

# User manual for the signatureFit command line interface

signature.tools.lib version: 2.3.0

latest edit: 25/01/2023

Andrea Degasperi, University of Cambridge, UK  
ad923@cam.ac.uk

## 1. Introduction

Mutational signature fit analysis attempts to identify the presence of a given set of mutational signatures in the somatic mutations of a cancer sample.

This document describes how to use the `signatureFit` command line script, which is a wrapper for the `signatureFit_pipeline` function in the `signature.tools.lib` R package, which in turn is an interface for the `Fit` and `FitMS` mutational signature fit analysis functions.

The `signatureFit_pipeline` function is a flexible interface for mutational signature fit analysis. Users can provide mutation calls as input or pre-built mutational catalogues, and then they can perform either an automated analysis, with very few options required such as the organ of origin of the sample, or use the options to perform a more tailored analysis.

## 2. Installation

The script `signatureFit` is included in the `signature.tools.lib` R package. Thus, in order to use it, one is required to install `signature.tools.lib`, which is available on GitHub:

<https://github.com/Nik-Zainal-Group/signature.tools.lib>

After the installation of `signature.tools.lib`, one can run the `signatureFit` script, which is located in the `scripts` folder in the github repository. For easy access, add a copy of, or a symbolic link to, the `signatureFit` script to a location in your command line `PATH`.

## 3. signatureFit options

The list of available options can be accessed by typing:

```
signatureFit --help
```

This is the current output:

This script runs the signature fit pipeline of the R package `signatures.tools.lib`, using the `Fit` or `FitMS` functions.

Run this script as follows:

```
signatureFit [OPTIONS]
```

Available options:

<code>-o, --outdir=DIR</code>	Name of the output directory. If omitted a name will be given automatically.
<code>-b, --bootstrap</code>	Request signature fit with bootstrap
<code>-c, --cataloguesfile=CFIL</code>	

CFILE is the name of a file containing mutational catalogues. Each sample catalogue is in a column, with sample names as column headers and channel names as row names in the first column with no header. Can be omitted if mutations are provided.

-x, --snvvcf=SNVVCF SNVVCF is a tab separated file containing two columns. The first column contains the sample names, while the second column contains the corresponding SNV vcf file names.

-X, --snvtab=SNVTAB SNVTAB is a tab separated file containing two columns. The first column contains the sample names, while the second column contains the corresponding SNV tab file names. Each SNV tab file should have a header with the following columns: chr, position, REF, ALT.

-y, --dnvvcf=DNVVCF DNVVCF is a tab separated file containing two columns. The first column contains the sample names, while the second column contains the corresponding DNV vcf file names. VCF files can contain both DNVs and SNVs and if two SNVs are adjacent they will be merged into additional DNVs.

-Y, --dnvtab=DNVTAB DNVTAB is a tab separated file containing two columns. The first column contains the sample names, while the second column contains the corresponding DNV tab file names. Each DNV tab file should have a header with the following columns: chr, position, REF, ALT. Tab files can contain both DNVs and SNVs and if two SNVs are adjacent they will be merged into additional DNVs.

-z, --svbedpe=SVBEDPE SVBEDPE is a tab separated file containing two columns. The first column contains the sample names, while the second column contains the corresponding BEDPE file names. Each BEDPE file is a tab separated file with header: chrom1, start1, end1, chrom2, start2, end2, and sample. In addition, another column with header svclass should indicate the type of SV: translocation, inversion, deletion, or tandem-duplication.

-w, --signaturesfile=SFILE SFILE is the name of a file containing mutational signatures. Each signature is in a column, with signature names as column headers and channel names as row names in the first column with no header. Each column must sum to 1. Use only to provide your own signatures. When fitmethod=FitMS, these signatures are considered common signatures.

-W, --raresignaturesfile=RSFILE RSFILE is the name of a file containing mutational signatures. Each signature is in a column, with signature names as column headers and channel names as row names in the first column with no header. Each column must sum to 1. Use only to provide your own signatures. When fitmethod=FitMS, these signatures are considered rare signatures.

