
Dye/DNA Plates Documentation

Release v0.7

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DYE / DNA PLATES

The purpose of this code is to enable reproduction and facilitate extension of the computational results associated with following work

DeJaco, R. F.; Majikes, J. M.; Liddle, J. A.; Kearsley, A. J. Temperature-dependent Thermodynamic and Photophysical Properties of SYTO-13 Dye Bound to DNA.

The dependencies required can be installed via

```
>>> pip install -r requirements.txt
```

The paper can be reproduced with

```
>>> pytest .
```

in this directory. Alternatively, input the commands used in *Reproduction of Manuscript* in a file.

The raw data can be found in the directory *data/*

The documentation can be found in the *doc/* directory. A pdf version of the documentation is available at *doc/manual.pdf*

REPRODUCTION OF MANUSCRIPT

2.1 Raw Data and Scaling

First, we read-in the data associated with each data set (see Table 1 in paper) and store as a `src.get_data.RawData` class.

```
>>> import sys, os; sys.path.append(os.getcwd())
>>> from src.get_data import RawData
```

The data for each replicate plate possessing single-stranded DNA with $D_k = 1 \times 10^{-6}$ mol/L is input into the following structures

```
>>> SS_A_1 = RawData(fluorescence_file_name="ssDNA_2-23-2021.xls", D_k=1e-6, t="SS", l="A")
>>> SS_B_1 = RawData(fluorescence_file_name="8-2-2021_GC_ssDNA.xls", D_k=1e-6, t="SS", l="B")
>>> SS_C_1 = RawData(fluorescence_file_name="1xSS_GC_11-3-2021_data.xls", D_k=1e-6, t="SS", l="C")
```

The data for each replicate plate possessing single-stranded DNA with $D_k = 2 \times 10^{-6}$ mol/L is input into the following structures

```
>>> SS_A_2 = RawData(fluorescence_file_name="2x_ssDNA_2-24-2021.xls", D_k=2e-6, t="SS", l="A")
>>> SS_B_2 = RawData(fluorescence_file_name="gc_2xssDNA_11-2-2021_data.xls", D_k=2e-6, t="SS", l="B")
```

The data for each replicate plate possessing double-stranded DNA with $D_k = 1 \times 10^{-6}$ mol/L is input into the following structures

```
>>> DS_A_1 = RawData(fluorescence_file_name="GC_0p5_dsDNA_11-2-2021.xls", D_k=1e-6, t="DS", l="A")
>>> DS_B_1 = RawData(fluorescence_file_name="gc_dsDNA_1uM_12-10-2021_data.xls", D_k=1e-6, t="DS", l="B")
```

The data for each replicate plate possessing double-stranded DNA with $D_k = 2 \times 10^{-6}$ mol/L is input into the following structures

```
>>> DS_A_2 = RawData(fluorescence_file_name="8-2-2021_GCdsDNA.xls", D_k=2e-6, t="DS", l="A")
>>> DS_B_2 = RawData(fluorescence_file_name="gc_dsDNA_2uM_12-9-2021_data.xls", D_k=2e-6, t="DS", l="B")
```

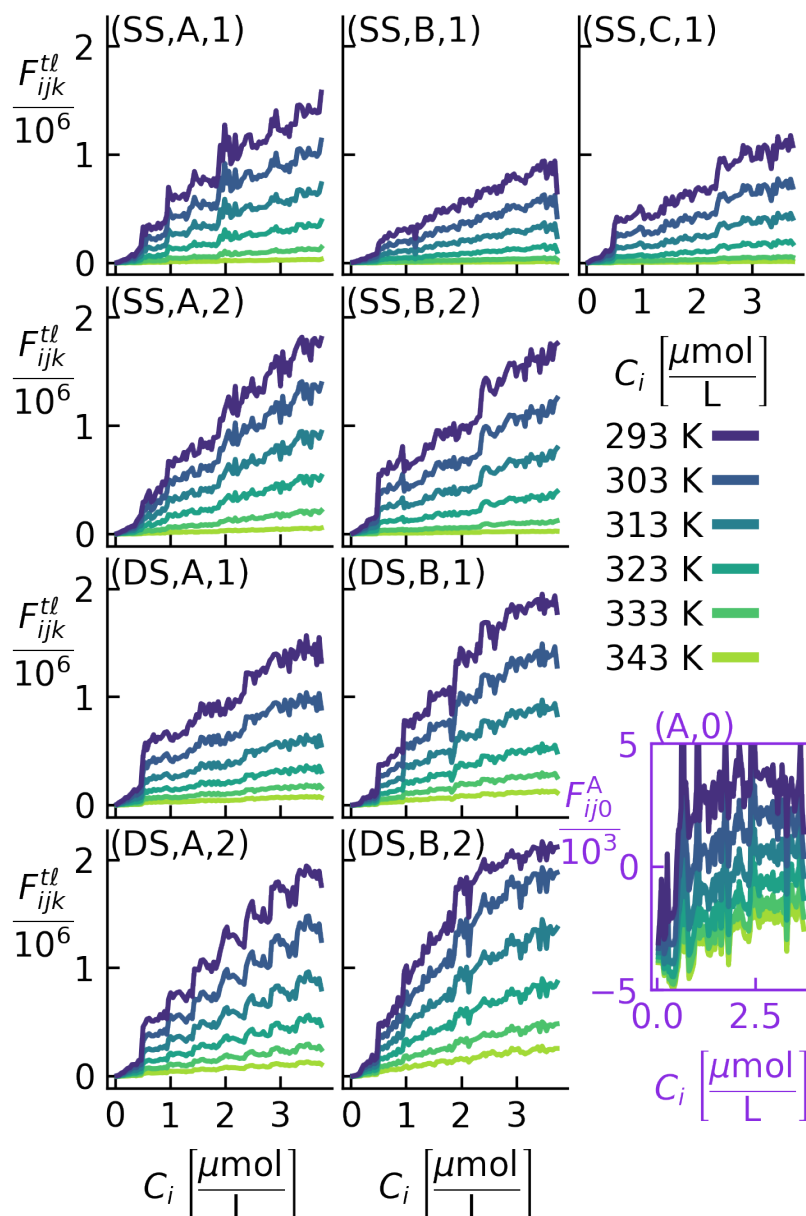
The data for the plate without DNA is input into the following structure

```
>>> A_1 = RawData(fluorescence_file_name="dyeOnly_11-6-2021_data.xls", D_k=0., t="None", l="A")
```

