



hu-neuro-pipeline

A Python-based and R-compatible pipeline
for processing single trial EEG data

Alexander Enge

Neuro Lab @ Humboldt-Universität zu Berlin

2023-07-12



The Frömer et al. (2018) pipeline

frontiers
in Neuroscience

ORIGINAL RESEARCH
doi: 10.3389/fnins.2018.00304

Group-Level EEG-Processing Pipeline for Flexible Single Trial-Based Analyses Including Linear Mixed Models

Romy Frömer^{1*}, Martin Häser², and Martin Abdoel Reichenbach¹

¹Göttingen Institute for Neurobiology, Göttingen University, Göttingen, N. Lichtensteigstrasse, 1, Hannoverstrasse 1, 37075 Göttingen, Germany; ²Barts School of Medicine, Queen Mary University of London, London, United Kingdom, Barts, London, United Kingdom

OPEN ACCESS

Edited by
Andrea Stefanidou,
Institute of Psychology,
University of Crete, Greece

審稿人
Carsten Pfeuffer,
Institute of Biostatistics,
Cognition (IBTC), Chair of
Medical Statistics, University of
Münster, Münster, Germany

Received: 12 January 2018
Accepted: 22 February 2018
Published: 27 March 2018
Keywords:
EEG analysis, behavioral
data, linear mixed models,
cluster-based permutation
tests, processing pipeline

Frömer R, Häser M, Reichenbach MA (2018) Group-Level EEG-Processing Pipeline for Flexible Single Trial-Based Analyses Including Linear Mixed Models. *Frontiers in Neuroscience* 10:304. doi: 10.3389/fnins.2018.00304

Copyright
© 2018 Frömer, Häser and Reichenbach. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forms is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted without the express permission of the copyright owner.

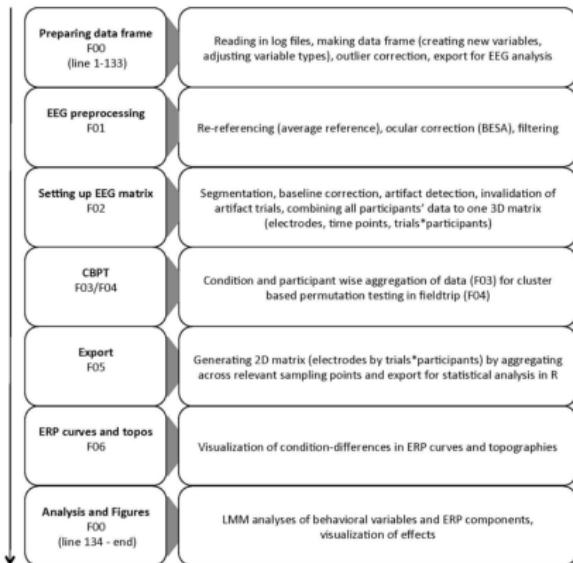
INTRODUCTION
I'll do something 100 times, until every time I can identify it as unique and fresh. I'll do something 100 times, until I'm sure that whatever has been said is true about it as well. How does one express in clear-cut words and plain shape my basic response to this task? How does variability in neural responses relate to variability in behavior?

Additional information: This article was submitted by the editorial team as a review article to ensure that it meets the standards for statistical methods, reproducibility and presentation that are required of all articles published in our journals. The peer review process for this article was overseen by an editor and handled exclusively by members of the editorial board. All review activity was conducted online. This includes manuscript management, tracking, reviewing and editorial decision making. As part of the peer review process, this article was also reviewed by an independent professional editor, or in some cases by two or more independent professionals, prior to being sent back to a member of the editorial board for final review. All editorial correspondence, decisions and next steps for this article are recorded in the review history attached to the article at the end of the document.

*Correspondence:
Romy Frömer,
Göttingen Institute for Neurobiology,
Göttingen University,
N. Lichtensteigstrasse 1,
Hannoverstrasse 1,
37075 Göttingen,
Germany.
E-mail: romy.froemer@uni-goettingen.de

Received: 12 January 2018; Accepted: 22 February 2018; Published: 27 March 2018
Copyright: © 2018 Frömer, Häser and Reichenbach. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forms is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted without the express permission of the copyright owner.

Frontiers in Neuroscience | www.frontiersin.org
8 | February 2018 | Volume 10 | Article 304



