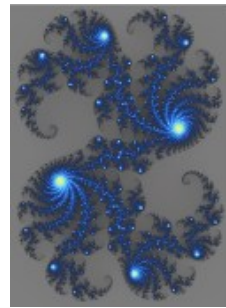


Using mass spectrometry to elucidate complex biological systems

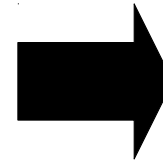
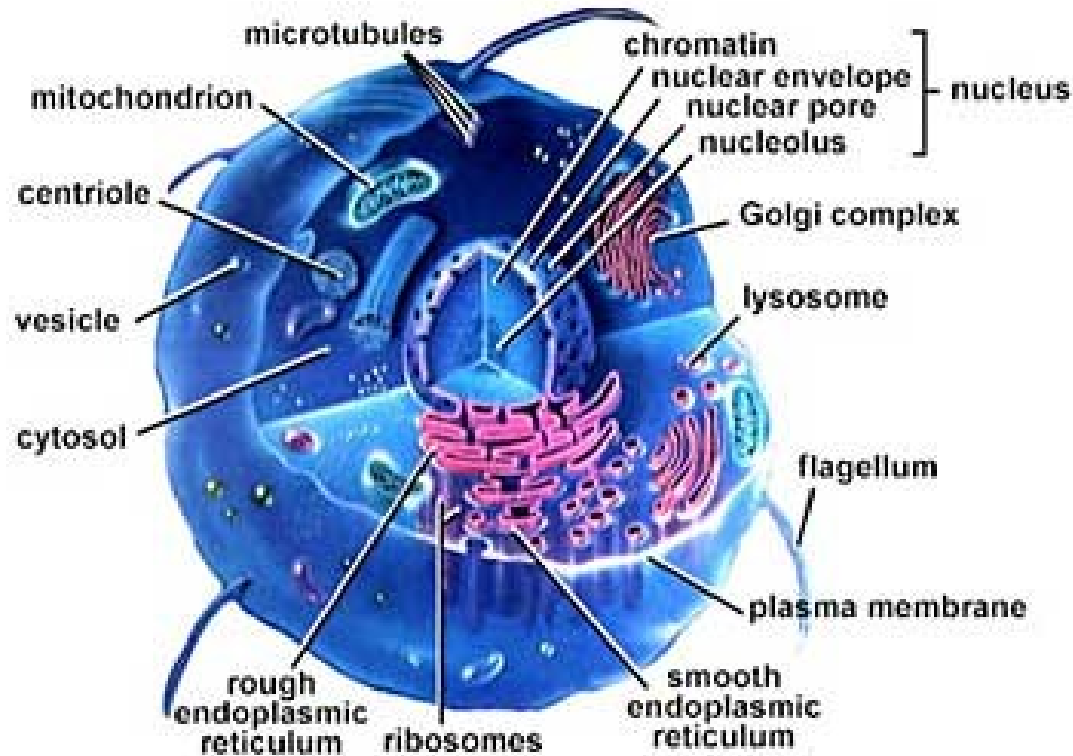


PRINCE
LAB

DEPARTMENT OF
CHEMISTRY AND BIOCHEMISTRY
BRIGHAM YOUNG UNIVERSITY

December 24, 2013

Biochemists are good at measuring the blueprints for cellular life (RNA, DNA), but we are not very good yet at measuring all the machinery and structures of which life is mostly composed (proteins, lipids, metabolites)!



proteins

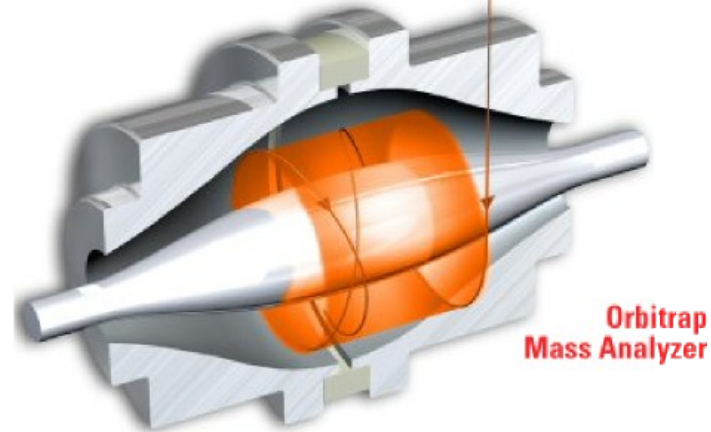
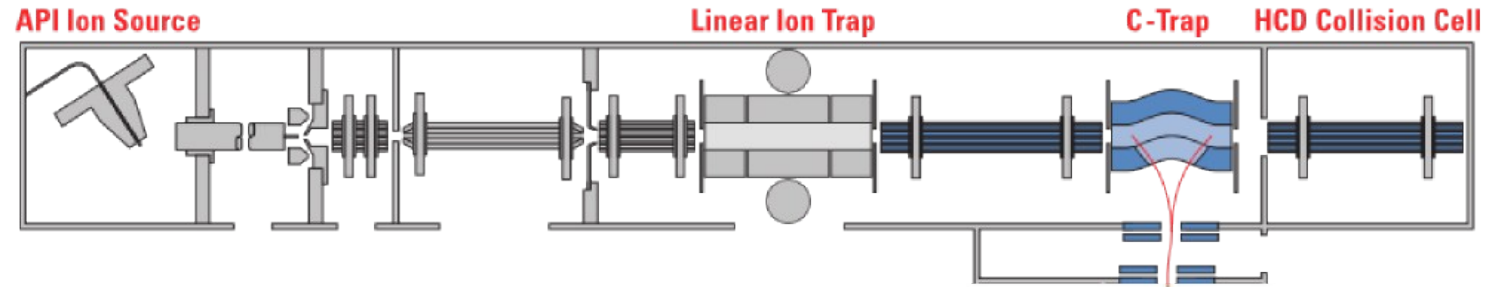
lipids

