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# Chapter 1

# Introduction

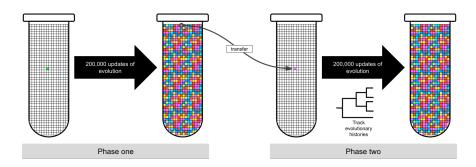


Figure 1.1: Experimental design overview

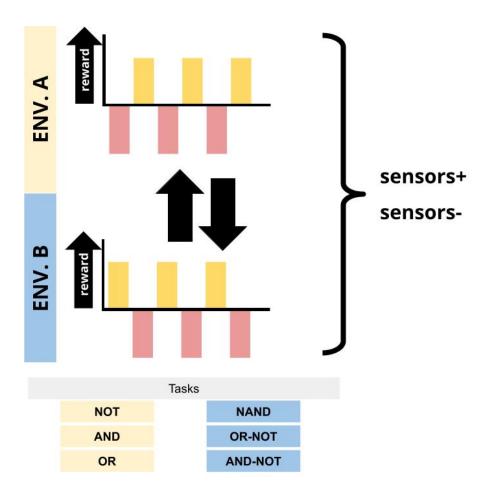


Figure 1.2: Fluctuating environment

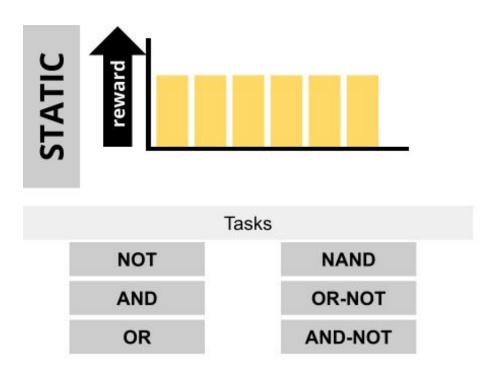


Figure 1.3: Static environment

### Chapter 2

## Validation experiment

In this experiment, we validate that (1) we observe the evolution of phenotypic plasticity in a changing environment when digital organisms have access to sensory instructions (capable of differentiating environmental states) and (2) that adaptive phenotypic plasticity does not evolve when populations lack access to sensory instructions.

#### 2.1 Overview

```
total_updates <- 200000
replicates <- 100

all_traits <- c("not", "nand", "ornot", "or", "andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-01-07-validation/analysis/"
# working directory <- "./"
# << For bookdown
# << For local analysis</pre>
```

We evolved populations of digital organisms under four conditions:

- 1. A fluctuating environment with access to sensory instructions
- 2. A fluctuating environment without access to sensory instructions (i.e., sensory instructions are no-operations)
- 3. A constant environment with access to sensory instructions
- 4. A constant environment without access to sensory instructions

In fluctuating environments, we alternate between rewarding and punishing different sets of computational tasks. In one environment, we reward tasks not,

and, or and punish tasks nand, ornot, andnot. In the alternative environment, we reward tasks nand, ornot, andnot and punish tasks not, and, or. In constant environments, we reward all tasks (not, nand, and, ornot, or, andnot).

For each replicate of each condition, we extract the dominant (i.e., most numerous) genotype at the end of the run to analyze further. We expect to observe the evolution of adaptive phenotypic plasticity in only the first experimental condition. In conditions without sensors, plasticity in any form should be unable to evolve.

#### 2.2 Analysis dependencies

Load all required R libraries.

```
library(ggplot2)
library(tidyverse)
library(cowplot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd9
```

These analyses were conducted/knitted with the following computing environment:

```
print(version)
```

```
x86 64-pc-linux-gnu
## platform
## arch
                  x86_64
## os
                  linux-gnu
## system
                  x86_64, linux-gnu
## status
                  4
## major
## minor
                  0.4
                  2021
## year
## month
                  02
                  15
## day
                  80002
## svn rev
## language
## version.string R version 4.0.4 (2021-02-15)
## nickname
                  Lost Library Book
```

### 2.3 Setup

```
data_loc <- paste0(working_directory, "data/aggregate.csv")
data <- read.csv(data_loc, na.strings="NONE")
data$DISABLE_REACTION_SENSORS <- as.factor(data$DISABLE_REACTION_SENSORS)</pre>
```

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```
data$chg_env <- as.factor(data$chg_env)</pre>
data$dom_plastic_odd_even <- as.factor(data$dom_plastic_odd_even)</pre>
data$sensors <- data$DISABLE_REACTION_SENSORS == "0"</pre>
data$is_plastic <- data$dom_plastic_odd_even == "True"</pre>
env_label_fun <- function(chg_env) {</pre>
 if (chg_env) {
    return("Fluctuating")
  } else {
    return("Constant")
 }
}
sensors_label_fun <- function(has_sensors) {</pre>
  if (has_sensors) {
   return("Sensors")
 } else {
    return("No sensors")
 }
}
# Count observed plasticity for each condition (I'm sure there's a 'tidier' way to do this..)
observed_plasticity <- data.frame(</pre>
  environment=character(),
  sensors=character(),
  plastic=integer(),
  nonplastic=integer(),
  plastic_adaptive=integer(),
  plastic optimal=integer(),
  plastic_nonadaptive=integer()
for (env_chg in levels(data$chg_env)) {
  for (disabled_sensors in levels(data$DISABLE_REACTION_SENSORS)) {
    cond_data <- filter(data, chg_env == env_chg & data$DISABLE_REACTION_SENSORS == disabled_sens
    environment_label <- env_label_fun(env_chg)</pre>
    sensors_label <- sensors_label_fun(disabled_sensors == "0")</pre>
    observed_plasticity <- observed_plasticity %>% add_row(
      environment=environment_label,
      sensors=sensors_label,
      plastic=nrow(filter(cond_data, is_plastic==TRUE)),
      nonplastic=nrow(filter(cond_data, is_plastic==FALSE)),
      plastic_adaptive=nrow(filter(cond_data, dom_adaptive_plasticity=="True")),
      plastic_optimal=nrow(filter(cond_data, dom_optimal_plastic=="True")),
      plastic_nonadaptive=nrow(filter(cond_data, is_plastic==TRUE & dom_adaptive_plasticity=="Fal
```

```
}

observed_plasticity <- pivot_longer(
  observed_plasticity,
  cols=c("plastic", "plastic_adaptive", "plastic_optimal", "plastic_nonadaptive", "nongonames_to="phenotype",
  values_to="phenotype_cnt"
)

####### misc ######

# Configure our default graphing theme
theme_set(theme_cowplot())
</pre>
```

#### 2.4 Evolution of phenotypic plasticity

For each experimental condition, do we observe the evolution of phenotypic plasticity? To test for phenotypic plasticity, we culture digital organisms in both environments from the fluctuating condition (including organisms evolved in a constant environment). Any plasticity that we observe from digital organisms evolved under constant conditions is cryptic variation (as these organisms were never exposed to these culturing environments).

```
ggplot(filter(observed_plasticity, phenotype %in% c("plastic", "nonplastic")), aes(x=p)
  geom_bar(
    stat="identity",
    position=position_dodge(0.9)
  geom_text(
    stat="identity",
   mapping=aes(label=phenotype_cnt),
   vjust=0.05
  ) +
 scale_fill_brewer(palette="Accent") +
  scale_x_discrete(
   name="Phenotype",
   limits=c("plastic", "nonplastic"),
   labels=c("Plastic", "Non-plastic")
  ) +
 facet_grid(sensors~environment) +
 theme(
    legend.position="none"
```



Indeed, we do not observe the evolution of phenotypic plasticity in any replicates in which digital organisms do not have access to sensory instructions. We do observe the evolution of plasticity (not necessarily adaptive plasticity) in both constant and fluctuating environments where sensors are enabled.

To what extent is the observed phenotypic plasticity adaptive?

```
ggplot(filter(observed_plasticity, environment=="Fluctuating" & sensors == "Sensors" & phenotype
  geom_bar(
   stat="identity",
   position=position_dodge(0.9)
  geom_text(
    stat="identity",
   mapping=aes(label=phenotype_cnt),
   vjust=0.05
  ) +
  scale_fill_brewer(palette="Accent") +
  scale_x_discrete(
   name="Phenotype",
   limits=c("plastic", "plastic_adaptive", "plastic_optimal", "plastic_nonadaptive"),
   labels=c("Total plastic", "Adaptive plasticity", "Optimal plasticity", "Non-adaptive plastic
  ) +
  facet_grid(sensors~environment) +
  theme(
    legend.position="none"
```



## Chapter 3

## Evolutionary change

The effect of adaptive phenotypic plasticity on evolutionary change.

#### 3.1 Overview

```
total_updates <- 200000
replicates <- 100
alpha <- 0.05

all_traits <- c("not", "nand", "ornot", "or", "andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-02-08-evo-dynamics/analysis/" # << For bookdown
# working_directory <- "./" # << For local analysis</pre>
```

### 3.2 Analysis dependencies

Load all required R libraries.

```
library(ggplot2)
library(rstatix)
library(ggsignif)
library(scales)
library(tidyverse)
library(cowplot)
library(RColorBrewer)
library(Hmisc)
```

```
library(boot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd9")
```

These analyses were conducted/knitted with the following computing environment:

```
print(version)
                  x86_64-pc-linux-gnu
## platform
## arch
                  x86_64
## os
                  linux-gnu
                  x86_64, linux-gnu
## system
## status
## major
## minor
                  0.4
## year
                  2021
## month
                  02
                  15
## day
## svn rev
                  80002
                  R
## language
## version.string R version 4.0.4 (2021-02-15)
## nickname
                Lost Library Book
```

#### 3.3 Setup

```
summary_data_loc <- paste0(working_directory, "data/aggregate.csv")</pre>
summary_data <- read.csv(summary_data_loc, na.strings="NONE")</pre>
summary_data$DISABLE_REACTION_SENSORS <- as.factor(summary_data$DISABLE_REACTION_SENSO
summary_data$chg_env <- summary_data$chg_env == "True"</pre>
summary_data$dominant_plastic_odd_even <- as.factor(summary_data$dominant_plastic_odd_
summary_data$sensors <- summary_data$DISABLE_REACTION_SENSORS == "0"</pre>
summary_data$is_plastic <- summary_data$dominant_plastic_odd_even == "True"
env_label_fun <- function(chg_env) {</pre>
  if (chg_env) {
   return("Fluctuating")
 } else {
    return("Constant")
  }
}
sensors_label_fun <- function(has_sensors) {</pre>
 if (has_sensors) {
```

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```
return("Sensors")
  } else {
    return("No sensors")
  }
}
# note that this labeler makes assumptions about how we set up our experiment
condition_label_fun <- function(has_sensors, env_chg) {</pre>
  if (has_sensors && env_chg) {
    return("PLASTIC")
  } else if (env_chg) {
    return("NON-PLASTIC")
  } else {
    return("STATIC")
  }
}
summary_data$env_label <- mapply(</pre>
  env_label_fun,
  summary_data$chg_env
)
summary_data$sensors_label <- mapply(</pre>
  sensors_label_fun,
  summary_data$sensors
summary_data$condition <- mapply(</pre>
  condition_label_fun,
  summary_data$sensors,
  summary_data$chg_env
condition_order = c(
  "STATIC",
  "NON-PLASTIC",
  "PLASTIC"
)
pairwise_comparisons <- list(</pre>
  c("STATIC", "NON-PLASTIC"),
  c("STATIC", "PLASTIC"),
  c("PLASTIC", "NON-PLASTIC")
p_label <- function(p_value) {</pre>
  threshold = 0.0001
  if (p_value < threshold) {</pre>
```

```
return(paste0("p < ", threshold))
} else {
    return(paste0("p = ", p_value))
}

####### misc ######

# Configure our default graphing theme
theme_set(theme_cowplot())

# Palette
cb_palette <- "Paired"

# Create a directory to store plots
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)

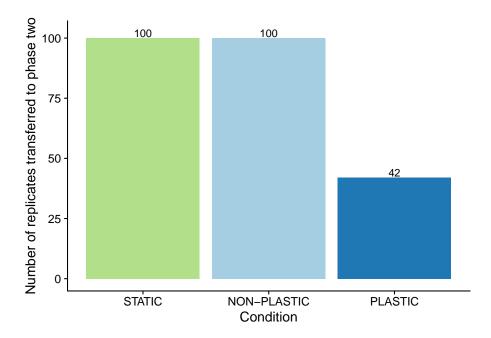
# Define sample mean function
samplemean <- function(x, d) {
    return(mean(x[d]))
}</pre>
```

#### 3.4 The evolution of phenotypic plasticity

For sensor-enabled populations in fluctuating environments, we only transfered populations containing an optimally plastic genotype to phase-two.

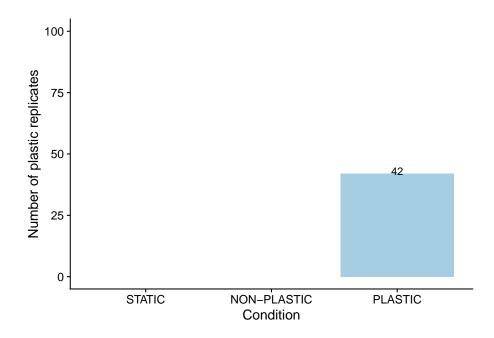
```
summary_data_grouped = dplyr::group_by(summary_data, condition)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())

ggplot(summary_data_group_counts, aes(x=condition, y=n, fill=condition)) +
    geom_col(position=position_dodge(0.9)) +
    geom_text(aes(label=n, y=n+2)) +
    scale_x_discrete(
    name="Condition",
    limits=condition_order
) +
    scale_fill_brewer(
    palette=cb_palette
) +
    scale_color_brewer(
    palette=cb_palette
) +
    ylab("Number of replicates transferred to phase two") +
    theme(
    legend.position="none"
)
```



We can confirm our expectation that the dominant genotypes in non-plastic conditions are not phenotypically plastic.

```
summary_data_grouped = dplyr::group_by(summary_data, condition, is_plastic)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
ggplot(filter(summary_data_group_counts, is_plastic), aes(x=condition, y=n, fill=condition)) +
  geom_col(
   position=position_dodge(0.9)
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  ) +
  scale_fill_brewer(
   palette=cb_palette
  scale_color_brewer(
   palette=cb_palette
  ) +
  geom_text(aes(label=n, y=n+1)) +
  ylab("Number of plastic replicates") +
  ylim(0, 100) +
  theme(
    legend.position="none"
```

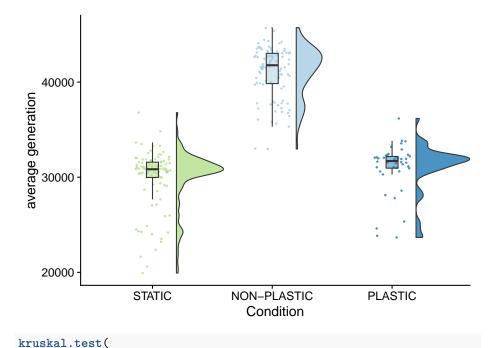


### 3.5 Average generation

```
ggplot(summary_data, aes(x=condition, y=time_average_generation, fill=condition)) +
 geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
   width = .1,
    outlier.shape = NA,
    alpha = 0.5
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  scale_fill_brewer(
```

```
palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
# coord_flip() +
ylab("average generation") +
theme(
  legend.position="none"
) +
ggsave(paste0(working_directory, "plots/", "average-generation.png"))
```

#### ## Saving $6.5 \times 4.5$ in image



```
formula=time_average_generation~condition,
  data=summary_data
)

##
## Kruskal-Wallis rank sum test
##
## data: time_average_generation by condition
## Kruskal-Wallis chi-squared = 177.33, df = 2, p-value < 2.2e-16</pre>
```

```
pairwise.wilcox.test(
  x=summary_data$time_average_generation,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$time_average_generation and summary_data$condition
##
           NON-PLASTIC PLASTIC
##
## PLASTIC <2e-16
## STATIC <2e-16
                       0.004
##
## P value adjustment method: bonferroni
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$time_average_generation)
  paste0(
    "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$time_average_generation)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC")$time_average_generation)
)
## [1] "PLASTIC median: 31697.65; STATIC median: 30839.75; NON-PLASTIC median: 41768.6
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)</pre>
  pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(
    formula=time_average_generation~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
```

```
)
    print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=9982"

## [1] "STATIC<-->PLASTIC: W=2818"

## [1] "PLASTIC<-->NON-PLASTIC: W=4186"
```

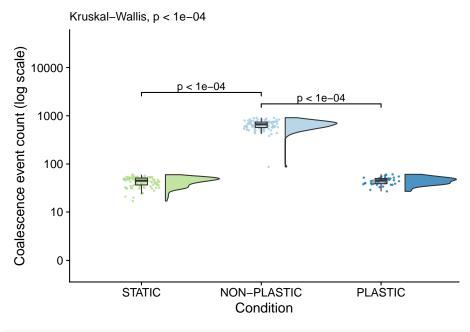
#### 3.6 Coalescence event count

The number of times the most recent common ancestor changes gives us the number of selective sweeps that occur during the experiment.

```
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(phylo_mrca_changes ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add xy position(x="condition",step.increase=1)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in
stat.test$manual_position <- log10(stat.test$y.position) * c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
coalescence_events_fig <- ggplot(</pre>
    summary_data,
    aes(x=condition, y=phylo_mrca_changes,fill=condition)
  ) +
  geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  ) +
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
  ) +
  scale_x_discrete(
   name="Condition",
   limits=condition_order,
```

```
labels=condition_order,
       breaks=condition_order
) +
scale_y_continuous(
      name="Coalescence event count (log scale)",
      trans=pseudo_log_trans(sigma = 1, base = 10),
      breaks=c(0, 10, 100, 1000, 10000),
      limits=c(-1, 35000)
) +
scale_fill_brewer(
      palette=cb_palette
) +
scale_color_brewer(
       palette=cb_palette
) +
labs(
       subtitle=paste0(
              "Kruskal-Wallis, ",
              p_label(signif(kruskal.test(formula=phylo_mrca_changes~condition, data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data=summary_data
       )
) +
ggsignif::geom_signif(
       data=filter(stat.test, p.adj <= alpha),</pre>
       aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
      manual=TRUE,
      inherit.aes=FALSE
) +
# coord_flip() +
theme(
       legend.position="none"
) +
ggsave(
       pasteO(working_directory, "plots/", "selective-sweeps.pdf"),
      width=5,
       height=5
)
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position
coalescence\_events\_fig



