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# **Dye/DNA Plates Documentation**

***Release v3.0.0***

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**Mar 15, 2023**



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## MODULES

## 1.1 Wells

`src.wells.column_row_to_well` (*ix: int, iy: int*) → str  
Convert indices to well

```
>>> column_row_to_well(0, 0)
'A1'
>>> column_row_to_well(0, 7)
'H1'
>>> column_row_to_well(11, 0)
'A12'
>>> column_row_to_well(11, 7)
'H12'
>>> well_to_row(column_row_to_well(11, 7))
7
>>> well_to_column(column_row_to_well(11, 7))
11
```

**Parameters**

- **ix** (*int*) – a index
- **iy** (*int*) – y index

**Returns well**

**Return type** str

`src.wells.number_to_column` (*number: int*) → int

```
>>> number_to_column(1)
1
>>> number_to_column(0)
0
>>> number_to_column(11)
11
>>> number_to_column(95)
11
>>> number_to_column(84)
0
```

`src.wells.number_to_row` (*number: int*) → int

```
>>> number_to_row(0)
0
>>> number_to_row(1)
0
>>> number_to_row(11)
0
>>> number_to_row(12)
1
>>> number_to_row(95)
7
>>> number_to_row(84)
7
```

`src.wells.number_to_well(number: int) → str`  
Return number of well

```
>>> number_to_well(0)
'A1'
>>> number_to_well(11)
'A12'
>>> number_to_well(95)
'H12'
>>> number_to_well(84)
'H1'
```

`src.wells.well_to_column(well: str) → int`  
Convert well name to column

```
>>> well_to_column('H100')
Traceback (most recent call last):
...
ValueError: '100' is not in list
>>> well_to_column('H1')
0
>>> well_to_column('A1')
0
>>> well_to_column('B12')
11
>>> well_to_column('B13')
Traceback (most recent call last):
...
ValueError: '13' is not in list
>>> well_to_column('Z1')
0
```

**Parameters** `well` (*str*) – name of well

**Returns** `ix` – a-index for well

**Return type** `int`

`src.wells.well_to_number(well: str) → int`  
Return number of well

```
>>> well_to_number("A1")
0
>>> well_to_number("A12")
```

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```

11
>>> well_to_number("H12")
95
>>> well_to_number("H1")
84

```

`src.wells.well_to_row(well: str) → int`  
Well to y index

