

R Exercise

RS-eco

09.04.2019

1. Piping

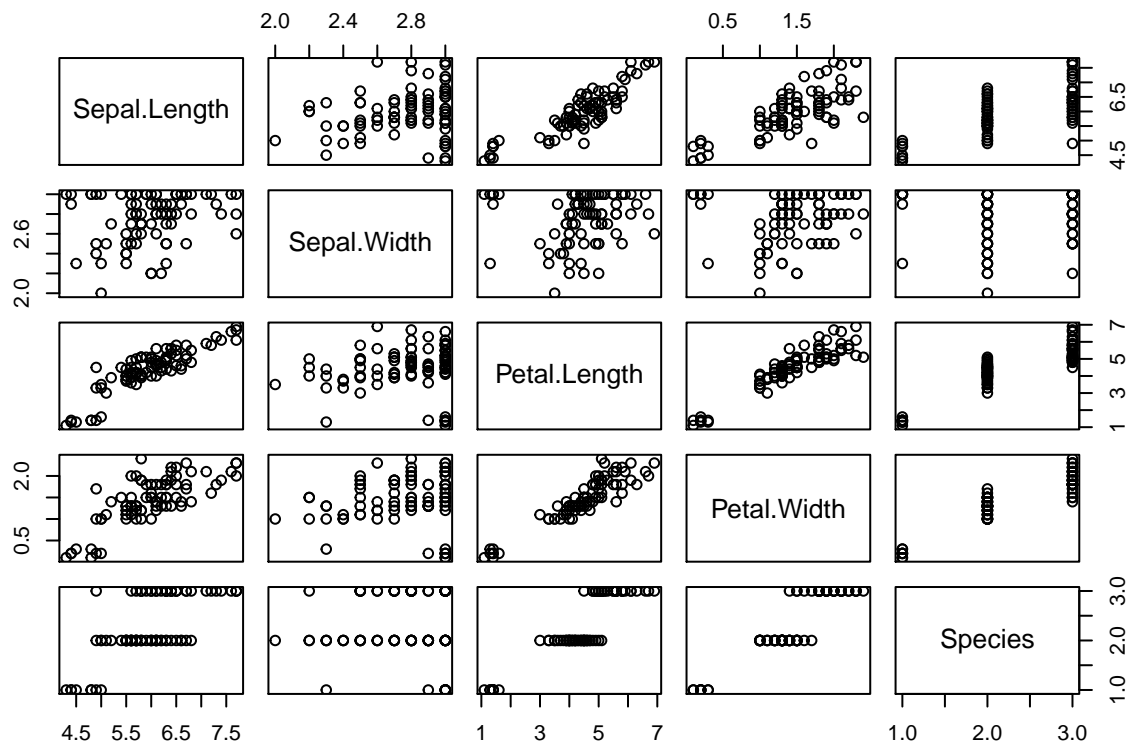
Install and load the magrittr package

```
#install.packages("magrittr", dep=T)
library(magrittr)
```

Turn the following code into a streamlined version using piping:

```
# Without piping
mean_width <- mean(iris$Sepal.Width)
iris_sub <- subset(iris, Sepal.Width < mean_width)
plot(iris_sub)
cor_iris <- cor(iris_sub$Sepal.Length,
               iris_sub$Sepal.Width)

# With piping
mean_width <- iris %>% mean(Sepal.Width)
iris %>%
  subset(Sepal.Width < mean_width) %>%
  plot()
```



```
cor_iris <- iris %>%
  subset(Sepal.Width < mean(Sepal.Width)) %>%
  plot %>% cor(Sepal.Length, Sepal.Width)
```

```
#####
```

2. Read data

Install and load the readr package

```
#install.packages(readr, dep=T)
library(readr)
```

Set working directory

```
#setwd("")
```

Read species_info.csv.xz into a data.frame called species_info Note: species_info contains information on more than 40000 vertebrate species

```
species_info <- read_csv("data/species_info.csv.xz")
library(readr)
species_info <- read_csv("data/species_info.csv.xz")
```

```
## Parsed with column specification:
## cols(
##   binomial = col_character(),
##   presence = col_double(),
##   origin = col_double(),
##   seasonal = col_double(),
##   kingdom_na = col_character(),
##   phylum_nam = col_character(),
##   class_name = col_character(),
##   order_name = col_character(),
##   family_nam = col_character(),
##   shape_Area = col_double()
## )
```

Check class and structure of species_info

```
class(species_info)
```

```
## [1] "spec_tbl_df" "tbl_df"      "tbl"        "data.frame"
```

```
str(species_info)
```

```
## Classes 'spec_tbl_df', 'tbl_df', 'tbl' and 'data.frame': 40584 obs. of  10 variables:
## $ binomial : chr "Abavorana luctuosa" "Acanthixalus sonjae" "Acanthixalus spinosus" "Acris crepitans" ...
## $ presence : num 1 1 1 1 1 1 1 1 1 1 ...
## $ origin : num 1 1 1 1 1 1 1 1 1 1 ...
## $ seasonal : num 1 1 1 1 1 1 1 1 1 1 ...
## $ kingdom_na: chr "ANIMALIA" "ANIMALIA" "ANIMALIA" "ANIMALIA" ...
## $ phylum_nam: chr "CHORDATA" "CHORDATA" "CHORDATA" "CHORDATA" ...
## $ class_name: chr "AMPHIBIA" "AMPHIBIA" "AMPHIBIA" "AMPHIBIA" ...
## $ order_name: chr "ANURA" "ANURA" "ANURA" "ANURA" ...
## $ family_nam: chr "RANIDAE" "HYPEROLIIDAE" "HYPEROLIIDAE" "HYLIDAE" ...
## $ shape_Area: num 9.399 0.465 98.687 322.578 63.883 ...
## - attr(*, "spec")=
## .. cols(
## .. binomial = col_character(),
## .. presence = col_double(),
```

```
## .. origin = col_double(),
## .. seasonal = col_double(),
## .. kingdom_na = col_character(),
## .. phylum_nam = col_character(),
## .. class_name = col_character(),
## .. order_name = col_character(),
## .. family_nam = col_character(),
## .. shape_Area = col_double()
## .. )
```

Read first 10 entries of species_info.csv.xz

```
species_info_first10 <- read_csv("data/species_info.csv.xz", n_max=10)
```

```
## Parsed with column specification:
## cols(
##   binomial = col_character(),
##   presence = col_double(),
##   origin = col_double(),
##   seasonal = col_double(),
##   kingdom_na = col_character(),
##   phylum_nam = col_character(),
##   class_name = col_character(),
##   order_name = col_character(),
##   family_nam = col_character(),
##   shape_Area = col_double()
## )
```

Read last 10 lines of species_info.csv.xz

```
col_sp_info <- colnames(read_csv("data/species_info.csv.xz",
                                n_max=1))
```

```
## Parsed with column specification:
## cols(
##   binomial = col_character(),
##   presence = col_double(),
##   origin = col_double(),
##   seasonal = col_double(),
##   kingdom_na = col_character(),
##   phylum_nam = col_character(),
##   class_name = col_character(),
##   order_name = col_character(),
##   family_nam = col_character(),
##   shape_Area = col_double()
## )
```

```
species_info_last10 <- read_csv("data/species_info.csv.xz",
                                col_names=col_sp_info,
                                skip=40574)
```

```
## Parsed with column specification:
## cols(
##   binomial = col_character(),
##   presence = col_logical(),
##   origin = col_logical(),
##   seasonal = col_logical(),
```

```
## kingdom_na = col_character(),
## phylum_nam = col_character(),
## class_name = col_character(),
## order_name = col_logical(),
## family_nam = col_logical(),
## shape_Area = col_double()
## )

# Skip the first 10 and skip the last 10 lines
species_info_other <- read_csv("data/species_info.csv.xz",
                              col_names=col_sp_info,
                              skip=10, n_max=40564)
```

```
## Parsed with column specification:
## cols(
##   binomial = col_character(),
##   presence = col_double(),
##   origin = col_double(),
##   seasonal = col_double(),
##   kingdom_na = col_character(),
##   phylum_nam = col_character(),
##   class_name = col_character(),
##   order_name = col_character(),
##   family_nam = col_character(),
##   shape_Area = col_double()
## )
```

```
#####
```

3. tidyr

Install and load the tidyr package

```
#install.packages("tidyr", dep=T)
library(tidyr)
```

Using species_info, replace NAs of presence, origin and season column with 0s. Complete binomial, presence, origin and season columns of species_info Drop missing values from species_info Separate binomial into genus and species, but keep binomial

```
# Are there any NAs?
species_info %>% anyNA()
```

```
## [1] TRUE
```

```
# How many rows with NAs in origin column?
species_info %>%
  subset(is.na(origin)==TRUE) %>%
  nrow
```

```
## [1] 10064
```

```
species_new <- species_info %>%
  replace_na(list(presence=0, origin=0, seasonal=0)) %>%
  complete(binomial, kingdom_na, presence, origin, seasonal) %>%
  drop_na() %>%
  separate(binomial,
```

```
into=c("genus", "species"),
remove=F)
```

```
## Warning: Expected 2 pieces. Additional pieces discarded in 7 rows [19706, 19981,
## 23353, 23746, 24166, 25971, 25973].
```

```
# colnames(species_new)
```

Why do you get an error message and how could we avoid it? Because some species also include information about the subspecies

```
species_new <- species_info %>%
  replace_na(list(presence=0, origin=0, seasonal=0)) %>%
  complete(binomial, kingdom_na, presence, origin, seasonal) %>%
  drop_na() %>%
  separate(binomial,
            into=c("genus", "species", "subspecies"),
            remove=F)
```

```
## Warning: Expected 3 pieces. Additional pieces discarded in 4 rows [19706, 19981,
## 23746, 24166].
```

```
## Warning: Expected 3 pieces. Missing pieces filled with `NA` in 30509 rows [1, 2,
## 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, ...].
```

Read species_data.csv.xz into a data.frame called species_data Note: species_data contains gridded species occurrence data for Bavaria at a spatial resolution of 0.5 degree

```
species_data <- read_csv("data/species_data.csv.xz")
```

```
## Parsed with column specification:
## cols(
##   .default = col_double()
## )
## See spec(...) for full column specifications.
```

2. Turn species_data from wide into long format, drop missing values and save to a data.frame called species_long

```
species_long <- species_data %>%
  gather(binomial, occurrence, -c(x,y)) %>% drop_na()

# Alternative
species_long <- gather(species_data,
                       colnames(species_data)[-c(1:2)],
                       key="binomial", value="occurrence") %>% drop_na()
```

Why do we have to drop NAs after we turn data into a long format? Because, we get lots of unmeaningful data entries when turning the data into a long format ##### Turn species_long back into wide format and write to a data.frame called species_wide

```
species_wide <- species_long %>%
  spread(binomial, occurrence)
```

```
#####
```

4. dplyr

```
# Install and load dplyr package
```

```
#install.packages("dplyr", dep=T)
library(dplyr)
```

Using species_long calculate the number of occurrences per species using group_by() and summarise

```
species_long %>% group_by(binomial) %>%
  summarise(sum=sum(occurrence))
```

```
## # A tibble: 322 x 2
##   binomial          sum
##   <chr>          <dbl>
## 1 Accipiter gentilis    189
## 2 Accipiter nisus      189
## 3 Acrocephalus arundinaceus  163
## 4 Acrocephalus dumetorum   121
## 5 Acrocephalus paludicola     6
## 6 Acrocephalus palustris   189
## 7 Acrocephalus schoenobaenus 156
## 8 Acrocephalus scirpaceus   189
## 9 Actitis hypoleucos    134
## 10 Aegithalos caudatus    189
## # ... with 312 more rows
```

```
species_long %>% group_by(binomial) %>%
  summarise(sum=n())
```

```
## # A tibble: 322 x 2
##   binomial          sum
##   <chr>          <int>
## 1 Accipiter gentilis    189
## 2 Accipiter nisus      189
## 3 Acrocephalus arundinaceus  163
## 4 Acrocephalus dumetorum   121
## 5 Acrocephalus paludicola     6
## 6 Acrocephalus palustris   189
## 7 Acrocephalus schoenobaenus 156
## 8 Acrocephalus scirpaceus   189
## 9 Actitis hypoleucos    134
## 10 Aegithalos caudatus    189
## # ... with 312 more rows
```

Of course, this could also be done with colSums() or count() ##### Using species_long, identify how many species have less than 10 occurrences?

```
species_long %>% group_by(binomial) %>%
  summarise(sum=sum(occurrence)) %>%
  arrange(desc(sum)) %>%
  filter(sum < 10)
```

```
## # A tibble: 34 x 2
##   binomial          sum
##   <chr>          <dbl>
## 1 Calidris alpina      9
```

```
## 2 Columba livia          9
## 3 Dryomys nitedula       9
## 4 Natrix tessellata      8
## 5 Pipistrellus kuhlii    8
## 6 Tichodroma muraria     8
## 7 Apodemus alpicola       7
## 8 Carduelis citrinella    7
## 9 Strix uralensis        7
## 10 Acrocephalus paludicola 6
## # ... with 24 more rows
```

Using species_long calculate species richness per grid cell

```
sr_xy <- species_long %>% group_by(x,y) %>%
  summarise(sr=sum(occurrence))
```

Extract data of the following 5 species from species_long:

```
species <- c("Anas crecca", "Lacerta agilis", "Mustela nivalis",
             "Pelophylax lessonae", "Vulpes vulpes")
# %in%
```

```
species_long %>% filter(binomial %in% species)
```

```
## # A tibble: 920 x 4
##       x     y binomial occurrence
##   <dbl> <dbl> <chr>         <dbl>
## 1  6.25  50.2 Anas crecca         1
## 2  6.25  50.8 Anas crecca         1
## 3  6.25  51.2 Anas crecca         1
## 4  6.25  51.8 Anas crecca         1
## 5  6.75  49.2 Anas crecca         1
## 6  6.75  49.8 Anas crecca         1
## 7  6.75  50.2 Anas crecca         1
## 8  6.75  50.8 Anas crecca         1
## 9  6.75  51.2 Anas crecca         1
## 10 6.75  51.8 Anas crecca         1
## # ... with 910 more rows
```

```
species_long %>%
  filter(binomial %in% c("Anas crecca", "Lacerta agilis"))
```

```
## # A tibble: 365 x 4
##       x     y binomial occurrence
##   <dbl> <dbl> <chr>         <dbl>
## 1  6.25  50.2 Anas crecca         1
## 2  6.25  50.8 Anas crecca         1
## 3  6.25  51.2 Anas crecca         1
## 4  6.25  51.8 Anas crecca         1
## 5  6.75  49.2 Anas crecca         1
## 6  6.75  49.8 Anas crecca         1
## 7  6.75  50.2 Anas crecca         1
## 8  6.75  50.8 Anas crecca         1
## 9  6.75  51.2 Anas crecca         1
## 10 6.75  51.8 Anas crecca         1
## # ... with 355 more rows
```

