)
<i>LD</i>	years	LeafDuration	Range duration of leaves (in years).
$S_{eco,i}^*$	°C	Sgdd	Degree days corresponding to leaf budburst (see section 4.1.2).
T_{eco}	°C	Tbgdd	Base temperature for the calculation of degree days to leaf budburst (see section 4.1.2).
$S_{sen,i}^*$	°C	Ssen	Degree days corresponding to leaf senescence (see section 4.1.3).
Ph_{sen}	hours	Phsen	Photoperiod corresponding to start counting senescence degree-days (see section 4.1.3).
T_{sen}	°C	Tbsen	Base temperature for the calculation of degree days to leaf senescence (see section 4.1.3).

A second set of functional parameters relate to light extinction and water interception (`paramsInterception`):

Symbol	Units	R	Description
$k_{PAR,i}$	(unitless)	kPAR	Extinction coefficient for diffuse PAR
$\alpha_{SWR,i}$	[0-1]	alphaSWR	Short-wave radiation leaf absorbance coefficient
$\gamma_{SWR,i}$	[0-1]	gammaSWR	Short-wave radiation leaf reflectance (albedo)
$s_{water,i}$	mm H ₂ O LAI ⁻¹		Crown water storage capacity (i.e. depth of water that can be retained by leaves and branches) per LAI unit

A third set includes parameters related to plant anatomic and morphological attributes (`paramsAnatomy`):

Symbol	Units	R param	Description
$H_{med,i}$	cm	Hmed	Median plant height
$1/H_{v,i}$	$m^2 \cdot m^{-2}$	A12As	Ratio of leaf area to sapwood area
LW_i	cm	LeafWidth	Leaf width
SLA_i	$m^2 \cdot kg^{-1}$	SLA	Specific leaf area
$\rho_{leaf,i}$	$g \cdot cm^{-3}$	LeafDensity	Leaf tissue density
$\rho_{wood,i}$	$g \cdot cm^{-3}$	WoodDensity	Wood tissue density
$\rho_{fineroot}$	$g \cdot cm^{-3}$	FineRootDensity	Fine root tissue density
SRL	$cm \cdot g^{-1}$	SRL	Specific root length
RLD	$cm \cdot cm^{-3}$	RLD	Fine root length density (i.e. density of root length per soil volume)
$r_{6.35}$		r635	Ratio between the weight of leaves plus branches and the weight of leaves alone for branches of 6.35 mm

A fourth set of parameters are related to transpiration and photosynthesis (`paramsTranspiration`):

Symbol	Units	R param	Description
$g_{swmin,i}$	$mol H_2O \cdot s^{-1} \cdot m^{-2}$	Gwmin	Minimum stomatal conductance to water vapour

Symbol	Units	R param	Description
$g_{swmax,i}$	$mol\ H_2O \cdot s^{-1} \cdot m^{-2}$	Gwmax	Maximum stomatal conductance to water vapour
$J_{max,298,i}$	$\mu mol\ e \cdot m^{-2} \cdot s^{-1}$	Jmax298	Maximum rate of electron transport at 298K
$V_{max,298,i}$	$\mu mol\ CO_2 \cdot s^{-1} \cdot m^{-2}$	Vmax298	Rubisco's maximum carboxylation rate at 298K
$K_{stem,max,ref}$	$kg \cdot s^{-1} \cdot m^{-1} \cdot MPa^{-1}$	Kmax_stemxylem	Maximum stem sapwood reference conductivity per leaf area unit
$K_{root,max,ref}$	$kg \cdot s^{-1} \cdot m^{-1} \cdot MPa^{-1}$	Kmax_rootxylem	Maximum root sapwood reference conductivity per leaf area unit
$k_{leaf,max,i}$	$mmol \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$	VCleaf_kmax	Maximum leaf conductance (per leaf area unit)
$k_{stem,max,i}$	$mmol \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$	VCstem_kmax	Maximum stem conductance (per leaf area unit)
$k_{root,max,i,s}$	$mmol \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$	VCroot_kmax	Maximum root conductance (per leaf area unit) for each soil layer
$k_{rhizo,max,i,s}$	$mmol \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$	VGrhizo_kmax	Maximum hydraulic conductance of the rhizosphere for each soil layer
$c_{leaf,i}$, $d_{leaf,i}$	(unitless), MPa	VCleaf_c, VCleaf_d	Parameters of the vulnerability curve for leaves
$c_{stem,i}$, $d_{stem,i}$	(unitless), MPa	VCstem_c, VCstem_d	Parameters of the vulnerability curve for stem xylem
$c_{root,i}$, $d_{root,i}$	(unitless), MPa	VCroot_c, VCroot_d	Parameters of the vulnerability curve for root xylem

It is important to mention that stem- and root-level hydraulic conductances (i.e. $k_{stem,max}$ and $k_{root,max}$) are not given as inputs by the user, but are the result from scaling tissue-level conductivity parameters (i.e. $K_{stem,max,ref}$ and $K_{root,max,ref}$) according to plant size (see A.4.1 and A.4.3 for details).

A fifth (final) set of parameters are related to water storage and water relations in plant tissues (`paramsWaterStorage`):

Symbol	Units	R	Description
$LFMC_{max}$	%	maxFMC	Maximum live fuel moisture content, corresponding to fine fuels (< 6.35 mm twigs and leaves).
$\epsilon_{leaf,i}$	MPa	LeafEPS	Modulus of elasticity of leaves
$\epsilon_{stem,i}$	MPa	StemEPS	Modulus of elasticity of symplastic xylem tissue
$\pi_{0,leaf,i}$	MPa	LeafPIO	Osmotic potential at full turgor of leaves
$\pi_{0,stem,i}$	MPa	StemPIO	Osmotic potential at full turgor of symplastic xylem tissue
$f_{apo,leaf,i}$	[0-1]	LeafAF	Apoplastic fraction in leaf tissues
$f_{apo,stem,i}$	[0-1]	StemAF	Apoplastic fraction in stem tissues
$V_{leaf,i}$	$l \cdot m^{-2}$	Vleaf	Leaf water capacity per leaf area unit
$V_{sapwood,i}$	$l \cdot m^{-2}$	Vsapwood	Sapwood water capacity per leaf area unit

Vegetation functional attributes are normally filled for each cohort by function `spwbInput()` or

`forest2spwbInput()` from species identity. However, different parameters can be specified for different cohorts of the same species if desired (see section 2.4.8).

7.5.3 Control parameters

Control parameters modulate the overall behaviour of water balance simulations (see section 2.6). There are three **key control parameters**:

1. `transpirationMode` defines the transpiration model. If `transpirationMode = "Sperry"` or `transpirationMode = "Cochard"` then the advanced water balance model is run with the corresponding plant hydraulics and stomatal regulation sub-models. In advanced water balance models, the water retention curve model is forced to be that of Van Genuchten, i.e. `soilFunctions = "VG"` (see section 2.3.2).
2. `capacitance` defines whether plant capacitance (hence, plant water balance) is considered (only relevant for `transpirationMode = "Sperry"`). By default it is set to `capacitance = FALSE`.
3. `ndailysteps` defines the number of daily steps. By default `ndailysteps = 24`, so that steps are hours, but users can increase the temporal resolution, for example to half hour substeps by setting `ndailysteps = 48`.
4. `multiLayerBalance` is a boolean flag to indicate that multi-layer energy balance is desired (by default `multiLayerBalance = FALSE`).
5. `nsubsteps` defines the number of substeps for multi-layer energy balance calculations. By default `nsubsteps = 3600`, so that substeps are 1 sec if `ndailysteps = 24`.

The other control parameters relevant for the advanced water balance model are:

- `defaultWindSpeed [= 2.5]`: Default value for wind speed (in $m \cdot s^{-1}$) when this is missing (only used for leaf fall, see section 4.1).
- `defaultCO2 [=386]`: Default atmospheric (abovecanopy) CO_2 concentration (in micromol $CO_2 \cdot mol^{-1} = ppm$). This value will be used whenever CO_2 concentration is not specified in the weather input.
- `snowpack [= TRUE]`: Whether dynamics of snow pack are included (see section 5.2).
- `leafPhenology [= TRUE]`: Whether leaf phenology is simulated for winter-deciduous species (see section 4.1). If `FALSE` then all species are assumed to be evergreen.
- `rockyLayerDrainage [= TRUE]`: Whether water exceeding the field capacity of the deepest layer can be drained via macropore flow towards a non-reachable compartment, only for layers with > 95% rocks (see section 5.4).
- `windMeasurementHeight [= 200]`: Distance (in cm) above the top of the canopy corresponding to the measurement of wind speed.
- `boundaryLayerSize [= 2000]`: Size of the canopy boundary layer (in cm). The boundary layer causes the model to define vertical layers above the top of the canopy. The parameter is only relevant when multi-layer canopy energy balance is performed.
- `unlimitedSoilWater [=FALSE]`: Boolean flag to indicate the simulation of plant transpiration assuming that soil water is always at field capacity.
- `plantWaterPools [=FALSE]`: Boolean flag to indicate the simulation of water balance assuming that each cohort has its own water pools.
- `cuticularTranspiration [=TRUE]`: Boolean flag to indicate the simulation of cuticular transpiration (at night or when stomata are fully closed) according to minimum stomatal conductance (see section 12.1.2).
- `thermalCapacityLAI [= 1000000]`: Canopy thermal capacitance per LAI unit (see section 13.1.3).
- `verticalLayerSize [= 100]`: The size of vertical layers (in cm) for leaf area distribution and light extinction calculations (see section 2.4.5).
- `cavitationRefill [= "total"]`: Controls recovery from cavitation. If "total" cavitation is completely reversible. If "none" the model operates in a irreversible cavitation mode. If "annual" plants

are assumed to recover from previous drought at the first day of the year and, finally, if "rate" cavitation recovery follows a daily recovery rate (see section 12.4).

- **refillMaximumRate** [= 0.05]: Maximum rate of conduit refilling (see section 12.4).
- **klatstem** [= 0.01]: Stem symplastic-apoplastic lateral conductance (in $\text{mmol} \cdot \text{s}^{-1} \cdot \text{um}^{-2} \cdot \text{MPa}^{-1}$). Only relevant when **capacitance** = TRUE (see section 10.5).
- **klatleaf** [= 0.01]: Leaf symplastic-apoplastic lateral conductance (in $\text{mmol} \cdot \text{s}^{-1} \cdot \text{um}^{-2} \cdot \text{MPa}^{-1}$). Only relevant when **capacitance** = TRUE (see section 10.5).
- **numericParams**: A list with params for numerical approximation routines.

In addition, a number of control parameters are used to modify how hydraulic parameters are estimated when calling function **spwbInput()**:

- **taper** [= TRUE]: Whether taper of xylem conduits is accounted for when estimating aboveground stem conductance from xylem conductivity.
- **fracRootResistance** [= 0.40]: Fraction of plant total resistance (leaf+stem+root) that corresponds to root system.
- **averageFracRhizosphereResistance** [= 0.15]: Fraction to total continuum (stem+root+rhizosphere) resistance that corresponds to rhizosphere (averaged across soil water potential values).

7.5.4 Model output

7.5.4.1 Daily output

Functions **spwb()** and **pwb()** with **transpirationMode** = "Sperry" return a list of a class with the same name. The first four elements of this list (i.e., **latitude**, **topography**, **weather** and **spwbInput**) are simply copies of model inputs. The next element is **spwbOutput**, which contains the state of the input object at the end of the simulation (this can be used to perform further simulations starting with current values of state variables). The remaining elements are:

Element	Description
WaterBalance	Climatic input and water balance flows (i.e. net precipitation, infiltration, runoff, transpiration...). All of them in $mm = l \cdot m^{-2}$.
EnergyBalance	Energy balance components (instantaneous values) for the soil and the canopy
Temperature	Minimum and maximum temperature values for topsoil, canopy and atmosphere
Soil	Soil variables for each soil layer: Moisture relative to field capacity (W_s), water potential (Ψ_s) and volumetric water content (V_s).
Stand	Stand-level variables, such as LAI_{stand}^ϕ , LAI_{stand}^{dead} , the water retention capacity of the canopy (S_{canopy}) or the fraction of light reaching the ground (L_{ground}^{PAR} and L_{ground}^{SWR}).
Plants	Plant-level variables, such as LAI_i^{phi} , plant transpiration, plant water potential, etc.
SunlitLeaves	Leaf-level output variables for sunlit leaves.
ShadeLeaves	Leaf-level output variables for shade leaves.
FireHazard	Fire hazard variables (fuel moisture, rate of spread, fire potentials, etc). Only returned if fireHazardResults = TRUE.

Compared to the output of the basic water balance 3.5.4, new output elements are produced. Many results are given as sub-elements of **Plants**, each of them a data frame with dates in rows and plant cohorts in columns:

Element	Symbol	Units	Description
LAI	LAI_i^ϕ	$m^2 \cdot m^{-2}$	Leaf area index (expanded).

Element	Symbol	Units	Description
LAIlive	LAI_i^{live}	$m^2 \cdot m^{-2}$	Leaf area index (live).
AbsorbedSWR		MJ	Shortwave radiation absorbed per ground area.
AbsorbedLWR		MJ	Longwave radiation absorbed per ground area.
Transpiration Tr_i		mm	Transpiration per ground area.
GrossPhotosynthesis G_i		$g C \cdot m^{-2}$	Gross photosynthesis per ground area.
NetPhotosynthesis N_i		$g C \cdot m^{-2}$	Net photosynthesis per ground area.
dEdP	$dE/d\Psi_i$	$mmol \cdot s^{-1} \cdot m^{-2}$	Derivative of the (aboveground) supply function.
PlantWaterBalance		MPa ⁻¹	
LeafPsiMin	$\Psi_{md,i}$	MPa	Minimum (midday) ‘average leaf’ water potential.
LeafPsiMax	$\Psi_{pd,i}$	MPa	Maximum (predawn) ‘average leaf’ water potential.
StemPsi	$\Psi_{stem,i}$	MPa	Minimum stem water potential.
RootPsi	$\Psi_{rootcrown,i}$	MPa	Minimum root-crown water potential.
RhizoPsi	$\Psi_{rootsurf,i,s}$	MPa	Minimum root surface water potential for each soil layer.
StemPLC	$PLC_{stem,i}$	MPa	Proportion of stem conductance loss.
LeafRWC	$RWC_{leaf,i}$	%	Mean leaf relative water content.
StemRWC	$RWC_{stem,i}$	%	Mean stem relative water content.
LFMC	$LFMC_i$	%	Live fuel moisture content (as percent of dry weight), corresponding to fine fuels (< 6.35 mm twigs and leaves).
PlantStress	DDS_i	[0-1]	Drought stress level suffered by each plant cohort (relative whole-plant conductance).

The following table describes the child elements of `SunlitLeaves` and `ShadeLeaves`, each of them a data frame with dates in rows and plant cohorts in columns:

Element	Symbol	Units	Description
LeafPsiMin	$\Psi_{md,i}^{sunlit}, \Psi_{md,i}^{shade}$	MPa	Minimum (midday) (sunlit or shade) leaf water potential.
LeafPsiMax	$\Psi_{pd,i}^{sunlit}, \Psi_{pd,i}^{shade}$	MPa	Maximum (predawn) (sunlit or shade) leaf water potential.
GW	$g_{w,i}^{sunlit}, g_{w,i}^{shade}$	$mol \cdot s^{-1} \cdot m^{-2}$	Stomatal conductance to water vapour.

The output of simulations can be inspected using `plot`, `shinyplot` and `summary` functions specific to `spwb` objects (examples are given in the corresponding package `vignette`).

7.5.4.2 Sub-daily output

Calling function `spwb_day()` with the advanced water balance model results in sub-daily results for the simulated date. When running the advanced water balance model with function `spwb()` one can request sub-daily results to be included with the function output. This is done via setting `subdailyResults = TRUE` in the control parameter list. In that case, the output list of `spwb()` will include an element `subdaily`, whose sub-elements correspond to the results of `spwb_day()` for each date simulated. Refer to the documentation of functions `spwb_day()` and `transp_transpirationSperry()` or `transp_transpirationCochard()` for details of sub-daily output.

Chapter 8

Sub-daily atmospheric temperature and radiation variation

After estimating leaf phenology (chapter 4) and simulating the main hydrological processes (chapter 5), the advanced water balance model (chapter 7) starts simulating *subdaily* processes, which include energy balances and photosynthesis/transpiration. In order to properly simulate these processes it is important to consider sub-daily variations in temperature and radiation. Here we detail how sub-daily estimates of above-canopy air temperature and atmospheric incoming radiation are derived from daily input values.

8.1 Above-canopy air temperature

Above-canopy air temperature (T_{atm} , in $^{\circ}C$) diurnal variations are modeled assuming a *sinusoidal pattern* with $T_{atm} = T_{\min}$ at sunrise and $T_{atm} = (T_{\min} + T_{\max})/2$ at sunset. Air temperature varies linearly between sunset and sunrise (McMurtrie *et al.* 1990). Sunrise and sunset hours are determined from latitude and sun declination (see section 4.2 of the reference manual for package **meteoland**).

8.2 Incoming diffuse and direct short-wave radiation

Daily global radiation (Rad , in $MJ \cdot m^{-2} \cdot d^{-1}$) is assumed to include both direct and diffuse short-wave radiation (SWR). Using latitude information and whether is a rainy day, this quantity is partitioned into instantaneous direct and diffuse SWR and PAR for different daily substeps. Values of instantaneous direct and diffuse SWR and PAR above the canopy (I_{beam} and I_{dif} , in $W \cdot m^{-2}$) are calculated using the methods described in Spitters *et al.* (1986), which involve comparing daily global radiation with daily potential radiation. All these calculations are performed using routines in package **meteoland** (see details in section 4.6 of the reference manual for this package).

8.3 Incoming long-wave radiation

Once the above-canopy air temperature for a given time step is known, instantaneous long-wave radiation (LWR) coming from the atmosphere (L_{atm} , in $W \cdot m^{-2}$) can be calculated following Campbell & Norman (1998):

$$L_{atm} = \epsilon_a \cdot \sigma \cdot (T_{atm} + 273.16)^{4.0} \quad (8.1)$$

where T_{atm} is air temperature, $\sigma = 5.67 \cdot 10^{-8.0} W \cdot K^{-4} \cdot m^{-2}$ is the Stephan-Boltzmann constant and ϵ_a is the emissivity of the atmosphere, calculated using:

$$\epsilon_a = (1 - 0.84 \cdot c) \cdot \epsilon_{ac} + 0.84 \cdot c \quad (8.2)$$

$$\epsilon_{ac} = 1.72 \cdot \left(\frac{e_{atm}}{T_{atm} + 273.16} \right)^{1/7} \quad (8.3)$$

where e_{atm} is the average daily water vapor pressure (in kPa; see 2.5) and c is the proportion of clouds ($c = 1$ in rainy days and $c = 0$ otherwise).

Chapter 9

Radiation transfer

In the previous chapter we detailed sub-daily variation in temperature and radiation inputs. Here we deal with the absorption of diffuse and direct shortwave radiation through the canopy as well as the long-wave radiation balance for canopy layers and the soil layer. These are necessary to estimate the different components of the canopy and soil energy balances (see eqs. (7.4), (7.5) and (7.6)) as well as to determine leaf energy balance, transpiration and photosynthesis (chapters 11 and 12). Short-wave radiation (SWR) absorption can be determined for all sub-daily steps once the variation of atmospheric inputs are known. In contrast, the long-wave radiation (LWR) balance for any surface depends on its temperature and therefore, net LWR for canopy and soil layers need to be determined at each sub-daily step after closing the energy balance of the previous step (see chapter 13).

The vertical distribution of leaves in the stand is a key element of radiation transfer. As explained in section 2.4.5, the canopy is divided into vertical layers (whose size is determined by the control parameter `verticalLayerSize`), and the expanded and dead leaf area index of each cohort within each layer is determined. Let l be the number of canopy layers, c the number of plant cohorts, and $LAI_{i,j}^{all} = LAI_{i,j}^\phi + LAI_{i,j}^{dead}$ be the leaf area index of cohort i in layer j .

9.1 Short-wave radiation

In section 8.2 we explained how the model estimates instantaneous and direct/diffuse SWR and PAR at the top of the canopy. Given an input direct and diffuse short-wave irradiance at the top of the canopy, the amount of light absorbed by sunlit leaves, shade leaves and the soil follows the model of Anten & Bastiaans (2016), which we detail in the following subsections.

9.1.1 Direct and diffuse irradiance across the canopy

The average irradiance reaching the top of each canopy layer j is calculated separately for direct beam and diffuse radiation (the same equations are valid for SWR or PAR, but with different coefficients):

$$I_{beam,j} = (1 - \gamma_i) \cdot I_{beam} \cdot \exp \left[\sum_{h=j+1}^l \sum_i^c -k_b \cdot \alpha_i^{0.5} \cdot LAI_{i,h}^{all} \right] \quad (9.1)$$

$$I_{dif,j} = (1 - \gamma_i) \cdot I_{dif} \cdot \exp \left[\sum_{h=j+1}^l \sum_i^c -k_{d,i} \cdot \alpha_i^{0.5} \cdot LAI_{i,h}^{all} \right] \quad (9.2)$$

where I_{beam} and I_{dif} are the direct and diffuse irradiance at the top of the canopy (in $W \cdot m^{-2}$), γ_i is the leaf reflectance ($\gamma_{SWR,i}$ is an input parameter, whereas $\gamma_{PAR,i} = 0.8 \cdot \gamma_{SWR,i}$), $k_b = 0.8$ is the extinction

coefficient for direct light, $k_{d,i}$ is the extinction coefficient of cohort i for diffuse light (i.e. $k_{PAR,i}$ or $k_{SWR,i}$) and α_i is the absorbance coefficient ($\alpha_{SWR,i}$ is an input parameter, whereas $\alpha_{PAR,i} = \alpha_{SWR,i} \cdot 1.35$).

9.1.2 Sunlit and shade leaves

It is generally accepted that sunlit and shade leaves need to be treated separately for photosynthesis calculations (De Pury & Farquhar 1997). This separation is necessary because photosynthesis of shade leaves has an essentially linear response to irradiance, while photosynthesis of leaves in sunflecks is often light saturated and independent of irradiance. The proportion of sunlit leaves, i.e. leaves in a canopy layer j that the direct light beams (sunflecks) reach is:

$$f_{SL,j} = \exp \left(\sum_{k>j}^l \sum_i^c -k_b \cdot LAI_{i,k}^{all} \right) \cdot \exp \left(\sum_i^c -k_b \cdot 0.5 \cdot LAI_{i,j}^{all} \right) \quad (9.3)$$

From this we can derive the (expanded) leaf area of each layer that is affected by direct light beams (i.e. the amount of sunlit and shade leaves):

$$LAI_{i,j}^{sunlit} = f_{SL,j} \cdot LAI_{i,j}^\phi \quad (9.4)$$

$$LAI_{i,j}^{shade} = (1 - f_{SL,j}) \cdot LAI_{i,j}^\phi \quad (9.5)$$

As an example we will consider a canopy of one species of $LAI = 2$, divided into ten layers with constant leaf density:

```
LAI = 2
nlayer = 10
LAIlayerlive = matrix(rep(LAI/nlayer,nlayer),nlayer,1)
LAIlayermax = matrix(rep(LAI/nlayer,nlayer),nlayer,1)
LAIlayerdead = matrix(0,nlayer,1)
kb = 0.8
kd_PAR = 0.5
kd_SWR = kd_PAR/1.35
alpha_PAR = 0.9
gamma_PAR = 0.04
gamma_SWR = 0.05
alpha_SWR = 0.7
```

This canopy definition leads to a percentage of the above-canopy irradiance reaching each layer (Anten & Bastiaans 2016). Extinction of direct radiation also defines the proportion of leaves of each layer that are affected by sunflecks (i.e. the proportion of sunlit leaves). Both outcomes are illustrated in the figure below:

9.1.3 Short-wave radiation absorbed by canopy elements

The amount of absorbed diffuse radiation per leaf area unit of cohort i within a given canopy layer j is calculated as:

$$\Phi_{dif,i,j} = I_{dif,j} \cdot k_{d,i} \cdot \alpha_i^{0.5} \exp \left[\sum_h^c -k_{d,h} \cdot \alpha_i^{0.5} \cdot 0.5 \cdot LAI_{h,j}^{all} \right] \quad (9.6)$$

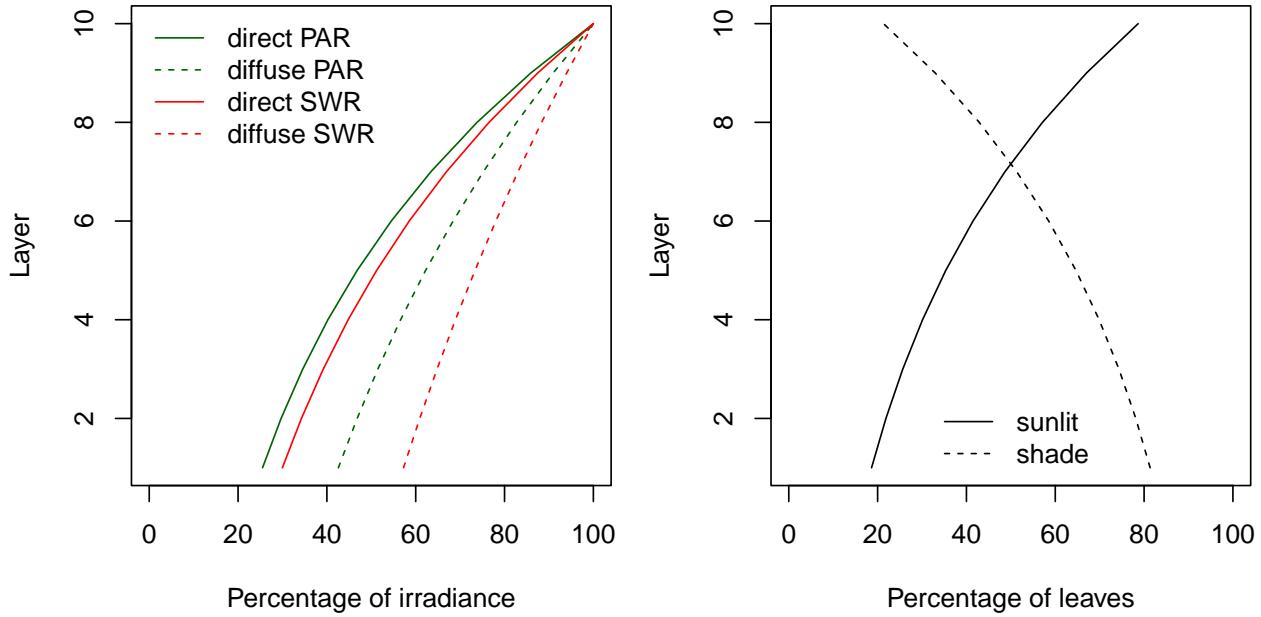


Figure 9.1: The left panel shows irradiance per layer of diffuse and direct PAR and SWR assuming a LAI of 2 equally distributed among layers (see function `light_layerIrradianceFraction()`). The right panel shows the corresponding proportions of sunlit and shade leaves in each layer (see function `light_layerSunlitFraction()`).

The amount of absorbed scattered beam radiation per leaf area unit of cohort i within a given canopy layer j is calculated as:

$$\Phi_{sca,i,j} = I_{b,j} \cdot k_b \cdot (A - B) \quad (9.7)$$

$$A = \alpha_i^{0.5} \cdot \exp \left(\sum_h^c -k_b \cdot \alpha_i \cdot 0.5 \cdot LAI_{h,i}^{all} \right) \quad (9.8)$$

$$B = \frac{\alpha_i}{(1 - \gamma_i)} \cdot \exp \left(\sum_h^c -k_b \cdot 0.5 \cdot LAI_{h,i}^{all} \right) \quad (9.9)$$

Finally, the direct radiation absorbed by a unit of sunlit leaf area of cohort i in a canopy layer j does not depend on the position of the canopy layer and is:

$$\Phi_{dir,i,j} = I_{beam} \cdot \alpha_i \cdot 0.5 / \sin \beta \quad (9.10)$$

where β is the solar elevation angle in radians, which changes throughout the day (see section 4.3 of the reference manual for package **meteoland**). The amount of light absorbed by sunlit/shaded foliage of cohort i in layer j per leaf area unit ($\Phi_{i,j}^{sunlit}$ and $\Phi_{i,j}^{shade}$, respectively) is:

$$\Phi_{i,j}^{sunlit} = \Phi_{dif,i,j} + \Phi_{sca,i,j} + \Phi_{dir,i,j} \quad (9.11)$$

$$\Phi_{i,j}^{shade} = \Phi_{dif,i,j} + \Phi_{sca,i,j} \quad (9.12)$$

Let us show how all this works in an example. Regarding incoming light, we assume the following direct and diffuse irradiance at the top of the canopy:

```
solarElevation = 0.67
SWR_direct = 1100
```

```
SWR_diffuse = 300
PAR_direct = 550
PAR_diffuse = 150
```

Solar elevation is the angle between the sun and the horizon (i.e. the complement of the zenith angle). Under these conditions, and for the same canopy used for fig. 9.1, the amount of PAR and SWR absorbed per unit of leaf area at each canopy layer is (Anten & Bastiaans 2016):

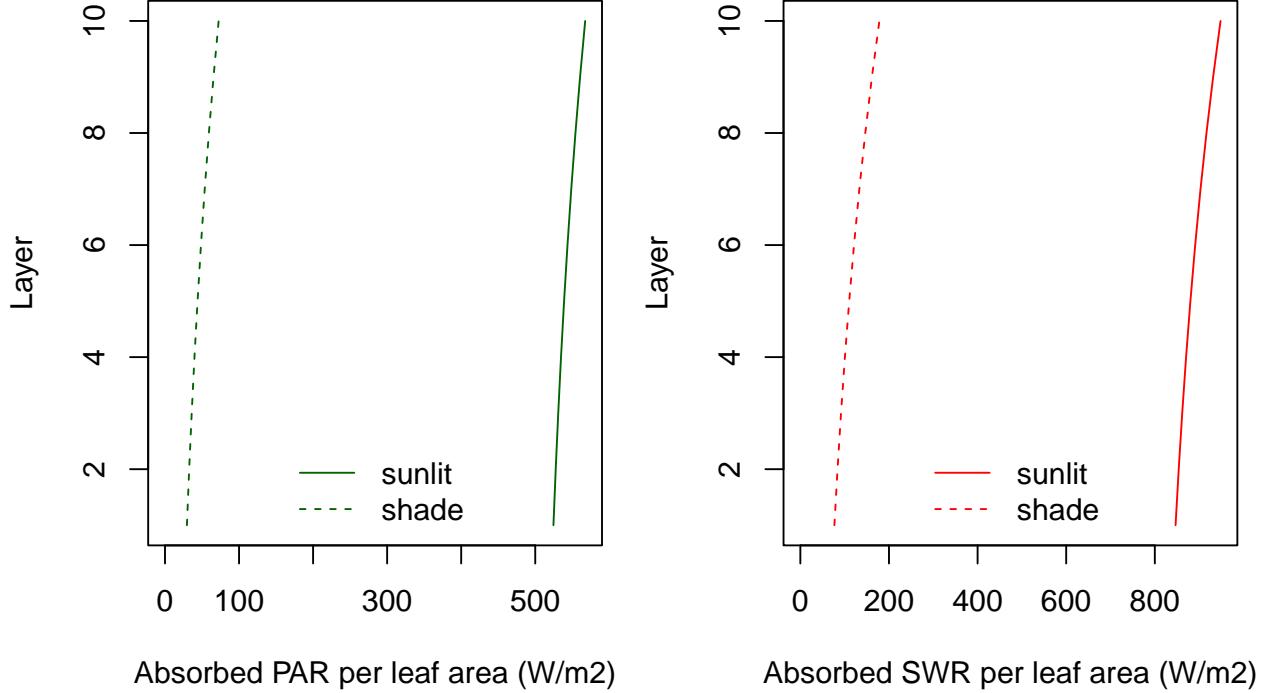


Figure 9.2: PAR (left) and SWR (right) absorbed per unit of sunlit/shade leaf area at each canopy layer ($I_{i,j}^{sunlit}$ and $I_{i,j}^{shade}$, respectively; see function `light_cohortSunlitShadeAbsorbedRadiation()`).

The SWR absorbed per ground area unit by sunlit and shade foliage of cohort i in canopy layer j is:

$$K_{abs,i,j}^{sunlit} = \Phi_{i,j}^{sunlit} \cdot LAI_{i,j}^{sunlit} \quad (9.13)$$

$$K_{abs,i,j}^{shade} = \Phi_{i,j}^{shade} \cdot LAI_{i,j}^{shade} \quad (9.14)$$

The previous quantities can be aggregated across cohorts or layers. The SWR absorbed per ground area unit by cohort i ($K_{abs,i}$) is found using:

$$K_{abs,i} = \sum_j^l K_{abs,i,j}^{sunlit} + K_{abs,i,j}^{shade} = K_{abs,i}^{sunlit} + K_{abs,i}^{shade} \quad (9.15)$$

where $K_{abs,i}^{sunlit}$ and $K_{abs,i}^{shade}$ are the SWR absorbed per ground area unit by sunlit and shade foliage, respectively, of cohort i . Similarly, the SWR absorbed by canopy layer j ($K_{abs,j}$) is found using:

$$K_{abs,j} = \sum_i^c K_{abs,i,j}^{sunlit} + K_{abs,i,j}^{shade} \quad (9.16)$$

and the SWR absorbed by the whole canopy ($K_{abs,ca}$) is:

$$K_{abs,ca} = \sum_j^l K_{abs,j} = \sum_i^c K_{abs,i} = \sum_i^c \sum_j^l K_{abs,i,j}^{sunlit} + K_{abs,i,j}^{shade} \quad (9.17)$$

9.1.4 Short-wave radiation absorbed by the soil

The instantaneous shortwave radiation reaching the soil is calculated separately for direct beam and diffuse radiation:

$$I_{beam,soil} = I_{beam} \cdot \exp \left[\sum_{h=j+1}^l \sum_i^c -k_b \cdot \alpha_i^{0.5} \cdot LAI_{i,h}^{all} \right] \quad (9.18)$$

$$I_{dif,soil} = I_{dif} \cdot \exp \left[\sum_{h=j+1}^l \sum_i^c -k_{d,i} \cdot LAI_{i,h}^{all} \right] \quad (9.19)$$

where I_{beam} and I_{dif} are the direct and diffuse irradiance at the top of the canopy, k_b is the extinction coefficient for direct light ($k_b = 0.8$) and $k_{d,i}$ is the extinction coefficient of cohort i for diffuse SWR. From these, the SWR absorbed by the soil ($K_{abs,soil}$) is found by:

$$K_{abs,soil} = (1 - \gamma_{SWR,soil}) \cdot (I_{beam,soil} + I_{dif,soil}) \quad (9.20)$$

where $\gamma_{SWR,soil} = 0.10$ is the SWR reflectance (10% albedo) of the soil.

9.2 Long-wave radiation

Long-wave radiation (LWR) transfer within the canopy is based on the SHAW model (Flerchinger *et al.* 2009).

9.2.1 Long-wave radiation balance by canopy layers

Assume canopy layers are ordered from $j = 1$ (in contact with the soil) to $j = l$ (in contact with the atmosphere above the boundary layer). The procedure first calculates downward LWR below each canopy layer, $L_{down,j}$, from $j = l$ to $j = 1$. The downward LWR below layer j is calculated as:

$$L_{down,j} = \tau_j \cdot L_{down,j+1} + (1 - \tau_j) \cdot \epsilon_{can} \cdot \sigma \cdot (T_{air,j} + 273.16)^4 \quad (9.21)$$

where $L_{down,j+1}$ is the downward LWR of the layer above (L_{atm} in the case of $j = l$), $T_{air,j}$ is the air temperature of layer j , τ_j is the diffuse transmissivity of layer j , $\sigma = 5.67 \cdot 10^{-8} W \cdot K^{-4} \cdot m^{-2}$ is the Stephan-Boltzmann constant and $\epsilon_{can} = 0.97$ is the canopy emissivity. Diffuse transmissivity values are defined by cohort i and layer j using:

$$\tau_{i,j} = \exp(-k_{LWR} \cdot LAI_{i,j}^{all}) \quad (9.22)$$

where $k_{LWR} = 0.7815$ is the extinction coefficient for LWR. The transmissivity of layer j is simply $\tau_j = \sum_i^c \tau_{i,j}$. Once we know $L_{down,1}$ we can calculate the upwards LWR from the soil ($L_{up,s}$) using:

$$L_{up,soil} = (1 - \epsilon_{soil}) \cdot L_{down,1} + \epsilon_{soil} \cdot \sigma \cdot (T_{soil,1} + 273.16)^4 \quad (9.23)$$

where $T_{soil,1}$ is the temperature of the topmost soil layer and $\epsilon_{soil} = 0.97$ is the soil emissivity. Now we calculate upward LWR from $j = 1$ to $j = J$:

$$L_{up,j} = \tau_j \cdot L_{up,j-1} + (1 - \tau_j) \cdot \epsilon_{can} \cdot \sigma \cdot (T_{air,j} + 273.16)^4 \quad (9.24)$$

where $L_{up,j-1}$ is the downward LWR of the layer below ($L_{up,soil}$ in the case of $j = 1$). Once we have downward and upward LWR fluxes, we can estimate the net LWR absorbed within canopy layer j using:

$$L_{net,j} = \epsilon_{can} \cdot (1 - \tau_j) \cdot (L_{down,j} + L_{up,j} - \sigma \cdot (T_{air,j} + 273.16)^4) \quad (9.25)$$

9.2.2 Net long-wave radiation of the whole canopy and the soil

Net LWR can be aggregated at the scale of the whole canopy. The net LWR of the whole canopy ($L_{net,can}$) is simply the sum across layers of net LWR:

$$L_{net,can} = \sum_j^l L_{net,j} \quad (9.26)$$

whereas the net LWR of the soil ($L_{net,soil}$) is:

$$L_{net,soil} = \epsilon_{soil} \cdot (L_{down,1} - \sigma \cdot (T_{soil,1} + 273.16)^4) \quad (9.27)$$

9.2.3 Long-wave radiation balance by plant cohorts

The net LWR per leaf area unit absorbed within canopy layer j can also be decomposed among plant cohorts. The net LWR of cohort i in layer j is found using the relative proportion of light absorbed by the cohort (i.e. the complement of its diffuse transmissivity):

$$L_{net,i,j} = L_{net,j} \cdot \frac{1 - \tau_{i,j}}{\sum_h 1 - \tau_{h,j}} \quad (9.28)$$

from which we can estimate $L_{net,i}^{sunlit}$ and $L_{net,i}^{shade}$, the average net LWR per leaf area unit for sunlit and shade leaves of cohort i , respectively:

$$L_{net,i}^{sunlit} = \frac{\sum_j^l L_{net,i,j} \cdot LAI_{i,j}^{sunlit}}{\sum_j^l LAI_{i,j}^{sunlit}} \quad L_{net,i}^{shade} = \frac{\sum_j^l L_{net,i,j} \cdot LAI_{i,j}^{shade}}{\sum_j^l LAI_{i,j}^{shade}} \quad (9.29)$$

Chapter 10

Plant hydraulics

We devote this chapter to describe the background necessary to understand how plant hydraulics is implemented in the advanced water balance model of **medfate**. Section 10.1 introduces the hydraulic networks from which to choose, depending on the sub-model of plant hydraulics employed (i.e. either `transpirationMode = "Sperry"` or `transpirationMode = "Cochard"`). After that, in sections 10.2 and 10.3 we present the building blocks of models of plant hydraulics, i.e. how to represent in a model the water content of plant tissues and the water flow through vascular elements. In sections 10.4 and 10.5 we describe in detail the plant hydraulics sub-models presented in De Cáceres *et al.* (2021). Finally, in 10.6 we describe the sub-model of plant hydraulics that was presented in Ruffault *et al.* (2022) and was added to **medfate**.

10.1 Hydraulic networks

The advanced water balance model of **medfate** implements different hydraulic networks depending on the sub-model of plant hydraulics:

- If `transpirationMode = "Sperry"` and `capacitance = FALSE`, the hydraulic network is described using $(S \times 2 + 2 + 1)$ resistance elements, with soil being represented in S different soil layers. For each soil layer there is a rhizosphere element in series with a root xylem element. The S soil layers are in parallel up to the root crown. From there there are two stem xylem segments and a final leaf segment, all in series (Fig. 10.1).
- If `transpirationMode = "Sperry"` and `capacitance = "TRUE"` To consider plant water storage compartments explicitly these need to be combined with supply functions in some way. The previous hydraulic network is divided into two parts: (1) the first is made of $(S \times 2 + 1)$ elements and draws water from the soil, pouring it into the stem xylem; and (2) the second is made of $(1 + 1)$ elements and draws water from stem xylem bringing it up to the leaves. Each part has its own supply function and the stem xylem segments exchange water with water storage compartments (Fig. 10.2).
- If `transpirationMode = "Cochard"`, ... [TO BE WRITTEN]

10.2 Vulnerability curves

Vulnerability curves form the basis of hydraulic calculations. Let us use k to denote hydraulic conductance, i.e. instantaneous flow rate per leaf surface unit and per pressure drop (in $\text{mmol} \cdot \text{s}^{-1} \cdot \text{m}^{-2} \cdot \text{MPa}^{-1}$). Each element of the hydraulic network has a vulnerability curve $k(\Psi)$ that starts at maximum hydraulic conductance ($k_{max} = k(0)$) and declines as water pressure (Ψ) becomes more negative.

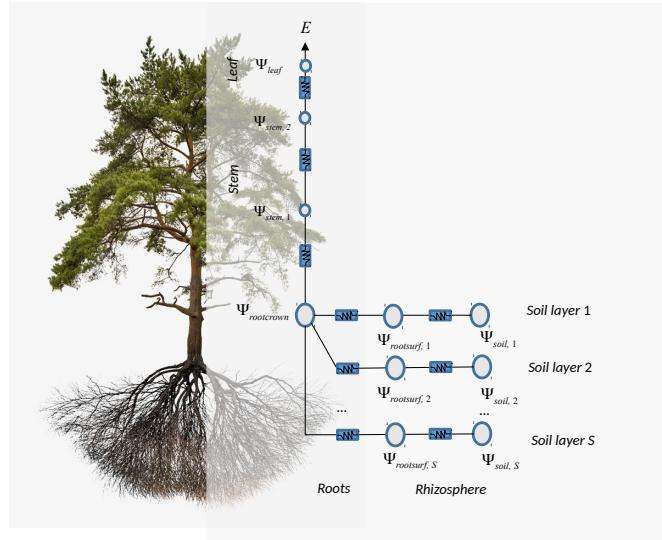


Figure 10.1: Schematic representation of hydraulics in a whole-plant network

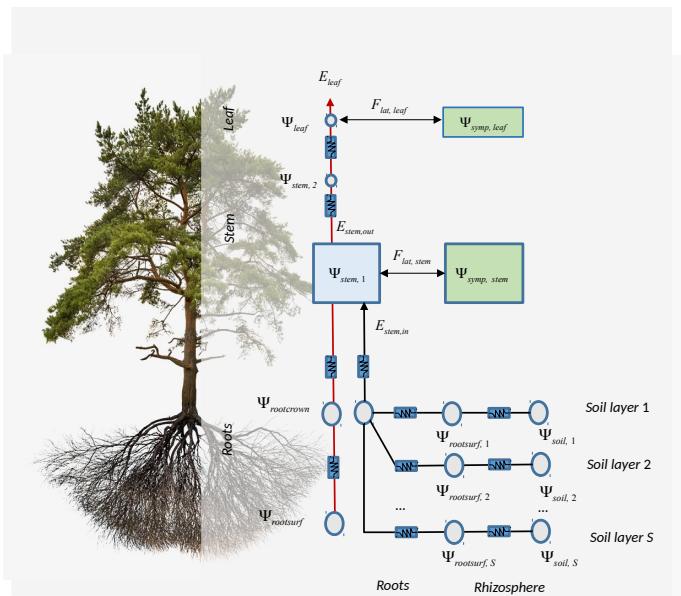


Figure 10.2: Schematic representation of hydraulics in a whole-plant network with water storage compartments

10.2.1 Xylem vulnerability curves

Xylem tissues are assigned a two-parameter Weibull function as the vulnerability curve $k(\Psi)$:

$$k(\Psi) = k_{max} \cdot e^{-((\Psi/d)^c)} \quad (10.1)$$

where k_{max} is the maximum hydraulic conductance, and c and d are species-specific and tissue-specific parameters. Note that parameter d is the water potential (in MPa) at which $k(\Psi)/k_{max} = e^{-1} = 0.367$, i.e. the water potential at which hydraulic conductance is 36.7% of its maximum value. Parameter c controls the shape of the vulnerability curve (*exponential* shape with no threshold has $c \leq 1$; and *sigmoidal* shape with threshold occurs when $c > 1$).

The concept of vulnerability curve can be used to specify the relationship between pressure and conductance in any portion of the flow path. For example, we define the following parameter values for a stem xylem ($k_{stem,max}$ and parameters c_{stem} and d_{stem} of the vulnerability curve):

```
kstemmax = 5.0 # mmol·m⁻²·s⁻¹·MPa⁻¹
stemc = 3
stemd = -3.0 # MPa
```

For root xylem ($k_{root,max}$), we may assume a higher conductance (i.e. higher efficiency) but also higher vulnerability to cavitation (defined by parameters c_{root} and d_{root}):

```
krootmax = 6.6 # mmol·m⁻²·s⁻¹·MPa⁻¹
rootc = 2
rootd = -2.5 # MPa
```

Leaf vulnerability curve $k_{leaf}(\Psi)$ can be modelled using the same equation as for stem xylem:

$$k_{leaf}(\Psi) = k_{leaf,max} \cdot e^{-((\Psi/d_{leaf})^c_{leaf})} \quad (10.2)$$

where $k_{leaf,max}$ is the leaf maximum hydraulic conductance. Values defined below specify higher conductance for leaves but also slightly higher vulnerability:

```
kleafmax = 10
leafc = 2
leafd = -2
```

With these parameter values, the vulnerability curves for root, stem and leaf are (see `hydraulics_xylemConductance()` and `hydraulics_vulnerabilityCurvePlot()`):

The dash-dot line between 0 and $\Psi_{cav} = -2.5$ MPa indicates the modification of the stem xylem vulnerability curve when cavitation has occurred (i.e., previous embolism limits the maximum conductance value), as indicated in Sperry *et al.* (2017). The corresponding proportion of stem conductance loss (PLC_{stem}) can be found using the stem vulnerability curve:

$$PLC_{stem}(\Psi_{cav}) = 1.0 - \frac{k_{stem}(\Psi_{cav})}{k_{stem,max}} = 1.0 - e^{-((\Psi_{cav}/d_{stem})^{c_{stem}})} \quad (10.3)$$

or, in R:

```
1.0 - exp(-(-2.5/stemd)^stemc)
```

```
## [1] 0.4393754
```

Although root xylem are more vulnerable to the formation of emboli for a given potential, it is generally accepted that the less negative potentials of root xylem compared to the stem lead to cavitation occurring more often in the stem. The constrain created by cavitation has an effect on the calculation of the flow rates and derived quantities (see below).

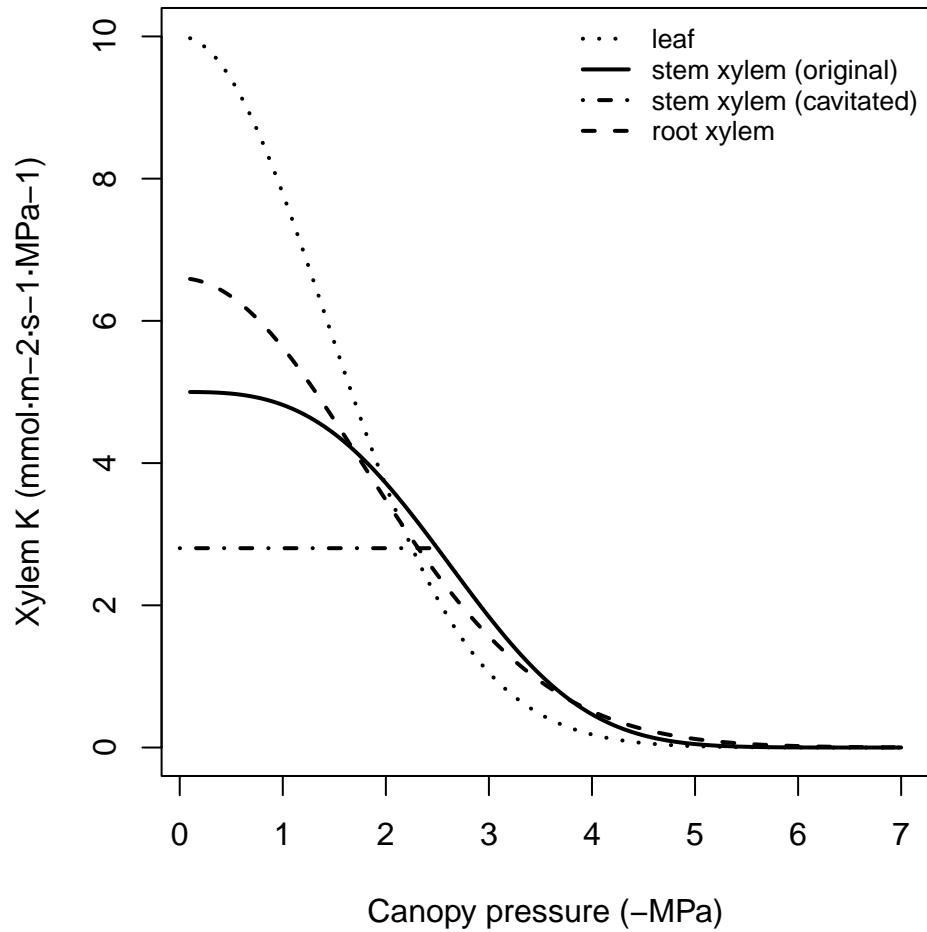


Figure 10.3: Example vulnerability curves corresponding to the parameters defined above for stem, root and leaf segments.

10.2.2 Rhizosphere vulnerability curve

The rhizosphere conductance function $k_{rhizo}(\Psi)$ is modelled as a van Genuchten function (Genuchten 1980) (this choice is the reason why the water retention curve also needs to be modelled with van Genuchten):

$$k_{rhizo}(\Psi) = k_{rhizo,max} \cdot v^{(n-1)/(2 \cdot n)} \cdot ((1-v)^{(n-1)/n} - 1)^2 \quad (10.4)$$

$$v = [(\alpha\Psi)^n + 1]^{-1} \quad (10.5)$$

where $k_{rhizo,max}$ is the maximum rhizosphere conductance, and n and α are texture-specific parameters (Carsel & Parrish 1988; Leij *et al.* 1996). These are automatically set by function `soil()` when initializing soil objects (see section 2.3.3), but we can use function `soil_vanGenuchtenParamsCarsel()` to derive them from texture types:

```

textures = c("Sandy loam", "Silt loam", "Clay")
#Textural parameters
#Sandy clay loam
p1 = soil_vanGenuchtenParamsCarsel(textures[1])
p1

##      alpha          n theta_res theta_sat      Ks
## 764.983    1.890     0.065    0.410 69513.524

alpha1 = p1[1]
n1 = p1[2]
#Silt loam
p2 = soil_vanGenuchtenParamsCarsel(textures[2])
p2

##      alpha          n theta_res theta_sat      Ks
## 203.9955   1.4100    0.0670    0.4500 7077.1687

alpha2 = p2[1]
n2 = p2[2]
#Silty clay
p3 = soil_vanGenuchtenParamsCarsel(textures[3])
p3

##      alpha          n theta_res theta_sat
## 81.59819   1.09000   0.06800    0.38000 3145.400

alpha3 = p3[1]
n3 = p3[2]

```

We can estimate maximum rhizosphere conductance values assuming that they account for an average percentage of the resistance (e.g. 15%) across the continuum (see functions `hydraulics_averageRhizosphereResistancePercent()` and `hydraulics_findRhizosphereMaximumConductance()`):

```

percentResistance = 15
#Sandy clay loam
krmax1 =hydraulics_findRhizosphereMaximumConductance(percentResistance,
            n1,alpha1, krootmax, rootc,rootd, kstemmax, stemc, stemd,
            kleafmax, leafc, leafd)
krmax1

```

```

## [1] 7.375648e+14

#Silt loam
krmax2 =hydraulics_findRhizosphereMaximumConductance(percentResistance,
n2,alpha2, krootmax, rootc,rootd, kstemmax, stemc, stemd,
kleafmax, leafc, leafd)
krmax2

## [1] 3420747735

#Silty clay
krmax3 =hydraulics_findRhizosphereMaximumConductance(percentResistance,
n3,alpha3, krootmax, rootc,rootd, kstemmax, stemc, stemd,
kleafmax, leafc, leafd)
krmax3

## [1] 36831905

```

With these parameters, the resulting $k_{rhizo}(\Psi)$ functions can be displayed using the function `hydraulics_vanGenuchtenConductance()`:

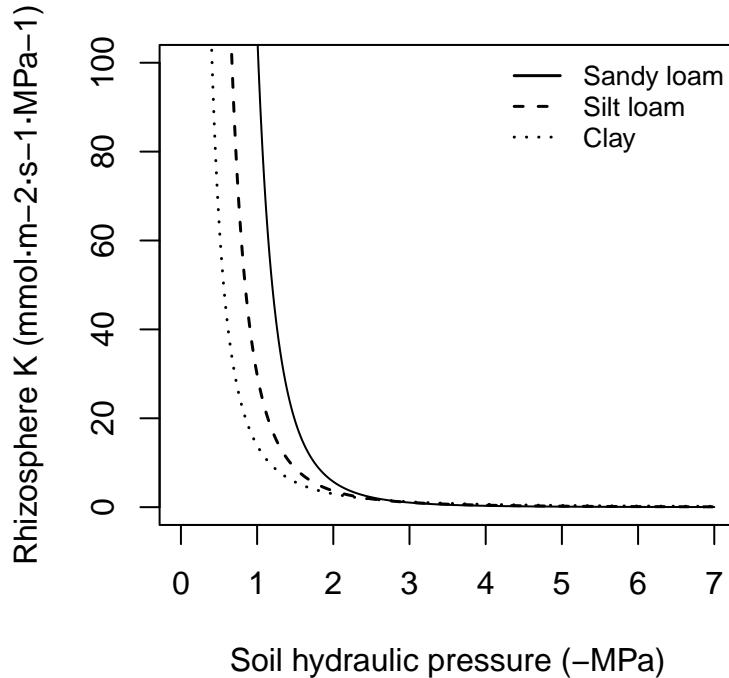


Figure 10.4: Example rhizosphere vulnerability curves (i.e. Van Genuchten functions) for three different soil textures.

10.2.3 Xylem sap viscosity

The ease of water flow (i.e. hydraulic conductance or resistance) is dependent on water viscosity, which fluctuates as a function of temperature and dissolved substances. Xylem sap can be considered to have

negligible amounts of solutes, but temperature is still relevant factor of viscosity. The dynamic viscosity of water V_w varies with temperature (relative to its viscosity at 20°C) according to the Vogel equation:

$$V_w(T) = \exp \left[-3.7188 + \frac{578.919}{-137.546 + T} \right] \quad (10.6)$$

where T is the temperature of the medium in Kelvin. Temperature of segments (roots, stem, leaves) is used to correct maximum conductances simply as follows:

$$k_{max,T} = \frac{k_{max}}{V_w(T)} \quad (10.7)$$

where k_{max} is the maximum conductance describing the vulnerability curve, assumed to be determined at 20°C.

10.3 Water content of plant tissues

Following Martin-StPaul *et al.* (2017), we consider two kind of tissues in (leaf and stem) plant segments (Tyree & Yang 1990). The first are conduits (tracheids or vessels), which will release water due to cavitation and may be refilled with water from adjacent living tissue or upstream segments. The second source of water is formed by more elastic living cells (i.e. parenchyma) and can potentially be a large source of water. This source can be described using the relative water content of a symplasmic tissue.

10.3.1 Relative water content of symplasmic tissues

A pressure-volume curve of a tissue relates its water potential against its *relative water content* (RWC ; $kg H_2O \cdot kg^{-1} H_2O$ at saturation). Pressure-volume theory is usually applied to leaf tissues (Bartlett *et al.* 2012), but it can also be applied to other tissues such as sapwood or cambium cells.

For living cells, the relationship between Ψ and RWC of the symplasmic fraction (RWC_{sym}) is achieved by separating Ψ into osmotic (solute) potential (Ψ_S) and the turgor potential (Ψ_P):

$$\Psi = \Psi_S + \Psi_P \quad (10.8)$$

The relationship for Ψ_P is:

$$\Psi_P = -\pi_0 - \epsilon \cdot (1.0 - RWC_{sym}) \quad (10.9)$$

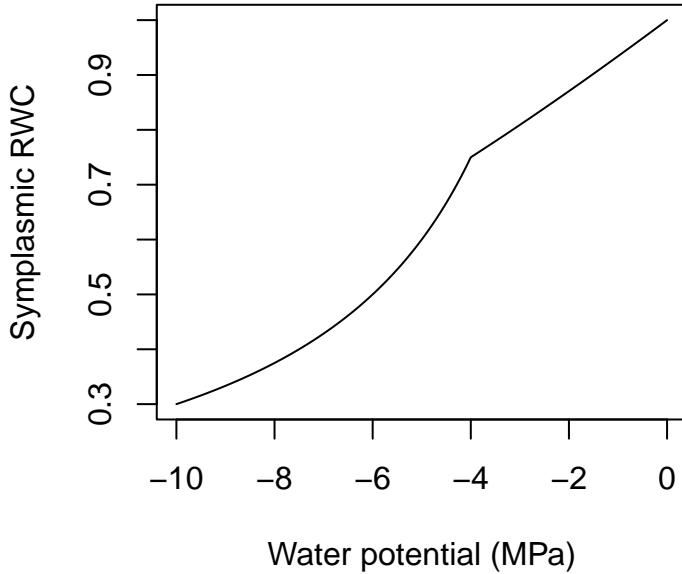
where π_0 (MPa) is the osmotic potential at full turgor (i.e. when $RWC_{sym} = 1$), and ϵ is the modulus of elasticity (i.e. the slope of the relationship). Assuming constant solute content, the relationship for Ψ_S is:

$$\Psi_S = \frac{-\pi_0}{RWC_{sym}} \quad (10.10)$$

When $\Psi \leq \Psi_{tlp}$, the water potential at turgor loss point, then $\Psi_P = 0$ and $\Psi = \Psi_S$. If $\Psi > \Psi_{tlp}$ then the two components are needed. The water potential at turgor loss point (Ψ_{tlp}) can be found by (Bartlett *et al.* 2012):

$$\Psi_{tlp} = \frac{\pi_0 \cdot \epsilon}{\pi_0 + \epsilon} \quad (10.11)$$

As an example, the following figure draws the pressure-volume curve for a tissue with $\epsilon = 12$ and $\pi_0 = -3.0$ MPa:



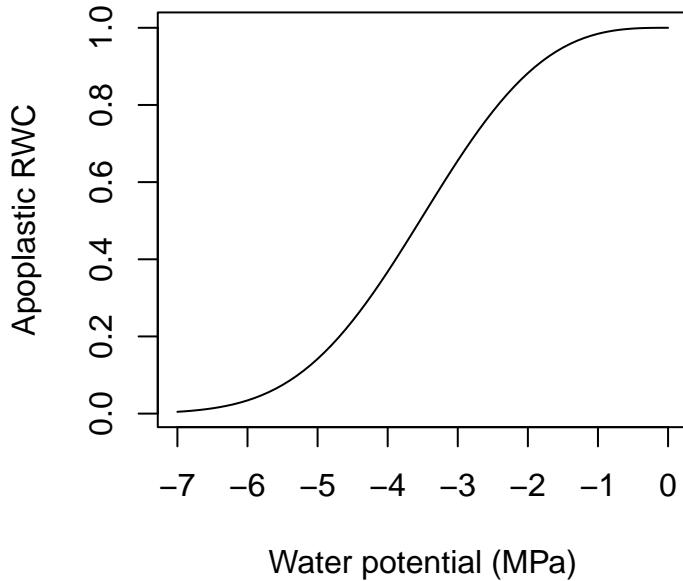
To calculate RWC_{sym} from the water potential of a tissue, the previous equations need to be combined and, after isolating RWC_{sym} , a quadratic relationship is obtained (Martin-StPaul *et al.* 2017).

10.3.2 Relative water content of apoplastic tissues

In **medfate** it is assumed that apoplastic water fraction of an organ corresponds to the water in its xylem conduits. Xylem conduits consist of inelastic cells that have very small changes of water volume in relation to changes in water potential. However, xylem conduits release their water to the transpiration stream following the formation of emboli. As in Hölttä *et al.* (2009), we equate the relative water content of the apoplastic reservoir of a segment (leaves or stem) to the proportion of conductance lost (relative to the maximum conductance) for a given water potential (see (10.1)):

$$RWC_{apo}(\Psi_{apo}) = \frac{k(\Psi_{apo})}{k_{max}} = e^{-((\Psi_{apo}/d)^c)} \quad (10.12)$$

Hence we assume that the relationship between Ψ_{apo} and the relative water content follows the same function as the relationship between Ψ_{apo} and the proportion of conductance loss (i.e. the hydraulic vulnerability curve, see (ref?)(vulnerabilitycurves)).



Since cavitation in leaves is assumed reversible we have that the relative water content of the apoplastic fraction in leaves ($RWC_{leaf,apo}$) will follow the leaf xylem water potential as indicated above. In stems, however, cavitation may be non-reversible, so $RWC_{stem,apo}$ will be dictated by the current level of embolisation, which is represented by PLC_{stem} and not by the current xylem water potential:

$$RWC_{stem,apo} = 1 - PLC_{stem} \quad (10.13)$$

If water compartments are not considered, PLC_{stem} is determined by $\Psi_{stem,cav}$, the minimum stem water potential experienced so far. When water compartments are considered, increases in PLC_{stem} follow decreases in stem water content and the water released can contribute to the transpirational stream (Martin-StPaul *et al.* 2017). This will be treated in the next section.

10.3.3 Average relative water content of a segment

The average relative water content in a given segment or organ (RWC) can be obtained by calculating RWC_{apo} and RWC_{sym} followed by assuming a constant apoplastic fraction f_{apo} :

$$RWC = RWC_{apo} \cdot f_{apo} + RWC_{sym} \cdot (1 - f_{apo}) \quad (10.14)$$

Normally the apoplastic fraction is large (~80%) in the stem and small (~15%) in leaves. Hence, the relative water content in stems will be mostly dictated by the level of cavitation, whereas that of leaves will mostly follow leaf symplastic water potential and the symplastic pressure-volume curve.

10.3.4 Live fuel moisture content

Given an average relative water content (RWC) of a plant organ, its live fuel moisture content ($LFMC$ in $gH_2O \cdot g^{-1}$ of dry tissue) can be calculated using:

$$LFMC = RWC \cdot \Theta \cdot \frac{\rho_{H_2O}}{\rho} = RWC \cdot LFMC_{max} \quad (10.15)$$

where Θ is the tissue porosity (cm^3 of water per cm^3 of tissue), ρ is the density of the tissue and ρ_{H_2O} is the density of water. In practice estimates of maximum fuel moisture content $LFMC_{max}$ are easier to

obtain than porosity estimates, so it is more straightforward to estimate $LFMC$ as the product of RWC and $LFMC_{max}$.

In medfate, $LFMC$ estimates are assumed to correspond to fine fuels (i.e. leaves and twigs of < 6.5 mm) but are estimated from leaf water content. The relative water content of leaves, and hence its $LFMC$, are estimated using (see 10.3.3):

$$LFMC = (RWC_{apo,leaf} \cdot f_{apo,leaf} + RWC_{sym,leaf} \cdot (1 - f_{apo,leaf})) \cdot LFMC_{max} \quad (10.16)$$

where $RWC_{apo,leaf}$ and $RWC_{sym,leaf}$ are the relative water content of apoplastic and symplastic leaf tissue and $f_{apo,leaf}$ is the fraction of apoplastic tissue in leaves.

The package allows estimating $LFMC$ from the output of `spwb()` or `growth()` using function `moisture_cohortFMC()`, but the post-processing function performs slightly different calculations depending on the complexity of the water balance. Basic soil water balance produces, among other outputs, the daily water potential of each plant cohort, Ψ_{plant} (representing an weighted average of soil layer water potentials using fine root distribution as weights) and an estimate of the level of stem cavitation, PLC_{stem} . Daily $LFMC$ estimates can be obtained from this output if we use $RWC_{apo,leaf} = 1 - PLC_{stem}$ and calculate $RWC_{sym,leaf}$ from Ψ_{plant} using symplasmic leaf pressure-volume curves (see 10.3.1). When using basic water balance, function `moisture_cohortFMC()` takes daily values Ψ_{plant} and PLC_{stem} from the simulation output, but parameters $\pi_{0,leaf}$, ϵ_{leaf} (i.e. pressure-volume curve parameters), $f_{apo,leaf}$ and $LFMC_{max}$ are drawn from the data frame `SpParams` of species parameter values. Unlike the basic water balance, the advanced water balance explicitly tracks the water potential in different plant segments and distinguishes between apoplastic and symplastic tissues. In this case, $RWC_{apo,leaf}$ and $RWC_{sym,leaf}$ are obtained directly from the plant water balance, and only $LFMC_{max}$ is drawn from the data frame `SpParams`.

10.4 Sperry's sub-model of plant hydraulics

The supply-loss theory of plant hydraulics, presented by Sperry & Love (2015) and used in Sperry *et al.* (2017), uses the physics of flow through soil and xylem to quantify how steady-state canopy water supply declines with drought and ceases by hydraulic failure. The theory builds on the hydraulic model of Sperry *et al.* (1998) and can be applied to different segmentations of the soil-plant continuum.

The supply function describes the steady-state rate of water supply (i.e. flow) for transpiration (E) as a function of water potential drop. The steady-state flow rate E_i through any i element of the continuum is related to the flow-induced drop in pressure across that element ($\Delta\Psi_i = \Psi_{down} - \Psi_{up}$) by the integral transform of the element's vulnerability curve $k_i(\Psi)$ (Sperry & Love 2015):

$$E_i = \int_{\Psi_{up}}^{\Psi_{down}} k_i(\Psi) d\Psi \quad (10.17)$$

where Ψ_{up} and Ψ_{down} are the upstream and downstream water potential values, respectively. The integral transform assumes infinite discretization of the flow path.

The supply function can be defined for individual elements of the continuum or for the whole soil-plant continuum using different hydraulic networks. Supply functions are used in the Sperry's sub-model to determine photosynthesis, stomatal conductance and transpiration (see chapters 11 and 12).

In the following subsections we illustrate the supply function for different cases.

10.4.1 Supply function for a single xylem element

In the case of a single xylem element, the supply function describes the steady-state flow rate as a function of pressure at the stem top (Ψ_{canopy}). It can be calculated by numerical integration or approximated using

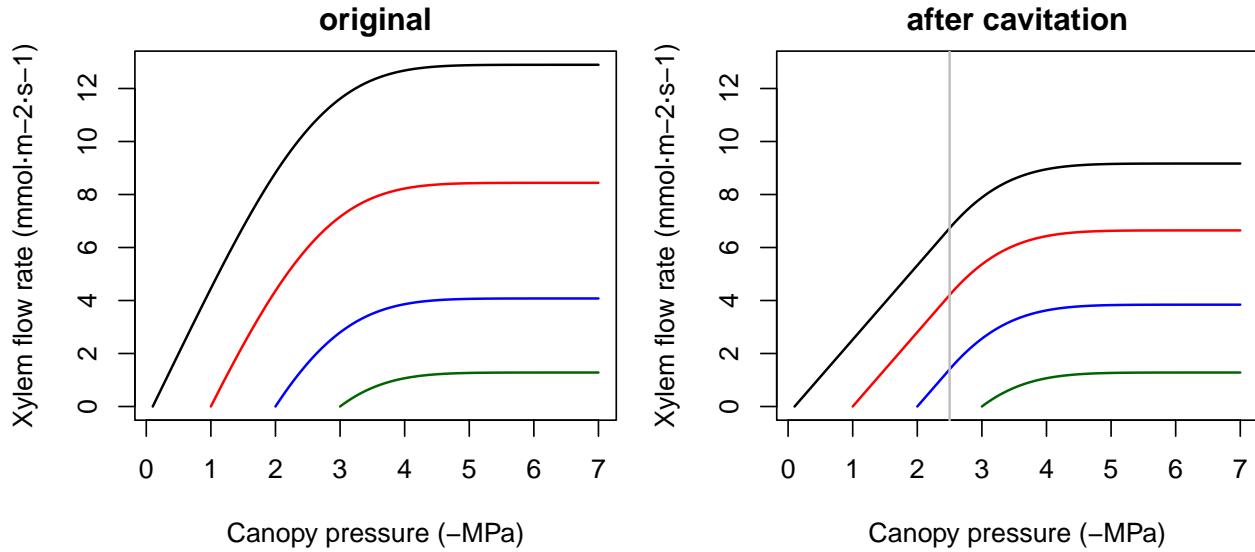


Figure 10.5: Supply function of a single xylem element starting at different root crown water potential values. Left pane shows the uncautitated supply functions and right pane shows the supply functions that are obtained in the case of a cavitated xylem (i.e. without refilling), assuming that the minimum water potential experienced so far was -2.5 MPa. Note the linear part of the flow rate between $\Psi_{rootcrown}$ and this limit.

an *incomplete gamma function*. The shape of the supply function starting at different root crown water potential values ($\Psi_{rootcrown}$) is (see function `hydraulics_ExYlem()`):

The supply function of a *rhizosphere* element relates the steady-state flow rate to the pressure inside the roots (Ψ_{root}). It is calculated by numerical integration of the van Genuchten function, for which we use the analytical approximation of Van Lier *et al.* (2009) (see function `hydraulics_EVanGenuchten()`). In the figure below, we draw examples of the supply function for the rhizosphere. The nearly vertical lines indicate that for many values of E_i the corresponding drop in water potential through the rhizosphere will be negligible. Only for increasingly negative soil water potential values the decrease in water potential through the rhizosphere becomes relevant. Both in the case of a xylem element or a rhizosphere element the derivative $dE_i/d\Psi$ of the supply function is equal to the corresponding vulnerability curve.

10.4.2 Supply function of two elements in series

Let us describe the soil-plant continuum is represented using *two* elements in series (rhizosphere + stem xylem). In this case, the supply function has to be calculated by using the previous supply functions sequentially. The E_i is identical for each element and equal to the canopy E . Since Ψ_{soil} is known, one first inverts the supply function of the rhizosphere to find Ψ_{root} (see function `hydraulics_E2psiVanGenuchten()`) and then inverts the supply function of the xylem to find Ψ_{canopy} (see function `hydraulics_E2psiXylem()`). The two operations can be summarized in a single supply function describing the potential rate of water supply for transpiration (E) as function of the canopy xylem pressure (Ψ_{canopy}), starting from different bulk soil (Ψ_{soil}) values (see function `hydraulics_supplyFunctionTwoElements()`):

The supply function for the whole continuum contains much information. The Ψ intercept at $E = 0$ represents the (predawn) canopy sap pressure which integrates the rooted soil moisture profile. As E increments from zero, the disproportionately greater drop in Ψ_{canopy} results from the loss of conductance. As the soil dries the differences in flow due to soil texture become more apparent.

The derivative of the whole continuum supply function, $dE/d\Psi$, is not equal to either of the vulnerability curves and it has to be obtained numerically. The derivative functions corresponding to the supply functions shown in the previous figure are:

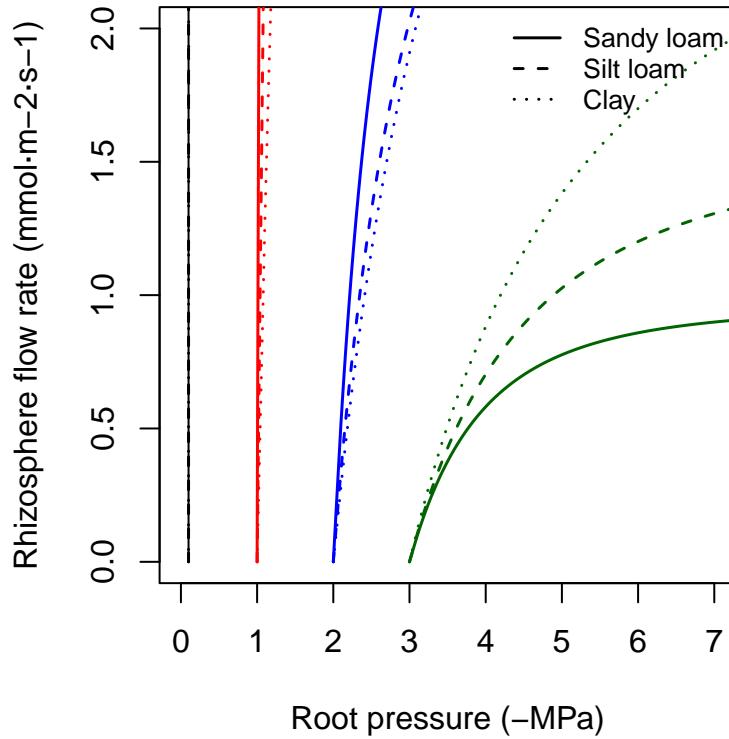


Figure 10.6: Supply functions of the rhizosphere starting at the four different values of bulk soil pressure (Ψ_{soil}) and for the same three texture types used for vulnerability curves.

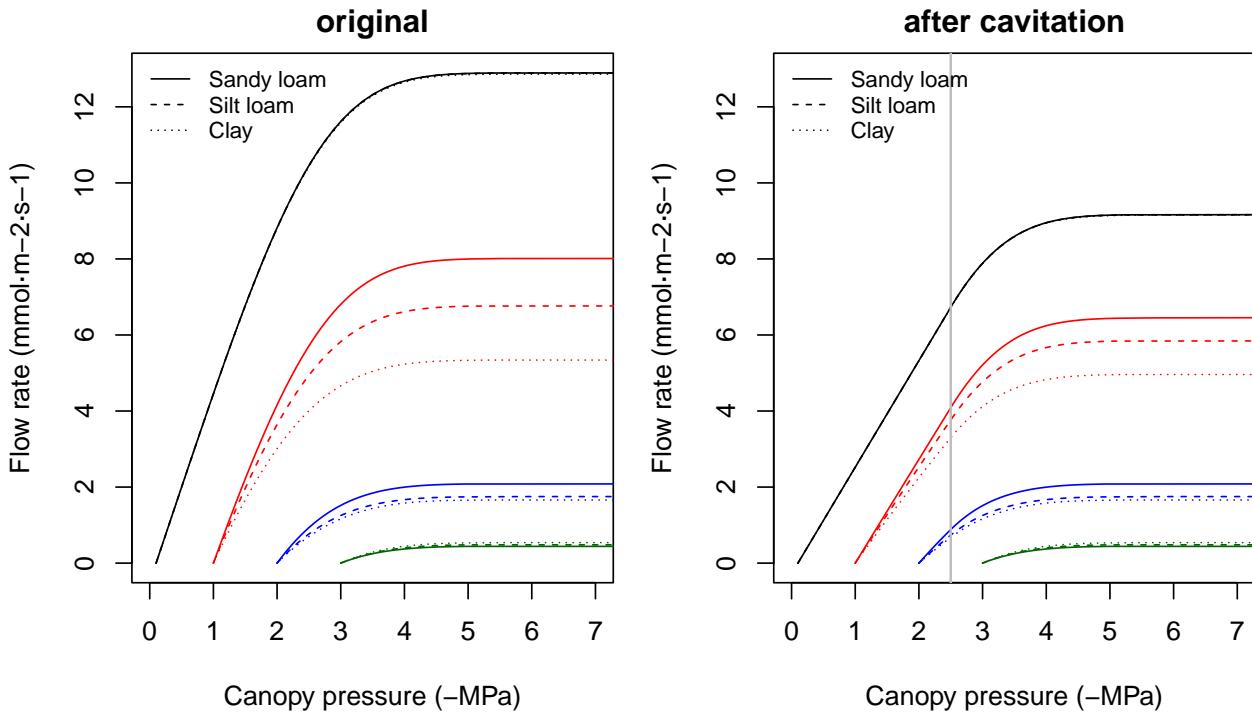


Figure 10.7: Example of two-element supply functions describing the potential rate of water supply for transpiration (E) as function of the canopy xylem pressure (Ψ_{canopy}), starting from different bulk soil (Ψ_{soil}) values and for different soil textures. Left/right panel shows uncavitated/cavitated supply functions.

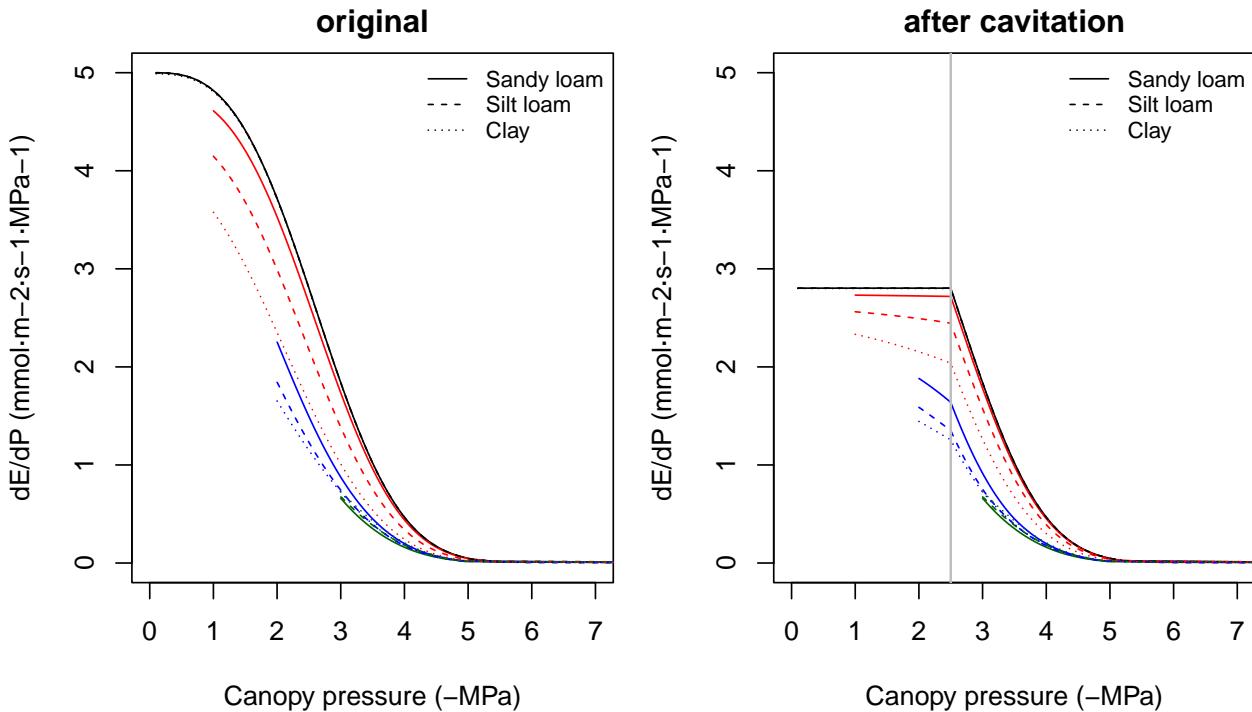


Figure 10.8: Derivatives of two-element supply functions corresponding to figure 10.7. Left/right panel shows the derivatives of uncavitated/cavitated supply functions.

