

hu-neuro-pipeline

A Python implementation of the single trial EEG pipeline by Frömer et al. (*Front. Neurosci.*, 2018)

Alexander Enge

Neuro Lab @ Humboldt-Universität zu Berlin

20/04/2022

Why Python?



Why MNE-Python?



- Versatile
 - EEG, MEG, ECoG, fNIRS
 - Preprocessing, statistics, time-frequency analysis, visualization, machine learning, connectivity, source localization, . . .
- Open source
 - 288 contributors on GitHub as of April 2022
 - Funding: NIH, NSF, ERC, Google, Amazon, ...
- Community standards
 - Code review, automatic tests, user forum, office hours, ...

Why the Frömer et al. (2018) pipeline?



- Allows single trial analysis of ERP amplitudes
 - Treat items as random effects (Bürki et al., 2018)
 - Model trial and item level covariates
 - Include continuous predictor variables
 - Handle unbalanced designs via partial pooling
 - Weaker assumptions than ANOVA

Why this re-implementation?



- User friendly, e.g.:
 - No MATLAB license; can be called from within R
 - Outputs readily usable for mixed models and plotting
- New features, e.g.:
 - Time-frequency analysis
 - Automatic ocular correction (ICA) + bad channel detection
- $\bullet \ \ Code \ standards + versioning \ {\scriptstyle (https://github.com/alexenge/hu-neuro-pipeline/)} \\$

And why not?



- More difficult to debug or modify
- Possibly not all features supported (e.g., RIDE)
- EEGLAB still more widely use than MNE-Python

Installation



For Python users:

```
# Install via the command line from the Python Packaging Index (PyPI) python3 -m pip install hu-neuro-pipeline
```

For R users:

```
# Install reticulate for interfacing with Python from R
install.packages("reticulate")

# Install the Miniconda Python distribution
reticulate::install_miniconda()

# Install the actual package
reticulate::py_install("hu-neuro-pipeline", pip = TRUE, python_version = "3.8")
```

General usage



```
# Import the Python package
pipeline <- reticulate::import("pipeline")

# Run the pipeline
res <- pipeline$group_pipeline(...)</pre>
```

A simple example



```
# Import the Puthon package
pipeline <- reticulate::import("pipeline")</pre>
# Run the pipeline
res <- pipeline$group_pipeline(
  # Input/output paths
 vhdr files = "data/raw".
 log_files = "data/log",
 output_dir = "output",
  # Preprocessing options
 ocular correction = "data/cali".
  # Epoching options
 triggers = c(201:208, 211:218).
 components = list(
   "name" = list("N2", "P3b"),
   "tmin" = list(0.25, 0.4),
    "tmax" = list(0.35, 0.55).
    "roi" = list(
     c("FC1", "FC2", "C1", "C2", "Cz"),
     c("CP3", "CP1", "CPz", "CP2", "CP4", "P3", "Pz", "P4", "P03", "P0z", "P04")
 ),
  # Averaging options
 average_by = c("n_b", "DeviantPosRL", "n_b/DeviantPosRL"),
```



```
# Input/output paths
vhdr_files = "data/raw",
log_files = "data/log",
output_dir = "output",
```

- Directory or list of raw EEG files (.vhdr)
- Directory or list of behavioral log files (.txt/.tsv/.csv)
- Output directory



```
# Preprocessing options
ocular_correction = "data/cali",
```

- Ocular correction:
 - Path or list of BESA files (.matrix) or
 - "auto" for independent component analysis (ICA)
- Default bandpass filter (0.1–40 Hz)



```
# Epoching options
triggers = c(201:208, 211:218),
components = list(
   "name" = list("N2", "P3b"),
   "tmin" = list(0.25, 0.4),
   "tmax" = list(0.35, 0.55),
   "roi" = list(
      c("FC1", "FC2", "C1", "C2", "Cz"),
      c("CP3", "CP1", "CP2", "CP2", "CP4", "P3", "P2", "P4", "P03", "P0z", "P04")
),
```

- List of numerical EEG triggers
- List of ERP component definitions:
 - name: Column names for each component
 - tmin + tmax: Onset and offset times (in s)
 - roi: List of channel names for each component



```
# Averaging options
average_by = c("n_b", "DeviantPosRL", "n_b/DeviantPosRL")
```

 List of column names (for main effects) and combinations of column names (for interaction effects, separated by "/")