```

>>> well_to_row('H100')
7
>>> well_to_row('H1')
7
>>> well_to_row('A1')
0
>>> well_to_row('B1')
1
>>> well_to_row('Z1')
Traceback (most recent call last):
...
ValueError: 'Z' is not in list

```

**Parameters** `well` (*str*) – well name

**Returns** `iy` – y-index of well

**Return type** `int`

## 1.2 Get Data

**class** `src.get_data.CombinedData` (\*replicates)

Combined data from several replicate plates. See changes to  $F$  and  $C$  in *Raw Data and Scaling* portion of *Results and Discussion*.

### Variables

- $\mathbf{F}$  (*np.ndarray*) – Fluorescence data,  $\mathbf{F}_D^t$  in Equation (13a)
- $\mathbf{C}$  (*np.ndarray*) – Dye concentrations,  $\mathbf{C}_D^t$  in Equation (13a)
- $\mathbf{D}$  (*float*) – DNA concentration,  $\mathbf{D}$  in Equation (13a)
- $\mathbf{t}$  (*str*) – Type of DNA,  $t$ , "SS" or "DS"
- $\mathbf{N}$  (*int*) – number of nucleobases per single strand,  $N$
- $\mathbf{M\_tls}$  (*np.array*) –  $\mathbf{M}^{\text{TLS}}$  in Equation (18a), set externally, defaults to `np.array([])`
- $\mathbf{C\_hat}$  (*np.array*) –  $\hat{\mathbf{C}}$  in Equation (18a), set externally, defaults to `np.array([])`
- $\mathbf{v\_M}$  (*np.array*) –  $V(\mathbf{M})$  (see section S1.3 in supporting material), defaults to `np.array([])`
- $\mathbf{v\_C}$  (*np.array*) –  $V(\mathbf{C})$  (see section S1.3 in supporting material), defaults to `np.array([])`
- $\mathbf{M\_std}$  (*np.array*) –  $\sqrt{V(\mathbf{M})}$ , defaults to `np.array([])`
- $\mathbf{C\_std}$  (*np.array*) –  $\sqrt{V(\mathbf{C})}$ , defaults to `np.array([])`

`__init__` (\*replicates) → None  
Combine replicate  $F_d^{t,l}$  and  $C$

**Parameters** `replicates` (`typing.List[Data]`) – list of plates to gather together as replicates

`make_subset` ( $F_{min}$ )

Make subset of data as described in *Raw Data and Scaling* portion of manuscript

---

**Note:**  $F$  and  $C$  are overwritten.

---

**class** `src.get_data.RawData` (`fluorescence_file_name`, `B_d`, `t`, `l`, `N`,  
`dye_conc_file_name='dye_conc_uM.csv'`)

Stores raw data.

#### Variables

- $F$  (`np.ndarray`) – Fluorescence data,  $F_d^{t,l}$  (see Eq. 11)
- $C$  (`np.ndarray`) – Dye concentrations,  $C$  (see Eq. 10)
- $B$  (`float`) – DNA concentration,  $B_d$  in mol/L
- $t$  (`str`) – Type of DNA,  $t$ , "SS" or "DS" or "None".
- $l$  (`str`) – Replicate name,  $l$  is "A", "B", or code: "C".
- $N$  (`int`) – number of nucleobases per single strand

`__init__` (`fluorescence_file_name`, `B_d`, `t`, `l`, `N`, `dye_conc_file_name='dye_conc_uM.csv'`)

Scale the data before interpolating/solving optimization problem.

#### Parameters

- `fluorescence_file_name` (`str`) – name of fluorescence file within data folder
- `B_d` (`float`) – Total concentration of nucleobases in mol/L  $B_d$
- `dye_conc_file_name` (`str`, `optional`) – name of dye concentration file name within data folder, defaults to "dye\_conc\_uM.csv"
- `t` (`str`) – Type of DNA, "SS", "DS", or "None".
- `l` (`str`) – Replicate name,  $l$  is A, B, or C
- `N` (`int`) – number of nucleobases per single strand

`src.get_data.excel_to_data` (`f_name: str`, `channel='GREEN'`)

Convert "Raw Data" sheet of excel file to pandas dataframe. Uses Equation (9) of manuscript to calculate temperature associated with each cycle.

#### Parameters

- `f_name` (`str`) – name of excel file
- `channel` (`str`, `optional`) – name of channel to investigate, defaults to "GREEN"

**Returns** Formatted data frame, with wells sorted from A1, A2 ... H11, H12

**Return type** `pd.DataFrame`

`src.get_data.get_C` (`file_name`)

Get total dye concentration associated with each well.



**Parameters** `file_name` (*str*) – CSV file containing values of concentrations of dye in units of  $\mu$  mol/L. Formatted like a 96-well plate:

Row	1	2	3	4	5	6	7	8	9	10	11	12
A	.	.	.	.	.	.	.	.	.	.	.	.
B	.	.	.	.	.	.	.	.	.	.	.	.
C	.	.	.	.	.	.	.	.	.	.	.	.
D	.	.	.	.	.	.	.	.	.	.	.	.
E	.	.	.	.	.	.	.	.	.	.	.	.
F	.	.	.	.	.	.	.	.	.	.	.	.
G	.	.	.	.	.	.	.	.	.	.	.	.
H	.	.	.	.	.	.	.	.	.	.	.	.

**Returns** Mapping of well name (“A1”,...) to dye concentration [units of mol/L]

**Return type** Dictionary

## 1.3 Quantifying Noise and Linearity

`src.noise_removal.compute_M_LS(F, C)`

Calculate M by least-squares approximation

**Returns**  $M^{LS}$ , see Eq. 17.

**Return type** np.array

`src.noise_removal.compute_M_plus(F, c_plus)`

Get updated guess for M

**Parameters**

- **F** (*np.ndarray*) – Fluorescence matrix **F**
- **c\_plus** (*np.array*) – Concentration matrix updated  $c_+$

**Returns**  $M_+$  by Eq. S3b (see supporting material).

**Return type** np.array

`src.noise_removal.compute_c_plus(F, C, M_minus, rho_squared)`

Compute updated guess of concentrations,  $c_+$

**Parameters**

- **F** (*np.ndarray*) – Fluorescence **F**
- **C** (*np.array*) – Dye Concentration **C**
- **M\_minus** (*np.array*) – Guess for M,  $M_-$
- **rho\_squared** (*float*) – Weight,  $\rho^2$

**Returns**  $c_+$  by Eq. S3a (see supporting material).

**Return type** np.ndarray

`src.noise_removal.predictor_corrector(F, C, rho_squared, maxiter=100000, print_iter=True)`

Solve Eqs. 18 with predictor–corrector algorithm

**Parameters**

- **F** (*np.ndarray*) – Fluorescence data **F**
- **C** (*np.ndarray*) – Dye concentration data **C**