metabolites

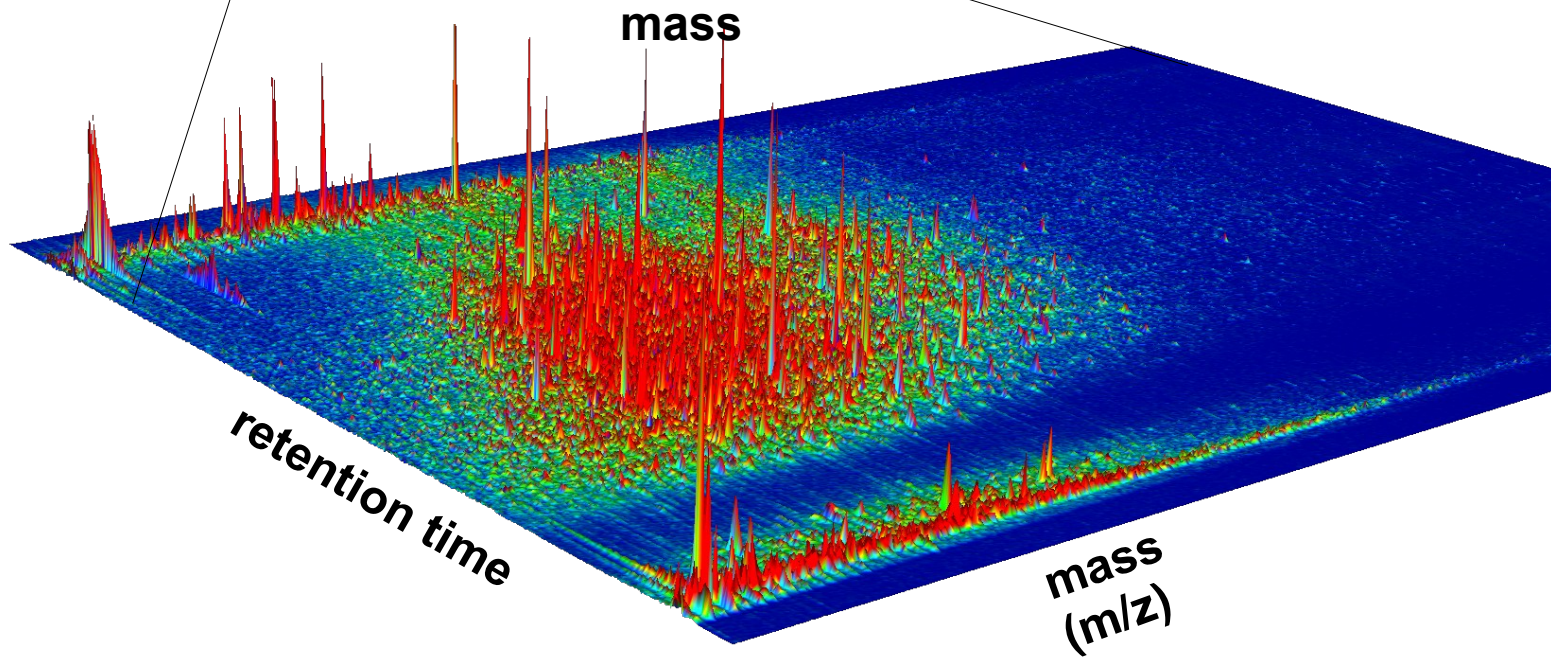
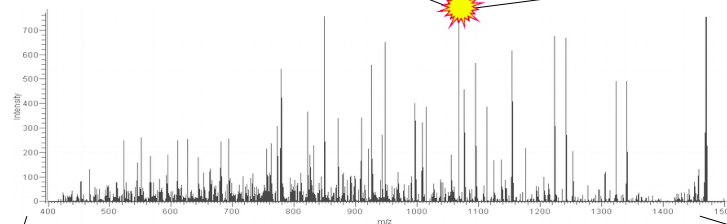
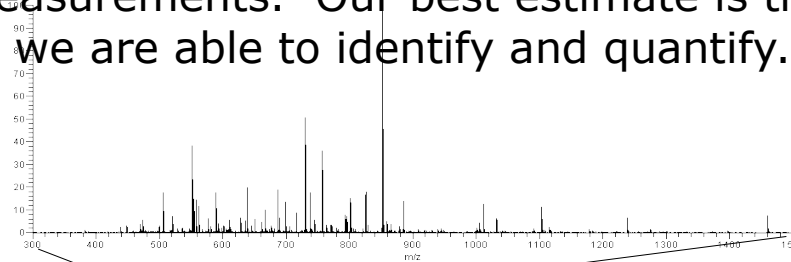
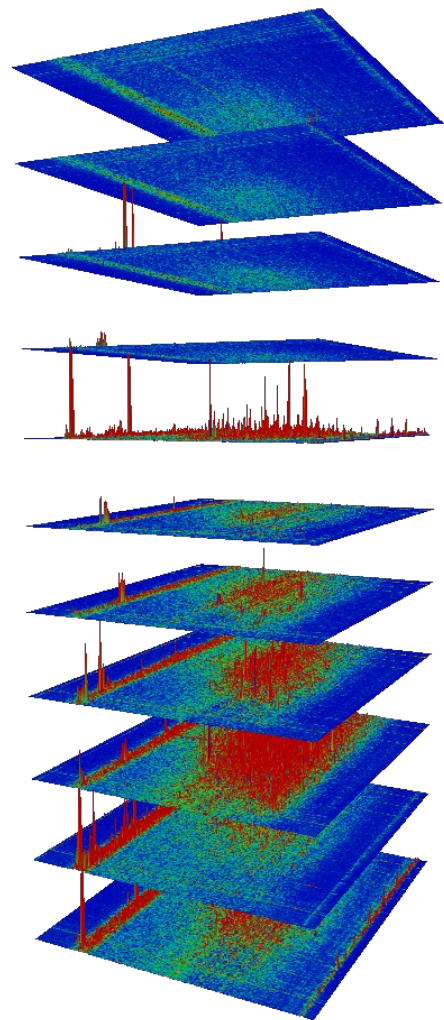
RNA

DNA

Mass spectrometry is the leading method for the high-throughput measurement of proteins, lipids, and metabolites. We can measure 100's to 10,000's of these biomolecules in a single experiment.