```
kruskal.test(
 formula=phylo_mrca_changes~condition,
  data=summary_data
)
##
   Kruskal-Wallis rank sum test
##
##
## data: phylo_mrca_changes by condition
## Kruskal-Wallis chi-squared = 175.46, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$phylo_mrca_changes,
  g=summary_data$condition,
 p.adjust.method="bonferroni",
)
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$phylo_mrca_changes and summary_data$condition
##
           NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
##
```

```
## P value adjustment method: bonferroni
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC")$phylo_mrca_changes)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC")$phylo_mrca_changes)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC")$phylo_mrca_changes)
  )
## [1] "PLASTIC median: 45.5; STATIC median: 45; NON-PLASTIC median: 663.5"
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)</pre>
  pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(</pre>
    formula=phylo_mrca_changes~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
}
## [1] "STATIC<-->NON-PLASTIC: W=10000"
## [1] "STATIC<-->PLASTIC: W=2215"
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"
```

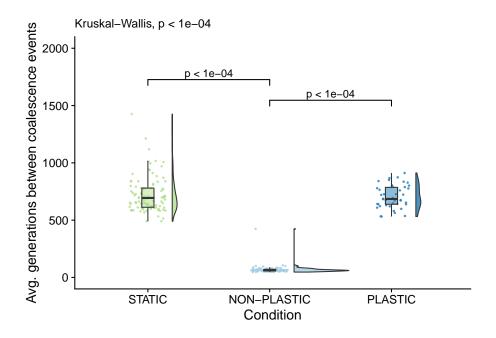
## 3.6.1 Average number of generations between coalescence events

```
# Compute frequency of coalescence events
summary_data$generations_per_mrca_change <- summary_data$time_average_generation / sum
# Compute manual labels for geom_signif</pre>
```

```
stat.test <- summary_data %>%
  wilcox_test(generations_per_mrca_change ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition")
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in
stat.test$manual_position <- stat.test$y.position</pre>
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
coalescence_events_freq_fig <- ggplot(</pre>
    summary_data,
    aes(x=condition, y=generations_per_mrca_change, fill=condition)
 ) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order,
    labels=condition_order
  ) +
  scale_y_continuous(
    name="Avg. generations between coalescence events",
    limits=c(0, 2000),
    breaks=seq(0, 2000, 500)
  ) +
  scale_fill_brewer(
    palette=cb_palette
  scale_color_brewer(
    palette=cb_palette
  # coord_flip() +
```

```
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=generations_per_mrca_change~condition, data=
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),</pre>
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/", "generations-between-selective-sweeps.png"),
  width=5,
  height=5
)
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position
coalescence\_events\_freq\_fig



```
kruskal.test(
 formula=generations_per_mrca_change~condition,
  data=summary_data
)
##
   Kruskal-Wallis rank sum test
##
##
## data: generations_per_mrca_change by condition
## Kruskal-Wallis chi-squared = 175.33, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
 x=summary_data$generations_per_mrca_change,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$generations_per_mrca_change and summary_data$condition
##
           NON-PLASTIC PLASTIC
##
## PLASTIC <2e-16
## STATIC <2e-16
                       1
##
## P value adjustment method: bonferroni
paste(
 sep="; ",
  paste0(
   "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$generations_per_mrca_change)
  paste0(
   "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$generations_per_mrca_change)
 ),
 paste0(
   "NON-PLASTIC median: ",
   median(filter(summary_data, condition=="NON-PLASTIC")$generations_per_mrca_change)
  )
)
```

## [1] "PLASTIC median: 685.001780758557; STATIC median: 693.676265008576; NON-PLASTIC median: 62

```
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"

for (pair in pairwise_comparisons) {
    pair_data <- filter(summary_data, condition %in% pair)
    pair_data$condition <- as.factor(pair_data$condition)
    wt <- wilcox.test(
        formula=generations_per_mrca_change~condition,
        data=pair_data,
        exact=FALSE,
        paired=FALSE
    )

    print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
}

## [1] "STATIC<-->PLASTIC: W=0"

## [1] "STATIC<-->PLASTIC: W=2151"

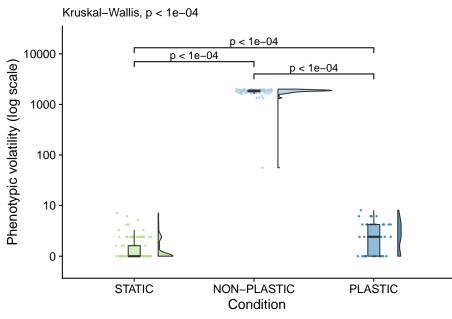
## [1] "PLASTIC<-->NON-PLASTIC: W=0"
```

# 3.7 Phenotypic volatility along the dominant lineage

```
# Compute manual labels for geom signif
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_trait_volatility ~ condition) %>%
 adjust_pvalue(method = "bonferroni") %>%
 add_significance() %>%
 add_xy_position(x="condition", step.increase=1)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad be
stat.test$manual_position <- log10(stat.test$y.position) * c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
phenotypic_volatility_fig <- ggplot(</pre>
    summary_data,
    aes(x=condition, y=dominant_lineage_trait_volatility, fill=condition)
 geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
 ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
```

```
alpha = 0.8
) +
geom_boxplot(
 width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order,
  labels=condition_order
) +
scale_y_continuous(
  name="Phenotypic volatility (log scale)",
  trans=pseudo_log_trans(sigma = 1, base = 10),
  breaks=c(0, 10, 100, 1000, 10000),
 limits=c(-1, 21000)
) +
scale_fill_brewer(
  palette=cb_palette
scale_color_brewer(
  palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_trait_volatility~condition, data=summa
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj<=alpha),</pre>
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
# coord_flip() +
theme(
  legend.position="none"
) +
  pasteO(working_directory, "plots/", "phenotypic-volatility.pdf"),
  width=5,
  height=5
)
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position
phenotypic\_volatility\_fig



```
kruskal.test(
  formula=dominant_lineage_trait_volatility~condition,
  data=summary_data
)

##

## Kruskal-Wallis rank sum test

##

## data: dominant_lineage_trait_volatility by condition

## Kruskal-Wallis chi-squared = 190.78, df = 2, p-value < 2.2e-16

pairwise.wilcox.test(
  x=summary_data$dominant_lineage_trait_volatility,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)</pre>
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_trait_volatility and summary_data$condition
##
## NON-PLASTIC PLASTIC
```

```
## PLASTIC < 2e-16
## STATIC < 2e-16
                      8.7e-07
##
## P value adjustment method: bonferroni
  sep="; ",
  paste0(
   "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_trait_volatility)
  ),
  paste0(
   "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$dominant_lineage_trait_volatility)
 ),
 paste0(
   "NON-PLASTIC median: ",
   median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_trait_volatility)
  )
)
## [1] "PLASTIC median: 2; STATIC median: 0; NON-PLASTIC median: 1868"
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
 pair_data <- filter(summary_data, condition %in% pair)</pre>
 pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(
   formula=dominant_lineage_trait_volatility~condition,
   data=pair data,
   exact=FALSE,
   paired=FALSE
 )
 print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=10000"
## [1] "STATIC<-->PLASTIC: W=3116.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"
```

# 3.7.1 Phenotypic volatility normalized by generations elapsed

```
summary_data$dominant_lineage_trait_volatility_per_generation <- summary_data$dominant
ggplot(summary_data, aes(x=condition, y=dominant_lineage_trait_volatility_per_generation)
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
   alpha = .8
 geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
 ) +
 geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
 scale_x_discrete(
   name="Condition",
   limits=condition_order
 scale_fill_brewer(
   palette=cb_palette
 scale_color_brewer(
   palette=cb_palette
 ) +
  # coord_flip() +
 theme(
   legend.position="none"
```

```
owinant lineage trait volument of the property of the property
```

4.2e-06

## STATIC < 2e-16

##

```
kruskal.test(
  formula=dominant_lineage_trait_volatility_per_generation~condition,
  data=summary_data
)
##
##
   Kruskal-Wallis rank sum test
## data: dominant_lineage_trait_volatility_per_generation by condition
## Kruskal-Wallis chi-squared = 189.62, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_trait_volatility_per_generation,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_trait_volatility_per_generation and summary_data$condition
##
           NON-PLASTIC PLASTIC
## PLASTIC < 2e-16
```

```
## P value adjustment method: bonferroni
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_trait_volatilit
 ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC")$dominant_lineage_trait_volatility
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_trait_volat
## [1] "PLASTIC median: 6.33339279717772e-05; STATIC median: 0; NON-PLASTIC median: 0.
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)</pre>
  pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(
    formula=dominant_lineage_trait_volatility_per_generation~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=10000"
## [1] "STATIC<-->PLASTIC: W=3061.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"
```

### 3.8 Phenotypic fidelity

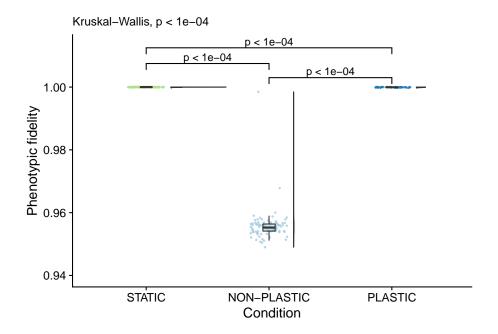
Frequency that an offspring's genotype is identical to a parent genotype (along the dominant lineage).

```
summary_data$dominant_lineage_trait_fidelity <- (summary_data$dominant_generation_born
# Compute manual labels for geom_signif</pre>
```

```
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_trait_fidelity ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition",step.increase=1.5)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in
stat.test$manual_position <- stat.test$y.position * c(1.0,1.0,1.0005)
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
phenotypic_fidelity_fig <- ggplot(</pre>
   summary_data,
   aes(x=condition, y=dominant_lineage_trait_fidelity, fill=condition)
 ) +
  geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
  ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  ) +
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
  ) +
  scale x discrete(
   name="Condition",
   limits=condition order,
   labels=condition_order
  ) +
  scale_y_continuous(
   name="Phenotypic fidelity",
   limits=c(0.94, 1.013),
   breaks=c(0.94, 0.96, 0.98, 1.0) #seq(0.94, 1.0, 0.01)
  ) +
  scale_fill_brewer(
   palette=cb_palette
  scale_color_brewer(
   palette=cb_palette
  # coord_flip() +
```

```
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_trait_fidelity~condition, data)
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),</pre>
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/", "phenotypic-fidelity.pdf"),
  width=5,
  height=5
)
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position
phenotypic\_fidelity\_fig



```
kruskal.test(
 formula=dominant_lineage_trait_fidelity~condition,
  data=summary_data
)
##
   Kruskal-Wallis rank sum test
##
##
## data: dominant_lineage_trait_fidelity by condition
## Kruskal-Wallis chi-squared = 189.62, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
 x=summary_data$dominant_lineage_trait_fidelity,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_trait_fidelity and summary_data$condition
##
##
           NON-PLASTIC PLASTIC
## PLASTIC < 2e-16
## STATIC < 2e-16
                       4.2e-06
##
## P value adjustment method: bonferroni
paste(
 sep="; ",
  paste0(
   "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_trait_fidelity)
  paste0(
   "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$dominant_lineage_trait_fidelity)
 ),
 paste0(
   "NON-PLASTIC median: ",
   median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_trait_fidelity)
  )
)
```

## [1] "PLASTIC median: 0.999936666072028; STATIC median: 1; NON-PLASTIC median: 0.95525598543618

```
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"

for (pair in pairwise_comparisons) {
    pair_data <- filter(summary_data, condition %in% pair)
    pair_data$condition <- as.factor(pair_data$condition)
    wt <- wilcox.test(
        formula=dominant_lineage_trait_fidelity~condition,
        data=pair_data,
        exact=FALSE,
        paired=FALSE
    )
    print(pasteO(pair[1], "<-->", pair[2], ": W=",wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=O"

## [1] "STATIC<-->PLASTIC: W=1138.5"

## [1] "PLASTIC<-->NON-PLASTIC: W=O"
```

## 3.9 Mutation count

```
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
 wilcox_test(dominant_lineage_total_mut_cnt ~ condition) %>%
 adjust_pvalue(method = "bonferroni") %>%
 add significance() %>%
 add_xy_position(x="condition",step.increase=1)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad be
stat.test\$manual_position <- log10(stat.test\$y.position) * c(1.0,1.0,1.03) # c(1.0,1.
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
mutation_count_fig <- ggplot(</pre>
    summary_data,
    aes(x=condition, y=dominant_lineage_total_mut_cnt, fill=condition)
 ) +
 geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
 ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
```

```
) +
 geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
 ) +
 scale_x_discrete(
   name="Condition",
   limits=condition_order,
   labels=condition_order
 ) +
 scale_y_continuous(
   name="Mutation count (log scale)",
   trans=pseudo_log_trans(sigma = 1, base = 10),
  breaks=c(0, 10, 100, 1000, 10000),
  limits=c(-1, 35000)
 ) +
 scale_fill_brewer(
   palette=cb_palette
 scale_color_brewer(
   palette=cb_palette
 ) +
 # coord_flip() +
 labs(
   subtitle=paste0(
     "Kruskal-Wallis, ",
     p_label(signif(kruskal.test(formula=dominant_lineage_total_mut_cnt~condition, data=summary
   )
 ) +
 ggsignif::geom_signif(
   data=filter(stat.test, p.adj <= alpha),</pre>
   aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
   manual=TRUE,
   inherit.aes=FALSE
 ) +
   legend.position="none"
 ) +
 ggsave(
   paste0(working_directory, "plots/", "mutation-accumulation.pdf"),
   width=5,
   height=4
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position
mutation\_count\_fig

```
| Non-Plastic Condition | Non-Plastic Plastic | Plastic
```

```
kruskal.test(
  formula=dominant_lineage_total_mut_cnt~condition,
  data=summary_data
)

##

## Kruskal-Wallis rank sum test

##

## data: dominant_lineage_total_mut_cnt by condition

## Kruskal-Wallis chi-squared = 179.33, df = 2, p-value < 2.2e-16

pairwise.wilcox.test(
  x=summary_data$dominant_lineage_total_mut_cnt,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)</pre>
```

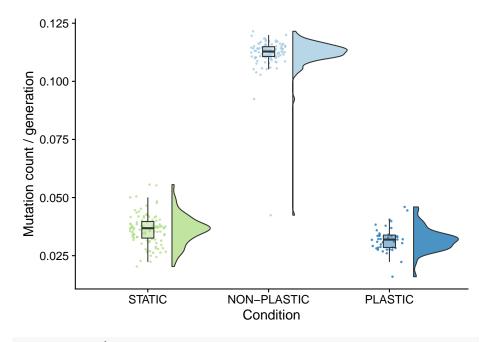
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary\_data\$dominant\_lineage\_total\_mut\_cnt and summary\_data\$condition
##
## NON-PLASTIC PLASTIC

```
## PLASTIC <2e-16
## STATIC <2e-16
                       0.0019
##
## P value adjustment method: bonferroni
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_total_mut_cnt)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC")$dominant_lineage_total_mut_cnt)
 ),
 paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_total_mut_cnt)
  )
)
## [1] "PLASTIC median: 998.5; STATIC median: 1100; NON-PLASTIC median: 4657.5"
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)</pre>
  pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(</pre>
    formula=dominant_lineage_total_mut_cnt~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=10000"
## [1] "STATIC<-->PLASTIC: W=1336.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"
```

### 3.9.1 Mutation count normalized by generations elapsed

```
summary_data$mutations_per_generation <- summary_data$dominant_lineage_total_mut_cnt / summary_data
ggplot(summary_data, aes(x=condition, y=mutations_per_generation, fill=condition)) +</pre>
```

```
geom_flat_violin(
  position = position_nudge(x = .2, y = 0),
 alpha = .8
) +
geom_point(
  mapping=aes(color=condition),
 position = position_jitter(width = .15),
 size = .5,
 alpha = 0.8
geom_boxplot(
 width = .1,
 outlier.shape = NA,
 alpha = 0.5
) +
scale_x_discrete(
 name="Condition",
 limits=condition_order
ylab("Mutation count / generation") +
scale_fill_brewer(
 palette=cb_palette
) +
scale_color_brewer(
 palette=cb_palette
# coord_flip() +
theme(
  legend.position="none"
```



```
kruskal.test(
  formula=mutations_per_generation~condition,
  data=summary_data
)
##
   Kruskal-Wallis rank sum test
##
## data: mutations_per_generation by condition
## Kruskal-Wallis chi-squared = 180.11, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$mutations_per_generation,
  g=summary_data$condition,
 p.adjust.method="bonferroni",
)
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$mutations_per_generation and summary_data$condition
##
           NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
                       2e-04
##
```

```
## P value adjustment method: bonferroni
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC")$mutations_per_generation)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC")$mutations_per_generation)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC")$mutations_per_generation)
## [1] "PLASTIC median: 0.0319267181456982; STATIC median: 0.0368157192941933; NON-PLA
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)</pre>
  pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(
    formula=mutations_per_generation~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=9987"
## [1] "STATIC<-->PLASTIC: W=1206"
## [1] "PLASTIC<-->NON-PLASTIC: W=4198"
```

## 3.10 Genotypic fidelity

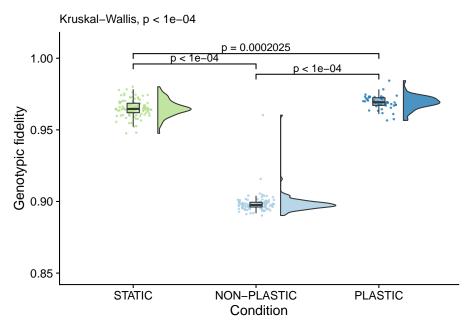
The frequency that an offspring's genotype is the same as a parent's genotype.

