# Counting visitors with wuepix

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The *wuepix* package counts visitor numbers using computer vision. Therefore three methods (Change Detection, HOG-Descriptor, YOLO-darknet) were wrapped into this package. Additional management tools, as a Ground-Truth-Data sampler, are also included here. This vignette demonstrates a typical workflow.

# **Packages**

```
# Installation
# devtools::install_github("georoen/wuepix")
library(wuepix)
library(tidyverse)
```

# Site configuration

#### Paths & Filenames

Define the directory paths and filename patterns implied by the data archive.

```
# Where to find Images?
## Raw data
img.folder_raw <- "IMG_raw/"
# Preprocessed (croped, scaled, enhanced,...)
img.folder <- "IMG/"
# Remove corrupted images by filesize (in byte)
threshold <- 10000
# How to grep date?
gsub.Date <- function(Filename){gsub("picam-", "", gsub(".jpg", "", Filename))}
# Date code
date.code <- "%Y%m%d-%H%M"

# Aggregation Scale
T_scale <- "20 mins"</pre>
```

#### Extent of interest

To speed up processing an extend of interest (EOI) should be selected. Using the Linux comandline tool ImageMagick, this can also include rotations as well as other image operations. However identifying the correct command involves visual interpretation of the results. To do so I proceeded as follows.

# 1. Using Gimp / Photoshop

Initially use GIMP to identify the pre-process routine (bounding-box, optional rotation).

Tipp: Overlay several images to cover different scenarios.

#### 2. Test comandline

After identifying the pre-process routine try to put the parameters into *ImageMagick* and test command on a single image using convert.

```
# Test
# SizeX x SizeY + PostionX + PositionY
convert.string <- "-crop 1600x800+0+1030"
cmd <- paste("convert extra/Ref_raw.jpg", convert.string, "extra/Ref.jpg")
system(cmd)
message("Please check cropped Ref.jpg, then proceed")</pre>
```

This results in the following extend of interest. Only this part of the image will be further analysed, so please only proceed if satisfied with the result.

#### 3. Preprocess image archive

Next all images will be pre-processed according to the routine developed above using mogrify. Please pay attention to the slightly different syntax of the mogrify -crop ... -path IMG/ IMG\_raw/\*.jpg. This will preprocess all images from IMG\_raw/ and save them in IMG/.

### List images

First all images need to be listed. The following chunk does so, plus enhances the data frame according to Site configuration: (1) due to external effects (eg. transmission) images can be corrupted. Here files with a file size smaller than the threshold will be exluded. (2) The Timestamp gets interpreted, therefore first the filenames are cropped with help of gsub.Date. Because filenames can be very different and the corresponding regular expression can very complex, it seemed easiest to do with a function. This also makes developing it more simple due better testing option. After cropping the timestamp it will be converted to a POSIXIt time object using date.code. (3) Last but not least the relative filepaths are reconstructed. Note, that this should also work with list.files(..., fullnames=TRUE) but I remeber then struggeling with grepping the datecode.

```
Files <- Files[order(Files$Timestamp),] # Order by Timestamp

# Full Filename
Files$Filename <- pasteO(img.folder, Files$Filename)
```

To get an overview about the data beeing processed, here some metadata summarys are promted.

```
## 21 files to analize
## Dates from 24.06.2017 12:30 to 24.06.2017 12:50
## Time difference of 20 mins
```

#### Ground-Truth-Data

To latter assess the classifiers accuracies, Ground-Truth-Data is mandatory. Use GTD\_list() to manually count pedestrians in Files\$Filename. Here all images (100%) got evaled, for sampling uncomment orange lines.

```
start <- Sys.time() # Get start time
#GTD <- GTD_list(sample(x = Files$Filename, size = 10))
#the.sample <- sample(c(1:nrow(Files)), size = 100)
#Files <- Files[the.sample,]
Files$GTD <- GTD_list(Files$Filename)
Files$GTD <- as.numeric(Files$GTD)
(Sys.time() - start) # Print runtime

save(Files, file = "Results/GTD.RData")
write.csv(Files, file = "Results/GTD.csv")</pre>
```

Sum of visitors in GTD.

To aggregate the time-series by T\_scale use fun\_Aggregation(). Use tidyverse gramar to select wished aggeration method (mean or sum).

```
# Aggregation
Files_res <- fun_Aggregation(Files$Timestamp, Files$GTD, T_scale) %>%
    select(-MEAN) %>%
    rename(GTD = SUM)
```

# Processing

Finally we can start processing the (preprocessed) image archive. For a detailed description of the methods, the reader is refered to the authors master thesis.

