

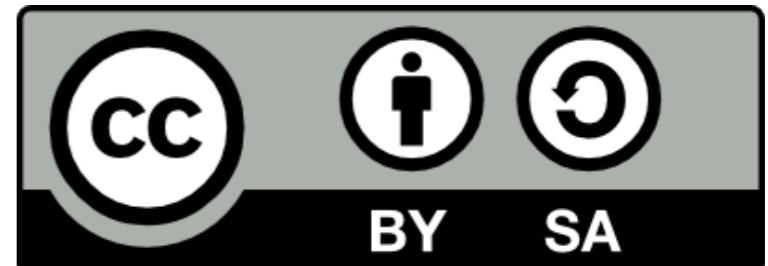
Genomics on the Half Shell: Making Science more Open

Steven B. Roberts
Associate Professor
School of Aquatic and Fishery Sciences
University of Washington

robertslab.info

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IP[y]: IPython
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oystergen.es/data



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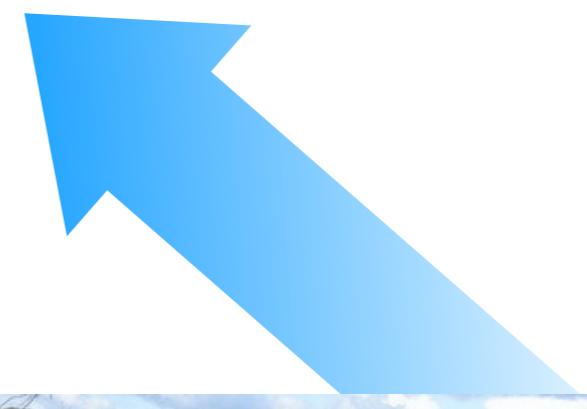
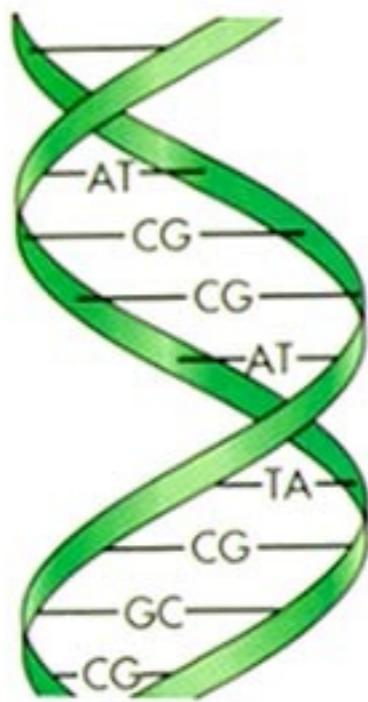
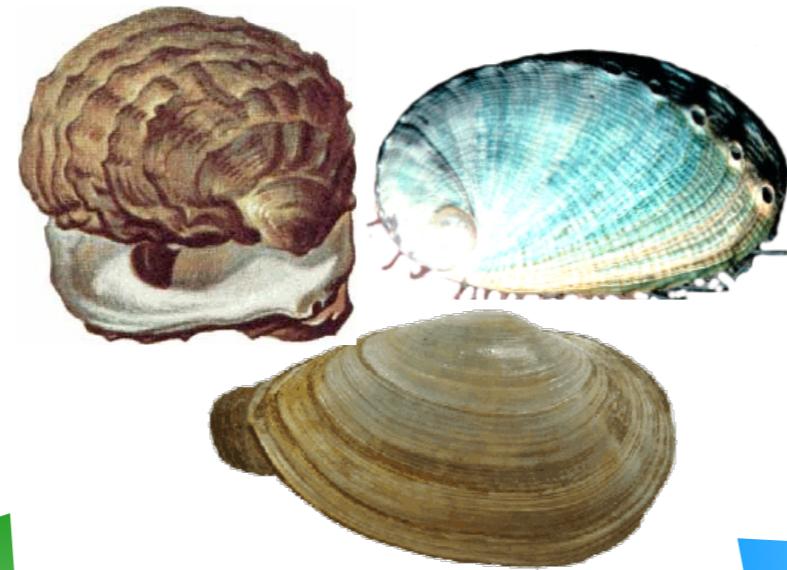
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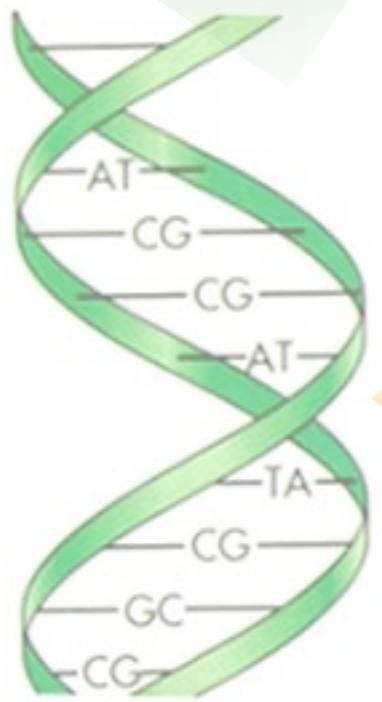


BIVALVE BANDSTAND



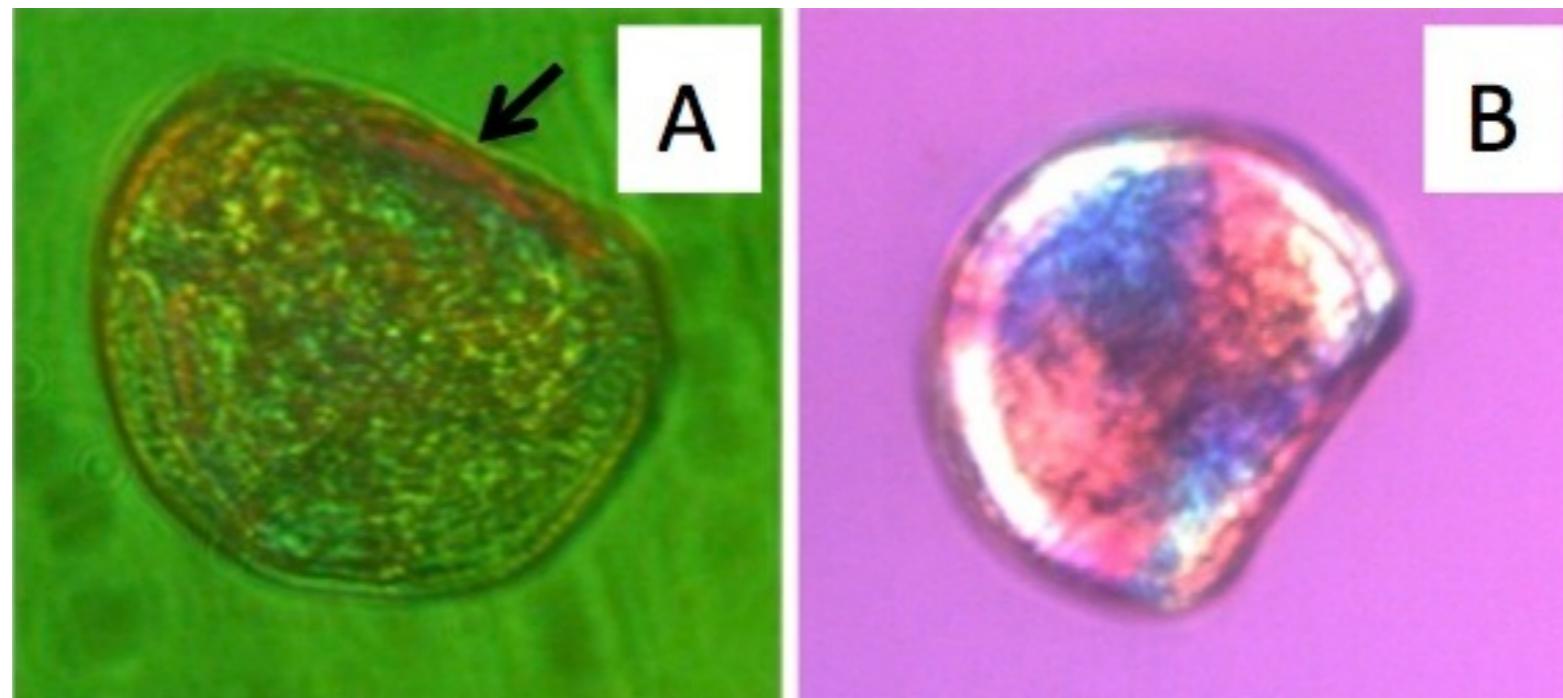






**Transcriptome
Proteome
DNA
Methylation**





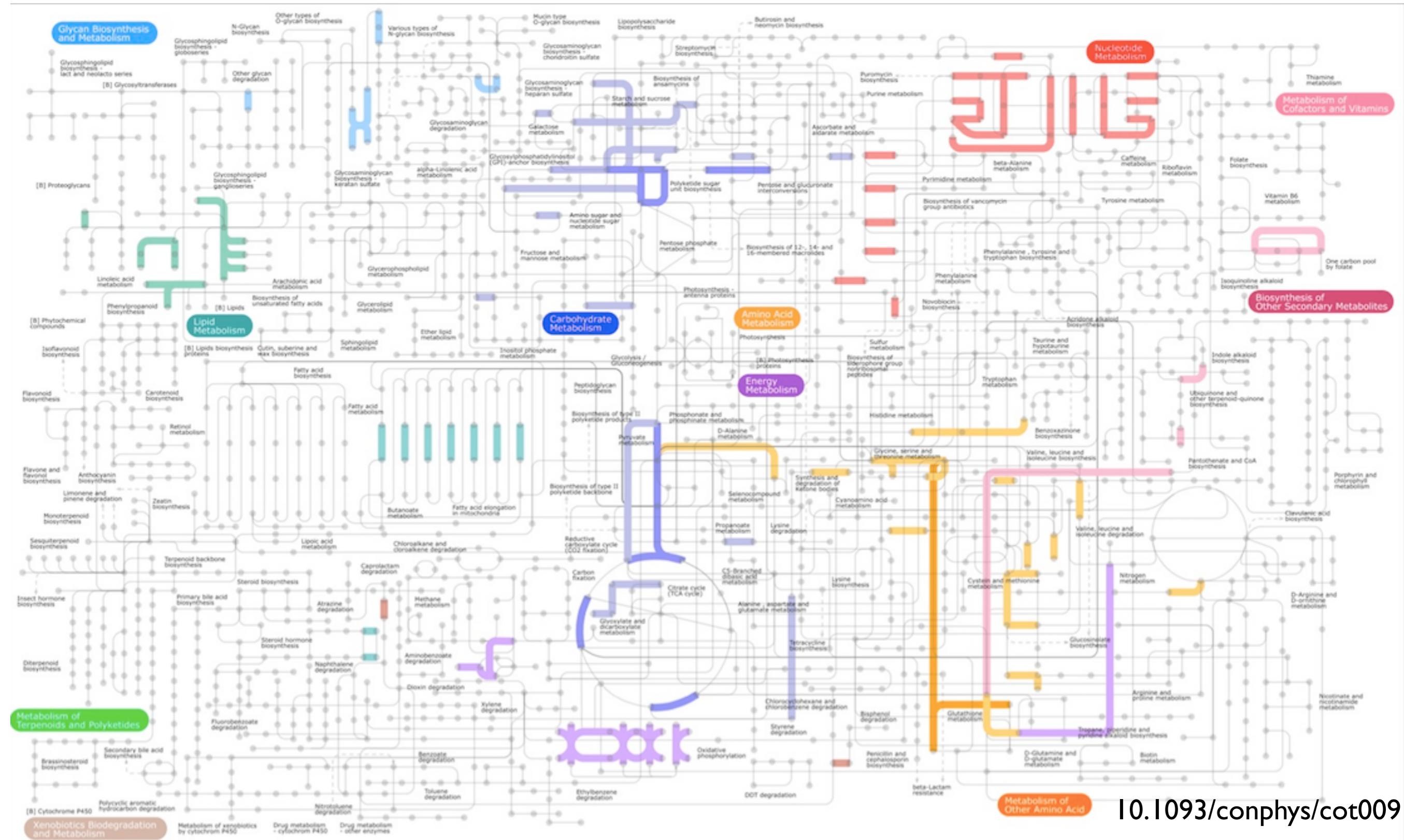
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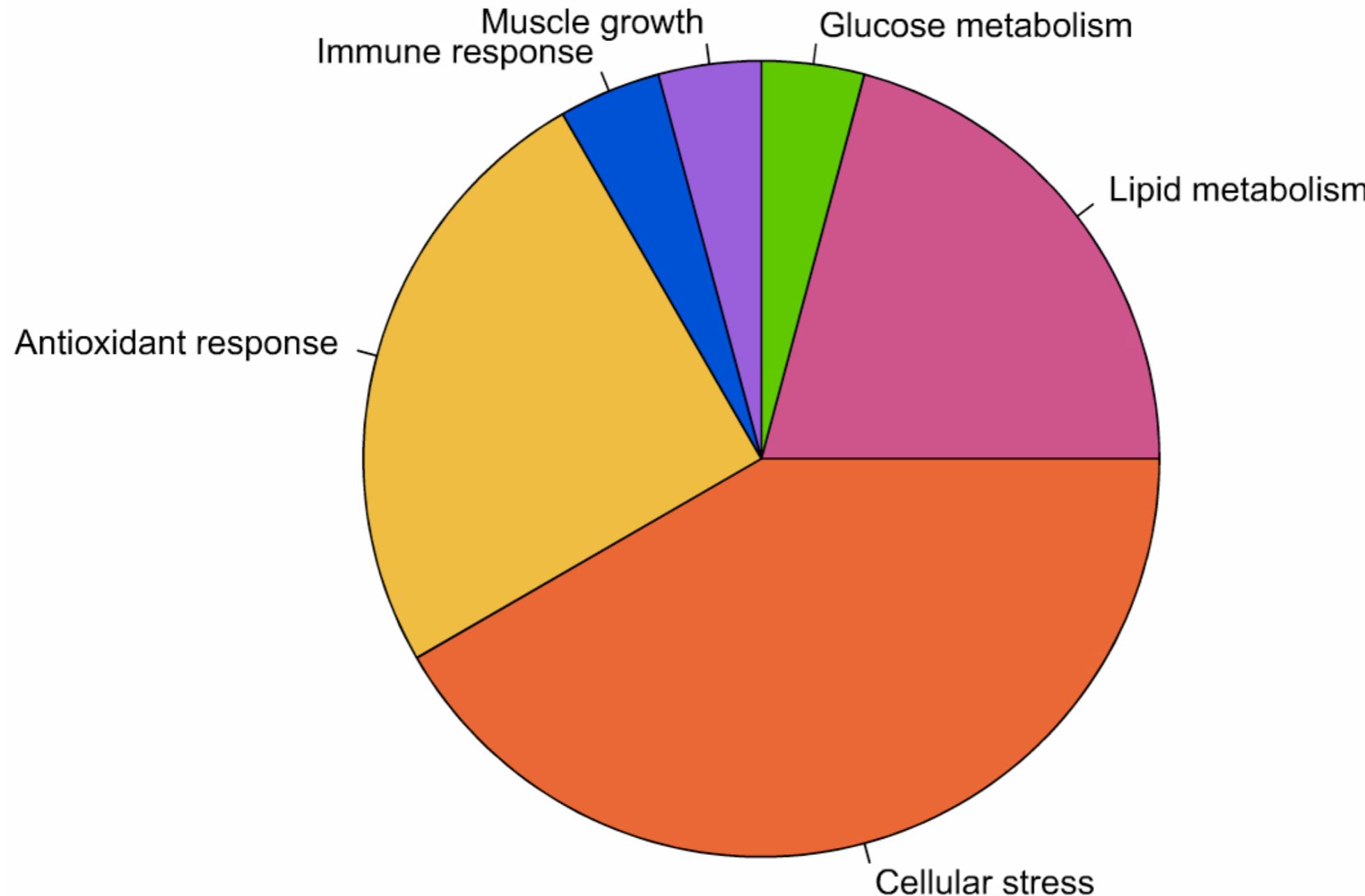
Elevated pCO₂ causes developmental delay in early larval Pacific oysters, *Crassostrea gigas*.
Timmings-Schiffman et al 2012

Shotgun Proteomics

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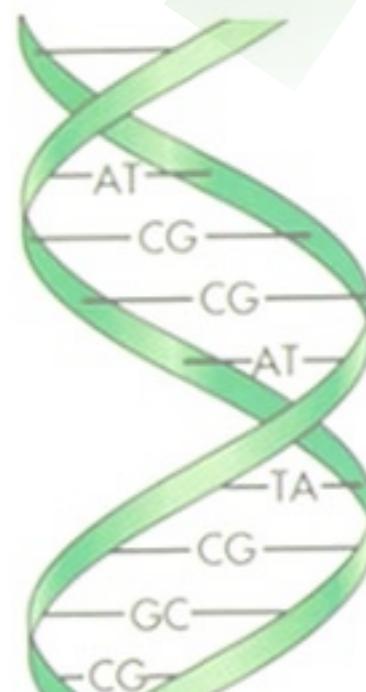
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Transcriptome Proteome

