

# Package ‘isolateR’

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**Type** Package

**Title** Automated processing of Sanger sequencing data, taxonomic profiling, and generation of microbial strain libraries

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**URL** <https://github.com/bdaisley/isolateR>

**BugReports** <https://github.com/bdaisley/isolateR/issues>

**Description** isolateR aims to enhance microbial isolation workflows and support the identification of novel taxa. It addresses the challenges of manual Sanger sequencing data processing and limitations of conventional BLAST searches, crucial for identifying microorganisms and creating strain libraries. The package offers a streamlined three-step process that automates quality trimming Sanger sequence files, taxonomic classification via global alignment against type strain databases, and efficient strain library creation based on customizable sequence similarity thresholds. It features interactive HTML output tables for easy data exploration and optional tools for generating phylogenetic trees to visualize microbial diversity.

**Citation**

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**VignetteBuilder** knitr**Additional\_repositories**

<http://R-Forge.R-project.org> <https://bioconductor.org/packages/3.18/bioc>

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class-isoLIB

*isoLIB Class Object***Description**

S4 wrapper for [isoLIB](#) function. Access data via S4 slot functions.

**Value**

Returns an class-isoLIB object.

**Slots**

`input` Character string containing input directory information.

`sequence_group` Character string containing list of group representative filenames.

`date` Character string containing run date from each of the input Sanger sequence .ab1 files ("YYYY\_MM\_DD" format).

`filename` Character string containing input filenames.

`phred_trim` Numeric string containing mean Phred scores after trimming.

`Ns_trim` Numeric string containing count of N's after trimming.

`length_trim` Numeric string containing sequence length after trimming.

`seqs_trim` Character string containing sequence after trimming.

`closest_match` Character string containing species + type strain no. of closest match from reference database.

`NCBI_acc` Character string containing NCBI accession number associated with closest match from reference database.

`ID` Numeric string containing pairwise similarity value for query vs database reference sequence. Calculation of ID is determined by isoTAX 'iddef' parameter (0-4, Default=2). See VSEARCH documentation for more details.

- (0) CD-HIT definition: (matching columns) / (shortest sequence length).
- (1) Edit distance: (matching columns) / (alignment length).
- (2) Edit distance excluding terminal gaps (default definition).
- (3) Marine Biological Lab definition counting each gap opening (internal or terminal) as a single mismatch, whether or not the gap was extended:  $1.0 - ((\text{mismatches} + \text{gap openings}) / (\text{longest sequence length}))$ .
- (4) BLAST definition, equivalent to --iddef 1 for global pairwise alignments.

`rank_phylum` Character string containing Phylum rank taxonomy

`rank_class` Character string containing Class rank taxonomy

`rank_order` Character string containing Order rank taxonomy

`rank_family` Character string containing Family rank taxonomy

`rank_genus` Character string containing Genus rank taxonomy

`rank_species` Character string containing Species rank taxonomy

`phylum_threshold` Numeric string containing Phylum-level sequence similarity threshold for rank demarcation

`class_threshold` Numeric string containing Class-level sequence similarity threshold for rank demarcation

`order_threshold` Numeric string containing Order-level sequence similarity threshold for rank demarcation

`family_threshold` Numeric string containing Family-level sequence similarity threshold for rank demarcation

`genus_threshold` Numeric string containing Genus-level sequence similarity threshold for rank demarcation

`species_threshold` Numeric string containing Species-level sequence similarity threshold for rank demarcation

**See Also**

[isoLIB](#)

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class-isoQC	<i>isoQC Class Object</i>
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## Description

S4 wrapper for [isoQC](#) function. Access data via S4 slot functions.

## Value

Returns an class-isoQC object.

## Slots

**date** Character string containing run date from each of the input Sanger sequence .ab1 files ("YYYY\_MM\_DD" format).

**filename** Character string containing input filenames.

**trim.start.pos** Numeric string containing trimming position start point.

**trim.end.pos** Numeric string containing trimming position end point.

**phred\_spark\_raw** List containing per nucleotide Phred score values for each sequence

**phred\_raw** Numeric string containing mean Phred scores before trimming.

**phred\_trim** Numeric string containing mean Phred scores after trimming.

**Ns\_raw** Numeric string containing count of N's before trimming.

**Ns\_trim** Numeric string containing count of N's after trimming.

**length\_raw** Numeric string containing sequence length before trimming.

**length\_trim** Numeric string containing sequence length after trimming.

**seqs\_raw** Character string containing sequences before trimming.

**seqs\_trim** Character string containing sequence after trimming.

**decision** Character string containing decision (PASS/FAIL) information based on [isoQC](#) 'min\_phred\_score' and 'min\_length\_cutoffs'.

**input** Character string containing input directory information.

## See Also

[isoQC](#)

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class-isoTAX	<i>isoTAX Class Object</i>
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## Description

S4 wrapper for [isoTAX](#) function. Access data via S4 slot functions.

## Value

Returns an class-isoTAX object.

## Slots

`input` Character string containing input directory information.

`warning` Character string containing list filenames of sequences that had poor alignment during taxonomic classification step.

`date` Character string containing run date from each of the input Sanger sequence .ab1 files ("YYYY\_MM\_DD" format).

`filename` Character string containing input filenames.

`phred_spark_raw` List containing per nucleotide Phred score values for each sequence

`phred_raw` Numeric string containing mean Phred scores before trimming.

`phred_trim` Numeric string containing mean Phred scores after trimming.

`Ns_raw` Numeric string containing count of N's before trimming.

`Ns_trim` Numeric string containing count of N's after trimming.

`length_raw` Numeric string containing sequence length before trimming.