More Pipeline inputs



- Downsampling (downsample_sfreq)
- Interpolate bad channels (bad_channels)
- Frequency filter (highpass_freq, lowpass_freq)
- Epoch duration (epochs_tmin, epochs_tmax)
- Baseline duration (baseline_tmin, baseline_tmax)
- Skip log file rows (skip_log_rows, skip_log_conditions)
- Threshold for artifact rejection (reject_peak_to_peak)

See https://github.com/alexenge/hu-neuro-pipeline/blob/main/docs/inputs.md



Extract directly from the pipeline run:

```
trials <- res[[1]] # Single trial data frame
evokeds <- res[[2]] # Evokeds data frame
config <- res[[3]] # List of pipeline options
```

Or read from the output directory:

```
library(tidyverse)
trials <- read_csv("output/trials.csv")
evokeds <- read_csv("output/ave.csv")
config <- jsonlite::read_json("output/config.json")</pre>
```

See https://github.com/alexenge/hu-neuro-pipeline/blob/main/docs/outputs.md

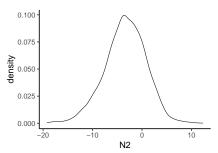


```
# Single trial data frame
print(trials)
## # A tibble: 3,840 x 33
     participant id VPNummer version
                                       wdh lfdNr n_b Standard Deviant Objektpaar
                               <dbl> <dbl> <dbl> <chr> <chr>
##
     <chr>>
                       <dbl>
                                                                <chr>
                                                                             <dh1>
  1 09
                                               1 norm~ objekt5~ objekt~
   2 09
                                               2 blurr objekt5~ objekt~
## 3.09
                                               3 blurr objekt3~ objekt~
  4 09
                                               4 blurr objekt4~ objekt~
                                                                                11
  5 09
                                            5 norm~ objekt3~ objekt~
                                                                                10
   6 09
                                           6 blurr objekt1~ objekt~
   7 09
                                           7 blurr objekt7~ objekt~
   8 09
                                           8 norm~ objekt3~ objekt~
## 9 09
                                               9 blurr objekt4~ objekt~
## 10 09
                                              10 norm~ objekt3~ objekt~
     ... with 3,830 more rows, and 24 more variables: BedCode_alt <chr>,
## #
      BedCode neu <chr>, bot <dbl>, DeviantPosRL <chr>, DeviantPosNR <dbl>,
## #
      BedCodeRL <chr>, kev <dbl>, ErrorCode <dbl>, RT <dbl>, Pos1 <chr>,
      Pos2 <chr>, Pos3 <chr>, Pos4 <chr>, Pos5 <chr>, Pos6 <chr>, Pos7 <chr>,
## #
      Pos8 <chr>, Pos9 <chr>, Pos10 <chr>, Pos11 <chr>, Pos12 <chr>, N2 <dbl>,
## #
## #
      P3b <dbl>, P1 <dbl>
```



```
# Single trial N2 mean amplitudes
ggplot(trials, aes(x = N2)) +
geom_density() +
theme_classic(base_size = 30)
```

Warning: Removed 73 rows containing non-finite values (stat_density).



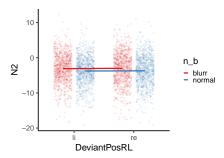
Linear mixed-effects model



```
form <- N2 ~ n_b * DeviantPosRL + (1 | participant_id)
mod <- lme4::lmer(form, trials)
summary(mod)
## Linear mixed model fit by REML ['lmerMod']
## Formula: N2 ~ n_b * DeviantPosRL + (1 | participant_id)
##
     Data: trials
##
## REML criterion at convergence: 21350.4
##
## Scaled residuals:
##
      Min
               1Q Median
                                      Max
## -3.6932 -0.6409 0.0206 0.6501 3.9292
##
## Random effects:
                              Variance Std.Dev.
## Groups
                  Name
## participant_id (Intercept) 1.141
                                      1.068
## Residual
                              16.900 4.111
## Number of obs: 3767, groups: participant_id, 2
##
## Fixed effects:
##
                           Estimate Std. Error t value
## (Intercept)
                           -3.09231 0.76708 -4.031
## n bnormal
                           -0.67552 0.18932 -3.568
## DeviantPosRLre
                          0.11436 0.18952 0.603
## n_bnormal:DeviantPosRLre -0.04093
                                      0.26792 -0.153
```



```
# Single trial N2 mean amplitudes by condition
ggplot(trials, aes(x = DeviantPosRL, y = N2, color = n_b, group = n_b)) +
geom_point(position = position_jitterdodge(0.3), alpha = 0.1) +
stat_summary(
    geom = "line",
    size = 2.,
    position = position_dodge(0.75)
) +
theme_classic(base_size = 30)
```