A key challenge in mass spectrometry is correctly inferring the identities and quantities of biomolecules from mass and fragment measurements. Our best estimate is that we collect data on 10X more biomolecules than we are able to identify and quantify.

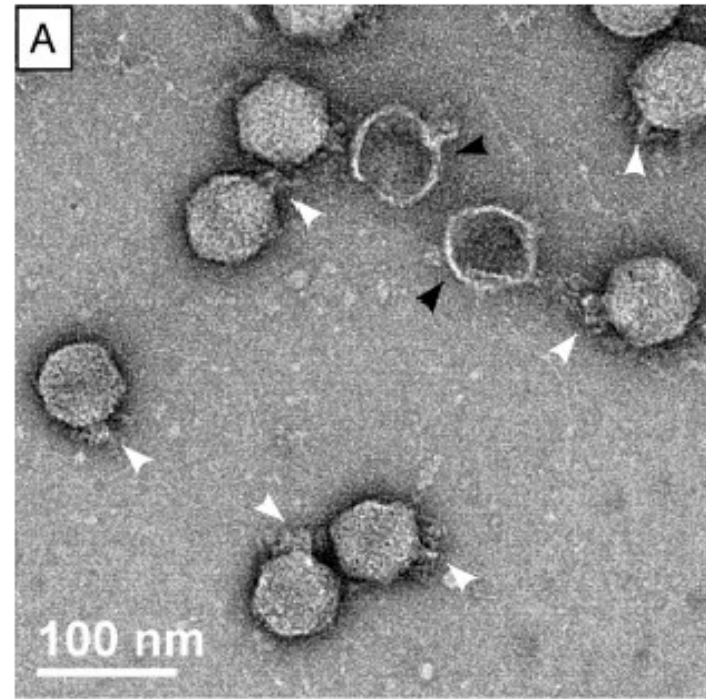
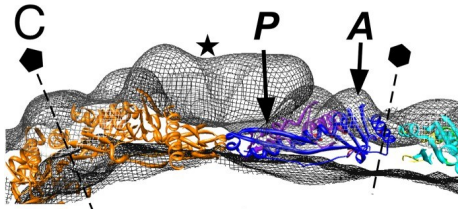
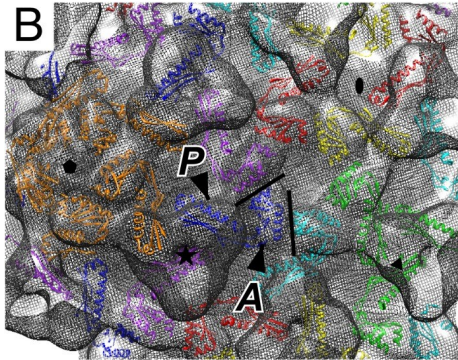
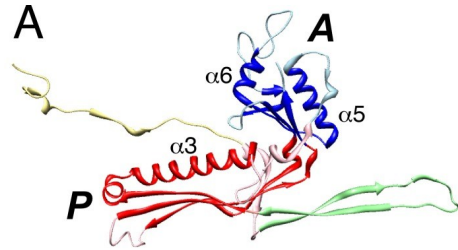


What kinds of things can we do right now with mass spectrometry?

We sequenced all the proteins in a Salt Lake bacteria virus.



In collaboration
with David Belnap

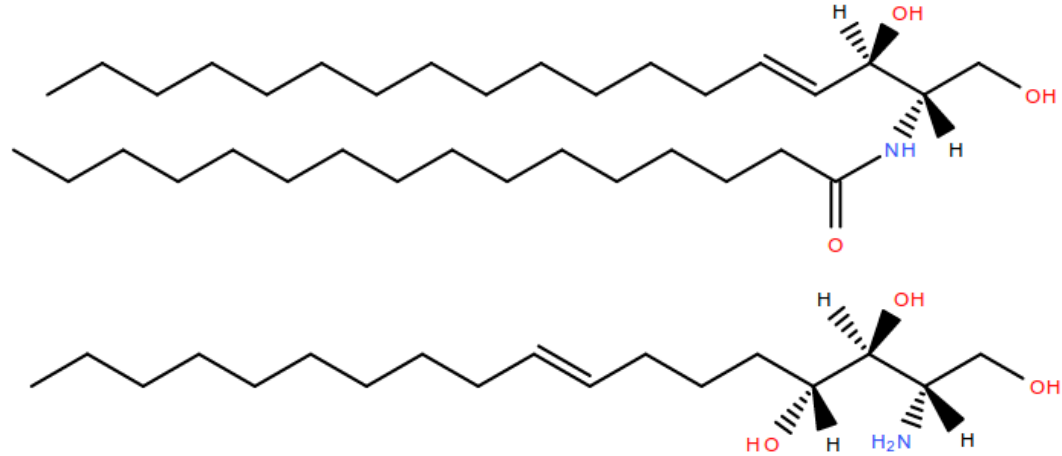
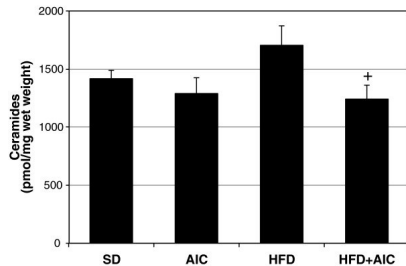
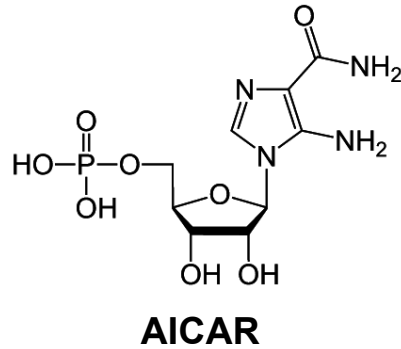


Shen PS, Domek MJ, Sanz-Gardia E, Makaju A, Taylor MJ, Hoggan R, Culumber MD, Oberg CJ, Breakwell DP, Prince JT, Belnap DM. *Journal of Virology*. **2012**

We quantified changing levels of ceramides, important lipids involved in metabolic diseases (obesity, heart disease, etc.)