`length_trim` Numeric string containing sequence length after trimming.

`seqs_raw` Character string containing sequences before trimming.

`seqs_trim` Character string containing sequence after trimming.

`closest_match` Character string containing species + type strain no. of closest match from reference database.

`NCBI_acc` Character string containing NCBI accession number associated with closest match from reference database.

`ID` Numeric string containing pairwise similarity value for query vs database reference sequence. Calculation of ID is determined by isoTAX 'iddef' parameter (0-4, Default=2). See VSEARCH documentation for more details.

- (0) CD-HIT definition: (matching columns) / (shortest sequence length).
- (1) Edit distance: (matching columns) / (alignment length).
- (2) Edit distance excluding terminal gaps (default definition).
- (3) Marine Biological Lab definition counting each gap opening (internal or terminal) as a single mismatch, whether or not the gap was extended: 1.0- ((mismatches + gap openings)/(longest sequence length)).
- (4) BLAST definition, equivalent to --iddef 1 for global pairwise alignments.

`rank_phylum` Character string containing Phylum rank taxonomy

`rank_class` Character string containing Class rank taxonomy

`rank_order` Character string containing Order rank taxonomy  
`rank_family` Character string containing Family rank taxonomy  
`rank_genus` Character string containing Genus rank taxonomy  
`rank_species` Character string containing Species rank taxonomy  
`phylum_threshold` Numeric string containing Phylum-level sequence similarity threshold for rank demarcation  
`class_threshold` Numeric string containing Class-level sequence similarity threshold for rank demarcation  
`order_threshold` Numeric string containing Order-level sequence similarity threshold for rank demarcation  
`family_threshold` Numeric string containing Family-level sequence similarity threshold for rank demarcation  
`genus_threshold` Numeric string containing Genus-level sequence similarity threshold for rank demarcation  
`species_threshold` Numeric string containing Species-level sequence similarity threshold for rank demarcation

## See Also

[isoTAX](#)

`df_to_isolib`      *Convert isoLIB .CSV output to isoLIB class object*

## Description

Helper function to convert isoLIB .CSV output to a [class-isolib](#) class object.

## Usage

```
df_to_isolib(df)
```

## Arguments

<code>df</code>	Dataframe in same format as .CSV output file from <a href="#">isolib</a> step.
-----------------	--

## Value

Returns an S4 [class-isolib](#) object that can be used to generate interactive HTML output tables.

<code>df_to_isoTAX</code>	<i>Convert isoTAX .CSV output to isoTAX class object</i>
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### Description

Helper function to convert isoTAX .CSV output to a [class-isoTAX](#) class object.

### Usage

```
df_to_isoTAX(df)
```

### Arguments

<code>df</code>	Dataframe in same format as .CSV output file from <a href="#">isoTAX</a> step.
-----------------	--

### Value

Returns an S4 [class-isoTAX](#) object that can be used to generate interactive HTML output tables.

<code>export_html</code>	<i>Export HTML for isoQC &gt; isoTAX &gt; isoLIB class objects</i>
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### Description

S4 wrapper functions to export interactive HTML tables from [isoQC](#), [isoTAX](#), or [isoLIB](#) class objects. Saves to HTML to current working directory and automatically opens.

### Usage

```
## S4 method for signature 'isoQC'
export_html(
  obj,
  min_phred_score = NULL,
  min_length = NULL,
  sliding_window_cutoff = NULL,
  sliding_window_size = NULL
)

## S4 method for signature 'isoTAX'
export_html(obj, quick_search = NULL, db = NULL)

## S4 method for signature 'isoLIB'
export_html(obj, method = NULL, group_cutoff = NULL)
```

### Arguments

<code>obj</code>	An S4 class object generated from one of <a href="#">isoQC</a> , <a href="#">isoTAX</a> , or <a href="#">isoLIB</a> steps
------------------	---

### Value

HTML output file saved to working directory.

**get\_db** *Download taxonomic reference database*

## Description

This function downloads taxonomic reference database and formats them for use.

## Usage

```
get_db(db = "16S_bac", force_update = FALSE, add_taxonomy = FALSE)
```

## Arguments

<b>db</b>	Database selection. One of "16S", "16S_arc", "18S", "ITS", or "cpn60"
<b>force_update</b>	Forces new databases to be downloaded.
<b>add_taxonomy</b>	Add full taxonomy to header to enable classification of sequences offline. This is primarily for cases of operation with no internet or when computing within an HPC cluster without administrator access. Results in a semicolon (;) delimited FASTA header as follows: Accession_no;d__Domain;p__Phylum;c__Class;o__Order;f__Family;g__Genus

## Value

Returns file path for database of interest

## Examples

```
db.path <- get_db(db="16S", force_update=FALSE)
```

**get\_os** *Determine user operating system.*

## Description

Determines the type of operating system being used.

## Usage

```
get_os()
```

## Value

Returns sysname as one of windows/osx-mac/linux

## Examples

```
#Example 1 on a Windows-based operating system
os.index <- get_os()
print(os.index)

#Example 2 on a Mac operating system
os.index <- get_os()
print(os.index)

#Example 3 on a Linux operating system
os.index <- get_os()
print(os.index)
```

---

get\_sanger\_date      *get\_sanger\_date function*

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## Description

Helper function to automatically retrieve run date from Sanger sequencing .ab1 files.