The derivative $dE/d\Psi_{canopy}$ is the conductance if the entire continuum was exposed to Ψ_{canopy} (Sperry & Love 2015). It corresponds to the local loss of hydraulic conductance at the downstream end of the flow path. It falls towards zero for asymptotic critical values (E_{crit}). For a cavitated system $dE/d\Psi_{canopy}$ can be rather flat, in accordance with the close to linear part of the supply function.

10.4.3 Supply function of three elements in series

If the soil-plant continuum is represented using *three* elements in series (rhizosphere + stem xylem + leaf), the resulting overall conductance and resistance fractions (under wet conditions) are:

```
rstemmin = 1/kstemmax
rleafmin = 1/kleafmax

#Percentages of minimum resistance
rvec = c(rstemmin,rleafmin)
100*rvec/sum(rvec)

## [1] 66.66667 33.33333

#Maximum overall conductance
1/sum(rvec)

## [1] 3.333333
```

As before, the supply function has to be calculated *sequentially*, knowing that E_i is identical through each element. Since Ψ_{soil} is known, one first inverts the supply function of the rhizosphere to find Ψ_{root} and

then inverts the supply function of the xylem to find Ψ_{stem} . Finally, one inverts the supply function of the leaf element to find Ψ_{leaf} . As before, the three operations can be summarized in a single supply function describing the potential rate of water supply for transpiration (E) as function of the leaf pressure (Ψ_{leaf}), starting from different bulk soil (Ψ_{soil}) values (see function `hydraulics_supplyFunctionThreeElements()`):

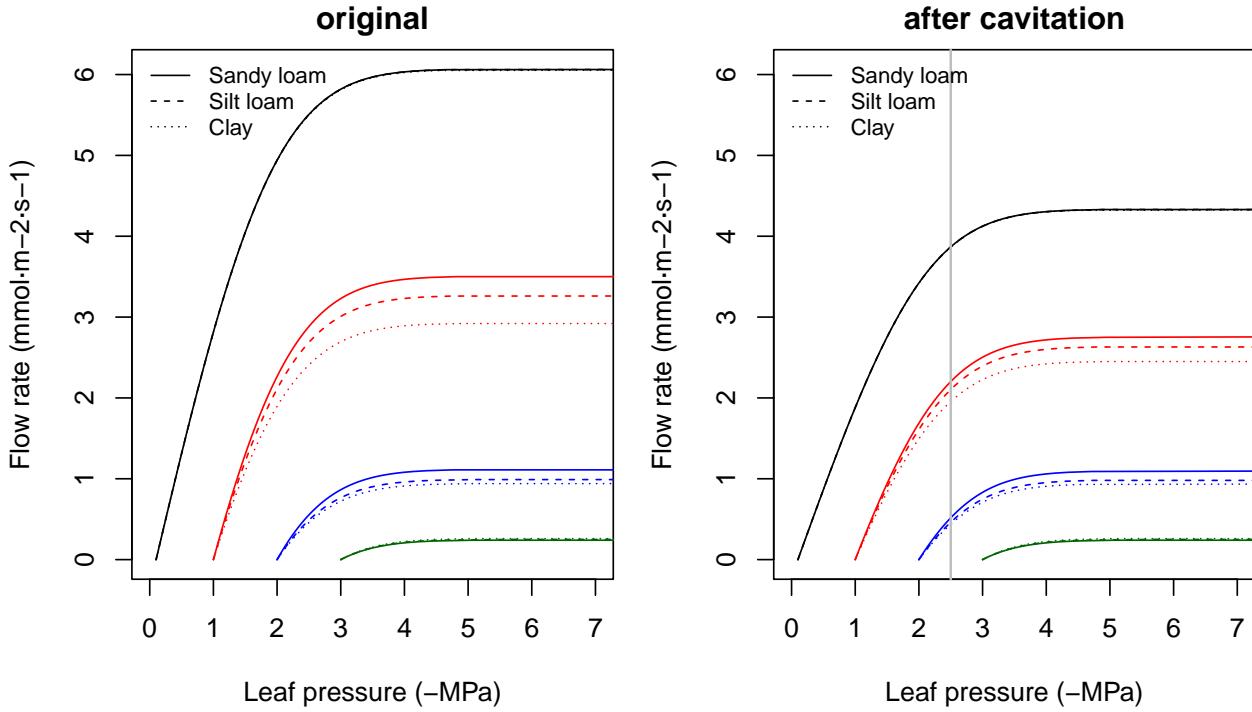


Figure 10.9: Example of three-element supply functions describing the potential rate of water supply for transpiration (E) as function of the leaf pressure (Ψ_{leaf}), starting from different bulk soil (Ψ_{soil}) values and for different soil textures. Left/right panel shows uncavirated/cavirated supply functions.

Note that overall conductance and the maximum flow of the supply function are smaller in this case than in the representation using two elements in series, because we added a new resistance (leaves). While the rhizosphere component only adds a significant resistance when the soil dries, considering the leaf segment (or a root xylem segment) substantially increases the overall resistance of the continuum. Higher vulnerability of leaves also makes the curve to saturate for less negative soil water potentials. The derivative functions corresponding to the supply functions shown in the previous figure are (note the highest value being equal to the overall maximum conductance):

10.4.4 Supply function of a root system

So far we considered supply functions of elements in series, but resistance elements will be in parallel if the soil is represented using $S > 1$ different layers. For each soil layer there is a rhizosphere element in series with a root xylem element. The S soil layers are in parallel up to the root crown.

Network of S rhizosphere components and root layers in parallel there are $S + 1$ unknown pressures: the S root surface pressures ($\Psi_{rootsurf,1}, \dots, \Psi_{rootsurf,S}$) and the root crown pressure at the downstream junction for all root components ($\Psi_{rootcrown}$). The unknown pressures are solved, for each specified total flow value E , using multidimensional Newton-Raphson on a set of equations for steady-state flow (Sperry *et al.* 2016):

$$E_{rhizo,s} - E_{root,s} = 0 \quad (10.18)$$

$$\sum_{s=1}^S E_{root,s} - E = 0 \quad (10.19)$$

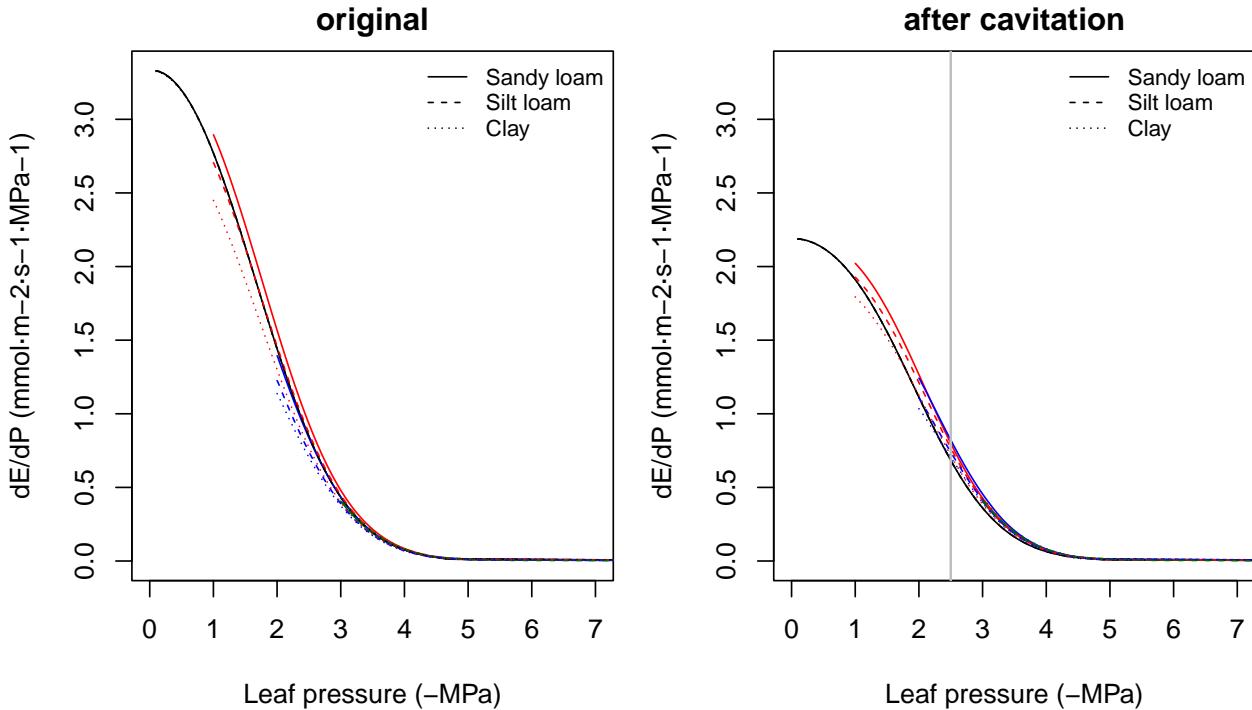


Figure 10.10: Derivatives of three-element supply functions corresponding to figure 10.9. Left/right panel shows the derivatives of uncavitated/cavitated supply functions.

where $E_{rhizo,s}$ and $E_{root,s}$ are steady-state supply flows calculated using the integrals of either van Genuchten or Weibull function as vulnerability curves, respectively. In the case of rhizosphere elements, $\Psi_{up} = \Psi_{soil,s}$ and in the case of root elements $\Psi_{up} = \Psi_{rootsurf,s}$. Solving the steady-state equations also provides values for flow across each of the parallel paths $E_{rhizo,s} = E_{root,s}$.

As an example, we start by defining the water potential of three soil layers corresponding to four situations (analogously with the soil water potentials defined above):

```
psiSoilLayers1 = c(-0.3,-0.2,-0.1)
psiSoilLayers2 = c(-1.3,-1.2,-1.1)
psiSoilLayers3 = c(-2.3,-2.2,-2.1)
psiSoilLayers4 = c(-3.3,-3.2,-3.1)
```

In a network of several soil layers, one has to divide the total rhizosphere and root xylem conductances among layers. Let layer widths (d_s) be:

```
d = c(300,700,3000) #Soil layer widths in mm
```

Now let FRP_1 , FRP_2 and FRP_3 be the proportion of fine root biomass in each soil layer (see section 2.4.6), which can be calculated using:

```
Z50 = 200 #Parameter of LDR root distribution
Z95 = 1200 #Parameter of LDR root distribution
FRP = root_ldrDistribution(Z50, Z95, d)
FRP
```

```
##          [,1]      [,2]      [,3]
## [1,] 0.6652935 0.2749944 0.05971209
```

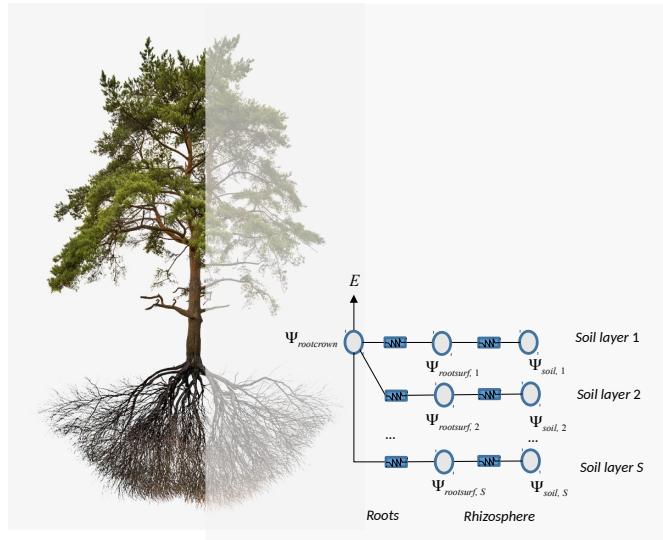


Figure 10.11: Schematic representation of hydraulics in a root network

In the case of the rhizosphere conductances, we can simply define them (for each soil texture type) as:

```
krhizomaxvec1 = krmmax1*FRP
krhizomaxvec2 = krmmax2*FRP
krhizomaxvec3 = krmmax3*FRP
```

To divide maximum root xylem conductance among soil layers we need weights inversely proportional to the length of transport distances (Sperry *et al.* 2016). Vertical transport lengths can be calculated from soil depths and radial spread can be calculated assuming cylinders with volume that has to be in accordance with the xylem root conductance (TO BE DESCRIBED). The whole process can be done using functions `root_coarseRootSoilVolumeFromConductance()` and `root_coarseRootLengthsFromVolume()`:

```
rfc = c(20,50,70)
Vol = root_coarseRootSoilVolumeFromConductance(1.0, 2500,krootmax,
                                               FRP,d, rfc)
lengths = root_coarseRootLengthsFromVolume(Vol, FRP, d, rfc)
lengths
```

```
## [1] 3621.093 2768.105 4513.795
```

```
weights = lengths/sum(lengths)
```

Transport weights are quite different than the fine root biomass proportions. This is because radial lengths are largest for the first (top) layer and vertical lengths are largest for the third (bottom) layer. The root xylem conductances are (in this case they do not depend on soil texture):

```
krootmaxvec = krootmax*weights
krootmaxvec
```

```
## [1] 2.191987 1.675640 2.732373
```

Having all these maximum conductances, we can now build the supply functions for each soil texture and starting from the different soil water potential configurations (see function `hydraulics_supplyFunctionBelowground()`):

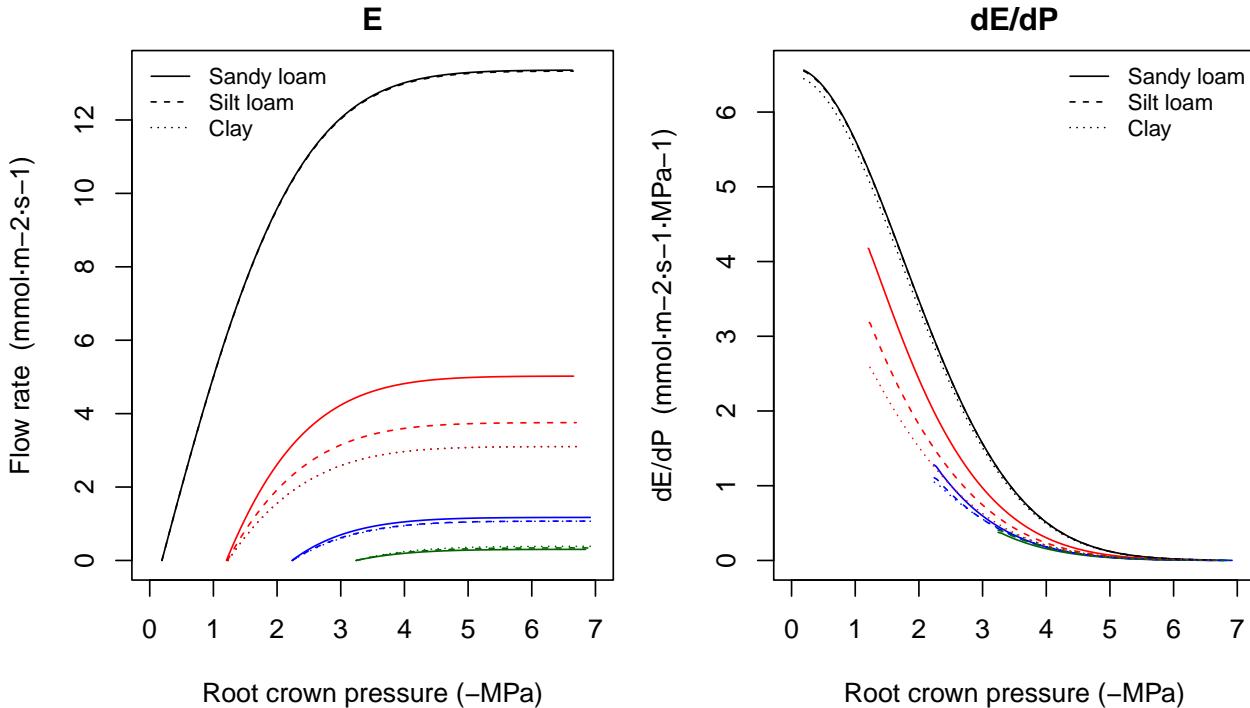


Figure 10.12: Example of supply function for a root system (left) and its derivative (right) under different soil textures and starting from different soil water potential vectors.

The derivative of $dE/d\Psi_{rootcrown}$ for the supply function of the root system is again obtained numerically.

Solving the previous system of equations provides the water potentials in different points of the root system. Here we plot them for the results of silt loam texture and the first and last soil potential vectors defined above:

Note that when soil is not dry (first situation) pressure drop in the rhizosphere is negligible, but not the pressure drop in the root xylem. For drier soils rhizosphere becomes more relevant.

We can also plot the flow rates across each of the parallel paths (again corresponding to the results of silt loam texture and for the four soil potential vectors):

Note that the contribution of each soil layer depends on the soil conditions and the total amount of flow. For a low total flow rate some layers may have negative flows if their potential is lower than others, which in a dynamic context will cause *hydraulic redistribution* of water among soil layers.

10.4.5 Supply function of the soil-plant continuum

As mentioned above, `medfate` uses a hydraulic network of $(S \times 2 + 2 + 1)$ resistance elements to represent the soil-plant continuum, with soil being represented in S different layers. As before, the S soil layers are in

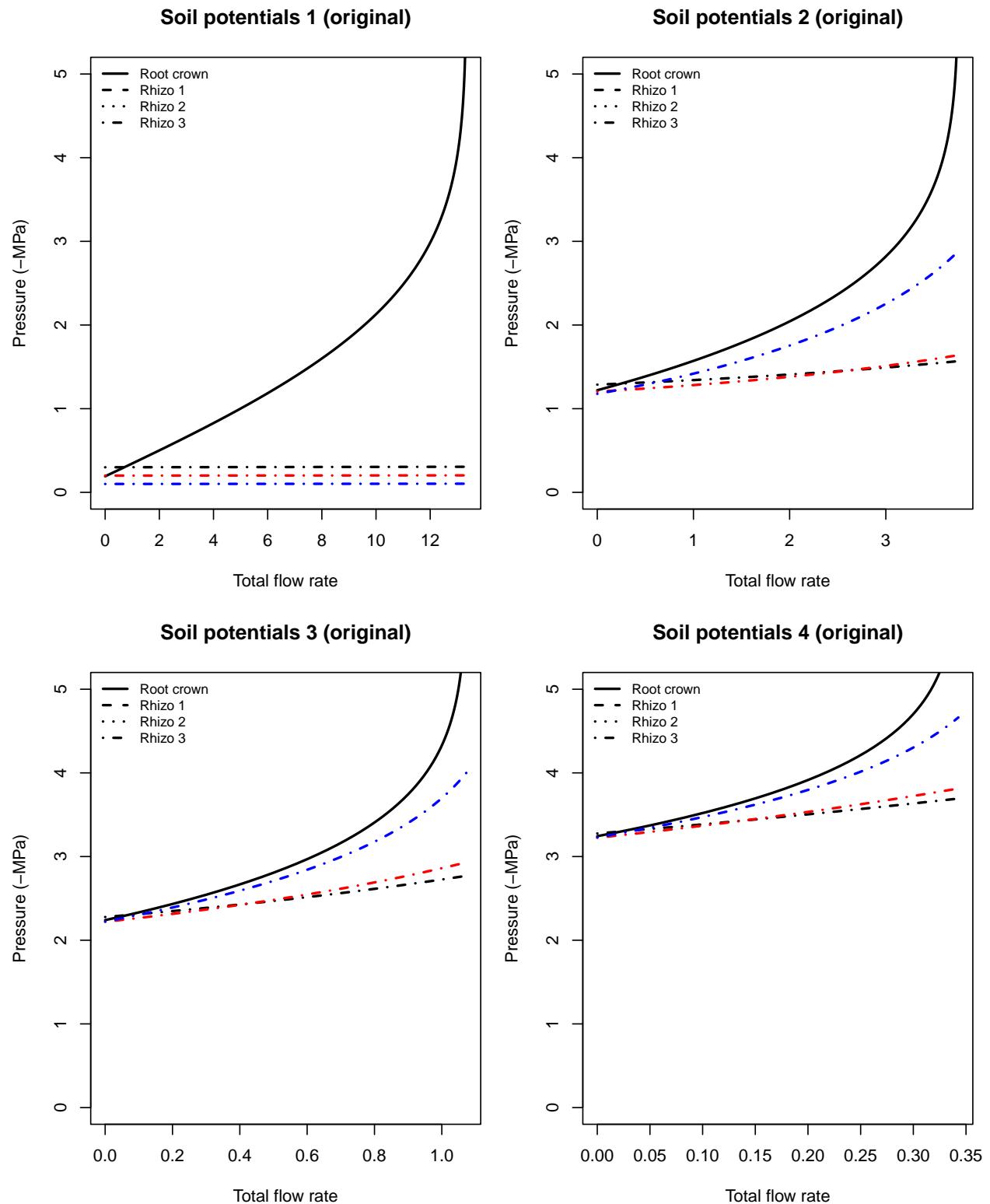


Figure 10.13: Water potentials inside fine roots and at the root crown, for overall flow rates corresponding to fig. 10.12.

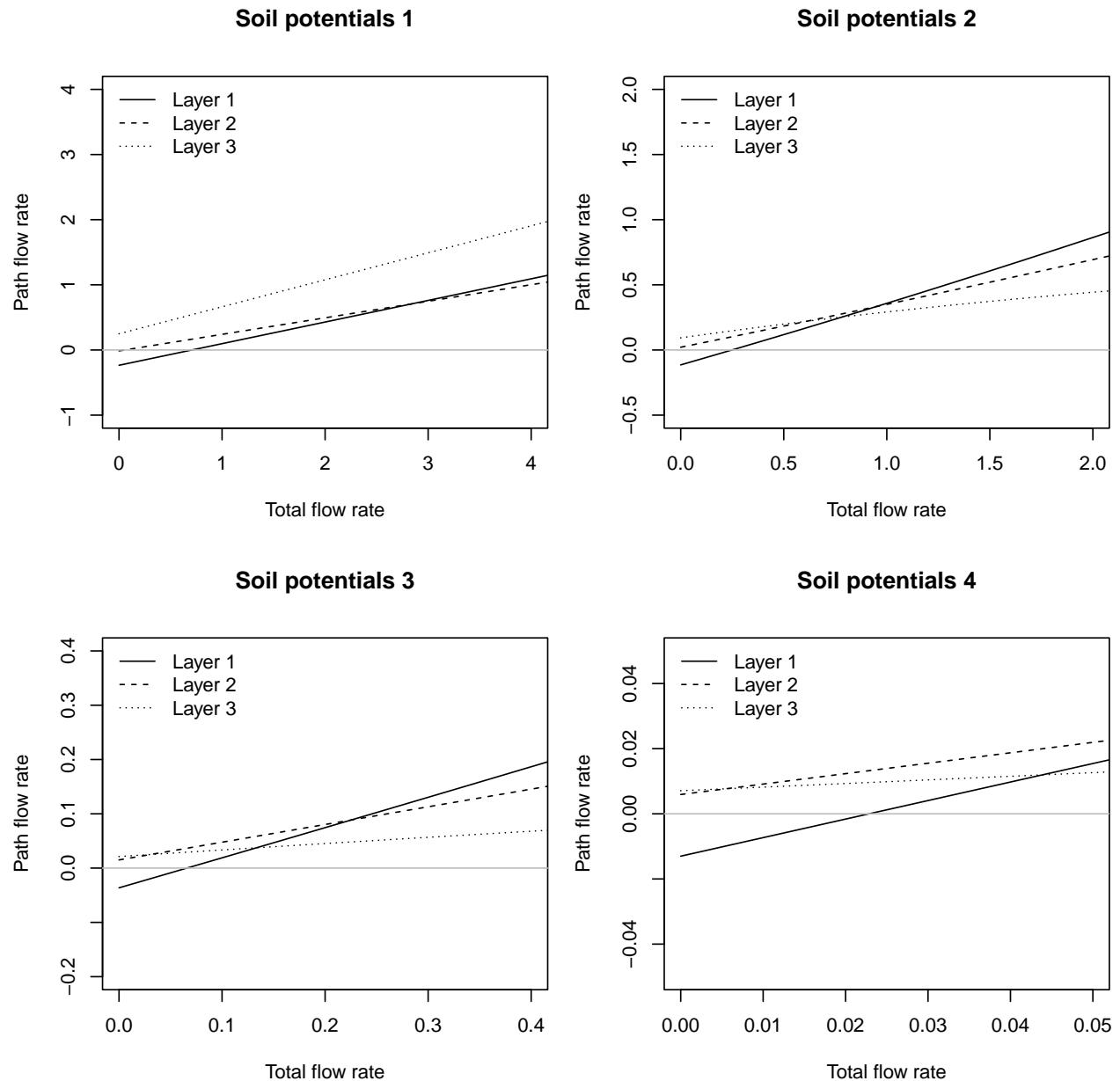


Figure 10.14: Flow rates inside roots corresponding to each soil layer, for overall flow rates corresponding to fig. 10.12.

parallel up to the root crown and each soil layer requires at least a rhizosphere and a root segment. From the root crown there are two stem xylem elements in series and a final leaf element (Fig. 10.1).

To build the supply function for the whole hydraulic network, we proceed by calculating water potentials in the network for each value of flow. For any given E value we start by calculating flows and potentials within the root system. After that, the water potential at the mid of the stem ($\Psi_{stem,1}$) is obtained using the inverse of the stem supply function and setting $\Psi_{up} = \Psi_{rootcrown}$, and the water potential at the upper end of the stem, $\Psi_{stem,2}$, is obtained similarly but setting $\Psi_{up} = \Psi_{stem,1}$. In both cases the maximum conductance for segments equal to $2 \cdot k_{stem,max,i}$. Leaf water potential (Ψ_{leaf}) is finally obtained using the inverse of the leaf supply function and setting $\Psi_{up} = \Psi_{stem,2}$ and assuming a steady-state flow E . The whole supply function $E(\Psi_{leaf})$ is obtained repeating these operations from $E = 0$ to a critical value E_{crit} .

The following figure shows network supply functions for each soil texture and starting from the different soil water potential configurations (see function `hydraulics_supplyFunctionNetwork()`):

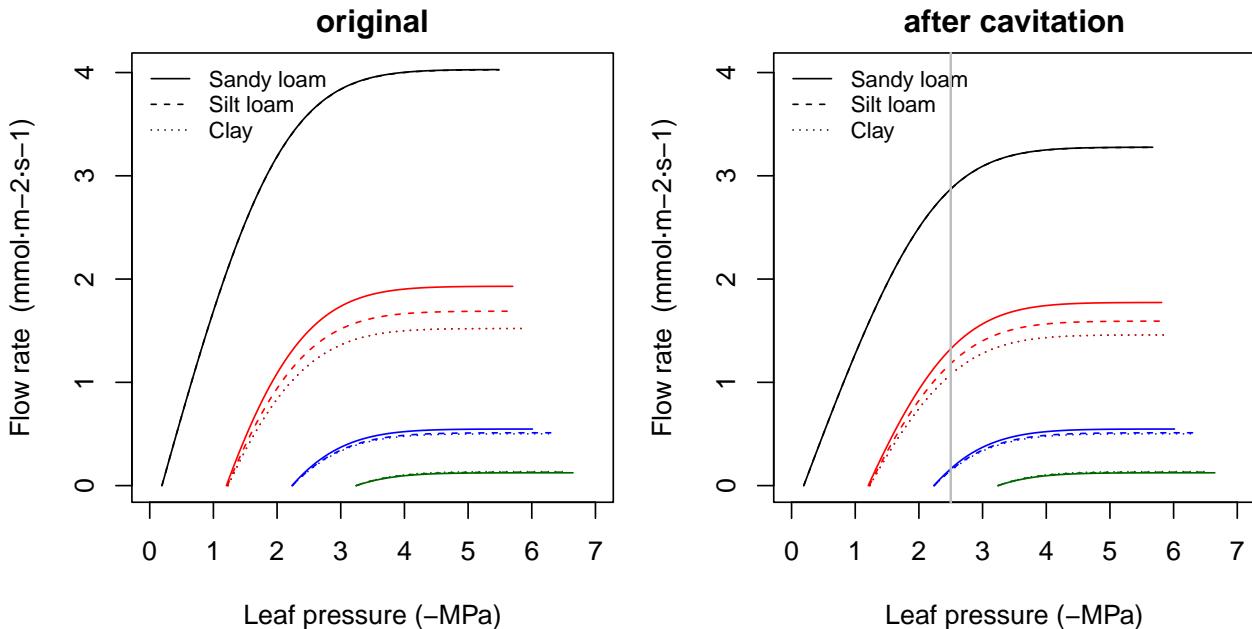


Figure 10.15: Examples of supply functions for a hydraulic network describing the potential rate of water supply for transpiration (E) as function of the leaf pressure (Ψ_{leaf}), starting from different bulk soil (Ψ_{soil}) values and for different soil textures. Left/right panel shows uncavitated/cavitated supply functions.

As with previous representations of the soil-plant continuum, the derivative of $dE/d\Psi_{leaf}$ for the network topology is obtained numerically:

As with the root system, we can know the water potentials in different points of the continuum. Here we plot them for the results of silt loam texture and the first and last soil potential vectors defined above:

A supply function of the soil-plant hydraulic network defines a set steady-state flows and corresponding water potentials given fixed values of soil moisture. Hence, supply functions need to be updated whenever soil moisture is updated, i.e. once a day in the water balance model (see subsection 7.4). This frequency should be enough in most cases, since soil water potentials change more slowly than plant water potentials.

10.5 Sperry's sub-model with water compartments

In `medfate` the relative water content of leaves and stems is tracked explicitly. When capacitance effects are not considered, the relative water content of both kinds of plant tissues follow the water potential

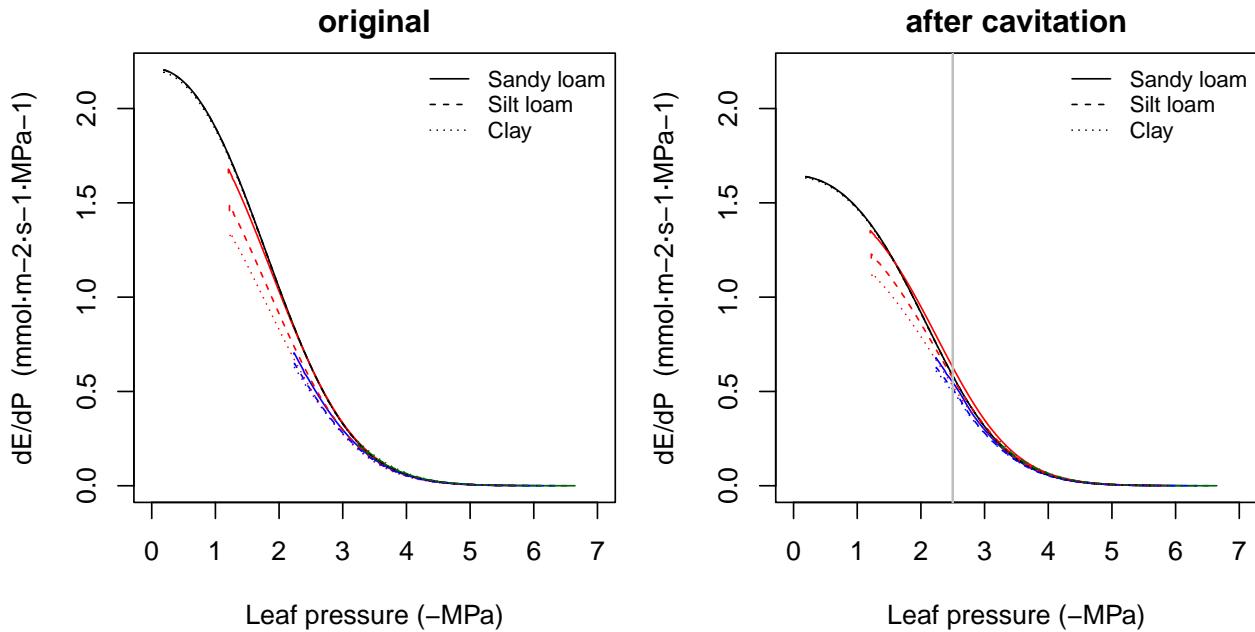


Figure 10.16: Derivatives of supply functions corresponding to figure 10.15. Left/right panel shows the derivatives of uncavitated/cavitated supply functions.

changes dictated by supply functions and stomatal regulation. In contrast, when plant capacitance effects are considered, leaf and stem symplastic tissues have their own water potential as state variable, they exchange water with leaf and stem xylem, respectively.

Supply functions are very useful to estimate steady-state transpiration flows, i.e. situations where a constant transpiration flow through the hydraulic network generates a drop in water potential along the pathway. However, with this modelling strategy the water potential of plants is at least as negative as that of the soil. In other words, plants are hydraulically *connected* to the soil at any time. In reality, evaporation of water in leaf intercellular spaces generates a tension that is transmitted along the xylem and generating hydraulic lift. Water within xylem and adjacent tissues is a source of water more readily accessible for transpiration than soil water.

Xylem conduits are quite inelastic (i.e. they have a very high elastic modulus) so their volumetric changes following diurnal changes in flow rate and water potential are small, and hence contribute little to capacitive water storage and retrieval. In contrast, more elastic living cells in the xylem and bark can potentially be a larger source of water during decreasing water potential. In addition, as xylem conduits cavitate they become filled with water vapour or gas, and nearly all the liquid water inside is freed to the transpiration stream. Cavitation can therefore affect the water status of a plant positively in the short-term due to this capacitive effect (Hölttä *et al.* 2009).

Considering plant water compartments explicitly opens the door to simulating an effective plant disconnection from the soil, because a plant can stop drawing soil water and rely on its own water reserves. Moreover, one can estimate the time needed for plant dessication by taking into account stem and leaf water reserves, which should be greater for larger plants.

10.5.1 Design

To consider plant water storage compartments **medfate** uses two supply functions (black and red arrows in figure below):

1. The first supply function is made of ($S\ddot{O}2 + 1$) elements and draws water from the soil, pouring it

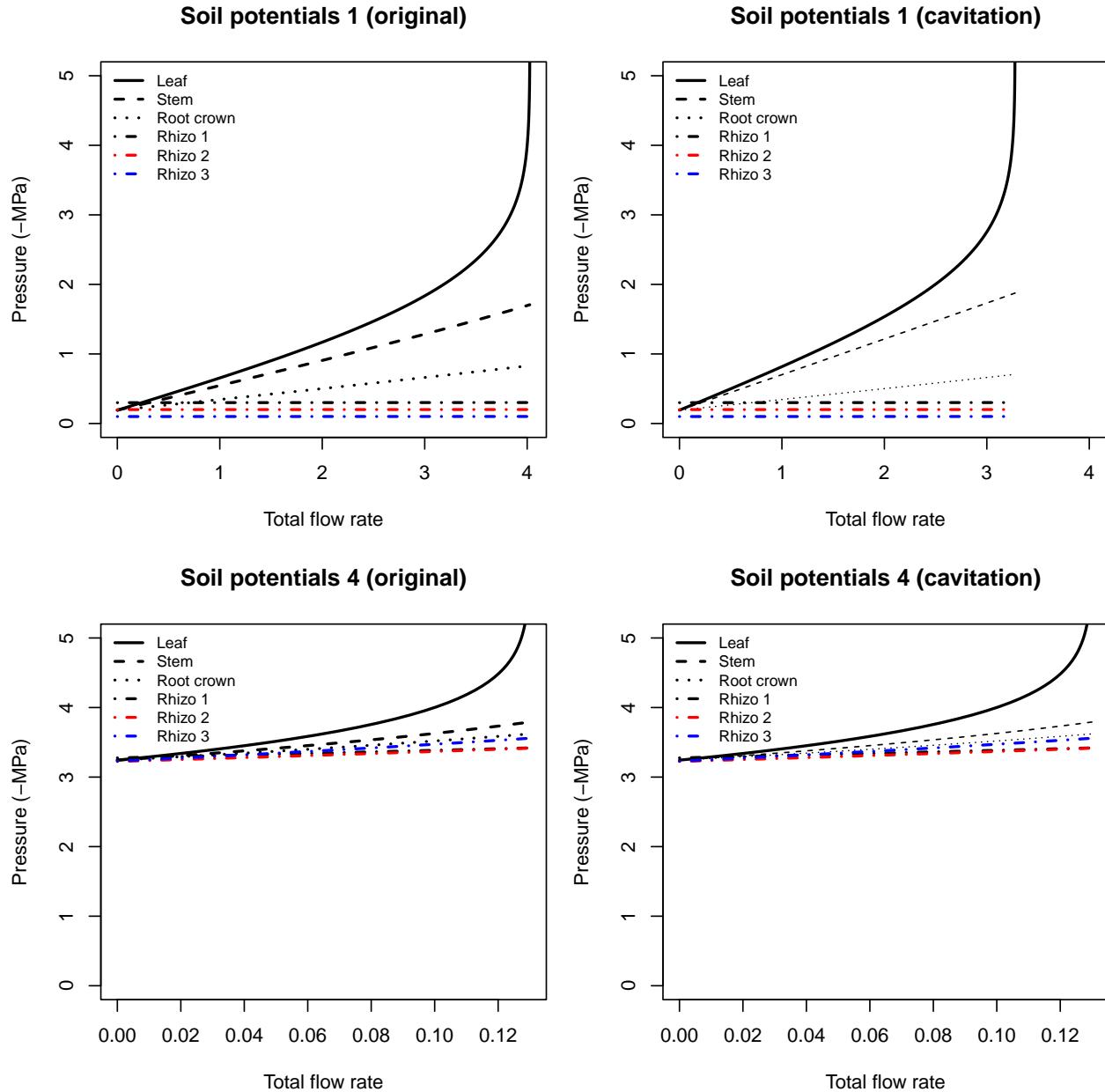


Figure 10.17: Water potential in network elements corresponding to overall flow rates of figure 10.15. Left/right panel shows the derivatives of uncavitated/cavitated supply functions.

into the stem xylem. This supply function is used to determine water uptake from each soil layer and defines an input instantaneous (steady-state) flow to the stem xylem compartment, $E_{stem,in}$.

2. The second supply is made of $(1 + 2 + 1)$ elements (representing plant resistances) and is used to determine the rate of water flowing from stem up to the leaves for transpiration and photosynthesis, as in the model without water compartments. It defines the transpirational flow $E_{leaf,out}$. It needs to be build from $\Psi_{rootsurf}$ to Ψ_{leaf} (and not from $\Psi_{stem,1}$ to Ψ_{leaf} only) because all plant resistances are needed to determine the relationship between $E_{leaf,out}$ and Ψ_{leaf} .

This division of water input and output from the stem allows considering the intermediate node, $\Psi_{stem,1}$, as the water potential of an explicit xylem water compartment from which water is drawn towards the leaves ($E_{stem,out}$) and where soil water is poured ($E_{stem,in}$). Following Hölttä *et al.* (2009), the xylem water compartment can be a source of water for transpiration when cavitation occurs. Following Steppe *et al.* (2006), two symplasmic water reserve compartments are also included, which can also provide water for transpiration. The xylem compartment is hydraulically connected with a stem symplastic compartment (whose moisture level is represented by the state variable $\Psi_{symp,stem}$). Another symplasmic compartment is considered at the leaf level (whose moisture level is represented by $\Psi_{symp,leaf}$) that adds or withdraws water from the transpirational stream depending on differences between leaf xylem and symplastic water potentials. As the supply function #2 assumes a steady state flow from the stem to the leaves and atmosphere, the leaf symplastic water compartment is used to modulate how much of the steady-state flow will actually be drawn from the stem xylem compartment. In other words, the leaf lateral flow simply modifies the water demand from the leaves to the stem (i.e., it makes $E_{stem,out}$ different from $E_{leaf,out}$).

The larger the three compartments in volume, the more water the plant will be able to transpire without needing a replacement of water from the soil. In case of edaphic drought and very much reduced soil-root conductance, the plant can be disconnected hydraulically from the soil and still loose small amounts of water (through cuticular transpiration) from these compartments.

10.5.2 Constitutive equations

The way the previous architecture is implemented is the following. Two supply functions provide flows $E_{stem,in}$ and $E_{leaf,out}$, respectively. Two instantaneous water flows, $F_{lat,stem}$ and $F_{lat,leaf}$, are defined from Darcy's law:

$$F_{lat,stem} = k_{lat,stem} \cdot (\Psi_{symp,stem} - \Psi_{stem,1}) \quad (10.20)$$

$$F_{lat,leaf} = k_{lat,leaf} \cdot (\Psi_{symp,leaf} - \Psi_{leaf}) \quad (10.21)$$

$$(10.22)$$

where $k_{lat,stem}$ and $k_{lat,leaf}$ are the lateral (between symplastic tissues and xylem) water conductance for stem and leaves, respectively. These conductance values are defined as control parameters. The flow leaving the stem is defined by the atmospheric demand minus the water supplied from the leaf symplastic tissues ($F_{lat,leaf}$ can be negative>):

$$E_{stem,out} = E_{leaf,out} - F_{lat,leaf} \quad (10.23)$$

And the mass balance in the stem xylem compartment is:

$$\frac{\delta m_{stem}}{\delta t} = E_{stem,in} - E_{stem,out} + F_{lat,stem} \quad (10.24)$$

However, the change in volume is composed of two terms: one arising from water movement in and out of the stem compartment and the other arising from the change in the gas volume caused by cavitation (Hölttä *et al.* 2009):

$$\frac{\delta V_{stem,apo}}{\delta t} = \frac{1}{\rho_w} \cdot \frac{\delta m_{stem}}{\delta t} + \frac{\delta V_{cav}}{\delta t} \quad (10.25)$$

where ρ_w is the density of water. If there has been an increase in PLC , this creates a source of available water proportional to this change (Hölttä *et al.* 2009):

$$\frac{\delta V_{cav}}{\delta t} = V_{stem,apo,max} \cdot \frac{\delta PLC_{stem}}{\delta t} \quad (10.26)$$

where $V_{stem,apo,max}$ is the capacity of the stem xylem compartment. So if $\delta PLC_{stem} = 0$, $\delta V_{cav}/\delta t = 0.0$. Since we model PLC using the vulnerability curve, the above formulation of water released by cavitation assumes that the relationship between stem water potential and the proportion of cavitated conduits follows the same function as the relationship between stem water potential and the proportion of conductance loss (i.e. the hydraulic vulnerability curve). The change in water pressure in the stem xylem compartment following the change in water volume is:

$$\frac{\delta \Psi_{stem,1}}{\delta t} = \epsilon_{xylem} \cdot \frac{1}{V_{stem,apo,max}} \cdot \frac{\delta V_{stem,apo}}{\delta t} = \epsilon_{xylem} \cdot \frac{\delta RWC_{stem,apo}}{\delta t} \quad (10.27)$$

Where $\epsilon_{xylem} = 10^3$ MPa is the xylem elastic modulus.

The water balance equations for the symplastic compartments are simply:

$$\frac{\delta V_{stem,symp}}{\delta t} = -\frac{1}{\rho_w} \cdot F_{lat,stem} \quad (10.28)$$

$$\frac{\delta V_{leaf,symp}}{\delta t} = -\frac{1}{\rho_w} \cdot F_{lat,leaf} \quad (10.29)$$

These are completed with maximum storage capacity values of the symplasmic water compartments ($V_{stem,symp,max}$ and $V_{leaf,symp,max}$), which allow determining changes in relative water content of symplasmic tissues, and the corresponding pressure-volume equations relating relative water content changes with water potentials $\Psi_{symp,stem}$ and $\Psi_{symp,leaf}$ (see section 10.3.1).

10.5.3 Implementation

The dynamic system is solved in the model by time discretization. Small time steps are needed when dealing with storage compartments, which poses additional challenges. The soil-to-stem supply function only needs to be updated when soil moisture has changed substantially, i.e. at the same rate as the hydraulic system without plant compartments. However, the second supply function needs to be recalculated more often, because $\Psi_{stem,1}$ changes faster than soil water potentials. In the model, the soil-to-stem supply function is recalculated once a day, whereas the stem-to-leaf supply function is recalculated following subdaily time steps (i.e. every Δt_{step} , by default $\Delta t_{step} = 1h = 3600s$). Water potentials of storage compartments, and lateral flows between water compartments ($F_{lat,stem}$ and $F_{lat,leaf}$) are recalculated every 1s time step. Instantaneous transpiration rate $E_{leaf,out}$ is considered constant within a given subdaily Δt_{step} , but $E_{stem,in}$ is re-estimated every second depending on the current value of $\Psi_{stem,1}$. Since the soil-to-stem supply function is already defined for a range of $\Psi_{stem,1}$ values, at each 1s time step the model only needs to determine the value of $E_{stem,in}$ in the supply function corresponding to the current value of $\Psi_{stem,1}$.

10.6 Cochard's sub-model of plant hydraulics

[TO BE WRITTEN]

Chapter 11

Leaf/canopy photosynthesis

In chapter 10 we introduced the necessary concepts of steady-state plant hydraulics to define the *hydraulic supply function for the soil-plant continuum*. The supply function specifies the flow rate, as per leaf area, for values of leaf water potential and given a soil moisture status. In the first section of this chapter we describe leaf photosynthesis functions, which define leaf gas exchange, photosynthesis and energy balance for values of leaf water potential. This requires knowing wind, water vapor and temperature conditions of the air surrounding leaves (section 13.1.3), the short-wave radiation absorbed by leaf elements (section 9.1.3) and the leaf long-wave radiation balance (section 9.2). In the second section of this chapter we discuss how to upscale photosynthesis values to the crown level.

11.1 Leaf energy balance, gas exchange and photosynthesis

The supply function specifies the transpiration flow rate E for values of leaf water potential Ψ_{leaf} . If we know air temperature, water vapor pressure and the light conditions in which leaves are, we can translate the supply function into a **leaf photosynthesis function** (Sperry *et al.* 2017). In a nutshell, E from the supply function is first used to calculate leaf temperature from an evaluation of the leaf energy balance. The diffusive conductances of the leaf to H_2O and CO_2 are obtained from water supply and leaf water vapor deficit. Finally, the gross/net assimilation rate is then obtained from the diffusive conductance and a modelled curve between assimilation and leaf internal CO_2 concentration. The following subsections detail each of these steps separately.

11.1.1 Leaf temperature

Leaf temperature (T_{leaf} ; in $^{\circ}C$) can be calculated for any given flow rate E using an energy balance equation (Campbell & Norman 1998):

$$T_{leaf} = T_{air} + \frac{\Phi_{SWR}^{leaf} + L_{net}^{leaf} - \lambda_v(T_{air}) \cdot E_{two-sided}}{C_p \cdot (g_r + g_{Ha})} \quad (11.1)$$

where Φ_{SWR}^{leaf} (in $W \cdot m^{-2}$) is the instantaneous shortwave radiation absorbed per leaf area unit, L_{net}^{leaf} is the instantaneous net long-wave radiation per leaf area unit, $E_{two-sided}$ is the flow rate E converted to $mol \cdot s^{-1} \cdot m^{-2}$ per two-sided leaf area basis, ϵ is longwave radiation emissivity (0.97), σ is the Stephan-Boltzman constant, T_{air} is the temperature of the air surrounding the leaf (in $^{\circ}C$), $C_p = 29.3 \text{ J} \cdot mol^{-1} \cdot ^{\circ}C^{-1}$ is the specific heat capacity of dry air at constant pressure and $\lambda_v(T_{air})$ is the latent heat of vaporization (in $J \cdot mol^{-1}$) corresponding to T_{air} (see utility functions in the **meteoland** reference manual). If canopy energy balance is performed assuming a single canopy layer T_{air} corresponds to T_{can} , the whole-canopy air

temperature resulting from the energy balance of the previous step (see section 13.1.3). If a multi-layer canopy energy balance is used, T_{air} corresponds to $T_{air,j}$, the air temperature of the canopy layer j that contains the leaf. Finally, g_r and g_{Ha} are the radiative and heat conductance values (in $\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$), respectively (Campbell & Norman 1998):

$$g_r = \frac{4 \cdot \epsilon \cdot \sigma \cdot (T_{air} + 273.15)^3}{C_p} \quad (11.2)$$

$$g_{Ha} = 0.189 \cdot (u_{leaf}/d)^{0.5} \quad (11.3)$$

where u_{leaf} is the leaf-level wind speed (in $\text{m} \cdot \text{s}^{-1}$) and d is 0.72 times the leaf width (species parameter LeafWidth in cm). As before, u_{leaf} corresponds to u_j , the wind-speed at the canopy layer j where the leaf is located (see 23.2).

By inspecting eq. (11.1), we can conclude that transpiration flow decreases leaf temperature, whereas radiation increases it and wind speed makes it more similar to the temperature of the surrounding air. The following figures illustrate the effect of varying wind speed and flow rate on T_{leaf} for two contrasted leaf widths (see function biophysics_leafTemperature):

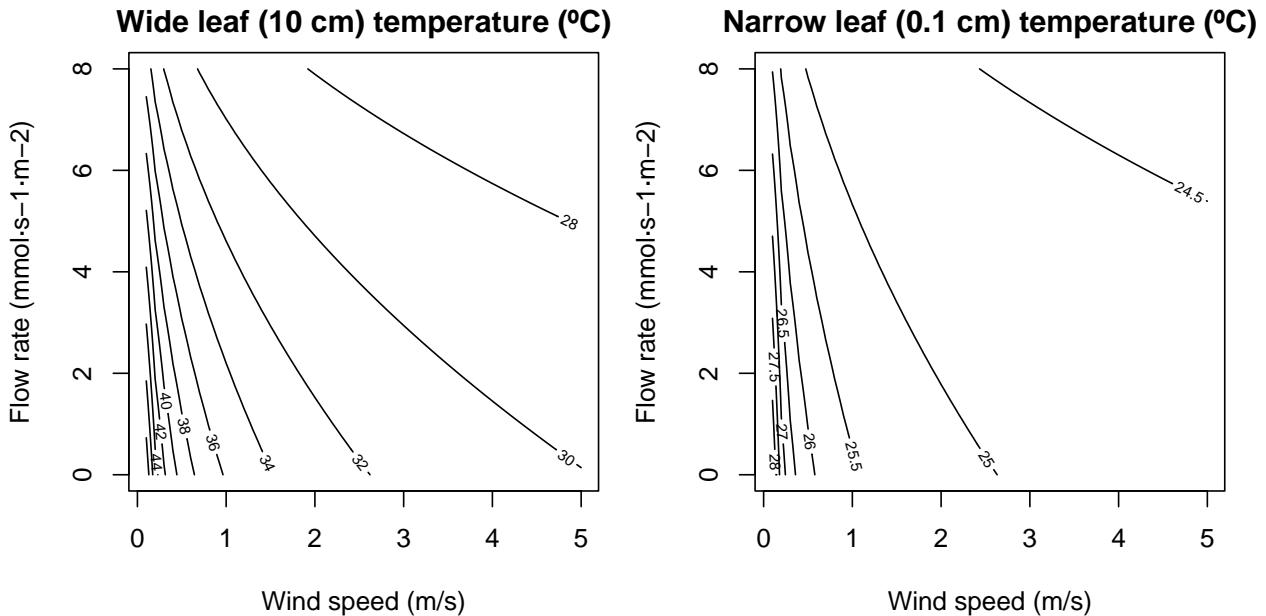


Figure 11.1: Values of T_{leaf} for two leaf widths and varying values of wind speed and flow rate, calculated for 24°C air temperature and 740 $\text{W} \cdot \text{m}^{-2}$ instantaneous absorbed radiation (including SWR and LWR).

Let us now fix wind speed at the leaf leavel to $u_{leaf} = 2 \text{ m/s}$. The application of the above equations to the $E(\Psi_{leaf})$ curves corresponding to the complete hydraulic network (10.4.5) yields the following $T_{leaf}(\Psi_{leaf})$ curves:

11.1.2 Leaf vapor pressure deficit

Vapor pressure deficit in the leaf (VPD_{leaf} , in kPa) is calculated as:

$$VPD_{leaf} = e_{leaf} - e_{air} \quad (11.4)$$

Where e_{air} is the water vapor pressure of the air surrounding the leaf (but beyond the leaf boundary layer) and e_{leaf} is the leaf vapor pressure, which can be calculated from leaf temperature (T_{leaf}) and leaf water potential (Ψ_{leaf}):

$$e_{leaf} = e_{sat}(T_{leaf}) \cdot \exp \left\{ \frac{2.17 \cdot \Psi_{leaf}}{T_{leaf} + 273.15} \right\} \quad (11.5)$$

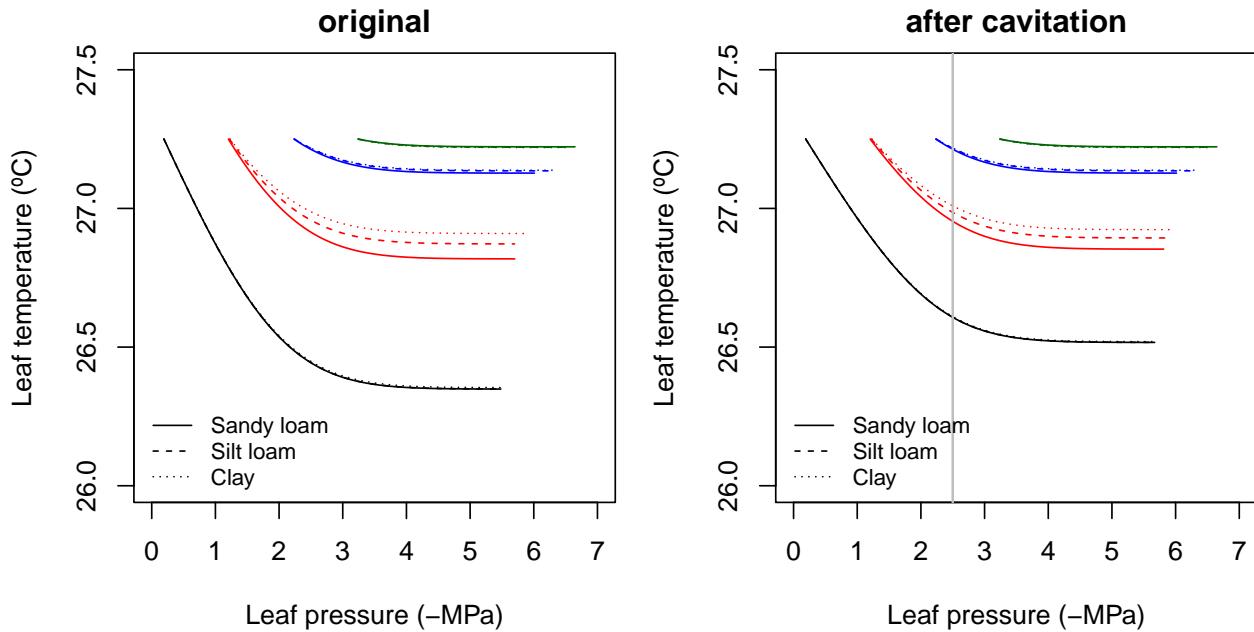


Figure 11.2: Examples of leaf temperature functions for a hydraulic network, corresponding to fig. 10.15 and for different soil textures. Left/right panel shows values for uncavitated/cavitated supply functions.

where $e_{sat}(T)$ is a function giving the saturated vapor pressure for temperature T (see utility functions of the **meteoland** reference manual). If a single-canopy energy balance is considered, the water vapor pressure of the air surrounding the leaf is taken as the average atmospheric water vapor pressure of the day, i.e. $e_{air} = e_{atm}$. When a multi-layer canopy energy balance is simulated, e_{air} will correspond to the water vapor pressure $e_{air,j}$ of the canopy layer j where the leaf is located.

Since e_{leaf} decreases when leaf temperature decreases in eq. (11.5), increasing transpiration decreases leaf VPD as a result of decreasing leaf temperature. To illustrate this effect, let us assume the following values of relative humidity, yielding a $e_{air} = e_{atm} = 1.91 \text{ kPa}$:

```
RHmin = 60
RHmax = 75
VPatm = meteoland::utils_averageDailyVP(Tmin, Tmax, RHmin, RHmax)
VPatm
```

```
## [1] 1.912181
```

The application of the above equation to the $T_{leaf}(\Psi_{leaf})$ curves of fig. 11.2 yields the following $VPD_{leaf}(\Psi_{leaf})$ curves:

Note that the VPD decreasing curves do not start at the same VPD_{leaf} value despite corresponding to the same T_{leaf} value, because of the effect of Ψ_{leaf} on e_{leaf} in eq. (11.5).

11.1.3 Leaf gas exchange

Leaf diffusive conductance to water vapor (g_w ; in $\text{mol H}_2\text{O} \cdot \text{s}^{-1} \cdot \text{m}^{-2}$) and to carbon dioxide (g_c ; in $\text{mol CO}_2 \cdot \text{s}^{-1} \cdot \text{m}^{-2}$) are obtained for each value of transpiration flow E (here expressed in $\text{mol H}_2\text{O} \cdot \text{s}^{-1} \cdot \text{m}^{-2}$)

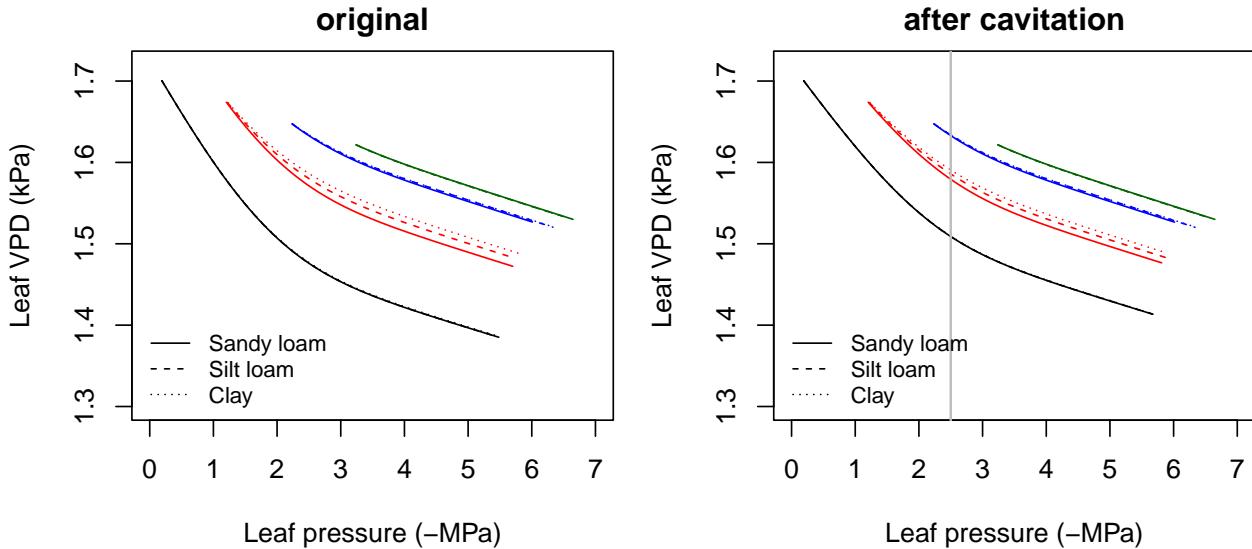


Figure 11.3: Examples of leaf vapour pressure deficit (VPD_{leaf}) functions for a hydraulic network, corresponding to fig. 10.15 and for different soil textures. Left/right panel shows values for uncavitated/cavitated supply functions.

and VPD_{leaf} using:

$$g_w = E \cdot \frac{P_{atm}}{VPD_{leaf}} \quad (11.6)$$

$$g_c = g_w/1.6 \quad (11.7)$$

Diffusive conductance to water vapour is assumed to be result of stomatal (g_{sw}) and boundary layer (g_{bw}) conductances (both in $\text{mol H}_2\text{O} \cdot \text{s}^{-1} \cdot \text{m}^{-2}$). If we estimate g_{bw} using:

$$g_{bw} = 0.397 \cdot (u_{leaf}/d)^{0.5} \quad (11.8)$$

then, stomatal conductance is:

$$g_{sw}^{-1} = g_w^{-1} - g_{bw}^{-1} \quad (11.9)$$

Ensuring here that $g_w \leq g_{bw}$ so that $g_w^{-1} \geq g_{bw}^{-1}$. The application of equations for g_w , g_{bw} and g_{sw} to the $VPD_{leaf}(\Psi_{leaf})$ curves yields the following stomatal conductance $g_{sw}(\Psi_{leaf})$ curves:

In the previous figure we have indicated the thresholds of $g_{sw,\min}$ and $g_{sw,\max}$, the species-specific minimum and maximum water vapour conductances (i.e. conductances when stomata are fully closed and fully open, respectively; see parameters **Gswmin** and **Gswmax** in **SpParamsMED**). g_{sw} cannot exceed $g_{sw,\max}$ so that some flow rates may not be possible (see stomatal regulation below). However, $g_{sw,\max}$ should quickly become non-limiting as soil dries (i.e. reducing E) or VPD_{leaf} increases (Sperry *et al.* 2017).

11.1.4 Leaf photosynthesis

Rubisco-limited photosynthesis rate A_c (in $\mu\text{mol CO}_2 \cdot \text{s}^{-1} \cdot \text{m}^{-2}$) is modelled using (Collatz *et al.* 1991; Medlyn *et al.* 2002):

$$A_c = \frac{V_{max} \cdot (C_i - \Gamma^*)}{C_i + K_c \cdot (1 + O_a/K_o)} \quad (11.10)$$

where V_{max} is Rubisco's maximum carboxylation rate (in $\mu\text{mol CO}_2 \cdot \text{s}^{-1} \cdot \text{m}^{-2}$), C_i is the internal carbon dioxide concentration (in $\mu\text{mol} \cdot \text{mol}^{-1}$), Γ^* is the compensation point (in $\mu\text{mol} \cdot \text{mol}^{-1}$), K_c (in $\mu\text{mol} \cdot \text{mol}^{-1}$) and K_o (in $\text{mmol} \cdot \text{mol}^{-1}$) are Michaelis-Menten constants for carboxylation and oxygenation, respectively,

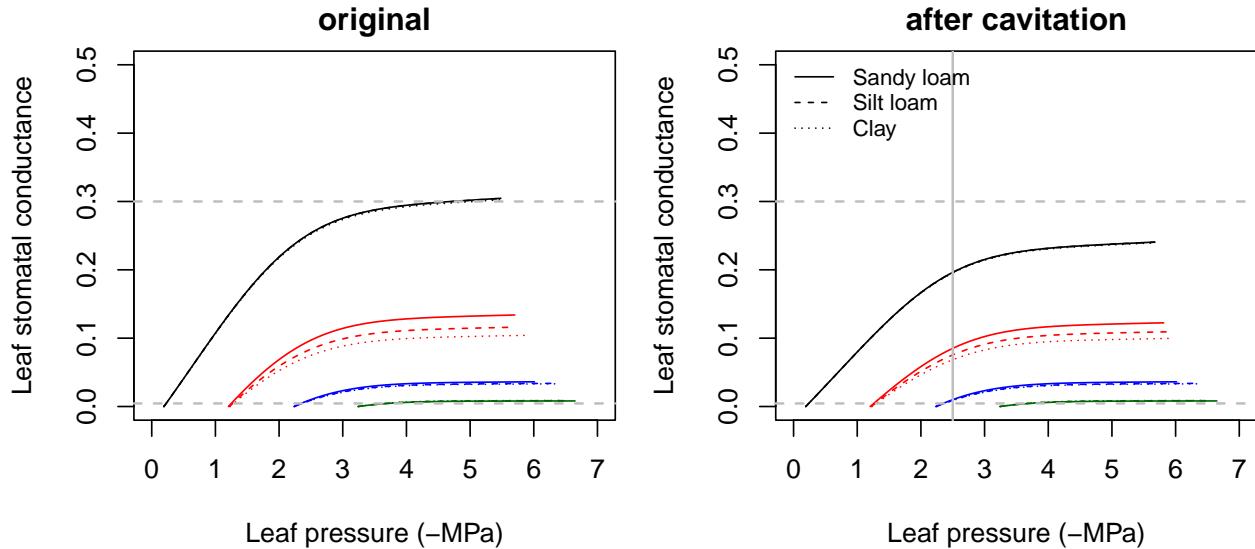


Figure 11.4: Examples of stomatal conductance to water vapour (g_{sw}) functions for a hydraulic network, corresponding to fig. 10.15 and for different soil textures. Left/right panel shows values for uncavitated/cavitated supply functions. Minimum and maximum conductance values ($g_{sw,\min} = 0.0045$ and $g_{sw,\max} = 0.3$) are indicated using dashed lines.

and O_a is the atmospheric oxygen concentration (i.e. $209 \text{ mmol} \cdot \text{mol}^{-1}$). Γ^* , K_c and K_o depend on leaf temperature (T_{leaf} , in $^{\circ}\text{C}$) (Bernacchi *et al.* 2001):

$$\Gamma^* = 42.75 \cdot e^{\frac{37830 \cdot (T_{leaf} - 25)}{298 \cdot R \cdot (T_{leaf} - 273)}} \quad (11.11)$$

$$K_c = 404.9 \cdot e^{\frac{79430 \cdot (T_{leaf} - 25)}{298 \cdot R \cdot (T_{leaf} - 273)}} \quad (11.12)$$

$$K_o = 278.4 \cdot e^{\frac{36380 \cdot (T_{leaf} - 25)}{298 \cdot R \cdot (T_{leaf} - 273)}} \quad (11.13)$$

Electron transport-limited photosynthesis A_e (in $\mu\text{mol CO}_2 \cdot \text{s}^{-1} \cdot \text{m}^{-2}$) was obtained from Medlyn *et al.* (2002):

$$A_e = \frac{J}{4} \cdot \frac{C_i - \Gamma^*}{C_i + 2 \cdot \Gamma^*} \quad (11.14)$$

$$J = \frac{(\alpha \cdot Q_{PAR}^{leaf} + J_{max}) - \sqrt{(\alpha \cdot Q_{PAR}^{leaf} + J_{max})^2 - 4.0 \cdot c \cdot \alpha \cdot Q_{PAR}^{leaf} \cdot J_{max}}}{2 \cdot c} \quad (11.15)$$

where $\alpha = 0.3 \text{ mol e} \cdot \text{mol photon}^{-1}$ is the quantum yield of electron transport, Q_{PAR}^{leaf} is the absorbed PAR photon flux density per leaf area ($\mu\text{mol photon} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$), which is calculated from absorbed PAR per leaf area (Φ_{PAR}^{leaf} ; in $\text{W} \cdot \text{m}^{-2}$):

$$Q_{PAR}^{leaf} = \Phi_{PAR}^{leaf} \cdot 546 \cdot 0.836 \cdot 10^{-2} \quad (11.16)$$

where 546 is the wavelength in nm. J_{max} and J are the maximum and actual rate of electron transport (both in $\mu\text{mol e} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$) and $c = 0.9$ defines the curvature of the light-response curve. The gross assimilation rate A at a given C_i is the minimum of A_e and A_c . To obtain a smooth A -vs- C_i curve we used (Collatz *et al.* 1991):

$$A = \frac{(A_c + A_e) - \sqrt{(A_c + A_e)^2 - 4.0 \cdot c' \cdot A_e \cdot A_c}}{2 \cdot c'} \quad (11.17)$$

where $c' = 0.98$ is a curvature factor. The temperature dependence of J_{max} and V_{max} relative to 25°C (i.e., relative to input parameters $J_{max,298}$ and $V_{max,298}$) is modelled using Leuning (2002) (his eq. 1 with

parameters from his Table 2). The internal CO_2 concentration, C_i , needs to be known to calculate A using the previous equations. Sperry *et al.* (2016) use a second equation for A which uses g_c , the diffusive conductance to CO_2 (which includes both the stomatal conductance and boundary layer conductance):

$$A = g_c \cdot (C_{air} - C_i) \quad (11.18)$$

where C_{air} is the CO_2 concentration (in $\mu mol \cdot mol^{-1}$) in the air surrounding the leaf (beyond the leaf boundary layer). If a single-layer canopy energy balance is used then $C_{air} = C_{atm}$, the atmospheric CO_2 concentration (see weather input and parameter C_{atm} in function `defaultControl()`), whereas if a multi-layer canopy energy balance is used C_{air} corresponds to $C_{air,j}$, the CO_2 concentration in the canopy layer j where the leaf occurs, analogously to T_{air} and e_{air} . Combining the two equations for A and finding the root of the resulting equation using Newton-Raphson method allows determining C_i and therefore A . Thus, after defining PAR photon flux density, atmosphere CO_2 concentration and maximum rate parameters:

```
Q = 2000
Catm = 386
Vmax298 = 100
Jmax298 = 1.67*Vmax298
```

one can obtain the following $A(\Psi_{leaf})$ curves from $T_{leaf}(\Psi_{leaf})$ and $g_{sw}(\Psi_{leaf})$:

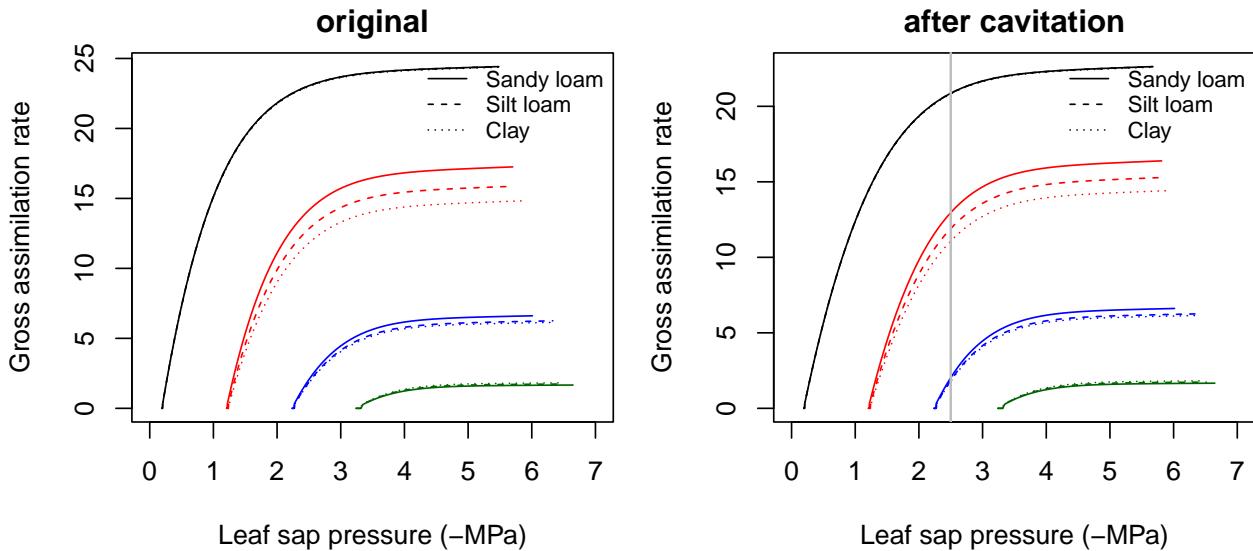


Figure 11.5: Examples of gross photosynthesis (A) functions for a hydraulic network, corresponding to fig. 10.15 and for different soil textures. Left/right panel shows values for uncavitated/cavitated supply functions.

Finally, leaf net photosynthesis (A_n ; i.e. accounting for autotrophic respiration) is calculated as:

$$A_n = A - 0.015 \cdot V_{max} \quad (11.19)$$

11.2 Crown/canopy photosynthesis

In the previous subsection we showed how to calculate photosynthesis at the leaf level, but gross or net assimilation needs to be up-scaled to the crown or canopy levels, while accounting for the variation of leaf photosynthetic conditions across canopies. In this section, we briefly describe different alternatives for canopy/crown photosynthesis and justify our modelling choice.

11.2.1 Multi-layer canopy photosynthesis model

Ideally, crown or canopy photosynthesis requires repeating the calculations of leaf temperature, leaf VPD, leaf gas conductance and photosynthesis for every leaf. Multi-layer canopy models, however, divide the canopy into vertical layers and determine photosynthesis for each one (and each plant species or plant cohort, if required). Multi-layer models can account for the fact that environmental variables usually vary across the crown, including direct/diffuse absorbed radiation, air temperature, wind speed, water vapor pressure and CO_2 concentration. The amount of SWR and PAR absorbed per unit of leaf area at each canopy layer was shown in fig. 9.2.

Not only environmental factors, but leaves themselves may be different across canopy layers. Importantly, it is generally accepted that sunlit and shade leaves need to be treated separately (De Pury & Farquhar 1997). Separating the two kinds of leaves acknowledges that they operate at different parts of the light-saturation curve. Following De Pury & Farquhar (1997), we further assume that maximum carboxylation and electron transport rates are highest for leaves at the top of the canopy and there is an exponential decrease from there towards the bottom, where maximum rates are 50% of those at the top:

$$V_{max,298,i,j} = V_{max,298,i} \cdot \exp(-0.713 \cdot \sum_{h>i} LAI_{h,j}^\phi / LAI_i^\phi) \quad (11.20)$$

$$J_{max,298,i,j} = J_{max,298,i} \cdot \exp(-0.713 \cdot \sum_{h>i} LAI_{h,j}^\phi / LAI_i^\phi) \quad (11.21)$$

where $LAI_{i,j}^\phi$ is the LAI value of the plant cohort i at a given canopy layer j and LAI_i^ϕ is the expanded LAI of the plant cohort. The following figure illustrates this decrease for the single-species canopy example of section 9.1.3:

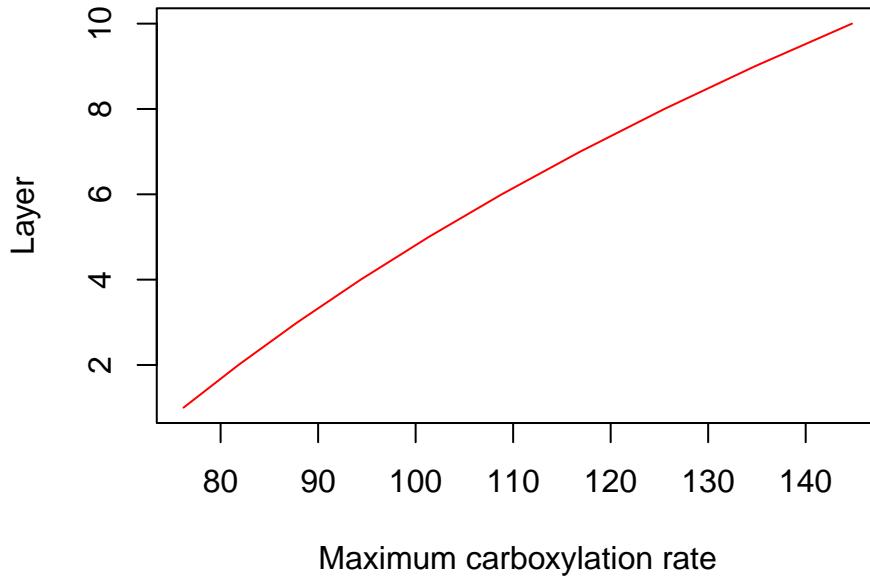


Figure 11.6: Decrease of Rubisco maximum carboxylation rate across the canopy

In a multi-layer canopy photosynthesis model, gross and net photosynthesis values (i.e. A and An) are determined for sunlit and shade leaves of each cohort in each canopy layer. Then, sunlit and shade photosynthesis values should be averaged across the crown for each plant cohort. Assuming that Ψ_{leaf} is equal for all leaves across the crown, the function $A(\Psi_{leaf})$ would be obtained for each plant cohort.

11.2.2 Big-leaf canopy photosynthesis model

Multi-layer canopy photosynthesis models allow evaluating leaf conditions, stomatal conductance and photosynthesis for different points of the canopy. However, this comes at high computational cost. For this reason, many models implement what is called the *big-leaf approximation*. Assuming that wind speed, temperature, water vapor pressure and CO_2 concentration are similar for all leaves and that the distribution of photosynthetic capacity between leaves is in proportion to the profile of absorbed irradiance then the equation describing leaf photosynthesis will also represent canopy photosynthesis (Sellers et al. 1992).

11.2.3 Sun-shade canopy photosynthesis model

An alternative between multi-layer and big-leaf canopy photosynthesis models is to collapse variation of photosynthetic conditions into two leaf classes: sunlit and shade leaves. While big-leaf canopy models are known to be unaccurate under some situations, sun-shade canopy models (De Pury & Farquhar 1997) provide estimates that are close to multiple layer models (Hikosaka *et al.* 2016). The sun-shade canopy photosynthesis model was adopted here. Assuming that wind speed, temperature, water vapor pressure and CO_2 concentration are similar for all leaves, sun-shade models involve the following steps:

- Aggregate the leaf area of sunlit/shade leaves across layers:

$$LAI_i^{sunlit} = \sum_{j=1}^l LAI_{i,j}^{sunlit} \quad (11.22)$$

$$LAI_i^{shade} = \sum_{j=1}^l LAI_{i,j}^{shade} \quad (11.23)$$

where $LAI_{i,j}^{sunlit}$ and $LAI_{i,j}^{shade}$ are the leaf area index of sunlit and shade leaves for cohort i in canopy layer j , from eq. (9.5).

- Average the SWR/PAR absorbed by leaves of each kind across layers. The average light absorbed by sunlit/shaded foliage of cohort i per ground area unit is found using:

$$\Phi_{abs,i}^{sunlit} = \frac{\sum_{j=1}^l K_{abs,i,j}^{sunlit}}{LAI_i^{sunlit}} \quad (11.24)$$

$$\Phi_{abs,i}^{shade} = \frac{\sum_{j=1}^l K_{abs,i,j}^{shade}}{LAI_i^{shade}} \quad (11.25)$$

where $K_{abs,i,j}^{sunlit}$ and $K_{abs,i,j}^{shade}$ are the light absorbed per ground area unit by sunlit/shade leaves of cohort i at layer j (see section 9.1.3). Analogous equations were already given for the net long-wave radiation balance of sunlit leaves ($L_{net,i}^{sunlit}$) and shade leaves ($L_{net,i}^{shade}$) in section 9.2.3.

- Average the maximum carboxylation (respectively, electron transport) rates across layers, again separating sunlit and shade leaves:

$$V_{max,298,i}^{sunlit} = \frac{\sum_{j=1}^l V_{max,298,i,j} \cdot LAI_{i,j}^{sunlit}}{LAI_i^{sunlit}} \quad (11.26)$$

$$V_{max,298,i}^{shade} = \frac{\sum_{j=1}^l V_{max,298,i,j} \cdot LAI_{i,j}^{shade}}{LAI_i^{shade}} \quad (11.27)$$

- Use $V_{max,298,i}^{sunlit}$ as $V_{max,298}$ in Leuning (2002) to obtain V_{max} for eq. (11.10); $\Phi_{SWR,i}^{sunlit}$ as Φ_{SWR}^{leaf} and $L_{net,i}^{sunlit}$ as L_{net}^{leaf} in eq. (11.1); and $\Phi_{PAR,i}^{sunlit}$ as Φ_{PAR}^{leaf} in eq. (11.16) to estimate sunlit leaf photosynthesis, which can be up-scaled to the crown level multiplying by LAI_i^{sunlit} . The same would be done for shade leaves. In a sun-shade canopy model one then calls the photosynthesis function twice (i.e. once for shade leaves and once for sunlit leaves) for each plant cohort i .