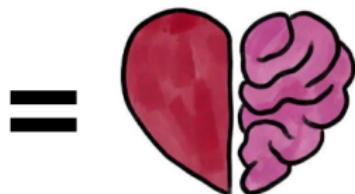
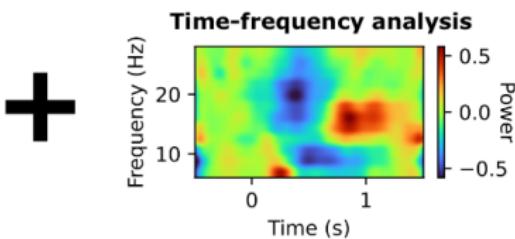
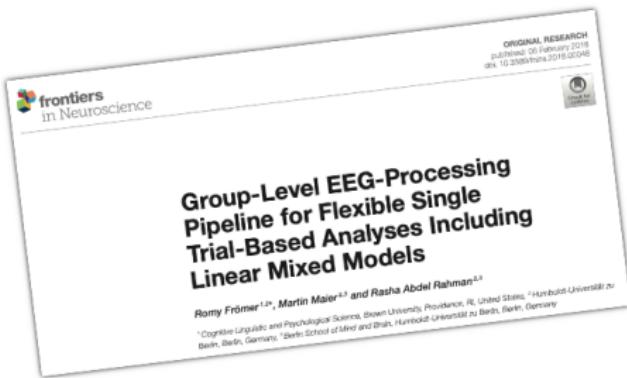
#2

The Frömer et al. (2018) pipeline

- Allows single trial analysis of ERP amplitudes
 - Random effects for items (Bürki et al., 2018)
 - Trial and item level covariates (Volpert-Esmond et al., 2021)
 - Continuous predictor variables
 - Unbalanced designs
 - Brain–behavior associations



Python implementation



#4



Python, I choose you!



Blog post: https://dominiquemakowski.github.io/post/2020-05-22-r_or_python

Online course: <https://swcarpentry.github.io/python-novice-inflammation>



MNE-Python

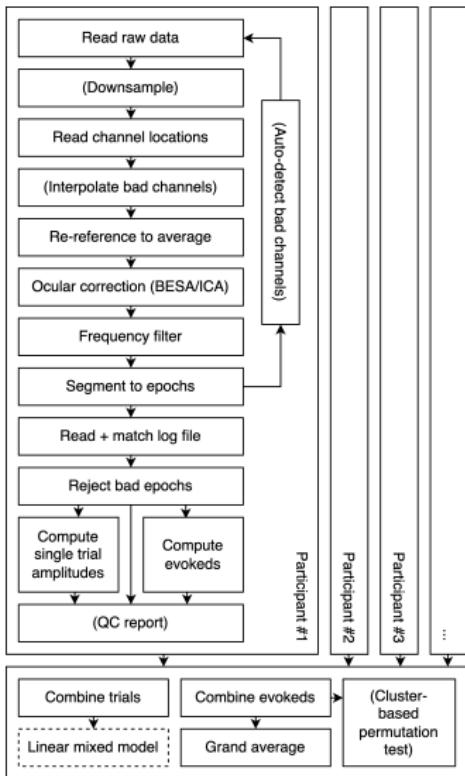
- Versatile
 - EEG, MEG, ECoG, fNIRS
 - Preprocessing, statistics, time-frequency analysis, visualization, machine learning, connectivity, source localization, ...
- Open source
 - > 300 contributors on GitHub (January 2023)
 - Funded by NIH, NSF, ERC, Google, Amazon, ...
 - Code review, automated tests, user forum, office hours, ...



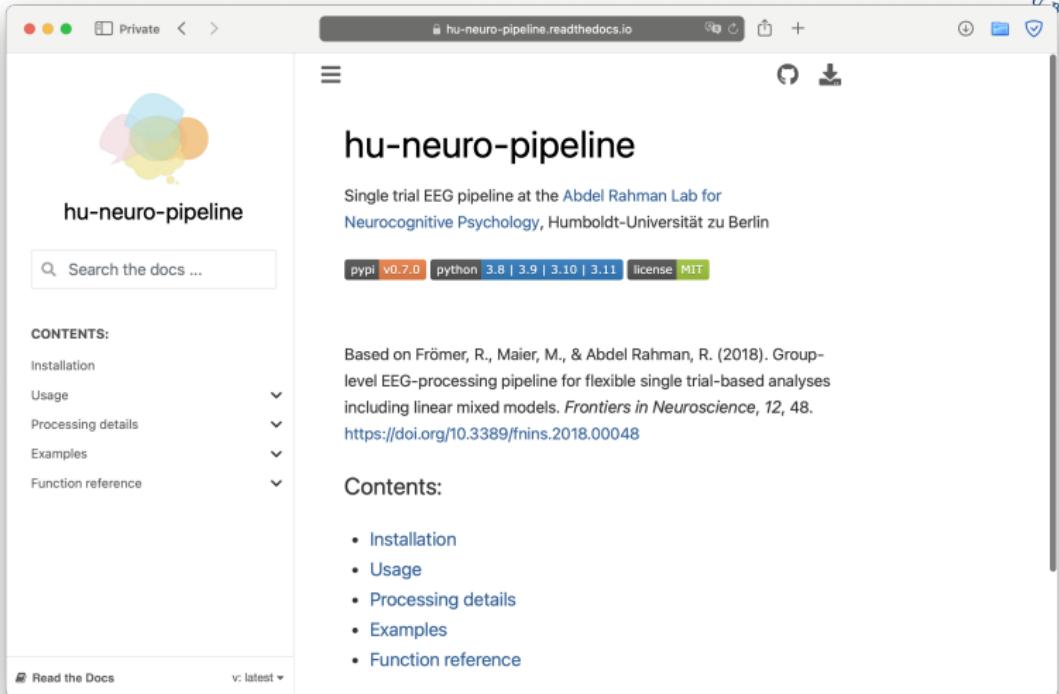
Python implementation

- No MATLAB required
- No Python skills required – can be called from R
- New features:
 - Time-frequency analysis
 - Fully automatic ocular correction (ICA)
 - Automatic bad channel detection
 - Automatic missing trial detection
- Code standards + version control
(<https://github.com/alexenge/hu-neuro-pipeline/>)