Having read-in the raw data, we plot it via

```
>>> from src.plot_raw_data import make_figure_2
>>> make_figure_2(SS_A_1, SS_B_1, SS_C_1, SS_A_2, SS_B_2, DS_A_1, DS_B_1, DS_A_2, DS_B_2, A_1)
```

which looks like



We combine the replicate plates by storing them as a `src.get_data.CombinedData` class.

```
>>> from src.get_data import CombinedData
```

The data for single-stranded DNA at $D = 1$ is


```
>>> SS_1 = CombinedData(SS_A_1, SS_B_1, SS_C_1)
```

The data for single-stranded DNA at $D = 2$ is

```
>>> SS_2 = CombinedData(SS_A_2, SS_B_2)
```

The data for double-stranded DNA at $D = 1$ is

```
>>> DS_1 = CombinedData(DS_A_1, DS_B_1)
```

The data for double-stranded DNA at $D = 2$ is

```
>>> DS_2 = CombinedData(DS_A_2, DS_B_2)
```

Having combined the data, we calculate F_{\min} via

```
>>> F_min = 0
>>> from src.get_data import C_REF, F_REF
>>> for dataset in (SS_1, SS_2, DS_1, DS_2):
...     wells = dataset.C*C_REF <= 0.5e-6
...     "Num wells <= 0.5e-6 mol/L for %s, %i is %d" % (dataset.t, int(dataset.D),
->len(dataset.C[wells]))
...     max_t_D = dataset.F[-1, wells].max()*F_REF
...     if max_t_D > F_min:
...         F_min = max_t_D
...
'Num wells <= 0.5e-6 mol/L for SS, 1 is 36'
'Num wells <= 0.5e-6 mol/L for SS, 2 is 24'
'Num wells <= 0.5e-6 mol/L for DS, 1 is 24'
'Num wells <= 0.5e-6 mol/L for DS, 2 is 24'
```

The value for F_{\min} is

```
>>> F_min
231432.0
```

The subsets of the data are made via

```
>>> for dataset in (SS_1, SS_2, DS_1, DS_2):
...     dataset.make_subset(F_min/F_REF)
...     "Max temperature for %s, %i is %g K" % (dataset.t, int(dataset.D), dataset.T.
->max())
...
'Max temperature for SS, 1 is 316.5 K'
'Max temperature for SS, 2 is 324.5 K'
'Max temperature for DS, 1 is 322 K'
'Max temperature for DS, 2 is 329 K'
```

2.2 Noise Removal

```
>>> from src.noise_removal import compute_M_LS
>>> import numpy as np
```

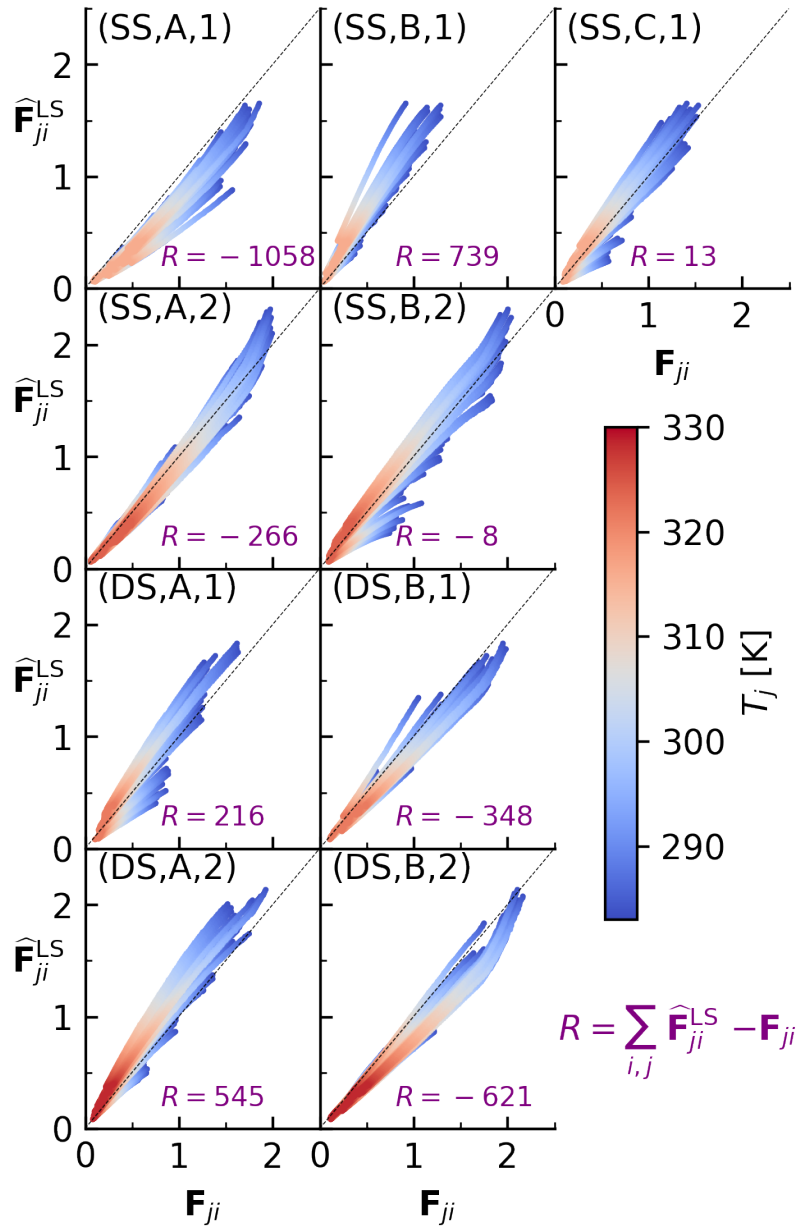
For each dataset, compute M^{LS} via Equation (21) and store the results,

```
>>> F_hats = []
>>> for dataset in (SS_1, SS_2, DS_1, DS_2):
...     M_LS = compute_M_LS(dataset.F, dataset.C)
...     F_hats.append(np.outer(M_LS, dataset.C))
... 
```