## Usage

```
get_sanger_date(file = NULL)
```

## Arguments

file	The .ab1 file in from which to retrieve the date information. (Must be in S4 abif format)
------	---

## Value

Returns date in "YYYY\_MM\_DD" format

## Examples

```
#Path to the first listed .ab1 file in example directory
fpath <- file.path(system.file("extdata/abif_examples/rocket_salad", package = "isolateR"),
                    list.files(system.file("extdata/abif_examples/rocket_salad", package = "isolateR"))[1])
#Read in the ab1 file to S4 format
ab1.S4 <- sangerseqR::read.abif(fpath)

#Retrieve date
get_sanger_date(ab1.S4)
```

`get_vsearch`*Download VSEARCH software reference database***Description**

This function downloads the VSEARCH software used querying sequences against taxonomic databases of interest.

**Usage**

```
get_vsearch(os = NULL)
```

**Arguments**

os	Operating system, one of: "windows", "osx-mac", or "linux". If blank (os=NULL) then will try to automatically determine operating system.
----	---

**Value**

Returns path for VSEARCH executable

**Examples**

```
#Example for automatically detecting operating system and downloading VSEARCH software
vsearch.path <- get_vsearch()
```

`isoALL`*Perform all commands in one step.***Description**

This function effectively wraps isoQC, isoTAX, and isoLIB steps into a single command for convenience. Input can be a single directory or a list of directories to process at once. If multiple directories are provided, the resultant libraries can be sequentially merged together by toggling the parameter 'merge=TRUE'. All other respective parameters from the wrapped functions can be passed through this command. . The The respective input parameters from the wrapped can be passed through this command with exception of the .creates a strain library by grouping closely related strains of interest based on sequence similarity. For adding new sequences to an already-established strain library, specify the .CSV file path of the older strain library using the 'old\_lib\_csv' parameter.

**Usage**

```
isoALL(
  input = NULL,
  export_html = TRUE,
  export_csv = TRUE,
  export_fasta = TRUE,
  export_fasta_revcomp = FALSE,
  export_blast_table = FALSE,
```

```

    quick_search = FALSE,
    db = "16S",
    db_path = NULL,
    iddef = 2,
    phylum_threshold = 75,
    class_threshold = 78.5,
    order_threshold = 82,
    family_threshold = 86.5,
    genus_threshold = 94.5,
    species_threshold = 98.7,
    include_warnings = FALSE,
    method = "dark_mode",
    group_cutoff = 0.995,
    keep_old_reps = TRUE,
    merge = FALSE
)

```

## Arguments

input	Directory path(s) containing .ab1 files. If more than one, provide as list (e.g. 'input=c("/path/to/directory1","/path/to/directory2")')
export_html	(Default=TRUE) Output the results as an HTML file
export_csv	(Default=TRUE) Output the results as a CSV file.
export_fasta	(Default=TRUE) Output the sequences in a FASTA file.
export_fasta_revcomp	(Default=FALSE) Output the sequences in reverse complement form in a fasta file. This is useful in cases where sequencing was done using the reverse primer and thus the orientation of input sequences needs reversing.
quick_search	(Default=FALSE) Whether or not to perform a comprehensive database search (i.e. optimal global alignment). If TRUE, performs quick search equivalent to setting VSEARCH parameters "-maxaccepts 100 -maxrejects 100". If FALSE, performs comprehensive search equivalent to setting VSEARCH parameters "-maxaccepts 0 -maxrejects 0"
db	(Default="16S") Select database option(s) including "16S" (for searching against the NCBI Refseq targeted loci 16S rRNA database), "ITS" (for searching against the NCBI Refseq targeted loci ITS database. For combined databases in cases where input sequences are derived from bacteria and fungi, select "16S ITS".
db_path	Path of FASTA-formatted database sequence file. Ignored if 'db' parameter is set to anything other than NULL or "custom". Expects a semicolon (;) delimited FASTA header as follows: Accession_no;d_Domain;p_Phylum;c_Class;o_Order;f_Family;g_Scientific_name. See <a href="#">get_db</a> function for examples and details on automatically generating custom databases for offline use or within an HPC cluster environment.
iddef	Set pairwise identity definition as per VSEARCH definitions (Default=2, and is recommended for highest taxonomic accuracy) (0) CD-HIT definition: (matching columns) / (shortest sequence length). (1) Edit distance: (matching columns) / (alignment length). (2) Edit distance excluding terminal gaps (default definition). (3) Marine Biological Lab definition counting each gap opening (internal or terminal) as a single mismatch, whether or not the gap was extended: 1.0-((mismatches + gap openings)/(longest sequence length)). (4) BLAST definition, equivalent to -iddef 1 for global pairwise alignments.