In collaboration
with Ben Bikman



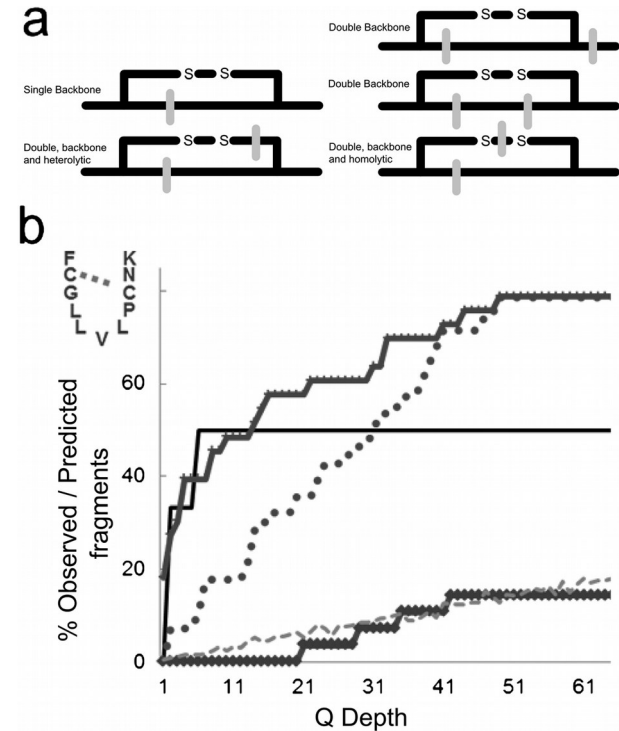
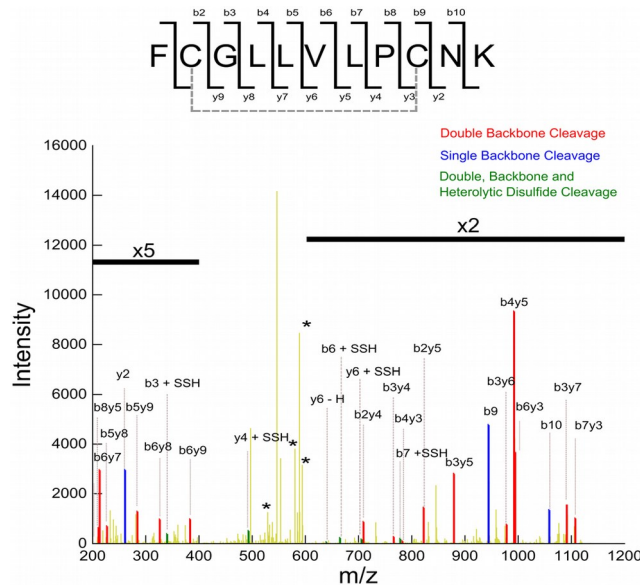
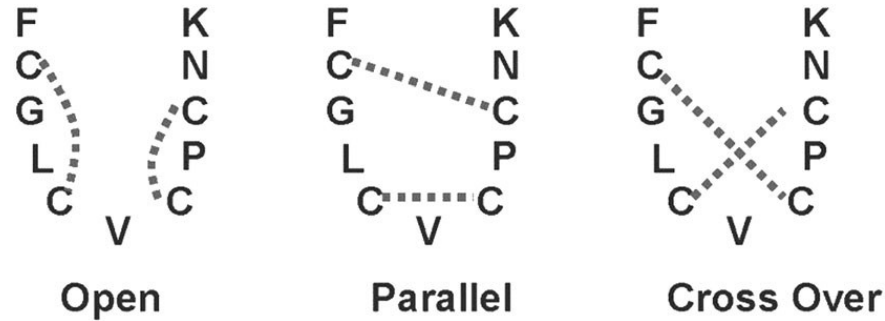
Erickson KA, Smith ME, Anthonymuthu TS, Evanson MJ, Brassfield ES, Hodson AE, Bressler MA, Tucker BJ, Thatcher MO, Prince JT, Hancock CR, Bikman BT. *Diabetol. Metab. Syndr.* **2012**

Smith ME, Tippetts TS, Brassfield ES, Tucker BJ, Ockey A, Swensen AC, Anthonymuthu TS, Washburn TD, Kane DA, Prince JT, Bikman BT. *Biochem J.* **2013.**

We deciphered the disulfide double bond configurations found in the vesicle fusion protein SNAP25, a key step in understanding how oxidative damage (e.g., aging) alters neurotransmission.