- **rho\_squared** (*float*) – Weighting factor for concentrations,  $\rho^2$  in Equation (18a)
- **maxiter** (*int, optional*) – maximum iterations allowed, by default 100000
- **print\_iter** (*bool, optional*) – whether or not to print the total number of iterations performed, by default True

**Returns** (**M**, **c**) – solution, ( $M^{TLS}$ ,  $\hat{C}$ )

**Return type** tuple(np.array, np.array)

## 1.4 Parameter Extraction

**class** `src.parameter_extraction.Parameters` (*cls1: src.get\_data.CombinedData, cls2: src.get\_data.CombinedData*)  
Stores multiple instances of CombinedData for one DNA type

### Notes

$M^{TLS}$  and  $\hat{C}$  are defined in Eq. 18a.

#### Variables

- **N** (*int*) – number of nucleobases,  $N$
- **M1** (*np.array*) –  $M^{TLS}$  associated with **D** = 1
- **M2** (*np.array*) –  $M^{TLS}$  associated with **D** = 2. Several high temperatures are removed to reflect smaller temperature range associated with **D** = 1
- **C1** (*np.array*) –  $\hat{C}$  associated with **D** = 1
- **C2** (*np.array*) –  $\hat{C}$  associated with **D** = 2
- **r** (*np.array*) –  $r$ , as defined in Equation (S7).
- **v\_C1** (*np.array*) –  $V(C)$  associated with **D** = 1
- **v\_C2** (*np.array*) –  $V(C)$  associated with **D** = 2
- **v\_M1** (*np.array*) –  $V(M)$  associated with **D** = 1
- **v\_M2** (*np.array*) –  $V(M)$  associated with **D** = 2. Several high temperatures are removed to reflect smaller temperature range associated with **D** = 1
- **dT** (*float*) – Change in temperature from one cycle to next,  $\Delta T$

**\_\_init\_\_** (*cls1: src.get\_data.CombinedData, cls2: src.get\_data.CombinedData*)  
Initialize data

---

**Note:** Since different temperature ranges for each, need to make subset of dataset that has more temperatures. Dataset with lower DNA concentration `cls1` always has less temperatures.

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#### Parameters

- **cls1** (`CombinedData`) – Data of DNA type at **D** = 1

- **cls2** (`CombinedData`) – Data of DNA type at  $D = 2$

**get\_K()** → numpy.array

Get **K** from vectorized version of Eq. 21.

**Returns** **K**

**Return type** np.array

**get\_K\_std()** → numpy.array

Get standard deviation estimate of **K**

**Returns**

$$\sqrt{\Delta M^2 (4V(M_1) + 2V(M_2)) + \frac{\Delta H^2}{8\Delta M^4} (V(M_1) + V(M_2))}$$

where  $\Delta H := 2M_1 - M_2$ ,  $\Delta M := M_2 - M_1$

**Return type** np.array

**get\_dg()** → numpy.array

Get free energy of dye binding,  $\Delta g$ , vectorized version of Eq. 25.

**Returns**

**Return type** np.array

**get\_dg\_std()** → numpy.array

Get estimate of standard deviation in  $\Delta g$ .

**Returns**

$$\frac{RT_j}{2\Delta M_j (2M_{j1} - M_{j2})} \sqrt{M_{j2}^2 V(M_{j1}) + M_{j1}^2 V(M_{j2})}$$

where  $\Delta M_j = M_{j2} - M_{j1}$ .

**Return type** np.array

**get\_dh()** → numpy.array

Get differential enthalpy of binding,  $\Delta h$  as the vectorized version of Eq. 26.

**Returns**

**Return type** np.array

**get\_dh\_std()** → numpy.array

Get estimate of error in  $\Delta h_j$

**Returns**

**Return type** np.array

**get\_f()** → numpy.array

Get **f** from vectorized version of Eq. 23.

**Returns** **f**

**Return type** np.array

**get\_f\_std()** → numpy.array

Get standard deviation estimate of **f**

**Returns**

$$\sqrt{V(\mathbf{M}_1) + V(\mathbf{M}_2) + \frac{E_B^2 V_A + E_A^2 V_B}{E_B^4}}$$

where

$$\begin{aligned} V_A &:= (V_B + H_+^2)(V(\mathbf{M}_1) + V(\mathbf{M}_2) + \Delta \mathbf{M}^2) - E_A^2 \\ V_B &:= 2V(\mathbf{M}_1) + V(\mathbf{M}_2) \\ E_A &:= \Delta \mathbf{M} H_+ \\ E_B &:= 2\mathbf{M}_1 - \mathbf{M}_2 \\ \Delta \mathbf{M} &:= \mathbf{M}_2 - \mathbf{M}_1 \\ H_+ &:= 2\mathbf{M}_1 + \mathbf{M}_2 \end{aligned}$$

**Return type** np.array

**get\_theta\_b\_all\_1** (*n*)

Returns  $\theta_{b,j,1}$  from Eq. S5 pointwise in *j*. The index *b* is the total number of wells.

**Returns**

**Return type** np.array

src.parameter\_extraction.**calculate\_relative\_brightness** (*f\_SS*, *f\_DS*)

Calculate relative brightness, Equation (24).

**Parameters**

- **f\_SS** (*np.array*) – Molar fluorescence of single-stranded DNA,  $f^{\text{SS}}$ .
- **f\_DS** (*np.array*) – Molar fluorescence of double-stranded DNA,  $f^{\text{DS}}$ .

**Returns** Relative brightness,  $f_j^{\text{DS}}/f_j^{\text{SS}}$  for each *j* associated with SS. (see Eq. 24).

**Return type** np.array

src.parameter\_extraction.**calculate\_relative\_brightness\_err** (*SS\_M1*, *SS\_M2*,  
*DS\_M1*, *DS\_M2*,  
*SS\_V\_M1*, *SS\_V\_M2*,  
*DS\_V\_M1*,  
*DS\_V\_M2*) →  
numpy.array

Estimate error in relative brightness (Eq. 24).

**Parameters**

- **SS\_M1** (*np.array*) –  $\mathbf{M}_1^{\text{SS}}$
- **SS\_M2** (*np.array*) –  $\mathbf{M}_2^{\text{SS}}$
- **DS\_M1** (*np.array*) –  $\mathbf{M}_1^{\text{DS}}$
- **DS\_M2** (*np.array*) –  $\mathbf{M}_2^{\text{DS}}$
- **SS\_V\_M1** (*np.array*) –  $V(\mathbf{M}_1^{\text{SS}})$  \_description\_
- **SS\_V\_M2** (*np.array*) –  $V(\mathbf{M}_2^{\text{SS}})$
- **DS\_V\_M1** (*np.array*) –  $V(\mathbf{M}_1^{\text{DS}})$
- **DS\_V\_M2** (*np.array*) –  $V(\mathbf{M}_2^{\text{DS}})$

**Returns** Estimate of error in relative brightness

**Return type** np.array

## 1.5 Plotting

### 1.5.1 Raw Data