```
summary_data$dominant_lineage_genotypic_fidelity <- (summary_data$dominant_generation_'
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
```

```
wilcox_test(dominant_lineage_genotypic_fidelity ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition",step.increase=0.2)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in
stat.test$manual_position <- stat.test$y.position * c(1.0,1.0,1.0)</pre>
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
genotypic_fidelity_fig <- ggplot(</pre>
    summary_data,
    aes(x=condition, y=dominant_lineage_genotypic_fidelity, fill=condition)
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order,
    labels=condition_order
  ) +
  scale_y_continuous(
    name="Genotypic fidelity",
    limits=c(0.85, 1.01),
    breaks=c(0.85, 0.90, 0.95, 1.0) #seq(0.85, 1.0, 0.02)
  scale_fill_brewer(
    palette=cb_palette
  scale_color_brewer(
    palette=cb_palette
  ) +
  # coord_flip() +
  labs(
```

```
subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_genotypic_fidelity~condition)
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),</pre>
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
ggsave(
  pasteO(working_directory, "plots/", "genotypic-fidelity.pdf"),
  width=5,
  height=4
)
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position
genotypic\_fidelity\_fig



kruskal.test(
 formula=dominant\_lineage\_genotypic\_fidelity~condition,

```
data=summary_data
)
##
##
   Kruskal-Wallis rank sum test
## data: dominant_lineage_genotypic_fidelity by condition
## Kruskal-Wallis chi-squared = 179.86, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_genotypic_fidelity,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: summary_data$dominant_lineage_genotypic_fidelity and summary_data$condition
##
           NON-PLASTIC PLASTIC
##
## PLASTIC <2e-16
                       2e-04
## STATIC <2e-16
## P value adjustment method: bonferroni
paste(
 sep="; ",
  paste0(
   "PLASTIC median: ",
   median(filter(summary data, condition=="PLASTIC")$dominant lineage genotypic fidelity)
 ),
  paste0(
   "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$dominant_lineage_genotypic_fidelity)
  ),
  paste0(
    "NON-PLASTIC median: ",
   median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_genotypic_fidelity)
  )
)
## [1] "PLASTIC median: 0.969286906891951; STATIC median: 0.964620594632577; NON-PLASTIC median:
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
```

```
for (pair in pairwise_comparisons) {
   pair_data <- filter(summary_data, condition %in% pair)
   pair_data$condition <- as.factor(pair_data$condition)
   wt <- wilcox.test(
      formula=dominant_lineage_genotypic_fidelity~condition,
      data=pair_data,
      exact=FALSE,
      paired=FALSE
)
   print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=18"
## [1] "STATIC<-->PLASTIC: W=2992"
## [1] "PLASTIC<-->NON-PLASTIC: W=2"
```

## 3.11 Characterizing variation along dominant lineages

## 3.11.1 What fraction of mutations along the lineage result in phenotypic changes?

```
summary_data$frac_phenotype_changing_mut_steps <- summary_data$dominant_lineage_num_mu
ggplot(summary_data, aes(x=condition, y=frac_phenotype_changing_mut_steps, fill=condit
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  ) +
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
    alpha = 0.5
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  scale_fill_brewer(
```

```
palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  # coord_flip() +
  theme(
    legend.position="none"
  ggsave(paste0(working_directory, "plots/", "frac_phenotype_changing_mutational_steps.png"))
## Saving 6.5 \times 4.5 in image
   0.5
 frac_phenotype_changing_mut_steps
   0.4
   0.2
   0.1
   0.0
                               NON-PLASTIC
               STATIC
                                                     PLASTIC
                                  Condition
kruskal.test(
  formula=frac_phenotype_changing_mut_steps~condition,
  data=summary_data
##
##
    Kruskal-Wallis rank sum test
##
## data: frac_phenotype_changing_mut_steps by condition
## Kruskal-Wallis chi-squared = 191.23, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$frac_phenotype_changing_mut_steps,
```

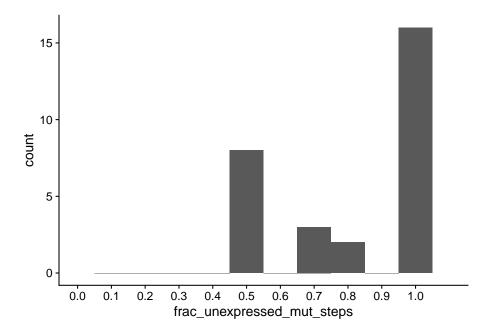
```
g=summary_data$condition,
 p.adjust.method="bonferroni",
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$frac_phenotype_changing_mut_steps and summary_data$condition
##
##
           NON-PLASTIC PLASTIC
## PLASTIC < 2e-16
## STATIC < 2e-16
                       2.3e-07
##
## P value adjustment method: bonferroni
paste(
 sep="; ",
 paste0(
    "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$frac_phenotype_changing_mut_step.
 ),
 paste0(
    "STATIC median: ",
   median(filter(summary_data, condition=="STATIC") $frac_phenotype_changing_mut_steps
 paste0(
    "NON-PLASTIC median: ",
   median(filter(summary_data, condition=="NON-PLASTIC") $frac_phenotype_changing_mut_
  )
)
## [1] "PLASTIC median: 0.00224941742616098; STATIC median: 0; NON-PLASTIC median: 0.4
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
 pair_data <- filter(summary_data, condition %in% pair)</pre>
 pair_data$condition <- as.factor(pair_data$condition)</pre>
 wt <- wilcox.test(
    formula=frac_phenotype_changing_mut_steps~condition,
   data=pair_data,
    exact=FALSE,
   paired=FALSE
 print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
```

```
## [1] "STATIC<-->NON-PLASTIC: W=10000"
## [1] "STATIC<-->PLASTIC: W=3172"
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"
```

# 3.11.2 For PLASTIC populations, what fraction of phenotype-altering mutations occurred in the unexpressed phenotype?

```
summary_data$frac_unexpressed_mut_steps <- summary_data$dominant_lineage_num_mut_steps_that_change
summary_data$frac_expressed_mut_steps <- summary_data$dominant_lineage_num_mut_steps_that_change
ggplot(filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_that_change_agg
geom_histogram(binwidth=0.1) +
scale_x_continuous(
   limits=c(0, 1.1),
   breaks=seq(0, 1.0, 0.1)
) +
theme(
legend.position="none"</pre>
```

## Warning: Removed 2 rows containing missing values (geom\_bar).



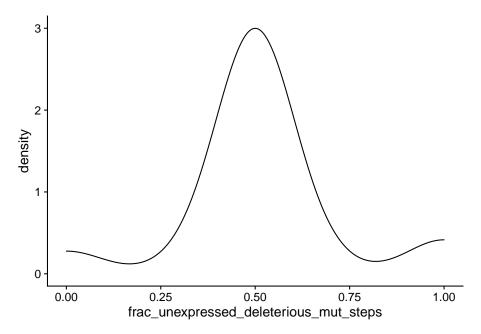
```
print(paste0("PLASTIC - Mean with bootstrapped 95% CI"))
## [1] "PLASTIC - Mean with bootstrapped 95% CI"
bo <- boot(filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_
print(bo)
##
## ORDINARY NONPARAMETRIC BOOTSTRAP
##
##
## Call:
## boot(data = filter(summary_data, condition == "PLASTIC" & dominant_lineage_num_mut_:
       0) $frac_unexpressed_mut_steps, statistic = samplemean, R = 10000)
##
##
## Bootstrap Statistics :
        original
                       bias
                               std. error
## t1* 0.8247126 0.0003025862 0.03989834
print(boot.ci(bo, conf=0.95, type="perc"))
## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
## Based on 10000 bootstrap replicates
##
## CALL :
## boot.ci(boot.out = bo, conf = 0.95, type = "perc")
##
## Intervals :
## Level
           Percentile
         (0.7443, 0.8994)
## 95%
## Calculations and Intervals on Original Scale
plastic_summary_data <- filter(summary_data, condition=="PLASTIC")</pre>
aggregate_frac_mut_steps_that_change_unexpressed_phenotype <- sum(plastic_summary_data
sum(plastic_summary_data$dominant_lineage_num_mut_steps_that_change_unexpressed_phenot
## [1] 83
sum(plastic_summary_data$dominant_lineage_num_mut_steps_that_change_aggregate_phenotype
## [1] 102
{\tt aggregate\_frac\_mut\_steps\_that\_change\_unexpressed\_phenotype}
## [1] 0.8137255
83 / 102 (0.8137255)
```

# 3.11.3 For PLASTIC populations, what fraction of mutations that affect the unexpressed phenotype are deleterious versus beneficial?

aggregate\_frac\_unexpressed\_deleterious\_mut\_steps <- sum(plastic\_summary\_data\$dominant\_lineage\_numaggregate\_frac\_unexpressed\_beneficial\_mut\_steps <- sum(plastic\_summary\_data\$dominant\_lineage\_num\_steps)

#### 3.11.3.1 Deleterious mutations

```
summary_data$frac_unexpressed_deleterious_mut_steps <- summary_data$dominant_lineage_num_mut_step
ggplot(
   filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_that_change_unexpress
   aes(x=frac_unexpressed_deleterious_mut_steps)
) +
   geom_density() +
   theme(
    legend.position="none"
)</pre>
```



bo <- boot(filter(summary\_data, condition=="PLASTIC" & dominant\_lineage\_num\_mut\_steps\_that\_change
print(bo)</pre>

```
##
## ORDINARY NONPARAMETRIC BOOTSTRAP
##
```

## Intervals :

## 95%

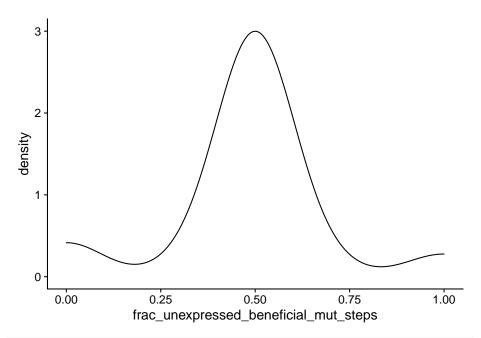
## Level Percentile

(0.4402, 0.5954)

## Calculations and Intervals on Original Scale

```
##
## Call:
## boot(data = filter(summary_data, condition == "PLASTIC" & dominant_lineage_num_mut_
       0)$frac_unexpressed_deleterious_mut_steps, statistic = samplemean,
       R = 10000
##
##
##
## Bootstrap Statistics :
                      bias std. error
        original
## t1* 0.5172414 -0.0003644828 0.03961947
print(boot.ci(bo, conf=0.95, type="perc"))
## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
## Based on 10000 bootstrap replicates
## boot.ci(boot.out = bo, conf = 0.95, type = "perc")
```

```
summary_data$frac_unexpressed_beneficial_mut_steps <- summary_data$dominant_lineage_num
ggplot(
   filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_that_changaes(x=frac_unexpressed_beneficial_mut_steps)
) +
   geom_density() +
   theme(
    legend.position="none"</pre>
```



bo <- boot(filter(summary\_data, condition=="PLASTIC" & dominant\_lineage\_num\_mut\_steps\_that\_change
print(bo)</pre>

```
## ORDINARY NONPARAMETRIC BOOTSTRAP
##
##
## Call:
## boot(data = filter(summary_data, condition == "PLASTIC" & dominant_lineage_num_mut_steps_that
##
       0)$frac_unexpressed_beneficial_mut_steps, statistic = samplemean,
       R = 10000
##
##
##
## Bootstrap Statistics :
        original
                       bias
                               std. error
## t1* 0.4827586 -8.16092e-05 0.03986791
print(boot.ci(bo, conf=0.95, type="perc"))
## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
## Based on 10000 bootstrap replicates
##
## boot.ci(boot.out = bo, conf = 0.95, type = "perc")
```

## Intervals :

```
## Level Percentile
## 95% ( 0.4046,  0.5598 )
## Calculations and Intervals on Original Scale
```

## 3.12 Manuscript figures

Figures styled for the paper.

```
grid <- plot_grid(</pre>
  coalescence_events_fig +
    theme(
      legend.position="none",
      # axis.ticks.x=element_blank(),
      # axis.text.x=element_blank(),
      axis.title.x=element_blank()
    ggtitle("Coalescence events count"),
 mutation_count_fig +
    theme(
      legend.position="none",
      # axis.ticks.x=element_blank(),
      # axis.text.x=element blank(),
      axis.title.x=element_blank()
    ggtitle("Mutation count"),
 phenotypic_volatility_fig +
    theme(
      legend.position="none",
      # axis.ticks.x=element blank(),
      # axis.text.x=element_blank(),
      axis.title.x=element_blank()
    ggtitle("Phenotypic volatility"),
  coalescence_events_freq_fig +
    theme(
      legend.position="none",
      axis.title.x=element_blank()
    ggtitle("Generations between coalescence events"),
  genotypic_fidelity_fig +
    theme(
      legend.position="none",
      axis.title.x=element blank()
    ggtitle("Genotypic fidelity"),
```

```
phenotypic_fidelity_fig +
    theme(
        legend.position="none",
        axis.title.x=element_blank()
    ) +
    ggtitle("Phenotypic fidelity"),
    nrow=2,
    ncol=3,
    align="v",
    labels="auto"
)
save_plot(
    paste0(working_directory, "plots/", "evolutionary-change-full-panel.pdf"),
    grid,
    # base_height=12,
    # base_asp=3/2
    base_height=12,
    base_asp=3/2
)
```

## Chapter 4

# Evolution and maintenance of novel traits

The effect of adaptive phenotypic plasticity on the evolution and maintenance of novel tasks.

#### 4.1 Overview

```
total_updates <- 200000
replicates <- 100
alpha <- 0.05
focal_traits <- c("not", "nand", "and", "ornot", "or", "andnot")</pre>
traits_set_a <- c("not", "and", "or")</pre>
traits_set_b <- c("nand", "ornot", "andnot")</pre>
extra_traits <- c(
  "nor", "xor", "equals",
  "logic_3aa", "logic_3ab", "logic_3ac",
  "logic_3ad","logic_3ae","logic_3af",
  "logic_3ag", "logic_3ah", "logic_3ai",
  "logic_3aj", "logic_3ak", "logic_3al",
  "logic_3am", "logic_3an", "logic_3ao",
  "logic_3ap", "logic_3aq", "logic_3ar",
  "logic_3as", "logic_3at", "logic_3au",
  "logic_3av", "logic_3aw", "logic_3ax",
  "logic_3ay", "logic_3az", "logic_3ba",
  "logic_3bb", "logic_3bc", "logic_3bd",
  "logic_3be", "logic_3bf", "logic_3bg",
  "logic_3bh","logic_3bi","logic_3bj",
```

```
"logic_3bk","logic_3bn","logic_3bm",
  "logic_3bn","logic_3bp","logic_3bs",
  "logic_3bt","logic_3bs","logic_3bs",
  "logic_3bt","logic_3bv","logic_3bv",
  "logic_3bz","logic_3cb","logic_3cb",
  "logic_3cz","logic_3cd","logic_3ce",
  "logic_3cf","logic_3cg","logic_3ch",
  "logic_3ci","logic_3cj","logic_3ck",
  "logic_3ci","logic_3cm","logic_3cn",
  "logic_3co","logic_3cm","logic_3cn",
  "logic_3co","logic_3cp"
)

# Relative location of data.
working_directory <- "experiments/2021-01-31-complex-features/analysis/" # << For book
# working_directory <- "./"</pre>
```

## 4.2 Analysis dependencies

2021

Load all required R libraries.

