

Supplemental Material

Alexander Lalejini, Austin J. Ferguson, Nkrumah Grant, and Charles Ofria

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Contents

1	Introduction	5
2	Validation experiment	9
2.1	Overview	9
2.2	Analysis dependencies	10
2.3	Setup	10
2.4	Evolution of phenotypic plasticity	12
3	Evolutionary change	15
3.1	Overview	15
3.2	Analysis dependencies	15
3.3	Setup	16
3.4	The evolution of phenotypic plasticity	18
3.5	Average generation	20
3.6	Coalescence event count	23
3.7	Phenotypic volatility along the dominant lineage	30
3.8	Phenotypic fidelity	37
3.9	Mutation count	40
3.10	Genotypic fidelity	46
3.11	Characterizing variation along dominant lineages	50
3.12	Manuscript figures	61
4	Evolution and maintenance of novel traits	63
4.1	Overview	63
4.2	Analysis dependencies	64
4.3	Setup	65
4.4	The evolution of phenotypic plasticity	67
4.5	Final novel task count (dominant genotype)	69
4.6	Novel task count (final population)	73
4.7	Novel task discovery (lineage)	75
4.8	Novel task discovery (population)	79
4.9	Novel task discovery frequency (lineage)	82
4.10	Novel tasks gained (lineage)	85
4.11	Novel task loss (lineage)	87

4.12	Frequency of novel task loss (lineage)	91
4.13	How many instances of novel trait loss co-occurred with changes in base phenotype?	94
4.14	Manuscript figures	98
4.15	Combined panel	98
5	Accumulation of deleterious instructions	101
5.1	Overview	101
5.2	Analysis dependencies	101
5.3	Setup	102
5.4	Evolution of phenotypic plasticity	105
5.5	Poison instruction execution	107
5.6	Characterizing mutations that increase poison instruction execution	118
5.7	What fraction of poison execution increases occur in unexpressed phenotype (as cryptic variation)?	136
5.8	Manuscript figures	137
6	Regulation in Avida	139
6.1	Overview	139
6.2	Analysis dependencies	139
6.3	Setup	140
6.4	How many instructions do plastic genomes toggle depending on environmental context?	141
6.5	What is the distribution of toggled sequence sizes?	142
7	Evolutionary change (variable length genomes)	145
7.1	Overview	145
7.2	Analysis dependencies	145
7.3	Setup	146
7.4	Evolution of phenotypic plasticity	147
7.5	Genome length	149
7.6	Average generation	152
7.7	Coalescence events	153
7.8	Phenotypic volatility along the dominant lineage	158
7.9	Mutation accumulation along the dominant lineage	161

Chapter 1

Introduction

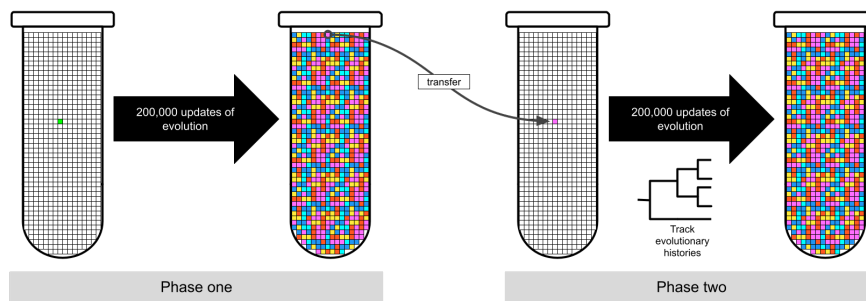


Figure 1.1: Experimental design overview

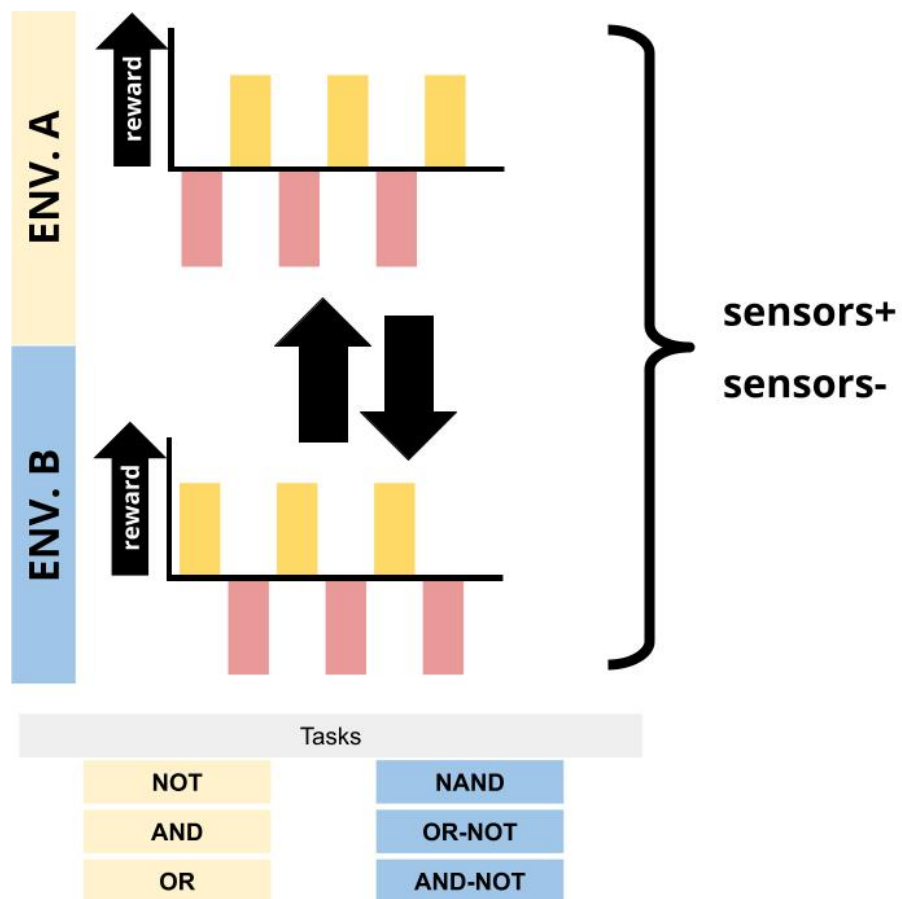


Figure 1.2: Fluctuating environment

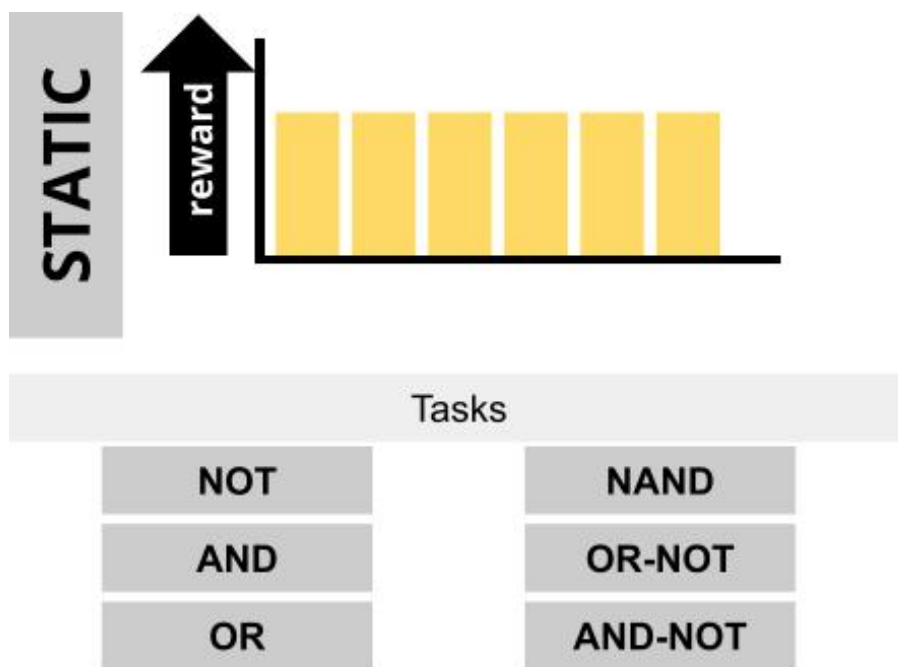


Figure 1.3: Static environment

Chapter 2

Validation experiment

In this experiment, we validate that (1) we observe the evolution of phenotypic plasticity in a changing environment when digital organisms have access to sensory instructions (capable of differentiating environmental states) and (2) that adaptive phenotypic plasticity does not evolve when populations lack access to sensory instructions.

2.1 Overview

```
total_updates <- 200000
replicates <- 100

all_traits <- c("not","nand","and","ornot","or","andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-01-07-validation/analysis/" # << For bookdown
# working_directory <- "./" # << For local analysis
```

We evolved populations of digital organisms under four conditions:

1. A fluctuating environment with access to sensory instructions
2. A fluctuating environment without access to sensory instructions (i.e., sensory instructions are no-operations)
3. A constant environment with access to sensory instructions
4. A constant environment without access to sensory instructions

In fluctuating environments, we alternate between rewarding and punishing different sets of computational tasks. In one environment, we reward tasks not,

and, or and punish tasks nand, ornot, andnot. In the alternative environment, we reward tasks nand, ornot, andnot and punish tasks not, and, or. In constant environments, we reward all tasks (not, nand, and, ornot, or, andnot).

For each replicate of each condition, we extract the dominant (i.e., most numerous) genotype at the end of the run to analyze further. We expect to observe the evolution of adaptive phenotypic plasticity in only the first experimental condition. In conditions without sensors, plasticity in any form should be unable to evolve.

2.2 Analysis dependencies

Load all required R libraries.

```
library(ggplot2)
library(tidyverse)
library(cowplot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd9")
```

These analyses were conducted/knitted with the following computing environment:

```
print(version)

##
## platform      x86_64-pc-linux-gnu
## arch          x86_64
## os            linux-gnu
## system        x86_64, linux-gnu
## status
## major         4
## minor         0.5
## year          2021
## month         03
## day           31
## svn rev       80133
## language      R
## version.string R version 4.0.5 (2021-03-31)
## nickname      Shake and Throw
```

2.3 Setup

```
data_loc <- paste0(working_directory, "data/aggregate.csv")
data <- read.csv(data_loc, na.strings="NONE")

data$DISABLE_REACTION_SENSORS <- as.factor(data$DISABLE_REACTION_SENSORS)
```

```

data$chg_env <- as.factor(data$chg_env)
data$dom_plastic_odd_even <- as.factor(data$dom_plastic_odd_even)
data$sensors <- data$DISABLE_REACTION_SENSORS == "0"
data$is_plastic <- data$dom_plastic_odd_even == "True"

env_label_fun <- function(chg_env) {
  if (chg_env) {
    return("Fluctuating")
  } else {
    return("Constant")
  }
}

sensors_label_fun <- function(has_sensors) {
  if (has_sensors) {
    return("Sensors")
  } else {
    return("No sensors")
  }
}

# Count observed plasticity for each condition (I'm sure there's a 'tidier' way to do this..)
observed_plasticity <- data.frame(
  environment=character(),
  sensors=character(),
  plastic=integer(),
  nonplastic=integer(),
  plastic_adaptive=integer(),
  plastic_optimal=integer(),
  plastic_nonadaptive=integer()
)

for (env_chg in levels(data$chg_env)) {
  for (disabled_sensors in levels(data$DISABLE_REACTION_SENSORS)) {
    cond_data <- filter(data, chg_env == env_chg & data$DISABLE_REACTION_SENSORS == disabled_sensors)
    environment_label <- env_label_fun(env_chg)
    sensors_label <- sensors_label_fun(disabled_sensors == "0")

    observed_plasticity <- observed_plasticity %>% add_row(
      environment=environment_label,
      sensors=sensors_label,
      plastic=nrow(filter(cond_data, is_plastic==TRUE)),
      nonplastic=nrow(filter(cond_data, is_plastic==FALSE)),
      plastic_adaptive=nrow(filter(cond_data, dom_adaptive_plasticity=="True")),
      plastic_optimal=nrow(filter(cond_data, dom_optimal_plastic=="True")),
      plastic_nonadaptive=nrow(filter(cond_data, is_plastic==TRUE & dom_adaptive_plasticity=="False"))
    )
  }
}

```

```

    )
  }
}

observed_plasticity <- pivot_longer(
  observed_plasticity,
  cols=c("plastic", "plastic_adaptive", "plastic_optimal", "plastic_nonadaptive", "nonplastic"),
  names_to="phenotype",
  values_to="phenotype_cnt"
)

##### misc #####
# Configure our default graphing theme
theme_set(theme_cowplot())

```

2.4 Evolution of phenotypic plasticity

For each experimental condition, do we observe the evolution of phenotypic plasticity? To test for phenotypic plasticity, we culture digital organisms in both environments from the fluctuating condition (including organisms evolved in a constant environment). Any plasticity that we observe from digital organisms evolved under constant conditions is cryptic variation (as these organisms were never exposed to these culturing environments).

```

ggplot(filter(observed_plasticity, phenotype %in% c("plastic", "nonplastic")), aes(x=phenotype, y=phenotype_cnt)) +
  geom_bar(
    stat="identity",
    position=position_dodge(0.9)
  ) +
  geom_text(
    stat="identity",
    mapping=aes(label=phenotype_cnt),
    vjust=0.05
  ) +
  scale_fill_brewer(palette="Accent") +
  scale_x_discrete(
    name="Phenotype",
    limits=c("plastic", "nonplastic"),
    labels=c("Plastic", "Non-plastic")
  ) +
  facet_grid(sensors~environment) +
  theme(
    legend.position="none"
  )

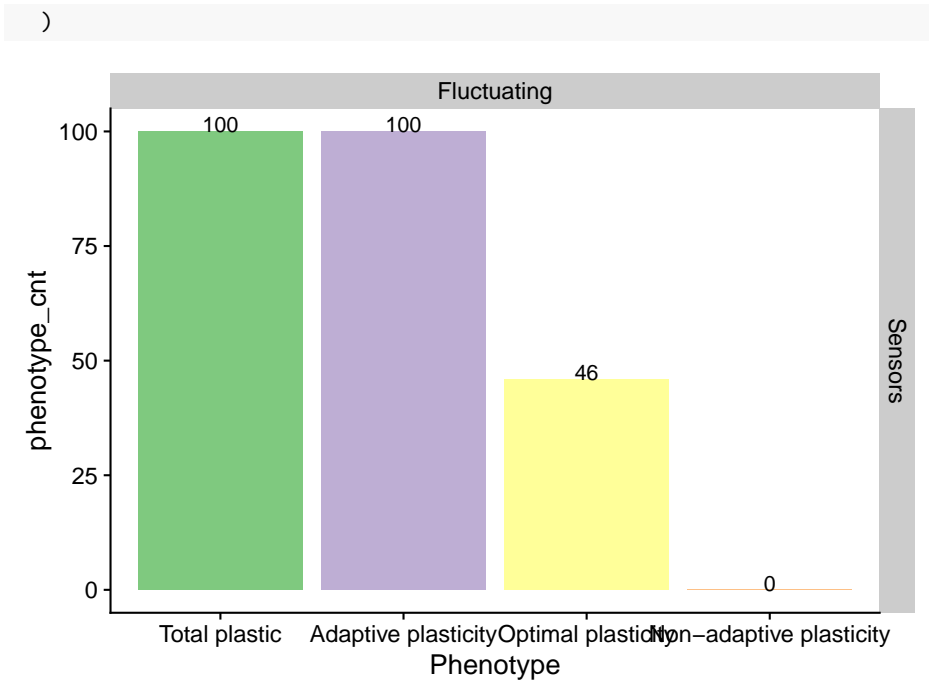
```



Indeed, we do not observe the evolution of phenotypic plasticity in any replicates in which digital organisms do not have access to sensory instructions. We do observe the evolution of plasticity (not necessarily adaptive plasticity) in both constant and fluctuating environments where sensors are enabled.

To what extent is the observed phenotypic plasticity adaptive?

```
ggplot(filter(observed_plasticity, environment=="Fluctuating" & sensors == "Sensors" & phenotype
  geom_bar(
    stat="identity",
    position=position_dodge(0.9)
  ) +
  geom_text(
    stat="identity",
    mapping=aes(label=phenotype_cnt),
    vjust=0.05
  ) +
  scale_fill_brewer(palette="Accent") +
  scale_x_discrete(
    name="Phenotype",
    limits=c("plastic", "plastic_adaptive", "plastic_optimal", "plastic_nonadaptive"),
    labels=c("Total plastic", "Adaptive plasticity", "Optimal plasticity", "Non-adaptive plasticity")
  ) +
  facet_grid(sensors~environment) +
  theme(
    legend.position="none"
```



Chapter 3

Evolutionary change

The effect of adaptive phenotypic plasticity on evolutionary change.

3.1 Overview

```
total_updates <- 200000
replicates <- 100
alpha <- 0.05

all_traits <- c("not","nand","and","ornot","or","andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-02-08-evo-dynamics/analysis/" # << For bookdown
# working_directory <- "./" # << For local analysis
```

3.2 Analysis dependencies

Load all required R libraries.

```
library(ggplot2)
library(rstatix)
library(ggsignif)
library(scales)
library(tidyverse)
library(cowplot)
library(RColorBrewer)
library(Hmisc)
```

```
library(boot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd9
```

These analyses were conducted/knitted with the following computing environment:

```
print(version)

##
## platform      _
## arch          x86_64-pc-linux-gnu
## os            linux-gnu
## system        x86_64, linux-gnu
## status
## major         4
## minor         0.5
## year          2021
## month         03
## day           31
## svn rev       80133
## language      R
## version.string R version 4.0.5 (2021-03-31)
## nickname      Shake and Throw
```

3.3 Setup

```
summary_data_loc <- paste0(working_directory, "data/aggregate.csv")
summary_data <- read.csv(summary_data_loc, na.strings="NONE")

summary_data$DISABLE_REACTION_SENSORS <- as.factor(summary_data$DISABLE_REACTION_SENSORS)
summary_data$chg_env <- summary_data$chg_env == "True"
summary_data$dominant_plastic_odd_even <- as.factor(summary_data$dominant_plastic_odd_even)
summary_data$sensors <- summary_data$DISABLE_REACTION_SENSORS == "0"
summary_data$is_plastic <- summary_data$dominant_plastic_odd_even == "True"

env_label_fun <- function(chg_env) {
  if (chg_env) {
    return("Fluctuating")
  } else {
    return("Constant")
  }
}

sensors_label_fun <- function(has_sensors) {
  if (has_sensors) {
```



```

    return("Sensors")
  } else {
    return("No sensors")
  }
}

# note that this labeler makes assumptions about how we set up our experiment
condition_label_fun <- function(has_sensors, env_chg) {
  if (has_sensors && env_chg) {
    return("PLASTIC")
  } else if (env_chg) {
    return("NON-PLASTIC")
  } else {
    return("STATIC")
  }
}

summary_data$env_label <- mapply(
  env_label_fun,
  summary_data$chg_env
)
summary_data$sensors_label <- mapply(
  sensors_label_fun,
  summary_data$sensors
)
summary_data$condition <- mapply(
  condition_label_fun,
  summary_data$sensors,
  summary_data$chg_env
)

condition_order = c(
  "STATIC",
  "NON-PLASTIC",
  "PLASTIC"
)

pairwise_comparisons <- list(
  c("STATIC", "NON-PLASTIC"),
  c("STATIC", "PLASTIC"),
  c("PLASTIC", "NON-PLASTIC")
)

p_label <- function(p_value) {
  threshold = 0.0001
  if (p_value < threshold) {

```

```

    return(paste0("p < ", threshold))
  } else {
    return(paste0("p = ", p_value))
  }
}

##### misc #####
# Configure our default graphing theme
theme_set(theme_cowplot())
# Palette
cb_palette <- "Paired"
# Create a directory to store plots
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)
# Define sample mean function
samplemean <- function(x, d) {
  return(mean(x[d]))
}

```

3.4 The evolution of phenotypic plasticity

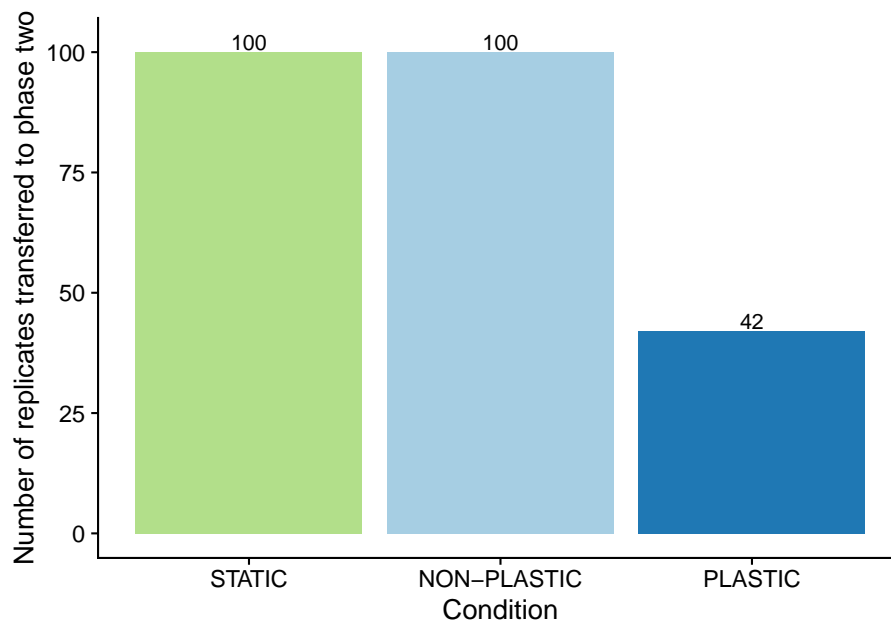
For sensor-enabled populations in fluctuating environments, we only transferred populations containing an optimally plastic genotype to phase-two.

```

summary_data_grouped = dplyr::group_by(summary_data, condition)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())

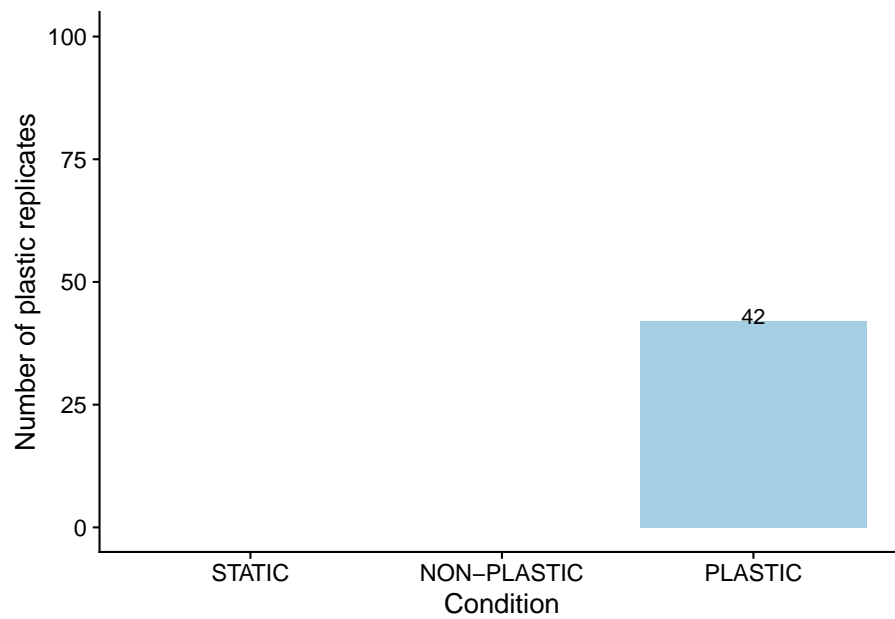
ggplot(summary_data_group_counts, aes(x=condition, y=n, fill=condition)) +
  geom_col(position=position_dodge(0.9)) +
  geom_text(aes(label=n, y=n+2)) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  ylab("Number of replicates transferred to phase two") +
  theme(
    legend.position="none"
  )

```



We can confirm our expectation that the dominant genotypes in non-plastic conditions are not phenotypically plastic.

```
summary_data_grouped = dplyr::group_by(summary_data, condition, is_plastic)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
ggplot(filter(summary_data_group_counts, is_plastic), aes(x=condition, y=n, fill=condition)) +
  geom_col(
    position=position_dodge(0.9)
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  geom_text(aes(label=n, y=n+1)) +
  ylab("Number of plastic replicates") +
  ylim(0, 100) +
  theme(
    legend.position="none"
  )
)
```



3.5 Average generation

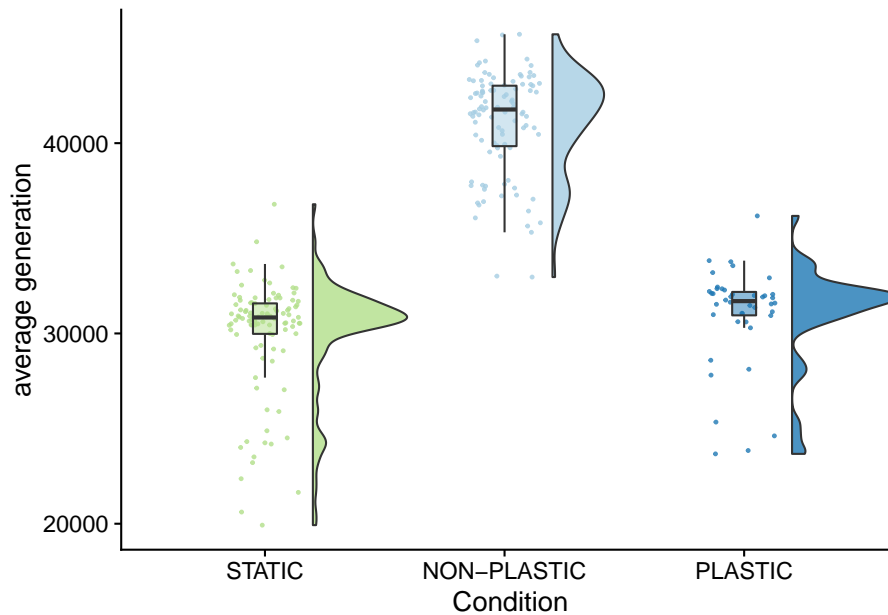
```
ggplot(summary_data, aes(x=condition, y=time_average_generation, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
```

```

    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  # coord_flip() +
  ylab("average generation") +
  theme(
    legend.position="none"
  ) +
  ggsave(paste0(working_directory, "plots/", "average-generation.png"))

```

```
## Saving 6.5 x 4.5 in image
```



```

kruskal.test(
  formula=time_average_generation~condition,
  data=summary_data
)

```

```

##
## Kruskal-Wallis rank sum test
##
## data: time_average_generation by condition
## Kruskal-Wallis chi-squared = 177.33, df = 2, p-value < 2.2e-16

```

```

pairwise.wilcox.test(
  x=summary_data$time_average_generation,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$time_average_generation and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16    0.004
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$time_average_generation)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$time_average_generation)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$time_average_generation)
  )
)

## [1] "PLASTIC median: 31697.65; STATIC median: 30839.75; NON-PLASTIC median: 41768.65"
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=time_average_generation~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
}

```

```

)
print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=9982"
## [1] "STATIC<-->PLASTIC: W=2818"
## [1] "PLASTIC<-->NON-PLASTIC: W=4186"

summary_data %>%
  group_by(condition) %>%
  summarise(mean=mean(time_average_generation), sd=sd(time_average_generation))

## # A tibble: 3 x 3
##   condition    mean    sd
##   <chr>      <dbl> <dbl>
## 1 NON-PLASTIC 41090. 2702.
## 2 PLASTIC    31016. 2615.
## 3 STATIC     30002. 3011.