```
src.plot_raw_data.make_figure_2(SS_A_1:      src.get_data.RawData,      SS_B_1:
                               src.get_data.RawData,  SS_C_1:      src.get_data.RawData,
                               SS_A_2:      src.get_data.RawData,      SS_B_2:
                               src.get_data.RawData,  DS_A_1:      src.get_data.RawData,
                               DS_B_1:      src.get_data.RawData,      DS_A_2:
                               src.get_data.RawData,  DS_B_2:      src.get_data.RawData,
                               A_1: src.get_data.RawData)
```

Makes figure 2

#### Parameters

- **SS\_A\_1** (*RawData*) – Data associated with  $(t, l, d) = (SS, A, 1)$  (see Table 1 of main text)
- **SS\_B\_1** (*RawData*) – Data associated with  $(t, l, d) = (SS, B, 1)$  (see Table 1 of main text)
- **SS\_C\_1** (*RawData*) – Data associated with  $(t, l, d) = (SS, C, 1)$  (see Table 1 of main text)
- **SS\_A\_2** (*RawData*) – Data associated with  $(t, l, d) = (SS, A, 2)$  (see Table 1 of main text)
- **SS\_B\_2** (*RawData*) – Data associated with  $(t, l, d) = (SS, B, 2)$  (see Table 1 of main text)
- **DS\_A\_1** (*RawData*) – Data associated with  $(t, l, d) = (DS, A, 1)$  (see Table 1 of main text)
- **DS\_B\_1** (*RawData*) – Data associated with  $(t, l, d) = (DS, B, 1)$  (see Table 1 of main text)
- **DS\_A\_2** (*RawData*) – Data associated with  $(t, l, d) = (DS, A, 2)$  (see Table 1 of main text)
- **DS\_B\_2** (*RawData*) – Data associated with  $(t, l, d) = (DS, B, 2)$  (see Table 1 of main text)
- **A\_1** (*RawData*) – Data associated with  $(l, d) = (A, 1)$  (without DNA, see Table 1 of main text)