Using species_wide, extract data for all species starting with “Ac”

```
species_ac <- species_long %>% spread(binomial, occurrence) %>%  
  select(starts_with("Ac"))  
nrow(species_ac); ncol(species_ac)
```

```
## [1] 189
```

```
## [1] 9
```

Why do you have to use species_wide here? Because starts_with() only works with select() and not with filter() and with select you can only subset columns and not rows #####
Calculate the number of species per class_name in species_info and output as table using the kable function of the knitr package

```
library(knitr)  
species_info %>% group_by(class_name) %>%  
  distinct(binomial) %>% summarise(no_species=n()) %>%  
  kable()
```

class_name	no_species
AMPHIBIA	6428
AVES	10066
MAMMALIA	5322
REPTILIA	10064

```
#####
```

5. dplyr - Part II

Calculate the number of species per class in species_data Hint: To do this, you first need to join species_data with species_info

```
species_info %<>%  
  distinct(binomial, class_name, order_name, family_name)  
#species_long %>%  
# left_join(species_info, by = "binomial") %>%  
# group_by(class_name) %>% distinct(binomial) %>%  
# summarise(no_species=n())  
species_long %>%  
  left_join(species_info, by = "binomial") %>%  
  group_by(class_name) %>%  
  summarise(no_species=length(unique(binomial)))
```

```
## # A tibble: 4 x 2  
##   class_name no_species  
##   <chr>      <int>  
## 1 AMPHIBIA      21  
## 2 AVES         213  
## 3 MAMMALIA      77  
## 4 REPTILIA     11
```

Calculate the number of species per grid cell for each class Hint: To do this, species_info must contain distinct species names (binomial)


```
species_long %>%
  left_join(species_info, by="binomial") %>%
  group_by(x,y,class_name) %>%
  summarise(sr=sum(occurrence))
```

```
## # A tibble: 756 x 4
## # Groups:   x, y [189]
##       x     y class_name    sr
##   <dbl> <dbl> <chr>      <dbl>
## 1  6.25  50.2 AMPHIBIA      16
## 2  6.25  50.2 AVES        126
## 3  6.25  50.2 MAMMALIA     54
## 4  6.25  50.2 REPTILIA      4
## 5  6.25  50.8 AMPHIBIA     17
## 6  6.25  50.8 AVES       132
## 7  6.25  50.8 MAMMALIA     54
## 8  6.25  50.8 REPTILIA      4
## 9  6.25  51.2 AMPHIBIA     17
## 10 6.25  51.2 AVES       133
## # ... with 746 more rows
```

Identify the most widely distributed species for each class

```
sp_wide <- species_long %>%
  left_join(species_info, by="binomial") %>%
  group_by(binomial, class_name, family_name) %>%
  summarise(sum=sum(occurrence)) %>%
  group_by(class_name) %>% top_n(1,sum)
```

Note: This should not be higher than the maximum grid cells of Germany (189)

```
#####
```

6. Making plots with ggplot2

Install and load ggplot2

```
#install.packages(ggplot2, dep=T)
library(ggplot2)
```

Plot barchart of the number of species per class

```
# Same as before
library(readr); library(dplyr);
library(tidyr); library(ggplot2)
species_long <- read_csv("data/species_data.csv.xz") %>%
  gather(binomial, occurrence, -c(x,y)) %>% drop_na()
```

```
## Parsed with column specification:
## cols(
##   .default = col_double()
## )
```

```
## See spec(...) for full column specifications.
```

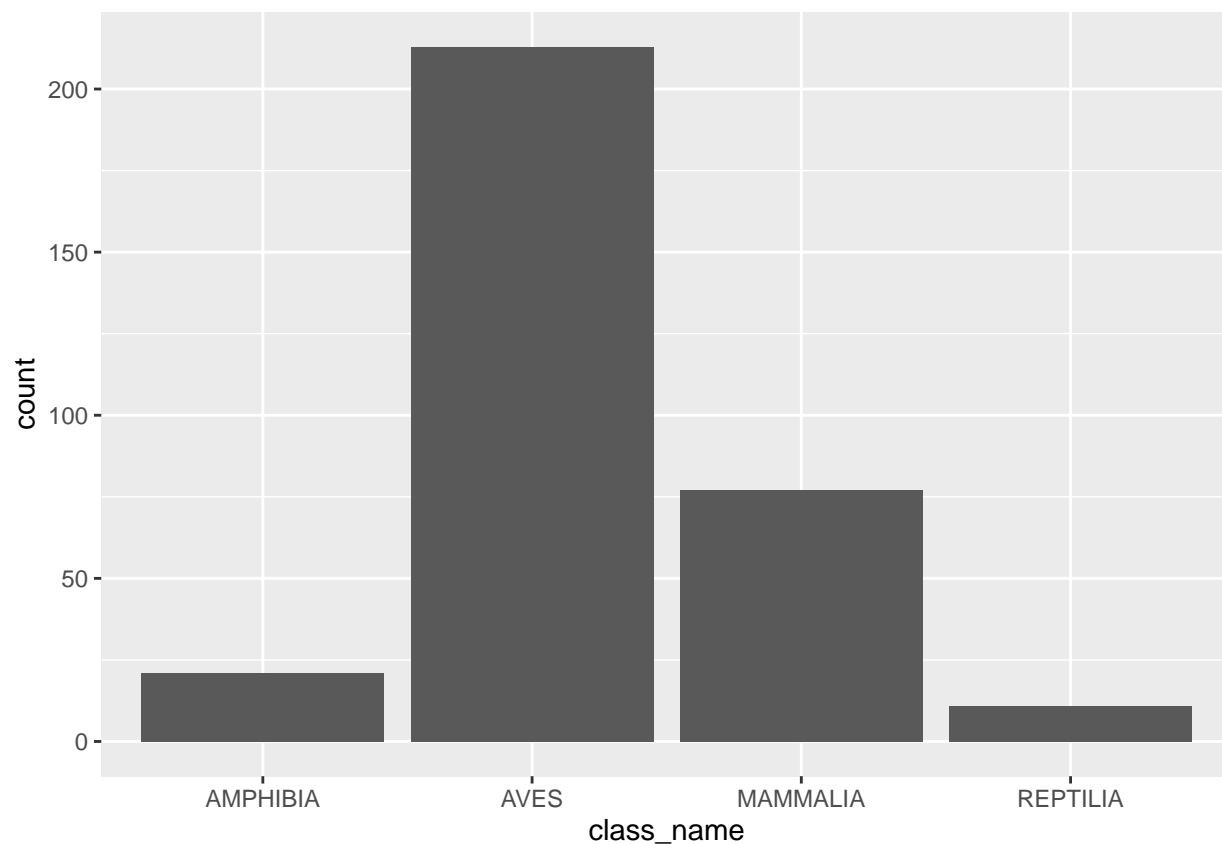
```
species_info <- read_csv("data/species_info.csv.xz")
```

```
## Parsed with column specification:
```

```
## cols(
##   binomial = col_character(),
##   presence = col_double(),
##   origin = col_double(),
##   seasonal = col_double(),
##   kingdom_na = col_character(),
##   phylum_name = col_character(),
##   class_name = col_character(),
##   order_name = col_character(),
##   family_name = col_character(),
##   shape_area = col_double()
## )

data_plot <- species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(class_name) %>% distinct(binomial) %>%
  summarise(no_species= n())

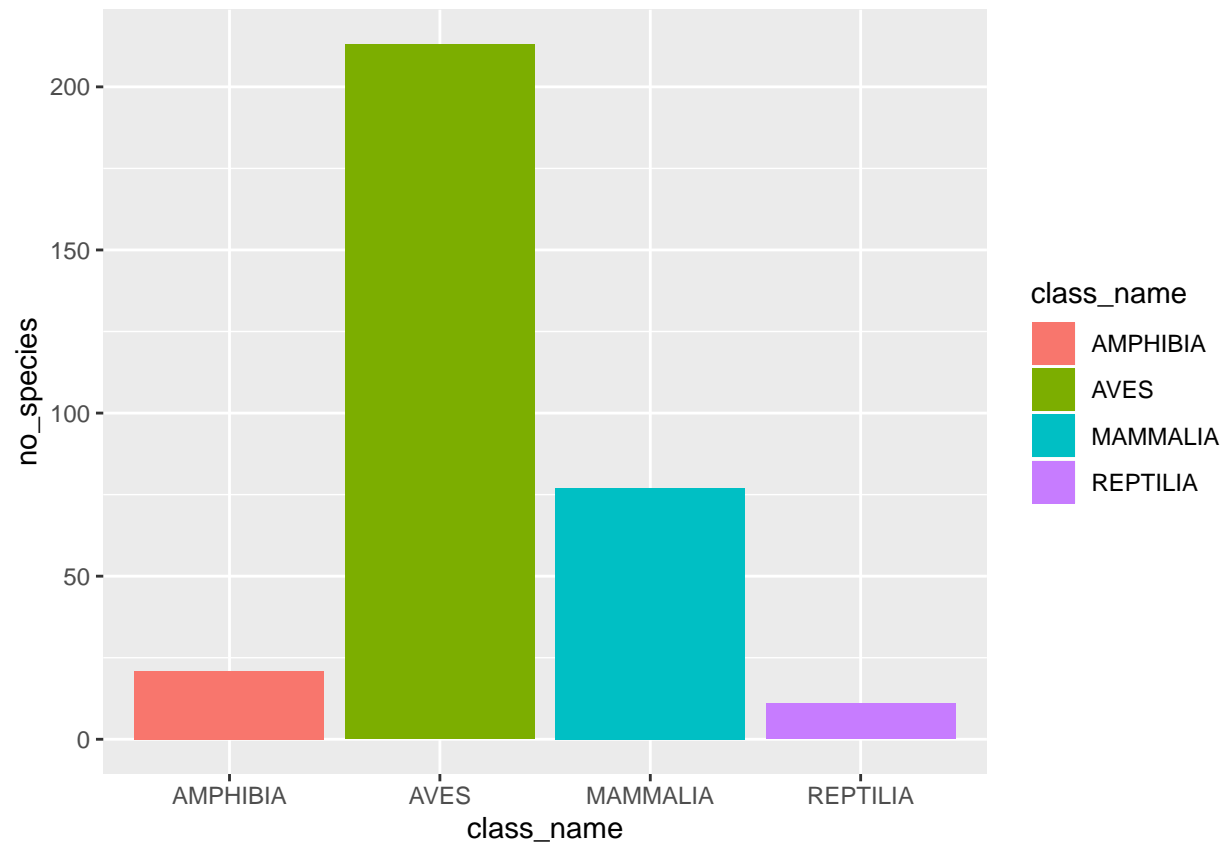
# Now, we add a plot
species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(class_name) %>% distinct(binomial) %>%
  ggplot(aes(class_name)) + geom_bar()
```



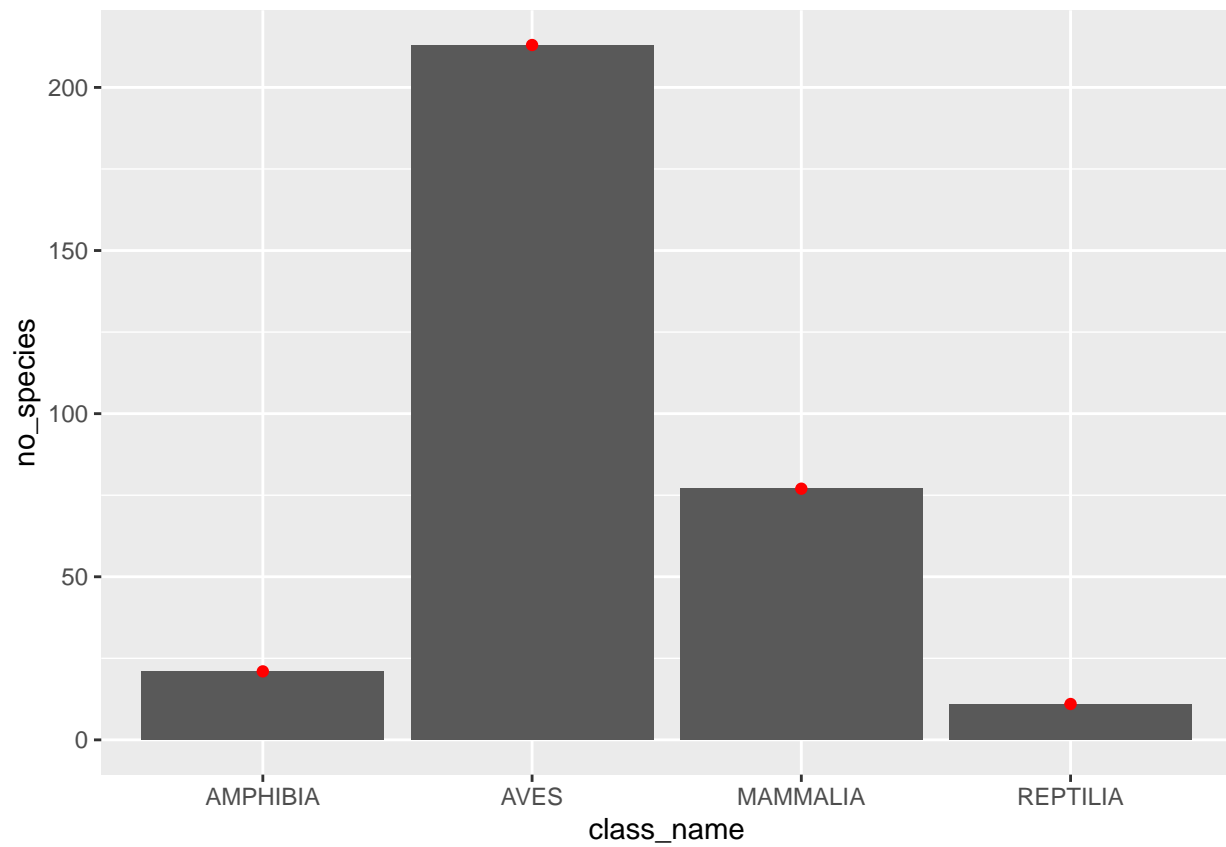
```
species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(class_name) %>% distinct(binomial) %>%
  summarise(no_species= n())
```

```
## # A tibble: 4 x 2
##   class_name no_species
##   <chr>      <int>
## 1 AMPHIBIA      21
## 2 AVES         213
## 3 MAMMALIA      77
## 4 REPTILIA      11
```