```

3.6 Coalescence event count

The number of times the most recent common ancestor changes gives us the number of selective sweeps that occur during the experiment.

```

# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(phylo_mrca_changes ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition", step.increase=1)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- log10(stat.test$y.position) * c(1.0, 1.0, 1.03)
stat.test$label <- mapapply(p_label, stat.test$p.adj)

coalescence_events_fig <- ggplot(
  summary_data,
  aes(x=condition, y=phylo_mrca_changes, fill=condition)
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,

```

```

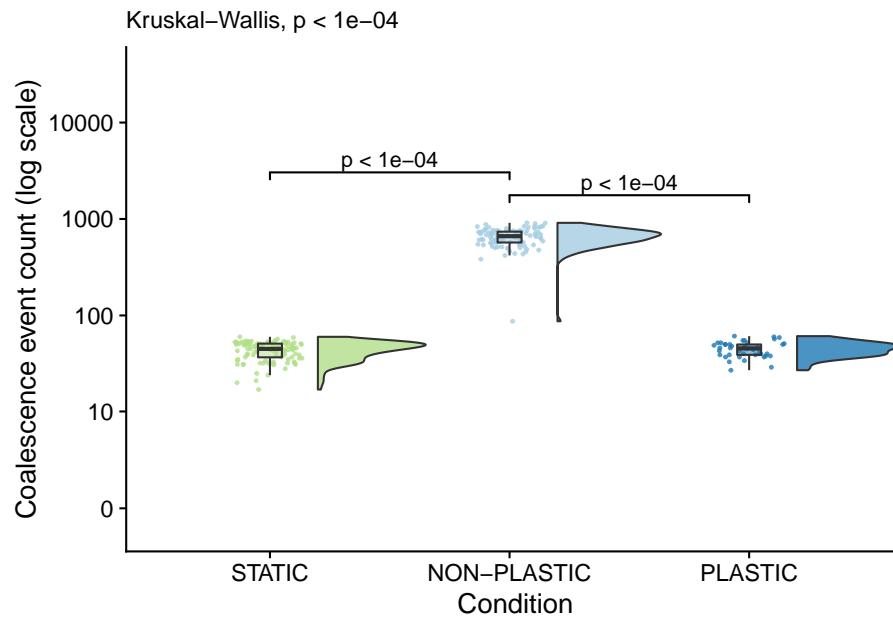
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order,
    labels=condition_order,
    breaks=condition_order
  ) +
  scale_y_continuous(
    name="Coalescence event count (log scale)",
    trans=pseudo_log_trans(sigma = 1, base = 10),
    breaks=c(0, 10, 100, 1000, 10000),
    limits=c(-1, 35000)
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  labs(
    subtitle=paste0(
      "Kruskal-Wallis, ",
      p_label(signif(kruskal.test(formula=phylo_mrca_changes~condition, data=summary_d
    )
  ) +
  ggsignif::geom_signif(
    data=filter(stat.test, p.adj <= alpha),
    aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
    manual=TRUE,
    inherit.aes=FALSE
  ) +
  # coord_flip() +
  theme(
    legend.position="none"
  ) +
  ggsave(
    paste0(working_directory, "plots/", "selective-sweeps.pdf"),
    width=5,
    height=5
  )

```



```
)

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
coalescence_events_fig
```



```
kruskal.test(
  formula=phylo_mrca_changes~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: phylo_mrca_changes by condition
## Kruskal-Wallis chi-squared = 175.46, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$phylo_mrca_changes,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$phylo_mrca_changes and summary_data$condition
```

```
##
##          NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      1
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$phylo_mrca_changes)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$phylo_mrca_changes)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$phylo_mrca_changes)
  )
)

## [1] "PLASTIC median: 45.5; STATIC median: 45; NON-PLASTIC median: 663.5"
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=phylo_mrca_changes~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=10000"
## [1] "STATIC<-->PLASTIC: W=2215"
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"
```

3.6.1 Average number of generations between coalescence events

```

# Compute frequency of coalescence events
summary_data$generations_per_mrca_change <- summary_data$time_average_generation / summary_data$P

# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(generations_per_mrca_change ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition")
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- stat.test$y.position
stat.test$label <- mapply(p_label,stat.test$p.adj)

coalescence_events_freq_fig <- ggplot(
  summary_data,
  aes(x=condition, y=generations_per_mrca_change, fill=condition)
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order,
    labels=condition_order
  ) +
  scale_y_continuous(
    name="Avg. generations between coalescence events",
    limits=c(0, 2000),
    breaks=seq(0, 2000, 500)
  ) +

```

```

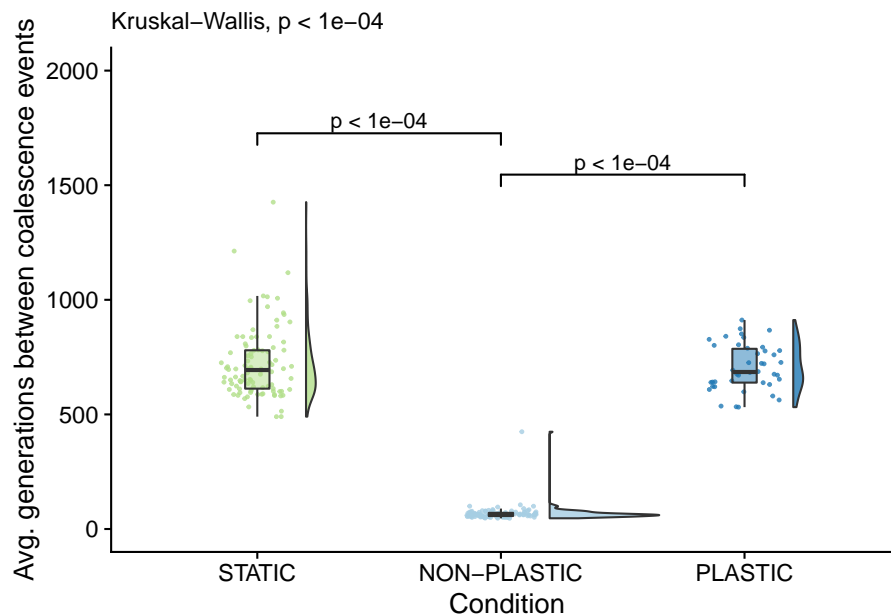
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
# coord_flip() +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=generations_per_mrca_change~condition, data=
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/", "generations-between-selective-sweeps.png"),
  width=5,
  height=5
)

```

```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
coalescence_events_freq_fig

```



```
kruskal.test(
  formula=generations_per_mrca_change~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: generations_per_mrca_change by condition
## Kruskal-Wallis chi-squared = 175.33, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$generations_per_mrca_change,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$generations_per_mrca_change and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      1
##
```

```
## P value adjustment method: bonferroni
```

```
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$generations_per_mrca_change)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$generations_per_mrca_change)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$generations_per_mrca_change)
  )
)
```

```
## [1] "PLASTIC median: 685.001780758557; STATIC median: 693.676265008576; NON-PLASTIC
print("Wilcox rank sum test statistics:")
```

```
## [1] "Wilcox rank sum test statistics:"
```

```
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=generations_per_mrca_change~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}
```

```
## [1] "STATIC<-->NON-PLASTIC: W=0"
```

```
## [1] "STATIC<-->PLASTIC: W=2151"
```

```
## [1] "PLASTIC<-->NON-PLASTIC: W=0"
```

3.7 Phenotypic volatility along the dominant lineage

```
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_trait_volatility ~ condition) %>%
```

3.7. PHENOTYPIC VOLATILITY ALONG THE DOMINANT LINEAGE31

```
adjust_pvalue(method = "bonferroni") %>%
add_significance() %>%
add_xy_position(x="condition", step.increase=1)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- log10(stat.test$y.position) * c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)

phenotypic_volatility_fig <- ggplot(
  summary_data,
  aes(x=condition, y=dominant_lineage_trait_volatility, fill=condition)
) +
geom_flat_violin(
  position = position_nudge(x = .2, y = 0),
  alpha = .8
) +
geom_point(
  mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order,
  labels=condition_order
) +
scale_y_continuous(
  name="Phenotypic volatility (log scale)",
  trans=pseudo_log_trans(sigma = 1, base = 10),
  breaks=c(0, 10, 100, 1000, 10000),
  limits=c(-1, 21000)
) +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
labs(
  subtitle=paste0(
```

```

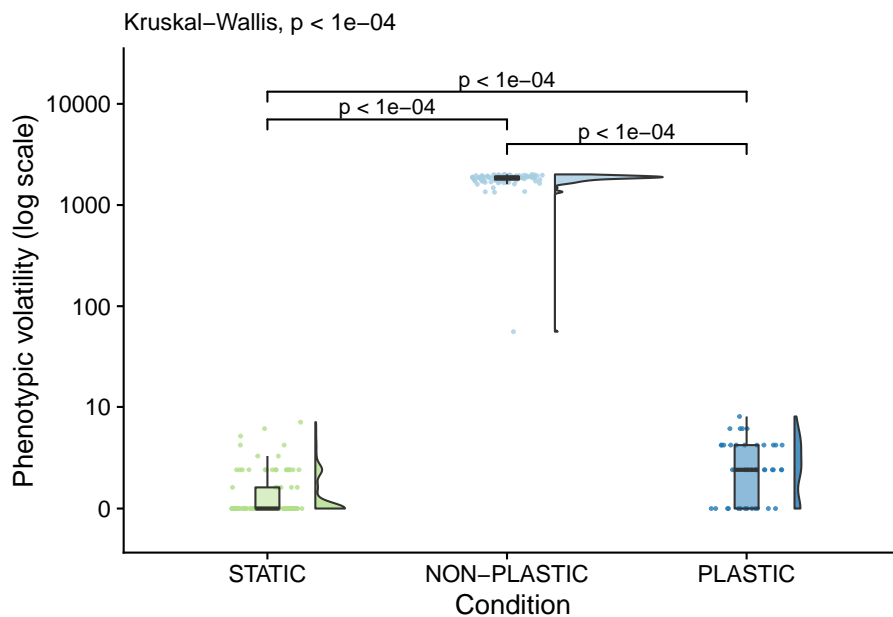
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_trait_volatility~condition,
    )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj<=alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
# coord_flip() +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/", "phenotypic-volatility.pdf"),
  width=5,
  height=5
)

```

```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
phenotypic_volatility_fig

```



```

kruskal.test(
  formula=dominant_lineage_trait_volatility~condition,

```


3.7. PHENOTYPIC VOLATILITY ALONG THE DOMINANT LINEAGE33

```
data=summary_data
)

##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_trait_volatility by condition
## Kruskal-Wallis chi-squared = 190.78, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_trait_volatility,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_trait_volatility and summary_data$condition
##
##          NON-PLASTIC PLASTIC
## PLASTIC < 2e-16      -
## STATIC  < 2e-16    8.7e-07
##
## P value adjustment method: bonferroni
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_trait_volatility)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_lineage_trait_volatility)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_trait_volatility)
  )
)

## [1] "PLASTIC median: 2; STATIC median: 0; NON-PLASTIC median: 1868"
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"
```

```

for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=dominant_lineage_trait_volatility~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

```

```

## [1] "STATIC<-->NON-PLASTIC: W=10000"
## [1] "STATIC<-->PLASTIC: W=3116.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"

```

3.7.1 Phenotypic volatility normalized by generations elapsed

```

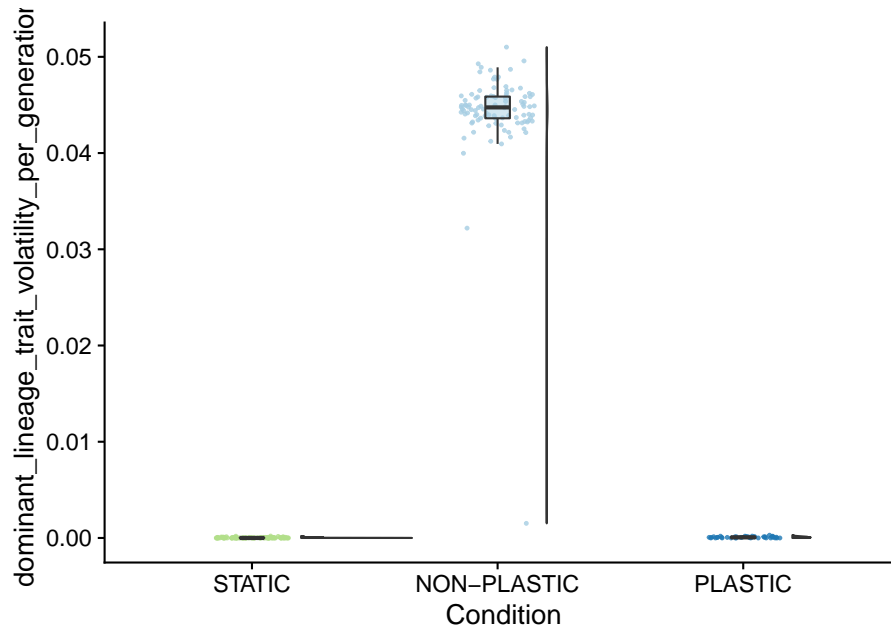
summary_data$dominant_lineage_trait_volatility_per_generation <- summary_data$dominant_lineage_trait_volatility / summary_data$generations_elapsed

ggplot(summary_data, aes(x=condition, y=dominant_lineage_trait_volatility_per_generation)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(

```

3.7. PHENOTYPIC VOLATILITY ALONG THE DOMINANT LINEAGE³⁵

```
palette=cb_palette
) +
# coord_flip() +
theme(
  legend.position="none"
)
```



```
kruskal.test(
  formula=dominant_lineage_trait_volatility_per_generation~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_trait_volatility_per_generation by condition
## Kruskal-Wallis chi-squared = 189.62, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_trait_volatility_per_generation,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
```

```
##
## data: summary_data$dominant_lineage_trait_volatility_per_generation and summary_da
##
##          NON-PLASTIC PLASTIC
## PLASTIC < 2e-16      -
## STATIC  < 2e-16      4.2e-06
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_trait_volatility
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_lineage_trait_volatility
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_trait_volatility
  )
)

## [1] "PLASTIC median: 6.33339279717772e-05; STATIC median: 0; NON-PLASTIC median: 0.
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=dominant_lineage_trait_volatility_per_generation~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=10000"
## [1] "STATIC<-->PLASTIC: W=3061.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"
```

3.8 Phenotypic fidelity

Frequency that an offspring's genotype is identical to a parent genotype (along the dominant lineage).

```
summary_data$dominant_lineage_trait_fidelity <- (summary_data$dominant_generation_born - summary_data$dominant_generation_born)

# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_trait_fidelity ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition", step.increase=1.5)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in ggplot2)
stat.test$manual_position <- stat.test$y.position * c(1.0, 1.0, 1.0005)
stat.test$label <- mapply(p_label, stat.test$p.adj)

phenotypic_fidelity_fig <- ggplot(
  summary_data,
  aes(x=condition, y=dominant_lineage_trait_fidelity, fill=condition)
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order,
    labels=condition_order
  ) +
  scale_y_continuous(
    name="Phenotypic fidelity",
    limits=c(0.94, 1.013),
    breaks=c(0.94, 0.96, 0.98, 1.0) #seq(0.94, 1.0, 0.01)
  ) +
```

```

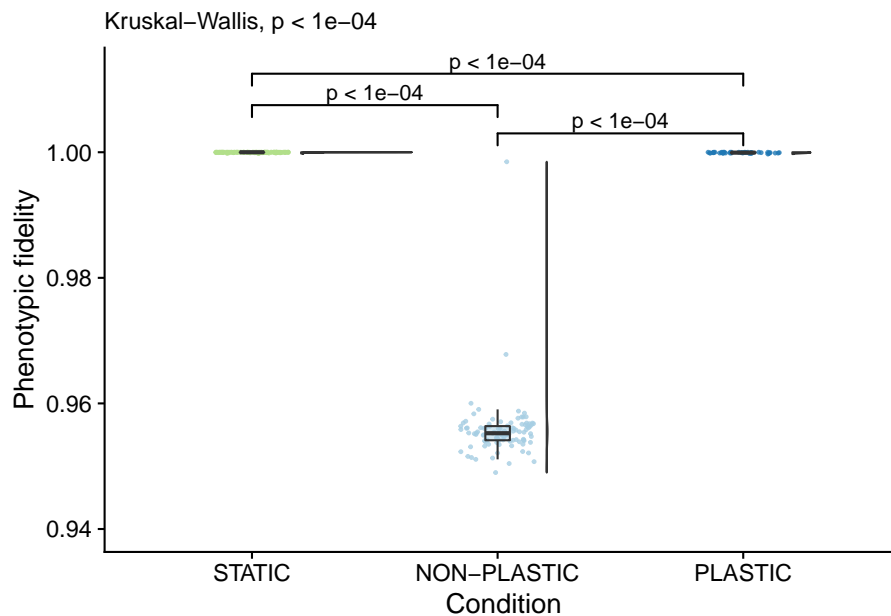
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
# coord_flip() +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_trait_fidelity~condition, d
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/", "phenotypic-fidelity.pdf"),
  width=5,
  height=5
)

```

```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
phenotypic_fidelity_fig

```



```
kruskal.test(
  formula=dominant_lineage_trait_fidelity~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_trait_fidelity by condition
## Kruskal-Wallis chi-squared = 189.62, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_trait_fidelity,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_trait_fidelity and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC < 2e-16      -
## STATIC  < 2e-16      4.2e-06
##
```

```
## P value adjustment method: bonferroni
```

```
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_trait_fidelity)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_lineage_trait_fidelity)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_trait_fidel.
  )
)
```

```
## [1] "PLASTIC median: 0.999936666072028; STATIC median: 1; NON-PLASTIC median: 0.955"
```

```
print("Wilcox rank sum test statistics:")
```

```
## [1] "Wilcox rank sum test statistics:"
```

```
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=dominant_lineage_trait_fidelity~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}
```

```
## [1] "STATIC<-->NON-PLASTIC: W=0"
```

```
## [1] "STATIC<-->PLASTIC: W=1138.5"
```

```
## [1] "PLASTIC<-->NON-PLASTIC: W=0"
```

3.9 Mutation count

```
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_total_mut_cnt ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
```



```

  add_xy_position(x="condition",step.increase=1)
  # Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- log10(stat.test$y.position) * c(1.0,1.0,1.03) # c(1.0,1.0,1.01)
stat.test$label <- mapply(p_label,stat.test$p.adj)

mutation_count_fig <- ggplot(
  summary_data,
  aes(x=condition, y=dominant_lineage_total_mut_cnt, fill=condition)
) +
geom_flat_violin(
  position = position_nudge(x = .2, y = 0),
  alpha = .8
) +
geom_point(
  mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order,
  labels=condition_order
) +
scale_y_continuous(
  name="Mutation count (log scale)",
  trans=pseudo_log_trans(sigma = 1, base = 10),
  breaks=c(0, 10, 100, 1000, 10000),
  limits=c(-1, 35000)
) +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
# coord_flip() +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",

```

```

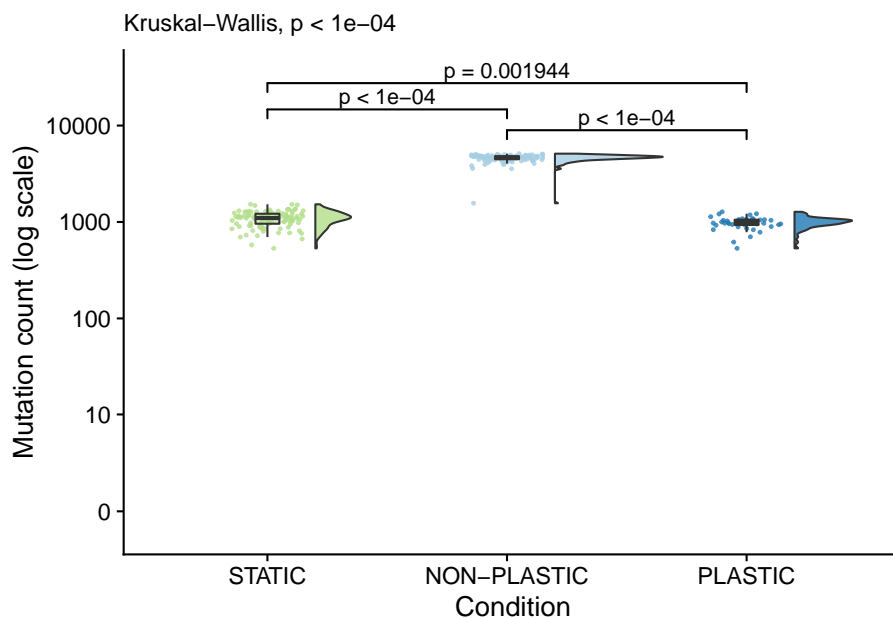
    p_label(signif(kruskal.test(formula=dominant_lineage_total_mut_cnt~condition, data=summary_data))
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/", "mutation-accumulation.pdf"),
  width=5,
  height=4
)

```

```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
mutation_count_fig

```



```

kruskal.test(
  formula=dominant_lineage_total_mut_cnt~condition,
  data=summary_data
)

```

```
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_total_mut_cnt by condition
## Kruskal-Wallis chi-squared = 179.33, df = 2, p-value < 2.2e-16

pairwise.wilcox.test(
  x=summary_data$dominant_lineage_total_mut_cnt,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_total_mut_cnt and summary_data$condition
##
##          NON-PLASTIC PLASTIC
## PLASTIC <2e-16          -
## STATIC  <2e-16          0.0019
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_total_mut_cnt
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_lineage_total_mut_cnt
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_total_mut_cnt
  )
)

## [1] "PLASTIC median: 998.5; STATIC median: 1100; NON-PLASTIC median: 4657.5"
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
```

```

wt <- wilcox.test(
  formula=dominant_lineage_total_mut_cnt~condition,
  data=pair_data,
  exact=FALSE,
  paired=FALSE
)
print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

```

```

## [1] "STATIC<-->NON-PLASTIC: W=10000"
## [1] "STATIC<-->PLASTIC: W=1336.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"

```

3.9.1 Mutation count normalized by generations elapsed

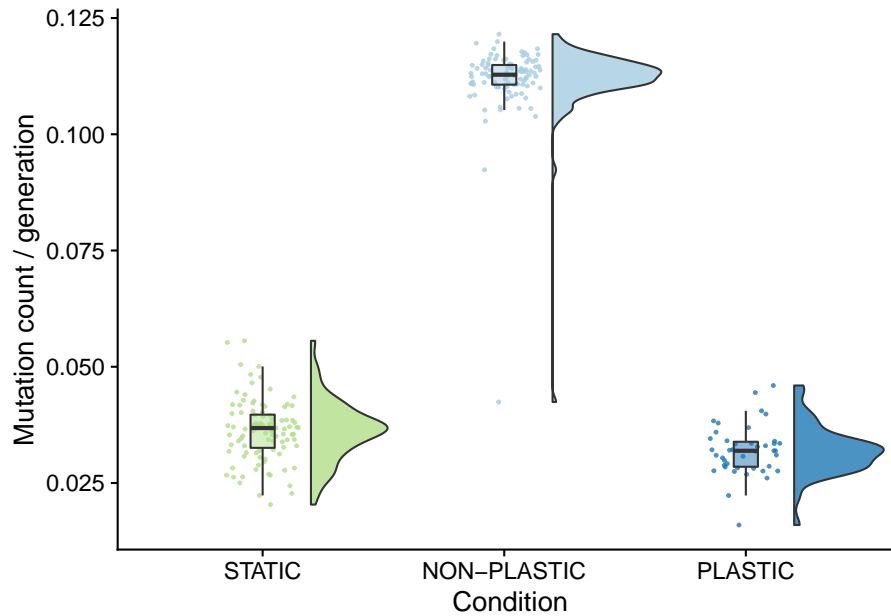
```

summary_data$mutations_per_generation <- summary_data$dominant_lineage_total_mut_cnt /

ggplot(summary_data, aes(x=condition, y=mutations_per_generation, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  ylab("Mutation count / generation") +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  # coord_flip() +

```

```
theme(
  legend.position="none"
)
```



```
kruskal.test(
  formula=mutations_per_generation~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: mutations_per_generation by condition
## Kruskal-Wallis chi-squared = 180.11, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$mutations_per_generation,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$mutations_per_generation and summary_data$condition
##
```

```
##          NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      2e-04
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$mutations_per_generation
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$mutations_per_generation
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$mutations_per_generation
  )
)

## [1] "PLASTIC median: 0.0319267181456982; STATIC median: 0.0368157192941933; NON-PLA
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"

for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=mutations_per_generation~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=9987"
## [1] "STATIC<-->PLASTIC: W=1206"
## [1] "PLASTIC<-->NON-PLASTIC: W=4198"
```

3.10 Genotypic fidelity

The frequency that an offspring's genotype is the same as a parent's genotype.

```

summary_data$dominant_lineage_genotypic_fidelity <- (summary_data$dominant_generation_born - summ

# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_genotypic_fidelity ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition",step.increase=0.2)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- stat.test$y.position * c(1.0,1.0,1.0)
stat.test$label <- mapply(p_label,stat.test$p.adj)

genotypic_fidelity_fig <- ggplot(
  summary_data,
  aes(x=condition, y=dominant_lineage_genotypic_fidelity, fill=condition)
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order,
    labels=condition_order
  ) +
  scale_y_continuous(
    name="Genotypic fidelity",
    limits=c(0.85, 1.01),
    breaks=c(0.85, 0.90, 0.95, 1.0) #seq(0.85, 1.0, 0.02)
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(

