

INDEED R package for cancer biomarker discovery

Yiming Zuo and Kian Ghaffari

2019-08-15

Introduction

Differential expression (DE) analysis is commonly used to identify biomarker candidates that have significant changes in their expression levels between distinct biological groups. One drawback of DE analysis is that it only considers the changes on single biomolecular level. In differential network (DN) analysis, network is typically built based on the correlation and biomarker candidates are selected by investigating the network topology. However, correlation tends to generate over-complicated networks and the selection of biomarker candidates purely based on network topology ignores the changes on single biomolecule level. Thus, we have proposed a novel method INDEED, which considers both the changes on single biomolecular and network levels by integrating DE and DN analysis. INDEED has been published in Methods journal (PMID: 27592383). This is the R package that implements the algorithm.

This R package will generate a list of dataframes containing information such as p-values, node degree and activity score for each biomolecule. A higher activity score indicates that the corresponding biomolecule has more neighbors connected in the differential network and their p-values are more statistically significant. It will also generate a csv file for the differential network created by INDEED.

Installation

You can install INDEED from github with:

```
# install.packages("devtools")
devtools::install_github("ressomlab/INDEED")
```

Load package

Load the package.

```
# load INDEED
library(INDEED)
# Loading required package: glasso
```

non-partial correlation data analysis function `non_partial_cor()`

- (**data**) A data frame that contains the expression level of individual biomolecule from two biologically disparate groups ($p \times n$)
- (**class_label**) A binary array with group 1 labeled as 0 and group 2 as 1
- (**id**) An array that includes the corresponding ID for each biomolecule
- (**method**) Type of correlation, user can choose between pearson and spearman correlation, default pearson
- (**permutation**) A positive integer number of permutation, default 1000
- (**p_val**) An optional dataframe containing p-values obtained from DE analysis

In non partial correlation method, user only need to run `non_partial_cor()` function. No rho number will be needed. In this method, user input dataframe with expression level, class label, id, method, number of permutation and p-value similar to partial correlation but in one step fashion. Result will be saved in a list

of two dataframes: `activity_score` and `diff_network`. `Activity_score` contains a dataframe with biomolecules ranked by activity score calculated from p-value and node degree. `Diff_network` contains a dataframe of connection weight between two nodes.

The following example demonstrates how to use `non_partial_cor()` function:

```
non_partial_cor(data = Met_GU, class_label = Met_Group_GU, id = Met_name_GU, method = "spearman")
```

partial correlation data preprocessing function `select_rho_partial()`

To use the function `select_rho_partial()`, users will need to have these data sets in hand:

- (**data**) A data frame that contains the expression level of individual biomolecule from two biologically disparate groups ($p * n$)
- (**class_label**) A binary array with group 1 labeled as 0 and group 2 as 1
- (**id**) An array that includes the corresponding ID for each biomolecule

partial correlation data analysis function `partial_cor()`

- (**data_list**) A list of dataframes output from `select_rho_partial` step
- (**rho_group1**) Rule to choose rho for first group of data, user can input “min” for minimum rule, “ste” as one standard error rule, or user can input a number of choice
- (**rho_group2**) Rule to choose rho for second group of data, user can input “min” for minimum rule, “ste” as one standard error rule, or user can input a number of choice
- (**permutation**) A positive integer number of permutation, default 1000
- (**p_val**) A dataframe containing p-values obtained from DE analysis (optional)

The demo data is presented in the **Demo Data** section below. It will be automatically loaded with the package.

In partial correlation method, user will need to preprocess the data using `select_rho_partial()` function, followed by using `partial_cor()` and choosing desired rho number and complete the analysis. User can also choose to provide p-value table and number of permutations in `partial_cor()` function. Result will be saved in a list of two dataframes: `activity_score` and `diff_network`. `Activity_score` contains a dataframe with biomolecules ranked by activity score calculated from p-value and node degree. `Diff_network` contains a dataframe of connection weight between two nodes.

The following example demonstrates how to use `select_rho_partial()` and `partial_cor()` function:

```
pre_data <- select_rho_partial(data = Met_GU, class_label = Met_Group_GU, id = Met_name_GU)
result <- partial_cor(data_list = pre_data, rho_group1 = 'min', rho_group2 = "min",
                      permutation = 1000, p_val = pvalue_M_GU, permutation_thres = 0.05)
```

In this case, the sparse differential network is based on partial correlation and p-value for each biomolecule is calculated for users. Also, **rho** is picked based on minimum rule and number of permutations is set to 1000.

Interactive Network Visualization function `network_display()`

- (**results**) This is the result from the calling either `non_partial_cor()` or `partial_cor()`.
- (**nodesize**) This parameter determines what the size of each node will represent. The options are ‘Node_Degree’, ‘Activity_Score’, ‘Z_Score’, and ‘P_Value’. The title of the resulting network will identify which parameter was selected to represent the node size. The default is Node_Degree.
- (**nodecolor**) This parameter determines what color each node will be, based on a yellow or red color gradient. The options are ‘Node_Degree’, ‘Activity_Score’, ‘Z_Score’, and ‘P_Value’. A color bar will be created based on which parameter is chosen.

- **(edgewidth)** This is a ‘YES’ or ‘NO’ option as to if the edgewidth should be representative of the weight value corresponding to the correlation between two nodes.
- **(layout)** User can choose from a handful of network visualization templates including: ‘nice’, ‘sphere’, ‘grid’, ‘star’, and ‘circle’.

An interactive tool to assist in the visualization of the results from INDEED functions `non_partial_corr()` or `partial_corr()`. The size and the color of each node can be adjusted by users to represent either the `Node_Degree`, `Activity_Score`, `Z_Score`, or `P_Value`. The color of the edge is based on the binary value of either 1 corresponding to a positive correlation depicted as green or a negative correlation of -1 depicted as red. The user also has the option of having the width of each edge be proportional to its weight value. The layout of the network can also be customized by choosing from the options: ‘nice’, ‘sphere’, ‘grid’, ‘star’, and ‘circle’. Nodes can be moved and zoomed in on. Each node and edge will display extra information when clicked on. Secondary interactions will be highlighted as well when a node is clicked on.

The following example demonstrates how to use the `network_display()` function:

```
network_display(result)
```

Examples

Example 1

```
pre_data <- select_rho_partial(data = Met_GU, class_label = Met_Group_GU, id = Met_name_GU)
partial_cor(data_list = pre_data, rho_group1 = 'min', rho_group2 = "min", permutation = 500,
            p_val = pvalue_M_GU, permutation_thres = 0.05)
```

Example 2

```
result2 <- non_partial_cor(data = Met_GU, class_label = Met_Group_GU, id = Met_name_GU,
                          method = "spearman", permutation_thres = 0.05)
```

Example 3

```
network_display(result2, layout = 'nice', nodesize = 'Node_Degree',
                nodecolor = 'Activity_Score', edgewidth = 'NO')
```