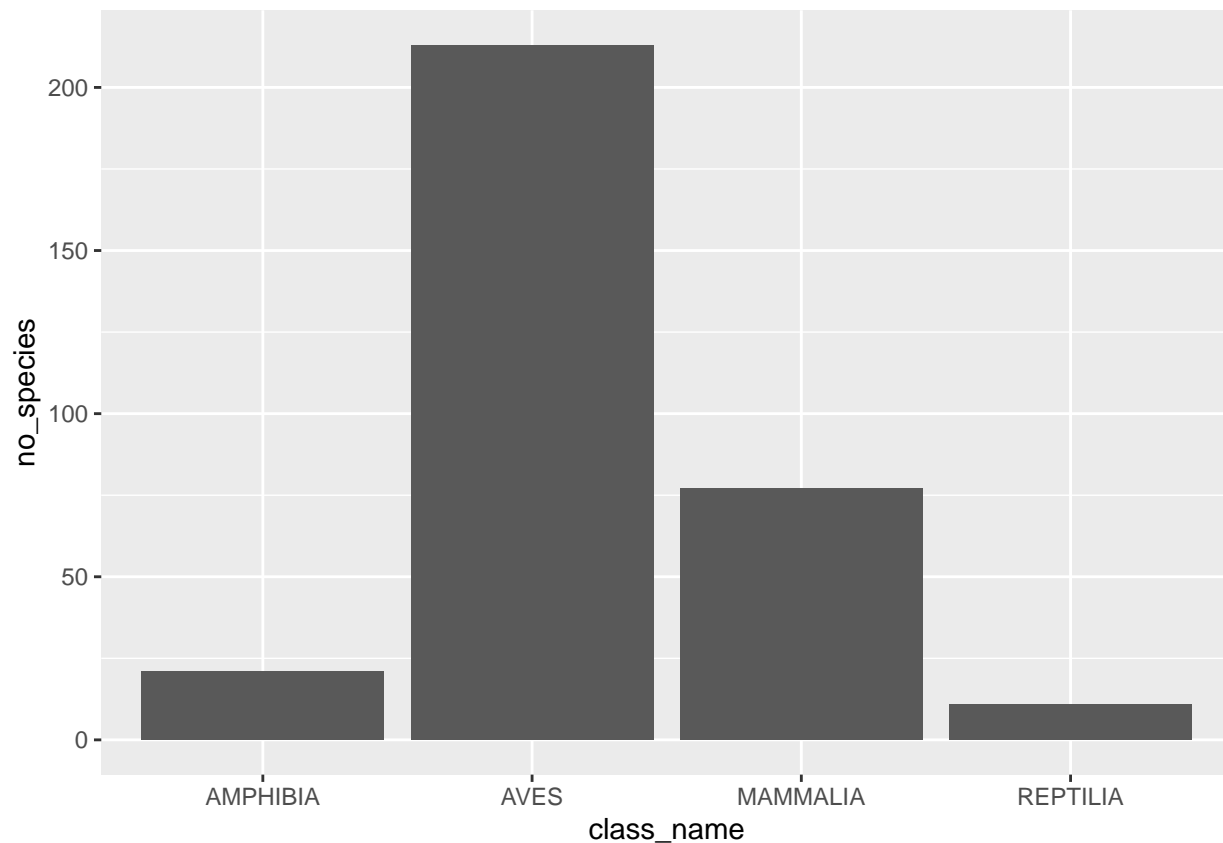
```
data_plot %>% ggplot(aes(class_name, no_species,
                          fill=class_name)) +
  geom_bar(stat="identity")
```



```
ggplot(data_plot, aes(class_name, no_species)) +
  geom_bar(stat="identity") + geom_point(colour="red")
```



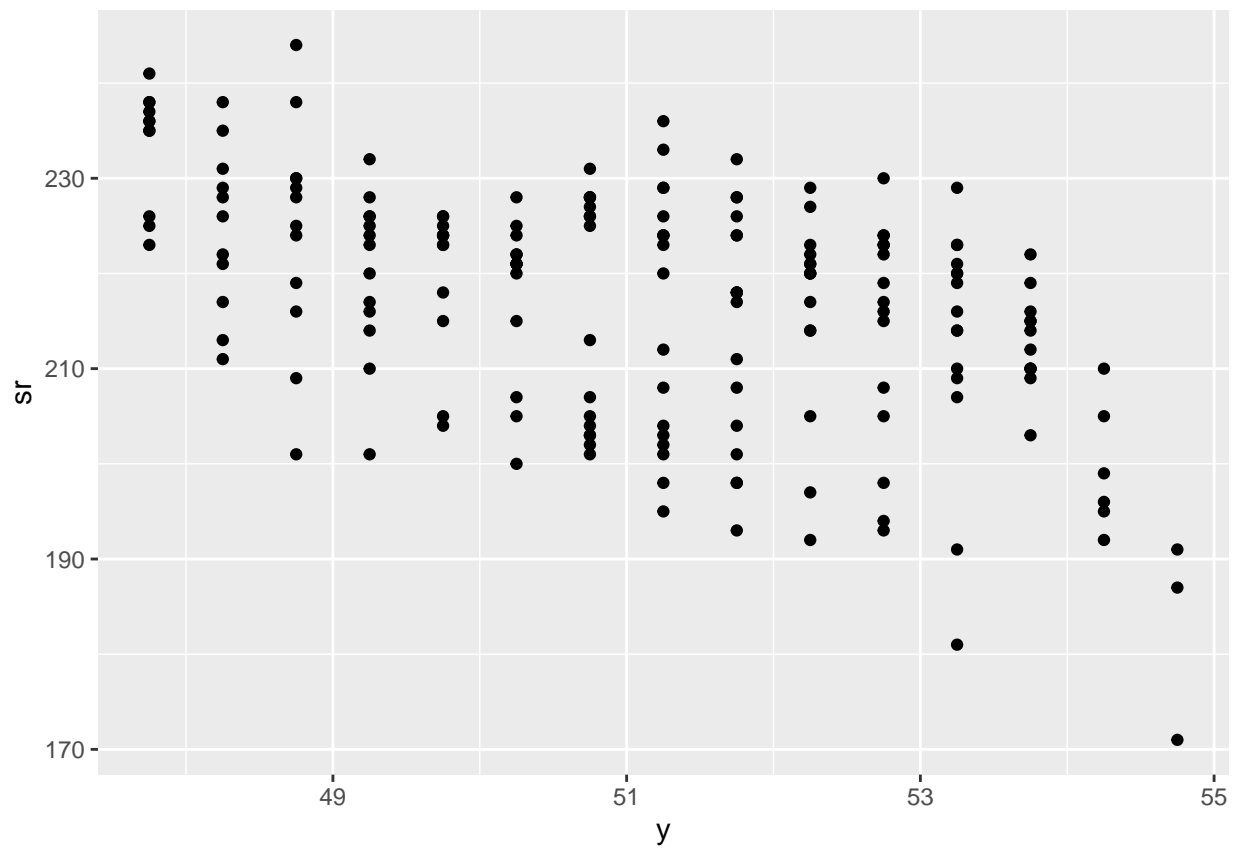
```
ggplot() +  
  geom_bar(data=data_plot, aes(class_name, no_species),  
           stat="identity")
```



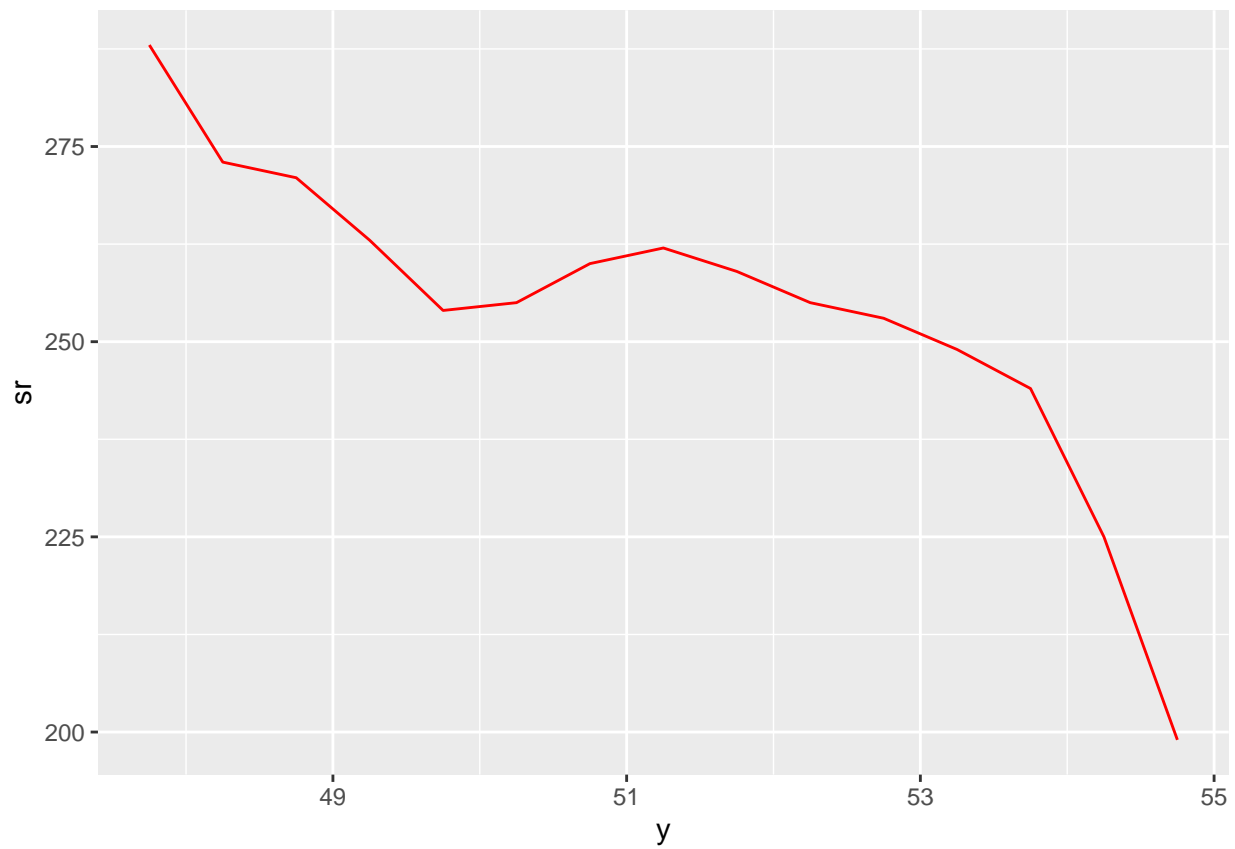
Plot number of occurrences for the 10 least widely distributed species

```
library(magrittr)
species_info %<>%
  distinct(binomial, class_name, order_name, family_nam)
species_long %>% drop_na() %>%
  left_join(species_info, by="binomial") %>%
  group_by(binomial, class_name, family_nam) %>%
  summarise(sum=sum(occurrence)) %>% ungroup() %>%
  top_n(-10) %>% # You can also use for example filter to extract the species
  ggplot(aes(binomial, sum, fill=class_name)) +
  geom_bar(stat="identity")
```

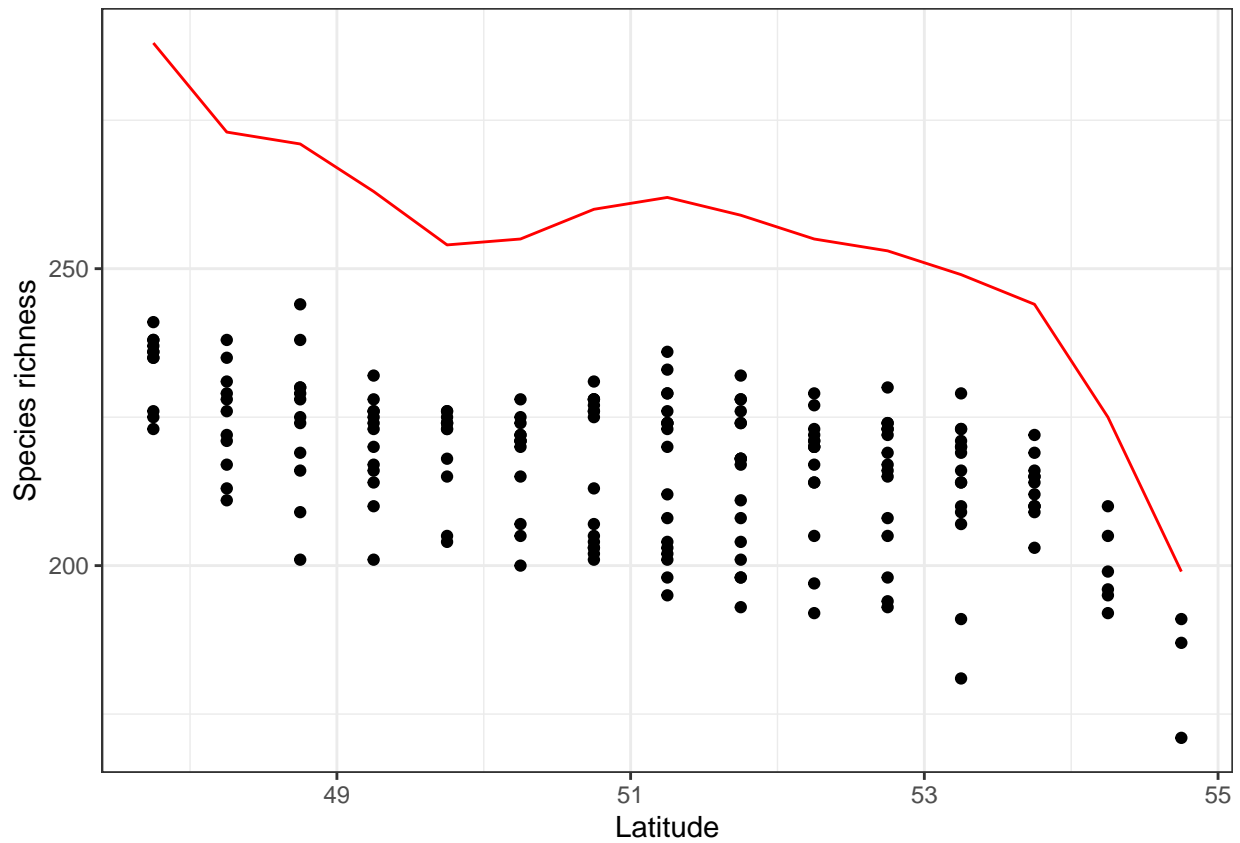
Selecting by sum



```
dat2 <- species_long %>% group_by(y) %>% distinct(binomial) %>%
  summarise(sr=n())
ggplot(dat2, aes(y,sr)) + geom_line(colour="red")
```