Python implementation



Python implementation



The screenshot shows a web browser window displaying the hu-neuro-pipeline.readthedocs.io documentation. The page features a header with the project logo (three overlapping colored circles) and title, a search bar, and a navigation menu. The main content area includes a brief introduction, installation instructions, usage examples, processing details, and function reference sections. A sidebar on the left provides a table of contents for the documentation.

hu-neuro-pipeline

Single trial EEG pipeline at the [Abdel Rahman Lab for Neurocognitive Psychology](#), Humboldt-Universität zu Berlin

pypi v0.7.0 python 3.8 | 3.9 | 3.10 | 3.11 license MIT

CONTENTS:

- Installation
- Usage
- Processing details
- Examples
- Function reference

Based on Frömer, R., Maier, M., & Abdel Rahman, R. (2018). Group-level EEG-processing pipeline for flexible single trial-based analyses including linear mixed models. *Frontiers in Neuroscience*, 12, 48.
<https://doi.org/10.3389/fnins.2018.00048>

Contents:

- Installation
- Usage
- Processing details
- Examples
- Function reference

Read the Docs v: latest

Installation

For Python users:

```
# Install via the command line from the Python Packaging Index (PyPI)
python3 -m pip install hu-neuro-pipeline
```

For R users:

```
# Install reticulate for interfacing with Python from R
install.packages("reticulate")

# Install Python (Miniconda distribution)
reticulate::install_miniconda()

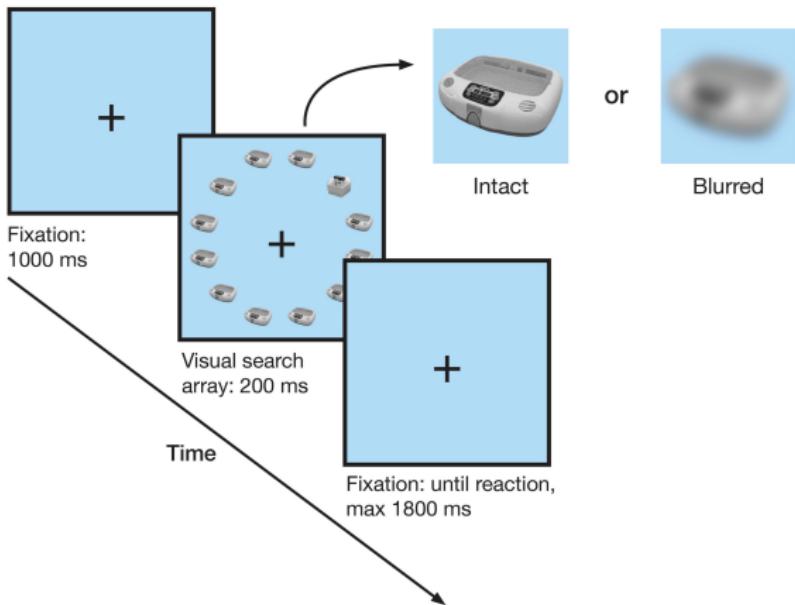
# Install the actual package from PyPI
reticulate::py_install("hu-neuro-pipeline", pip = TRUE, python_version = "3.8")
```

General usage

```
# Import the Python package
pipeline <- reticulate::import("pipeline")

# Run the pipeline
res <- pipeline$group_pipeline(...)
```

Minimal example



For details, see Frömer et al. (2018), <https://doi.org/10.3389/fnins.2018.00048>

Minimal example

```
# Run the pipeline
res <- pipeline$group_pipeline(
  # Input/output paths
  vhdr_files = "data/raw",
  log_files = "data/log",
  output_dir = "output",
  # Preprocessing options
  besa_files = "data/cali",
  # Epoching options
  triggers = c(201:208, 211:218),
  components = list(
    "name" = list("N2", "P3b"),
    "tmin" = list(0.25, 0.4),
    "tmax" = list(0.35, 0.55),
    "roi" = list(
      c("FC1", "FC2", "C1", "C2", "Cz"),
      c("CP3", "CP1", "CPz", "CP2", "CP4", "P3", "Pz", "P4", "P03", "P0z", "P04")
    )
  ),
  # Averaging options
  average_by = list(
    blurr_left = "n_b == 'blurr' and DeviantPosRL == 'li' and RT > 200",
    blurr_right = "n_b == 'blurr' and DeviantPosRL == 're' and RT > 200",
    normal_right = "n_b == 'normal' and DeviantPosRL == 're' and RT > 200",
    normal_left = "n_b == 'normal' and DeviantPosRL == 're' and RT > 200"
  )
)
```