```

```

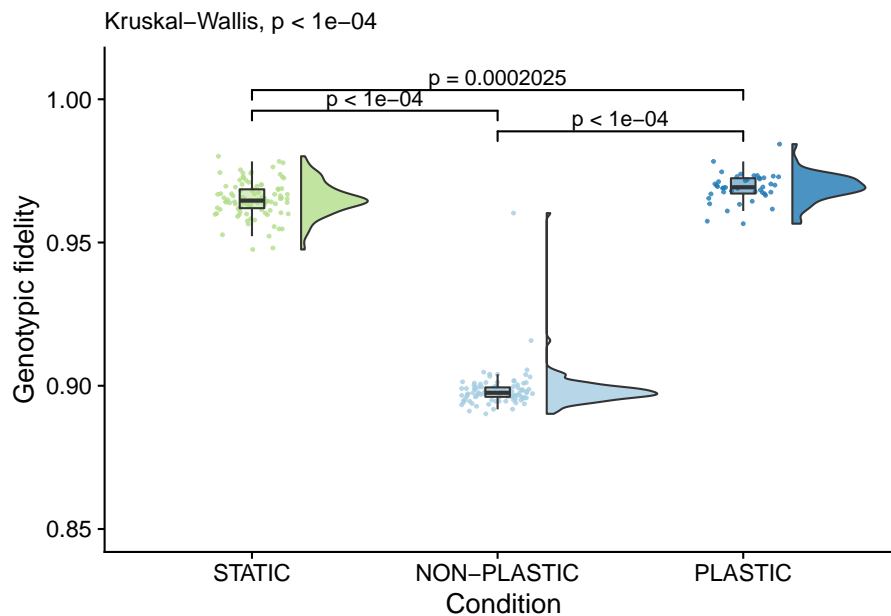
    palette=cb_palette
  ) +
  # coord_flip() +
  labs(
    subtitle=paste0(
      "Kruskal-Wallis, ",
      p_label(signif(kruskal.test(formula=dominant_lineage_genotypic_fidelity~condition
    )
  ) +
  ggsignif::geom_signif(
    data=filter(stat.test, p.adj <= alpha),
    aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
    manual=TRUE,
    inherit.aes=FALSE
  ) +
  theme(
    legend.position="none"
  ) +
  ggsave(
    paste0(working_directory, "plots/", "genotypic-fidelity.pdf"),
    width=5,
    height=4
  )

```

```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
genotypic_fidelity_fig

```

```
kruskal.test(
  formula=dominant_lineage_genotypic_fidelity~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_genotypic_fidelity by condition
## Kruskal-Wallis chi-squared = 179.86, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_genotypic_fidelity,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_genotypic_fidelity and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      2e-04
##
```

```
## P value adjustment method: bonferroni
```

```
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_genotypic_fidel
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_lineage_genotypic_fideli
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_genotypic_f
  )
)
```

```
## [1] "PLASTIC median: 0.969286906891951; STATIC median: 0.964620594632577; NON-PLASTIC median: 0.964620594632577"
```

```
print("Wilcox rank sum test statistics:")
```

```
## [1] "Wilcox rank sum test statistics:"
```

```
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=dominant_lineage_genotypic_fidelity~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}
```

```
## [1] "STATIC<-->NON-PLASTIC: W=18"
```

```
## [1] "STATIC<-->PLASTIC: W=2992"
```

```
## [1] "PLASTIC<-->NON-PLASTIC: W=2"
```

3.11 Characterizing variation along dominant lineages

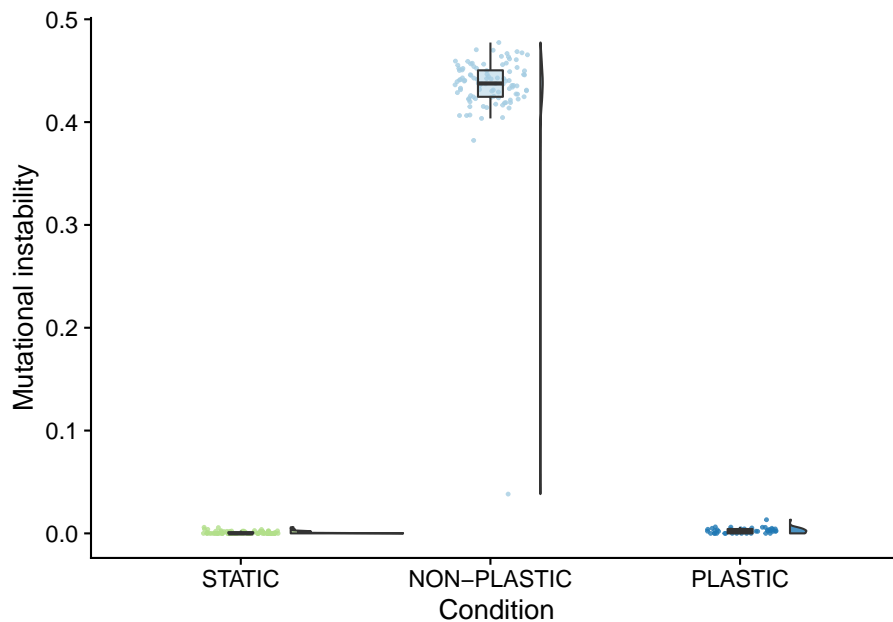
3.11.1 Mutational instability

```
summary_data$frac_phenotype_changing_mut_steps <- summary_data$dominant_lineage_num_mutations /
summary_data$frac_phenotype_stable_mut_steps <- 1 - summary_data$frac_phenotype_changing_mut_steps
```

3.11. CHARACTERIZING VARIATION ALONG DOMINANT LINEAGES⁵¹

```
ggplot(summary_data, aes(x=condition, y=frac_phenotype_changing_mut_steps, fill=condition)) +  
  geom_flat_violin(  
    position = position_nudge(x = .2, y = 0),  
    alpha = .8  
  ) +  
  geom_point(  
    mapping=aes(color=condition),  
    position = position_jitter(width = .15),  
    size = .5,  
    alpha = 0.8  
  ) +  
  geom_boxplot(  
    width = .1,  
    outlier.shape = NA,  
    alpha = 0.5  
  ) +  
  scale_x_discrete(  
    name="Condition",  
    limits=condition_order  
  ) +  
  ylab("Mutational instability") +  
  scale_fill_brewer(  
    palette=cb_palette  
  ) +  
  scale_color_brewer(  
    palette=cb_palette  
  ) +  
  # coord_flip() +  
  theme(  
    legend.position="none"  
  ) +  
  ggsave(paste0(working_directory, "plots/", "frac_phenotype_changing_mutational_steps.png"))
```

Saving 6.5 x 4.5 in image



```
kruskal.test(
  formula=frac_phenotype_changing_mut_steps~condition,
  data=summary_data
)
```

```
##
##  Kruskal-Wallis rank sum test
##
## data:  frac_phenotype_changing_mut_steps by condition
## Kruskal-Wallis chi-squared = 191.23, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$frac_phenotype_changing_mut_steps,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
```

```
##
##  Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  summary_data$frac_phenotype_changing_mut_steps and summary_data$condition
##
##           NON-PLASTIC PLASTIC
## PLASTIC < 2e-16      -
## STATIC  < 2e-16      2.3e-07
##
```

3.11. CHARACTERIZING VARIATION ALONG DOMINANT LINEAGES⁵³

```
## P value adjustment method: bonferroni
```

```
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$frac_phenotype_changing_mut_steps)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$frac_phenotype_changing_mut_steps)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$frac_phenotype_changing_mut_steps)
  )
)
```

```
## [1] "PLASTIC median: 0.00224941742616098; STATIC median: 0; NON-PLASTIC median: 0.437583018324"
```

```
print("Wilcox rank sum test statistics:")
```

```
## [1] "Wilcox rank sum test statistics:"
```

```
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=frac_phenotype_changing_mut_steps~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}
```

```
## [1] "STATIC<-->NON-PLASTIC: W=10000"
```

```
## [1] "STATIC<-->PLASTIC: W=3172"
```

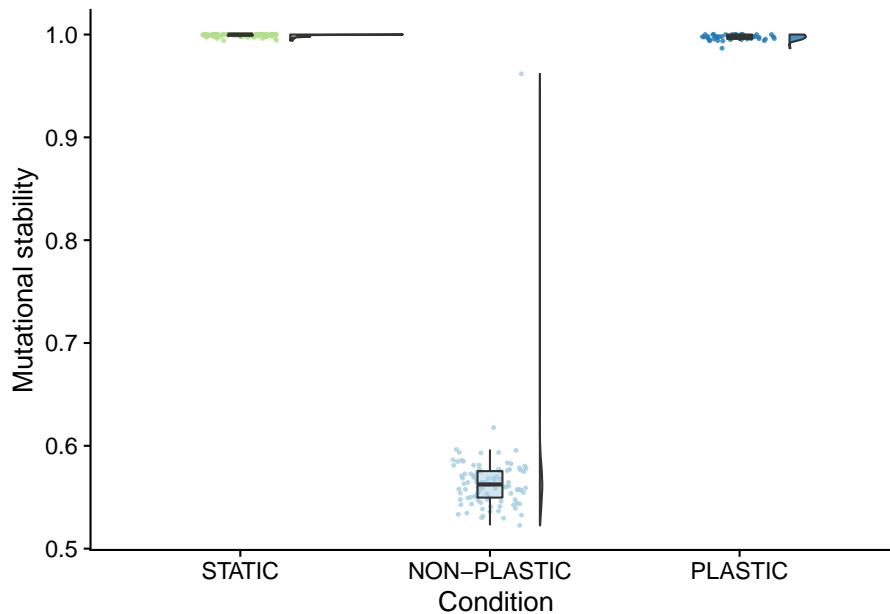
```
## [1] "PLASTIC<-->NON-PLASTIC: W=4200"
```

3.11.2 Mutational stability

```
ggplot(summary_data, aes(x=condition, y=frac_phenotype_stable_mut_steps, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
```

```
mapping=aes(color=condition),
position = position_jitter(width = .15),
size = .5,
alpha = 0.8
) +
geom_boxplot(
width = .1,
outlier.shape = NA,
alpha = 0.5
) +
scale_x_discrete(
name="Condition",
limits=condition_order
) +
ylab("Mutational stability") +
scale_fill_brewer(
palette=cb_palette
) +
scale_color_brewer(
palette=cb_palette
) +
# coord_flip() +
theme(
legend.position="none"
)
```

3.11. CHARACTERIZING VARIATION ALONG DOMINANT LINEAGES55



```
kruskal.test(
  formula=frac_phenotype_stable_mut_steps~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: frac_phenotype_stable_mut_steps by condition
## Kruskal-Wallis chi-squared = 191.23, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$frac_phenotype_stable_mut_steps,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$frac_phenotype_stable_mut_steps and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC < 2e-16      -
## STATIC  < 2e-16      2.3e-07
##
```

```
## P value adjustment method: bonferroni
```

```
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$frac_phenotype_stable_mut_steps)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$frac_phenotype_stable_mut_steps)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$frac_phenotype_stable_mut_steps)
  )
)
```

```
## [1] "PLASTIC median: 0.997750582573839; STATIC median: 1; NON-PLASTIC median: 0.562"
```

```
print("Wilcox rank sum test statistics:")
```

```
## [1] "Wilcox rank sum test statistics:"
```

```
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=frac_phenotype_stable_mut_steps~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}
```

```
## [1] "STATIC<-->NON-PLASTIC: W=0"
```

```
## [1] "STATIC<-->PLASTIC: W=1028"
```

```
## [1] "PLASTIC<-->NON-PLASTIC: W=0"
```

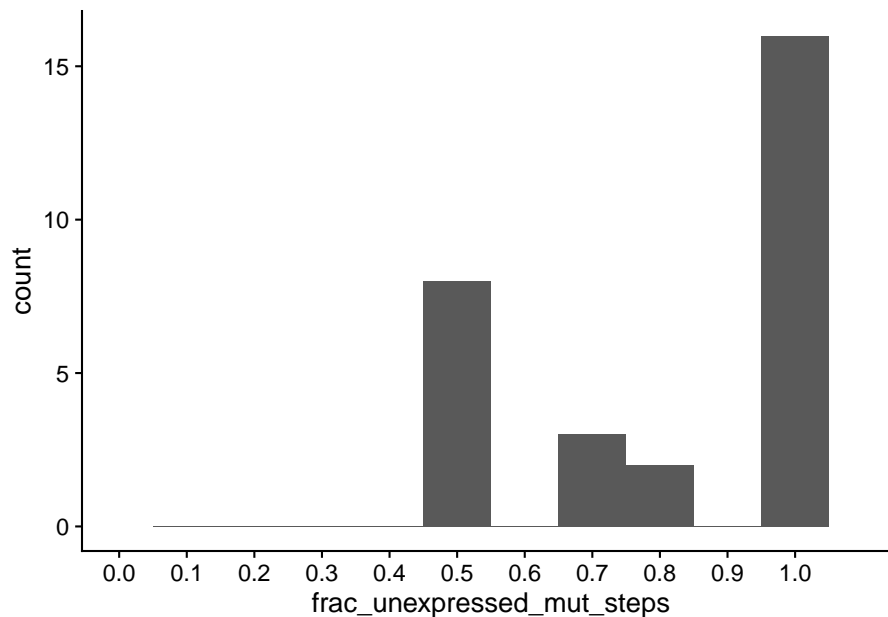
3.11.3 For PLASTIC populations, what fraction of phenotype-altering mutations occurred in the unexpressed phenotype?

```
summary_data$frac_unexpressed_mut_steps <- summary_data$dominant_lineage_num_mut_steps /
summary_data$frac_expressed_mut_steps <- summary_data$dominant_lineage_num_mut_steps /
```


3.11. CHARACTERIZING VARIATION ALONG DOMINANT LINEAGES⁵⁷

```
ggplot(filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_that_change_aggr
  geom_histogram(binwidth=0.1) +
  scale_x_continuous(
    limits=c(0, 1.1),
    breaks=seq(0, 1.0, 0.1)
  ) +
  theme(
    legend.position="none"
  )
)
```

```
## Warning: Removed 2 rows containing missing values (geom_bar).
```



```
print(paste0("PLASTIC - Mean with bootstrapped 95% CI"))

## [1] "PLASTIC - Mean with bootstrapped 95% CI"
bo <- boot(filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_that_change_aggr
print(bo)

##
## ORDINARY NONPARAMETRIC BOOTSTRAP
##
##
## Call:
## boot(data = filter(summary_data, condition == "PLASTIC" & dominant_lineage_num_mut_steps_that_change_aggr
##       0)$frac_unexpressed_mut_steps, statistic = samplemean, R = 10000)
```

```
##
##
## Bootstrap Statistics :
##      original      bias      std. error
## t1* 0.8247126 4.597701e-06 0.03989852
print(boot.ci(bo, conf=0.95, type="perc"))

## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
## Based on 10000 bootstrap replicates
##
## CALL :
## boot.ci(boot.out = bo, conf = 0.95, type = "perc")
##
## Intervals :
## Level      Percentile
## 95%      ( 0.7443,  0.8994 )
## Calculations and Intervals on Original Scale
plastic_summary_data <- filter(summary_data, condition=="PLASTIC")
aggregate_frac_mut_steps_that_change_unexpressed_phenotype <- sum(plastic_summary_data$dominant_lineage_num_mut_steps_that_change_unexpressed_phenotype)
sum(plastic_summary_data$dominant_lineage_num_mut_steps_that_change_unexpressed_phenotype)

## [1] 83
sum(plastic_summary_data$dominant_lineage_num_mut_steps_that_change_aggregate_phenotype)

## [1] 102
aggregate_frac_mut_steps_that_change_unexpressed_phenotype

## [1] 0.8137255
83 / 102 (0.8137255)
```

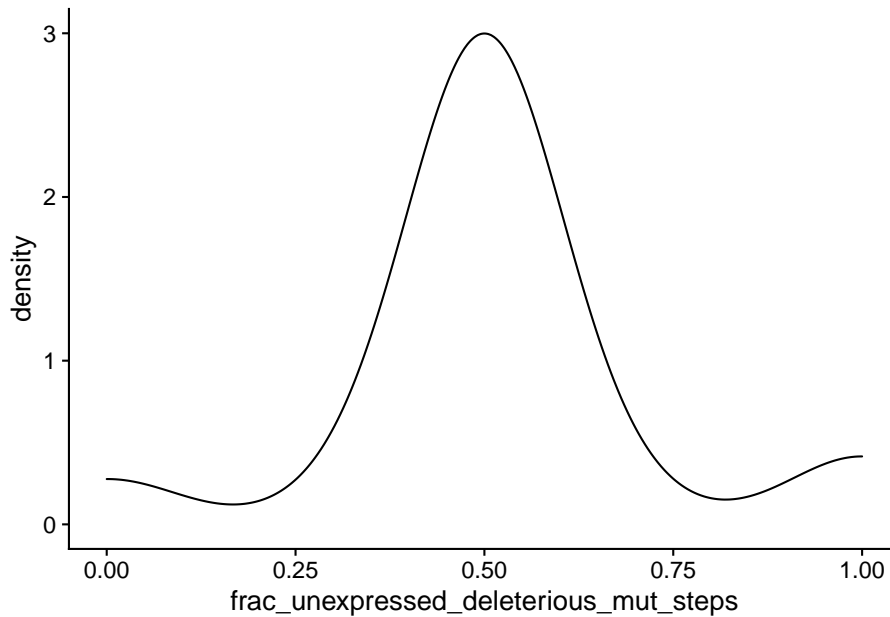
3.11.4 For PLASTIC populations, what fraction of mutations that affect the unexpressed phenotype are deleterious versus beneficial?

```
aggregate_frac_unexpressed_deleterious_mut_steps <- sum(plastic_summary_data$dominant_lineage_num_mut_steps_that_change_unexpressed_phenotype)
aggregate_frac_unexpressed_beneficial_mut_steps <- sum(plastic_summary_data$dominant_lineage_num_mut_steps_that_change_unexpressed_phenotype)
```

3.11.4.1 Deleterious mutations

```
summary_data$frac_unexpressed_deleterious_mut_steps <- summary_data$dominant_lineage_num_mut_steps_that_change_unexpressed_phenotype / aggregate_frac_unexpressed_deleterious_mut_steps
ggplot(
  filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_that_change_unexpressed_phenotype > 0)
  aes(x=frac_unexpressed_deleterious_mut_steps)
```

```
) +
geom_density() +
theme(
  legend.position="none"
)
```



```
bo <- boot(filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_that_change == 0)$frac_unexpressed_deleterious_mut_steps, statistic = samplemean,
print(bo)
```

```
##
## ORDINARY NONPARAMETRIC BOOTSTRAP
##
##
## Call:
## boot(data = filter(summary_data, condition == "PLASTIC" & dominant_lineage_num_mut_steps_that_change == 0)$frac_unexpressed_deleterious_mut_steps, statistic = samplemean,
##       R = 10000)
##
##
## Bootstrap Statistics :
##      original      bias    std. error
## t1* 0.5172414 7.16092e-05 0.03958966
print(boot.ci(bo, conf=0.95, type="perc"))

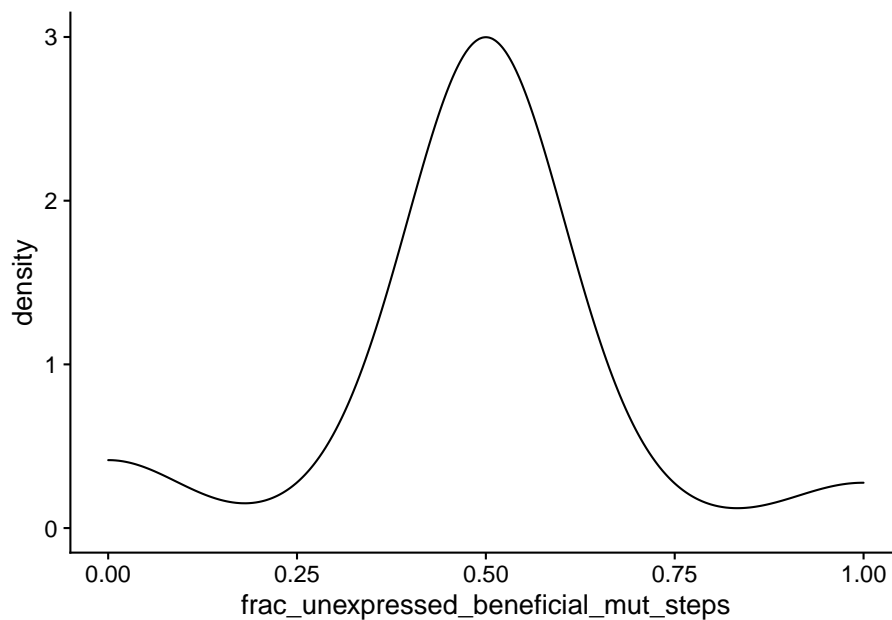
## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
```

```
## Based on 10000 bootstrap replicates
##
## CALL :
## boot.ci(boot.out = bo, conf = 0.95, type = "perc")
##
## Intervals :
## Level      Percentile
## 95%      ( 0.4414,  0.5954 )
## Calculations and Intervals on Original Scale
```

3.11.4.2 Beneficial mutations

```
summary_data$frac_unexpressed_beneficial_mut_steps <- summary_data$dominant_lineage_num_mut_steps_that_changed / summary_data$dominant_lineage_num_mut_steps

ggplot(
  filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_that_changed > 0)
  aes(x=frac_unexpressed_beneficial_mut_steps)
) +
  geom_density() +
  theme(
    legend.position="none"
  )
```



```
bo <- boot(filter(summary_data, condition=="PLASTIC" & dominant_lineage_num_mut_steps_that_changed > 0),
  print(bo)
```

```
##
## ORDINARY NONPARAMETRIC BOOTSTRAP
##
##
## Call:
## boot(data = filter(summary_data, condition == "PLASTIC" & dominant_lineage_num_mut_steps_that_
##       0)$frac_unexpressed_beneficial_mut_steps, statistic = samplemean,
##       R = 10000)
##
##
## Bootstrap Statistics :
##      original      bias      std. error
## t1* 0.4827586 7.45977e-05 0.03909157
print(boot.ci(bo, conf=0.95, type="perc"))

## BOOTSTRAP CONFIDENCE INTERVAL CALCULATIONS
## Based on 10000 bootstrap replicates
##
## CALL :
## boot.ci(boot.out = bo, conf = 0.95, type = "perc")
##
## Intervals :
## Level      Percentile
## 95%      ( 0.4057,  0.5575 )
## Calculations and Intervals on Original Scale
```

3.12 Manuscript figures

Figures styled for the paper.

```
grid <- plot_grid(
  coalescence_events_fig +
    theme(
      legend.position="none",
      # axis.ticks.x=element_blank(),
      # axis.text.x=element_blank(),
      axis.title.x=element_blank()
    ) +
    ggtitle("Coalescence events count"),
  mutation_count_fig +
    theme(
      legend.position="none",
      # axis.ticks.x=element_blank(),
      # axis.text.x=element_blank(),
      axis.title.x=element_blank()
    )
)
```

```

    ) +
    ggtitle("Mutation count"),
  phenotypic_volatility_fig +
  theme(
    legend.position="none",
    # axis.ticks.x=element_blank(),
    # axis.text.x=element_blank(),
    axis.title.x=element_blank()
  ) +
  ggtitle("Phenotypic volatility"),
  coalescence_events_freq_fig +
  theme(
    legend.position="none",
    axis.title.x=element_blank()
  ) +
  ggtitle("Generations between coalescence events"),
  genotypic_fidelity_fig +
  theme(
    legend.position="none",
    axis.title.x=element_blank()
  ) +
  ggtitle("Genotypic fidelity"),
  phenotypic_fidelity_fig +
  theme(
    legend.position="none",
    axis.title.x=element_blank()
  ) +
  ggtitle("Phenotypic fidelity"),
  nrow=2,
  ncol=3,
  align="v",
  labels="auto"
)
save_plot(
  paste0(working_directory, "plots/", "evolutionary-change-full-panel.pdf"),
  grid,
  # base_height=12,
  # base_asp=3/2
  base_height=12,
  base_asp=3/2
)

```

Chapter 4

Evolution and maintenance of novel traits

The effect of adaptive phenotypic plasticity on the evolution and maintenance of novel tasks.