DNA Methylation





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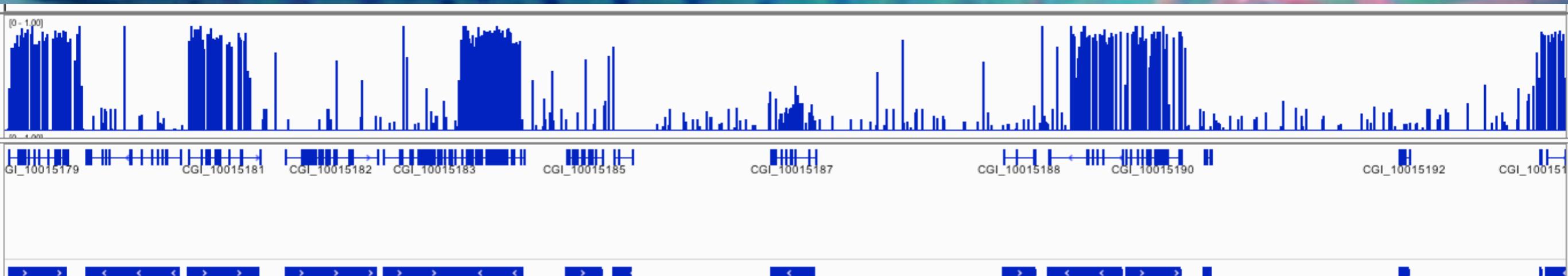
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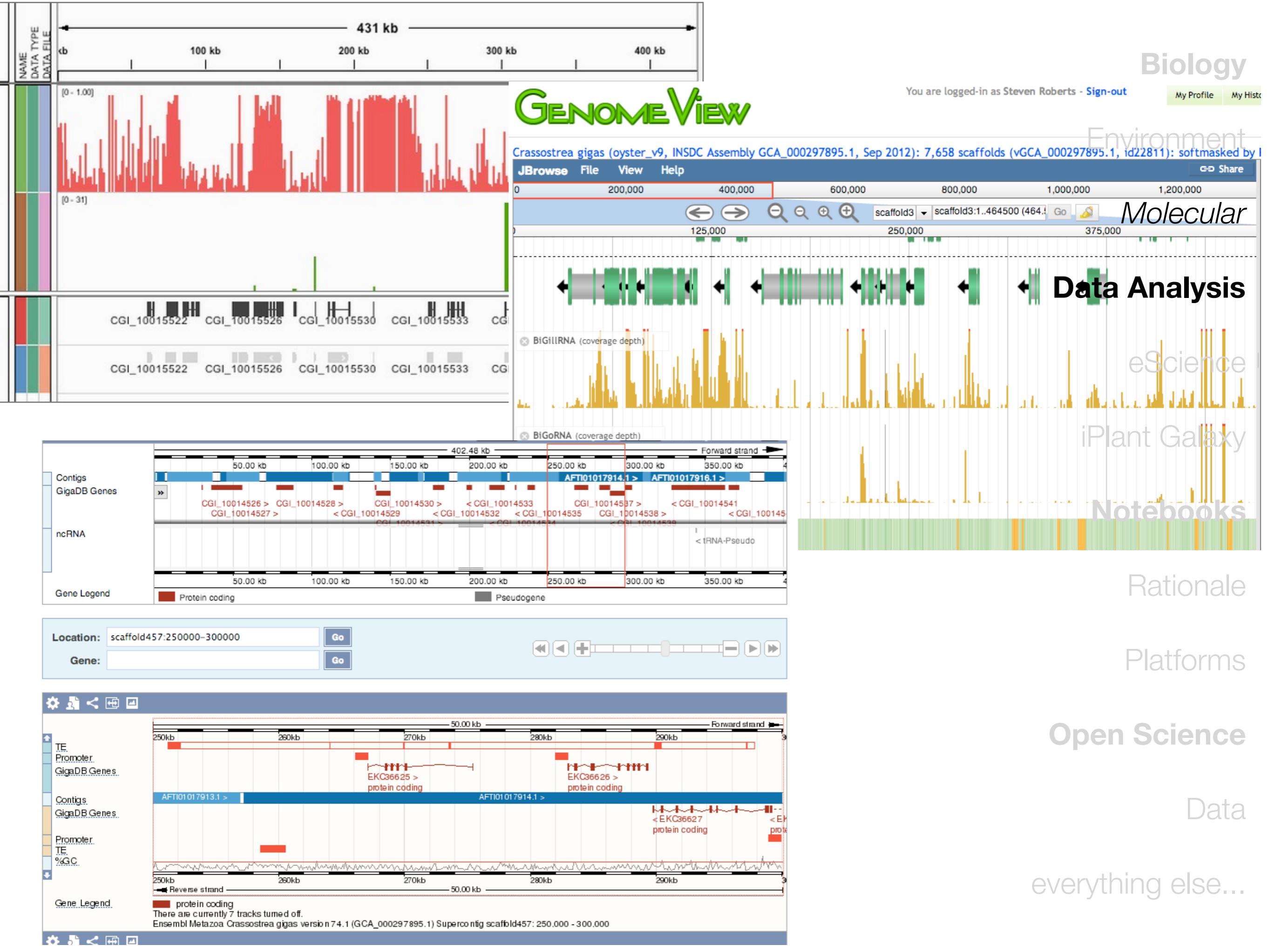
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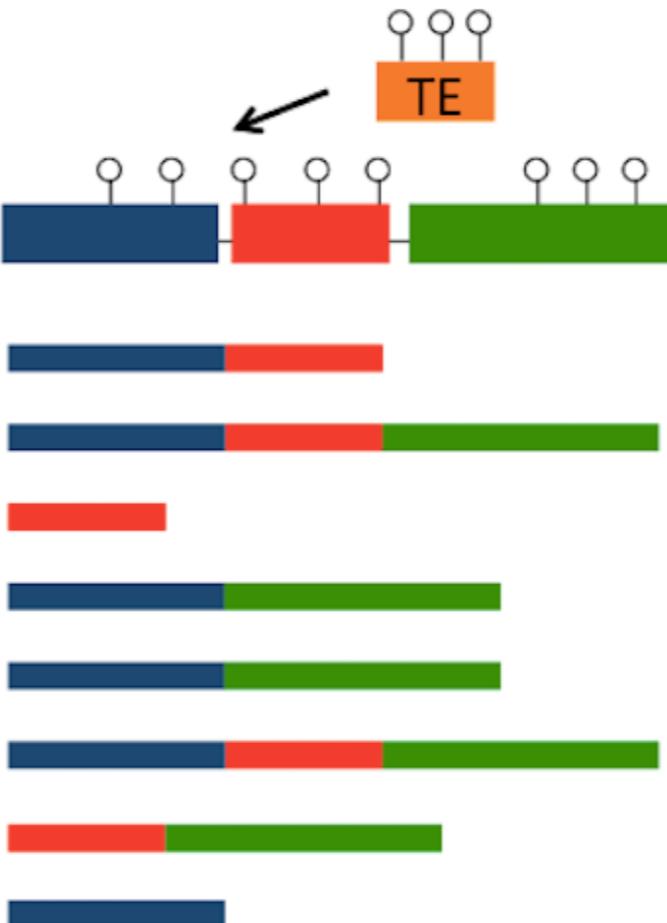
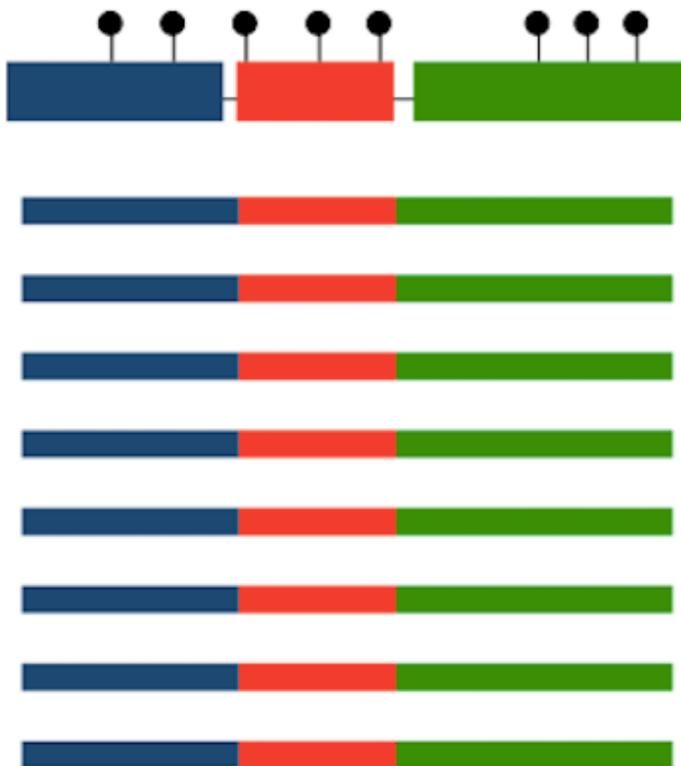
associated with gene bodies



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Stochastic Variation

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**A context dependent role for
DNA methylation in bivalves**

Mackenzie R. Gavery and Steven B. Roberts

10.1093/bfgp/elt054
10.6084/m9.figshare.880763

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 **Peter Cock**
@pjacock

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Biology

Or “advanced file format conversion”? RT

@pathogenomenick:

Bioinformatics... Or ‘advanced file copying’ as I like to call it.

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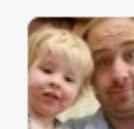
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FAVORITES

2



11:53 AM - 29 Jan 2014



John Chilton @jmchilton · Jan 29

@pjacock @pathogenomenick I'm jealous, all I've ever done is convert & copy but I've never succeeded to the point I'd call it advanced.

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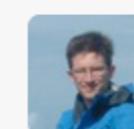


Nick Loman @pathogenomenick · Jan 29

@jmchilton @pjacock today it was all about the copying .. but I had to use all my skills!

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Peter Cock @pjacock · Jan 29

@jmchilton @pathogenomenick if you have to write your own parser & output code, especially for binary formats, then I'd call it advanced ;)

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Chris Cole @drchriscole · Jan 30

@pjacock @pathogenomenick I prefer, advanced data compression. Here's your 80gig of fastqs reduced to a bunch R plots.

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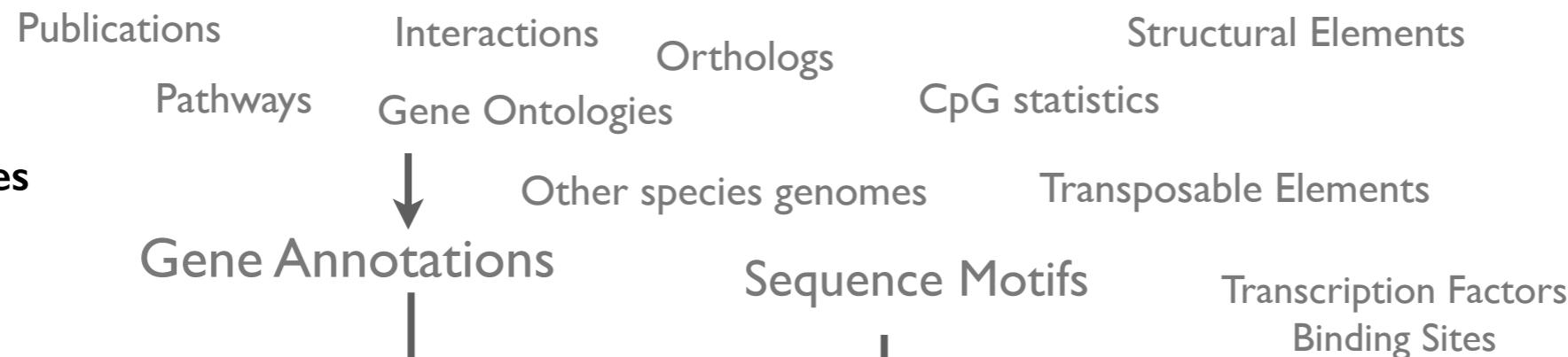
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raw - 70G
mapping - 60G
tables - 40G

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static

Data Tables**Genome***transcripts*

dynamic

Primary Data Table Groupings

- Size
- Growth
- Location
- Environment
- Stage
- Treatment
- Tissue
- Trait
- Strain

Genomic Data Types

RNA-Sequencing	Single Nucleotide Polymorphisms	DNA Methylation
Expressed Sequence Tags	Amplified Fragment Length Polymorphisms	Histone Modification
Expression Microarrays	Simple Sequence Repeats	miRNA Expression

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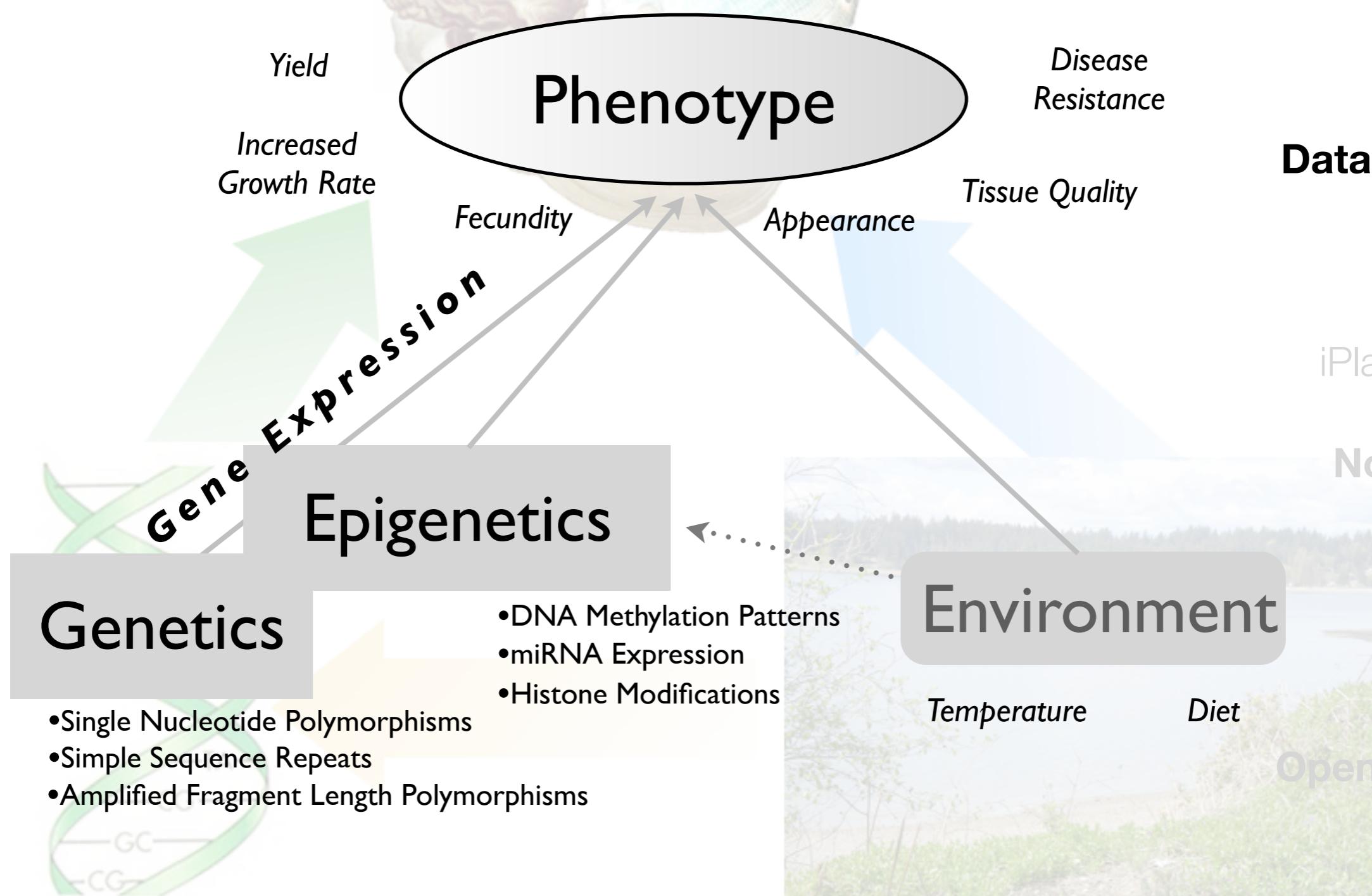
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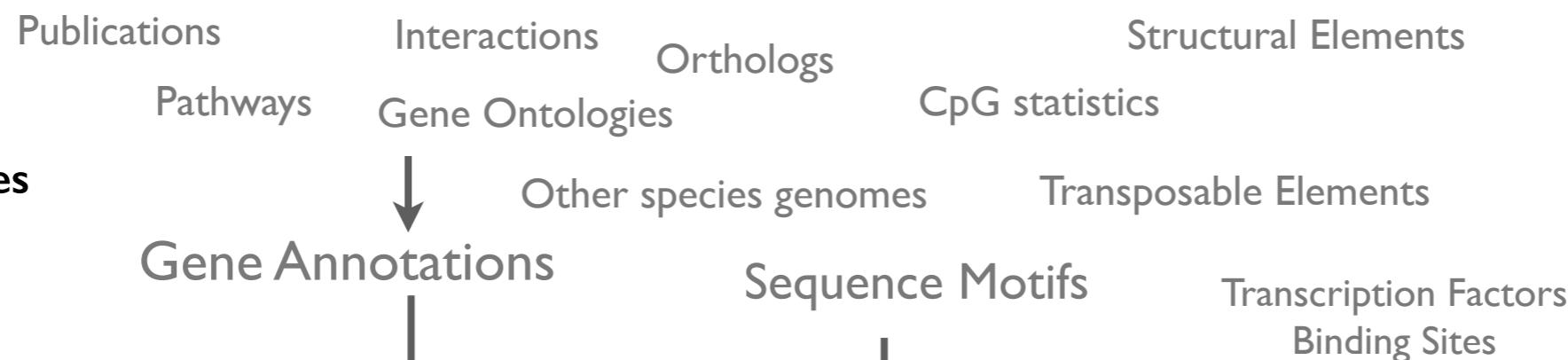
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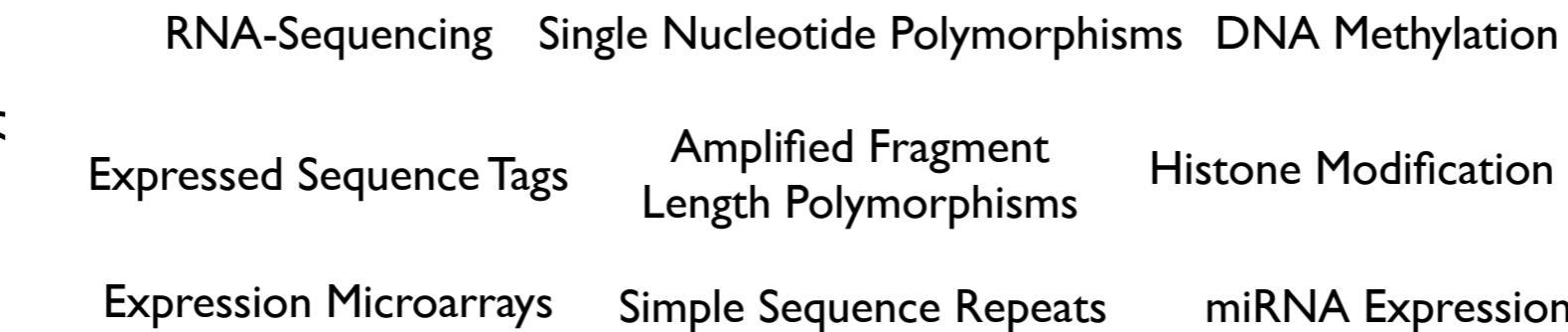
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dynamic

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- Size
- Growth
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- Trait
- Strain

Genomic Data Types

Preliminary Phase: Aggregating Datasets

Using SQLShare as a platform we have already begun to aggregate and format data. Anyone can view (and contribute) using the tag "qdod". Below is a table describing some of the relevant datasets. "Snapshot" provides you with a screenshot of the data in SQLShare and "Direct Link" brings you directly to the data in SQLShare. You can also [open the table in a new webpage](#).

qDOD online

qDOD_Cgigas_gene_fasta	sequence fasta file. Exon only.	http://goo.gl/oqCxl	https://sqlshare.escapevelocity.net/datasets/qdod/qDOD_Cgigas_gene_fasta
qDOD_Zhang_Gil_gene_RNA-seq	Gill RNA-seq data (gene based)	http://goo.gl/8oISR	https://sqlshare.escapevelocity.net/datasets/qdod/qDOD_Zhang_Gil_gene_RNA-seq
qDOD_Zhang_Mgo_gene_RNA-seq	Male Gonad RNA-seq data (gene based)	http://goo.gl/6buVz	https://sqlshare.escapevelocity.net/datasets/qdod/qDOD_Zhang_Mgo_gene_RNA-seq

everything else...