and then plot \mathbf{F} vs $\widehat{\mathbf{F}}$ for each combination via

```
>>> from src.plot_noise_removal import plot_Fhat_vs_F
>>> plot_Fhat_vs_F(
...     (SS_1.F, SS_2.F, DS_1.F, DS_2.F),
...     tuple(F_hats),
...     (SS_1.T, SS_2.T, DS_1.T, DS_2.T),
...     "figure3.png",
...     sname=r"$\widehat{\mathbf{F}}_{ji}^{\mathrm{LS}}$")
... 
```

This is Figure 3 in the main text, which looks like



Subsequently, Equation (22) is solved using `src.noise_removal.predictor_corrector()` and $V(\mathbf{M})$ and $V(\mathbf{C})$ are calculated

```
>>> from src.noise_removal import predictor_corrector
>>> RHO_SQUARED = 0.1
>>> for dataset in (SS_1, SS_2, DS_1, DS_2):
...     dataset.M_tls, dataset.C_hat = predictor_corrector(
...         dataset.F, dataset.C, RHO_SQUARED
...     )
...     dataset.Fhat_tls = np.outer(dataset.M_tls, dataset.C_hat.T)
...
...     m, n = dataset.F.shape
...     H = np.vstack([
...         np.hstack([np.eye(n) * (RHO_SQUARED + np.inner(dataset.M_tls, dataset.M_
... ↪ tls)), np.zeros((n, m))]),
```

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```

...         np.hstack([np.zeros((m, n)), np.eye(m)*np.inner(dataset.C_hat,
↪dataset.C_hat)])
...     ])
...     dF = dataset.Fhat_tls - dataset.F
...     dC = dataset.C_hat - dataset.C
...     f_star = (dF*dF).sum() + RHO_SQUARED*(dC*dC).sum()
...     bbV = f_star / (m*(n-1))*np.linalg.inv(H)
...     dataset.V_C = np.array([bbV[i, i] for i in range(n)])
...     dataset.V_M = np.array([bbV[j, j] for j in range(n, n + m)])
...
...     dataset.M_std = np.sqrt(dataset.V_M)
...     dataset.C_std = np.sqrt(dataset.V_C)
...
Total number of iterations was 756
Total number of iterations was 1347
Total number of iterations was 1928
Total number of iterations was 3073

```

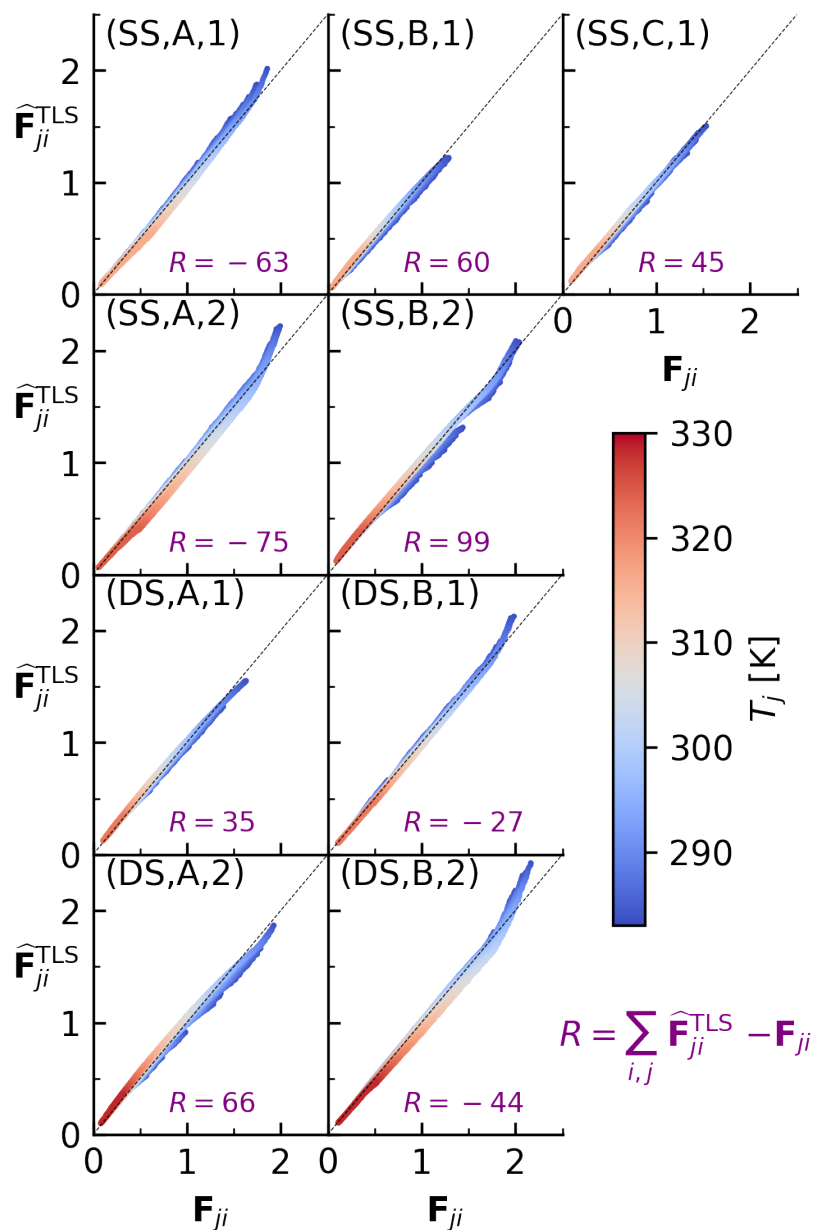
The results are plotted via Figure 4