4.1 Overview

```
total_updates <- 200000
replicates <- 100
alpha <- 0.05

focal_traits <- c("not", "nand", "and", "ornot", "or", "andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")
extra_traits <- c(
  "nor", "xor", "equals",
  "logic_3aa", "logic_3ab", "logic_3ac",
  "logic_3ad", "logic_3ae", "logic_3af",
  "logic_3ag", "logic_3ah", "logic_3ai",
  "logic_3aj", "logic_3ak", "logic_3al",
  "logic_3am", "logic_3an", "logic_3ao",
  "logic_3ap", "logic_3aq", "logic_3ar",
  "logic_3as", "logic_3at", "logic_3au",
  "logic_3av", "logic_3aw", "logic_3ax",
  "logic_3ay", "logic_3az", "logic_3ba",
  "logic_3bb", "logic_3bc", "logic_3bd",
  "logic_3be", "logic_3bf", "logic_3bg",
  "logic_3bh", "logic_3bi", "logic_3bj",
```

```

    "logic_3bk", "logic_3bl", "logic_3bm",
    "logic_3bn", "logic_3bo", "logic_3bp",
    "logic_3bq", "logic_3br", "logic_3bs",
    "logic_3bt", "logic_3bu", "logic_3bv",
    "logic_3bw", "logic_3bx", "logic_3by",
    "logic_3bz", "logic_3ca", "logic_3cb",
    "logic_3cc", "logic_3cd", "logic_3ce",
    "logic_3cf", "logic_3cg", "logic_3ch",
    "logic_3ci", "logic_3cj", "logic_3ck",
    "logic_3cl", "logic_3cm", "logic_3cn",
    "logic_3co", "logic_3cp"
  )

# Relative location of data.
working_directory <- "experiments/2021-01-31-complex-features/analysis/" # << For book
# working_directory <- "/"

```

4.2 Analysis dependencies

Load all required R libraries.

```

library(ggplot2)
library(rstatix)
library(ggsignif)
library(scales)
library(tidyverse)
library(cowplot)
library(RColorBrewer)
library(Hmisc)
library(boot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd9")

```

These analyses were conducted/knitted with the following computing environment:

```

print(version)

##
## platform      x86_64-pc-linux-gnu
## arch          x86_64
## os            linux-gnu
## system        x86_64, linux-gnu
## status
## major         4
## minor         0.5
## year          2021

```



```
## month      03
## day        31
## svn rev    80133
## language   R
## version.string R version 4.0.5 (2021-03-31)
## nickname   Shake and Throw
```

4.3 Setup

```
##### summary data #####
summary_data_loc <- paste0(working_directory, "data/aggregate.csv")
summary_data <- read.csv(summary_data_loc, na.strings="NONE")

summary_data$DISABLE_REACTION_SENSORS <- as.factor(summary_data$DISABLE_REACTION_SENSORS)
summary_data$chg_env <- summary_data$chg_env == "True"
summary_data$dominant_plastic_odd_even <- as.factor(summary_data$dominant_plastic_odd_even)
summary_data$sensors <- summary_data$DISABLE_REACTION_SENSORS == "0"
summary_data$is_plastic <- summary_data$dominant_plastic_odd_even == "True"
summary_data$extra_task_value <- as.factor(summary_data$extra_task_value)
summary_data <- filter(summary_data, extra_task_value == 0.1)

env_label_fun <- function(chg_env) {
  if (chg_env) {
    return("Fluctuating")
  } else {
    return("Constant")
  }
}

sensors_label_fun <- function(has_sensors) {
  if (has_sensors) {
    return("Sensors")
  } else {
    return("No sensors")
  }
}

condition_label_fun <- function(has_sensors, env_chg) {
  if (has_sensors && env_chg) {
    return("PLASTIC")
  } else if (env_chg) {
    return("NON-PLASTIC")
  } else {
    return("STATIC")
  }
}
```

```

    }
  }

summary_data$env_label <- mapply(
  env_label_fun,
  summary_data$chg_env
)
summary_data$sensors_label <- mapply(
  sensors_label_fun,
  summary_data$sensors
)
summary_data$condition <- mapply(
  condition_label_fun,
  summary_data$sensors,
  summary_data$chg_env
)

condition_order = c(
  "STATIC",
  "NON-PLASTIC",
  "PLASTIC"
)

pairwise_comparisons <- list(
  c("STATIC", "NON-PLASTIC"),
  c("STATIC", "PLASTIC"),
  c("PLASTIC", "NON-PLASTIC")
)

p_label <- function(p_value) {
  threshold = 0.0001
  if (p_value < threshold) {
    return(paste0("p < ", threshold))
  } else {
    return(paste0("p = ", p_value))
  }
}

##### time series #####
lineage_time_series_data_loc <- paste0(working_directory, "data/lineage_series.csv")
lineage_time_series_data <- read.csv(lineage_time_series_data_loc)

lineage_time_series_data$DISABLE_REACTION_SENSORS <- as.factor(lineage_time_series_data$DISABLE_REACTION_SENSORS)
lineage_time_series_data$chg_env <- lineage_time_series_data$chg_env == "True"
lineage_time_series_data$sensors <- lineage_time_series_data$DISABLE_REACTION_SENSORS
lineage_time_series_data$extra_task_value <- as.factor(lineage_time_series_data$extra_task_value)

```

```

lineage_time_series_data$env_label <- mapapply(
  env_label_fun,
  lineage_time_series_data$chg_env
)
lineage_time_series_data$sensors_label <- mapapply(
  sensors_label_fun,
  lineage_time_series_data$sensors
)
lineage_time_series_data$condition <- mapapply(
  condition_label_fun,
  lineage_time_series_data$sensors,
  lineage_time_series_data$chg_env
)

##### misc #####
# Configure our default graphing theme
theme_set(theme_cowplot())
# Palette
cb_palette <- "Paired"
# Create directory to dump plots
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)
# Sample mean function
samplemean <- function(x, d) {
  return(mean(x[d]))
}

```

4.4 The evolution of phenotypic plasticity

For sensor-enabled populations in fluctuating environments, we only transferred populations containing an optimally plastic genotype to phase two.

```

summary_data_grouped = dplyr::group_by(summary_data, sensors, env_label, condition, extra_task_val)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())

```

```

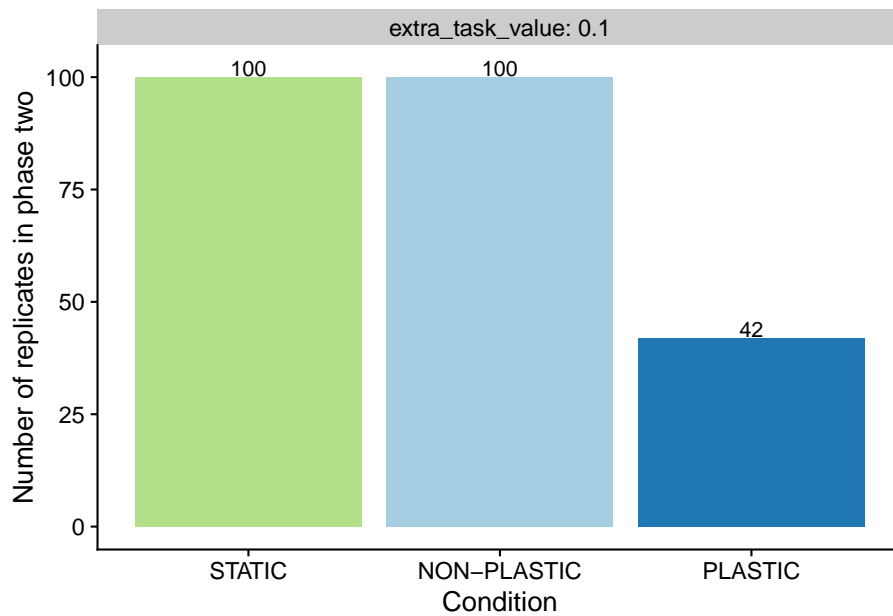
ggplot(summary_data_group_counts, aes(x=condition, y=n, fill=condition)) +
  geom_col(position=position_dodge(0.9)) +
  geom_text(aes(label=n, y=n+2)) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +

```

```

scale_color_brewer(
  palette=cb_palette
) +
ylab("Number of replicates in phase two") +
facet_wrap(~extra_task_value, labeller=label_both) +
theme(
  legend.position="none"
)

```



We can confirm our expectation that the dominant genotypes in non-plastic conditions are not phenotypically plastic.

```

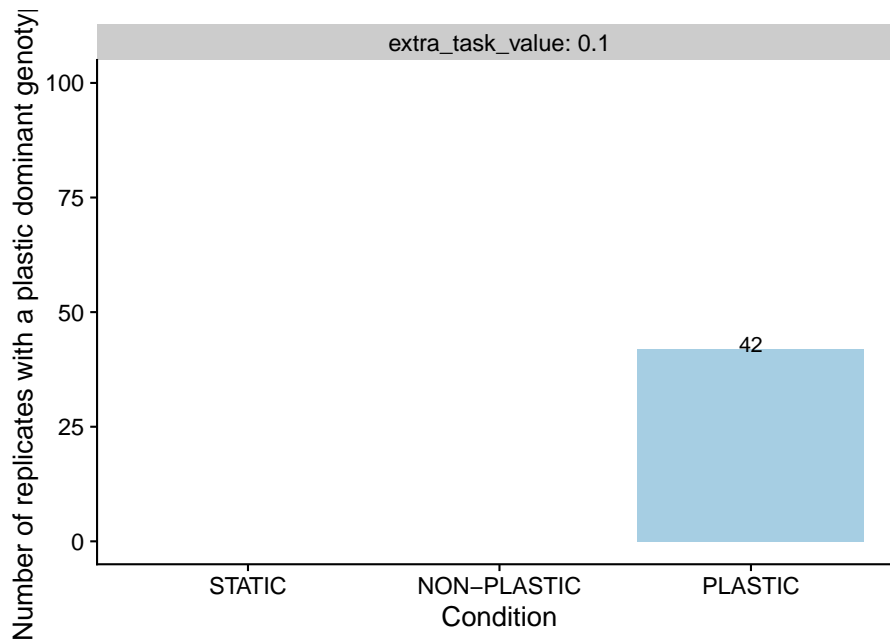
summary_data_grouped = dplyr::group_by(summary_data, condition, is_plastic, extra_task_value)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
ggplot(filter(summary_data_group_counts, is_plastic), aes(x=condition, y=n, fill=condition)) +
  geom_col(position=position_dodge(0.9)) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  )

```

```

) +
ylim(0, 100) +
geom_text(aes(label=n, y=n+1)) +
ylab("Number of replicates with a plastic dominant genotype") +
facet_wrap(~extra_task_value, labeller=label_both) +
theme(
  legend.position="none"
)

```



4.5 Final novel task count (dominant genotype)

How many novel tasks do final dominant genotypes perform?

```

# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(dominant_extra_tasks ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition") # ,step.increase=1
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- stat.test$y.position ## c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)

final_novel_task_count_fig <- ggplot(

```

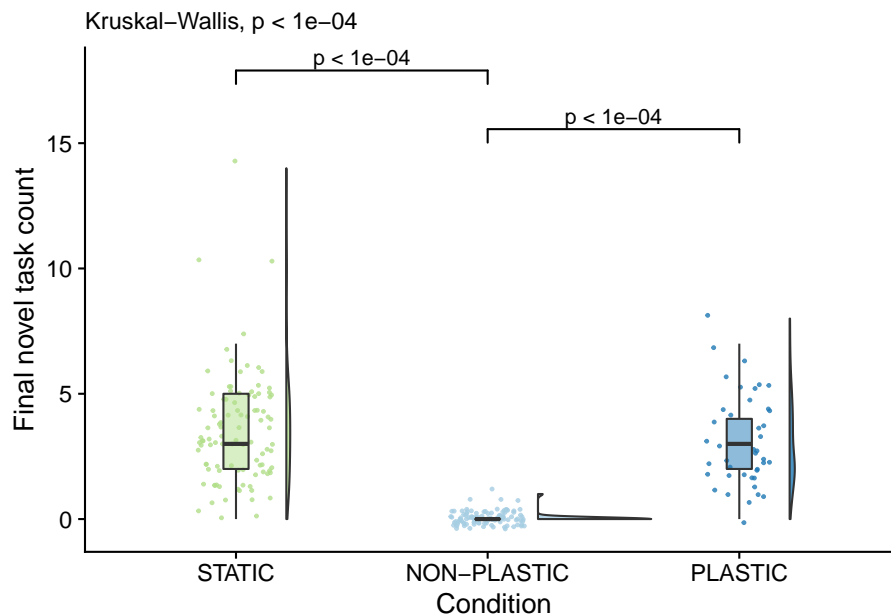
```

summary_data,
aes(x=condition, y=dominant_extra_tasks, fill=condition)
) +
geom_flat_violin(
  position = position_nudge(x = .2, y = 0),
  alpha = .8
) +
geom_point(
  mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order,
  labels=condition_order
) +
scale_y_continuous(
  name="Final novel task count"
) +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_extra_tasks~condition, data=summary_data))
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1, xmax=group2, annotations=label, y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
# coord_flip()

```

```
theme(
  legend.position="none"
)
```

```
## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
final_novel_task_count_fig
```



```
kruskal.test(
  formula=dominant_extra_tasks~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: dominant_extra_tasks by condition
## Kruskal-Wallis chi-squared = 177.17, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$dominant_extra_tasks,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_extra_tasks and summary_data$condition
##
##          NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      0.9
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_extra_tasks)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_extra_tasks)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_extra_tasks)
  )
)

## [1] "PLASTIC median: 3; STATIC median: 3; NON-PLASTIC median: 0"
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"

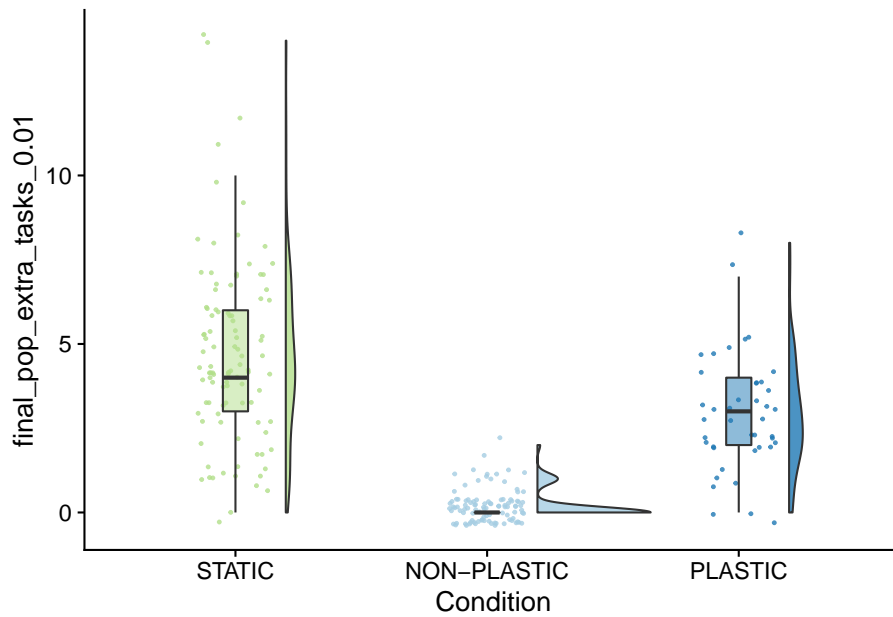
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=dominant_extra_tasks~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=184"
## [1] "STATIC<-->PLASTIC: W=1871"
## [1] "PLASTIC<-->NON-PLASTIC: W=64"
```


4.6 Novel task count (final population)

How many novel tasks are performed across the final population (1% of organisms must perform to count)?

```
ggplot(summary_data, aes(x=condition, y=final_pop_extra_tasks_0.01, fill=condition)) +  
  geom_flat_violin(  
    position = position_nudge(x = .2, y = 0),  
    alpha = .8  
  ) +  
  geom_point(  
    mapping=aes(color=condition),  
    position = position_jitter(width = .15),  
    size = .5,  
    alpha = 0.8  
  ) +  
  geom_boxplot(  
    width = .1,  
    outlier.shape = NA,  
    alpha = 0.5  
  ) +  
  scale_x_discrete(  
    name="Condition",  
    limits=condition_order  
  ) +  
  scale_fill_brewer(  
    palette=cb_palette  
  ) +  
  scale_color_brewer(  
    palette=cb_palette  
  ) +  
  # coord_flip() +  
  theme(  
    legend.position="none"  
  )
```



```
kruskal.test(
  formula=final_pop_extra_tasks_0.01~condition,
  data=summary_data
)
```

```
##
##  Kruskal-Wallis rank sum test
##
## data:  final_pop_extra_tasks_0.01 by condition
## Kruskal-Wallis chi-squared = 169.47, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$final_pop_extra_tasks_0.01,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
)
```

```
##
##  Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  summary_data$final_pop_extra_tasks_0.01 and summary_data$condition
##
##          NON-PLASTIC PLASTIC
## PLASTIC < 2e-16      -
```

```
## STATIC < 2e-16      0.00016
##
## P value adjustment method: bonferroni
paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$final_pop_extra_tasks_0.01)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$final_pop_extra_tasks_0.01)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$final_pop_extra_tasks_0.01)
  )
)

## [1] "PLASTIC median: 3; STATIC median: 4; NON-PLASTIC median: 0"
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=final_pop_extra_tasks_0.01~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=227.5"
## [1] "STATIC<-->PLASTIC: W=1203"
## [1] "PLASTIC<-->NON-PLASTIC: W=225.5"
```

4.7 Novel task discovery (lineage)

```
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_extra_traits_discovered ~ condition) %>%
```

```

adjust_pvalue(method = "bonferroni") %>%
add_significance() %>%
add_xy_position(x="condition") # ,step.increase=1
# Tweak y.position manually to account for scaled axis (edge case that triggers bad be
stat.test$manual_position <- stat.test$y.position ** c(1.0,1.0,1.03)
stat.test$label <- mapapply(p_label,stat.test$p.adj)

lineage_novel_task_discovery_fig <- ggplot(
  summary_data,
  aes(x=condition, y=dominant_lineage_extra_traits_discovered, fill=condition)
) +
geom_flat_violin(
  position = position_nudge(x = .2, y = 0),
  alpha = .8
) +
geom_point(
  mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order,
  labels=condition_order
) +
scale_y_continuous(
  name="Novel task discovery"
) +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_extra_traits_discovered~con
  )

```

```

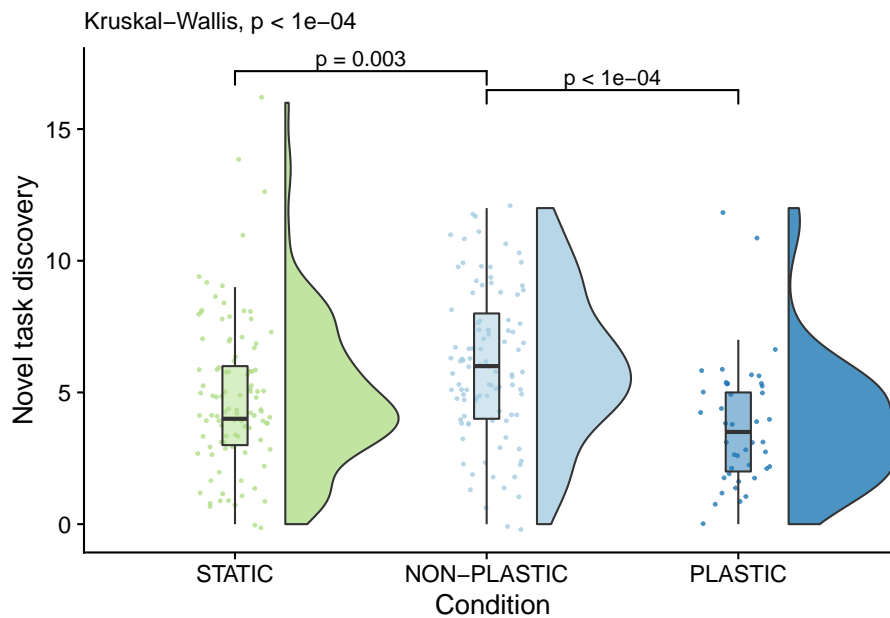
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
# coord_flip()
theme(
  legend.position="none"
)

```

```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
lineage_novel_task_discovery_fig

```



```

kruskal.test(
  formula=dominant_lineage_extra_traits_discovered~condition,
  data=summary_data
)

```

```

##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_extra_traits_discovered by condition
## Kruskal-Wallis chi-squared = 24.099, df = 2, p-value = 5.846e-06

```

```

pairwise.wilcox.test(
  x=summary_data$dominant_lineage_extra_traits_discovered,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_extra_traits_discovered and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC 1.7e-05      -
## STATIC  0.0035      0.0561
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_extra_traits_discovered
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_lineage_extra_traits_discovered
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_extra_traits_discovered
  )
)

## [1] "PLASTIC median: 3.5; STATIC median: 4; NON-PLASTIC median: 6"
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=dominant_lineage_extra_traits_discovered~condition,
    data=pair_data,

```

```

    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

```

```

## [1] "STATIC<-->NON-PLASTIC: W=6319.5"
## [1] "STATIC<-->PLASTIC: W=1578"
## [1] "PLASTIC<-->NON-PLASTIC: W=3110.5"

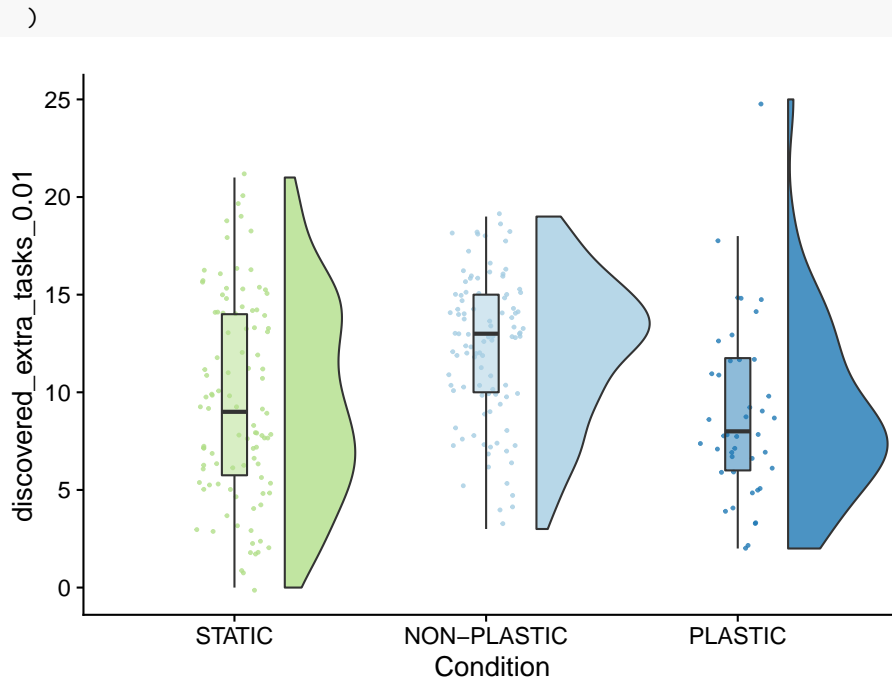
```

4.8 Novel task discovery (population)

```

ggplot(
  summary_data,
  aes(x=condition, y=discovered_extra_tasks_0.01, fill=condition)
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  # coord_flip() +
  theme(
    legend.position="none"
  )

```



```
kruskal.test(
  formula=discovered_extra_tasks_0.01~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: discovered_extra_tasks_0.01 by condition
## Kruskal-Wallis chi-squared = 24.271, df = 2, p-value = 5.365e-06
```

```
pairwise.wilcox.test(
  x=summary_data$discovered_extra_tasks_0.01,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$discovered_extra_tasks_0.01 and summary_data$condition
##
```



```
##          NON-PLASTIC PLASTIC
## PLASTIC 2.4e-05      -
## STATIC 0.00035      1.00000
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$discovered_extra_tasks_0.01)
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$discovered_extra_tasks_0.01)
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$discovered_extra_tasks_0.01)
  )
)
```

```
## [1] "PLASTIC median: 8; STATIC median: 9; NON-PLASTIC median: 13"
print("Wilcox rank sum test statistics:")
```

```
## [1] "Wilcox rank sum test statistics:"

for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=discovered_extra_tasks_0.01~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}
```

```
## [1] "STATIC<-->NON-PLASTIC: W=6573.5"
## [1] "STATIC<-->PLASTIC: W=1918.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=3096"
```

4.9 Novel task discovery frequency (lineage)

```
summary_data$dominant_lineage_extra_traits_discovered_per_generation <- summary_data$d
summary_data$dominant_lineage_extra_traits_generations_per_discovery <- summary_data$d

# Compute manual labels for geom_signif
# stat.test <- filter(summary_data, dominant_lineage_extra_traits_discovered > 0) %>%
#   wilcox_test(dominant_lineage_extra_traits_generations_per_discovery ~ condition) %>%
#   adjust_pvalue(method = "bonferroni") %>%
#   add_significance() %>%
#   add_xy_position(x="condition") # ,step.increase=1
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_extra_traits_discovered_per_generation ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition", step.increase=0.0001) # ,step.increase=1
# Tweak y.position manually to account for scaled axis (edge case that triggers bad be
stat.test$manual_position <- stat.test$y.position ## c(1.0,1.0,1.03)
stat.test$label <- mapply(p_label,stat.test$p.adj)

lineage_novel_task_discovery_freq_fig <- ggplot(
  # filter(summary_data, dominant_lineage_extra_traits_discovered > 0),
  summary_data,
  aes(x=condition, y=dominant_lineage_extra_traits_discovered_per_generation, fill=c
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  ylab("Novel task discovery frequency") +
```

```

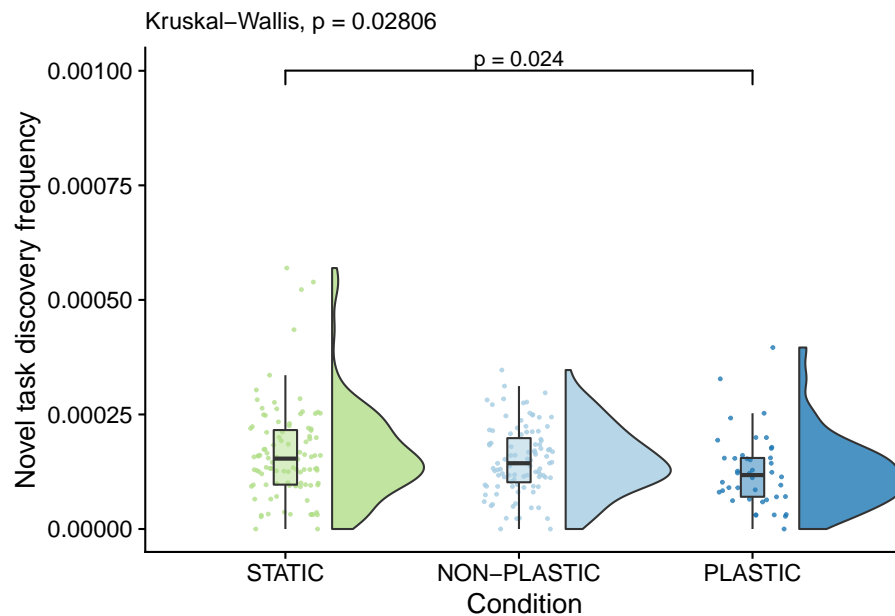
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_extra_traits_discovered_per_generation
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
# coord_flip() +
theme(
  legend.position="none"
)

```

```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
lineage_novel_task_discovery_freq_fig

```



```
kruskal.test(
  formula=dominant_lineage_extra_traits_discovered_per_generation~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_extra_traits_discovered_per_generation by condition
## Kruskal-Wallis chi-squared = 7.1465, df = 2, p-value = 0.02806
```

```
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_extra_traits_discovered_per_generation,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_extra_traits_discovered_per_generation and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC 0.092      -
```

```
## STATIC 1.000 0.025
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_extra_traits_discovered_per
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_lineage_extra_traits_discovered_per
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_extra_traits_discovered_per
  )
)

## [1] "PLASTIC median: 0.000117695011124939; STATIC median: 0.00015363220504867; NON-PLASTIC median: 0.000117695011124939"
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"

for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=dominant_lineage_extra_traits_discovered_per_generation~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=4751"
## [1] "STATIC<-->PLASTIC: W=1510.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=2584"
```

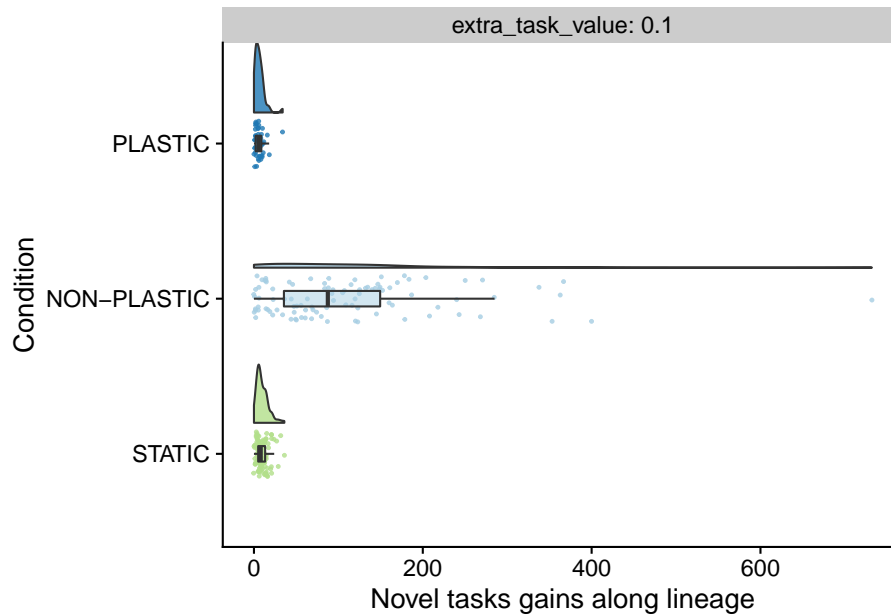
4.10 Novel tasks gained (lineage)

```
ggplot(
  summary_data,
  aes(x=condition, y=dominant_lineage_extra_traits_gained, fill=condition)
```

```

) +
geom_flat_violin(
  position = position_nudge(x = .2, y = 0),
  alpha = .8
) +
geom_point(
  mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order
) +
ylab("Novel tasks gains along lineage") +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
coord_flip() +
facet_wrap(
  ~extra_task_value,
  labeller=label_both
) +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/dominant-lineage-extra-tasks-gained.pdf"),
  width=15,
  height=10
)