11.2.4 Comparison of big-leaf, sun-shade and multi-canopy photosynthesis models

The figure below provides the canopy photosynthesis functions obtained using the multi-layer canopy photosynthesis model (top), a sunshade canopy photosynthesis model (center) or a big-leaf photosynthesis model (bottom). These were generated using functions `photo_multilayerPhotosynthesisFunction()`, `photo_sunshadePhotosynthesisFunction()` and `photo_leafPhotosynthesisFunction()`, respectively, and assuming homogeneous wind, temperature and water vapor pressure through the canopy. Thus, only absorbed radiation varied across layers and leaf types. Note the coincidence between the multi-layer and the sun-shade models.

11.2.5 Within-canopy variation in environmental conditions

When presenting the sun-shade photosynthesis model, we assumed constant wind, temperature, water vapor pressure and CO_2 concentration values through the canopy. If a single-layer canopy energy balance is used, CO_2 concentration and vapor pressure are assumed equal to the atmosphere (i.e., $e_{air} = e_{atm}$ and $C_{air} = C_{atm}$), whereas air temperature is that of the canopy (i.e., $T_{air} = T_{can}$), but leaf-level wind speed (u_{leaf}) can still be different for different canopy layers. If a multi-layer canopy energy balance is used, all four environmental variables can differ between canopy layers (i.e. $T_{air,j}$, $e_{air,j}$, $C_{air,j}$ and u_j). In the sun-shade photosynthesis model, the question arises on how to determine layer j for sunlit or shade leaves of a given cohort i . The choice is done by calculating the height corresponding to the mass center of sunlit leaves or shade leaves of each cohort i . The canopy layer j where this mass center height is contained is chosen as the layer from which environmental conditions will be taken. For any given plant cohort i , sunlit leaves will take their environmental conditions from layers above (or equal to) those corresponding to shade leaves.

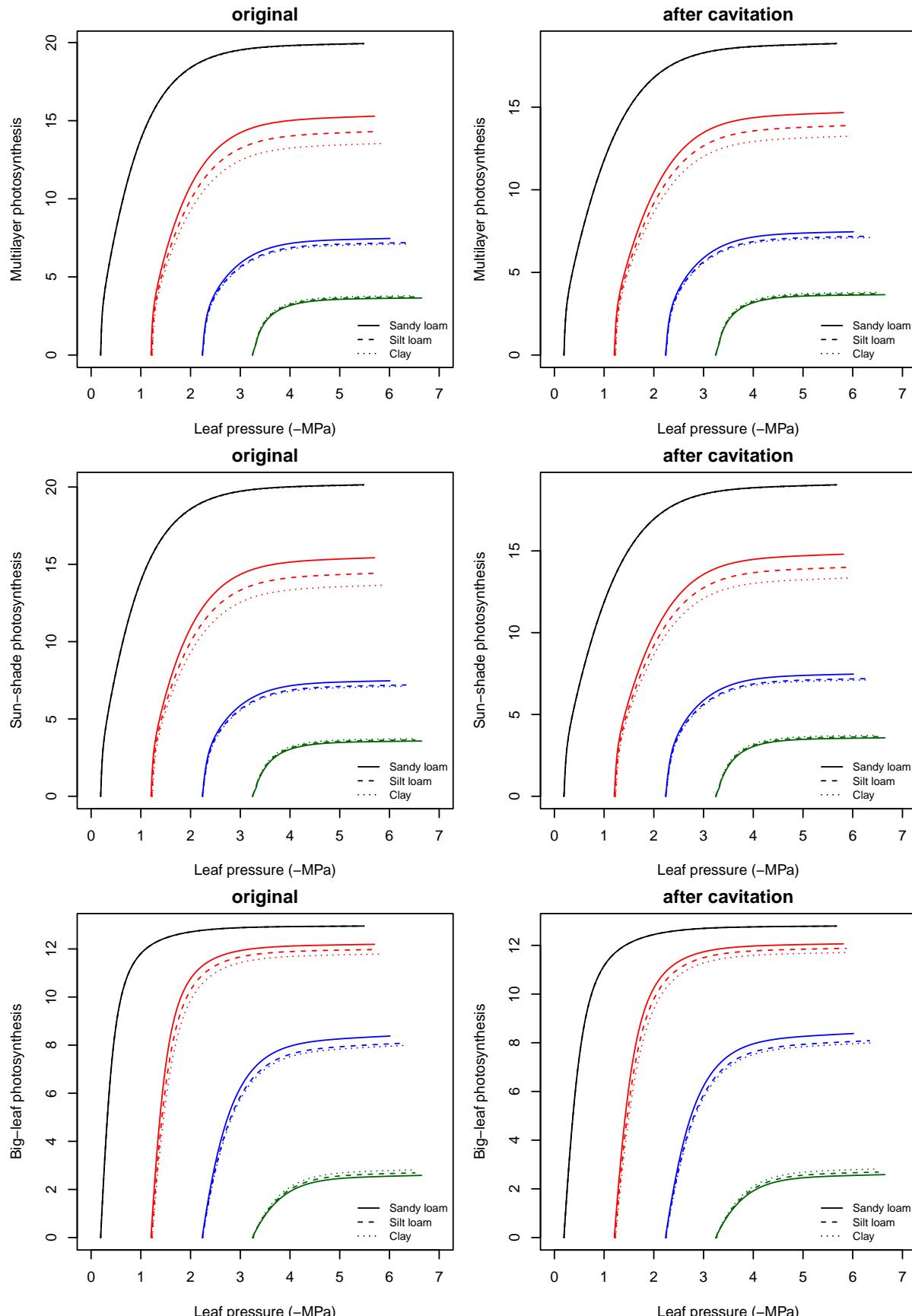


Figure 11.7: Whole-canopy photosynthesis functions obtained for a hydraulic network, corresponding to fig. 10.15 and different soil textures, using the multi-layer canopy photosynthesis model (top), a sunshade canopy photosynthesis model (center) or a big-leaf photosynthesis model (bottom). Left/right panel shows values for uncavitated/cavitated supply functions.

Chapter 12

Stomatal regulation, transpiration and photosynthesis

Plants must open their stomata to acquire CO_2 and perform photosynthesis, but doing so promotes water loss. This trade-off has resulted in a tight coordination between capacity to supply and transpire water (hydraulic conductance and diffusive conductance to water vapor) and the maximum capacity for photosynthesis (carboxylation rate and electron-transport rate). For modelling purposes, this carbon-for-water trade-off means that hydraulics, stomatal conductance, transpiration and photosynthesis need to be estimated simultaneously. In chapters 10 and 11 we introduced plant hydraulics and photosynthesis, respectively, but we did not explain how are actual transpiration and photosynthesis values determined. This depends on the sub-model chosen and is the subject of the present chapter.

12.1 Stomatal regulation under Sperry's sub-model

The framework of Sperry *et al.* (2017) suggests estimating stomatal conductance from the instantaneous maximization of profit, defined as the difference between photosynthesis gain and hydraulic cost (both normalized for comparability). First, transpiration and photosynthesis is estimated separately for sunlit and shade leaves according to this framework. An average transpiration rate is then determined depending on sunlit/shade transpiration rates and their leaf area contributions. Finally, the instantaneous transpiration and assimilation rates of each time step are scaled to the duration of the time step and to the leaf area of the plant cohort. The following sections provide details for all these steps.

12.1.1 Water supply and sunlit/shade photosynthesis functions

Let us start by summarizing the concepts introduced in the last two chapters. The **supply function** described in chapter 10 describes the rate of transpiration flow as a function of the pressure drop between the soil and the leaf, and incorporates both soil, xylem and leaf hydraulic constrains (Sperry *et al.* 1998, 2016; Sperry & Love 2015). Here we assume that hydraulic conductance k is in $mmol\ H_2O \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$ of leaf area, transpiration rate E_{leaf} in $mmol\ H_2O \cdot s^{-1} \cdot m^{-2}$ of leaf area and leaf water potential Ψ_{leaf} is in MPa.

According to section 11.1, each E_{leaf} value implies an energy balance at the leaf level, stomatal conductance to water vapour and a particular value of leaf photosynthesis. More specifically, for each pair of E_{leaf} and Ψ_{leaf} values, we have a corresponding leaf temperature (T_{leaf} ; in $^{\circ}C$), leaf-to-air vapor pressure deficit (VPD_{leaf} ; in kPa), leaf water vapor conductance (g_{sw} ; in $mol\ H_2O \cdot s^{-1} \cdot um^{-2}$) and, finally the leaf gross and net (i.e. after accounting for autotrophic respiration) photosynthesis assimilation rates (A_g and A_n ;

both in $\mu\text{mol CO}_2 \text{us}^{-1} \text{um}^{-2}$). In short, the supply function generates a **photosynthesis function**. Since the model deals with canopies and not single leaves, however, different parts of the crowns of plant cohorts may be in different canopy positions. Calculating photosynthesis at the canopy level requires dividing the canopy into vertical layers, differentiating between *sunlit* and *shade* leaves and determining **photosynthesis functions for sunlit and shade leaves separately**, as explained in section 11.2.

12.1.2 Leaf stomatal regulation by profit maximization

Wolf *et al.* (2016) proposed the **carbon maximization criterion**, which states that at each instant in time the stomata regulate canopy gas exchange and pressure to achieve the maximum profit, which is the maximum difference between photosynthetic gains and costs, the latter associated with hydraulic vulnerability attained with low water potentials. Such approach has been shown to be supported by data from global forest biomes (Anderegg *et al.* 2018). Building on this approach, Sperry *et al.* (2017) presented a profit maximization function where hydraulic costs of opening the stomata are compared against photosynthetic gains. Details of their formulation are given in this section. Stomatal regulation is performed in **medfate** separately for sunlit and shade leaves.

12.1.2.1 Cost and gain functions

The hydraulic supply function is used to derive a transpirational *cost function* $\theta_1(\Psi_{leaf})$ that reflects the increasing damage from cavitation and the greater difficulty of moving water along the continuum (Sperry *et al.* 2016):

$$\theta(\Psi_{leaf}) = \frac{k_{c,max} - k_c(\Psi_{leaf})}{k_{c,max} - k_{crit}} \quad (12.1)$$

where $k_c(\Psi_{leaf}) = dE/d\Psi(\Psi)$ is the slope of the supply function corresponding to (leaf) water potential Ψ_{leaf} , $k_{c,max}$ is the maximum slope of the supply function (which occurs when $E_{leaf} = 0$), i.e. the maximum whole-plant conductance for the current soil moisture conditions, and $k_{crit} = k_c(\Psi_{crit})$ is the slope of the supply function at $E_{leaf} = E_{crit}$ the critical flow beyond which hydraulic failure occurs.

The following figures illustrate the $\theta(\Psi_{leaf})$ curves corresponding to the supply functions:

The normalized photosynthetic *gain function* $\beta(\Psi_{leaf})$ reflects the actual assimilation rate with respect to the maximum (Sperry *et al.* 2016):

$$\beta(\Psi_{leaf}) = \frac{A(\Psi_{leaf})}{A_{max}} \quad (12.2)$$

where A_{max} is the instantaneous maximum (gross) assimilation rate estimated over the full Ψ_{leaf} range (including values that imply a stomatal conductance larger than the maximum).

The following figures illustrate the $\beta(\Psi_{leaf})$ curves corresponding to the supply and assimilation functions:

12.1.2.2 Profit maximization at the leaf level

Sperry *et al.* (2017) suggested that stomatal regulation can be effectively estimated by determining the maximum of the *profit function* ($Profit(\Psi_{leaf})$):

$$Profit(\Psi_{leaf}) = \beta(\Psi_{leaf}) - \theta(\Psi_{leaf}) \quad (12.3)$$

The maximization is achieved when the slopes of the gain and cost functions are equal:

$$\frac{\delta\beta(\Psi_{leaf})}{\delta\Psi_{leaf}} = \frac{\delta\theta(\Psi_{leaf})}{\delta\Psi_{leaf}} \quad (12.4)$$

Instantaneous profit maximization assumes a ‘use it or lose it’ reality with regards to available soil water. Because the gain function accelerates more quickly from zero and reaches 1 sooner than the cost function, their

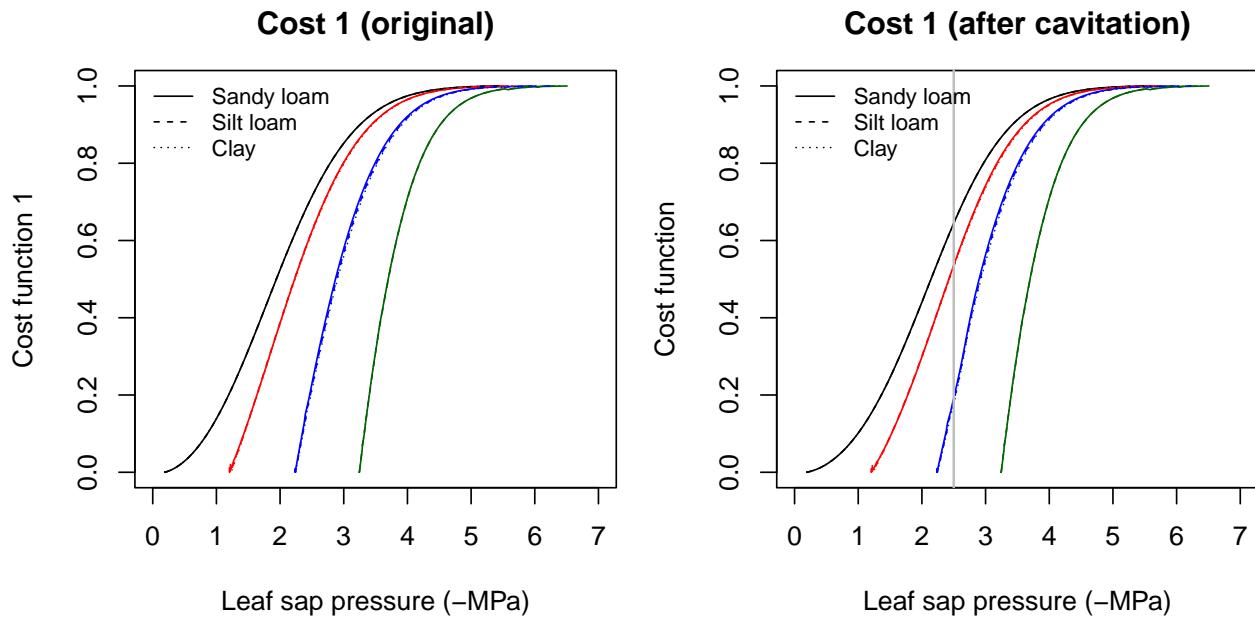


Figure 12.1: Cost functions (i.e. $\theta(\Psi_{leaf})$) obtained for a hydraulic network, corresponding to fig. 10.15 and different soil textures and soil water potentials. Left/right panels show values for uncavitated/cavitated supply functions, respectively.

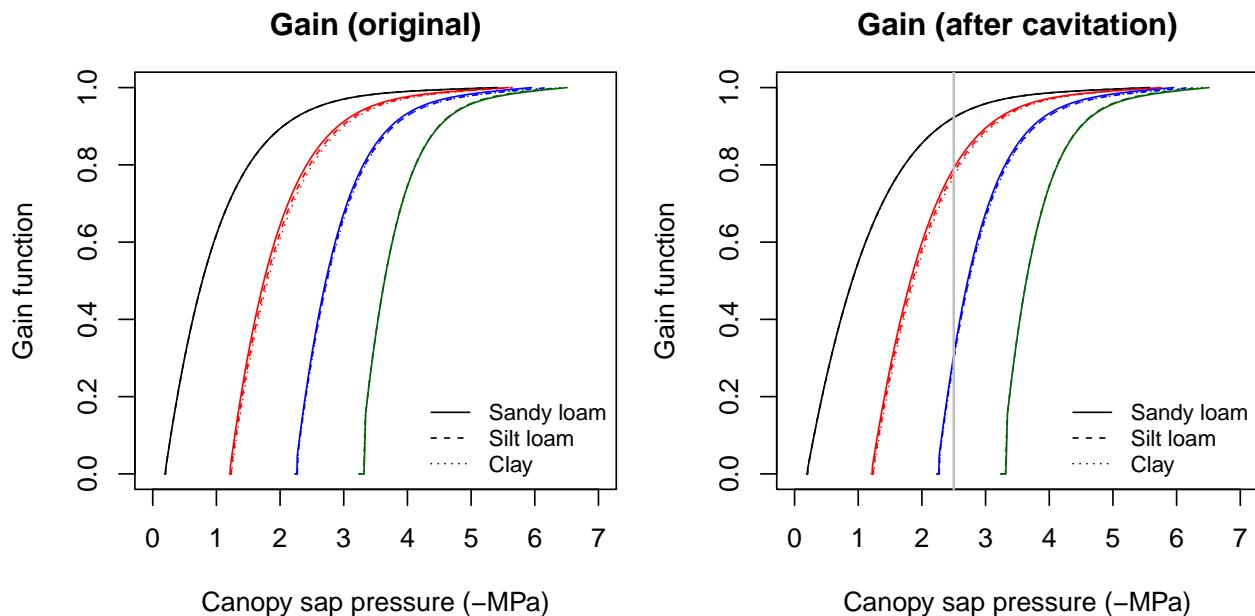


Figure 12.2: Gain function ($\beta(\Psi_{leaf})$) obtained for a hydraulic network, corresponding to fig. 10.15 and different soil textures and soil water potentials. Left/right panels show values for uncavitated/cavitated supply functions, respectively.

maximum difference occurs at intermediate Ψ_{leaf} values. Once Ψ_{leaf} that maximizes profit is determined, the values of the remaining variables are also determined. At this point, it may happen that $g_{sw}(\Psi_{leaf})$ is lower than the minimum (i.e. cuticular) water vapor conductance ($g_{sw,min}$) or larger than the maximum water vapor conductance ($g_{sw,max}$). These thresholds need to be taken into account when determining the maximum of the profit function.

The following figures illustrate the $Profit(\Psi_{leaf})$ curves corresponding to the previous cost and gain curves:

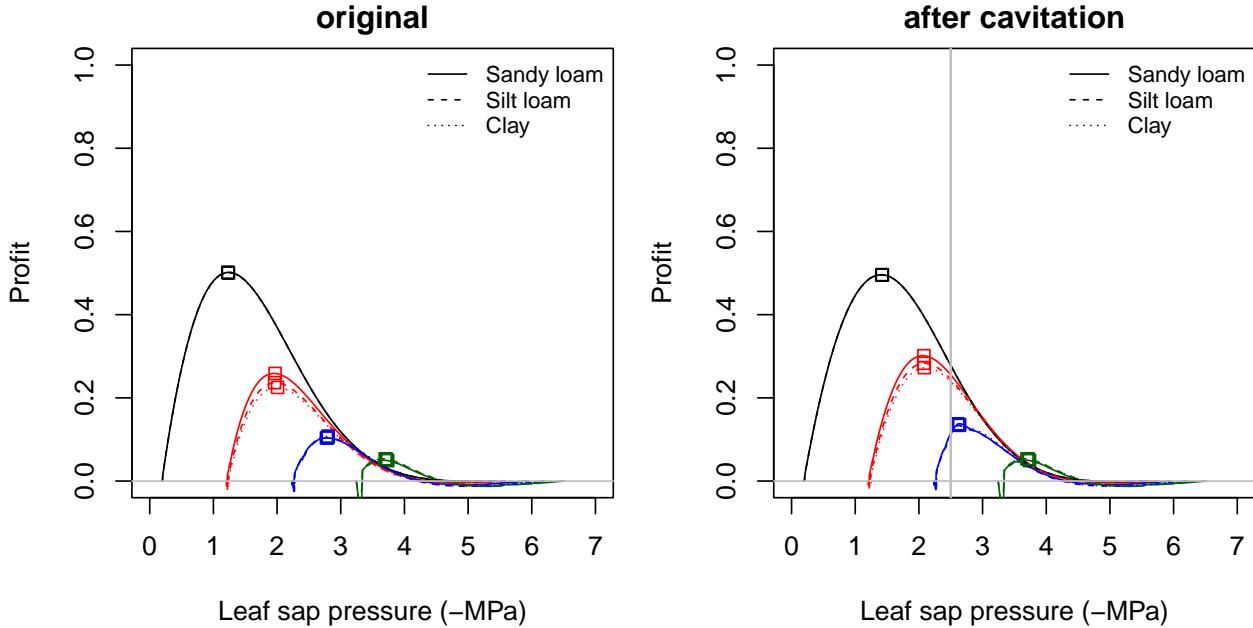


Figure 12.3: Profit functions (i.e. $Profit(\Psi_{leaf})$) obtained for a hydraulic network, corresponding to fig. 10.15 and different soil textures and soil water potentials. Left/right panels show values for uncavitated/cavitated supply functions, respectively.

Squares in the previous figures indicate the maximum profit points in each situation. The drier the soil, the closer is the maximum profit Ψ_{leaf} to soil water potential as one would expect intuitively (i.e. a smaller drop in water potential along the hydraulic pathway). Note that when the soil is very dry the squares are to the right of the true maximum. This is because the `transp_profitMaximization()` function takes into account the minimum and maximum stomatal conductance and, in this case, does not allow optimum stomatal conductances below the minimum (cuticular) value.

Note that θ , β and $Profit$ functions can be expressed as a function of stomatal conductance, instead of leaf water potential. This allows visualizing more clearly the effect of $g_{sw,min}$ and $g_{sw,max}$ thresholds on the maximum profit optimization strategy, as illustrated in the following figures.

12.2 Stomatal regulation under Cochard's sub-model

[TO BE DESCRIBED]

12.3 Cohort-level transpiration and photosynthesis

In the previous section, we considered stomatal regulation at the leaf level only. At the plant cohort level, the gain function could be build from a crown photosynthesis function $A(\Psi_{leaf})$, as shown in section 11.2.

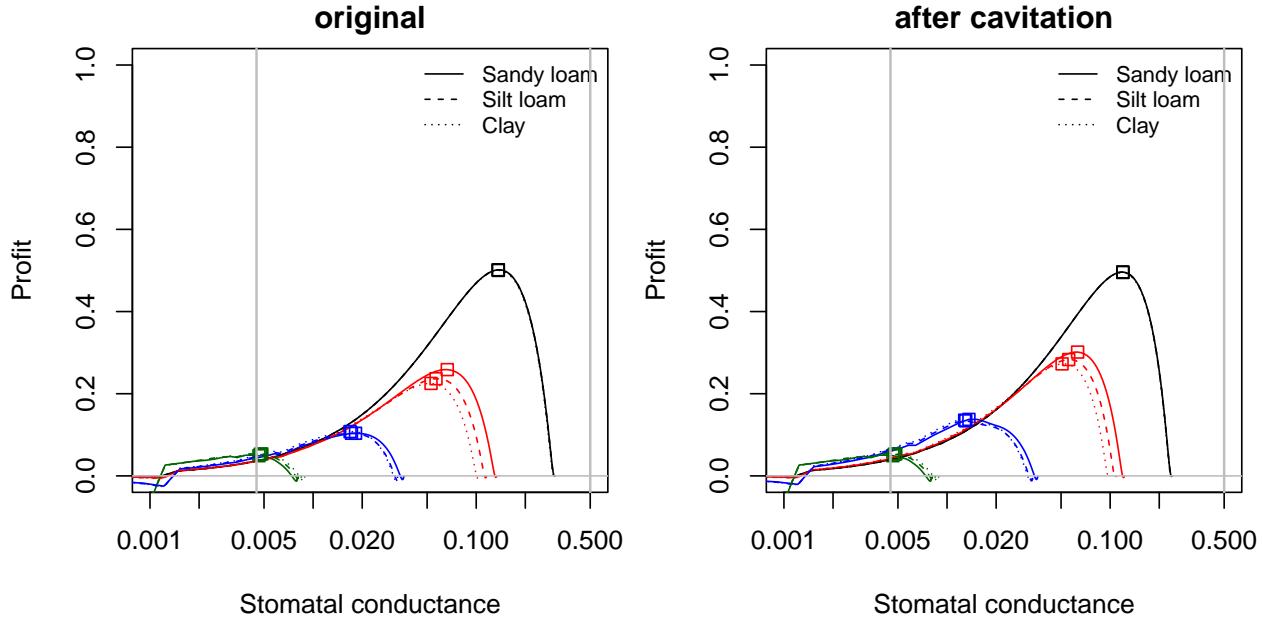


Figure 12.4: Profit function as a function of stomatal conductance, corresponding to fig. 12.3 and different soil textures and soil water potentials. Left/right panels show values for uncavitated/cavitated supply functions, respectively.

However, applying the profit maximization approach for a single crown photosynthesis function would imply the assumption that the same stomatal aperture occurs in all leaves of the plant cohort, independently of whether they are in shade or sunlit. A more realistic approach is to determine stomatal regulation by profit maximization for sunlit and shade leaves separately. The gain function and profit maximization calculations conducted for each leaf type yield instantaneous leaf water potentials Ψ_{leaf}^{sunlit} and Ψ_{leaf}^{shade} and instantaneous flow values E^{shade} and E^{sunlit} from the supply functions. The corresponding photosynthesis functions allow determining values for leaf temperatures T_{leaf}^{sunlit} and T_{leaf}^{shade} , vapour pressure deficits VPD_{leaf}^{sunlit} and VPD_{leaf}^{shade} , stomatal conductance g_{sw}^{sunlit} and g_{sw}^{shade} and net photosynthesis rates A_n^{sunlit} and A_n^{shade} . This is a lot of useful information at the leaf level for each plant cohort i , but we also need transpiration and photosynthesis values at the plant cohort-level.

The average instantaneous flow rate ($E_i^{average}$, in $mmol H_2O \cdot s^{-1} \cdot m^{-2}$) per leaf area unit of plant cohort i is the weighed average:

$$E_i^{average} = \frac{E_i^{shade} \cdot LAI_i^{sunlit} + E_i^{sunlit} \cdot LAI_i^{shade}}{LAI_i^\phi} \quad (12.5)$$

where LAI_i^{sunlit} and LAI_i^{shade} are the cohorts LAI values for sunlit and shade leaves, from eq. (9.5), and LAI_i^ϕ is the leaf area index of plant cohort i . The model then uses the hydraulic supply function to find the transpiration rate E_i numerically closest to $E_i^{average}$ (remember that the supply function is build in discrete steps). Finding the E_i value numerically closest to $E_i^{average}$ determines the $E_{i,t}$ the instantaneous transpiration flow for current time step t , and also leads to setting values for t of several other variables, such as water potentials ($\Psi_{leaf,i,t}$, $\Psi_{stem,i,t}$, $\Psi_{rootcrown,i,t}$, ...), the slope of the supply function ($(dE/d\Psi_i)_t$), and instantaneous soil water extraction rates ($E_{i,s,t}$).

The amount of transpiration from the plant cohort in a given time step t ($Tr_{i,t}$, in $l \cdot m^{-2}$ of ground area, i.e. in mm) is:

$$Tr_{i,t} = E_{i,t} \cdot LAI_i^\phi \cdot 10^{-3} \cdot 0.01802 \cdot \Delta t_{step} \quad (12.6)$$

where 0.01802 is the molar weight (in $kg = l$) of water and $\Delta t_{step} = 86400/n_t$ the size of the time step in seconds, being n_t the number of time steps. Soil extraction rates (Ex) are scaled to the cohort level as done

for transpiration:

$$Ex_{i,s,t} = E_{i,s,t} \cdot LAI_i^\phi \cdot 10^{-3} \cdot 0.01802 \cdot \Delta t_{step} \quad (12.7)$$

If plant water storage is not considered, one should have that soil extraction equals transpiration:

$$Tr_{i,t} = Ex_{i,t} = \sum_s Ex_{i,s,t} \quad (12.8)$$

Instantaneous net photosynthesis per leaf area of sunlit and shade leaves in a given time step t (i.e. $A_{n,i,t}^{sunlit}$ and $A_{n,i,t}^{shade}$) are also aggregated into $A_{n,i,t}$, the net photosynthesis of the plant cohort i for the time step t , in $\text{g C} \cdot \text{m}^{-2}$ of ground area:

$$A_{n,i,t} = (A_{n,i,t}^{sunlit} \cdot LAI_i^{sunlit} + A_{n,i,t}^{shade} \cdot LAI_i^{shade}) \cdot 10^{-6} \cdot 12.01017 \cdot \Delta t_{step} \quad (12.9)$$

12.4 Stem cavitation and conduit refilling

Like with the ‘Granier’ transpiration mode, the water balance model with ‘Sperry’ transpiration mode is normally run assuming that although soil drought may reduce transpiration, embolized xylem conduits are automatically refilled when soil moisture recovers (i.e. `cavitation = "total"`). Automatic refilling is always assumed for leaf and root segments, but not for stems. There are three other options to deal with stem xylem cavitation besides automatic refilling. Any of them cause the model to keep track of the proportion of lost conductance in the stem xylem of the plant cohort i ($PLC_{stem,i}$) at successive time steps:

$$PLC_{stem,i,t} = \max\{PLC_{stem,i,t-1}, 1 - k_{stem,i}(\Psi_{stem})/k_{max,stem,i}\} \quad (12.10)$$

When water compartments are not considered, this equation is evaluated every hour time step, whereas it is evaluated every 1s with water compartments, as part of the constitutive equations (see 10.5.2).

For simulations of less than one year one can use `cavitationRefill = "none"` to keep track of the maximum drought stress. However, for simulations of several years, it is normally advisable to allow recovery. If `cavitation = "annual"`, $PLC_{stem,i}$ values are set to zero at the beginning of each year, assuming that embolized plants overcome the conductance loss by creating new xylem tissue. Finally, if `cavitationRefill = "rate"` the model simulates stem refilling at daily steps as a function of symplasmic water potential. First, a daily recovery rate (r_{refill} ; in $\text{cm}^2 \cdot \text{m}^{-2} \cdot \text{day}^{-1}$) is estimated as a function of $\Psi_{symp,stem}$:

$$r_{refill}(\Psi_{symp,stem}) = r_{max,refill} \cdot \max\{0.0, (\Psi_{symp,stem} + 1.5)/1.5\} \quad (12.11)$$

Where $r_{max,refill}$ is the control parameter `refillMaximumRate` indicating a maximum refill rate. The right part of the equation normalizes the water potential, so that $r_{refill} = r_{refill,max}$ if $\Psi_{symp,stem} = 0$ and $r_{refill} = 0$ if $\Psi_{symp,stem} <= -1.5 \text{ MPa}$. The proportion of conductance lost is then updated using:

$$PLC_{stem,i} = \max\{0.0, PLC_{stem,i} - (r_{refill}/H_v)\} \quad (12.12)$$

where H_v is the Huber value in units of $\text{cm}^2 \cdot \text{m}^{-2}$.

Chapter 13

Closing energy balances

In chapter 9 we described the details of radiation energy transfer. Transpiration is an important latent heat component of the energy balance, and the details of how plant transpiration is determined were given in chapters 10, 11 and 12. This chapter explains how to calculate the remaining components of the canopy energy balance, while distinguishing between assuming a single-layer canopy or considering a multiple-layer canopy.

13.1 Single-layer canopy

The first part of the section describes the calculation of latent heat and convective energy exchanges between the soil, canopy and atmosphere layers. These are the remaining energy components that are needed to close the energy balances of the canopy (eq. (7.4)) and soil layers (eqs. (7.6) and (7.7)). Temperature changes applied to the canopy and soil layers are also described.

13.1.1 Latent heat

Canopy exchanges of latent heat include latent heat exchanged from plant transpiration and evaporation of rain intercepted by the canopy. In the single-layer canopy energy balance, latent heat exchange for the whole canopy (LE_{can} ; in $W \cdot m^{-2}$) is calculated as:

$$LE_{can} = \lambda_v(T_{can}) \cdot \left(\frac{In_t + \sum_{i=1}^c Tr_{i,t}}{\Delta t_{step}} \right) \quad (13.1)$$

where $\lambda_v(T_{can})$ is the latent heat of vaporization at temperature T_{can} (in $J \cdot kg^{-1}$), $Tr_{i,t}$ is the amount (in $mm = kg \cdot m^{-2}$) of transpiration for cohort i during time step t (12.6), and In_t is the amount of intercepted water evaporated from the canopy for time step t (total daily interception In is divided into timesteps according to the proportion of shortwave radiation absorbed by the canopy at each time step).

The latent heat exchange between the soil and the atmosphere (LE_{soil} ; in $W \cdot m^{-2}$) comes either from water evaporated from the soil surface or snow melting (both in $mm = kg \cdot m^{-2}$). They do not occur simultaneously, but can be expressed in the same equation:

$$LE_{soil} = \lambda_v(T_{soil}) \cdot \left(\frac{Es_t + \lambda_f \cdot Sm_t}{\Delta t_{step}} \right) \quad (13.2)$$

where $\lambda_v(T_{soil})$ is the latent heat of vaporization at temperature T_{soil} and $\lambda_f = 0.33355 J \cdot kg^{-1}$ is the latent heat of fusion. As before, daily evaporation from bare soil (Es) and snow melt water equivalent (Sm) are divided into time steps according to the proportion of shortwave radiation absorbed by the soil layer each step.

13.1.2 Convective energy

Convective energy fluxes between atmosphere and the canopy ($H_{can,atm}$) and between the canopy and the soil ($H_{can,soil}$) are determined as follows:

$$H_{can,atm} = \frac{\rho_{atm} \cdot c_p}{r_{can,atm}} \cdot (T_{can} - T_{atm}) \quad (13.3)$$

$$H_{can,soil} = \frac{\rho_{can} \cdot c_p}{r_{can,soil}} \cdot (T_{can} - T_{soil,1}) \quad (13.4)$$

where ρ_{atm} and ρ_{can} are the air density above-canopy and inside-canopy, respectively, calculated from the corresponding temperatures (see utility functions in meteoland reference manual); $c_p = 1013.86 \text{ J} \cdot \text{kg}^{-1} \cdot \text{K}^{-1}$ is the specific heat capacity of the air. $r_{can,atm}$ and $r_{can,soil}$ are the atmosphere-canopy and canopy-soil aerodynamic resistances (in $\text{s} \cdot \text{m}^{-1}$). These, in turn, are calculated using the FAO56 calculation procedure and canopy height, total LAI and above-canopy and below-canopy wind speeds. Wind speed below the canopy is calculated as explained in 23.2 assuming a height of 2 m.

13.1.3 Canopy temperature changes

After evaluating the canopy energy balance equation (7.4) one has to translate energy balance into temperature variation of the canopy. Rearranging the same equation and expressing it in discrete time steps we have:

$$\Delta T_{can} = \Delta t_{step} \cdot \frac{K_{abs,can} + L_{net,can} - LE_{can} - H_{can,atm} - H_{can,soil}}{TC_{can}} \quad (13.5)$$

where TC_{can} is the canopy thermal capacitance (in $\text{J} \cdot \text{m}^{-2} \cdot \text{K}^{-1}$) and $\Delta t_{step} = 86400/n_t$ is the size of the time step in seconds. Canopy thermal capacitance depends on the leaf area index of the stand:

$$TC_{can} = TC_{LAI} \cdot \frac{0.8 \cdot LAI_{stand} + 1.2 \cdot (LAI_{stand}^{phi} + LAI_{stand}^{dead})}{2} \quad (13.6)$$

where TC_{LAI} is the thermal capacitance per LAI unit, which is specified by the control parameter `thermalCapacityLAI`. By using both the maximum leaf area of the stand, LAI_{stand} , and its current live/dead leaf area ($LAI_{stand}^{phi} + LAI_{stand}^{dead}$) it is assumed that part of thermal capacitance corresponds to stems and branches, so that capacitance does not drop to zero for deciduous canopies.

13.1.4 Soil temperature changes

Analogously to the canopy, the change in temperature for a given soil layer s in the current time step is given by rearranging eq. (7.7):

$$\Delta T_{soil,s} = \Delta t_{step} \cdot \frac{G_{s-1,s} - G_{s,s+1}}{C_{soil,s}} \quad (13.7)$$

where the energy balance on each soil layer s depends on the balance between energy coming from above ($G_{s-1,s}$) and energy going to below ($G_{s,s+1}$) and $C_{soil,s}$, the thermal capacitance of soil layer s . Energy inflow to the first (uppermost) layer (i.e. $G_{0,1}$) is the result of energy exchanges between the soil layer and the canopy and atmosphere, i.e. from eq. (7.6):

$$G_{0,1} = K_{soil} + L_{net,soil} + H_{can,soil} - LE_{soil} \quad (13.8)$$

Heat conduction between layers s and $s+1$ (i.e. $G_{s,s+1}$) depend on the soil temperature gradient (see function `soil_temperatureGradient()`):

$$G_{s,s+1} = \lambda_{soil,s} \cdot \frac{\delta T_{soil,s}}{\delta z} = \lambda_{soil,s} \cdot \frac{T_{soil,s} - T_{soil,s+1}}{(Z_{s-1} - Z_{s+1})/2} \quad (13.9)$$

where Z_{s-1} and Z_{s+1} are expressed in m and $\lambda_{soil,s}$ is the *thermal conductivity* of layer s , calculated from soil moisture and texture following Dharssi *et al.* (2009) (see function `soil_thermalconductivity()`). The gradient in the lowermost layer is calculated assuming a temperature of the earth (at 10 m) of 15.5 Celsius.

Finally, $C_{soil,s}$ the thermal capacitance of soil layer s is calculated as:

$$C_{soil,s} = VTC_{soil,s} \cdot d_s \quad (13.10)$$

where d_s is the soil width of layer s (expressed in m) and VTC_s is the *volumetric thermal capacity* of soil layer s (in $J \cdot m^{-3} \cdot K^{-1}$), calculated from soil moisture and texture following a simplification of Cox *et al.* (1999) (see function `soil_thermalcapacity()`).

13.2 Multi-layer canopy

A multi-layer canopy energy balance requires the calculation of sensible heat and latent heat exchanges between the air in each canopy layer and the leaves it contains. In addition, one has to consider heat exchanges between each layer and the ones below/above, derived from turbulent flow.

13.2.1 Latent heat

The latent heat for a given canopy layer j (LE_j ; in $W \cdot m^{-2}$) includes transpiration and evaporation of intercepted rain, and is estimated analogously to the single-canopy case (eq. (13.1)):

$$LE_j = \lambda_v(T_{air,j}) \cdot \left(\frac{In_{j,t}}{\Delta t_{step}} + E_{j,t} \right) \quad (13.11)$$

where $\lambda_v(T_{air,j})$ is the latent heat of vaporization at temperature $T_{air,j}$ (in $J \cdot kg^{-1}$), $In_{j,t}$ is the intercepted water that is evaporated from the canopy layer, determined by dividing In_t among canopy layers according to the fraction of total LAI in each one:

$$In_{j,t} = In_t \cdot \frac{\sum_{i=1}^c LAI_{i,j}^\phi}{LAI_{stand}^\phi} \quad (13.12)$$

and $E_{j,t}$ is the instantaneous transpiration rate in layer j (in $kg H_2O \cdot m^{-2} \cdot s^{-1}$), given by:

$$E_{j,t} = 10^{-3} \cdot 0.01802 \cdot \sum_{i=1}^c E_{i,t}^{sunlit} \cdot LAI_{i,j}^{sunlit} + E_{i,t}^{shade} \cdot LAI_{i,j}^{shade} \quad (13.13)$$

where $E_{i,t}^{sunlit}$ and $E_{i,t}^{shade}$ are the instantaneous transpiration rates (in $mmol H_2O \cdot m^{-2} \cdot s^{-1}$) of sunlit and shade leaves, respectively, of cohort i during time step t ; and $LAI_{i,j}^{sunlit}$ and $LAI_{i,j}^{shade}$ are the leaf area index of sunlit and shade leaves, respectively, of cohort i in layer j (9.5).

Latent heat derived from soil water evaporation of snow melting (LE_{soil} ; in $W \cdot m^{-2}$) is estimated as in the single-canopy case, using eq. (13.2).

13.2.2 Sensible heat between leaves and the canopy air space

The sensible heat exchanged between the air of canopy layer j and the leaves within it ($H_{leaf,j}$; $W \cdot m^{-2}$) is obtained using:

$$H_{leaf,j} = \sum_{i=1}^c LAI_{i,j}^{sunlit} \cdot H_{i,j}^{sunlit} + LAI_{i,j}^{shade} \cdot H_{i,j}^{shade} \quad (13.14)$$

where $LAI_{i,j}^{sunlit}$ and $LAI_{i,j}^{shade}$ are the leaf area index of sunlit and shade leaves of plant cohort i in layer j , respectively (see eq. (9.5)). $H_{i,j}^{sunlit}$ and $H_{i,j}^{shade}$ are the sensible heat exchange between layer j and sunlit or shade leaves of cohort i (Ma & Liu 2019):

$$H_{i,j}^{sunlit} = -2 \cdot C_p \cdot (T_{leaf,i}^{sunlit} - T_{air,j}) \cdot g_{Ha,i,j} \quad (13.15)$$

$$H_{i,j}^{shade} = -2 \cdot C_p \cdot (T_{leaf,i}^{shade} - T_{air,j}) \cdot g_{Ha,i,j} \quad (13.16)$$

where $T_{leaf,i}^{sunlit}$ and $T_{leaf,i}^{shade}$ are, respectively, the sunlit and shade leaf temperatures for cohort i (determined using eq. (11.1)); $T_{air,j}$ is the current air temperature in canopy layer j , $C_p = 29.37152 \text{ J} \cdot \text{mol}^{-1} \cdot \text{zC}^{-1}$ is the specific heat capacity of dry air at constant pressure; and $g_{Ha,i,j}$ is the leaf boundary layer conductance for heat ($\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$) for leaves of cohort i and wind speed in layer j (see eq. (11.3)).

13.2.3 Turbulent heat exchange

Turbulent heat exchanges occur between the soil and the bottom canopy layer (i.e. $j = 1$):

$$H_{1,soil} = \frac{\rho_1 \cdot c_p}{r_{1,soil}} \cdot (T_{air,1} - T_{soil,1}) \quad (13.17)$$

where $c_p = 1013.86 \text{ J} \cdot \text{kg}^{-1} \cdot \text{zC}^{-1}$ is the specific heat capacity of the air, ρ_1 is the air density at $j = 1$, $T_{air,1}$ is the air temperature of layer 1, $T_{soil,1}$ is the temperature of the top soil layer and $r_{1,soil}$ is the aerodynamic resistance between the soil and canopy layer 1 (in sm^{-1}). The bottom layer also have turbulent heat exchanges with the layer above, determined by the canopy turbulence model. The turbulent heat flux gradient for layer $j = 1$ is:

$$(\delta F_H / \delta z)_1 = \frac{-c_p \cdot \rho_1 \cdot (T_{air,2} - T_{air,1}) \cdot u\bar{w}_1}{\Delta z \cdot (\delta u / \delta z)_1} - H_{1,soil} \quad (13.18)$$

where Δz is the size of vertical layers (in m); and $(\delta u / \delta z)_1$ and $u\bar{w}_1$ are the wind speed gradient and Reynolds stress at $j = 1$.

For all intermediate layers (i.e. $1 < j < l$) we have turbulent exchanges with the neighboring layers $j - 1$ and $j + 1$, which results in the following turbulent heat flux gradient:

$$(\delta F_H / \delta z)_j = \frac{-c_p \cdot \rho_j \cdot (T_{air,j+1} - T_{air,j-1}) \cdot u\bar{w}_j}{2 \cdot \Delta z \cdot (\delta u / \delta z)_j} \quad (13.19)$$

And, finally, the topmost layer (i.e. $j = l$) has turbulent heat exchanges with the layer below $l - 1$ and the atmosphere, so that:

$$(\delta F_H / \delta z)_l = \frac{-c_p \cdot \rho_l \cdot (T_{atm} - T_{air,l-1}) \cdot u\bar{w}_l}{2 \cdot \Delta z \cdot (\delta u / \delta z)_l} \quad (13.20)$$

13.2.4 Canopy and soil temperature changes

Evaluation of the canopy layer energy balance equation (7.5) is done taking into account that sensible heat exchange for a given layer j includes heat exchange with leaves (i.e. $H_{leaf,j}$) and the turbulent heat flux gradient (i.e. $(\delta F_H / \delta z)_j$), so that the variation of air temperature in layer j for a given time interval $\Delta t_{substep}$ is:

$$\Delta T_{air,j} = \Delta t_{substep} \cdot \frac{K_{abs,j} + L_{net,j} - LE_j + H_{leaf,j} + (\delta F_H / \delta z)_j}{TC_j} \quad (13.21)$$

where TC_j is the thermal capacitance of canopy layer j (in $\text{J} \cdot \text{m}^{-2} \cdot \text{K}^{-1}$), found analogously to TC_{can} (eq. (13.6)):

$$TC_j = TC_{LAI} \cdot \frac{0.8 \cdot LAI_j + 1.2 \cdot (LAI_j^{phi} + LAI_j^{dead})}{2} \quad (13.22)$$

$\Delta t_{substep}$ is the time step for closing the energy balance, which has to be smaller than Δt_{step} to avoid numerical instabilities due to turbulent flow.

When simulating a multiple-layer energy balance, we have to replace $H_{can,soil}$ by $H_{1,soil}$ in the energy inflow of soil layer 1: (13.8):

$$G_{0,1} = K_{soil} + L_{net,soil} + H_{1,soil} - LE_{soil} \quad (13.23)$$

Soil temperature changes are estimated in the same way as for single-canopy energy balance, except that a smaller temporal step is used:

$$\Delta T_{soil,s} = \Delta t_{substep} \cdot \frac{G_{s-1,s} - G_{s,s+1}}{C_{soil,s}} \quad (13.24)$$

13.2.5 Within-canopy changes in water vapor and CO_2

Multi-layer canopy energy balance also entails the possibility of considering gradients of scalars such as water vapor or CO_2 . Analogously to the turbulent heat exchange, wind turbulence leads to changes in concentration of water vapor or CO_2 within canopy layers. In addition, plant physiology also alters the concentration of these gases through transpiration in photosynthesis. To model temporal changes in these scalars for canopy layers, we first have to estimate CO_2 concentration in $mg \cdot m^{-3}$ and H_2O concentration in $kg \cdot m^{-3}$ from $C_{air,j}$ (ppm) and $e_{air,j}$ (kPa) using:

$$[CO_2]_j = 0.409 \cdot 44.01 \cdot C_{air,j} \quad (13.25)$$

$$[H_2O]_j = 0.622 \cdot \frac{e_{air,j} \cdot \rho_j}{P_{atm}} \quad (13.26)$$

The one-dimensional scalar flux balance for an homogeneous turbulent flow can be described by the following conservation equations (Lai *et al.* 2000):

$$(\delta[CO_2]/\delta t)_j = (\delta F_{[CO_2]}/\delta z)_j + S_{[CO_2],j} \quad (13.27)$$

$$(\delta[H_2O]/\delta t)_j = (\delta F_{[H_2O]}/\delta z)_j + S_{[H_2O],j} \quad (13.28)$$

where $(\delta F_{[CO_2]}/\delta z)_j$ and $(\delta F_{[H_2O]}/\delta z)_j$ are the gradients of vertical flux of CO_2 and H_2O in layer j , respectively; and $S_{[H_2O],j}$ and $S_{[CO_2],j}$ are the source strengths for $[H_2O]$ (in $kg H_2O \cdot m^{-3} \cdot s^{-1}$) and $[CO_2]$ (in $mg CO_2 \cdot m^{-3} \cdot s^{-1}$) in layer j , respectively, which derive from transpiration (source of water) and photosynthesis (sink of CO_2):

$$S_{[H_2O],j} = \frac{(In_{j,t}/\Delta t) + E_{j,t}}{\Delta z} \quad (13.29)$$

$$S_{[CO_2],j} = -\frac{A_{n,j,t}}{\Delta z} \quad (13.30)$$

where $E_{j,t}$ and $A_{n,j,t}$ are the instantaneous transpiration and photosynthesis rates in layer j . The former was already defined in eq. (13.13), whereas the latter ($A_{n,j,t}$; in $mg CO_2 \cdot m^{-2} \cdot s^{-1}$) is defined analogously:

$$A_{n,j,t} = 10^{-3} \cdot 44.01 \cdot \sum_{i=1}^c A_{n,i,t}^{sunlit} \cdot LAI_{i,j}^{sunlit} + A_{n,i,t}^{shade} \cdot LAI_{i,j}^{shade} \quad (13.31)$$

where $A_{n,i,t}^{sunlit}$ and $A_{n,i,t}^{shade}$ are the instantaneous net photosynthesis rates (in $\mu mol CO_2 \cdot m^{-2} \cdot s^{-1}$) of sunlit and shade leaves, respectively, of cohort i during time step t ; and $LAI_{i,j}^{sunlit}$ and $LAI_{i,j}^{shade}$ are the leaf area index of sunlit and shade leaves, respectively of cohort i in layer j (9.5). For the special case of the lowest canopy layer, water vapor needs to include the evaporation from soil, i.e.:

$$S_{[H_2O],1} = \frac{((In_{1,t} + Es_t)/\Delta t) + E_{1,t}}{\Delta z} \quad (13.32)$$

For any layer $1 < j < l$ the gradients of vertical scalar fluxes are found using:

$$(\delta F_{[H_2O]}/\delta z)_j = -\frac{c_p \cdot \rho_j \cdot ([H_2O]_{j+1} - [H_2O]_{j-1}) \cdot u \bar{w}_j}{2 \cdot \Delta z \cdot (\delta u/\delta z)_j} \quad (13.33)$$

$$(\delta F_{[CO_2]}/\delta z)_j = -\frac{c_p \cdot \rho_j \cdot ([CO_2]_{j+1} - [CO_2]_{j-1}) \cdot u \bar{w}_j}{2 \cdot \Delta z \cdot (\delta u/\delta z)_j} \quad (13.34)$$

whereas for the bottom layer ($j = 1$) we have:

$$(\delta F_{[H_2O]}/\delta z)_1 = -\frac{c_p \cdot \rho_1 \cdot ([H_2O]_2 - [H_2O]_1) \cdot u \bar{w}_1}{\Delta z \cdot (\delta u/\delta z)_1} \quad (13.35)$$

$$(\delta F_{[CO_2]}/\delta z)_1 = -\frac{c_p \cdot \rho_1 \cdot ([CO_2]_2 - [CO_2]_1) \cdot u \bar{w}_1}{\Delta z \cdot (\delta u/\delta z)_1} \quad (13.36)$$

and for the top layer ($j = l$) we have:

$$(\delta F_{[H_2O]}/\delta z)_l = -\frac{c_p \cdot \rho_l \cdot ([H_2O]_{atm} - [H_2O]_{l-1}) \cdot u \bar{w}_l}{2 \cdot \Delta z \cdot (\delta u/\delta z)_l} \quad (13.37)$$

$$(\delta F_{[CO_2]}/\delta z)_l = -\frac{c_p \cdot \rho_l \cdot ([CO_2]_{atm} - [CO_2]_{l-1}) \cdot u \bar{w}_l}{2 \cdot \Delta z \cdot (\delta u/\delta z)_l} \quad (13.38)$$

Equation (13.28) is solved via discretization both in terms of space (vertical layers) and time, the latter using $\Delta t_{substep}$ as for the energy balance.

Chapter 14

Day-level results

In the preceding chapters we detailed how the model performs canopy and soil energy balances for subdaily time steps and how transpiration and photosynthesis values are determined for any given substep. This chapter indicates how these are aggregated at the daily scale and how other day-level model outputs are calculated.

14.1 Photosynthesis and transpiration

Cohort's transpiration $Tr_{i,t}$ (eq. (12.6)) are added across subdaily steps to yield daily transpiration (Tr_i , in $mm H_2O$):

$$Tr_i = \sum_{t=1}^{n_t} Tr_{i,t} \quad (14.1)$$

An the same for water extraction $Ex_{i,s,t}$ for each soil layer s (eq.(12.7)):

$$Ex_{i,s} = \sum_{t=1}^{n_t} Ex_{i,s,t} \quad (14.2)$$

$Ex_{i,s}$ are substracted from the water content of the corresponding soil layer, closing the soil water balance of the day (eq. (7.1)).

Daily values of net carbon assimilation for plant cohorts are obtained similarly. $A_{n,i,t}$ (eq. (12.9)) are added across subdaily steps to obtain $A_{n,i}$, the daily net assimilation at the cohort level (in $g C \cdot m^{-2}$):

$$A_{n,i} = \sum_{t=1}^{n_t} A_{n,i,t} \quad (14.3)$$

14.2 Plant water potentials and relative water contents

Because the model determines optimum transpiration for every subdaily time step, this leads to a daily sequence of leaf water potential ($\Psi_{leaf,i,t}$), stem water potential ($\Psi_{stem,i,t}$), root crown water potential ($\Psi_{rootcrown,i,t}$) and root surface water potential ($\Psi_{rootcrown,i,s,t}$) values for each plant cohort i (and soil layer s in the last case). The model defines the following daily water potentials for every cohort i :

- *Pre-dawn leaf water potential* ($\Psi_{pd,i}$): the maximum of $\Psi_{leaf,i,t}$ values.
- *Pre-dawn shade leaf water potential* ($\Psi_{pd,i}^{shade}$): the maximum of $\Psi_{leaf,i,t}^{shade}$ values.

- *Pre-dawn sunlit leaf water potential ($\Psi_{pd,i}^{sunlit}$)*: the maximum of $\Psi_{leaf,i,t}^{sunlit}$ values.
- *Mid-day leaf water potential ($\Psi_{md,i}$)*: the minimum of $\Psi_{leaf,i,t}$ values.
- *Mid-day shade leaf water potential ($\Psi_{md,i}^{shade}$)*: the minimum of $\Psi_{leaf,i,t}^{shade}$ values.
- *Mid-day sunlit leaf water potential ($\Psi_{md,i}^{sunlit}$)*: the minimum of $\Psi_{leaf,i,t}^{sunlit}$ values.
- *Stem water potential ($\Psi_{stem,i}$)*: the minimum of $\Psi_{stem,i,t}$ values.
- *Root-crown water potential ($\Psi_{rootcrown,i}$)*: the minimum of $\Psi_{rootcrown,i,t}$ values.
- *Root surface water potentials ($\Psi_{rootsurf,i,s}$)*: the minimum of $\Psi_{rootsurf,i,s,t}$ values for each soil layer s .

Analogously, relative water content of stems and leaves is known for every subdaily time step, which results in a daily sequence of leaf relative water content ($RWC_{leaf,i,t}$) and stem relative water content ($RWC_{stem,i,t}$). These are summarized at the daily level for each plant cohort i :

- *Leaf relative water content ($RWC_{leaf,i}$)*: the mean $RWC_{leaf,i,t}$ values.
- *Stem relative water content ($RWC_{stem,i}$)*: the mean of $RWC_{stem,i,t}$ values.

Finally, the daily sequence of slopes of the supply function ($dE/d\Psi_{i,t}$) is also averaged at the daily level:

- *Slope of the supply function ($dE/d\Psi_i$)*: the mean $dE/d\Psi_{i,t}$ values.

14.3 Plant drought stress

In order to have an estimate of daily drought stress for the plant cohort, the model uses the stem vulnerability curve of the plant to find the conductance relative to maximum stem conductance and turns it into its complement:

$$DDS_i = \phi_i \cdot \left(1.0 - \frac{k_{stem,i}(\Psi_{rootcrown,i})}{k_{\max stem,i}} \right) = \phi_i \cdot \left(1.0 - e^{-(\Psi_{rootcrown,i}/d_{stem})^{c_{stem}}} \right) \quad (14.4)$$

where ϕ_i is the leaf phenological status. Note the use of $\Psi_{rootcrown,i}$ (and not $\Psi_{leaf,i}$) to determine drought stress index. Thus the model tracks the degree of conductance decrease at the beginning of the stem as a measure of drought stress. This choice makes daily drought stress values of the Simple and Complex transpiration modes more comparable (because leaf mid-day water potentials are usually much more negative than soil water potentials) and is a sensible choice if one wants to run the model in irreversible cavitation mode (see below).

Part IV

Forest growth modelling

Chapter 15

Forest growth model

This chapter provides an overview of a forest growth model, which builds on the previous two models and allows simulating carbon and water balance as well as growth and mortality on a daily step basis for a given forest stand and for the period corresponding to input weather data. The model was described in De Cáceres *et al.* (2023) and is run using function `growth()`, for a set of days, or function `growth_day()`, for a single day.

The forest growth model can be run in two different levels of complexity, depending on the water balance submodel employed (basic and advanced, corresponding to chapters 3 and 7). In this and the following chapters, we indicate those design and formulation elements where the forest growth model building on the basic water balance differs from that building on the advanced one.

The information provided in the current chapter should be enough to understand what the growth model does, but the reading previous chapters and the following ones should provide a more detailed understanding on how the different processes are designed and formulated.

15.1 Design principles

15.1.1 Overview

Like in the preceding models, the physical structure of the stand is represented in one (vertical) dimension (i.e. the horizontal coordinates of plants are not explicit). The model is *cohort-based*, meaning that similar plant individuals are represented using a single entity with average characteristics (e.g. tree height and diameter at breast height) and a density variable is used to scale from individual level to the cohort level. Processes are modelled either at the cohort level (water balance, photosynthesis, or plant mortality) or at the “average” individual level (carbon balance, tissue growth and senescence). Water fluxes, soil water balance and plant photosynthesis processes follow the design of the soil water balance model and will not be repeated here (see chapters 3 and 7). The growth model tries to reproduce the physical and physiological processes that modulate leaf area changes, sapwood area changes and fine root area changes (the latter only under the advanced water balance submodel), while accounting for both source and sink limitations to tissue growth. Formation of new sapwood leads to diameter and height increases, so that physiological processes involve structural changes. Since the model does not implement all processes and drivers (such as nutrient availability) potentially affecting growth, user input parameters are used to empirically constrain growth rates and plant size, i.e. maximum relative sapwood area growth rates, maximum plant height and diameter-height ratios. The forest growth model also implements tree mortality, which reduces cohort density, arising from different processes. A baseline mortality rate is incremented either because of exhaustion of carbon reserves (i.e. plant starvation) or dehydration of plant tissues (i.e. plant desiccation). Additionally, fire behavior and effects (i.e. fire severity) can be simulated. Fires can partially or totally burn crown leaves and buds. Fire heat can also induce necrosis in the cambium, leading to full above-ground mortality.

15.1.2 Carbon compartments, transport and carbon balance

Three *carbon compartments* are considered separately: **leaves**, **sapwood** (including stem, branches and coarse roots) and **fine roots**. Each compartments has one or more *carbon pools* where carbon is in different form. Carbon pools are differentiated between **structural** and **labile**. In turn, labile carbon involves **metabolic** and **storage** (Richardson *et al.* 2013; Dietze *et al.* 2014), with a conversion rate regulated by the need to maintain, as much as possible, an equilibrium in the amount of metabolic carbon to sustain cell metabolism and for osmotic purposes. Similar designs of two non-structural carbon compartments have been implemented in other forest ecosystem models (Cho *et al.* 2022).

Details of the carbon pools in different compartments are as follows (Fig. 15.1):

- **Structural carbon** corresponds carbon forming cell membranes, cell walls and the cytosolic machinery (in the case of living cells). The amount of structural carbon in a given compartment is used to determine *maintenance respiration* costs (together with labile carbon), and the structural carbon per tissue unit determines *growth respiration* costs. Structural carbon occurs in the three compartments, *leaves*, *sapwood* and *fine roots*, which however have different maintenance and growth costs. Sapwood carbon includes branches, stem and coarse roots. Only live (parenchymatic) sapwood cells, accounting for between 5 and 40% of sapwood volume (Plavcová & Jansen 2015), contribute to estimate sapwood maintenance respiration costs. The fraction of embolized sapwood is also discounted from sapwood respiratory biomass. Structural carbon is lost via tissue senescence (in the case of sapwood due to conversion to heartwood).
- **Metabolic carbon** is a labile carbon form assumed to correspond to **sugars** (e.g., glucose or fructose) used to sustain cell functioning (i.e. provide carbon for maintenance respiration). Moreover, changes in metabolic carbon are coupled to variations in osmotic potential. Metabolic carbon occurs in leaves and sapwood compartments only. *Leaf metabolic carbon* is generated from photosynthesis or leaf starch degradation and used in different functions, including leaf maintenance respiration and export to sapwood via phloem transport. An excess of leaf metabolic carbon can inhibit photosynthesis. Sugar concentration in the phloem is assumed to be (passively) in equilibrium with leaf metabolic sugar concentration. *Sapwood metabolic carbon* comes from phloem transport, or sapwood starch degradation, and is used to directly sustain maintenance respiration of sapwood and fine roots.
- **Storage carbon** is a labile carbon form assumed to loosely correspond to **starch**. *Leaf storage carbon* is intended to represent short-term storage (mainly starch within chloroplasts) and its inclusion in the model follows the need to buffer daily variations in leaf metabolic carbon as well as in the corresponding leaf osmotic potential. *Sapwood storage carbon* in parenchymatic tissues of branches, stems and coarse roots conforms a much larger carbon pool and is intended to serve as carbon source for growth and long-term (seasonal and inter-annual) storage (Plavcová & Jansen 2015).

Different processes contribute to the carbon balance of structural and labile compartments:

- Carbon balance of **labile carbon pools** is performed by considering **photosynthesis**, **maintenance respiration**, **growth costs**, **sugar-starch conversion**, **root exudation** and **phloem transport**. Photosynthesis estimates come from the water balance submodel. Respiration is modelled following the *growth-maintenance* paradigm, but we acknowledge that such approach has limitations and that others have been proposed such as assuming a constant *carbon use efficiency* or the *process/residual approach* (Cannell & Thornley 2000; Gifford 2003). Maintenance respiration demand of leaves and sapwood depends on both structural and metabolic carbon, whereas only structural carbon counts for fine root maintenance respiration because they lack labile carbon pools. Maintenance respiration of leaves and fine roots can be species-specific rates, but can also be estimated from species-specific values of nitrogen concentration (Reich *et al.* 2008). In contrast, the species-specific maintenance respiration per sapwood unit mass (in fact, only parenchymatic tissues contribute to sapwood respiration) is not estimated from nitrogen concentration and should be calibrated. In all three cases, however, temperature dependencies are included in the actual respiration rates (Tjoelker *et al.* 2001). Growth costs include the carbon cost of building new structures as well as the respiration expenses of growth. Sugar-starch conversion is

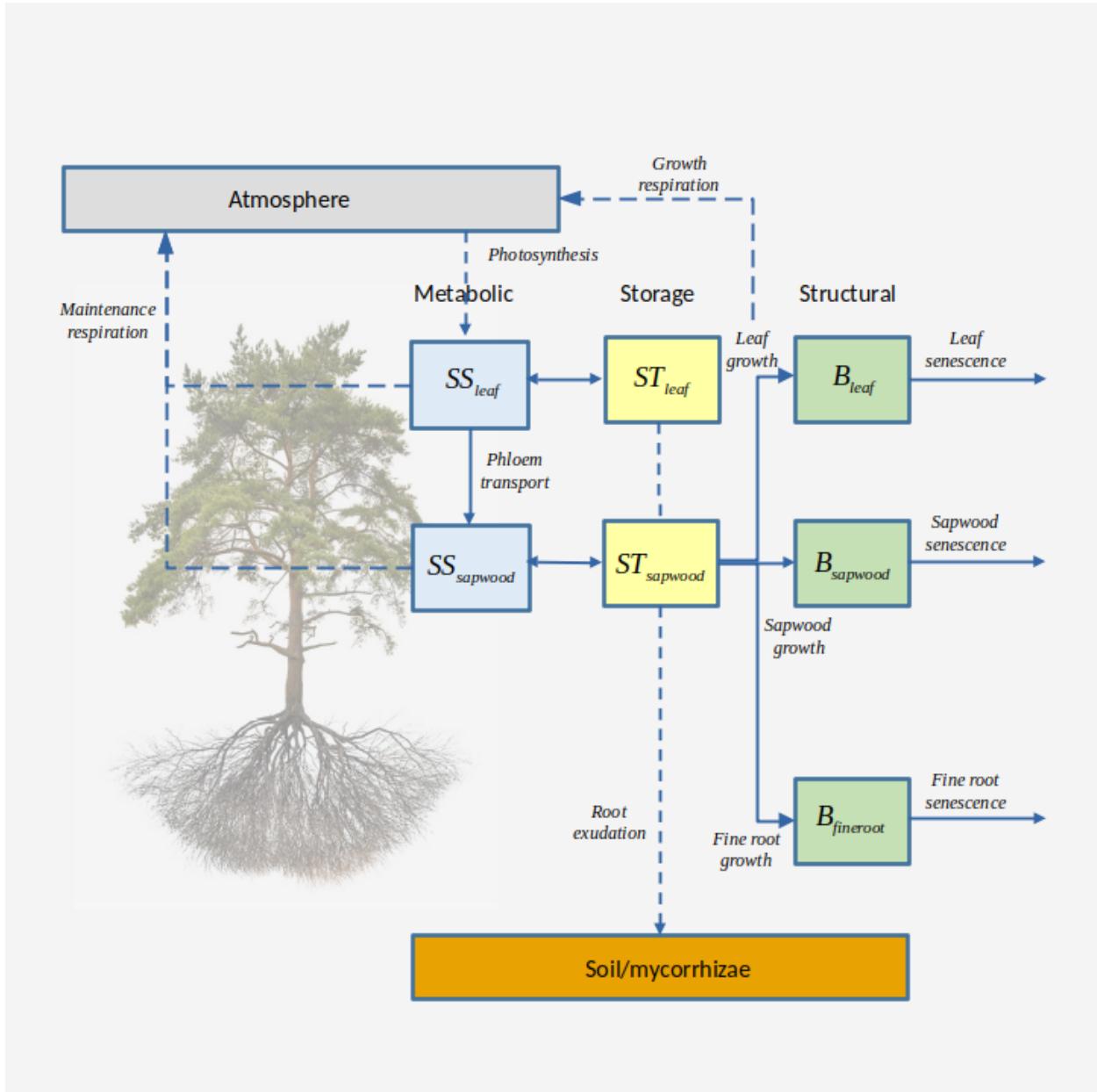


Figure 15.1: Schematic representation of carbon compartments, carbon pools and carbon balance components (translocation of reserves during senescence not included)

designed to maintain an equilibrium in the concentration of metabolic sugar. Any surplus in sapwood storage carbon over storage capacity is assumed to be diverted towards root exudation. Hence, root exudation is not modeled as a process actively competing for metabolic carbon, but as a consequence of plant C storage capacity being surpassed (Prescott *et al.* 2020).

- Biomass balance in **structural pools** comes from the interplay between **growth** (i.e. new tissue formation) and **senescence** (see below). Structural growth rates are lower than growth costs for labile pools, since growth costs also include growth respiration expenses. During leaf senescence it is assumed that non-structural carbon sources (i.e. labile carbon pools) are recycled and relocated to sapwood storage. Analogously, when sapwood is converted into heartwood, labile carbon in the protoplasm of parenchyma cells is assumed to be re-absorbed by neighboring living cells as storage (Spicer 2005).

Further design details and formulation regarding the estimation of carbon pool sizes and carbon balance components are given in chapter 16.

15.1.3 Tissue growth, senescence and allocation

The forest growth model allows simulating the variation of leaf area, sapwood area and fine root area via the following processes (see fig. 15.2):

- Leaf growth:** Whereas in the water balance models expanded leaf area corresponded to the portion of live leaf area that was unfolded according to the phenological status (see 4.1), in the growth model expanded leaf area results from explicit simulation of growth. However, leaf phenology submodel still controls the duration of phenophases corresponding budburst, leaf development and senescence. Leaf area growth occurs during *leaf unfolding* periods only, whereas *bud formation* periods are required to update the target leaf area (in accordance to an allocation strategy, see below). Storage carbon levels need to be at more than 10% of their capacity to allow leaf growth. Moreover, leaf area target is constrained by stand structure (i.e. competition) via allometric equations.
- Leaf senescence:** Leaf senescence (and the corresponding increases in dead leaf area) can occur in the model due to *leaf aging* in evergreen species, due to *programmed leaf senescence* in deciduous species, or due to *leaf dessication* (as a result of xylem cavitation and/or low leaf relative water content). In addition, fires can induce *foliage burning*, which also leads to a decrease in leaf area (but where non-structural carbon reallocation is not possible and dead leaf area does not increase).
- Sapwood growth:** Formation of new sapwood can only occur if unfolded leaves are present, assuming that hormonal signals controlling cambium division are mainly synthesized within leaves. Sapwood formation corresponds mostly to secondary growth (but primary growth occurs at the same time due to diameter-height relationships), and is only allowed if storage carbon levels are above a given species-specific threshold (e.g. 50%). These levels are much higher than required for leaf or fine root growth. Thus, when carbon levels are low, sapwood growth rates fall to zero, whereas replacement of leaves and fine roots continues to sustain metabolism. This design was chosen to stop sapwood growth much before emptying storage carbon reserves, so that maintenance respiration can be sustained longer whenever photosynthesis is low, as in shadow-tolerant species.
- Sapwood senescence:** Losses in sapwood area (i.e. conversion to heartwood) occur via tissue *aging* (i.e. assuming a daily rate of conversion). Similarly to 3-PG (Landsberg & Waring 1997) we assume that the relative turnover rate of sapwood is smallest for young plants, and it increases up to a maximum value. However, recent reviews indicate that heartwood formation should be viewed as an active developmental program during which a conductive but vulnerable sapwood is transformed in a non-conductive but durable heartwood (Spicer 2005). Tree sapwood senescence also occurs whenever the sapwood area exceeds the amount required to sustain the current leaf area target. Note that stem *cavitation* reduces the area of sapwood that is effectively functional for water transport, but does not lead to sapwood senescence per se.
- Fine root growth:** Fine root growth can in principle occur any time of the year (i.e. the model does not explicitly consider root phenology), but since fine root biomass targets depend on leaf area,

seasonal fine root area patterns may follow phenological changes in leaf area. Like in leaf growth, storage carbon levels need to be at more than 10% of their capacity to allow fine root growth.

- vi. **Fine root senescence:** Fine root senescence occurs via *aging* with a turnover rate depending on soil temperature.

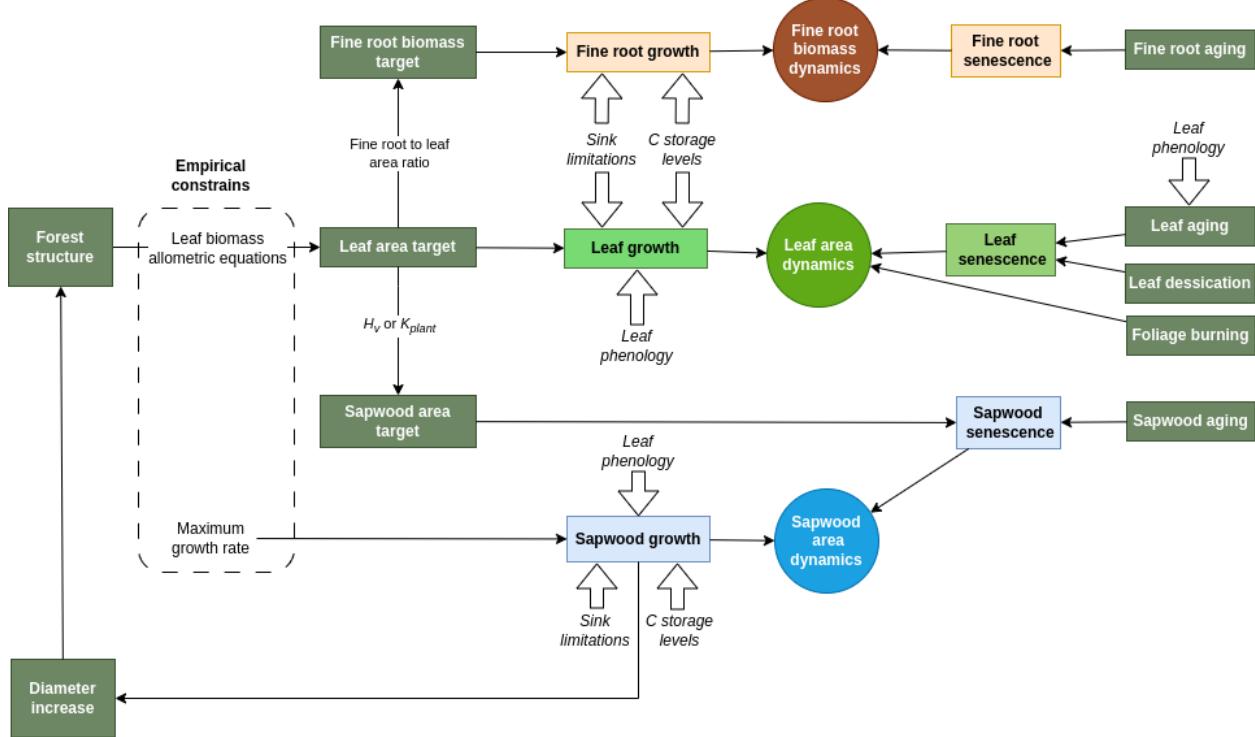


Figure 15.2: Schematic representation of tissue growth and senescence drivers for trees

Formation of new leaf, sapwood and fine root tissues is determined taking into account both **source** (i.e. carbon availability) and **sink** (i.e. biophysical constraints on plant tissue expansion) limitations (Fatichi *et al.* 2014; Guillemot *et al.* 2015; Körner 2015). Temperature and turgor sink limitations are implemented following Cabon *et al.* (2020a) and Cabon *et al.* (2020b). These authors developed their approach for tracheid production and enlargement only, whereas we apply it to different kinds of tissues. In the case of fine roots, differences in soil water potential between layers can lead to different degrees of turgor limitation for the growth of fine roots, resulting in adaptation of fine root distribution.

The growth rates in all three compartments are modulated by parameters specifying *maximum daily growth rates relative to the amount of sapwood area*. Nevertheless, tree leaf area and fine root biomass targets are constrained by leaf biomass allometric equations, which limits leaf and fine root growth to the replacement of aging tissues, unless there are changes in forest structure, i.e. competition is released or the target tree increases in diameter (see fig. 15.2). In contrast, the sapwood area target modulates sapwood senescence (i.e. the conversion to heartwood), but not sapwood growth. Sapwood growth rates are mainly determined by maximum relative growth rates (which can be parameterized with empirical observations), carbon availability and sink limitations. Since sapwood formation leads to tree diameter increases, maximum sapwood growth rate indirectly determine the rate of growth of leaves and fine roots at seasonal and annual scales, hence determining the whole-plant maximum growth rate.

Two options are possible regarding the relationship between structural carbon of leaves and sapwood:

1. *Stable Huber value:* In this strategy, growth of leaf area and sapwood area need are coordinated so that the ratio of sapwood area to leaf area (i.e. Huber value) is in dynamic equilibrium around its initial

value. To this aim, sapwood senescence is scheduled whenever the current Huber value is larger than the target (i.e. there is an excess of sapwood area). This approach is similar to the LPG model (Sitch *et al.* 2003), where different turnover rates are applied for different tissues, but then the model tries to satisfy the pipe model (Shinozaki *et al.* 1964) by allocating C where it is more limiting. Note that in this strategy the whole-plant conductance will progressively decrease with the increase in plant height, because of the increase in the length of stem xylem pathway.

2. *Stable whole-plant conductance:* Ratios of sapwood area to leaf area (i.e. Huber values) can vary within species, due to environmental conditions (Mencuccini & Grace 1995). In this allocation strategy, growth of leaf area, sapwood area and height needs to be coordinated so that whole-plant conductance (i.e. from roots to leaves) is in dynamic equilibrium around its initial value. Sapwood senescence is scheduled whenever the current whole-plant conductance is above the target value. In this strategy, the Huber value will tend to increase with the increase in plant height, thereby increasing the sapwood respiratory demands.

When running the growth model along with the basic water balance submodel, a stable Huber value is the only strategy available and fine root area growth also follows the need to achieve a stable ratio between leaf area and fine root area. In contrast, the two strategies are possible when simulating growth along with the advanced water balance. In this case, fine root area growth is coordinated with the formation of the other tissues via the need to ensure an *average resistance fraction in the rhizosphere*, which is used to define the fine root area target and, hence, the allocation to fine roots.

Further details and formulation of growth, senescence and allocation are given in chapter 17.

15.1.4 Coordination between water and carbon balances

The growth model allows coordinating several aspects of carbon and water balances through different state variables (Fig. 15.3), most of them applying to the simulation using the advanced transpiration model:

- a. Transpiration and photosynthesis are coordinated via the *stomatal regulation* submodel (see chapter 11).
- b. Hydrostatic water potential of leaf symplastic tissues influences the turgor pressure necessary for leaf growth (17.1.2), while (metabolic) leaf sugar concentration determines changes in osmotic potential at full turgor and hence, the relative water content of leaf symplasmic tissue.
- c. Analogously, water potential of stem symplastic tissues determines the turgor in the cambium and the sapwood formation rate (17.1.3), while the sugar concentration in the sapwood determines the osmotic potential at full turgor of stem symplastic tissues and its relative water content.
- d. Leaf growth and senescence influence hydraulic conductances of several tissues (fine roots, coarse root sapwood and stem sapwood), as they are defined on a leaf area basis (17.3.1).
- e. Sapwood growth and senescence influence hydraulic conductance of stems via changes in sapwood area, and indirectly those of coarse roots and the rhizosphere (17.3.1).
- f. Fine root biomass dynamics determines fine root distribution and rhizosphere conductance across soil layers (17.3.1).