<b>phylum_threshold</b>	Percent cutoff for phylum rank demarcation
<b>class_threshold</b>	Percent cutoff for class rank demarcation
<b>order_threshold</b>	Percent cutoff for order rank demarcation
<b>family_threshold</b>	Percent cutoff for family rank demarcation
<b>genus_threshold</b>	Percent cutoff for genus rank demarcation
<b>species_threshold</b>	Percent cutoff for species rank demarcation
<b>include_warnings</b>	(Default=FALSE) Whether or not to keep sequences with poor alignment warnings from Step 2 'isoTAX' function. Set TRUE to keep warning sequences, and FALSE to remove warning sequences.
<b>method</b>	Method used for grouping sequences. Either 1) "dark_mode", or 2) "closest_species" (Default="dark_mode"). <ul style="list-style-type: none"><li>• Method 1 ("dark_mode") performs agglomerative hierarchical-based clustering to group similar sequences based on pairwise identity (see 'id' parameter) and then within each group, attempts to assign the longest sequence with the most top hits as the group representative. This method is tailored for capturing distinct strains which may represent novel taxa (i.e. microbial dark matter) during isolation workflows. As such, the sequence representatives chosen in each group will not always have the highest % identity to the closest matching type strain. In some cases, sequence members within a group may also have different taxonomic classifications due to them having close to equidistant % identities to different matching type strain material – indicative of a potentially novel taxonomic grouping.</li><li>• Method 2 ("closest_species") groups similar sequences based on their closest matching type strain. For each unique grouping, this results in all sequence members having the same taxonomic classification. The longest sequence with the highest % identity to the closest matching type strain will be assigned as the group representative. Note: The "id" parameter is only used for Method 1 ("dark_mode") and otherwise ignored if using Method 2 ("closest_species").</li></ul>
<b>group_cutoff</b>	(Default=0.995) Similarity threshold based on pairwise identity (0-1) for delineating between sequence groups. 1 = 100% identical/0.995=0.5% difference/0.95=5.0% difference/etc. Used only if method="dark_mode", otherwise ignored.
<b>keep_old_reps</b>	(Default=TRUE) If TRUE, original sequence representatives from old library will be kept when merging with new library. If FALSE, sequence group representatives will be recalculated after combining old and new libraries. Ignored if old_lib_csv=NULL.
<b>merge</b>	If TRUE, combines isoLIB output files consecutively in the order they are listed. Default=FALSE performs all the steps (isoQC/isoTAX/isoLIB) on each directory separately.
<b>verbose</b>	(Default=FALSE) Output progress while script is running.
<b>files_manual</b>	(Default=NULL) For testing purposes only. Specify a list of files to run as file-names without extensions, rather than the whole directory format. Primarily used for testing, use at your own risk.

exclude	(Default=NULL) For testing purposes only. Excludes files of interest from input directory.
min_phred_score	(Default=20) Do not accept trimmed sequences with a mean Phred score below this cutoff
min_length	(Default=200) Do not accept trimmed sequences with sequence length below this number
sliding_window_cutoff	(Default=NULL) Quality trimming parameter (M2) for wrapping SangerRead function in sangeranalyseR package. If NULL, implements auto cutoff for Phred score (recommended), otherwise set between 1-60.
sliding_window_size	(Default=15) Quality trimming parameter (M2) for wrapping SangerRead function in sangeranalyseR package. Recommended range between 5-30.
date	Set date "YYYY_MM_DD" format. If NULL, attempts to parse date from .ab1 file

### Value

Returns a list of [class-isoLIB](#) class objects.

### See Also

[isoQC](#), [isoTAX](#), [isoLIB](#)

### Examples

```
#Set path to directory containing example .ab1 files
fpath1 <- system.file("extdata/abif_examples/rocket_salad", package = "isolateR")

#Run isoALL function with default settings
isoALL(input=fpath1)
```

**isoLIB**

*Generate new strain library or add to existing one.*

### Description

This function creates a strain library by grouping closely related strains of interest based on sequence similarity. For adding new sequences to an already-established strain library, specify the .CSV file path of the older strain library using the 'old\_lib\_csv' parameter.

### Usage

```
isoLIB(
  input = NULL,
  old_lib_csv = NULL,
  method = "dark_mode",
  group_cutoff = 0.995,
  keep_old_reps = TRUE,
  export_html = TRUE,
```

```

    export_csv = TRUE,
    include_warnings = TRUE,
    vsearch_path = NULL,
    phylum_threshold = 75,
    class_threshold = 78.5,
    order_threshold = 82,
    family_threshold = 86.5,
    genus_threshold = 94.5,
    species_threshold = 98.7
)

```

## Arguments

<code>input</code>	Path of CSV output file from isoTAX step.
<code>old_lib_csv</code>	Optional: Path of CSV output isoLIB file or combined isoLIB file from previous run(s)
<code>method</code>	Method used for grouping sequences. Either 1) "dark_mode", or 2) "closest_species" (Default="dark_mode"). <ul style="list-style-type: none"> <li>• Method 1 ("dark_mode") performs agglomerative hierarchical-based clustering to group similar sequences based on pairwise identity (see 'id' parameter) and then within each group, attempts to assign the longest sequence with the most top hits as the group representative. This method is tailored for capturing distinct strains which may represent novel taxa (i.e. microbial dark matter) during isolation workflows. As such, the sequence representatives chosen in each group will not always have the highest % identity to the closest matching type strain. In some cases, sequence members within a group may also have different taxonomic classifications due to them having close to equidistant % identities to different matching type strain material – indicative of a potentially novel taxonomic grouping.</li> <li>• Method 2 ("closest_species") groups similar sequences based on their closest matching type strain. For each unique grouping, this results in all sequence members having the same taxonomic classification. The longest sequence with the highest % identity to the closest matching type strain will be assigned as the group representative. Note: The "id" parameter is only used for Method 1 ("dark_mode") and otherwise ignored if using Method 2 ("closest_species").</li> </ul>
<code>group_cutoff</code>	(Default=0.995) Similarity threshold based on pairwise identity (0-1) for delineating between sequence groups. 1 = 100% identical/0.995=0.5% difference/0.95=5.0% difference/etc. Used only if method="dark_mode", otherwise ignored.
<code>keep_old_reps</code>	(Default=TRUE) If TRUE, original sequence representatives from old library will be kept when merging with new library. If FALSE, sequence group representatives will be recalculated after combining old and new libraries. Ignored if <code>old_lib_csv=NULL</code> .
<code>export_html</code>	(Default=TRUE) Output the results as an HTML file
<code>export_csv</code>	(Default=TRUE) Output the results as a CSV file.
<code>include_warnings</code>	(Default=FALSE) Whether or not to keep sequences with poor alignment warnings from Step 2 'isoTAX' function. Set TRUE to keep warning sequences, and FALSE to remove warning sequences.