## year

```
library(ggplot2)
library(rstatix)
library(ggsignif)
library(scales)
library(tidyverse)
library(cowplot)
library(RColorBrewer)
library(Hmisc)
library(boot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd9")
```

These analyses were conducted/knitted with the following computing environment:

4.3. SETUP 63

## 4.3 Setup

```
###### summary data ######
summary_data_loc <- paste0(working_directory, "data/aggregate.csv")</pre>
summary_data <- read.csv(summary_data_loc, na.strings="NONE")</pre>
summary_data$DISABLE_REACTION_SENSORS <- as.factor(summary_data$DISABLE_REACTION_SENSORS)</pre>
summary_data$chg_env <- summary_data$chg_env == "True"</pre>
summary_data$dominant_plastic_odd_even <- as.factor(summary_data$dominant_plastic_odd_even)</pre>
summary_data$sensors <- summary_data$DISABLE_REACTION_SENSORS == "0"</pre>
summary_data$is_plastic <- summary_data$dominant_plastic_odd_even == "True"</pre>
summary_data$extra_task_value <- as.factor(summary_data$extra_task_value)
summary_data <- filter(summary_data, extra_task_value == 0.1)</pre>
env_label_fun <- function(chg_env) {</pre>
  if (chg env) {
    return("Fluctuating")
  } else {
    return("Constant")
  }
}
sensors_label_fun <- function(has_sensors) {</pre>
  if (has_sensors) {
    return("Sensors")
  } else {
    return("No sensors")
}
condition_label_fun <- function(has_sensors, env_chg) {</pre>
  if (has_sensors && env_chg) {
    return("PLASTIC")
  } else if (env_chg) {
    return("NON-PLASTIC")
  } else {
 return("STATIC")
```

```
}
}
summary_data$env_label <- mapply(</pre>
  env_label_fun,
  summary_data$chg_env
summary_data$sensors_label <- mapply(</pre>
  sensors_label_fun,
  summary_data$sensors
summary_data$condition <- mapply(</pre>
  condition_label_fun,
  summary_data$sensors,
  summary_data$chg_env
condition_order = c(
  "STATIC",
  "NON-PLASTIC",
  "PLASTIC"
pairwise_comparisons <- list(</pre>
  c("STATIC", "NON-PLASTIC"),
  c("STATIC", "PLASTIC"),
  c("PLASTIC", "NON-PLASTIC")
)
p_label <- function(p_value) {</pre>
  threshold = 0.0001
  if (p_value < threshold) {</pre>
    return(paste0("p < ", threshold))</pre>
  } else {
    return(paste0("p = ", p_value))
  }
}
##### time series #####
lineage_time_series_data_loc <- pasteO(working_directory, "data/lineage_series.csv")</pre>
lineage_time_series_data <- read.csv(lineage_time_series_data_loc)</pre>
lineage_time_series_data$DISABLE_REACTION_SENSORS <- as.factor(lineage_time_series_data
lineage_time_series_data$chg_env <- lineage_time_series_data$chg_env == "True"</pre>
lineage_time_series_data$sensors <- lineage_time_series_data$DISABLE_REACTION_SENSORS</pre>
lineage_time_series_data$extra_task_value <- as.factor(lineage_time_series_data$extra_
```

```
lineage_time_series_data$env_label <- mapply(</pre>
  env_label_fun,
  lineage_time_series_data$chg_env
lineage_time_series_data$sensors_label <- mapply(</pre>
  sensors_label_fun,
  lineage_time_series_data$sensors
lineage_time_series_data$condition <- mapply(</pre>
  condition_label_fun,
  lineage_time_series_data$sensors,
  lineage_time_series_data$chg_env
###### misc ######
# Configure our default graphing theme
theme_set(theme_cowplot())
# Palette
cb_palette <- "Paired"</pre>
# Create directory to dump plots
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)
# Sample mean function
samplemean <- function(x, d) {</pre>
  return(mean(x[d]))
}
```

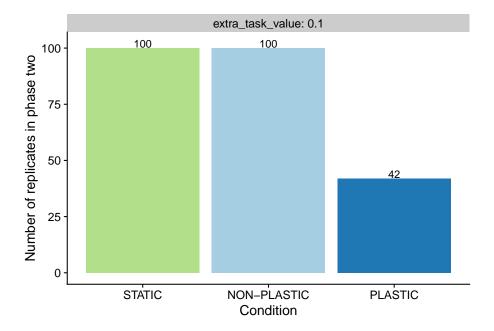
## 4.4 The evolution of phenotypic plasticity

For sensor-enabled populations in fluctuating environments, we only transfered populations containing an optimally plastic genotype to phase two.

```
summary_data_grouped = dplyr::group_by(summary_data, sensors, env_label, condition, extra_task_vasummary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())

ggplot(summary_data_group_counts, aes(x=condition, y=n, fill=condition)) +
    geom_col(position=position_dodge(0.9)) +
    geom_text(aes(label=n, y=n+2)) +
    scale_x_discrete(
    name="Condition",
    limits=condition_order
) +
    scale_fill_brewer(
    palette=cb_palette
) +
```

```
scale_color_brewer(
   palette=cb_palette
) +
ylab("Number of replicates in phase two") +
facet_wrap(~extra_task_value, labeller=label_both) +
theme(
  legend.position="none"
)
```

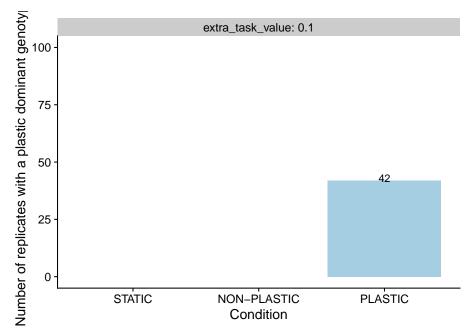


We can confirm our expectation that the dominant genotypes in non-plastic conditions are not phenotypically plastic.

palette=cb\_palette

```
summary_data_grouped = dplyr::group_by(summary_data, condition, is_plastic, extra_task
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
ggplot(filter(summary_data_group_counts, is_plastic), aes(x=condition, y=n, fill=condition)
geom_col(position=position_dodge(0.9)) +
scale_x_discrete(
    name="Condition",
    limits=condition_order
) +
scale_fill_brewer(
    palette=cb_palette
) +
scale_color_brewer(
```

```
) +
ylim(0, 100) +
geom_text(aes(label=n, y=n+1)) +
ylab("Number of replicates with a plastic dominant genotype") +
facet_wrap(~extra_task_value, labeller=label_both) +
theme(
   legend.position="none"
)
```



## 4.5 Final novel task count (dominant genotype)

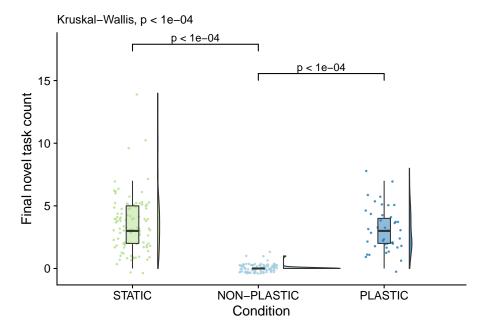
How many novel tasks do final dominant genotypes perform?

```
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(dominant_extra_tasks ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition") # ,step.increase=1
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in grant stat.test$manual_position <- stat.test$y.position #* c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)
final_novel_task_count_fig <- ggplot(</pre>
```

```
summary_data,
  aes(x=condition, y=dominant_extra_tasks, fill=condition)
) +
geom_flat_violin(
  position = position_nudge(x = .2, y = 0),
 alpha = .8
) +
geom_point(
 mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
 alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
 limits=condition_order,
 labels=condition_order
scale y continuous(
 name="Final novel task count"
scale_fill_brewer(
 palette=cb_palette
) +
scale_color_brewer(
 palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_extra_tasks~condition, data=summary
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),</pre>
  aes(xmin=group1,xmax=group2,annotations=label,y position=manual position),
 manual=TRUE,
 inherit.aes=FALSE
) +
# coord_flip()
```

```
theme(
  legend.position="none"
)
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position
final\_novel\_task\_count\_fig



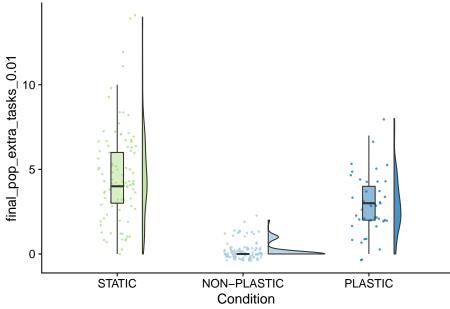
```
kruskal.test(
  formula=dominant_extra_tasks~condition,
  data=summary_data
)
##
##
   Kruskal-Wallis rank sum test
##
## data: dominant_extra_tasks by condition
## Kruskal-Wallis chi-squared = 177.17, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$dominant_extra_tasks,
  g=summary_data$condition,
 p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
```

```
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
##
## data: summary_data$dominant_extra_tasks and summary_data$condition
##
          NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
                       0.9
## P value adjustment method: bonferroni
paste(
 sep="; ",
 paste0(
    "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$dominant_extra_tasks)
 ),
 paste0(
    "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$dominant_extra_tasks)
 ),
 paste0(
    "NON-PLASTIC median: ",
   median(filter(summary_data, condition=="NON-PLASTIC")$dominant_extra_tasks)
  )
)
## [1] "PLASTIC median: 3; STATIC median: 3; NON-PLASTIC median: 0"
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
 pair_data <- filter(summary_data, condition %in% pair)</pre>
 pair_data$condition <- as.factor(pair_data$condition)</pre>
 wt <- wilcox.test(
   formula=dominant_extra_tasks~condition,
   data=pair_data,
   exact=FALSE,
    paired=FALSE
 print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=184"
## [1] "STATIC<-->PLASTIC: W=1871"
## [1] "PLASTIC<-->NON-PLASTIC: W=64"
```

## 4.6 Novel task count (final population)

How many novel tasks are performed across the final population (1% of organisms must perform to count)?

```
ggplot(summary_data, aes(x=condition, y=final_pop_extra_tasks_0.01, fill=condition)) +
  geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
 ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  ) +
  scale_fill_brewer(
   palette=cb_palette
  scale_color_brewer(
   palette=cb_palette
  ) +
  # coord_flip() +
  theme(
   legend.position="none"
```



```
kruskal.test(
  formula=final_pop_extra_tasks_0.01~condition,
  data=summary_data
)
##
##
   Kruskal-Wallis rank sum test
##
## data: final_pop_extra_tasks_0.01 by condition
## Kruskal-Wallis chi-squared = 169.47, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$final_pop_extra_tasks_0.01,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
)
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
```

## data: summary\_data\$final\_pop\_extra\_tasks\_0.01 and summary\_data\$condition

NON-PLASTIC PLASTIC

##

##

## PLASTIC < 2e-16

```
## STATIC < 2e-16
                       0.00016
## P value adjustment method: bonferroni
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$final_pop_extra_tasks_0.01)
  ),
  paste0(
   "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$final_pop_extra_tasks_0.01)
 ),
  paste0(
   "NON-PLASTIC median: ",
   median(filter(summary_data, condition=="NON-PLASTIC")$final_pop_extra_tasks_0.01)
  )
)
## [1] "PLASTIC median: 3; STATIC median: 4; NON-PLASTIC median: 0"
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
 pair_data <- filter(summary_data, condition %in% pair)</pre>
 pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(
   formula=final_pop_extra_tasks_0.01~condition,
   data=pair data,
   exact=FALSE,
   paired=FALSE
  print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=227.5"
## [1] "STATIC<-->PLASTIC: W=1203"
## [1] "PLASTIC<-->NON-PLASTIC: W=225.5"
```

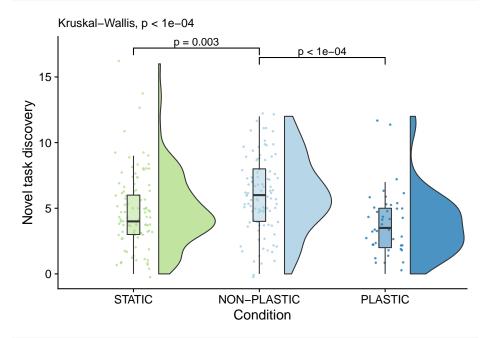
### 4.7 Novel task discovery (lineage)

```
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
wilcox_test(dominant_lineage_extra_traits_discovered ~ condition) %>%
```

```
adjust_pvalue(method = "bonferroni") %>%
 add_significance() %>%
  add_xy_position(x="condition") # ,step.increase=1
# Tweak y.position manually to account for scaled axis (edge case that triggers bad be
stat.test$manual_position <- stat.test$y.position #* c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
lineage_novel_task_discovery_fig <- ggplot(</pre>
    summary_data,
    aes(x=condition, y=dominant_lineage_extra_traits_discovered, fill=condition)
 geom flat violin(
   position = position_nudge(x = .2, y = 0),
    alpha = .8
 ) +
 geom_point(
   mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
   alpha = 0.8
 ) +
 geom_boxplot(
   width = .1,
    outlier.shape = NA,
    alpha = 0.5
  scale_x_discrete(
   name="Condition",
   limits=condition order,
   labels=condition_order
 ) +
 scale_y_continuous(
   name="Novel task discovery"
  scale_fill_brewer(
    palette=cb_palette
  scale_color_brewer(
   palette=cb_palette
  ) +
 labs(
    subtitle=paste0(
      "Kruskal-Wallis, ",
      p_label(signif(kruskal.test(formula=dominant_lineage_extra_traits_discovered~con-
```

```
gsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
# coord_flip()
theme(
  legend.position="none"
)
</pre>
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position lineage\_novel\_task\_discovery\_fig



```
kruskal.test(
  formula=dominant_lineage_extra_traits_discovered~condition,
  data=summary_data
)

##

##

Kruskal-Wallis rank sum test
##
```

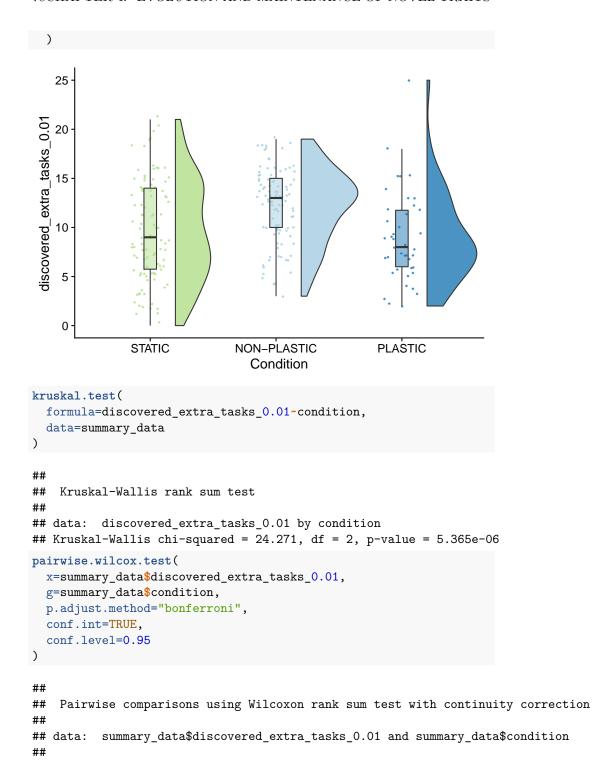
## data: dominant\_lineage\_extra\_traits\_discovered by condition
## Kruskal-Wallis chi-squared = 24.099, df = 2, p-value = 5.846e-06

```
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_extra_traits_discovered,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_extra_traits_discovered and summary_data$condi
##
           NON-PLASTIC PLASTIC
## PLASTIC 1.7e-05
## STATIC 0.0035
                       0.0561
## P value adjustment method: bonferroni
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_extra_traits_di
  ),
  paste0(
    "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$dominant_lineage_extra_traits_dis
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_extra_trait
  )
)
## [1] "PLASTIC median: 3.5; STATIC median: 4; NON-PLASTIC median: 6"
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)</pre>
  pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(
   formula=dominant_lineage_extra_traits_discovered~condition,
    data=pair_data,
```

```
exact=FALSE,
  paired=FALSE
)
  print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
}
## [1] "STATIC<-->NON-PLASTIC: W=6319.5"
## [1] "STATIC<-->PLASTIC: W=1578"
## [1] "PLASTIC<-->NON-PLASTIC: W=3110.5"
```

## 4.8 Novel task discovery (population)

```
ggplot(
   summary_data,
   aes(x=condition, y=discovered_extra_tasks_0.01, fill=condition)
  geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
  ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  ) +
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
 ) +
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  ) +
  scale_fill_brewer(
   palette=cb_palette
  scale_color_brewer(
   palette=cb_palette
  ) +
  # coord_flip() +
 theme(
   legend.position="none"
```



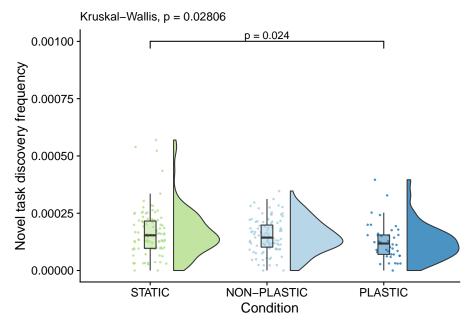
```
##
           NON-PLASTIC PLASTIC
## PLASTIC 2.4e-05
## STATIC 0.00035
                       1.00000
## P value adjustment method: bonferroni
paste(
 sep="; ",
 paste0(
   "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$discovered_extra_tasks_0.01)
 ),
  paste0(
    "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$discovered_extra_tasks_0.01)
 ),
 paste0(
   "NON-PLASTIC median: ",
   median(filter(summary_data, condition=="NON-PLASTIC")$discovered_extra_tasks_0.01)
 )
)
## [1] "PLASTIC median: 8; STATIC median: 9; NON-PLASTIC median: 13"
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
 pair_data <- filter(summary_data, condition %in% pair)</pre>
 pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(
   formula=discovered_extra_tasks_0.01~condition,
   data=pair_data,
   exact=FALSE,
   paired=FALSE
 )
 print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=6573.5"
## [1] "STATIC<-->PLASTIC: W=1918.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=3096"
```

#### 4.9 Novel task discovery frequency (lineage)

```
summary_data$dominant_lineage_extra_traits_discovered_per_generation <- summary_data$d
summary_data$dominant_lineage_extra_traits_generations_per_discovery <- summary_data$d
# Compute manual labels for geom_signif
# stat.test <- filter(summary_data, dominant_lineage_extra_traits_discovered > 0) %>%
   wilcox test(dominant lineage extra traits generations per discovery ~ condition) %
   adjust_pvalue(method = "bonferroni") %>%
   add significance() %>%
   add_xy_position(x="condition") # ,step.increase=1
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_extra_traits_discovered_per_generation ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
 add_significance() %>%
  add_xy_position(x="condition", step.increase=0.0001) # , step.increase=1
# Tweak y.position manually to account for scaled axis (edge case that triggers bad be
stat.test$manual_position <- stat.test$y.position #* c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
lineage_novel_task_discovery_freq_fig <- ggplot(</pre>
    # filter(summary_data, dominant_lineage_extra_traits_discovered > 0),
    summary_data,
    aes(x=condition, y=dominant_lineage_extra_traits_discovered_per_generation, fill=c
 ) +
 geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  geom_boxplot(
   width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
   name="Condition",
   limits=condition_order
 ylab("Novel task discovery frequency") +
```

```
scale_fill_brewer(
 palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
   p_label(signif(kruskal.test(formula=dominant_lineage_extra_traits_discovered_per_generation)
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),</pre>
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
 manual=TRUE,
 inherit.aes=FALSE
# coord_flip() +
theme(
  legend.position="none"
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position
lineage\_novel\_task\_discovery\_freq\_fig



