2024 SISBID Graphical Models Lab

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Load packages

```
library("igraph")
library("huge")
library("glasso")
library("glmnet")
library("ggplot2")
```

Read the Sachs et al data: Flow cytometry proteomics in single cells, p=11 proteins measured in n=6466 cells.

```
load("UnsupL_SISBID_2024.Rdata")
p <- ncol(sachsdat)
n <- nrow(sachsdat)
dim(sachsdat)</pre>
```

```
## [1] 7466 11
```

head(sachsdat)

```
##
              praf
                         pmek
                                   plcg
                                             PIP2
                                                        PIP3
                                                                    P44
## [1,]
        -97.67193 -132.18100 -46.03364 -132.8207
                                                   31.765040 -20.021190 -64.16721
## [2,]
        -88.17193 -128.88100 -42.55364 -134.3207 -18.904960 -8.031193 -48.66721
## [3,]
        -64.67193 -101.28100 -40.25364 -140.9207 -14.034960 -11.731190 -48.66721
## [4,]
        -51.07193 -62.58096 -31.75364 -137.6207 -25.744960 -20.801190 -69.36721
## [5,]
        -90.37193 -125.58100 -49.66364 -141.3907
                                                  -2.234962 -5.531193 -35.06721
##
  [6,] -105.27190 -141.63100 -37.25364 -129.0207 -16.134960 -14.731190 -55.46721
##
               PKA
                         PKC
                                   P38
## [1,] -211.75860 -13.34166
                             -90.1145 -33.267500
## [2,] -273.75860 -26.97166 -118.5145 -11.767500
## [3,] -222.75860 -18.94166 -103.1145 -53.767500
## [4,]
        -97.75859 -16.64166 -106.4145 -50.167500
## [5,] -320.75860 -25.68166 -109.3145
        -15.75859 -16.64166 -85.9145 -15.467500
```

Coexpression network

simple thresholding of correlations, at a cutoff chosen to give similar number of edges to partial correlation methods

• a randomly chosen threshold

```
sachscor = cor(sachsdat)
tau <- 0.1
A1 <- abs(sachscor) > tau
diag(A1) <- 0
sum(A1)/2</pre>
```

[1] 43

testing for nonzero correlations

• testing for nonzero correlation, using Fisher Z-transform

```
fisherzs <- atanh(sachscor)
fisherps <- 2*pnorm(abs(fisherzs), 0, 1/sqrt(n-3), lower.tail=FALSE)
A2 <- fisherps < (0.01/(p*(p-1)/2))
diag(A2) <- 0
sum(A2)/2</pre>
```

[1] 45

```
g1 <- graph.adjacency(A1, mode="undirected")</pre>
```

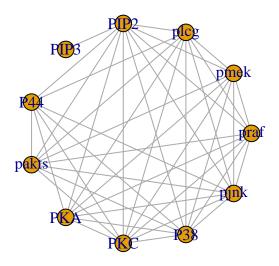
plot the two networks

```
## Warning: 'graph.adjacency()' was deprecated in igraph 2.0.0.
## i Please use 'graph_from_adjacency_matrix()' instead.
## This warning is displayed once every 8 hours.
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was
## generated.
```

```
g2 <- graph.adjacency(A2, mode="undirected")</pre>
```

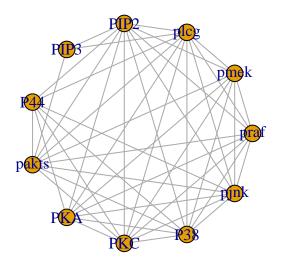
plot(g1,layout=layout.circle(g1), main='simple thresholding of correlations')

simple thresholding of correlations



plot(g2,layout=layout.circle(g2), main='testing for nonzero correlations')

testing for nonzero correlations



Partial correlation networks

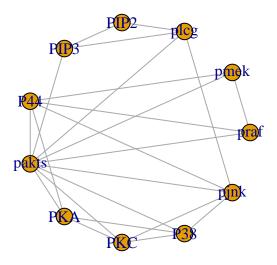
inverse covariance matrix

```
invcov <- abs(round(solve(sachscor),3))
invcor <- cov2cor(invcov)
A1 <- 1*(invcor > 0.05)
diag(A1) <- 0
sum(A1)/2</pre>
```