Pipeline inputs

```
# Input/output paths
vhdr_files = "data/raw",
log_files = "data/log",
output_dir = "output",
```

- Directory or list of raw EEG files (.vhdr)
- Directory or list of behavioral log files (.txt/.tsv/.csv)
- Output directory

Pipeline inputs

```
# Preprocessing options
besa_files = "data/cali",
```

- Directory path or list of BESA files (.matrix)
- Default bandpass filter (0.1–40 Hz)

Pipeline inputs

```
# Epoching options
triggers = c(201:208, 211:218),
components = list(
  "name" = list("N2", "P3b"),
  "tmin" = list(0.25, 0.4),
  "tmax" = list(0.35, 0.55),
  "roi" = list(
    c("FC1", "FC2", "C1", "C2", "Cz"),
    c("CP3", "CP1", "CPz", "CP2", "CP4", "P3", "Pz", "P4", "P03", "POz", "P04")
  )
),
```

- List of numerical EEG triggers
- List of ERP component definitions:
 - name: Column names for each component
 - tmin + tmax: Onset and offset times (in s)
 - roi: List of channel names for each component

Pipeline inputs

```
# Averaging options
average_by = list(
    blurr_left = "n_b == 'blurr' and DeviantPosRL == 'li' and RT > 200",
    blurr_right = "n_b == 'blurr' and DeviantPosRL == 're' and RT > 200",
    normal_right = "n_b == 'normal' and DeviantPosRL == 're' and RT > 200",
    normal_left = "n_b == 'normal' and DeviantPosRL == 're' and RT > 200"
)
```

- List with all (combinations of) conditions to create by-participant averages for:
 - List keys are custom labels for each average
 - List values are query strings to select log file rows (trials/epochs)

More pipeline inputs

- Downsampling (`downsample_sfreq`)
- Interpolate bad channels (`bad_channels`)
- Frequency filter (`highpass_freq`, `lowpass_freq`)
- Epoch duration (`epochs_tmin`, `epochs_tmax`)
- Baseline duration (`baseline`)
- Skip log file rows (`skip_log_rows`, `skip_log_conditions`)
- Threshold for artifact rejection (`reject_peak_to_peak`)
- ...

See https://hu-neuro-pipeline.readthedocs.io/en/latest/usage_inputs.html

Pipeline outputs

Extract directly from the pipeline run:

```
trials <- res[[1]]    # Single trial data frame
evokeds <- res[[2]]   # Evokeds data frame
config <- res[[3]]    # List of pipeline options
```

Or read from the output directory:

```
library(tidyverse)
trials <- read_csv("output/trials.csv")
evokeds <- read_csv("output/ave.csv")
config <- jsonlite::read_json("output/config.json")
```

See https://hu-neuro-pipeline.readthedocs.io/en/latest/usage_outputs.html

Pipeline outputs

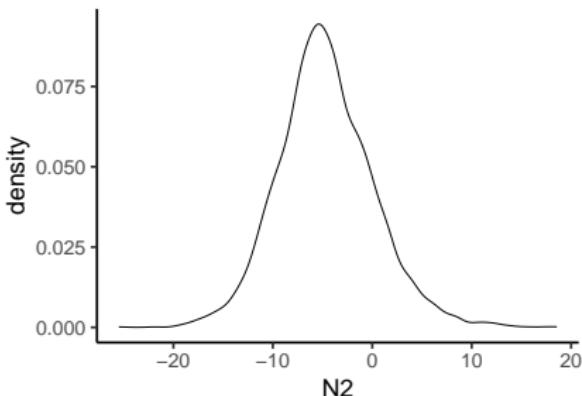
```
# Single trial data frame
print(trials)

## # A tibble: 3,840 x 32
##   participant_id VPNNummer version    wdh lfdNr n_b    Standard Deviant Objektpaar
##   <chr>           <dbl> <dbl> <dbl> <chr> <chr>    <chr>      <dbl>
## 1 05              5     1     1     1 norm~ objekt7~ objekt~       8
## 2 05              5     1     1     2 blurr objekt3~ objekt~      10
## 3 05              5     1     1     3 blurr objekt3~ objekt~      10
## 4 05              5     1     1     4 norm~ objekt2~ objekt~       3
## 5 05              5     1     1     5 norm~ objekt8~ objekt~      15
## 6 05              5     1     1     6 norm~ objekt6~ objekt~       7
## 7 05              5     1     1     7 blurr objekt1~ objekt~       2
## 8 05              5     1     1     8 norm~ objekt8~ objekt~      16
## 9 05              5     1     1     9 norm~ objekt2~ objekt~       9
## 10 05             5     1     1    10 norm~ objekt4~ objekt~      11
## # i 3,830 more rows
## # i 23 more variables: BedCode_alt <chr>, BedCode_neu <chr>, bot <dbl>,
## #   DeviantPosRL <chr>, DeviantPosNR <dbl>, BedCodeRL <chr>, key <dbl>,
## #   ErrorCode <dbl>, RT <dbl>, Pos1 <chr>, Pos2 <chr>, Pos3 <chr>, Pos4 <chr>,
## #   Pos5 <chr>, Pos6 <chr>, Pos7 <chr>, Pos8 <chr>, Pos9 <chr>, Pos10 <chr>,
## #   Pos11 <chr>, Pos12 <chr>, N2 <dbl>, P3b <dbl>
```