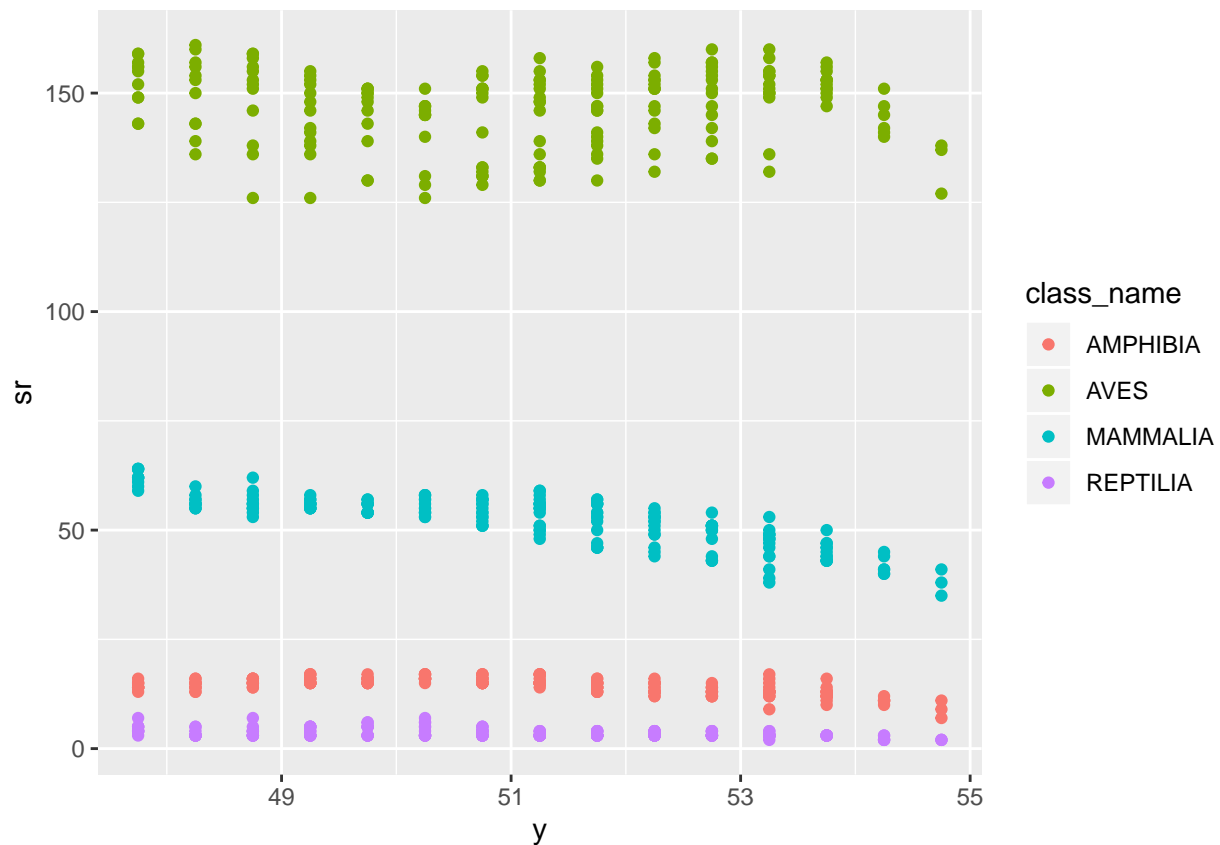


```
ggplot() + geom_point(data=dat1, aes(y, sr)) +  
  geom_line(data=dat2, aes(y,sr), colour="red") +  
  theme_bw() + labs(x="Latitude", y="Species richness")
```

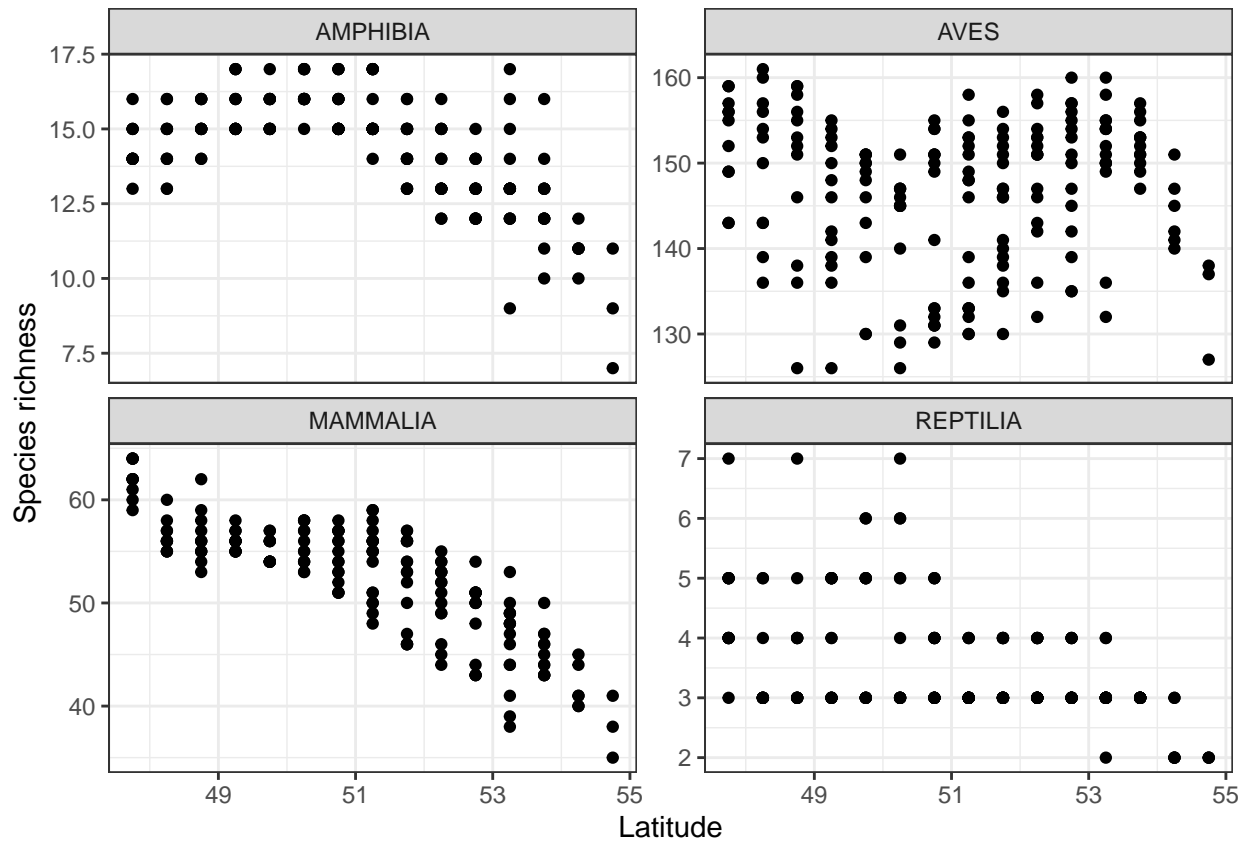



Use faceting to split the previous plots by class

```
species_long %>% drop_na() %>%
  left_join(species_info, by="binomial") %>%
  group_by(x,y,class_name) %>%
  summarise(sr=sum(occurrence)) %>%
  ggplot(aes(y, sr, colour=class_name)) + geom_point()
```



```
species_long %>% drop_na() %>%
  left_join(species_info, by="binomial") %>%
  group_by(x,y,class_name) %>%
  summarise(sr=sum(occurrence)) %>%
  ggplot(aes(y, sr)) + geom_point() +
  facet_wrap(~class_name, scales = "free_y") +
  theme_bw() + labs(x="Latitude", y="Species richness")
```

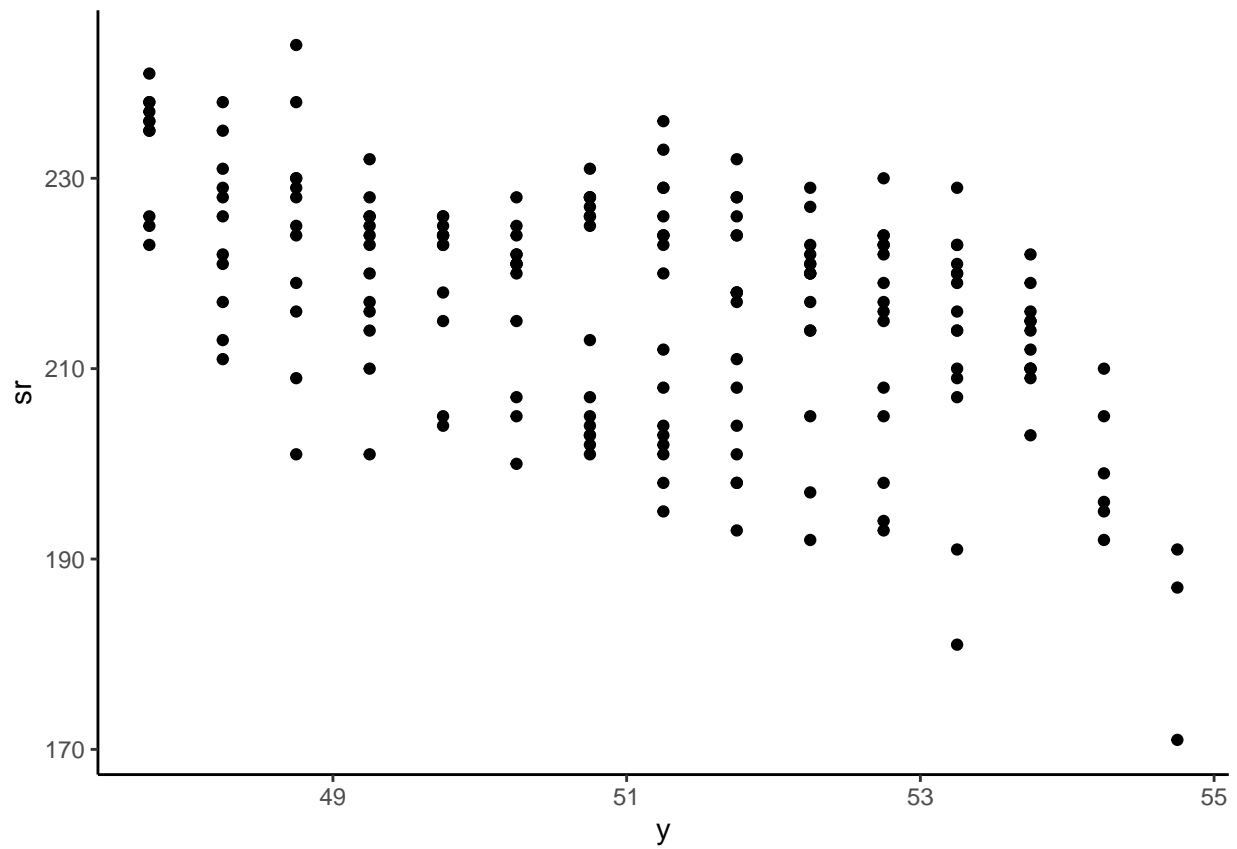


Alternatively, you can also use facet_grid()

Use patchwork to create individual plots and combine them into one figure Install and load the patchwork package

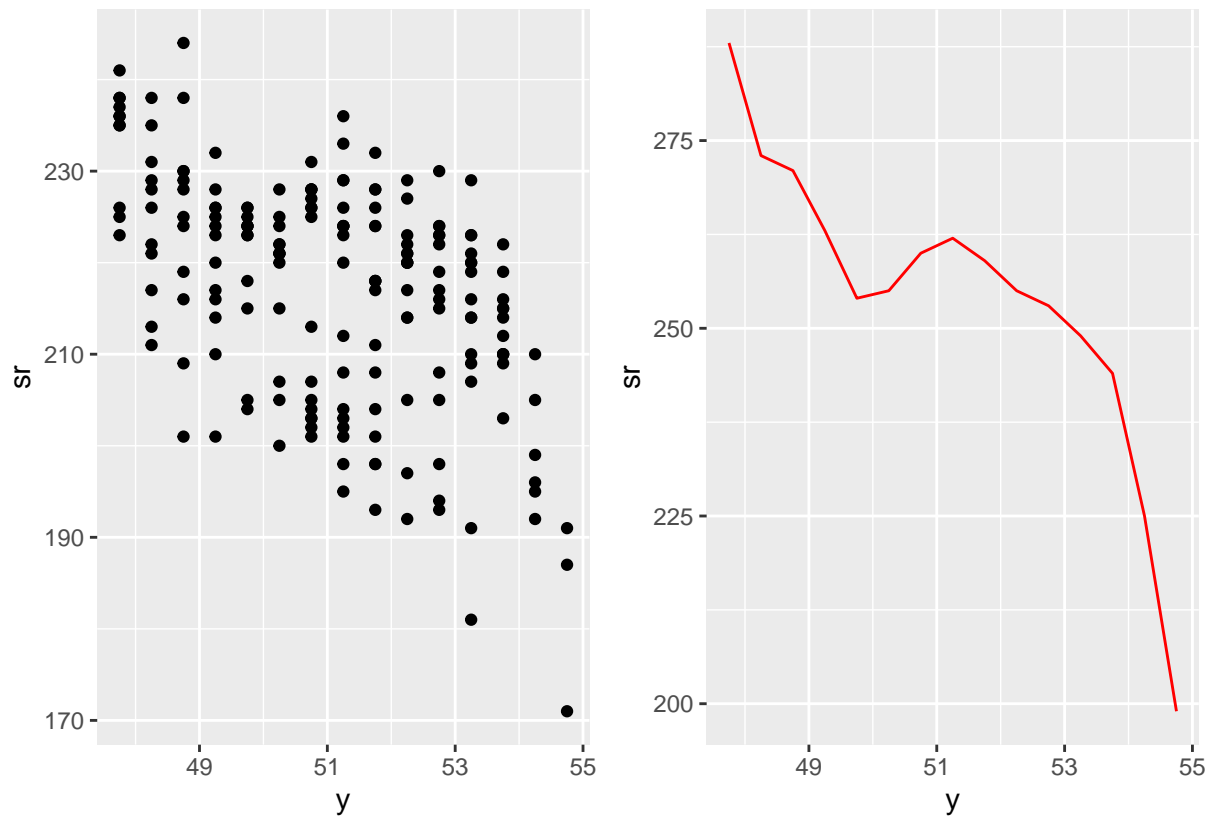
```
#install.packages("devtools")
#devtools::install_github("thomasp85/patchwork")
library(patchwork)

dat1 <- species_long %>% group_by(x,y) %>%
  summarise(sr=sum(occurrence))
p1 <- ggplot(dat1, aes(y, sr)) + geom_point()
p1 + theme_classic()
```



```
dat2 <- species_long %>% group_by(y) %>% distinct(binomial) %>%
  summarise(sr=n())
p2 <- ggplot(dat2, aes(y,sr)) + geom_line(colour="red")

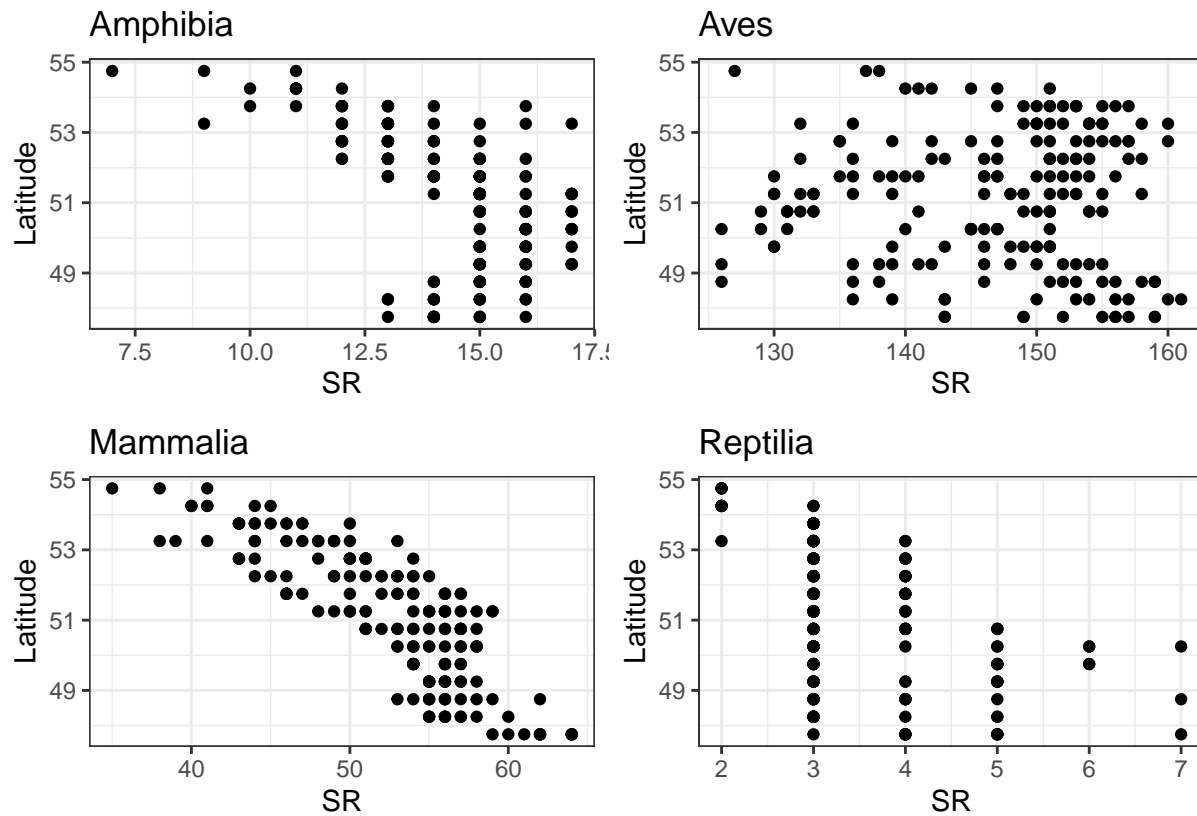
p1 + p2
```