15.1.5 Plant mortality

Mortality pathways

Dynamic vegetation models implement mortality processes in very different ways, from purely-empirical to process-based (Hawkes 2000; Keane *et al.* 2001; Bugmann *et al.* 2019). The fact that proximal causes of mortality are complex, co-occurring, interrelated and variable has prompted the question of whether a mechanistic approach will improve prediction ability (Adams *et al.* 2013). In medfate, (above-ground parts of) woody plants can die following a combination of empirical and process-based approaches:

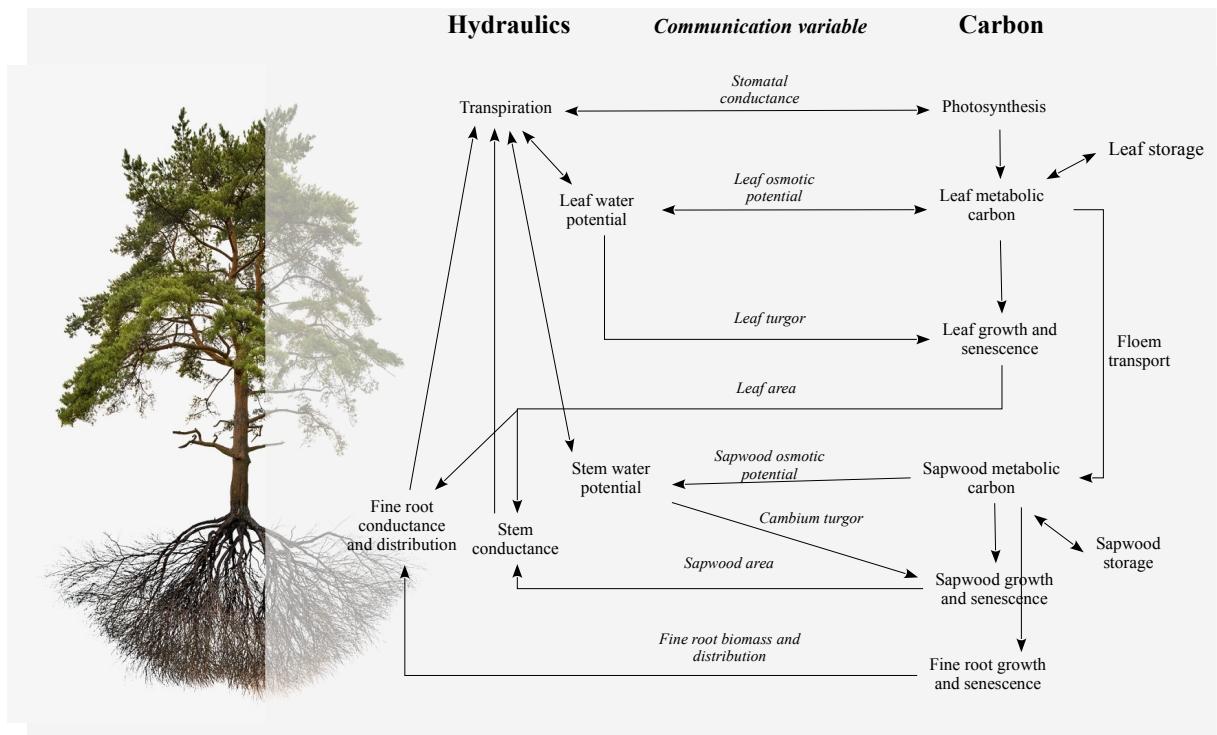


Figure 15.3: Schematic representation of relationships between water balance (advanced model) and carbon balance. Relationships within water or carbon balances have been omitted

1. Trees with $DBH < 7.5$ cm (i.e. saplings) are assumed to suffer mortality due to a self-thinning process of unspecified cause. Specifically, the model represents the self-thinning process occurring between small trees (e.g. 1 cm DBH) resulting from seed establishment (or resprouting) and trees achieving the size (e.g. 7.5 cm) corresponding to inclusion as individual in forest inventories (e.g. ingrowth size). Although implemented here, this self-thinning process is most relevant for simulations of mid- and long-term forest dynamics (see 18).
2. All woody cohorts are assumed to die at a basal rate due to processes not included in the model (e.g. biotic attacks). This rate is empirically determined and can be a constant value or, in the case of trees, can depend on the degree of competition (i.e. basal area). This latter option is important to ensure a limit in stand basal area, which may not occur as a result of mechanistic mortality processes.
3. Mortality rates can be higher than the basal rates whenever physiological thresholds presumed to lead to plant mortality are surpassed (Hawkes 2000). The model allows plants to die from either *starvation* (if metabolic carbon is exhausted) or *desiccation* (tissue dehydration) (McDowell *et al.* 2008; McDowell 2011). Allowing for starvation is important to simulate death due to shading by other plant cohorts, whereas plant desiccation is important to represent situations of low soil water availability.

With respect to intra-cohort variability, the model can be run assuming that all individuals within plant cohorts are functionally equal, so that mortality applies to the entire plant cohort. Alternatively, mortality can be applied to a fraction of the plant cohort, in proportion to the mortality likelihood, which implicitly acknowledges intra-cohort differences. As common in other models, mortality process can be simulated in a deterministic or stochastic fashion.

Physiological underpinnings of drought-related mortality

Multiple mechanisms determine plant survival or mortality under drought (McDowell *et al.* 2008; McDowell *et al.* 2022). However, hydraulic failure is widely recognized as one of the key mechanisms of drought-induced mortality in woody plants. Lethal water potential have been defined as a critical threshold beyond which trees are unable to recover from drought, even after rewatering. Experimental work by Urli *et al.* (2013) determined that irreversible damage in angiosperm saplings occurred only after 88% losses in stem hydraulic conductance. Similarly, Hammond *et al.* (2019) found probability of mortality to strongly increase beyond 80% of loss in hydraulic conductivity in saplings a gymnosperm species. In a multi-species synthesis, Adams *et al.* (2017) assessed found that all studied species showed at least 60% or higher loss of xylem hydraulic conductivity, whereas evidence for carbon starvation was more common for gymnosperms. While loss of hydraulic function seems ubiquitous in drought-related mortality, not all researchers agree that hydraulic failure be the primary cause of mortality. Körner (2019) argued that hydraulic failure often correlates with a critical degree of tree dehydration and that the disruption of capillary continuum from soil to the root because of soil dehydration is more important because it forces trees to drop their leaves and rely on their own water reserves. Trees will die when the symplast surpasses its dehydration tolerance, and Körner (2019) considers xylem embolism as a byproduct rather than the cause of critical tree dehydration under severe drought, so that mechanistic models should focus on the dehydration process. Other authors have stressed the importance of the interplay between water reserves and water loss rates through cuticle after stomatal closure (Blackman *et al.* 2016; Martin-StPaul *et al.* 2017). Regardless of causality, loss of hydraulic function could still be a good indicator of mortality likelihood. However, recently Martinez-Vilalta *et al.* (2019) suggested use the relative water content as an indicator of mortality risk, as it integrates several aspects related to hydraulics, stomatal responses and carbon economy during drought.

Starvation thresholds

We use the amount of metabolic sugar in the stem as indicator of carbon starvation. Martínez-Vilalta *et al.* (2016) found that plants can have very distinct amounts of non-structural carbon concentration but minimum seasonal values are around 46% of maximum values. Accordingly, we define the indicator of starvation in relative terms (see control variable `mortalityRelativeSugarThreshold`, set to 30% by default) with respect to the stem equilibrium sugar concentration.

Dessication thresholds

Following Martinez-Vilalta *et al.* (2019), we primarily use the relative water content (RWC) in the stem symplastic tissues as the stress variable increasing dessication likelihood under drought. Whereas the stem

symplastic pressure-volume curve is always used to determine RWC from the stem water potential, in the basic water balance model the latter is assumed to be equal to the estimated plant water potential (i.e. $\Psi_{sym,stem} = \Psi_{plant}$; see 6.2.1). Moreover, in the advanced water balance model there is no distinction between stem apoplastic and symplastic (i.e. parenchymatic) water potentials (i.e. $\Psi_{sym,stem} = \Psi_{apo,stem}$) unless plant water compartments are explicitly simulated.

Using stem relative water content as indicator of dessication has the inconvenient, however, that the parameters $\pi_{0,stem}$ and ϵ_{stem} specifying the stem pressure-volume curve are often not known, so that estimates have to come from its covariation with other traits (here wood density, see @ref{inputationPV}). The uncertainty coming from this parameterization issue can be avoided if one combines the use of RWC with the loss of xylem hydraulic conductivity, since parameters of the hydraulic vulnerability curve are much more frequently available. For this reason, we use the **maximum of stem relative water content and relative xylem hydraulic conductance** as the drought stress variable to determine dessication.

The critical threshold for dessication is specified in the control variable `mortalityRWCThreshold`, set to 40% by default. Note that most studies focus on leaf RWC, for which Kursar *et al.* (2009) indicate ranges between 16% and 61%, and Körner (2019) advocates for values around 50%.

Details of the implementation of plant mortality are given in section 17.4.

15.1.6 Fire occurrence and severity

The occurrence of fires is represented in the growth model in a forcing variable that describes the daily probability that a wildfire strikes the stand. Whenever a fire occurs, the model first simulates fire behavior according to the conditions of the stand (see chapter 24) and then fire severity is evaluated.

The degree to which vegetation is impacted by fire depends on the heat fluxes incident on plant tissues and organs, which is an outcome of various fire behavior variables (such as fireline intensity and residence time) and the position of plant organs in the stand, and the physiological responses triggered thereafter. Fire effects on plants can be classified as two types: first- and second-order effects (Hood *et al.* 2018; Bär *et al.* 2019). First-order effects comprise the immediate impacts of heat transfer on plant tissues. Nonlethal first-order heat injuries can trigger second-order effects, such as physiological limitations in carbon and water relations or increased susceptibility to insect attacks and pathogenic infections. The growth model currently deals with first-order effects only. Moreover, the model focuses on effects in above-ground tissues (leaves, buds and cambium), disregarding fire effects on below-ground structures such as seed banks, or below-ground meristems, which are also important for the resilience of ecosystems (Stephan *et al.* 2010).

Fire-induced plant injury and mortality models can be grouped into two categories: (1) statistical models that involve regression equations relating injury and mortality to observable indicators; and (2) process models that explicitly simulate the underlying thermal and biophysical processes occurring when a plant is heated (Butler & Dickinson 2010; Hood *et al.* 2018). The design of first-order fire effects on vegetation follows, with some modifications, the process-based model proposed by Michaletz & Johnson (2008) for surface fires, and it is outlined in fig. 15.3. The surface fire behavior variables required for severity evaluation are the residence time and fireline intensity. With them, the model estimates the air temperature gradient of the plume, which is then compared to the distribution of foliage for the determination of the proportion of the crown foliage and crown buds that die due to heat accumulation. In addition, the model determines the effects on the cambium by estimating the radial necrosis depth and comparing it with bark thickness. If enough heat reaches the cambium, the tree is considered girdled, which leads to above-ground mortality. Otherwise, the degree of crown bud necrosis will determine whether the plant can resprout from the crown (and rebuild the leaf area lost) or not.

Crown fires will be considered to impact the forest stand whenever surface fireline intensity is larger than the critical value considered necessary for crowning (Scott & Reinhardt 2002). Crown fires are considered to burn for all plant cohorts, 100% of crown foliage and, most likely, 100% of crown buds (although very fast crown fires could spare buds if sufficiently protected, (Michaletz & Johnson 2008)). When crown fires do not occur at the stand level, torching may still occur for particular plant cohorts depending on its crown base height, with the same effects.

If 100% of crown buds have died but the cambium is alive and the species has dormant buds beneath the bark, then the plant has suffered crown kill but it can resprout epicormically without losing height (Hood *et al.* 2018). If the cambium is dead, however, all above-ground plant organs will be considered dead (i.e. topkill; Hood *et al.* (2018)). At this point, the plant may still exhibit basal resprouting from the root crown or below-ground meristems, but is dealt in the forest dynamics model (see chapter 18).

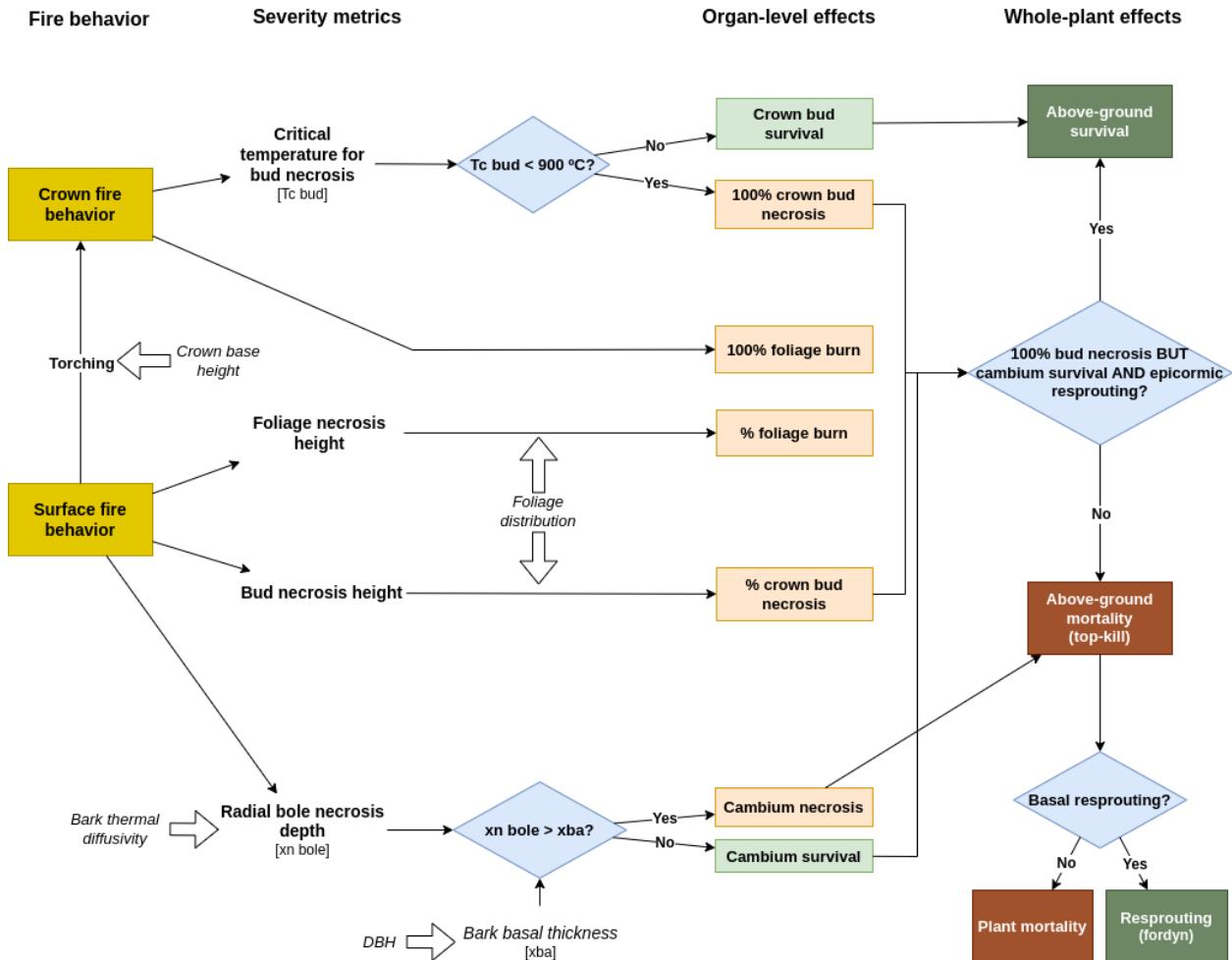


Figure 15.4: Schematic representation of the design of fire effects on foliage, buds and cambium.

While the severity sub-model described above is based on heat transfer and accumulation in tissues, causing necrosis, it is important to note that a growing number of studies increasingly have been documenting fire impacts on xylem hydraulic function, both via increased vapor pressure deficit (causing cavitation) and via structural damage in the xylem, affecting its efficiency and vulnerability (Hood *et al.* 2018; Bär *et al.* 2019). These first-order effects are not considered here.

Details of the implementation of fire severity are given in section 17.5.

15.1.7 Tree diameter, tree height and shrub cover updates

Tree diameter increment follows the creation of sapwood area, whereas sapwood area decreases do not affect diameter, as they imply conversion to heartwood. Tree height increments are estimated from diameter increments following diameter-height ratios that are modulated by the proportion of light available to the plant cohort, as frequently done in forest gap models (Lindner *et al.* 1997). Tree crown ratios are updated

using static allometric relationships, whereas shrub height and cover are estimated from new leaf area values by inverting static leaf biomass allometries.

Further details and formulation of structural update are given in chapter 17.6.

15.2 State variables

Many state variables are inherited by the corresponding water balance model and depend on its level of complexity (see 3.2 and 7.2). Several *cohort-level state variables* were treated as fixed parameters in the water balance models. For each plant cohort:

- Leaf area index of live leaves, LAI^{live} , and the actual expanded leaf area, LAI^ϕ .
- Diameter at breast height for trees, DBH (cohort average).
- Percent cover of shrubs, $Cover$.
- Density of individuals in the plant cohort, N .
- Plant height, H (cohort average).
- Crown ratio, CR .
- The proportion of fine roots, FRP_s , in each soil layer s .
- The radial length of coarse roots $L_{radial,s}$ in each soil layer s .

Additional *individual-level state variables* are needed in growth model simulations:

- Leaf area, LA (m^2), and target leaf area, LA_{target} (m^2).
- Sapwood area, SA (cm^2), and target sapwood area, SA_{target} (cm^2).
- The percentage of the crown with active buds ($PCAB$).
- Fine root biomass, $B_{fineroot,s}$ ($g\ dry$), and the target fine root biomass, $B_{fineroot,target,s}$ ($g\ dry$), in each soil layer s .
- Sugar concentration in leaves, SS_{leaf} ($mol\ gluc \cdot L^{-1}$).
- Starch concentration in leaves, ST_{leaf} ($mol\ gluc \cdot L^{-1}$).
- Sugar concentration in sapwood (stem, branches and coarse roots), $SS_{sapwood}$ ($mol\ gluc \cdot L^{-1}$).
- Starch concentration in sapwood (stem, branches and coarse roots), $ST_{sapwood}$ ($mol\ gluc \cdot L^{-1}$).

Note that LA is the same state variable as LAI^ϕ , but expressed at the individual level, and the same occurs between LA_{target} and LAI^{live} . These pairs of state variables are related by cohort density (N).

Other functional variables can be derived from the true state variables and, hence, hence do not need to be treated as state variables themselves but are often used in equations instead of the true state variables:

- Leaf biomass (B_{leaf} ; in $g\ dry$) from LA .
- Sapwood biomass ($B_{sapwood}$; in $g\ dry$) from SA , H and FRP_s and $L_{radial,s}$.
- Leaf storage volume ($V_{storage,leaf}$; in L) from B_{leaf} .
- Sapwood storage volume ($V_{storage,sapwood}$; in L) from SA , H and FRP_s and $L_{radial,s}$.
- Fine root area (FRA_s ; in m^2) in each soil layer s , from $B_{fineroot,s}$.

Some belowground variables (such as the radial length of coarse roots) are only considered dynamic when simulating growth with advance water balance.

15.3 Mass and energy balances

15.3.1 Water and energy balance

Since the growth model builds on soil and plant water balance models, the water and energy balances explained in sections 3.3 or 7.3 (depending on `transpirationMode`) also hold for this model.

15.3.2 Labile carbon balance

The model performs daily carbon balance of *labile carbon* in leaves and sapwood (stem, branches and coarse roots) for each plant cohort. In each of these two compartments carbon balance is performed for the two labile carbon forms, dissolved sugars (i.e. *metabolic carbon*) and starch (i.e. *storage carbon*). In the following all carbon flows (i.e. the components of right hand side of equations) are assumed to be in units of $g\text{ gluc}\cdot day^{-1}$. At the leaf level, the carbon balance equations for dissolved sugars and starch are (*i* subscripts omitted):

$$\Delta SS_{leaf} \cdot V_{storage,leaf} = A - M_{leaf} - F_{phloem} - SC_{leaf} - TS_{leaf} \quad (15.1)$$

$$\Delta ST_{leaf} \cdot V_{storage,leaf} = SC_{leaf} - TT_{leaf} \quad (15.2)$$

where A is (gross) photosynthesis, M_{leaf} is leaf maintenance respiration, F_{phloem} is the sugar flow via phloem towards the stem and roots, SC_{leaf} is the conversion from leaf sugar to leaf starch, TS_{leaf} and TT_{leaf} are the translocation of leaf sugars and starch into sapwood storage, respectively, during leaf senescence (see (15.6) below) or if leaf storage capacity is surpassed.

The corresponding carbon balance equations for sapwood dissolved sugars and starch are:

$$\Delta SS_{sapwood} \cdot V_{storage,sapwood} = F_{phloem} - M_{sapwood} - M_{fineroot} \quad (15.3)$$

$$-SC_{sapwood} - TS_{sapwood} \quad (15.4)$$

$$\Delta ST_{sapwood} \cdot V_{storage,sapwood} = SC_{sapwood} + TS_{leaf} + TT_{leaf} + TS_{sapwood} \quad (15.5)$$

$$-G_{sapwood} - G_{leaf} - G_{fineroot} - RE_{sapwood} \quad (15.6)$$

where F_{phloem} is the sugar inflow via phloem transport, $M_{sapwood}$ is the sum of maintenance respiration of branches, stem and coarse roots, $M_{fineroot}$ is the maintenance respiration of fine roots, G_{leaf} , $G_{sapwood}$ and $G_{fineroot}$ are the carbon costs of forming new leaves, sapwood or fine roots (respectively), $SC_{sapwood}$ is the conversion of sapwood sugar to sapwood starch, $TS_{sapwood}$ is the translocation of sapwood sugars into starch during leaf senescence and $RE_{sapwood}$ is the root exudation of sapwood carbon.

If we define S_{labile} as the total amount of labile carbon in the plant (in $g\text{ gluc}$):

$$S_{labile} = (SS_{leaf} + ST_{leaf}) \cdot V_{storage,leaf} + (SS_{sapwood} + ST_{sapwood}) \cdot V_{storage,sapwood} \quad (15.7)$$

then labile carbon balance at the plant level reduces to:

$$\Delta S_{labile} = A - M - G - RE \quad (15.8)$$

where $M = M_{leaf} + M_{sapwood} + M_{fineroot}$ is plant-level maintenance respiration, $G = G_{leaf} + G_{sapwood} + G_{fineroot}$ is plant-level growth costs and $RE = RE_{sapwood}$ is plant-level root exudation.

15.3.3 Biomass balance

In addition to changes in labile carbon pools, the model also implements changes in the size of carbon compartments (leaves, sapwood and fine roots), as a result of the interplay between growth and senescence processes. Hence, we can define the following structural equations:

$$\Delta B_{leaves} = B_{leaves,growth} - B_{leaves,senescence} \quad (15.9)$$

$$\Delta B_{sapwood} = B_{sapwood,growth} - B_{sapwood,senescence} \quad (15.10)$$

$$\Delta B_{fineroot} = B_{fineroot,growth} - B_{fineroot,senescence} \quad (15.11)$$

where all variables would be in units of $g\text{ dry}\cdot day^{-1}$. And the following equation defines the structural biomass balance at the plant level:

$$\Delta B_{structure} = \Delta B_{leaves} + \Delta B_{sapwood} + \Delta B_{fineroot} \quad (15.12)$$

The biomass balance at the **individual level** is completed if we add the labile biomass balance to the structural biomass balance:

$$\Delta B_{plant} = \Delta B_{structure} + \Delta S_{labile} \quad (15.13)$$

At the **cohort level**, however, we need to take into account that some individuals will die, so that the biomass balance needs to incorporate mortality losses:

$$\Delta B_{cohort} = \Delta B_{plant} \cdot N_{cohort} - B_{mortality} \quad (15.14)$$

where N_{cohort} is the initial cohort density (before mortality occurred) and $B_{mortality}$ is the biomass loss due to mortality of individuals, which in order to close the balance has to be defined as:

$$B_{mortality} = (B_{plant} + \Delta B_{plant}) \cdot N_{dead} \quad (15.15)$$

where N_{dead} is the density of dead individuals and B_{plant} is the initial plant biomass.

15.3.4 Stand-level carbon balance

With the aim to produce outputs to be compared with Eddy-covariance flux data, the model produces aggregates of gross photosynthesis and respiration at the stand level, all expressed using units of $g C \cdot m^{-2}$.

- *Gross primary production (GPP)* is the sum of gross photosynthesis across plant cohorts.
- *Maintenance respiration* is the sum across cohorts of the carbon emitted to the atmosphere as a result of maintenance respiration, i.e. M .
- *Synthesis respiration* is the sum across cohorts of the carbon returned to the atmosphere as a result of the process of synthesizing new structures. Note that this is less than G , as it does not include the carbon allocated to the new structures.
- *Net primary production (NPP)* is GPP minus the maintenance and synthesis respiration terms.

15.4 Process scheduling

Growth simulations are conducted in **daily** time steps. If using the basic water balance sub-model, processes are scheduled following the *regular scheduling*. In contrast, if using the advanced water balance model, the growth processes can be simulated using a *detailed scheduling*.

15.4.1 Regular scheduling

For every day to be simulated, first executes soil water balance, transpiration and photosynthesis calculations by calling the basic water balance model (3.4). After that, the model performs the following steps for each plant cohort:

1. Determine the amount of live structural biomass (in leaves, sapwood and fine root compartments), storage volume (in leaves and sapwood) and storage capacity (16.1).
2. Calculate gross photosynthesis per individual (A), maintenance respiration for leaves (M_{leaf}), sapwood ($M_{sapwood}$) and fine roots ($M_{fineroot}$) (16.2).
3. If phenology indicates leaf unfolding, evaluate leaf temperature and turgor sink limitations, growth costs (G_{leaf}) and leaf area increase (17.1.2).
4. If sapwood growth is permitted, evaluate sapwood temperature and turgor sink limitations, growth costs ($G_{sapwood}$) and sapwood area increase (17.1.3).
5. Evaluate fineroot temperature and turgor sink limitations, growth costs ($G_{fineroot}$) and fineroot biomass increase (17.1.4).

6. Update changes in sugar and starch compartments due to photosynthesis, maintenance respiration and growth. Apply transfer from leaf metabolic to sapwood metabolic carbon (16.2.4) and conversions between storage and metabolic carbon (16.2.5).
7. Determine leaf senescence due to aging or cavitation effects (17.2.1).
8. Determine sapwood area senescence by aging (17.2.2).
9. Determine fine root senescence by aging (17.2.3).
10. Evaluate labile carbon translocation due to leaf and sapwood senescence (TS_{leaf} , TT_{leaf} and $TS_{sapwood}$). If leaf storage carbon is surpassed, add the excess to TT_{leaf} . Add the sum of translocated carbon to the starch sapwood pool.
11. Move any surplus in sapwood storage carbon to root exudation ($RE_{sapwood}$). This completes the daily carbon balance for leaf labile pools (15.2), sapwood labile pools (15.6), and plant-level labile carbon. (15.8).
12. Update leaf area, sapwood area, fine root area and derived variables (e.g., leaf area index and Huber value) (17.3.1).
13. Determine if a wildfire stroke the forest stand and, if so, fire severity effects (17.5).
14. Determine if plant death by starvation or desiccation occurred (17.4).
15. Update fine root area target and, if phenology indicates bud formation, update the leaf area target (17.3.2).
16. Translate sapwood area growth into structural variables (i.e., plant height, tree DBH, tree crown ratio and shrub cover), (17.6) and update storage volumes and the concentration of labile pools.

15.4.2 Advanced scheduling

For every day to be simulated, the model first executes soil water balance, energy balance, transpiration and photosynthesis calculations in subdaily times steps, by calling the advanced water balance model (7.4). After that, the model performs the following steps for each plant cohort:

1. Determine the amount of live structural biomass (in leaves, sapwood and fine root compartments), storage volume (in leaves and sapwood) and storage capacity (16.1).
2. For every **subdaily** step, perform carbon balance as follows:
 - a. Calculate gross photosynthesis per individual (A) and maintenance respiration for leaves (M_{leaf}), sapwood ($M_{sapwood}$) and fine roots ($M_{fineroot}$), taking into account both live structural biomass and labile sugar biomass (see 16.2).
 - b. If phenology indicates leaf unfolding, evaluate temperature and turgor sink limitations for growth of leaves, estimate growth costs (G_{leaf}) and record leaf area increases (17.1.2).
 - c. If sapwood growth is permitted, evaluate temperature and turgor sink limitations for growth of sapwood, estimate growth costs ($G_{sapwood}$) and record sapwood area increases (17.1.3).
 - d. Evaluate temperature and turgor sink limitations for growth of fine roots, estimate growth costs ($G_{fineroot}$) and record fine root biomass increases (17.1.4).
 - e. Evaluate phloem transport (F_{phloem}) and sugar-starch dynamics in the leaf and sapwood compartments **at 1s time steps** (16.2.4), including photosynthesis, maintenance respiration and growth respiration as well as diversion towards root exudation if maximum leaf or sapwood starch capacity is attained (RE_{leaf} and $RE_{sapwood}$). This completes the sub-daily carbon balance for plant-level labile carbon (15.8).
3. Determine leaf senescence due to aging or cavitation effects (17.2.1).
4. Determine sapwood area senescence by aging (17.2.2).
5. Determine fine root senescence by aging (17.2.3).
6. Evaluate labile carbon translocation due to leaf and sapwood senescence (TS_{leaf} , TT_{leaf} and $TS_{sapwood}$), adding the resulting carbon to the starch sapwood pool. This completes the daily carbon balance for leaf labile pools (15.2), sapwood labile pools (15.6), and plant-level labile carbon.

7. Update leaf area, sapwood area, fine root biomass and distribution. Update derived variables (e.g., leaf area index, Huber values, hydraulic conductance of stems, roots and rhizosphere, osmotic potential at full turgor of leaves and sapwood) (17.3.1).
8. Determine if plant death by starvation or desiccation occurred (17.4).
9. If phenology indicates bud formation, update the leaf area target (17.3.2). Update the fine root biomass target, according to the hydraulic conductance in the rhizosphere (17.3.2).
10. Translate sapwood area growth into structural variables (i.e., plant height, tree DBH, tree crown ratio and shrub cover), (17.6) and update storage volumes and the concentration of labile pools.

15.5 Inputs and outputs

15.5.1 Soil, vegetation and meteorology

Soil

Soil input requirements are the same as for the basic water balance model and were fully described in section 2.3. Genuchten (1980) water retention curves are forced for this model, because rhizosphere conductance is modelled using van Genuchten hydraulic equations (see subsection 10.2).

Vegetation

Vegetation input requirements were fully described in section 2.4. Trees and shrubs do not need to be characterized with different variables in soil water balance calculations, since both kinds of cohort have a *LAI* value. In most cases, users only need to estimate the leaf area index corresponding to live leaves, i.e. LAI^{live} , because one can assume no dead leaves in the canopy (i.e., $LAI^{dead} = 0$). Note that vegetation characteristics are modified by simulations in this model.

Metereological input

The minimum weather variables required to run the model are min/max temperatures (T_{min} and T_{max}), min/max relative humidity (RH_{min} and RH_{max}), precipitation (P) and solar radiation (Rad). Wind speed (u) is also needed, but the user may use missing values if not available (a default value will be used in this case). Wind speed is assumed to have been measured at a specific height above the canopy (by default at 2 m). Atmospheric CO_2 concentration (C_{atm}) may also be specified, but if missing a default constant value is assumed, which is taken from the control parameters. If wildfire occurrence and severity is to be simulated, the user has to specify daily fire probability (P_{fire}) in an additional column. Definitions and units of these variables were given in section 2.5.

15.5.2 Vegetation functional parameters

The growth model requires many functional parameters to be specified for plant cohorts. Some of them depend on whether the basic or advanced water balance is adopted. Here we report functional parameters needed in addition to those necessary for the water balance submodel.

A first set of parameters refers to anatomical features (`paramsAnatomy`), most of them already required by water balance model simulations:

Symbol	Units	R	Description
H_{max}	cm	Hmax	Maximum plant height
H_{med}	cm	Hmed	Median plant height
$1/H_v$	$m^2 \cdot m^{-2}$	A12As	Leaf area to sapwood area ratio
RLR	$m^2 \cdot m^{-2}$	Ar2A1	Fine root area to leaf area ratio
SLA	$m^2 \cdot kg^{-1}$	SLA	Specific leaf area
LW	cm	LeafWidth	Leaf width

Symbol	Units	R	Description
ρ_{leaf}	$g \cdot cm^{-3}$	LeafDensity	Dry tissue density
ρ_{wood}	$g \cdot cm^{-3}$	WoodDensity	Dry tissue density
$\rho_{fineroot}$	$g \cdot cm^{-3}$	FineRootDensity	Dry tissue density
$f_{conduits}$	[0-1]	conduitProportion	Proportion of sapwood corresponding to xylem conduits (the remaining corresponds to parenchymatic tissues)
SRL	$cm \cdot g^{-1}$	SRL	Specific root length
RLD	$cm \cdot cm^{-3}$	RLD	Fine root length density (i.e. density of root length per soil volume)
$r_{6.35}$		r635	Ratio between the weight of leaves plus branches and the weight of leaves alone for branches of 6.35 mm

A second set of parameters refers to allometric coefficients needed for structural updates (`paramsAllometries`).

Symbol	Units	R param	Description
a_{ash}		Aash	Coefficient relating the square of shrub height with shrub area
a_{bsh}, b_{bsh}		Absh, Bbsh	Coefficients relating crown volume with dry weight of shrub individuals
bt_{sh}		BTsh	Bark thickness in shrubs
$a_{cr}, b_{1cr}, b_{2cr}, b_{3cr}, c_{1cr}, c_{2cr}$		B1cr, B2cr, B3cr, C1cr, C2cr	Coefficients to calculate crown ratio of trees
a_{cw}, b_{cw}		Acw, Bcw	Regression coefficients used to calculate the crown width of trees (as intermediary step to obtain the crown ratio).
a_{bt}, b_{bt}		Abt, Bbt	Regression coefficients used to calculate the bark thickness of trees

A third set of parameters refers to growth parameters (`paramsGrowth`).

Symbol	Units	R param	Description
N_{leaf}	$mg N \cdot g dry^{-1}$	Nleaf	Leaf nitrogen concentration per dry mass
$N_{sapwood}$	$mg N \cdot g dry^{-1}$	Nsapwood	Sapwood nitrogen concentration per dry mass
$N_{fineroot}$	$mg N \cdot g dry^{-1}$	Nfineroot	Fine root nitrogen concentration per dry mass
MR_{leaf}	$g gluc \cdot g dry^{-1} \cdot day^{-1}$	RERleaf	Leaf respiration rate at 20 °C. Normally estimated via N_{leaf} .
$MR_{sapwood}$	$g gluc \cdot g dry^{-1} \cdot day^{-1}$	RERsapwood	Living sapwood (parenchymatic tissue) respiration rate at 20 °C. Normally estimated via $N_{sapwood}$.
$MR_{fineroot}$	$g gluc \cdot g dry^{-1} \cdot day^{-1}$	RERfineroot	Fine root respiration rate at 20 °C. Normally estimated via $N_{fineroot}$.
MR_{leaf}	$g gluc \cdot g dry^{-1} \cdot day^{-1}$	RERleaf	Leaf respiration rate at 20 °C

Symbol	Units	R param	Description
$MR_{sapwood}$	$g\text{ gluc} \cdot g\text{ dry}^{-1} \cdot day^{-1}$	RERsapwood	Living sapwood (parenchymatic tissue) respiration rate at 20 °C
$MR_{fineroot}$	$g\text{ gluc} \cdot g\text{ dry}^{-1} \cdot day^{-1}$	RERfineroot	Fine root respiration rate at 20 °C
$RGR_{leaf,max}$	$m^2 \cdot cm^{-2} \cdot day^{-1}$	RGRleafmax	Maximum leaf area daily growth rate, relative to sapwood area
$RGR_{cambium,max}$	$cm^2 \cdot cm^{-1} \cdot day^{-1}$	RGRcambiummax	Maximum daily tree sapwood growth rate relative to cambium perimeter length
$RGR_{sapwood,max}$	$cm^2 \cdot cm^{-2} \cdot day^{-1}$	RGRsapwoodmax	Maximum daily shrub sapwood growth rate relative to sapwood area
$RGR_{fineroot,max}$	$g\text{ dry} \cdot g\text{ dry}^{-1} \cdot day^{-1}$	RGRfinerootmax	Maximum daily fine root relative growth rate
CC_{leaf}	$g\text{ gluc} \cdot g\text{ dry}^{-1}$	CCleaf	Leaf construction cost
$CC_{sapwood}$	$g\text{ gluc} \cdot g\text{ dry}^{-1}$	CCsapwood	Sapwood construction cost
$CC_{fineroot}$	$g\text{ gluc} \cdot g\text{ dry}^{-1}$	CCfineroot	Fine root construction cost
$SR_{sapwood}$	day^{-1}	SRsapwood	Daily sapwood senescence rate
$SR_{fineroot}$	day^{-1}	SRfineroot	Daily fine root senescence rate
$RSSG$	[0-1]	RSSG	Minimum relative starch for sapwood growth to occur.
$f_{HD,min}$	$cm \cdot cm^{-1}$	fHDmin	Minimum height-to-diameter ratio
$f_{HD,max}$	$cm \cdot cm^{-1}$	fHDmax	Maximum height-to-diameter ratio
	$g\text{ C} \cdot g^{-1}\text{ dry}$	WoodC	Wood carbon content per dry weight

A third set of parameters refers to mortality/regeneration parameters (`paramsMortalityRegeneration`).

Symbol	Units	R param	Description
$P_{mort,base}$	[0 – 1]	MortalityBaselineRate	Deterministic proportion or probability specifying the baseline reduction of cohort's density occurring in a year as a result of processes not explicitly modelled
TS_{surv}	[0 – 1]	SurvivalModelStep	Time step in years of the empirical survival model depending on stand basal area (e.g. 10)
$\beta_{surv,0}$	[0 – 1]	SurvivalB0	Intercept of the logistic baseline survival model depending on stand basal area
$\beta_{surv,1}$	[0 – 1]	SurvivalB1	Slope of the logistic baseline survival model depending on stand basal area
$N_{tree,recr}$	$ind \cdot ha^{-1}$	RecrTreeDensity	Density of tree recruits from seeds.
$N_{tree,ingrowth}$	$ind \cdot ha^{-1}$	IngrowthTreeDensity	Density of trees reaching ingrowth DBH.
$DBH_{tree,recr}$	cm	RecrTreeDBH	DBH for tree recruits from seeds or resprouting (e.g. 1 cm).
$DBH_{tree,ingrowth}$	cm	IngrowthTreeDBH	Ingrowth DBH for trees (e.g. 7.5 cm).

Vegetation functional attributes are normally filled for each cohort by function `growthInput()` or `forest2growthInput()` from species identity. However, different parameters can be specified for different cohorts of the same species if desired (see section 2.4.8).

15.5.3 Control parameters

Control parameters modulate the overall behaviour of growth simulations (see section 2.6). Parameters controlling water balance have been already described in sections 3.5.3 and 7.5.3. Here we only mention those control parameters specific to the `growth()` function.

First, there are flags that regulate the application of processes:

- `allowDessication [=TRUE]`: Boolean flag to indicate that mortality by dessication is allowed.
- `allowStarvation [=TRUE]`: Boolean flag to indicate that mortality by starvation is allowed.
- `sinkLimitation [=TRUE]`: Boolean flag to indicate that temperature and turgor limitations to growth are applied.
- `shrubDynamics [= TRUE]`: Boolean flag to allow the application of demographic processes to shrubs.
- `herbDynamics [= TRUE]`: Boolean flag to allow dynamic herb leaf area as a function of shading due to leaf area of woody cohorts.

An important parameter concerns the allocation strategy:

- `allocationStrategy [= "Plant_kmax"]` defines the allocation strategy (investment in leaf area vs. sapwood area) of the growth model. If `allocationStrategy = "Plant_kmax"` then the target leaf area is set to maintain the input whole-plant maximum conductance. If `allocationStrategy = "A12As"` then the target leaf area is set to maintain the input Huber value.

A number of control parameters regulate carbon balance:

- `phloemConductanceFactor [= 0.2]`: Factor to transform stem xylem conductance to phloem conductance (only for `transpirationMode = "Sperry"`).
- `nonSugarConcentration [=0.25]`: A constant specifying non-sugar (inorganic) solute concentration ($mol \cdot L^{-1}$) in cells.
- `equilibriumOsmoticConcentration [= c(leaf = 0.8, sapwood = 0.6)]`: Equilibrium osmotic concentrations ($mol \cdot L^{-1}$) for leaf and sapwood cells. The difference between leaf and sapwood values helps maintaining phloem transport. The equilibrium sugar concentration is found using `equilibriumOsmoticConcentration - nonSugarConcentration` and defaults to `[= c(leaf = 0.55, sapwood = 0.35)]`.
- `minimumRelativeStarchForGrowth [= 0.50]`: Default minimum concentration of storage carbon (starch), relative to the maximum storage capacity, for sapwood growth to occur. This default value is used when species-specific values are not specified via `SpParams`.
- `senescenceRates [= c(sapwood = 0.0001261398, fineroot = 0.001897231)]`: Relative senescence rates (day^{-1}) for sapwood and fine roots. Default are equivalent to 4.5% and 50% annual turnover for sapwood and fine roots, respectively. Leaf senescence rates depend on leaf lifespan (i.e. leaf duration) for evergreen species. These are default values used when relative growth rates are not specified via species-specific `SpParams`.
- `constructionCosts [= c(leaf = 1.5, sapwood = 1.47, fineroot = 1.30)]`: Default construction costs, including respiration and structural carbon, per dry weight of new tissue ($g\ gluc \cdot g\ dry^{-1}$). These are default values used when relative growth rates are not specified via species-specific `SpParams`.
- `maximumRelativeGrowthRates [= c(leaf = 0.01, cambium = 0.005, sapwood = 0.002, fineroot = 0.1)]`: Default maximum relative growth rates for leaves ($m^2 \cdot cm^{-2} \cdot day^{-1}$), tree sapwood ($cm^2 \cdot cm^{-1} \cdot day^{-1}$), shrub sapwood ($cm^2 \cdot cm^{-2} \cdot day^{-1}$) and fine roots ($g\ dry \cdot g\ dry^{-1} \cdot day^{-1}$). These are default values used when relative growth rates are not specified via species-specific `SpParams`.

Finally, we have the following options regarding mortality:

- `mortalityMode` [= "density/deterministic"]: String describing how mortality is applied. Current accepted values are combinations of "cohort" vs "density" (for whole-cohort mortality vs reduction of stem density) and "deterministic" vs. "stochastic".
- `mortalityBaselineRate` [= 0.0015]: Default deterministic proportion or probability specifying the baseline reduction of cohort's density occurring in a year as a result of processes not explicitly modelled (for `mortalityMode` = "density/deterministic" or "density/stochastic").
- `mortalityRelativeSugarThreshold` [= 0.4]: Threshold of stem sugar concentration relative to equilibrium values (by default 40%), resulting in an increased mortality rate/probability whenever levels are below.
- `mortalityRWCThreshold` [= 0.4]: Threshold of stem relative water content resulting in increased mortality rate/probability whenever levels are below (by default 40%).

15.5.4 Model output

15.5.4.1 Daily output

Functions `growth()` returns a list of a class with the same name. The first four elements of this list (i.e., `latitude`, `topography`, `weather` and `growthInput`) are simply copies of model inputs. The next element is `growthOutput`, which contains the state of the input object at the end of the simulation (this can be used to perform further simulations starting with current values of state variables). The remaining list elements are:

Element	Description
<code>WaterBalance</code>	Climatic input and water balance flows (i.e. net precipitation, infiltration, runoff, transpiration...). All of them in $mm = L \cdot m^{-2}$.
<code>EnergyBalance</code>	Energy balance components (instantaneous values) for the soil and the canopy
<code>CarbonBalance</code>	Stand-level carbon balance components (gross primary production, maintenance respiration, synthesis respiration and net primary production). All of them in $g C \cdot m^{-2}$.
<code>BiomassBalance</code>	Stand-level biomass balance components. All of them in $g dry \cdot m^{-2}$.
<code>Temperature</code>	Minimum and maximum temperature values for topsoil, canopy and atmosphere (if <code>transpirationMode</code> = "Sperry")
<code>Soil</code>	Soil variables for each soil layer: Moisture relative to field capacity (W_s), water potential (Ψ_s) and volumetric water content (V_s).
<code>Stand</code>	Stand-level variables, such as $LAI_{stand}^\phi = LAI_{stand}^{live}$, LAI_{stand}^{dead} , the water retention capacity of the canopy (S_{canopy}) or the fraction of light reaching the ground (L_{ground}^{PAR} and L_{ground}^{SWR}).
<code>Plants</code>	Plant-level variables, such as LAI_i^{phi} , plant transpiration, plant water potential, etc.
<code>SunlitLeaves</code>	Leaf-level output variables for sunlit leaves (<code>transpirationMode</code> = "Sperry").
<code>ShadeLeaves</code>	Leaf-level output variables for shade leaves (<code>transpirationMode</code> = "Sperry").
<code>LabileCarbonBalance</code>	A list with the daily results of labile carbon balance components and carbon pools.
<code>PlantBiomassBalance</code>	A list with the daily results of biomass balance components for plant cohorts.
<code>PlantStructure</code>	A list with daily results of plant compartment sizes, in terms of area and biomass.
<code>GrowthMortality</code>	A list with daily results of growth rates for leaf, sapwood and fine roots; as well as daily mortality rates.

Elements `WaterBalance` to `ShadeLeaves` are equal to the output of function `spwb()` and were described in previous chapters (see 3.5.4 and 7.5.4).

The following table describes the child elements of `LabileCarbonBalance`, each of them a data frame with dates in rows and plant cohorts in columns:

Element	Symbol	Units	Description
GrossPhotosynthesis	A_{B_i}	$g\text{ gluc} \cdot g\text{ dry}^{-1} \cdot day^{-1}$	Daily gross photosynthesis per dry weight of living biomass.
MaintenaceRespiration	R_{B_i}	$g\text{ gluc} \cdot g\text{ dry}^{-1} \cdot day^{-1}$	Daily maintenance respiration (including leaves, sapwood and fine roots) per dry weight of living biomass.
GrowthCosts	G_i/B_i	$g\text{ gluc} \cdot g\text{ dry}^{-1} \cdot day^{-1}$	Daily growth costs (including leaves, sapwood and fine roots) per dry weight of living biomass.
RootExudation	RE_i	$g\text{ gluc} \cdot g\text{ dry}^{-1} \cdot day^{-1}$	Daily root exudation per dry weight of living biomass.
CarbonBalance	$\Delta S_i/B_i$	$g\text{ gluc} \cdot g\text{ dry}^{-1}$	Daily labile carbon balance (eq. (15.8)) per dry weight of living biomass.
SugarLeaf	$SS_{leaf,i}$	$mol\text{ gluc} \cdot L^{-1}$	Sugar concentration in leaves.
StarchLeaf	$ST_{leaf,i}$	$mol\text{ gluc} \cdot L^{-1}$	Starch concentration in leaves.
SugarSapwood	$SS_{sapwood,i}$	$mol\text{ gluc} \cdot L^{-1}$	Sugar concentration in sapwood.
StarchSapwood	$ST_{sapwood,i}$	$mol\text{ gluc} \cdot L^{-1}$	Starch concentration in sapwood.
SugarTransport	F_{phloem}	$mol\text{ gluc} \cdot s^{-1}$	Average instantaneous rate of carbon transferred between leaves and stem compartments via phloem.

The following table describes the child elements of `PlantCarbonBalance`, each of them a data frame with dates in rows and plant cohorts in columns:

Element	Symbol	Units	Description
StructuralBiomassBalance		$g\text{ dry} \cdot m^{-2} \cdot day^{-1}$	Daily structural biomass balance.
LabileBiomassBalance		$g\text{ dry} \cdot m^{-2} \cdot day^{-1}$	Daily labile biomass balance.
PlantBiomassBalance		$g\text{ dry} \cdot m^{-2} \cdot day^{-1}$	Daily plant biomass balance.
MortalityBiomassLoss		$g\text{ dry} \cdot m^{-2} \cdot day^{-1}$	Daily biomass loss due to mortality.
CohortBiomassBalance		$g\text{ dry} \cdot m^{-2} \cdot day^{-1}$	Daily cohort biomass balance (includes plant biomass balance and mortality).

The following table describes the child elements of `PlantStructure`, again each of them a data frame with dates in rows and plant cohorts in columns:

Element	Symbol	Units	Description
LeafBiomass	$B_{leaf,i}$	$g \text{ dry} \cdot ind^{-1}$	Daily leaf structural biomass for an average individual.
SapwoodBiomass	$B_{sapwood,i}$	$g \text{ dry} \cdot ind^{-1}$	Daily sapwood structural biomass for an average individual.
FineRootBiomass	$B_{fineroot,i}$	$g \text{ dry} \cdot ind^{-1}$	Daily fine root biomass for an average individual.
LeafArea	LA_i	m^2	Daily amount of leaf area for an average individual.
SapwoodArea	SA_i	cm^2	Daily amount of sapwood area for an average individual.
FineRootArea	FRA_i	m^2	Daily amount of fine root area for an average individual.
HuberValue	H_v	$cm^2 \cdot m^{-2}$	The ratio of sapwood area to (target) leaf area
RootAreaLeafArea		$m^2 \cdot m^{-2}$	The ratio of fine root area to (target) leaf area
DBH	DBH_i	cm	Daily diameter at breast height for an average individual.
Height	H_i	cm	Daily total height for an average individual.

The following table describes the child elements of `GrowthMortality`, again each of them a data frame with dates in rows and plant cohorts in columns:

Element	Symbol	Units	Description
LAGrowth	$\Delta LA_i / SA_i$	$m^2 \cdot cm^{-2} \cdot day^{-1}$	Leaf area growth relative to sapwood area for an average individual.
SAgrowth	$\Delta SA_i / SA_i$	$cm^2 \cdot cm^{-2} \cdot day^{-1}$	Leaf area growth relative to sapwood area for an average individual.
FRAgrowth	$\Delta FRA_i / SA_i$	$m^2 \cdot cm^{-2} \cdot day^{-1}$	Fine root area growth relative to sapwood area for an average individual.
StarvationRate		$ind \cdot ind^{-1} \cdot day^{-1}$	Daily mortality rate from starvation.
DessicationRate		$ind \cdot ind^{-1} \cdot day^{-1}$	Daily mortality rate from dessication.
MortalityRate		$ind \cdot ind^{-1} \cdot day^{-1}$	Daily mortality rate (any cause).

15.5.4.2 Subdaily output

As with function `spwb()`, subdaily output can also be generated with function `growth()`. This is done via setting `subdailyResults = TRUE` in the control parameter list. In that case, the output list of `growth()` will include an element `subdaily`, whose subelements correspond to the results of `growth_day()` for each date simulated. Refer to the documentation of function `growth_day()` for details of sub-daily output.

Chapter 16

Carbon pools and components of the carbon balance

16.1 Size of carbon pools

Biomass of leaves, sapwood and fine roots is needed in the model to estimate respiratory costs and the size of the C storage pools.

16.1.1 Leaf structural, metabolic and storage biomass

The live leaf area LA^{live} per individual (in m^2) is:

$$LA^{live} = 10000 \cdot LAI^{live}/N \quad (16.1)$$

where N is the density of the cohort and LAI^{live} is the leaf area index. The actual (expanded) leaf area LA^ϕ is estimated analogously from LAI^ϕ :

$$LA^\phi = 10000 \cdot LAI^\phi/N \quad (16.2)$$

Structural leaf biomass per individual (B_{leaf} ; $g\ dry\ ünd^{-1}$) is the result of dividing leaf area by SLA ($m^2 \cdot kg^{-1}$), the specific leaf area of the species:

$$B_{leaf} = 1000 \cdot LA^\phi/SLA \quad (16.3)$$

where a factor 1000 is used to convert from kg to g. Hence, only expanded leaf area has respiratory cost (i.e. winter resistance buds do not). Leaf storage volume for an individual ($V_{storage,leaf}$; in $L \cdot ind^{-1}$) is:

$$V_{storage,leaf} = LA^\phi \cdot V_{leaf} \quad (16.4)$$

where V_{leaf} is the water storage capacity of leaf tissue per leaf area unit (in $L \cdot m^{-2}$) (see A.4.5). Leaf storage volume is necessary to estimate the biomass of metabolic (sugars) or storage (starch) from the concentration of these substances in leaves (SS_{leaf} and ST_{leaf} , which are expressed in $mol\ gluc \cdot L^{-1}$), and conversely. For example:

$$B_{sugar,leaf} = SS_{leaf} \cdot V_{storage,leaf} \cdot m_{gluc} \quad (16.5)$$

where m_{gluc} is the molar mass of glucose. To estimate leaf starch storage capacity, the model assumes that up to 10% of leaf tissue volume can be occupied with starch, so that the leaf starch storage capacity (ST_{leaf}^{max} ; in $mol \cdot ind^{-1}$) is:

$$ST_{leaf}^{max} = 0.1 \cdot 1000 \cdot \frac{V_{storage,leaf} \cdot \rho_{starch}}{m_{starch}} \quad (16.6)$$

where ρ_{starch} is the density of starch and m_{starch} is its molar mass.

16.1.2 Sapwood structural, metabolic and storage biomass

Sapwood volume ($V_{sapwood}$; in $L \cdot \text{ind}^{-1}$) is defined as the product of sapwood area and the sum of aboveground height and belowground coarse root length:

$$V_{sapwood} = 1000 \cdot SA \cdot (H + \sum_s FRP_s \cdot L_s) \quad (16.7)$$

where SA is sapwood area, H is plant height, FRP_s is the proportion of fine roots in soil layer s and L_s is the coarse root length in layer s (all lengths expressed here in m). Sapwood structural biomass per individual ($B_{sapwood}$; $\text{g dry} \cdot \text{ind}^{-1}$) represents the sapwood biomass of sapwood of trunks, branches and coarse roots. It is defined as the product of $V_{sapwood}$ and wood density (ρ_{wood} ; $\text{gdry} \cdot \text{cm}^{-3}$):

$$B_{sapwood} = 1000 \cdot V_{sapwood} \cdot \rho_{wood} \quad (16.8)$$

While $B_{sapwood}$ represents the structural sapwood biomass, it cannot be used to estimate sapwood respiration, since only xylem axial or radial parenchymatic rays (and not dead cells like tracheids or vessels) contribute to sapwood respiration. In addition, we assume that embolized parts of sapwood do not contribute to sapwood respiration. Hence, sapwood living (respiratory) biomass is estimated using:

$$B_{living,sapwood} = 1000 \cdot (1 - PLC_{stem}) \cdot V_{sapwood} \cdot \rho_{wood} \cdot (1 - f_{conduits}) \quad (16.9)$$

where PLC_{stem} is the current proportion of stem conductance lost and $f_{conduits}$ is the fraction of sapwood volume that corresponds to dead conduits, being the complement of the fraction of parenchymatic tissue. The volume available for metabolic or storage carbon within sapwood is limited by both the sapwood fraction that corresponds to parenchyma and wood porosity (i.e. $(1 - (\rho_{wood}/1.54))$). Sapwood storage volume ($V_{storage,sapwood}$; in $L \cdot \text{ind}^{-1}$) is:

$$V_{storage,sapwood} = V_{sapwood} \cdot (1 - (\rho_{wood}/1.54)) \cdot (1 - f_{conduits}) \quad (16.10)$$

Analogously to leaf carbon, $V_{storage,sapwood}$ is used to estimate the biomass of metabolic (sugars) or storage (starch) from the concentration of these substances in sapwood ($SS_{sapwood}$ and $ST_{sapwood}$, which are expressed in $\text{mol gluc} \cdot L^{-1}$), and conversely. For example:

$$B_{sugar,sapwood} = SS_{sapwood} \cdot V_{storage,sapwood} \cdot m_{gluc} \quad (16.11)$$

where m_{gluc} is the molar mass of glucose. To estimate sapwood storage capacity, the model assumes that up to 50% of volume of parenchymatic cells can be occupied with starch, so that the sapwood starch storage capacity ($ST_{sapwood}^{max}$; in $\text{mol} \cdot \text{ind}^{-1}$) is:

$$ST_{sapwood}^{max} = 0.5 \cdot 1000 \cdot \frac{V_{storage,sapwood} \cdot \rho_{starch}}{m_{starch}} \quad (16.12)$$

16.1.3 Fine root area and biomass

The fine root compartment does not have labile carbon (i.e. the model supplies labile carbon for fine roots from sapwood compartment), so that only the structural biomass of fine roots needs to be estimated. If growth is simulated using the *basic* water balance model, the initial area of fine roots (FRA , m^2) for layer s is estimated from leaf area using:

$$FRA = FRP_s \cdot LA^\phi \cdot RLR \quad (16.13)$$

where FRP_s is the proportion of fine roots in layer s and RLR is the species-specific root area to leaf area ratio. Biomass of fine roots per individual for a given layer s ($B_{fineroot,s}$; $\text{g dry} \cdot \text{ind}^{-1}$) is then estimated from FRA_s using:

$$B_{fineroot,s} = \frac{10^4 \cdot FRA_s}{2.0 \cdot \sqrt{\frac{\pi \cdot SRL}{\rho_{fineroot}}}} \quad (16.14)$$

where $\rho_{fineroot}$ is fine root tissue density ($g \text{ dry} \cdot cm^{-3}$) and SRL ($cm \cdot g \text{ dry}^{-1}$) is the specific root length. Fine root biomass per individual ($g \text{ dry} \cdot ind^{-1}$) is simply the sum across layers:

$$B_{fineroot} = \sum_s B_{fineroot,s} \quad (16.15)$$

If growth is simulated using the *advanced* water balance model, the initial fine root length per area in each soil layer (FLA_s ; $m \cdot m^{-2}$) is estimated from rhizosphere conductance, assuming a cylindrical flow geometry in the rhizosphere. The initial fine root biomass for a given layer s ($g \text{ dry} \cdot ind^{-1}$) is then calculated using:

$$B_{fineroot,s} = \frac{10000 \cdot FLA_s}{N \cdot 0.01 \cdot SRL} \quad (16.16)$$

where N is the density of individuals. Finally, initial fine root area in each soil layer (FRA_s) is then calculated from $B_{fineroot}$ by inverting the equation presented above:

$$FRA_s = 10^{-4} \cdot B_{fineroot,s} \cdot 2.0 \cdot \sqrt{\frac{\pi \cdot SRL}{\rho_{fineroot}}} \quad (16.17)$$

16.1.4 Total living biomass

Total living biomass per individual (i.e. not accounting for heartwood or xylem conduits) is:

$$\begin{aligned} B_{total} = & B_{leaf} + B_{living,sapwood} + B_{fineroot} + \\ & + B_{sugar,leaf} + B_{starch,leaf} + B_{sugar,sapwood} + B_{starch,sapwood} \end{aligned}$$

B_{total} is used to express carbon balance components per dry weight of living biomass in the model output, which allows a better comparison across plant cohorts of different size and species.

16.2 Components of labile carbon balance

Here we provide the details of labile carbon balance calculations, i.e. how the components of eqs. (15.2) and (15.6) are determined.

16.2.1 Gross photosynthesis

The soil water balance submodel provides values of photosynthesis calculated at the plant cohort level, but note that the user output of function `spwb` and `pwb` refers to net photosynthesis (A_n) after accounting for an estimate of leaf respiration. In the basic water balance model there is no way to make the distinction between gross and net photosynthesis, so we will assume here that $A_g = A_n$ (in $g C \cdot m^{-2}$) as calculated in section 6.1.6. In the advance water balance model the output is also net photosynthesis, but the distinction is possible (see section 11.1). In any case, here we start with a A_g value in units of carbon per ground area unit, i.e. $g C \cdot m^{-2}$, and to enter the carbon balance we need scale photosynthesis to units of glucose per individual (A ; in $g \text{ gluc} \cdot ind^{-1}$). This is done using:

$$A = \frac{10000 \cdot A_g}{N} \cdot \frac{m_{gluc}}{m_{carbon} \cdot 6} \quad (16.18)$$

where N is the density of individuals per hectare, m_{gluc} is the glucose molar mass and m_{carbon} is the carbon molar mass.

16.2.2 Maintenance respiration

Maintenance respiration (in $g\text{ glucind}^{-1}$) is calculated for each of the three compartments (leaves, sapwood, and fine roots) individually (Mouillot *et al.* 2001), and differs slightly depending on the complexity of the transpiration submodel.

When simulating growth with the basic water balance, the model uses a Q_{10} relationship of baseline respiration with temperature, which means that for every 10 °C change in temperature there is a Q_{10} factor change in respiration:

$$M_{leaf} = (B_{leaf} + B_{sugar,leaf}) \cdot MR_{leaf} \cdot Q_{10}^{(T_{mean}-20)/10} \cdot (L^{PAR})^{WUE_{decay}} \quad (16.19)$$

$$M_{sapwood} = (B_{living,sapwood} + B_{sugar,sapwood}) \cdot MR_{sapwood} \cdot Q_{10}^{(T_{mean}-20)/10} \quad (16.20)$$

$$M_{fineroot} = B_{fineroot} \cdot MR_{fineroot} \cdot Q_{10}^{(T_{mean}-20)/10} \cdot (LA_i/LA_i^{live}) \quad (16.21)$$

where T_{mean} is the average daily temperature (in °C). Baseline maintenance respiration rates per dry biomass (MR_{leaf} , $MR_{sapwood}$ and $MR_{fineroot}$ for leaves, parenchymatic sapwood and fine roots, respectively; in $g\text{ gluc} \cdot g\text{ dry}^{-1} \cdot day^{-1}$) are can be supplied via species-specific parameters and should refer to 20 °C. Leaf and fine root rates can also be estimated from nitrogen concentration, following Reich *et al.* (2008) (see section A.3.17). Factor Q_{10} has been found to decrease with temperature (Tjoelker *et al.* 2001). For example, a 10 °C increase in measurement temperature at low measurement temperatures from 5 to 15 °C results in an approximate 2.8 fold increase in respiration rate, whereas an increase in temperature from 25 to 35 °C results in a less than two-fold (1.8) increase in rate. The general temperature relationship proposed by Tjoelker *et al.* (2001) is used here:

$$Q_{10} = 3.22 - 0.046 \cdot T_{mean} \quad (16.22)$$

Factor $(L^{PAR})^{WUE_{decay}}$ is added to reduce respiration rates in leaves under shade, analogously to the reduction of water use efficiency explained in 6.1.6. Thus, it is assumed that the carbon use efficiency of the leaves is more or less constant. Factor LA_i/LA_i^{live} is added to the maintenance respiration of fine roots to reduce respiration rates during winter in winter-deciduous species, assuming a decrease in respiration rates of fine roots parallel to that of leaves.

When simulating growth with the advanced model, sub-daily leaf maintenance respiration rates are estimated as the difference between gross and net photosynthesis (see 11.1.4). Sapwood and fine root respiration rates are also estimated at sub-daily steps and, hence, sub-daily canopy temperature variation is taken into account (i.e. sub-daily canopy T_{can} values are used instead of the daily temperature mean, T_{mean}).

16.2.3 Growth respiration

Construction costs per unit dry weight of new tissue (CC_{leaf} , $CC_{sapwood}$ and $CC_{fineroot}$ for leaves, sapwood and fine roots; in $g\text{ gluc} \cdot g\text{ dry}^{-1}$) are specified as control parameters (i.e. they are not species-specific). These unitary costs include both the carbon used in respiration during growth and structural carbon. Given an increase in leaf area ΔLA , leaf growth respiration is:

$$G_{leaf} = \Delta LA \cdot CC_{leaf} \cdot 1000/SLA \quad (16.23)$$

Analogously, given an increase in sapwood area ΔSA , sapwood growth respiration is:

$$G_{sapwood} = \Delta SA \cdot CC_{sapwood} \cdot (H + \sum_s FRP_s \cdot L_s) \cdot \rho_{wood} \quad (16.24)$$

Finally, and given an increase in fine root biomass $\Delta B_{fineroot}$:

$$G_{fineroot} = \Delta B_{fineroot} \cdot CC_{fineroot} \quad (16.25)$$

16.2.4 Phloem transport

When growth is simulated using the basic water balance model, phloem transport between leaf and sapwood (F_{phloem}) is simply modelled as the flow needed to make the concentration of metabolic carbon equal in both compartments. When the advanced water balance submodel is used, F_{phloem} is modelled following Hölttä *et al.* (2017). Specifically, the instantaneous phloem flow per leaf area basis ($\text{mol gluc} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$) depends on k_{phloem} , i.e. the phloem conductance per leaf area basis ($\text{L} \cdot \text{m}^{-2} \cdot \text{MPa}^{-1} \cdot \text{s}^{-1}$), sap viscosity relative to water and the difference in turgor between the sieve cells of leaf and sapwood compartments. k_{phloem} is estimated as linear function of stem maximum conductance, $k_{stem,max}$:

$$k_{phloem} = f_{phloem} \cdot k_{stem,max} \quad (16.26)$$

where factor f_{phloem} is specified by the control parameter `phloemConductanceFactor`. Sap viscosity is calculated following Först *et al.* (2002), which takes into account temperature and the average sugar concentration between the two compartments (no phloem flow can occur if temperature is below zero). Turgor in sieve cells of each compartment depends on symplastic water potential and osmotic water potential, with Van't Hoff's equation being used to calculate the osmotic water potential, based on temperature, sugar concentration and the concentration of other solutes (control parameter `nonSugarConcentration`). Note that here leaf or sapwood sugar concentration is modulated to account for the effect of the relative water content of the compartment on osmotic water potential.

16.2.5 Sugar-starch dynamics

When growth is simulated using the *basic* water balance submodel, sugar-starch dynamics (SC_{leaf} and $SC_{sapwood}$) are simply defined as the conversion between metabolic and storage carbon needed to keep the sugar concentration equal to an equilibrium required for metabolic functioning, which is specified by control parameters `equilibriumOsmoticConcentration` (including two values, for leaves and sapwood respectively) and `nonSugarConcentration`. For example, if leaf osmotic equilibrium concentration is $0.8 \text{ mol gluc} \cdot \text{L}^{-1}$ and non-sugar concentration is $0.3 \text{ mol gluc} \cdot \text{L}^{-1}$, the model will simulate sugar-starch dynamics to stabilize sugar concentration in leaves to $0.5 \text{ mol gluc} \cdot \text{L}^{-1}$. A lower equilibrium osmotic concentration is required for sapwood, by default $0.6 \text{ mol gluc} \cdot \text{L}^{-1}$, so that sugar equilibrium concentration is $0.1 \text{ mol gluc} \cdot \text{L}^{-1}$. That lower equilibrium concentrations are required for sapwood than for leaves is necessary to sustain the phloem transport of sugars from leaves towards sapwood.

When growth is simulated using the *advanced* water balance submodel, sugar-starch dynamics are similar to the previous case, in the sense that equilibrium sugar concentrations are sought for leaves and sapwood. However, instantaneous conversion rates are calculated depending on equations regulating starch synthesis (occurring when sugar concentration is higher than the equilibrium concentration) and starch hydrolysis (occurring when sugar concentration is lower than the equilibrium concentration). Synthesis is modelled as a Michaelis-Menten function, whereas hydrolysis rate is simply a linear function of starch concentration. Maximum synthesis rates and hydrolysis linear factors are larger (i.e. faster sugar-starch dynamics) in leaves than sapwood.

16.2.6 Root exudation

Whereas surplus in leaf storage capacity is relocated into sapwood storage, surplus in sapwood storage carbon is diverted towards root exudation ($RE_{sapwood}$). Hence, root exudation is not a process competing for metabolic carbon, but only a consequence of plant C storage capacity being surpassed (Prescott *et al.* 2020). Root exudation may happen in the model when growth is restricted due to sink limitations but photosynthesis continues and storage carbon levels have attained the storage capacity in the sapwood compartment ($ST_{sapwood} > ST_{sapwood}^{max}$).

Chapter 17

Growth, senescence and mortality

17.1 Growth

17.1.1 Temperature and turgor sink limitations

Sink limitations due to temperature and turgor effects on growth are modelled following Cabon *et al.* (2020a) and Cabon *et al.* (2020b). These authors suggested equations to model sink limitations on cambium cell division and tracheid expansion, but we apply the same approach for simulating growth of leaves, sapwood and fine roots. Cell relative expansion rate (r_{cell}) is central to the approach by Cabon et al. and is defined as the relative time derivative of cell volume:

$$r_{cell} = \frac{dV}{Vdt} \quad (17.1)$$

Carbon *et al.* (2020a) first suggested to model the dependence of r_{cell} on cell turgor using Lockhart's equation:

$$r_{cell}(\Psi, \pi_0) = \phi_{max} \cdot (\Psi - \pi_0 - Y_P) \quad (17.2)$$

where Ψ is the water potential, π_0 is the osmotic water potential at full turgor, Y_P is the turgor yield threshold and ϕ_{max} is the maximum cell wall extensibility. Later, Cabon *et al.* (2020b) suggested to account for both turgor and temperature limitations on r_{cell} using the following expanded equation:

$$r_{cell}(T, \Psi, \pi_0) = \phi_{max} \cdot (\Psi - \pi_0 - Y_P) \cdot \frac{f_{met}(T_K)}{f_{met}(288.15)} \cdot f_{micro}(T, T_{thr}) \quad (17.3)$$

where T is temperature, $f_{met}(T_K)$ is a function modulating the effect of temperature T_K in Kelvin, on metabolic rate, and $f_{micro}(T, T_{thr})$ is a sigmoidal function modulating the effect of temperature on micro-tubule stability, depending on a temperature threshold T_{thr} . Function $f_{met}(T_K)$ is defined as:

$$f_{met}(T_K) = \frac{T_K \cdot \exp \left\{ \frac{\Delta H_A}{R^n \cdot T_K} \right\}}{1 + \exp \left\{ \frac{\Delta S_D}{R^n} \cdot \left(1 - \frac{\Delta H_D}{\Delta S_D \cdot T_K} \right) \right\}} \quad (17.4)$$

where R^n is the ideal gas constant, $\Delta H_A = 87500$ the enthalpy of activation and $\Delta H_D = 333000$ and $\Delta S_D = 1090$ the enthalpy and entropy difference (respectively) between the catalytically active and inactive states of the enzymatic system).

Scaling from the cell to the tissue level is conducted by assuming that maximum cell-level expansion rates correspond to maximum tissue-level relative growth rates.

17.1.2 Leaf growth

Leaf area increment ΔLA only occurs when phenological state is *unfolding*, and is defined as the minimum of three values expressing three corresponding constraints:

$$\Delta LA = \min(\Delta LA_{alloc}, \Delta LA_{source}, \Delta LA_{sink}) \quad (17.5)$$

First, ΔLA_{alloc} is the maximum leaf area increment allowed by the leaf area target set by the allocation rule, LA^{target} , in comparison with current leaf area LA^ϕ :

$$\Delta LA_{alloc} = \max(LA^{target} - LA^\phi, 0) \quad (17.6)$$

Remember that leaf area target is updated during *bud formation* (see 17.3.2).

Second, ΔLA_{source} represents the leaf area increment expected given carbon source limitations and is calculated:

$$\Delta LA_{source} = \frac{ST_{sapwood} \cdot m_{gluc} \cdot V_{sapwood,leaf}}{1000 \cdot CC_{leaf}/SLA} \quad (17.7)$$

where $ST_{sapwood}$ is the current concentration of sapwood storage carbon (starch), $V_{storage,sapwood}$ is the sapwood storage volume and m_{gluc} is glucose molar mass and the denominator contains the construction costs per leaf area unit, see eq. (16.23).

Finally, ΔLA_{sink} represents the leaf area increment expected by taking into account the maximum leaf tissue growth rate relative to sapwood area ($RGR_{leaf,max}$; in $m^2 \cdot cm^{-2} \cdot day^{-1}$), the percentage of the crown with active buds ($PCAB$) and the relative cell expansion rate given T_{day} , $\Psi_{symp,leaf}$ and $\pi_{0,leaf}$:

$$\Delta LA_{sink} = SA \cdot (PCAB/100) \cdot RGR_{leaf,max} \cdot \frac{r_{cell}(T_{day}, \Psi_{symp,leaf}, \pi_{0,leaf})}{r_{cell,max}} \quad (17.8)$$

$$= SA \cdot (PCAB/100) \cdot RGR_{leaf,max} \cdot \frac{(\Psi_{symp,leaf} - \pi_{0,leaf} - Y_P)}{(2 - Y_P)} \cdot \frac{f_{met}(T_{day})}{f_{met}(30)} \quad (17.9)$$

where cell relative expansion rate is divided by the maximum relative cell expansion rate $r_{cell,max} = r_{cell}(30, 0, -2)$ so that $RGR_{leaf,max}$ is attained when sink conditions are optimal. Other parameters for eq. (17.3) are $\phi_{max} = 0.5$ (which becomes irrelevant), $Y_P = 0.05 MPa$ and $T_{thr} = 5^\circ C$. Note that the final expression of ΔLA_{sink} reduces to a product of the maximum leaf area growth times two factors (for turgor and temperature) bounded between 0 and 1.

17.1.3 Sapwood growth

Sapwood area increment ΔSA can only occur if $LA^{live} > 0$. Unlike leaf area or fine root biomass, it is not explicitly constrained by any allocation rule. However, newly assimilated carbon is preferentially allocated to leaves and fine roots whenever storage levels are low. ΔSA is defined as the minimum of two values expressing source and sink constraints:

$$\Delta SA = \min(\Delta SA_{source}, \Delta SA_{sink}) \quad (17.10)$$

ΔSA_{source} represents the sapwood area increment expected given carbon source limitations and is calculated using:

$$\Delta SA_{source} = \frac{\max(ST_{sapwood} - ST_{sapwood,growth}, 0) \cdot m_{gluc} \cdot V_{storage,sapwood}}{CC_{sapwood} \cdot (H + \sum_s FRP_s \cdot L_s) \cdot \rho_{wood}} \quad (17.11)$$

where $ST_{sapwood}$ is the current starch concentration, $ST_{sapwood,growth}$ is the minimum starch concentration required for sapwood growth, $V_{storage,sapwood}$ is the sapwood storage volume, m_{gluc} is glucose molar mass and the denominator contains the construction costs per sapwood area unit, see eq. (16.24). $ST_{sapwood,growth}$ is related to the minimum relative starch concentration for growth ($RSSG$), which is specified via the species-specific parameter `RSSG` or the control parameter `minimumRelativeStarchForGrowth`. This parameter is

important because it allows specifying to which extent a given plant species stops growth and saves carbon to ensure survival (e.g. shadow tolerant species), as opposed to a species strongly investing in growth to reach the top of the canopy and have access to high light levels (e.g. light-demanding species).

Sink limitations to sapwood growth are simulated using a set of $N_c = 10$ cambium cells that undergo division and the new cells are added to a ring (i.e. a vector) of vessels that later suffer enlargement and maturation (Friend *et al.* 2019). Assuming that cambium cell volume must double before division, that cell-doubling time is the variable limiting cell division and that each division instantly results in a cell exiting the cambium and initiating differentiation into a vessel, production rate ($PR_{cambium}$) of new vessels cells can be related to the cambial cell expansion rate $r_{cambium}$ such that (Cabon *et al.* 2020b):

$$PR_{cambium} = \frac{N_c}{\ln(2)} \cdot r_{cambium} \quad (17.12)$$

where $r_{cambium} = r_{cell}(T_{day}, \Psi_{symp,stem}, -0.8)$, with other parameters of eq. (17.3) being $\phi_{max} = 0.13$, $Y_P = 0.05 \text{ MPa}$ and $Y_{thr} = 8 \text{ }^{\circ}\text{C}$. The ring growth model is implemented at daily time steps and daily $PR_{cambium}$ values are accumulated (i.e. $P_{cambium,t} = P_{cambium,t-1} + PR_{cambium,t}$). A new vessel is formed and added to the ring every time $P_{cambium}$ advances one integer. All vessel cells in the ring undergo daily enlargement following eq. (17.3), again with $Y_P = 0.05 \text{ MPa}$ and $Y_{thr} = 8 \text{ }^{\circ}\text{C}$. During cell later stages of enlargement, the cell wall becomes thicker and more rigid as a consequence of cell wall deposition. Consequently, $\phi_{max} = 0.13$ for newly formed cells, but then variations in ϕ_{max} are modelled as (Cabon *et al.* 2020a):

$$\frac{d\phi_{max}}{\phi_{max} \cdot dt} = s \cdot r_{cell} - h \cdot \frac{f_{met}(T_K)}{f_{met}(288.15)} \quad (17.13)$$

where $s = 1.8$ and $h = 0.043 \cdot 1.8$ are a softening factor and a constant hardening rate, respectively, and the hardening rate is modulated by temperature effects. Cell radial diameter is initially $8.3 \mu\text{m}$ and tangential diameter is assumed constant and equal to $20 \mu\text{m}$, so that volume increases in ring cells can be translated to daily ring area increases, ΔA_{ring} (in μm^2).

Assuming that the maximum relative sapwood growth rate ($RGR_{cambium,max}$ or $RGR_{sapwood,max}$) corresponds to a maximum rate of daily ring area increase, we have that the daily increase in sapwood area according to sink limitations, ΔSA_{sink} , is defined as:

$$\Delta SA_{sink} = \pi \cdot DBH \cdot RGR_{cambium,max} \cdot \frac{\Delta A_{ring}}{\Delta A_{ring,max}} \quad (17.14)$$

for trees, and as:

$$\Delta SA_{sink} = SA \cdot RGR_{sapwood,max} \cdot \frac{\Delta A_{ring}}{\Delta A_{ring,max}} \quad (17.15)$$

for shrubs. Sapwood growth in trees is proportional to the cambium length (in cm), whereas in trees it is proportional to the current sapwood area (in cm^2). Hence, two different maximum relative growth rates are defined for trees ($RGR_{cambium,max}$) and shrubs ($RGR_{sapwood,max}$). In the previous equations, $\Delta A_{ring,max} = N_c \cdot 950 \mu\text{m}^2 \cdot \text{day}^{-1}$, which was obtained by simulating a ring growth under optimum temperature ($T_{day} = 30 \text{ }^{\circ}\text{C}$) and water ($\Psi_{symp,stem} = -0.033 \text{ MPa}$) conditions and observing the steady-state ring area increase.

17.1.4 Fine root growth

Fine root biomass increment is modelled for each soil layer separately, and is defined analogously to leaf area increment:

$$\Delta B_{fineroot} = \min(\Delta B_{fineroot,alloc}, \Delta B_{fineroot,source}, \Delta B_{fineroot,sink}) \quad (17.16)$$

First, $\Delta B_{fineroot,alloc}$ is the maximum fine root biomass increment allowed by the biomass target set by the allocation rule, $B_{fineroot,target}$ (see 17.3.2), in comparison with current biomass, $B_{fineroot}$:

$$\Delta B_{fineroot,alloc} = \max(B_{fineroot,target} - B_{fineroot}, 0) \quad (17.17)$$

Second, $\Delta B_{fineroot,source}$ represents the biomass increment expected given the available storage carbon (starch):

$$\Delta B_{fineroot,source} = \frac{ST_{sapwood} \cdot m_{gluc} \cdot V_{storage,sapwood}}{CC_{fineroot}} \quad (17.18)$$

where $ST_{sapwood}$ is the current sapwood concentration of storage carbon, $V_{storage,sapwood}$ is the sapwood storage volume, m_{gluc} is glucose molar mass and $CC_{fineroot}$ is the construction costs per fine root biomass unit.

Finally, $\Delta B_{fineroot,sink}$ represents the biomass increment expected by taking into account maximum tissue growth rate ($RGR_{fineroot,max}$; in $g\text{ dry}\cdot g\text{ dry}^{-1}\cdot day^{-1}$) and the relative cell expansion rate given temperature (T_{day}), water potential in the rhizosphere ($\Psi_{rhizo,s}$) and the stem osmotic potential at full turgor ($\pi_{0,stem}$):

$$\Delta B_{fineroot,sink} = B_{fineroot} \cdot RGR_{fineroot,max} \cdot \frac{r_{cell}(T_{day}, \Psi_{rhizo,s}, \pi_{0,stem})}{r_{cell,max}} \quad (17.19)$$

$$= B_{fineroot} \cdot RGR_{fineroot,max} \cdot \frac{(\Psi_{rhizo,s} - \pi_{0,stem} - Y_P)}{(2 - Y_P)} \cdot \frac{f_{met}(T_{day})}{f_{met}(30)} \quad (17.20)$$

cell relative expansion rate is divided by the maximum relative cell expansion rate $r_{cell,max} = r_{cell}(30, 0, -2)$ so that $RGR_{fineroot,max}$ is attained when sink conditions are optimal. Other parameters for eq. (17.3) are like for leaf growth, $\phi_{max} = 0.5$, $Y_P = 0.05\text{ MPa}$ and $T_{thr} = 5^\circ\text{C}$.