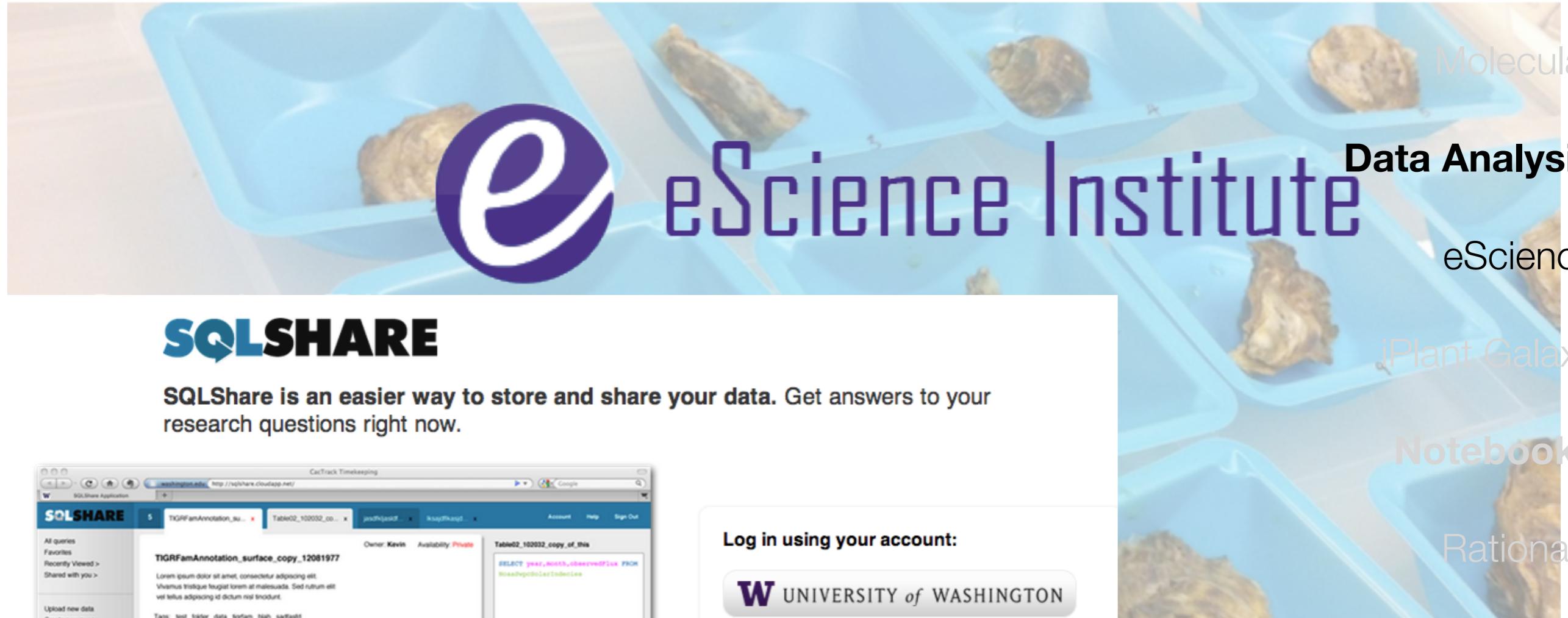
The background of the slide features a photograph of several oysters in blue plastic trays, arranged in a grid pattern. The oysters are partially open, showing their meat. The trays are set against a light-colored wooden surface.

eScience Institute

Querying Disparate Oyster Datasets | qDOD

The goal of this project is to produce a web-based interface for querying and visualizing *Crassostrea gigas* genomic datasets. This site serves as a portal for documenting our efforts, providing user access, as well as a means to gather feedback.

One can view (and contribute) using the
ides you with a screenshot of the data in
en the table in a new webpage.



SQLSHARE

SQLShare is an easier way to store and share your data. Get answers to your research questions right now.

Upload

Upload any tabular data and start analyzing instantly. No need to install, configure, or design a database.

Modify

Exercise the full power of SQL even with zero programming experience: joins, subqueries, set operations.

Share

Analyze and compare your data collaboratively. Derive new datasets and share them with your colleagues.

<https://sqlshare.eso.org/>
<https://sqlshare.eso.org/>
[seq](https://sqlshare.eso.org/)
<https://sqlshare.eso.org/>
[seq](https://sqlshare.eso.org/)

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BiGo_lar_T3D5	10	

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skyline	24	
oceanography	23	
ssgcid	18	
qdod2	18	
qdod	18	
swissprot	17	
suna	16	
tsg	16	

Your Datasets		
Name	Sharing / Owner	Modified
qdod_proteome_blast_mouse	sr320@washington.edu	Jan 2
qDOD_v9_gene GFF format file of oyster genes ~28k gene	sr320@washington.edu	Nov 1
_qdod_goslim_graphtest	sr320@washington.edu	Oct 2
SNP_RNAseqLibrary_SB_BiGill SNP table from RNA-seq library - SB gill tissue pool (BiGill complement) qdod2	sr320@washington.edu	Oct 2
BiGill_meth_Zhang_exp Gene-centric data including length, CG, percent methylation (gill) and tissue specific RPKM data from Zhang et al a qdod2	sr320@washington.edu	Oct 2
qDOD_Cgigas_gene_fasta Tabular format of Cgigas gene sequence fasta file Derived using Dataset: Genomic data from the Pacific oyster qdod2	sr320@washington.edu	Oct 2
qDOD_Cgigas_Gene_Descriptions (Swiss-prot) Description and evalutes associated with Cgigas 28k genes Derived using Dataset: Genomic blast	sr320@washington.edu	Oct 2
file0	sr320@washington.edu	Aug 1
BiGill meth with SP	sr320@washington.edu	Aug 1
SPID and GO Numbers Swiss-Prot IDs and corresponding GO numbers qdod	sr320@washington.edu	Aug 1
Cgigas_larvae_RNAseq_OsHV_GO	sr320@washington.edu	Jul 2
qDOD_Cgigas_GO_GOsliM_DISTINCT	sr320@washington.edu	Jul 2
Cgigas Larvae RNA-Seq OsHV UR10 RNA-seq data with descriptions of larvae exposed to OsHV. (>/= 10 UniqueReads) oyster	sr320@washington.edu	Jul 1
Cgigas Larvae RNA-Seq OsHV RNA-seq data with descriptions of larvae exposed to OsHV oshv	sr320@washington.edu	Jul 1
Zhang_Mgo_gene_RNA-seq_IGV sperm	sr320@washington.edu	Jun 2
Zhang_Gil_gene_RNA-seq_IGV IGV format rna-seq	sr320@washington.edu	Jun 2
BiGill_methratio_Gene_Genomic_GFF GFF formatted file indicated DNA methylation on oyster genes qdod	sr320@washington.edu	May
TJGR_GeneBased_CDS_GFF GFF format file with exons indicated for genes in oyster genome qdod	sr320@washington.edu	May
BiGill_Gene_Methratio_VD	sr320@washington.edu	May
oyster_v9_mRNA_GFF GFF (gene) from Zhang et al. Column9 modified for Joining qdod	sr320@washington.edu	May
Cgigas_gene_length CDS only	sr320@washington.edu	May

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qdod	18	
swissprot	17	
suna	16	
tsg	16	

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Name	Sharing / Owner	Modi...
qdod_proteome_blast_mouse	🔒 sr320@washington.edu	Jan 2
qDOD_v9_gene GFF format file of oyster genes ~28k gene	🔒 sr320@washington.edu	Nov 1
_qdod_goslim_graphtest		Oct 2
SNP_RNAseqLibrary_SB_BiGill SNP table from RNA-seq library - SB qdod2		Oct 2
BiGill_meth_Zhang_exp Gene-centric data including length, CG, perc qdod2		Oct 2
qDOD_Cgigas_gene_fasta Tabular format of Cgigas gene sequence qdod2		Oct 2
qDOD Cgigas Gene Descriptions (Swiss-prot) Description and evalua blast		Oct 2
file0	🔒 sr320@washington.edu	Aug 1
BiGill meth with SP	🔒 sr320@washington.edu	Aug 1
SPID and GO Numbers Swiss-Prot IDs and corresponding GO numbers qdod	↳ sr320@washington.edu	Aug 1
Cgigas_larvae_RNAseq_OsHV_GO	↳ sr320@washington.edu	Jul 2
qDOD_Cgigas_GO_GOsli..._DISTINCT	↳ sr320@washington.edu	Jul 2
Cgigas Larvae RNA-Seq OsHV UR10 RNA-seq data with descriptions of larvae exposed to OsHV. (>/= 10 UniqueReads) oyster	↳ sr320@washington.edu	Jul 1
Cgigas Larvae RNA-Seq OsHV RNA-seq data with descriptions of larvae exposed to OsHV oshv	↳ sr320@washington.edu	Jul 1
Zhang_Mgo_gene_RNA-seq_IGV sperm	↳ sr320@washington.edu	Jun 2
Zhang_Gil_gene_RNA-seq_IGV IGV format rna-seq	↳ sr320@washington.edu	Jun 2
BiGill_methratio_Gene_Genomic_GFF GFF formatted file indicated DNA methylation on oyster genes qdod	↳ sr320@washington.edu	May
TJGR_GeneBased_CDS_GFF GFF format file with exons indicated for genes in oyster genome qdod	↳ sr320@washington.edu	May
BiGill_Gene_Methratio_VD	↳ sr320@washington.edu	May
oyster_v9_mRNA_GFF GFF (gene) from Zhang et al. Column9 modified for Joining qdod	↳ sr320@washington.edu	May
Cgigas_gene_length CDS only	↳ sr320@washington.edu	May

Use Cases

- Joining on Annotations
- File Conversion
- Querying Gene Tables

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BiGill_meth_Zhang_exp Gene-centric data including length, CG, perc qdod2		Oct 2
qDOD_Cgigas_gene_fasta Tabular format of Cgigas gene sequence qdod2		Oct 2
qDOD Cgigas Gene Descriptions (Swiss-prot) Description and evalua blast		Oct 2
Predominant intragenic methylation is associated with gene expression charact Predominant intragenic methylation is associated with gene expression charact	sr320@washington.edu	Aug 2
	sr320@washington.edu	Jul 2
	sr320@washington.edu	Jul 2
	sr320@washington.edu	Jul 1
	sr320@washington.edu	Jul 1
	sr320@washington.edu	Jul 1
	sr320@washington.edu	Jun 2
	sr320@washington.edu	May

Use Cases

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- File Conversion
- Querying Gene Tables

Predominant intragenic methylation is associated with gene expression characteristics in a bivalve mollusc

Mackenzie R. Gavery, Steven B. Roberts

oshv	
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Zhang_Gil_gene_RNA-seq_IGV IGV format	
	sr320@washington.edu
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	sr320@washington.edu
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Predominant intragenic methylation is associated with gene expression character

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qDOD_Cgigas_Gene Descriptions (Swiss-prot) Description and evaluates associations with Cgigas 28k genes Derived using Dataset: Genomic	sr320@washington.edu	Oct 2
blast	sr320@washington.edu	Aug 2

DNA methylation landscape of genomic features

In order to examine relationships between DNA methylation and genomic features, data from BSMAP (i.e., *methratio*) was converted to genomic feature tracks (i.e., generic feature format [GFF] files). Conversion was done using SQLShare ([Howe et al., 2011](#)), with the files and corresponding query language published ([Gavery & Roberts, 2013](#)).

The distribution of methylated CpGs with respect to specific genomic features was determined using BEDtools (i.e., *intersectBED*) ([Quinlan & Hall, 2010](#)). For this analysis, a CpG locus was considered methylated if at least half of the reads remained unconverted after bisulfite treatment. Genomic features that were examined include: exons and introns ([Fang et al., 2012](#)), putative promoters (defined as 1 kb upstream of open reading frames), and transposable elements. Putative transposable elements were

Use Cases

- Joining on Annotations
- File Conversion
- Querying Gene Tables

Gavery M, Roberts S. 2013. Crassostrea gigas high-throughput bisulfite sequencing (gill tissue). [figshare](#). Retrieved 18:44, Oct 21, 2013 (GMT)

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Predominant intragenic

DNA methylation landscape of genomic features

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BiGill_CpG_methylation.igv	preview	download
BiGill_exon_clc_rpkm.igv	preview	download
BiGill_ivg_charlie.xml	preview	download
Query to derive_CG_AllData_IGV.txt	preview	download

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Filesize in total is 92.16 MB

Categories

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Authors

Mackenzie Gavery
Steven Roberts

Tags

- epigenetics
- Gene regulation
- invertebrate
- oyster
- dna methylation

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*The embed functionality can only be used for non commercial purposes... [more](#)

Description

This fileset contains genomic feature tracks from methylation-enriched high-throughput bisulfite sequencing and RNA-seq analysis for Pacific oyster (*Crassostrea gigas*) gill tissue. Feature tracks were developed to be viewed with Integrative Genomics Viewer (<http://www.broadinstitute.org/igv/>) in conjunction with the *C. gigas* genome (Fang et al. 2012). All data and instructions are also available at <http://oystergen.es/bigill>.

File descriptions:

BiGill_CpG_methylation.igv - Location and proportion of methylation for all analyzed CpG dinucleotides with greater than 5x coverage.

BiGill_exon_clc_rpkm.igv - Exon-specific gene expression values (RPKM) from RNA-seq analysis.

BiGill_ivg_charlie.xml - A session file, which loads methylation and RNA-seq feature tracks as well as the location of *C. gigas* genome features.

Query to derive_CG_AllData_IGV.txt - Query (SQLShare) used to derive the methylation feature track from the original methratio output (<http://goo.gl/5LGq9Q>)

Reference:

Fang X, Li L, Luo R, Xu F, Wang X, Zhu Y, Yang L, Huang Z. 2012. Genomic data from the Pacific

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Filter dataset by keyword:

Name	Sharing / Owner	Modi...
qdod_proteome_blast_mouse	sr320@washington.edu	Jan 2
qDOD_v9_gene GFF format file of oyster genes ~28k gene	sr320@washington.edu	Nov 1
_qdod_goslim_graphtest	sr320@washington.edu	Oct 2
SNP_RNAseqLibary_SB_BiGill SNP table from RNA-seq library - SB gill tissue pool / BiGill complement	sr320@washington.edu	Oct 2
BiGill_meth_Zhang_exp Gene-centric data including length, CG, percent methylation (gill) and tissue specific RPKM data from Zhang et al	sr320@washington.edu	Oct 2
qDOD_Cgigas_gene_fasta Tabular format of Cgigas gene sequence fasta file Derived using Dataset: Genomic data from the Pacific oyster	sr320@washington.edu	Oct 2
qDOD Cgigas Gene Descriptions (Swiss-prot) Description and evaluates associations with Cgigas (~28k genes) Derived using Dataset: Genomic data from the Pacific oyster	sr320@washington.edu	Oct 2

Secondary stress: proteomics

Original input file had some peptides of charge state >2, so had to redo everything with fixed input file.