```
kruskal.test(
  formula=dominant_lineage_extra_traits_discovered_per_generation~condition,
  data=summary_data
)
##
   Kruskal-Wallis rank sum test
##
##
## data: dominant_lineage_extra_traits_discovered_per_generation by condition
## Kruskal-Wallis chi-squared = 7.1465, df = 2, p-value = 0.02806
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_extra_traits_discovered_per_generation,
  g=summary_data$condition,
 p.adjust.method="bonferroni",
 conf.int=TRUE,
  conf.level=0.95
)
##
```

Pairwise comparisons using Wilcoxon rank sum test with continuity correction

summary\_data\$dominant\_lineage\_extra\_traits\_discovered\_per\_generation and sum

##

## ##

##

##

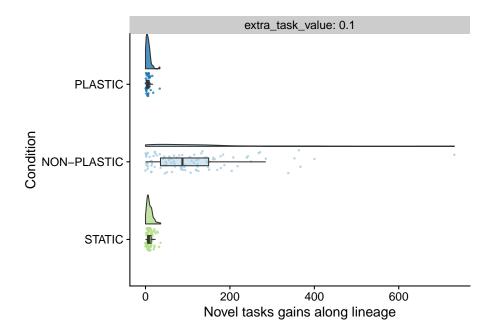
## PLASTIC 0.092

NON-PLASTIC PLASTIC

```
## STATIC 1.000
                     0.025
## P value adjustment method: bonferroni
paste(
 sep="; ",
 paste0(
   "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_extra_traits_discovered_pe
 ),
 paste0(
   "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$dominant_lineage_extra_traits_discovered_per
 ),
 paste0(
   "NON-PLASTIC median: ",
   median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_extra_traits_discovered
 )
)
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
 pair_data <- filter(summary_data, condition %in% pair)</pre>
 pair_data$condition <- as.factor(pair_data$condition)</pre>
 wt <- wilcox.test(
   formula=dominant_lineage_extra_traits_discovered_per_generation~condition,
   data=pair data,
   exact=FALSE,
   paired=FALSE
 print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=4751"
## [1] "STATIC<-->PLASTIC: W=1510.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=2584"
       Novel tasks gained (lineage)
```

```
ggplot(
    summary_data,
    aes(x=condition, y=dominant_lineage_extra_traits_gained, fill=condition)
```

```
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
   size = .5,
    alpha = 0.8
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
   alpha = 0.5
  ) +
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  ylab("Novel tasks gains along lineage") +
  scale_fill_brewer(
   palette=cb_palette
  ) +
  scale_color_brewer(
   palette=cb_palette
  ) +
  coord_flip() +
  facet wrap(
    ~extra_task_value,
   labeller=label_both
  ) +
  theme(
    legend.position="none"
  ggsave(
    paste0(working_directory, "plots/dominant-lineage-extra-tasks-gained.pdf"),
    width=15,
    height=10
```



#### 4.11 Novel task loss (lineage)

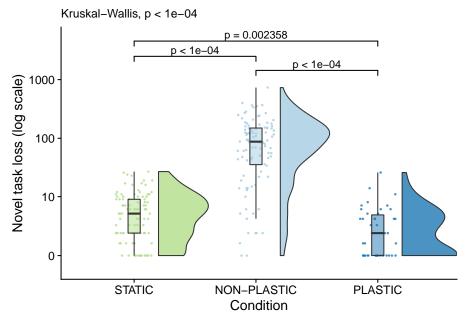
```
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_extra_traits_lost ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition", step.increase=1)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in
stat.test$manual_position <- log10(stat.test$y.position) * c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
lineage_novel_task_loss_fig <- ggplot(</pre>
    summary_data,
    aes(x=condition, y=dominant_lineage_extra_traits_lost, fill=condition)
  ) +
  geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
  ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
```

```
size = .5,
  alpha = 0.8
) +
geom_boxplot(
 width = .1,
 outlier.shape = NA,
 alpha = 0.5
) +
scale_x_discrete(
 name="Condition",
 limits=condition_order,
 labels=condition_order
) +
scale_y_continuous(
  name="Novel task loss (log scale)",
  trans=pseudo_log_trans(sigma=1, base=10),
  breaks=c(0,10,100,1000),
 limits=c(-1,5000)
scale_fill_brewer(
 palette=cb_palette
) +
scale_color_brewer(
 palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_extra_traits_lost~condition
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj<=alpha),</pre>
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
 manual=TRUE,
 inherit.aes=FALSE
) +
# coord_flip()
theme(
  legend.position="none"
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position

#### lineage\_novel\_task\_loss\_fig

##



```
kruskal.test(
  formula=dominant_lineage_extra_traits_lost~condition,
  data=summary_data
)
##
##
   Kruskal-Wallis rank sum test
##
## data: dominant_lineage_extra_traits_lost by condition
## Kruskal-Wallis chi-squared = 129.06, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_extra_traits_lost,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
)
```

## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary\_data\$dominant\_lineage\_extra\_traits\_lost and summary\_data\$condition
##

```
##
           NON-PLASTIC PLASTIC
## PLASTIC 2.7e-16
## STATIC < 2e-16
                       0.0024
## P value adjustment method: bonferroni
paste(
 sep="; ",
  paste0(
    "PLASTIC median: ",
   median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_extra_traits_loc
 ),
  paste0(
    "STATIC median: ",
   median(filter(summary_data, condition=="STATIC")$dominant_lineage_extra_traits_los
  ),
  paste0(
   "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_extra_trait
  )
)
## [1] "PLASTIC median: 2; STATIC median: 5; NON-PLASTIC median: 87.5"
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
 pair_data <- filter(summary_data, condition %in% pair)</pre>
  pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(
    formula=dominant_lineage_extra_traits_lost~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
 print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=9105"
## [1] "STATIC<-->PLASTIC: W=1353.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=3959"
```

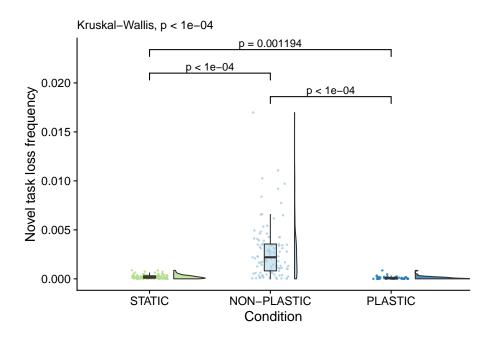
ylab("Novel task loss frequency") +

#### 4.12 Frequency of novel task loss (lineage)

```
summary_data$dominant_lineage_extra_traits_lost_per_generation <- summary_data$dominant_lineage_e
summary_data$dominant_lineage_extra_traits_generations_per_loss <- summary_data$dominant_generations_
# Compute manual labels for geom_signif
\# stat.test <- filter(summary_data, dominant_lineage_extra_traits_lost > 0) %>%
# wilcox_test(dominant_lineage_extra_traits_generations_per_loss ~ condition) %>%
# adjust_pvalue(method = "bonferroni") %>%
# add significance() %>%
# add_xy_position(x="condition", step.increase=1)
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_extra_traits_lost_per_generation ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition", step.increase=.1)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
\text{stat.test\$manual\_position} \leftarrow \text{stat.test\$y.position} \ \#* \ c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)</pre>
lineage_novel_task_loss_freq_fig <- ggplot(</pre>
    # filter(summary_data, dominant_lineage_extra_traits_lost > 0),
    summary_data,
    aes(x=condition, y=dominant_lineage_extra_traits_lost_per_generation, fill=condition)
  ) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  scale_x_discrete(
    name="Condition",
    limits=condition_order
```

```
scale_fill_brewer(
  palette=cb_palette
scale_color_brewer(
  palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_extra_traits_lost_per_gener
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj<=alpha),</pre>
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y\_position
lineage\_novel\_task\_loss\_freq\_fig



```
kruskal.test(
     formula=dominant_lineage_extra_traits_lost_per_generation~condition,
     data=summary_data
)
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_extra_traits_lost_per_generation by condition
## Kruskal-Wallis chi-squared = 121.41, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
     x=summary_data$dominant_lineage_extra_traits_lost_per_generation,
     g=summary_data$condition,
     p.adjust.method="bonferroni",
     conf.int=TRUE,
     conf.level=0.95
)
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_extra_traits_lost_per_generation and summary_data$condit
##
                              NON-PLASTIC PLASTIC
##
## PLASTIC 1.1e-15
## STATIC < 2e-16
                                                               0.0012
## P value adjustment method: bonferroni
paste(
     sep="; ",
     paste0(
           "PLASTIC median: ",
          median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_extra_traits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general_netraits_lost_per_general
     ),
     paste0(
           "STATIC median: ",
          median(filter(summary_data, condition=="STATIC")$dominant_lineage_extra_traits_lost_per_gener
     ),
     paste0(
           "NON-PLASTIC median: ",
          median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_extra_traits_lost_per_
     )
)
```

## [1] "PLASTIC median: 6.25141973661864e-05; STATIC median: 0.000161396283669756; NON-PLASTIC median: 0.000161396283669756;

```
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"

for (pair in pairwise_comparisons) {
    pair_data <- filter(summary_data, condition %in% pair)
    pair_data$condition <- as.factor(pair_data$condition)
    wt <- wilcox.test(
        formula=dominant_lineage_extra_traits_lost_per_generation~condition,
        data=pair_data,
        exact=FALSE,
        paired=FALSE
    )
    print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=8940"

## [1] "STATIC<-->PLASTIC: W=1311"

## [1] "PLASTIC<-->NON-PLASTIC: W=3922"
```

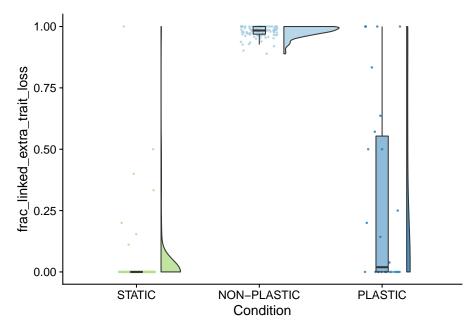
## 4.13 How many instances of novel trait loss cooccurred with changes in base phenotype?

Task loss linked with primary trait changes.

```
lost_traits_summary_data <- filter(summary_data, extra_task_value==0.1 & dominant_line)
lost_traits_summary_data$frac_linked_extra_trait_loss <- lost_traits_summary_data$dominant_line)</pre>
```

```
ggplot(lost_traits_summary_data, aes(x=condition, y=frac_linked_extra_trait_loss, fill=
 geom_flat_violin(
   position = position nudge(x = .2, y = 0),
   alpha = .8
 ) +
 geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
 ) +
 scale_x_discrete(
   name="Condition",
```

```
limits=condition_order
) +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
# coord_flip() +
theme(
  legend.position="none"
)
```



```
kruskal.test(
  formula=frac_linked_extra_trait_loss~condition,
  data=lost_traits_summary_data
)

##

## Kruskal-Wallis rank sum test

##

## data: frac_linked_extra_trait_loss by condition

## Kruskal-Wallis chi-squared = 153.68, df = 2, p-value < 2.2e-16

pairwise.wilcox.test(
  x=lost_traits_summary_data$frac_linked_extra_trait_loss,</pre>
```

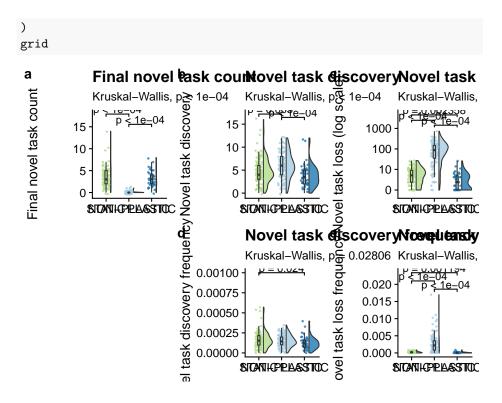
```
g=lost_traits_summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: lost_traits_summary_data$frac_linked_extra_trait_loss and lost_traits_summary
##
##
           NON-PLASTIC PLASTIC
## PLASTIC 1.9e-08
## STATIC < 2e-16
                       1.8e-06
##
## P value adjustment method: bonferroni
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(lost_traits_summary_data, condition=="PLASTIC")$frac_linked_extra_traits_summary_data
  paste0(
    "STATIC median: ",
    median(filter(lost_traits_summary_data, condition=="STATIC")$frac_linked_extra_tra
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(lost_traits_summary_data, condition=="NON-PLASTIC")$frac_linked_extra
)
## [1] "PLASTIC median: 0.0192307692307692; STATIC median: 0; NON-PLASTIC median: 0.98
print("Wilcox rank sum test statistics:")
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(lost_traits_summary_data, condition %in% pair)</pre>
  pair_data$condition <- as.factor(pair_data$condition)</pre>
  wt <- wilcox.test(
    formula=frac_linked_extra_trait_loss~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
```

```
print(paste0(pair[1], "<-->", pair[2], ": W=",wt$statistic))
## [1] "STATIC<-->NON-PLASTIC: W=8344"
## [1] "STATIC<-->PLASTIC: W=1602"
## [1] "PLASTIC<-->NON-PLASTIC: W=2212"
sum(filter(lost_traits_summary_data, condition=="NON-PLASTIC")$dominant_lineage_extra_traits_lost
## [1] 10998
sum(filter(lost_traits_summary_data, condition=="NON-PLASTIC")$dominant_lineage_extra_traits_lost
## [1] 11229
aggregate_frac_linked_extra_trait_loss_nonplastic <- sum(filter(lost_traits_summary_data, condition))
aggregate_frac_linked_extra_trait_loss_nonplastic
## [1] 0.9794283
sum(filter(lost_traits_summary_data, condition=="PLASTIC")$dominant_lineage_extra_traits_lost_lin
## [1] 29
sum(filter(lost_traits_summary_data, condition=="PLASTIC")$dominant_lineage_extra_traits_lost)
## [1] 142
aggregate_frac_linked_extra_trait_loss_plastic <- sum(filter(lost_traits_summary_data, condition=
aggregate_frac_linked_extra_trait_loss_plastic
## [1] 0.2042254
sum(filter(lost_traits_summary_data, condition=="STATIC")$dominant_lineage_extra_traits_lost_link
## [1] 13
sum(filter(lost_traits_summary_data, condition=="STATIC")$dominant_lineage_extra_traits_lost)
## [1] 631
aggregate_frac_linked_extra_trait_loss_nonplastic <- sum(filter(lost_traits_summary_data, condition))
{\tt aggregate\_frac\_linked\_extra\_trait\_loss\_nonplastic}
## [1] 0.02060222
```

#### 4.14 Manuscript figures

#### 4.15 Combined panel

```
grid <- plot_grid(</pre>
 final_novel_task_count_fig +
    theme(
      axis.title.x=element blank()
    ggtitle("Final novel task count"),
 lineage_novel_task_discovery_fig +
      axis.title.x=element_blank(),
      # axis.text.x=element_blank(),
      # axis.ticks.x=element_blank()
    ggtitle("Novel task discovery"),
 lineage_novel_task_loss_fig +
    theme(
      axis.title.x=element_blank(),
      # axis.text.x=element_blank(),
      # axis.ticks.x=element blank()
   ) +
    ggtitle("Novel task loss"),
 NULL,
 lineage_novel_task_discovery_freq_fig +
    theme(
      axis.title.x=element_blank()
    ) +
    ggtitle("Novel task discovery frequency"),
 lineage_novel_task_loss_freq_fig +
    theme(
      axis.title.x=element_blank()
    ) +
    ggtitle("Novel task loss frequency"),
 nrow=2,
 align="v",
  # labels="auto"
 labels=c("a", "b", "c", "", "d", "e")
save_plot(
  pasteO(working_directory, "plots/", "complex-traits-panel.pdf"),
  grid,
  base height=12,
  base_asp=3/2
```



#### 98CHAPTER 4. EVOLUTION AND MAINTENANCE OF NOVEL TRAITS

## Chapter 5

# Accumulation of deleterious instructions

The effect of adaptive phenotypic plasticity on the accumulation of deleterious genes.

#### 5.1 Overview

```
total_updates <- 200000
replicates <- 100

focal_traits <- c("not","nand","and","ornot","or","andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-02-05-hitchhiking/analysis/" # << For bookdown
# working_directory <- "./"</pre>
```

#### 5.2 Analysis dependencies

Load all required R libraries.