17.2 Senescence

17.2.1 Leaf senescence and cavitation effects on leaves and buds

Leaf senescence can occur due to two processes: aging or dessication. Senescence due to advanced leaf age is assumed to be programmed (Ca+ accumulation?). In deciduous species all live leaf area turns to death leaf area when the phenology submodel indicates it (4.1.3). In evergreen species the proportion of leaf area that undergoes senescence each day is determined by the species-specific leaf duration parameter (LD):

$$p_{aging,leaf} = \frac{1}{365.25 \cdot LD} \quad (17.21)$$

Senescence due to drought effects occurs as a consequence of stem cavitation. If the proportion of conductance loss has increased with respect to the preceeding day, the model determines the leaf area corresponding to the current level of cavitation:

$$LA^{cavitation} = (1 - PLC_{stem}) \cdot LA^{target} \quad (17.22)$$

if $LA^{cavitation} < LA^\phi$ then the corresponding proportion $p_{cavitation,leaf}$ is estimated. The maximum of $p_{aging,leaf}$ and $p_{cavitation,leaf}$ is applied as a reduction of leaf area. If growth is simulated using the advanced water model, it also checks whether the relative water content in leaves becomes less than 50% (i.e. $RWC_{leaf} < 0.5$) and, if this happens, the plant cohort is completely defoliated.

Xylem cavitation not only causes defoliation but also mortality of buds, which has the effect of slowing down post-drought leaf area recovery. The percentage of the crown with active buds ($PCAB$) is reduced whenever the proportion of conductance loss has increased with respect to the preceeding day.

$$PCAB = \min(PLC_{stem}, 100 \cdot (1 - PLC_{stem})) \quad (17.23)$$

17.2.2 Sapwood senescence

The daily rate of sapwood senescence is specified via the species-specific parameter $SR_{sapwood}$ or, when missing, via a control parameter. Prentice *et al.* (1993) assumed a constant annual rate of 4% for the

conversion from sapwood to heartwood. Similarly, Sitch *et al.* (2003) assumed a sapwood annual turnover rate of 5% for all biomes. In our model, the proportion of sapwood area that is transformed into heartwood daily is estimated using:

$$p_{aging,sapwood} = \frac{SR_{sapwood}}{1 + 15 \cdot e^{-0.01 \cdot H}} \cdot \frac{\max(T_{day} - 5, 0)}{20} \quad (17.24)$$

where $SR_{sapwood}$ is a species-specific parameter, T_{day} is the average day temperature and 0.01 is a constant causing shorter plants to have slower senescence rates. It is important to mention that, while stem cavitation PLC_{stem} reduces the amount of functional sapwood in with respect to hydraulics (and therefore transpiration and photosynthesis), it does not increase the rate of sapwood senescence. Hence, when xylem embolism occurs air bubbles are formed within vessels but surrounding parenchymatic cells (as well as the storage carbon they contain) are assumed unaffected.

17.2.3 Fine root senescence

Aging is the only process leading to fine root senescence. The daily turnover proportion for fine roots ($SR_{fineroot}$) is assumed to correspond to a temperature of 25 °C, is specified via species-specific parameters or the control parameters `turnoverRates`. Actual turnover proportion for a given soil layer ($p_{aging,fineroot}$) decreases linearly with soil temperature down to zero at 5 °C:

$$p_{aging,fineroot} = SR_{fineroot} \cdot \frac{\max(T_{soil,s} - 5, 0)}{20} \quad (17.25)$$

where $T_{soil,s}$ is the temperature of soil layer s . When $SR_{fineroot}$ is missing from species parameter, a default control value for $SR_{fineroot}$ is taken, which produces an annual 50% turnover of fine roots.

17.3 Update of plant traits and allocation targets

17.3.1 Plant traits

Multiple anatomic and physiological parameters are updated every day after applying changes in the size of leaf, sapwood and fine root compartments, which creates a feedback to those hydraulic and physiological processes simulated in the water balance submodel (15.1.4). Hence, the growth model allows emulating plant acclimation to environmental cues, mediated by growth and senescence of plant tissues.

First leaf area index (LAI^ϕ) is updated from leaf area (LA^ϕ) inverting eq. (16.2). The Huber value (H_v , the sapwood area to leaf area ratio; in $m^2 \cdot m^{-2}$) is affected by changes in $LA^{live} = LA_{target}$ and sapwood area SA :

$$H_v = \frac{SA/10000}{LA^{live}} \quad (17.26)$$

Note that we specify $LA^{live} = LA_{target}$ and not LA^ϕ to avoid variations in Huber value due to leaf phenology. By definition, fine root biomass changes in each soil layer ($B_{fineroot,s}$) lead to updates in the proportion of fine roots in each layer (FRP_s):

$$FRP_s = \frac{B_{fineroot,s}}{\sum_l B_{fineroot,l}} \quad (17.27)$$

When we simulate growth with the advanced water balance model other parameters are subject to acclimation. Stem maximum conductance per leaf area unit ($k_{stem,max}$; in $mmol \cdot m^{-2} \cdot s^{-1} \cdot MPa^{-1}$) is determined as a function of species-specific xylem conductivity ($K_{xylem,max}$; in $kg \cdot m^{-1} \cdot s^{-1} \cdot MPa^{-1}$), leaf area, sapwood area and tree height (Christoffersen *et al.* 2016):

$$k_{stem,max} = \frac{1000}{0.018} \cdot \frac{K_{xylem,max} \cdot H_v}{(H/100)} \cdot \chi_{taper} \quad (17.28)$$

where χ_{taper} is a factor to account for taper of xylem conduit with height (Savage *et al.* 2010; Christoffersen *et al.* 2016), 0.018 is the molar weight of water (in $kg \cdot mol^{-1}$). Both an increase in SA or a decrease in LA^{live} (i.e. an increase in H_v) increase $k_{stem,max}$ and, hence, alleviate drought effects (i.e. a lower decrease in water potential across the stem for the same flow). In contrast, an increase in plant height will decrease stem conductance and increase drought stress. Changes in stem maximum conductance has cascade effects on root maximum conductance. First, coarse root minimum resistance is defined as a fixed proportion of whole-plant minimum resistance, so an increase in stem maximum conductance will increase whole-plant conductance and coarse root conductance, $k_{root,max}$.

Rhizosphere maximum conductance per leaf area unit in a given soil layer s ($k_{rhizo,max,s}$; in $mmol \cdot m^{-2} \cdot s^{-1} \cdot MPa^{-1}$) depends on fine root biomass in this layer ($B_{fineroot,s}$) and on leaf area (i.e. LA^{live}). The equations regulating these relationships are modulated by several soil and species parameters, such as soil saturated hydraulic conductance, species-specific root length, root length density and density of fine roots.

The proportion of conductance loss due to cavitation (PLC_{stem}) is reduced whenever sapwood area growth occurs (provided control parameter `cavitationRefill = "growth"`):

$$PLC_{stem,t+1} = \min(PLC_{stem,t} - \frac{\Delta SA}{SA}, 0) \quad (17.29)$$

This allows a progressive increase in functional sapwood area. Similarly, if previous drought or fire impacts had reduced the percentage of the crown with active buds ($PCAB$), this percentage is increased along with the formation of new sapwood area (until it achieves 100%):

$$PCAB_{t+1} = \min(PCAB_t + 100 \cdot \frac{\Delta SA}{SA}, 100) \quad (17.30)$$

17.3.2 Leaf area and fine root biomass targets

Leaf area target ($LA^{live} = LA_{target}$) is updated when phenological phase is *bud formation*. If the allocation strategy pursues a **stable Huber value**, the model tries to bring H_v close to an initial value $H_{v,target}$, and the leaf area target is defined as:

$$LA^{live} = LA_{target} = \frac{SA}{10000 \cdot H_{v,target}} \quad (17.31)$$

Note that the specification of LA_{target} will cause sapwood area increases to be followed by leaf area increases, as long as $H_v > H_{v,target}$. On the contrary, if $H_v < H_{v,target}$ then leaf area growth is inhibited and sapwood area growth will progressively increase H_v . Fine root biomass target is set using:

$$B_{fineroot,target} = \frac{10^4 \cdot LA_{target} \cdot RLR}{2.0 \cdot \sqrt{\frac{\pi \cdot SRL}{\rho_{fineroot}}}} \quad (17.32)$$

where RLR is the root area to leaf area ratio, $\rho_{fineroot}$ is fine root tissue density ($g dry \cdot cm^{-3}$) and SRL ($cm \cdot g dry^{-1}$) is the specific root length.

If the allocation strategy pursues a **stable whole-plant conductance**, the model tries to keep k_{plant} close to an initial value $k_{plant,target}$, and here the leaf area target is defined as:

$$LA_{target} = LA^{live} \cdot \frac{k_{plant,max}}{k_{plant,target}} \quad (17.33)$$

In this strategy, increases in leaf area will be scheduled whenever the current whole-plant conductance is above target value (i.e. $k_{plant,max} > k_{plant,target}$). Analogously to the previous strategy, if $k_{plant,max} < k_{plant,target}$ then leaf area growth is inhibited and sapwood area growth will progressively increase $k_{plant,max}$.

While leaf area target depends on the allocation strategy, the target of fine root biomass for any given soil layer s ($B_{fineroot,target,s}$) directly follows changes in maximum whole-plant conductance. The average resistance in

the rhizosphere is assumed to correspond to a fixed percentage of total soil-plant resistance. Hence, changes in the conductance of leaves, stem or coarse roots will entail a variation in the absolute rhizosphere maximum conductance to be targeted ($k_{rhizo,max,target,s}$), which in turn will determine $B_{fineroot,target,s}$. For example $k_{rhizo,max,target,s}$ will increase as a consequence of sapwood area growth. The model thus first estimate $k_{rhizo,max,target,s}$ for each layer s and then translates $k_{rhizo,max,target,s}$ values to $B_{fineroot,target,s}$ using the relationships based on soil saturated hydraulic conductance, species-specific root length, root length density and density of fine roots mentioned above.

17.4 Plant mortality

17.4.1 Self-thinning of small trees

As explained in the mortality design section (15.1.5), the model implements a mortality process for young individual trees, meant to represent a self-thinning process occurring between trees of small diameter ($DBH_{tree,recr}$, typically 1 cm) and trees having a diameter corresponding to inclusion as individual in forest inventories ($DBH_{tree,ingrowth}$, typically 7.5 cm). The aim is to ensure that the tree density is progressively reduced until the tree cohort reaches $DBH_{tree,ingrowth}$, where the density should be $N_{tree,ingrowth}$. The following relationship between tree diameter and density is used:

$$N = a_{st} \cdot DBH^{b_{st}} \quad (17.34)$$

where a_{st} and b_{st} are parameters regulating the speed of the self-thinning process. Note that if we know the diameter and density of recruitment (i.e. $DBH_{tree,recr}$ and $N_{tree,recr}$), as well as $N_{tree,ingrowth}$, the density we want to ensure when the tree reaches $DBH_{tree,ingrowth}$, the self-thinning curve is completely determined. Hence, we can estimate b_{st} using:

$$b_{st} = \frac{\log(N_{tree,ingrowth}/N_{tree,recr})}{\log(DBH_{tree,ingrowth}/DBH_{tree,recr})} \quad (17.35)$$

whereas a_{st} can be estimated using:

$$a_{st} = \frac{N_{tree,ingrowth}}{DBH_{tree,ingrowth}^{b_{st}}} \quad (17.36)$$

Once we know the parameters of the self-thinning curve, we can determine the maximum cohort density allowed for any cohort of diameter DBH_i . If the tree cohort has density N_i , the density decrease due to self-thinning mortality can be estimated using:

$$N_{dead,i} = N_i - \min(N_i, a_{st} \cdot DBH_i^{b_{st}}) \quad (17.37)$$

Note that the self-thinning process does not distinguish between tree resprouts or trees recruited from seeds.

17.4.2 Basal mortality rates

Adult trees ($DBH > 7.5$ cm) and shrubs die at a basal rate due to unspecific causes. For shrubs, the basal rate is always a species-specific constant ($P_{mort,base}$). For trees, the basal rate can alternatively depend on the degree of (symmetric) competition, which is measured using the tree basal area BA ($m^2 \cdot ha^{-1}$) of the stand, through a simple survival logistic model:

$$P_{surv,base} = \text{logit}^{-1}(\beta_{surv,0} + \beta_{surv,1} \cdot \sqrt{BA}) = \frac{\exp(\beta_{surv,0} + \beta_{surv,1} \cdot \sqrt{BA})}{1 + \exp(\beta_{surv,0} + \beta_{surv,1} \cdot \sqrt{BA})} \quad (17.38)$$

where $\beta_{surv,0}$ and $\beta_{surv,1}$ are species-specific coefficients obtained by fitting Generalized Linear Models on repeated forest inventory plot data. These models can be developed for arbitrary time steps (normally 5 or 10 years), but the basal mortality rate has to be finally expressed as a daily probability ($P_{base,daily}$).

17.4.3 Mortality due to starvation

The starvation stress indicator is $ST_{relative}^{sapwood}$, the starch concentration in sapwood relative to the equilibrium value:

$$ST_{relative}^{sapwood} = \frac{ST_{sapwood}}{ST_{equilibrium}^{sapwood}} \quad (17.39)$$

where $ST_{sapwood}$ ($mol\ gluc\cdot l^{-1}$) is the current starch concentration in sapwood and $ST_{equilibrium}^{sapwood}$ is the equilibrium concentration (by default $ST_{equilibrium}^{sapwood} = 0.35\ mol\ gluc\cdot l^{-1}$). Assuming that a threshold of relative starch concentration ($ST_{relative}^{thresh}$) corresponds to 50% annual mortality due to starvation, a logistic sigmoidal function is used to estimate the probability of annual starvation from a given $ST_{relative}^{sapwood}$:

$$P_{starv,annual} = 1.0 - \frac{\exp(40 \cdot (ST_{relative}^{sapwood} - ST_{relative}^{thresh}))}{1.0 + \exp(40 \cdot (ST_{relative}^{sapwood} - ST_{relative}^{thresh}))} \quad (17.40)$$

Since the model operates at the daily temporal resolution, $P_{starv,annual}$ is re-expressed as a daily probability using:

$$P_{starv,daily} = 1.0 - \exp(\log(1.0 - P_{starv,annual})/356) \quad (17.41)$$

Assuming a relative threshold $ST_{relative}^{thresh} = 0.4$, the figure below illustrates the shape of the sigmoidal function (top) and the corresponding daily probability function.

17.4.4 Mortality due to dessication

As explained in the section 15.1.5, the dessication stress indicator (D_{stem}) is the maximum of relative water content in the stem (RWC_{stem}) and relative hydraulic conductance in the stem, i.e. the complement of PLC_{stem}

$$D_{stem} = (RWC_{stem}, 1.0 - PLC_{stem})/2.0 \quad (17.42)$$

Assuming that a threshold of relative water content (RWC_{thresh}) corresponds to 50% annual mortality due to dessication, its probability is estimated analogously to starvation. First, a logistic sigmoidal function is used to estimate the probability of annual dessication from a given D_{stem} value:

$$P_{dessic,annual} = 1.0 - \frac{\exp(40 \cdot (D_{stem} - RWC_{thresh}))}{1.0 + \exp(40 \cdot (D_{stem} - RWC_{thresh}))} \quad (17.43)$$

Since the model operates at the daily temporal resolution, $P_{dessic,annual}$ is re-expressed as a daily probability using:

$$P_{dessic,daily} = 1.0 - \exp(\log(1.0 - P_{dessic,annual})/356) \quad (17.44)$$

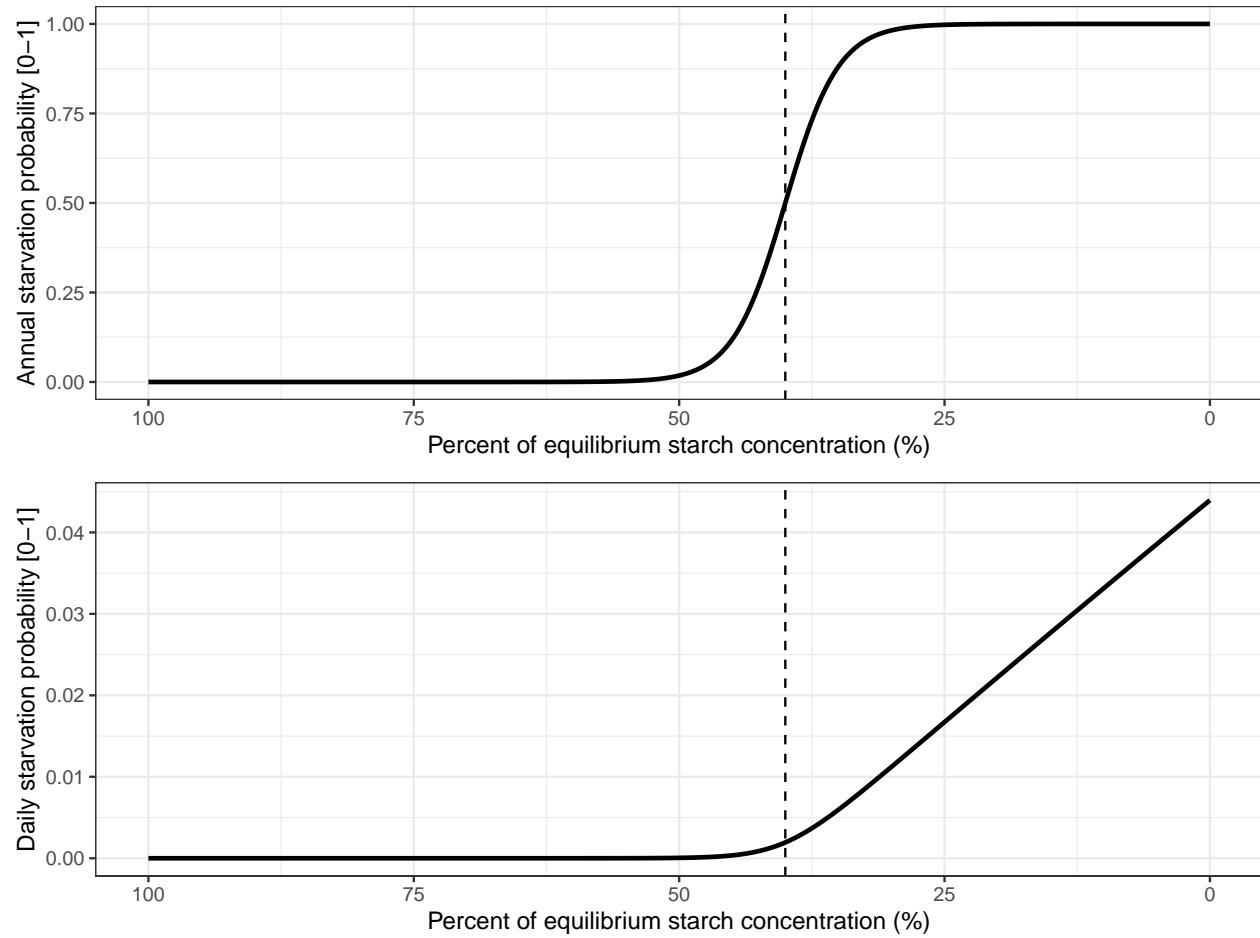
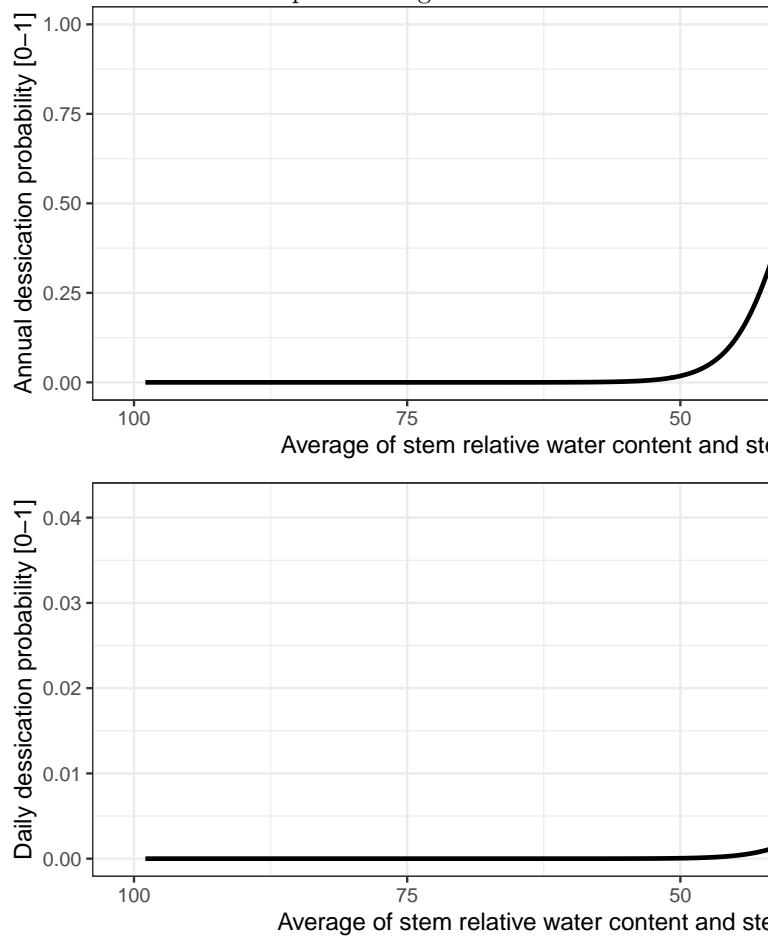


Figure 17.1: Annual (top) and daily (bottom) probability of starvation as a function of sapwood starch concentration relative to its equilibrium value.

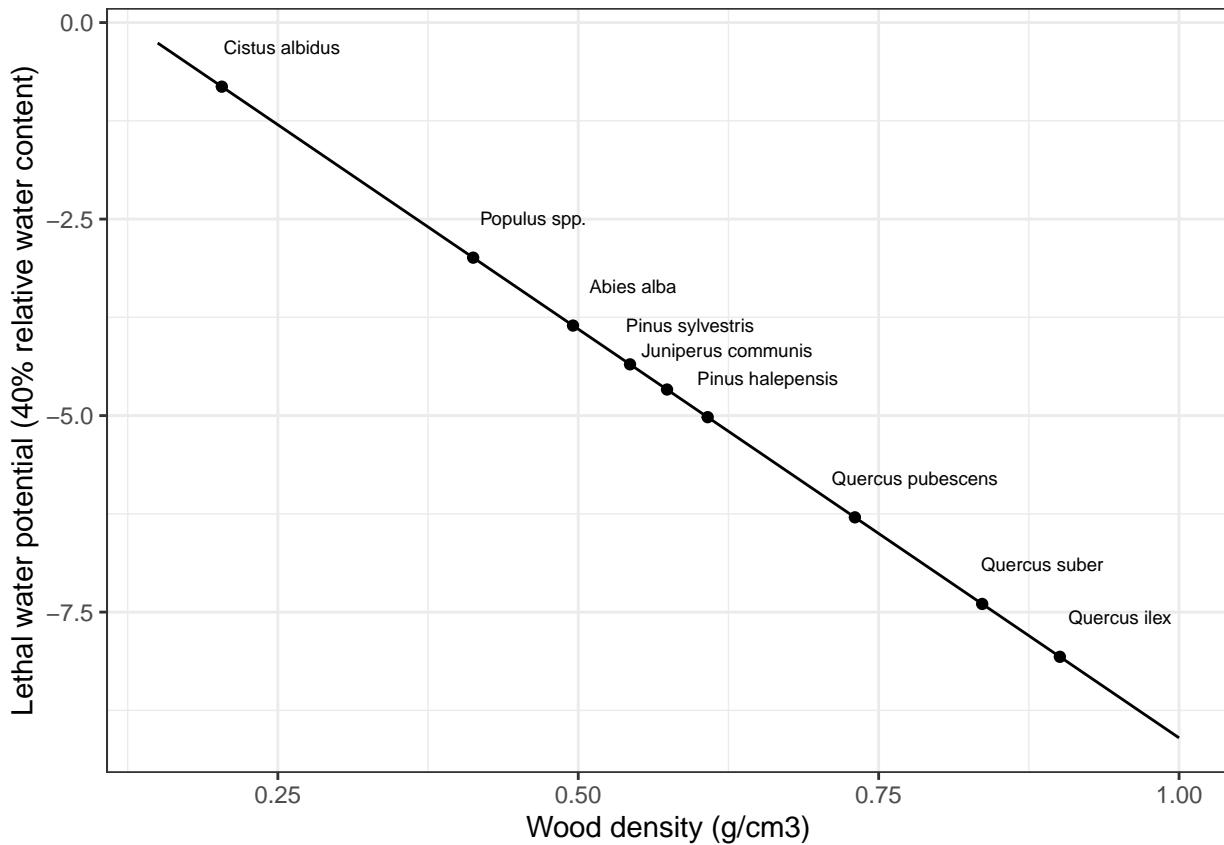
Assuming a relative threshold $RWC_{thresh} = 0.4$, the figure below illustrates the shape of the sigmoidal func-



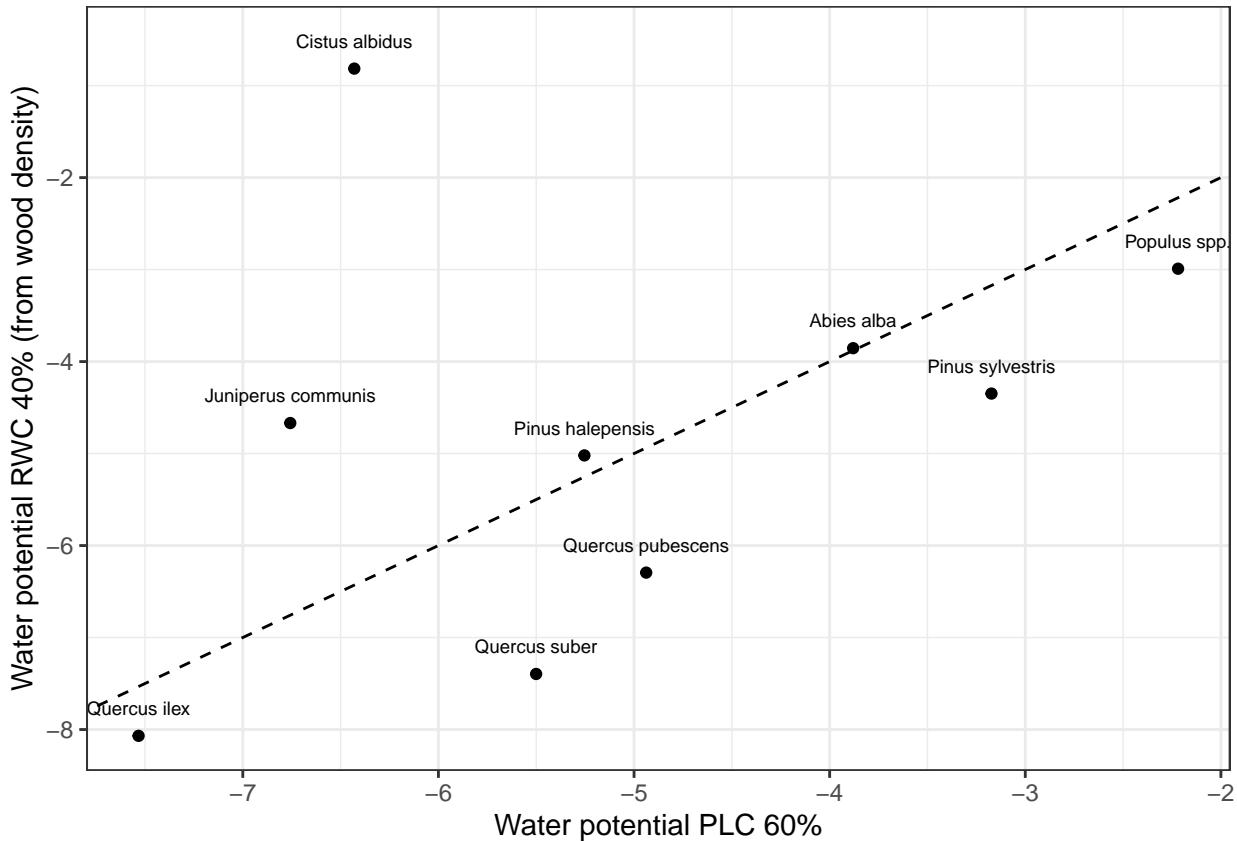
tion (top) and the corresponding daily probability function.

Uncertainty of stem pressure-volume parameters

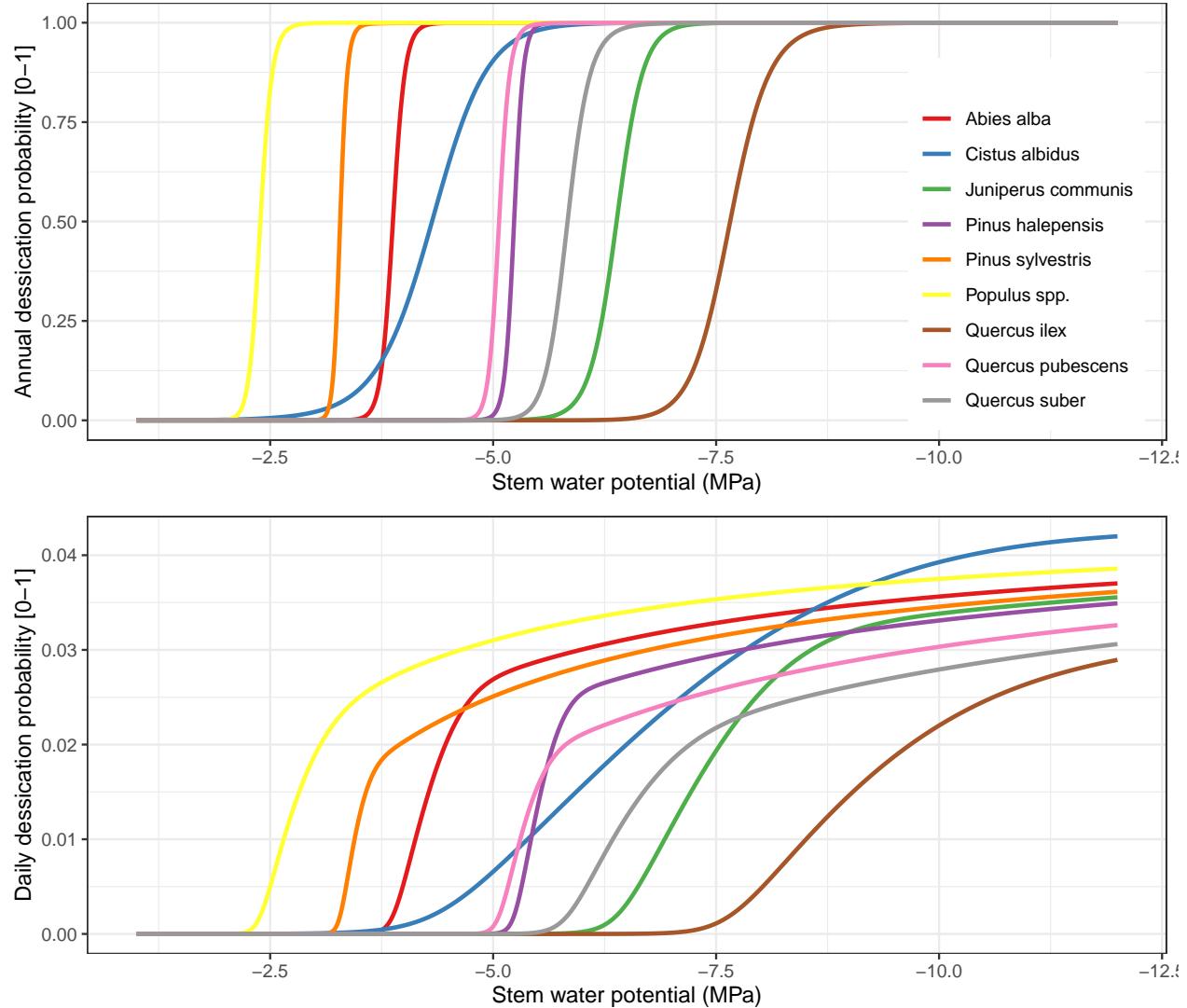
As explained in 15.1.5, using stem relative water content as indicator of dessication has the inconvenient that the parameters $\pi_{0,stem}$ and ϵ_{stem} specifying the pressure-volume curve are often not known, so that estimates have to come from wood density (see section A.3.12). The following figure shows the water potential corresponding to the dessication threshold for species with different wood density:



If the relationship between wood density and the stem pressure-volume curve (A.3.12) does not hold for a particular species the mortality rate may be under or overestimated. Here an example would be *Cistus albidus*, which has a low wood density but high desiccation resistance. This is illustrated when we draw a scatter plot between the water potential corresponding to 60% stem PLC (i.e. 40% stem relative hydraulic conductance) against the water potential corresponding to 40% stem RWC:



As explained in 15.1.5, one can avoid the potential uncertainty in desiccation mortality by defining the desiccation stress indicator (D_{stem}) as the maximum of relative water content in the stem (RWC_{stem}) and relative hydraulic conductance in the stem. When we do so, the following curves are obtained for each of the previous species:



It is obvious in the former figure that using D_{stem} as indicator results in curves that are not continuous in all their range. However, we prefer this inconvenient to the potential overestimation of mortality due to an uncertainty in the pressure-volume curve parameters.

17.4.5 Overall mortality probability

Every day the model determines for woody cohorts the overall probability of mortality ($P_{mort,daily}$) as the maximum of the basal probability, starvation probability and desiccation probability:

$$P_{mort,daily} = \max(P_{base,daily}, P_{starv,daily}, P_{desicc,daily}) \quad (17.45)$$

At this point $P_{mort,daily}$ may be used deterministically (i.e., as a proportion of N to kill) or stochastically (i.e. throwing a random number between 0 and 1 to determine the mortality event).

17.5 Fire severity

Fire effects on plants follow, with some modifications, the model by Michaletz & Johnson (2008). Buoyant plume theory is first used to estimate the vertical plume temperature T_{plume} distribution that will drive heat transfer to the vascular cambium and vegetative organs. Heat transfer theory is then used to calculate the depth of vascular cambium necrosis and height of crown foliage and crown bud necrosis. Finally, these severity metrics are used to define the fate of the plant cohort, as described in section 15.1.6.

17.5.1 Plume temperature distribution

The surface fire is assumed to occur at the time of the day where temperature is T_{max} is the maximum daily temperature. For a line-source plume in a quiescent atmosphere (no wind), the plume temperature T_{plume} at height z (in m) can be estimated using (Michaletz & Johnson 2008):

$$T_{plume}(z) = \min \left[900, C_{plume} \cdot \left(\frac{1}{z} \right) \cdot \left(\frac{T_{max} + 273.15}{g} \right)^{1/3} \cdot \left(\frac{I_B}{c_{p,air} \cdot \rho_{air}} \right)^{2/3} + T_{max} \right] \quad (17.46)$$

where $C_{plume} = 2.6$ is the plume proportionality constant, $g = 9.8 \text{ m} \cdot \text{s}^{-2}$ is the gravity constant, I_B is Byram's surface fireline intensity (eq. (24.131); expressed in $\text{kW} \cdot \text{m}^{-1}$), $c_{p,air} = 1.007 \text{ J} \cdot \text{kg}^{-1} \cdot {}^\circ\text{C}^{-1}$ is the specific heat capacity of the air and ρ_{air} is the air density at temperature T_{max} . Because this similarity analysis fails at heights comparable with the fireline width, T_{plume} is constrained to a maximum flame temperature of 900 °C.

17.5.2 Foliage and crown bud necrosis

The model predicting the distribution of plume temperature and the residence time of surface fires t_R (eq. (24.132)), are used to determine the height of necrosis for plant organs, here foliage or crown buds, depending also on their heat capacitance. The estimate (foliage or bud) necrosis height z_n (in m) is given by (Michaletz & Johnson 2008):

$$z_n = C_{plume} \cdot \left(\frac{1}{T_{crit} - T_{max}} \right) \cdot \left(\frac{T_{max} + 273.15}{g} \right)^{1/3} \cdot \left(\frac{I_B}{c_{p,air} \cdot \rho_{air}} \right)^{2/3} \quad (17.47)$$

where T_{crit} is the critical temperature of the plume required for organ necrosis, given a residence time t_R , which is estimated using:

$$T_{crit} = \frac{T_n - \theta_T \cdot T_{max}}{1 - \theta_T} \quad (17.48)$$

Here, T_n is the temperature leading to necrosis, assumed to be 60°C and θ_T is defined as the excess temperature ratio, which depends on the organ thermal factor (TF) and t_R :

$$\theta_T = \exp(-TF \cdot t_R) \quad (17.49)$$

The thermal factor TF is normally larger for leaves than buds, given their higher surface area to volume ratio, resulting in lower values of T_{crit} and, therefore, higher values of necrosis height. In the case of leaves, the thermal factor (TF_{leaves}) is estimated from specific leaf area, SLA , using (Michaletz & Johnson 2006; Michaletz & Johnson 2008):

$$TF_{leaves} = SLA \cdot (h_{leaves}/c_{leaves}) \quad (17.50)$$

where $h_{leaves} = 130$ stands for the convection heat transfer coefficient of leaves and $c_{leaves} = 2500$ is the specific heat capacity of leaves. Typical values would be $TF_{leaves} = 0.208$ for $SLA = 4$ and $TF_{leaves} = 0.624$ for $SLA = 12$. Currently, for crown buds a constant $TF_{buds} = 0.130$ is assumed, on the basis of values given in Michaletz & Johnson (2006), but we acknowledge that TF_{buds} should be at least species-specific. Once

z_n is defined, for either leaves or crown buds, depending on their TF value, the vertical leaf distribution described in 2.4.5 is used to determine the proportion of crown leaves or crown buds with necrosis.

If the case of a crown fires, i.e. if surface fireline intensity is larger than van Wagner's critical intensity; eq. (24.160), then all crown foliage is burned. The same happens if torching occurs for the target plant cohort, which is determined by calculating a cohort-specific critical intensity. Crown bud necrosis is decided in those cases by calculating T_{crit} using the crown fire residence time and comparing it to the flame temperature, i.e. 900°C. Despite these calculations, in practice all crown buds will also normally suffer necrosis under a crown fire.

17.5.3 Cambium necrosis

Vascular cambium necrosis is a one-dimensional transient conduction problem depending the thermal diffusivity of the bark, α_{bark} , and the surface fire residence time t_R . Assuming that the bark surface temperature is equal to the plume temperature at $z = 0.1$, i.e. $T_{plume}(0.1)$, the radial bole necrosis depth, x_n (in m), can be estimated using:

$$x_n = 2 \cdot (\alpha_{bark} \cdot t_R)^{1/2} \cdot \text{erf}^{-1} \left(\frac{T_n - T_{plume}(0.1)}{T_{max} - T_{plume}(0.1)} \right) \quad (17.51)$$

where erf^{-1} is the inverse error function. Bark diffusivity (α_{bark}) is estimated following the equations given in Michaletz & Johnson (2008), which depend on bark's tissue density, moisture content (estimated assuming a fine dead fuel) and temperature. Cambium necrosis is determined by comparing x_n (in m) with x_{ba} (in mm) the bark thickness of the plant, which in case of trees depends on DBH:

$$x_{ba} = a_{bt} \cdot DBH^{b_{bt}} \quad (17.52)$$

and in the case of shrubs x_{ba} is assumed to be equal to a fixed species-specific value bt_{sh} .

17.6 Update of structural variables

17.6.1 Tree diameter, height and crown ratio

In the case of tree cohorts, the new sapwood area (ΔSA , in cm^2) is translated to an increment in DBH (ΔDBH , in cm) following:

$$\Delta DBH = 2 \cdot \sqrt{(DBH/2)^2 + (\Delta SA/\pi)} - DBH \quad (17.53)$$

Furthermore, the model assumes that increments in height are linearly related to increments in diameter through a function f_{HD} (Lindner et al. 1997):

$$\Delta H = f_{HD} \cdot \Delta DBH \quad (17.54)$$

Hence, f_{HD} represents the height increment (in cm) per each cm of diameter increment. It was customary in forest gap models to prevent height from being larger than a species-specific value H_{max} , so that beyond some point trees only grew in size by increasing their diameter. Moreover, light conditions influence growth in height with trees living under the shade of others generally showing larger increases in height than trees living in open conditions. Hence, our formulation for f_{HD} is (Lindner et al. 1997; Rasche et al. 2012):

$$f_{HD} = [f_{HD,min} \cdot L^{PAR} + f_{HD,max} \cdot (1 - L^{PAR})] \cdot \left(1 - \frac{H - 137}{H_{max} - 137} \right) \quad (17.55)$$

where $f_{HD,min}$ would be the height-diameter ratio for a tree of 137 cm height growing in full light and $f_{HD,max}$ would be the same ratio for a tree of the same height growing in the shadow, and L^{PAR} is the proportion

of photosynthetically active radiation available at mid-crown height (4.2). This formulation seems slightly easier to calibrate than that presented in Rasche *et al.* (2012). H_{\max} could be dependent on environmental conditions, but we skip this here, because environmental conditions already affect growth rate and carbon balance.

After updating tree diameter (DBH) and tree height (H), the model updates tree crown ratio (CR) by applying allometric relationships that take into account tree size and competition (see details in 21.2.3).

17.6.2 Shrub height and cover

Since shrub structural variables are height and cover, shrub growth is done in a way somewhat different from trees. Shrubs are often multi-stemmed (some trees also are), so that increases in sapwood area are not easily related to diameter growth. Since leaf biomass is related to sapwood area, one may model shrub growth assuming an allometric relationship between phytovolume of individual shrub crowns and photosynthetic biomass. This strategy entails that shrubs may grow or shrink in size depending on their C balance, in the same way that tree crowns would become denser or sparser depending on their C balance. Hence, shrubs can be understood as crowns in the floor.

Starting from live leaf area ($m^2 \text{ um}^{-2}$) we can calculate the foliar weight per shrub individual (in $kg \text{ um}^{-1}$):

$$W_{leaves} = \frac{LAI^{live}}{(N/10000) \cdot SLA} \quad (17.56)$$

An allometric relationship relating the biomass of leaves plus small branches and crown phytovolume (PV ; in $m^3 \text{ um}^{-1}$) can be drawn from fuel calculations:

$$W_{leaves+branches} = W_{leaves} \cdot r_{6.35} = a_{bsh} \cdot PV^{b_{bsh}} \quad (17.57)$$

where a_{bsh} and b_{bsh} are allometric relationships and $r_{6.35}$ is a species-specific ratio relating the dry weight of leaves plus small branches to the dry weight of leaves. Inverting this relationship we obtain an expression of shrub crown phytovolume:

$$PV = \left[\frac{W_{leaves} \cdot r_{6.35}}{a_{bsh}} \right]^{1/b_{bsh}} \quad (17.58)$$

Phytovolume is defined as the volume occupied by the shrub individual, i.e.:

$$PV = (A_{sh}/10000) \cdot (H/100) \quad (17.59)$$

where A_{sh} is the area of a single shrub individual (in cm^2). If we use the following quadratic relationship between A_{sh} and H :

$$A_{sh} = a_{ash} \cdot H^{b_{ash}} \quad (17.60)$$

we can calculate shrub height from phytovolume using:

$$H = \left[\frac{10^6 \cdot PV}{a_{ash}} \right]^{1/(1+b_{ash})} \quad (17.61)$$

Finally, the new value for shrub cover (in percent) can be obtained from H and N (in $\text{ind} \cdot \text{ha}^{-1}$):

$$Cover = 100 \cdot (N/10000) \cdot (A_{sh}/10000) = \frac{N \cdot a_{ash} \cdot H^2}{10^6} \quad (17.62)$$

Note that crown ratio for shrubs is assumed constant in the model. Like for trees, shrub height is limited to a maximum height H_{\max} . However, unlike trees, shrubs are not allowed to continue growing once this maximum size is attained.

Part V

Forest dynamics modelling

Chapter 18

Forest dynamics model

This chapter provides an overview of a forest dynamics model, which builds on the previous models and allows simulating the recruitment, growth, mortality and management of woody plant cohorts in a forest stand. The model was described in De Cáceres *et al.* (2023) and is run using function `fordyn()` for a set of years.

18.1 Design principles

The design of the forest dynamics model is to a large degree inherited from the water balance and growth models it builds on. Readers should refer to former sections to learn the design of the basic water balance (see 3.1), advanced water/energy balance (see 7.1) or growth/mortality processes (see 15.1). Regeneration, from either seed germination and establishment or via resprouting, and forest management are explicitly simulated at the level of `fordyn()` and hence will be the focus of this chapter.

18.1.1 Recruitment from seeds

Recruitment of saplings involves a number of processes (flowering and pollination, fruit/seed production, dispersal, storage, seed predation, germination, seedling establishment and survival until the sapling stage). All these processes have their own biotic and abiotic drivers (Price *et al.* 2001), so that modelling becomes a challenging task. Processes leading to sapling recruitment are frequently extremely simplified or their mechanisms ignored in many forest models (Price *et al.* 2001). The design of the forest dynamics model with respect to recruitment follows that of many gap models. Local seed production is considered in a binary way, where plants are considered fertile and able to produce viable seeds if they reach a given height (different for shrubs and trees). Alternatively, the user can specify a set of species whose seeds arrive to the target stand via simulation control parameters or dispersal processes (see chapter 20). A seed bank exists in the forest stand, where seed levels are relative (up to 100%) and result from the interplay between seed rain and seed mortality. Actual recruitment from seeds depends on a set of regeneration *thresholds* are used to determine whether recruitment of new saplings (i.e. ingrowth into a given diameter class for trees or size for shrubs) is possible. Typically, regeneration thresholds concern environmental conditions, although some models also consider ungulate browsing (Wehrli *et al.* 2007). In our case we focus on three environmental drivers limiting the transition from seedlings to saplings:

1. Tolerance to low temperatures, indicated by the mean temperature of the coldest month.
2. Drought tolerance, indicated by the annual moisture index (annual precipitation divided by annual evapotranspiration).
3. Shade tolerance, indicated by the percentage of photosynthetically active radiation reaching the ground.

A target species can recruit on a given year if current conditions for all three environmental indicators are above the species tolerance thresholds. A constant probability of recruitment determines actual recruitment within these bioclimatic limits. Recruitment densities and plant size of recruited individuals are specified via species parameters. Trees recruited from seed are subject to self-thinning processes before attaining the diameter of ingrowth (see 17.4.1).

18.1.2 Resprouting

Resprouting is a common feature of Mediterranean species. Most, if not all, Mediterranean broadleaved species are able to resprout from buds protected in branches, subterranean structures (e.g. burl, taproot, lateral roots) or in the root collar after a disturbance destroys the aerial part. Yet, some differences in the resprouting ability (i.e. survival) exist regarding the type of disturbance and the species affected.

In medfate, resprouting is assumed happen from buds at the stump/stool level after a given disturbance has led to aboveground plant die-back. Resprouting can occur as a response to different disturbances. For example, resprouting occurs after clipping (e.g. if management is applied), after a fire, or after cavitation has led to dessication of above-ground plant organs.

Regarding disturbance type, differences in survivorship through resprouting mostly depends on the intensity of the disturbance experienced, with an increasing occurrence of mortality after disturbances in the following order: *browsing, cutting, drought* and *fire* (Espelta *et al.* 1999). Fire is the disturbance that causes greatest mortality, probably because it may physically destroy part of the bud bank due to lethal temperatures or directly charring it (Pausas *et al.* 2016). Moreover, resprouting vigor also depends on the size of the plant (Moreira *et al.* 2012). According to different literature sources, the number of resprouts initially produced is approximately 2 per 1 cm² of stump area (Retana *et al.* 1992; García-Jiménez *et al.* 2017). Hence, in medfate the final density of resprouts depends on the disturbance that caused resprouting (via survivorship) and on the diameter of the *parent* plant, but in all cases the resprouts inherit the root system of their parent.

Like saplings originated from seeds, resprouts are subject to self-thinning processes before attaining the diameter of ingrowth (see 17.4.1).

18.1.3 Forest management

Forest management is an optional process in `fordyn()` simulations. Furthermore, management actions need to be defined in an external function to be supplied by the user. This design was chosen because there are multiple potential management strategies, and different traditions are followed in different countries. To work properly within `fordyn()` simulations, the supplied management function needs to accept a `forest` object as input, as well as a list of management parameters, and it has to return the reduction in tree density, the reduction in shrub cover and the management parameters to be applied in subsequent calls.

Although the former design allows users to tailor management functions to their simulation needs, the package includes `defaultManagementFunction()` to facilitate the simulation of these process in many situations, and which is briefly described here. The function implements two different management models:

- *Irregular management*: An uneven-aged stand is managed using thinning operations each time a threshold value of a chosen stand-level metric (such as basal area or Hart-Becking index) is trespassed. Thinning operations can focus on trees of specific diameter classes.
- *Regular management*: An even-aged (monospecific) stand is managed in cycles where thinning (preparatory) cuts are followed by final (regeneration) cuts. Thinning operations are similar as those of irregular management. One or several final cuts can be scheduled, the first starting whenever mean diameter surpasses a chosen threshold, and the following after a chosen number of years. Optionally, tree planting (of a chosen species) can be scheduled to occur after the last final cut.

The former models apply cuts on tree cohorts irrespective of their species. The default management function applies shrub clearing each time there is an operation on trees, resulting in removal of any shrub cover above a given maximum value.

18.2 State variables

The main state variables of the forest dynamic model are those conforming the structure and composition of the forest stand, i.e. the set of woody cohorts (either trees or shrubs) and their attributes (height, density, DBH, cover, etc.). Additionally, the seed species identity and relative abundance in the seed bank are also state variables of the model. Since the model performs calls to the `growth()` model, many other state variables are defined for intra-annual simulations (see 15.4). When including forest management action, additional state variables may be defined as management parameters.

18.3 Process scheduling

The `fordyn()` model divides the period to be simulated in years, which is the top-level time step of simulations. Given an input `forest` object, the function first initializes the input for function `growth()`. For each year to be simulated the model performs the following steps:

1. Calls function `growth()` to simulate daily water/carbon balance, growth and mortality processes (sub-daily processes may also be involved in `transpirationMode = "Sperry"` or `transpirationMode = "Cochard"`). See section 15.4 for details of growth scheduling.
2. If a management function is supplied, calls this function and apply the resulting reductions in tree density and shrub cover.
3. If required, simulates seed production, seed bank mortality, seed rain, recruitment from seeds and/or resprouting. In a spatial context, seed rain will include seeds dispersed from other forest stands 20.
4. Removes tree (or shrub) cohorts whose remaining density (resp. cover) is lower than a specified threshold.
5. Merges surviving cohorts with recruitment in the `forest` object and prepares the input of function `growth()` for the next annual time step.
6. Store current status of the `forest` object and update output tables/summaries.

18.4 Inputs and outputs

An important difference between `fordyn()` and the previous simulation functions is that it does not require a specific input object, as in `spwb()` or `growth()` functions. In other words, soil, vegetation, meteorology and control inputs are directly introduced as parameters to the function call to `fordyn()`.

18.4.1 Soil, vegetation and meteorology

Soil

Soil input requirements are the same as for the former models and were fully described in section 2.3.

Vegetation

Unlike the former models, vegetation input for `fordyn` are objects of the class `forest`, which were described in section 2.4.4.

Metereological input

The minimum weather variables required to run the model are min/max temperatures (T_{min} and T_{max}), min/max relative humidity (RH_{min} and RH_{max}), precipitation (P) and solar radiation (Rad). Wind speed (u) is also needed, but the user may use missing values if not available (a default value will be used in this case). Wind speed is assumed to have been measured at a specific height above the canopy (by default at 2 m). Atmospheric CO_2 concentration (C_{atm}) may also be specified, but if missing a default constant value is assumed, which is taken from the control parameters. Definitions and units of these variables were given in section 2.5.

18.4.2 Vegetation functional parameters

The forest dynamics model requires many functional parameters to be specified for plant cohorts. Some of them depend on whether the basic or advanced water balance is adopted, whereas others are inherited from the growth model. Here we report functional parameters needed in addition to those necessary for the growth model (see 15.5.2).

All of them concern the simulation of recruitment and are specified in the species parameter table (i.e. `SpParams`).

Symbol	Units	R	Description
H_{seed}	cm	SeedProdHeight	Minimum height for seed production
SM	mg	SeedMass	Seed dry mass
SL	yr	SeedLongevity	Average longevity
$Dispdist$	m	DispersalDistance	Parameter for dispersal kernel
$Dispshape$		DispersalShape	Parameter for dispersal kernel
P_{recr}	[0-1]	ProbRecr	Probability of recruitment from seeds within the bioclimatic limits imposed by temperature, moisture and light thresholds
TCM_{recr}	°C	MinTemp	Minimum average temperature (Celsius) of the coldest month for successful recruitment from seeds
MI_{recr}		MinMoisture	Minimum value of the moisture index (annual precipitation over annual PET) for successful recruitment from seeds
$FPAR_{recr}$	%	MinFPAR	Minimum percentage of PAR at the ground level for successful recruitment from seeds
$N_{tree,recr}$	$ind \cdot ha^{-1}$	RecrTreeDensity	Density of tree recruits from seeds.
$N_{tree,ingrowth}$	$ind \cdot ha^{-1}$	IngrowthTreeDensity	Density of trees reaching ingrowth DBH.
$DBH_{tree,recr}$	cm	RecrTreeDBH	DBH for tree recruits from seeds or resprouting (e.g. 1 cm).
$DBH_{tree,ingrowth}$	cm	IngrowthTreeDBH	DBH for trees (e.g. 7.5 cm).
$H_{tree,recr}$	cm	RecrTreeHeight	Height for tree recruits from seeds or resprouting
$Cover_{shrub,recr}$	%	RecrShrubCover	Vegetation cover for shrubs
$H_{shrub,recr}$	cm	RecrShrubHeight	Height for shrubs
$Z50_{recr}$	mm	RecrZ50	Soil depth corresponding to 50% of fine roots for recruitment
$Z95_{recr}$	mm	RecrZ95	Soil depth corresponding to 95% of fine roots for recruitment
$Resp_{fire}$		RespFire	Number of resprouts per stem after fire disturbance
$Resp_{dist}$		RespDist	Number of resprouts per stem after undefined disturbance (typically desiccation)
$Resp_{clip}$		RespClip	Number of resprouts per stem after clipping

18.4.3 Control parameters

Control parameters modulate the overall behavior of `fordyn` simulations, which extend the parameters used for growth simulations (see section 15.5.3). First, there are parameters that regulate the application of seed production, seed bank dynamics, recruitment from seeds, resprouting and the removal of cohorts with few individuals:

- `applySeedBankDynamics [= TRUE]`: Boolean flag to indicate that seed bank dynamics (seed production, seed bank mortality and seed rain) need to be simulated.
- `applyRecruitment [= TRUE]`: Boolean flag to indicate that recruitment from seeds is allowed.
- `applyResprouting [= TRUE]`: Boolean flag to indicate that resprouting is allowed.
- `recruitmentMode [= "stochastic"]`: String describing how recruitment from seeds is applied. Current accepted values are “deterministic” or “stochastic”.
- `removeEmptyCohorts [= FALSE]`: Boolean flag to indicate the removal of cohorts whose density is too low.
- `minimumTreeCohortDensity [= 1]`: Threshold of tree density resulting in cohort removal.
- `minimumShrubCohortCover [= 0.01]`: Threshold of shrub cover resulting in cohort removal.
- `dynamicallyMergeCohorts [= TRUE]`: Boolean flag to indicate that cohorts should be merged when possible. This option speeds up calculations but results in a loss of cohort identity and reinitialization of many state variables.

Next, a few parameters control the production/arrival of seeds:

- `seedRain [= NULL]`: Vector of species names whose seed rain is to be added to seed bank, regardless of local seed production.
- `seedProductionTreeHeight [= 300]`: Default minimum tree height for producing seeds (when species parameter `SeedProductionHeight` is missing).
- `seedProductionShrubHeight [= 30]`: Default minimum shrub height for producing seeds (when species parameter `SeedProductionHeight` is missing).

Then we have default parameters determining whether recruitment occurs:

- `probRecr [= 0.05]`: Default annual probability of recruitment (when species parameter `ProbRecr` is missing).
- `minTempRecr [= 0]`: Default threshold of minimum average temperature of the coldest month necessary for recruiting (when species parameter `MinTempRecr` is missing).
- `minMoistureRecr [= 0.3]`: Default threshold of minimum moisture index (annual precipitation over annual ETP) necessary for recruiting (when species parameter `MinMoistureRecr` is missing).
- `minFPARRecr [= 10]`: Default threshold of minimum fraction of PAR (in %) reaching the ground necessary for recruiting (when species parameter `MinFPARRecr` is missing).

Finally, there are a set of parameters specifying default values for recruited cohort attributes:

- `recrTreeDBH [= 1]`: Default DBH (cm) for recruited trees (when species parameter `RecrTreeDBH` is missing).
- `recrTreeDensity [= 100]`: Default density (ind · ha⁻¹) for recruited trees (when species parameter `RecrTreeDensity` is missing).
- `recrTreeHeight [= 100]`: Default height (cm) for recruited trees (when species parameter `RecrTreeHeight` is missing).
- `recrShrubCover [= 1]`: Default cover (%) for recruited shrubs (when species parameter `RecrShrubCover` is missing).
- `recrShrubHeight [= 100]`: Default height (cm) for recruited shrubs (when species parameter `RecrShrubHeight` is missing).
- `recrTreeZ50 [= 100]`: Default value for Z50 (mm) in recruited trees (when species parameter `RecrZ50` is missing).
- `recrShrubZ50 [= 50]`: Default value for Z50 (mm) in recruited shrubs (when species parameter `RecrZ50` is missing).
- `recrTreeZ95 [= 1000]`: Default value for Z95 (mm) in recruited trees (when species parameter `RecrZ50` is missing).
- `recrShrubZ50 [= 500]`: Default value for Z95 (mm) in recruited shrubs (when species parameter `RecrZ50` is missing).

18.4.4 Model output

Element	Description
<code>StandSummary</code>	A data frame with stand-level summaries (leaf area index, tree basal area, tree density, shrub cover, etc.) at the beginning of the simulation and after each simulated year.
<code>SpeciesSummary</code>	A data frame with species-level summaries (leaf area index, tree basal area, tree density, shrub cover, etc.) at the beginning of the simulation and after each simulated year.
<code>CohortSummary</code>	A data frame with cohort-level summaries (leaf area index, tree basal area, tree density, shrub cover, etc.) at the beginning of the simulation and after each simulated year.
<code>TreeTable</code>	A data frame with tree-cohort data (species, density, diameter, height, etc.) at the beginning of the simulation (if any) and after each simulated year.
<code>DeadTreeTable</code>	A data frame with dead tree-cohort data (species, density, diameter, height, etc.) at the beginning of the simulation and after each simulated year.
<code>CutTreeTable</code>	A data frame with cut tree-cohort data (species, density, diameter, height, etc.) per each simulated year.
<code>ShrubTable</code>	A data frame with shrub-cohort data (species, density, cover, height, etc.) at the beginning of the simulation and after each simulated year.
<code>DeadShrubTable</code>	A data frame with dead shrub-cohort data (species, density, cover, height, etc.) at the beginning of the simulation (if any) and after each simulated year.
<code>CutShrubTable</code>	A data frame with cut shrub-cohort data (species, density, cover, height, etc.) per each simulated year.
<code>ForestStructures</code>	A list with the <code>forest</code> object of the stand at the beginning of the simulation and after each simulated year.
<code>GrowthResults</code>	A list with the results of calling function <code>growth</code> (i.e., see 15.5.4) for each simulated year.
<code>ManagementArgs</code>	If management is considered, a list of management arguments to be used in another call to <code>fordyn()</code> .
<code>NextInputObject</code>	An object of class <code>growthInput</code> to be used in a subsequent simulation.
<code>NextForestObject</code>	An object of class <code>forest</code> to be used in a subsequent simulation.

18.5 Process details

18.5.1 Seed production and seed bank dynamics

The model considers **mortality of seeds** in the seed bank before adding new seeds. Annual seed bank mortality is simulated for each species using an exponential decay function, driven by the species-specific seed longevity (SL):

$$Seeds_{t+1} = Seeds_t \cdot \exp(-1/SL) \quad (18.1)$$

To determine seed production, the model determines the **seed rain** in the stand by determining which cohorts have total plant heights above maturity thresholds for trees and shrubs. This is done, for each cohort, by comparing its height with the species-specific parameter seed production height (H_{seed}). Missing values of H_{seed} are given values from control variables `seedProductionTreeHeight` and `seedProductionShrubHeight` for trees and shrubs, respectively. In addition to locally produced seeds, the user can use control parameter `seedRain` to specify a list of species names whose seeds arrive to the stand. Despite the origin, in non-spatial simulations the seed bank relative amount of species with seed rain is set to 100%, assuming that there is enough seed production to allow normal recruitment. However, when simulations are conducted in a spatial context, the species identity and relative amount of seeds is determined via dispersal sub-model (see chapter 20).

18.5.2 Recruitment from seeds

Actual **sapling recruitment** (i.e. recruitment of small trees, typically 1 cm diameter) depends on environmental conditions in the stand. Specifically, the model calculates, for the year that ended, the mean temperature of the coldest month (TCM), the moisture index (MI) and the fraction of photosynthetic active radiation reaching the ground, given the current structure ($FPAR$). These values are compared to species specific parameter thresholds TCM_{recr} , MI_{recr} and $FPAR_{recr}$, respectively. More specifically, a given species j can recruit only if $TCM > TCM_{recr,j}$, $MI > MI_{recr,j}$ and $FPAR > FPAR_{recr,j}$. These filters do not ensure recruitment, as it is assumed that multiple processes can further determine the death of recruits. A probability of recruitment P_{recr} is used to represent these additional processes. When simulation of recruitment is *stochastic*, a given species will recruit if (in addition to the bioclimatic limits) a Bernouilli draw falls below P_{recr} and, if so, the initial density or cover will be fixed. If recruitment is deterministic, then the probability of recruitment is used to multiply the recruitment density or cover.

Tree recruitment density, diameter and height are determined by parameters $N_{tree,recr}$, $DBH_{tree,recr}$ and $H_{tree,recr}$, respectively; whereas cover and height of shrub recruitment is determined by parameters $Cover_{recr}$ and $H_{shrub,recr}$, respectively. Density/cover of recruits is reduced depending on the relative amount of seeds in the seed bank (i.e. maximum density or cover will occur if seed bank levels are 100%). If stochastic simulation of recruitment is requested, then density or cover values are considered mean values of a Poisson distribution. Note that density of tree recruits will decrease during the years after recruitment due to a **self-thinning** process, until DBH reaches $DBH_{tree,ingrowth}$ where N will be $N_{tree,ingrowth}$, as explained in 17.4.1.

18.5.3 Resprouting

Currently, resprouting only occurs in the model if plant cohorts have been **cut**, **burned** or they died of **desiccation** (future model versions will include resprouting after fire impacts). In other words, resprouting does not occur for baseline mortality, self-thinning of recruits or due to starvation.

Resprouting survivorship

The model first determines **resprouting survivorship**, which depends on the disturbance type and, in the case of fire, on species identity. Unfortunately, we lack information on the mortality caused in resprouting by cutting or drought for the vast majority of species except for holm oak (*Quercus ilex*). Thus, we suggest applying tentatively to all species the mortality values obtained in different studies for this species: 2.5% after browsing (Espelta *et al.* 2006), 4% after cutting (Retana *et al.* 1992), 5% after drought (Lloret *et al.* 2004). Larger differences among species have been reported for fire (Espelta *et al.* 2012). This pattern may be linked to inter-specific differences in the size of the bud bank, the degree of bud protection or the amount of stored resources in belowground organs (burl, taproot). Accordingly, default values for survivorship are $Resp_{clip} = 0.96$ (for cutting) and $Resp_{dist} = 0.95$ (for dessication), whereas no default value is given for $Resp_{fire}$. Naturally, the number of surviving trees (stumps) is given by the number of trees affected by the disturbance multiplied by the survivorship rate:

$$N_{surv} = N_{mort,dist} \cdot Resp_{dist} \quad (18.2)$$

for a generic disturbance, but the same equation would apply for cutting, dessication or fire.

Resprouting vigor

For those individuals that survive the disturbance event and are able to resprout, the number of resprouts present in a given moment is a matter of the stump area and age. According to different literature sources, the number of resprouts initially produced is approximately 1.82 per 1 cm² of stump area, but owing to the intra- and inter-individual competition (self-thinning), the number of resprouts will decrease with time following an exponential equation (Retana *et al.* 1999. Espelta *et al.* 1999, 2013):

$$N_{resp} = N_{surv} \cdot \pi \cdot (DBH/2)^2 \cdot 1.82 \cdot 10^{-0.053 \cdot x} \quad (18.3)$$

where DBH is diameter at breast height of the *parent* tree cohort, in cm, $x = 5$ is time in years, N_{resp} is the number of resprouts, N_{surv} is the number of parent trees that survived.

Initial characteristics and self-thinning

Tree resprouts have all a starting diameter $DBH_{tree,recr}$ and a height $H_{tree,recr}$, but the root system is the same of the *parent* tree cohort. Analogously to recruitment from seeds, the initial density of tree resprout will decrease during the years after the initial resprouting due to a **self-thinning** process, until DBH reaches $DBH_{tree,ingrowth}$ where N will be $N_{tree,ingrowth}$, as explained in 17.4.1.

Resprouting in shrubs

In the case of shrub species, the same operations are done, but on cover values instead of density values. Shrub height of resprouts is assumed to be $H_{shrub,recr}$.

Part VI

Landscape/regional modelling

Chapter 19

Watershed hydrology

This chapter describes the distributed watershed water balance sub-model implemented in functions `spwb_land()`, `growth_land()` and `fordyn_land()` of package **medfateland** (Fig. 1.3).

19.1 Design principles

Watersheds are described in raster (i.e. gridded) mode, each cell representing a patch of vegetation (or another land cover) in a catchment. Land cover types are:

- a. *wildland*: forests, shrublands or grasslands.
- b. *agriculture*: agricultural lands.
- c. *rock*: rock outcrops.
- d. *artificial*: urban areas.
- e. *water*: water bodies.

Watershed water balance simulations extend the simulation of soil water balance to the landscape scale. Hence, the design of most vertical hydrological processes is the same as those of forest water balance. However, additional water compartments and processes are represented in watershed water balance simulations. Each cell in the watershed may include the following: a snow-pack compartment, one or several soil layers (including a rocky layer down to several meters depth), and a groundwater compartment assumed to be beyond the reach of plant roots. Similar to other models such as TETIS (Francés *et al.* 2007), three lateral flows are considered between adjacent cells:

- (i) Overland surface flows from upper elevation cells.
- (ii) Lateral saturated soil flows (i.e. interflow) between adjacent cells.
- (iii) Lateral groundwater flow (i.e. baseflow) between adjacent cells.

Overland surface flows are modeled following T-HYDRO Ostendorf & Reynolds (1993). Subsurface flows, including both soil saturated flow and groundwater flow, are modeled following the kinematic wave approach of DSHVM Wigmosta *et al.* (1994).

Split-parameter parametrization following Francés *et al.* (2007).

19.2 State variables

Distributed simulation of rainfall-runoff processes implies that state variables are defined for each cell of the watershed. For this kind of simulations, state variables describing the water content of different compartments are the most important. In the case of *wildland* and *agriculture* cells these are:

- W_s , the proportion of soil moisture in relation to field capacity for each soil layer s in the cell.
- S_{snow} the snow water equivalent (mm) of the snow pack storage over the cell surface.
- $S_{aquifer}$ the water content (mm) in the cell's aquifer beyond the reach of plant roots.

Additional cell state variables in *wildland* cells concern the water status (or other state variables) of plant cohorts in the cell, and they were described in chapters 3 and 7. In the case of cells of other land cover types (i.e. *rock*, *artificial* or *water*) soils are not defined, and neither are soil state variables.

19.3 Water balance

In distributed watershed simulations water balance can be defined both at the cell level and at the watershed level.

19.3.1 Cell level water balance

At the cell level, daily variations in water content can occur in the *snowpack*, *soil* or *groundwater* compartments:

1. Variations in snowpack water equivalent content follows eq. (3.2).
2. Variations in soil water content (ΔV_{soil}) need to account for additional flows and are summarized as (compare to (3.1)):

$$\Delta V_{soil} = Pr + Sm + Ro + Ad - In - Ru - Dd - Es - Tr + \Delta S_{sat} \quad (19.1)$$

where Pr is precipitation as rainfall, Sm is snowmelt, Ro is surface *runon* water entering the cell from neighboring cells at higher elevation, Ad is aquifer discharge, In is rainfall interception loss, Ru is cell surface runoff, Dd is deep drainage towards the aquifer, Es is soil evaporation, Tr is plant transpiration and ΔS_{sat} is the variation in soil water content derived from the balance between saturated soil lateral inputs and outputs.

3. Variations in the aquifer water content of a cell are summarized by:

$$\Delta S_{aquifer} = Dd - Ad - \Delta S_{base} \quad (19.2)$$

where ΔS_{base} is the variation in water content of the aquifer derived from the balance between groundwater lateral inputs and outputs.

19.3.2 Watershed water balance

At the watershed level, separate water balances can again be defined for the average water content of snow pack, soil or groundwater compartments. These result from averaging water balances across cells. Additionally, a water balance is defined regarding the overall water content in the watershed.

1. Changes in the average snow pack water equivalent over cells is the result of balancing precipitation as snow (P_s) and snow melt (Sm), both flows averaged over cells:

$$\Delta \hat{S}_{snow} = \hat{P}_s - \hat{Sm} \quad (19.3)$$

where \hat{P}_s and \hat{Sm} are the average snow fall and snow melt over cells.

2. Changes in the average soil moisture ($\Delta\hat{V}_{soil}$) are the result of pooling soil inputs and outputs over cells, which yields:

$$\Delta\hat{V}_{soil} = \hat{P}r + \hat{S}m + \hat{A}d + \hat{R}o - \hat{R}u - \hat{I}n - \hat{D}d - \hat{E}s - \hat{T}r \quad (19.4)$$

where $\hat{P}r$, $\hat{A}d$, $\hat{R}o$, $\hat{R}u$, $\hat{I}n$, $\hat{D}d$, $\hat{E}s$ and $\hat{T}r$ are the average (over cells) of precipitation (including rain and snow), aquifer discharge, lateral surface water input (runon), runoff, rainfall interception loss, deep drainage, soil evaporation and plant transpiration, respectively. Lateral saturated soil flows are not included as they cancel out at the watershed level.

3. Changes in the average of cell aquifer water content ($\Delta\hat{S}_{aquifer}$) are the result of balancing deep drainage (Dd) from soils and aquifer discharge (Ad):

$$\Delta\hat{S}_{aquifer} = \hat{D}d - \hat{A}d \quad (19.5)$$

4. If we integrate the three water compartments, water balance at the watershed level is given by:

$$\Delta S_{watershed} = \hat{P}r - \hat{I}n - \hat{E}s - \hat{T}r - \hat{E}x \quad (19.6)$$

where $\Delta S_{watershed} = \Delta\hat{S}_{snow} + \Delta\hat{V}_{soil} + \Delta\hat{S}_{aquifer}$ is the change in water content in the watershed and $\hat{E}x$ is the water exported as runoff from cells without neighbors (i.e. catchment outlet cells).

19.4 Process scheduling

For every day to be simulated, the model performs the following steps:

1. Calculation of soil and groundwater hydraulic heads for each cell (see 19.6.1).
2. Calculation of soil and groundwater lateral flows between adjacent cells, according to hydraulic gradients (see 19.6.1).
3. Apply changes in cell soil moisture content due to soil lateral flows (i.e. determine ΔS_{sub} for each cell), including the possibility of a return flow to the surface if the soil becomes saturated (i.e. saturation excess flow).
4. Apply changes in cell aquifer water content, including the possibility of discharge from the aquifer to the soil and a saturation excess flow (eq. (19.2)).
5. Determine remaining cell flows by processing watershed cells in order of decreasing elevation (i.e. cells at higher elevation are processed before cells at lower elevation). For each cell to be processed:
 - a. For *wildland* or *agriculture* cells, determine snow-pack dynamics, rainfall interception loss, infiltration and runoff, transpiration, soil evaporation processes as described in 3.4, while including saturation excess flow as well as potential runon (Ro) from upslope cells as additional water inputs. For *rock* and *artificial* cells, snow pack dynamics is still processed in these cells, but all runon or rainfall input becomes runoff to be passed downhill, and when the level of the aquifer reaches the surface any discharge also becomes runoff. In the case of *water* cells, snow pack dynamics is also processed, but evaporation from the water surface is not modelled. All liquid water inputs are poured onto the aquifer water content, and runoff can still occur when the aquifer reaches the surface.
 - b. Distribute surface runoff among cells downhill (19.6.2). If a cell does not have downhill neighbors (i.e outlet cell) its runoff becomes water exported from the watershed.

19.5 Inputs and outputs

19.5.1 Gridded inputs

Hydrological distributed models demand large amounts of data, information and parameters in order to accurately represent the spatial variability of the main hydrological processes and weather inputs. As mentioned in design principles, watersheds are described in raster (i.e. gridded) mode, each cell representing

a patch of vegetation (or another land cover) in a catchment. Rasters do not need to be (should not be) complete, since the model is intended to be used on individual catchments with defined boundaries.

Given a set of grid cells representing the watershed at a given spatial resolution, input data for `spwb_land()` and `growth_land()` should be specified using `sf` objects. These were described in section 2.1.2.

19.5.2 Watershed meteorology input

Simulation of watershed hydrology requires specifying weather variables for all cells and the simulation period considered. Functions `spwb_land()` and `growth_land()` allow weather to be specified in three ways:

- If a single data frame of daily weather is provided, the model assumes that all watershed cells experience the same weather.
- A different data frame of daily weather may be provided for each cell, using column `meteo` of the `sf` object (see 2.1.2).
- If an interpolator object of class `stars` is provided, the model will call function `interpolate_data()` for every day to be simulated (see `meteoland` package documentation).

19.5.3 Watershed hydraulic correction parameters

Distributed hydrological models suffer from the need to specify many parameters. Frances et al. (2007) demonstrated that by using a proper parameter structure, it is possible to obtain an excellent automatic calibration of a distributed conceptual model while maintaining the spatial variability of parameters. In particular, they showed how to define effective model parameters for those quantities normally estimated at the point scale, so that model parameters and estimates usually suffer from being *uncommensurable*. Effective model parameters can in this situation be defined as a *correction function* of the corresponding hydrological characteristic. If the correction function takes into account the model and input errors, the temporal and spatial scale effects and also the hydrological characteristics estimation error, it is reasonable to assume the correction function for each parameter will be common to all cells within the watershed. Frances et al. (2007) suggest using a correction factor for each hydraulic parameter, in a strategy called *split-parameter*. From the calibration point of view, a very important consequence of the split-parameter approach is that the number of variables to be adjusted is reduced dramatically.

Our model follows Frances et al. (2007) in that correction factors are defined for three hydraulic parameters with spatial variation across the watershed:

- K_{drain} vertical saturated soil conductivity for deep drainage.
- K_{sat} horizontal saturated soil conductivity for soil lateral flows.
- K_{base} bedrock conductivity for groundwater lateral flows.

The effective model parameter for each of these is defined for the spatial (depending on the grid resolution) and temporal (daily) scales of the model. Each effective parameter is split into the input value, assumed to have been estimated at a given spatial and temporal scale, and the corresponding correction factor, i.e. R_{drain} , R_{sat} and R_{base} . The values for these correction factors should be the target of calibration procedures, assuming that the spatial heterogeneity of effective parameters is adequately covered by the soil and bedrock spatial inputs.

19.5.4 Model outputs

Distributed hydrological models can produce very detailed spatial and temporal outputs, hence consuming lots of memory resources. While the model runs at a daily scale, functions `spwb_land()` and `growth_land()`

can be asked to retain spatial outputs and summaries at coarser temporal scales (i.e. weekly, monthly, etc.). The output of functions `spwb_land()` and `growth_land()` is an S3 list with the following data elements:

Element	Description
<code>sf</code>	An object of class <code>sf</code> containing the geometries, final state and summary of the landscape cells.
<code>watershed_balance</code>	Data frame with averaged (over cell) components of the watershed balance corresponding to each summary.
<code>watershed_soilBalance</code>	Average (over cell) components of the watershed balance, including only cells that have defined soils (i.e. <i>wildland</i> and <i>agriculture</i> cells), corresponding to each summary.
<code>daily_runoff</code>	A matrix with daily runoff (in m ³ /day) at each of the outlet cells of the landscape.

19.6 Process details

19.6.1 Subsurface flows

Both soil saturated lateral flow and groundwater lateral flow follow the saturated subsurface flow model of Wigmosta et al. (1994), which is based on a kinematic wave approximation. Each grid cell can exchange water with its eight adjacent neighbors. Local hydraulic gradients are approximated by local ground surface slopes, so that each cell will generally receive water from its upslope neighbors and discharge to its downslope neighbors. The rate of saturated flow from cell i to its downgradient neighbors is equal to (Wigmosta & Lettenmaier 1999):

$$q_{ij} = -T_i \cdot \tan(\beta_{i,j}) \cdot w_j \quad \beta_{i,j} < 0 \quad (19.7)$$

$$q_{ij} = 0 \quad \beta_{i,j} \geq 0 \quad (19.8)$$

where $q_{i,k}$ is the flow rate from cell i in the direction towards neighbor j , T_i is the transmissivity at cell i , $\beta_{i,j}$ is the water table slope in the direction of j . The transmissivity function for soil saturated lateral flow in cell i ($T_{sat,i}$) is specified using:

$$T_{sat,i} = \frac{R_{sat} \cdot K_{sat,i} \cdot Z_{soil,i}}{n_{sat}} \cdot (1 - Z_{wt,i}/Z_{soil,i})^{n_{sat}} \quad (19.9)$$

where $K_{sat,i}$ is the saturated conductivity of the first (top) soil layer in cell i , $Z_{soil,i}$ is the soil depth at cell i , $Z_{wt,i}$ is the water table depth at cell i and n_{sat} is a power exponent. R_{sat} is a correction factor (a watershed parameter) for the fact that $K_{sat,i}$ is assessed from soil samples, whereas the parameter in the model is defined at the scale of a grid cell (Francés *et al.* 2007). The transmissivity function for groundwater lateral flow in cell i ($T_{base,i}$) is analogous:

$$T_{base,i} = \frac{R_{base} \cdot K_{base,i} \cdot Z_{bedrock,i}}{n_{base}} \cdot (1 - Z_{aquifer,i}/Z_{bedrock,i})^{n_{base}} \quad (19.10)$$

where $K_{base,i}$ is the bedrock conductivity at cell i , $Z_{bedrock,i}$ is the depth of the unaltered bedrock, $Z_{aquifer,i}$ is the depth of the aquifer water table and n_{base} is again a power exponent.