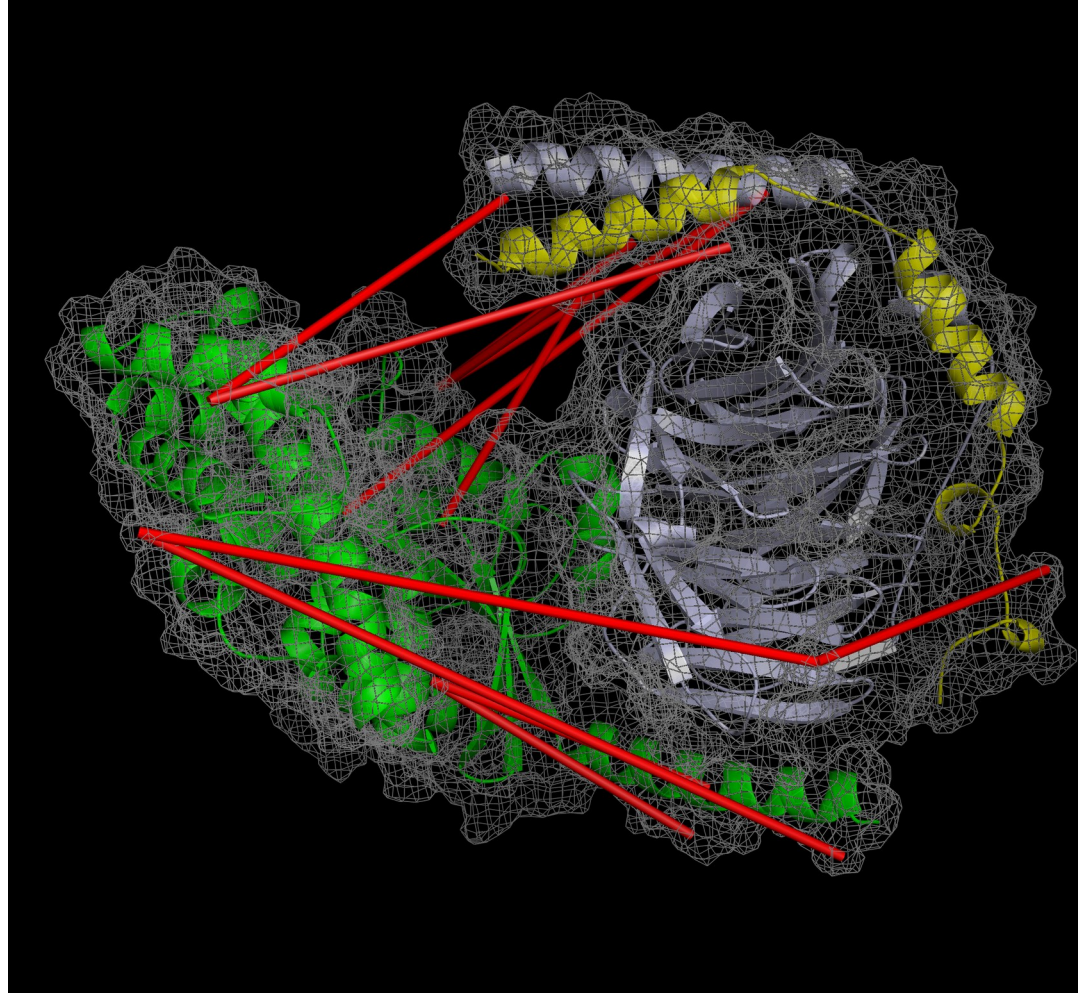
In collaboration with
Dixon Woodbury



We are using chemical crosslinks and mass spectrometry along with cryoelectron microscopy to understand protein complexes important in a variety of disease (e.g., Bardet-Biedl syndrome)



In collaboration with
Barry Willardson

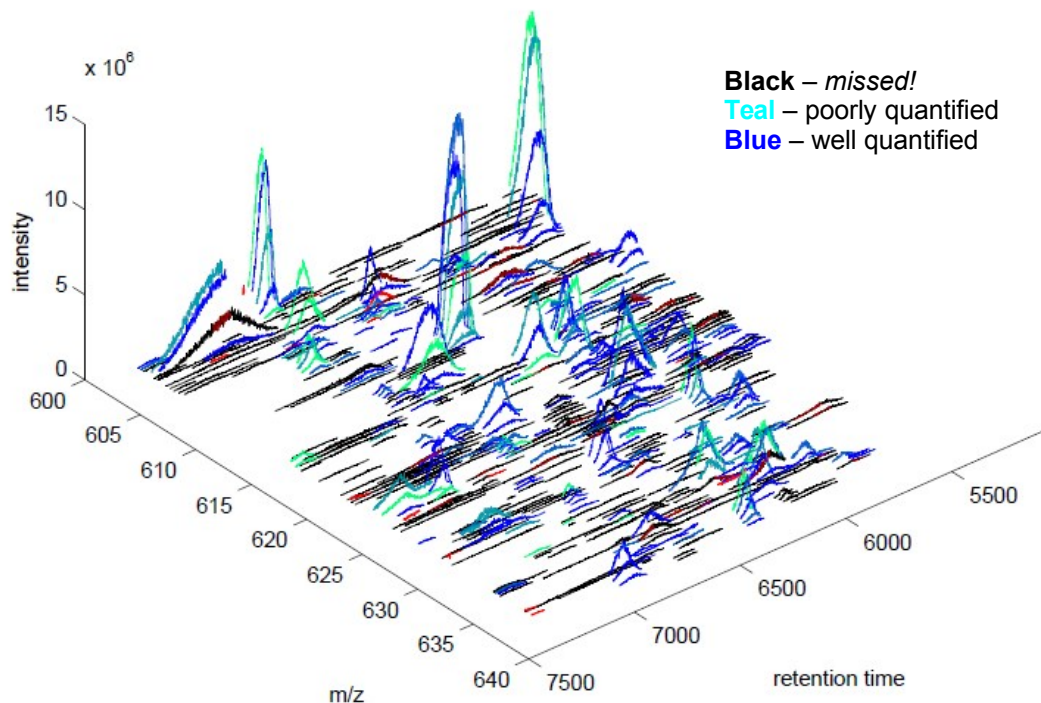


The Prince Lab is working on methods to extract useful information to increase the reach of current approaches and develop new ways of understanding disease.