Create individual plots for each class and using patchwork display all plots combined in one

```
p1 <- species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(x,y,class_name) %>% filter(class_name == "AMPHIBIA") %>%
  summarise(sum=sum(occurrence)) %>% ggplot() +
  geom_point(aes(x=sum, y=y)) + theme_bw() +
  labs(x="SR", y="Latitude", title="Amphibia")
p2 <- species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(x,y,class_name) %>% filter(class_name == "AVES") %>%
  summarise(sum=sum(occurrence)) %>% ggplot() +
  geom_point(aes(x=sum, y=y)) + theme_bw() +
  labs(x="SR", y="Latitude", title="Aves")
p3 <- species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(x,y,class_name) %>% filter(class_name == "MAMMALIA") %>%
  summarise(sum=sum(occurrence)) %>% ggplot() +
  geom_point(aes(x=sum, y=y)) + theme_bw() +
  labs(x="SR", y="Latitude", title="Mammalia")
p4 <- species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(x,y,class_name) %>% filter(class_name == "REPTILIA") %>%
  summarise(sum=sum(occurrence)) %>% ggplot() +
  geom_point(aes(x=sum, y=y)) + theme_bw() +
  labs(x="SR", y="Latitude", title="Reptilia")

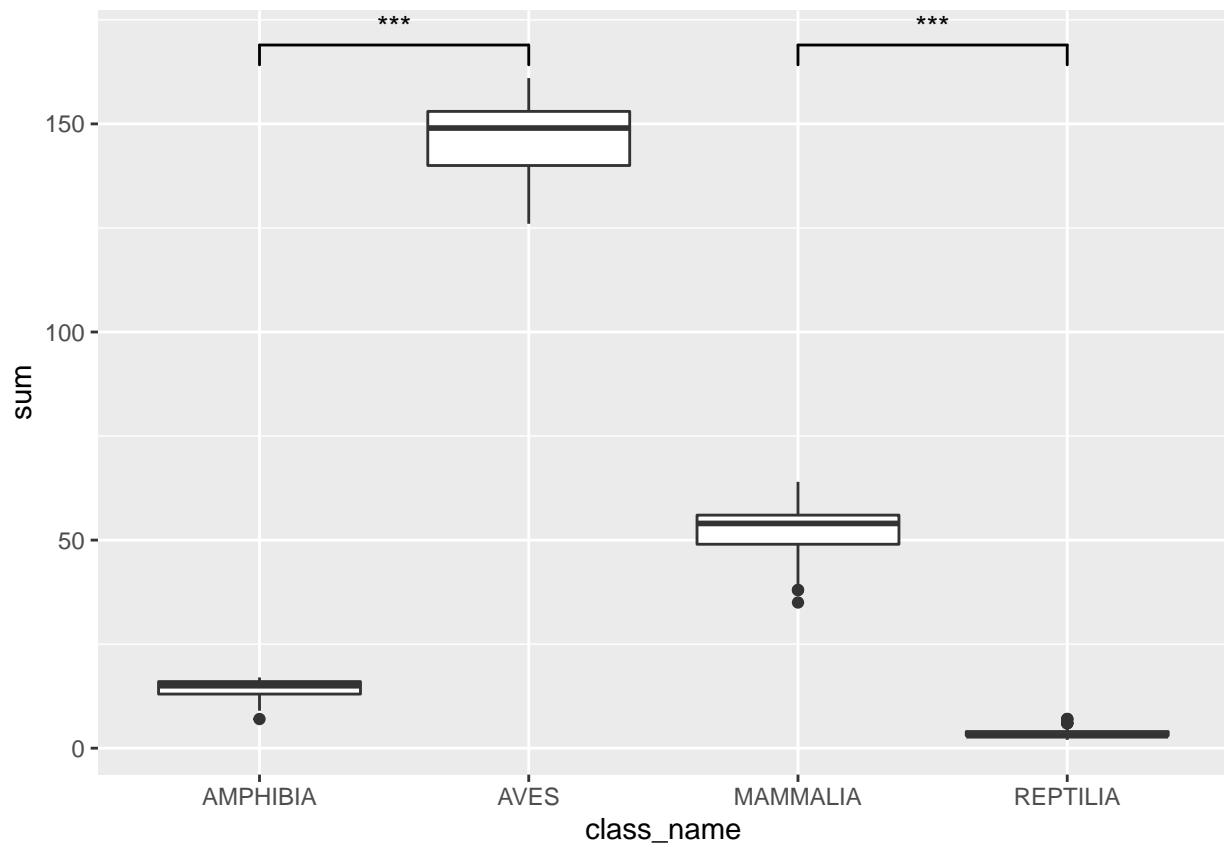
p1 + p2 + p3 + p4
```



Plot boxplot of species richness against class and add significant differences with `geom_signif()` from the `ggsignif` package

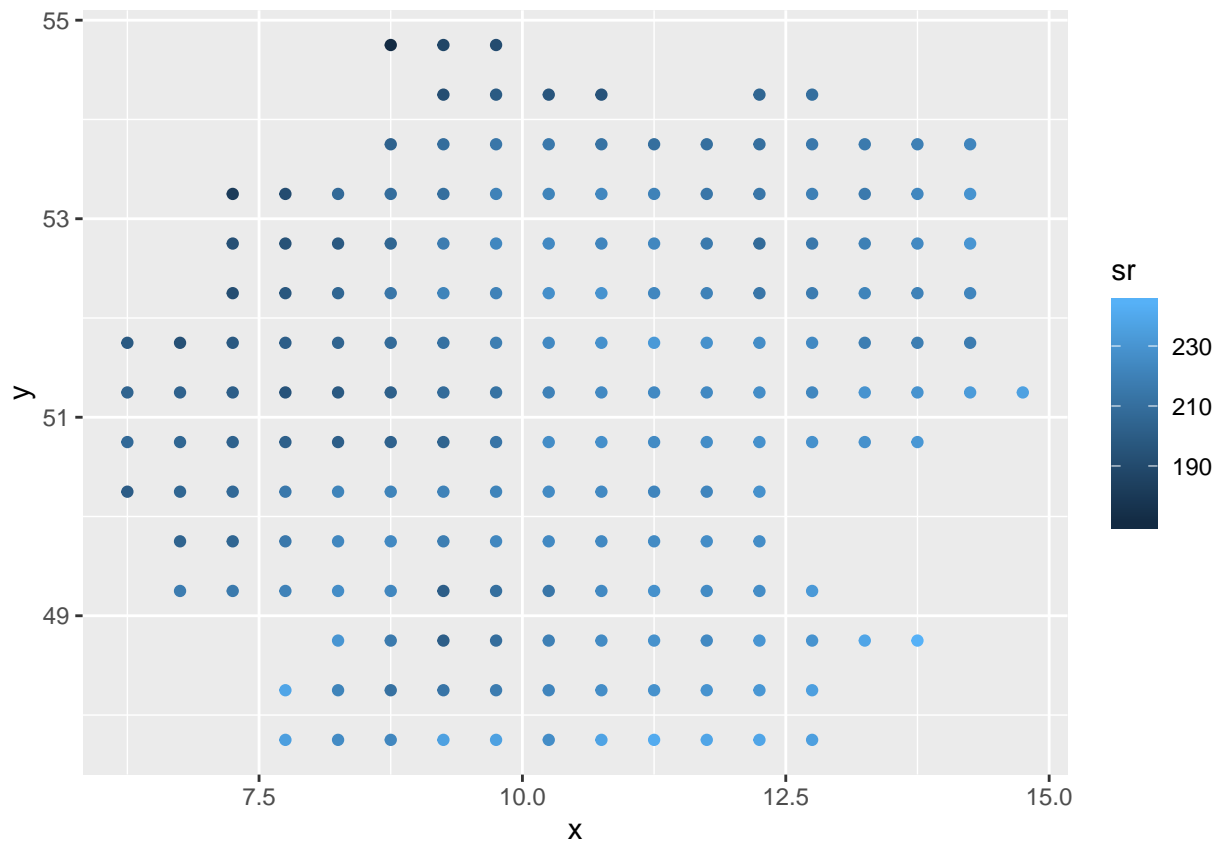
```
#install.packages("ggsignif", dep=T)
library(ggsignif)

species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(x,y,class_name) %>%
  summarise(sum=sum(occurrence)) %>%
  ggplot(aes(x=class_name, y=sum)) +
  geom_boxplot() +
  geom_signif(comparisons = list(c("AMPHIBIA", "AVES"),
                                c("REPTILIA", "MAMMALIA")),
             map_signif_level=TRUE)
```

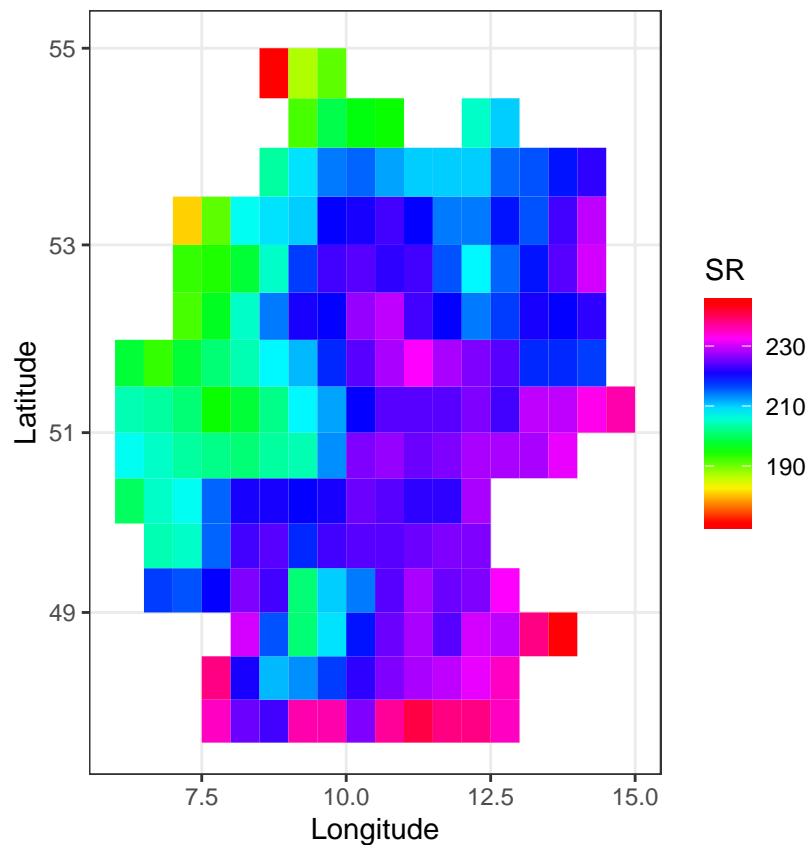


Make map of species richness

```
species_long %>% group_by(x,y) %>%
  summarise(sr=sum(occurrence)) %>%
  ggplot() + geom_point(aes(x,y,colour=sr))
```



```
species_long %>% group_by(x,y) %>%
  summarise(sr=sum(occurrence)) %>%
  ggplot() + geom_tile(aes(x,y,fill=sr)) +
  scale_fill_gradientn(name="SR", colours=rainbow(255)) +
  theme_bw() + coord_map() +
  labs(x="Longitude", y="Latitude")
```