Saturated lateral flow inputs and outputs for a given cell i are balanced to determine increase or decrease in soil moisture:

$$\Delta S_{sat,i} = \sum_j q_{sat,ji} - \sum_j q_{sat,ij} \quad (19.11)$$

and the same occurs for groundwater lateral flows and aquifer balance:

$$\Delta S_{base,i} = \sum_j q_{base,ji} - \sum_j q_{base,ij} \quad (19.12)$$

19.6.2 Overland flows

To simulate surface runoff routing from one cell to the other, the approach of Ostendorf & Reynolds (1993) is used, as in SIERRA (Mouillot *et al.* 2001). Overland water lateral transport for a given day occurs instantaneously (i.e. no velocities are calculated) and depends on topography only. The model determines cell neighbors following the queen rule (up to eight neighbors per cell). The proportion of surface water runoff of cell i that will be added to the infiltration input (runon) of a neighboring cell j is Ostendorf & Reynolds (1993):

$$q_{ij} = \frac{\Delta z_{ij}/L_{ij}}{\sum_j \Delta z_{ij}/L_{ij}} \quad (19.13)$$

if $\Delta z_{ij} = z_i - z_j > 0$, that is, if the difference in elevation between the two cells is positive (i.e. if $z_j < z_i$). Otherwise there is no overland transport from i to j , i.e. $q_{ij} = 0$. L_{ij} indicates the distance between cell i and j (which depends on cell size and on whether the neighboring cell j is diagonal to cell i). The summation of the denominator is done only for neighbors at lower elevation, so that $\sum_i q_{ij} = 1$. The table of q_{ij} values is calculated when initializing distributed watershed objects.

Every day, cells are processed in order from higher to lower elevation. After the daily water balance of a given cell i , water runoff Ru_i is divided among the neighboring cells at lower elevation. The runon of a neighbor j , Ro_j is updated as:

$$Ro_j = Ro_j + Ru_i \cdot q_{ij} \quad (19.14)$$

Note that a given cell j can receive overland flow from more than one neighbor.

19.6.3 Watershed runoff

A special situation arises when processing cells that do not have downhill neighbors defined (i.e. where $q_{ij} = 0$ for all j), either because they are in flat surfaces or at the watershed boundary. In both cases, these cells should correspond to water bodies or streams connecting to the catchment outlet. Hence, these cells are called *outlet cells* in the model, and the runoff they generate becomes watershed runoff.

Chapter 20

Seed dispersal

This chapter describes the seed dispersal sub-model implemented in functions `fordyn_scenario()` and `fordyn_land()` of package **medfateLand** (Fig. 1.3).

20.1 Design principles

Given their sessile lifestyle, dispersal is the only process through which plants can spread in a landscape. Different parts of plants can be dispersed (seeds, fruits, branches or even the entire plant), but here it is assumed that seeds are the dispersed elements. Mathematically, dispersal can be addressed via process-based models (e.g. Nathan *et al.* (2001)). However, it is more commonly addressed via empirical dispersal kernels, which describe the statistical distribution of dispersal distances in a population (Nathan *et al.* 2012). More specifically, the dispersal kernel is a probability density function describing the distribution of post-dispersal locations relative to a source point. While many different equations can be used and compared to describe dispersal (Nathan *et al.* 2012), here we focus on the exponential power kernel proposed by Clark *et al.* (1998), which has shown a good performance compared to other kernels (Bullock *et al.* 2017) and has been used for other dynamic vegetation models (Snell 2014). The kernel is isotropic, meaning that is equal in all directions. Therefore, one can consider dispersal either from source or sink locations.

When implementing a dispersal kernel in a dynamic forest simulation model, an important issue to overcome is the fact that forest stands in the target landscape are often separated by distances that are larger (e.g. 500 m, 1 km or even more) than the average area that they represent (normally forest plots have a radius between 10 and 25 m). This entails that there is a uncertainty in the model regarding the presence of seed sources from forest stands at distances between the two scales. In other words, the model does not know whether there are forest patches that can act as seed sources, between a given target forest stand and its nearest neighbors (Fig. 20.1). Ignoring this fact can lead to underestimating colonization. We address this issue by considering that seed sources from unavailable forest stands can be estimated from the set of stands that are represented in the data set, using weights inverse to their distance to the location of the unavailable stand. This is illustrated in Fig. 20.1 below with the area with unavailable stands in grey and distances represented using red and black arrows:

20.2 Process scheduling

Seed dispersal is considered once a year. Process scheduling in the dispersal sub-model is rather straightforward:

1. Mortality of seeds in first performed for all stands (see 18.5.1).

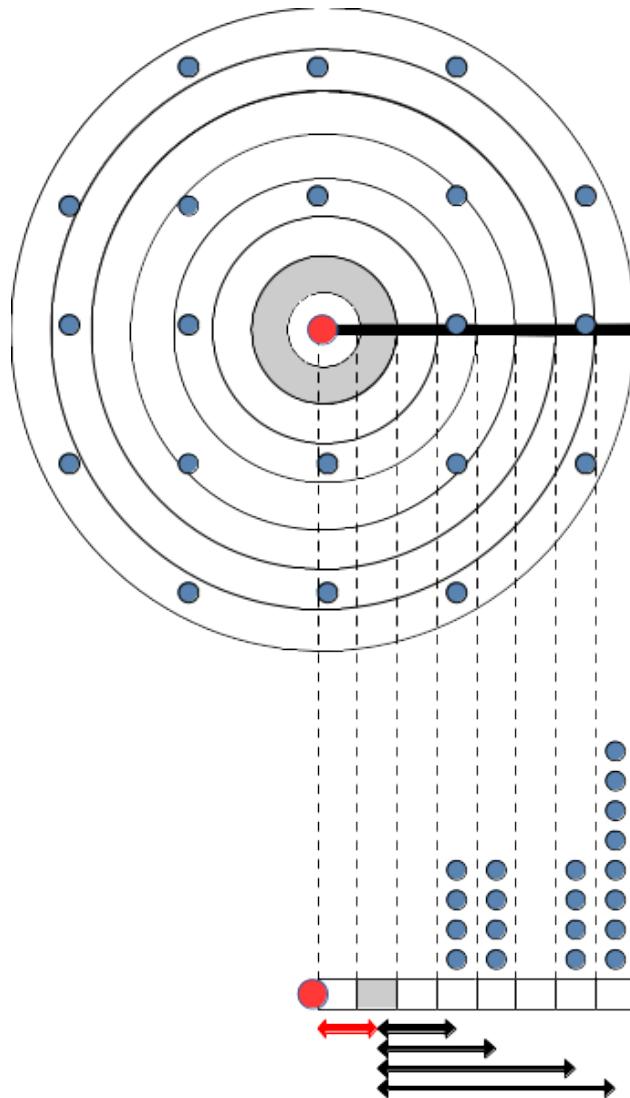


Figure 20.1: Example of sparse forest stand distribution and how to deal with dispersal in this situation. The target stand where seed rain is to be determined is represented using a red dot. Blue dots are the remaining forest stands. The upper figure shows the spatial distribution of stands, whereas the figure below represents the number of stands per distance classes. In both cases, grey is used to identify an area with unavailable data.

2. Seed production is simulated in all stands with woody vegetation (see 18.5.1).
3. For each target forest stand, seed dispersal (including local seed rain) is simulated, leading to a replenishment of its seed bank.

20.3 Process details

The exponential power kernel depends on species-specific parameters $Disp_{dist}$ and $Disp_{shape}$ and is calculated for a given distance r using (Clark *et al.* 1998):

$$f(r) = \frac{1}{N} \cdot \exp \left[- \left(\frac{r}{Disp_{dist}} \right)^{Disp_{shape}} \right] \quad (20.1)$$

where N is the normalizing constant:

$$N = \frac{2 \cdot \pi \cdot Disp_{dist}^2 \cdot \Gamma(2/Disp_{shape})}{Disp_{shape}} \quad (20.2)$$

and $\Gamma()$ is the Gamma function.

Let i be the stand corresponding to seed sink location (e.g. the red dot in Fig. 20.1). We begin by computing the geographic distances between i and all the other stands using geographic coordinates x and y (assuming these are in meters).

$$d(i, j) = \sqrt{(x_i - x_j)^2 + (y_i - y_j)^2} \quad (20.3)$$

Let us now consider a given distance value $r > 0$ from i (e.g. the grey band in Fig. 20.1). We define the probability of choosing a given stand j for a location at this distance from i as follows:

$$p_i(r, j) = \frac{1}{M} \cdot \frac{1}{|r - d(i, j)|} \quad (20.4)$$

where M is an appropriate normalizing factor so that:

$$\sum_{j=1}^n p_i(r, j) = 1 \quad (20.5)$$

Note that this last equation assumes that the density of spatial points is even across space. If not, the sum should be lower than one.

Given the above definitions and assuming that seed bank of a given species should be refilled to 100% if and only if all plots are seed sources for that species, the percentage of seeds of species s reaching the target location i , i.e. $P_{s,i}$, is estimated using:

$$P_{s,i} = 100 \cdot 2\pi \cdot \int_{r>0}^{r_{\max}} r \cdot f(r) \cdot \left(\sum_{j=1}^n I(s, j) \cdot p_i(r, j) \right) \cdot \delta r \quad (20.6)$$

where r_{\max} is the maximum allowed dispersal distance and $I(s, j) \in \{0, 1\}$ is an indicator variable of the availability of seed sources of species s in stand j .

Part VII

Static modules

Chapter 21

Allometric models

The purpose of this chapter is to describe how certain plant structural attributes are estimated in **medfate** using allometric relationships. These static models are used to initialize vegetation input parameters from forest plot data before simulations (in functions `forest2spwbInput()` and `forest2growthInput()`), to update vegetation structure during growth simulations or to calculate foliar biomass for fuel assessments.

21.1 Input data

21.1.1 Forest plot data

As explained in section 2.4, **medfate** has been specially designed to work with forest inventory plots with data in form of plant cohorts. The tree/shrub cohort attributes required to apply allometric models are the following:

Symbol	Units	R	Description	trees	shrubs
SP_i		Species	Species identity	Y	Y
H_i	cm	Height	Average tree or shrub height	Y	Y
N_i	$ind\cdot ha^{-1}$	N	Density of tree individuals	Y	N
DBH_i	cm	DBH	Tree diameter at breast height	Y	N
$Cover_i$	%	Cover	Shrub percent cover	N	Y

Most allometric functions require data in form of a `forest` object (see section 2.4.4).

21.1.2 Species parameters

The following table shows the allometric coefficients needed for the calculations detailed in this chapter.

Symbol	Units	R param	Description
$a_{fbt}, b_{fbt}, c_{fbt}$		<code>a_fbt, b_fbt, c_fbt</code>	Coefficients to calculate foliar biomass of an individual tree
a_{ash}, b_{ash}		<code>a_ash, b_ash</code>	Coefficients relating the square of shrub height with shrub area
a_{bsh}, b_{bsh}		<code>a_bsh, b_bsh</code>	Coefficients relating crown volume with dry weight of shrub individuals

Symbol	Units	R param	Description
$a_{cr}, b_{1cr}, b_{2cr}, b_{3cr}, c_{1cr}, c_{2cr}$		$a_cr, b_1cr, b_2cr, b_3cr, c_1cr, c_2cr$	Coefficients to calculate crown ratio of trees
cr	[0-1]	cr	Ratio between crown length and total height for shrubs
$r_{6.35}$		$r635$	Ratio between the weight of leaves plus branches and the weight of leaves alone for branches of 6.35 mm
SLA	$m^2 \cdot kg^{-1}$	SLA	Specific leaf area

21.2 Allometric relationships

21.2.1 Leaf biomass

Leaf biomass calculations are done differently for trees and shrub cohorts. They are calculated from `forest` objects using function `plant_foliarBiomass()`.

Tree cohorts

Foliar biomass for a single tree of cohort i ($FB_{tree,i}$; in kg) is calculated using:

$$FB_{tree,i} = a_{fbt} \cdot DBH_i^{b_{fbt}} \cdot e^{c_{fbt} \cdot BAL_i} \cdot e^{-0.0001 \cdot N_i} \quad (21.1)$$

where DBH_i is the diameter of the tree (in cm), BAL_i is the cumulative basal area ($m^2 \cdot ha^{-1}$) of trees having an equal or larger diameter (including the target tree), N_i is the density of the cohort, and a_{fbt} , b_{fbt} and c_{fbt} are species-specific regression coefficients. The first two determine the relationship between tree diameter and its foliar biomass, whereas the third one leads to reductions of foliar biomass because of over-shading and self-shading. Factor $e^{-0.0001 \cdot N_i}$ reduces foliar biomass for cohorts with very high density (such as very dense regeneration after a fire). The foliar biomass of the whole tree cohort (FB_i ; in $kg \cdot m^{-2}$) is obtained multiplying tree foliar biomass by tree density (N_i ; in $ind. \cdot ha^{-1}$):

$$FB_i = FB_{tree,i} \cdot (N_i / 10000) \quad (21.2)$$

Shrub cohorts

To calculate the leaf biomass of a shrub cohort, we first determine $A_{sh,i}$, the area (in cm^2) occupied by one average individual of height H_i (in cm), using the relationship:

$$A_{sh,i} = a_{ash} \cdot H_i^{b_{ash}} \quad (21.3)$$

where a_{ash} and b_{ash} are species-specific parameters. The model then estimates the dry weight of leaves and branches up to 6.35mm in diameter ($B_{sh,i}$, in kg) of this average individual (i.e. fine fuel biomass), using an allometric relationship with shrub crown phytovolume assuming a cylinder (in cm^3):

$$B_{sh,i} = a_{bsh} \cdot (A_{sh,i} \cdot H_i)^{b_{bsh}} \cdot e^{-0.235 \cdot LAI_{trees}^{live}} \quad (21.4)$$

where a_{bsh} and b_{bsh} are species-specific parameters. Here LAI_{trees}^{live} ($m^2 \cdot m^{-2}$) is the sum of leaf area index values of tree cohorts, which induces a reduction of $B_{sh,i}$ similar to the one described for tree foliar biomass by BAL_i . This reduction is important for biomass and leaf area estimation of shrubs in forests, since it has been observed that light availability affects the growth, architecture and bulk density of the understory (Pimont *et al.* 2018).

Shrub density (N_i ; in $ind. \cdot m^{-2}$) can be grossly estimated from percent cover (C_i , in percent) and $A_{sh,i}$ (in cm^{-2}):

$$N_i = \frac{C_i / 100}{A_{sh,i} / 10000} \quad (21.5)$$

The fine fuel biomass of a shrub cohort (W_i , in $kg \cdot m^{-2}$) is simply the product of $B_{sh,i}$ (kg of dry weight) and N_i :

$$W_i = B_{sh,i} \cdot N_i \quad (21.6)$$

Foliar biomass (in $kg \cdot m^{-2}$) can be obtained using the species-specific ratio $r_{6.35,i}$:

$$FB_i = W_i / r_{6.35,i} \quad (21.7)$$

If not known, $r_{6.35,i}$ can be set to a default value of 2 (equivalent to 50% of weight corresponding to leaves).

Herbaceous layer

Foliar biomass of the herbaceous layer (which is equal to its fine fuel loading) is estimated from herbaceous cover ($Cover_{herb}$; in %) and height (H_{herb} ; in cm) assuming a constant bulk density coefficient:

$$FB_{herb} = 0.014 \cdot Cover_{herb} \cdot (H_{herb}/100.0) \cdot e^{-0.235 \cdot LAI_{woody}^{live}} \quad (21.8)$$

where the factor $e^{-0.235 \cdot LAI_{woody}^{live}}$ is analogous to that of shrubs (eq. (21.4)), but using the overall leaf area index of woody cohorts as driving variable.

21.2.2 Leaf area and LAI

Leaf area index (LAI in $m^2 \cdot m^{-2}$) of a given cohort i can be calculated from its foliar biomass (in $kg \cdot m^{-2}$) by using a *specific leaf area* coefficient (SLA , in $m^2 \cdot kg^{-1}$) that in **medfate** is species-specific:

$$LAI_i = FB_i \cdot SLA \quad (21.9)$$

The leaf area (LA in m^2) of an individual of the cohort is found by dividing it by plant density (N_i , in $ind \cdot ha^{-1}$) and multiplying by 10000 $m^2 \cdot ha^{-1}$:

$$LA_i = 10000 \cdot LAI_i / N_i \quad (21.10)$$

LAI values of woody cohorts are calculated from **forest** objects using function **plant_LAI()**.

Leaf area index of the herbaceous layer (LAI_{herb}) is estimated from FB_{herb} by applying a $SLA = 9$ value, is limited to $LAI_{herb} = 2 m^2 \cdot m^{-2}$, and can be calculated from **forest** objects using function **herb_LAI()**.

21.2.3 Crown vertical dimensions

Crown base height, i.e. the height where the first living branch of trees or shrubs occurs, is an important parameter to know the vertical distribution of leaves (see section 2.4.5). Crown base height of a plant cohort i (i.e., $H_{crown,i}$, in cm) is related to the total height of the cohort (H_i , in cm) and its crown ratio (CR_i), which is the ratio between crown length and plant height:

$$H_{crown,i} = (1 - CR_i) \cdot H_i \quad (21.11)$$

In the case of shrubs the crown ratio CR_i is normally an input (species-specific) parameter (cr). In the case of trees, the crown ratio is modelled as a function of tree size and stand competition, following a modification of the logistic equation of Hasenauer & Monserud (1996) :

$$CR_i = \frac{1}{1 + e^{-(a_{cr} + b_{1cr} \cdot HD_i + b_{2cr} \cdot (H_i/100) + b_{3cr} \cdot DBH_i^2 + c_{1cr} \cdot BAL_i + c_{2cr} \cdot \ln(CCF_i))}} \quad (21.12)$$

where $HD_i = H_i/(100 \cdot DBH_i)$ is the height to diameter ratio (in $m \cdot cm^{-1}$), H_i is the tree height, DBH_i is the diameter, CCF_i is the crown competition factor and a_{cr} , b_{1cr} , b_{2cr} , b_{3cr} , c_{1cr} and c_{2cr} are species-specific parameters. The crown competition factor is in turn calculated using (Krajicek *et al.* 1961):

$$CCF_i = \sum_i N_i \cdot \pi \cdot (CW_i/2)^2 / 100 \quad (21.13)$$

where N_i is the tree density and CW_i is the crown width (in m) assuming an open-grown tree, estimated from an allometric relationship with tree diameter:

$$CW_i = a_{cw} \cdot DBH_i^{b_{cw}} \quad (21.14)$$

where again a_{cw} and b_{cw} are species-specific parameters.

Crown ratios can be calculated from forest objects using function `plant_crownRatio()`. Similar calculations are performed to estimate crown length and crown base height, with functions `plant_crownLength()` and `plant_crownBaseHeight()`, respectively.

Chapter 22

Post-simulation procedures

22.1 Water use efficiency

Chapter 23

Wind extinction

In this chapter we describe a few wind extinction models that are implemented in **medfate**. Wind extinction is relevant: (a) to determine convective heat exchanges between the soil and the canopy; (b) to estimate wind speed and energy balance at the leaf level for different plant cohorts; and (c) for fire behaviour calculations.

23.1 Wind speed at the top of the canopy

Input wind speed (u) is assumed to represent the speed of wind at 6 m (20 feet) above the canopy. Following Albini & Baughman (1979) the wind speed (in $m\text{s}^{-1}$) at the top of the canopy is:

$$u_{top} = \frac{(1.01857 \cdot u) \cdot 0.4265092 \cdot H_{canopy}}{\log(20 + 1.181102 \cdot H_{top})} \quad (23.1)$$

where H_{top} is the canopy top height in m .

23.2 Wind extinction profile

The wind extinction profile, i.e the wind speed at any height z in m , can be calculated following Massman (1987):

$$\beta_{stand} = \frac{4.0 \cdot 0.2 \cdot LAI_{stand}^{phi}}{0.16 \cdot 1.5^2} \quad (23.2)$$

$$u(z) = u_{top} \cdot \sqrt{\frac{\cosh(\beta_{stand} \cdot z/H_{top})}{\cosh(\beta_{stand})}} \quad (23.3)$$

Chapter 24

Fuel characteristics and fire behaviour

24.1 Overview

Functions `fuel_FCCS()` and `fire_FCCS()` allow calculating potential fire behaviour for forest inventory plots. Formulation of fuel characteristics and fire behaviour is an adaptation of the Fuel Characteristics Classification System [FCCS; Prichard *et al.* (2013)]. In FCCS, fuelbed is divided into six strata, including canopy, shrub, herbaceous vegetation, dead woody materials, leaf litter and ground fuels. All except ground fuels are considered here. The intensity of burning depends on several factors, including topography, wind conditions, fuel structure and its moisture content, which is determined from antecedent and current meteorological conditions. A modification of the Rothermel's (1972) model is used to calculate the intensity of surface fire reaction (in kW/m^2) and the rate of fire spread (in m/min) of surface fires assuming a steady-state fire. Both quantities are dependent on fuel characteristics, windspeed and direction, and topographic slope and aspect.

Fuel and fire behavior calculations provide the following results:

- 1) Fuel characteristics by stratum.
- 2) Surface fire behavior (i.e. reaction intensity, rate of spread, fireline intensity and flame length);
- 3) Crown fire behavior.
- 4) Fire potential ratings of surface fire behavior and crown fire behavior.

The following figure provide an overview of the steps to calculate surface/crown fire behavior and fire potentials from fuel characteristics.

24.2 Input data

24.2.1 Forest plot data

As explained in section 2.4, `medfate` has been specially designed to work with forest inventory plots. The tree/shrub cohort attributes required to apply allometric models are same as for chapter 21:

Symbol	Units	R	Description	trees	shrubs
SP_i		Species	Species identity	Y	Y
H_i	cm	Height	Average tree or shrub height	Y	Y
N_i	$ind\cdot ha^{-1}$	N	Density of tree individuals	Y	N

Symbol	Units	R	Description	trees	shrubs
DBH_i	cm	DBH	Tree diameter at breast height	Y	N
$Cover_i$	%	Cover	Shrub percent cover	N	Y

Cohorts are not distinguished for the herbaceous stratum, and the variables needed are:

Symbol	Units	R	Description
C_{he}	%	herbCover	Herbaceous percent cover
H_{he}	cm	herbHeight	Mean herb height

Finally, the model also requires the percent cover of trees in the canopy (C_{ca}). This is easily available from forest inventory data, but could also be derived from the description of tree cohorts.

Fire behaviour functions require data in form of a `forest` object (see section 2.4.4).

24.2.2 Species parameters

The following functional parameters are required for each species:

Symbol	Units	R	Description
L_{shape}	Categorical	LeafShape	Leaf shape: “Broad”, “Needle”, “Linear”, “Scale”, “Spines” or “Succulent”
L_{size}	Categorical	LeafSize	Leaf size: “Small” (< 225 mm), “Medium” (> 225 mm & < 2025 mm) or “Large” (> 2025 mm)
$r_{6.35}$		r635	Ratio between the weight of leaves plus branches and the weight of leaves alone for branches of 6.35 mm
ρ_{wood}	$g \cdot cm^{-3}$	WoodDens	Density of wood tissue
ρ_{leaf}	$g \cdot cm^{-3}$	LeafDens	Density of leaf tissue
σ_i	$m^2 \cdot m^{-3}$	SAV	Surface-area-to-volume ratio of the small fuel (1h) fraction (leaves and branches < 6.35mm)
h	$kJ \cdot kg^{-1}$	HeatContent	High fuel heat content.
LD	years	LeafDuration	Leaf duration (in years)
LI	%	PercentLignin	Percentage of lignin in leaves

Leaf shape and size categories are used to determine leaf litter fuel types.

24.2.3 Other inputs

Other inputs may be given by expert opinion or they may be calculated from another model. Specifically, for each plant cohort (and for any day of application) the fire behaviour model requires:

- P_{dead} : Proportion of the plant that is dead.
- FMC : Fuel moisture content (in percent of dry weight).

Analogously, the same variables are needed for the herbaceous stratum.

- $P_{dead,he}$: Proportion of herb fuels that respond to humidity changes as 1-h dead fuels.
- $FMC_{live,he}$: Fuel moisture content of live herb fuels (in percent of dry weight).

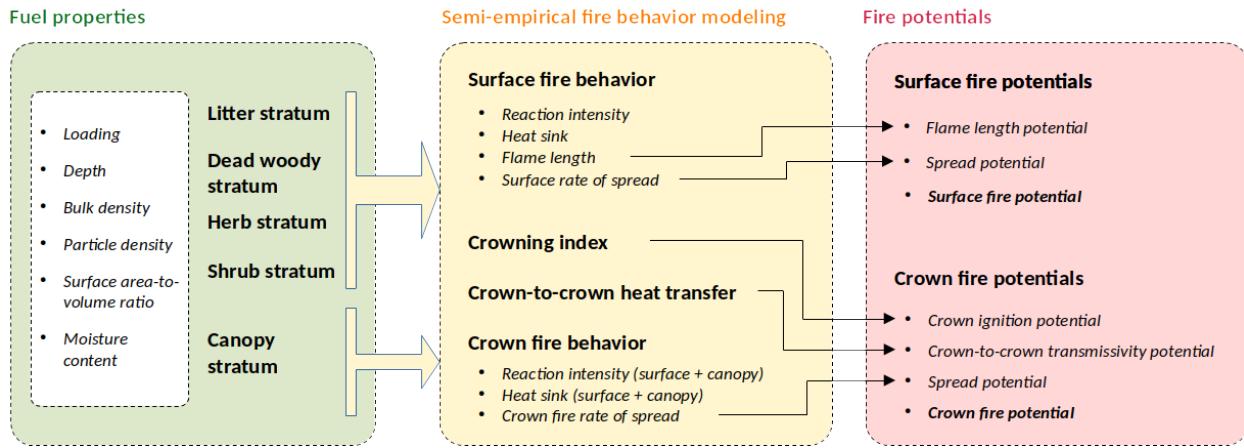


Figure 24.1: Overview of the steps involved in fire behavior and fire potential calculations starting from fuel properties

The model also needs the following input parameters:

- FMC_{dead} : the moisture content of 1-h dead fuels (in percent of dry weight).
- U : Midflame windspeed (in $m \cdot s^{-1}$).
- S : Slope (in percent).

24.3 Fuel characteristics

24.3.1 Fuel strata

The Fuel Characteristics Classification System (FCCS) on which this document is based, defines six fuel strata (Prichard *et al.* 2013):

- *Canopy*: Trees, snags and ladder fuels.
- *Shrubs*: Primary and secondary layers.
- *Non-woody vegetation (herbs)*: grasses, sedges, rushes and forbs.
- *Woody fuels*: All downed and dead wood, sound wood, rotten wood and stumps.
- *Litter-lichen-moss*: Lichen, litter and moss layers.
- *Ground fuels*: Duff, basal accumulation and squirrel middens.

Shrubs, herbs and woody fuels are constitute the **upper surface fuels**, whereas herbs and woody fuels alone constitute the **lower surface fuels**. FCCS summarizes and calculates characteristics for each fuelbed stratum and layer. Our model estimates fuel loading and characteristics for canopy, shrub, non-woody vegetation, as well as fine (1h) woody fuels and litter fuels. Larger woody fuels (10h or 100h) could be considered if information about forest management actions is available. Ground fuels are not included here.

24.3.2 Cohort fuel loading

Here we consider as burnable fuels foliage and branches up to 6.35 mm = 0.25 in in diameter. The same consideration applies to both trees and shrubs. They are calculated from `forest` objects using function `plant_fuel()`.

Tree cohorts

Fine fuel loading for a tree cohort (W_i ; in $kg \cdot m^{-2}$), including its leaves and branches with diameter up to 6.35 mm = 0.25 in, is calculated from foliar biomass (FB_i , see eq. (21.2)) using:

$$W_i = r_{6.35,i} \cdot FB_i \quad (24.1)$$

where $r_{6.35,i}$ is the ratio between the weight of leaves plus branches and the weight of leaves alone for branches of 6.35 mm in diameter for the species of cohort i . The biomass corresponding to branches of less than < 6.35 mm (SBB_i , also in $kg \cdot m^{-2}$) is obtained by subtraction:

$$SBB_i = (r_{6.35,i} - 1) \cdot FB_i \quad (24.2)$$

Whereas W_i is the cohort loading variable influencing fire behavior, FB_i and SBB_i are cohort variables used to estimate fine dead woody and leaf litter loadings.

Shrub cohorts

Our procedure to estimate shrub fuel loading differs from Prichard *et al.* (2013) because they calculate first total biomass of the shrub species and then consider the percentage of total weight that corresponds to leaves and small branches. In our case, we estimate fine fuel loading (W_i , in $kg \cdot m^{-2}$) and foliar biomass (FB_i , in $kg \cdot m^{-2}$) of shrubs from eqs. (21.6) and (21.7). Biomass of small branches (in $kg \cdot m^{-2}$) can be obtained from :

$$SBB_i = W_i - FB_i \quad (24.3)$$

24.3.3 Vertical distribution of cohort fuels

Vertical distribution of fine fuels are distributed between the crown base height ($H_{crown,i}$; in cm) and the total height (H_i , in cm) following a **truncated Gaussian distribution**, as done for the distribution of leaves (see section 2.4.5). Crown base height of trees is calculated as explained in 21.2.3. The loading of a cohort that occurs within a given height interval of limits H_1 and H_2 is calculated as:

$$W_i(H_1, H_2) = W_i \cdot p_i(H_1, H_2) \quad (24.4)$$

where $p_i(H_1, H_2)$ is the proportion of the crown of cohort i that corresponds to the height interval (H_1, H_2).

24.3.4 Fuel bulk density profile

Knowing at which height fuels are placed, the **fuel bulk density profile** (Reinhardt *et al.* 2006) is defined for any given interval (H_1, H_2) as the bulk density (kg/m^3) of fine fuels corresponding to that interval:

$$BDP(H_1, H_2) = \frac{\sum_i W_i(H_1, H_2)}{H_2 - H_1} \quad (24.5)$$

Canopy bulk density normally ranges between 0 and 0.4 kg/m^3 (Scott & Reinhardt 2002). Sando & Wick (1972) arbitrarily defined canopy base height as the lower vertical 0.3-m section with a weight greater than $0.01124 kg/m^3$. A user-defined threshold t_{BDP} (in kg/m^3) in 0.1-m sections is used to differentiate the surface fuelbed from canopy fuels. Using this threshold the model calculates the following three heights (Reinhardt *et al.* 2006):

- *Shrub stratum base height, H_{sb}* (in cm): the minimum height between 0 and 2 m where fuel bulk density is larger than t_{BDP} .
- *Shrub stratum top height, H_{st}* (in cm): the maximum height between 0 and 2 m where fuel bulk density is larger than t_{BDP} . With this definition h_s cannot be higher than 2 m (corresponding to fuel model 4 in Anderson 1982).

- *Canopy base height, H_{cb} (in cm):* In terms of its consequences to crown fire initiation, canopy base height can be defined as the lowest height above the ground at which there is sufficient canopy fuel to propagate fire vertically through the canopy. It is calculated as the minimum height over H_{st} when the bulk density starts again to be larger than t_{BDP} .
- *Canopy top height, H_{ct} (in cm):* the maximum height where bulk density is larger than t_{BDP} .
- *Canopy gap, H_{gap} (in cm):* the difference between H_{cb} and H_{st} . The canopy gap is used to calculate crown initiation potential.

Figure 24.2 illustrates the definition and analysis of the fuel bulk density profile for a given forest stand (see function `vprofile_fuelBulkDensity()`). Following Mitsopoulos & Dimitrakopoulos (2007), a threshold $t_{BDP} = 0.04$ is used to determine shrub and canopy heights.

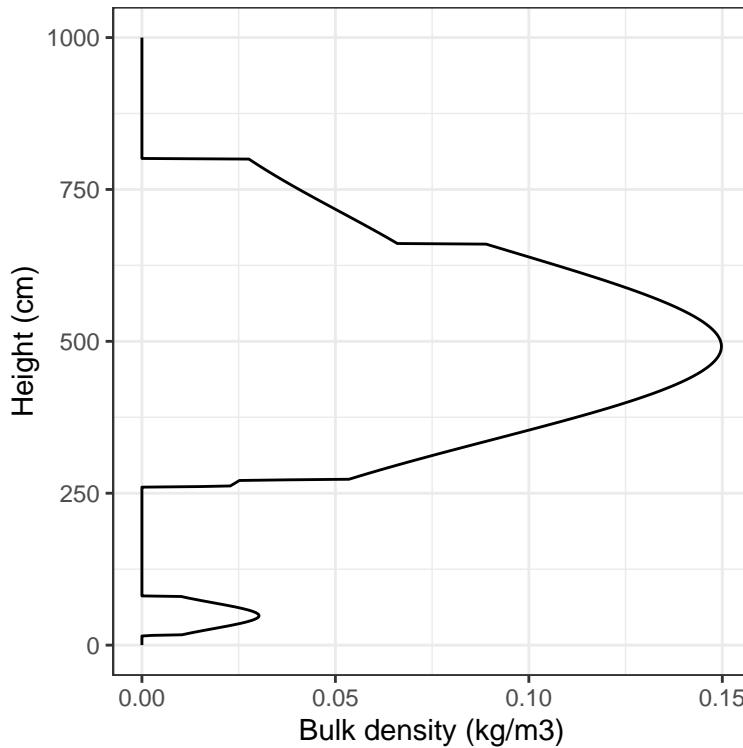


Figure 24.2: Bulk density profile of an example forest

24.3.5 Fuel loading (w) and fuel depth (δ)

24.3.5.1 Canopy stratum

Canopy loading (in $kg \cdot m^{-2}$) is the sum of (tree and shrub) cohort loadings above 2 m (i.e. 200 cm):

$$w_{ca} = \sum_i w_{i,ca} = \sum_i W_i(200, \infty) \quad (24.6)$$

where $w_{i,ca}$ is the canopy stratum loading of cohort i . Canopy depth (in m) is defined as the average of tree (or shrub) crown lengths above 2 m, weighted by the loadings of cohorts in the canopy:

$$\delta_{ca} = \frac{1}{100} \cdot \frac{\sum_i w_{i,ca} \cdot (H_i - H_{b,i}) \cdot p_{i,ca}}{\sum_i w_{i,ca}} \quad (24.7)$$

where the proportion of a tree (or shrub) cohort in the canopy stratum is $p_{i,ca} = p_i(200, \infty)$.

24.3.5.2 Shrub stratum

Shrub loading (in $kg \cdot m^{-2}$) is the sum of (tree and shrub) cohort loadings between the ground and 2 m (i.e. 200 cm):

$$w_{sh} = \sum_i w_{i,sh} = \sum_i W_i(0, 200) \quad (24.8)$$

where $w_{i,sh}$ is the shrub stratum loading of cohort i . The depth of the shrub stratum (in m) is defined as the average of tree (or shrub) crown lengths below 2 m, weighted by the loadings of cohorts in the shrub stratum:

$$\delta_{sh} = \frac{1}{100} \cdot \frac{\sum_i w_{i,sh} \cdot (H_i - H_{b,i}) \cdot p_{i,sh}}{\sum_i w_{i,sh}} \quad (24.9)$$

where the proportion of a shrub (or tree) cohort in the shrub stratum is $p_{i,sh} = p_i(0, 200)$.

24.3.5.3 Non-woody stratum

Herb percent cover and average herb height are transformed into herbaceous loading ($kg \cdot m^{-2}$) using (ref piropinos):

$$w_{he} = 0.014 \cdot C_{he} \cdot (H_{he}/100) \quad (24.10)$$

The depth of the herbaceous stratum (in m) is simply the mean height of herbs:

$$\delta_{he} = H_{he}/100 \quad (24.11)$$

24.3.5.4 Woody and litter strata

In FCCS (Prichard *et al.* 2013), woody surface loading includes several fuel sizes. However, when calculating surface fire behavior w_{wo} includes 100% of 1h fuels, 25% of 10h fuels and 12.5% of 100h fuels, which represents the material available for flaming combustion. Obtaining loading estimates for 10h- and 100h-fuels is very difficult without field fuel sampling. However, we might estimate 1h woody fuels and leaf litter from standing biomass of small branches (< 6.35mm) and leaves for trees and shrubs. Hence, our treatment of surface woody fuels includes only fine (1h) fuels.

Assuming a continuous input of litter, the variation in accumulated litter is described by a simple differential equation (Birk & Simpson 1980):

$$\frac{dX}{dt} = L - k \cdot X \quad (24.12)$$

where k is the decay constant, L is the rate of litterfall and X is the litter mass accumulated in the forest floor. Assuming that litter mass has reached a steady state, X can be estimated as the ratio between L and k . If litterfall is estimated as the total foliar biomass divided by leaf duration, the amount of steady state leaf litter corresponding to each tree and shrub cohort can be estimated using:

$$w_{li,i} = \frac{FB_i}{LD(SP_i) \cdot k_i} \quad (24.13)$$

where FB_i is the foliar biomass of cohort i , $LD(SP_i)$ is the species-specific average leaf duration (in years) and k_i is the rate of decay of leaves of cohort i , which is given by the regression model of Meentemeyer (1978):

$$k_i = (-0.5365 + 0.00241 \cdot AET) - (-0.01586 + 0.000056 \cdot AET) \cdot LI(SP_i) \quad (24.14)$$

where $LI(SP_i)$ is the species-specific percentage of lignin content in leaves and AET is actual evapotranspiration (default $AET = 1000mm$). Litter loadings are summed for four litter types (short pine needles, long pine needles, other conifers, broadleaves). In the case of fine dead woody materials (small fallen branches),

loading of small branches is taken as woody litter and it is assumed that small branch litterfall occurs at the same time as leaf litterfall (i.e. according to leaf duration):

$$w_{wo} = \sum_i w_{wo,i} = \sum_i \frac{SBB_i}{LD(SP_i) \cdot k_{wo}} \quad (24.15)$$

where $k_{wo} = 0.95y^{-1}$ is a constant rate of decomposition for small branches.

In FCCS, the depth of woody and LLM strata are inputs. In our case the depth of the woody and litter strata are estimated from the corresponding fuel loadings:

$$\delta_{wo} = w_{wo}/\rho_{b,wo} \quad (24.16)$$

$$\delta_{li} = w_{li}/\rho_{b,li} \quad (24.17)$$

where $\rho_{b,wo}$, $\rho_{b,li}$ are the woody and litter bulk density (in $kg \cdot m^{-3}$), respectively. Litter bulk density $\rho_{b,li}$ is calculated as a weighted average of litter types:

$$\rho_{b,li} = \frac{\sum_k \rho_{b,k} \cdot w_{li,k}}{\sum_k w_{li,k}} \quad (24.18)$$

where k indicates litter type. The bulk density for litter types are [Prichard *et al.* (2013); Table 1]:

$$\rho_{b,shortneedlepine} = \rho_{b,longneedlepine} = \rho_{b,otherconifer} = 1.65lb \cdot ft^{-3} = 26.43kg \cdot m^{-3} \quad (24.19)$$

$$\rho_{b,hardwood} = 0.83lb \cdot ft^{-3} = 13.30kg \cdot m^{-3} \quad (24.20)$$

24.3.6 Other fuel characteristics

All the following characteristics are calculated in metric units (although British units are indicated to qualify specific values for compatibility).

24.3.6.1 Particle density (ρ_p)

Particle density is the ratio of dry weight over volume for fuel particles (in $kg \cdot m^{-3}$). When species have different values, particle density averages for shrub and canopy strata can be obtained as:

$$\rho_{p,sh} = \frac{\sum_i w_{i,sh} \cdot \rho_p(SP_i)}{\sum_i w_{i,sh}} \quad (24.21)$$

$$\rho_{p,ca} = \frac{\sum_i w_{i,ca} \cdot \rho_p(SP_i)}{\sum_i w_{i,ca}} \quad (24.22)$$

where $\rho_p(SP_i)$ is the species-specific particle density (in $kg \cdot m^{-3}$), which can be obtained from wood tissue (ρ_{wood}) and leaf tissue (ρ_{leaf}) densities, using $r_{6.35}$ to weight them:

$$\rho_p(SP_i) = 1000 \cdot (\rho_{leaf}(SP_i) \cdot f_{leaves,vol} + \rho_{wood}(SP_i) \cdot (1 - f_{leaves,vol})) \quad (24.23)$$

where $f_{leaves,vol}$ is the volumetric fraction of leaves with respect to branchlets:

$$f_{leaves,vol} = \left[1.0 + (r_{6.35}(SP_i) - 1.0) \cdot \frac{\rho_{wood}(SP_i)}{\rho_{leaf}(SP_i)} \right]^{-1} \quad (24.24)$$

Dead woody and litter particle densities, i.e. $\rho_{p,wo}$ and $\rho_{p,li}$, are obtained averaging wood tissue and leaf tissue densities across cohorts:

$$\rho_{p,wo} = \frac{\sum_i w_i \cdot \rho_{wood}(SP_i)}{\sum_i w_i} \quad (24.25)$$

$$\rho_{p,li} = \frac{\sum_i w_i \cdot \rho_{leaf}(SP_i)}{\sum_i w_i} \quad (24.26)$$

$$(24.27)$$

Finally, herb particle density, $\rho_{p,he}$, is set to a default value $\rho_{p,he} = 400kg \cdot m^{-3} = 25lb \cdot ft^{-3}$ (Prichard *et al.* 2013).

24.3.6.2 Particle volume (PV)

Particle volume is defined as the volume of particles per surface area (in $m^3 \cdot m^{-2}$). Is calculated as dry weight loading divided by particle density. If species have different particle density values, the particle volume for canopy (PV_{ca}) and shrub(PV_{sh}) strata can be calculated using:

$$PV_{ca} = \sum_i PV_{i,ca} = \sum_i w_{i,ca}/\rho_p(SP_i) \quad (24.28)$$

$$PV_{sh} = \sum_i PV_{i,sh} = \sum_i w_{i,sh}/\rho_p(SP_i) \quad (24.29)$$

where $PV_{i,ca}$ and $PV_{i,sh}$ are the particle volume of cohort i in the canopy and shrub strata, respectively. The particle volume for woody and herb strata are simply:

$$PV_{wo} = w_{wo}/\rho_{p,wo} \quad (24.30)$$

$$PV_{he} = w_{he}/\rho_{p,he} \quad (24.31)$$

The particle volume for the litter stratum is the sum of particle volume of litter components:

$$PV_{li} = \sum_i PV_{li,k} = \sum_i w_{li,k}/\rho_{p,li} \quad (24.32)$$

24.3.6.3 Packing ratio (β)

The proportion of fuelbed stratum volume occupied by fuel particles is an important factor to predict fire behavior. At low packing ratios (low particle density) fire intensity is limited by excessive heat loss. At high packing ratios (high particle density), lack of oxygen limits combustion. The packing ratios for the canopy and shrub stratum (β_{ca} and β_{sh} ; dimensionless) are given by:

$$\beta_{ca} = \frac{PV_{ca}}{\delta_{ca}} \quad (24.33)$$

$$\beta_{sh} = \frac{PV_{sh}}{\delta_{sh}} \quad (24.34)$$

where $w_{i,ca}$ and $w_{i,sh}$ are the contribution of cohort i to canopy and shrub strata loading (in $kg \cdot m^{-2}$), respectively, and $\rho_p(SP_i)$ is the particle density (in $kg \cdot m^{-3}$) of fuels in cohort i . The packing ratio for the herbaceous, woody and litter strata are:

$$\beta_{he} = \frac{PV_{he}}{\delta_{he}} \quad (24.35)$$

$$\beta_{wo} = \frac{PV_{wo}}{\delta_{wo}} = \frac{\rho_{b,wo}}{\rho_{p,wo}} \quad (24.36)$$

$$\beta_{li} = \frac{PV_{li}}{\delta_{li}} = \frac{\rho_{b,li}}{\rho_{p,li}} \quad (24.37)$$

Note that the packing ratio expressions for woody and litter strata as a ratio of bulk and particle density arises as a consequence of how fuel depth and particle volume are estimated.

24.3.6.4 Surface-area-to-volume ratio (σ)

The surface-area-to-volume ratio (in $m^2 \cdot m^{-3}$) for the canopy or shrub strata are calculated using weighted averages:

$$\sigma_{ca} = \frac{\sum_i w_{i,ca} \cdot \sigma(SP_i)}{\sum_i w_{i,ca}} \quad (24.38)$$

$$\sigma_{sh} = \frac{\sum_i w_{i,sh} \cdot \sigma(SP_i)}{\sum_i w_{i,sh}} \quad (24.39)$$

where $w_{i,ca}$ and $w_{i,sh}$ are the contribution of cohort i to canopy and shrub strata loading (in $kg \cdot m^{-2}$), respectively, and $\sigma(SP_i)$ is the species-specific surface-area-to-volume ratio. The surface-area-to-volume ratio of herbs is assumed constant $\sigma_{he} = 11483m^2 \cdot m^{-3} = 3500ft^2 \cdot ft^{-3}$ and that of small (1-h) woody fuels is $\sigma_{wo} = 1601.05m^2 \cdot m^{-3} = 488ft^2 \cdot ft^{-3}$. The surface-area-to-volume ratio for the litter stratum is:

$$\sigma_{li} = \frac{\sum_k w_{li,k} \cdot \sigma_k}{\sum_k w_{li,k}} \quad (24.40)$$

and the surface-area-to-volume ratio for litter types are:

$$\sigma_{shortneedlepine} = 6562m^2 \cdot m^{-3} = 2000ft^2 \cdot ft^{-3} \quad (24.41)$$

$$\sigma_{longneedlepine} = 4921m^2 \cdot m^{-3} = 1500ft^2 \cdot ft^{-3} \quad (24.42)$$

$$\sigma_{otherconifer} = 8202m^2 \cdot m^{-3} = 2500ft^2 \cdot ft^{-3} \quad (24.43)$$

$$\sigma_{hardwood} = 8202m^2 \cdot m^{-3} = 2500ft^2 \cdot ft^{-3} \quad (24.44)$$

24.3.6.5 Fuel area index (FAI)

The fuel area index (FAI) is the total fuel surface area per unit of ground area (unitless). It is analogous to leave area index (LAI), and it is used to calculate FCCS fire potentials (Schaaf *et al.* 2007). For shrub and canopy strata, FAI is calculated as:

$$FAI_{ca} = \sum_i FAI_{i,ca} = \sum_i PV_{i,ca} \cdot \sigma(SP_i) \quad (24.45)$$

$$FAI_{sh} = \sum_i FAI_{i,sh} = \sum_i PV_{i,sh} \cdot \sigma(SP_i) \quad (24.46)$$

where $FAI_{i,ca}$ and $FAI_{i,sh}$ are the FAI of cohort i in the canopy and shrub strata, respectively. The FAI of herbs and woody strata are given by:

$$FAI_{he} = PV_{he} \cdot \sigma_{he} \quad (24.47)$$

$$FAI_{wo} = PV_{wo} \cdot \sigma_{wo} \quad (24.48)$$

For the litter layer, FAI is calculated as a sum of FAI for litter components:

$$FAI_{li} = \sum_k FAI_{li,k} = \sum_k PV_{li,k} \cdot \sigma_k \quad (24.49)$$

24.3.6.6 Moisture content (FMC)

Fuel moisture content (FMC in percent of dry weight) is averaged across cohorts composing the shrub or canopy strata, to obtain $FMC_{live,sh}$ and $FMC_{live,ca}$:

$$FMC_{live,sh} = \frac{\sum_i w_{i,sh} \cdot FMC_i}{\sum_i w_{i,sh}} \quad (24.50)$$

$$FMC_{live,ca} = \frac{\sum_i w_{i,ca} \cdot FMC_i}{\sum_i w_{i,ca}} \quad (24.51)$$

Live fuel moisture of herb stratum ($FMC_{live,he}$) is an input. The moisture of dead plant in the canopy and shrub layers ($FMC_{dead,ca}$ and $FMC_{dead,sh}$), the moisture of dead herbs ($FMC_{dead,he}$), as well as that of litter (FMC_{li}) and woody (FMC_{wo}) strata are all assumed equal to the moisture of 1-h dead fuels, which is an input of the model.

24.3.6.7 Proportion of dead fuel (P_{dead})

Woody and litter strata are dead fuels, but for canopy, shrub and herb strata the proportion of fuels that are dead are variable. The proportion of dead fuels in the herbaceous stratum ($P_{dead,he}$) is an input of the model, but for the shrub and canopy strata these are calculated from the proportion of dead fuels in each cohort:

$$P_{dead,sh} = \frac{\sum_i w_{i,sh} \cdot P_{dead,i}}{\sum_i w_{i,sh}} \quad (24.52)$$

$$P_{dead,ca} = \frac{\sum_i w_{i,ca} \cdot P_{dead,i}}{\sum_i w_{i,ca}} \quad (24.53)$$

24.3.6.8 Low heat content (h)

The low fuel heat content of each surface fuel stratum (in $\text{kJ} \cdot \text{kg}^{-1}$) is used for the calculation of reaction intensity. Heat content values are adjusted for live foliar moisture content in canopy, shrub and herb strata; and are left to the default value for woody and litter strata:

$$h_{ca} = h_{ca,def} - (M_{live,ca}/100) \cdot V \quad (24.54)$$

$$h_{sh} = h_{sh,def} - (M_{live,sh}/100) \cdot V \quad (24.55)$$

$$h_{he} = h_{def} - (M_{live,he}/100) \cdot V \quad (24.56)$$

$$h_{wo} = h_{li} = h_{def} \quad (24.57)$$

where $h_{def} = 18608 \text{ kJ} \cdot \text{kg}^{-1} = 8000 \text{ Btu} \cdot \text{lb}^{-1}$ is the default low heat content value for herbs, woody and litter strata, and $V = 2596 \text{ kJ} \cdot \text{kg}^{-1} = 1116 \text{ Btu} \cdot \text{lb}$ is the latent heat of vaporisation of water. The default low heat of contents for the canopy and shrub strata ($h_{ca,def}$ and $h_{sh,def}$) are calculated as a weighted average across cohorts:

$$h_{ca,def} = \frac{\sum_i w_{i,ca} \cdot h(SP_i)}{\sum_i w_{i,ca}} \quad (24.58)$$

$$h_{sh,def} = \frac{\sum_i w_{i,sh} \cdot h(SP_i)}{\sum_i w_{i,sh}} \quad (24.59)$$

where $h(SP_i)$ is a species-specific low heat content value.

24.3.6.9 Reactive volume (RV)

The volume per surface unit ($\text{m}^3 \cdot \text{m}^{-2}$) that would be involved in flaming combustion.

$$RV_{sh} = w_{shrub}/\rho_{p,sh} \quad (24.60)$$

$$RV_{he} = w_{he}/\rho_{p,he} \quad (24.61)$$

$$RV_{wo} = w_{wo}/\rho_{p,wo} \quad (24.62)$$

$$RV_{li} = \min(w_{li}, w_{max,li})/\rho_{p,li} \quad (24.63)$$

In the case of litter, the flame loading is limited by $w_{max,li}$, the maximum loading that would be consumed in the flaming stage of combustion, calculated as a weighted average of litter types:

$$w_{max,li} = \frac{\sum_k w_{max,k} \cdot w_{li,k}}{\sum_k w_{li,k}} \quad (24.64)$$

where k indicates litter type. The maximum combustion loadings for litter types are [Prichard *et al.* (2013); Table 2]:

$$w_{max,shortneedlepine} = w_{max,otherconifer} = 0.3248 \text{ kg} \cdot \text{m}^{-2} = 2900 \text{ lb} \cdot \text{ac}^{-1} \quad (24.65)$$

$$w_{max,longneedlepine} = 0.6496 \text{ kg} \cdot \text{m}^{-2} = 5800 \text{ lb} \cdot \text{ac}^{-1} \quad (24.66)$$

$$w_{max,hardwood} = 0.3472 \text{ kg} \cdot \text{m}^{-2} = 3100 \text{ lb} \cdot \text{ac}^{-1} \quad (24.67)$$

24.3.7 Unit conversion of fuel characteristics

FCCS calculations employ empirical equations that were derived in British units system. Hence, all the fuel characteristics and model inputs that are in metric units have to be translated into British units prior to fire behaviour calculations:

- Loading: $1kg \cdot m^{-2} = 0.204918lb \cdot ft^{-2}$
- Depths: $1m = 3.2808399ft$
- Particle density and bulk density: $1kg \cdot m^{-3} = 0.06242796lb \cdot ft^{-3}$
- Particle volume and reactive volume: $1m^3 \cdot m^{-2} = 3.2808399ft^3 \cdot ft^{-2}$
- Surface-to-area-volume ratio: $1m^2 \cdot m^{-3} = 0.3048ft^2 \cdot ft^{-3}$
- Heat content: $1kJ \cdot kg^{-1} = 0.429922614Btu \cdot lb^{-1}$
- Wind speed: $1m \cdot s^{-1} = 2.23693629mph$

24.4 Surface fire behavior

24.4.1 Surface rate of spread (R)

In the Rothermel (1972) model, surface rate of spread is defined as the ratio of heat source (i.e. the surface fire energy propagated to unburned fuels) to surface fuel heat sink (i.e. the energy required to preheat fuels). Owing to the difference in packing ratio between the litter stratum and the other surface fuels, litter-dominated fuelbeds may have substantially different spread rates than other fuelbeds. For this reason, in FCCS the rate of spread (in $ft \cdot min^{-1}$) is calculated separately for litter stratum and the final rate of spread is the maximum of the rate of spread of all surface fuels and that of the litter stratum. Rate of spread is also limited to a maximum based in windspeed and slope.

$$R = \min(WindSlopeCap, \max(R_{surf}, R_{litter})) \quad (24.68)$$

The surface fuel and litter fuel rates of spread are given by the application of Rothermel's (1972) equation to each case:

$$R_{surf} = \frac{I_{R,surf} \cdot \xi_{surf} \cdot (1 + \phi_W + \phi_S)}{q_{surf}} \quad (24.69)$$

$$R_{litter} = \frac{I_{R,litter} \cdot \xi_{litter} \cdot (1 + \phi_W + \phi_S)}{q_{litter}} \quad (24.70)$$

where $I_{R,surf}$ and $I_{R,litter}$ are the reaction intensities (in $Btu \cdot ft^{-2} \cdot min^{-1}$), ξ_{surf} and ξ_{litter} are the propagating flux ratios, q_{surf} and q_{litter} are the heat sinks. Finally, ϕ_W and ϕ_S are the slope and wind modifiers. All of them are explained in the following sections. The maximum rate of spread calculated from windspeed and slope is:

$$WindSlopeCap = 88 \cdot U \cdot (1 + \phi_S) \quad (24.71)$$

where U is windspeed (in mph) and 88 is a conversion factor (from mph to ft/min).

24.4.1.1 Reaction intensity (I_R)

Reaction intensity of surface fuels (in $Btu \cdot ft^{-2} \cdot min^{-1}$) is calculated as the sum of component reaction intensities of the four different surface fuel strata, whereas the reaction intensity in the litter uses this strata alone:

$$I_{R,surf} = I_{R,sh} + I_{R,he} + I_{R,wo} + I_{R,li} \quad (24.72)$$

$$I_{R,litter} = I_{R,li} \quad (24.73)$$

Each component reaction intensity is calculated using:

$$I_{R,sh} = (\eta_{\beta'_{allsurf}})^{A_{sh}} \cdot \Gamma'_{max,sh} \cdot w_{sh} \cdot h_{sh} \cdot \eta_{FMC,sh} \cdot \eta_{K,sh} \quad (24.74)$$

$$I_{R,he} = (\eta_{\beta'_{lowsurf}})^{A_{he}} \cdot \Gamma'_{max,he} \cdot w_{he} \cdot h_{he} \cdot \eta_{FMC,he} \cdot \eta_{K,he} \quad (24.75)$$

$$I_{R,wo} = (\eta_{\beta'_{lowsurf}})^{A_{wo}} \cdot \Gamma'_{max,wo} \cdot w_{wo} \cdot h_{wo} \cdot \eta_{FMC,wo} \cdot \eta_{K,wo} \quad (24.76)$$

$$I_{R,li} = (\eta_{\beta'_{litter}})^{A_{li}} \cdot \Gamma'_{max,li} \cdot w_{li} \cdot h_{li} \cdot \eta_{FMC,li} \cdot \eta_{K,li} \quad (24.77)$$

In the above equations, w_{sh} , w_{he} , w_{wo} and w_{li} are the loadings of the corresponding shrub, herb, woody and litter strata, respectively. These quantities were defined in previous sections, as were the corresponding low heat fuel contents (h_{sh} , h_{he} , h_{wo} and h_{li}). Mineral damping coefficient (η_K ; dimensionless) is set to the same value (corresponding to the conventional value for silica-free ash content of 1%) for all strata:

$$\eta_{K,sh} = \eta_{K,he} = \eta_{K,wo} = \eta_{K,li} = 0.42 \quad (24.78)$$

In the following subsections, we describe the calculation of the remaining variables for each stratum: reaction efficiency ($\eta_{\beta'}$), Rothermel's A parameter, maximum reaction velocity (Γ'_{max}) and moisture damping coefficient (η_{FMC}).

24.4.1.2 Reaction efficiency ($\eta_{\beta'}$)

Reaction efficiency (between 0 and 1) represents the damping effect of inefficiently packed fuels in the reaction intensity. Because shrubs rarely burn without lower surface fuels, the reaction efficiency of the surface layer ($\eta_{\beta'_{allsurf}}$) includes shrubs, herbs and woody fuels. Low surface fuels may carry flames without involving shrubs, so are assumed to burn with a single reaction efficiency ($\eta_{\beta'_{lowsurf}}$) determined by the combined characteristics of herb and woody fuel strata. Both are calculated similarly:

$$\eta_{\beta'_{allsurf}} = \beta'_{allsurf} \cdot e^{1-\beta'_{allsurf}} \quad (24.79)$$

$$\eta_{\beta'_{lowsurf}} = \beta'_{lowsurf} \cdot e^{1-\beta'_{lowsurf}} \quad (24.80)$$

where $\beta'_{allsurf}$ and $\beta'_{lowsurf}$ are the relative packing ratios corresponding to all surface fuels and low surface fuels, respectively. Relative packing ratios (β' ; dimensionless) are defined as the ratio of optimum depth (δ_{opt}) to effective depth (δ_{eff}):

$$\beta'_{allsurf} = \delta_{opt,allsurf} / \delta_{eff,allsurf} \quad (24.81)$$

$$\beta'_{lowsurf} = \delta_{opt,lowsurf} / \delta_{eff,lowsurf} \quad (24.82)$$

Optimum depth is the depth (in ft) at which fuels are optimally packed for maximum reaction intensity:

$$\delta_{opt,allsurf} = PV_{allsurf} + OptAirVol_{allsurf} \quad (24.83)$$

$$\delta_{opt,lowsurf} = PV_{lowsurf} + OptAirVol_{lowsurf} \quad (24.84)$$

where $PV_{allsurf}$ and $PV_{lowsurf}$ are the volume of particles (in $ft^3 \cdot ft^{-2}$) for all surface fuels and low surface fuels, respectively, given by:

$$PV_{allsurf} = PV_{sh} + PV_{he} + PV_{wo} \quad (24.85)$$

$$PV_{lowsurf} = PV_{he} + PV_{wo} \quad (24.86)$$

$OptAirVol_{allsurf}$ and $OptAirVol_{lowsurf}$ are the volume of air space (in $ft^3 \cdot ft^{-2}$) between fuel particles that would result in maximum reaction intensity:

$$OptAirVol_{allsurf} = 45 \cdot (RV_{sh} + RV_{he} + RV_{wo}) \quad (24.87)$$

$$OptAirVol_{lowsurf} = 45 \cdot (RV_{he} + RV_{wo}) \quad (24.88)$$

On the other hand, effective depths of all surface fuels and low surface fuels (in ft) are calculated as their depth, weighted by the reactive volume (and percentage cover in FCCS):

$$\delta_{eff,allsurf} = \frac{(RV_{sh} \cdot \delta_{sh}) + (RV_{he} \cdot \delta_{he}) + (RV_{wo} \cdot \delta_{wo})}{RV_{sh} + RV_{he} + RV_{wo}} \quad (24.89)$$

$$\delta_{eff,lowsurf} = \frac{(RV_{he} \cdot \delta_{he}) + (RV_{wo} \cdot \delta_{wo})}{RV_{he} + RV_{wo}} \quad (24.90)$$

Reaction efficiency of the litter stratum is determined separately from the other strata. It is defined as the average of reaction efficiency across litter types, calculated using loadings as weights:

$$\eta_{\beta'_{litter}} = \frac{\sum_k \eta_{\beta'_k} \cdot w_{li,k}}{\sum_k w_{li,k}} \quad (24.91)$$

where k indicates litter type. The reaction efficiencies of litter types are [Prichard *et al.* (2013); Table 2]:

$$\eta_{\beta'_{shortneedlepine}} = \eta_{\beta'_{otherconifer}} = 0.18 \quad (24.92)$$

$$\eta_{\beta'_{longneedlepine}} = 0.27 \quad (24.93)$$

$$\eta_{\beta'_{hardwood}} = 0.11 \quad (24.94)$$

24.4.1.3 Rothermel's A

A dimensionless coefficient that modifies reaction's efficiency (eq. (24.80)) to account for lower sensitivity of reaction efficiency to relative packing ratio in flash fuels:

$$A_{wo} = A_{li} = 1.0 \quad (24.95)$$

$$A_{sh} = 133 \cdot \sigma_{sh}^{-0.7913} \quad (24.96)$$

$$A_{he} = 133 \cdot \sigma_{he}^{-0.7913} \quad (24.97)$$

where σ_{sh} and σ_{he} have to be expressed in $ft^2 \cdot ft^{-3}$; Values 133 and -0.7913 are empirical constants (Rothermel 1972).

24.4.1.4 Maximum reaction velocity (Γ'_{max})

The reaction velocity (in min^{-1}) that would exist at optimum fuelbed depth with no fuel moisture or mineral content.

$$\Gamma'_{max,sh} = 9.495 \cdot \frac{\sigma_{sh}}{\sigma_{wo}} \quad (24.98)$$

$$\Gamma'_{max,he} = 9.495 \cdot \frac{\sigma_{he}}{\sigma_{wo}} \quad (24.99)$$

$$\Gamma'_{max,wo} = 9.495 \quad (24.100)$$

$$\Gamma'_{max,li} = 15 \quad (24.101)$$

where $\sigma_{wo} = 488 ft^2 \cdot ft^{-3} = 1601.05 m^2 \cdot m^{-3}$ is the surface-to-area-volume ratio typical of small woody fuels. In Prichard *et al.* (2013) σ_{sh} is defined as the average of shrub foliar surface-to-area-volume ratio and σ_{wo} , but in our case $\sigma_{(SP_i)}$ for each species includes both leaves and small branches. Eq. (24.101) represent a significant departure from Rothermel (1972) maximum reaction velocity, and are also different from Sandberg *et al.* (2007).

24.4.1.5 Moisture damping coefficient (η_{FMC})

Moisture damping reduces reaction velocity and hence reaction intensity (eq. (24.73)). It is calculated for each stratum using the following regression equations:

$$\eta_{FMC, live, sh} = \left[1 - 2.59 \cdot \left(\frac{FMC_{live, sh}}{X_{live, sh}} \right) \right] + \left[5.11 \cdot \left(\frac{FMC_{live, sh}}{X_{live, sh}} \right)^2 \right] - \left[3.52 \cdot \left(\frac{FMC_{live, sh}}{X_{live, sh}} \right)^3 \right]^{24} 102 \quad (24.102)$$

$$\eta_{FMC, dead, sh} = \left[1 - 2.59 \cdot \left(\frac{FMC_{dead, sh}}{X_{dead, sh}} \right) \right] + \left[5.11 \cdot \left(\frac{FMC_{dead, sh}}{X_{dead, sh}} \right)^2 \right] - \left[3.52 \cdot \left(\frac{FMC_{dead, sh}}{X_{dead, sh}} \right)^3 \right]^{24} 103 \quad (24.103)$$

$$\eta_{FMC, live, he} = \left[1 - 2.59 \cdot \left(\frac{FMC_{live, he}}{X_{live, he}} \right) \right] + \left[5.11 \cdot \left(\frac{FMC_{live, he}}{X_{live, he}} \right)^2 \right] - \left[3.52 \cdot \left(\frac{FMC_{live, he}}{X_{live, he}} \right)^3 \right]^{24} 104 \quad (24.104)$$

$$\eta_{FMC, dead, he} = \left[1 - 2.59 \cdot \left(\frac{FMC_{dead, he}}{X_{dead, he}} \right) \right] + \left[5.11 \cdot \left(\frac{FMC_{dead, he}}{X_{dead, he}} \right)^2 \right] - \left[3.52 \cdot \left(\frac{FMC_{dead, he}}{X_{dead, he}} \right)^3 \right]^{24} 105 \quad (24.105)$$

$$\eta_{FMC, wo} = \left[1 - 2.59 \cdot \left(\frac{FMC_{wo}}{X_{wo}} \right) \right] + \left[5.11 \cdot \left(\frac{FMC_{wo}}{X_{wo}} \right)^2 \right] - \left[3.52 \cdot \left(\frac{FMC_{wo}}{X_{wo}} \right)^3 \right] \quad (24.106)$$

$$\eta_{FMC, li} = \left[1 - 2.59 \cdot \left(\frac{FMC_{li}}{X_{li}} \right) \right] + \left[5.11 \cdot \left(\frac{FMC_{li}}{X_{li}} \right)^2 \right] - \left[3.52 \cdot \left(\frac{FMC_{li}}{X_{li}} \right)^3 \right] \quad (24.107)$$

where moisture contents of extinctions were arbitrarily set to $X_{dead, sh} = X_{dead, he} = X_{wo} = X_{li} = 25$, $X_{live, sh} = 180$ and $X_{live, he} = 120$ in Sandberg et al. (2007). As it can be seen in the equations above, in the case of shrub and herb strata, moisture damping of live and dead fuels are differentiated. Average values are found after accounting for the proportion of live and dead material:

$$\eta_{FMC, sh} = \eta_{FMC, live, sh} \cdot (1 - P_{dead, sh}) + \eta_{FMC, dead, sh} \cdot P_{dead, sh} \quad (24.108)$$

$$\eta_{FMC, he} = \eta_{FMC, live, he} \cdot (1 - P_{dead, he}) + \eta_{FMC, dead, he} \cdot P_{dead, he} \quad (24.109)$$

24.4.1.6 Propagating flux ratio (ξ)

The propagating flux ratio (dimensionless) is the proportion of the reaction intensity (eq. (24.73)) that contributes to the forward rate of spread, estimated using an empirical regression:

$$\xi_{surf} = 0.03 + 2.5 \cdot \min \left[0.06, \frac{RV_{sh} + RV_{he} + RV_{wo} + RV_{li}}{\delta_{surfheatssink}} \right] \quad (24.110)$$

$$\xi_{litter} = 0.03 + 2.5 \cdot \min \left[0.06, \frac{RV_{li}}{\delta_{li}} \right] \quad (24.111)$$

where $\delta_{surfheatssink}$ is the depth of surface heat sink (in ft), which in Prichard et al. (2013) is calculated as the sum of strata depths weighted by their relative cover. In our case we weighted stratum depths as in the calculation of effective depth ($\delta_{eff, allsurf}$), but considering all four strata:

$$\delta_{surfheatssink} = \frac{(RV_{sh} \cdot \delta_{sh}) + (RV_{he} \cdot \delta_{he}) + (RV_{wo} \cdot \delta_{wo}) + (RV_{li} \cdot \delta_{li})}{RV_{sh} + RV_{he} + RV_{wo} + RV_{li}} \quad (24.112)$$

24.4.2 Heat sink (q)

Like reaction intensity, the heat sink term (in $Btu \cdot ft^{-3}$) of the rate of spread equation is calculated in FCCS for each fuel stratum and then summed:

$$q_{surf} = q_{sh} + q_{he} + q_{wo} + q_{li} \quad (24.113)$$

$$q_{litter} = q_{li} \quad (24.114)$$

where the heat sink for each stratum is:

$$q_{sh} = \eta_{\beta'_{surf}} \cdot \frac{RV_{sh} \cdot \rho_{p,sh} \cdot Qig_{sh}}{\min(\delta_{sh}, 1ft)} \quad (24.115)$$

$$q_{he} = \eta_{\beta'_{lowsurf}} \cdot \frac{RV_{he} \cdot \rho_{p,he} \cdot Qig_{he}}{\min(\delta_{he}, 1ft)} \quad (24.116)$$

$$q_{wo} = \eta_{\beta'_{lowsurf}} \cdot \frac{RV_{wo} \cdot \rho_{p,wo} \cdot Qig_{wo}}{\min(\delta_{wo}, 1ft)} \quad (24.117)$$

$$q_{li} = \eta_{\beta'_{li}} \cdot \frac{RV_{li} \cdot \rho_{p,li} \cdot Qig_{li}}{\min(\delta_{li}, 1ft)} \quad (24.118)$$

Where $\rho_{p,sh}$, $\rho_{p,he}$, $\rho_{p,wo}$ and $\rho_{p,li}$ are the particle densities (in $lb \cdot ft^{-3}$) of each fuel stratum; and RV_{sh} , RV_{he} , RV_{wo} , and RV_{li} are the reactive volumes of each fuel stratum. Unlike in Sandberg et al. (2007), the calculated heat sink is corrected by the reaction-efficiency term ($\eta_{\beta'_{surf}}$, $\eta_{\beta'_{lowsurf}}$ or $\eta_{\beta'_{li}}$), and the effective depth of each stratum included is limited to 1ft, based on the assumption that it is not necessary to preheat more than one 1ft of depth within a stratum to achieve ignition.

Heat of pre-ignition (Qig ; in $Btu \cdot lb^{-1}$) is the amount of heat required to ignite 1lb of fuel. It is calculated by stratum as a weighted average of live and dead fuels in shrubs and herbs.

$$Qig_{sh} = Qig_{live,sh} \cdot (1 - P_{dead,sh}) + Qig_{dead,sh} \cdot P_{dead,sh} \quad (24.119)$$

$$Qig_{he} = Qig_{live,he} \cdot (1 - P_{dead,he}) + Qig_{dead,he} \cdot P_{dead,he} \quad (24.120)$$

Whereas $Qig_{live,sh}$ and $Qig_{live,he}$ are corrected by fuel moisture, $Qig_{dead,sh}$, $Qig_{dead,he}$ and the other strata (Qig_{wo} and Qig_{li}) are assumed a constant value:

$$Qig_{live,sh} = 250 + (V \cdot (M_{live,sh}/100)) \quad (24.121)$$

$$Qig_{live,he} = 250 + (V \cdot (M_{live,he}/100)) \quad (24.122)$$

$$Qig_{dead,sh} = Qig_{dead,he} = Qig_{wo} = Qig_{li} = 250 \quad (24.123)$$

where $250Btu/lb$ is the heat of preignition of dry cellulose and $V = 1116Btu/lb$ is the latent heat of vaporization.

24.4.3 Wind and slope coefficients (ϕ_W and ϕ_S)

Wind and slope coefficients modify the heat source term of the rate of spread equation. Owing to differences in fuel characteristics and boundary conditions between the litter stratum and other surface fuel strata, in FCCS wind and slope coefficients are calculated separately for the litter stratum. The wind and slope coefficients terms in the rate of spread equation are a weighted average of litter and surface wind and slope coefficients using the relative contribution to reaction intensity as weights:

$$\phi_W = (1 - I_{R,litter}/I_{R,surf}) \cdot \phi_{W,surf} + (I_{R,litter}/I_{R,surf}) \cdot \phi_{W,litter} \quad (24.124)$$

$$\phi_S = (1 - I_{R,litter}/I_{R,surf}) \cdot \phi_{S,surf} + (I_{R,litter}/I_{R,surf}) \cdot \phi_{S,litter} \quad (24.125)$$

Wind coefficients are calculated using:

$$\phi_{W,surf} = 8.8 \cdot \beta'_{surf}^{-E} \cdot (U/BMU)^B \quad (24.126)$$

$$\phi_{W,litter} = 8.8 \cdot \beta_{litter}^{-E} \cdot (U/BMU)^B \quad (24.127)$$

where U is the input midflame windspeed (in $ft \cdot min^{-1}$), $BMU = 352ft \cdot min^{-1}$ is the benchmark midflame windspeed, β'_{surf} is the relative packing ratio (eq. 24.82), B is the exponential response of wind coefficient to windspeed ($B = 1.2$ in Sandberg et al. (2007)), and E is the exponential term representing the mild effect of large fuels in reducing the accelerating effect of wind on fire spread by attenuating wind flow, given by:

$$E = 0.55 - 0.2 \cdot \frac{FAI_{sh} + FAI_{he}}{FAI_{sh} + FAI_{he} + FAI_{wo}} \quad (24.128)$$

E is assumed to be the same for both all surface fuels and litter fuels.

Slope coefficients are calculated using the empirical equation of Rothermel (1972), applied to all surface fuels and litter fuels:

$$\phi_{S,surf} = 5.275 \cdot (S/100)^2 \cdot (\beta_{sh} + \beta_{he} + \beta_{wo})^{-0.3} \quad (24.129)$$

$$\phi_{S,litter} = 5.275 \cdot (S/100)^2 \cdot \beta_{li}^{-0.3} \quad (24.130)$$

where S is the slope (in percent) and β is the packing ratio (not relative!) of fuels.

24.4.4 Fireline intensity (I_B) and flame length (FL)

Byram's fireline intensity (I_B) is the rate of heat release per unit of fire edge (in $Btu \cdot ft^{-1} \cdot min^{-1}$), and in FCCS is calculated as (Albini 1976):

$$I_B = I_{R,surf} \cdot (R \cdot t_R) \quad (24.131)$$

where $I_{R,surf}$ is the surface reaction intensity, R is the rate of spread and t_R is the flame residence time, which is defined as the time (in min) fuels contribute to propagating flux and is estimated as Albini (1976):

$$t_R = 192 \cdot \frac{(I_{R,sh} \cdot RT_{sh}) + (I_{R,he} \cdot RT_{he}) + (I_{R,wo} \cdot RT_{wo}) + (I_{R,li} \cdot RT_{li})}{I_{R,surf}} \quad (24.132)$$

where RT is the reaction thickness, the approximate thickness (in ft) of a fuel element shell that contributes to reaction intensity. In FCCS, reaction thickness is estimated as $RT = 0.0028ft$ for thermally thick fuel elements (Sandberg *et al.* 2007). When the diameter of a fuel element is less than twice the reaction thickness, the entire fuel element contributes to reaction intensity. Reaction thickness values for each stratum are given by:

$$RT_{sh} = \min(0.0028, 2/\sigma_{sh}) \quad (24.133)$$

$$RT_{he} = \min(0.0028, 2/\sigma_{he}) \quad (24.134)$$

$$RT_{wo} = \min(0.0028, 2/\sigma_{wo}) \quad (24.135)$$

$$RT_{li} = \min(0.0028, 2/\sigma_{li}) \quad (24.136)$$

Flame length is defined as the distance (in ft) between the flame tip and the midpoint of the flame depth at the base of the flame, and is calculated (Byram 1959):

$$FL = 0.45 \cdot (I_B/60)^{0.46} \quad (24.137)$$

where 60.0 is a factor to convert from $Btu \cdot ft^{-1} \cdot min^{-1}$ to $Btu \cdot ft^{-1} \cdot s^{-1}$.

24.5 Crown fire behavior

Crown fire behavior is difficult to model and actual rates of spread are not possible to predict. Here we mainly follow the approach given in FCCS (Prichard *et al.* 2013), although in our case the canopy is not subdivided into layers (overstory, midstory and understory).

24.5.1 Crown fire rate of spread (R_{crown})

The rate of spread of crown fires is estimated by using a modification of Rothermel's equation:

$$R_{crown} = \frac{I_{R,crown} \cdot \xi_{crown} \cdot WAF}{q_{crown}} = \frac{(I_{R,surf} + I_{R,ca}) \cdot \xi_{crown} \cdot WAF}{q_{surf} + q_{ca}} \quad (24.138)$$

where $I_{R,surf}$ is the surface reaction intensity, $I_{R,ca}$ is the canopy reaction intensity, ξ_{crown} is the propagating flux ratio in the canopy, q_{surf} is the surface heat sink and q_{ca} is the canopy heat sink. Note that reaction intensities and heat sinks of canopy and surface fuels are added for the application of Rothermel's equation. Other modifications include the exclusion of slope effects and the consideration of wind effects through a wind adjustment factor (WAF).

Crown propagating flux ratio (ξ_{crown} ; in $Btu \cdot ft^{-3}$) represents the proportion of the crown reaction intensity that contributes to crown fire's forward rate of spread:

$$\xi_{crown} = 1 - e^{(-\frac{FAI_{ca}}{4 \cdot \delta_{ca}})} \quad (24.139)$$

where FAI_{ca} is the fuel area index of the canopy, and δ_{ca} is the canopy depth (in ft). Wind adjustment factor (WAF) is defined as:

$$WAF = \frac{U/\sqrt{U^2 + VS^2}}{BMU/\sqrt{BMU^2 + VS^2}} \quad (24.140)$$

where U is the input (midflame) windspeed (in $ft \cdot min^{-1}$), $BMU = 352 ft \cdot min^{-1}$ is the benchmark windspeed and $VS = 900 ft \cdot min^{-1}$ is the vertical stack velocity.

The following two subsections detail the calculation of canopy reaction intensity ($I_{R,ca}$) and canopy heat sink (q_{ca}).

24.5.2 Canopy reaction intensity ($I_{R,ca}$)

Reaction intensity of canopy fuels (in $Btu \cdot ft^{-2} \cdot min^{-1}$) is estimated as:

$$I_{R,ca} = (\eta_{\beta'_{ca}})^{A_{ca}} \cdot \Gamma'_{max,ca} \cdot w_{ca} \cdot h_{ca} \cdot \eta_{FMC,ca} \cdot \eta_{K,ca} \quad (24.141)$$

where $A_{ca} = 133 \cdot \sigma_{ca}^{-0.7913}$ is Rothermel's A coefficient, $\Gamma'_{max,ca} = 15 min^{-1}$ is the maximum reaction velocity of the canopy, w_{ca} is the loading of canopy fuels (in $lb \cdot ft^{-2}$), h_{ca} is the heat content of the canopy fuels (in $Btu \cdot lb^{-1}$) and $\eta_{K,ca} = 0.42$ is the mineral damping coefficient of the canopy stratum. Moisture damping coefficient for the canopy ($\eta_{FMC,ca}$) is estimated as done for shrub and herb strata:

$$\eta_{FMC, live, ca} = \left[1 - 2.59 \cdot \left(\frac{FMC_{live, ca}}{X_{live, ca}} \right) \right] + \left[5.11 \cdot \left(\frac{FMC_{live, ca}}{X_{live, ca}} \right)^2 \right] - \left[3.52 \cdot \left(\frac{FMC_{live, ca}}{X_{live, ca}} \right)^3 \right] \quad (24.142)$$

$$\eta_{FMC, dead, ca} = \left[1 - 2.59 \cdot \left(\frac{FMC_{dead, ca}}{X_{dead, ca}} \right) \right] + \left[5.11 \cdot \left(\frac{FMC_{dead, ca}}{X_{dead, ca}} \right)^2 \right] - \left[3.52 \cdot \left(\frac{FMC_{dead, ca}}{X_{dead, ca}} \right)^3 \right] \quad (24.143)$$

$$\eta_{FMC, ca} = \eta_{FMC, live, ca} \cdot (1 - P_{dead, ca}) + \eta_{M, dead, ca} \cdot P_{dead, ca} \quad (24.144)$$

where moisture contents of extinctions were arbitrarily set to $X_{dead, ca} = 25$ and $X_{live, ca} = 180$.

