- Title: DateLife: Leveraging databases and analytical tools to reveal the dated Tree of Life
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abstract.- The combination of new analytical techniques, availability of more fossil and molecular data, and better practices in data sharing has resulted in a steady accumulation of chronograms in public and open databases such as TreeBASE, Dryad, and Open Tree of Life for a large quantity and diversity of organisms in the last few decades. However, getting a tree with branch lengths proportional to time remains difficult for many biologists and the non-academic community, despite its importance in many areas of research, 11 education, and science communication. datelife is a service implemented via an R package and a web site 12 (http://www.datelife.org/) for efficient reuse, summary and reanalysis of published data on lineage divergence 13 times. The main workflow starts with at least two taxon names as input, either as tip labels on a tree, or as a simple comma separated character string. A name search is then performed across the chronogram database and positively identified source trees are pruned to maintain queried taxa only and stored as a named list of patristic distance matrices. Source chronogram data can be summarised using branch length summary statistics or variance minimizing approaches to generate a single summary chronogram. Source chronogram data can also be used as calibration points to date a tree containing some or all names from the initial query. If there is no information available for any queried taxa, data can be simulated. All source and summary chronograms can be saved in formats that permit easy reuse and reanalysis. Summary and newly generated trees are potentially useful to evaluate evolutionary hypothesis in different areas of research 22 in biology. How well this trees work for this purpose still needs to be tested. datelife will be useful to increase awereness on the existing variation in expert time of divergence data, and might foster exploration of the effect of alternative divergence time hypothesis on the results of analyses, nurturing a culture of more cautious interpretation of evolutionary results.

Keywords: Tree; Phylogeny; Scaling; Dating; Ages; Divergence times; Open Science; Congruification;

28 Supertree; Calibrations

29 INTRODUCTION

Clade ages represent a fundamental piece of information for evolutionary understanding in many areas of research, from developmental to conservation biology (Felsenstein 1985; Webb 2000), from historical biogeography to species diversification studies (Posadas et al. 2006; Morlon 2014). The primary information needed for these time estimates comes from the fossil record. Coupled with phylogenies with branch lengths based on molecular and/or morphological data, the time of divergence of extant and extinct lineages can be reconstructed with molecular dating methods. The number of studies publishing phylogenies with branch lengths proportional to geological time (hereafter chronograms) have constantly increased in number for the last two decades (Kumar et al. 2017). Still, generating a chronogram is not an easy task unless you have specialized training: it requires inferring a tree, understanding what fossil data are available and their limits, and where fossils go on the tree. That is why there has been an urge for promoting and facilitating reuse of the vast amount of phylogenetic and time of lineage divergence data that has been generated and made available in publications, for the advantage of research relying on this information (Webb and Donoghue 2005; Stoltzfus et al. 2013).

Wide interest from the scientific community to make information from phylogenies in general and chronograms in particular available for consultation and reuse has spurred the creation of public platforms with various goals and characteristics. TreeBASE (Morell 1996; Piel et al. 2002), the Dryad repository (http://datadryad.org/), and the Open Tree of Life (OToL; Hinchliff et al. 2015) platforms store and make available published phylogenies and chronograms for easy scientific reuse. All of them can be queried using automatised web procedures, which permit personalized, large scale queries that are also very fast. OToL stores trees with branch length information from a wide range of living organisms, implementing a metadata structure that stores the branch length units (i.e., time or relative susbtitution rates). Treebase and Dryad repositories also contain trees from all groups of life, but the former did not store branch length information until recently (and lacks consistent metadata on what any branchlengths stored mean) and Dryad stores many other types of biological data using metadata that does not allow automatic distinction of types of trees and branch length units, impairing the automatised access to time of lineage divergence information.

Besides keeping a repository to easily store and share expert phylogenetic and chronogram knowledge, OToL also has the primary goal of synthesising all trees in their repository to expose to the community a single tree of all life depicting the phylogenetic relationships among known lineages. All or parts of this synthetic tree 57 can be reused for any purpose. However, it currently only focus on synthesizing tree topology, meaning that it does not expose branch length data of any type. The Timetree of Life project focuses on the synthesis of a single chronogram of life (Hedges et al. 2006) and presents a very accessible, attractive interface. However, the thousands of chronograms this NSF-funded project have compiled for synthesis are only publicly available for visual examination in their website or for download as images, but large scale download remains prohibited by their site. The latest version of their synthetic chronogram (Kumar et al. 2017) can be queried only through their website in a non-automatised fashion, and only subsets of it can be reused for analyses with the permission of the authors. Other platforms such as SuperSmart (Antonelli et al. 2017) and phylogenerator (Pearse and Purvis 2013) are focused in automatised de novo chronogram inference, by reusing DNA sequence data to reconstruct phylogenetic trees. However, expert fossil information necessary for subsequent molecular dating analyses still needs to be compiled and curated by the user, rendering them a challenging tool to obtain data on time of lineage divergence for the non-specialist. Moreover, these tools do not provide information from already created expert chronograms.