```
library(RColorBrewer)
library(ggplot2)
library(tidyverse)
library(cowplot)
library(Hmisc)
```

```
library(fmsb)
library(knitr)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd9")
These analyses were conducted/knitted with the following computing environ-
ment:
print(version)
##
## platform
                  x86_64-pc-linux-gnu
## arch
                  x86 64
## os
                  linux-gnu
## system
                  x86_64, linux-gnu
## status
## major
                  0.4
## minor
                  2021
## year
```

#### 5.3 Setup

## month

## svn rev

## language

## nickname

## day

02

15

R

80002

## version.string R version 4.0.4 (2021-02-15)

Lost Library Book

library(boot)

```
###### summary data ######
summary_data_loc <- pasteO(working_directory, "data/aggregate.csv")
summary_data <- read.csv(summary_data_loc, na.strings="NONE")

summary_data$DISABLE_REACTION_SENSORS <- as.factor(summary_data$DISABLE_REACTION_SENSOR)
summary_data$chg_env <- summary_data$chg_env == "True"
summary_data$dominant_plastic_odd_even <- as.factor(summary_data$dominant_plastic_odd_even mary_data$sensors <- summary_data$DISABLE_REACTION_SENSORS == "O"
summary_data$is_plastic <- summary_data$dominant_plastic_odd_even == "True"
summary_data$POISON_PENALTY <- as.factor(summary_data$POISON_PENALTY)

env_label_fun <- function(chg_env) {
    return("Fluctuating")
} else {
    return("Constant")
}</pre>
```

5.3. SETUP 101

```
sensors_label_fun <- function(has_sensors) {</pre>
  if (has_sensors) {
    return("Sensors")
 } else {
    return("No sensors")
  }
}
condition_label_fun <- function(has_sensors, env_chg) {</pre>
  if (has_sensors && env_chg) {
    return("PLASTIC")
 } else if (env_chg) {
    return("NON-PLASTIC")
 } else {
    return("STATIC")
}
summary_data$env_label <- mapply(</pre>
  env_label_fun,
  summary_data$chg_env
summary_data$sensors_label <- mapply(</pre>
  sensors_label_fun,
  summary_data$sensors
summary_data$condition <- mapply(</pre>
  condition_label_fun,
  summary_data$sensors,
  summary_data$chg_env
condition_order = c(
  "STATIC",
  "NON-PLASTIC",
  "PLASTIC"
)
##### time series #####
lineage_time_series_data_loc <- paste0(working_directory, "data/lineage_series.csv")</pre>
lineage_time_series_data <- read.csv(lineage_time_series_data_loc)</pre>
lineage_time_series_data$DISABLE_REACTION_SENSORS <- as.factor(lineage_time_series_data$DISABLE_I
```

```
lineage_time_series_data$chg_env <- lineage_time_series_data$chg_env == "True"</pre>
lineage_time_series_data$sensors <- lineage_time_series_data$DISABLE_REACTION_SENSORS
lineage_time_series_data$POISON_PENALTY <- as.factor(lineage_time_series_data$POISON_V</pre>
lineage_time_series_data$env_label <- mapply(</pre>
  env_label_fun,
  lineage_time_series_data$chg_env
lineage_time_series_data$sensors_label <- mapply(</pre>
  sensors_label_fun,
 lineage_time_series_data$sensors
lineage_time_series_data$condition <- mapply(</pre>
  condition_label_fun,
  lineage_time_series_data$sensors,
  lineage_time_series_data$chg_env
###### misc ######
# Configure our default graphing theme
theme_set(theme_cowplot())
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)
samplemean <- function(x, d) {</pre>
 return(mean(x[d]))
}
```

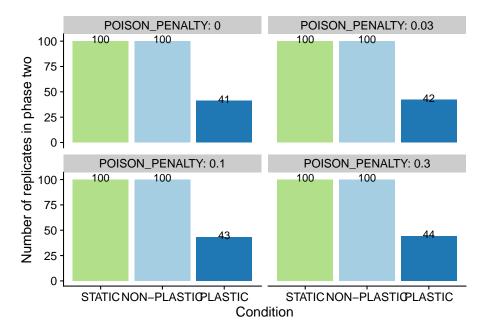
#### 5.4 Evolution of phenotypic plasticity

For sensor-enabled populations in fluctuating environments, we only transferred populations containing an optimally plastic genotype to phase-two.

```
summary_data_grouped = dplyr::group_by(summary_data, sensors, env_label, condition, PO
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())

ggplot(summary_data_group_counts, aes(x=condition, y=n, fill=condition)) +
    geom_col(position=position_dodge(0.9)) +
    geom_text(aes(label=n, y=n+2)) +
    scale_x_discrete(
    name="Condition",
    limits=condition_order
) +
    scale_fill_brewer(
    palette="Paired"
) +
    scale_color_brewer(
```

```
palette="Paired"
) +
ylab("Number of replicates in phase two") +
facet_wrap(~POISON_PENALTY, labeller=label_both) +
theme(
  legend.position="none"
)
```

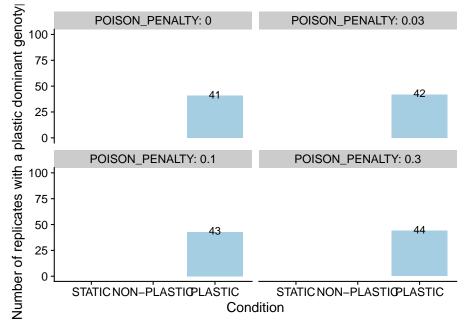


We can confirm our expectation that the dominant genotypes in non-plastic conditions are not phenotypically plastic.

```
summary_data_grouped = dplyr::group_by(summary_data, condition, is_plastic, POISON_PENALTY)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
```

```
## `summarise()` has grouped output by 'condition', 'is_plastic'. You can override using the `.gr
ggplot(filter(summary_data_group_counts, is_plastic), aes(x=condition, y=n, fill=condition)) +
    geom_col(position=position_dodge(0.9)) +
    scale_x_discrete(
        name="Condition",
        limits=condition_order
) +
    geom_text(aes(label=n, y=n+1)) +
    scale_fill_brewer(
        palette="Paired"
) +
```

```
scale_color_brewer(
   palette="Paired"
) +
ylab("Number of replicates with a plastic dominant genotype") +
ylim(0, 100) +
facet_wrap(~POISON_PENALTY, labeller=label_both) +
theme(
   legend.position="none"
)
```



#### 5.5 Poison instruction execution

## 5.5.1 Number of replicates where final dominant genotype executes the poison instruction

```
length(filter(summary_data, POISON_PENALTY==poison_penalty & condition=="PLASTIC")$RANDOM_SEED)
  length(filter(summary_data, POISON_PENALTY==poison_penalty & condition=="STATIC" )$RANDOM_SEED)
names(trials) <- c(</pre>
  "NON-PLASTIC",
  "PLASTIC",
  "STATIC"
)
names(occurrences) <- c(</pre>
 "NON-PLASTIC",
  "PLASTIC",
  "STATIC"
)
poison_exec_table <- data.frame(</pre>
  executes.poison=occurrences,
 replicates=trials
kable(poison_exec_table)
```

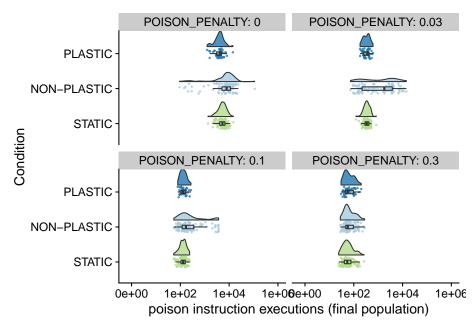
|             | executes.poison | replicates |
|-------------|-----------------|------------|
| NON-PLASTIC | 14              | 100        |
| PLASTIC     | 0               | 43         |
| STATIC      | 0               | 100        |

```
pairwise.fisher.test(x=occurrences, n=trials, p.adjust.method="bonferroni")
##
## Pairwise comparisons using Pairwise comparison of proportions (Fisher)
##
## data: occurrences out of trials
##
## NON-PLASTIC PLASTIC
## PLASTIC 0.03212 -
## STATIC 0.00022 1.00000
##
## P value adjustment method: bonferroni
```

#### 5.5.2 Poison instruction execution in final population

```
ggplot(summary_data, aes(x=condition, y=final_population_poison, fill=condition)) +
  geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
    alpha = .8
) +
  geom_point(
```

```
mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
 width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
 name="Condition",
 limits=condition_order
scale_y_continuous(
 name="poison instruction executions (final population)",
  trans="pseudo_log",
 breaks=c(0,100,10000,1000000),
 limits=c(-1,1000000)
) +
scale_fill_brewer(
 palette="Paired"
) +
scale_color_brewer(
 palette="Paired"
facet_wrap(
  ~POISON_PENALTY,
  labeller=label both
) +
coord_flip() +
theme(
 legend.position="none"
) +
ggsave(
 pasteO(working_directory, "plots/final-population-poison-log.pdf"),
 width=15,
 height=10
```



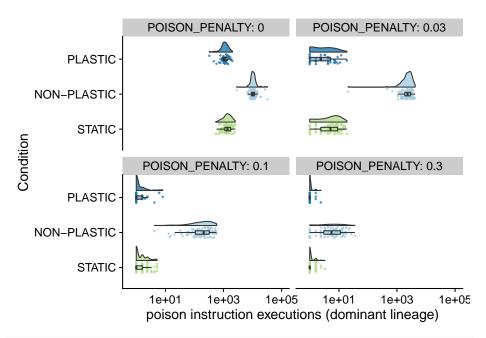
```
penalties <- levels(summary_data$POISON_PENALTY)</pre>
for (penalty in penalties) {
  stat_data <- filter(summary_data, POISON_PENALTY==penalty)</pre>
  print(
    paste0(
      "PENALTY: ", penalty
  kt <- kruskal.test(</pre>
      formula=final_population_poison~condition,
      data=stat_data
    )
 print(
    kt
  if (is.na(kt$p.value)) { next }
  if (kt$p.value > 0.05) { next }
  print(
    pairwise.wilcox.test(
      x=stat_data$final_population_poison,
      g=stat_data$condition,
      p.adjust.method="bonferroni"
    )
}
```

```
## [1] "PENALTY: O"
##
## Kruskal-Wallis rank sum test
##
## data: final_population_poison by condition
## Kruskal-Wallis chi-squared = 43.589, df = 2, p-value = 3.426e-10
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$final_population_poison and stat_data$condition
##
          NON-PLASTIC PLASTIC
## PLASTIC 8.7e-07
## STATIC 9.8e-07
                     0.00074
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
## Kruskal-Wallis rank sum test
##
## data: final_population_poison by condition
## Kruskal-Wallis chi-squared = 20.74, df = 2, p-value = 3.136e-05
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat_data$final_population_poison and stat_data$condition
##
          NON-PLASTIC PLASTIC
##
## PLASTIC 0.003
## STATIC 1e-04
                      1.000
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
## Kruskal-Wallis rank sum test
##
## data: final_population_poison by condition
## Kruskal-Wallis chi-squared = 20.608, df = 2, p-value = 3.35e-05
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat_data$final_population_poison and stat_data$condition
##
```

## 5.5.3 Poison instruction execution along final dominant lineage (cumulative)

```
ggplot(summary_data, aes(x=condition, y=dominant_lineage_times_poison_executed, fill=condition))
  geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
  ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  ) +
  scale_y_continuous(
   name="poison instruction executions (dominant lineage)",
   trans="pseudo_log",
   breaks=c(10,1000,100000),
   limits=c(-1,100000)
  ) +
  facet_wrap(
    ~POISON PENALTY,
   labeller=label_both
```

```
scale_fill_brewer(
    palette="Paired"
) +
scale_color_brewer(
    palette="Paired"
) +
coord_flip() +
theme(
    legend.position="none"
) +
ggsave(
    paste0(working_directory, "plots/final-dominant-lineage-poison-log.pdf"),
    width=15,
    height=10
)
```



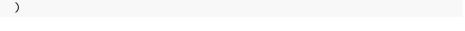
```
penalties <- levels(summary_data$POISON_PENALTY)
for (penalty in penalties) {
   stat_data <- filter(summary_data, POISON_PENALTY==penalty)
   print(
     paste0(
        "PENALTY: ", penalty
     )
   )
   kt <- kruskal.test(</pre>
```

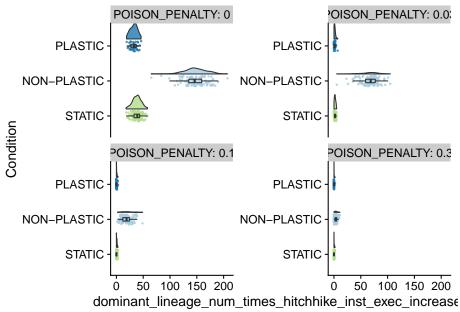
```
formula=dominant_lineage_times_poison_executed~condition,
      data=stat_data
   )
 print(
   kt
  if (is.na(kt$p.value)) { next }
  if (kt$p.value > 0.05) { next }
 print(
   pairwise.wilcox.test(
     x=stat_data$dominant_lineage_times_poison_executed,
      g=stat_data$condition,
     p.adjust.method="bonferroni"
   )
  )
}
## [1] "PENALTY: O"
##
## Kruskal-Wallis rank sum test
## data: dominant_lineage_times_poison_executed by condition
## Kruskal-Wallis chi-squared = 178.84, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat_data$dominant_lineage_times_poison_executed and stat_data$condition
          NON-PLASTIC PLASTIC
##
## PLASTIC <2e-16
## STATIC <2e-16
                       0.0018
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
##
## Kruskal-Wallis rank sum test
## data: dominant_lineage_times_poison_executed by condition
## Kruskal-Wallis chi-squared = 178.62, df = 2, p-value < 2.2e-16
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat_data$dominant_lineage_times_poison_executed and stat_data$condition
```

```
##
##
          NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
                     0.011
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_times_poison_executed by condition
## Kruskal-Wallis chi-squared = 184.83, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_times_poison_executed and stat_data$condition
##
##
          NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
                      0.21
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.3"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_times_poison_executed by condition
## Kruskal-Wallis chi-squared = 149.48, df = 2, p-value < 2.2e-16
##
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_times_poison_executed and stat_data$condition
##
          NON-PLASTIC PLASTIC
## PLASTIC 4.4e-16
## STATIC < 2e-16
                      0.84
## P value adjustment method: bonferroni
```

- 5.6 Characterizing mutations that increase poison instruction execution
- 5.6.1 Number of offspring along dominant lineage with increase in poison instruction execution

```
ggplot(summary_data, aes(x=condition, y=dominant_lineage_num_times_hitchhike_inst_exec_increases.
  geom flat violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
 ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
  ) +
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  scale fill brewer(
   palette="Paired"
  ) +
  scale_color_brewer(
   palette="Paired"
  coord_flip() +
  facet_wrap(
    ~POISON_PENALTY,
   labeller=label_both,
   scales="free_y"
  ) +
  theme(
   legend.position="none"
  ) +
   paste0(working_directory, "plots/final-dominant-lineage-poison-increase-num-mutants-log.png")
   width=15,
   height=10
```





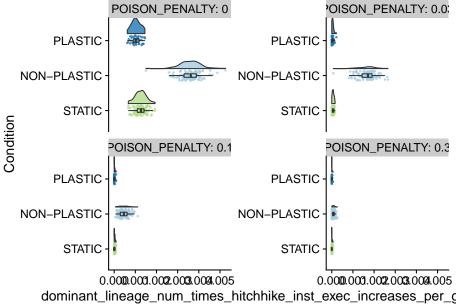
```
penalties <- levels(summary_data$POISON_PENALTY)</pre>
for (penalty in penalties) {
  stat_data <- filter(summary_data, POISON_PENALTY==penalty)</pre>
  print(
    paste0(
      "PENALTY: ", penalty
  kt <- kruskal.test(</pre>
      formula=dominant_lineage_num_times_hitchhike_inst_exec_increases~condition,
      data=stat_data
  print(
    kt
  if (is.na(kt$p.value)) { next }
  if (kt$p.value > 0.05) { next }
  print(
    pairwise.wilcox.test(
      x=stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases,
      g=stat_data$condition,
      p.adjust.method="bonferroni"
```

```
)
## [1] "PENALTY: O"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases by condition
## Kruskal-Wallis chi-squared = 179.79, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases and stat_data$condit
##
           NON-PLASTIC PLASTIC
##
## PLASTIC < 2e-16
## STATIC < 2e-16
                       0.00046
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
## Kruskal-Wallis rank sum test
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases by condition
## Kruskal-Wallis chi-squared = 179.35, df = 2, p-value < 2.2e-16
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases and stat_data$condit
##
           NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
                       0.03
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
##
## Kruskal-Wallis rank sum test
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases by condition
## Kruskal-Wallis chi-squared = 185.34, df = 2, p-value < 2.2e-16
##
##
```

```
Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases and stat_o
##
##
          NON-PLASTIC PLASTIC
## PLASTIC <2e-16
                       0.27
## STATIC <2e-16
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.3"
##
##
   Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases by condition
## Kruskal-Wallis chi-squared = 146.35, df = 2, p-value < 2.2e-16
##
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases and stat_
##
          NON-PLASTIC PLASTIC
##
## PLASTIC 7.8e-16
## STATIC < 2e-16
                       0.86
## P value adjustment method: bonferroni
sum(filter(summary_data, condition=="NON-PLASTIC" & POISON_PENALTY==0.1)$dominant_line
## [1] 1916
sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_:
## [1] 18
sum(filter(summary_data, condition=="STATIC" & POISON_PENALTY==0.1)$dominant_lineage_n
## [1] 58
# sum(filter(summary_data, condition=="NON-PLASTIC" & POISON_PENALTY==0.1)$dominant_li
# sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_lineaq
# sum(filter(summary_data, condition=="STATIC" & POISON_PENALTY==0.1)$dominant_lineage
```

### 5.6.2 Per-generation increases in poison instruction execution

```
summary_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation <- summary_data$
ggplot(summary_data, aes(x=condition, y=dominant_lineage_num_times_hitchhike_inst_exec_increases
  geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
 ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  ) +
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
 ) +
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  scale_fill_brewer(
   palette="Paired"
  scale_color_brewer(
   palette="Paired"
  ) +
  facet_wrap(
    ~POISON_PENALTY,
   labeller=label_both,
   scales="free_y"
  ) +
  coord_flip() +
  theme(
   legend.position="none"
 ) +
   paste0(working_directory, "plots/final-dominant-lineage-poison-increase-per-generation.png");
   width=15,
   height=10
```