-s, --sigversion=SIGVERSION Either COSMICv2, COSMICv3.2, RefSigv1 or RefSigv2. If not specified SIGVERSION=RefSigv2.

-O, --organ=ORGAN When using RefSigv1 or RefSigv2 as SIGVERSION, organ-specific signatures will be used. If SIGVERSION is COSMICv2 or COSMICv3.2, then a selection of signatures found in the given organ will be used. Available organs depend on the selected SIGVERSION. For RefSigv1 or RefSigv2: Biliary, Bladder, Bone\_SoftTissue, Breast, Cervix (v1 only), CNS, Colorectal, Esophagus, Head\_neck, Kidney, Liver, Lung, Lymphoid, NET (v2 only), Oral\_Oropharyngeal (v2 only), Ovary, Pancreas, Prostate, Skin, Stomach, Uterus.

-l, --signames=SIGNAMES If no ORGAN is specified, SIGNAMES can be used to provide a comma separated list of signature names

to select from the COSMIC or reference signatures, depending on the SIGVERSION requested. For example, for COSMICv3.2 use: SBS1,SBS2,SBS3.

-e, --genomev=GENOMEV Genome version to be used: hg19, hg38 or mm10.  
If not specified GENOMEV=hg19.

-m, --fitmethod=FITMETHOD Either Fit or FitMS. If not specified FITMETHOD=FitMS

-M, --optmethod=OPTMETHOD Optimisation objective function, either KLD or NNLS.  
If not specified OPTMETHOD=KLD.

-t, --filtertype=FTYPE FTYPE is either fixedThreshold or giniScaledThreshold.  
When using fixedThreshold, exposures will be removed based on a fixed percentage with respect to the total number of mutations (THRPERC will be used). When using giniScaledThreshold each signature will use a different threshold calculated as  $(1 - \text{Gini}(\text{signature})) * \text{GINISCALING}$ . If not specified then FTYPE=fixedThreshold

-p, --thresholdperc=THRPERC THRPERC is a threshold in percentage of total mutations in a sample, only exposures larger than THRPERC are considered. If not specified THRPERC=5.

-d, --giniscaling=GINISCALING GINISCALING is a scaling factor for the threshold type giniScaledThreshold, which is based on the Gini score of a signature. If not specified GINISCALING=10.

-u, --thresholdpval=THRPVAL THRPVAL is a p-value to determine whether an exposure is above the THRPERC or the threshold calculated with Gini scaling, when using bootstrap. In other words, this is the empirical probability that the exposure is lower than the threshold. If not specified then THRPVAL=0.05.

-a, --fitmsmode=FMSMODE FMSMODE is either constrainedFit, partialNMF, errorReduction, or cossimIncrease. If not specified FMSMODE=errorReduction.

-q, --commonsigtier=CSTIER CSTIER is either T1, T2 or T3. For each organ, T1 indicates to use the common organ-specific signatures, while T2 indicates to use the corresponding reference signatures. In general, T1 should be more appropriate for organs where there are no mixed organ-specific signatures, e.g. SBS1+18 or SBS2+13, while T2 might be more suitable for when such mixed signatures are present, so that each signature can be fitted, e.g. fitting the two signatures SBS1 and SBS18, instead of a single SBS1+18. T3 is a combination of T1 and T2, where only the mixed organ signatures are replaced with the corresponding reference signatures. If not specified CSTIER=T1.

-T, --raresigtier=RTIER RTIER is either T0, T1, T2, T3 or T4. For each organ, T0 are rare signatures that were observed in the requested organ, including low quality signatures (QC amber and red signatures). T1 are high quality (QC green) rare signatures that were observed in the requested organ. T2-T4 signatures extend the rare signatures set to what has been observed also in other organs. T2 includes all QC green signatures that were classified as rare at least twice (SBS only) in Degasperis et al. 2022 Science. T3 includes all QC green signatures (if not SBS, T3=T2). T4 includes all signatures including QC amber and red. In general we advise to use T2 signatures. If not specified RTIER=T2.