SR discovered that for some proteins, a peptide was sequenced multiple times and so had multiple expression values. From the unique protein associations file in SQLshare, I summed the expression values for all identical peptides.

```
SELECT [peptide sequence], SUM([2_01 TotalArea]) AS CG2_01, SUM([2_02 TotalArea]) AS CG2_02, SUM([2_03
TotalArea]) AS CG2_03, SUM([5_01 TotalArea]) AS CG5_01, SUM([5_02 TotalArea]) AS CG5_02, SUM([5_03
TotalArea]) AS CG5_03, SUM([8_01 TotalArea]) AS CG8_01, SUM([8_02 TotalArea]) AS CG8_02, SUM([8_03
TotalArea]) AS CG8_03, SUM([11_01 TotalArea]) AS CG11_01, SUM([11_02 TotalArea]) AS CG11_02, SUM([11_03
TotalArea]) AS CG11_03, SUM([26_01 TotalArea]) AS CG26_01, SUM([26_02 TotalArea]) AS CG26_02, SUM([26_03
TotalArea]) AS CG26_03, SUM([29_01 TotalArea]) AS CG29_01, SUM([29_02 TotalArea]) AS CG29_02, SUM([29_03
TotalArea]) AS CG29_03, SUM([32_01 TotalArea]) AS CG32_01, SUM([32_02 TotalArea]) AS CG32_02, SUM([32_03
TotalArea]) AS CG32_03, SUM([35_01 TotalArea]) AS CG35_01, SUM([35_02 TotalArea]) AS CG35_02, SUM([35_03
TotalArea]) AS CG35_03, SUM([221_01 TotalArea]) AS CG221_01, SUM([221_02 TotalArea]) AS CG221_02,
SUM([221_03 TotalArea]) AS CG221_03, SUM([224_01 TotalArea]) AS CG224_01, SUM([224_02 TotalArea]) AS
CG224_02, SUM([224_03 TotalArea]) AS CG224_03, SUM([227_01 TotalArea]) AS CG227_01, SUM([227_02
TotalArea]) AS CG227_02, SUM([227_03 TotalArea]) AS CG227_03, SUM([230_01 TotalArea]) AS CG230_01,
SUM([230_02 TotalArea]) AS CG230_02, SUM([230_03 TotalArea]) AS CG230_03,
SUM([242_01 TotalArea]) AS CG242_01, SUM([242_02 TotalArea]) AS CG242_02, SUM([242_03 TotalArea]) AS
CG242_03, SUM([245_01 TotalArea]) AS CG245_01, SUM([245_02 TotalArea]) AS CG245_02, SUM([245_03
TotalArea]) AS CG245_03, SUM([248_01 TotalArea]) AS CG248_01, SUM([248_02 TotalArea]) AS CG248_02,
SUM([248_03 TotalArea]) AS CG248_03, SUM([251_01 TotalArea]) AS CG251_01, SUM([251_02 TotalArea]) AS
CG251_02, SUM([251_03 TotalArea]) AS CG251_03
```

Use Cases

- Joining on Annotations
- File Conversion
- Querying Gene Tables

Visualize Dataset

SQLSHARE

Graphs

 SQL
 Share
 </> Embed

```
SELECT TOP 100 * FROM (SELECT * FROM [che625@washington.edu].[BiGO_MethylatedCGs_oysterv9]) x
```

Click to edit

Data Analysis

Logged in: sr320@washington.edu

eScience

Plant Galaxy

Notebooks

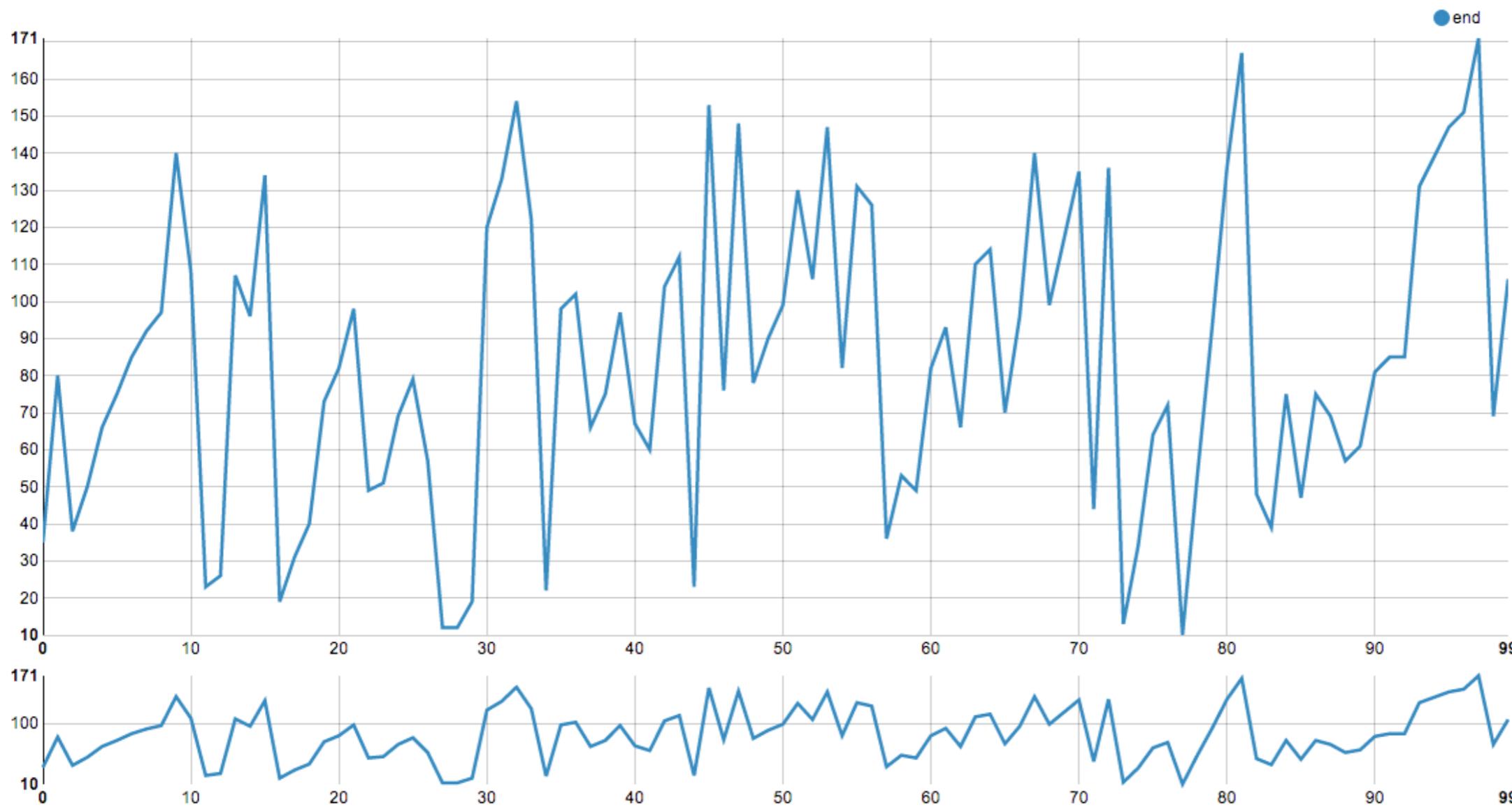
Rationale

Platforms

on Science

Data

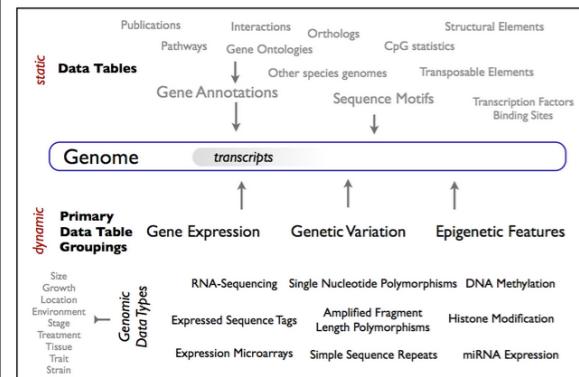
ything else...

Graph  Share</> Embed

qdod: Querying Disparate Oyster Datasets

This repository provides access to genomic data and workflows (IPython notebooks) that are being integrated as part of effort to increase efficiency of biological discovery. The [wiki](#) associated with this repository will serve as the *primary means for documentation*. Currently the documentation is focused on 1) **describing current datasets** and 2) **providing workflow tutorials**.

In brief, data in the form of delimited text files is aggregated into [SQLShare](#) where they can be easily queried. Below is schematic representation of the different types of datasets.



During the initial phases the focus is on the Pacific oyster and primary data from the [Roberts Lab](#).

Select IPython Notebooks

- [Static Data Tables - Universal](#)
- [Static Data Tables - Annotations](#)

Biology

Environment

Molecular

Data Analysis

eScience

iPlant Galaxy

Notebooks

Rationale

Platforms

Open Science

Data

everything else...

Currently the documentation is focused on 1) **describing current datasets** and 2) **providing workflow tutorials**.

A. Raw Data

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B. Datasets in SQLShare

- [Universal](#)
- [Generic Oyster Datasets](#)
- [Tissue Specific Oyster Datasets](#)

C. Tutorials

- [Simple Gene Search](#)
- [Standard SQLShare Queries](#)
- [Annotating Genes](#)
- [File Format Conversions](#)

D. Genome Browser Feature Tracks

- [Canonical Tracks](#)
- [Bisulfite sequencing \(gill tissue\)](#)
- [Reference Genome Files](#)

Please use [GitHub's Issue feature](#) to ask question, report problems, or suggest features.

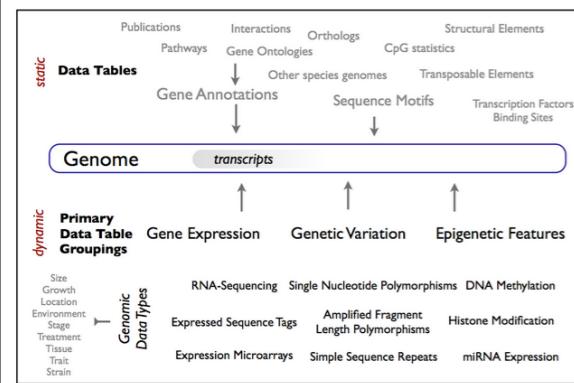
Last edited by sr320, 9 days ago



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Select Genomic Data

ID	Platform	Molecule	Tissue	Length	Files
BB3	SOLiD	RNA	gill	25 x 1	csfasta ; qual
DH3	SOLiD	RNA	gill	25 x 1	csfasta ; qual
DH2	SOLiD	RNA	gill	25 x 1	csfasta ; qual
GE	SOLiD	RNA	larvae	50 x 1	csfasta ; qual
GC	SOLiD	RNA	larvae	50 x 1	csfasta ; qual
SBunmeth	SOLiD	DNA	gill	25 x 1	csfasta ; qual
SBmeth	SOLiD	DNA	gill	25 x 1	csfasta ; qual
BSseqGill	Illumina	DNA	gill	36 x 1	fastq
ETStagseq	Illumina	RNA	gill		zip
BSseqSperm	Illumina	DNA	sperm	72 x 2	fastq1 ; fastq2
BiGillRNA	Illumina	RNA	gill	50 x 2	fastq1 ; fastq2
BiGoRNA	Illumina	RNA	sperm	50 x 2	fastq1 ; fastq2

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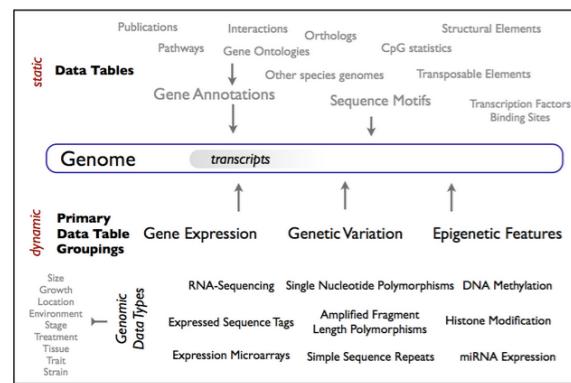
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BB3	SOLiD	RNA	gill	25 x 1	csfasta ; qual
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Data Snapshots

Select datasets available from SQLShare. Tag: [qdod2](#)
<https://sqlshare.esciencewashington.edu/sqlshare/#s=tag/qdod2>

Universal

Dataset	Screenshot	more
UniprotProtNamesReviewed_yes20130610	<p>All entries in UniProt with protein names, gene names, species and Swiss-Prot IDs that have been reviewed. File was generated in the following fashion:

 Go here: https://www.uniprot.org/uniprot/reviewed

 Then, towards the top-right side of the page, there's a "Download" button. Clicking on the "Download" button will take you to a new page that will allow you to download the file in either FASTA or tab-delimited text format.</p> <pre> SELECT SPID, Status, (Protein Name) AS ProteinName, (Gene Name) AS GeneName, Organism, (Length) FROM [zenelab@localhost:5432].(stable_UniprotProtNamesReviewed_yes20130610) </pre>	etc

SPID and GO Number...	more																		
<p>SPID and GO Numbers -> Visible by everyone Swiss-Prot IDs and corresponding GO numbers gox_ipdb swiss-prot</p> <pre> SELECT Distinct AGA000 AS SPID,"GO:0003824" AS GOID FROM [dhalperi@washington.edu].[SPID_GOnumber.txt] </pre> <p>DATASET PREVIEW Rows: 1 - 100 of 8887152 Columns: 2 / 2</p> <table border="1"> <thead> <tr> <th>SPID</th><th>GOID</th></tr> </thead> <tbody> <tr><td>AGA000</td><td>GO:0003824</td></tr> <tr><td>AGA000</td><td>GO:0003870</td></tr> <tr><td>AGA000</td><td>GO:0009058</td></tr> <tr><td>AGA000</td><td>GO:0016769</td></tr> <tr><td>AGA000</td><td>GO:0030170</td></tr> <tr><td>AGA000</td><td>GO:0033014</td></tr> <tr><td>AGA001</td><td>GO:0001666</td></tr> <tr><td>AGA001</td><td>GO:0005524</td></tr> </tbody> </table>	SPID	GOID	AGA000	GO:0003824	AGA000	GO:0003870	AGA000	GO:0009058	AGA000	GO:0016769	AGA000	GO:0030170	AGA000	GO:0033014	AGA001	GO:0001666	AGA001	GO:0005524	etc
SPID	GOID																		
AGA000	GO:0003824																		
AGA000	GO:0003870																		
AGA000	GO:0009058																		
AGA000	GO:0016769																		
AGA000	GO:0030170																		
AGA000	GO:0033014																		
AGA001	GO:0001666																		
AGA001	GO:0005524																		

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Biology

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science

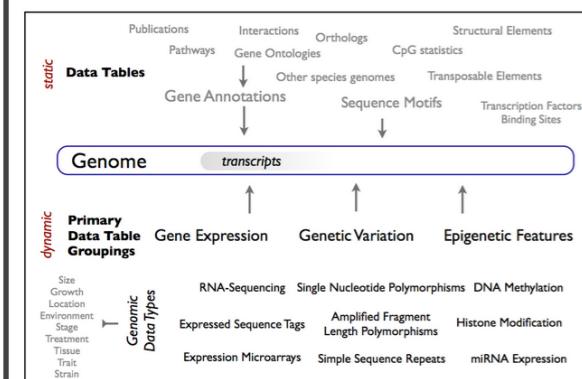
Data

g else...

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Data Snapshots

Select datasets available from SQLShare. Tag: [qdod2](#)
<https://sqlshare.esciencewashington.edu/sqlshare/#s=tag/qdod2>

Universal

The screenshot shows the 'Dataset' section for the 'Universal' dataset. It includes a 'Screenshot' preview of a table with protein names and IDs, and a 'Dataset Preview' table with 100 rows of data. The preview table has columns: SPID, Status, ProteinName, GenName, Organism, and Length. Several rows are listed, such as ASA163 reviewed, and ASA163 reviewed. The 'SPID and GO Number...' section is also visible.

Workflow 1: Annotating Oyster Genes

This workflow will take focus on taking a simple SQLShare table that has gene IDs and associated expression data and will take you through the steps of figuring out the name, function, etc of each gene.