A tool for efficient reuse of expert, published data on time of lineage divergence should have an open and fully public chronogram database storing data in a format suitable for scientific reuse, an automatised way of accessing the information, and straightforward means of comparing and summarizing chronogram information as needed by the user. A prototype service aiming to meet this characteristics was developed over a series of hackathons at the National Evolutionary Synthesis Center (Stoltzfus et al. 2013). In here we present the formal description and implementation of the datelife service, constituted by an R package and a web site (http://www.datelife.org/). There is still much room for improvement, and flaws and limitations are addressed below. We strived for the current implementation of datelife to perform the basic tasks described above, featuring a system for maintenance of an open database of chronograms pulled from public repositories, methods to summarize and compare source chronograms, and new functions to visualize and graphically compare source and summary chronograms.

DESCRIPTION

- The basic datelife workflow is shown in figure 1 and consists of:
- 1. A user providing at least two taxon names as input, either as tip labels on a tree, or as a simple comma separated character string. The tree can be in newick or phylo format, and can be with or without branch lengths.
- 2. A name search is then performed across the chronogram database; source trees with at least two
 matching input names are identified; all other taxa that do not match the original query are then
 dropped from the positively identified source trees. These pruned chronograms are hereafter referred as
 source chronograms. Finally, each source chronogram is transformed to a patristic matrix named by
 the citation of the original study. This format facilitates and greatly speeds up all further analyses and
 summarizing algorithms.
- 3. The user can obtain different summary information from the source chronograms including: a) all source chronograms, b) maximum ages of source chronograms, c) citations of studies where source chronograms were originally published, d) a summary table with all of the above, e) a single summary tree of all or a subset of source chronograms, f) a report of successful matches of input taxon names across source chronograms, and g) the single source chronogram with the greatest number of taxa. Summary information can be used to make decisions on the next steps of the workflow.
- 4. Source chronogram data can be used as calibration points to date a tree with or without branch lengths containing some or all names from the initial query.
- 5. If there is no information available for any queried taxa, users can also create both age and phylogenetic data for this missing taxa with a variety of algorithms described below.
- 6. Finally, users can easily save all source and summary chronograms in formats that permit easy reuse and reanalyses (newick and R 'phylo' format), as well as view and compare results graphically, or construct their own graphs using inbuilt 'datelife' graphic generation functions.

To gather, process, and present information, datelife builds up from functions available in several R
packages including rotl (Michonneau et al. 2016), ape (Paradis et al. 2004), geiger (Harmon et al. 2008),
paleotree (Bapst 2012), bold (Chamberlain 2018), phytools (Revell 2012), taxize (Chamberlain and Szöcs
2013; Chamberlain 2018), phyloch (Heibl 2008), phylocomr (Ooms and Chamberlain 2018) and rphylotastic
(O'Meara et al. 2019).

A datelife search currently accepts scientific names only. It can be any named clade or binomial specific. 111 Chronogram search is performed at the species level, so when input names correspond to named clades, 112 datelife pulls all accepted species names within the clade from OToL's reference taxonomy to perform the 113 search. Searches at the infraspecies level are not currently allowed, so input names belonging to subspecies or 114 any other infraspecific category are collapsed to the species level. datelife processes input names with the 115 taxon name resolution service (TNRS; Boyle et al. 2013), which corrects potentially misspelled names and 116 typos, and standardizes spelling variations and synonyms, increasing the probability of correctly finding the 117 queried taxa in datelife's chronogram database. 118

The chronogram search is performed across datelife's chronogram database which is assembled from OToL's
tree repository. Compared to other existing open tree repositories OToL's metadata rich tree store is the
only one that supports search, identification, and handling of chronograms in an automatised fashion. Also,
all their chronograms come from peer-reviewed published studies generated by specialists in the targeted
lineages, arguably representing expert knowledge on time of lineage divergence.

Information from source chronograms can be summarised with a summary statistic of tree branch lengths, such as median or mean. A much slower, but possibly more accurate Super Distance Matrix (SDM) approach for supertree reconstruction with branch lengths (Criscuolo et al. 2006) is also implemented via the ape package (Paradis et al. 2004). The resulting summary patristic distance matrix could be clustered with classic algorithms to return a tree. However, we noticed that the resulting trees are often non-ultrameric and do not reflect the source chronogram data (see datelife_examples package). Instead, we obtained a distribution of age data from the summary matrix available for nodes on a consensus tree. The Branch Length Adjuster (BLADJ) algorithm (Webb et al. 2008) was then used to distribute node ages evenly over the consensus tree,

minimizing age variance in the resulting chronogram.