[1] 23

```
ginv <- graph.adjacency(A1, mode="undirected")
plot(ginv,layout=layout.circle(ginv),main = "Partial correlation networks")</pre>
```

Partial correlation networks



Graphical lasso

Calculate lambda, based on formula in the slides (the third method)

```
alpha <- 0.01
num <- qt(p=alpha/(2*(p^2)),df=n-2, lower.tail=F)
lambda <- num / sqrt(n-2 + num)</pre>
```

Apply glasso

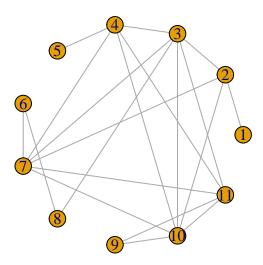
Neighborhood selection

```
ns.est <- glasso(s=sachscor, rho=lambda, approx=TRUE, penalize.diagonal=FALSE)
A3 <- abs(ns.est$wi) > 1E-16; diag(A3) <- 0
gns <- graph.adjacency(A3, mode="undirected")</pre>
```

Compare graph estimates:

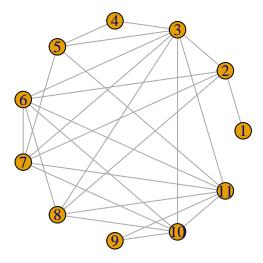
```
plot(gglasso,layout=layout.circle(g1), main='Graphical Lasso')
```

Graphical Lasso



plot(gns,layout=layout.circle(g2), main='Neighborhood Selection')

Neighborhood Selection



Hint: Try changing λ and see how the graph estimates change.

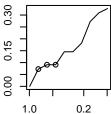
Neighborhood selection estimate with huge (Stability selection for the value of λ)

```
X <- data.matrix(scale(sachsdat))
neth = huge(X,method="mb")</pre>
```

Conducting Meinshausen & Buhlmann graph estimation (mb)....done

plot(neth)

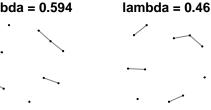
parsity vs. Regularization



lambda = 0.767



lambda = 0.594



Regularization Parameter

```
## stability selection with huge
net.s <- huge.select(neth, criterion="stars")</pre>
```

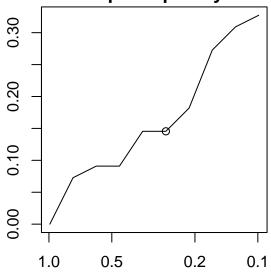
Conducting Subsampling....in progress:5% Conducting Subsampling....in progress:10% Conducting Subsampling....

net.s

```
## Model: Meinshausen & Buhlmann Graph Estimation (mb)
## selection criterion: stars
## Graph dimension: 11
## sparsity level 0.1454545
```

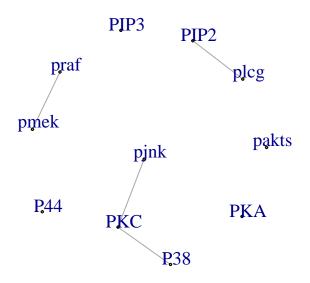
plot(net.s)