```

>>> plot_Fhat_vs_F(
...     (SS_1.F, SS_2.F, DS_1.F, DS_2.F),
...     (SS_1.Fhat_tls, SS_2.Fhat_tls, DS_1.Fhat_tls, DS_2.Fhat_tls),
...     (SS_1.T, SS_2.T, DS_1.T, DS_2.T),
...     "figure4.png",
...     sname=r"$\widehat{\mathbf{F}}_{ji}^{\mathrm{TLS}}$")
...

```

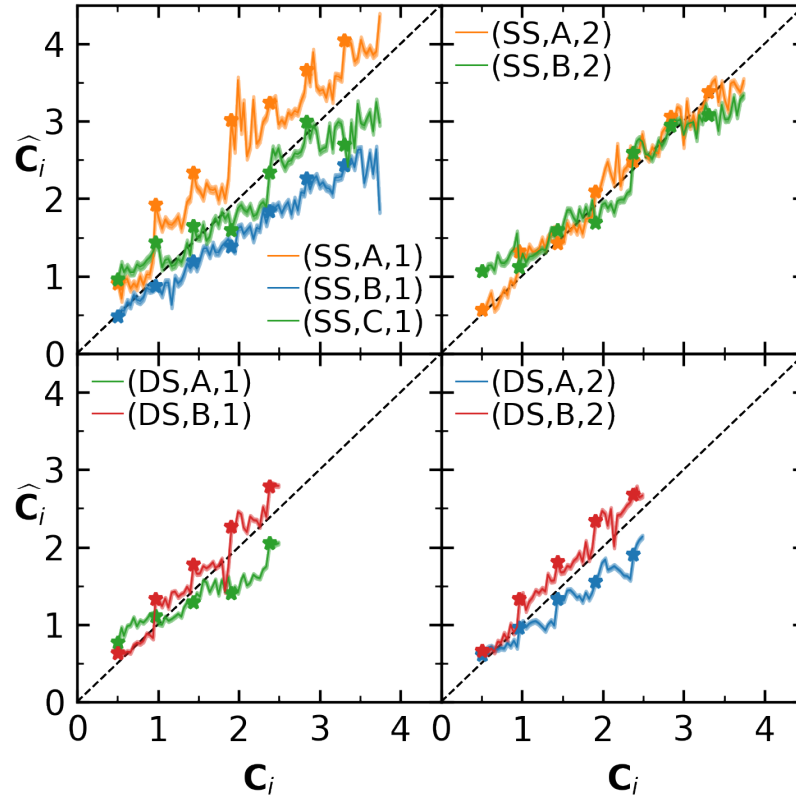
which looks like



and Figure 5,

```
>>> from src.plot_noise_removal import plot_Chat_vs_C
>>> plot_Chat_vs_C(
...     (SS_1.C, SS_2.C, DS_1.C, DS_2.C),
...     (SS_1.C_hat, SS_2.C_hat, DS_1.C_hat, DS_2.C_hat),
...     (SS_1.C_std, SS_2.C_std, DS_1.C_std, DS_2.C_std),
...     "figure5.png"
... )
... 
```

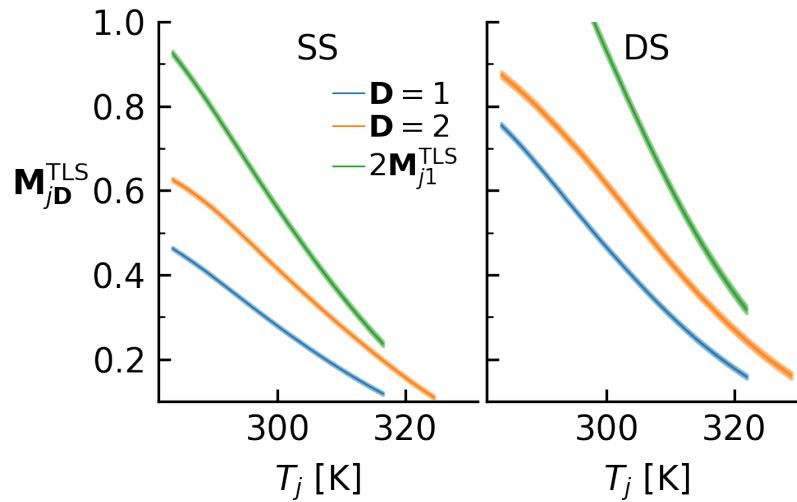
which looks like



and Figure 6,