Initial Data Table: Oyster larvae RNA-seq - OsHV exposure

SCREENSHOT

[solid0078_20091105_RobertsLab_GE_F3 trimmed RNA-Seq.txt](#) Viewable by everyone

OsHV RNA-seq on Version 9 transcriptome

[Click here to add a tag](#)

```

SELECT
    ["Feature ID"] as ID,
    ["Unique gene reads"] as UniqueReads,
    ["Total gene reads"] as TotalReads,
    ["RPKM"] as RPKM
  FROM [sr320@washington.edu].[table_solid0078_20091105_RobertsLab_GE_F3 trim]

```

DATASET PREVIEW Rows 1 - 100 of 28027 | Columns 4 of 4

ID	UniqueReads	TotalReads	RPKM
CGI_10000001	0	10	5.23
CGI_10000002	5	5	2.756
CGI_10000003	0	0	0
CGI_10000004	0	0	0
CGI_10000005	0	0	0
CGI_10000006	0	0	0





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plant science



Discovery Environment



Data



Apps



Analyses

Data Analysis

Discovery Environment

Data

Upload New Refresh Download Edit Share Trash

Navigation

- sr320
- labshare
- qdod
- BiGo_larvae
- Cgigas_v9
 - TopHat2-PE_BiGill_Bi...
 - Cuffdiff2-analysis1-2
 - Cuffmerge2_analysis
 - Cufflinks2_analysis1
 - TopHat2-SE_analysis
 - Uncompress_files_wi...
 - logs
- bamout
- bisque_data
- iplant_ws
- coge_data
- OlyO_PacBio
- cacheServiceTempDir
- OlympiaOyster
- fish546
- analyses
- AltSplice
- OlyO_SNPhunt
- Black_Abalone
- ce
- Community Data
- Shared With Me
- Trash

Details

Select a file or folder to view its details

Apps

Create Use App Request Tool Edit Submit Search Apps

Categories

- Workspace (8)
 - Apps under development (6)
 - Favorite Apps (2)
 - My public apps (0)
- Public Apps (434)
 - Archive (27)
 - BEDTools 2.15.0 (22)
 - Beta (66)
 - BLAST (2)
 - Data Sources (2)
 - Experimental (28)
 - Functional Analysis (7)
 - Functional RNA Prediction (5)
 - General Utilities (73)
 - Compress and Decompress
 - Conversion (1)
 - EMBOSS (50)
 - Text and Tabular Data (16)
 - List Analysis (1)
 - Metagenomics (6)
 - microRNA Analysis (3)
 - Molecular Structure (1)
 - Motif Identification (2)
 - NGS (111)
 - Aligners (16)
 - Assemblers (17)
 - Assembly Annotation (9)
 - Bisulfite (3)
 - ChIPseq (4)
 - QC and Processing (21)
 - SAMTools (6)
 - Transcriptome Profiling (11)
 - Misc RNASeq Tools (2)
 - Tuxedo RNA-Seq 1 (4)
 - Tuxedo RNA-Seq 2 (5)
 - Utilities (16)
 - Variant Identification (8)

Analyses

Delete Refresh Cancel View Parameters Relaunch

Name	App	Start Date	End Date	Status
genomeCoverageBed_Bi...	genomeCov...	2014 Feb 15 05:01...	2014 Feb 15 05:11...	Completed
genomeCoverageBed_Bi...	genomeCov...	2014 Feb 15 04:14...	2014 Feb 15 04:18...	Completed
RNAseq2bedgraph_anal...	RNAseq2be...	2014 Feb 15 04:07...	2014 Feb 15 09:55...	Completed
RNAseq2bedgraph SE_BB3	RNAseq2be...	2014 Feb 12 14:49...	2014 Feb 12 14:49...	Failed
RNAseq2bedgraph_BiGo...	RNAseq2be...	2014 Feb 12 14:36...	2014 Feb 12 20:00...	Completed
genomeCoverageBed_a...	genomeCov...	2014 Feb 12 11:22...	2014 Feb 12 11:28...	Completed
bamToBed_analysis1	bamToBed	2014 Feb 12 07:50...	2014 Feb 12 07:54...	Completed
VCF to GFF3_analysis1	VCF to GFF3	2014 Feb 11 06:58...	2014 Feb 11 06:59...	Completed
Find SNPs - mpileup_analysis1	Find SNPs - mpileup	2014 Feb 11 04:13...	2014 Feb 11 06:37...	Completed
TopHat2-PE_analysis1	TopHat2-PE	2014 Feb 10 17:23...	2014 Feb 10 21:38...	Completed

Data Analysis

The screenshot displays the Discovery Environment interface, divided into two main sections: Data and Apps.

Data Section:

- Navigation:** A tree view of user folders:
 - sr320
 - labshare
 - qdod
 - BIGo_larvae
 - Cgigas_v9
 - TopHat2-PE_BiGill_Bi
 - Cuffdiff2_analysis1-2
 - Cuffmerge2_analysis
 - Cufflinks2_analysis1-
 - TopHat2-SE_analysis
 - bam
 - logs
 - U1_out
 - T2_out
 - T1_out
 - U2_out
 - Uncompress_files_wi
 - logs
 - bamout
 - bisque_data
 - iplant_ws
 - coge_data
 - OlyO_PacBio

Details: A table showing file and folder details. The first row is highlighted in yellow. The columns are: Name, Last Modified, Size, and Details. The 'Details' column shows a placeholder message: "Select a file or folder to view its details".

Name	Last M...	...	Details
condor-0-input...	2014 Fe...	0...	Select a file or folder to view its details
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-0-input...	2014 Fe...	3...	
condor-0-input...	2014 Fe...	0...	
condor-stderr-0	2014 Fe...	2...	
condor-stdout-0	2014 Fe...	1...	
iplant.cmd	2014 Fe...	6...	
iplant.sh	2014 Fe...	5...	
output-last-stderr	2014 Fe...	0...	
output-last-std...	2014 Fe...	6...	
script-condor-log	2014 Fe...	3...	

Apps Section:

 - Categories:** A list of categories: Workspace (8), Apps under development (6), Favorite Apps (2), My public apps (0), Public Apps (434), and Archive (27).
 - Workspace:** A table showing integrated applications and their details. One entry is highlighted in yellow.

Name	Integrated by	Rating
Uncompress files wi...	Matthew Vaughn	★★★★★
RNAseq2bedgraph	Steven Roberts	★★★★★
RNAseq2bedgraph SE	Steven Roberts	★★★★★
FastQC 0.10.1 (mul...	Matthew Vaughn	★★★★★
 - RNAseq2bedgraph:** A detailed configuration panel for the RNAseq2bedgraph application. It includes fields for Analysis Name, TopHat2-PE for workflows - README, TopHat2-PE for workflows - Input data, TopHat2-PE for workflows - Reference Genome (Mandatory), and a note about selecting a reference genome. It also includes a dropdown menu for choosing a reference genome from a list and a note for non-matching species.
 - TopHat2-PE for workflows - Reference Annotations:** A section for providing a reference genome file in FASTA format, with a "Select a file" input field and a "Browse" button.

Username:
sr320
Password:

Log In Enter As Guest
Forgot Password? Register



The screenshot shows the iPlant Galaxy interface for selecting a Docker image:

- Select an Image:** Search bar: "Search by name, emi, tag, d".
- Featured Images:**
 - iPlant Centos 6.5 NoGUI Base (selected)
 - Ubuntu 12.04 - iPlant Base
 - NGS Viewers v3.2 (03-15-2013)
 - IBP Workbench v5
- My Projected Resource Usage:** 0% (You are using 0 of 168 allotted AUs), 6% (You will use 1 of 16 allotted CPUs), 3% (You will use 4 of 128 allotted GBs).
- Configuration:** First, select an image on the left. Then you can rename it, change its size, or tag it.
- Instance Size:** Name: Description: tiny1 (1 CPUs, 4 GB memory, 30 GB disk) (selected), tiny2 (1 CPUs, 8 GB memory, 60 GB disk), small1 (2 CPUs, 8 GB memory, 60 GB disk), small2 (2 CPUs, 16 GB memory, 120 GB disk), medium1 (4 CPUs, 8 GB memory, 80 GB disk), medium2 (4 CPUs, 16 GB memory, 160 GB disk), medium3 (4 CPUs, 32 GB memory, 240 GB disk), large1 (8 CPUs, 16 GB memory, 160 GB disk) (selected), large2 (8 CPUs, 48 GB memory, 320 GB disk), large3 (8 CPUs, 64 GB memory, 480 GB disk), large4 (16 CPUs, 96 GB memory, 400 GB disk).

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everything else...

Galaxy

Analyze Data Workflow Shared Data Visualization Cloud Help User Using 13%

Tools Load Data

search tools

[Get Data](#)

[Send Data](#)

[Lift-Over](#)

[Text Manipulation](#)

[Convert Formats](#)

[FASTA manipulation](#)

[Filter and Sort](#)

[Join, Subtract and Group](#)

[Extract Features](#)

[Fetch Sequences](#)

Extract Genomic DNA using coordinates from assembled/unassembled genomes

[Fetch Alignments](#)

[Get Genomic Scores](#)

[Operate on Genomic Intervals](#)

[Statistics](#)

[Graph/Display Data](#)

[Regional Variation](#)

[Multiple regression](#)

[Multivariate Analysis](#)

[Evolution](#)

[Motif Tools](#)

[Multiple Alignments](#)

[Metagenomic analyses](#)

[Genome Diversity](#)

[NGS TOOLBOX BETA](#)

[Phenotype Association](#)

[NGS: QC and manipulation](#)

[NGS: Mapping](#)

Bowtie2 is a short-read aligner

Lastz map short reads against reference sequence

Map with Bowtie for SOLID

Map with Bowtie for SOLID (version 1.1.2)

Will you select a reference genome from your history or use a built-in index?:
Use a built-in index

Built-ins were indexed using default options

Select the reference genome:
Arabidopsis lyrata: Araly1

If your genome of interest is not listed – contact Galaxy team

Is this library mate-paired?:
Single-end

FASTQ file:
Must have Sanger-scaled quality values with ASCII offset 33

Bowtie settings to use:
Commonly used

For most mapping needs use Commonly used settings. If you want full control use Full parameter list

Suppress the header in the output SAM file:
 Bowtie produces SAM with several lines of header information by default

Execute

What it does

Bowtie is a short read aligner designed to be ultrafast and memory-efficient. It is developed by Ben Langmead and Cole Trapnell. Please cite: Langmead B, Trapnell C, Pop M, Salzberg SL. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. Genome Biology 10:R25.

Know what you are doing

⚠ There is no such thing (yet) as an automated gearshift in short read mapping. It is all like stick-shift driving in San Francisco. In other words = running this tool with default parameters will probably not give you meaningful results. A way to deal with this is to understand the parameters by carefully reading the documentation and experimenting. Fortunately, Galaxy makes experimenting easy.

Input formats

Bowtie accepts files in Sanger FASTQ format. Use the FASTQ Groomer to prepare your files.

History

Ensembl
735.8 MB

3: Crassostrea gigas.GC
A 000297895.1.21.gff3

2: Crassostrea gigas.GC
A 000297895.1.21.gtf

~420,000 lines
format: gtf, database: cgigas_E_sm

uploaded gtf file

1: Seqname Z: Source 3: Feature

Seqname	Z: Source	3: Feature
scaffold39040	protein_coding	exon
scaffold39040	protein_coding	CDS
scaffold39040	protein_coding	start_cod
scaffold39040	protein_coding	exon
scaffold39040	protein_coding	CDS
scaffold39040	protein_coding	exon

1: Crassostrea gigas.GC
A 000297895.1.21.dna_sm.genome.f

a

7,658 sequences
format: fasta, database: cgigas_alpha_v012

uploaded fasta file

>C12722 dna_sm:supercontig supercontig
TTTTCTTATTTTATTGTACAGTTAACCTGTGAG
ccgtttttagtctttctgTTGACTTTATTATTCTC
TATGAATTATTATGTGTATCTTCATGGGATATTTC
ACACTTGCTTTCGGTGTAT
>C12722 dna_sm:supercontig supercontig

Galaxy Analyze Data Workflow Shared Data Visualization Cloud Help User Using 13%

Published Workflows | rna-seq-helin-group | RNA-seq differential analysis replicates)

Galaxy Workflow 'RNA-seq differential analysis replicates'

Annotation: Workflow based on Tophat and cuffdiff. Inputs: 4 fastq files (ex. Outputs: bam, bigwig, xls, and html reports with quality controls.

Step

Step 1: Input dataset
Gene Annotations
select at runtime

Step 2: Input dataset
Group1 R1
select at runtime

Step 3: Input dataset
Group1 R2
select at runtime

Step 4: Input dataset
Group2 R1
select at runtime

Step 5: Input dataset
Group2 R2
select at runtime

Step 6: Input dataset
Pseudo reads BAM file
select at runtime

Step 7: Tophat for Illumina
RNA-Seq FASTQ file
Output dataset 'output' from step 2
Use a built in reference genome or own from your history
Use a built in genome
<https://usegalaxy.org/library/index>

Annotation

Data Lib Access published resources (2 conditions, 2 replicates) + About this Workflow

Data Libraries Beta

Published Histories Published Workflows Published Visualizations Published Pages

reads, 2 conditions, 2
reads, 1 gtf file (annotations).

Author
rna-seq-helin-group

Related Workflows

All published workflows Published workflows by rna-seq-helin-group

Rating

Community (3 ratings, 5.0 average) ★★★★★

Yours ★★★★★

Tags

Community: illumina cuffdiff rnaseq tophat

Yours:

Biology

Environment

Molecular

Data Analysis

eScience

iPlant Galaxy

Notebooks

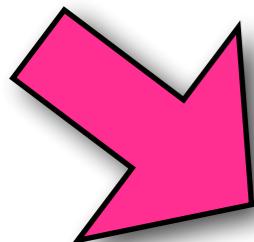
Rationale

Platforms

Open Science

Data

everything else...



SQLSHARE

fGalaxy

**iPlant
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Hyak



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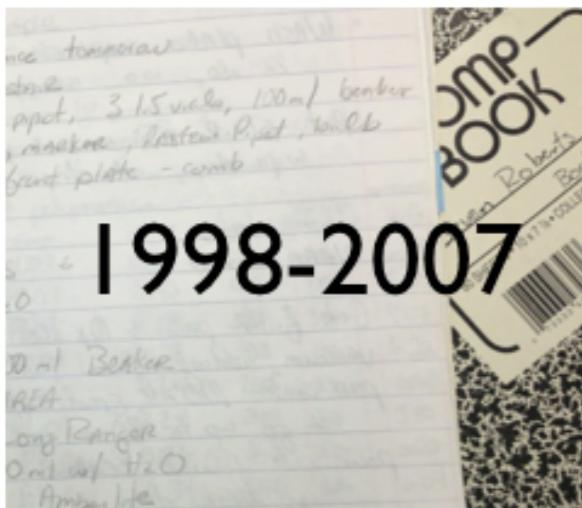
everything else...



iPlant

		TUESDAY, FEB 11 2014	Z05.0 MID
■	BiGoRNA_genomecov.bedgraph	Feb 13, 2014, 5:47 PM	278.2 MB
■	BiGo_egg_redo4.sam	Feb 13, 2014, 5:26 PM	98.4 MB
■	BiGo_egg_redo3.sam	Feb 13, 2014, 1:18 PM	1.46 GB
■	solid0078_20091105_BB3 mapping.bam	Feb 13, 2014, 1:15 PM	4.99 GB
■	solid0078_20091105_BB3 mapping.sam	Feb 13, 2014, 1:14 PM	706.3 MB
■	solid0078_20091105_BB3 RNA-Seq.bam	Feb 13, 2014, 1:13 PM	2.65 GB
■	solid0078_20091105_BB3 RNA-Seq.sam	Feb 12, 2014, 3:31 PM	269.8 MB
■	BiGoRNA_bg1_sorted.bedgraph	Feb 12, 2014, 3:26 PM	117 KB
■	cgigasgenome3.txt	Feb 12, 2014, 11:49 AM	4.46 GB
■	solid0078_20091105_BB3.1.fastq	Feb 12, 2014, 9:43 AM	1.6 MB
■	BiGoRNA_GTGTCTAC_1.sorted.bam.bai	Feb 12, 2014, 9:31 AM	269.8 MB
■	BiGoRNA_bg1.bedgraph	Feb 12, 2014, 8:33 AM	117 KB
■	cgigasgenome3.tab	Feb 12, 2014, 8:31 AM	558.4 MB
■	cgigasgenome2.tab	Feb 12, 2014, 8:31 AM	558.3 MB
■	cgigasgenome1.tab	Feb 12, 2014, 8:29 AM	1.42 GB
■	BiGoRNA_GTGTCTAC_1.sorted.bam	Feb 12, 2014, 8:28 AM	2 KB
■	igv_session_cgigas_ensembl_1.xml	Feb 12, 2014, 8:16 AM	1.42 GB
■	BiGoRNA_GTGTCTAC_1.bam	Feb 11, 2014, 8:52 AM	Zero bytes
■	filtered_106A_Female_Mix_GATCAG_L004_R1.bam.fai	Feb 11, 2014, 4:35 AM	9.1 MB
■	filtered_106A_Female_Mix_GATCAG_L004_R1.bam.bai	Feb 11, 2014, 4:24 AM	8.58 GB
■	filtered_106A_Female_Mix_GATCAG_L004_R1.bam	Feb 10, 2014, 1:32 PM	23.98 GB
■	fish546_of_BowtieOut.sam	Feb 10, 2014, 12:56 PM	3.7 MB
■	fish546_of_BowtieOut.sorted.sam	Feb 10, 2014, 12:53 PM	4 bytes
■	fish546_of_BowtieOut.sorted.sam.sai	Feb 10, 2014, 12:37 PM	4.3 MB
■	fish546_14_Trinity.fasta.fai	Feb 10, 2014, 12:02 PM	88.1 MB
■	fishfe6_oCD-HITout.fa	Feb 10, 2014, 6:19 AM	112.3 MB
■	Emma_Defense_Talk.mp3	Feb 9, 2014, 12:53 PM	163.3 MB
■	fish546_14_Trinity_ls_length2.tab	Feb 9, 2014, 12:26 PM	162.9 MB
■	fish546_14_Trinity_ls_length.tab	Feb 9, 2014, 12:16 PM	162.8 MB
■	fish546_14_Trinity.tab	Feb 9, 2014, 10:25 AM	121 bytes
■	data.csv	Feb 9, 2014, 10:24 AM	165.1 MB
■	fish546_14_Trinity.fasta	Feb 9, 2014, 2:00 PM	112.1 MB
■	282014Emma_Defense_Talk.m4a	Feb 8, 2014, 10:46 AM	7 KB
■	f546_bubREVIGO.r	Feb 8, 2014, 10:45 AM	6 KB
■	f546REVIGO_treemap.r	Feb 8, 2014, 10:35 AM	30 KB
■	f546REVIGO.xqmmi		

The Evolution of My Lab Notebook



1998-2007

A screenshot of a software interface designed to look like a handwritten lab notebook. It shows a timestamp of 1/23/2009 at 10:43 AM. The main area contains handwritten text about a fish assembly and sequencing. A sidebar on the left lists '103009' and '103009_H'.

2008-2009

A screenshot of a digital lab notebook interface. It features a dashboard with various tabs: TIGRA, Oly annotation, Oly annotation accuracy, Oly collection map, Pois Al Dente analysis, Mutation Genome Wide Comparison, Partition Assembly tracks for v200, and Genomics Gateway. There are also sections for H3K27ac notes and Journals.

2010-2013

A screenshot of a Jupyter Notebook interface titled 'IP(y) Notebook BiGo_methratio'. It shows a table of methylation data for CpG sites across five samples (C17314). Below the table are two code snippets: one for reading data from a CSV file into a pandas DataFrame and another for creating a histogram of CDS values.

now

Open Notebook Science

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Data

... there is a URL to a laboratory notebook that is freely available and indexed on common search engines. It does not necessarily have to look like a paper notebook but it is essential that all of the information available to the researchers to make their conclusions is equally available to the rest of the world.