```



4.11 Novel task loss (lineage)

```
# Compute manual labels for geom_signif
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_extra_traits_lost ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition", step.increase=1)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- log10(stat.test$y.position) * c(1.0,1.0,1.03)
stat.test$label <- mapapply(p_label,stat.test$p.adj)

lineage_novel_task_loss_fig <- ggplot(
  summary_data,
  aes(x=condition, y=dominant_lineage_extra_traits_lost, fill=condition)
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
```

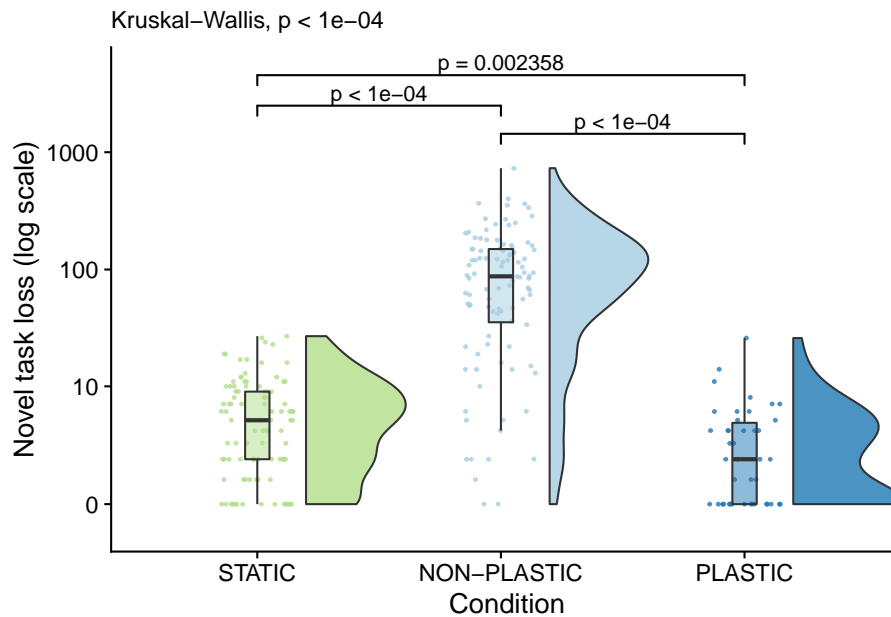
```

    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order,
    labels=condition_order
  ) +
  scale_y_continuous(
    name="Novel task loss (log scale)",
    trans=pseudo_log_trans(sigma=1, base=10),
    breaks=c(0,10,100,1000),
    limits=c(-1,5000)
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  labs(
    subtitle=paste0(
      "Kruskal-Wallis, ",
      p_label(signif(kruskal.test(formula=dominant_lineage_extra_traits_lost~condition
    )
  ) +
  ggsignif::geom_signif(
    data=filter(stat.test, p.adj<=alpha),
    aes(xmin=group1,xmax=group2,annotations=label,y_position>manual_position),
    manual=TRUE,
    inherit.aes=FALSE
  ) +
  # coord_flip()
  theme(
    legend.position="none"
  )

```

```
## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
```


lineage_novel_task_loss_fig



```
kruskal.test(
  formula=dominant_lineage_extra_traits_lost~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_extra_traits_lost by condition
## Kruskal-Wallis chi-squared = 129.06, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_extra_traits_lost,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_extra_traits_lost and summary_data$condition
##
```

```
##          NON-PLASTIC PLASTIC
## PLASTIC 2.7e-16      -
## STATIC  < 2e-16      0.0024
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_extra_traits_lost
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_lineage_extra_traits_lost
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_extra_traits_lost
  )
)
```

```
## [1] "PLASTIC median: 2; STATIC median: 5; NON-PLASTIC median: 87.5"
print("Wilcox rank sum test statistics:")
```

```
## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=dominant_lineage_extra_traits_lost~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}
```

```
## [1] "STATIC<-->NON-PLASTIC: W=9105"
## [1] "STATIC<-->PLASTIC: W=1353.5"
## [1] "PLASTIC<-->NON-PLASTIC: W=3959"
```

4.12 Frequency of novel task loss (lineage)

```
summary_data$dominant_lineage_extra_traits_lost_per_generation <- summary_data$dominant_lineage_e
summary_data$dominant_lineage_extra_traits_generations_per_loss <- summary_data$dominant_generati

# Compute manual labels for geom_signif
# stat.test <- filter(summary_data, dominant_lineage_extra_traits_lost > 0) %>%
#   wilcox_test(dominant_lineage_extra_traits_generations_per_loss ~ condition) %>%
#   adjust_pvalue(method = "bonferroni") %>%
#   add_significance() %>%
#   add_xy_position(x="condition", step.increase=1)
stat.test <- summary_data %>%
  wilcox_test(dominant_lineage_extra_traits_lost_per_generation ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition", step.increase=.1)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- stat.test$y.position ## c(1.0,1.0,1.03)
stat.test$label <- mapapply(p_label,stat.test$p.adj)

lineage_novel_task_loss_freq_fig <- ggplot(
  # filter(summary_data, dominant_lineage_extra_traits_lost > 0),
  summary_data,
  aes(x=condition, y=dominant_lineage_extra_traits_lost_per_generation, fill=condition)
) +
geom_flat_violin(
  position = position_nudge(x = .2, y = 0),
  alpha = .8
) +
geom_point(
  mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order
) +
ylab("Novel task loss frequency") +
```

```

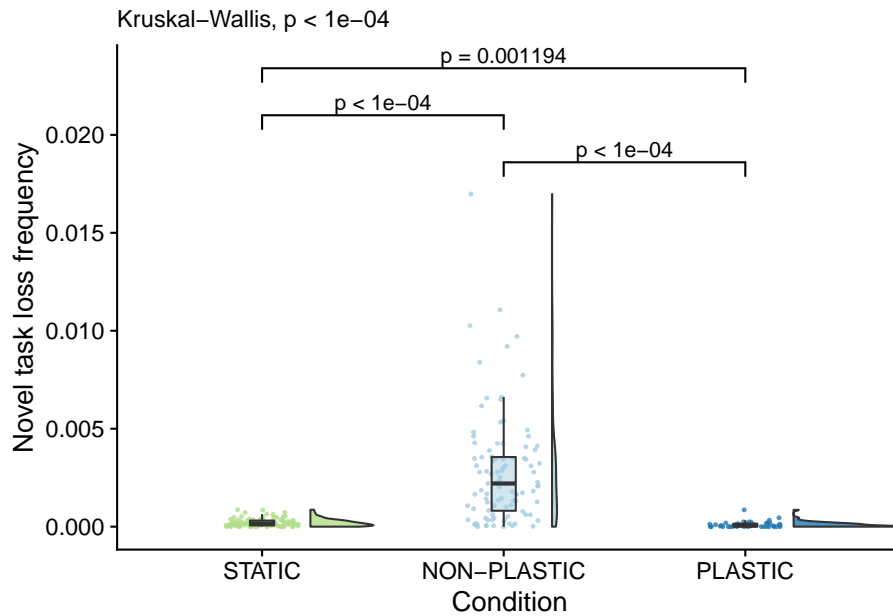
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_extra_traits_lost_per_gener
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj<=alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
)

```

```

## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
lineage_novel_task_loss_freq_fig

```



```

kruskal.test(
  formula=dominant_lineage_extra_traits_lost_per_generation~condition,
  data=summary_data
)

##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_extra_traits_lost_per_generation by condition
## Kruskal-Wallis chi-squared = 121.41, df = 2, p-value < 2.2e-16

pairwise.wilcox.test(
  x=summary_data$dominant_lineage_extra_traits_lost_per_generation,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
  conf.int=TRUE,
  conf.level=0.95
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_extra_traits_lost_per_generation and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC 1.1e-15      -
## STATIC  < 2e-16    0.0012
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_extra_traits_lost_per_generation
  ),
  paste0(
    "STATIC median: ",
    median(filter(summary_data, condition=="STATIC"))$dominant_lineage_extra_traits_lost_per_generation
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_extra_traits_lost_per_generation
  )
)

## [1] "PLASTIC median: 6.25141973661864e-05; STATIC median: 0.000161396283669756; NON-PLASTIC median: 0.000161396283669756"

```

```

print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=dominant_lineage_extra_traits_lost_per_generation~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
  print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))
}

## [1] "STATIC<-->NON-PLASTIC: W=8940"
## [1] "STATIC<-->PLASTIC: W=1311"
## [1] "PLASTIC<-->NON-PLASTIC: W=3922"

```

4.13 How many instances of novel trait loss co-occurred with changes in base phenotype?

Task loss linked with primary trait changes.

```

lost_traits_summary_data <- filter(summary_data, extra_task_value==0.1 & dominant_lineage_extra_traits_lost_per_generation>0.1)
lost_traits_summary_data$frac_linked_extra_trait_loss <- lost_traits_summary_data$dominant_lineage_extra_traits_lost_per_generation / lost_traits_summary_data$extra_task_value

ggplot(lost_traits_summary_data, aes(x=condition, y=frac_linked_extra_trait_loss, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",

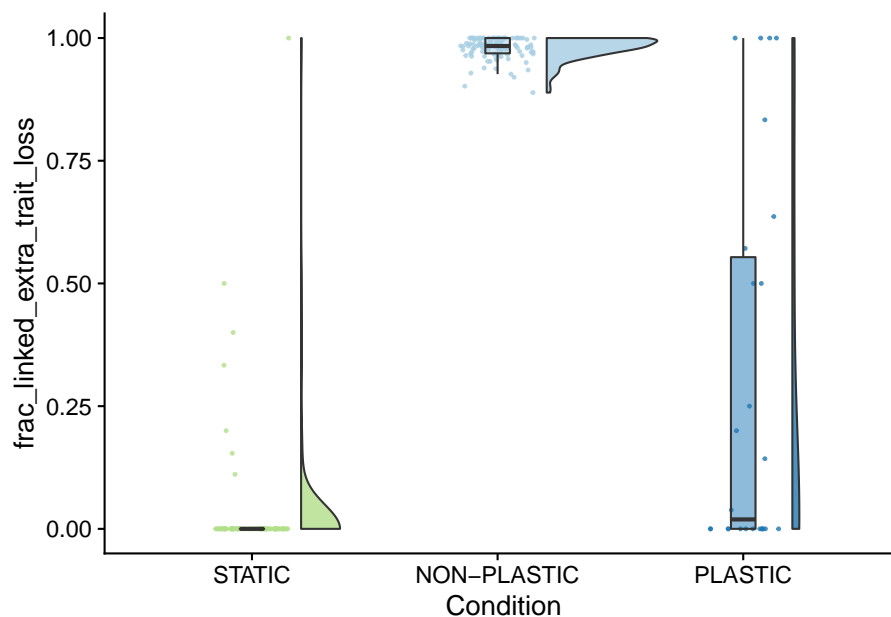
```

4.13. HOW MANY INSTANCES OF NOVEL TRAIT LOSS CO-OCCURRED WITH CHANGES IN BASE PHENO

```

    limits=condition_order
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  # coord_flip() +
  theme(
    legend.position="none"
  )

```



```

kruskal.test(
  formula=frac_linked_extra_trait_loss~condition,
  data=lost_traits_summary_data
)

```

```

##
##  Kruskal-Wallis rank sum test
##
## data:  frac_linked_extra_trait_loss by condition
## Kruskal-Wallis chi-squared = 153.68, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=lost_traits_summary_data$frac_linked_extra_trait_loss,

```

```

g=lost_traits_summary_data$condition,
p.adjust.method="bonferroni",
conf.int=TRUE,
conf.level=0.95
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: lost_traits_summary_data$frac_linked_extra_trait_loss and lost_traits_summary_data$frac_linked_extra_trait_loss
##
##      NON-PLASTIC PLASTIC
## PLASTIC 1.9e-08      -
## STATIC  < 2e-16      1.8e-06
##
## P value adjustment method: bonferroni

paste(
  sep="; ",
  paste0(
    "PLASTIC median: ",
    median(filter(lost_traits_summary_data, condition=="PLASTIC"))$frac_linked_extra_trait_loss
  ),
  paste0(
    "STATIC median: ",
    median(filter(lost_traits_summary_data, condition=="STATIC"))$frac_linked_extra_trait_loss
  ),
  paste0(
    "NON-PLASTIC median: ",
    median(filter(lost_traits_summary_data, condition=="NON-PLASTIC"))$frac_linked_extra_trait_loss
  )
)

## [1] "PLASTIC median: 0.0192307692307692; STATIC median: 0; NON-PLASTIC median: 0.983333333333333"
print("Wilcox rank sum test statistics:")

## [1] "Wilcox rank sum test statistics:"
for (pair in pairwise_comparisons) {
  pair_data <- filter(lost_traits_summary_data, condition %in% pair)
  pair_data$condition <- as.factor(pair_data$condition)
  wt <- wilcox.test(
    formula=frac_linked_extra_trait_loss~condition,
    data=pair_data,
    exact=FALSE,
    paired=FALSE
  )
}

```


4.13. HOW MANY INSTANCES OF NOVEL TRAIT LOSS CO-OCCURRED WITH CHANGES IN BASE PHENO

```
)  
print(paste0(pair[1], "<-->", pair[2], ": W=", wt$statistic))  
}  
  
## [1] "STATIC<-->NON-PLASTIC: W=8344"  
## [1] "STATIC<-->PLASTIC: W=1602"  
## [1] "PLASTIC<-->NON-PLASTIC: W=2212"  
  
sum(filter(lost_traits_summary_data, condition=="NON-PLASTIC"))$dominant_lineage_extra_traits_lost  
  
## [1] 10998  
sum(filter(lost_traits_summary_data, condition=="NON-PLASTIC"))$dominant_lineage_extra_traits_lost  
  
## [1] 11229  
aggregate_frac_linked_extra_trait_loss_nonplastic <- sum(filter(lost_traits_summary_data, condition=="NON-PLASTIC"))$dominant_lineage_extra_traits_lost  
aggregate_frac_linked_extra_trait_loss_nonplastic  
  
## [1] 0.9794283  
sum(filter(lost_traits_summary_data, condition=="PLASTIC"))$dominant_lineage_extra_traits_lost  
  
## [1] 29  
sum(filter(lost_traits_summary_data, condition=="PLASTIC"))$dominant_lineage_extra_traits_lost  
  
## [1] 142  
aggregate_frac_linked_extra_trait_loss_plastic <- sum(filter(lost_traits_summary_data, condition=="PLASTIC"))$dominant_lineage_extra_traits_lost  
aggregate_frac_linked_extra_trait_loss_plastic  
  
## [1] 0.2042254  
sum(filter(lost_traits_summary_data, condition=="STATIC"))$dominant_lineage_extra_traits_lost  
  
## [1] 13  
sum(filter(lost_traits_summary_data, condition=="STATIC"))$dominant_lineage_extra_traits_lost  
  
## [1] 631  
aggregate_frac_linked_extra_trait_loss_nonplastic <- sum(filter(lost_traits_summary_data, condition=="NON-PLASTIC"))$dominant_lineage_extra_traits_lost  
aggregate_frac_linked_extra_trait_loss_nonplastic  
  
## [1] 0.02060222
```

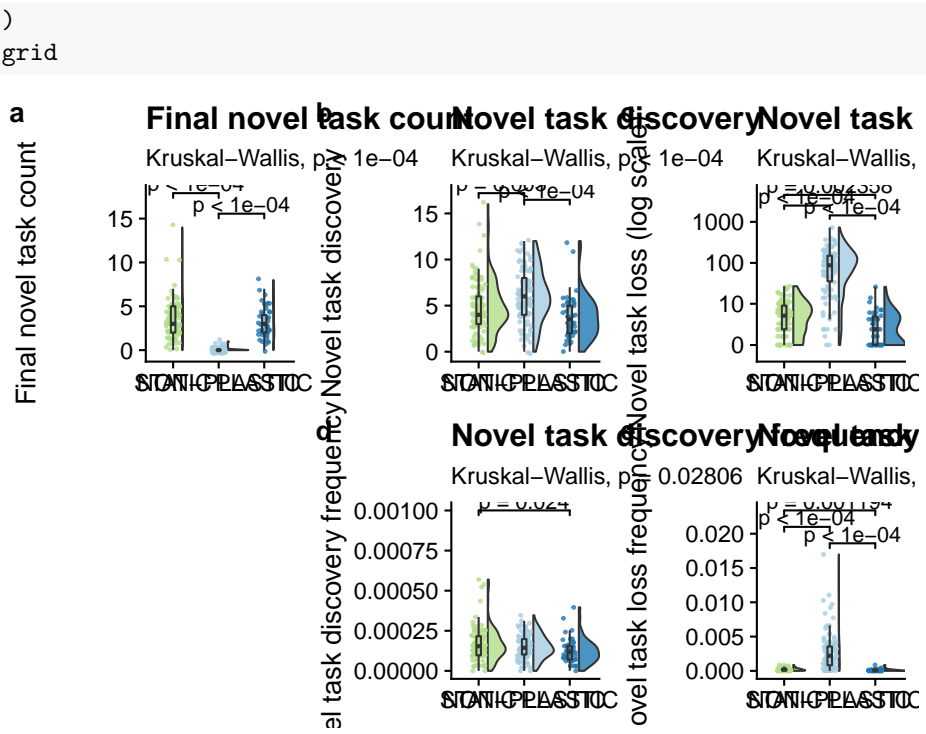
4.14 Manuscript figures

4.15 Combined panel

```

grid <- plot_grid(
  final_novel_task_count_fig +
    theme(
      axis.title.x=element_blank()
    ) +
    ggtitle("Final novel task count"),
  lineage_novel_task_discovery_fig +
    theme(
      axis.title.x=element_blank(),
      # axis.text.x=element_blank(),
      # axis.ticks.x=element_blank()
    ) +
    ggtitle("Novel task discovery"),
  lineage_novel_task_loss_fig +
    theme(
      axis.title.x=element_blank(),
      # axis.text.x=element_blank(),
      # axis.ticks.x=element_blank()
    ) +
    ggtitle("Novel task loss"),
  NULL,
  lineage_novel_task_discovery_freq_fig +
    theme(
      axis.title.x=element_blank()
    ) +
    ggtitle("Novel task discovery frequency"),
  lineage_novel_task_loss_freq_fig +
    theme(
      axis.title.x=element_blank()
    ) +
    ggtitle("Novel task loss frequency"),
  nrow=2,
  align="v",
  # labels="auto"
  labels=c("a", "b", "c", "", "d", "e")
)
save_plot(
  paste0(working_directory, "plots/", "complex-traits-panel.pdf"),
  grid,
  base_height=12,
  base_asp=3/2
)

```



Chapter 5

Accumulation of deleterious instructions

The effect of adaptive phenotypic plasticity on the accumulation of deleterious genes.

5.1 Overview

```
total_updates <- 200000
replicates <- 100
alpha <- 0.05
focal_poison_penalty <- 0.1

focal_traits <- c("not", "nand", "and", "ornot", "or", "andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-02-05-hitchhiking/analysis/" # << For bookdown
# working_directory <- "./"
```

5.2 Analysis dependencies

Load all required R libraries.

```
library(RColorBrewer)
library(ggplot2)
library(rstatix)
```

```
library(ggsignif)
library(scales)
library(tidyverse)
library(cowplot)
library(Hmisc)
library(boot)
library(fmsb)
library(knitr)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd9")
```

These analyses were conducted/knitted with the following computing environment:

```
print(version)

##
## platform      _
## arch          x86_64-pc-linux-gnu
## os            linux-gnu
## system        x86_64, linux-gnu
## status
## major         4
## minor         0.5
## year          2021
## month         03
## day           31
## svn rev       80133
## language      R
## version.string R version 4.0.5 (2021-03-31)
## nickname      Shake and Throw
```

5.3 Setup

```
##### summary data #####
summary_data_loc <- paste0(working_directory, "data/aggregate.csv")
summary_data <- read.csv(summary_data_loc, na.strings="NONE")

summary_data$DISABLE_REACTION_SENSORS <- as.factor(summary_data$DISABLE_REACTION_SENSORS)
summary_data$chg_env <- summary_data$chg_env == "True"
summary_data$dominant_plastic_odd_even <- as.factor(summary_data$dominant_plastic_odd_even)
summary_data$sensors <- summary_data$DISABLE_REACTION_SENSORS == "0"
summary_data$is_plastic <- summary_data$dominant_plastic_odd_even == "True"
summary_data$POISON_PENALTY <- as.factor(summary_data$POISON_PENALTY)

summary_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation <-
```

```

summary_data$frac_hitchhiking_linked_trait_change <- summary_data$dominant_lineage_num_times_hitchhike
summary_data$frac_unexpressed_hitchhiker_inc <- summary_data$dominant_lineage_num_times_hitchhike
summary_data$frac_expressed_hitchhiker_inc <- summary_data$dominant_lineage_num_times_hitchhike_in

env_label_fun <- function(chg_env) {
  if (chg_env) {
    return("Fluctuating")
  } else {
    return("Constant")
  }
}

sensors_label_fun <- function(has_sensors) {
  if (has_sensors) {
    return("Sensors")
  } else {
    return("No sensors")
  }
}

condition_label_fun <- function(has_sensors, env_chg) {
  if (has_sensors && env_chg) {
    return("PLASTIC")
  } else if (env_chg) {
    return("NON-PLASTIC")
  } else {
    return("STATIC")
  }
}

summary_data$env_label <- mapply(
  env_label_fun,
  summary_data$chg_env
)

summary_data$sensors_label <- mapply(
  sensors_label_fun,
  summary_data$sensors
)

summary_data$condition <- mapply(
  condition_label_fun,
  summary_data$sensors,
  summary_data$chg_env
)

condition_order = c(

```

```

    "STATIC",
    "NON-PLASTIC",
    "PLASTIC"
  )

pairwise_comparisons <- list(
  c("STATIC", "NON-PLASTIC"),
  c("STATIC", "PLASTIC"),
  c("PLASTIC", "NON-PLASTIC")
)

p_label <- function(p_value) {
  threshold = 0.0001
  if (p_value < threshold) {
    return(paste0("p < ", threshold))
  } else {
    return(paste0("p = ", p_value))
  }
}

poison_penalties <- levels(summary_data$POISON_PENALTY)

##### time series #####
lineage_time_series_data_loc <- paste0(working_directory, "data/lineage_series.csv")
lineage_time_series_data <- read.csv(lineage_time_series_data_loc)

lineage_time_series_data$DISABLE_REACTION_SENSORS <- as.factor(lineage_time_series_data$DISABLE_REACTION_SENSORS == "True")
lineage_time_series_data$chg_env <- lineage_time_series_data$chg_env == "True"
lineage_time_series_data$sensors <- lineage_time_series_data$DISABLE_REACTION_SENSORS == "True"
lineage_time_series_data$POISON_PENALTY <- as.factor(lineage_time_series_data$POISON_PENALTY)

lineage_time_series_data$env_label <- mapapply(
  env_label_fun,
  lineage_time_series_data$chg_env
)

lineage_time_series_data$sensors_label <- mapapply(
  sensors_label_fun,
  lineage_time_series_data$sensors
)

lineage_time_series_data$condition <- mapapply(
  condition_label_fun,
  lineage_time_series_data$sensors,
  lineage_time_series_data$chg_env
)

```



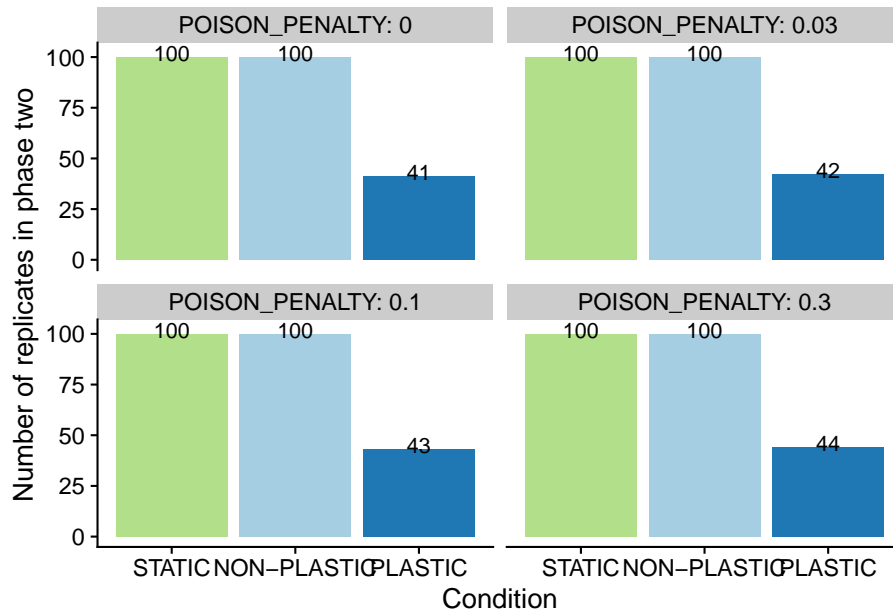
```
##### misc #####
# Configure our default graphing theme
focal_summary_data <- filter(summary_data, POISON_PENALTY==focal_poison_penalty)
theme_set(theme_cowplot())
cb_palette <- "Paired"
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)
samplemean <- function(x, d) {
  return(mean(x[d]))
}
```

5.4 Evolution of phenotypic plasticity

For sensor-enabled populations in fluctuating environments, we only transferred populations containing an optimally plastic genotype to phase-two.

```
summary_data_grouped = dplyr::group_by(summary_data, sensors, env_label, condition, POISON_PENALTY)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
```

```
ggplot(summary_data_group_counts, aes(x=condition, y=n, fill=condition)) +
  geom_col(position=position_dodge(0.9)) +
  geom_text(aes(label=n, y=n+2)) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  ylab("Number of replicates in phase two") +
  facet_wrap(~POISON_PENALTY, labeller=label_both) +
  theme(
    legend.position="none"
  )
```