For tree dating, the congruification algorithm described by Eastman et al. (2013) is implemented to find shared nodes between trees (congruent nodes). The ages of these nodes are then used as calibrations to date any given tree. Currently implemented methods for tree dating are BLADJ, MrBayes (Huelsenbeck and Ronquist 2001; Ronquist and Huelsenbeck 2003) and PATHd8 (Britton et al. 2007), a non-clock, rate-smoothing dating method.

138 BENCHMARK

datelife's code speed was tested on an Apple iMac with one 3.4 GHz Intel Core i5 processor. We registered 139 variation in computing time of query processing and search through the database relative to number of 140 queried taxon names. Query processing increases roughly linearly with number of input taxon names, and 141 increases considerably if TNRS service is activated. Up to ten thousand names can be processed and searched 142 in less than 30 minutes. A name search through the chronogram database with an already processed query can be performed in less than a minute, even with a very large number of taxon names (Fig. 2). datelife's 144 code performance was evaluated with a set of unit tests designed and implemented with the R package 145 testthat (R Core Team 2018) that were run locally –using the devtools package (R Core Team 2018), and on a public server -via GitHub, using the continuous integration tool Travis CI (https://travis-ci.org). At present, unit tests cover more than 50% of datelife's code (https://codecov.io/gh/phylotastic/datelife).

EXAMPLE EXAMPLE

In this section we demonstrate the types of outputs that can be obtained with datelife, using as an example
the bird family Fringillidae of true finches. We performed a higher-taxon search to obtain all data on
lineage divergence available in datelife's database for all recognised species within the Fringillidae (475 spp.
according to the Open Tree of Life taxonomy). There are 13 chronograms containing at least two Fringillidae
species, published in 9 different studies (Fig. 3). Data from these source chronograms was used to generate
two types of summary chronograms, median and SDM. As explained in the Description, data from source
chronograms was first summarised into a single distance matrix (using either the median or the SDM method)

and then the available node ages were used as calibrations points over a consensus tree topology, to obtain 157 a dated tree with the program BLADJ (Fig. 4). Median summary chronograms are older and have wider 158 variation in maximum ages than chronograms obtained with SDM. In both cases, ages are generally consistent 159 with source ages. Different source chronograms often show substantial variation in ages for clades (see, for 160 example, the ongoing debate about crown group age of angiosperms (Ramshaw et al. 1972; Sanderson and 161 Doyle 2001; Magallón et al. 2015; Barba-Montoya et al. 2018). For some studies, especially ones based 162 on branch lengths (studies of diversification, timing of events, trait evolution, and more), using a different 163 chronogram may return different results (Title and Rabosky 2016). Stitching together these chronograms can create a larger tree and uses information from multiple studies, but the effect of uncertainties and errors here on downstream analyses still requires more research.

Data from source chronograms was also used to date tree topologies with no branch length information and 167 trees with branch lengths in relative substitution rates (Figs. 5 and 6). As a form of cross validation, we used 168 tree topologies from each study and calibrated them using information from all other source chronograms. In the absence of branch length data, the ages of internal nodes were approximately recovered in almost 170 all cases (except for studies 3, and 5; Fig. 5). Maximum tree ages were only approximately recovered in 171 one case (study 2; Fig. 5). Branch lengths were successfully generated using the BOLD database for all 172 source chronograms. However, dating with PATHd8 (using congruified calibrations) was only successful in three cases (studies 3, 5, and 9; Fig. 6). From these, two trees have a different sampling than the 174 original source chronogram, mainly because DNA data for some species is absent from the BOLD. Maximum ages are quite different from source chronograms, but this might be explained also by the differences in sampling between source chronograms and BOLD trees. More examples and details can be consulted in https://github.com/LunaSare/datelife examples.

Flaws, Limitations and Prospects

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The main goal of datelife is to make expert information on time of lineage divergence easily accesible for comparison, reuse, and reanalysis, to researchers in all areas of science and with all levels of expertise in the matter. It is a very fast tool that fulfills the quality of openness and does not require any expert

biological knowledge from users –besides the names of the organisms they want to work with– for any of its functionalities. However, it has many flaws. Some of them can be overcome, some of them might represent limitations.

At the moment, datelife's chronogram database is not very large, storing 231 chronograms up to the time 186 the manuscript was written. This represents 5.79% of the largest existing chronogram database, which is 187 not open for scientific reuse nor automatised data mining (Kumar et al. 2017). OToL is the only public 188 tree repository from where datelife can currently pull chronograms to construct its database. A previous 189 version of TimeTree's synthetic chronogram (Hedges et al. 2015) was made available in the OToL repository, 190 hence the amount of lineages represented in datelife's database is at least as substantial as TimeTree's. This 191 ensures that some information will be available for any given query, but it does not ensure that the full state 192 of knowledge of time of divergence data will be available for any given lineage. Thus, incorporation of more 193 published chronograms deposited in OpenTree or perhaps pulled from Dryad directly, to datelife's database 194 is crucial to improve its services. Methods to automatically mine chronogram data from the Dryad repository could be designed and implemented. However, the unit of branch lengths would still need to be determined by hand. Consequently, we would like to emphasize on the importance of sharing chronogram data for the 197 scientific community, in repositories that require expert input and manual curation, such as OToL's tree repository.