—Jean-Claude Bradley

everything else...

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xtra Credit

paper02 Draft

All Reactions

Edit 2 2



List of all experiments

NOTE: All solubility experiments by the Bradley group past this point are recorded on the [challenge wiki](#).

[EXP286](#) Synthesis of aldol condensation product of phenanthrene-9-carboxaldehyde and NaOH as the catalyst. -Matthew McBride

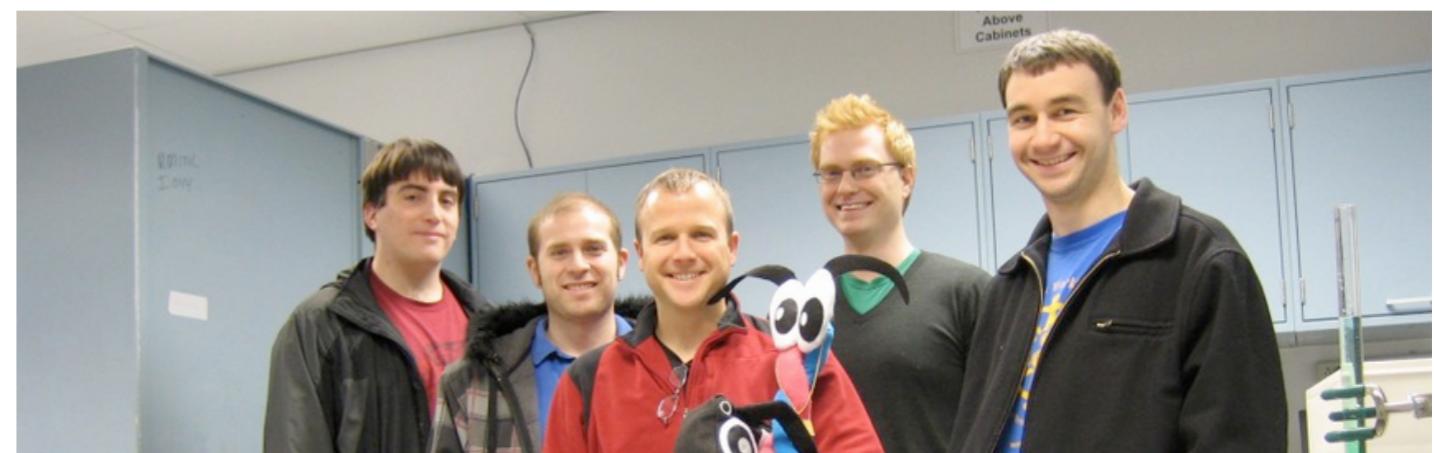
[EXP285](#) Synthesis of trans-dibenzalacetone in ethanol/water (1:1 vol) using NaOH as the catalyst. -Matthew McBride

[EXP284](#) Synthesis of trans-dibenzalacetone in ethanol/water (1:1 vol) using NaOH as the catalyst. -Matthew McBride

IheartAnthony's Research

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[Repeating Crumley](#)

[Experiment Introduction:](#) Steve asked me to try and repeat an experiment by

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- Extra Credit
- paper03 Draft

All Reactions



List of all experiments

NOTE: All solubility experiments by the Bradley group past this point are recorded on the challenge wiki.

[EXP286](#) Synthesis of aldol condensation product of phenanthrene-9-carboxaldehyde and NaOH as the catalyst. -Matthew McBride

[EXP285](#) Synthesis of trans-dibenzalacetone in ethanol/water (1:1 vol) using NaOH as the catalyst. -Matthew McBride

[EXP284](#) Synthesis of trans-dibenzalacetone in ethanol/water (1:1 vol) using NaOH as the catalyst. -Matthew McBride

IheartAnthony's Research

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Optical tweezers pages

Active Experiments

Repeating Crumley

Experiment Introduction: Steve asked me to try and repeat an experiment by

Open Notebook Science

Lab Notebook

(Introduction)

Entries

Notes

31 Jan 2014

pageviews: (not calculated)

Scratch notes on uncertainty.

Function definition: [multiple_uncertainty.m](#)

Settings:

```
f=@(x, h) max( (x-h) * (1 - (x-h) ./ 100)
+ (x-h), 0);
x_grid = [0:5:100];
h_grid = x_grid; % Must be same dimension
% as x_grid, or L91 errors...
Tmax = 5;
sigma_q = 0.1;
```

[Read more](#)

Notes

09 Jan 2014

pageviews: 8

- fishbase API discussions continue
- some discussion / work on rfigshare continues (As Karthik writes comprehensive test suite, yay).
- Possible ontological term for simulated data? modeling and simulation operation looking for example using this term now...
- Request for term on OBI
- Moore DDD investigators, full call

@NOAA gives a glimpse of what the future of
[Read more](#)

Carl Boettiger

Theoretical Ecology and Evolution

I'm Carl Boettiger, a post-doctoral researcher with Marc Mangel and Steve Munch in the Center for Stock Assessment Research, at UC Santa Cruz, working on regime shifts in ecology and evolution. Such shifts mark the most dramatic events in complex systems. What causes these transitions to occur? Can we detect or forecast these changes? How do we robustly manage ecosystems that experience such tipping points? Understanding and forecasting these events challenges the capacity of both our statistical methods and the data available. My research aims to extend and quantify the limits of understanding through nonparametric Bayesian inference, high performance computing and big data



Notes

07 Jan 2014

pageviews: 6

Coding

- fishbase API discussions
- rfishbase updates pushed to CRAN
- Merges and minor bug fixes for rfigshare
- RNeXML: After some cooperative sleuth-work, we successfully resolved issue #23, uncovering a bug (missing feature really) in xmllint.

as usual, detailed activity on github.

[Read more](#)

Notes

06 Jan 2014

pageviews: 3

- fishbase API discussions
- knitcitations (push previous updates to CRAN)
- data extraction from sardine collapse story (see EML in data/), news article

Open Science Literature Highlights

02 Jan 2014

pageviews: 40

A student recently asked me for some recommendations for an article on an open science topic for a journal club. Since I haven't jotted these down in one place before, though I might copy my reply here for

Notes

10 Dec 2013

pageviews: 64

Consider the model

$$dX_t = \alpha(\theta - X_t)dt + \sigma dB_t \quad (1)$$

carlboettiger.info/lab-notebook

Open Notebook Science



genefish.wikispaces.com

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[Steven's Notebook](#)

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This wiki has been developed as a resource for lab personnel and students to access information and publish research activities using an [open notebook science](#) based system. All lab notebooks can be accessed via the side menu. The Roberts Lab is in the [School of Aquatic and Fishery Sciences](#) within the [College of Environment](#) at the [University of Washington](#). More information can be found concerning [research](#), [personnel](#), and [outreach](#) on the [Roberts Lab Official Webpage](#).

Laboratory Reference Material

- [Data and Resource Sharing Plan](#)
- [Laboratory Protocols](#)
- [Emergency Contact Information](#)
- [UW Lab Safety Manual](#)
- [Code Repository](#)
- [UW Biosafety Manual](#)
- [Chemical Inventory](#)

Lab Activity and Communication

- [Lab Meetings](#)
- [IPUS: Information for Prospective Undergraduate Students](#)
- [Lab Calendar](#)

Data Repositories

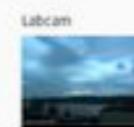
- [The Eagle](#)
- [CLC Genomics Server](#) (password protected)
- [Primer Database](#)
- [NGS Library Info](#)
- [crassostreome](#)
- [UW HTSEQ](#) (gycalMS)



Research
Research in our lab focuses on characterizing physiological responses of marine organisms to environmental change. Using integrative approaches we strive to examine impacts and adaptive potential from the nucleotide to organismal level. A core component of this includes investigating the functional relationship of genetics, epigenetics, and transcription.

Outreach
We are continually looking for new ways to engage and interact with students, scientists, and the general public. Everyone can follow us on tumblr, facebook, youtube, twitter, wikispace, flickr, and figshare. Web-based resources we have developed include an online version of the Galapagos Shellfish Collection, crassostreome, Research Notes on Ocean Acidification and oysters genes.

News and Notes
- Congrats to MacKenzie Gavry, who won a Student Spotlight Award at Aquaculture 2013
- "Papers" page rebranded as "Products".
- Claire Ellis launches her website - check it out! (and is now on twitter!)
- Archive



Labcam
Connect:
@n320
206.681.3342

Access

Funding

Roberts Lab Website

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Katie's Notebook

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0

75

February 4, 2014

This weekend, I downloaded Colony onto Thomas's computer (since it is Windows). I have been running the 2011 data on it since Saturday afternoon. As of this morning, it was making some pretty good progress. I will post the results on here when they are done! Tonight, I am going to download "HP-Rare" onto his computer so I can look at the rarefaction curve of this data. I am also working on organizing the south sound data from David Stick so that I can analyze that and compare it to the data I am working on now.

I made a Public Folder in my dropbox where I will keep all of the Microsat data so that there is a live file anyone can look at. Here is the link:

https://www.dropbox.com/sh/5839mey4mgn2y28/cydXmRe_zd

So far, there are the allele frequencies from David Stick's south sound wild Oly's and another file which will have the GenAIEx relatedness values.

I am working on Excel with this data and doing the same things that I have already done to the restoration grade data. (I will also put this through Colony when I get the other files through).

I also started running the 2010 restoration grade oly data through Colony downstairs in Jake's office.

January 30, 2014

In FSH 207 running Colony. I *think* I am now about 30% of the way through with the 2011 data.



Oly.MidResult5

[Details](#) [Download](#) 1 KB

January 29, 2014

Open Notebook Science

Search

che625's notebook

Combining technical replicates for methylkit
Yesterday Female correlation file.list <-
list('/Volumes/web/Mollusk/bs_larvae_exp/methylkit_ files/round2/CgF_nov.txt','/Volumes/web/Mollusk/bs_

methylkit troubleshooting
Yesterday Used Giles' code for formatting my files for methyl kit. Also used the R code listed below but kept getting errors. Will keep troubleshooting and try re-formatting my files. Used the files listed in

Query for using methylkit
Yesterday SELECT chr as chr, pos as start, '+' as strand, cast (CT_count as float) as CT_count, cast (C_count as float) as C_count, cast (C_count as float) / cast (CT_count as float) as

methyl kit with larvae data
Yesterday library(methylKit) library(data.table)
library(GenomicRanges) file.list <-
list('/Volumes/web/Mollusk/bs_larvae_exp/methylkit_

Filtering Larvae data
6 days ago Ran all files through BSMAP and methratio. Now need to filter so context= CG and a coverage cutoff of 5x or greater. Here is the query I am using to filter the data, and am changing the file

Bioinformatics 2014- Module 6
6 days ago See Module 1 for De novo assembly with Trinity using oyster gonad RNA-Seq data:
<https://www.evernote.com/shard/s232/sh/0596b21b->

Untitled Note
Last week For next figure look at this file:
/Volumes/web/cnidarian/BiGo_gene_pmeth_expression.csv
Porzio<- read.csv("/Users/claireolson/Desktop/Porzio\ et al 2011\

Beautiful graphics in R- experimenting with plotly
Last week # Required packages from CRAN .pkgs =
c("RJSONIO", "RCurl", "devtools") # Install required packages from CRAN (if not) .inst <- .pkgs %in% installed.packages()

Figure 1 for beautiful graphics in R
Last week With labels: Exon<-
read.table("/Volumes/web/Mollusk/174gm_analysis/
Whole_Genome_Analyses/intersectBedfiles_Steven

Bioinformatics 2014- Module 4
Last week My file of p-values for all genes in gill v gonad: http://eagle.fish.washington.edu/Mollusk/FIS_H546_2014/DESeq%20results%20for%20Tissue%2

che625's notebook

Created: Feb 18, 2014 Modified: Feb 18, 2014

Larvae T1D5 correlation

```
file.list <- list('/Volumes/web/Mollusk/bs_larvae_exp/methylkit_files/round2/CgLarv_T1D5_nov.txt', '/Volumes/web/Mollusk/bs_larvae_exp/methylkit_files/round2/CgLarv_T1D5_sept.txt')  
  
myobj<-read(file.list,pipeline=list(fraction=TRUE,header=T, chr.col=1,start.col=2,end.col=3,strand.col=4, coverage.col=freqT.col=7),  
          sample.id=list("T1D5_nov", "T1D5_sept"),assembly="v9",treatment=c(1,0))  
  
meth<-unite(myobj,destrand=FALSE)  
  
head(meth)  
  
nrow(meth)  
  
getCorrelation(meth,plot=T)
```

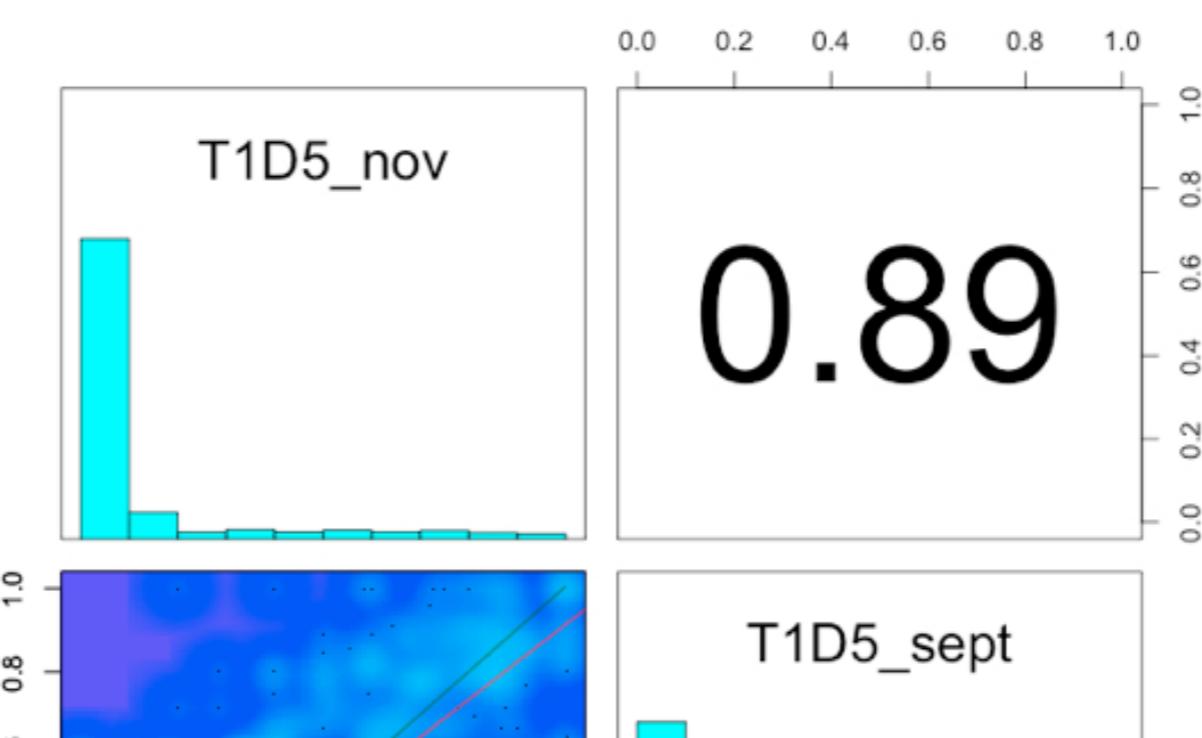
2373 loci make up this plot:

CpG base pearson cor.

T1D5_nov

0.89

T1D5_sept



Open Notebook Science

IP[y]: Notebook

BiGo_methratio Last Checkpoint: Sep 13 09:53 (autosaved)

File Edit View Insert Cell Kernel Help



Code

Cell Toolbar:

None

DNA methylation of Oyster Sperm based on Genomic Features

methratio file in SQLShare https://sqlshare.escience.washington.edu/sqlshare#s=query/sr320%2540washington.edu/clean_BiGo_methratio_v1

BiGO methratio GFF

```
SELECT
    chr as seqname,
    'methratio' as source,
    'CpG' as feature,
    pos as start,
    pos + 1 as [end],
    ratio as score,
    strand,
    '.' as frame,
    '.' as attribute
FROM [sr320@washington.edu].[clean_BiGo_methratio_v1]
where
context like '__CG_'
and
CT_Count >= 5
```

https://sqlshare.escience.washington.edu/sqlshare#s=query/sr320%2540washington.edu/BiGo_methratio_GFF_boop&q=

```
In [13]: # ie python fetchdata.py -d "[sr320@washington.edu].[BiGO_Methylation_oysterv9_GFF]*" -f tsv -o /Volumes/web/cnidarian/BiGo_methratio_GFF_boop.tsv
# running on commandline because cannot get to work in IPython
```

```
python fetchdata.py -d "[sr320@washington.edu].[BiGo_methratio_GFF_boop]*" -f tsv -o /Volumes/web/cnidarian/BiGo_methratio_boop.gff
```

```
In [15]: #fetchdata failed
```

Open Notebook Science



Automating a Workflow: Beyond Blast - to GO Slim

The concept is that you can take a fasta file in a working directory and end up with GO slim information all within a single notebook that is automated. Currently this work by writing (and overwriting) a scratch file to SQLShare. Assumptions are that you are working in a directory with fasta file named `query.fa`. And blast algorithms are in PATH.

```
In [13]: #allows plots to be shown inline
%pylab inline

Populating the interactive namespace from numpy and matplotlib

In [4]: #Setting Working Directory
wd="/Volumes/web/whale/fish546/qpx_go_val"
#Setting directory of Blast Databases
dbd="/Volumes/Bay3/Software/ncbi-blast-2.2.29+/db/"
#Database name
dbn="uniprot_sprot_r2013_12"
#Blast algorithm
ba="blastx"
#Location of SQLShare python tools: you can empty ("") if tools are in PATH
spd="/Users/sr320/sqlshare-pythonclient/tools/"

In [5]: cd {wd}
/Volumes/web/whale/fish546/qpx_go_val

In [5]: !{ba} -query query.fa -db {dbd}{dbn} -out {dbn}_{ba}_out.tab -evalue 1E-50 -num_threads 4 -max_hsps_per_subject 1
BLAST Database error: No alias or index file found for protein database [/Volumes/Bay3/Software/ncbi-blast-2.2.29+
/db/uniprot_sprot_r2013_12] in search path [/Volumes/web/whale/fish546/pipeline_test_dir4::]

In [6]: !head -1 {dbn}_{ba}_out.tab
QPX_transcriptome_v1_Contig_2    sp|P52712|CBPX_ORYSJ    43.75   416    213    12    2095    869    6    40
7      3e-98    326

In [17]: #Translate pipes to tab so SPID is in separate column for Joining
!tr '|'"\t" <{dbn}_{ba}_out.tab> {dbn}_{ba}_out2.tab

In [18]: !head -1 {dbn}_{ba}_out2.tab

In [8]: #Uploads formatted blast table to SQLshare; currently has generic name and meant to be temporary: Warning will over
!python {spd}singleupload.py -d scratchblast_out {dbn}_{ba}_out2.tab
...

In [9]: !python {spd}fetchdata.py -s "SELECT * FROM [sr320@washington.edu].[scratchblast_out]blast Left Join [sr320@washing

In [10]: !head -2 {dbn}_join2goslim.txt
...

In [11]: !python {spd}singleupload.py -d scratchjoin.slim {dbn}_join2goslim.txt
processing chunk line 0 to 18037 (0.0718240737915 s elapsed)
pushing uniprot_sprot_r2013_12_join2goslim.txt...
parsing 9A18D989...
finished scratchjoin.slim

In [12]: #Sets GO aspect
!python {spd}fetchdata.py -s "SELECT Distinct Column1 as query, Column3 as SPID, GOSlim_bin FROM [sr320@washin

In [13]: !head justslim.txt
...

In [15]: from pandas import *
```

Set some variables

blast

convert file format

upload to SQLShare
(python client)

join in SQLShare -
download

read in pandas

matplotlib generates
graph of GOslim

Open Notebook Science

https://github.com/sr320/ipython_nb

This repository has 206 commits, 2 branches, 0 releases, and 1 contributor. The master branch is selected.