```
>>> from src.plot_noise_removal import plot_figure6
>>> plot_figure6(
...     (SS_1.M_tls, SS_2.M_tls, DS_1.M_tls, DS_2.M_tls),
...     (SS_1.M_std, SS_2.M_std, DS_1.M_std, DS_2.M_std),
...     (SS_1.T, SS_2.T, DS_1.T, DS_2.T)
... )
... 
```

which looks like



2.3 Parameter Extraction

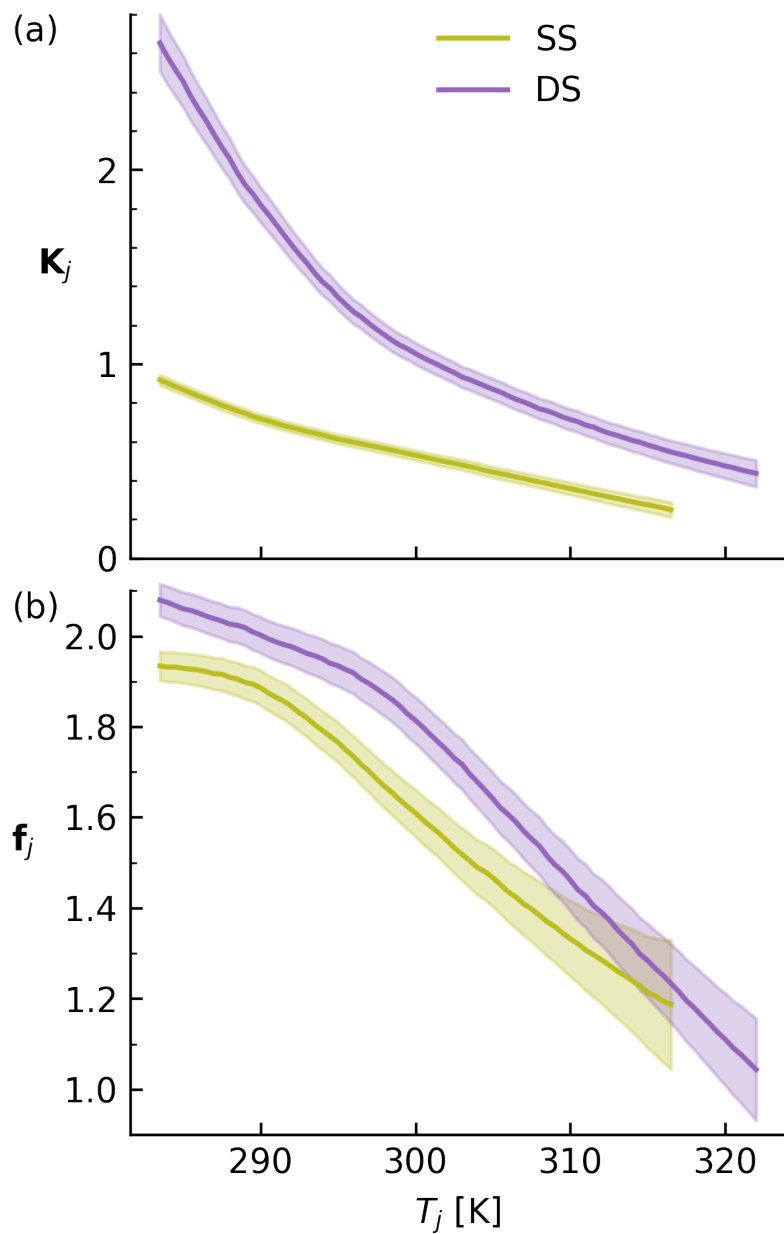
First, we combine the DNA concentrations associated with each DNA type into an instance of `src.parameter_extraction.Parameters`

```
>>> from src.parameter_extraction import Parameters
>>> SS = Parameters(SS_1, SS_2)
>>> DS = Parameters(DS_1, DS_2)
```

These instances now perform all parameter calculations; we can readily plot the Figure 7 as

```
>>> from src.plot_params import plot_figure7
>>> plot_figure7(
...     SS.T, SS.get_f(), SS.get_K(), SS.get_f_std(), SS.get_K_std(),
...     DS.T, DS.get_f(), DS.get_K(), DS.get_f_std(), DS.get_K_std(),
... )
...
```

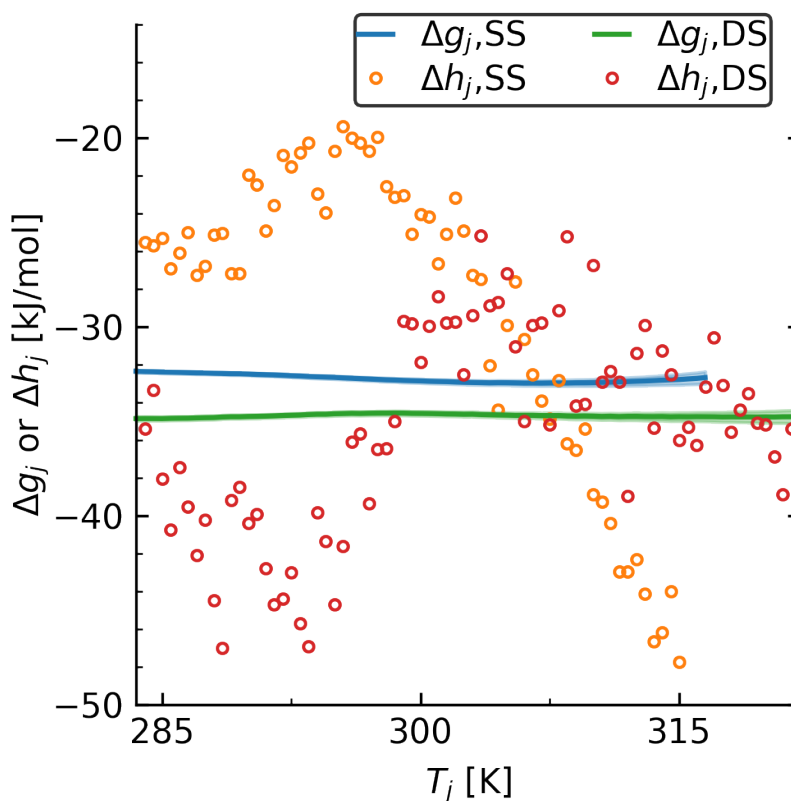
which looks like



and Figure 8,