The reaction efficiency in the canopy ($\eta_{\beta'_{canopy}}$) represents the damping effect of inefficiently packed fuels in the canopy:

$$\eta_{\beta'_{canopy}} = \beta'_{canopy} \cdot e^{1 - \beta'_{canopy}} \quad (24.145)$$

where β'_{canopy} is the relative packing ratio in the canopy:

$$\beta'_{canopy} = \delta_{opt, canopy} / \delta_{eff, canopy} \quad (24.146)$$

where the effective depth is $\delta_{eff, canopy} = \delta_{ca}$ (in ft) and the optimum canopy depth is calculated using:

$$\delta_{opt, canopy} = 0.4 \cdot FAI_{ca} + \beta_{ca} \cdot (\delta_{ca} \cdot C_{ca} / 100) \quad (24.147)$$

where C_{ca} is the percent cover of the canopy, FAI_{ca} is the fuel area index of the canopy, β_{ca} is the packing ratio of canopy fuels and δ_{ca} is the canopy depth (in ft).

24.5.3 Canopy heat sink (q_{ca})

Canopy heat sink (in $Btu \cdot ft^{-3}$) is estimated using:

$$q_{ca} = \frac{0.5 \cdot FAI_{ca} \cdot RT_{ca} \cdot \rho_{p,ca} \cdot Qig_{ca}}{(C_{ca}/100) \cdot \delta_{ca}} \quad (24.148)$$

where C_{ca} is the percent cover of the canopy, FAI_{ca} is the fuel area index of the canopy, $RT_{ca} = \min(0.0028, 2/\sigma_{ca})$ is the reaction thickness of the canopy stratum (in ft), $\rho_{p,ca}$ is the particle density of the canopy (in $lb \cdot ft^{-3}$), δ_{ca} is the canopy depth (in ft) and Qig_{ca} is the heat of pre-ignition of the canopy stratum (in $Btu \cdot lb^{-1}$), which is calculated as a weighted average of live and dead fuels:

$$Qig_{live,ca} = 250 + (V \cdot (M_{live,ca}/100)) \quad (24.149)$$

$$Qig_{dead,ca} = 250 \quad (24.150)$$

$$Qig_{ca} = Qig_{live,ca} \cdot (1 - P_{dead,ca}) + Qig_{dead,ca} \cdot P_{dead,ca} \quad (24.151)$$

where $250Btu/lb$ is the heat of preignition of dry cellulose and $V = 1116Btu/lb$ is the latent heat of vaporization of water.

24.5.4 Fireline intensity ($I_{B,crown}$) and flame length (FL_{crown})

Byram's fireline intensity for crown fires is estimated using:

$$I_{B,crown} = I_{R,crown} \cdot (R_{crown} \cdot t_{R,crown}) \quad (24.152)$$

where $I_{R,crown}$ is the crown reaction intensity (i.e. the sum of canopy and surface reaction intensities), R_{crown} is the rate of crown fire spread and $t_{R,crown}$ is the flame residence time, estimated as:

$$t_R = 192 \cdot RT_{ca} \quad (24.153)$$

where $RT_{ca} = \min(0.0028, 2/\sigma_{ca})$ is the reaction thickness of the canopy. As for surface fires, flame length is calculated using:

$$FL_{crown} = 0.45 \cdot (I_{B,crown}/60)^{0.46} \quad (24.154)$$

where 60.0 is a factor to convert from $Btu \cdot ft^{-1} \cdot min^{-1}$ to $Btu \cdot ft^{-1} \cdot s^{-1}$.

24.6 Fire potentials

24.6.1 Surface fire behavior potentials

The **surface fire behavior potential** (SFP ; between 0 and 9) is an index defined as the maximum of spread potential (SP) and flame length (FL) potential indices (both between 0 and 9):

$$SFP = \max(SP, FP) \quad (24.155)$$

Spread potential is derived from R (in $ft \cdot min^{-1}$), and *flame length potential* is derived from FL (in ft), both quantities being calculated at benchmark environmental conditions:

$$SP = \min\left[9, R^{1/2}\right] \quad (24.156)$$

$$FP = \min\left[9, FL^{1/2}\right] \quad (24.157)$$

24.6.2 Crown fire behavior potentials

The *crown fire summary potential (CPF)* combines three subpotentials into a single index value between 0 and 9. It places more emphasis on crown fire initiation (*IC*) and rate of spread (*RC*) than to crown-to-crown transmissivity (*TC*):

$$CPF = 0.4286 \cdot (IC + (TC/3) + RC) \quad (24.158)$$

where 0.4286 is used to limit *CPF* between 0 and 9.

The *crown fire initiation potential (IC)* represents the likelihood of a surface fire torching into single or multiple trees. *IC* is based on the work by Van Wagner. If $FAI_{ca} = 0$ then $IC = 0$. Otherwise it is calculated as:

$$IC = \min \left[9, 4 \cdot \left(\frac{I_B/60}{I_c} \right)^{0.2} \right] \quad (24.159)$$

where I_B is the surface fireline intensity (60 is used to convert it to $Btu \cdot ft^{-1} \cdot s^{-1}$) and I_c is Van Wagner's critical fireline intensity (Scott & Reinhardt 2002):

$$I_c = 0.288894658 \cdot [0.01 \cdot (H_{gap}/100) \cdot (460 + 25.9 \cdot M_{live,ca})]^{1.5} \quad (24.160)$$

where $M_{live,ca}$ is the moisture content of the canopy (in percent of dry weight), 0.288894658 is used to convert from $kJ \cdot m^{-1} \cdot s^{-1}$ to $Btu \cdot ft^{-1} \cdot s^{-1}$ and H_{gap} is the canopy gap (in cm) determined from the analysis of the bulk density profile.

The *crown-to-crown transmittivity potential (TC)* is set to zero if $FAI_{ca} < TFAI/(3 \cdot \pi)$, where *TFAI* is a threshold for FAI calculated as:

$$TFAI = A_q \cdot e^{-0.0019 \cdot U} \quad (24.161)$$

where $A_q = 3.2868$ if $\sigma_{ca} > 2000 ft^2 \cdot ft^{-3}$ and $A_q = 2.6296$ otherwise. If $FAI_{ca} > TFAI/(3 \cdot \pi)$, then *TC* is calculated as:

$$TC = \min [9, 10 \cdot TC_q] \quad (24.162)$$

where TC_q is the efficiency of crown-to-crown heat transfer, as a proportion of maximum efficiency at 100 % canopy cover:

$$TC_q = \frac{(\max(0.0, C_{ca} \cdot WAF - 40))^{0.3}}{(100 \cdot WAF - 40)^{0.3}} \quad (24.163)$$

In this last equation 40 represents the threshold of canopy cover necessary to initiate dependent crown spread, and 0.3 is a coefficient describing the assumed effect of crown cover on transmittivity at benchmark windspeed. The canopy adjustment ratio *WAF* is added to modulate transmittivity depending on windspeed.

Finally, the *crown fire rate of spread potential (RC)* is defined as:

$$RC = \min \left[9, 2.5 \cdot R_{crown}^{1/e} \right] \quad (24.164)$$

where R_{crown} is the rate of spread (in $ft \cdot min^{-1}$) of the crown fire.

24.7 Unit conversion of outputs

The following factors are used to express fire behavior outputs to metric units:

- *Fire spread rates:* $1ft \cdot min^{-1} = 0.3048m \cdot min^{-1}$
- *Flame length:* $1ft = 0.3048m$
- *Reaction intensity:* $1Btu \cdot ft^{-2} \cdot min^{-1} = 11.3484kJ \cdot m^{-2} \cdot min^{-1}$
- *Heat sink:* $1Btu \cdot ft^{-3} = 37.2589458kJ \cdot m^{-3}$
- *Fireline intensity:* $1Btu \cdot ft^{-1} \cdot min^{-1} = 0.0576911555kW \cdot m^{-1}$

Appendix A

Inbuilt parameter estimation

A.1 Introduction

Package **medfate** has been designed to allow simulations requiring a minimum set of vegetation functional parameters. This entails that several other parameters have to be estimated automatically (via inbuilt procedures) before starting simulations. Inbuilt parameter estimation is done in functions `spwbInput()`, `growthInput()`, `forest2spwbInput()` and `forest2growthInput()`, with the user controlling the process through the species parameter table input (e.g., `SpParamsMED`) and the object `control` (see default control values in `defaultControl()`).

A.2 Strict, scaled and imputable parameters

Different kinds of vegetation functional parameters can be distinguished according to whether inbuilt parameter estimation is possible and how it is conducted:

- **Strictly-required parameters** are those for which there are no inbuilt estimation procedures implemented in the initialization functions. Hence, either values in the species parameter table input are non-missing or suitable values need to be specified before running simulation models. Since **medfate ver. 2.3**, only plant/leaf classification parameters and plant size parameters are strict. The remaining ones can be estimated from other parameters. This facilitates having a functional species parameter table, because only a set of parameters have to be strictly filled, from either soft trait databases or forest inventory data.
- **Scaled parameters** are functional parameters that cannot be defined at the species level, because they need to be estimated taking into account the size and structure of the plant cohort. These are not normally defined at the level of species parameter table. Specific `control` parameters are used to determine how scaling is performed.
- **Imputable parameters** parameters are those for which the initialization routines can provide default values or estimations derived from relationships with other parameters. Parameter imputation is conducted if control parameter `fillMissingSpParams = TRUE`. Sometimes, default parameter values are also specified in the `control` object.

The following tables describe how the different functional parameters are dealt with, grouped by function. Links are given to the chapter subsections where scaling and/or imputation procedures are described.

Plant/leaf classification

Symbol	R	Description	Strict	Scaled	Imputable
<i>GF</i>	GrowthForm	Growth form, defined depending on the treatment in forest inventory plots (Tree, Shrub or Tree/Shrub)	Yes	No	No
<i>LF</i>	LifeForm	Raunkiaer life form	Yes	No	No
<i>Lshape</i>	LeafShape	Leaf type (Linear, Needle, Broad, Scale, Spines or Succulent)	Yes	No	No
<i>Lsize</i>	LeafSize	Leaf size (Small, Medium, Large)	Yes	No	No
<i>Lpheno</i>	PhenologyType	Phenology type	Yes	No	No

Plant size

Symbol	R	Description	Strict	Scaled	Imputable
<i>H_{max}</i>	Hmed	Maximum plant height	Yes	No	No
<i>H_{med}</i>	Hmed	Median plant height	Yes	No	No
<i>f_{HD,min}</i>	fHDmin	Minimum height-to-diameter ratio	Yes	No	No
<i>f_{HD,max}</i>	fHDmax	Maximum height-to-diameter ratio	Yes	No	No

Allometric coefficients

Symbol	R	Description	Strict	Scaled	Imputable
<i>a_{ash}, b_{ash}</i>	a_ash, b_ash	Coefficients relating the square of shrub height with shrub area	No	No	A.3.1
<i>a_{bsh}, b_{bsh}</i>	a_bsh, b_bsh	Coefficients relating crown volume with dry weight of shrub individuals	No	No	A.3.1
<i>cr</i>	cr	Ratio between crown length and total height for shrubs	No	No	A.3.1
<i>a_{fbt}, b_{fbt}, c_{fbt}</i>	a_fbt, b_fbt, c_fbt	Coefficients to calculate foliar biomass of an individual tree	No	No	A.3.2
<i>a_{cr}, b_{1cr}, b_{2cr}, b_{3cr}, c_{1cr}, c_{2cr}</i>	a_cr, b_1cr, b_2cr, b_3cr, c_1cr, c_2cr	Coefficients to calculate crown ratio of trees	No	No	A.3.2
<i>a_{cw}, b_{cw}</i>	a_cw, b_cw	Regression coefficients used to calculate the crown width of trees	No	No	A.3.2

Leaf phenology

Symbol	R	Description	Strict	Scaled	Imputable
<i>LD</i>	LeafDuration	Average duration of leaves	No	No	A.3.8
<i>t_{0,eco}</i>	t0gdd	Degree days corresponding to leaf budburst	No	No	A.3.8
<i>S_{eco}*</i>	Sgdd	Degree days corresponding to leaf budburst	No	No	A.3.8
<i>T_{eco}</i>	Tbgdd	Base temperature for the calculation of degree days to leaf budburst	No	No	A.3.8
<i>S_{sen}*</i>	Ssen	Degree days corresponding to leaf senescence	No	No	A.3.8

Symbol	R	Description	Strict	Scaled	Imputable
Ph_{sen}	<code>Phsen</code>	Photoperiod corresponding to start counting senescence degree-days	No	No	A.3.8
T_{sen}	<code>Tbsen</code>	Base temperature for the calculation of degree days to leaf senescence	No	No	A.3.8
x_{sen}	<code>xsen</code>	Discrete values, to allow for any absent/proportional/more than proportional effects of temperature on senescence	No	No	A.3.8
y_{sen}	<code>ysen</code>	Discrete values, to allow for any absent/proportional/more than proportional effects of photoperiod on senescence	No	No	A.3.8

Plant anatomy

Symbol	R	Description	Strict	Scaled	Imputable
$1/H_v$	<code>A12As</code>	Ratio of leaf area to sapwood area	No	No	A.3.6
RLR	<code>Ar2Al</code>	Fine root area to leaf area ratio	No	No	A.3.7
LW	<code>LeafWidth</code>	Leaf width	No	No	A.3.3
SLA	<code>SLA</code>	Specific leaf area	No	No	A.3.3
ρ_{leaf}	<code>LeafDensity</code>	Leaf tissue density	No	No	A.3.4
ρ_{wood}	<code>WoodDensity</code>	Wood tissue density	No	No	A.3.4
$\rho_{fineroot}$	<code>FineRootDensity</code>	Fine root tissue density	No	No	A.3.4
$f_{conduits}$	<code>conduit2sapwood</code>	Proportion of sapwood corresponding to xylem conduits	No	No	A.3.6
SRL	<code>SRL</code>	Specific fine root length	No	No	A.3.5
RLD	<code>RLD</code>	Fine root length density	No	No	A.3.5
$r_{6.35}$	<code>r635</code>	Ratio between the weight of leaves plus branches and the weight of leaves alone for branches of 6.35 mm	No	No	A.3.3

Radiation balance and water interception

Symbol	R	Description	Strict	Scaled	Imputable
k_{PAR}	<code>kPAR</code>	PAR extinction coefficient	No	No	A.3.10
α_{SWR}	<code>alphaSWR</code>	Short-wave radiation leaf absorbance coefficient	No	No	A.3.10
γ_{SWR}	<code>gammaSWR</code>	Short-wave radiation leaf reflectance (albedo)	No	No	A.3.10
s_{water}	<code>g</code>	Crown water storage capacity	No	No	A.3.10

Hydraulics, transpiration, photosynthesis

Symbol	R	Description	Strict	Scaled	Imputable
$T_{max,LAI}$	<code>Tmax_LAI</code>	Empirical coefficient relating LAI with the ratio of maximum transpiration over potential evapotranspiration	No	No	A.3.9

Symbol	R	Description	Strict	Scaled	Imputable
$T_{max,sqLAI}$	T_{max_LAI}	Empirical coefficient relating squared LAI with the ratio of maximum transpiration over potential evapotranspiration	No	No	A.3.9
WUE_{max}	WUE	Water use efficiency at VPD = 1kPa and without light or CO ₂ limitations	No	No	A.3.9
WUE_{PAR}	WUE_{par}	Coefficient describing the progressive decay of WUE with lower light levels	No	No	A.3.9
WUE_{CO2}	WUE_{co2}	Coefficient for WUE dependency on atmospheric CO ₂ concentration	No	No	A.3.9
WUE_{VPD}	WUE_{vpd}	Coefficient for WUE dependency on vapor pressure deficit	No	No	A.3.9
$\Psi_{extract}$	$\Psi_{extract}$	The water potential at which plant transpiration is 50% of its maximum	No	No	A.3.9
Ψ_{critic}	Ψ_{critic}	The water potential corresponding to 50% of stem xylem cavitation	No	No	A.3.15
g_{swmin}	G_{wmin}	Minimum stomatal conductance to water vapour	No	No	A.3.11
g_{swmax}	G_{wmax}	Maximum stomatal conductance to water vapour	No	No	A.3.11
$J_{max,298}$	J_{max298}	Maximum rate of electron transport at 298K	No	No	A.3.16
$V_{max,298}$	V_{max298}	Rubisco's maximum carboxylation rate at 298K	No	No	A.3.16
$K_{stem,max,ref}$	K_{max_stem}	Maximum stem sapwood reference conductivity per leaf area unit	No	No	A.3.13
$K_{root,max,ref}$	K_{max_root}	Maximum root sapwood reference conductivity per leaf area unit	No	No	A.3.13
$k_{leaf,max}$	VC_{leaf_km}	Maximum leaf hydraulic conductance	No	A.4.2	A.3.14
$k_{stem,max}$	VC_{stem_km}	Maximum stem hydraulic conductance	No	A.4.1	No
$k_{root,max,s}$	VC_{root_km}	Maximum root hydraulic conductance for each soil layer	No	A.4.3	No
$k_{rhizo,max,s}$	VG_{rhizo_km}	Maximum hydraulic conductance of the rhizosphere for each soil layer	No	A.4.4	No
$c_{leaf},$ d_{leaf}	VC_{leaf_c} , VC_{leaf_d}	Parameters of the vulnerability curve for leaves	No	No	A.3.15
$c_{stem},$ d_{stem}	VC_{stem_c} , VC_{stem_d}	Parameters of the vulnerability curve for stem xylem	No	No	A.3.15
$c_{root},$ d_{root}	VC_{root_c} , VC_{root_d}	Parameters of the vulnerability curve for root xylem	No	No	A.3.15

Plant water storage

Symbol	R	Description	Strict	Scaled	Imputable
ϵ_{leaf}	$LeafEPS$	Modulus of elasticity of leaves	No	No	A.3.12
ϵ_{stem}	$StemEPS$	Modulus of elasticity of symplastic xylem tissue	No	No	A.3.12
$\pi_{0,leaf}$	$LeafPIO$	Osmotic potential at full turgor of leaves	No	No	A.3.12
$\pi_{0,stem}$	$StemPIO$	Osmotic potential at full turgor of symplastic xylem tissue	No	No	A.3.12
$f_{apo,leaf}$	$LeafAF$	Apoplastic fraction in leaf tissues	No	No	A.3.12
$f_{apo,stem}$	$StemAF$	Aploplastic fraction in stem tissues	No	No	A.3.12

Symbol	R	Description	Strict	Scaled	Imputable
V_{leaf}	V_{leaf}	Leaf water capacity per leaf area unit	No	A.4.5	No
$V_{sapwood}$	$V_{sapwood}$	Sapwood water capacity per leaf area unit	No	A.4.5	No

Growth and mortality

Symbol	R	Description	Strict	Scaled	Imputable
N_{leaf}	N_{leaf}	Leaf nitrogen concentration per dry mass	No	No	A.3.17
$N_{sapwood}$	$N_{sapwood}$	Sapwood nitrogen concentration per dry mass	No	No	A.3.17
$N_{fineroot}$	$N_{fineroot}$	Fine root nitrogen concentration per dry mass	No	No	A.3.17
MR_{leaf}	MR_{leaf}	Leaf respiration rate at 20 °C	No	No	A.3.17
$MR_{sapwood}$	$MR_{sapwood}$	Dying sapwood (parenchymatic tissue) respiration rate at 20 °C	No	No	A.3.17
$MR_{fineroot}$	$MR_{fineroot}$	Fine root respiration rate at 20 °C	No	No	A.3.17
$RGR_{leaf,max}$	$RGR_{leaf,max}$	Maximum leaf area daily growth rate, relative to sapwood area	No	No	A.3.18
$RGR_{cambium,max}$	$RGR_{cambium,max}$	Maximum tree daily sapwood growth rate relative to cambium perimeter length	No	No	A.3.18
$RGR_{sapwood,max}$	$RGR_{sapwood,max}$	Maximum shrub daily sapwood growth rate relative to sapwood area	No	No	A.3.18
$RGR_{fineroot,max}$	$RGR_{fineroot,max}$	Maximum daily fine root relative growth rate	No	No	A.3.18
$SR_{sapwood}$	$SR_{sapwood}$	Daily sapwood senescence rate	No	No	A.3.19
$SR_{fineroot}$	$SR_{fineroot}$	Daily fine root senescence rate	No	No	A.3.19
$RSSG$	$RSSG$	Minimum relative starch for sapwood growth	No	No	A.3.20
C_{wood}	C_{wood}	Wood carbon content per dry weight	No	No	A.3.21
$P_{mort,base}$	$P_{mort,base}$	Baseline realistic proportion or probability specifying the baseline reduction of cohort's density occurring in a year	No	No	A.3.22

Recruitment

Symbol	R	Description	Strict	Scaled	Imputable
H_{seed}	H_{seed}	Height for seed production	No	No	A.3.23
TCM_{recr}	TCM_{recr}	Minimum average temperature (Celsius) of the coldest month for successful recruitment	No	No	A.3.23
MI_{recr}	MI_{recr}	Minimum value of the moisture index for successful recruitment	No	No	A.3.23
$FPAR_{recr}$	$FPAR_{recr}$	Minimum percentage of PAR at the ground level for successful recruitment	No	No	A.3.23
DBH_{recr}	DBH_{recr}	DBH for trees	No	No	A.3.23
$H_{tree,recr}$	$H_{tree,recr}$	Height for trees	No	No	A.3.23
$N_{tree,recr}$	$N_{tree,recr}$	Density for trees	No	No	A.3.23
$Cover_{shrub,recr}$	$Cover_{shrub,recr}$	Cover for shrubs	No	No	A.3.23
$H_{shrub,recr}$	$H_{shrub,recr}$	Height for shrubs	No	No	A.3.23

Symbol	R	Description	Strict	Scaled	Imputable
$Z_{50_{recr}}$	RecrZ50	Soil depth corresponding to 50% of fine roots for recruitment	No	No	A.3.23
$Z_{95_{recr}}$	RecrZ95	Soil depth corresponding to 95% of fine roots for recruitment	No	No	A.3.23

Flammability

Symbol	R	Description	Strict	Scaled	Imputable
ρ_p	PD	Density of fuel particles	No	No	A.3.24
σ	SAV	Surface-area-to-volume ratio of the small fuel (1h) fraction (leaves and branches < 6.35mm)	No	No	A.3.24
h	HeatContHmt	High fuel heat content.	No	No	A.3.24
LI	PercentLignin	Percentage of lignin in leaves	No	No	A.3.24

A.3 Imputation of missing values

The following figure summarizes the percentage of missing values in SpParamsMED for different model parameters and the other model parameters used for the imputation of missing values:

A.3.1 Shrub allometric coefficients

Missing shrub allometric coefficients are filled using information from Raunkiaer's life form and maximum plant height (H_{max}).

Life form	H_{max}	a_{ash}	b_{ash}	a_{bsh}	b_{bsh}	cr
Chamaephyte	[any]	24.5888	1.1662	0.7963	0.3762	0.8076
Phanerophyte	< 300 cm	1.0083	1.8700	0.7900	0.6942	0.6630
Phanerophyte	> 300 cm	5.8458	1.4944	0.3596	0.7138	0.7190
(Hemi)cryptophyte	[any]	24.5888	1.1662	0.7963	0.3762	0.9500

Allometric coefficients were taken from De Cáceres *et al.* (2019).

A.3.2 Tree allometric coefficients

Missing tree allometric coefficients are replaced with values depending on whether the plant species is a gymnosperm or an angiosperm:

Parameter	Gymnosperm	Angiosperm
a_{fbt}	0.1300	0.0527
b_{fbt}	1.2285	1.5782
c_{fbt}	-0.0147	-0.0066
a_{cw}	0.747	0.839
b_{cw}	0.672	0.735
a_{cr}	1.995	1.506

Parameter	Gymnosperm	Angiosperm
b_{1cr}	-0.649	-0.706
b_{2cr}	-0.020	-0.078
b_{3cr}	-0.00012	0.00018
c_{1cr}	-0.004	-0.007
c_{2cr}	-0.159	0.000

A.3.3 Leaf width, specific leaf area and fine foliar ratio

Leaf width (LW), specific leaf area (SLA) and the ratio between the weight of leaves plus branches and the weight of leaves alone for branches of 6.35 mm ($r_{6.35}$) are key anatomical parameters. When missing from species parameter table, default estimates for these parameters are obtained from combinations of leaf shape and leaf size:

Leaf shape	Leaf size	SLA	LW	$r_{6.35}$
Broad	Large	16.039	6.898	2.278
Broad	Medium	11.499	3.054	2.359
Broad	Small	9.540	0.644	3.026
Linear	Large	5.522	0.639	3.261
Linear	Medium	4.144	0.639	3.261
Linear	Small	13.189	0.639	3.261
Needle	[any]	9.024	0.379	1.716
Scale	[any]	4.544	0.101	1.483

These estimates have been obtained by averaging species-level values across combinations of the categorical variables.

A.3.4 Tissue density

Default values for the dry weight density of leaves and wood (in $g \cdot cm^{-3}$) are determined from taxonomic family using an internal data set (`medfate:::trait_family_means`):

If the family is not any of those in the table, default values are $\rho_{leaf} = 0.7$ and $\rho_{wood} = 0.652$. The default value for fine root density is always $\rho_{fineroot} = 0.165$.

A.3.5 Specific root length and root length density

Default values for specific fine root length and fine root length density are $3870 \text{ cm} \cdot g^{-1}$ and $10 \text{ cm} \cdot cm^{-3}$, respectively. [JUSTIFICATION MISSING]

A.3.6 Huber value and ratio of conduits to sapwood

Missing values for A12As, the inverse of the Huber value ($1/Hv$) are determined from taxonomic family using an internal data set (`medfate:::trait_family_means`):

If there is no information derived from taxonomic family for A12As, a default value is given depending on leaf shape and leaf size:

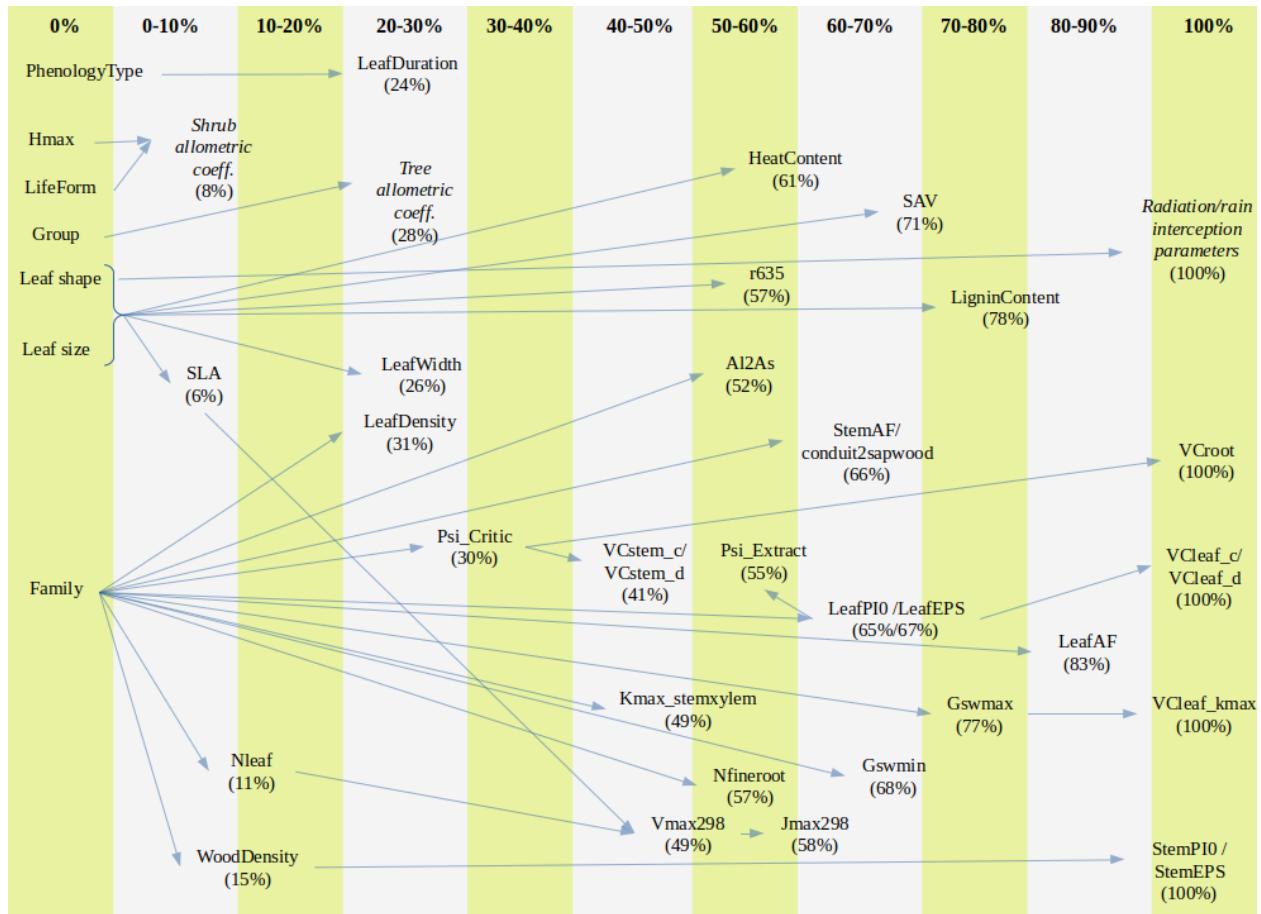


Figure A.1: Representation of imputation relationships between model parameters. The percentage of missing parameter values increases from left to right. Left-most parameters are strict.

Table A.15: Default leaf density and wood density values by taxonomic family.

	LeafDensity	WoodDensity
Acanthaceae	0.2959003	0.5684693
Achariaceae	0.2552495	0.6052036
Acoraceae	0.1000000	NA
Actinidiaceae	0.4439776	0.4092320
Adoxaceae	0.3916194	0.5157416
Aextoxicaceae	NA	0.5666667
Aizoaceae	0.0824125	NA
Akaniaceae	NA	0.5547825
Alismataceae	0.1914208	NA
Altingiaceae	0.6833971	0.6010948
Amaranthaceae	0.2045831	0.6315739
Amaryllidaceae	0.1220704	NA
Amphorogynaceae	NA	0.6097400
Anacardiaceae	0.4568088	0.5685583
Anisophylleaceae	NA	0.6734780
Annonaceae	0.3757811	0.5642062
Aphloiaceae	0.4627060	0.6205200
Apiaceae	0.2874756	0.2561785
Apocynaceae	0.2981148	0.5683635
Aptandraceae	0.3318492	0.7076756
Aquifoliaceae	0.4812626	0.5579305
Araceae	0.1659746	NA
Araliaceae	0.3118821	0.4142687
Araucariaceae	0.3447902	0.4641456
Arecaceae	0.4393589	0.5913967
Aristolochiaceae	0.2681434	0.2900000
Asparagaceae	0.1425099	0.4254258
Asphodelaceae	0.5512472	0.3951990
Aspleniaceae	0.2644557	NA
Asteraceae	0.2511093	0.4822337
Asteropeiaceae	NA	0.7554862
Atherospermataceae	0.2451357	0.4767621
Athyriaceae	0.2270615	NA
Austrobaileyaceae	0.2620992	NA
Balanopaceae	NA	0.7348976
Balsaminaceae	0.3948094	NA
Begoniaceae	0.1626789	NA
Berberidaceae	0.3599453	0.7028850
Betulaceae	0.4446528	0.5381493
Bignoniaceae	0.3572590	0.6256030
Bixaceae	0.4400000	0.3546357
Blechnaceae	0.3147993	NA
Bonnetiaceae	0.2797829	0.8400000
Boraginaceae	0.2850462	0.4987559
Brassicaceae	0.2267605	0.4516377
Bromeliaceae	0.1677139	NA
Brunelliaceae	NA	0.3112500
Bruniaceae	NA	0.5636500
Burseraceae	0.4226787	0.5205008
Buxaceae	0.2706294	0.7314511
Cactaceae	0.1819372	0.6187500
Calophyllaceae	0.4065092	0.6067855
Calycanthaceae	0.3204073	0.6500550
Calyceraceae	0.1689366	NA
Campanulaceae	0.2396318	NA

Table A.16: Default leaf area to sapwood area (m²/m²) / Huber value (cm²/m²) and fraction of sapwood corresponding to conduits by taxonomic family.

	Al2As	Hv	conduit2sapwood
Acanthaceae	3070.43202	3.2568707	0.6300000
Adoxaceae	4889.57295	2.0451684	NA
Altingiaceae	5129.90090	1.9493554	0.8226667
Amaranthaceae	973.13500	10.2760665	NA
Amborellaceae	4255.31915	2.3500000	NA
Anacardiaceae	19581.70525	0.5106808	0.7155889
Annonaceae	10266.65654	0.9740269	0.5685000
Apiaceae	81.15013	123.2283950	NA
Apocynaceae	19766.80134	0.5058987	0.7112500
Aquifoliaceae	4886.36183	2.0465124	0.6528500
Araliaceae	3928.39877	2.5455664	0.7785000
Araucariaceae	3846.15385	2.6000000	0.9375000
Arecaceae	5492.53731	1.8206522	NA
Asteraceae	2421.76507	4.1292197	0.7219423
Atherospermataceae	2435.95630	4.1051640	0.7560000
Austrobaileyaceae	15384.61538	0.6500000	NA
Berberidaceae	570.06271	17.5419298	NA
Betulaceae	6158.73417	1.6237103	0.8444000
Bignoniaceae	12439.80827	0.8038709	0.6360476
Bixaceae	12274.01424	0.8147294	NA
Burseraceae	12218.54705	0.8184279	0.8204286
Buxaceae	NA	NA	0.8330000
Cactaceae	2554.61304	3.9144872	0.3636905
Calophyllaceae	2662.52127	3.7558385	0.7335000
Calycanthaceae	NA	NA	0.6535500
Cannabaceae	29406.04715	0.3400661	0.7598125
Capparaceae	37525.87992	0.2664828	0.7005000
Caprifoliaceae	7568.54261	1.3212583	NA
Cardiopteridaceae	NA	NA	0.6300000
Caryocaraceae	5183.13611	1.9293339	NA
Casuarinaceae	3647.15784	2.7418610	NA
Celastraceae	8199.20056	1.2196311	NA
Chrysobalanaceae	10531.83851	0.9495018	0.5876000
Cistaceae	2129.37624	4.6962109	NA
Clusiaceae	6707.44573	1.4908805	0.5005000
Cochlospermaceae	1757.46925	5.6900000	0.7070000
Combretaceae	23650.62103	0.4228219	0.6416667
Cordiaceae	10754.86431	0.9298118	0.6300000
Cornaceae	7478.58356	1.3371516	0.6740000
Coulaceae	10676.38668	0.9366465	0.6520000
Cunoniaceae	4781.22975	2.0915121	0.7110000
Cupressaceae	1793.78915	5.5747912	0.9242857
Dichapetalaceae	7505.55229	1.3323470	NA
Dilleniaceae	7707.14493	1.2974973	0.5432500
Dipterocarpaceae	NA	NA	0.7043750
Ebenaceae	5746.58263	1.7401647	0.6300000
Ehretiaceae	3401.36054	2.9400000	NA
Elaeagnaceae	NA	NA	0.5641000
Elaeocarpaceae	8949.89085	1.1173321	0.6970000
Ericaceae	2717.89086	3.6793236	0.7423400
Erythropalaceae	11359.73085	0.8803025	NA
Erythroxylaceae	16409.39633	0.6094069	NA
Escalloniaceae	NA	NA	0.5755000
Euphorbiaceae	9648.31966	1.0364499	0.6630417

Leaf shape	Leaf size	A12As
Broad	Large	4768.7
Broad	Medium	2446.1
Broad	Small	2284.9
Linear	Large	2156.0
Linear	Medium	2156.0
Linear	Small	2156.0
Needle	[any]	2751.7
Scale	[any]	1696.6

Missing values for $f_{conduits}$, the fraction of sapwood corresponding to conduits are derived from taxonomic family (see table above). If information from taxonomic family is missing, default values are $f_{conduits} = 0.7$ (i.e. 30% of parenchyma) for angiosperms, and $f_{conduits} = 0.925$ (i.e. 7.5% of parenchyma) for gymnosperms (Plavcová & Jansen 2015).

A.3.7 Fine root to leaf area ratio

When missing, the fine root area to leaf area ratio is given a default value of $RLR = 1 \text{ m}^2 \cdot \text{m}^{-2}$.

A.3.8 Leaf phenology

When missing, leaf duration is assigned a value of 1 year for winter-deciduous species and 2.41 years for the remaining leaf phenology types.

Default values for leaf phenological parameters are the same regardless of the leaf phenology type:

Phenology type	t0gdd	Sgdd	Tbgdd	Ssen	Phsen	Tbsen	xsen	ysen
One-flush evergreen	50	200	0	8268	12.5	28.5	2	2
Winter deciduous	50	200	0	8268	12.5	28.5	2	2
Winter semi-deciduous	50	200	0	8268	12.5	28.5	2	2
Drought deciduous	50	200	0	8268	12.5	28.5	2	2

Leaf senescence values were derived for deciduous broad-leaved forests by Delpierre *et al.* (2009).

A.3.9 Basic transpiration and water-use efficiency

When the basic soil water balance model is used, $T_{max,LAI}$ and $T_{max,sqLAI}$ are species-specific parameters that regulate the maximum transpiration of plant cohorts (see 6.1.1). When these parameters are missing from `SpParams` table, they are given default values $T_{max,LAI} = 0.134$ and $T_{max,sqLAI} = -0.006$, according to Granier *et al.* (1999).

When maximum water use efficiency (WUE_{max}) is missing, it is given a value of $WUE_{max} = 7.55$. By default, the coefficient describing the decay of water use efficiency with lower light levels is given a default value of $WUE_{PAR} = 0.2812$, and the coefficient regulating the relationship between gross photosynthesis and CO₂ concentration is given a default $WUE_{CO2} = 0.0028$.

When missing, the water potential corresponding to 50% of transpiration ($\Psi_{extract}$) is estimated by calculating the water potential corresponding to the loss leaf turgor (Ψ_{tlp}), using equation (10.11) from Bartlett

et al. (2012). The parameters of the leaf pressure-volume curve needed for applying equation (10.11) may be themselves estimated (see A.3.12). Note that Ψ_{tlp} has been found to be highly correlated to Ψ_{gs50} , the water potential corresponding to 50% of stomatal conductance (Bartlett *et al.* 2016).

A.3.10 Radiation balance and water interception

Default values for radiation extinction and water interception parameters depend on the leaf shape:

Leaf shape	k_{PAR}	α_{SWR}	γ_{SWR}	s_{water}
Broad	0.55	0.70	0.18	0.5
Linear	0.45	0.70	0.15	0.8
Needle/Scale	0.50	0.70	0.14	1.0

where k_{PAR} is the PAR extinction coefficient, α_{SWR} is the short-wave radiation leaf absorbance coefficient, γ_{SWR} is the short-wave radiation leaf reflectance (albedo) and s_{water} is the crown water storage capacity per LAI unit.

A.3.11 Stomatal conductance

Default values for minimum and maximum conductance to water vapour (g_{swmin} and g_{swmax} ; in $\text{mol H}_2\text{O} \cdot \text{s}^{-1} \cdot \text{m}^{-2}$) were defined depending on taxonomic family, from Duursma *et al.* (2018) and Hoshika *et al.* (2018), and stored in an internal data set (`medfate:::trait_family_means`):

If there is no information derived from taxonomic family, $g_{swmin} = 0.0049$ and $g_{swmax} = 0.200$.

A.3.12 Pressure-volume curves

Parameters of the pressure-volume curve (i.e. $\pi_{0,stem}$ and ϵ_{stem}) for leaf and stem symplastic tissue are required for each species.

When parameters for stem tissue are missing, **medfate** estimates them from wood density following Christofersen *et al.* (2016):

$$\pi_{0,stem} = 0.52 - 4.16 \cdot \rho_{wood} \quad (\text{A.1})$$

$$\epsilon_{stem} = \sqrt{1.02 \cdot e^{8.5 \cdot \rho_{wood}} - 2.89} \quad (\text{A.2})$$

while the apoplastic fraction of stem is assumed $f_{apo,stem} = f_{conducts}$ (see A.3.6).

Default values for leaf pressure-volume parameters, i.e. $\pi_{0,leaf}$, ϵ_{leaf} and $f_{apo,leaf}$, are determined from taxonomic family using an internal data set (`medfate:::trait_family_means`):

If family-level values are missing, following Bartlett *et al.* (2012) average values for Mediterranean climate leaves are taken as defaults, i.e. $\pi_{0,leaf} = -2$ MPa, $\epsilon_{leaf} = 17$, whereas a 29% leaf apoplastic fraction is assumed (i.e. $f_{apo,leaf} = 0.29$).

A.3.13 Stem and root maximum hydraulic conductivity

Tissue-level maximum conductivity parameters (i.e. $K_{stem,max,ref}$ and $K_{root,max,ref}$) are not direct parameters to simulation functions. Instead, they are scaled to estimate stem- and root-level hydraulic conductances (i.e. $k_{stem,max}$ and $k_{root,max}$) using plant size (see A.4.1 and A.4.3 for details). $K_{stem,max,ref}$ and $K_{root,max,ref}$ are supplied via species parameter table and missing values can therefore occur.

Table A.20: Default minimum and maximum conductance to water vapour by taxonomic family.

	Gswmin	Gswmax
Acanthaceae	NA	0.2500000
Altingiaceae	NA	0.5000000
Amaranthaceae	NA	0.0750000
Amaryllidaceae	0.0180400	NA
Anacardiaceae	0.0122299	0.3148333
Apiaceae	0.0015873	NA
Aquifoliaceae	0.0005740	0.2600000
Araliaceae	0.0003902	NA
Araucariaceae	0.0018786	NA
Arecaceae	0.0004500	NA
Aristolochiaceae	0.0035896	NA
Aspleniaceae	0.0082000	NA
Asteraceae	0.0105288	0.1275000
Balsaminaceae	0.0126573	NA
Berberidaceae	0.0016500	NA
Betulaceae	0.0029731	0.3965000
Boraginaceae	0.0057402	NA
Brassicaceae	0.0106600	NA
Cactaceae	0.0000243	NA
Calophyllaceae	NA	0.1350000
Cannabaceae	NA	0.3300000
Caryophyllaceae	0.0018519	NA
Cephalotaxaceae	0.0022700	NA
Cercidiphyllaceae	NA	0.4000000
Chrysobalanaceae	NA	0.1740000
Cistaceae	0.0082820	NA
Clethraceae	NA	0.2500000
Combretaceae	0.0131200	NA
Convolvulaceae	0.0041967	NA
Cordiaceae	0.0065600	NA
Cornaceae	NA	0.3000000
Crassulaceae	0.0022751	NA
Cupressaceae	0.0063192	0.0840000
Cyperaceae	0.0190600	NA
Dipterocarpaceae	NA	0.3874722
Ehretiaceae	NA	0.2900000
Elaeagnaceae	NA	0.1700000
Ericaceae	0.0041455	0.1723333
Euphorbiaceae	0.0026500	0.2616667
Fabaceae	0.0085178	0.3066667
Fagaceae	0.0060137	0.3127601
Geraniaceae	0.0043050	NA
Hamamelidaceae	NA	0.2600000
Juglandaceae	0.0039355	0.4900000
Krameriaceae	NA	0.1600000
Lamiaceae	0.0052377	0.7300000
Lauraceae	NA	0.2212500
Magnoliaceae	0.0038800	0.3075000
Malvaceae	0.0064056	0.5532500
Meliaceae	NA	0.1600000
Moraceae	0.0064020	0.3835000
Myristicaceae	NA	0.0880000
Myrtaceae	0.0078149	0.2310769
Oleaceae	0.0046740	NA
Onagraceae	0.0169506	NA

Table A.21: Default leaf pressure-volume parameters by taxonomic family.

	LeafPI0	LeafEPS	LeafAF
Acanthaceae	-3.395000	23.230000	0.1270000
Adoxaceae	-1.560000	12.790000	NA
Amaranthaceae	-2.250000	NA	NA
Anacardiaceae	-1.700000	12.760000	NA
Annonaceae	-2.160000	23.710000	NA
Apocynaceae	-2.390000	20.940000	NA
Aquifoliaceae	-2.300000	20.700000	0.4000000
Araliaceae	-1.528503	11.231462	0.4065000
Arecaceae	-3.400000	73.400000	0.2000000
Aspleniaceae	-1.240000	35.300000	NA
Asteraceae	-1.471389	14.491429	0.2435000
Atherospermataceae	-1.340000	8.380000	NA
Betulaceae	-1.246984	5.498667	NA
Bignoniaceae	-1.990000	17.610000	0.1770000
Boraginaceae	-1.140000	NA	NA
Brassicaceae	-1.485000	7.710000	0.2320000
Burseraceae	-1.435000	14.980000	NA
Cactaceae	NA	8.700000	NA
Cannabaceae	-1.580000	5.180000	NA
Capparaceae	-2.840000	14.750000	0.1723333
Caryocaraceae	-1.710000	11.340000	NA
Celastraceae	-2.600000	19.060000	NA
Combretaceae	-2.110000	6.830000	NA
Connaraceae	-2.250000	18.010000	NA
Convolvulaceae	-1.320000	NA	NA
Cordiaceae	-1.647500	11.295938	NA
Cucurbitaceae	-0.980000	NA	NA
Cupressaceae	-1.480000	12.600000	0.2720000
Dipterocarpaceae	-1.131429	23.630000	0.4634286
Dryopteridaceae	-1.425000	48.950000	NA
Ebenaceae	-2.110000	14.650000	NA
Ericaceae	-1.670000	14.382000	0.4660000
Erythroxylaceae	-1.916667	16.993333	NA
Euphorbiaceae	-1.260000	17.570000	0.4745000
Fabaceae	-1.641724	13.577917	0.2609091
Fagaceae	-1.994080	18.132800	0.2206667
Geraniaceae	-0.730000	3.740000	NA
Goodeniaceae	-1.500000	NA	NA
Grossulariaceae	-1.845000	NA	NA
Hydrophyllaceae	-1.260000	NA	NA
Irvingiaceae	-1.690000	38.380000	0.4760000
Lamiaceae	-1.184000	6.120000	0.2200000
Lauraceae	-2.074728	16.763373	0.1760000
Lindsaeaceae	-1.970000	7.340000	0.1700000
Lythraceae	-1.535000	6.055000	NA
Magnoliaceae	-1.430000	9.140000	0.1560000
Malpighiaceae	-1.540000	9.450000	NA
Malvaceae	-2.110000	17.730000	NA
Melastomataceae	-1.754000	12.306667	NA
Moraceae	-1.353563	13.575000	NA
Myrtaceae	-1.852385	14.462948	0.4006000
Nothofagaceae	-1.480000	8.380000	NA
Nyctaginaceae	-1.280000	NA	NA
Oleaceae	-2.060557	14.205556	NA
Onagraceae	-1.200000	NA	NA

Default values for $K_{stem,max,ref}$ are determined from taxonomic family using an internal data set (`medfate:::trait_family_means`):

If family-level values are missing, suitable $K_{stem,max,ref}$ values are decided according to combinations of taxon group (either Angiosperm or Gymnosperm), growth form (either tree or shrub) and leaf phenology (Maherali *et al.* 2004):

Group	Growth form	Leaf phenology	$K_{stem,max,ref}$
Angiosperm	Tree	Winter-(semi)deciduous	1.58
Angiosperm	Shrub	Winter-(semi)deciduous	1.55
Angiosperm	Tree/Shrub	Evergreen	2.43
Gymnosperm	Tree	any	0.48
Gymnosperm	Shrub	any	0.24

Following Oliveras *et al.* (2003), missing values for $K_{root,max,ref}$ are assumed to be four-times the values given or estimated for $K_{stem,max,ref}$.

A.3.14 Leaf maximum hydraulic conductance

Leaf maximum hydraulic conductance ($k_{l,max}$, in $\text{mmol} \cdot \text{m}^{-2} \cdot \text{s}^{-1} \cdot \text{MPa}^{-1}$) is an input parameter that should be provided for each species. When missing, leaf maximum hydraulic conductance can be estimated from maximum stomatal conductance (g_{swmax}), following Franks (2006) (original coefficients were modified for better fit):

$$k_{l,max} = (g_{swmax}/0.015)^{1/1.3} \quad (\text{A.3})$$

Note that values for g_{swmax} may also be imputed (see A.3.11).

A.3.15 Xylem vulnerability

Default values for $\Psi_{50,stem}$ are determined from taxonomic family using an internal data set (`medfate:::trait_family_means`):

If family-level values is missing, a suitable estimate of $\Psi_{50,stem}$ the water potential corresponding to 50% of conductance loss, can be obtained from Maherali *et al.* (2004) according to combinations of taxon group (either Angiosperm or Gymnosperm), growth form (either tree or shrub) and leaf phenology:

Group	Growth form	Leaf phenology	$\Psi_{50,stem}$
Angiosperm	Tree/Shrub	Winter-(semi)deciduous	-2.34
Angiosperm	Tree	Evergreen	-1.51
Angiosperm	Shrub	Evergreen	-5.09
Gymnosperm	Tree	any	-4.17
Gymnosperm	Shrub	any	-8.95

$\Psi_{50,stem}$ estimates are taken for parameter Ψ_{critic} , in the case of the basic water balance model.

Vulnerability curves in the advanced model need to be specified for leaves, stem and root segments via the two parameters of the Weibull function (see 10.2). When any of the parameters of the stem vulnerability curve is missing, a regression equation using data from Choat *et al.* (2012) can be used to estimate $\Psi_{88,stem}$ from $\Psi_{50,stem}$:

$$\Psi_{88,stem} = -1.4264 + 1.2593 \cdot \Psi_{50,stem} \quad (\text{A.4})$$

Finally, estimates for c_{stem} and d_{stem} are obtained from $\Psi_{50,stem}$ and $\Psi_{88,stem}$ via function `hydraulics_psi2Weibull()`.

Table A.22: Default maximum stem hydraulic conductivity by taxonomic family.

	Kmax_stemxylem
Adoxaceae	4.0535575
Altingiaceae	0.5050000
Amaranthaceae	0.0819000
Amborellaceae	0.5400000
Anacardiaceae	4.0772016
Annonaceae	5.2706667
Apiaceae	0.5150000
Apocynaceae	2.5651667
Aquifoliaceae	0.2255757
Araliaceae	1.6809011
Araucariaceae	0.7322500
Asteraceae	0.4986571
Austrobaileyaceae	2.3000000
Berberidaceae	0.0873333
Betulaceae	2.8733374
Bignoniaceae	2.1014900
Bruniaceae	0.2515000
Burseraceae	3.4050000
Cactaceae	1.8688095
Calophyllaceae	0.8982857
Cannabaceae	4.3396105
Capparaceae	0.5572667
Caprifoliaceae	0.2521010
Caryocaraceae	1.7587273
Casuarinaceae	1.8954048
Cistaceae	0.3958482
Clusiaceae	0.5950000
Cochlospermaceae	7.7500000
Combretaceae	5.7436667
Convolvulaceae	2.2000000
Cordiaceae	4.4607389
Cornaceae	2.3439777
Cupressaceae	0.9325131
Daphniphyllaceae	0.4600000
Dennstaedtiaceae	21.9800000
Dilleniaceae	1.2500000
Dipterocarpaceae	8.8857143
Ebenaceae	1.5527759
Ehretiaceae	0.4100000
Ericaceae	0.6151806
Erythroxylaceae	0.5352000
Euphorbiaceae	3.3816717
Eupomatiaceae	1.1239700
Fabaceae	2.9075403
Fagaceae	2.5306061
Garryaceae	1.9333333
Gnetaceae	1.2900000
Iteaceae	0.4500000
Juglandaceae	4.2050000
Lamiaceae	4.8686077
Lauraceae	1.1201255
Lecythidaceae	6.5443450
Loranthaceae	0.2817696
Lythraceae	6.1800000
Malpighiaceae	11.0500000

Table A.24: Default stem P50 values by taxonomic family.

	P50
Adoxaceae	-3.0384833
Altingiaceae	-2.0370147
Amaranthaceae	-2.4252402
Amborellaceae	-3.0000000
Anacardiaceae	-2.6535235
Annonaceae	-2.5276068
Apiaceae	-5.7000000
Apocynaceae	-2.4864334
Aquifoliaceae	-3.6437782
Araliaceae	-1.6530859
Araucariaceae	-2.6183226
Arecaceae	-1.8100000
Asparagaceae	-1.6960000
Asteraceae	-3.2565860
Atherospermataceae	-3.0063333
Austrobaileyaceae	-0.4990000
Berberidaceae	-4.5000000
Betulaceae	-2.1017591
Bignoniaceae	-0.8616667
Boraginaceae	-3.5677066
Bruniaceae	-3.3883558
Burseraceae	-1.3054970
Buxaceae	-8.0000000
Cactaceae	-1.2875000
Calophyllaceae	-1.5400000
Calycanthaceae	-1.2808475
Canellaceae	-0.2320000
Cannabaceae	-1.5325304
Capparaceae	-2.3615234
Caprifoliaceae	-5.5144833
Caryocaraceae	-1.6766667
Casuarinaceae	-2.1750000
Celastraceae	-3.4679167
Chloranthaceae	-1.7228571
Chrysobalanaceae	-2.2000000
Cistaceae	-7.1951892
Cleomaceae	-2.1842678
Clusiaceae	-1.3172327
Cochlospermaceae	-1.4400000
Combretaceae	-1.8483333
Convolvulaceae	-1.5987102
Cordiaceae	-2.3307795
Cornaceae	-4.1378833
Cunoniaceae	-1.1500000
Cupressaceae	-8.3049767
Daphniphyllaceae	-0.5985705
Dennstaedtiaceae	-1.9900000
Dilleniaceae	-1.4112676
Dipterocarpaceae	-0.3628571
Dryopteridaceae	-2.5797602
Ebenaceae	-1.5093287
Ehretiaceae	-3.8200000
Ericaceae	-2.8142685
Euphorbiaceae	-1.4638127
Eupomatiaceae	-0.3950000

Vulnerability curves for root xylem are less common than for stem xylem. If these values are missing, $\Psi_{50,stem}$ is first estimated according to its definition and the stem vulnerability curve parameters, c_{stem} and d_{stem} . Then, a relationship from Bartlett *et al.* (2016) is used to estimate $\Psi_{50,root}$ from $\Psi_{50,stem}$:

$$\Psi_{50,root} = 0.4892 + 0.742 \cdot \Psi_{50,stem} \quad (\text{A.5})$$

Finally, $\Psi_{88,stem}$ and the Weibull vulnerability parameters are obtained as explained for stems.

Vulnerability curves for leaf xylem are also less common than for stem xylem. If these values are missing, the water potential at turgor loss point Ψ_{tlp} is first estimated from π_{leaf} and ϵ_{leaf} according to eq. (10.11). Then, a relationship calibrated with data from Bartlett *et al.* (2016) is used to estimate $\Psi_{50,leaf}$ from Ψ_{tlp} :

$$\Psi_{50,leaf} = 0.2486 + 0.9944 \cdot \Psi_{tlp} \quad (\text{A.6})$$

Finally, $\Psi_{88,leaf}$ and the Weibull vulnerability parameters are obtained as explained for stems.

A.3.16 Photosynthesis rates

Rubisco's maximum carboxylation rate at 25°C ($V_{max,298}$, in $\mu\text{molCO}_2 \cdot s^{-1} \cdot m^{-2}$) is a required input parameter for each species ($V_{max,298}$). When missing, the work by Walker *et al.* (2014) suggests that suitable estimates can be derived from SLA and N_{area} , the latter being the nitrogen concentration per leaf area:

$$V_{max,298} = e^{1.993+2.555 \cdot \log(N_{area}) - 0.372 \cdot \log(SLA) + 0.422 \cdot \log(N_{area}) \cdot \log(SLA)} \quad (\text{A.7})$$

In turn, imputation for SLA is explained in A.3.3, whereas values for N_{area} are determined from N_{leaf} and SLA , being N_{leaf} estimated as indicated in A.3.17. Would N_{leaf} and SLA values be both unavailable, a default value of 100 $\mu\text{molCO}_2 \cdot s^{-1} \cdot m^{-2}$ is used for $V_{max,298}$ imputation.

When the maximum rate of electron transport at the same temperature ($J_{max,298}$) is not provided by the user, it can be estimated from $V_{max,298}$ using (Walker *et al.* 2014):

$$J_{max,298} = e^{1.197+0.847 \cdot \log(V_{max,298})} \quad (\text{A.8})$$

A.3.17 Maintenance respiration rates

When missing at the species parameter table, maintenance respiration rates for leaves, sapwood and fine roots (RER_{leaf} , $RER_{sapwood}$ and $RER_{fineroot}$; all in $g\text{gluc} \cdot g\text{dry}^{-1} \cdot day^{-1}$) are estimated from corresponding tissue nitrogen concentrations (N_{leaf} , $N_{sapwood}$ and $N_{fineroot}$; all in $mg\text{N} \cdot g\text{dry}^{-1}$) following the equations given by Reich *et al.* (2008) (after appropriate unit conversion):

$$RER_{leaf} = e^{0.691+1.639 \cdot \log(N_{leaf})} \quad (\text{A.9})$$

$$RER_{sapwood} = e^{1.024+1.344 \cdot \log(N_{sapwood})} \quad (\text{A.10})$$

$$RER_{fineroot} = e^{0.980+1.352 \cdot \log(N_{fineroot})} \quad (\text{A.11})$$

where in the previous equations nitrogen concentrations are in $mmol\text{N} \cdot g\text{dry}^{-1}$ and respiration rates in $nmol\text{CO}_2 \cdot g\text{dry}^{-1} \cdot s^{-1}$.

In turn, when tissue nitrogen concentrations are missing they are estimated from taxonomic family using an internal data set (`medfate:::trait_family_means`):

When family values are also missing, default tissue-averaged nitrogen concentrations are given: $N_{leaf} = 20.088$, $N_{sapwood} = 3.9791$ and $N_{fineroot} = 12.207$.

Default control values ($MR_{leaf} = 0.00260274$), sapwood ($MR_{sapwood} = 6.849315e - 05$) and fine roots ($MR_{fineroot} = 0.002054795$) were used in previous model versions, derived from Ogle & Pacala (2009), but these are no longer used because of easier parameterization using tissue nitrogen concentration.

Table A.26: Default nitrogen concentration per dry mass in different tissues by taxonomic family.

	Nleaf	Nsapwood	Nfineroot
Acanthaceae	27.122403	NA	5.580000
Achariaceae	22.162815	NA	NA
Acoraceae	18.000000	NA	NA
Actinidiaceae	20.183321	NA	NA
Adoxaceae	20.566364	NA	11.013750
Aextoxicaceae	9.621429	NA	NA
Aizoaceae	14.800000	NA	NA
Alismataceae	27.287834	NA	NA
Alstroemeriaeae	19.402857	NA	NA
Altingiaceae	15.462547	NA	7.550000
Amaranthaceae	23.740694	NA	12.411441
Amaryllidaceae	28.734076	NA	11.440000
Amphorogynaceae	25.236766	NA	NA
Anacardiaceae	17.914439	NA	10.539737
Annonaceae	23.593695	NA	23.830391
Apiaceae	24.746891	NA	10.787434
Apocynaceae	21.754119	NA	16.845575
Aptandraceae	28.435524	NA	NA
Aquifoliaceae	14.645818	NA	14.324450
Araceae	22.255347	NA	NA
Araliaceae	18.093569	NA	20.500000
Araucariaceae	12.622035	NA	13.000000
Arecaceae	18.349681	NA	13.216667
Aristolochiaceae	31.726525	NA	NA
Asparagaceae	22.875870	NA	NA
Asphodelaceae	12.352222	NA	NA
Aspleniaceae	28.260000	NA	NA
Asteraceae	22.036543	NA	9.346061
Atherospermataceae	17.869722	NA	23.200000
Athyriaceae	26.901037	NA	NA
Aulacomniaceae	8.000000	NA	NA
Balsaminaceae	36.003950	NA	NA
Begoniaceae	34.200000	NA	NA
Berberidaceae	17.997372	NA	21.087500
Betulaceae	24.194029	14.505149	13.430951
Bignoniaceae	23.830902	6.575973	19.915047
Bixaceae	25.531043	NA	NA
Blechnaceae	11.749050	NA	NA
Bonnetiaceae	9.800000	NA	NA
Boraginaceae	23.032555	NA	NA
Brassicaceae	34.316821	NA	18.501306
Bromeliaceae	9.559591	NA	NA
Brunelliaceae	21.006960	NA	NA
Bruniaceae	7.781667	NA	NA
Burseraceae	19.028712	NA	10.964427
Butomaceae	42.600000	NA	NA
Buxaceae	22.691830	NA	NA
Cabombaceae	19.500000	NA	NA
Cactaceae	16.988641	NA	NA
Calophyllaceae	12.424790	NA	NA
Calycanthaceae	17.400000	NA	NA
Calyceraceae	43.000000	NA	NA
Campanulaceae	27.682767	NA	5.721013
Cannabaceae	28.941055	NA	NA
Cannaceae	39.700000	NA	NA

A.3.18 Relative growth rates

When missing at the species parameter table, maximum relative growth rates for leaves, sapwood and fine roots are taken from `control` parameters. Default values are provided in 15.5.3.

A.3.19 Senescence rates

When missing at the species parameter table, senescence rates for sapwood and fine roots are taken from `control` parameters.

A.3.20 Relative starch for sapwood growth

When missing at the species parameter table, the minimum relative starch for sapwood growth is taken from `control` parameters. Default value is provided in 15.5.3.

A.3.21 Wood carbon

Default values for C_{wood} are determined from taxonomic family using an internal data set (`medfate:::trait_family_means`). If family-level values are missing, default value of $C_{wood} = 0.5 \text{ g C} \cdot \text{g dry}^{-1}$ is used.

A.3.22 Mortality baseline rate

When missing at the species parameter table, the mortality baseline rate is taken from `control` parameters. Default value is provided in 15.5.3.

A.3.23 Recruitment

Imputation of missing values for recruitment is specified via `control` parameters. Default values are provided in 18.4.3.

A.3.24 Flammability

Default values for the surface-area-to-volume ratio (σ), fuel heat content (h) and lignin percent (LI) are defined from leaf size and leaf shape as follows:

Leaf shape	Leaf size	σ	h	LI
Broad	Large	5740	19740	15.50
Broad	Medium	4039	19825	20.21
Broad	Small	4386	20062	22.32
Linear/Needle	Large	3620	18250	24.52
Linear/Needle	Medium	4758	21182	24.52
Linear/Needle	Small	6697	21888	18.55
Spines	[any]	6750	20433	14.55
Scale	[any]	1120	20504	14.55

Default value for the density of fuel particles (ρ_p) is $400 \text{ kg} \cdot \text{m}^{-3}$.

Table A.27: Default wood carbon content by taxonomic family.

	WoodC
Acanthaceae	0.4242167
Altingiaceae	0.4435000
Amaranthaceae	0.4197662
Amaryllidaceae	0.4508400
Anacardiaceae	0.4566867
Annonaceae	0.4726000
Apiaceae	0.4413076
Apocynaceae	0.4924750
Aquifoliaceae	0.4400000
Araliaceae	0.4540667
Arecaceae	0.4556000
Asparagaceae	0.4631000
Asteraceae	0.4375956
Betulaceae	0.4721962
Bignoniaceae	0.4618077
Boraginaceae	0.4153500
Brassicaceae	0.4274775
Burseraceae	0.4548544
Calophyllaceae	0.4663000
Campanulaceae	0.4498475
Cannabaceae	0.4683583
Caprifoliaceae	0.4475552
Caryophyllaceae	0.4147683
Casuarinaceae	0.4200000
Celastraceae	0.4900000
Chrysobalanaceae	0.4886000
Cistaceae	0.4698925
Clethraceae	0.4400000
Combretaceae	0.4678282
Convolvulaceae	0.4200000
Cordiaceae	0.4652500
Corynocarpaceae	0.4520000
Cupressaceae	0.5006151
Cyperaceae	0.4639737
Dipterocarpaceae	0.4744750
Ebenaceae	0.4828000
Ehretiaceae	0.4500000
Elaeocarpaceae	0.4350000
Ericaceae	0.4865107
Euphorbiaceae	0.4715052
Fabaceae	0.4559793
Fagaceae	0.4642838
Gentianaceae	0.4654867
Geraniaceae	0.4381400
Haematoxilaceae	0.4400000
Hypericaceae	0.4547167
Juglandaceae	0.4898900
Juncaceae	0.4238725
Juncaginaceae	0.4056355
Lamiaceae	0.4650444
Lauraceae	0.4677907
Lecythidaceae	0.4618750
Lentibulariaceae	0.4450200
Loganiaceae	0.4943000
Lythraceae	0.4219000

A.4 Scaling size-related parameters

A.4.1 Stem xylem maximum conductance

Estimation of maximum stem conductance ($k_{s,max}$, in $\text{mmol} \cdot \text{m}^{-2} \cdot \text{s}^{-1} \cdot \text{MPa}^{-1}$) is done by function `hydraulics_maximumStemHydraulicConductance()` and follows the work by Savage *et al.* (2010), Olson *et al.* (2014) and Christoffersen *et al.* (2016). Calculations are based on tree height and two species-specific parameters: maximum sapwood reference conductivity ($K_{s,max,ref}$) and the ratio of leaf area to sapwood area (A_l/A_s ; `A12As` in `SpParamsMED`), i.e. the inverse of the Huber value H_v .

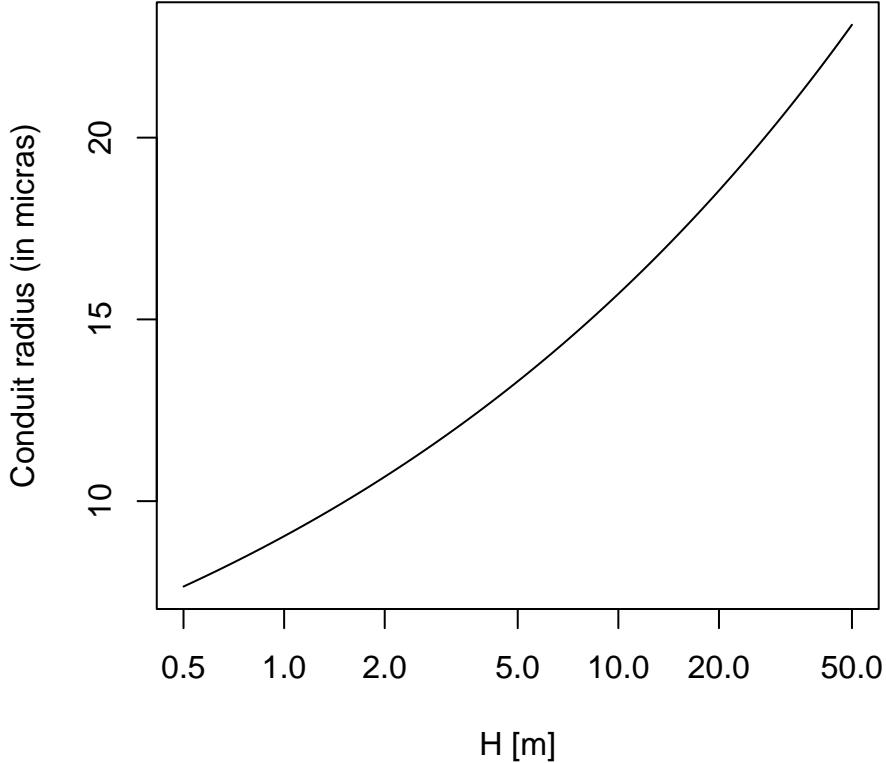
The reference value for maximum sapwood conductivity $K_{s,max,ref}$ is assumed to have been measured on a *terminal branch* of a plant of known height H_{ref} . If our target plant is very different in height, the conduits of terminal branches will have different radius and hence conductivity. We correct the reference conductivity to the target plant height using the following empirical relationship, developed by Olson *et al.* (2014) between tree height and diameter of conduits for angiosperms and the equation described by Christoffersen *et al.* (2016):

$$2 \cdot r_{int,H} = 10^{1.257 + (0.24 \cdot \log_{10}(H))} \quad (\text{A.12})$$

$$2 \cdot r_{int,ref} = 10^{1.257 + (0.24 \cdot \log_{10}(H_{ref}))} \quad (\text{A.13})$$

$$K_{s,max,cor} = K_{s,max,ref} \cdot (r_{int,H}/r_{int,ref})^2 \quad (\text{A.14})$$

Where $r_{int,H}$ is the radius of conduits for a terminal branch of a tree of height H and $r_{int,ref}$ is the corresponding radius for a tree of height H_{ref} (H and H_{ref} are measured in m). The form of the empirical relationship by Olson *et al.* (2014) is:



Let's consider an example for a *Quercus ilex* target tree of 4m height and where species-specific conductivity $K_{s,max,ref} = 0.77$ is the apical value for trees of $H_{ref} = 6.6$ m (in `medfate`, values of H_{ref} are taken from median height values; see parameter `Hmed` in `SpParamsMED`). The corrected conductivity for a tree of height 4 m will be a bit lower than that of the reference height:

```

xylem_kmax = 0.77
H = 400 # in cm
Href = 660 # in cm
f = hydraulics_referenceConductivityHeightFactor(Href, H);
f

## [1] 0.7863352

xylem_kmax_cor = xylem_kmax * f
xylem_kmax_cor

## [1] 0.6054781

```

Once the reference conductivity is corrected, the maximum stem conductance without accounting for conduit taper is:

$$k_{s,max,notaper} = \frac{1000}{0.018} \frac{K_{s,max,cor} \cdot A_s}{H \cdot A_l} \quad (\text{A.15})$$

where H is the tree height (here in m), A_s is the sapwood area, A_l is the leaf area and $1000/0.018$ is a factor used to go from kg to mmol. The ratio $A_l/A_s = 1/H_v$ is a fixed species parameter in soil water balance calculations (see parameter `A12As`), but becomes variable when simulating plant growth. Let's assume that *Quercus ilex* the leaf to sapwood area ratio is $A_l/A_s = 2512$. The maximum (leaf-specific) stem conductance without taper ($k_{s,max,notaper}$) for the tree of 4 m height is then:

```

A12As = 2512

kstemmax = hydraulics_maximumStemHydraulicConductance(xylem_kmax,
              Href, A12As, H, taper = FALSE)
kstemmax

## [1] 3.347698

```

In order to consider taper of xylem conduits we calculate the whole-tree conductance per unit leaf area ($k_{s,max,taper}$) as described in Christoffersen *et al.* (2016):

$$k_{s,max,taper} = \frac{1000}{0.018} \cdot \frac{K_{s,max,pet} \cdot A_s}{H \cdot A_l} \cdot \chi_{\text{tap:notap,ag}}(H) \quad (\text{A.16})$$

where $K_{s,max,pet}$ is the conductivity at the petiole level and $\chi_{\text{tap:notap,ag}}(H)$ is the taper factor accounting for the decrease in the xylem conduits diameter with the height, from the petiole to base of the trunk, which mitigates the negative effects of height on the hydraulic safety. The conductivity at the petiole level is obtained from $K_{s,max,ref}$ using again:

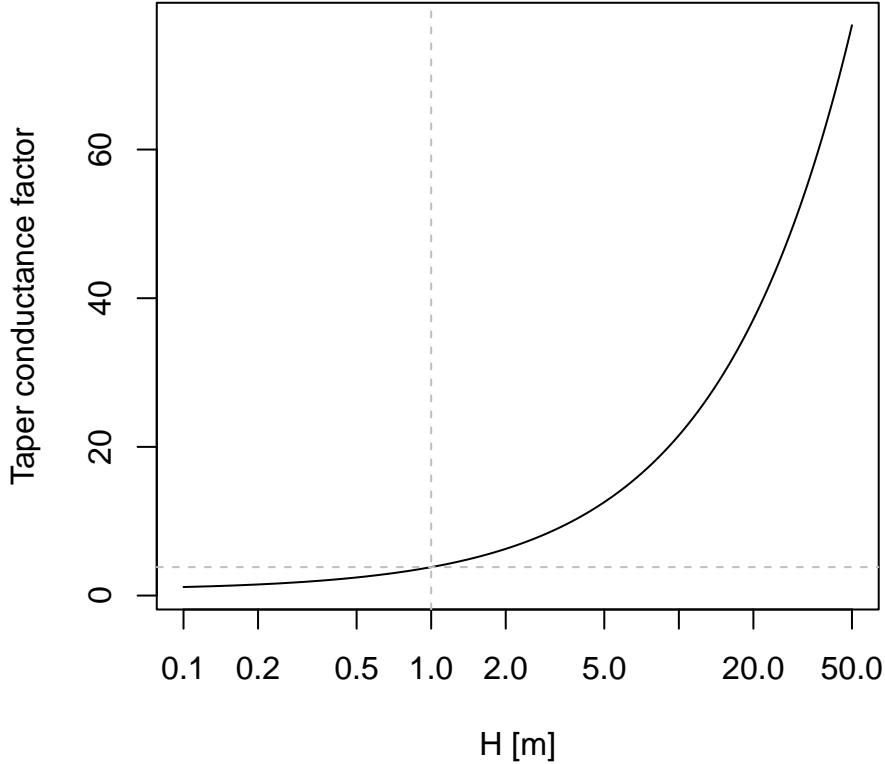
$$K_{s,max,pet} = K_{s,max,ref} \cdot (r_{int,pet}/r_{int,ref})^2 \quad (\text{A.17})$$

where $r_{int,pet}$ is the radius of the petiole in the model of Savage *et al.* (2010). Christoffersen *et al.* (2016) use $r_{int,pet} = 10 \mu\text{m}$ but we define it as the radius of apical conduits in a tree of 1 m height:

```
hydraulics_terminalConduitRadius(100.0)
```

```
## [1] 9.035871
```

$\chi_{\text{tap:notap,ag}}(H)$ is calculated as described in the Appendix 1 section of Christoffersen *et al.* (2016) (see also Savage *et al.* (2010)). The following figure shows the value of $\chi_{\text{tap:notap,ag}}$ for different heights:



Note that, since $\chi_{tap:notap,ag}(1) = 3.82$ (indicated using grey dashed lines in the last figure), the equation of maximum conductance with taper would give a higher conductance than the equation without taper for a tree of 1 m height, which is supposed to have a conductance equal to conductivity. To solve this issue we define the taper factor as $\chi_{tap:notap,ag}(H)/\chi_{tap:notap,ag}(1)$:

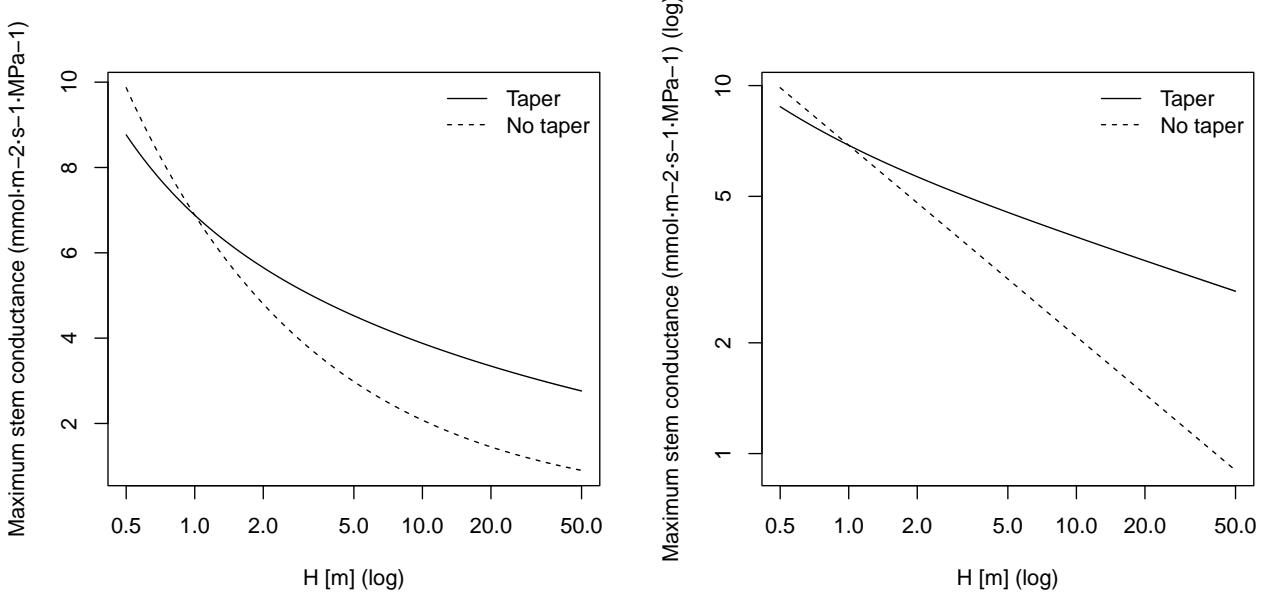
$$k_{s,max,taper} = \frac{1000}{0.018} \cdot \frac{K_{s,max,pet} \cdot A_s}{H \cdot A_l} \cdot \frac{\chi_{tap:notap,ag}(H)}{\chi_{tap:notap,ag}(1)} \quad (\text{A.18})$$

The maximum stem conductance with taper ($k_{s,max,taper}$) of a *Q. ilex* tree of 4 m height, calculated with this second equation, is:

```
kstemmax_tap = hydraulics_maximumStemHydraulicConductance(xylem_kmax,
                                                               Href, Al2As, H, taper = TRUE)
kstemmax_tap
```

```
## [1] 4.764396
```

The next two plots show the variation of $k_{s,max}$ for *Q. ilex* depending on the tree height and with/without considering taper of conduits. The plot on the right (both axes in log) show the slope of the dependency of conductance with height in both cases:



A.4.2 Leaf maximum hydraulic conductance

A.4.3 Root xylem maximum hydraulic conductance

To obtain maximum root xylem conductance ($k_{r,max}$, in $mmol \cdot m^{-2} \cdot s^{-1} \cdot MPa^{-1}$), one option taken by Christoffersen *et al.* (2016) is to assume that minimum stem resistance (inverse of maximum conductance) represents a fixed proportion of the minimum total tree (stem+root) resistance. A value 0.625 (i.e. 62.5%) suggested by these authors leads to maximum total tree conductance for our *Q. ilex* tree being:

```
ktot = kstemmax*0.625
ktot
```

```
## [1] 2.092311
```

and the maximum root xylem conductance would be therefore:

```
krootmax = 1/((1/ktot)-(1/kstemmax))
krootmax
```

```
## [1] 5.579497
```

Now, we need to divide total maximum conductance of the root system xylem among soil layers we need weights inversely proportional to the length of transport distances (Sperry *et al.* 2017). Vertical transport lengths can be calculated from soil depths and radial spread can be calculated assuming cylinders with volume proportional to the proportions of fine root biomass. Let's assume a soil with three layers:

```
s = soil(defaultSoilParams(3))
d = s$dVec
d
```

```
## [1] 300 700 1000
```

The proportion of fine roots in each layer, assuming a linear dose response model, will be:

```
Z50 = 200
Z95 = 1500
v1 = root_ldrDistribution(Z50,Z95, d)
v1

##          [,1]      [,2]      [,3]
## [1,] 0.6661036 0.2784153 0.05548106
```

Having this information, the calculation of root length (i.e. the sum of vertical and radial lengths) to each layer (L_j) is done using function `root_coarseRootLengths()`:

```
rfc = c(20,50,70)
Vol = root_coarseRootSoilVolumeFromConductance(1.0, 2500,krootmax,
                                                v1, d, rfc)
l = root_coarseRootLengthsFromVolume(Vol, v1, d, rfc)
l

## [1] 4503.445 3250.363 3085.055
```

where lengths are in mm. The proportion of total root xylem conductance corresponding to each layer (w_j) is given by `root_xylemConductanceProportions()`:

```
w1 = hydraulics_rootxylemConductanceProportions(v1, l)

## [1] 0.09131268 0.15767645 0.75101087
```

Xylem conductance proportions can be quite different than the fine root biomass proportions. This is because radial lengths are largest for the first top layers and vertical lengths are largest for the bottom layers. The maximum root xylem conductances of each layer will be the product of maximum total conductance of root xylem and weights:

```
w1*krootmax

## [1] 0.5094788 0.8797553 4.1902629
```

The maximum root xylem conductances of each layer would be:

```
krootmaxvec = w1*krootmax
krootmaxvec

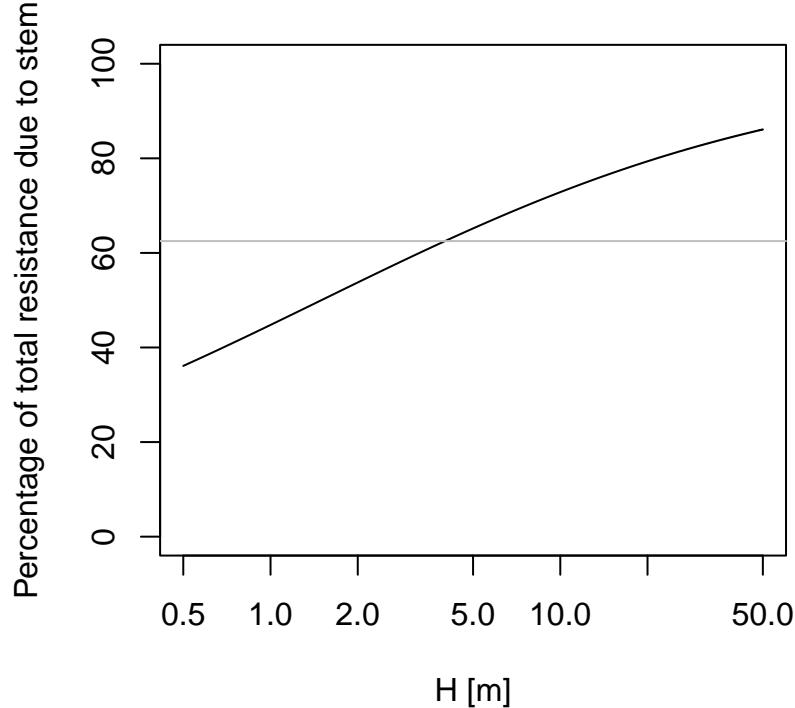
## [1] 0.5094788 0.8797553 4.1902629
```

and the fraction of total xylem resistance due to stem would be:

```
(1/kstemmax)/((1/kstemmax)+(1/krootmax))

## [1] 0.625
```

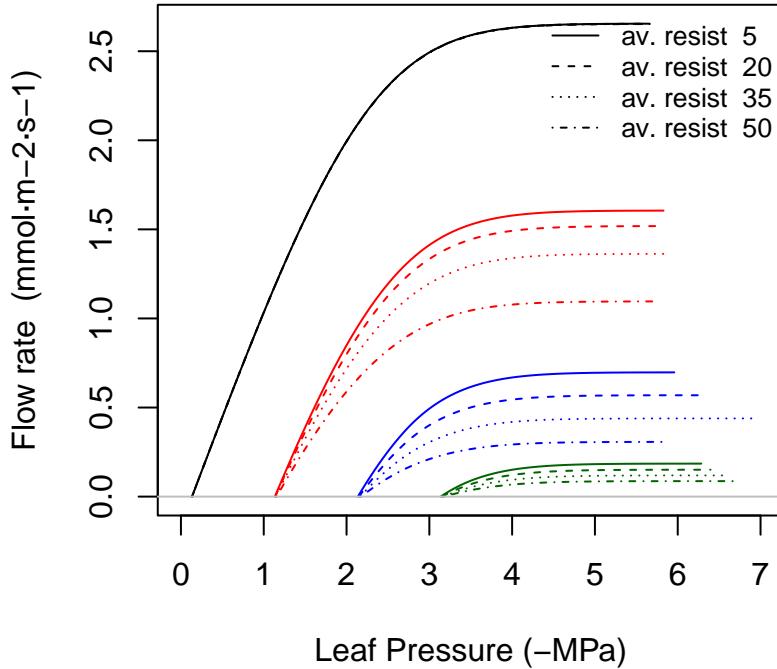
In contrast with the approach of Christoffersen *et al.* (2016), in this approach the root maximum conductance depends root length and distribution, and is not a fixed fraction of stem maximum conductance. Assuming constant root length, then the proportion of total resistance due to the stem will increase with tree height (Magnani *et al.* 2000):



where the horizontal gray line indicates the value of 62.5%. Of course rooting depth also increases with tree age, but young trees have higher root-to-shoot ratios than older ones. Hence, a root maximum conductance that is not fixed but increases with age seems a priori more realistic. Moreover, Christoffersen *et al.* (2016) justify the value of 62.5% from a study which quantified total aboveground and belowground resistance in tropical trees (Fisher *et al.* 2006) under near-saturated (wet season) conditions, but values of belowground resistance reported in this study for wet conditions and trees of 30 m height are around 13%, which equals to 87% fraction of aboveground resistance. On the other hand, while rooting depths are limited by soil depth, lateral root length increases with age and, hence, the model could be made more realistic if this is taken into account and the curve above would probably saturate at lower percentages.