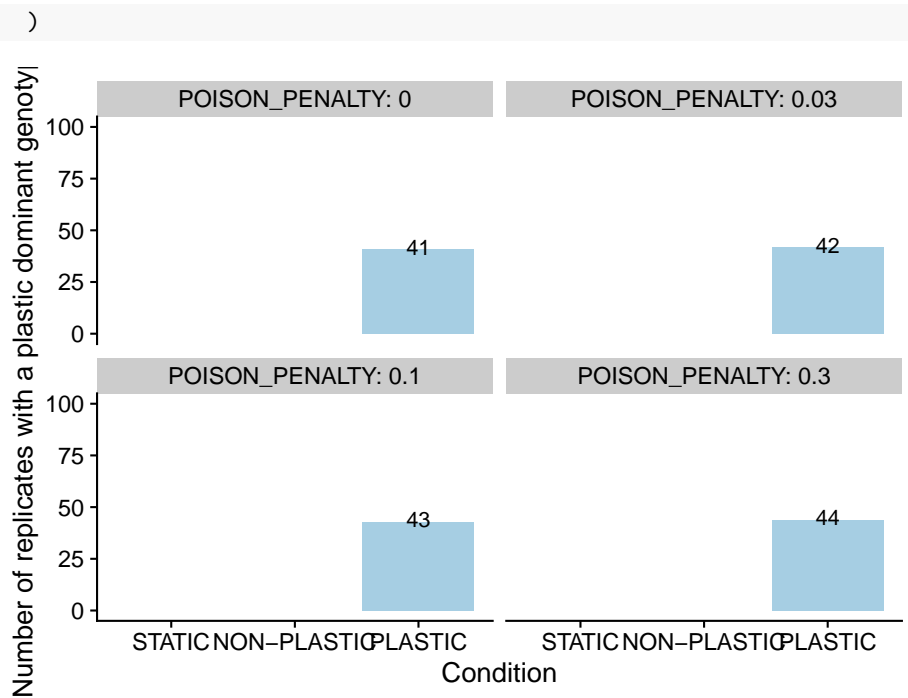


We can confirm our expectation that the dominant genotypes in non-plastic conditions are not phenotypically plastic.

```
summary_data_grouped = dplyr::group_by(summary_data, condition, is_plastic, POISON_PENALTY)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
```

`summarise()` has grouped output by 'condition', 'is_plastic'. You can override using `ungroup()`.

```
ggplot(filter(summary_data_group_counts, is_plastic), aes(x=condition, y=n, fill=condition)) +
  geom_col(position=position_dodge(0.9)) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  geom_text(aes(label=n, y=n+1)) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  ylab("Number of replicates with a plastic dominant genotype") +
  ylim(0, 100) +
  facet_wrap(~POISON_PENALTY, labeller=label_both) +
  theme(
    legend.position="none"
```



5.5 Poison instruction execution

5.5.1 Number of replicates where final dominant genotype executes the poison instruction

```
for (penalty in poison_penalties) {
  occurrences <- c(
    length(filter(summary_data, POISON_PENALTY==penalty & condition=="NON-PLASTIC" & dominant_times_p
    length(filter(summary_data, POISON_PENALTY==penalty & condition=="PLASTIC" & dominant_times_p
    length(filter(summary_data, POISON_PENALTY==penalty & condition=="STATIC" & dominant_times_p
  )
  trials <- c(
    length(filter(summary_data, POISON_PENALTY==penalty & condition=="NON-PLASTIC")$RANDOM_SEED),
    length(filter(summary_data, POISON_PENALTY==penalty & condition=="PLASTIC")$RANDOM_SEED),
    length(filter(summary_data, POISON_PENALTY==penalty & condition=="STATIC" )$RANDOM_SEED)
  )
  names(trials) <- c(
    "NON-PLASTIC",
    "PLASTIC",
    "STATIC"
  )
}
```

```

names(occurrences) <- c(
  "NON-PLASTIC",
  "PLASTIC",
  "STATIC"
)
poison_exec_table <- data.frame(
  executes.poison=occurrences,
  replicates=trials
)
cat(paste0("#### Penalty: ", penalty, "\n"))
cat(print(kable(poison_exec_table)))
cat("\n")
ft <- pairwise.fisher.test(x=occurrences, n=trials, p.adjust.method="bonferroni")
print(ft)
cat("\n\n")
}

```

```

## #### Penalty: 0
##
## \begin{tabular}{l|r|r}
## \hline
##   & executes.poison & replicates\\
## \hline
## NON-PLASTIC & 86 & 100\\
## \hline
## PLASTIC & 27 & 41\\
## \hline
## STATIC & 85 & 100\\
## \hline
## \end{tabular}
##
##
## Pairwise comparisons using Pairwise comparison of proportions (Fisher)
##
## data:  occurrences out of trials
##
##           NON-PLASTIC PLASTIC
## PLASTIC 0.03           -
## STATIC  1.00           0.06
##
## P value adjustment method: bonferroni
##
##
## #### Penalty: 0.03
##

```

```

## \begin{tabular}{l|r|r}
## \hline
##   & executes.poison & replicates\\
## \hline
## NON-PLASTIC & 46 & 100\\
## \hline
## PLASTIC & 1 & 42\\
## \hline
## STATIC & 1 & 100\\
## \hline
## \end{tabular}
##
##
## Pairwise comparisons using Pairwise comparison of proportions (Fisher)
##
## data:  occurrences out of trials
##
##           NON-PLASTIC PLASTIC
## PLASTIC 1.2e-07      -
## STATIC  2.9e-15      1
##
## P value adjustment method: bonferroni
##
##
## ##### Penalty: 0.1
##
## \begin{tabular}{l|r|r}
## \hline
##   & executes.poison & replicates\\
## \hline
## NON-PLASTIC & 14 & 100\\
## \hline
## PLASTIC & 0 & 43\\
## \hline
## STATIC & 0 & 100\\
## \hline
## \end{tabular}
##
##
## Pairwise comparisons using Pairwise comparison of proportions (Fisher)
##
## data:  occurrences out of trials
##
##           NON-PLASTIC PLASTIC
## PLASTIC 0.03212      -
## STATIC  0.00022      1.00000

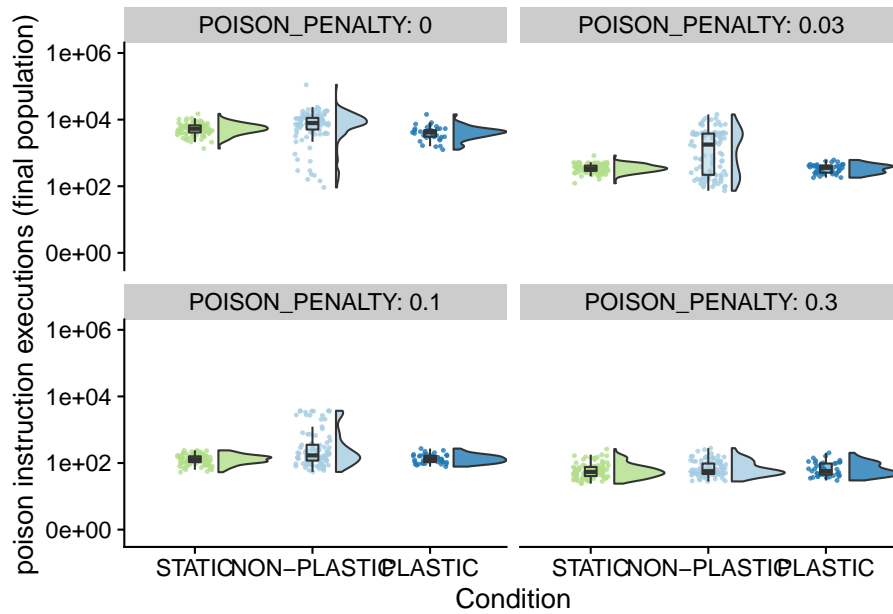
```

```
##
## P value adjustment method: bonferroni
##
## ##### Penalty: 0.3
##
## \begin{tabular}{l|r|r}
## \hline
## & executes.poison & replicates\\
## \hline
## NON-PLASTIC & 0 & 100\\
## \hline
## PLASTIC & 0 & 44\\
## \hline
## STATIC & 0 & 100\\
## \hline
## \end{tabular}
##
##
## Pairwise comparisons using Pairwise comparison of proportions (Fisher)
##
## data: occurrences out of trials
##
##          NON-PLASTIC PLASTIC
## PLASTIC 1             -
## STATIC  1             1
##
## P value adjustment method: bonferroni
```

5.5.2 Poison instruction execution (final population)

```
ggplot(summary_data, aes(x=condition, y=final_population_poison, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
```

```
alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order
) +
scale_y_continuous(
  name="poison instruction executions (final population)",
  trans=pseudo_log_trans(sigma=1,base=10),
  breaks=c(0,100,10000,1000000),
  limits=c(-1,1000000)
) +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
facet_wrap(
  ~POISON_PENALTY,
  labeller=label_both
) +
# coord_flip() +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/final-population-poison-log.pdf"),
  width=15,
  height=10
)
```



```

for (penalty in poison_penalties) {
  stat_data <- filter(summary_data, POISON_PENALTY==penalty)
  print(
    paste0(
      "PENALTY: ", penalty
    )
  )
  kt <- kruskal.test(
    formula=final_population_poison~condition,
    data=stat_data
  )
  print(
    kt
  )
  if (is.na(kt$p.value)) { next }
  if (kt$p.value > 0.05) { next }
  print(
    pairwise.wilcox.test(
      x=stat_data$final_population_poison,
      g=stat_data$condition,
      p.adjust.method="bonferroni"
    )
  )
}

```



```
## [1] "PENALTY: 0"
##
## Kruskal-Wallis rank sum test
##
## data: final_population_poison by condition
## Kruskal-Wallis chi-squared = 43.589, df = 2, p-value = 3.426e-10
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$final_population_poison and stat_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC 8.7e-07      -
## STATIC  9.8e-07      0.00074
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
##
## Kruskal-Wallis rank sum test
##
## data: final_population_poison by condition
## Kruskal-Wallis chi-squared = 20.74, df = 2, p-value = 3.136e-05
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$final_population_poison and stat_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC 0.003      -
## STATIC  1e-04      1.000
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
##
## Kruskal-Wallis rank sum test
##
## data: final_population_poison by condition
## Kruskal-Wallis chi-squared = 20.608, df = 2, p-value = 3.35e-05
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$final_population_poison and stat_data$condition
##
```

```
##          NON-PLASTIC PLASTIC
## PLASTIC 0.0093      -
## STATIC 4.9e-05     1.0000
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.3"
##
## Kruskal-Wallis rank sum test
##
## data: final_population_poison by condition
## Kruskal-Wallis chi-squared = 3.3994, df = 2, p-value = 0.1827
```

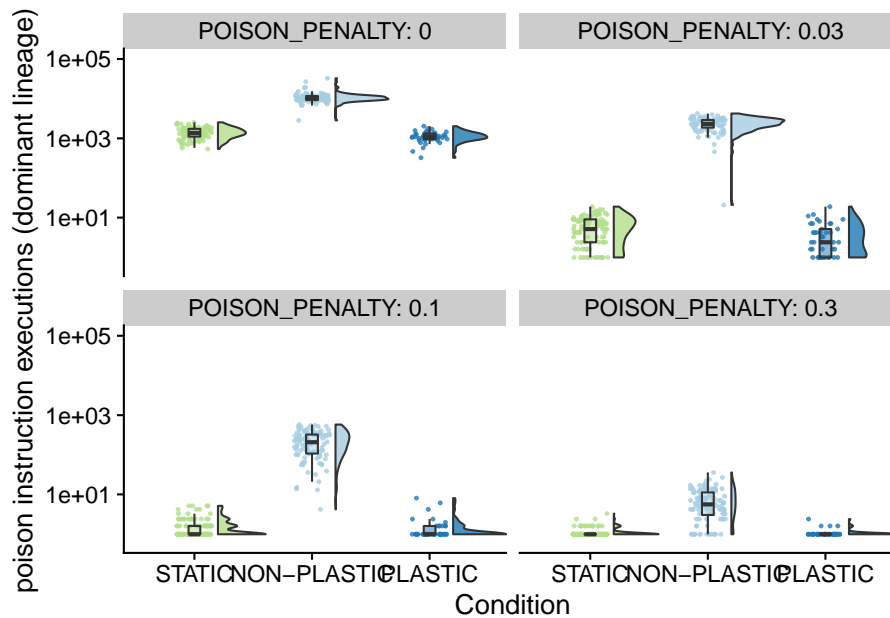
5.5.3 Cumulative poison instruction execution along final dominant lineages

```
ggplot(summary_data, aes(x=condition, y=dominant_lineage_times_poison_executed, fill=c
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_y_continuous(
    name="poison instruction executions (dominant lineage)",
    trans=pseudo_log_trans(sigma = 1, base = 10),
    breaks=c(10,1000,100000),
    limits=c(-1,100000)
  ) +
  facet_wrap(
    ~POISON_PENALTY,
    labeller=label_both
  ) +
```

```

scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/final-dominant-lineage-poison-log.pdf"),
  width=15,
  height=10
)

```



```

for (penalty in poison_penalties) {
  stat_data <- filter(summary_data, POISON_PENALTY==penalty)
  print(
    paste0(
      "PENALTY: ", penalty
    )
  )
  kt <- kruskal.test(
    formula=dominant_lineage_times_poison_executed~condition,
    data=stat_data
  )
}

```

```

    )
    print(
      kt
    )
    if (is.na(kt$p.value)) { next }
    if (kt$p.value > 0.05) { next }
    print(
      pairwise.wilcox.test(
        x=stat_data$dominant_lineage_times_poison_executed,
        g=stat_data$condition,
        p.adjust.method="bonferroni"
      )
    )
  }
}

```

```

## [1] "PENALTY: 0"
##
##  Kruskal-Wallis rank sum test
##
## data:  dominant_lineage_times_poison_executed by condition
## Kruskal-Wallis chi-squared = 178.84, df = 2, p-value < 2.2e-16
##
##
##  Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  stat_data$dominant_lineage_times_poison_executed and stat_data$condition
##
##          NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      0.0018
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
##
##  Kruskal-Wallis rank sum test
##
## data:  dominant_lineage_times_poison_executed by condition
## Kruskal-Wallis chi-squared = 178.62, df = 2, p-value < 2.2e-16
##
##
##  Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  stat_data$dominant_lineage_times_poison_executed and stat_data$condition
##
##          NON-PLASTIC PLASTIC

```

```

## PLASTIC <2e-16      -
## STATIC  <2e-16      0.011
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
##
## Kruskal-Wallis rank sum test
##
## data:  dominant_lineage_times_poison_executed by condition
## Kruskal-Wallis chi-squared = 184.83, df = 2, p-value < 2.2e-16
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  stat_data$dominant_lineage_times_poison_executed and stat_data$condition
##
##          NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      0.21
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.3"
##
## Kruskal-Wallis rank sum test
##
## data:  dominant_lineage_times_poison_executed by condition
## Kruskal-Wallis chi-squared = 149.48, df = 2, p-value < 2.2e-16
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  stat_data$dominant_lineage_times_poison_executed and stat_data$condition
##
##          NON-PLASTIC PLASTIC
## PLASTIC 4.4e-16      -
## STATIC  < 2e-16      0.84
##
## P value adjustment method: bonferroni

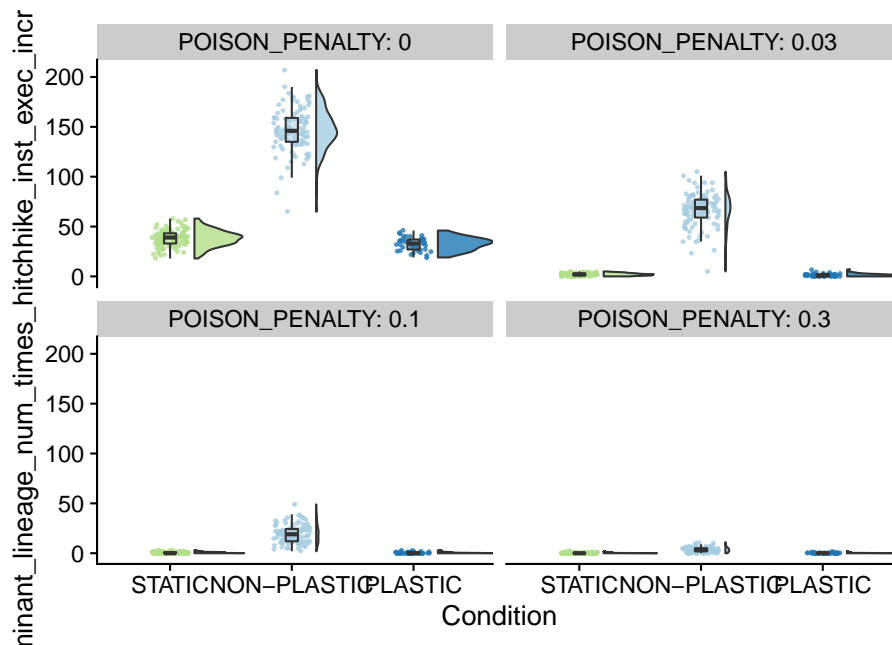
```

5.6 Characterizing mutations that increase poison instruction execution

```

ggplot(summary_data, aes(x=condition, y=dominant_lineage_num_times_hitchhike_inst_exec.
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  facet_wrap(
    ~POISON_PENALTY,
    labeller=label_both
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  theme(
    legend.position="none"
  )

```



```
for (penalty in poison_penalties) {
  stat_data <- filter(summary_data, POISON_PENALTY==penalty)
  print(
    paste0(
      "PENALTY: ", penalty
    )
  )
  kt <- kruskal.test(
    formula=dominant_lineage_num_times_hitchhike_inst_exec_increases~condition,
    data=stat_data
  )
  print(
    kt
  )
  if (is.na(kt$p.value)) { next }
  if (kt$p.value > 0.05) { next }
  print(
    pairwise.wilcox.test(
      x=stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases,
      g=stat_data$condition,
      p.adjust.method="bonferroni"
    )
  )
}
```

```

## [1] "PENALTY: 0"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases by condition
## Kruskal-Wallis chi-squared = 179.79, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases and stat_c
##
##          NON-PLASTIC PLASTIC
## PLASTIC < 2e-16      -
## STATIC  < 2e-16      0.00046
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases by condition
## Kruskal-Wallis chi-squared = 179.35, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases and stat_c
##
##          NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      0.03
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases by condition
## Kruskal-Wallis chi-squared = 185.34, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases and stat_c
##

```


5.6. CHARACTERIZING MUTATIONS THAT INCREASE POISON INSTRUCTION EXECUTION121

```
##          NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      0.27
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.3"
##
## Kruskal-Wallis rank sum test
##
## data:  dominant_lineage_num_times_hitchhike_inst_exec_increases by condition
## Kruskal-Wallis chi-squared = 146.35, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases and stat_data$condit
##
##          NON-PLASTIC PLASTIC
## PLASTIC 7.8e-16      -
## STATIC  < 2e-16      0.86
##
## P value adjustment method: bonferroni
# sum(filter(summary_data, condition=="NON-PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num_t
# sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num_times
# sum(filter(summary_data, condition=="STATIC" & POISON_PENALTY==0.1)$dominant_lineage_num_times
```

Focal figure for the manuscript:

```
# Compute manual labels for geom_signif
stat.test <- focal_summary_data %>%
  wilcox_test(dominant_lineage_num_times_hitchhike_inst_exec_increases ~ condition) %>%
  adjust_pvalue(method = "bonferroni") %>%
  add_significance() %>%
  add_xy_position(x="condition")
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- stat.test$y.position
stat.test$label <- mapapply(p_label,stat.test$p.adj)

poison_increases_fig <- ggplot(
  focal_summary_data,
  aes(x=condition, y=dominant_lineage_num_times_hitchhike_inst_exec_increases, fill=condition)
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
```

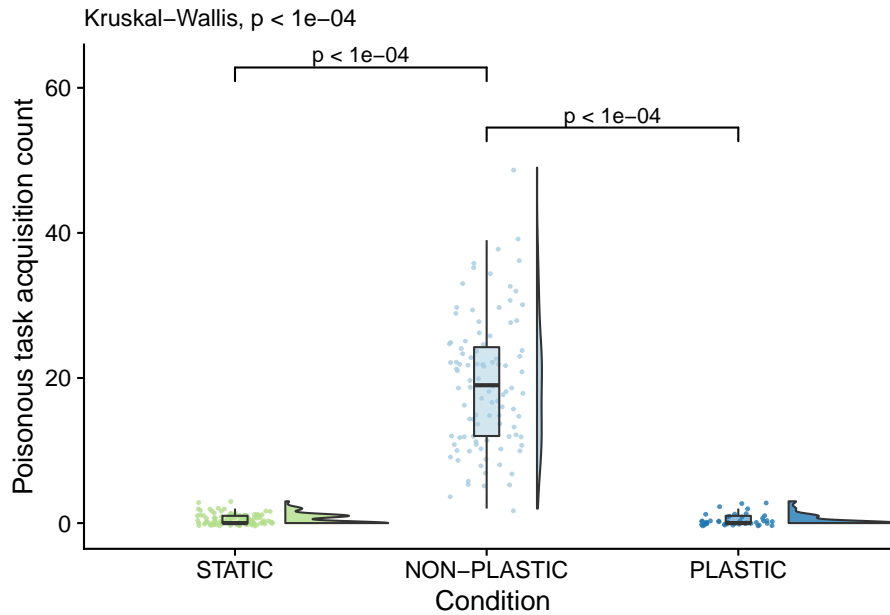
```

geom_point(
  mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order,
  labels=condition_order
) +
scale_y_continuous(
  name="Poisonous task acquisition count",
) +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
# coord_flip()
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_num_times_hitchhike_inst_ex
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position>manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
)

```

```
## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
```

poison_increases_fig



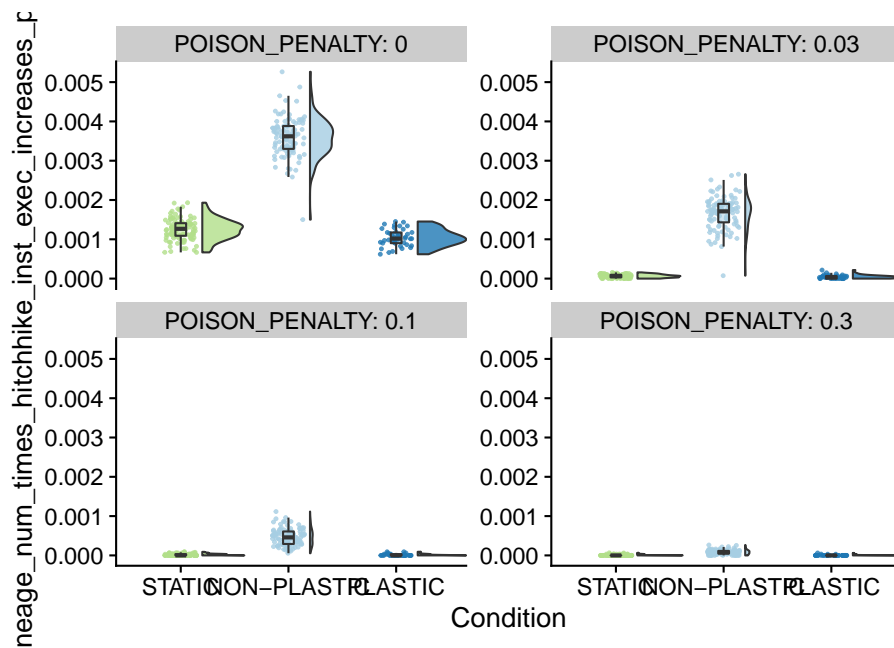
5.6.2 Frequency of increases in poison instruction execution (lineage)

```
ggplot(summary_data, aes(x=condition, y=dominant_lineage_num_times_hitchhike_inst_exec_increases_
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  )
```

```

) +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
facet_wrap(
  ~POISON_PENALTY,
  labeller=label_both,
  scales="free_y"
) +
# coord_flip() +
theme(
  legend.position="none"
) +
ggsave(
  paste0(working_directory, "plots/final-dominant-lineage-poison-increase-per-genera
  width=15,
  height=10
)

```



```

for (penalty in poison_penalties) {
  stat_data <- filter(summary_data, POISON_PENALTY==penalty)
  print(

```

5.6. CHARACTERIZING MUTATIONS THAT INCREASE POISON INSTRUCTION EXECUTION 125

```

    paste0(
      "PENALTY: ", penalty
    )
  )
  kt <- kruskal.test(
    formula=dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation~condition,
    data=stat_data
  )
  print(
    kt
  )
  if (is.na(kt$p.value)) { next }
  if (kt$p.value > 0.05) { next }
  print(
    pairwise.wilcox.test(
      x=stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation,
      g=stat_data$condition,
      p.adjust.method="bonferroni"
    )
  )
}

```

```

## [1] "PENALTY: 0"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation by condition
## Kruskal-Wallis chi-squared = 180.05, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation and s
##
##      NON-PLASTIC PLASTIC
## PLASTIC < 2e-16      -
## STATIC  < 2e-16      7.8e-05
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation by condition
## Kruskal-Wallis chi-squared = 176.25, df = 2, p-value < 2.2e-16

```

```

##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation
##
##      NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      0.019
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation by condition
## Kruskal-Wallis chi-squared = 184.17, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation
##
##      NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      0.2
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.3"
##
## Kruskal-Wallis rank sum test
##
## data: dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation by condition
## Kruskal-Wallis chi-squared = 140.99, df = 2, p-value < 2.2e-16
##
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation
##
##      NON-PLASTIC PLASTIC
## PLASTIC 2.2e-15      -
## STATIC  < 2e-16      0.79
##
## P value adjustment method: bonferroni

```

Figure for the manuscript:

```
# Compute manual labels for geom_signif
stat.test <- focal_summary_data %>%
  wilcox_test(dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation ~ condition,
    adjust_pvalue(method = "bonferroni") %>%
    add_significance() %>%
    add_xy_position(x="condition", step.increase=0.2)
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior in g
stat.test$manual_position <- stat.test$y.position
stat.test$label <- mapply(p_label,stat.test$p.adj)

poison_increases_per_gen_fig <- ggplot(
  focal_summary_data,
  aes(x=condition, y=dominant_lineage_num_times_hitchhike_inst_exec_increases_per_generation, f
) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order,
    labels=condition_order
  ) +
  scale_y_continuous(
    name="Poisonous task acquisition frequency",
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  # coord_flip()
```

```

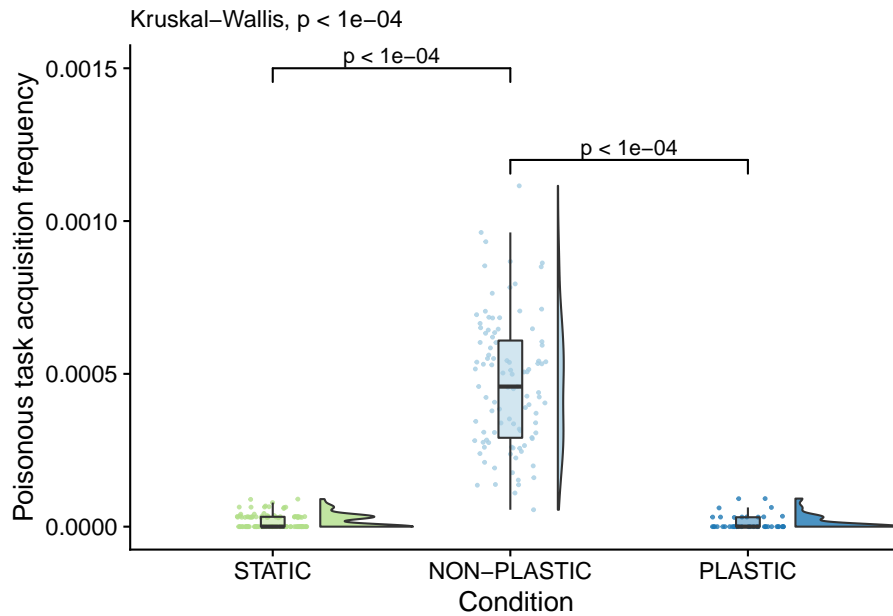
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=dominant_lineage_num_times_hitchhike_inst_ex
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
)