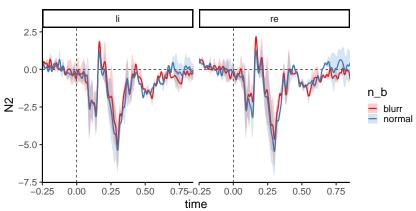
```
# Evokeds by participant and condition
print(evokeds)
```

```
## # A tibble: 12,000 x 71
##
     participant id average by n b
                                     time
                                               Fp1
                                                      Fpz
                                                            Fp2 AF7
##
     <chr>>
                    <chr>>
                                      <chr>
                    n_b
                              normal -0.5
                                            0.790 -0.478 -0.539 0.353 0.459
##
  1 09
  2 09
                    n b
                              normal -0.498 0.765 -0.438 -0.623 0.405 0.544
## 3.09
                    n b
                              normal -0.496 0.648 -0.376 -0.647 0.446 0.537
## 4 09
                              normal -0.494 0.466 -0.315 -0.626 0.477 0.460
                    n_b
## 5 09
                    n_b
                              normal -0.492 0.252 -0.277 -0.577 0.492 0.347
##
  6 09
                    n b
                              normal -0.49
                                            0.0393 -0.273 -0.512 0.487 0.237
                             normal -0.488 -0.145 -0.300 -0.439 0.458 0.163
  7 09
                    n_b
   8 09
                    n_b
                            normal -0.486 -0.286 -0.345 -0.357 0.402 0.140
## 9 09
                    n b
                              normal -0.484 -0.380 -0.389 -0.265 0.318 0.162
## 10 09
                    n b
                              normal -0.482 -0.437 -0.413 -0.160 0.210 0.206
    ... with 11,990 more rows, and 62 more variables: AFz <dbl>, AF4 <dbl>,
## #
      AF8 <dbl>, F9 <dbl>, F7 <dbl>, F5 <dbl>, F3 <dbl>, Fz <dbl>, F4 <dbl>,
## #
      F6 <dbl>, F8 <dbl>, F10 <dbl>, FT7 <dbl>, FC5 <dbl>, FC3 <dbl>, FC1 <dbl>,
      FC2 <dbl>, FC4 <dbl>, FC6 <dbl>, FT8 <dbl>, T7 <dbl>, C5 <dbl>, C3 <dbl>,
## #
## #
      C1 <dbl>, Cz <dbl>, C2 <dbl>, C4 <dbl>, C6 <dbl>, T8 <dbl>, TP9 <dbl>,
      TP7 <dbl>, CP5 <dbl>, CP3 <dbl>, CP1 <dbl>, CPz <dbl>, CP2 <dbl>,
## #
      CP4 <dbl>, CP6 <dbl>, TP8 <dbl>, TP10 <dbl>, P7 <dbl>, P5 <dbl>, ...
## #
```



```
# Evokeds by participant/condition
evokeds %>%
 filter(average_by == "n_b/DeviantPosRL") %>%
 Rmisc::summarvSEwithin(
   measurevar = "N2".
   withinvars = c("time", "n_b", "DeviantPosRL"),
   idvar = "participant id"
 ) %>%
 mutate(time = as.numeric(levels(time))[time]) %>%
 ggplot(aes(
   x = time.
   v = N2
   ymin = N2 - se,
   vmax = N2 + se.
   color = n b.
   fill = n_b
 )) +
 facet_wrap(~DeviantPosRL) +
 geom_hline(vintercept = 0, linetype = "dashed") +
 geom_vline(xintercept = 0, linetype = "dashed") +
 geom line(size = 1) +
  geom_ribbon(color = NA, alpha = 0.2) +
 coord_cartesian(xlim = c(-0.2, 0.8)) +
 theme classic(base size = 20)
```







```
# List of pipeline options
names(config)
  [1] "vhdr_files"
                               "log_files"
                                                     "output_dir"
## [4] "clean dir"
                               "epochs dir"
                                                     "report dir"
## [7] "to df"
                               "downsample_sfreq"
                                                     "veog channels"
## [10] "heog_channels"
                               "montage"
                                                     "bad_channels"
## [13] "ocular correction"
                               "highpass_freq"
                                                     "lowpass_freq"
## [16] "triggers"
                               "triggers_column"
                                                     "epochs_tmin"
## [19] "epochs_tmax"
                                                     "skip_log_rows"
                               "baseline"
## [22] "skip_log_conditions" "reject_peak_to_peak" "components"
## [25] "average_by"
                               "perform tfr"
                                                     "tfr subtract evoked"
## [28] "tfr_freqs"
                               "tfr_cycles"
                                                     "tfr_baseline"
## [31] "tfr_components"
                               "perm_contrasts"
                                                     "perm_tmin"
## [34] "perm_tmax"
                               "perm_channels"
                                                     "perm fmin"
## [37] "perm_fmax"
                               "n iobs"
                                                     "rejected_epochs"
# Number of rejected epochs per participant
lengths(config$rejected epochs)
```

```
## 09 47
## 66 7
```