A.4.4 Rhizosphere maximum hydraulic conductance

Maximum rhizosphere conductance ($k_{rh,max}$, in $\text{mmol} \cdot \text{m}^{-2} \cdot \text{s}^{-1} \cdot \text{MPa}^{-1}$) is difficult to measure directly, as it depends on the rhizosphere (i.e. fine root) surface in each soil layer, and will probably always be a parameter to be calibrated. Instead of trying to estimate rhizosphere surface from root architecture (Sperry *et al.* 1998), we follow Sperry *et al.* (2016) and determine the maximum rhizosphere conductance in each layer from an inputted ‘average percentage rhizosphere resistance’. The percentage of continuum resistance corresponding to the rhizosphere is calculated from the vulnerability curves of stem, root and rhizosphere at the same water potential. The average resistance is found by evaluating the percentage for water potential values between 0 and Ψ_{crit} . The following figure illustrates how the supply function, for different soil water potentials, is affected by increasing values of the average percentage of rhizosphere resistance:



Sperry *et al.* (2016) found average percentages of rhizosphere resistance around 67%, but these exceptionally-high values were probably a consequence of using an unsegmented supply function (i.e. single vulnerability curve for roots, stem and leaves). If we specify a 15% of average resistance in the rhizosphere (see parameter `averageFracRhizosphereResistance` in function `defaultControl()`), the maximum rhizosphere conductance values for the three layers are found calling:

```

krmax = rep(0,3)
krmax[1] = hydraulics_findRhizosphereMaximumConductance(15,
  s$VG_n[1],s$VG_alpha[1],
  krootmax, rootc,rootd,
  kstemmax, stemc, stemd,
  kleafmax, leafc, leafd)
krmax[2] = hydraulics_findRhizosphereMaximumConductance(15,
  s$VG_n[2],s$VG_alpha[2],
  krootmax, rootc,rootd,
  kstemmax, stemc, stemd,
  kleafmax, leafc, leafd)
krmax[3] = hydraulics_findRhizosphereMaximumConductance(15,
  s$VG_n[3],s$VG_alpha[3],
  krootmax, rootc,rootd,
  kstemmax, stemc, stemd,
  kleafmax, leafc, leafd)
krmax

## [1] 240174188 85636744 85636744

```

The values are the same because the texture of the three layers is the same in this case. If we take into account root distribution, actual maximum rhizosphere conductance values are:

```
krmax*v1

##           [,1]      [,2]      [,3]
## [1,] 159980896 23842581 4751218
```

A.4.5 Plant water storage capacity

The water storage capacity of sapwood tissue per leaf area unit ($V_{sapwood}$; in $l \cdot m^{-2}$) can be estimated as the product of stem height (H in m) and Huber value (H_v ; ratio of sapwood area to leaf area in $m^2 \cdot m^{-2}$) times a factor to account for the non-cylindrical shape (<http://www.fao.org/forestry/17109/en/>):

$$V_{sapwood} = 10^3 \cdot 0.48 \cdot H \cdot H_v \cdot \Theta_{sapwood} \quad (\text{A.19})$$

$\Theta_{sapwood}$ is sapwood porosity (cm^3 of water per cm^3 of sapwood tissue), which can be estimated from wood density (ρ_{wood} ; in $g \cdot cm^{-3}$):

$$\Theta_{sapwood} = 1 - (\rho_{wood}/1.54) \quad (\text{A.20})$$

where the density of wood substance can be assumed to be fixed and equal to $1.54 g \cdot cm^{-3}$ (Dunlap 1914). For example, wood densities ranging from 0.443 to $1.000 g \cdot cm^{-3}$ result in sapwood porosity values between 0.35 and 0.71 .

Water storage capacity of leaf tissue per leaf area unit (V_{leaf} ; in $l \cdot m^{-2}$) can be estimated as the product of specific leaf area (SLA; in $m^2 \cdot kg^{-1}$) and leaf density (ρ_{leaf} ; in $g \cdot cm^{-3}$):

$$V_{leaf} = \frac{1}{SLA \cdot \rho_{leaf}} \cdot \Theta_{leaf} \quad (\text{A.21})$$

Θ_{leaf} is leaf porosity (cm^3 of water per cm^3 of leaf tissue), which can be estimated from leaf density:

$$\Theta_{leaf} = 1 - (\rho_{leaf}/1.54) \quad (\text{A.22})$$

where the density of wood substance can be assumed to be fixed and equal to $1.54 g \cdot cm^{-3}$ (Dunlap 1914).

For example, let's calculate the stem and leaf water capacity for a Q. ilex tree of 15 m height:

```
ld = 0.7
SLA = 5.870
moisture_leafWaterCapacity(SLA, ld)
```

```
## [1] 0.1327463
```

A.4.6 Horizontal root overlap

A.4.6.1 Basic water balance

A given plant cohort i will have its roots in layer s partitioned among different water pools. We thus need to define $fr_{i,s,j}$, the (horizontal) proportion of fine roots of cohort i in layer s of the water pool j , with the restriction that:

$$\sum_j fr_{i,s,j} = 1 \quad \forall i, s \quad (\text{A.23})$$

It is important to realize that proper estimation of $fr_{i,s,j}$ is challenging when we do not have explicit plant coordinates, root lateral widths, etc. For this reason, an intuitive approach is followed here based on the following two premises:

- The amount of overlap between roots of different plants should monotonically increase along with the LAI of the stand (i.e. LAI_{stand}^{live}).
- The amount of overlap between roots of different plants at a given soil depth should increase/decrease with the vertical proportion of roots at that depth.

The specific formulation we chose for $fr_{i,s,j}$ is:

$$fr_{i,s,j} = f_{pool,j} \cdot (1 - \exp(-f_{overlap} \cdot LAI_{stand}^{live})) \cdot \left(\frac{\sqrt{FRP_{i,s} \cdot FRP_{j,s}}}{\max_s(FRP_{i,s})} \right) \quad (\text{A.24})$$

for all $j \neq i$, where $f_{overlap}$ is an overlap factor (a control parameter called `poolOverlapFactor`). For $j = i$ then we simply have:

$$fr_{i,s,i} = 1 - \sum_j fr_{i,s,j} \quad (\text{A.25})$$

Note that if $f_{overlap} = 0$ then $fr_{i,s,j} = 1$ if $j = i$ and zero otherwise (i.e. plants exploit their corresponding water pools only). For very large values of LAI_{stand}^{live} and/or $f_{overlap}$ we have that $(1 - \exp(-f_{overlap} \cdot LAI_{stand}^{live})) = 0$ and $fr_{i,s,j} = f_{pool,j}$ (neglecting vertical differences), so that plants exploit the each water pool in the same proportion as the fraction of stand occupied by the pool (i.e. overlap is maximum).

Appendix B

Symbols

The following tables list all symbols used in this document, along with their units and definition. When symbols are input for **medfate** model functions, the R name of those parameters in the package (either in data frame `SpParamsMED`, soil input data frame, or the output of functions such as `spwbInput()`) is also indicated.

B.1 Dimensions

Symbol	Units	R	Description
Δt_{step}	s	86400/ndailysteps	Temporal step
$\Delta t_{substep}$	s	86400/(ndailysteps*ndailysubsteps)	Daily substep
c	(count)		Number of plant cohorts
l	(count)		Number of vertical layers
Δz	m	verticalLayerSizeWidth	Width of vertical layers
S	(count)		Number of soil layers
d_s	mm	widths	Width of soil layer s
Z_s	mm	widths	Depth of soil layer s
Z_{soil}	mm		Overall soil depth

B.2 Soils

Symbol	Units	R	Description
$P_{clay,s}$	%	clay	Percent of clay in soil layer s
$P_{sand,s}$	%	sand	Percent of sand in soil layer s
OM_s	%	om	Percentage of organic mater per dry weight in soil layer s
BD_s	$g \cdot cm^{-3}$	bd	Bulk density in soil layer s
$P_{rocks,s}$	%	rfc	Percentage of rock fragment content in soil layer s
θ_s	$m^3 \cdot m^{-3}$		Volumetric moisture in soil layer s
Ψ_s	MPa		Water potential in soil layer s
Ψ_{fc}	MPa		Water potential at field capacity
A_s, B_s			Parameters of the Saxton pedotransfer functions for soil layer s
$\theta_{sat,s}$	$m^3 \cdot m^{-3}$	VG_theta_sat	Volumetric moisture at saturation for soil layer s

Symbol	Units	R	Description
$\theta_{fc,s}$	$m^3 \cdot m^{-3}$		Volumetric moisture at field capacity (-0.033 MPa) for soil layer s
$\theta_{wp,s}$	$m^3 \cdot m^{-3}$		Volumetric moisture at wilting point (-1.5 MPa) for soil layer s
$\theta_{res,s}$	$m^3 \cdot m^{-3}$	VG_theta_res	Residual volumetric moisture for soil layer s
n_s		VG_n	Parameter n of the Van Genuchten (1980) model for soil layer s
α_s		VG_alpha	Parameter α of the Van Genuchten (1980) model for soil layer s
$K_{sat,s}$	$mmol \cdot s^{-1} \cdot m^{-1} \cdot MPa^{-1}$	Ksat	Saturated soil conductivity for soil layer s .
$K_{unsat,s}$	$mmol \cdot s^{-1} \cdot m^{-1} \cdot MPa^{-1}$	Kunsat	Unsaturated soil conductivity for soil layer s .
K_{drain}	$mm \cdot d^{-1}$	Kdrain	Hydraulic conductivity for deep drainage.
$P_{macro,s}$	%	macro	Percentage of macroporosity corresponding to soil layer s .
γ_{soil}	$mm \cdot day^{-1}$	Gsoil	The maximum daily evaporation from soil
κ_{soil}	(unitless)	Ksoil	Extinction parameter to regulate the amount of water evaporated from each soil layer
Z_{wt}	mm		Water table depth
V_s	mm		Water volume in soil layer s
V_{soil}	mm		Overall water volume in the soil
$V_{fc,soil}$	mm		Water retention capacity of the whole soil
W_s	[0-1]	W	Proportion of moisture in relation to field capacity in soil layer s
$W_{i,s}$	[0-1]	W	Proportion of moisture in relation to field capacity in soil layer s within fraction of stand area covered by cohort i
$W_{rhizo,i,s}$	[0-1]		Proportion of moisture in relation to field capacity in soil layer s within the rhizosphere of cohort i
S_{snow}	mm	SWE	Snow water equivalent of the snow pack storage over the soil surface

B.3 Plant/leaf classification

Symbol	Units	R	Description
GF	Categorical	GrowthForm	“Tree”, “Shrub” or “Tree/Shrub”
LF	Categorical	LifeForm	Raunkiaer life form
L_{shape}	Categorical	LeafShape	Leaf shape: “Broad”, “Needle”, “Linear”, “Scale”, “Spines” or “Succulent”
L_{size}	Categorical	LeafSize	Leaf size: “Small” (< 225 mm), “Medium” (> 225 mm & < 2025 mm) or “Large” (> 2025 mm)
L_{pheno}	Categorical	PhenologyType	Leaf phenology type

B.4 Vegetation

Symbol	Units	R	Description
SP_i	(count)	Species	Species identity of cohort i
$A_{sh,i}$	cm^2		Area occupied by an average shrub individual of cohort i
$B_{sh,i}$	kg		Fine-fuel biomass of an average shrub individual of cohort i
H_i	cm	Height	Average tree or shrub height of cohort i
$H_{crown,i}$	cm		Crown base height (i.e. the height corresponding to the first living branch) of cohort i
CCF_i			Crown competition factor of cohort i
CW_i	m		Crown width that a tree of cohort i would have in open-ground conditions
CR_i	[0-1]	CR	Crown ratio (i.e. ratio between crown length and plant height) of cohort i
N_i	$ind \cdot ha^{-1}$	N	Density of tree individuals of cohort i
DBH_i	cm	DBH	Diameter at breast height of trees in cohort i
$Cover_i$	%	Cover	Percent cover of shrubs in cohort i
H_{herb}	%	herbHeight	Average height (in cm) of herbaceous vegetation
$Cover_{herb}$	%	herbCover	Percent cover of herbaceous vegetation
FB_i	$kg \cdot m^{-2}$		Foliar biomass of cohort i
$FB_{tree,i}$	kg		Foliar biomass of a single tree of cohort i
ϕ_i	[0-1]		Fraction of maximum leaf area currently expanded for cohort i
LAI_i^{live}	$m^2 \cdot m^{-2}$	LAI_live	(Maximum) leaf area index (one-side leaf area per surface area of the stand) of woody cohort i
LAI_i^{dead}	$m^2 \cdot m^{-2}$	LAI_dead	Dead leaf area index (one-side dead leaf area per surface area of the stand) of woody cohort i
LAI_i^ϕ	$m^2 \cdot m^{-2}$	LAI_expanded	Current expanded leaf area index (one-side expanded leaf area per surface area of the stand) of woody cohort i
LAI_i^{all}	$m^2 \cdot m^{-2}$		Total leaf area index (live or dead) of woody cohort i
LAI_{woody}^{live}	$m^2 \cdot m^{-2}$		Maximum leaf area index of living leaves of woody vegetation in the stand
LAI_{woody}^{dead}	$m^2 \cdot m^{-2}$		Leaf area index of dead leaves of woody vegetation in the stand
LAI_{woody}^ϕ	$m^2 \cdot m^{-2}$		Current expanded leaf area index of live leaves of woody vegetation in the stand
LAI_{woody}	$m^2 \cdot m^{-2}$		Total leaf area index (live or dead) of woody vegetation in the stand
LAI_{herb}	$m^2 \cdot m^{-2}$		Leaf area index of herbaceous vegetation in the stand
LAI_{stand}	$m^2 \cdot m^{-2}$		Total leaf area index of the stand
LA_i	m^2		Leaf area of an individual of cohort i
$FRP_{i,s}$	[0-1]	V[i,s]	The proportion of fine roots of cohort i in soil layer s
$FRA_{i,s}$	m^2		Fine root area of cohort i in soil layer s
$L_{radial,i,s}$	m		Radial length of coarse roots of cohort i in soil layer s
W_i	$kg \cdot m^{-2}$		Fine fuel biomass of cohort i
$Z_{50,i}$	mm	Z50	Depth above which 50% of the fine root mass of cohort i is located
$Z_{95,i}$	mm	Z95	Depth above which 95% of the fine root mass of cohort i is located

B.5 Meteorology

Symbol	Units	R	Description
DOY		DOY	Day of the year
T_{mean}	$^{\circ}\text{C}$	MeanTemperature	Mean daily temperature
T_{min}	$^{\circ}\text{C}$	MinTemperature	Minimum daily temperature
T_{max}	$^{\circ}\text{C}$	MaxTemperature	Maximum daily temperature
T_{base}	$^{\circ}\text{C}$		Base temperature for calculation of GDD
RH_{min}	%	MinRelativeHumidity	Minimum daily relative humidity
RH_{max}	%	MaxRelativeHumidity	Maximum daily relative humidity
P	$L \cdot m^{-2} = mm$	Precipitation	Precipitation (including rainfall and snow)
Pr	$L \cdot m^{-2} = mm$	Rainfall	Liquid water precipitation (rainfall)
Ps	$L \cdot m^{-2} = mm$	Snow	Snow precipitation
PET	$L \cdot m^{-2} = mm$	PET	Potential evapotranspiration, preferably calculated using Penman's equation
Rad	$MJ \cdot m^{-2}$	Radiation	Solar radiation after accounting for clouds
u	$m \cdot s^{-1}$	WindSpeed	Wind speed
P_{atm}	kPa		Atmospheric pressure
ρ_{air}	$kg \cdot m^{-3}$		Air density
T_{atm}	$^{\circ}\text{C}$	Tatm	Atmospheric (above-canopy) air temperature
T_{can}	$^{\circ}\text{C}$	Tcan	Canopy air temperature
$T_{air,j}$	$^{\circ}\text{C}$	Tair	Air temperature in canopy layer j
$T_{soil,s}$	$^{\circ}\text{C}$	Tsoil.s	Temperature of soil layer s
e_{atm}	kPa		Atmospheric (above-canopy) water vapor pressure
$e_{air,j}$	kPa	VPair	Water vapor pressure in canopy layer j
C_{atm}	kPa	Catm	Atmospheric (above-canopy) CO_2 concentration
$C_{air,j}$	kPa	VPair	CO_2 concentration in canopy layer j
u_j	$m \cdot s^{-1}$		Wind speed at canopy layer j

B.6 Anatomy

Symbol	Units	R	Description
H_v	$m^2 \cdot m^{-2}$	1/Al2As	Huber value (ratio of sapwood area to leaf area)
LW	cm	LeafWidth	Leaf width
SLA	$m^2 \cdot kg^{-1}$	SLA	Specific leaf area
ρ_{leaf}	$g \cdot cm^{-3}$	LeafDensity	Leaf tissue density
ρ_{wood}	$g \cdot cm^{-3}$	WoodDensity	Wood tissue density
$\rho_{fineroot}$	$g \cdot cm^{-3}$	FineRootDensity	Fine root tissue density
$\Theta_{sapwood}$	$m^3 \cdot m^{-3}$		Sapwood porosity (volume of empty spaces over total volume)
Θ_{leaf}	$m^3 \cdot m^{-3}$		Leaf porosity (volume of empty spaces over total volume)
SRL	$cm \cdot g^{-1}$	SRL	Specific root length
RLD	$cm \cdot cm^{-3}$	RLD	Fine root length density (i.e. density of root length per soil volume)
$r_{6.35}$		r635	Ratio between the weight of leaves plus branches and the weight of leaves alone for branches of 6.35 mm

B.7 Radiation

Symbol	Units	R	Description
$k_{PAR,i}$	(unitless)	kPAR	PAR extinction coefficient for cohort i
$k_{SWR,i}$	(unitless)		SWR extinction coefficient for cohort i
k_{LWR}	(unitless)		LWR extinction coefficient
k_b	(unitless)		Extinction coefficient for direct light for cohort i
$k_{d,i}$	(unitless)		Extinction coefficient for diffuse light for cohort i (equal to $k_{PAR,i}$ or $k_{SWR,i}$)
$\alpha_{SWR,i}$	[0-1]	alphaSWR	Short-wave radiation absorbance coefficient for cohort i
$\alpha_{PAR,i}$	[0-1]		PAR absorbance coefficient for cohort i
$\gamma_{SWR,i}$	[0-1]	gammaSWR	Short-wave radiation leaf reflectance (albedo) for cohort i
$\gamma_{SWR,soil}$	[0-1]		Short-wave radiation soil reflectance
$\gamma_{PAR,i}$	[0-1]		PAR leaf reflectance for cohort i
L_{PAR}^{ground}	[0-1]	LgroundPAR	Fraction of PAR reaching the ground
L_{SWR}^{ground}	[0-1]	LgroundSWR	Fraction of SWR reaching the ground
I_{beam}	$W\text{ um}^{-2}$		Instantaneous direct shortwave irradiance from the atmosphere
I_{dif}	$W\text{ um}^{-2}$		Instantaneous diffuse shortwave irradiance from the atmosphere
L_{atm}	$W\text{ um}^{-2}$		Instantaneous longwave radiation from the atmosphere
$I_{beam,j}$	$W\text{ um}^{-2}$		Instantaneous direct shortwave irradiance at the top of canopy layer j
$I_{dif,j}$	$W\text{ um}^{-2}$		Instantaneous diffuse shortwave irradiance at the top of canopy layer j
$I_{beam,soil}$	$W\text{ um}^{-2}$		Instantaneous direct shortwave irradiance reaching the soil
$I_{dif,soil}$	$W\text{ um}^{-2}$		Instantaneous diffuse shortwave irradiance reaching the soil
$LAI_{i,j}^{sunlit}$	$m^2\text{ um}^{-2}$		Leaf area index of sunlit leaves of cohort i in layer j .
$LAI_{i,j}^{shade}$	$m^2\text{ um}^{-2}$		Leaf area index of shade leaves of cohort i in layer j .
$\Phi_{i,j}^{sunlit}$	$W\text{ um}^{-2}$		Short-wave radiation absorbed by sunlit leaves of cohort i in layer j , per leaf area unit.
$\Phi_{i,j}^{shade}$	$W\text{ um}^{-2}$		Short-wave radiation absorbed by sunlit leaves of cohort i in layer j , per leaf area unit.
$K_{abs,i,j}^{sunlit}$	$W\text{ um}^{-2}$		Short-wave radiation absorbed by sunlit foliage of cohort i in layer j , per ground area unit.
$K_{abs,i,j}^{shade}$	$W\text{ um}^{-2}$		Short-wave radiation absorbed by sunlit foliage of cohort i in layer j , per ground area unit.

B.8 Water balance

Symbol	Units	R	Description
P_s	mm	Snow	Precipitation as snow
P_r	mm	Rain	Precipitation as rainfall
Sm	mm	Snowmelt	Snowmelt
P_{net}	mm	NetRain	Net rainfall
In	mm	Interception	Interception loss
Ru	mm	Runoff	Water exported from the stand as runoff
Ro	mm	Runon	Water imported to the stand as runon

Symbol	Units	R	Description
Dd	mm	DeepDrainage	Water exported from the stand as deep drainage
Es	mm	SoilEvaporation	Evaporation from the soil surface
Tr_{woody}	mm	Transpiration	Woody plant transpiration
Tr_{herb}	mm	HerbTranspiration	Herbaceous plant transpiration
Tr_i	mm		Transpiration of woody plant cohort i

B.9 Energy balance

Symbol	Units	R	Description
$K_{abs,can}$	$W \cdot m^{-2}$	SWRcan	Atmosphere shortwave radiation absorbed by the canopy
$K_{abs,j}$	$W \cdot m^{-2}$		Atmosphere shortwave radiation absorbed by canopy layer j
$K_{abs,soil}$	$W \cdot m^{-2}$	SWRsoil	Atmosphere shortwave radiation absorbed by the soil
$L_{net,can}$	$W \cdot m^{-2}$	LWRcan	Canopy net longwave radiation
$L_{net,j}$	$W \cdot m^{-2}$		Net long-wave radiation of canopy layer j
$L_{net,soil}$	$W \cdot m^{-2}$	LWRsoil	Soil net longwave radiation
$H_{can,atm}$	$W \cdot m^{-2}$	Hcan	Turbulent heat exchange between the canopy and the atmosphere
$H_{can,soil}$	$W \cdot m^{-2}$	Hcanoil	Turbulent heat exchange between the canopy and the soil
H_j	$W \cdot m^{-2}$		Sensible heat flux between canopy layer j and the leaves it contains
LE_{can}	$W \cdot m^{-2}$	LEcan	Energy released as latent heat from the canopy towards the atmosphere
LE_j	$W \cdot m^{-2}$		Energy released as latent heat by canopy layer j
LE_{soil}	$W \cdot m^{-2}$	LEsoil	Energy released as latent heat from the soil
TC_{LAI}	$J \cdot m^{-2} \cdot K^{-1}$	thermalcapacity _{LAI}	Canopy thermal capacitance per LAI unit
TC_{can}	$J \cdot m^{-2} \cdot K^{-1}$		Canopy thermal capacitance
TC_j	$J \cdot m^{-2} \cdot K^{-1}$		Thermal capacitance of canopy layer j
$TC_{soil,s}$	$J \cdot m^{-2} \cdot K^{-1}$		Thermal capacitance of soil layer s
$VHC_{soil,s}$	$J \cdot m^{-3} \cdot K^{-1}$		Volumetric heat capacity of soil in layer s

B.10 Forest hydrology

Symbol	Units	R	Description
P_G	mm		Amount of rainfall needed to saturate the canopy for a given event
S_{canopy}	mm	Cm	Canopy water storage capacity
ER_{ratio}	(unitless)		Ratio between evaporation rate and rainfall rate
C_{canopy}	[0-1]		Canopy cover
PE_{soil}	mm		Evaporation demand from the soil
SE_{soil}	mm		Evaporation supply from the soil

Symbol	Units	R	Description
t_{soil}	day		Time needed to evaporate the current water deficit in the first soil layer
PET_{herb}	mm		Potential evapotranspiration in the herb layer

B.11 Plant hydraulics

Symbol	Units	R	Description
$K_{stem,max,ref}$	$kg \cdot s^{-1} \cdot m^{-1} \cdot MPa^{-1}$	<code>Kmax_stemxylem</code>	Maximum stem sapwood reference conductivity per leaf area unit
$K_{root,max,ref}$	$kg \cdot s^{-1} \cdot m^{-1} \cdot MPa^{-1}$	<code>Kmax_rootxylem</code>	Maximum root sapwood reference conductivity per leaf area unit
$k_{stem,max}$	$mmol \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$	<code>VCstem_kmax</code>	Maximum whole-stem conductance (per leaf area unit)
$k_{root,max}$	$mmol \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$	<code>VCroot_kmax</code>	Maximum root conductance (per leaf area unit)
$k_{rhizo,max}$	$mmol \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$		Maximum rhizosphere conductance (per leaf area unit)
$k_{leaf,max}$	$mmol \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$	<code>VCleaf_kmax</code>	Maximum leaf conductance (per leaf area unit)
c_{leaf}, d_{leaf}	(unitless), MPa	<code>VCleaf_c</code> , <code>VCleaf_d</code>	Parameters of the vulnerability curve for leaves
c_{root}, d_{root}	(unitless), MPa	<code>VCroot_c</code> , <code>VCroot_d</code>	Parameters of the vulnerability curve for root xylem
c_{stem}, d_{stem}	(unitless), MPa	<code>VCstem_c</code> , <code>VCstem_d</code>	Parameters of the vulnerability curve for stem xylem
Ψ	MPa		Water potential in a given water compartment/segment
Ψ_P	MPa		Turgor water potential in a given water compartment/segment
Ψ_S	MPa		Osmotic (solute) water potential in a given water compartment
Ψ_{cav}	MPa		Minimum water potential experienced by xylem in previous steps (cavitation)
Ψ_{canopy}	MPa		Canopy water potential
Ψ_{leaf}	MPa		Leaf water potential
$\Psi_{rootcrown}$	MPa		Water potential at the root crown
Ψ_{stem}	MPa		Water potential at the end (highest part) of the stem
PLC	[0-1]		Proportion of conductance loss
PLC_{stem}	[0-1]		Proportion of conductance loss in stem xylem tissue
p_{root}	[0-1]	<code>pRootDisc</code>	Relative root conductance leading to hydraulic disconnection from a soil layer
E_i	$mmol \cdot s^{-1} \cdot m^{-2}$		Steady-state water flow through a hydraulic segment i

Symbol	Units	R	Description
k_i	$mmol \cdot s^{-1} \cdot m^{-2} \cdot MPa^{-1}$		Hydraulic conductance function for segment i
Ψ_{up}	MPa		Upstream water potential
Ψ_{down}	MPa		Downstream water potential

B.12 Photosynthesis

Symbol	Units	R	Description
WUE_{max}	$g C \cdot mm^{-1}$	WUE	Water use efficiency at VPD = 1kPa and without light or CO ₂ limitations
WUE_{PAR}		WUE_par	Coefficient describing the progressive decay of WUE with lower light levels
WUE_{CO2}		WUE_co2	Coefficient for WUE dependency on atmospheric CO ₂ concentration
WUE_{VPD}		WUE_vpd	Coefficient for WUE dependency on vapor pressure deficit
J_{max}	$\mu mol e \cdot m^{-2} \cdot s^{-1}$	J_{max}	Maximum rate of electron transport at current leaf temperature
$V_{max,298}$	$\mu mol CO_2 \cdot s^{-1} \cdot m^{-2}$	V_{max}	Rubisco's maximum carboxylation rate at current leaf temperature
$J_{max,298}$	$\mu mol e \cdot m^{-2} \cdot s^{-1}$	$J_{max,298}$	Maximum rate of electron transport at 298K
$V_{max,298}$	$\mu mol CO_2 \cdot s^{-1} \cdot m^{-2}$	$V_{max,298}$	Rubisco's maximum carboxylation rate at 298K
T_{leaf}	°C		Leaf temperature.
u_{leaf}	$m \cdot s^{-1}$		Leaf-level wind speed.
VPD_{leaf}	kPa		Leaf vapour pressure deficit.
g_w	$mol H_2O \cdot s^{-1} \cdot m^{-2}$		Leaf diffusive conductance to water vapor
g_c	$mol CO_2 \cdot s^{-1} \cdot m^{-2}$		Leaf diffusive conductance to carbon dioxide
g_{sw}	$mol H_2O \cdot s^{-1} \cdot m^{-2}$		Leaf stomatal conductance to water vapour
$g_{sw,min}$	$mol H_2O \cdot s^{-1} \cdot m^{-2}$	G_{swmin}	Minimum stomatal conductance to water vapour
$g_{sw,max}$	$mol H_2O \cdot s^{-1} \cdot m^{-2}$	G_{swmax}	Maximum stomatal conductance to water vapour
Φ_{PAR}^{leaf}	$W \cdot m^{-2}$		Photosynthetically active radiation absorbed per leaf area
Q_{PAR}^{leaf}	$\mu mol photon \cdot m^{-2} \cdot s^{-1}$		PAR photon flux density per leaf area
A_c	$\mu mol CO_2 \cdot s^{-1} \cdot m^{-2}$		Rubisco-limited photosynthesis rate
A_e	$\mu mol CO_2 \cdot s^{-1} \cdot m^{-2}$		Electron transport-limited photosynthesis rate
A	$\mu mol CO_2 \cdot s^{-1} \cdot m^{-2}$		Leaf gross photosynthesis rate
A_n	$\mu mol CO_2 \cdot s^{-1} \cdot m^{-2}$		Leaf net photosynthesis rate

B.13 Plant water content

Symbol	Units	R	Description
$LFMC_{\max}$	%	maxFMC	Maximum live fuel moisture content, corresponding to fine fuels (< 6.35 mm twigs and leaves).
ϵ_{leaf}	MPa	LeafEPS	Modulus of elasticity of leaves
ϵ_{stem}	MPa	StemEPS	Modulus of elasticity of symplastic xylem tissue
$\pi_{0,leaf}$	MPa	LeafPIO	Osmotic potential at full turgor of leaves
$\pi_{0,stem}$	MPa	StemPIO	Osmotic potential at full turgor of symplastic xylem tissue
RWC	[0-1]		Relative water content
RWC_{sym}	[0-1]		Relative water content in the symplasm fraction of a tissue
RWC_{apo}	[0-1]		Relative water content in the apoplasm fraction of a tissue
$V_{segment}$	$l \cdot m^{-2}$		Water capacity of a segment (leaf or stem)
V_{leaf}	$l \cdot m^{-2}$	Vleaf	Leaf water capacity per leaf area unit
$V_{sapwood}$	$l \cdot m^{-2}$	Vsapwood	Sapwood water capacity per leaf area unit

B.14 Stomatal regulation

Symbol	Units	R	Description
$\theta_1(\Psi_{leaf})$, $\theta_2(\Psi_{leaf})$	[0-1]		Cost functions 1 and 2
$\beta(\Psi_{leaf})$	[0-1]		Gain function
$Profit_1(\Psi_{leaf})$, $Profit_2(\Psi_{leaf})$	[0-1]		Profit functions 1 and 2
E_{sunlit}	$mmol \cdot s^{-1} \cdot m^{-2}$		Instantaneous transpiration flow rate for sunlit leaves
E_{shade}	$mmol \cdot s^{-1} \cdot m^{-2}$		Instantaneous transpiration flow rate for shade leaves

B.15 Carbon pools

Symbol	Units	R	Description
$V_{storage,leaf,i}$	$L \cdot ind^{-1}$		Leaf (carbon) storage volume for an individual of cohort i
$V_{storage,sapwood,i}$	$L \cdot ind^{-1}$		Sapwood (carbon) storage volume for an individual of cohort i
$SS_{leaf,i}$	$mol \text{ gluc} \cdot L^{-1}$	SugarLeaf	Sugar concentration in leaves of cohort i
$SS_{sapwood,i}$	$mol \text{ gluc} \cdot L^{-1}$	SugarSapwood	Sugar concentration in sapwood of cohort i
$ST_{leaf,i}$	$mol \text{ gluc} \cdot L^{-1}$	StarchLeaf	Starch concentration in leaves of cohort i
$ST_{sapwood,i}$	$mol \text{ gluc} \cdot L^{-1}$	StarchSapwood	Starch concentration in sapwood of cohort i
S_{plant}	$mol \text{ gluc} \cdot ind^{-1}$		Total labile carbon storage in a plant individual

Symbol	Units	R	Description
B_{leaf}	$g \text{ dry}\cdot\text{ind}^{-1}$		Leaf dry biomass in a plant individual (for respiration costs)
$B_{sapwood}$	$g \text{ dry}\cdot\text{ind}^{-1}$		Sapwood dry biomass in a plant individual
$B_{living,sapwood}$	$g \text{ dry}\cdot\text{ind}^{-1}$		Sapwood dry biomass corresponding to (living) parenchymatic tissues in a plant individual (for respiration costs)
$B_{fineroot}$	$g \text{ dry}\cdot\text{ind}^{-1}$		Fine root dry biomass in a plant individual (for respiration costs)

B.16 Carbon balance

Symbol	Units	R	Description
M_{leaf}	$\text{mol gluc}\cdot\text{day}^{-1}$		Leaf daily maintenance respiration in a plant individual
$M_{sapwood}$	$\text{mol gluc}\cdot\text{day}^{-1}$		Sapwood daily maintenance respiration in a plant individual
$M_{fineroot}$	$\text{mol gluc}\cdot\text{day}^{-1}$		Fine root daily maintenance respiration in a plant individual
MR_{leaf}	$\text{g gluc}\cdot\text{g dry}^{-1}\cdot\text{day}^{-1}$	RERleaf	Leaf respiration rate at 20 °C
$MR_{sapwood}$	$\text{g gluc}\cdot\text{g dry}^{-1}\cdot\text{day}^{-1}$	RERsapwood	Living sapwood (parenchymatic tissue) respiration rate at 20 °C
$MR_{fineroot}$	$\text{g gluc}\cdot\text{g dry}^{-1}\cdot\text{day}^{-1}$	RERfineroot	Fine root respiration rate at 20 °C
G_{leaf}	$\text{mol gluc}\cdot\text{day}^{-1}$		Leaf daily growth respiration in a plant individual
$G_{sapwood}$	$\text{mol gluc}\cdot\text{day}^{-1}$		Sapwood daily growth respiration in a plant individual
$G_{fineroot}$	$\text{mol gluc}\cdot\text{day}^{-1}$		Fine root daily growth respiration in a plant individual
F_{phloem}	$\text{mol gluc}\cdot\text{day}^{-1}$		Daily phloem transport of sugars from leaves to sapwood
SC_{leaf}	$\text{mol gluc}\cdot\text{day}^{-1}$		Daily conversion from leaf sugar to leaf starch
$SC_{sapwood}$	$\text{mol gluc}\cdot\text{day}^{-1}$		Daily conversion from sapwood sugar to sapwood starch
TS_{leaf}	$\text{mol gluc}\cdot\text{day}^{-1}$		Daily translocation of leaf sugars (prior to senescence)
$TS_{sapwood}$	$\text{mol gluc}\cdot\text{day}^{-1}$		Daily translocation of sapwood sugars (prior to senescence)
TT_{leaf}	$\text{mol gluc}\cdot\text{day}^{-1}$		Daily translocation of leaf starch (prior to senescence)
RE_{leaf}	$\text{mol gluc}\cdot\text{day}^{-1}$		Daily root exudation of leaf carbon
$RE_{sapwood}$	$\text{mol gluc}\cdot\text{day}^{-1}$		Daily root exudation of sapwood carbon

B.17 Growth, senescence and mortality

Symbol	Units	R	Description
$RGR_{leaf,max}$	$m^2 \cdot cm^{-2} \cdot day^{-1}$	$RGRleafmax$	Maximum leaf area daily growth rate, relative to sapwood area
$RGR_{sapwood,max}$	$cm^2 \cdot cm^{-2} \cdot day^{-1}$	$RGRsapwoodmax$	Maximum daily sapwood relative growth rate (in sapwood area basis)
$RGR_{fineroot,max}$	$g dry^{-1} \cdot day^{-1}$	$RGRfinerootmax$	Maximum daily fine root relative growth rate
LA_{target}	m^2		Leaf area allocation target
ΔLA_{alloc}	m^2		Leaf area increase dictated by the difference between leaf area allocation target and current leaf area
ΔLA_{source}	m^2		Leaf area increase according to leaf metabolic carbon availability
ΔLA_{sink}	m^2		Leaf area increase dictated by sink limitations (leaf relative growth rate and relative cell expansion rate)
ΔLA	m^2		Actual leaf area increase
ΔSA_{source}	cm^2		Sapwood area increase according to sapwood storage carbon availability
ΔSA_{sink}	cm^2		Sapwood area increase dictated by sink limitations
ΔSA	cm^2		Actual sapwood area increase
$B_{fineroot,target}$	$g dry$		Fine root biomass allocation target
$\Delta B_{fineroot,alloc}$	$g dry$		Fine root biomass increase dictated by the difference between fine root biomass allocation target and current fine root biomass
$\Delta B_{fineroot,source}$	$g dry$		Fine root biomass increase according to sapwood metabolic carbon availability
$\Delta B_{fineroot,sink}$	$g dry$		Fine root biomass increase dictated by sink limitations (fine root relative growth rate and relative cell expansion rate)
$\Delta B_{fineroot}$	$g dry$		Actual fine root biomass increase

- Adams, H.D., Park Williams, A., Xu, C., Rauscher, S.A., Jiang, X. & McDowell, N.G. (2013). Empirical and process-based approaches to climate-induced forest mortality models. *Frontiers in Plant Science*, 4, 1–5.
- Adams, H.D., Zeppel, M.J.B., Anderegg, W.R.L., Hartmann, H., Landhäusser, S.M., Tissue, D.T., *et al.* (2017). A multi-species synthesis of physiological mechanisms in drought-induced tree mortality. *Nature Ecology and Evolution*, 1, 1285–1291.
- Albini, F.A. (1976). Estimating wildfire behavior and effects. *For. Serv. Gen. Tech. Rep. INT-30*, 4235, 44 ST—Guidelines and sample protocol for sampli.
- Albini, F. & Baughman, R. (1979). Estimating windspeeds for predicting wildland fire behavior. *USDA Forest Service, Intermountain Forest and Range Experiment Station, Research Paper*, INT-RP-221.
- Ameztegui, A., Cabon, A., De Caceres, M. & Coll, L. (2017). Managing stand density to enhance the adaptability of Scots pine stands to climate change: A modelling approach. *Ecological Modelling*, 356, 141–150.
- Anderegg, W.R.L., Wolf, A., Arango-Velez, A., Choat, B., Chmura, D.J., Jansen, S., *et al.* (2018). Woody plants optimise stomatal behaviour relative to hydraulic risk. *Ecology Letters*, 21, 968–977.
- Anten, N.P.R. & Bastiaans, L. (2016). The use of canopy models to analyze light competition among plants. In: *Canopy photosynthesis: From basics to application* (eds. Hikosaka, K., Niinemets, Ü. & Anten, N.P.R.). Springer, pp. 379–398.
- Bär, A., Michaletz, S.T. & Mayr, S. (2019). Fire effects on tree physiology. *New Phytologist*, 223, 1728–1741.
- Bartlett, M.K., Klein, T., Jansen, S., Choat, B. & Sack, L. (2016). The correlations and sequence of plant

- stomatal, hydraulic, and wilting responses to drought. *Proceedings of the National Academy of Sciences of the United States of America*, 113, 13098–13103.
- Bartlett, M.K., Scoffoni, C. & Sack, L. (2012). The determinants of leaf turgor loss point and prediction of drought tolerance of species and biomes: A global meta-analysis. *Ecology letters*, 15, 393–405.
- Bernacchi, C.J., Singsaas, E.L., Pimentel, C., Portis, A.R. & Long, S.P. (2001). Improved temperature response functions for models of Rubisco-limited photosynthesis. *Plant, Cell and Environment*, 24, 253–259.
- Best, M.J., Pryor, M., Clark, D.B., Rooney, G.G., Essery, R.L.H., Ménard, C.B., et al. (2011). The Joint UK Land Environment Simulator (JULES), model description – Part 1: Energy and water fluxes. *Geoscientific Model Development*, 4, 677–699.
- Birk, E.M. & Simpson, R.W. (1980). Steady state and the continuous input model of litter accumulation and decomposition in Australian eucalypt forests. *Ecology*, 61, 481–485.
- Blackman, C.J., Pfautsch, S., Choat, B., Delzon, S., Gleason, S.M. & Duursma, R.A. (2016). Toward an index of desiccation time to tree mortality under drought. *Plant Cell and Environment*, 39, 2342–2345.
- Bonan, G.B., Williams, M., Fisher, R.A. & Oleson, K.W. (2014). Modeling stomatal conductance in the earth system: Linking leaf water-use efficiency and water transport along the soil-plant-atmosphere continuum. *Geoscientific Model Development*, 7, 2193–2222.
- Boughton, W. (1989). A review of the USDA SCS curve number method. *Australian Journal of Soil Research*, 27, 511–523.
- Bugmann, H., Seidl, R., Hartig, F., Bohn, F., Brůna, J., Cailleret, M., et al. (2019). Tree mortality submodels drive simulated long-term forest dynamics: Assessing 15 models from the stand to global scale. *Ecosphere*, 10, e02616.
- Bullock, J.M., Mallada González, L., Tamme, R., Götzenberger, L., White, S.M., Pärtel, M., et al. (2017). A synthesis of empirical plant dispersal kernels. *Journal of Ecology*, 105, 6–19.
- Butler, B.W. & Dickinson, M.B. (2010). Tree Injury and Mortality in Fires: Developing Process-Based Models. *Fire Ecology*, 6, 55–79.
- Byram, G.M. (1959). Combustion of forest fuels. In: *Forest fire: Control and use* (ed. Davis, K.P.). McGraw-Hill, New York, pp. 61–89.
- Cabon, A., Fernández-de-Uña, L., Gea-Izquierdo, G., Meinzer, F.C., Woodruff, D.R., Martínez-Vilalta, J., et al. (2020a). Water potential control of turgor-driven tracheid enlargement in Scots pine at its xeric distribution edge. *New Phytologist*, 225, 209–221.
- Cabon, A., Peters, R.L., Fonti, P., Martínez-Vilalta, J. & De Cáceres, M. (2020b). Temperature and water potential co-limit stem cambial activity along a steep elevational gradient. *New Phytologist*, 226, 1325–1340.
- Campbell, G.S. & Norman, J.M. (1998). *An introduction to environmental biophysics*. Springer-Verlag, New York.
- Cannell, M.G.R. & Thornley, J.H.M. (2000). Modelling the Components of Plant Respiration: Some Guiding Principles. *Annals of Botany*, 85, 45–54.
- Carsel, R.F. & Parrish, R.S. (1988). Developing joint probability distributions of soil water retention characteristics. *Water Resources Research*, 24, 755–769.
- Cho, N., Kang, S., Agossou, C., Kim, E. & Lim, J.-H. (2022). Modeling temporal variations of non-structural carbohydrate (NSC) storages across biomes. *Forest Ecology and Management*, 508, 120033.
- Choat, B., Jansen, S., Brodribb, T.J., Cochard, H., Delzon, S., Bhaskar, R., et al. (2012). Global convergence in the vulnerability of forests to drought. *Nature*, 491, 752–755.
- Christoffersen, B.O., Gloor, M., Fauset, S., Fyllas, N.M., Galbraith, D.R., Baker, T.R., et al. (2016). Linking hydraulic traits to tropical forest function in a size-structured and trait-driven model (TFS v.1-Hydro). *Geoscientific Model Development Discussions*, 9, 4227–4255.
- Chuine, I., Garcia de Cortazar-Atauri, I., Kramer, K. & Hänninen, H. (2013). Plant Development Models. In: *Phenology: An Integrative Environmental Science* (ed. Schwartz, M.D.). Springer Science, Dordrecht, pp. 275–293.
- Clark, J.S., Macklin, E. & Wood, L. (1998). Stages and spatial scales of recruitment limitation in Southern Appalachian forests. *Ecological Monographs*, 68, 213–235.
- Collatz, G.J., Ball, J.T., Grivet, C. & Berry, J.A. (1991). Physiological and environmental regulation of stomatal conductance, photosynthesis and transpiration: A model that includes a laminar boundary

- layer. *Agricultural and Forest Meteorology*, 54, 107–136.
- Collins, D.B.G. & Bras, R.L. (2007). Plant rooting strategies in water-limited ecosystems. *Water Resources Research*, 43, W06407.
- Cox, P.M., Betts, R.A., Bunton, C.B., Essery, R.L.H., Rowntree, P.R. & Smith, J. (1999). The impact of new land surface physics on the GCM simulation of climate and climate sensitivity. *Climate Dynamics*, 15, 183–203.
- De Cáceres, M., Casals, P., Gabriel, E. & Castro, X. (2019). Scaling-up individual-level allometric equations to predict stand-level fuel loading in Mediterranean shrublands. *Annals of Forest Science*, 76, 87.
- De Cáceres, M., Martínez-Vilalta, J., Coll, L., Llorens, P., Casals, P., Poyatos, R., et al. (2015). Coupling a water balance model with forest inventory data to predict drought stress: The role of forest structural changes vs. Climate changes. *Agricultural and Forest Meteorology*, 213, 77–90.
- De Cáceres, M., Martin-StPaul, N., Turco, M., Cabon, A. & Granda, V. (2018). Estimating daily meteorological data and downscaling climate models over landscapes. *Environmental Modelling & Software*, 108, 186–196.
- De Cáceres, M., Mencuccini, M., Martin-StPaul, N., Limousin, J.-M., Coll, L., Poyatos, R., et al. (2021). Unravelling the effect of species mixing on water use and drought stress in Mediterranean forests: A modelling approach. *Agricultural and Forest Meteorology*, 296, 108233.
- De Cáceres, M., Molowny-Horas, R., Cabon, A., Martínez-Vilalta, J., Mencuccini, M., García-Valdés, R., et al. (2023). MEDFATE 2.9.3: A trait-enabled model to simulate Mediterranean forest function and dynamics at regional scales. *Geoscientific Model Development*, 16, 3165–3201.
- De Pury, D.G.G. & Farquhar, G.D. (1997). Simple scaling of photosynthesis from leaves to canopies without the errors of big-leaf models. *Plant, Cell and Environment*, 20, 537–557.
- Deguchi, A., Hattori, S. & Park, H.-T. (2006). The influence of seasonal changes in canopy structure on interception loss: Application of the revised Gash model. *Journal of Hydrology*, 318, 80–102.
- Delpierre, N., Dufrêne, E., Soudani, K., Ulrich, E., Cecchini, S., Boé, J., et al. (2009). Modelling interannual and spatial variability of leaf senescence for three deciduous tree species in France. *Agricultural and Forest Meteorology*, 149, 938–948.
- Dharssi, I., Vidale, P.L., Verhoef, A., MacPherson, B., Jones, C. & Best, M. (2009). New soil physical properties implemented in the Unified Model at PS18, 9–12.
- Dietze, M.C., Sala, A., Carbone, M.S., Czimczik, C.I., Mantooth, J.A., Richardson, A.D., et al. (2014). Nonstructural Carbon in Woody Plants. *Annual Review of Plant Biology*, 65, 667–687.
- Dunlap, F. (1914). Density of wood substance and porosity of wood. *Journal of Agricultural Research*, II, 423–428.
- Duursma, R.A., Blackman, C.J., Lopéz, R., Martin-StPaul, N.K., Cochard, H. & Medlyn, B.E. (2018). On the minimum leaf conductance: Its role in models of plant water use, and ecological and environmental controls. *New Phytologist*, 221, 693–705.
- Espelta, J.M., Barbatí, A., Quevedo, L., Tárrega, R., Navascués, P., Bonfil, C., et al. (2012). Post-Fire Management of Mediterranean Broadleaved Forests. In: *Post-Fire Management and Restoration of Southern European Forests*, Managing Forest Ecosystems (eds. Moreira, F., Arianoutsou, M., Corona, P. & De las Heras, J.). Springer Netherlands, Dordrecht, pp. 171–194.
- Espelta, J.M., Habrouk, A. & Retana, J. (2006). Response to natural and simulated browsing of two Mediterranean oaks with contrasting leaf habit after a wildfire. *Annals of Forest Science*, 63, 441–447.
- Espelta, J.M., Sabaté, S. & Retana, J. (1999). Resprouting Dynamics. In: *Ecology of Mediterranean Evergreen Oak Forests*, Ecological Studies (eds. Rodà, F., Retana, J., Gracia, C.A. & Bellot, J.). Springer, Berlin, Heidelberg, pp. 61–73.
- Faticchi, S., Leuzinger, S. & Körner, C. (2014). Moving beyond photosynthesis: From carbon source to sink-driven vegetation modeling. *New Phytologist*, 201, 1086–1095.
- Federer, C. (1982). Transpirational supply and demand: Plant, soil, and atmospheric effects evaluated by simulation. *Water Resources Research*, 18, 355–362.
- Fisher, R.A., Williams, M., Do Vale, L.R., Da Costa, A.L. & Meir, P. (2006). Evidence from Amazonian forests is consistent with a model of isohydric control of leaf water potential. *Plant, Cell and Environment*, 29, 151–165.
- Flerchinger, G.N., Xiao, W., Sauer, T.J. & Yu, Q. (2009). Simulation of within-canopy radiation exchange. *NJAS - Wageningen Journal of Life Sciences*, 57, 5–15.

- Först, P., Werner, F. & Delgado, A. (2002). On the pressure dependence of the viscosity of aqueous sugar solutions. *Rheologica Acta*, 41, 369–374.
- Francés, F., Vélez, J.I. & Vélez, J.J. (2007). Split-parameter structure for the automatic calibration of distributed hydrological models. *Journal of Hydrology*, 332, 226–240.
- Franks, P.J. (2006). Higher rates of leaf gas exchange are associated with higher leaf hydrodynamic pressure gradients. *Plant, Cell and Environment*, 29, 584–592.
- Friend, A.D., Eckes-Shephard, A.H., Fonti, P., Rademacher, T.T., Rathgeber, C.B.K., Richardson, A.D., *et al.* (2019). On the need to consider wood formation processes in global vegetation models and a suggested approach. *Annals of Forest Science*, 76, 49.
- Friend, A.D., Stevens, A.K., Knox, R.G. & Cannell, M.G.R. (1997). A process-based, terrestrial biosphere model of ecosystem dynamics (Hybrid v3.0). *Ecological Modelling*, 95, 249–287.
- Fyllas, N.M. & Troumbis, A.Y. (2009). Simulating vegetation shifts in north-eastern Mediterranean mountain forests under climatic change scenarios. *Global Ecology and Biogeography*, 18, 64–77.
- García-Jiménez, R., Palmero-Iniesta, M. & Espelta, J. (2017). Contrasting Effects of Fire Severity on the Regeneration of *Pinus halepensis* Mill. And Resprouter Species in Recently Thinned Thickets. *Forests*, 8, 55.
- Gash, J., Lloyd, C. & Lachaud, G. (1995). Estimating sparse forest rainfall interception with an analytical model. *Journal of Hydrology*, 170.
- Genuchten, M.V. (1980). A closed-form equation for predicting the hydraulic conductivity of unsaturated soils. *Soil science society of America journal*, 44, 892–898.
- Gifford, R.M. (2003). Plant respiration in productivity models: Conceptualisation, representation and issues for global terrestrial carbon-cycle research. *Functional Plant Biology*, 30, 171–186.
- Granier, A., Bréda, N., Biron, P. & Villette, S. (1999). A lumped water balance model to evaluate duration and intensity of drought constraints in forest stands. *Ecological Modelling*, 116, 269–283.
- Granier, A., Reichstein, M., Bréda, N., Janssens, I.A., Falge, E., Ciais, P., *et al.* (2007). Evidence for soil water control on carbon and water dynamics in European forests during the extremely dry year: 2003. *Agricultural and Forest Meteorology*, 143, 123–145.
- Guillemot, J., Martin-Stpaul, N.K., Dufrêne, E., François, C., Soudani, K., Ourcival, J.M., *et al.* (2015). The dynamic of the annual carbon allocation to wood in European tree species is consistent with a combined source-sink limitation of growth: Implications for modelling. *Biogeosciences*, 12, 2773–2790.
- Hammond, W.M., Yu, Kailiang.L., Wilson, L.A., Will, R.E., Anderegg, W.R.L. & Adams, H.D. (2019). Dead or dying? Quantifying the point of no return from hydraulic failure in drought-induced tree mortality. *New Phytologist*.
- Hasenauer, H. & Monserud, R.A. (1996). A crown ratio model for Austrian Forests. *Forest Ecology and Management*, 84, 49–60.
- Hawkes, C. (2000). Woody plant mortality algorithms: Description, problems and progress. *Ecological Modelling*, 126, 225–248.
- Hikosaka, K., Kumagai, T. & Ito, A. (2016). Modeling Canopy Photosynthesis. pp. 239–268.
- Hölttä, T., Cochard, H., Nikinmaa, E. & Mencuccini, M. (2009). Capacitive effect of cavitation in xylem conduits: Results from a dynamic model. *Plant, Cell and Environment*, 32, 10–21.
- Hölttä, T., Lintunen, A., Chan, T., Mäkelä, A. & Nikinmaa, E. (2017). A steady-state stomatal model of balanced leaf gas exchange, hydraulics and maximal source-sink flux. *Tree physiology*, 37, 851–868.
- Hood, S.M., Varner, J.M., Van Mantgem, P. & Cansler, C.A. (2018). Fire and tree death: Understanding and improving modeling of fire-induced tree mortality. *Environmental Research Letters*, 13.
- Hoshika, Y., Osada, Y., Marco, A. de, Peñuelas, J. & Paoletti, E. (2018). Global diurnal and nocturnal parameters of stomatal conductance in woody plants and major crops. *Global Ecology and Biogeography*, 27, 257–275.
- Karavani, A., De Cáceres, M., Martínez de Aragón, J., Bonet, J.A. & de-Miguel, S. (2018). Effect of climatic and soil moisture conditions on mushroom productivity and related ecosystem services in Mediterranean pine stands facing climate change. *Agricultural and Forest Meteorology*, 248.
- Katul, G.G., Mahrt, L., Poggi, D. & Sanz, C. (2004). One- and two-equation models for canopy turbulence. *Boundary-Layer Meteorology*, 113, 81–109.
- Keane, R., Austin, M., Field, C. & Huth, A. (2001). Tree mortality in gap models: Application to climate change. *Climatic Change*, 51, 509–540.

- Kergoat, L. (1998). A model for hydrological equilibrium of leaf area index on a global scale. *Journal of Hydrology*, 212–213, 268–286.
- Körner, C. (2015). Paradigm shift in plant growth control. *Current Opinion in Plant Biology*, 25, 107–114.
- Körner, C. (2019). No need for pipes when the well is dry—a comment on hydraulic failure in trees. *Tree Physiology*, 39, 695–700.
- Krajicek, J., Brinkman, K. & Gingrich, S. (1961). Crown competition—a measure of density. *Forest Science*, 7, 35–42.
- Kursar, T.A., Engelbrecht, B.M.J., Burke, A., Tyree, M.T., El Omari, B. & Giraldo, J.P. (2009). Tolerance to low leaf water status of tropical tree seedlings is related to drought performance and distribution. *Functional Ecology*, 23, 93–102.
- Lai, C., Katul, G., Oren, R., Ellsworth, D. & Schäfer, K. (2000). Modeling CO₂ and water vapor turbulent flux distributions within a forest canopy. *Journal of Geophysical Research: Atmospheres*, 105, 26333–26351.
- Landsberg, J.J. & Waring, R.H. (1997). A generalised model of forest productivity using simplified concepts of radiation-use efficiency, carbon balance and partitioning. *Forest Ecology and Management*, 95, 209–228.
- Leij, F.J., Alves, W.J., Genuchten, M.Th.V. & Williams, J.R. (1996). *The UNSODA Unsaturated Soil Hydraulic Database User's Manual Version 1.0*.
- Leuning, R. (2002). Temperature dependence of two parameters in a photosynthesis model. *Plant, Cell and Environment*, 25, 1205–1210.
- Lindner, M., Sievänen, R. & Pretzsch, H. (1997). Improving the simulation of stand structure in a forest gap model. *Forest Ecology and Management*, 95, 183–195.
- Lloret, F., Siscart, D. & Dalmases, C. (2004). Canopy recovery after drought dieback in holm-oak Mediterranean forests of Catalonia (NE Spain). *Global Change Biology*, 10, 2092–2099.
- Ma, Y. & Liu, H. (2019). An Advanced Multiple-Layer Canopy Model in the WRF Model With Large-Eddy Simulations to Simulate Canopy Flows and Scalar Transport Under Different Stability Conditions. *Journal of Advances in Modeling Earth Systems*, 11, 2330–2351.
- Magnani, F., Mencuccini, M. & Grace, J. (2000). Age-related decline in stand productivity: The role of structural acclimation under hydraulic constraints. *Plant, Cell and Environment*, 23, 251–263.
- Maherali, H., Pockman, W. & Jackson, R. (2004). Adaptive variation in the vulnerability of woody plants to xylem cavitation. *Ecology*, 85, 2184–2199.
- Manoli, G., Huang, C., Bonetti, S., Domec, J., Marani, M. & Katul, G. (2017). Competition for light and water in a coupled soil-plant system. *Advances in Water Resources*, 108, 216–230.
- Martinez-Vilalta, J., Anderegg, W.R.L., Sapes, G. & Sala, A. (2019). Greater focus on water pools may improve our ability to understand and anticipate drought-induced mortality in plants. *New Phytologist*, 223, 22–32.
- Martínez-Vilalta, J., Sala, A., Asensio, D., Galiano, L., Hoch, G., Palacio, S., et al. (2016). Dynamics of non-structural carbohydrates in terrestrial plants: A global synthesis. *Ecological Monographs*, 86, 495–516.
- Martin-StPaul, N., Delzon, S. & Cochard, H. (2017). Plant resistance to drought depends on timely stomatal closure. *Ecology Letters*, 20, 1437–1447.
- Massman, W. (1987). A comparative study of some mathematical models of the mean wind structure and aerodynamic drag of plant canopies. *Boundary-Layer Meteorology*, 40, 179–197.
- McDowell, N.G. (2011). Mechanisms linking drought, hydraulics, carbon metabolism, and vegetation mortality. *Plant Physiology*, 155, 1051–1059.
- McDowell, N.G., Sapes, G., Pivovaroff, A., Adams, H.D., Allen, C.D., Anderegg, W.R.L., et al. (2022). Mechanisms of woody-plant mortality under rising drought, CO₂ and vapour pressure deficit. *Nature Reviews Earth & Environment*, 0123456789, 41–44.
- McDowell, N., Pockman, W.T., Allen, C.D., Breshears, D.D., Cobb, N., Kolb, T., et al. (2008). Mechanisms of plant survival and mortality during drought: Why do some plants survive while others succumb to drought? *The New Phytologist*, 178, 719–39.
- McMurtrie, R.E., Rook, D.A. & Kelliher, F.M. (1990). Modelling the yield of *Pinus radiata* on a site limited by water and nitrogen. *Forest Ecology and Management*, 30, 381–413.
- Medlyn, B.E., Loustau, D. & Delzon, S. (2002). Temperature response of parameters of a biochemically based

- model of photosynthesis. I. Seasonal changes in mature maritime pine (*Pinus pinaster* Ait.). *Plant, Cell and Environment*, 25, 1155–1165.
- Mencuccini, M. & Grace, J. (1995). Climate influences the leaf area/sapwood area ratio in Scots pine. *Tree Physiology*, 15, 1–10.
- Michaletz, S.T. & Johnson, E.A. (2006). A heat transfer model of crown scorch in forest fires. *Canadian Journal of Forest Research*, 36, 2839–2851.
- Michaletz, S.T. & Johnson, E.A. (2008). A biophysical process model of tree mortality in surface fires. *Canadian Journal of Forest Research*, 38, 2013–2029.
- Miralles, D.G., Gash, J.H., Holmes, T.R.H., Jeu, R.A.M. de & Dolman, A.J. (2010). Global canopy interception from satellite observations. *Journal of Geophysical Research*, 115, D16122.
- Mitsopoulos, I.D. & Dimitrakopoulos, A.P. (2007). Allometric equations for crown fuel biomass of Aleppo pine (*Pinus halepensis* Mill.) in Greece. *International Journal of Wildland Fire*, 16, 642–647.
- Moreira, B., Tormo, J. & Pausas, J.G. (2012). To resprout or not to resprout: Factors driving intraspecific variability in resprouting. *Oikos*, 121, 1577–1584.
- Mouillet, F., Rambal, S. & Joffre, R. (2002). Simulating climate change impacts on fire frequency and vegetation dynamics in a Mediterranean-type ecosystem. *Global Change Biology*, 8, 423–437.
- Mouillet, F., Rambal, S. & Lavorel, S. (2001). A generic process-based Simulator for mediterranean landcApes (SIERRA): Design and validation exercises. *Forest Ecology and Management*, 147, 75–97.
- Nathan, R., Klein, E., Robledo-Arnuncio, J.J. & Revilla, E. (2012). Dispersal kernels: review. In: *Dispersal Ecology and Evolution* (eds. Clobert, J., Baguette, M., Benton, T.G. & Bullock, J.M.). Oxford University Press, pp. 186–210.
- Nathan, R., Safriel, U.N. & Noy-Meir, I. (2001). Field validation and sensitivity analysis of a mechanistic model for tree seed dispersal by wind. *Ecology*, 82, 374–388.
- Ogle, K. & Pacala, S.W. (2009). A modeling framework for inferring tree growth and allocation from physiological, morphological and allometric traits. *Tree Physiology*, 29, 587–605.
- Oliveras, I., Martínez-Vilalta, J., Jimenez-Ortiz, T., Lledó, M.J., Escarré, A. & Piñol, J. (2003). Hydraulic architecture of *Pinus halepensis*, *P. pinea* and *Tetraclinis articulata* in a dune ecosystem of Eastern Spain. *Plant Ecology*, 131–141.
- Olson, M.E., Anfodillo, T., Rosell, J.A., Petit, G., Crivellaro, A., Isnard, S., et al. (2014). Universal hydraulics of the flowering plants: Vessel diameter scales with stem length across angiosperm lineages, habits and climates. *Ecology Letters*, 17, 988–997.
- Ostendorf, B. & Reynolds, J.F. (1993). Relationships between a terrain-based hydrologic model and patch-scale vegetation patterns in an arctic tundra landscape. *Landscape Ecology*, 8, 229–237.
- Pausas, J.G., Pratt, R.B., Keeley, J.E., Jacobsen, A.L., Ramirez, A.R., Vilagrosa, A., et al. (2016). Towards understanding resprouting at the global scale. *New Phytologist*, 209, 945–954.
- Pimont, F., Dupuy, J.L. & Rigolot, E. (2018). A simple model for shrub-strata-fuel dynamics in *Quercus coccifera* L. communities. *Annals of Forest Science*, 75, 1–9.
- Plavcová, L. & Jansen, S. (2015). The Role of Xylem Parenchyma in the Storage and Utilization of Nonstructural Carbohydrates. In: *Functional and Ecological Xylem Anatomy*. Springer International Publishing, Cham, pp. 209–234.
- Prentice, I.C., Sykes, M.T. & Cramer, W. (1993). A simulation model for the transient effects of climate change on forest landscapes. *Ecological Modelling*, 65, 51–70.
- Prescott, C.E., Grayston, S.J., Helmisaari, H.-S., Kaštovská, E., Körner, C., Lambers, H., et al. (2020). Surplus Carbon Drives Allocation and Plant–Soil Interactions. *Trends in Ecology & Evolution*, 35, 1110–1118.
- Price, D., Zimmermann, N. & Meer, P.V.D. (2001). Regeneration in gap models: Priority issues for studying forest responses to climate change. *Climatic Change*, 51, 475–508.
- Prichard, S.J., Sandberg, D.V., Ottmar, R.D., Eberhardt, E., Andreu, A., Eagle, P., et al. (2013). *Classification System Version 3.0: Technical Documentation*.
- Rasche, L., Fahse, L., Zingg, A. & Bugmann, H. (2012). Enhancing gap model accuracy by modeling dynamic height growth and dynamic maximum tree height. *Ecological Modelling*, 232, 133–143.
- Reich, P.B., Tjoelker, M.G., Pregitzer, K.S., Wright, I.J., Oleksyn, J. & Machado, J.L. (2008). Scaling of respiration to nitrogen in leaves, stems and roots of higher land plants. *Ecology Letters*, 11, 793–801.
- Reinhardt, E., Lutes, D. & Scott, J. (2006). *FuelCalc: A method for estimating fuel characteristics*.

- Retana, J., Riba, M., Castell, C. & Espelta, J.M. (1992). Regeneration by sprouting of holm-oak (*Quercus ilex*) stands exploited by selection thinning. *Vegetatio*, 99, 355–364.
- Richardson, A.D., Carbone, M.S., Keenan, T.F., Czimczik, C.I., Hollinger, D.Y., Murakami, P., *et al.* (2013). Seasonal dynamics and age of stemwood nonstructural carbohydrates in temperate forest trees. *New Phytologist*, 197, 850–861.
- Ritchie, J. (1972). Model for predicting evaporation from a row crop with incomplete cover. *Water resources research*, 8, 1204–1213.
- Rolo, V. & Moreno, G. (2019). Shrub encroachment and climate change increase the exposure to drought of Mediterranean wood-pastures. *Science of The Total Environment*, 660, 550–558.
- Rothermel, R.C. (1972). A mathematical model for predicting fire spread in wildland fuels. *USDA Forest Service Research Paper INT USA*.
- Rötzer, T., Häberle, K.H., Kallenbach, C., Matyssek, R., Schütze, G. & Pretzsch, H. (2017). Tree species and size drive water consumption of beech/spruce forests - a simulation study highlighting growth under water limitation. *Plant and Soil*, 418, 337–356.
- Ruffault, J., Martin-StPaul, N.K., Duffet, C., Goge, F. & Mouillot, F. (2014). Projecting future drought in Mediterranean forests: Bias correction of climate models matters! *Theoretical and Applied Climatology*, 117, 113–122.
- Ruffault, J., Martin-StPaul, N.K., Rambal, S. & Mouillot, F. (2013). Differential regional responses in drought length, intensity and timing to recent climate changes in a Mediterranean forested ecosystem. *Climatic Change*, 117, 103–117.
- Ruffault, J., Pimont, F., Cochard, H., Dupuy, J.-L. & Martin-StPaul, N. (2022). SurEau-Ecos v2.0: A trait-based plant hydraulics model for simulations of plant water status and drought-induced mortality at the ecosystem level. *Geoscientific Model Development*, 15, 5593–5626.
- Sandberg, D.V., Riccardi, C.L. & Schaaf, M.D. (2007). Reformulation of Rothermel's wildland fire behaviour model for heterogeneous fuelbeds. *Canadian Journal of Forest Research*, 37, 2438–2455.
- Sando, R.W. & Wick, C.H. (1972). A method of evaluating crown fuels in forest stands. Research Paper NC-84. *USDA Forest Service Research Paper NC-84*, 10 pp.
- Savage, V.M., Bentley, L.P., Enquist, B.J., Sperry, J.S., Smith, D.D., Reich, P.B., *et al.* (2010). Hydraulic trade-offs and space filling enable better predictions of vascular structure and function in plants. *Proceedings of the National Academy of Sciences of the United States of America*, 107, 22722–7.
- Saxton, K.E. & Rawls, W.J. (2006). Soil water characteristic estimates by texture and organic matter for hydrologic solutions. *Soil Science Society of America Journal*, 70, 1569.
- Saxton, K.E., Rawls, W.J., Romberger, J.S. & Papendick, R.I. (1986). Estimating generalized soil-water characteristics from texture. *Soil Science Society of America Journal*, 50, 1031–1036.
- Schaaf, M.D., Sandberg, D.V., Schreuder, M.D. & Riccardi, C.L. (2007). A conceptual framework for ranking crown fire potential in wildland fuelbeds. *Canadian Journal of Forest Research*, 37, 2464–2478.
- Schenk, H.J. & Jackson, R.B. (2002). Rooting depths, lateral root spreads and below-ground/above-ground allometries of plants in water-limited ecosystems. *Journal of Ecology*, 90, 480–494.
- Schume, H., Jost, G. & Hager, H. (2004). Soil water depletion and recharge patterns in mixed and pure forest stands of European beech and Norway spruce. *Journal of Hydrology*, 289, 258–274.
- Scott, J.H. & Reinhardt, E.D. (2002). Estimating canopy fuels in conifer forests. *Fire Management Today*, 62, 45–50.
- Shinozaki, K., Yoda, K., Hozumi, K. & Kira, T. (1964). A quantitative analysis of plant form - the pipe model theory. I. Basic analysis. *Japanese Journal of Ecology*, 14, 97–105.
- Sitch, S., Smith, B., Prentice, I.C., Arneth, a., Bondeau, a., Cramer, W., *et al.* (2003). Evaluation of ecosystem dynamics, plant geography and terrestrial carbon cycling in the LPJ dynamic global vegetation model. *Global Change Biology*, 9, 161–185.
- Snell, R.S. (2014). Simulating long-distance seed dispersal in a dynamic vegetation model. *Global Ecology and Biogeography*, 23, 89–98.
- Sperry, J.S., Adler, F.R., Campbell, G.S. & Comstock, J.P. (1998). Limitation of plant water use by rhizosphere and xylem conductance: Results from a model. *Plant, Cell & Environment*, 21, 347–359.
- Sperry, J.S. & Love, D.M. (2015). What plant hydraulics can tell us about responses to climate-change droughts. *New Phytologist*, 207, 14–27.
- Sperry, J.S., Venturas, M.D., Anderegg, W.R.L., Mencuccini, M., Mackay, D.S., Wang, Y., *et al.* (2017).

- Predicting stomatal responses to the environment from the optimization of photosynthetic gain and hydraulic cost. *Plant, Cell & Environment*, 40, 816–830.
- Sperry, J.S., Wang, Y., Wolfe, B.T., Mackay, D.S., Anderegg, W.R.L., McDowell, N.G., *et al.* (2016). Pragmatic hydraulic theory predicts stomatal responses to climatic water deficits. *New Phytologist*, 212, 577–589.
- Spicer, R. (2005). Senescence in Secondary Xylem: Heartwood Formation as an Active Developmental Program. *Vascular Transport in Plants*, 457–475.
- Spitters, C.J.T., Toussaint, H.A.J.M. & Goudriaan, J. (1986). Separating the diffuse and direct components of global radiation and its implications for modeling canopy photosynthesis. I. Components of incoming radiation. *Agricultural and Forest Meteorology*, 38, 231–242.
- Stephan, K., Miller, M. & Dickinson, M.B. (2010). First-Order Fire Effects on Herbs and Shrubs: Present Knowledge and Process Modeling Needs. *Fire Ecology*, 6, 95–114.
- Steppe, K., Lemeur, R. & Vanrolleghem, A. (2006). A mathematical model linking tree sap flow dynamics to dialy stem diameter fluctuation and radial stem growth. *Tree Physiology*, 26, 257–273.
- Stolf, R., Thurler, Á., Oliveira, O., Bacchi, S. & Reichardt, K. (2011). Method to estimate soil macroporosity and microporosity based on sand content and bulk density. *Revista Brasileira de Ciencias do Solo*, 35, 447–459.
- Tjoelker, M.G., Oleksyn, J. & Reich, P.B. (2001). Modelling respiration of vegetation: Evidence for a general temperature-dependent Q₁₀. *Global Change Biology*, 7, 223–230.
- Tóth, B., Weynants, M., Nemes, A., Makó, A., Bilas, G. & Tóth, G. (2015). New generation of hydraulic pedotransfer functions for Europe. *European Journal of Soil Science*, 66, 226–238.
- Tyre, M.T. & Yang, S. (1990). Water-storage capacity of Thuja, Tsuga and Acer stems measured by dehydration isotherms - The contribution of capillary water and cavitation. *Planta*, 182, 420–426.
- Urlı, M., Porte, A.J., Cochard, H., Guengant, Y., Burlett, R. & Delzon, S. (2013). Xylem embolism threshold for catastrophic hydraulic failure in angiosperm trees. *Tree Physiology*, 33, 672–683.
- Van Lier, Q.D.J., Neto, D.D. & Metselaar, K. (2009). Modeling of transpiration reduction in van genuchten-mualem type soils. *Water Resources Research*, 45, 1–9.
- Walker, A.P., Beckerman, A.P., Gu, L., Kattge, J., Cernusak, L.A., Domingues, T.F., *et al.* (2014). The relationship of leaf photosynthetic traits - V_{cmax} and J_{max} - to leaf nitrogen, leaf phosphorus, and specific leaf area: A meta-analysis and modeling study. *Ecology and Evolution*, 4, 3218–3235.
- Watanabe, T. & Mizutani, K. (1996). Model study on micrometeorological aspects of rainfall interception over an evergreen broad-leaved. *Agricultural and Forest Meteorology*, 80, 195–214.
- Wehrli, A., Weisberg, P.J., Schönenberger, W., Brang, P. & Bugmann, H. (2007). Improving the establishment submodel of a forest patch model to assess the long-term protective effect of mountain forests. *European Journal of Forest Research*, 126, 131–145.
- Wigmsta, M.S. & Lettenmaier, D.P. (1999). A comparison of simplified methods for routing topographically driven subsurface flow. *Water Resources Research*, 35, 255–264.
- Wigmsta, M., Vail, L. & Lettenmaier, D. (1994). A distributed hydrology-vegetation model for complex terrain. *Water Resources Research*, 30, 1665–1679.
- Wolf, A., Anderegg, W.R.L. & Pacala, S.W. (2016). Optimal stomatal behavior with competition for water and risk of hydraulic impairment. *Proceedings of the National Academy of Sciences*, 113, E7222–E7230.