Solution path sparsity levels



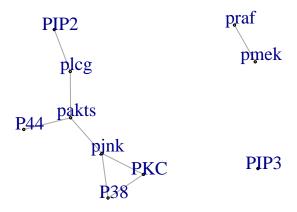
Regularization Parameter

```
#larger lambda
mat <- neth$path[[2]]</pre>
neti <- as.undirected(graph_from_adjacency_matrix(mat))</pre>
plot(neti,vertex.label=colnames(X),vertex.size=2,vertex.label.cex=1.2,vertex.label.dist=1,layout=layout
```



```
#smaller lambda
mat = neth$path[[5]]
neti = as.undirected(graph_from_adjacency_matrix(mat))
plot(neti,vertex.label=colnames(X),vertex.size=2,vertex.label.cex=1.2,vertex.label.dist=1,layout=layout
```

PKA



Nonparanormal Models: rank-based correlation

```
scor <- cor(sachsdat,method='spearman')
scor <- 2*sin(scor*pi/6)
npn.est <- glasso(s=scor, rho=lambda, approx=FALSE, penalize.diagonal=FALSE)
A4 <- abs(npn.est$wi) > 1E-16
diag(A4) <- 0
gnp1 <- graph.adjacency(A4, mode="undirected")</pre>
```

Nonparanormal Models – alternative estiamtion

```
npn.cor <- huge.npn(x=sachsdat, npn.func="skeptic", npn.thresh=NULL, verbose=FALSE)
npn.est <- glasso(s=npn.cor, rho=lambda, penalize.diagonal=FALSE)
A5 <- abs(npn.est$wi) > 1E-16
diag(A5) <- 0
gnp2 <- graph.adjacency(A5, mode="undirected")</pre>
```

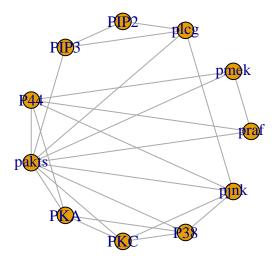
binary network estimation

```
sachsbin \leftarrow 1*(sachsdat > 0) + -1*(sachsdat \leftarrow 0)
head(sachsbin)
##
       praf pmek plcg PIP2 PIP3 P44 pakts PKA PKC P38 pjnk
                   -1
## [1,]
              -1
                              1 -1
                                        -1 -1 -1 -1
         -1
                         -1
                                               -1
## [2,]
          -1
               -1
                    -1
                         -1
                              -1
                                  -1
                                        -1
                                           -1
                                           -1
## [3,]
         -1
                       -1
                             -1 -1
                                        -1
                                               -1
                                                         -1
              -1
                  -1
                                                   -1
## [4,]
         -1
              -1 -1
                       -1
                            -1 -1
                                        -1 -1 -1 -1
## [5,]
         -1
              -1
                   -1
                        -1
                             -1 -1
                                        -1 -1 -1 -1
                                                         1
## [6,]
                              -1 -1
                                        -1 -1 -1 -1
bin.est <- matrix(0,p,p)</pre>
## estiamte the neighborhood for each node
for(j in 1:p){
  ## this is the same method used in neighborhood selection, the only difference is 'family'
  nbr <- glmnet(x=sachsbin[,-j], y=sachsbin[,j], family='binomial', lambda=lambda)</pre>
  bin.est[j,-j] <- 1*(abs(as(nbr$beta, "matrix")) > 0) #store the estimates in jth row of matrix
}
A6 <- bin.est
diag(A6) <- 0
sum(A6)/2
## [1] 20
gising <- graph.adjacency(A6, mode="undirected")</pre>
```

plot the networks

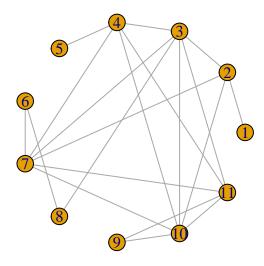
```
plot(ginv,layout=layout.circle(ginv), main='Partial correlation networks')
```

Partial correlation networks



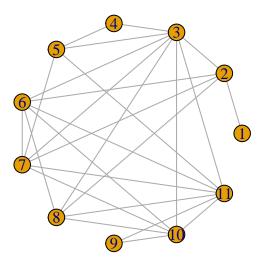
plot(gglasso,layout=layout.circle(gglasso), main='Glasso')