```

```

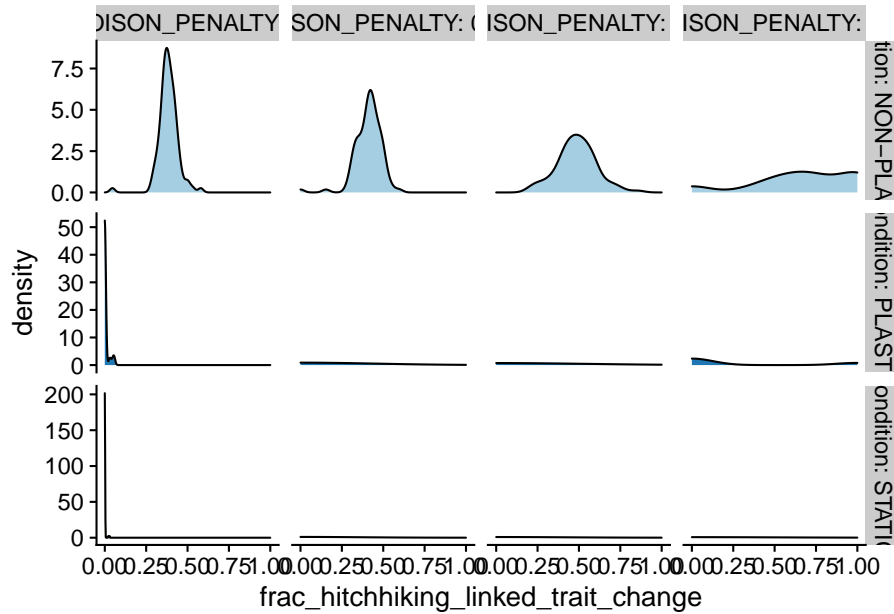
## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
poison_increases_per_gen_fig

```



5.6.3 What fraction of mutations that increase poison instruction execution co-occur with base trait changes?

```
ggplot(filter(summary_data, dominant_lineage_num_times_hitchhike_inst_exec_increases>0), aes(x=fr
  geom_density() +
  facet_grid(
    condition~POISON_PENALTY,
    labeller=label_both,
    scales="free_y"
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  ) +
  theme(
    legend.position="none"
  ) +
  ggsave(
    paste0(working_directory, "plots/dominant-lineage-frac_hitchhiking_linked_trait_change.png"),
    width=15,
    height=10
  )
```

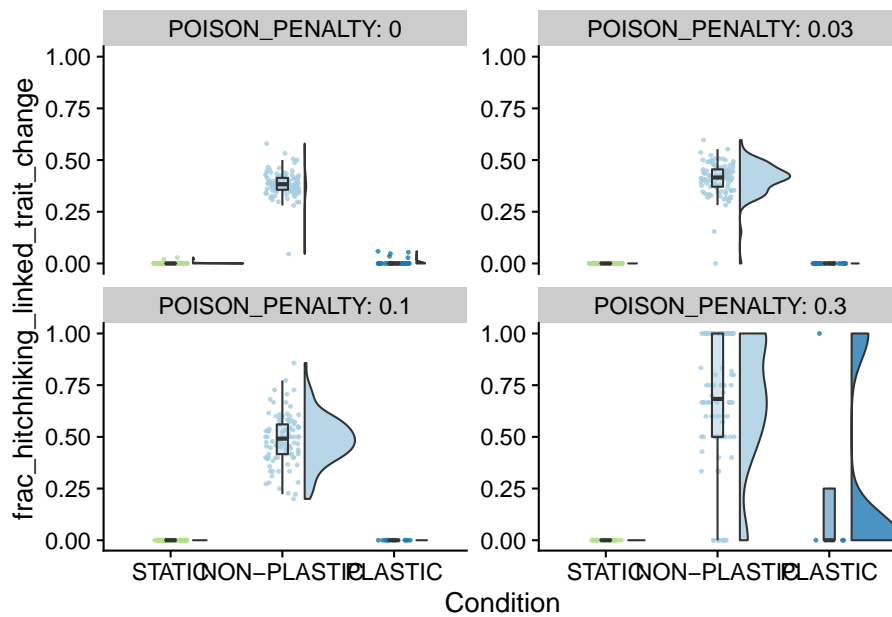


```
ggplot(filter(summary_data, dominant_lineage_num_times_hitchhike_inst_exec_increases>0)
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette=cb_palette
  ) +
  scale_color_brewer(
    palette=cb_palette
  )
```

```

) +
facet_wrap(
  ~POISON_PENALTY,
  labeller=label_both,
  scales="free_y"
) +
# coord_flip() +
theme(
  legend.position="none"
)

```



```

for (penalty in poison_penalties) {
  stat_data <- filter(summary_data, POISON_PENALTY==penalty & dominant_lineage_num_times_hitchhik
  print(
    paste0(
      "PENALTY: ", penalty
    )
  )
  kt <- kruskal.test(
    formula=frac_hitchhiking_linked_trait_change~condition,
    data=stat_data
  )
  print(
    kt
  )
}

```

```

if (is.na(kt$p.value)) { next }
if (kt$p.value > 0.05) { next }
print(
  pairwise.wilcox.test(
    x=stat_data$frac_hitchhiking_linked_trait_change,
    g=stat_data$condition,
    p.adjust.method="bonferroni",
    exact=FALSE
  )
)
}

```

```

## [1] "PENALTY: 0"
##
##   Kruskal-Wallis rank sum test
##
## data:  frac_hitchhiking_linked_trait_change by condition
## Kruskal-Wallis chi-squared = 211.29, df = 2, p-value < 2.2e-16
##
##
##   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  stat_data$frac_hitchhiking_linked_trait_change and stat_data$condition
##
##           NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      0.031
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.03"
##
##   Kruskal-Wallis rank sum test
##
## data:  frac_hitchhiking_linked_trait_change by condition
## Kruskal-Wallis chi-squared = 186.88, df = 2, p-value < 2.2e-16
##
##
##   Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  stat_data$frac_hitchhiking_linked_trait_change and stat_data$condition
##
##           NON-PLASTIC PLASTIC
## PLASTIC 2.9e-16      -
## STATIC  < 2e-16      -
##

```

5.6. CHARACTERIZING MUTATIONS THAT INCREASE POISON INSTRUCTION EXECUTION133

```
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.1"
##
## Kruskal-Wallis rank sum test
##
## data: frac_hitchhiking_linked_trait_change by condition
## Kruskal-Wallis chi-squared = 113.72, df = 2, p-value < 2.2e-16
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$frac_hitchhiking_linked_trait_change and stat_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC 3.3e-08      -
## STATIC  < 2e-16      -
##
## P value adjustment method: bonferroni
## [1] "PENALTY: 0.3"
##
## Kruskal-Wallis rank sum test
##
## data: frac_hitchhiking_linked_trait_change by condition
## Kruskal-Wallis chi-squared = 34.791, df = 2, p-value = 2.788e-08
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: stat_data$frac_hitchhiking_linked_trait_change and stat_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC 0.26      -
## STATIC  2.4e-08    0.18
##
## P value adjustment method: bonferroni
denom <- sum(filter(summary_data, condition=="NON-PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num)
num <- sum(filter(summary_data, condition=="NON-PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num)
paste0("NON-PLASTIC (0.1 penalty): ", num/denom, " (", num, "/", denom, ")")

## [1] "NON-PLASTIC (0.1 penalty): 0.498956158663883(956/1916)"
denom <- sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num)
num <- sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num)
paste0("PLASTIC (0.1 penalty): ", num/denom, " (", num, "/", denom, ")")

## [1] "PLASTIC (0.1 penalty): 0 (0/18)"
```

```
denom <- sum(filter(summary_data, condition=="STATIC" & POISON_PENALTY==0.1)$dominant_lineage_num_times_hitchhike_inst_exec_increases)
num <- sum(filter(summary_data, condition=="STATIC" & POISON_PENALTY==0.1)$dominant_lineage_num_times_hitchhike_inst_exec_increases)
paste0("STATIC (0.1 penalty): ", num/denom, " (", num, "/", denom, ")")
```

```
## [1] "STATIC (0.1 penalty): 0 (0/58)"
```

Focal figure for the manuscript:

```
# Compute manual labels for geom_signif
stat.test <- filter(focal_summary_data, dominant_lineage_num_times_hitchhike_inst_exec_increases)
wilcox_test(frac_hitchhiking_linked_trait_change ~ condition, comparisons=list(c("PL", "D"))
adjust_pvalue(method = "bonferroni") %>%
add_significance() %>%
add_xy_position(x="condition")
# Tweak y.position manually to account for scaled axis (edge case that triggers bad behavior)
stat.test$manual_position <- stat.test$y.position
stat.test$label <- mapply(p_label, stat.test$p.adj)

linked_trait_change_fig <- ggplot(
  filter(focal_summary_data, dominant_lineage_num_times_hitchhike_inst_exec_increases)
  aes(x=condition, y=frac_hitchhiking_linked_trait_change, fill=condition)
) +
geom_flat_violin(
  position = position_nudge(x = .2, y = 0),
  alpha = .8
) +
geom_point(
  mapping=aes(color=condition),
  position = position_jitter(width = .15),
  size = .5,
  alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order,
  labels=condition_order
) +
scale_y_continuous(
  name="Fraction of linked poisonous task acquisition",
  limits=c(-0.01, 1.2),
  breaks=c(0, 0.25, 0.50, 0.75, 1.0)
```

```

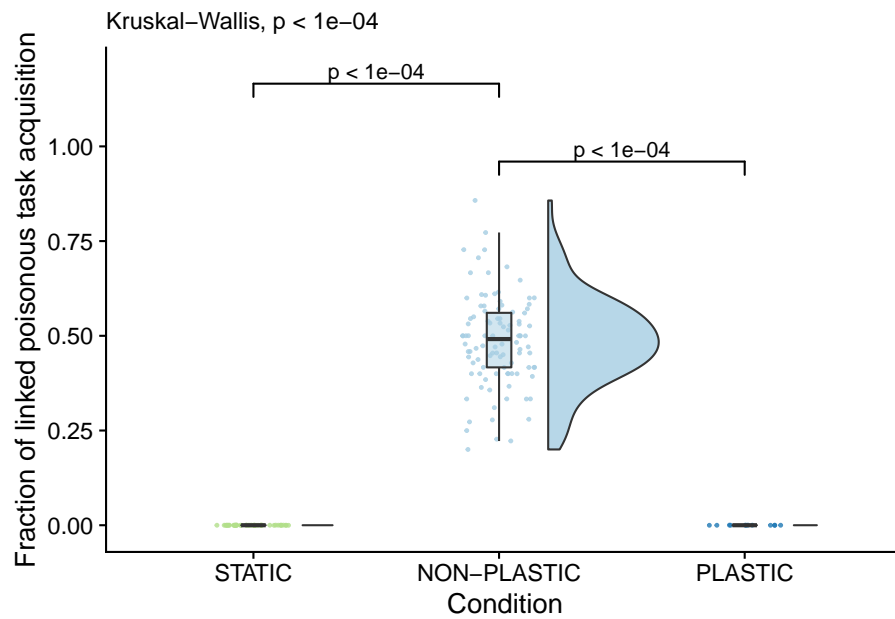
) +
scale_fill_brewer(
  palette=cb_palette
) +
scale_color_brewer(
  palette=cb_palette
) +
labs(
  subtitle=paste0(
    "Kruskal-Wallis, ",
    p_label(signif(kruskal.test(formula=frac_hitchhiking_linked_trait_change~condition, data=f
  )
) +
ggsignif::geom_signif(
  data=filter(stat.test, p.adj <= alpha),
  aes(xmin=group1,xmax=group2,annotations=label,y_position=manual_position),
  manual=TRUE,
  inherit.aes=FALSE
) +
theme(
  legend.position="none"
)

```

```

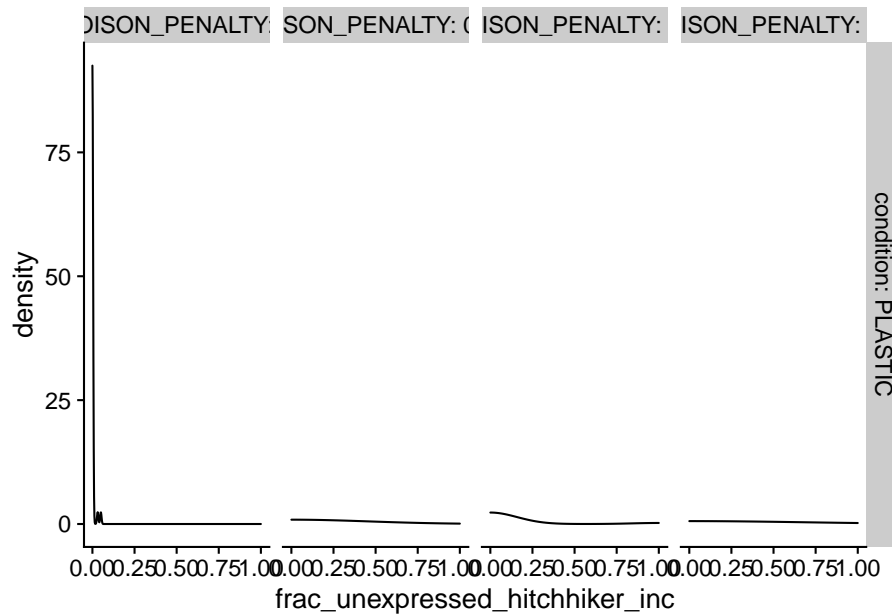
## Warning: Ignoring unknown aesthetics: xmin, xmax, annotations, y_position
linked_trait_change_fig

```



5.7 What fraction of poison execution increases

```
ggplot(filter(summary_data, dominant_lineage_num_times_hitchhike_inst_exec_increases > 0)) +
  geom_density() +
  facet_grid(
    condition ~ POISON_PENALTY,
    labeller = label_both,
    scales = "free_y"
  ) +
  theme(
    legend.position = "none"
  )
```

```
denom <- sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num)
num <- sum(filter(summary_data, condition=="PLASTIC" & POISON_PENALTY==0.1)$dominant_lineage_num)
paste0("PLASTIC: ", num/denom, " (", num, "/", denom, ")")
```

```
## [1] "PLASTIC: 0.0555555555555556 (1/18)"
```

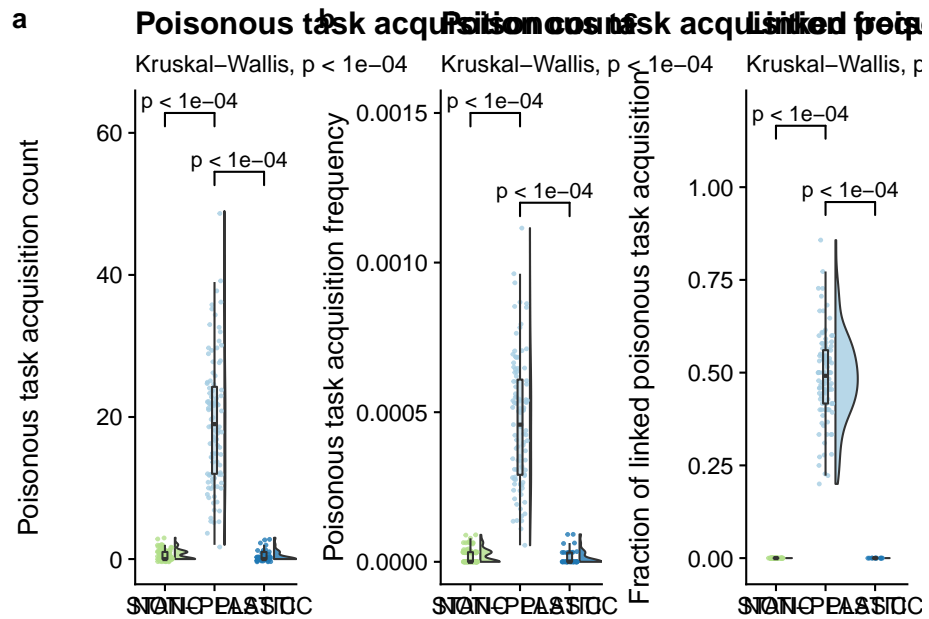
5.8 Manuscript figures

```
grid <- plot_grid(
  poison_increases_fig +
    theme(
      axis.title.x=element_blank()
    ) +
    ggtitle("Poisonous task acquisition count"),
  poison_increases_per_gen_fig +
    theme(
      axis.title.x=element_blank()
    ) +
    ggtitle("Poisonous task acquisition frequency"),
  linked_trait_change_fig +
    theme(
      axis.title.x=element_blank()
    ) +
    ggtitle("Linked poisonous task acquisition"),
```

```

nrow=1,
align="v",
labels="auto"
)
save_plot(
  paste0(working_directory, "plots/", "poison-accumulation-panel.pdf"),
  grid,
  base_height=6,
  base_asp=3/1
)
grid

```



Chapter 6

Regulation in Avida

6.1 Overview

```
total_updates <- 200000
replicates <- 100

all_traits <- c("not","nand","and","ornot","or","andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-02-08-evo-dynamics/analysis/" # << For bookdown
# working_directory <- "./" # << For local analysis
```

6.2 Analysis dependencies

Load all required R libraries.

```
library(ggplot2)
library(tidyverse)
library(cowplot)
library(RColorBrewer)
library(Hmisc)
library(boot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd97121f7f9ce9")
```

These analyses were conducted/knitted with the following computing environment:

```
print(version)

##
## platform      x86_64-pc-linux-gnu
## arch          x86_64
## os            linux-gnu
## system        x86_64, linux-gnu
## status
## major         4
## minor         0.5
## year          2021
## month         03
## day           31
## svn rev       80133
## language      R
## version.string R version 4.0.5 (2021-03-31)
## nickname      Shake and Throw
```

6.3 Setup

```
trace_summary_data_loc <- paste0(working_directory, "data/trace_summary.csv")
trace_summary_data <- read.csv(trace_summary_data_loc, na.strings="NONE")

trace_summary_data$DISABLE_REACTION_SENSORS <- as.factor(trace_summary_data$DISABLE_RE
trace_summary_data$chg_env <- trace_summary_data$chg_env == "True"
trace_summary_data$sensors <- trace_summary_data$DISABLE_REACTION_SENSORS == "0"

env_label_fun <- function(chg_env) {
  if (chg_env) {
    return("Fluctuating")
  } else {
    return("Constant")
  }
}

sensors_label_fun <- function(has_sensors) {
  if (has_sensors) {
    return("Sensors")
  } else {
    return("No sensors")
  }
}
```

6.4. HOW MANY INSTRUCTIONS DO PLASTIC GENOMES TOGGLE DEPENDING ON ENVIRONMENTAL C

```
# note that this labeler makes assumptions about how we set up our experiment
condition_label_fun <- function(has_sensors, env_chg) {
  if (has_sensors && env_chg) {
    return("PLASTIC")
  } else if (env_chg) {
    return("NON-PLASTIC")
  } else {
    return("STATIC")
  }
}

trace_summary_data$env_label <- mapply(
  env_label_fun,
  trace_summary_data$chg_env
)
trace_summary_data$sensors_label <- mapply(
  sensors_label_fun,
  trace_summary_data$sensors
)
trace_summary_data$condition <- mapply(
  condition_label_fun,
  trace_summary_data$sensors,
  trace_summary_data$chg_env
)

##### misc #####
# Configure our default graphing theme
theme_set(theme_cowplot())
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)
```

6.4 How many instructions do plastic genomes toggle depending on environmental context?

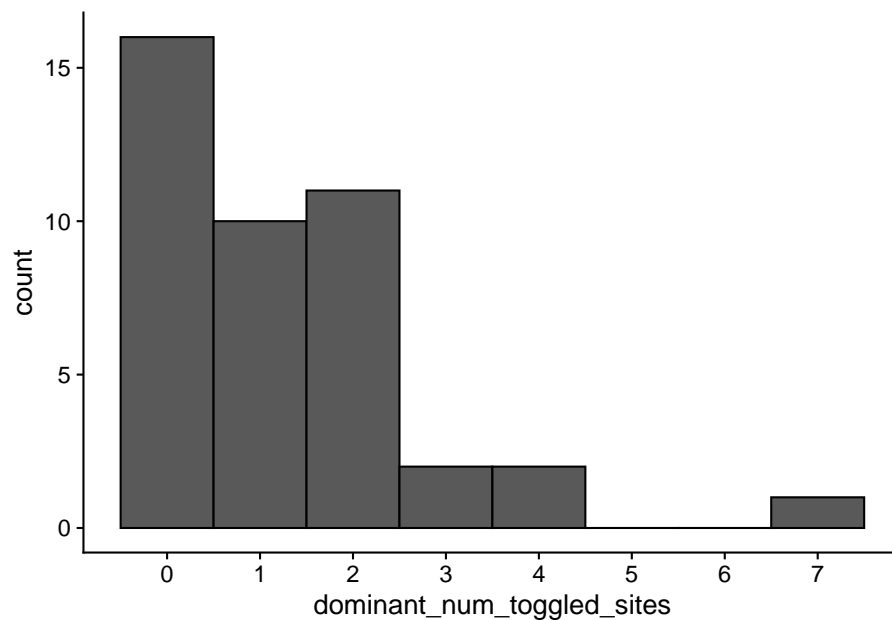
```
ggplot(trace_summary_data, aes(x=dominant_num_toggled_sites)) +
  geom_histogram(
    binwidth=1,
    color="black"
  ) +
  scale_fill_brewer(
    palette="Paired"
  ) +
  scale_color_brewer(
    palette="Paired"
```

```

) +
scale_x_continuous(
  breaks=seq(0, max(trace_summary_data$dominant_num_toggled_sites)+1)
) +
theme(
  legend.position="none"
) +
ggsave(paste0(working_directory, "plots/", "toggled-sites.png"))

## Saving 6.5 x 4.5 in image

```



6.5 What is the distribution of toggled sequence sizes?

```

chunk_sizes <- data.frame(
  size=integer()
)
for (sizes in trace_summary_data$dominant_toggled_chunk_sizes) {
  if (sizes == "") { next }
  sizes <- unlist(lapply(str_split(sizes, ';'), as.integer))
  chunk_sizes <- rbind(chunk_sizes, data.frame(size=c(sizes)))
}

```

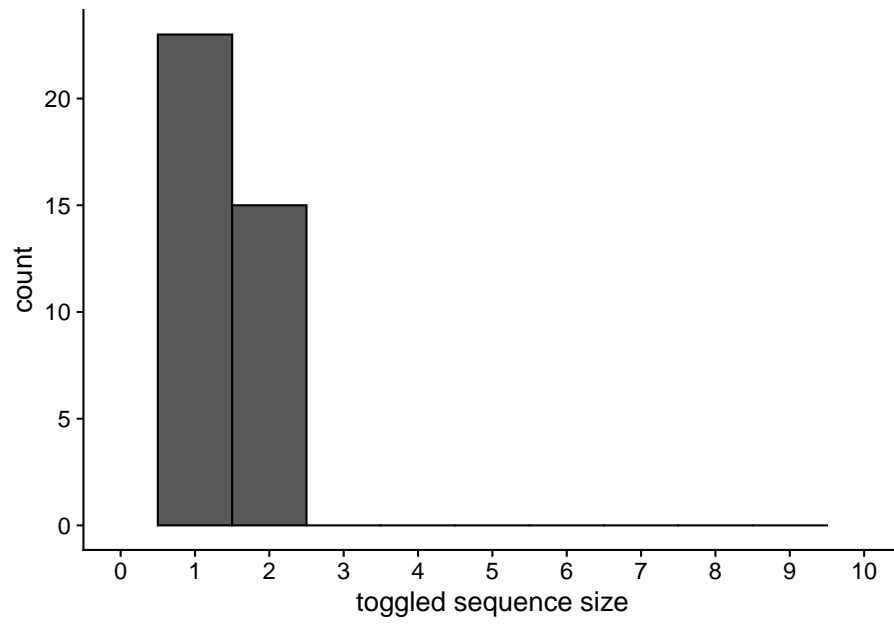
6.5. WHAT IS THE DISTRIBUTION OF TOGGLED SEQUENCE SIZES?143

```
ggplot(chunk_sizes, aes(x=size)) +  
  geom_histogram(  
    binwidth=1,  
    color="black"  
  ) +  
  scale_fill_brewer(  
    palette="Paired"  
  ) +  
  scale_color_brewer(  
    palette="Paired"  
  ) +  
  scale_x_continuous(  
    name="toggled sequence size",  
    breaks=seq(0, 10),  
    limits=c(0, 10)  
  ) +  
  theme(  
    legend.position="none"  
  ) +  
  ggsave(paste0(working_directory, "plots/", "toggled-chunk-sizes.png"))
```

```
## Saving 6.5 x 4.5 in image
```

```
## Warning: Removed 2 rows containing missing values (geom_bar).
```

```
## Warning: Removed 2 rows containing missing values (geom_bar).
```



Chapter 7

Evolutionary change (variable length genomes)

7.1 Overview

```
total_updates <- 200000
replicates <- 100

all_traits <- c("not","nand","and","ornot","or","andnot")
traits_set_a <- c("not", "and", "or")
traits_set_b <- c("nand", "ornot", "andnot")

# Relative location of data.
working_directory <- "experiments/2021-01-30-evo-dynamics/analysis/" # << For bookdown
# working_directory <- "./" # << For local analysis
```

7.2 Analysis dependencies

Load all required R libraries.

```
library(ggplot2)
library(tidyverse)
library(cowplot)
library(RColorBrewer)
library(Hmisc)
library(boot)
source("https://gist.githubusercontent.com/benmarwick/2a1bb0133ff568cbe28d/raw/fb53bd97121f7f9ce9")
```

These analyses were conducted/knitted with the following computing environ-

ment:

```
print(version)

##
## platform      x86_64-pc-linux-gnu
## arch          x86_64
## os            linux-gnu
## system        x86_64, linux-gnu
## status
## major         4
## minor         0.5
## year          2021
## month         03
## day           31
## svn rev       80133
## language      R
## version.string R version 4.0.5 (2021-03-31)
## nickname      Shake and Throw
```