Another potential concern comes from summary chronograms. We currently summarize all source chronogram data by default. Users can subset source data if they have reasons to favor some or one source chronogram over others. Strictly speaking, a good chronogram should reflect the real time of lineage divergence accurately and precisely. To our knowledge, there is no objective way to determine if an expert chronogram is better than other. Some criteria that have been put forward are the level of lineage sampling and the number of calibrations used. Scientists usually also favor chronograms coming from studies with primary calibrations to ones from secondary calibrations. It has been observed with simulations that divergence times inferred with secondary calibrations are significantly younger than those inferred with primary calibrations in analyses performed with bayesian inference methods when priors are implemented in similar ways in both analyses

(Schenk 2016). Yet, there are different ways to use secondary calibrations and the bias might not be encountered with other dating methods that do not require prior assumptions (such as ML methods). This remains to be tested.

Furthermore, even chronograms obtained with primary data can be very different, as observed from the comparison of source chronograms in the Fringillidae example. A large discrepancy in time of lineage 213 divergence across expert knowledge is well known for different groups of organisms (e.g., angiosperms; 214 Magallón et al. 2015). Comparison of available chronogram data for a wide range of organisms shown here 215 suggest that this is a widespread phenomenon that requires further attention. Characteristics of the data 216 used for dating analyses as well as from the output chronogram itself, such as quality of alignment (missing 217 data, GC content), lineage sampling strategy and proportion, phylogenetic and dating inference method, 218 number of fossils used as calibrations, support for nodes and ages, and confidence intervals could be used to 219 score quality of source chronograms. To facilitate subsetting of source chronograms following different criteria, 220 this information should be included as metadata manually entered by curators (as is done in OToL) in the 221 future. Still, even if all source chronograms have been generated by excellent standards and using similar 222 methods, the evolutionary history they depict might be very different. Hence, summarizing chronograms 223 might imply summarizing evolutionary hypothesis. This could be good from certain point of view, since it could help to get a single global evolutionary history for a lineage. But it could also be bad, since we could be loosing part of the evolutionary history that is only being reflected in some chronograms and not from the summary chronogram. Ideally, we should still rely on time of lineage divergence data obtained from a single analysis using fossil data as primary sources of calibrations, and using fossils that have already been curated as calibrations to date other trees, which should reflect a more homogeneous evolutionary history (Antonelli et al. 2017). This will be implemented in future datelife versions.

In other areas of biological research, such as ecology and conservation biology, it has been indicated that at
least some data on lineage divergence represents a relevant improvement for testing alternative hypothesis
using phylogenetic distance. Hence, we allow accepted ways of creating branch lengths in the absence
of starting branch length information (such as BLADJ (Webb et al. 2008)) for several taxa lacking this

information. Making up branch lengths in this or other ways is accepted in scientific publications: Rabosky et al. (2018) created a time-calibrated tree of 31,536 ray-finned fishes, of which only 37% had molecular data; Jetz et al. (2012), created a time-calibrated tree of all 9,993 bird species, where 67% had molecular data; Smith and Brown (2018) constructed a tree of 353,185 seed plants where only 23% had molecular data. Taken to the extreme, one could make a fully resolved, calibrated tree of all modern and extinct taxa using a single taxonomy and a single calibration with the polytomy resolution and branch imputation methods. There has yet to be a thorough analysis of what can go wrong when one goes beyond the data in this way, so we urge caution; we also urge readers to follow the example of many of the large tree papers cited above and make sure results are substantially similar between trees containing only taxa with molecular or other data and trees that combine those taxa with taxa from taxonomy alone.

245 CONCLUSIONS

Divergence time information is key to many areas of evolutionary studies: trait evolution, diversification,
biogeography, macroecology and more. Generating this information is difficult, especially for those who want
to use phylogenies but who are not systematists, or do not have the time to acquire and develop the necessary
knowledge and data curation skills to produce chronograms de novo. Knowledge on clade ages is also crucial
for science communication and education.

datelife allows an easy and fast obtention, as well as comparison of publicly available information on time
of lineage divergence, providing a straightforward way to get an informed idea on the state of knowledge
of the time frame of evolution of different regions of the tree of life, allowing identification of regions that
require more research or that have conflicting information. Both summary and newly generated trees are
potentially useful to evaluate evolutionary hypothesis in different areas of research. datelife helps with
awereness on the existing variation in expert time of divergence data, and might foster exploration of the
effect of alternative divergence time hypothesis on the results of analyses, nurturing a culture of more cautious
interpretation of evolutionary results.