```
>>> from src.plot_params import plot_figure8
>>> plot_figure8(SS, DS)
dg_SS at 295.00 K is -32.682584 +/- 0.088140
dg_DS at 295.00 K is -34.608172 +/- 0.108761
```

which looks like



2.4 Supplementary Figures

Figure S1 is made via

```
>>> from src.plot_raw_data import make_figure_S1
>>> make_figure_S1()
```

Figures S2, S3, S4 are made via

```
>>> from src.plot_params import plot_figure7, plot_figure8, plot_figure_S2, plot_
↳figure_S3, plot_figure_S4, plot_figure_S5
>>> plot_figure_S2(SS_1, SS_2, DS_1, DS_2)
>>> plot_figure_S3(SS, DS)
>>> plot_figure_S4(SS, DS)
```

Figure S5 is made via

```
>>> from src.parameter_extraction import calculate_relative_brightness, calculate_
↳relative_brightness_err
>>> rb = calculate_relative_brightness(SS.get_f(), DS.get_f())
>>> d_rb = calculate_relative_brightness_err(SS.M1, SS.M2, DS.M1, DS.M2,
...     SS.V_M1, SS.V_M2, DS.V_M1, DS.V_M2)
>>> plot_figure_S5(SS.T, rb, d_rb)
```


3.1 Wells

`src.wells.column_row_to_well` (*ix: int, iy: int*) → str
Convert indices to well

```
>>> column_row_to_well(0, 0)
'A1'
>>> column_row_to_well(0, 7)
'H1'
>>> column_row_to_well(11, 0)
'A12'
>>> column_row_to_well(11, 7)
'H12'
>>> well_to_row(column_row_to_well(11, 7))
7
>>> well_to_column(column_row_to_well(11, 7))
11
```

Parameters

- **ix** (*int*) – a index
- **iy** (*int*) – y index

Returns well

Return type str

`src.wells.number_to_column` (*number: int*) → int

```
>>> number_to_column(1)
1
>>> number_to_column(0)
0
>>> number_to_column(11)
11
>>> number_to_column(95)
11
>>> number_to_column(84)
0
```

`src.wells.number_to_row` (*number: int*) → int

```
>>> number_to_row(0)
0
>>> number_to_row(1)
0
>>> number_to_row(11)
0
>>> number_to_row(12)
1
>>> number_to_row(95)
7
>>> number_to_row(84)
7
```

`src.wells.number_to_well(number: int) → str`
Return number of well

```
>>> number_to_well(0)
'A1'
>>> number_to_well(11)
'A12'
>>> number_to_well(95)
'H12'
>>> number_to_well(84)
'H1'
```

`src.wells.well_to_column(well: str) → int`
Convert well name to column

```
>>> well_to_column('H100')
Traceback (most recent call last):
...
ValueError: '100' is not in list
>>> well_to_column('H1')
0
>>> well_to_column('A1')
0
>>> well_to_column('B12')
11
>>> well_to_column('B13')
Traceback (most recent call last):
...
ValueError: '13' is not in list
>>> well_to_column('Z1')
0
```

Parameters `well` (*str*) – name of well

Returns `ix` – a-index for well

Return type `int`

`src.wells.well_to_number(well: str) → int`
Return number of well

```
>>> well_to_number("A1")
0
>>> well_to_number("A12")
```

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```

11
>>> well_to_number("H12")
95
>>> well_to_number("H1")
84

```

`src.wells.well_to_row(well: str) → int`
Well to y index

```

>>> well_to_row('H100')
7
>>> well_to_row('H1')
7
>>> well_to_row('A1')
0
>>> well_to_row('B1')
1
>>> well_to_row('Z1')
Traceback (most recent call last):
...
ValueError: 'Z' is not in list

```

Parameters `well` (*str*) – well name

Returns `iy` – y-index of well

Return type `int`

3.2 Get Data

class `src.get_data.CombinedData(*replicates)`

Combined data from several replicate plates. See changes to F and C in *Raw Data and Scaling* portion of *Results and Discussion*.

Variables

- **\mathbf{F}** (*np.ndarray*) – Fluorescence data, \mathbf{F}_k^t in Equation (16a)
- **\mathbf{C}** (*np.ndarray*) – Dye concentrations, \mathbf{C}_k^t in Equation (16a)
- **\mathbf{D}** (*float*) – DNA concentration, \mathbf{D}_k in Equation (16a)
- **\mathbf{t}** (*str*) – Type of DNA, t , "SS" or "DS"
- **$\mathbf{M_tls}$** (*np.array*) – \mathbf{M}^{TLS} , set externally, defaults to `np.array([])`
- **$\mathbf{C_hat}$** (*np.array*) – $\hat{\mathbf{C}}$, set externally, defaults to `np.array([])`
- **$\mathbf{v_M}$** (*np.array*) – $V(\mathbf{M})$ (see Section S1.2), defaults to `np.array([])`
- **$\mathbf{v_C}$** (*np.array*) – $V(\mathbf{C})$ (see Section S1.2), defaults to `np.array([])`
- **$\mathbf{M_std}$** (*np.array*) – $\sqrt{V(\mathbf{M})}$ (see Section S1.2), defaults to `np.array([])`
- **$\mathbf{C_std}$** (*np.array*) – $\sqrt{V(\mathbf{C})}$ (see Section S1.2), defaults to `np.array([])`

`__init__(*replicates) → None`
Combine replicate F and C

Parameters **replicates** (*typing.List[Data]*) – list of plates to gather together as replicates

make_subset (*F_min*)

Make subset of data as described in *Raw Data and Scaling* portion of manuscript

Note: **F** and **C** are overwritten.

class `src.get_data.RawData` (*fluorescence_file_name*, *D_k*, *t*, *l*,
dye_conc_file_name='dye_conc_uM.csv')

Stores raw data.