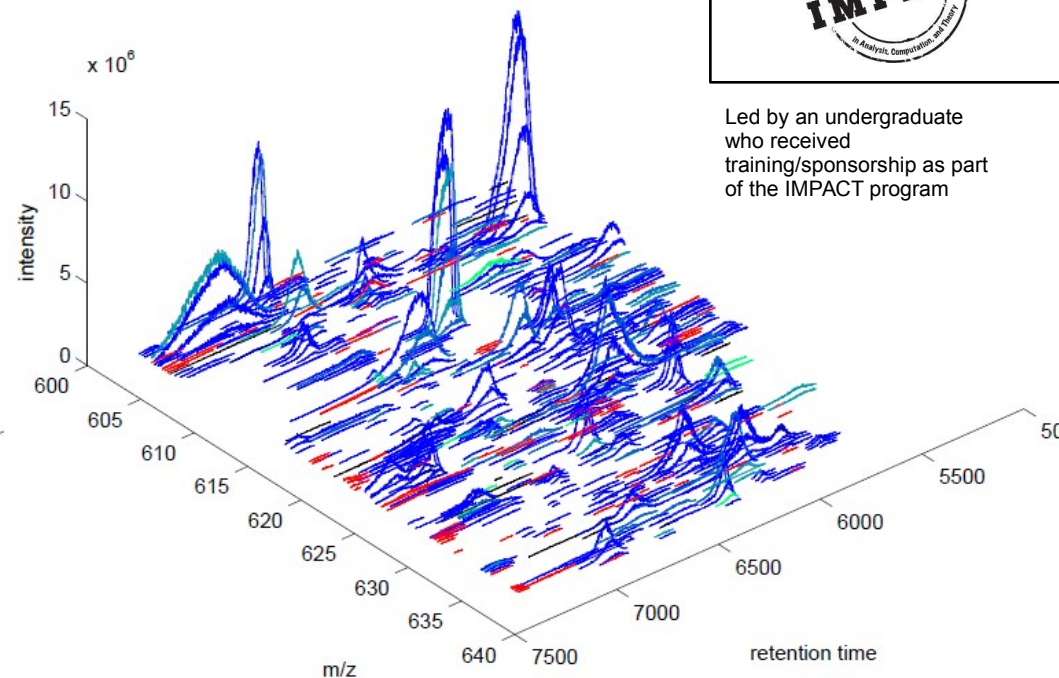
We wrote and characterized software that vastly increases the number of biomolecules that can be detected.



Led by an undergraduate who received training/sponsorship as part of the IMPACT program