AVAILABILITY

datelife is free and open source and it can be used through its current website http://www.datelife.org/
query/, through its R package, and through Phylotastic's project web portal http://phylo.cs.nmsu.edu:3000/.
datelife's website is maintained using RStudio's shiny server and the shiny package open infrastructure, as well as Docker. datelife's R package stable version will be available for installation from the CRAN repository (https://cran.r-project.org/package=datelife) using the command
install.packages(pkgs = "datelife") from within R. Development versions are available from the
GitHub repository (https://github.com/phylotastic/datelife) and can be installed using the command
devtools::install_github("phylotastic/datelife").

SUPPLEMENTARY MATERIAL

Code used to generate all versions of this manuscript, the biological examples, as well as the benchmark of functionalities are respectively in the datelife_paper1, datelife_examples, and datelife_benchmark repositories in LLSR GitHub account.

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285 REFERENCES

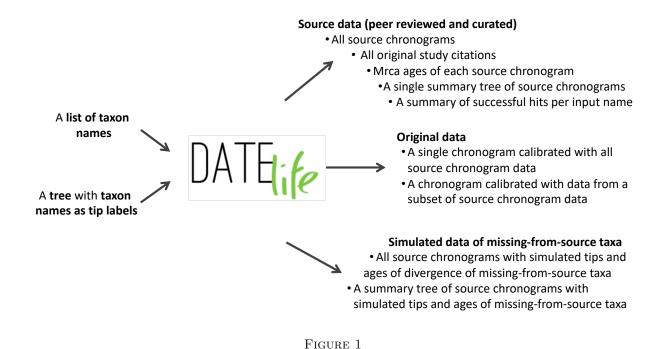
- ²⁸⁶ Antonelli A., Hettling H., Condamine F.L., Vos K., Nilsson R.H., Sanderson M.J., Sauquet H., Scharn
- 287 R., Silvestro D., Töpel M., Bacon C.D., Oxelman B., Vos R.A. 2017. Toward a self-updating platform for
- estimating rates of speciation and migration, ages, and relationships of Taxa. Systematic Biology. 66:153–166.
- ²⁸⁹ Bapst D.W. 2012. Paleotree: An R package for paleontological and phylogenetic analyses of evolution.
- 290 Methods in Ecology and Evolution. 3:803–807.
- ²⁹¹ Barba-Montoya J., Reis M. dos, Schneider H., Donoghue P.C., Yang Z. 2018. Constraining uncertainty in the
- timescale of angiosperm evolution and the veracity of a cretaceous terrestrial revolution. New Phytologist.
- 293 218:819-834.
- Barker F.K., Burns K.J., Klicka J., Lanyon S.M., Lovette I.J. 2012. Going to extremes: Contrasting rates of
- diversification in a recent radiation of new world passerine birds. Systematic biology. 62:298–320.
- Barker F.K., Burns K.J., Klicka J., Lanyon S.M., Lovette I.J. 2015. New insights into new world biogeography:
- ²⁹⁷ An integrated view from the phylogeny of blackbirds, cardinals, sparrows, tanagers, warblers, and allies. The
- ²⁹⁸ Auk: Ornithological Advances. 132:333–348.
- Boyle B., Hopkins N., Lu Z., Raygoza Garay J.A., Mozzherin D., Rees T., Matasci N., Narro M.L., Piel
- W.H., Mckay S.J., Lowry S., Freeland C., Peet R.K., Enquist B.J. 2013. The taxonomic name resolution
- ₃₀₁ service: An online tool for automated standardization of plant names. BMC Bioinformatics. 14.
- Britton T., Anderson C.L., Jacquet D., Lundqvist S., Bremer K. 2007. Estimating Divergence Times in Large
- Phylogenetic Trees. Systematic Biology. 56:741–752.
- Burns K.J., Shultz A.J., Title P.O., Mason N.A., Barker F.K., Klicka J., Lanyon S.M., Lovette I.J. 2014.
- ³⁰⁵ Phylogenetics and diversification of tanagers (passeriformes: Thraupidae), the largest radiation of neotropical
- songbirds. Molecular Phylogenetics and Evolution. 75:41–77.
- Chamberlain S. 2018. bold: Interface to Bold Systems API..