```
penalties <- levels(summary_data$POISON_PENALTY)</pre>
for (penalty in penalties) {
  stat_data <- filter(summary_data, POISON_PENALTY==penalty)</pre>
 print(
    paste0(
      "PENALTY: ", penalty
 kt <- kruskal.test(</pre>
      formula=dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation~
      data=stat_data
    )
 print(
    kt
  )
  if (is.na(kt$p.value)) { next }
  if (kt$p.value > 0.05) { next }
 print(
    pairwise.wilcox.test(
      x=stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generat
      g=stat_data$condition,
      p.adjust.method="bonferroni"
  )
}
```

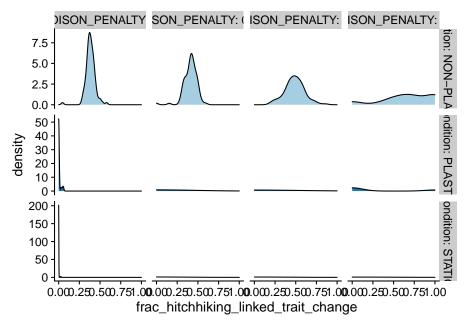
```
## [1] "PENALTY: O"
##
## Kruskal-Wallis rank sum test
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation by condition
## Kruskal-Wallis chi-squared = 180.05, df = 2, p-value < 2.2e-16
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation and a
          NON-PLASTIC PLASTIC
## PLASTIC < 2e-16
                      7.8e-05
## STATIC < 2e-16
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
##
## Kruskal-Wallis rank sum test
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation by condition
## Kruskal-Wallis chi-squared = 176.25, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat data$dominant lineage num times hitchhike inst exec increases per generation and s
##
          NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
                       0.019
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
## Kruskal-Wallis rank sum test
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation by condition
## Kruskal-Wallis chi-squared = 184.17, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation and a
##
```

```
##
           NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
                       0.2
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.3"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation by c
## Kruskal-Wallis chi-squared = 140.99, df = 2, p-value < 2.2e-16
##
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_gener
##
##
           NON-PLASTIC PLASTIC
## PLASTIC 2.2e-15
## STATIC < 2e-16
                       0.79
## P value adjustment method: bonferroni
```

# 5.6.3 What fraction of mutations that increase poison instruction execution co-occur with base trait changes?

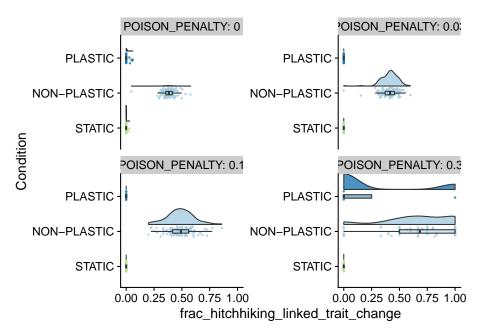
```
summary_data\frac_hitchhiking_linked_trait_change <- summary_data\frac_hitchhiking_trait_change <- summary_data\frac_hitchhiking_trait_change <- summary_data\frac_hitchhiking_trait_change <- summary_data\frac_hitchhiking_trait_change <- summary_data\frac_hitchhiking_trait_change <- summary_data\f
ggplot(filter(summary_data, dominant_lineage_num_times_hitchhike_inst_exec_increases>0
           geom_density() +
           facet_grid(
                         condition~POISON_PENALTY,
                        labeller=label_both,
                         scales="free_y"
           ) +
           scale_fill_brewer(
                       palette="Paired"
           ) +
            scale_color_brewer(
                        palette="Paired"
           ) +
           theme(
                        legend.position="none"
```

```
ggsave(
   paste0(working_directory, "plots/dominant-lineage-frac_hitchhiking_linked_trait_change.png");
   width=15,
   height=10
)
```



```
ggplot(filter(summary_data, dominant_lineage_num_times_hitchhike_inst_exec_increases>0 ), aes(x=o
  geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  geom_boxplot(
   width = .1,
    outlier.shape = NA,
   alpha = 0.5
  scale_x_discrete(
   name="Condition",
   limits=condition_order
```

```
) +
scale_fill_brewer(
   palette="Paired"
) +
scale_color_brewer(
   palette="Paired"
) +
facet_wrap(
   ~POISON_PENALTY,
   labeller=label_both,
   scales="free_y"
) +
coord_flip() +
theme(
  legend.position="none"
)
```



```
penalties <- levels(summary_data$POISON_PENALTY)
for (penalty in penalties) {
   stat_data <- filter(summary_data, POISON_PENALTY==penalty & dominant_lineage_num_time
   print(
     pasteO(
        "PENALTY: ", penalty
    )
   )
}</pre>
```

kt <- kruskal.test(</pre>

```
formula=frac_hitchhiking_linked_trait_change~condition,
      data=stat_data
   )
  print(
   kt
  if (is.na(kt$p.value)) { next }
  if (kt$p.value > 0.05) { next }
  print(
   pairwise.wilcox.test(
     x=stat_data$frac_hitchhiking_linked_trait_change,
     g=stat_data$condition,
     p.adjust.method="bonferroni",
     exact=FALSE
   )
 )
}
## [1] "PENALTY: O"
##
## Kruskal-Wallis rank sum test
## data: frac_hitchhiking_linked_trait_change by condition
## Kruskal-Wallis chi-squared = 211.29, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: stat_data$frac_hitchhiking_linked_trait_change and stat_data$condition
##
           NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
                       0.031
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
##
## Kruskal-Wallis rank sum test
##
## data: frac_hitchhiking_linked_trait_change by condition
## Kruskal-Wallis chi-squared = 186.88, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
```

```
##
## data: stat_data$frac_hitchhiking_linked_trait_change and stat_data$condition
##
          NON-PLASTIC PLASTIC
## PLASTIC 2.9e-16
## STATIC < 2e-16
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
##
##
   Kruskal-Wallis rank sum test
##
## data: frac_hitchhiking_linked_trait_change by condition
## Kruskal-Wallis chi-squared = 113.72, df = 2, p-value < 2.2e-16
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
##
## data: stat_data$frac_hitchhiking_linked_trait_change and stat_data$condition
##
          NON-PLASTIC PLASTIC
##
## PLASTIC 3.3e-08
## STATIC < 2e-16
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.3"
##
## Kruskal-Wallis rank sum test
##
## data: frac_hitchhiking_linked_trait_change by condition
## Kruskal-Wallis chi-squared = 34.791, df = 2, p-value = 2.788e-08
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
##
## data: stat_data$frac_hitchhiking_linked_trait_change and stat_data$condition
##
##
          NON-PLASTIC PLASTIC
## PLASTIC 0.26
## STATIC 2.4e-08
                       0.18
##
## P value adjustment method: bonferroni
denom <- sum(filter(summary_data, condition=="NON-PLASTIC" & POISON_PENALTY==0.1)$domi:
num <- sum(filter(summary_data, condition=="NON-PLASTIC" & POISON_PENALTY==0.1)$dominates
pasteO("NON-PLASTIC: ", num/denom, "(", num, "/", denom, ")")
```

```
## [1] "NON-PLASTIC: 0.498956158663883(956/1916)"
denom <- sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num
num <- sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num
pasteO("PLASTIC: ", num/denom, " (", num, "/", denom, ")")

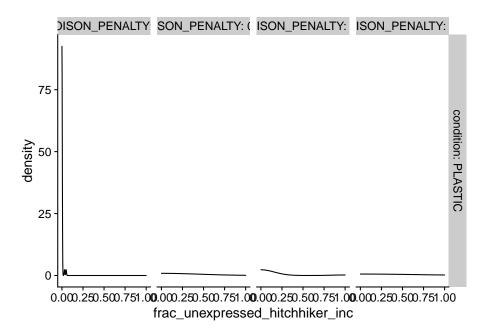
## [1] "PLASTIC: 0 (0/18)"

denom <- sum(filter(summary_data, condition=="STATIC" & POISON_PENALTY==0.1)$dominant_lineage_num
num <- sum(filter(summary_data, condition=="STATIC" & POISON_PENALTY==0.1)$dominant_lineage_num_num <- sum(filter(summary_data, sondition=="STATIC" & POISON_PENALTY==0.1
```

# 5.7 What fraction of poison execution increases occur in unexpressed phenotype (as cryptic

```
summary_data$frac_unexpressed_hitchhiker_inc <- summary_data$dominant_lineage_num_times_hitchhike
summary_data$frac_expressed_hitchiker_inc <- summary_data$dominant_lineage_num_times_hitchhike_ir

ggplot(filter(summary_data, dominant_lineage_num_times_hitchhike_inst_exec_increases>0 & condition
geom_density() +
facet_grid(
    condition~POISON_PENALTY,
    labeller=label_both,
    scales="free_y"
) +
theme(
    legend.position="none"
```



```
denom <- sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant
num <- sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_1
paste0("PLASTIC: ", num/denom, " (", num, "/", denom, ")")</pre>
```

#### 5.8 Manuscript figures

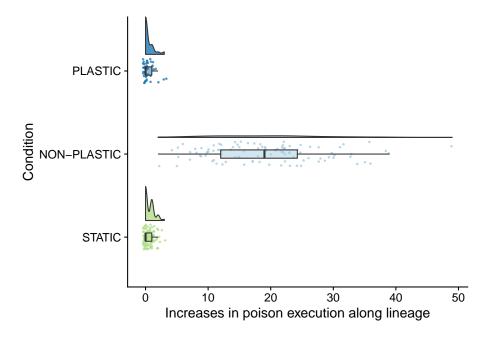
## [1] "PLASTIC: 0.0555555555556 (1/18)"

```
poison_penalty <- 0.1
```

#### 5.8.1 Total poison execution increases along lineage.

```
poison_increases_fig <- ggplot(
    filter(summary_data, POISON_PENALTY==poison_penalty),
    aes(x=condition, y=dominant_lineage_num_times_hitchhike_inst_exec_increases, fill=
) +
    geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
) +
    geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,</pre>
```

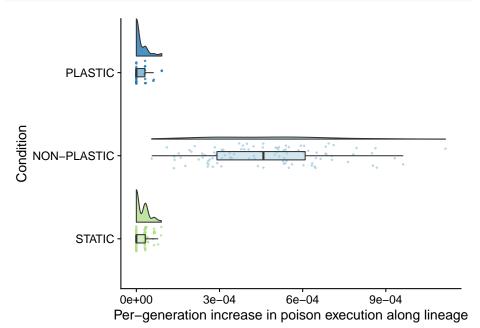
```
alpha = 0.8
 ) +
 geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
 ) +
 scale_x_discrete(
   name="Condition",
   limits=condition_order,
   labels=condition_order
 ) +
  scale_y_continuous(
   name="Increases in poison execution along lineage",
 scale_fill_brewer(
   palette="Paired"
 scale_color_brewer(
   palette="Paired"
 ) +
 theme(
   legend.position="none"
 ) +
 coord_flip()
poison_increases_fig
```



# 5.8.2 Per-generation poison execution increases along lineage

```
poison_increases_per_gen_fig <- ggplot(</pre>
    filter(summary_data, POISON_PENALTY==poison_penalty),
    aes(x=condition, y=dominant_lineage_num_times_hitchhike_inst_exec_increases_per_get
  ) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
```

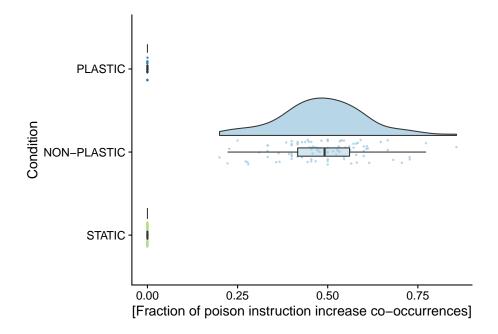
```
limits=condition_order,
  labels=condition_order
) +
scale_y_continuous(
  name="Per-generation increase in poison execution along lineage",
) +
scale_fill_brewer(
  palette="Paired"
) +
scale_color_brewer(
  palette="Paired"
) +
theme(
  legend.position="none"
) +
coord_flip()
```



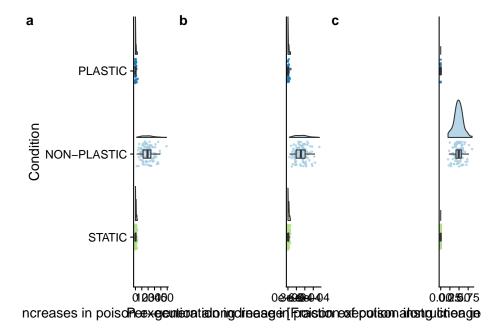
#### 5.8.3 Co-occurrence

```
linked_trait_change_fig <- ggplot(
    filter(summary_data, POISON_PENALTY==poison_penalty & dominant_lineage_num_times_hitchhike_ir
    aes(x=condition, y=frac_hitchhiking_linked_trait_change, fill=condition)</pre>
```

```
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order,
   labels=condition_order
  ) +
  scale_y_continuous(
   name="[Fraction of poison instruction increase co-occurrences]",
  scale_fill_brewer(
   palette="Paired"
  ) +
  scale_color_brewer(
    palette="Paired"
  ) +
  theme(
    legend.position="none"
  coord_flip()
linked_trait_change_fig
```



```
grid <- plot_grid(
  poison_increases_fig,
  poison_increases_per_gen_fig + theme(axis.ticks.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(),axis.text.y=element_blank(
```



### Chapter 6

### Regulation in Avida

#### 6.1 Overview

```
total_updates <- 200000
replicates <- 100

all_traits <- c("not", "nand", "and", "ornot", "or", "andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-02-08-evo-dynamics/analysis/" # << For bookdown
# working_directory <- "./" # << For local analysis</pre>
```

#### 6.2 Analysis dependencies

Load all required R libraries.

```
library(ggplot2)
library(tidyverse)
library(cowplot)
library(RColorBrewer)
library(Hmisc)
library(boot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd97121f7f9ce9
```

These analyses were conducted/knitted with the following computing environment:

```
print(version)
##
## platform
                  x86_64-pc-linux-gnu
## arch
                  x86 64
## os
                  linux-gnu
## system
                  x86_64, linux-gnu
## status
## major
                  0.4
## minor
## year
                  2021
## month
                  02
## day
                  15
                  80002
## svn rev
## language
                  R
## version.string R version 4.0.4 (2021-02-15)
                 Lost Library Book
## nickname
```

#### 6.3 Setup

```
trace_summary_data_loc <- paste0(working_directory, "data/trace_summary.csv")</pre>
trace_summary_data <- read.csv(trace_summary_data_loc, na.strings="NONE")</pre>
trace_summary_data$DISABLE_REACTION_SENSORS <- as.factor(trace_summary_data$DISABLE_RE.
trace_summary_data$chg_env <- trace_summary_data$chg_env == "True"</pre>
trace_summary_data$sensors <- trace_summary_data$DISABLE_REACTION_SENSORS == "0"</pre>
env_label_fun <- function(chg_env) {</pre>
  if (chg_env) {
    return("Fluctuating")
  } else {
    return("Constant")
  }
}
sensors_label_fun <- function(has_sensors) {</pre>
  if (has_sensors) {
    return("Sensors")
  } else {
    return("No sensors")
  }
}
```

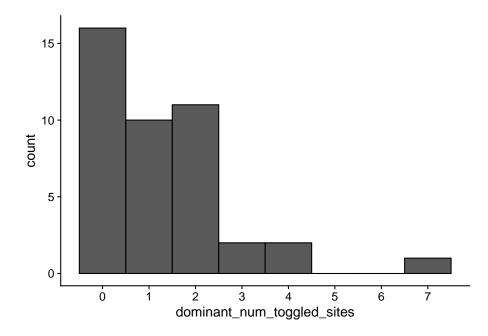
```
# note that this labeler makes assumptions about how we set up our experiment
condition_label_fun <- function(has_sensors, env_chg) {</pre>
  if (has_sensors && env_chg) {
    return("PLASTIC")
 } else if (env_chg) {
    return("NON-PLASTIC")
  } else {
    return("STATIC")
 }
}
trace_summary_data$env_label <- mapply(</pre>
  env_label_fun,
  trace_summary_data$chg_env
trace_summary_data$sensors_label <- mapply(</pre>
  sensors_label_fun,
  trace_summary_data$sensors
trace_summary_data$condition <- mapply(</pre>
  condition_label_fun,
  trace_summary_data$sensors,
  trace_summary_data$chg_env
###### misc ######
# Configure our default graphing theme
theme_set(theme_cowplot())
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)
```

# 6.4 How many instructions do plastic genomes toggle depending on environmental context?

```
ggplot(trace_summary_data, aes(x=dominant_num_toggled_sites)) +
    geom_histogram(
        binwidth=1,
        color="black"
    ) +
    scale_fill_brewer(
        palette="Paired"
    ) +
    scale_color_brewer(
        palette="Paired"
```

```
) +
scale_x_continuous(
   breaks=seq(0, max(trace_summary_data$dominant_num_toggled_sites)+1)
) +
theme(
  legend.position="none"
) +
ggsave(paste0(working_directory, "plots/", "toggled-sites.png"))
```

## Saving 6.5 x 4.5 in image



# 6.5 What is the distribution of toggled sequence sizes?

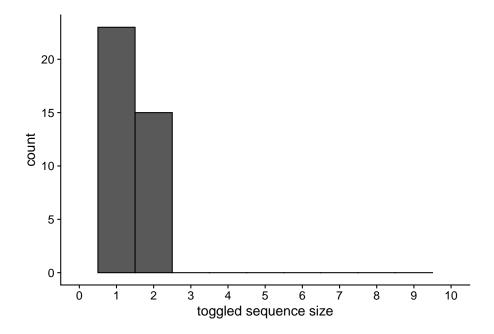
```
chunk_sizes <- data.frame(
    size=integer()
)
for (sizes in trace_summary_data$dominant_toggled_chunk_sizes) {
    if (sizes == "") { next }
        sizes <- unlist(lapply(str_split(sizes, ';'), as.integer))
        chunk_sizes <- rbind(chunk_sizes, data.frame(size=c(sizes)))
}</pre>
```

#### 6.5. WHAT IS THE DISTRUBUTION OF TOGGLED SEQUENCE SIZES?137

```
ggplot(chunk_sizes, aes(x=size)) +
   geom_histogram(
     binwidth=1,
     color="black"
   scale_fill_brewer(
     palette="Paired"
   ) +
   scale_color_brewer(
     palette="Paired"
   scale_x_continuous(
     name="toggled sequence size",
     breaks=seq(0, 10),
     limits=c(0, 10)
   ) +
   theme(
     legend.position="none"
   ggsave(paste0(working_directory, "plots/", "toggled-chunk-sizes.png"))
```

```
## Saving 6.5 \times 4.5 in image
```

```
## Warning: Removed 2 rows containing missing values (geom_bar).
## Warning: Removed 2 rows containing missing values (geom_bar).
```



### Chapter 7

# Evolutionary change (variable length genomes)

#### 7.1 Overview

```
total_updates <- 200000
replicates <- 100

all_traits <- c("not", "nand", "ornot", "or", "andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-01-30-evo-dynamics/analysis/" # << For bookdown
# working_directory <- "./"
# << For local analysis</pre>
```

#### 7.2 Analysis dependencies

Load all required R libraries.