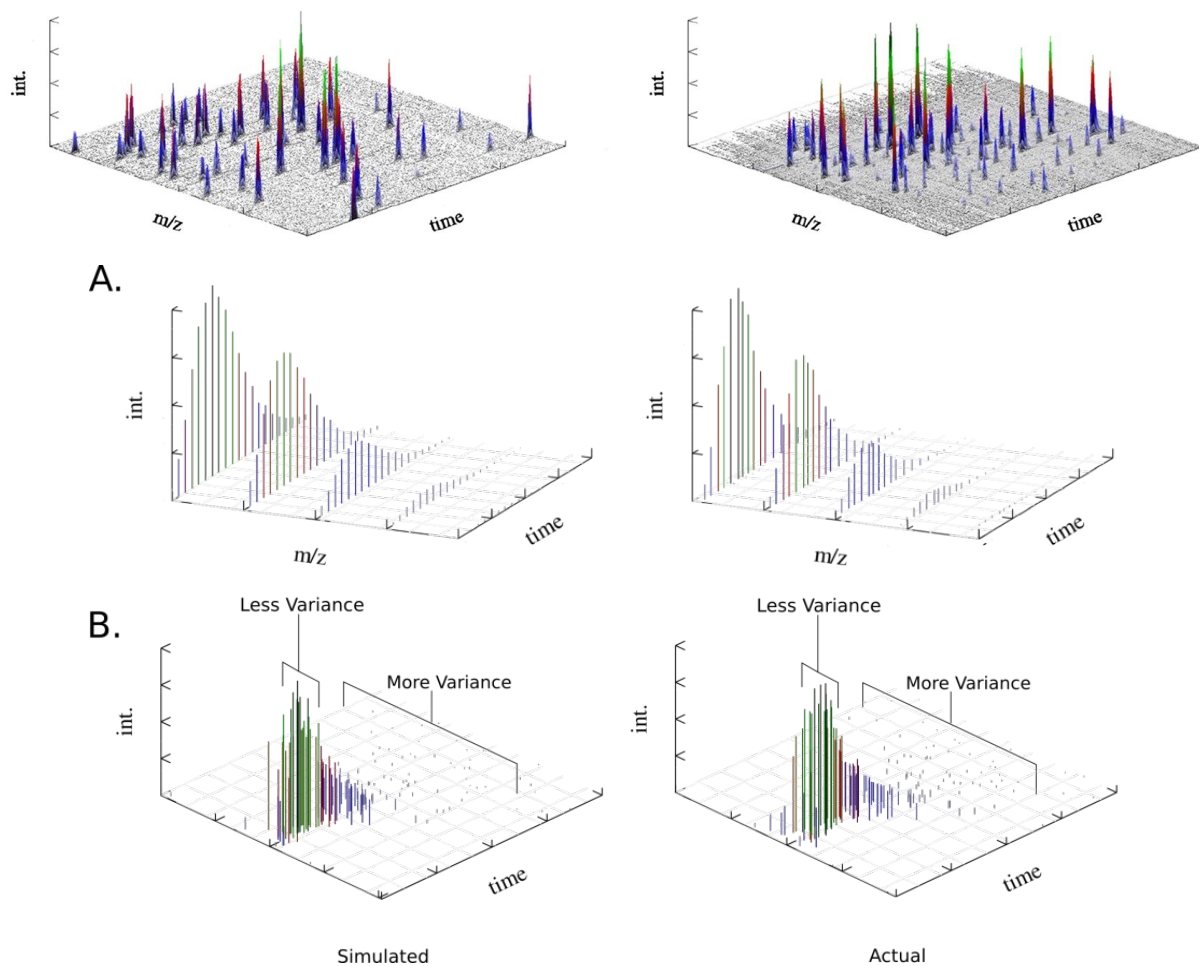


centwave

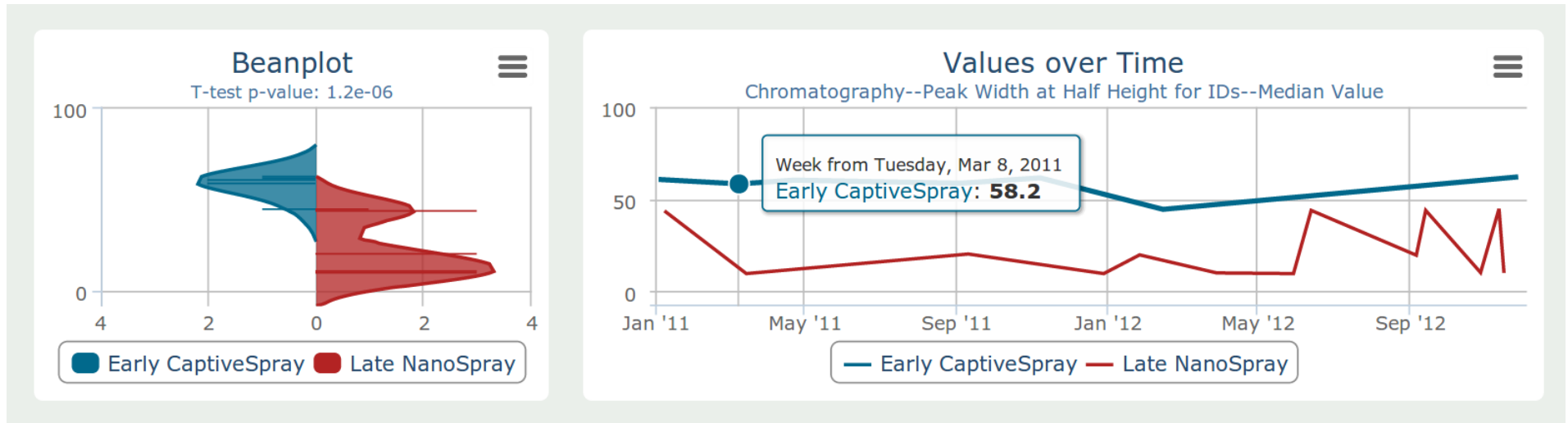


massifquant

We created a state-of-the-art mass spectrometry data simulator to help test software that extracts biomolecular signals.



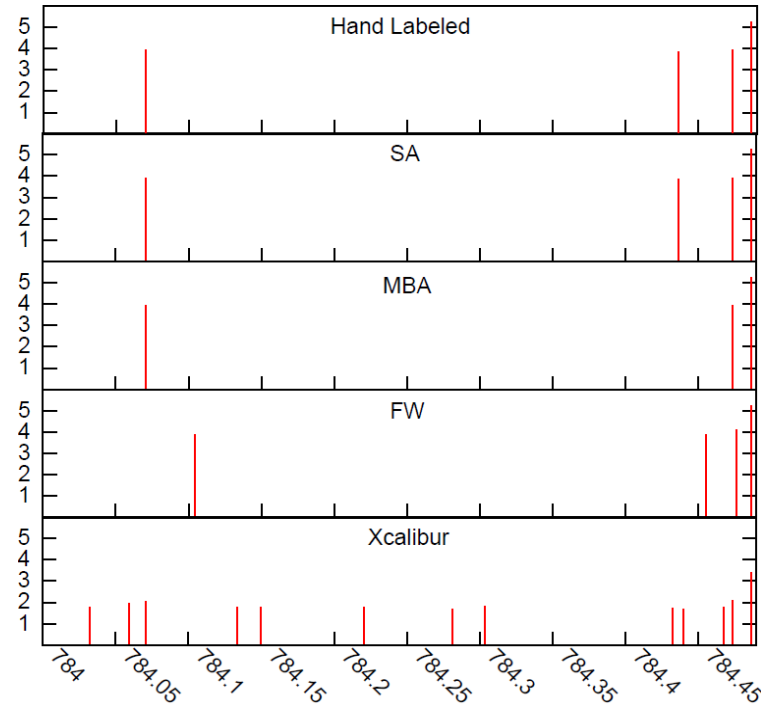
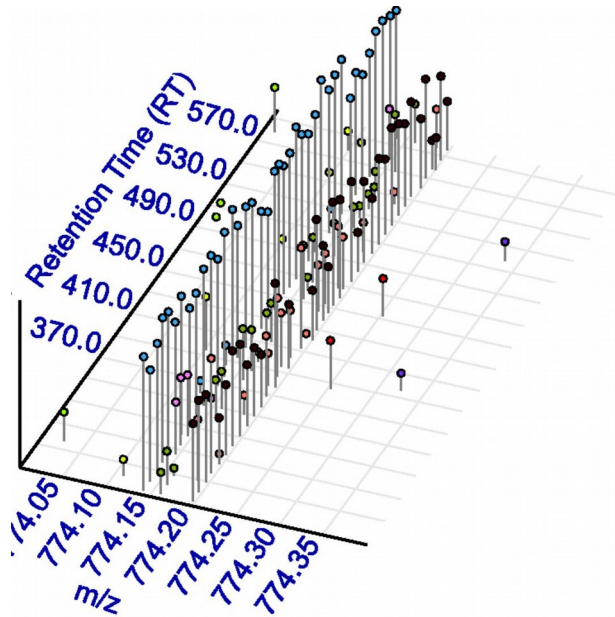
We developed a quality control web application that allows users to track over 300 quality metrics and alert them if performance is degrading.



We created software that is far more accurate at extracting lipid signals than current methods.



In collaboration with
Dan Ventura and
Rob Smith in
Computer Science



Smith R, Anthonymuthu TS, Ventura D, Prince JT. *Bioinformatics*. **2013**.

Smith R, Ventura D, Prince JT. *Brief Bioinform*. **2013**.