- ³⁰⁸ Chamberlain S.A., Szöcs E. 2013. taxize: taxonomic search and retrieval in R [version 2; referees: 3 approved].
- 309 F1000Research. 2:1-29.
- 310 Claramunt S., Cracraft J. 2015. A new time tree reveals earth history's imprint on the evolution of modern
- birds. Science advances. 1:e1501005.
- ³¹² Criscuolo A., Berry V., Douzery E.J., Gascuel O. 2006. SDM: A fast distance-based approach for (super)tree
- building in phylogenomics. Systematic Biology. 55:740–755.
- Eastman J.M., Harmon L.J., Tank D.C. 2013. Congruification: Support for time scaling large phylogenetic
- trees. Methods in Ecology and Evolution. 4:688–691.
- Felsenstein J. 1985. Phylogenies and the Comparative Method. The American Naturalist. 125:1–15.
- Gibb G.C., England R., Hartig G., McLenachan P.A., Taylor Smith B.L., McComish B.J., Cooper A., Penny
- 318 D. 2015. New zealand passerines help clarify the diversification of major songbird lineages during the oligocene.
- Genome biology and evolution. 7:2983–2995.
- Harmon L., Weir J., Brock C., Glor R., Challenger W. 2008. GEIGER: investigating evolutionary radiations.
- 321 Bioinformatics. 24:129–131.
- Hedges S.B., Dudley J., Kumar S. 2006. TimeTree: A public knowledge-base of divergence times among
- organisms. Bioinformatics. 22:2971–2972.
- Hedges S.B., Marin J., Suleski M., Paymer M., Kumar S. 2015. Tree of life reveals clock-like speciation and
- diversification. Molecular Biology and Evolution. 32:835–845.
- ₃₂₆ Heibl C. 2008. PHYLOCH: R language tree plotting tools and interfaces to diverse phylogenetic software
- packages..
- Hinchliff C.E., Smith S.A., Allman J.F., Burleigh J.G., Chaudhary R., Coghill L.M., Crandall K.A., Deng J.,
- Drew B.T., Gazis R., Gude K., Hibbett D.S., Katz L.A., Laughinghouse H.D., McTavish E.J., Midford P.E.,

- owen C.L., Ree R.H., Rees J.A., Soltis D.E., Williams T., Cranston K.A. 2015. Synthesis of phylogeny and
- taxonomy into a comprehensive tree of life. Proceedings of the National Academy of Sciences. 112:12764–12769.
- Hooper D.M., Price T.D. 2017. Chromosomal inversion differences correlate with range overlap in passerine
- birds. Nature ecology & evolution. 1:1526.
- Huelsenbeck J.P., Ronquist F. 2001. MRBAYES: Bayesian inference of phylogenetic trees. Bioinformatics.
- 17:754-755.
- Jetz W., Thomas G., Joy J.J., Hartmann K., Mooers A. 2012. The global diversity of birds in space and
- 337 time. Nature. 491:444-448.
- ³³⁸ Kumar S., Stecher G., Suleski M., Hedges S.B. 2017. TimeTree: A Resource for Timelines, Timetrees, and
- Divergence Times. Molecular biology and evolution. 34:1812–1819.
- Magallón S., Gómez-Acevedo S., Sánchez-Reves L.L., Hernández-Hernández T. 2015. A metacalibrated
- time-tree documents the early rise of flowering plant phylogenetic diversity. New Phytologist. 207:437–453.
- Michonneau F., Brown J.W., Winter D.J. 2016. rotl: an R package to interact with the Open Tree of Life
- data. Methods in Ecology and Evolution. 7:1476–1481.
- Morell V. 1996. The roots of phylogeny. Science. 273:569.
- Morlon H. 2014. Phylogenetic approaches for studying diversification. Ecology Letters. 17:508–525.
- O'Meara B., Md Tayeen A.S., Sanchez Reyes L.L. 2019. Rphylotastic: An r interface to 'phylotastic' web
- services..
- Ooms J., Chamberlain S. 2018. Phylocomr: Interface to 'phylocom'..
- Paradis E., Claude J., Strimmer K. 2004. APE: analyses of phylogenetics and evolution in R language.
- 350 Bioinformatics. 20:289–290.

- Pearse W.D., Purvis A. 2013. PhyloGenerator: An automated phylogeny generation tool for ecologists.
- Methods in Ecology and Evolution. 4:692–698.
- ³⁵³ Piel W.H., Donoghue M., Sanderson M. 2002. TreeBASE: A database of phylogenetic information. In:
- Shimura J., Wilson K., Gordon D., editors. To the interoperable "catalog of life" with partners. Tsukuba,
- Japan: National Institute for Environmental Studies. p. 41–47.
- Posadas P., Crisci J.V., Katinas L. 2006. Historical biogeography: A review of its basic concepts and critical
- issues. Journal of Arid Environments. 66:389–403.
- ³⁵⁸ Price T.D., Hooper D.M., Buchanan C.D., Johansson U.S., Tietze D.T., Alström P., Olsson U., Ghosh-Harihar
- M., Ishtiaq F., Gupta S.K., others. 2014. Niche filling slows the diversification of himalayan songbirds.
- 360 Nature. 509:222.
- Rabosky D.L., Chang J., Title P.O., Cowman P.F., Sallan L., Friedman M., Kaschner K., Garilao C., Near
- 362 T.J., Coll M., others. 2018. An inverse latitudinal gradient in speciation rate for marine fishes. Nature.
- 363 559:392.
- Ramshaw J., Richardson D., Meatyard B., Brown R., Richardson M., Thompson E., Boulter D. 1972. The
- time of origin of the flowering plants determined by using amino acid sequence data of cytochrome c. New
- ³⁶⁶ Phytologist. 71:773–779.
- R Core Team. 2018. R: a language and environment for statistical computing. Vienna, Austria: R Foundation
- 368 for Statistical Computing.
- Revell L.J. 2012. Phytools: An r package for phylogenetic comparative biology (and other things). Methods
- in Ecology and Evolution. 3:217–223.
- Ronquist F., Huelsenbeck J.P. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models.
- 372 Bioinformatics. 19:1572–1574.
- ₃₇₃ Sanderson M.J., Doyle J.A. 2001. Sources of error and confidence intervals in estimating the age of angiosperms