Commit	Message	Date
sm		latest commit 717a24e7f6
sr320 authored 3 days ago		3 days ago
.ipynb_checkpoints	testing post	4 days ago
fish546	new tools	4 days ago
img	catching up with class	a month ago
tools	sm	3 days ago
.gitignore	en ignore	7 months ago
BiGill_Gene_Methylation.ipynb	sm	6 months ago
BiGill_RNAseq.ipynb	DMG annotations	2 months ago
BiGill_Tran_Elements.ipynb	Adding all ipynb files from cnidarian	7 months ago
BiGill_array.ipynb	DMG annotations	2 months ago
BiGo - methratio error.ipynb	Adding all ipynb files from cnidarian	7 months ago
BiGo_GFF_dev.ipynb	Adding all ipynb files from cnidarian	7 months ago
BiGo_RNAseq.ipynb	sm	a month ago
BiGo_larvae.ipynb	DMG annotations	2 months ago
BiGo_larvae_2.ipynb	bsmap on egg QC	4 days ago

Biology

Environment

Molecular

Data Analysis

eScience

iPlant Galaxy

Notebooks

Rationale

Platforms

Open Science

Data

everything else...

Open Notebook Science

The screenshot shows a GitHub repository page for 'sr320 / ipython_nb'. The top navigation bar includes links for 'Explore', 'Gist', 'Blog', and 'Help'. The repository stats show 206 commits, 2 branches, 0 releases, and 1 contributor. The 'branch: master' dropdown is set to 'ipython_nb'. The main area displays a list of files and their commit history:

File	Commit Message	Date
sm	sr320 authored 3 days ago	latest commit 717a24e7f6
.ipynb_checkpoints	testing post	4 days ago
fish546	new tools	4 days ago
img	catching up with class	a month ago
tools	sm	3 days ago
.gitignore	en ignore	7 months ago
BiGill_Gene_Methylation.ipynb	sm	6 months ago
BiGill_RNAseq.ipynb	DMG annotations	
BiGill_Tran_Elements.ipynb	Adding all ipynb files from cnidarian	
BiGill_array.ipynb	DMG annotations	
BiGo - methratio error.ipynb	Adding all ipynb files from cnidarian	
BiGo_GFF_dev.ipynb	Adding all ipynb files from cnidarian	
BiGo_RNAseq.ipynb	sm	
BiGo_larvae.ipynb	DMG annotations	
BiGo_larvae_2.ipynb	bsmap on egg QC	

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IPython Notebook

Viewer Open Science

A simple way to share your IPython Notebooks

Share your own notebook, and browse others'

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a very new experiment

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Open Notebook Science

a very new experiment

The screenshot shows a web-based application for managing research experiments. At the top right are links to "Biology", "Environment", "Molecular", "Data Analysis", "eScience", "iPlant Galaxy", "Notebooks", "Rationale", "Platforms", "Open Science", and "Data". On the left, a vertical timeline shows experiments posted on February 14 and 13. Each experiment entry includes a title, a timestamp ("Posted 2 days ago"), and tags ("nb, ipython"). The sidebar on the right contains links to "About", "Home", "Ask me anything", "Archive", and "The Details of my Lab Notebook". It also features a search bar and an RSS feed link.

FEB 14

qDOD RNAseq[0214_0646]
Posted 2 days ago
nb, ipython,

w6_denovo[0214_0643]
Posted 2 days ago
N nb, ipython,

TJGR_Eensembl[0214_0627]
Posted 2 days ago
nb, ipython,

FEB 13

Notes
Worked on BSMAP pipeline

About
This is an experiment by [Steven Roberts](#) to develop a single portal that automatically captures research activity in real time.
This includes

- [IPython Notebook snapshots](#)
- [iPlant DE alerts](#)
- [GitHub commits](#)
- and much more

Please [contact me](#) if you have any comments, suggestions, or questions.

[Home](#)
[Ask me anything](#)
[Archive](#)

[The Details of my Lab Notebook](#)

SEARCH RSS

Open Notebook Science

Biology

a very new experiment

Environment

Molecular

February 2014 81 posts

IFT_05_uniprot_sprot_r2013_12_join2goslim.txt
eagle.fish.washington.edu

notes
Running Ift_05 through fasta2slim pipeline

notes
looking at new assembly of all libraries- and new trim

notes
Only 1.7% map of BSgill to Ensembl v

notes
BiGII_CpG_Eensembl[0215_0710]
ift.tt

notes
BSMAP needs redo on new genome

notes
grep "[ATCG][ATCG]CG[ATCG]" <file>
When run on what you gave me it gives me
C12722 104 + TACGT
1.000 1.00 1 1 1
1 0.207 1.000
C12722 134 + TGGCG
1.000 1.00 1 1 1
1 0.207 1.000

github: sr320 pushed to master at sr320/ipython_nb
ift.tt

notes
Used some one-liners to help Hannah convert csv data- still mystery non-integer

w3_RNAseq[0214_1323]
ift.tt

notes
posting ip? —

iPlant DE Alert:
RNAseq2bedgraph_BiGoRNA

notes
Haliotis 26,246 7 2 87 Click on organism name to get more information.
• Haliotis australis 5,666 1 13
• Haliotis australis (Australian abalone)
• Haliotis corrugata
• Haliotis discus
• Haliotis corrugata (pink abalone)
• Haliotis cracherodii (black abalone)
• Haliotis discus 2,033 13
• Haliotis discus discus (disc abalone) 829 1
• Haliotis discus discus hamata (Japanese disc abalone) 2,787 3
• Haliotis diversicolor 2,291 13
• Haliotis diversicolor abalones
• Haliotis diversicolor diversicolor
• Haliotis diversicolor diversicolor superosa 208 6
• Haliotis dentifera
• Haliotis gigantea
• Haliotis laevigata 1 13
• Haliotis laevigata (green abalone)
• Haliotis gigantea
• Haliotis laevigata
• Haliotis jacchus
• Haliotis kamtschatkana 1 13
• Haliotis kamtschatkana kamtschatkana (banded abalone)
• Haliotis hirayamai (greenlip abalone)
• Haliotis microtis
• Haliotis midae (yellowlip abalone) 40 1 13

notes
Run v4 Black Ab assembly

BlackAb_Annot[0215_1334]
ift.tt

IFT_BlackAbalone_sp_go_path
eagle.fish.washington.edu

The Details of my Lab Notebook
I use IPython Notebook as my lab notebook, hosted on GitHub, with the working directories in Dropbox. I use one primary repository which currently has subdirectories for fish546 (class) and 'tools' which represent reusable workflows. I maintain a second repository for a former variant.

sr320 pushed to master at sr320/qdod
ift.tt

bb3 bam uploading, and bedgraph
Feb 15, 2014
ift.tt

http://ift.tt/1f0caCH
ift.tt

http://ift.tt/1f0c9Pf
ift.tt

notes
Trying RNAseq2Bedgraph with BiGoRNA

notes
Relook at Black Abalone transcriptome

BlackAb_Annot[0214_1530]
ift.tt

notes
Notes
Worked on BSMAP pipeline

notes
Worked on automating IPython snapshots.

qDOD RNAseq[0214_0646]
ift.tt

github: sr320 pushed to master at sr320/ipython_nb
ift.tt

BiGo_larvae_2.html
ift.tt

w6_denovo[0214_0643]
ift.tt

iPlant DE Alert:
bamToBed_analysis1 status

CC_ampk.html
ift.tt

iPlant DE Alert:
RNAseq2bedgraph_BiGoRNA

iPlant DE Alert:
RNAseq2bedgraph_SE_BB3

iPlant DE Alert:
genomeCoverageBed_analys

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sr320.info

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about science

Open Science

about science
web-native scholarship

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Sharing



Photo credit: Flickr, Creative Commons, speechless

Example



Public Sigenae Contig Browser Oyster

Search e! Oyster: Anything Go

e.g. [BQ426804.p.cg.8](#), [snp_EW777925_683](#)

Based on Ensembl release 40 - Aug 2006

Use Ensembl to...

- Search Contig Browser
- Data mining [BioMart]
- Export data
- Download data
- Digital Differential Display
- Venn Diagrams

Docs and downloads

- About Contig Browser
- Contig Browser data

Other links

- Home
- Sigenae
- Agenae



Browse the *Crassostrea gigas* contigs database

About the *Crassostrea gigas* contigs database



Assembly

The Oyster EST contig browser aims to produce and maintain an automatic annotation of Oyster EST libraries. This database [GigasDatabase](#) was initiated within the frame of the [AquaFirst](#) European project, it now gathers EST sequences produced by a [Marine Genomics Europe](#) project (GOCE-CT-2004-505403) and a [Genoscope project](#). [GigasDatabase](#) is regularly updated in the context of the ANR project "Gametogenes" (ANR-08-GENM-041).

Annotation

Contigs were annotated searching sequence homologies against following databases:

- ▶ UniProtKB/Swiss-Prot Release 2011_03 of 08-Mar-2011
- ▶ RefSeq Protein Index Blast of 09-Jan-2011
- ▶ Pfam Release 24.0 of Jul-2009
- ▶ Sigenae Oyster Contigs V8
- ▶ RefSeq RNA Index Blast of 09-Jan-2011
- ▶ TIGR Fugu FGI 3.0

Statistics

Assembly: pcg8, Mar 2011

Number of base pairs (Contigs): 67,875,621

Number of sequences (EST/mRNA): 1,013,570

Total number of contigs: 82,312

Number of singlets: 20,148

Polymorphism: Mar 2011

Number of putative SNPs: 29,493

Number of contigs including SNP: 10,392

Example Data Points

Jump directly to the example entry points:

Contig: [BQ426804.p.cg.8](#)

SNP: [snp_EW777925_683](#)

Example

S!  **Public Sigenae Contig Browser Oyster** Search e| Oyster: Anything Go e.g. [BQ426804.p.cg.8](#), [snp_EW777925_683](#)

Based on Ensembl release 40 - Aug 2006

Use Ensembl to... Browse the *Crassostrea gigas* contigs database

Search Contig Browser

Table 1. Summary of assembly and RNA-Seq statistics for de novo assembly and reference mapping (GigasDatabase v8).

	De novo assembly	Reference mapping	
Assembly	Mapped reads	8,407,963	29,107,760
	Unmapped reads	36,944,698	16,244,901
	Contigs	18,510	77,433
	Average contig length	276	554
	Average contig coverage	62	16
	Contigs annotated to GO Slim	3931	7296
RNA-Seq	Differentially expressed genes	2991	427
	Enriched GO biological process	15	3

About the *Crassostrea gigas* contigs

Statistics

Assembly: pcg8, Mar 2011

Number of base pairs (Contigs): 67,875,621

Number of sequences (EST/mRNA): 1,013,570

Total number of contigs: 82,312

Number of singlets: 20,148

Polymorphism: Mar 2011

Number of putative SNPs: 29,493

Number of contigs including SNP: 10,392

Contigs were annotated searching sequence homologies against following databases:

Characterizing short read sequencing for gene discovery and RNA-Seq analysis in *Crassostrea gigas*

Mackenzie R. Gavery, Steven B. Roberts *

INRA

▶ Sigenae Oyster Contigs V8
▶ RefSeq RNA Index Blast of 09-Jan-2011
▶ TIGR Fugu FGI 3.0

Example Data Points

Jump directly to the example entry points:

Contig: [BQ426804.p.cg.8](#)

SNP: [snp_EW777925_683](#)

Example

S!  **Public Sigenae**

Based on Ensembl release 40 - Aug 2006

Use Ensembl to... [Search Contig Browser](#) [Browse the Crassostrea gigas genome](#) [About the Crassostrea gigas genome](#)

Table 1. Summary of assembly and RNA-Seq statistics for de novo assembly (v8).

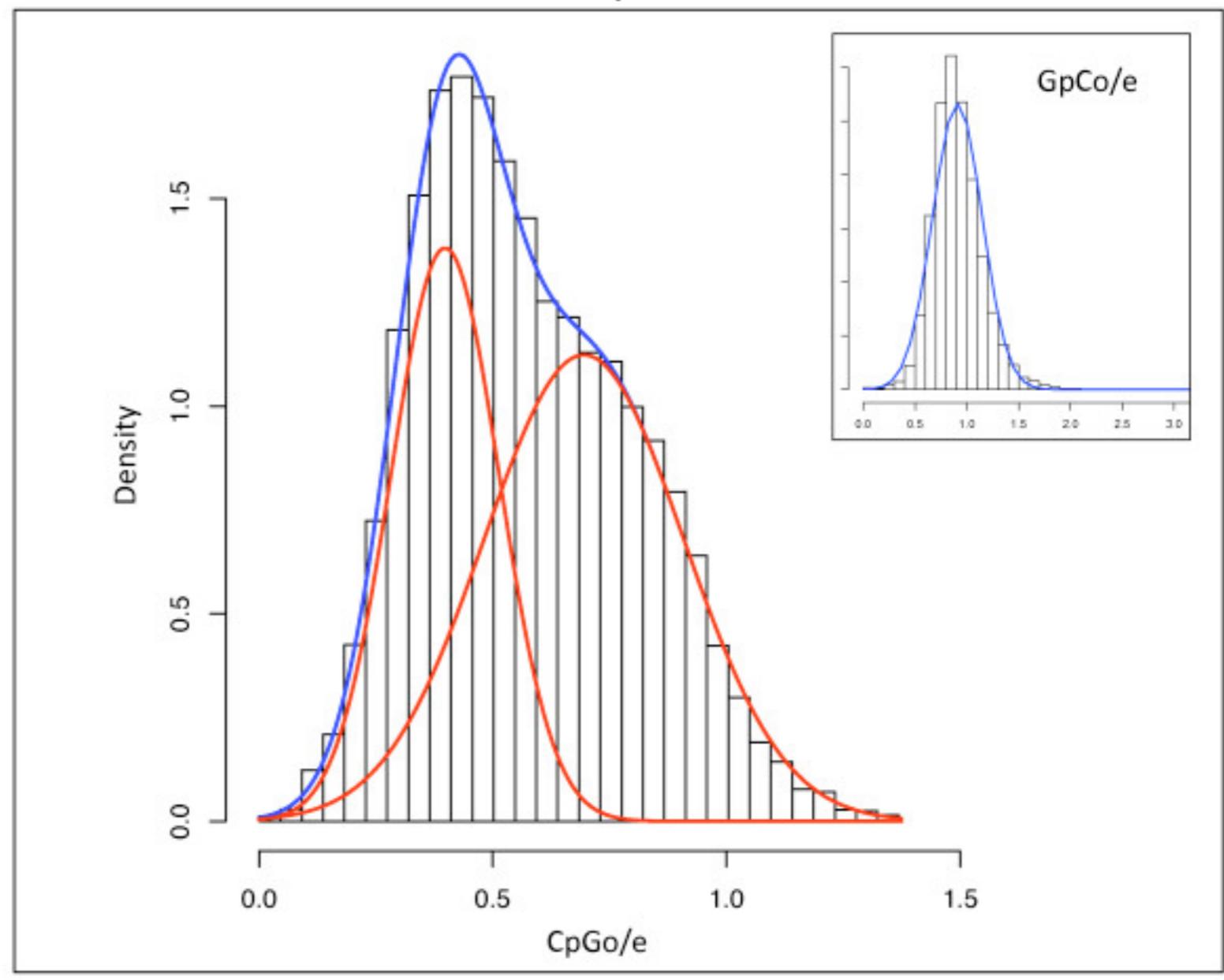
	De novo assembly	Referer
Assembly	Mapped reads	8,407,963
	Unmapped reads	36,944,698
	Contigs	18,510
	Average contig length	276
	Average contig coverage	62
	Contigs annotated to GO Slim	3931
RNA-Seq	Differentially expressed genes	2991
	Enriched GO biological process	15

Contigs were annotated search against following databases:

- UniProtKB/Swiss-Prot Reference Proteins 2011
- RefSeq Protein Index Blast
- Pfam Release 24.0 of June 2011
- Sigenae Oyster Contigs 1.0
- RefSeq RNA Index Blast
- TIGR Fugu FGI 3.0

[Sigenae](#) [Agenae](#)



Research article **Highly accessed** **Open Access**

DNA methylation patterns provide insight into epigenetic regulation in the Pacific oyster (*Crassostrea gigas*)

Mackenzie R Gavery and Steven B Roberts*

DOI: <https://doi.org/10.1186/1471-2164-12-503>

Living in an Ivory Basement

Stochastic thoughts on science, testing, and programming.