Automated QC reports



```
# Input/output paths
report_dir = "output/qc_reports",
```

Cluster-based permutation tests



```
# Permutation test options
perm_contrasts = list(
 c("blurr", "normal"),
 c("blurr/re", "blurr/li"),
 c("normal/re", "normal/li")
# Permutation test outputs
clusters <- read csv("output/clusters.csv") # or clusters <- res[[4]]
print(na.omit(clusters))
## # A tibble: 4.991 x 6
##
   contrast
                  time channel t_obs cluster p_val
  <chr>
                  <dbl> <chr> <dbl> <chr>
                                              <db1>
## 1 blurr - normal 0 FT7
                                58.3 pos_1
## 2 blurr - normal 0 P6
                               -14.8 neg 92
## 3 blurr - normal 0.002 T8
                               -90.9 neg_94
## 4 blurr - normal 0.002 CP1
                                 -17.6 neg_101
## 5 blurr - normal 0.004 T8
                                 -22.0 neg 94
## 6 blurr - normal 0.004 CP4
                                 -26.1 neg 95
## 7 blurr - normal 0.004 CP6
                                 -18.0 neg_95
## 8 blurr - normal 0.006 T8
                                 -30.6 neg 94
## 9 blurr - normal 0.006 P7
                                 71.9 pos 101
## 10 blurr - normal 0.008 TP9
                                 110. pos_102
## # ... with 4,981 more rows
```

Automated tools



- Reject bad epochs (reject_peak_to_peak = 200)
 - Using per-channel peak-to-peak amplitudes
- Ocular correction (ocular_correction = "auto")
 - FastICA (Hyvärinen, 1999) + correlation with HEOG/VEOG
- Interpolate bad channels (bad_channel = "auto")
 - Based on per-channel standard error across epochs

One more thing



Auto-match log files to triggers
triggers_column = "trigger",

- Have such a column? Great!
- If not, create in R and pass data frames as log_files

Plans



- Improve documentation + tests
- More detailed QC reports
- Mixed models with pymer4 (?)
- Better permutation tests (Frossard & Renaud, 2021, 2022)
- BIDS interface
- Your ideas + contributions?

Thanks



References



- Bürki, A., Frossard, J., & Renaud, O. (2018). Accounting for stimulus and participant effects in event-related potential analyses to increase the replicability of studies. *Journal of Neuroscience Methods*, 309, 218–227. https://doi.org/10.1016/j.jneumeth.2018.09.016
- Frömer, R., Maier, M., & Abdel Rahman, R. (2018). Group-level EEG-processing pipeline for flexible single trial-based analyses including linear mixed models. Frontiers in Neuroscience, 12, 48. https://doi.org/10.3389/fnins.2018.00048
- Frossard, J., & Renaud, O. (2021). Permutation tests for regression, ANOVA, and comparison of signals: The permuco package. *Journal of Statistical Software*, *99*, 1–32. https://doi.org/10.18637/jss.v099.i15
- Frossard, J., & Renaud, O. (2022). The cluster depth tests: Toward point-wise strong control of the family-wise error rate in massively univariate tests with application to M/EEG. *NeuroImage*, *247*, 118824.
 - https://doi.org/10.1016/j.neuroimage.2021.118824
- Hyvärinen, A. (1999). Fast and robust fixed-point algorithms for independent component analysis. *IEEE Transactions on Neural Networks*, *10*(3), 626–634. https://doi.org/10.1109/72.761722