#####

7. Data analysis with dplyr

Create linear model using `do()`, i.e. of species richness against latitude for each `class_name`

```
library(magrittr)
species_long <- read_csv("data/species_data.csv.xz") %>%
  gather(binomial, occurrence, -c(x,y)) %>% drop_na()
```

```
## Parsed with column specification:
## cols(
##   .default = col_double()
## )
## See spec(...) for full column specifications.
```

```
species_info <- read_csv("data/species_info.csv.xz")
```

```
## Parsed with column specification:
## cols(
##   binomial = col_character(),
##   presence = col_double(),
##   origin = col_double(),
##   seasonal = col_double(),
##   kingdom_na = col_character(),
##   phylum_na = col_character(),
##   class_name = col_character(),
```

```
##   order_name = col_character(),
##   family_nam = col_character(),
##   shape_Area = col_double()
## )

species_info %<>%
  distinct(binomial, class_name, order_name, family_nam)

models <- species_long %>% left_join(species_info, by="binomial") %>%
  group_by(x,y,class_name) %>% summarise(sr=sum(occurrence)) %>%
  group_by(class_name) %>% do(model=lm(sr ~ y, data=.) )
```

Extract coefficients of linear model

```
ctl <- c(4.17,5.58,5.18,6.11,4.50,4.61,5.17,4.53,5.33,5.14)
trt <- c(4.81,4.17,4.41,3.59,5.87,3.83,6.03,4.89,4.32,4.69)
group <- gl(2, 10, 20, labels = c("Ctl", "Trt"))
weight <- c(ctl, trt)
mod <- lm(weight ~ group)
mod
```

```
##
## Call:
## lm(formula = weight ~ group)
##
## Coefficients:
## (Intercept)      groupTrt
##          5.032         -0.371
```

```
summary(mod)
```

```
##
## Call:
## lm(formula = weight ~ group)
##
## Residuals:
##      Min       1Q   Median       3Q      Max
## -1.0710 -0.4938  0.0685  0.2462  1.3690
##
## Coefficients:
##              Estimate Std. Error t value Pr(>|t|)
## (Intercept)    5.0320     0.2202  22.850 9.55e-15 ***
## groupTrt       -0.3710     0.3114  -1.191   0.249
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
##
## Residual standard error: 0.6964 on 18 degrees of freedom
## Multiple R-squared:  0.07308,    Adjusted R-squared:  0.02158
## F-statistic: 1.419 on 1 and 18 DF,  p-value: 0.249
```

```
#models$model[[1]]$coefficients
#coef(summary(models$model[[1]]))
```

```
models %>% do(data.frame(class = .$class_name,
                        var=names(coef(.$model)),
                        coef(.$model)))
```

```
## Warning in bind_rows_(x, .id): Unequal factor levels: coercing to character
## Warning in bind_rows_(x, .id): binding character and factor vector, coercing
## into character vector

## Warning in bind_rows_(x, .id): binding character and factor vector, coercing
## into character vector

## Warning in bind_rows_(x, .id): binding character and factor vector, coercing
## into character vector

## Source: local data frame [8 x 3]
## Groups: <by row>
##
## # A tibble: 8 x 3
##   class      var      coef...model.
## * <chr>    <fct>      <dbl>
## 1 AMPHIBIA (Intercept)    41.9
## 2 AMPHIBIA y             -0.539
## 3 AVES      (Intercept)   151.
## 4 AVES      y             -0.0814
## 5 MAMMALIA (Intercept)   179.
## 6 MAMMALIA y             -2.48
## 7 REPTILIA (Intercept)   15.2
## 8 REPTILIA y             -0.230
```

Alternatively

```
library(broom)
# glance(), tidy(), augment()
tidy(models$model[[1]])
```

```
## # A tibble: 2 x 5
##   term      estimate std.error statistic  p.value
##   <chr>      <dbl>    <dbl>    <dbl>    <dbl>
## 1 (Intercept)  41.9      2.85     14.7  4.40e-33
## 2 y          -0.539    0.0557    -9.68  3.24e-18
```

```
glance(models$model[[1]])
```

```
## # A tibble: 1 x 11
##   r.squared adj.r.squared sigma statistic  p.value    df logLik   AIC   BIC
##   <dbl>      <dbl> <dbl>    <dbl>    <dbl> <int> <dbl> <dbl> <dbl>
## 1    0.334    0.330  1.45     93.7  3.24e-18     2  -338.  681.  691.
## # ... with 2 more variables: deviance <dbl>, df.residual <int>
```

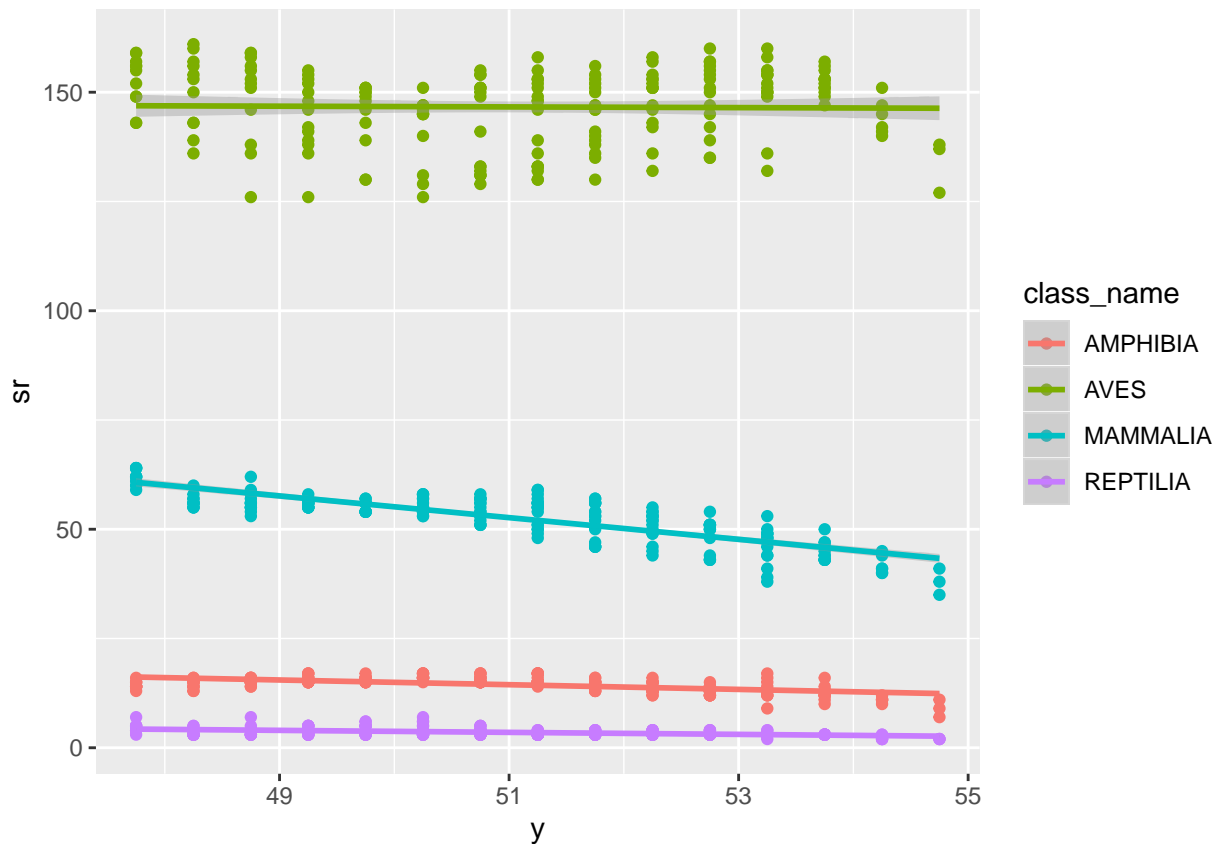
```
models %>% do(tidy(.$model))
```

```
## Source: local data frame [8 x 5]
## Groups: <by row>
##
## # A tibble: 8 x 5
##   term      estimate std.error statistic  p.value
## * <chr>      <dbl>    <dbl>    <dbl>    <dbl>
## 1 (Intercept)  41.9      2.85     14.7  4.40e-33
```

```
## 2 y          -0.539      0.0557    -9.68  3.24e-18
## 3 (Intercept) 151.        17.2      8.77  1.08e-15
## 4 y          -0.0814     0.336    -0.242 8.09e- 1
## 5 (Intercept) 179.        6.39     28.1   5.47e-69
## 6 y          -2.48       0.125    -19.9   5.43e-48
## 7 (Intercept) 15.2        1.63      9.34  2.93e-17
## 8 y          -0.230      0.0319    -7.20  1.44e-11
```

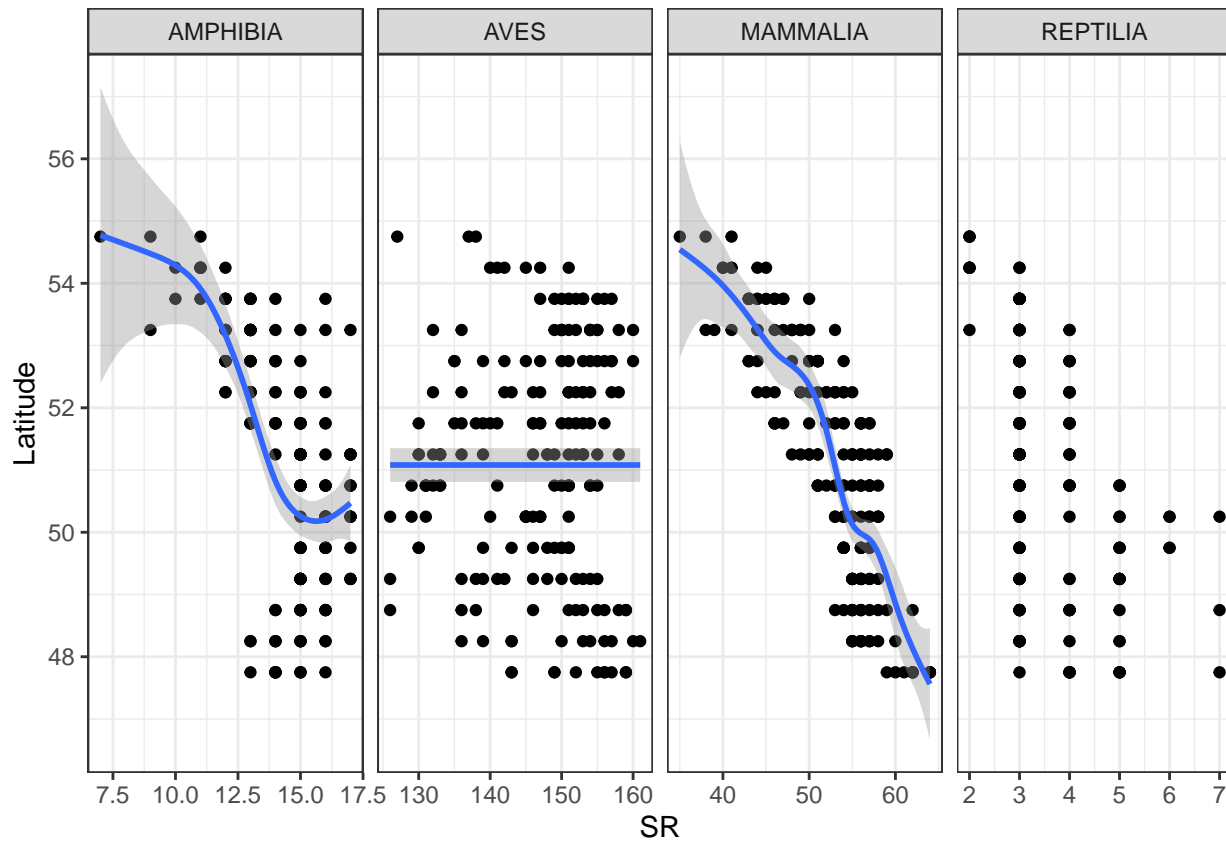
Run and plot gam of species richness against latitude for each class using ggplot2 (geom_smooth)

```
species_long %>% left_join(species_info, by="binomial") %>%
  group_by(x,y,class_name) %>% summarise(sr=sum(occurrence)) %>%
  ggplot(aes(y,sr, colour=class_name)) +
  geom_point() + geom_smooth(method="gam")
```



```
species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(x,y,class_name) %>% summarise(sr=sum(occurrence)) %>%
  ggplot(aes(x=sr, y=y)) + geom_point() +
  facet_grid(.~class_name, scales="free_x") +
  geom_smooth(method="gam", formula = y ~ s(x, bs = "cs")) +
  labs(x="SR", y="Latitude") + theme_bw()
```

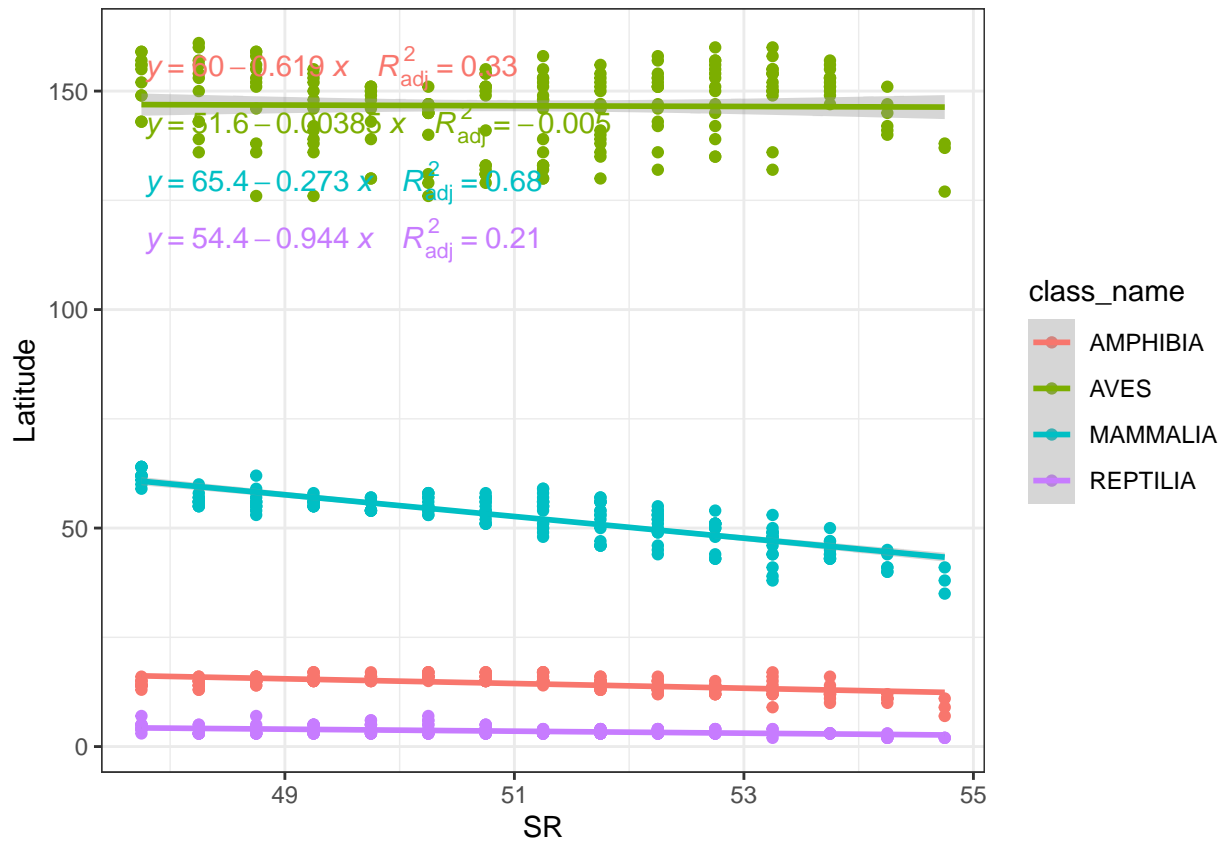
```
## Warning: Computation failed in `stat_smooth()`:
## x has insufficient unique values to support 10 knots: reduce k.
```