vsearch\_path Path of VSEARCH software if manually downloaded in a custom directory. If NULL (Default), will attempt automatic download.

phylum\_threshold Percent sequence similarity threshold for phylum rank demarcation

class\_threshold Percent sequence similarity threshold for class rank demarcation

order\_threshold Percent sequence similarity threshold for order rank demarcation

family\_threshold Percent sequence similarity threshold for family rank demarcation

genus\_threshold Percent sequence similarity threshold for genus rank demarcation

species\_threshold Percent sequence similarity threshold for species rank demarcation

### Value

Returns an isoLIB class object. Default taxonomic cutoffs for phylum (75.0), class (78.5), order (82.0), family (86.5), genus (94.5), and species (98.7) demarcation are based on Yarza et al. 2014, Nature Reviews Microbiology (DOI:10.1038/nrmicro3330)

### See Also

[isoTAX](#), [isoLIB](#)

### Examples

```
#Set path to directory containing example .ab1 files
fpath1 <- system.file("extdata/abif_examples/rocket_salad", package = "isolateR")

#Step 1: Run isoQC function with default settings
isoQC.S4 <- isoQC(input=fpath1)

#Step 2: Run isoTAX function with default settings
fpath2 <- file.path(fpath1, "isolateR_output/01_isoQC_trimmed_sequences_PASS.csv")
isoTAX.S4 <- isoTAX(input=fpath2)

#Step 3: Run isoLIB function with default settings
fpath3 <- file.path(fpath1, "isolateR_output/02_isoTAX_results.csv")
isoLIB.S4 <- isoLIB(input=fpath3)

#Show summary statistics
isoLIB.S4
```

isoQC

*Perform automated quality trimming of input .ab1 files*

### Description

This function loads in ABIF files (.ab1 extension) and performs automatic quality trimming in batch mode.

## Usage

```
isoQC(
  input = NULL,
  export_html = TRUE,
  export_csv = TRUE,
  export_fasta = TRUE,
  export_fasta_revcomp = FALSE,
  verbose = FALSE,
  exclude = NULL,
  min_phred_score = 20,
  min_length = 200,
  sliding_window_cutoff = NULL,
  sliding_window_size = 15,
  date = NULL,
  files_manual = NULL
)
```

## Arguments

<code>input</code>	Path to directory with .ab1 files.
<code>export_html</code>	(Default=TRUE) Output the results as an HTML file
<code>export_csv</code>	(Default=TRUE) Output the results as a CSV file.
<code>export_fasta</code>	(Default=TRUE) Output the sequences in a FASTA file.
<code>export_fasta_revcomp</code>	(Default=FALSE) Output the sequences in reverse complement form in a fasta file. This is useful in cases where sequencing was done using the reverse primer and thus the orientation of input sequences needs reversing.
<code>verbose</code>	(Default =FALSE) Output progress while script is running, FALSE for simplified progress, TRUE for file-by-file details
<code>exclude</code>	(Default=NULL) For testing purposes only. Excludes files of interest from input directory.
<code>min_phred_score</code>	(Default=20) Do not accept trimmed sequences with a mean Phred score below this cutoff
<code>min_length</code>	(Default=200) Do not accept trimmed sequences with sequence length below this number
<code>sliding_window_cutoff</code>	(Default=NULL) Quality trimming parameter (M2) for wrapping SangerRead function in sangeranalyseR package. If NULL, implements auto cutoff for Phred score (recommended), otherwise set between 1-60.
<code>sliding_window_size</code>	(Default=15) Quality trimming parameter (M2) for wrapping SangerRead function in sangeranalyseR package. Recommended range between 5-30.
<code>date</code>	Set date "YYYY_MM_DD" format. If NULL, attempts to parse date from .ab1 file
<code>files_manual</code>	(Default=NULL) For testing purposes only. Specify a list of files to run as file-names without extensions, rather than the whole directory format. Primarily used for testing, use at your own risk.

**Value**

Returns quality trimmed Sanger sequences in FASTA format.

**See Also**

[isoTAX](#), [isoLIB](#)

**Examples**

```
#Set path to directory containing example .ab1 files
fpath1 <- system.file("extdata/abif_examples/rocket_salad", package = "isolateR")

#Step 1: Run isoQC function with default settings
isoQC.S4 <- isoQC(input=fpath1)

#Show summary statistics
isoQC.S4
```

---

isoTAX

*Classify taxonomy of sequences after quality trimming steps.*

---

**Description**

This function performs taxonomic classification steps by searching query Sanger sequences against specified database of interest. Takes CSV input files, extracts FASTA-formatted query sequences and performs global alignment against specified database of interest via Needleman-Wunsch algorithm by wrapping the –usearch\_global command implemented in VSEARCH. Default taxonomic rank cutoffs for 16S rRNA gene sequences are based on Yarza et al. 2014, Nat Rev Microbiol.