Pipeline outputs

```
# Single trial N2 mean amplitudes
ggplot(trials, aes(x = N2)) +
  geom_density() +
  theme_classic(base_size = 30)
```

```
## Warning: Removed 7 rows containing non-finite values ('stat_density()').
```



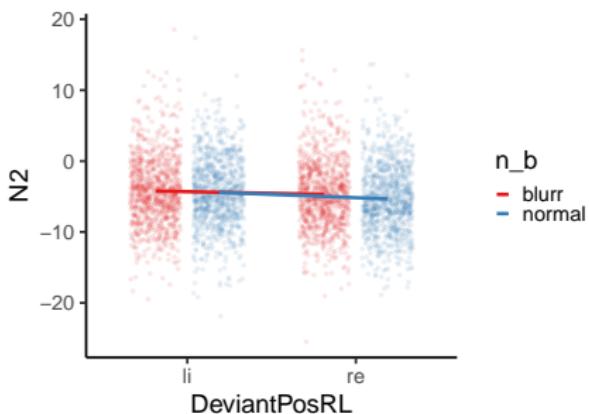
Pipeline outputs

```
# Linear mixed-effects model
form <- N2 ~ n_b * DeviantPosRL + (1 | participant_id)
mod <- lme4::lmer(form, trials)
summary(mod)

## Linear mixed model fit by REML ['lmerMod']
## Formula: N2 ~ n_b * DeviantPosRL + (1 | participant_id)
##   Data: trials
##
## REML criterion at convergence: 22696.1
##
## Scaled residuals:
##     Min      1Q  Median      3Q     Max
## -4.6856 -0.6130 -0.0002  0.6148  5.1121
##
## Random effects:
##   Groups      Name        Variance Std.Dev.
##   participant_id (Intercept) 2.287    1.512
##   Residual           21.780    4.667
## Number of obs: 3833, groups: participant_id, 2
##
## Fixed effects:
##                   Estimate Std. Error t value
## (Intercept)      -4.2285    1.0800 -3.915
## n_bnrmal       -0.1796    0.2132 -0.842
## DeviantPosRLre  -0.4587    0.2132 -2.151
## n_bnrmal:DeviantPosRLre -0.4732    0.3015 -1.569
##
## Correlation of Fixed Effects:
```

Pipeline outputs

```
# Single trial N2 mean amplitudes by condition
ggplot(trials, aes(x = DeviantPosRL, y = N2, color = n_b, group = n_b)) +
  geom_point(position = position_jitterdodge(0.3), alpha = 0.1) +
  stat_summary(
    geom = "line",
    linewidth = 2.0,
    position = position_dodge(0.75)
  ) +
  theme_classic(base_size = 30)
```