```
library(ggplot2)
library(tidyverse)
library(cowplot)
library(RColorBrewer)
library(Hmisc)
library(boot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd97121f7f9ce8
```

These analyses were conducted/knitted with the following computing environ-

ment:

```
print(version)
##
                  x86_64-pc-linux-gnu
## platform
## arch
                  x86_64
## os
                  linux-gnu
                  x86_64, linux-gnu
## system
## status
## major
## minor
                  0.4
                  2021
## year
## month
                 02
## day
                 15
## svn rev
                 80002
## language
                 R
## version.string R version 4.0.4 (2021-02-15)
## nickname
                 Lost Library Book
```

#### 7.3 Setup

```
summary_data_loc <- paste0(working_directory, "data/aggregate.csv")</pre>
summary_data <- read.csv(summary_data_loc, na.strings="NONE")</pre>
summary_data$DISABLE_REACTION_SENSORS <- as.factor(summary_data$DISABLE_REACTION_SENSO
summary_data$chg_env <- summary_data$chg_env == "True"</pre>
summary_data$dominant_plastic_odd_even <- as.factor(summary_data$dominant_plastic_odd_.
summary_data$sensors <- summary_data$DISABLE_REACTION_SENSORS == "0"</pre>
summary_data$is_plastic <- summary_data$dominant_plastic_odd_even == "True"
env_label_fun <- function(chg_env) {</pre>
  if (chg_env) {
   return("Fluctuating")
 } else {
    return("Constant")
  }
sensors_label_fun <- function(has_sensors) {</pre>
  if (has sensors) {
   return("Sensors")
 } else {
    return("No sensors")
```

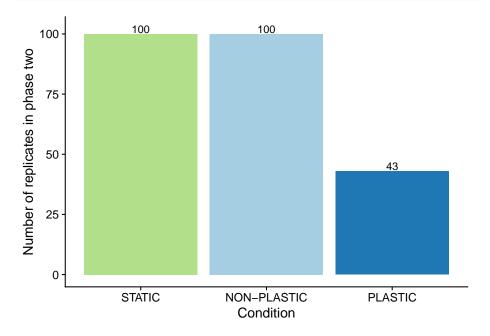
```
# note that this labeler makes assumptions about how we set up our experiment
condition_label_fun <- function(has_sensors, env_chg) {</pre>
  if (has_sensors && env_chg) {
    return("PLASTIC")
  } else if (env_chg) {
    return("NON-PLASTIC")
  } else {
    return("STATIC")
 }
}
summary_data$env_label <- mapply(</pre>
  env_label_fun,
  summary_data$chg_env
)
summary_data$sensors_label <- mapply(</pre>
  sensors_label_fun,
  summary_data$sensors
)
summary_data$condition <- mapply(</pre>
  condition_label_fun,
  summary_data$sensors,
  summary_data$chg_env
condition_order = c(
  "STATIC",
  "NON-PLASTIC",
  "PLASTIC"
)
###### misc ######
# Configure our default graphing theme
theme_set(theme_cowplot())
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)
```

#### 7.4 Evolution of phenotypic plasticity

For sensor-enabled populations in fluctuating environments, we only transfered populations containing an optimally plastic genotype to phase-two.

```
summary_data_grouped = dplyr::group_by(summary_data, sensors, env_label, condition)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
```

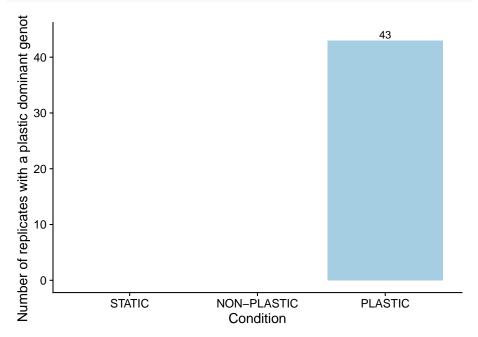
```
## `summarise()` has grouped output by 'sensors', 'env_label'. You can override using 'ggplot(summary_data_group_counts, aes(x=condition, y=n, fill=condition)) +
    geom_col(position=position_dodge(0.9)) +
    geom_text(aes(label=n, y=n+2)) +
    scale_x_discrete(
        name="Condition",
        limits=condition_order
) +
    scale_fill_brewer(
        palette="Paired"
) +
    scale_color_brewer(
        palette="Paired"
) +
    ylab("Number of replicates in phase two") +
    theme(
        legend.position="none"
)
```



We can confirm our expectation that the dominant genotypes in non-plastic conditions are not phenotypically plastic.

```
summary_data_grouped = dplyr::group_by(summary_data, condition, is_plastic)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
```

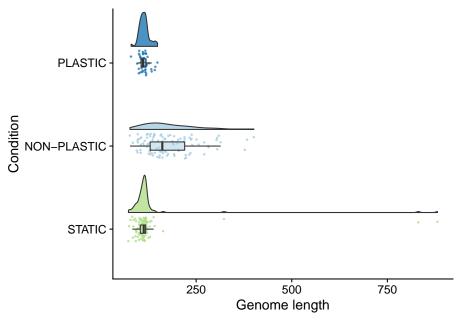
```
ggplot(filter(summary_data_group_counts, is_plastic), aes(x=condition, y=n, fill=condition)) +
  geom_col(position=position_dodge(0.9)) +
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  ) +
  scale_fill_brewer(
   palette="Paired"
  scale_color_brewer(
   palette="Paired"
  ) +
  geom_text(aes(label=n, y=n+1)) +
  ylab("Number of replicates with a plastic dominant genotype") +
  theme(
   legend.position="none"
 )
```



#### 7.5 Genome length

Single-instruction insertions and deletions were possible for this experiment, so genome size also evolved.

```
ggplot(summary_data, aes(x=condition, y=dominant_genome_length, fill=condition)) +
 geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
 ) +
 geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
 scale_x_discrete(
   name="Condition",
   limits=condition_order
  scale_fill_brewer(
   palette="Paired"
 ) +
  scale_color_brewer(
   palette="Paired"
  coord_flip() +
 ylab("Genome length") +
 theme(
    legend.position="none"
```



```
kruskal.test(
 formula=dominant_genome_length~condition,
  data=summary_data
)
##
   Kruskal-Wallis rank sum test
##
##
## data: dominant_genome_length by condition
## Kruskal-Wallis chi-squared = 82.798, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
 x=summary_data$dominant_genome_length,
  g=summary_data$condition,
 p.adjust.method="bonferroni",
)
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_genome_length and summary_data$condition
##
           NON-PLASTIC PLASTIC
## PLASTIC 1.8e-10
## STATIC < 2e-16
##
```

```
## P value adjustment method: bonferroni
median(filter(summary_data, condition=="PLASTIC")$phylo_mrca_changes)

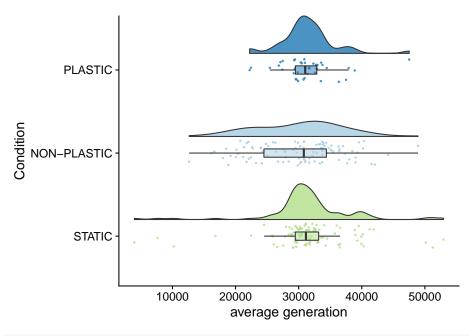
## [1] 45
median(filter(summary_data, condition=="STATIC")$phylo_mrca_changes)

## [1] 47
median(filter(summary_data, condition=="NON-PLASTIC")$phylo_mrca_changes)

## [1] 393
```

### 7.6 Average generation

```
ggplot(summary_data, aes(x=condition, y=time_average_generation, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
   alpha = .8
  ) +
 geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  ) +
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
  scale_x_discrete(
   name="Condition",
   limits=condition_order
  scale_fill_brewer(
   palette="Paired"
 scale_color_brewer(
   palette="Paired"
  coord flip() +
 ylab("average generation") +
 theme(
    legend.position="none"
```

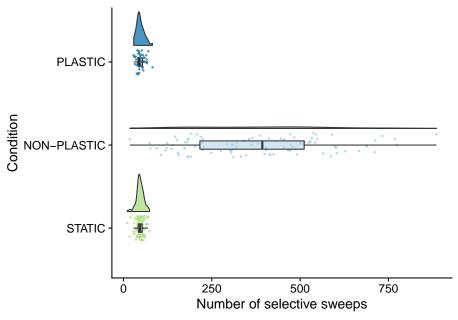


```
median(filter(summary_data, condition=="PLASTIC")$time_average_generation)
## [1] 31028.6
median(filter(summary_data, condition=="STATIC")$time_average_generation)
## [1] 31147.5
median(filter(summary_data, condition=="NON-PLASTIC")$time_average_generation)
## [1] 30817.95
kruskal.test(
  formula=time_average_generation~condition,
  data=summary_data
)
##
##
   Kruskal-Wallis rank sum test
##
## data: time_average_generation by condition
## Kruskal-Wallis chi-squared = 1.3804, df = 2, p-value = 0.5015
```

#### 7.7 Coalescence events

The number of times the most recent common ancestor changes gives us the number of selective sweeps that occur during the experiment.

```
ggplot(summary_data, aes(x=condition, y=phylo_mrca_changes, fill=condition)) +
 geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
 ) +
 geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
 geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
  scale_fill_brewer(
   palette="Paired"
  scale_color_brewer(
    palette="Paired"
  ) +
  coord_flip() +
 scale_x_discrete(
   name="Condition",
   limits=condition_order
 ylab("Number of selective sweeps") +
 theme(
    legend.position="none"
```



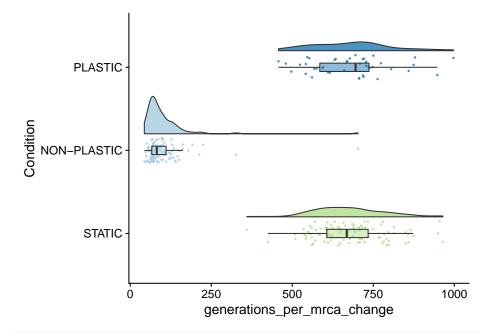
```
paste0(
 "PLASTIC: ",
  median(filter(summary_data, condition=="PLASTIC")$phylo_mrca_changes)
## [1] "PLASTIC: 45"
paste0(
  "STATIC: ",
  median(filter(summary_data, condition=="STATIC")$phylo_mrca_changes)
)
## [1] "STATIC: 47"
paste0(
  "NON-PLASTIC: ",
  median(filter(summary_data, condition=="NON-PLASTIC")$phylo_mrca_changes)
## [1] "NON-PLASTIC: 393"
kruskal.test(
  formula=phylo_mrca_changes~condition,
  data=summary_data
)
##
## Kruskal-Wallis rank sum test
```

```
##
## data: phylo_mrca_changes by condition
## Kruskal-Wallis chi-squared = 168.89, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$phylo_mrca_changes,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
##
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary data$phylo mrca changes and summary data$condition
##
##
           NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
## P value adjustment method: bonferroni
```

## 7.7.1 Average number of generations between selective sweeps

```
summary data$generations per mrca change <- summary data$time average generation / sum
ggplot(summary_data, aes(x=condition, y=generations_per_mrca_change, fill=condition))
 geom_flat_violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
 ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
   width = .1,
    outlier.shape = NA,
   alpha = 0.5
 ) +
  scale_x_discrete(
   name="Condition",
   limits=condition_order
```

```
scale_fill_brewer(
   palette="Paired"
) +
scale_color_brewer(
   palette="Paired"
) +
coord_flip() +
theme(
   legend.position="none"
)
```



```
paste0(
   "PLASTIC: ",
   median(filter(summary_data, condition=="PLASTIC")$generations_per_mrca_change)
)

## [1] "PLASTIC: 695.504761904762"

paste0(
   "STATIC: ",
   median(filter(summary_data, condition=="STATIC")$generations_per_mrca_change)
)
```

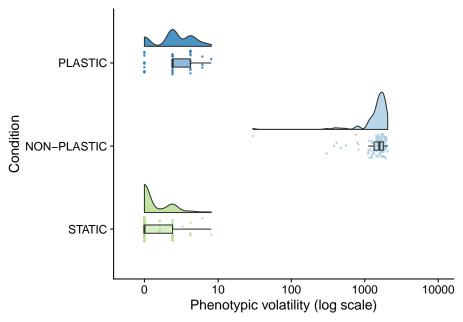
## [1] "STATIC: 668.25523255814"

```
paste0(
  "NON-PLASTIC: ",
  median(filter(summary_data, condition=="NON-PLASTIC")$generations_per_mrca_change)
## [1] "NON-PLASTIC: 81.9208459944751"
kruskal.test(
  formula=generations_per_mrca_change~condition,
  data=summary_data
##
##
   Kruskal-Wallis rank sum test
## data: generations_per_mrca_change by condition
## Kruskal-Wallis chi-squared = 171.73, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$generations_per_mrca_change,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
##
   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$generations_per_mrca_change and summary_data$condition
##
           NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
## P value adjustment method: bonferroni
```

# 7.8 Phenotypic volatility along the dominant lineage

```
ggplot(summary_data, aes(x=condition, y=dominant_lineage_trait_volatility, fill=condit
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
) +
  geom_point(
    mapping=aes(color=condition),
```

```
position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
 width = .1,
  outlier.shape = NA,
 alpha = 0.5
scale_x_discrete(
 name="Condition",
  limits=condition_order
) +
scale_y_continuous(
 name="Phenotypic volatility (log scale)",
 trans="pseudo_log",
 breaks=c(0, 10, 100, 1000, 10000),
 limits=c(-1,10000)
) +
scale_fill_brewer(
  palette="Paired"
) +
scale_color_brewer(
 palette="Paired"
) +
coord_flip() +
theme(
  legend.position="none"
```



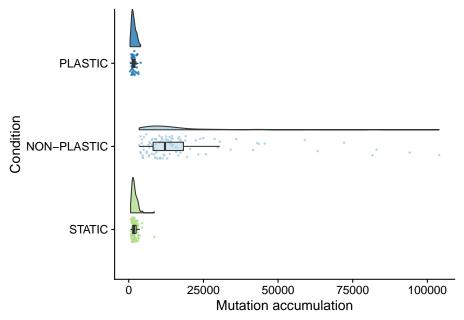
```
paste0(
  "PLASTIC: ",
  median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_trait_volatility)
## [1] "PLASTIC: 2"
paste0(
  "STATIC: ",
  median(filter(summary_data, condition=="STATIC")$dominant_lineage_trait_volatility)
## [1] "STATIC: O"
paste0(
  "NON-PLASTIC: ",
  median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_trait_volatil
## [1] "NON-PLASTIC: 1580"
kruskal.test(
  formula=dominant_lineage_trait_volatility~condition,
  data=summary_data
)
##
## Kruskal-Wallis rank sum test
```

```
##
## data: dominant_lineage_trait_volatility by condition
## Kruskal-Wallis chi-squared = 191.98, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_trait_volatility,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
## data: summary data$dominant lineage trait volatility and summary data$condition
##
          NON-PLASTIC PLASTIC
##
## PLASTIC < 2e-16
## STATIC < 2e-16
                       5.2e-08
## P value adjustment method: bonferroni
```

# 7.9 Mutation accumulation along the dominant lineage

```
ggplot(summary_data, aes(x=condition, y=dominant_lineage_total_mut_cnt, fill=condition)) +
  geom flat violin(
   position = position_nudge(x = .2, y = 0),
   alpha = .8
  ) +
  geom_point(
   mapping=aes(color=condition),
   position = position_jitter(width = .15),
   size = .5,
   alpha = 0.8
  ) +
  geom_boxplot(
   width = .1,
   outlier.shape = NA,
   alpha = 0.5
  ylab("Mutation accumulation") +
  scale_x_discrete(
   name="Condition",
   limits=condition_order
```

```
scale_fill_brewer(
   palette="Paired"
) +
scale_color_brewer(
   palette="Paired"
) +
coord_flip() +
theme(
   legend.position="none"
)
```



```
paste0(
   "PLASTIC: ",
   median(filter(summary_data, condition=="PLASTIC")$dominant_lineage_total_mut_cnt)
)

## [1] "PLASTIC: 1552"

paste0(
   "STATIC: ",
   median(filter(summary_data, condition=="STATIC")$dominant_lineage_total_mut_cnt)
)

## [1] "STATIC: 1724.5"
```

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```
paste0(
 "NON-PLASTIC: ",
 median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_total_mut_cnt)
## [1] "NON-PLASTIC: 12123"
kruskal.test(
  formula=dominant_lineage_total_mut_cnt~condition,
  data=summary_data
)
##
   Kruskal-Wallis rank sum test
##
## data: dominant_lineage_total_mut_cnt by condition
## Kruskal-Wallis chi-squared = 174.38, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
 x=summary_data$dominant_lineage_total_mut_cnt,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_total_mut_cnt and summary_data$condition
##
           NON-PLASTIC PLASTIC
## PLASTIC <2e-16
## STATIC <2e-16
                       0.57
##
## P value adjustment method: bonferroni
```