Glasso



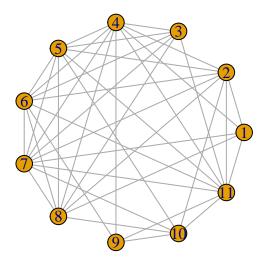
plot(gns,layout=layout.circle(gns), main='Neighborhood selection')

Neighborhood selection



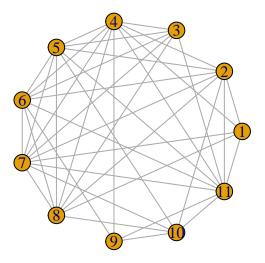
plot(gnp1,layout=layout.circle(gnp1), main='nonparanormal')

nonparanormal



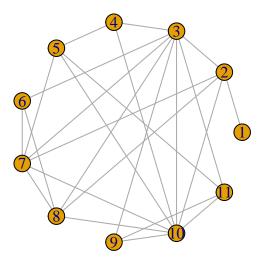
plot(gnp2,layout=layout.circle(gnp2), main='nonparanormal - v2')

nonparanormal - v2



```
plot(gising,layout=layout.circle(gising), main='Binary')
```

Binary



Community detection with stochastic block models

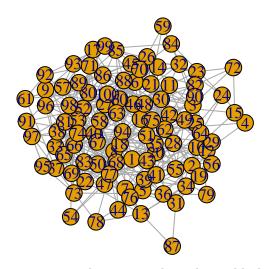
```
# generation function
gen.A.from.B <- function(B,n,c,undirected=TRUE){
    g <- vector()
    K <- length(c)
    for(i in 1:(K-1)){
        g <- c(g,rep(i,c[i]*n))
    }
    g <- c(g,rep(K,n - length(g)))
    Z <- matrix(0,n,K)</pre>
```

```
Z[cbind(1:n,g)] <- 1</pre>
  P <- Z%*%B%*%t(Z)
  n \leftarrow nrow(P)
  if(undirected){
    upper.tri.index <- which(upper.tri(P))</pre>
    tmp.rand <- runif(n=length(upper.tri.index))</pre>
    \#A \leftarrow matrix(0,n,n)
    A <- rsparsematrix(n,n,0)
    A[upper.tri.index[tmp.rand<P[upper.tri.index]]] <- 1
    A \leftarrow A+t(A)
    diag(A) \leftarrow 0
    return(list(A=A,g=g))
  }else{
    A <- matrix(0,n,n)
    r.seq <- runif(n=length(P))</pre>
    A[r.seq < as.numeric(P)] <- 1
    diag(A) <- 0
    return(list(A=A,g=g))
  }
}
```

visualization with a small network

```
n <- 100
K <- 2
B <- matrix(0.05,K,K)
diag(B) <- 0.1
c <- rep(1/K,K)
graph <- gen.A.from.B(B,n,c)
A <- graph$A
true_label <- graph$g
neti = as.undirected(graph_from_adjacency_matrix(A))
plot(neti,layout=layout_with_kk, main='stochastic block models')</pre>
```

stochastic block models



community detection with stochastic block models

```
n <- 1000
K <- 2
B <- matrix(0.05,K,K)
diag(B) <- 0.1
c <- rep(1/K,K)
graph <- gen.A.from.B(B,n,c)
A <- graph$A
true_label <- graph$g
evA <- RSpectra::eigs(A,k = K)
clusterA <- kmeans(evA$vectors,K)
estimated_label <- clusterA$cluster</pre>
```

plot

```
newdata <- data.frame(v1 = evA$vectors[,1], v2 = evA$vectors[,2], true_label = as.factor(true_label), e
ggplot(newdata)+
  geom_point(aes(x = v1,y = v2,colour = estimated_label, shape = true_label))</pre>
```