7.3 Setup

```
summary_data_loc <- paste0(working_directory, "data/aggregate.csv")
summary_data <- read.csv(summary_data_loc, na.strings="NONE")

summary_data$DISABLE_REACTION_SENSORS <- as.factor(summary_data$DISABLE_REACTION_SENSORS)
summary_data$chg_env <- summary_data$chg_env == "True"
summary_data$dominant_plastic_odd_even <- as.factor(summary_data$dominant_plastic_odd_even)
summary_data$sensors <- summary_data$DISABLE_REACTION_SENSORS == "0"
summary_data$is_plastic <- summary_data$dominant_plastic_odd_even == "True"

env_label_fun <- function(chg_env) {
  if (chg_env) {
    return("Fluctuating")
  } else {
    return("Constant")
  }
}

sensors_label_fun <- function(has_sensors) {
  if (has_sensors) {
    return("Sensors")
  } else {
    return("No sensors")
  }
}
```

```

}

# note that this labeler makes assumptions about how we set up our experiment
condition_label_fun <- function(has_sensors, env_chg) {
  if (has_sensors && env_chg) {
    return("PLASTIC")
  } else if (env_chg) {
    return("NON-PLASTIC")
  } else {
    return("STATIC")
  }
}

summary_data$env_label <- mapply(
  env_label_fun,
  summary_data$chg_env
)
summary_data$sensors_label <- mapply(
  sensors_label_fun,
  summary_data$sensors
)
summary_data$condition <- mapply(
  condition_label_fun,
  summary_data$sensors,
  summary_data$chg_env
)

condition_order = c(
  "STATIC",
  "NON-PLASTIC",
  "PLASTIC"
)

##### misc #####
# Configure our default graphing theme
theme_set(theme_cowplot())
dir.create(paste0(working_directory, "plots"), showWarnings=FALSE)

```

7.4 Evolution of phenotypic plasticity

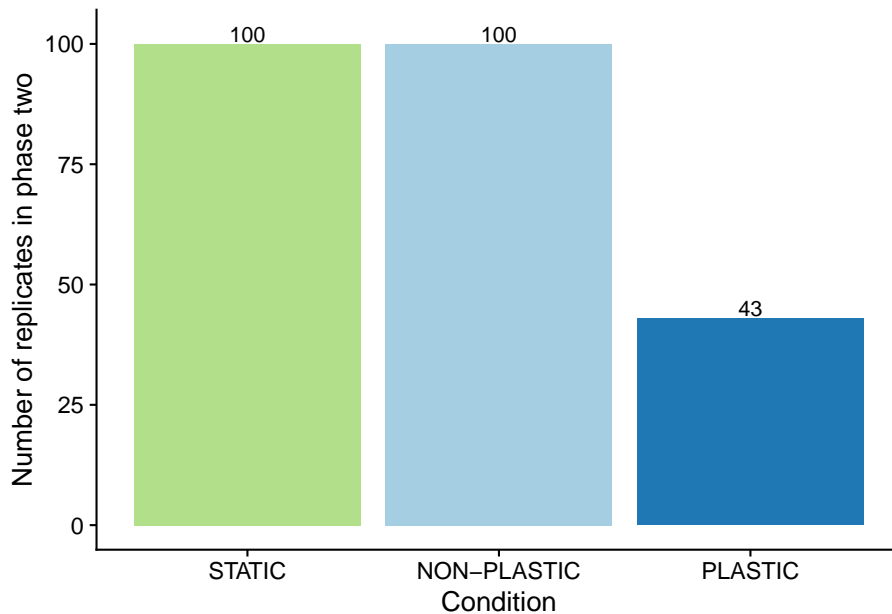
For sensor-enabled populations in fluctuating environments, we only transferred populations containing an optimally plastic genotype to phase-two.

```

summary_data_grouped = dplyr::group_by(summary_data, sensors, env_label, condition)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())

```

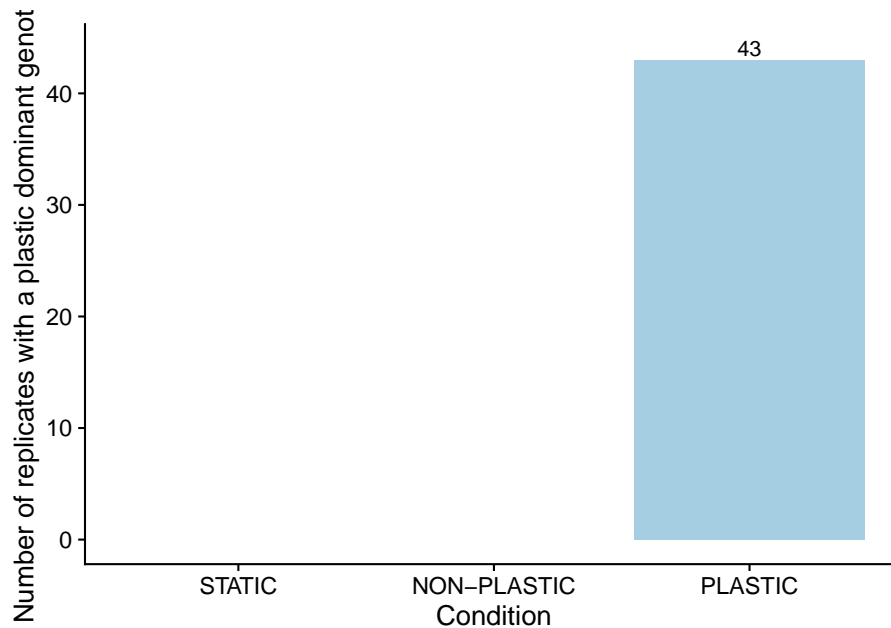
```
## `summarise()` has grouped output by 'sensors', 'env_label'. You can override using `ungroup()`
ggplot(summary_data_group_counts, aes(x=condition, y=n, fill=condition)) +
  geom_col(position=position_dodge(0.9)) +
  geom_text(aes(label=n, y=n+2)) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette="Paired"
  ) +
  scale_color_brewer(
    palette="Paired"
  ) +
  ylab("Number of replicates in phase two") +
  theme(
    legend.position="none"
  )
```



We can confirm our expectation that the dominant genotypes in non-plastic conditions are not phenotypically plastic.

```
summary_data_grouped = dplyr::group_by(summary_data, condition, is_plastic)
summary_data_group_counts = dplyr::summarize(summary_data_grouped, n=dplyr::n())
```

```
ggplot(filter(summary_data_group_counts, is_plastic), aes(x=condition, y=n, fill=condition)) +
  geom_col(position=position_dodge(0.9)) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette="Paired"
  ) +
  scale_color_brewer(
    palette="Paired"
  ) +
  geom_text(aes(label=n, y=n+1)) +
  ylab("Number of replicates with a plastic dominant genotype") +
  theme(
    legend.position="none"
  )
)
```



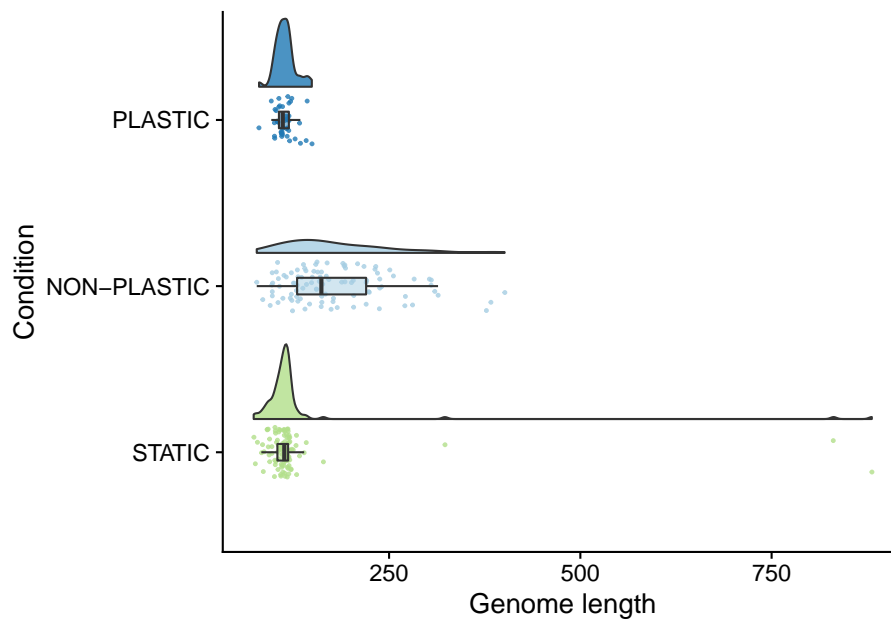
7.5 Genome length

Single-instruction insertions and deletions were possible for this experiment, so genome size also evolved.

```

ggplot(summary_data, aes(x=condition, y=dominant_genome_length, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette="Paired"
  ) +
  scale_color_brewer(
    palette="Paired"
  ) +
  coord_flip() +
  ylab("Genome length") +
  theme(
    legend.position="none"
  )

```



```
kruskal.test(
  formula=dominant_genome_length~condition,
  data=summary_data
)
```

```
##
## Kruskal-Wallis rank sum test
##
## data:  dominant_genome_length by condition
## Kruskal-Wallis chi-squared = 82.798, df = 2, p-value < 2.2e-16
```

```
pairwise.wilcox.test(
  x=summary_data$dominant_genome_length,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)
```

```
##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  summary_data$dominant_genome_length and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC 1.8e-10      -
## STATIC  < 2e-16      1
##
```

```
## P value adjustment method: bonferroni
median(filter(summary_data, condition=="PLASTIC")$phylo_mrca_changes)

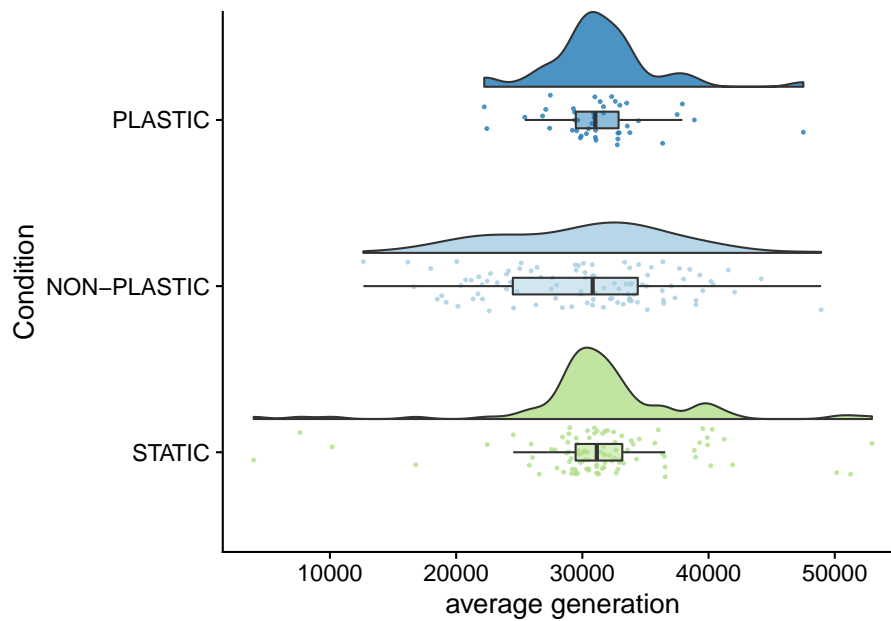
## [1] 45
median(filter(summary_data, condition=="STATIC")$phylo_mrca_changes)

## [1] 47
median(filter(summary_data, condition=="NON-PLASTIC")$phylo_mrca_changes)

## [1] 393
```

7.6 Average generation

```
ggplot(summary_data, aes(x=condition, y=time_average_generation, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  scale_fill_brewer(
    palette="Paired"
  ) +
  scale_color_brewer(
    palette="Paired"
  ) +
  coord_flip() +
  ylab("average generation") +
  theme(
    legend.position="none"
  )
```

```

median(filter(summary_data, condition=="PLASTIC")$time_average_generation)

## [1] 31028.6
median(filter(summary_data, condition=="STATIC")$time_average_generation)

## [1] 31147.5
median(filter(summary_data, condition=="NON-PLASTIC")$time_average_generation)

## [1] 30817.95
kruskal.test(
  formula=time_average_generation~condition,
  data=summary_data
)

##
## Kruskal-Wallis rank sum test
##
## data: time_average_generation by condition
## Kruskal-Wallis chi-squared = 1.3804, df = 2, p-value = 0.5015

```

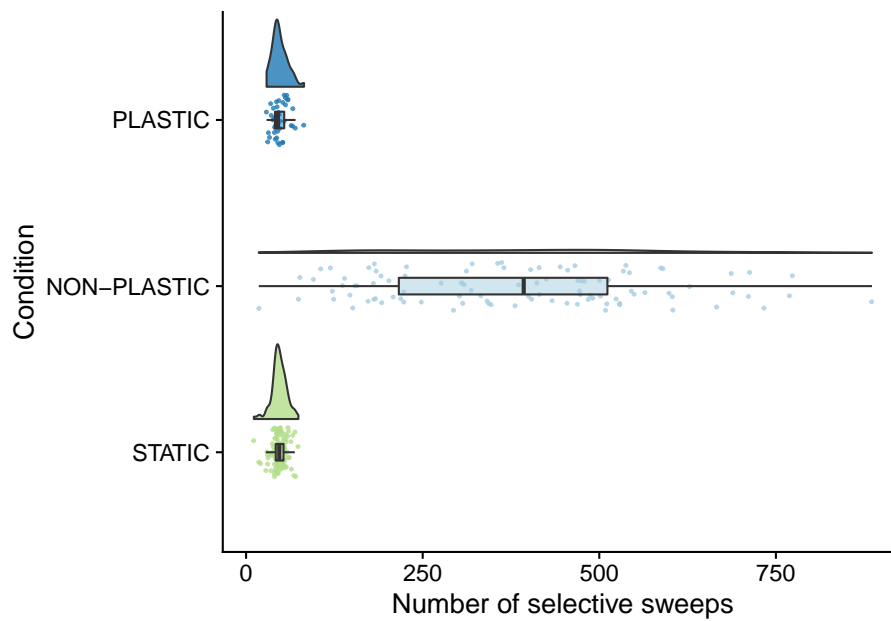
7.7 Coalescence events

The number of times the most recent common ancestor changes gives us the number of selective sweeps that occur during the experiment.

```

ggplot(summary_data, aes(x=condition, y=phylo_mrca_changes, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_fill_brewer(
    palette="Paired"
  ) +
  scale_color_brewer(
    palette="Paired"
  ) +
  coord_flip() +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
  ylab("Number of selective sweeps") +
  theme(
    legend.position="none"
  )

```



```

paste0(
  "PLASTIC: ",
  median(filter(summary_data, condition=="PLASTIC")$phylo_mrca_changes)
)

## [1] "PLASTIC: 45"

paste0(
  "STATIC: ",
  median(filter(summary_data, condition=="STATIC")$phylo_mrca_changes)
)

## [1] "STATIC: 47"

paste0(
  "NON-PLASTIC: ",
  median(filter(summary_data, condition=="NON-PLASTIC")$phylo_mrca_changes)
)

## [1] "NON-PLASTIC: 393"

kruskal.test(
  formula=phylo_mrca_changes~condition,
  data=summary_data
)

##
## Kruskal-Wallis rank sum test

```

```
##
## data: phylo_mrca_changes by condition
## Kruskal-Wallis chi-squared = 168.89, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$phylo_mrca_changes,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$phylo_mrca_changes and summary_data$condition
##
##          NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      1
##
## P value adjustment method: bonferroni
```

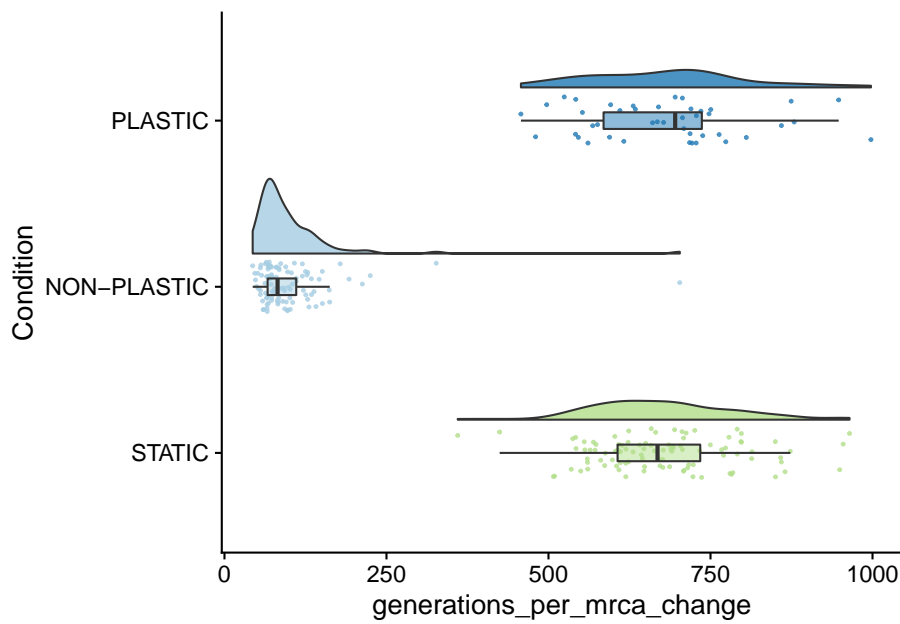
7.7.1 Average number of generations between selective sweeps

```
summary_data$generations_per_mrca_change <- summary_data$time_average_generation / sum
ggplot(summary_data, aes(x=condition, y=generations_per_mrca_change, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
```

```

scale_fill_brewer(
  palette="Paired"
) +
scale_color_brewer(
  palette="Paired"
) +
coord_flip() +
theme(
  legend.position="none"
)

```



```

paste0(
  "PLASTIC: ",
  median(filter(summary_data, condition=="PLASTIC"))$generations_per_mrca_change)
)

```

```
## [1] "PLASTIC: 695.504761904762"
```

```

paste0(
  "STATIC: ",
  median(filter(summary_data, condition=="STATIC"))$generations_per_mrca_change)
)

```

```
## [1] "STATIC: 668.25523255814"
```

```

paste0(
  "NON-PLASTIC: ",
  median(filter(summary_data, condition=="NON-PLASTIC")$generations_per_mrca_change)
)

## [1] "NON-PLASTIC: 81.9208459944751"

kruskal.test(
  formula=generations_per_mrca_change~condition,
  data=summary_data
)

##
## Kruskal-Wallis rank sum test
##
## data: generations_per_mrca_change by condition
## Kruskal-Wallis chi-squared = 171.73, df = 2, p-value < 2.2e-16

pairwise.wilcox.test(
  x=summary_data$generations_per_mrca_change,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$generations_per_mrca_change and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      1
##
## P value adjustment method: bonferroni

```

7.8 Phenotypic volatility along the dominant lineage

```

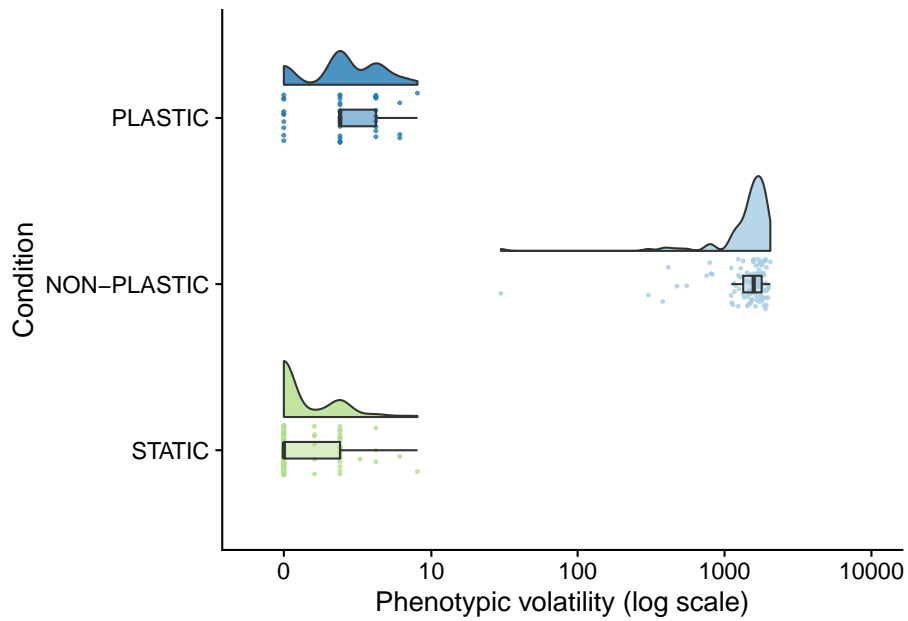
ggplot(summary_data, aes(x=condition, y=dominant_lineage_trait_volatility, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),

```

```

    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
) +
geom_boxplot(
  width = .1,
  outlier.shape = NA,
  alpha = 0.5
) +
scale_x_discrete(
  name="Condition",
  limits=condition_order
) +
scale_y_continuous(
  name="Phenotypic volatility (log scale)",
  trans="pseudo_log",
  breaks=c(0, 10, 100, 1000, 10000),
  limits=c(-1,10000)
) +
scale_fill_brewer(
  palette="Paired"
) +
scale_color_brewer(
  palette="Paired"
) +
coord_flip() +
theme(
  legend.position="none"
)

```



```
paste0(
  "PLASTIC: ",
  median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_trait_volatility)
)
```

```
## [1] "PLASTIC: 2"
```

```
paste0(
  "STATIC: ",
  median(filter(summary_data, condition=="STATIC"))$dominant_lineage_trait_volatility)
)
```

```
## [1] "STATIC: 0"
```

```
paste0(
  "NON-PLASTIC: ",
  median(filter(summary_data, condition=="NON-PLASTIC"))$dominant_lineage_trait_volatility)
)
```

```
## [1] "NON-PLASTIC: 1580"
```

```
kruskal.test(
  formula=dominant_lineage_trait_volatility~condition,
  data=summary_data
)
```

```
##
```

```
## Kruskal-Wallis rank sum test
```


7.9. MUTATION ACCUMULATION ALONG THE DOMINANT LINEAGE161

```
##
## data: dominant_lineage_trait_volatility by condition
## Kruskal-Wallis chi-squared = 191.98, df = 2, p-value < 2.2e-16
pairwise.wilcox.test(
  x=summary_data$dominant_lineage_trait_volatility,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data: summary_data$dominant_lineage_trait_volatility and summary_data$condition
##
##          NON-PLASTIC PLASTIC
## PLASTIC < 2e-16      -
## STATIC  < 2e-16      5.2e-08
##
## P value adjustment method: bonferroni
```

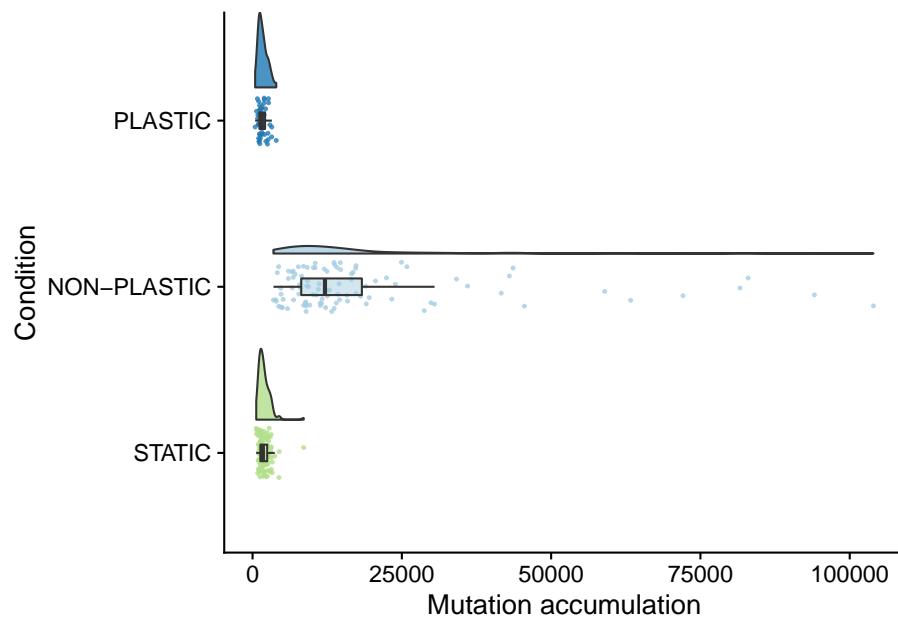
7.9 Mutation accumulation along the dominant lineage

```
ggplot(summary_data, aes(x=condition, y=dominant_lineage_total_mut_cnt, fill=condition)) +
  geom_flat_violin(
    position = position_nudge(x = .2, y = 0),
    alpha = .8
  ) +
  geom_point(
    mapping=aes(color=condition),
    position = position_jitter(width = .15),
    size = .5,
    alpha = 0.8
  ) +
  geom_boxplot(
    width = .1,
    outlier.shape = NA,
    alpha = 0.5
  ) +
  ylab("Mutation accumulation") +
  scale_x_discrete(
    name="Condition",
    limits=condition_order
  ) +
```

```

scale_fill_brewer(
  palette="Paired"
) +
scale_color_brewer(
  palette="Paired"
) +
coord_flip() +
theme(
  legend.position="none"
)

```



```

paste0(
  "PLASTIC: ",
  median(filter(summary_data, condition=="PLASTIC"))$dominant_lineage_total_mut_cnt)
)

```

```
## [1] "PLASTIC: 1552"
```

```

paste0(
  "STATIC: ",
  median(filter(summary_data, condition=="STATIC"))$dominant_lineage_total_mut_cnt)
)

```

```
## [1] "STATIC: 1724.5"
```

7.9. MUTATION ACCUMULATION ALONG THE DOMINANT LINEAGE163

```
paste0(
  "NON-PLASTIC: ",
  median(filter(summary_data, condition=="NON-PLASTIC")$dominant_lineage_total_mut_cnt)
)

## [1] "NON-PLASTIC: 12123"

kruskal.test(
  formula=dominant_lineage_total_mut_cnt~condition,
  data=summary_data
)

##
## Kruskal-Wallis rank sum test
##
## data:  dominant_lineage_total_mut_cnt by condition
## Kruskal-Wallis chi-squared = 174.38, df = 2, p-value < 2.2e-16

pairwise.wilcox.test(
  x=summary_data$dominant_lineage_total_mut_cnt,
  g=summary_data$condition,
  p.adjust.method="bonferroni",
)

##
## Pairwise comparisons using Wilcoxon rank sum test with continuity correction
##
## data:  summary_data$dominant_lineage_total_mut_cnt and summary_data$condition
##
##      NON-PLASTIC PLASTIC
## PLASTIC <2e-16      -
## STATIC  <2e-16      0.57
##
## P value adjustment method: bonferroni
```