-i, --residualnegprop=RNP RNP is the maximum proportion of mutations (w.r.t. total mutations in a sample) that can be in the negative part of a residual when using the constrained least squares fit when fitMS mode is FMSMODE=constrainedFit. If not specified then RNP=0.003.

-R, --minresidualmut=MINRM MINRM is the minimum number of mutations in a residual when FMSMODE=constrainedFit or FMSMODE=partialNMF.

Deactivated by default (MINRM=NULL).

-C, --mincossimraresigs=MINCSRS  
MINCSRS is the minimum cosine similarity between a residual and a rare signature for considering the rare signature as a candidate in a sample when FMSMODE=constrainedFit or FMSMODE=partialNMF. If not specified, MINCSRS=0.8.

-E, --minerrorredperc=MINERPERC  
MINERPERC is the minimum percentage of error reduction for a rare signature to be considered as candidate in a sample when FMSMODE=errorReduction. The error is computed as mean absolute deviation. If not specified MINERPERC=15.

-I, --mincossimincr=MINCSINCR  
MINCSINCR is the minimum cosine similarity increase for a rare signature to be considered as candidate in a sample when FMSMODE=cossimIncrease. If not specified MINCSINCR=0.02.

-k, --maxraresigs=MAXRS  
MAXRS is the maximum number of rare signatures that are allowed to be present in each sample. If not specified MAXRS=1.

-S, --rarecandidatesel=RARESEL  
RARESEL is either MaxCosSim or MinError. This is a FitMS parameter. Whenever there is more than one rare signature candidate for a sample, then the best candidate rare signature is automatically selected using the RARESEL criteria. Error is computed as the mean absolute deviation of channels. allowed to be present in each sample. If not specified RARESEL=MaxCosSim.

-n, --nparallel=NPARALLEL  
Number of parallel CPUs to be used.

-f, --nboot=NBOOT  
Number of bootstrap to be used when bootstrap is requested (-b), if not specified, NBOOT=200.

-A, --writeannotations  
Write the annotated mutations to file. This is off by default, because annotated mutations are saved as a flat text file containing mutations from all samples and may occupy a lot of space if many samples and many mutations are processed.

-J, --writejson  
Write the signature fit object to a JSON file, which will save all fit results and options used.

-r, --randomSeed=SEED  
Specify a random seed to obtain always the same identical results.

-v, --verbose  
Verbose option for additional output.

-h, --help  
Show this explanation.

## 4. Using the signatureFit command line interface

### 4.1 Mutational catalogues

A mutational catalogue is a vector that contains counts of specific classes (channels) of mutations, which depend on the type of mutations considered. For example, SNV mutations will use the 96-channel catalogue type, which contain substitution counts in their trinucleotide context (e.g., A[C>T]A).

Using the signatureFit script, it is possible to provide a mutational catalogue directly, or, alternatively, provide a list of mutation files that will be automatically converted into catalogues. Only one list of mutation files of one single type of mutations can be provided, e.g., a list of vcf files containing SNVs can be provided using the --snvvcf option. If a catalogue is provided directly (option --cataloguesfile), the mutation type will be inferred by the row names, which are the catalogue channels.

It is also possible to provide both a catalogues file and a list of mutation files. Provided that the type of mutations is the same, the new catalogues built from the mutation files will be added to the catalogues provided.

## 4.2 Mutational signatures

The `signatureFit` script can use different strategies to estimate how many mutations in a catalogue are associated with a certain mutational signature. To do so, it requires an *a priori* set of mutational signatures to use.

With `signatureFit`, it is possible to provide files containing the mutational signatures to use, or use available options to select a set of signatures automatically. If mutational signatures are provided manually, this will have the priority, and the options used to select signatures automatically, such as `--organ`, `--sigversion` and `--signames`, will be ignored.

Depending on the fit method, one or two sets of signatures is required. If `--fitmethod=Fit`, then just one set of signatures will be used, which can be specified manually with `--signaturesfile`, while if `--fitmethod=FitMS`, then two sets of signatures are required, one set of common signatures (`--signaturesfile`) and one of rare signatures (`--raresignaturesfile`).