[misc](#) [personal](#) [python](#) [science](#) [teaching](#) [testing](#)

Loligo pealeii (squid) data dump

A few months back, I announced [the khmer protocols project](#), an effort to write down an explicit, open protocol for transcriptome and metagenome assembly. This project was started [during the summer of 2013 at the Woods Hole Marine Biological Lab](#), in collaboration with [Joshua Rosenthal](#). I'd met Josh at the [CephSeq meeting](#) the year before, and we reconnected at MBL in 2013; there, motivated in part by discussions at CephSeq, I started writing the eel-pond protocol.

During the summer, Josh and I found that we were both incredibly frustrated by the general failure of researchers to share data pre-pub, and so we started to chat seriously about some ideas we'd independently had about incentivizing the opening of transcriptome data. This eventually led to [the Open Marine Transcriptome project](#), which is still in its formative stages.

However, in the meantime I am happy to report that Josh assembled a bunch of squid transcriptomes and is willing to make them available to all. In addition, Josh did some skim Illumina sequencing (~40x coverage) of the squid genome, and my lab assembled it; we are making a really basic draft genome available as well. (Here, by squid, we mean *Loligo pealeii*, or the [Longfin inshore squid](#) -- "loligo" for short. It is also sometimes called "Doryteuthis pealeii".)

It should be noted that one of the transcriptomes was made from carefully dissected Giant Fiber Lobe neurons. These are the cell bodies for the well-studied squid giant axon. Thus this transcriptome should provide "molecular support" for all the cell biology and neurophysiology that has focused on this most famous of preps.

We are pleased to announce the availability of this data, in two formats.

First, you can download the assembled and annotated transcriptome data in FASTA format:

- [Loligo pealeii buccal ganglion transcriptome \(v1.0\)](#)
- [Loligo pealeii giant fiber lobe \(v1.0\)](#)
- [Loligo pealeii optic lobe \(v1.0\)](#)
- [Loligo pealeii stellate ganglion \(without giant fiber lobe\) \(v1.0\)](#)
- [Loligo pealeii vertical lobe \(v1.0\)](#)

Living in an Ivory Basement

Stochastic thoughts on science, testing, and programming.

misc personal python science teaching testing

Loligo pealeii (squid) data dump

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Mon 17 February 2014

By [C. Titus Brown](#)

In [science](#).

tags: [open dataf-yeah](#)

Why aren't you trying to get a Science or Nature paper out of all this? Aren't your reputations going to suffer for doing all this work without trying to milk the data for all it's worth?

We're pretty sure our reputations won't suffer from making a bunch of useful data available. Heck, we're pretty sure **your reputation** wouldn't suffer from making a bunch of useful data available (hint).

First, you can download the assembled and annotated transcriptome data in FASTA format:

- [Loligo pealeii buccal ganglion transcriptome \(v1.0\)](#)
- [Loligo pealeii giant fiber lobe \(v1.0\)](#)
- [Loligo pealeii optic lobe \(v1.0\)](#)
- [Loligo pealeii stellate ganglion \(without giant fiber lobe\) \(v1.0\)](#)
- [Loligo pealeii vertical lobe \(v1.0\)](#)



The Olympia oyster (*Ostrea lurida*) is the only native oyster on the west coast of the US.

Alleviating Regulatory Impediments To Native Shellfish Aquaculture

Our approach is to simultaneously address local adaptation in three genetically differentiated populations of Olympia oysters by evaluating genotype-by-environment interactions. We will reciprocally transplant seed produced from wild parents collected from contrasting environments into all environments. This very large reciprocal transplant experiment can test for a home field advantage in survival, maturation and growth in Olympia oysters. The overall goals of this project are to increase our knowledge of local adaptation in Olympia oysters to address concerns that interbreeding between potentially maladapted cultured and wild stocks could negatively impact wild populations. Accordingly, in order to attain these goals, the specific objectives of this proposal are to 1) Evaluate fitness components and performance of seed from different origins in a reciprocal transplant experiment and 2) Characterize genetic and epigenetic

Check out [our wiki](#) *for up to date data and more details.*

Genomic Resources

Transcriptomes

[O.lurida transcriptome version3 \[10/13\]](#)

[O.lurida transcriptome version2](#)

[O.lurida transcriptome version1](#)

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Steven Roberts

Associate Professor ([Marine Biology](#))
University of Washington



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Timmins-
Schiffman
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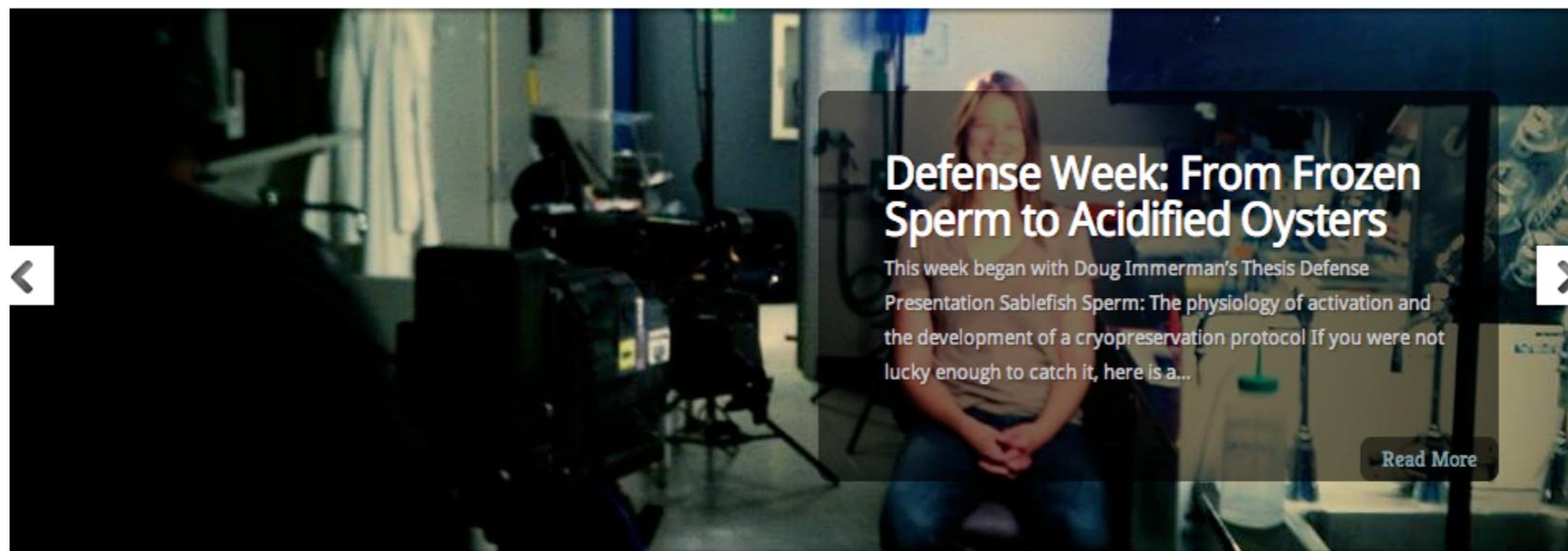
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[Bay scallop population structure on Cape Cod](#) Fileset 662 views 14 shares

[Hard clam transcriptome contigs](#) Dataset 453 views 5 shares

[DNA methylation as a mechanism to increase adaptive potential in invertebrates](#) Paper 412 views 1 shares

[Development of genetic markers to assess disease resistance in the eastern oyster](#) Paper 320 views 9 shares



• • • •

Research

Research in our lab focuses on characterizing physiological responses of marine organisms to environmental change. Using integrative approaches we strive to examine impacts and adaptive potential from the nucleotide to organismal level. A core component of this includes investigating the *functional relationship of genetics, epigenetics, and transcription*.

Open Science

We practice open science with lab members maintaining online electronic lab notebooks. In addition we have developed a Data and Resource Sharing Plan and are in the process of providing a list of resources we use for open science. To learn more about what we are doing, be sure to view our weekly, live lab meetings on YouTube, follow us on Facebook, or check out one or more of our blogs.

News and Notes

- Congrats to Emma Timmins-Schiffman! Successfully Passed her PhD Final Exam! [2/14]
- Congrats to Doug Immerman! Successfully Passed his Masters Final Exam! [2/14]
- Archive

Labcam



Notebook Updates

- Sam's Working Notebook Jan - July 2014 (kubu4)
- sr320 Notebook (sr320)
- sr320 Notebook (sr320)
- sr320 Notebook (sr320)

Fresh from the lab...

Big Data!
Exploring the Biology of Oysters
a few million base pairs at a time



Connect

- Roberts Lab
- Roberts Lab
- @sr320
- 206.685.3742

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Open Science Philosophy

Transparency with limited effort

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Molecular

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Open Science Philosophy

Transparency with limited effort

will try just about anything

Biology

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iPlant Galaxy

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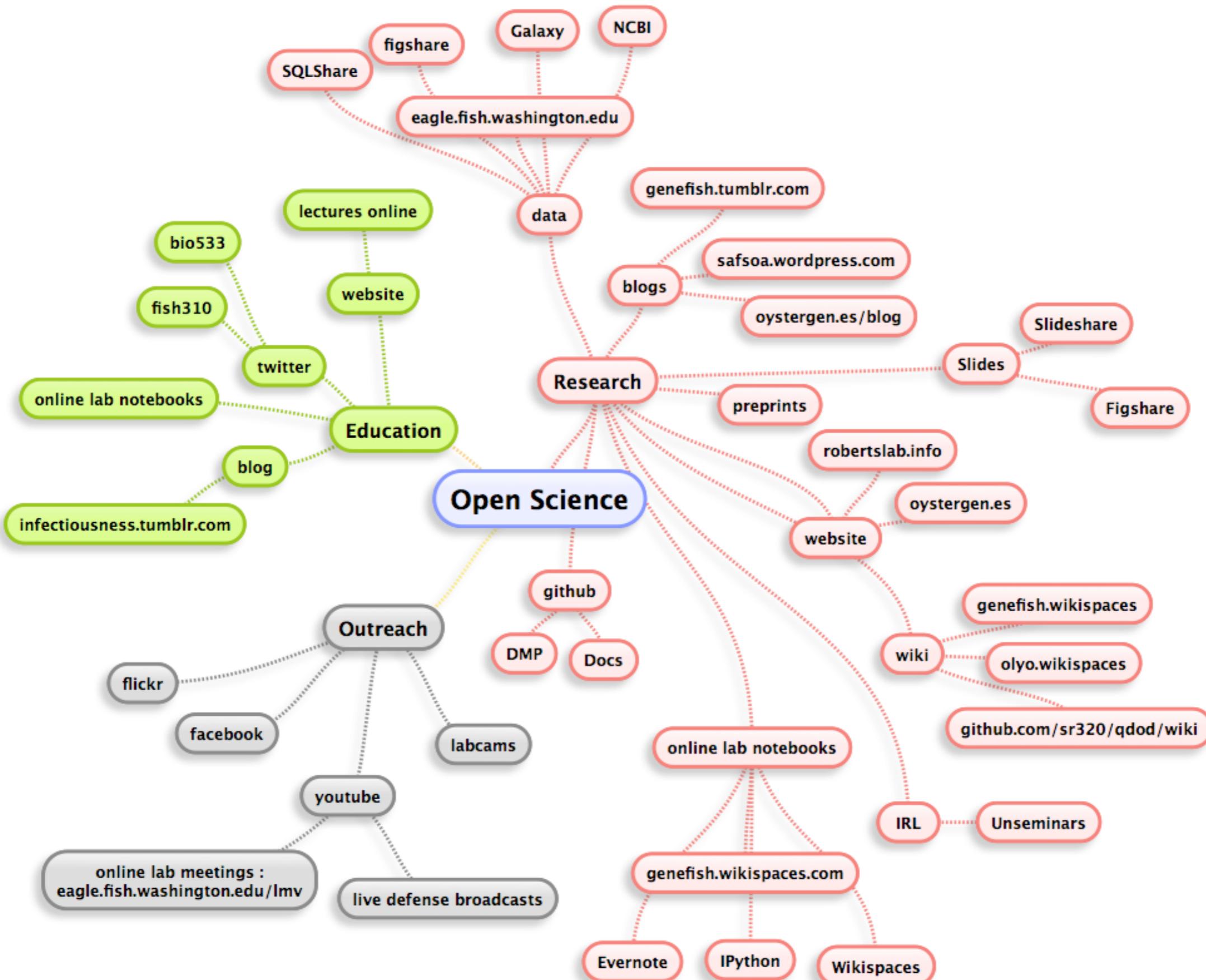
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Wednesday, 19 February 2014

In the ERA of science communication, Why you need Twitter, Professional Blog and ImpactStory?

Molecular

Yasset Perez-Riverol en Wednesday, February 19, 2014

Data Analysis

eScience

iPlant Galaxy

Notebooks

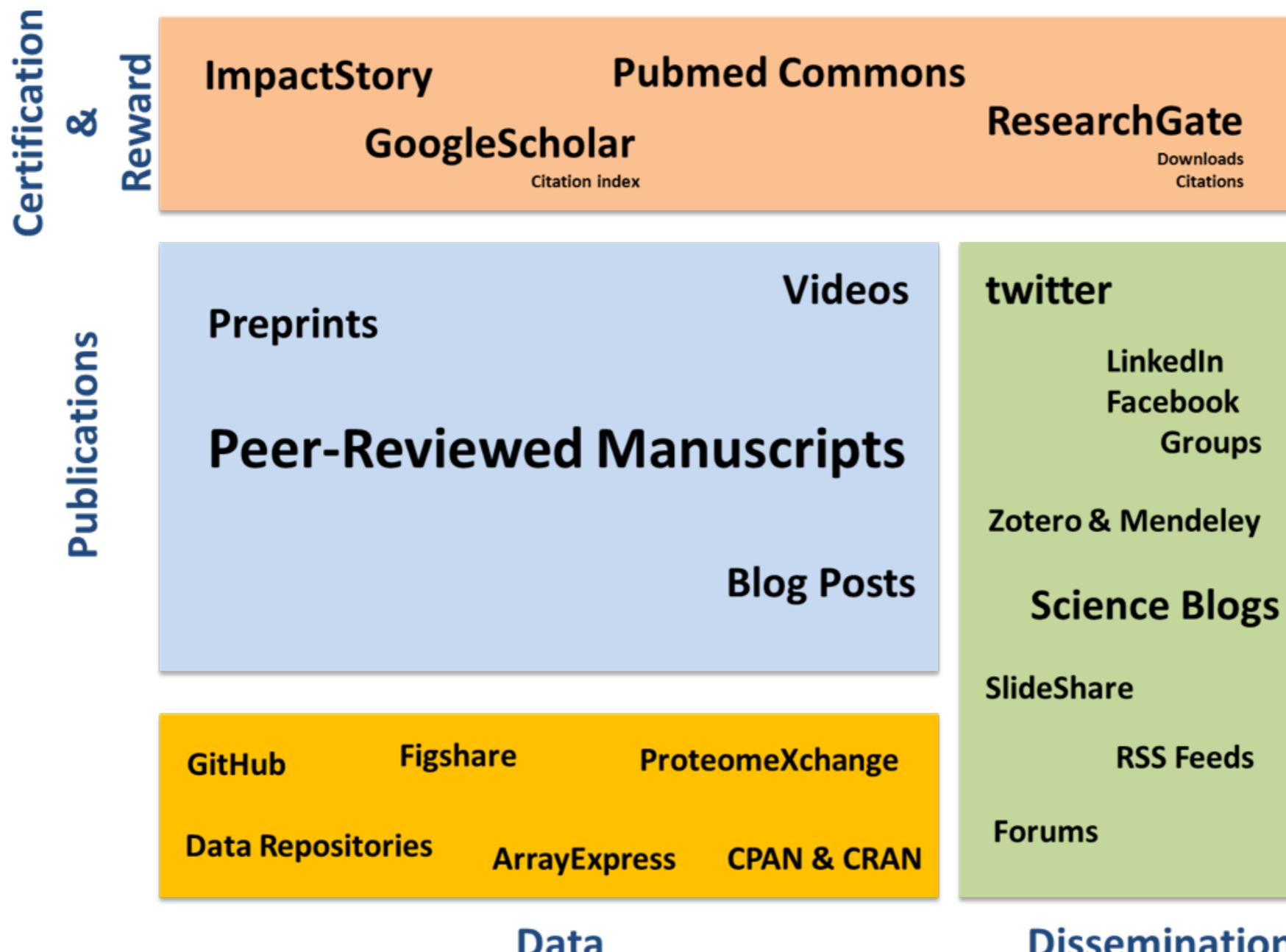
Rationale

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Start them early





Acknowledgements

Emma Timmins-Schiffman

Mackenzie Gavery
Claire Olson

Sam White
Brent Vadopalas
Jake Heare

Bill Howe
Dan Halperin



oystergen.es/data



DNA methylation



EPA
STAR



Aquaculture Program



acidification

Saltonstall-Kennedy