Pipeline outputs

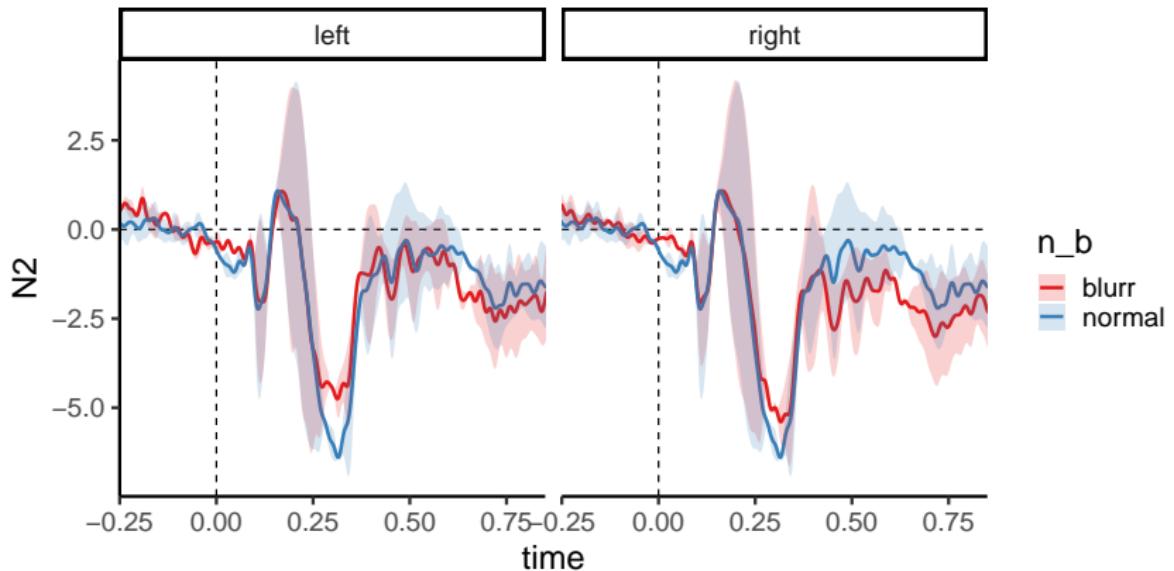
```
# Evokeds by participant and condition
print(evokeds)

## # A tibble: 8,000 x 69
##   participant_id label      query    time     Fp1     Fpz     Fp2     AF7     AF3     AFz
##   <chr>          <chr>    <chr>    <dbl>    <dbl>    <dbl>    <dbl>    <dbl>    <dbl>    <dbl>
## 1 05            blurr_left n_b ~ -0.5    -0.559   -1.41   -1.45   -0.617   -1.12   -1.49
## 2 05            blurr_left n_b ~ -0.498   -0.550   -1.37   -1.35   -0.640   -0.930   -1.45
## 3 05            blurr_left n_b ~ -0.496   -0.528   -1.34   -1.28   -0.666   -0.788   -1.43
## 4 05            blurr_left n_b ~ -0.494   -0.486   -1.31   -1.23   -0.689   -0.722   -1.42
## 5 05            blurr_left n_b ~ -0.492   -0.427   -1.29   -1.21   -0.701   -0.743   -1.40
## 6 05            blurr_left n_b ~ -0.49    -0.355   -1.27   -1.22   -0.698   -0.838   -1.39
## 7 05            blurr_left n_b ~ -0.488   -0.283   -1.26   -1.24   -0.675   -0.977   -1.38
## 8 05            blurr_left n_b ~ -0.486   -0.222   -1.26   -1.27   -0.633   -1.12    -1.37
## 9 05            blurr_left n_b ~ -0.484   -0.187   -1.26   -1.30   -0.577   -1.24   -1.36
## 10 05           blurr_left n_b ~ -0.482   -0.185   -1.26   -1.34   -0.515   -1.31   -1.35
## # i 7,990 more rows
## # i 59 more variables: AF4 <dbl>, AF8 <dbl>, F9 <dbl>, F7 <dbl>, F5 <dbl>,
## #   F3 <dbl>, Fz <dbl>, F4 <dbl>, F6 <dbl>, F8 <dbl>, F10 <dbl>, FT7 <dbl>,
## #   FC5 <dbl>, FC3 <dbl>, FC1 <dbl>, FC2 <dbl>, FC4 <dbl>, FC6 <dbl>,
## #   FT8 <dbl>, T7 <dbl>, C5 <dbl>, C3 <dbl>, C1 <dbl>, Cz <dbl>, C2 <dbl>,
## #   C4 <dbl>, C6 <dbl>, T8 <dbl>, TP9 <dbl>, TP7 <dbl>, CP5 <dbl>, CP3 <dbl>,
## #   CP1 <dbl>, CPz <dbl>, CP2 <dbl>, CP4 <dbl>, CP6 <dbl>, TP8 <dbl>, ...
```

Pipeline outputs

```
# Evokeds by participant/condition
evokeds |>
  separate_wider_delim(label, delim = "_", names = c("n_b", "DeviantPosRL")) |>
  Rmisc::summarySEwithin(
    measurevar = "N2",
    withinvars = c("time", "n_b", "DeviantPosRL"),
    idvar = "participant_id"
  ) |>
  mutate(time = as.numeric(levels(time))[time]) |>
  ggplot(aes(
    x = time,
    y = N2,
    ymin = N2 - se,
    ymax = N2 + se,
    color = n_b,
    fill = n_b
  )) +
  facet_wrap(~DeviantPosRL) +
  geom_hline(yintercept = 0, linetype = "dashed") +
  geom_vline(xintercept = 0, linetype = "dashed") +
  geom_line(linewidth = 1) +
  geom_ribbon(color = NA, alpha = 0.2) +
  coord_cartesian(xlim = c(-0.2, 0.8)) +
  theme_classic(base_size = 20)
```

Pipeline outputs



Pipeline outputs

```
# List of pipeline options
```

```
names(config)
```

```
## [1] "vhdr_files"           "log_files"          "output_dir"  
## [4] "clean_dir"            "epochs_dir"         "report_dir"  
## [7] "to_df"                "downsample_sfreq"   "veog_channels"  
## [10] "heog_channels"        "montage"           "bad_channels"  
## [13] "besa_files"           "ica_method"        "ica_n_components"  
## [16] "highpass_freq"        "lowpass_freq"      "triggers"  
## [19] "triggers_column"      "epochs_tmin"       "epochs_tmax"  
## [22] "baseline"             "skip_log_rows"     "skip_log_conditions"  
## [25] "reject_peak_to_peak"  "components"        "average_by"  
## [28] "perform_tfr"          "tfr_subtract_evoked" "tfr_freqs"  
## [31] "tfr_cycles"           "tfr_mode"          "tfr_baseline"  
## [34] "tfr_components"        "perm_contrasts"    "perm_tmin"  
## [37] "perm_tmax"            "perm_channels"     "perm_fmin"  
## [40] "perm_fmax"             "n_jobs"            "auto_rejected_epochs"  
## [43] "package_versions"
```

```
# Number of rejected epochs per participant
```

```
lengths(config$auto_rejected_epochs)
```

```
## 05 07
```

```
## 7 0
```