**Usage**

```
isoTAX(
  input = NULL,
  export_html = TRUE,
  export_csv = TRUE,
  export_blast_table = FALSE,
  quick_search = FALSE,
  db = "16S_bac",
  db_path = NULL,
  vsearch_path = NULL,
  iddef = 2,
  phylum_threshold = 75,
  class_threshold = 78.5,
  order_threshold = 82,
  family_threshold = 86.5,
  genus_threshold = 94.5,
  species_threshold = 98.7
)
```

## Arguments

<code>input</code>	Path of either 1) CSV output file from isoQC step, or 2) a FASTA formatted file. If input is a FASTA file, the sequence(s) will be converted and saved as an isoQC-formatted output file in the current working directory ("isolateR_output/01_isoQC_mock_table"). Sequence date, name, length, and number of ambiguous bases (Ns) will be calculated from the input file and used to populate the relevant columns. Phred quality scores (phred_trim) will be set to the maximum value (60) and the remaining columns will be populated with mock data to allow compatibility with the isoTAX function. The main purpose of this output file is for flexibility and to allow users to edit/modify the sequence metadata before continuing with subsequent steps.
<code>export_html</code>	(Default=TRUE) Output the results as an HTML file
<code>export_csv</code>	(Default=TRUE) Output the results as a CSV file.
<code>export_blast_table</code>	(Default=FALSE) Output the results as a tab-separated BLAST-like hits table.
<code>quick_search</code>	(Default=FALSE) Whether or not to perform a comprehensive database search (i.e. optimal global alignment). If TRUE, performs quick search equivalent to setting VSEARCH parameters "--maxaccepts 100 --maxrejects 100". If FALSE, performs comprehensive search equivalent to setting VSEARCH parameters "--maxaccepts 0 --maxrejects 0"
<code>db</code>	(Default="16S_bac") Select database option(s) including "16S" (for searching against the NCBI Refseq targeted loci 16S rRNA database), "ITS" (for searching against the NCBI Refseq targeted loci ITS database. For combined databases in cases where input sequences are derived from bacteria and fungi, select "16SITS". Setting to anything other than db=NULL or db="custom" causes 'db.path' parameter to be ignored.
<code>db_path</code>	Path of FASTA-formatted database sequence file. Ignored if 'db' parameter is set to anything other than NULL or "custom". Expects a semicolon (;) delimited FASTA header as follows: Accession_no;d_Domain;p_Phylum;c_Class;o_Order;f_Family;g_Species. See <a href="#">get_db</a> function for examples and details on automatically generating custom databases for offline use or within an HPC cluster environment.
<code>vsearch_path</code>	Path of VSEARCH software if manually downloaded in a custom directory. If NULL (Default), will attempt automatic download.
<code>iddef</code>	Set pairwise identity definition as per VSEARCH definitions (Default=2, and is recommended for highest taxonomic accuracy) (0) CD-HIT definition: (matching columns) / (shortest sequence length). (1) Edit distance: (matching columns) / (alignment length). (2) Edit distance excluding terminal gaps (default definition). (3) Marine Biological Lab definition counting each gap opening (internal or terminal) as a single mismatch, whether or not the gap was extended: 1.0-((mismatches + gap openings)/(longest sequence length)). (4) BLAST definition, equivalent to --iddef 1 for global pairwise alignments.
<code>phylum_threshold</code>	Percent sequence similarity threshold for phylum rank demarcation
<code>class_threshold</code>	Percent sequence similarity threshold for class rank demarcation
<code>order_threshold</code>	Percent sequence similarity threshold for order rank demarcation
<code>family_threshold</code>	Percent sequence similarity threshold for family rank demarcation

```

genus_threshold
Percent sequence similarity threshold for genus rank demarcation
species_threshold
Percent sequence similarity threshold for species rank demarcation

```

**Value**

Returns taxonomic classification table of class isoTAX. Default taxonomic cutoffs for phylum (75.0), class (78.5), order (82.0), family (86.5), genus (94.5), and species (98.7) demarcation are based on Yarza et al. 2014, Nature Reviews Microbiology (DOI:10.1038/nrmicro3330)

**See Also**

[isoQC](#), [isoLIB](#), [search\\_db](#)

**Examples**

```

#Set path to directory containing example .ab1 files
fpath1 <- system.file("extdata/abif_examples/rocket_salad", package = "isolateR")

#Step 1: Run isoQC function with default settings
isoQC.S4 <- isoQC(input=fpath1)

#Step 2: Run isoTAX function with default settings
fpath2 <- file.path(fpath1, "isolateR_output/01_isoQC_trimmed_sequences_PASS.csv")
isoTAX.S4 <- isoTAX(input=fpath2)
#Show summary statistics
isoTAX.S4

```

**make\_fasta**

*Convert CSV file containing sequences to FASTA format*

**Description**

This function extracts sequences from a table in CSV format and converts them to FASTA format. Requires two columns, one with sequences and one with sequence names.

**Usage**

```

make_fasta(
  csv_file = NULL,
  col_names = "ID",
  col_seqs = "Sequence",
  output = "output.fasta"
)

```

**Arguments**

csv_file	Filename (or path and filename if not in working directory) of the table from which you would like to generate a FASTA file.
col_names	Column name with the unique names/identifiers. (Default="ID")
col_seqs	Column name with the sequences. (Default="Sequence")
output	Desired filename for output FASTA file (Default = "output.fasta")

**Value**

Returns sequences in FASTA format.

**Examples**

```
#Set path to directory containing example .ab1 files
fpath1 <- system.file("extdata/abif_examples/rocket_salad", package = "isolateR")

#Run isoQC function with default settings to generate CSV file
isoQC.S4 <- isoQC(input=fpath1)

#Set path of CSV output file from isoQC step
csv.path <- file.path(fpath1, "isolateR_output/01_isoQC_trimmed_sequences_PASS.csv")

#Run make_fasta function
make_fasta(csv_file= csv.path, col_names="filename", col_seqs="seqs_trim", output="output.fasta")
```

make\_tree

*Generate a phylogenetic tree from an isoLIB output file*

**Description**

This script will help the user make a simple phylogenetic tree from a strain library. It will allow the user to colour the tree by taxonomic rank only. See [ggtree](#) documentation for more information on customization options available.