- from rbcL and 18S rDNA data. American Journal of Botany. 88:1499–1516.
- 375 Schenk J.J. 2016. Consequences of secondary calibrations on divergence time estimates. PLoS ONE. 11.
- Smith S.A., Brown J.W. 2018. Constructing a broadly inclusive seed plant phylogeny. American Journal of
- 377 Botany. 105:302-314.
- Stoltzfus A., Lapp H., Matasci N., Deus H., Sidlauskas B., Zmasek C.M., Vaidya G., Pontelli E., Cranston
- K., Vos R., Webb C.O., Harmon L.J., Pirrung M., O'Meara B., Pennell M.W., Mirarab S., Rosenberg M.S.,
- Balhoff J.P., Bik H.M., Heath T.A., Midford P.E., Brown J.W., McTavish E.J., Sukumaran J., Westneat M.,
- Alfaro M.E., Steele A., Jordan G. 2013. Phylotastic! Making tree-of-life knowledge accessible, reusable and
- 382 convenient. BMC Bioinformatics. 14.
- Title P.O., Rabosky D.L. 2016. Do Macrophylogenies Yield Stable Macroevolutionary Inferences? An
- Example from Squamate Reptiles. Systematic Biology.:syw102.
- Webb C.O. 2000. Exploring the Phylogenetic Structure of Ecological Communities: An Example for Rain
- Forest Trees. The American Naturalist. 156:145–155.
- Webb C.O., Ackerly D.D., Kembel S.W. 2008. Phylocom: Software for the analysis of phylogenetic community
- structure and trait evolution. Bioinformatics. 24:2098–2100.
- Webb C.O., Donoghue M.J. 2005. Phylomatic: Tree assembly for applied phylogenetics. Molecular Ecology
- 390 Notes. 5:181–183.



Stylized DateLife workflow. This shows the general worflows and analyses that can be performed with

datelife, via the R package or through the website. Details on the functions involved on each workflow are

shown in datelife's R package vignette.

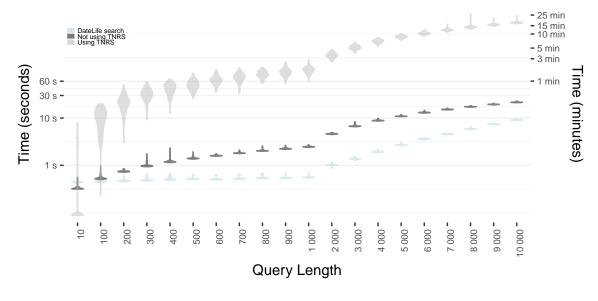


Figure 2

Computation time of query processing and search across datelife's chronogram database relative to number
of input taxon names. We sampled N names from the class Aves for each cohort 100 times and then performed
a search with query processing not using the Taxon Names Resoultion Service (TNRS; dark gray), and using
TNRS (light gray). We also performed a search using the already processed query for comparison (light blue).

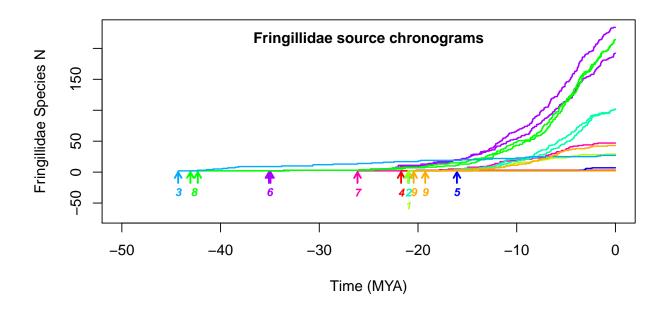


FIGURE 3

Lineage through time (LTT) plots of source chronograms containing all or a subset of species from the bird family Fringillidae of true finches. Arrows indicate maximum age of each chronogram. Numbers reference to chronograms' original publications 1: Barker et al. (2012), 2: Barker et al. (2015), 3: Burns et al. (2014), 4: Claramunt and Cracraft (2015), 5: Gibb et al. (2015), 6: Hedges et al. (2015), 7: Hooper and Price (2017), 8: Jetz et al. (2012), 9: Price et al. (2014).