```
src.plot_raw_data.make_figure_S1()
```

Makes Figure S1.

```
src.plot_raw_data.make_figure_S3()
```

Makes Figure S3.

```
src.plot_raw_data.plot_linemap(cls:      src.get_data.RawData,      ax,      colorbar=False,
                               get_ticks=False, ordered_by_row=True)
```

Plot  $F_d^t(T_j, C_i)$  vs  $C_i$  for various temperatures  $T_j$  (colors)

#### Parameters

- **cls** (*Data*) – Instance of data (i.e., a dataset)
- **ax** (*axis*) – Matplotlib axis to plot on

- **colorbar** (*bool, optional*) – whether or not to plot colorbar, in which case the axis is colorbar axis, by default False
- **get\_ticks** (*bool, optional*) – whether or not to return list of ticks, by default False
- **ordered\_by\_row** (*bool, optional*) – whether or not well concentrations are ordered by row, by default True

**Returns** Only returns list of `get_ticks=True`.

**Return type** None or list

## 1.5.2 Quantifying Noise and Linearity

`src.plot_noise_removal.plot_Chat_vs_C(Cs, C_hats, C_stds, fname)`

Plot comparison between  $\mathbf{C}$  and  $\hat{\mathbf{C}}$ . Used to plot Fig. 6

### Parameters

- **Cs** (*tuple of np.arrays*) – each element of the tuple corresponds to  $\mathbf{C}$  for a specific  $\mathbf{D}$  and type.
- **C\_hats** (*tuple of np.arrays*) – each element of the tuple corresponds to  $\hat{\mathbf{C}}$  for a specific  $\mathbf{D}$  and type.
- **C\_stds** (*tuple of np.arrays*) – each element of the tuple corresponds to the estimate for the standard deviation in  $\hat{\mathbf{C}}$  for a specific  $\mathbf{D}$  and type.
- **fname** (*str*) – file name to save as figure (relative path).

`src.plot_noise_removal.plot_Fhat_vs_F(Fs, Fhats, Ts, fname, sname= "\widehat{\mathbf{F}}_{ji}^{\mathrm{LS}}")`

### Parameters

- **Fs** (*tuple of np.array*) – each element of the tuple corresponds to  $\mathbf{F}$  for a specific  $\mathbf{D}$  and type.
- **Fhats** (*tuple of np.array*) – each element of the tuple corresponds to  $\hat{\mathbf{F}}$  for a specific  $\mathbf{D}$  and type.
- **Ts** (*tuple of np.array*) – each element of the tuple corresponds to the temperatures associated with a specific  $\mathbf{D}$  and type.
- **fname** (*str*) – name of figure to plot (relative path)
- **sname** (*str*) – name of symbol in latex

`src.plot_noise_removal.plot_error_C(ax, C, Chat, Chat_std=None, **kwargs)`

Plot Errors in Dye concentration

### Parameters

- **ax** (*matplotlib.pyplot.axis*) – plots on this axis
- **C** (*np.array*) – Nominal dye concentrations determined experimentally (dimensionless)
- **Chat** (*np.array*) – Dye concentrations determined from total least squares
- **Chat\_std** (*np.array*) – Estimate of standard deviations in dye concentrations determined from total least squares
- **kwargs** – Used to determine color if not provided. Also used when calling `ax.plot`.

`src.plot_noise_removal.plot_error_F(ax, F_k_tl, F_hat_k_tl, T)`

Plot the predicted fluorescence  $\hat{\mathbf{F}}_{ijk}^{t\ell}$  against  $\mathbf{F}_{ijk}^{t\ell}$  for  $i = 1, \dots, n$  and  $j = 1, \dots, n$ .

#### Parameters

- **ax** (*matplotlib.pyplot.axes*) – axes to plot on
- **F\_k\_l** (*np.ndarray*) – Experimentally measured fluorescence  $\mathbf{F}_k^{t\ell}$ .
- **F\_hat\_k\_l** (*np.ndarray*) – Predicted fluorescence  $\hat{\mathbf{F}}_k^{t\ell}$ .
- **T** (*np.array*) – Temperatures in K

**Returns** image for colorbar

**Return type** `img`

`src.plot_noise_removal.plot_figure7(M_mean, M_std, T, color_D1='C0', color_D2='C1')`

Plot figure 7

#### Parameters

- **M\_mean** (*tuple of np.array*) – each element of the tuple corresponds to  $\mathbf{M}^{\text{TLS}}$  for a specific **D** and type.
- **M\_std** (*tuple of np.array*) – each element of the tuple corresponds to standard deviations in  $\mathbf{M}^{\text{TLS}}$  for a specific **D** and type.
- **T** (*tuple of np.array*) – each element of the tuple corresponds to the temperatures associated with a specific **D** and type.
- **color\_D1** (*str*) – name of color for data associated with **D** = 1
- **color\_D2** – name of color for data associated with **D** = 2

### 1.5.3 Parameters

`src.plot_params.plot_figure8(T_SS, f_SS, K_SS, f_std_SS, K_std_SS, T_DS, f_DS, K_DS, f_std_DS, K_std_DS)`

Plot figure 8

#### Parameters

- **T\_SS** (*np.array*) – temperatures for single-stranded DNA
- **f\_SS** (*np.array*) – molar fluorescence for single-stranded DNA
- **K\_SS** (*np.array*) – partition coefficients for single-stranded DNA
- **f\_std\_SS** (*np.array*) – standard deviation in molar fluorescence of single-stranded DNA
- **K\_std\_SS** (*np.array*) – standard deviation in partition coefficients for single-stranded DNA
- **T\_DS** (*np.array*) – temperatures for double-stranded DNA
- **f\_DS** (*np.array*) – molar fluorescence for double-stranded dna
- **K\_DS** (*np.array*) – partition coefficients for single-stranded DNA
- **f\_std\_DS** (*np.array*) – standard deviation in molar fluorescence of double-stranded DNA
- **K\_std\_DS** (*np.array*) – standard deviation in partition coefficients for double-stranded DNA