**Usage**

```
make_tree(input = NULL)
```

**Arguments**

input	Full path to isoLIB strain library output file in .CSV format.
-------	--

**Value**

Returns a [ggtree](#) class object

**See Also**

[isoLIB](#)

**Examples**

```
#Set path to directory containing example .ab1 files
fpath1 <- system.file("extdata/abif_examples/rocket_salad", package = "isolateR")

#Step 1: Run isoQC function with default settings
isoQC.S4 <- isoQC(input=fpath1)

#Step 2: Run isoTAX function with default settings
fpath2 <- file.path(fpath1, "isolateR_output/01_isoQC_trimmed_sequences_PASS.csv")
isoTAX.S4 <- isoTAX(input=fpath2)
```

```
#Step 3: Run isoLIB function with default settings
fpath3 <- file.path(fpath1, "isolateR_output/02_isоТAX_results.csv")
isoLIB.S4 <- isoLIB(input=fpath3)
#Step 4: Make a tree from isoLIB output CSV file
fpath4 <- file.path(fpath1, "isolateR_output/03_isоЛIB_results.csv")
make_tree(input= fpath4)
```

method-isoLIB

*setMethod functions for isoLIB***Description**

Initiation of isoLIB functions.

**Usage**

```
## S4 method for signature 'missing'
isoLIB(
  input = NULL,
  old_lib_csv = NULL,
  method = "dark_mode",
  group_cutoff = 0.995,
  keep_old_reps = TRUE,
  export_html = TRUE,
  export_csv = TRUE,
  include_warnings = TRUE,
  vsearch_path = NULL,
  phylum_threshold = 75,
  class_threshold = 78.5,
  order_threshold = 82,
  family_threshold = 86.5,
  genus_threshold = 94.5,
  species_threshold = 98.7
)
```

method-isoQC

*setMethod functions for isoQC***Description**

Initiation of isoQC functions.

**Usage**

```
## S4 method for signature 'missing'
isoQC(
  input = NULL,
  export_html = TRUE,
  export_csv = TRUE,
```

```

  export_fasta = TRUE,
  export_fasta_revcomp = FALSE,
  verbose = FALSE,
  exclude = NULL,
  min_phred_score = 20,
  min_length = 200,
  sliding_window_cutoff = NULL,
  sliding_window_size = 15,
  date = NULL,
  files_manual = NULL
)

```

**method-isoTAX***setMethod functions for isoTAX***Description**

Initiation of isoTAX functions.

**Usage**

```

## S4 method for signature 'missing'
isoTAX(
  input = NULL,
  export_html = TRUE,
  export_csv = TRUE,
  export_blast_table = FALSE,
  quick_search = FALSE,
  db = "16S_bac",
  db_path = NULL,
  vsearch_path = NULL,
  iddef = 2,
  phylum_threshold = 75,
  class_threshold = 78.5,
  order_threshold = 82,
  family_threshold = 86.5,
  genus_threshold = 94.5,
  species_threshold = 98.7
)

```

**S4\_to\_dataframe***Converts S4 objects (isoQC, isoTAX, or isoLIB) to dataframe***Description**

Helper function to convert S4 class objects ([isoQC](#), [isoTAX](#), or [isoLIB](#)) to dataframe

**Usage**

```
S4_to_dataframe(obj)
```

**Arguments**

**obj** S4 object generated from [isoQC](#), [isoTAX](#), or [isoLIB](#) steps

**Value**

Returns a dataframe containing sequence information in columns.

sanger\_assembly

*Overlap multiple paired Sanger sequences in batch.*

**Description**

This function loads in the CSV results table from isoQC and merges related sequences based on user input. Original file names before isoQC step need to have a common prefix and differentiating suffixes. (e.g. SAMPLE\_01\_F.ab1, SAMPLE\_01\_R.ab1). After aligning paired sequences, the consensus sequence is extracted and priority is given to the read with higher quality. Phred quality scores are reassigned in the final output table in a basic way by taking the mean of both input sequences.

Note: This function is designed to be used after the isoQC step and before the isoTAX step.

**Usage**

```
sanger_assembly(input = NULL, suffix = "_F.ab1|_R.ab1")
```

**Arguments**

<b>input</b>	Path of CSV output file from isoQC step.
<b>suffix</b>	Regex-friendly suffix for denoting filename groupings. Default="\_F.ab1 _R.ab1" for the common scenario of Sanger sequencing a marker gene in forward and reverse. Direction of sequences including reverse complements will be automatically detected.

**Value**

Returns merged pairs of Sanger sequences in FASTA format.

**See Also**

[isoQC](#), [isoTAX](#)

**Examples**

```
#Load package
library(isolateR)

#Step 1: Set path to directory containing paired .ab1 files
fpath <- system.file("extdata/abif_examples/drosophila_paired", package="isolateR")

#Step 2: Run isoQC function to trim poor quality regions (Phred score <20) before assembly
isoQC.S4 <- isoQC(input=fpath, sliding_window_cutoff = 20)

#Step 3: Assemble paired sequences
```

```
sanger_assembly(input = file.path(fpath,"isolateR_output", "01_isoQC_trimmed_sequences_PASS.csv"),
                 suffix = "_F.ab1|_R.ab1")

#Detected 3 unique group(s) with suffix provided.
#Group      Individual filenames
#-----
# 1        DRO-1-isolate_F.ab1 | DRO-1-isolate_R.ab1
# 2        DRO-2-isolate_F.ab1 | DRO-2-isolate_R.ab1
# 3        DRO-3-isolate_F.ab1 | DRO-3-isolate_R.ab1
```

**search\_db**

*Perform global alignment pairwise identity search using VSEARCH and type strain database of interest.*

**Description**

Performs global alignment between FASTA-formatted query sequences and the specified database of interest. Uses the Needleman-Wunsch algorithm by wrapping the –usearch\_global command implemented in VSEARCH.