Smith R, Ventura D, Prince JT. *Bioinformatics*. **2013**.

In order to dramatically boost the number of lipids that we can identify, we are developing software to predict how a lipid will fragment inside a mass spectrometer. As a first step, we developed a high-level chemistry library to help us model lipid fragmentation with greater ease.



In collaboration with
Dan Ventura and
Rob Smith in
Computer Science

Draw

```
mol = Rubabel["NCC(O)C(=O)O"]  
mol.write("file.svg")
```

Add

```
mol = Rubabel["OCC"]  
# adds a carbon, then an oxygen to the previous carbon  
mol << 6 << 8 # #<Mol "OCCCCO">  
mol << :C << :O # same thing
```

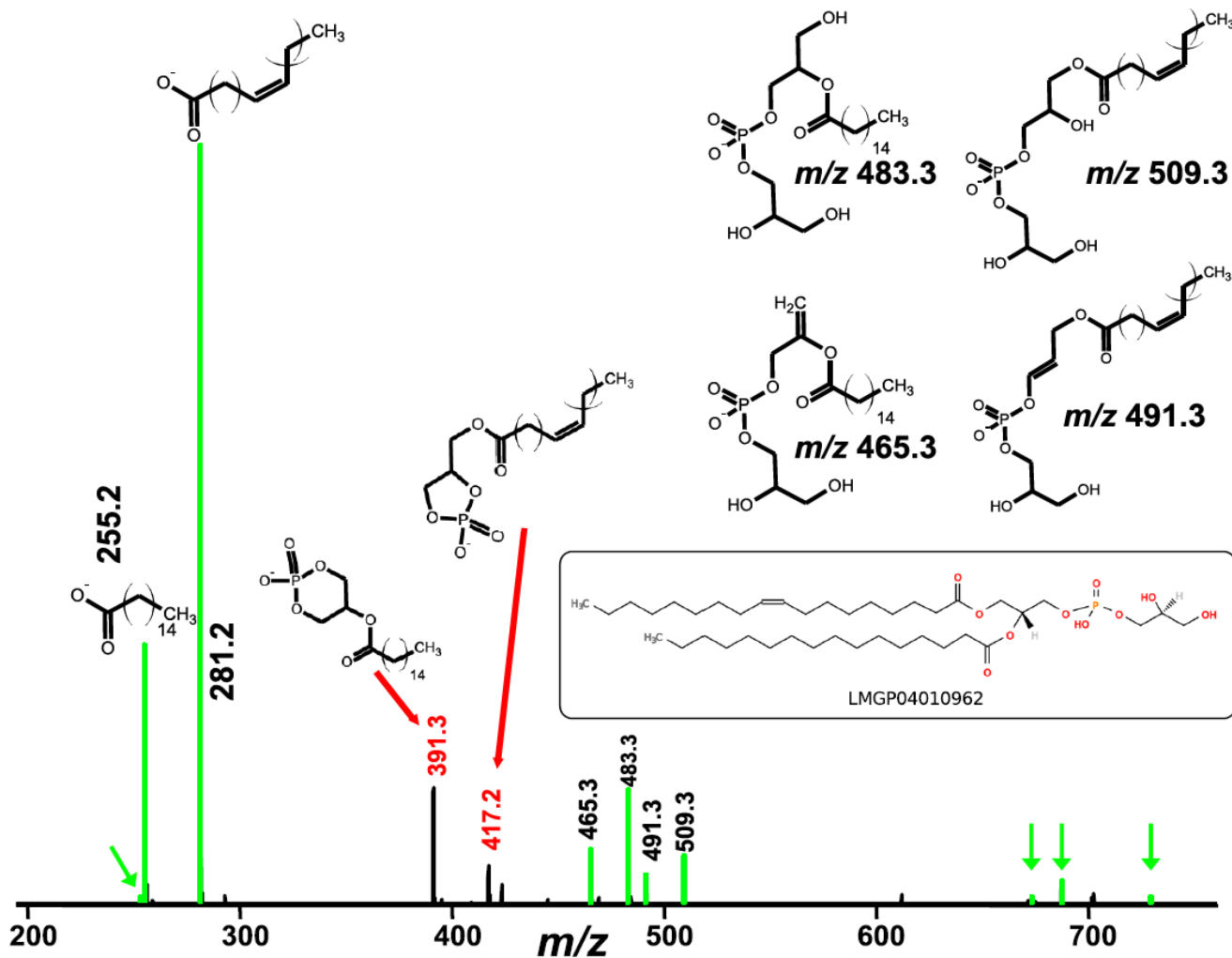
Search

```
mol = Rubabel["NCC(O)C(=O)O"]  
mol.each_match("CO") do |match|  
  # match is just an array of atoms that matched  
  match.first.el # => :C  
  match.last.el # => :O  
end
```

Split

```
bonds = mol.matches("CO").map {|c, o| c.get_bond(o) }  
mol.split(*bonds) # splits between every carbon single bonded to oxygen
```

Our software is now allowing us to predict how lipids will fragment in a mass spectrometer. We correctly predicted all the green peaks for this glycerophosphoglyceride. Just this last month we modeled the rearrangement events that allow us to predict the two fragments pointed to in red.



All three of my graduate students are scheduled to work as teaching assistants this semester. Support would allow them to work full-time on their projects and help bridge the gap in our lab between startup and NIH/NSF funding which we are currently seeking

- Adam Swensen is working to create a panel of lipid signatures for cancer cell lines, optimize methods for improving lipid detection, and characterize the way in which lipids are involved in pancreatic cancer.
- Huan Kang is working to measure the mitochondrial proteome and lipidome to create a model capable of predicting which chronic lymphocytic leukemia patients will respond to the anticancer drug flavopyridol.
- Ryan Taylor is working to model how lipids will fragment in a mass spectrometer.
- Many undergraduates are directly involved in these and similar projects (undergrads were the primary authors on two of the publications from this year and another half dozen were co-authors).