More pipeline outputs

- Cleaned continuous data (`clean_dir`)
- Epoched data (`epochs_dir`)
- Automated QC reports (`reports_dir`)

Cluster-based permutation tests

```
# Permutation test input
perm_contrasts = list(
  c("blurr_left", "normal_left"),
  c("blurr_right", "normal_right")
)

# Permutation test outputs
clusters <- read_csv("output/clusters.csv") # or clusters <- res[[4]]
print(na.omit(clusters))

## # A tibble: 3,954 x 6
##   contrast              time channel t_obs cluster p_val
##   <chr>                <dbl> <chr>   <dbl> <chr>    <dbl>
## 1 blurr_left - normal_left 0     T7     735. pos_2      1
## 2 blurr_left - normal_left 0     TP9     13.1 pos_2      1
## 3 blurr_left - normal_left 0.002 AF8    -17.3 neg_285    1
## 4 blurr_left - normal_left 0.002 FC3    381. pos_96     1
## 5 blurr_left - normal_left 0.002 C3     115. pos_96     1
## 6 blurr_left - normal_left 0.002 TP9    91.4 pos_2      1
## 7 blurr_left - normal_left 0.002 TP7    15.6 pos_2      1
## 8 blurr_left - normal_left 0.004 AF8    -20.3 neg_285    1
## 9 blurr_left - normal_left 0.004 F3     23.1 pos_95     1
## 10 blurr_left - normal_left 0.004 TP9    53.8 pos_2      1
## # i 3,944 more rows
```

Artifact correction

- **Multiple source eye correction (MSEC)**
 - Requires .matrix files from BESA

Artifact correction

- **Independent component analysis (ICA)**
 - Different algorithms available
(e.g., `ica_method = "fastica"`)
 - Can specify initial number of principal components with
`ica_n_components`
 - Automatic detection + exclusion of eye movement components
based on correlation with HEOG and VEOG (see https://mne.tools/stable/generated/mne.preprocessing.ICA.html#mne.preprocessing.ICA.find_bads_eog)
 - Verify in QC reports
 - Other selection methods (manual selection, ICLabel) not yet implemented

Artifact rejection

- Per-channel peak-to-peak amplitude threshold via `reject_peak_to_peak` (default: 200.0)
- In addition to or instead of BESA or ICA

Repairing bad channels

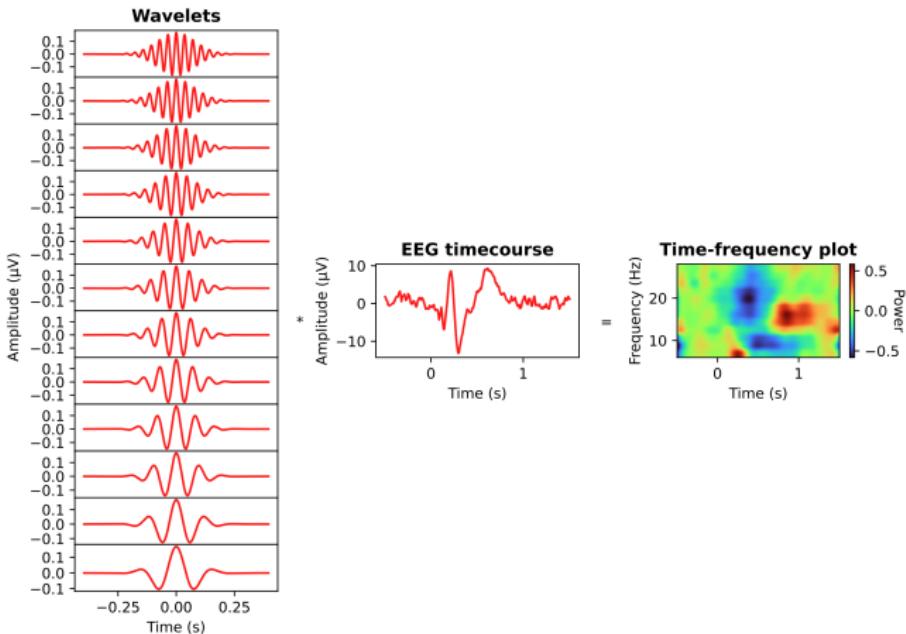
- Pass participant-specific vectors of bad channel labels
 - E.g., `bad_channels = list("05" = c("C3", "P7"), ...)`
- Uses spherical spline interpolation
- Experimental: Automatic bad channel detection
(`bad_channels = "auto"`)
 - Based on channel *SDs* across epochs