Variables

- **F** (*np.ndarray*) – Fluorescence data, $F_k^{t\ell}$ (see Equation 14 of main text)
- **C** (*np.ndarray*) – Dye concentrations, C (see Equation 13 of main text)
- **D** (*float*) – DNA concentration, D_k in mol/L
- **t** (*str*) – Type of DNA, t , "SS" or "DS" or "None".
- **l** (*str*) – Replicate name, ℓ is A, B, or C

__init__ (*fluorescence_file_name*, *D_k*, *t*, *l*, *dye_conc_file_name*='dye_conc_uM.csv')

Scale the data before interpolating/solving optimization problem.

Parameters

- **fluorescence_file_name** (*str*) – name of fluorescence file within data folder
- **D_k** (*float*) – Total concentration of DNA in mol/L D_k
- **dye_conc_file_name** (*str*, *optional*) – name of dye concentration file name within data folder, defaults to "dye_conc_uM.csv"
- **t** (*str*) – Type of DNA, "SS", "DS", or "None".
- **l** (*str*) – Replicate name, ℓ is A, B, or C

`src.get_data.excel_to_data` (*f_name*: *str*, *channel*='GREEN')

Convert "Raw Data" sheet of excel file to pandas dataframe. Uses Equation (12) of manuscript to calculate temperature associated with each cycle.

Parameters

- **f_name** (*str*) – name of excel file
- **channel** (*str*, *optional*) – name of channel to investigate, defaults to "GREEN"

Returns Formatted data frame, with wells sorted from A1, A2 ... H11, H12

Return type `pd.DataFrame`

`src.get_data.get_C` (*file_name*)

Get total dye concentration associated with each well.

Parameters **file_name** (*str*) – CSV file formatted like a 96-well plate. The top left corner looks like

Row	1	2
A	.	.
B	.	.
C	.	.

The values are concentrations of dye in units of mol/L

Returns Mapping of well name (“A1”,...) to dye concentration [units of mol/L]

Return type Dictionary

3.3 Noise Removal

`src.noise_removal.compute_M_LS(F, C)`

Calculate M by least-squares approximation

Returns M^{LS} , see Equation (21)

Return type `np.array`

`src.noise_removal.compute_M_plus(F, c_plus)`

Get updated guess for M

Parameters

- **F** (`np.ndarray`) – Fluorescence matrix F
- **c_plus** (`np.array`) – Concentration matrix updated c_+

Returns M_+ by Equation (S3b)

Return type `np.array`

`src.noise_removal.compute_c_plus(F, C, M_minus, rho_squared)`

Compute updated guess of concentrations, c_+

Parameters

- **F** (`np.ndarray`) – Fluorescence F
- **C** (`np.array`) – Dye Concentration C
- **M_minus** (`np.array`) – Guess for M , M_-
- **rho_squared** (`float`) – Weight, ρ^2

Returns c_+ by Equation (S3a)

Return type `np.ndarray`

`src.noise_removal.predictor_corrector(F, C, rho_squared, maxiter=100000, print_iter=True)`

Solve Equation (22) with predictor–corrector algorithm

Parameters

- **F** (`np.ndarray`) – Fluorescence data F
- **C** (`np.ndarray`) – Dye concentration data C
- **rho_squared** (`float`) – Weighting factor for concentrations, ρ^2 in Equation (22a)
- **maxiter** (`int`, *optional*) – maximum iterations allowed, by default 100000
- **print_iter** (`bool`, *optional*) – whether or not to print the total number of iterations performed, by default True

Returns (M, c) – solution, (M^{TLS}, \hat{C})

Return type `tuple(np.array, np.array)`

3.4 Parameter Extraction

class `src.parameter_extraction.Parameters` (*cls1*: `src.get_data.CombinedData`, *cls2*: `src.get_data.CombinedData`)

Stores multiple instances of `CombinedData` for one DNA type

Variables

- **M1** (`np.array`) – M^{TLS} associated with $D = 1$
- **M2** (`np.array`) – M^{TLS} associated with $D = 2$. Several high temperatures are removed to reflect smaller temperature range associated with $D = 1$
- **C1** (`np.array`) – \hat{C} associated with $D = 1$
- **C2** (`np.array`) – \hat{C} associated with $D = 2$
- **r** (`np.array`) – r , as defined in Equation (S7).
- **v_C1** (`np.array`) – $V(C)$ associated with $D = 1$
- **v_C2** (`np.array`) – $V(C)$ associated with $D = 2$
- **v_M1** (`np.array`) – $V(M)$ associated with $D = 1$
- **v_M2** (`np.array`) – $V(\textit{mathbf{M}})$ associated with $D = 2$. Several high temperatures are removed to reflect smaller temperature range associated with $D = 1$
- **dT** (`float`) – Change in temperature from one cycle to next, ΔT

__init__ (*cls1*: `src.get_data.CombinedData`, *cls2*: `src.get_data.CombinedData`)

Initialize data

Note: Since different temperature ranges for each, need to make subset of dataset that has more temperatures. Dataset with lower DNA concentration `cls1` always has less temperatures.

Parameters

- **cls1** (`CombinedData`) – Data of DNA type at $D = 1$
- **cls2** (`CombinedData`) – Data of DNA type at $D = 2$

get_K () → `numpy.array`

Get **K** from vectorized version of Equation (24)

Returns **K**

Return type `np.array`

get_K_std () → `numpy.array`

Get standard deviation estimate of **K**

Returns

$$\sqrt{\Delta M^2 (4V(M_1) + 2V(M_2)) + \frac{\Delta H^2}{8\Delta M^4} (V(M_1) + V(M_2))}$$

where $\Delta H := 2M_1 - M_2$, $\Delta M := M_2 - M_1$

Return type `np.array`

get_dg () → `numpy.array`

Get free energy of dye binding, Δg , vectorized version of Equation (29).