- Fringillidae source chronogramsFringillidae summary chronograms

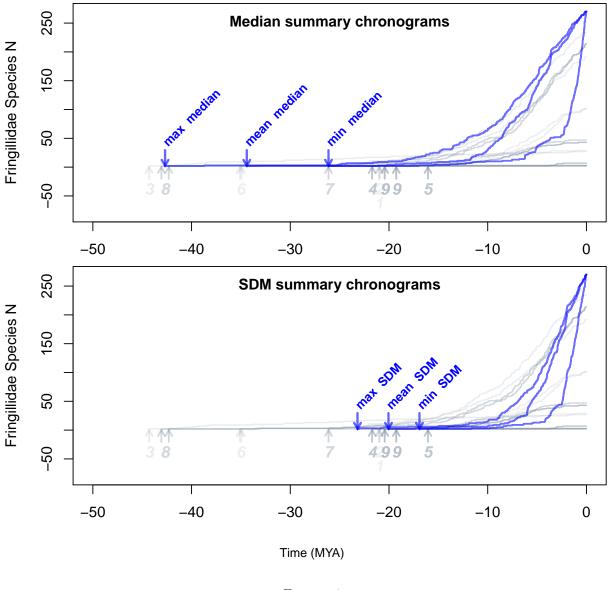


Figure 4

- LTT plots of median (top) and Supermatrix Distance Method (SDM; bottom) chronograms summarising
- information from source chronograms found for the Fringillidae. Arrows indicate tree maximum age.

- Fringillidae source chronograms used as calibrations
 Fringillidae source chronograms used only as topology
 Fringillidae new chronograms generated with BLADJ

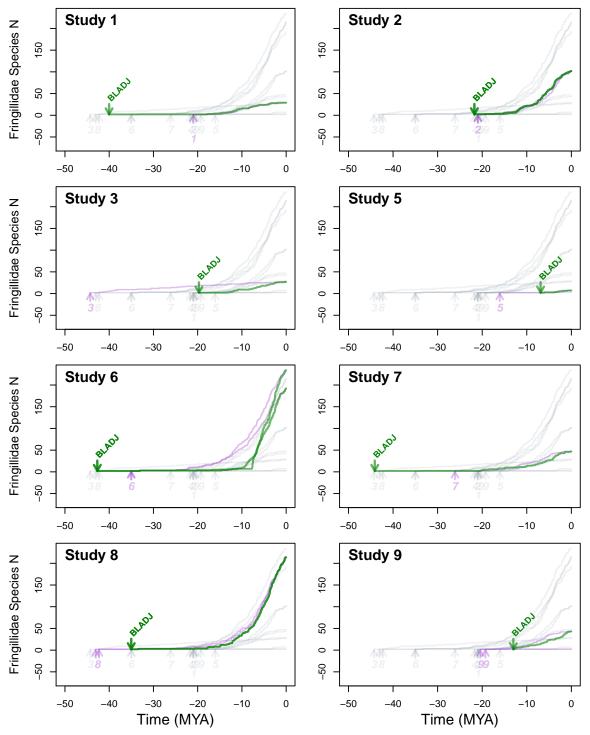


FIGURE 5

- $_{405}$ LTT plots showing results from the cross-validation analyses of trees without branch lengths dated using
- ⁴⁰⁶ BLADJ. The dating analysis can only be performed in trees with more than 2 tips, thus excluding chronogram
- from study 4; its data was still used as calibration for the other source chronograms.

- Fringillidae source chronograms used as calibrations
- Fringillidae source chronograms used only as topology
- Fringillidae new chronograms generated with PATHd8

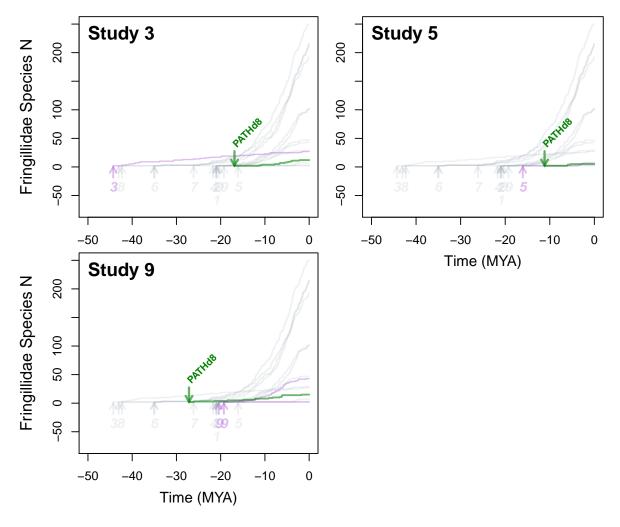


Figure 6

LTT plots showing results from the cross-validation analyses of trees with branch length reconstructed with
data from the Barcode of Life Database (BOLD) dated using PATHd8. We could construct a tree with
branch lengths for all source chronograms. However, dating with PATHd8 was only successful in three source
chronograms shown here.