**Usage**

```
search_db(
  query_path = NULL,
  uc.out = "VSEARCH_output.uc",
  b6.out = "VSEARCH_output.b6o",
  path = getwd(),
  quick_search = FALSE,
  db = NULL,
  db_path = NULL,
  vsearch_path = NULL,
  keep_temp_files = FALSE,
  iddef = 2
)
```

**Arguments**

<code>query_path</code>	Path of FASTA-formatted query sequence file.
<code>uc.out</code>	Path of UC-formatted results output table.
<code>b6.out</code>	Path of blast6-formatted results output table.
<code>path</code>	Working path directory (Default is set to current working directory via 'getwd()')
<code>quick_search</code>	(Default=FALSE) Whether or not to perform a comprehensive database search (i.e. optimal global alignment). If TRUE, performs quick search equivalent to setting VSEARCH parameters "-maxaccepts 100 -maxrejects 100". If FALSE, performs comprehensive search equivalent to setting VSEARCH parameters "-maxaccepts 0 -maxrejects 0" Note: This option is provided for convenience and rough approximation of taxonomy only, set to FALSE for accurate % pairwise identity results.

db	Optional: Select any of the standard database option(s) including "16S" (for searching against the NCBI Refseq targeted loci 16S rRNA database), "ITS" (for searching against the NCBI Refseq targeted loci ITS database. For combined databases in cases where input sequences are derived from bacteria and fungi, select "16SIITS". Setting to anything other than db=NULL or db="custom" causes 'db.path' parameter to be ignored.
vsearch_path	Path of VSEARCH software if manually downloaded in a custom directory. If NULL (Default), will attempt automatic download.
keep_temp_files	Toggle (TRUE/FALSE). If TRUE, temporary .uc and .b6o output files are kept from VSEARCH –uc and –blast6out commands, respectively. If FALSE, temporary files are removed.
iddef	Set pairwise identity definition as per VSEARCH definitions (Default=2, and is recommended for highest taxonomic accuracy) (0) CD-HIT definition: (matching columns) / (shortest sequence length). (1) Edit distance: (matching columns) / (alignment length). (2) Edit distance excluding terminal gaps (default definition). (3) Marine Biological Lab definition counting each gap opening (internal or terminal) as a single mismatch, whether or not the gap was extended: 1.0-((mismatches + gap openings)/(longest sequence length)). (4) BLAST definition, equivalent to –iddef 1 for global pairwise alignments.
db.path	Path of FASTA-formatted database sequence file. Ignored if 'db' parameter is set to anything other than "custom"

### Value

Returns a dataframe matching the UC-formatted output table from VSEARCH. Query sequences are automatically added to the final column. Summary of column information. See VSEARCH documentation for more details.

- V1 = Record type of hit (H) or no hit (N)
- V2 = Ordinal number of the target sequence (based on input order, starting from zero). Set to '\*' for N.
- V3 = Sequence length. Set to '\*' for N.
- V4 = Percentage of similarity with the target sequence. Set to '\*' for N.
- V5 = Match orientation + or - . Set to '.' for N.
- V6 = Not used, always set to zero for H, or '\*' for N.
- V7 = Not used, always set to zero for H, or '\*' for N.
- V8 = Compact representation of the pairwise alignment using the CIGAR format (Compact Idiosyncratic Gapped Alignment Report): M (match/mismatch), D (deletion) and I (insertion). The equal sign '=' indicates that the query is identical to the centroid sequence. Set to '\*' for N.
- V9 = Label of the query sequence. Equivalent to 'filename' slot of isolateR class objects (e.g. isoQC, isoTAX, isoLIB).
- V10 = Label of the target centroid sequence. Set to '\*' for N.

### See Also

[isoTAX](#)

## Examples

```
#Set path to directory containing example .ab1 files
fpath1 <- system.file("extdata/abif_examples/rocket_salad", package = "isolateR")

#Run isoQC function with default settings
isoQC.S4 <- isoQC(input=fpath1)

#Set path of CSV output file containing PASS sequences from isoQC step
fasta.path <- "01_isoQC_trimmed_sequences_PASS.fasta"

#Set paths
output.path <- file.path(fpath1, "isolateR_output")

#Run search_db function
uc.df <- search_db(query.path=fasta.path, path=output.path, quick_search=TRUE, db="16S")

#Inspect results
uc.df[1:10,1:10]
```

show

*Generic show method for S4 class objects*

## Description

Generic show method for S4 class objects.

## Usage

```
## S4 method for signature 'isoQC'
show(object)

## S4 method for signature 'isoTAX'
show(object)

## S4 method for signature 'isoLIB'
show(object)
```

valid\_tax\_check

*Validate species name via API client of LPSN*

## Description

This function will determine if each species in a CSV file is validly published or not. Result file will be a CSV with the results appended to the input data. This function requires the user to have an LPSN API account setup. For more details and to register, see here: <https://api.lpsn.dsmz.de/>

## Usage

```
valid_tax_check(input = NULL, col_species = "rank_species", export_csv = TRUE)
```

**Arguments**

input	CSV file path. Expects full path if CSV file is not in the current working directory.
col_species	Specify the column containing the binomial species names (e.g. "Akkermansia muciniphila")
export_csv	Toggle (TRUE/FALSE). Set TRUE to automatically write .CSV file of results to current directory. (Default=TRUE)

**Value**

Returns a CSV saved in working directory

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