Detecting missing epochs

- Requires log file column (`triggers_column`) with the EEG trigger for every trial
- Pipeline magically detects and deletes log file trials with missing EEG



Time-frequency analysis



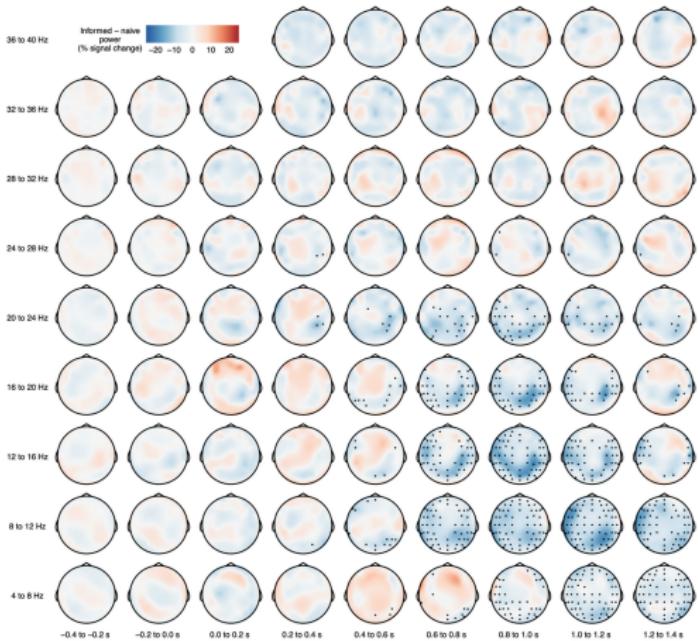
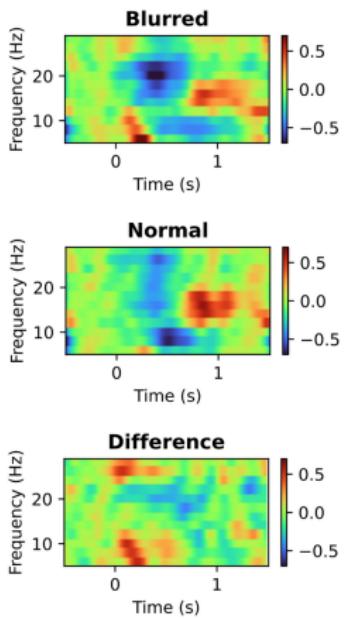
See <https://github.com/alexenge/tfr-workshop>

Time-frequency analysis

```
# Time-frequency analysis options
perform_tfr = TRUE,
tfr_components = list(
  "name" = list("alpha"),
  "tmin" = list(0.0), "tmax" = list(0.2),
  "fmin" = list(8.0), "fmax" = list(14.0),
  "roi" = list(c("P09", "P07", "P03", "POz", "P04", "P08", "P010", "O1", "Oz", "O2"))
)
```

- `tfr_components` extracts single trial power values
- Additional options:
 - Morlet frequencies (`tfr_freqs`, default 4, 5, 6, ..., 40 Hz)
 - Morlet no. of cycles (`tfr_cycles`, default 2, 2.5, 3, ..., 20)
 - Baseline window (`tfr_baseline`, default -450 ms to -50 ms)
 - Baseline method (`tfr_method`, default percent signal change)

Time-frequency analysis



Plans

- Enhance documentation (examples, boilerplate, preprint)
- Unit tests
- RIDE correction for speech artifacts
- Mixed models with `pymer4` or `bambi`
- Better permutation tests (Frossard & Renaud, 2021, 2022)
- BIDS interface
- Your ideas + contributions?

See <https://github.com/alexenge/hu-neuro-pipeline/issues>

Thanks



References

- Bürki, A., Frossard, J., & Renaud, O. (2018). Accounting for stimulus and participant effects in event-related potential analyses to increase the replicability of studies. *Journal of Neuroscience Methods*, 309, 218–227.
<https://doi.org/10.1016/j.jneumeth.2018.09.016>
- Frömer, R., Maier, M., & Abdel Rahman, R. (2018). Group-level EEG-processing pipeline for flexible single trial-based analyses including linear mixed models. *Frontiers in Neuroscience*, 12, 48. <https://doi.org/10.3389/fnins.2018.00048>
- Frossard, J., & Renaud, O. (2021). Permutation tests for regression, ANOVA, and comparison of signals: The permuco package. *Journal of Statistical Software*, 99, 1–32. <https://doi.org/10.18637/jss.v099.i15>
- Frossard, J., & Renaud, O. (2022). The cluster depth tests: Toward point-wise strong control of the family-wise error rate in massively univariate tests with application to M/EEG. *NeuroImage*, 247, 118824.
<https://doi.org/10.1016/j.neuroimage.2021.118824>
- Volpert-Esmond, H. I., Page-Gould, E., & Bartholow, B. D. (2021). Using multilevel models for the analysis of event-related potentials. *International Journal of Psychophysiology*, 162, 145–156. <https://doi.org/10.1016/j.ijpsycho.2021.02.006>