If no signatures have been provided manually, then `signatureFit` will check if the `--organ` option has been used, and select the appropriate signatures based on the `--sigversion` and `--fitmethod` options, as well as the inferred mutation type.

If no signatures have been provided manually and also no organ was specified, then `signatureFit` will select signatures based on the `--sigversion` and `--signames` parameters and the mutation type. For example, one could use `--sigversion=COSMICv3.2` and `--signames=SBS1,SBS2,SBS3` to fit only three COSMIC v3.2 signatures into a catalogue, provided it is an SNV catalogue.

When using `--sigversion=RefSigv1` or `--sigversion=RefSigv2`, `signatureFit` will check the `--organ` option to select organ-specific mutational signatures. If the organ is not specified, then `signatureFit` will use the reference signatures instead, which are obtained as average of multiple similar organ-specific signatures.

## 5. Examples

### 5.1 FitMS example

In this example we run a signature fit analysis with `FitMS`, using `vcf` single nucleotide variant (SNV) files as input, assuming that the `vcf` are obtained from breast cancer samples.

```
signatureFit --organ Breast -b -o outfolder -x snvvcf.tsv
```

Note that `FitMS` is the default fit method, so there is no need to specify `--fitmethod=FitMS`. `FitMS` will use the latest `RefSigv2` signatures, which include the common and rare SBS signatures identified in the analysis of the Genomics England WGS cancer dataset. The flag `-b` requests a bootstrap analysis, `-o` indicates the output folder, and `-x` indicates the location of a tab separated file containing a list of sample names and corresponding `vcf` locations. The content of `snvvcf.tsv` could be as follows:

```
Sample1    sample1_snv.vcf
```

```
Sample2    sample2_snv.vcf
Sample3    sample3_snv.vcf
...
```

Finally, note that all the mutations in the input vcf files will be used, so they should already be filtered, e.g. containing only PASS variants.

## 5.2 Fitting COSMIC v3.2 signatures

In this example we show how to fit a specific set of SBS COSMIC signatures from the COSMIC v3.2 set.

```
signatureFit -b -o outfolder -x snvvcf.tsv -s COSMICv3.2 -m Fit -l
SBS1,SBS2,SBS3,SBS5,SBS8,SBS13,SBS17a,SBS17b,SBS18
```

In this case, we did not specify an organ, but rather used the `-s` option to request the use of COSMIC v3.2 signatures and the `-l` option to supply a list of signatures to use. We also specified to use the Fit algorithm with the option `-m`.

## 5.3 Fitting organ-specific rearrangement signatures

In this example, we fit organ-specific ovarian cancer rearrangement signatures, from the Degasperi et al 2020 Nature Cancer paper.

```
signatureFit --organ Ovary -b -o outfolder -s RefSigv1 -m Fit -z svbedpe.tsv
```

In this case, we provided a list of bedpe files using the option `-z`. The content of `svbedpe` could be as follows:

```
Sample1    sample1_sv.bedpe
Sample2    sample2_sv.bedpe
Sample3    sample3_sv.bedpe
...
```

Note that we specified the option `-s RefSigv1`, because the latest organ-specific rearrangement signatures belong to this signature version, although, this could have been omitted, implying the default option `-s RefSigv2`, and `signatureFit` would have switched to `-s RefSigv1` automatically with a warning message, so that the latest organ-specific rearrangement signatures available could be selected.

When some of the organ-specific signatures are mixed, one might prefer to replace the mixed organ-specific signatures with the corresponding reference signatures. For example, replace `Ovary_B` (RefSig R6a+R6b) with the signatures RefSig R6a and RefSig R6b. This can be done automatically using the `signatureFit` parameter `commonsigtier`. Selecting common signature tier T3 will replace only the mixed signatures, while T2 will replace all the organ-specific signatures with the corresponding reference signatures. For example:

```
signatureFit --organ Ovary -b -o outfolder --commonsigtier T2 -z svbedpe.tsv
```

Notice that the above command will issue some warnings, informing the user that the default options for signature version and fitting method have been changed to RefSigv1 and Fit respectively.