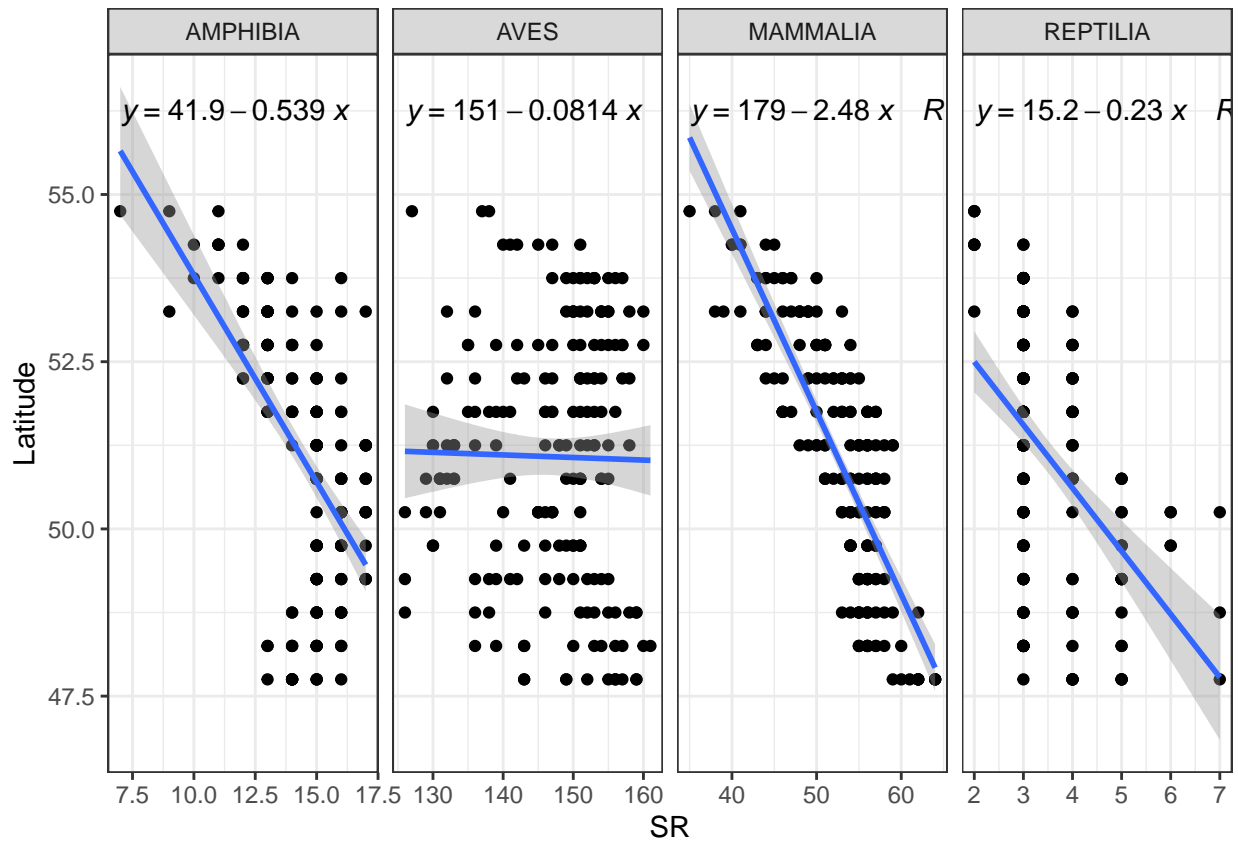
Run and plot lm of species richness against latitude for each class using ggplot2 and add equation to plot using `stat_poly_eq()` of the `ggpmisc` package

```
#install.packages("ggpmisc", dep=T)
library(ggpmisc)

# One plot with different colours
species_long %>% left_join(species_info, by="binomial") %>%
  group_by(x,y,class_name) %>% summarise(sr=sum(occurrence)) %>%
  ggplot(aes(y,sr, colour=class_name)) +
  geom_point() + geom_smooth(method="lm") +
  stat_poly_eq(formula = x~y,
               aes(colour=class_name,
                   label = paste(stat(eq.label),
                                stat(adj.rr.label), sep = "~~~"),
                   parse = TRUE) +
  labs(x="SR", y="Latitude") + theme_bw()
```

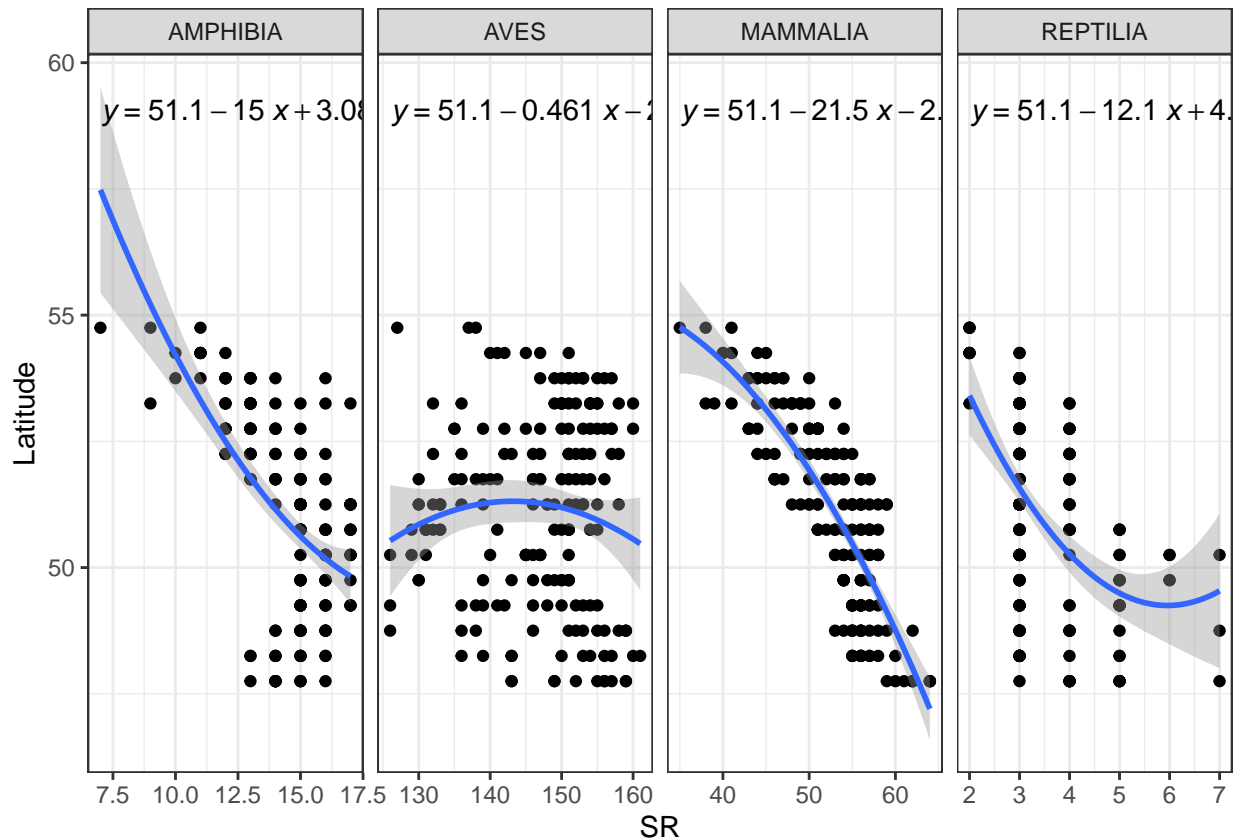


```
# Panel plot with different panels
species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(x,y,class_name) %>% summarise(sr=sum(occurrence)) %>%
  ggplot(aes(x=sr, y=y)) + geom_point() +
  facet_grid(.~class_name, scales="free_x") +
  geom_smooth(method="lm") +
  stat_poly_eq(formula = x~y,
               aes(label = paste(stat(eq.label),
                                stat(adj.rr.label), sep = "~~~")),
               parse = TRUE) +
  labs(x="SR", y="Latitude") + theme_bw()
```



We can do the same using a quadratic term

```
species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(x,y,class_name) %>% summarise(sr=sum(occurrence)) %>%
  ggplot(aes(x=sr, y=y)) + geom_point() +
  facet_grid(.~class_name, scales="free_x") +
  geom_smooth(method="lm", formula= y ~ poly(x, 2)) +
  stat_poly_eq(formula = y ~ poly(x, 2),
               aes(label = paste(stat(eq.label),
                                stat(adj.rr.label), sep = "~~~"),
                   parse = TRUE) +
  labs(x="SR", y="Latitude") + theme_bw()
```



Note: Then you have to specify the quadratic term in the formula argument of both functions, `geom_smooth` and `stat_poly_eq`. ##### Run ANOVA to compare linear models of linear versus quadratic term using the `tidy()` function of the `broom` package

```
#install.packages("broom", dep=T)
library(broom)

species_long %>%
  left_join(species_info, by=("binomial")) %>%
  group_by(x,y,class_name) %>% summarise(sr=sum(occurrence)) %>%
  group_by(class_name) %>% do(
    mod_linear = lm(sr ~ y, data = .),
    mod_quad = lm(sr ~ poly(y, 2), data = .) %>%
    do(aov = anova(. $mod_linear, . $mod_quad)) %>%
    rowwise %>% do(tidy(. $aov)) %>% tidyr::drop_na()
```

```
## Warning: Unknown or uninitialised column: 'term'.
```

```
## Warning: Unknown or uninitialised column: 'term'.
```

```
## Warning: Unknown or uninitialised column: 'term'.
```

```
## Warning: Unknown or uninitialised column: 'term'.
```

```
## # A tibble: 4 x 6
##   res.df    rss    df sumsq statistic p.value
##   <dbl> <dbl> <dbl> <dbl>      <dbl>   <dbl>
## 1     186    231.     1 162.      131. 2.88e-23
```



```
## 2      186 13695.      1 648.      8.80 3.40e- 3
## 3      186 1764.      1 215.      22.7 3.87e- 6
## 4      186 128.      1 1.55      2.25 1.35e- 1
```

Note: For the Anova, you also need the rowwise function, which you do not need for extracting the output of `lm()`.

```
#####
```

8. Spatio-temporal data with stars and sf

Read `tas_ewembi_deu_1981_2010.nc` using the `read_ncdf()` function of the `stars` package, turn into `data.frame` and drop NAs `tas_ewembi_deu_1981_2010.nc` contains daily temperature data for Germany at a spatial resolution of 0.5 degree from 1981 - 2010

```
#install.packages("stars", dep=T)
library(stars)

env <- read_ncdf("data/tas_ewembi_deu_1981_2010.nc") %>%
  as.data.frame() %>% drop_na()
```

Define timestamp of date variable (z) using `as.Date()`

```
# Start date 1/01/1981, End date 31/12/2010
env$z <- factor(env$z, labels=seq(as.Date("1981/01/01"),
                                   as.Date("2010/12/31"), "days"))
head(env)
```

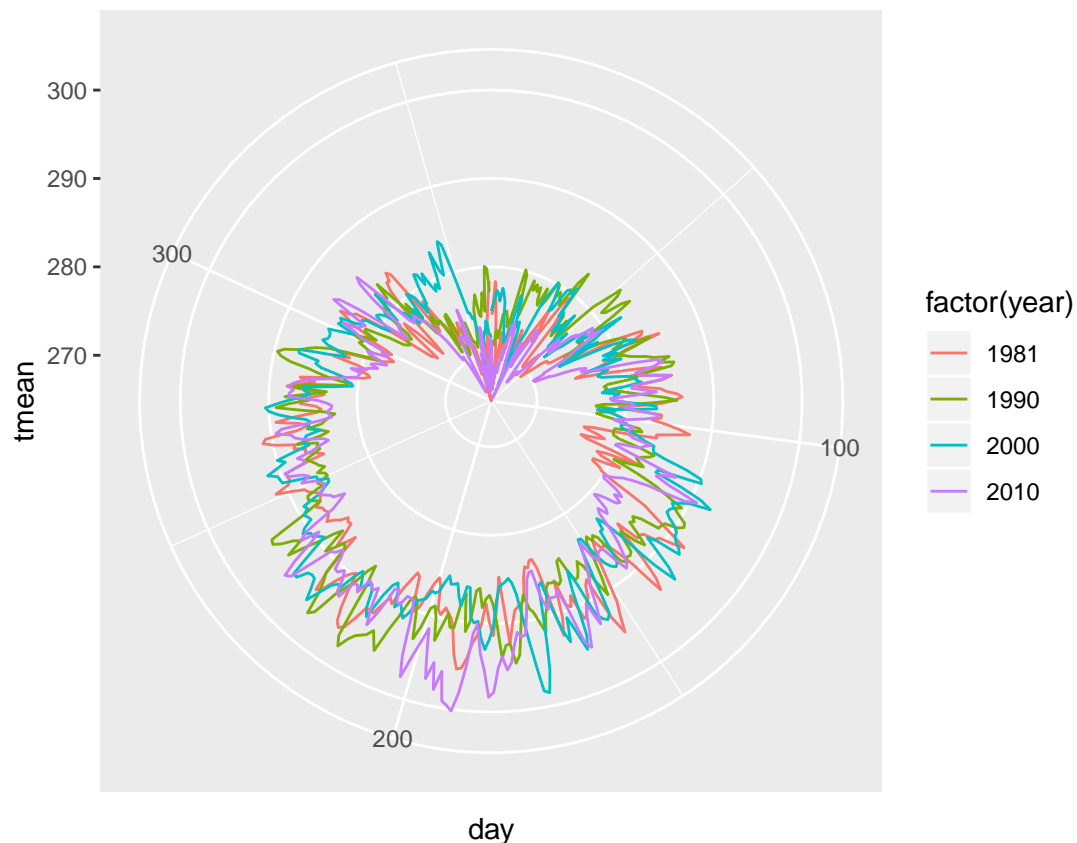
```
##      longitude latitude      z variable
## 6          8.75    54.75 1981-01-01 275.2337
## 7          9.25    54.75 1981-01-01 274.9547
## 8          9.75    54.75 1981-01-01 275.2550
## 25         9.25    54.25 1981-01-01 275.1317
## 26         9.75    54.25 1981-01-01 275.2247
## 27        10.25    54.25 1981-01-01 275.4385
```