Returns**Return type** np.array**get_dg_std**() → numpy.arrayGet estimate of standard deviation in Δg .**Returns**

$$\frac{RT_j}{2\Delta M_j (2M_{j1} - M_{j2})} \sqrt{M_{j2}^2 V(M_{j1}) + M_{j1}^2 V(M_{j2})}$$

where $\Delta M_j = M_{j2} - M_{j1}$.**Return type** np.array**get_dh**() → numpy.arrayGet differential enthalpy of binding, Δh as the vectorized version of Equation (30).**Returns****Return type** np.array**get_dh_std**() → numpy.arrayGet estimate of error in Δh_j **Returns****Return type** np.array**get_f**() → numpy.array

Get f from vectorized version of Equation (27)

Returns f**Return type** np.array**get_f_std**() → numpy.array

Get standard deviation estimate of f

Returns

$$\sqrt{V(M_1) + V(M_2) + \frac{E_B^2 V_A + E_A^2 V_B}{E_B^4}}$$

where

$$V_A := (V_B + H_+^2)(V(M_1) + V(M_2) + \Delta M^2) - E_A^2$$

$$V_B := 2V(M_1) + V(M_2)$$

$$E_A := \Delta M H_+$$

$$E_B := 2M_1 - M_2$$

$$\Delta M := M_2 - M_1$$

$$H_+ := 2M_1 + M_2$$

Return type np.array**get_phi_1**() → numpy.arrayGet φ_1 , vectorized version of Equation (S6a)**Returns****Return type** np.array

get_phi_2()

Get φ_2 , vectorized version of Equation (S6b)

Returns

Return type np.array

get_std_phi_1() → numpy.array

Get estimate of standard deviation in φ_1

Returns

$$\frac{2}{M_{2j}} \sqrt{V(M_{1j}) + r_j^2 V(M_{2j})}$$

Return type np.array

get_std_phi_2()

Get estimate of standard deviation in φ_2

Returns

$$\frac{1}{M_{1j}} \sqrt{V(M_{2j}) + r_j^{-2} V(M_{1j})}$$

Return type np.array

src.parameter_extraction.calculate_relative_brightness(*f_SS, f_DS*)

Calculate relative brightness, Equation (28).

Parameters

- **f_SS** (*np.array*) – Molar fluorescence of single-stranded DNA, f^{SS} .
- **f_DS** (*np.array*) – Molar fluorescence of double-stranded DNA, f^{DS} .

Returns Relative brightness, f_j^{DS}/f_j^{SS} for each j associated with SS.

Return type np.array

src.parameter_extraction.calculate_relative_brightness_err (*SS_M1, SS_M2, DS_M1, DS_M2, SS_V_M1, SS_V_M2, DS_V_M1, DS_V_M2*) → numpy.array

Estimate error in relative brightness, Equation (28).

Parameters

- **SS_M1** (*np.array*) – M_1^{SS}
- **SS_M2** (*np.array*) – M_2^{SS}
- **DS_M1** (*np.array*) – M_1^{DS}
- **DS_M2** (*np.array*) – M_2^{DS}
- **SS_V_M1** (*np.array*) – $V(M_1^{SS})$ _description_
- **SS_V_M2** (*np.array*) – $V(M_2^{SS})$
- **DS_V_M1** (*np.array*) – $V(M_1^{DS})$
- **DS_V_M2** (*np.array*) – $V(M_2^{DS})$

Returns Estimate of error in relative brightness

Return type np.array

3.5 Plotting

3.5.1 Raw Data

```
src.plot_raw_data.make_figure_2(SS_A_1:      src.get_data.RawData,      SS_B_1:
                                src.get_data.RawData, SS_C_1:      src.get_data.RawData,
                                SS_A_2:      src.get_data.RawData,      SS_B_2:
                                src.get_data.RawData, DS_A_1:      src.get_data.RawData,
                                DS_B_1:      src.get_data.RawData,      DS_A_2:
                                src.get_data.RawData, DS_B_2:      src.get_data.RawData,
                                A_1: src.get_data.RawData)
```

Makes Figure 2

```
src.plot_raw_data.make_figure_S1()
```

Makes Figure S1.

```
src.plot_raw_data.plot_linemap(cls:      src.get_data.RawData,      ax,      colorbar=False,
                                get_ticks=False, ordered_by_row=True)
```

Plot F vs C for various temperatures (colors)

Parameters

- **cls** (*Data*) – Instance of data (i.e., a dataset)
- **ax** (*axis*) – Matplotlib axis to plot on
- **colorbar** (*bool, optional*) – whether or not to plot colorbar, in which case the axis is colorbar axis, by default False
- **get_ticks** (*bool, optional*) – whether or not to return list of ticks, by default False
- **ordered_by_row** (*bool, optional*) – whether or not well concentrations are ordered by row, by default True

Returns Only returns list of `get_ticks=True`.

Return type None or list

3.5.2 Noise Removal

3.5.3 Parameters

```
src.plot_params.plot_figure_S5(T, rb, d_rb)
```

Plot figure S5, relative brightness

Parameters

- **T** (*np.array*) – array of temperatures
- **rb** (*np.array*) – Relative brightness, $f_j^{\text{DS}}/f_j^{\text{SS}}$
- **d_rb** (*np.array*) – standard deviation in relative brightness

3.6 Util

`src.util.figure_name_to_abspath(fname: str) → str`

Figure name to absolute path

Parameters `fname` (*str*) – name of figure

Returns absolute path to name of figure

Return type `str`

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