### Method 1: Change detection

This approach is inspired by methods used remote sensing and biotech. Using algebra, two pictures are applied against each other, revealing changes. Use CD\_list() for processing a list of images, including parallel processing. See ?CD\_single() for the available parameters.

```
# Processing
start <- Sys.time() # Get start time
CD <- CD_list(Files$Filename, Min = 0.9, method = "ratio",</pre>
```

```
predictions = "CD_Predictions")
(Sys.time() - start)
                      # Print runtime
## Time difference of 6.885636 secs
Files$Hum <- CD
# Aggregation
Files_res <- fun_Aggregation(Files$Timestamp, Files$Hum, T_scale) %>%
  select(-SUM) %>%
 rename(CD = MEAN) %>%
 left_join(Files_res)
## Joining, by = "Timestamp"
Now convert the number of changed pixels into visitor numbers by calibrating them.
# Calibration
lm_cal <- lm(GTD ~ 0+CD, data = Files_res)</pre>
summary(lm_cal)
##
## Call:
## lm(formula = GTD ~ 0 + CD, data = Files_res)
##
## Residuals:
##
        1
## -3.155 3.442
## Coefficients:
      Estimate Std. Error t value Pr(>|t|)
## CD 0.07309
                  0.03516
                             2.079
                                      0.285
##
## Residual standard error: 4.669 on 1 degrees of freedom
## Multiple R-squared: 0.8121, Adjusted R-squared: 0.6242
## F-statistic: 4.322 on 1 and 1 DF, p-value: 0.2854
Files_res$CD_pred <- round(predict(lm_cal, select(Files_res, -GTD)))</pre>
```

#### Method 2: Histogramms of Oriented Gradients

The second method uses *Histogramms of Oriented Gradients* to detect pedestrians [DALAL\_2005]. The HOG-Descriptor, focuses on a feature class of the same name. Here the *OpenCV*-Python implementation was wrapped into hog\_list(). For an installation guide please see ?hog\_install(). First however upscale the images according to the trainingset.

## Finished preprocessing

```
Files_resized <- gsub("IMG/", "IMG_resize/", Files$Filename)</pre>
# Processing
start <- Sys.time() # Get start time</pre>
HOG <- hog_list(Files_resized, resize = 1, padding = 24, winStride = 2,
                      Mscale = 1.05, predictions = "HOG Predictions/")
(Sys.time() - start) # Print runtime
## Time difference of 1.459361 secs
To access the object-based accuracies, run GTD_truePositives(). It will cor.test() the input for you and
returns the miss-rate, FPPW and so forth.
Files$HOG <- HOG
GTD_truePositives(Files$GTD, Files$HOG)
##
##
   Pearson's product-moment correlation
##
## data: GTD and PRD
## t = 9.7321, df = 19, p-value = 8.136e-09
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## 0.7936218 0.9643845
## sample estimates:
##
         cor
## 0.9126413
   FN TP FP
                     MR. FPPW
## 1 6 8 0 0.4285714
                            0 0.9126413
# Aggregation
Files_res <- fun_Aggregation(Files$Timestamp, Files$HOG, T_scale) %>%
  select(-MEAN) %>%
  rename(HOG = SUM) %>%
 left_join(Files_res)
```

### Method 3: Convolutional-Neural-Network

## Joining, by = "Timestamp"

The third and last object detector discussed in this study bases on Convolutional-Neural-Networks. Here YOLO [REDMON\_2016] was utilized, as it comes pre-trained and has a open license. To install it use yolo\_install(). Attention, after every wuepix installation it is necessary to run yolo\_update() as well, as this write a small file into the package installation linking to the YOLO installation. This links aids running the wrapper functions conveniently.

```
# yolo_install("~/Programmierung/YOLO")
yolo_update("~/Programmierung/YOLO")

## Changing working directory during update!

## Working directory has been resetted

## NULL
```

YOLO was wrapped into yolo\_list() and yolo\_single(), whereby the first function runs faster as the weights only get loaded once, while only the latter is capable of saving the predictions. Using sapply()

however it is possible to process a list of images and saving the predictions.

```
# Processing
start <- Sys.time() # Get start time</pre>
YOLO <- yolo_list(Files$Filename)
# YOLO <- sapply(Files$Filename, yolo_single, predictions = "YOLO_Predictions")
(Sys.time() - start) # Print runtime
## Time difference of 8.931464 mins
You can also use GTD_truePositives() here.
Files$YOLO <- YOLO
GTD_truePositives(Files$GTD, Files$YOLO)
##
##
   Pearson's product-moment correlation
##
## data: GTD and PRD
## t = 25.457, df = 19, p-value = 3.811e-16
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## 0.9642537 0.9942813
## sample estimates:
##
         cor
## 0.9856558
   FN TP FP MR
                      FPPW
                                  cor
## 1 0 14 2 0 0.0952381 0.9856558
# Aggregation
Files_res <- fun_Aggregation(Files$Timestamp, Files$YOLO, T_scale) %>%
  select(-MEAN) %>%
  rename(YOLO = SUM) %>%
 left_join(Files_res)
## Joining, by = "Timestamp"
As YOLO detects a lot of objects, yolo_list() logs a complete list of all detections to a separate file
yolo_detections.txt. To read it in a tidy way use yolo_Read().
# Read, group and count detections
yolo.results <- yolo_Read("yolo_detections.txt")</pre>
unique(yolo.results$Class)
## [1] "backpack"
                      "bicycle"
                                    "person"
                                                   "umbrella"
                                                                 "sports ball"
```

## Save Results

Last but not least, save the R environment for further studies.

```
save.image(file = "Results/Environment.RData")
```