Calculate mean temperature per date Split date into yday and year using the `lubridate` package And make a polar plot of daily mean temperature coloured by year

```
#install.packages("lubridate", dep=T)
library(lubridate)
```

```
##
## Attaching package: 'lubridate'
##
## The following object is masked from 'package:base':
##
##      date
```

```
env %>% group_by(z) %>%
  summarise(tmean=mean(variable)) %>%
  mutate(day = yday(z), year = year(z)) %>%
  filter(year %in% c(1981, 1990, 2000, 2010)) %>%
  ggplot(aes(x=day, y=tmean, colour=factor(year))) +
  geom_line() + coord_polar()
```



Plot map of 30-year average temperature Read `gadm36_DEU_adm1.shp` into R using `st_read()` of the `sf` package Add polygon of Germany to map using `geom_sf`

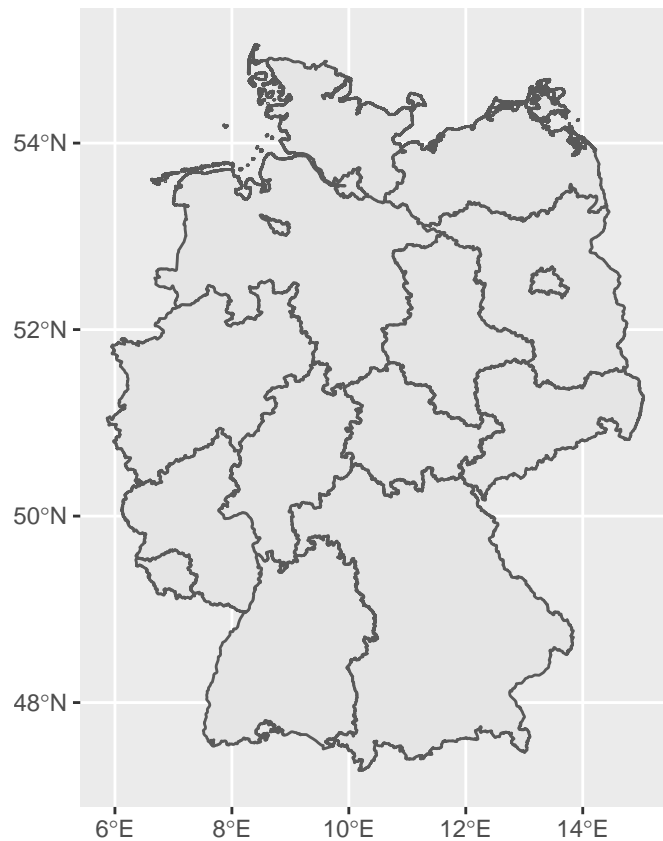
```
#install.packages("sf", dep=T)
library(sf)
```

```
# Read shapefile of Germany
deu <- st_read("data/gadm36_DEU_adm1.shp")
```

```
## Reading layer `gadm36_DEU_adm1' from data source `/home/matt/Documents/Github/dmvR/data/gadm36_DEU_a
## Simple feature collection with 16 features and 0 fields
## geometry type:  MULTIPOLYGON
## dimension:      XY
## bbox:           xmin: 5.866251 ymin: 47.27012 xmax: 15.04181 ymax: 55.05653
## epsg (SRID):    4326
## proj4string:     +proj=longlat +datum=WGS84 +no_defs
```

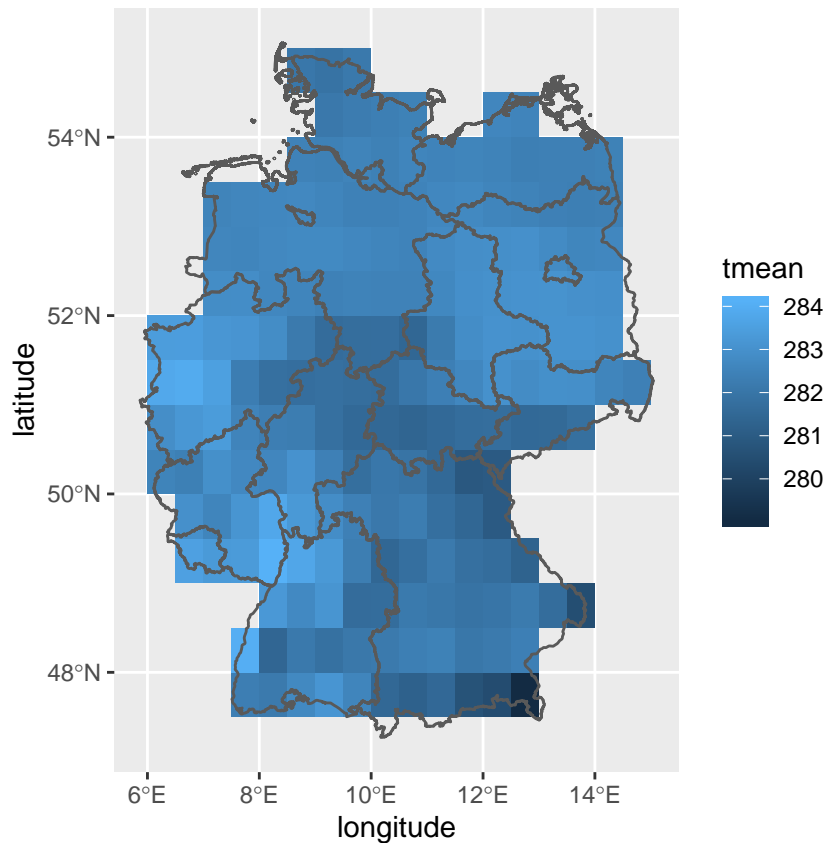
```
#bav <- deu %>% filter(NAME_1=="Bayern")
```

```
# Plot the shapefile
deu %>% ggplot() + geom_sf()
```



Note: `geom_sf()` automatically plots the shapefile with a black outline and grey filling, which is not meaningful if we want to overlay the shapefile onto our data, thus we need to specify the fill argument, in the plot below

```
env %>% group_by(longitude,latitude) %>%
  summarise(tmean=mean(variable)) %>%
  ggplot() + geom_tile(aes(x=longitude, y=latitude,
                           fill=tmean)) +
  geom_sf(data=deu, fill="transparent")
```



For every grid cell calculate monthly mean temperature per year using `as.yearmon()` from the `zoo` package. Make a linear model using `do()` and extract Intercept, Slope and R2 values per grid cell.

```
#install.packages("zoo", dep=T)
library(zoo)

##
## Attaching package: 'zoo'

## The following objects are masked from 'package:base':
##
##   as.Date, as.Date.numeric

mod_lm <- env %>% mutate(yearmon = as.yearmon(z)) %>%
  group_by(yearmon, latitude, longitude) %>%
  summarise(tmean=mean(variable)) %>%
  group_by(longitude,latitude) %>%
  do(mod = lm(tmean ~ yearmon, data = .)) %>%
  mutate(Intercept = summary(mod)$coeff[1],
         Slope = summary(mod)$coeff[2],
         R2 = summary(mod)$r.squared) %>%
  dplyr::select(-mod)
```

Plot map of slope and R2 With `ggspatial`, you can add a scale bar to your map using the `annotation_scale()` function

```
#install.packages("ggspatial", dep=T)
library(ggspatial)
```

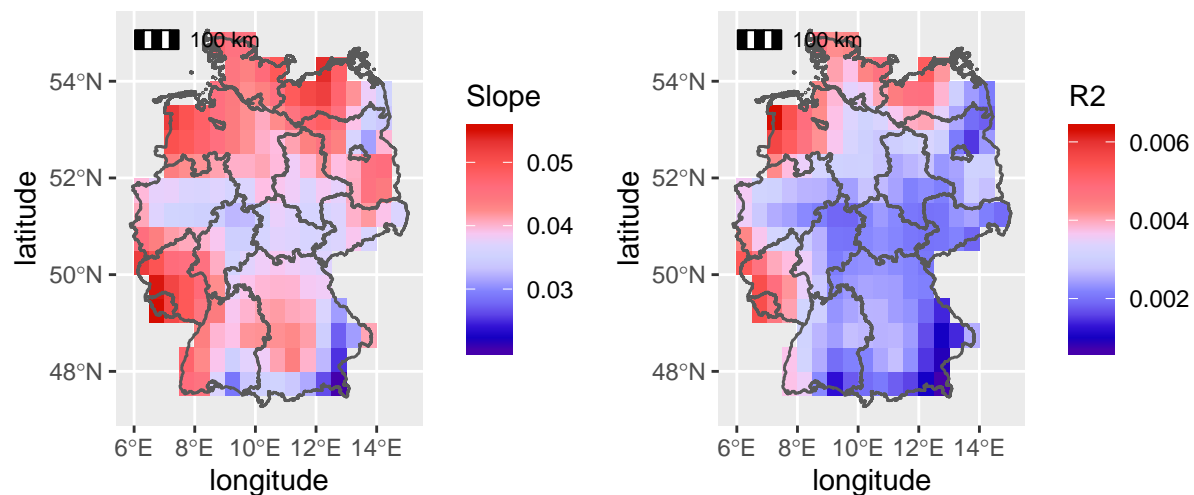
The `ggsci` package, provides easy to use scientific color palettes for `ggplot2`

```
#install.packages("ggsci", dep=T)
library(ggsci)

p1 <- mod_lm %>% ggplot() +
  geom_tile(aes(x=longitude, y=latitude, fill=Slope)) +
  scale_fill_gsea() +
  geom_sf(data=deu, fill="NA") +
  annotation_scale(location="tl") +
  coord_sf()
p2 <- mod_lm %>% ggplot() +
  geom_tile(aes(x=longitude, y=latitude, fill=R2)) +
  scale_fill_gsea() +
  geom_sf(data=deu, fill="NA") +
  annotation_scale(location="tl") +
  coord_sf()

p1 + p2
```

Scale on map varies by more than 10%, scale bar may be inaccurate
 ## Scale on map varies by more than 10%, scale bar may be inaccurate



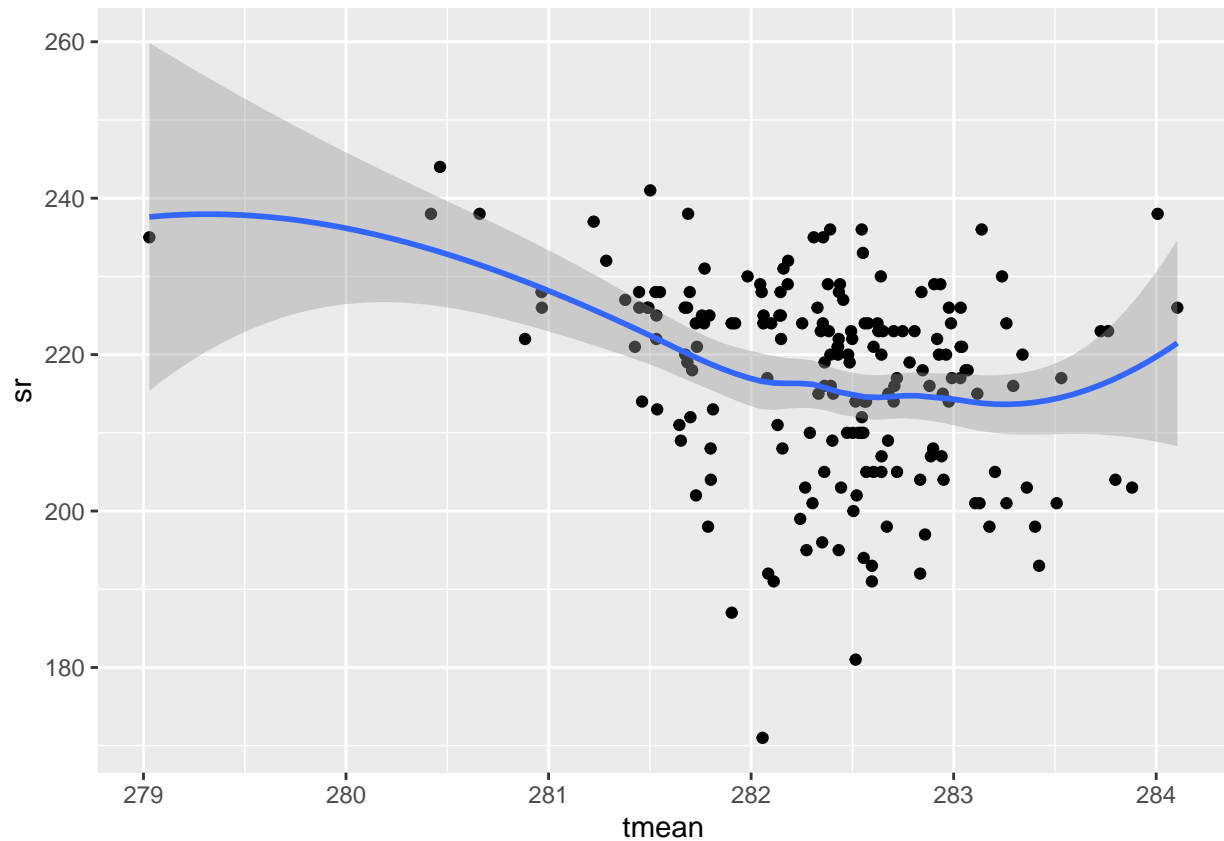
Join species_long with species_info and then join 30-year average temperature per year with species data

```
species_all <- species_long %>%
  left_join(species_info, by=("binomial"))
env_sp <- env %>% mutate(yearmon = as.yearmon(z)) %>%
  group_by(latitude, longitude) %>%
  summarise(tmean=mean(variable)) %>%
  right_join(species_all,
    by=c("longitude"="x", "latitude"="y"))
```

Plot richness against temperature

```
env_sp %>% group_by(longitude,latitude) %>%
  summarise(sr=sum(occurrence), tmean=mean(tmean)) %>%
  ggplot(aes(x=tmean, y=sr)) + geom_point() + geom_smooth()
```

`geom_smooth()` using method = 'loess' and formula 'y ~ x'



Plot richness against temperature per class

```
env_sp %>% group_by(longitude,latitude, class_name) %>%
  summarise(sr=sum(occurrence), tmean=mean(tmean)) %>%
  ggplot(aes(x=tmean, y=sr)) + geom_point() +
  labs(x="Mean temperature (C)", y="Species richness") +
  geom_smooth() + facet_wrap(. ~ class_name, scales="free")

## `geom_smooth()` using method = 'loess' and formula 'y ~ x'
```