```
src.plot_params.plot_figure9(SS:          src.parameter_extraction.Parameters,      DS:
                             src.parameter_extraction.Parameters)
```

**Parameters**

- **SS** (*Parameters*) – object containing all single-stranded DNA input and methods to perform calculations
- **DS** (*Parameters*) – object containing all single-stranded DNA input and methods to perform calculations

```
src.plot_params.plot_figure_S2(SS_1:          src.get_data.CombinedData,
                               SS_2:          src.get_data.CombinedData,
                               DS_1:          src.get_data.CombinedData,      DS_2:
                               src.get_data.CombinedData)
```

Plot figure S2

**Parameters**

- **SS\_1** (*CombinedData*) – object containing all replicate plates for single-stranded DNA with  $D = 1$
- **SS\_2** (*CombinedData*) – object containing all replicate plates for single-stranded DNA with  $D = 2$
- **DS\_1** (*CombinedData*) – object containing all replicate plates for double-stranded DNA with  $D = 1$
- **DS\_2** (*CombinedData*) – object containing all replicate plates for double-stranded DNA with  $D = 2$

```
src.plot_params.plot_figure_S4(SS_data:      src.parameter_extraction.Parameters,  DS_data:
                             src.parameter_extraction.Parameters)
```

Plot Figure S4

**Parameters**

- **SS\_data** (*Parameters*) – parameters class that has method to calculate  $\theta_{b,j,1}$  pointwise in  $j$
- **DS\_data** (*Parameters*) – parameters class that has method to calculate  $\theta_{b,j,1}$  pointwise in  $j$

```
src.plot_params.plot_figure_S5(T, rb, d_rb)
```

Plot figure S5, relative brightness

**Parameters**

- **T** (*np.array*) – array of temperatures
- **rb** (*np.array*) – Relative brightness,  $f_j^{\text{DS}}/f_j^{\text{SS}}$
- **d\_rb** (*np.array*) – standard deviation in relative brightness

## 1.6 Util

```
src.util.figure_name_to_abspath(fname: str) → str
```

Figure name to absolute path

**Parameters** **fname** (*str*) – name of figure

**Returns** absolute path to name of figure



**Return type** str



## INDICES AND TABLES

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