

# Using lm with asremlPlus for the Ladybird example from Welham et al. (2014)

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## Introduction

This vignette shows how to use `asremlPlus` (Brien, 2025a), and `dae` (Brien, 2025b), for exploring and presenting predictions (estimated marginal means: EMMs) from a linear mixed model analysis, the predictions having been produced using `lmerTest` (Kuznetsova et al., 2017), `lm` and `emmeans` (Lenth, 2023). Here, `asremlPlus`, `dae`, `lmerTest` and `emmeans` are packages for the R Statistical Computing environment (R Core Team, 2025) and `lm` is available from `stats` and is included in R.

The context is a three-factor factorial experiment on ladybirds (Welham et al., 2014, Example 8.2) that aims to answer the question “Will ladybirds transfer fungus to aphids on plants?” The experiment consists of 2 runs of 36 containers, each with a plant and aphids. There are three factors that results in 12 treatments: Host plant (beans, trefoil), infected Cadavers (5, 10, 20), Ladybird (-, +). These are randomized to the containers within a run so that each is replicated 3 times within a run. The response to be analysed is the logit of the proportion of live aphids that were infected.

## Initialize

```
library(knitr)
opts_chunk$set("tidy" = FALSE, comment = NA)
suppressMessages(library(lmerTest))
packageVersion("lmerTest")
```

```
## [1] '3.1.3'
```

```
suppressMessages(library(emmeans))
packageVersion("emmeans")
```

```
## [1] '1.11.2.8'
```

```
suppressMessages(library(asremlPlus))
packageVersion("asremlPlus")
```

```
## [1] '4.4.55'
```

```

suppressMessages(library(dae))
packageVersion("dae")

## [1] '3.2.33'

options(width = 95, show.signif.stars = FALSE)

```

## Get data available in asremlPlus

```

data("Ladybird.dat")

```

## Do an ANOVA of logits

```

Ladybird.aov <- aov(logitP ~ Host*Cadavers*Ladybird + Error(Run/Plant),
                      data=Ladybird.dat)
summary(Ladybird.aov)

```

```

Error: Run
      Df  Sum Sq Mean Sq F value Pr(>F)
Residuals 1 0.06766 0.06766

```

```

Error: Run:Plant
      Df  Sum Sq Mean Sq F value    Pr(>F)
Host          1 13.599 13.599 59.172 1.82e-10
Cadavers      2 17.027  8.514 37.044 3.78e-11
Ladybird       1 11.091 11.091 48.257 3.33e-09
Host:Cadavers 2  0.308  0.154  0.670  0.5158
Host:Ladybird   1  0.228  0.228  0.992  0.3234
Cadavers:Ladybird 2  1.735  0.867  3.774  0.0287
Host:Cadavers:Ladybird 2  0.200  0.100  0.435  0.6493
Residuals      59 13.560  0.230

```

The anova table gives the F-tests for the three-factor effects and interactions. Note the `Residuals` `Mean Sq` value for `Run:Plant` of 0.230. Also, it is clear that the `Run` component is negative, given that the `Residuals` `Mean Sq` value for `Run` is less than that for `Run:Plant`; it is  $(0.06766 - 0.230) / 36$ . From the table it is seen that the only significant interaction is `Cadavers:Ladybird` and that the `Host` main effect is significant.

## Use lmerTest and lm to analyse the logits

### Mixed model analysis of logits

```

m1.lmer <- lmerTest::lmer(logitP ~ Host*Cadavers*Ladybird + (1|Run),
                           data=Ladybird.dat)

```

```
boundary (singular) fit: see help('isSingular')
```

```
summary(m1.lmer)
```

```
Linear mixed model fit by REML. t-tests use Satterthwaite's method [lmerModLmerTest]
Formula: logitP ~ Host * Cadavers * Ladybird + (1 | Run)
Data: Ladybird.dat
```

```
REML criterion at convergence: 102.8
```

```
Scaled residuals:
```

Min	1Q	Median	3Q	Max
-1.9633	-0.5217	0.1360	0.5789	2.1896

```
Random effects:
```

Groups	Name	Variance	Std.Dev.
Run	(Intercept)	0.0000	0.0000
Residual		0.2271	0.4766

```
Number of obs: 72, groups: Run, 2
```

```
Fixed effects:
```

	Estimate	Std. Error	df	t value	Pr(> t )
(Intercept)	-1.603097	0.194560	60.000000	-8.240	1.91e-11
Hosttrefoil	-0.870675	0.275149	60.000000	-3.164	0.00244
Cadavers10	0.564771	0.275149	60.000000	2.053	0.04448
Cadavers20	0.919229	0.275149	60.000000	3.341	0.00144
Ladybird+	0.547710	0.275149	60.000000	1.991	0.05109
Hosttrefoil:Cadavers10	-0.212735	0.389120	60.000000	-0.547	0.58661
Hosttrefoil:Cadavers20	-0.120410	0.389120	60.000000	-0.309	0.75806
Hosttrefoil:Ladybird+	0.073153	0.389120	60.000000	0.188	0.85151
Cadavers10:Ladybird+	-0.040048	0.389120	60.000000	-0.103	0.91837
Cadavers20:Ladybird+	0.414204	0.389120	60.000000	1.064	0.29138
Hosttrefoil:Cadavers10:Ladybird+	0.005698	0.550299	60.000000	0.010	0.99177
Hosttrefoil:Cadavers20:Ladybird+	0.449979	0.550299	60.000000	0.818	0.41676

```
Correlation of Fixed Effects:
```

	(Intr)	Hsttrf	Cdvr10	Cdvr20	Ldybr+	Hs:C10	Hs:C20	Hst:L+	C10:L+	C20:L+	H:C10:
Hosttrefoil	-0.707										
Cadavers10	-0.707	0.500									
Cadavers20	-0.707	0.500	0.500								
Ladybird+	-0.707	0.500	0.500	0.500							
Hsttrfl:C10	0.500	-0.707	-0.707	-0.354	-0.354						
Hsttrfl:C20	0.500	-0.707	-0.354	-0.707	-0.354	0.500					
Hsttrfl:Ld+	0.500	-0.707	-0.354	-0.354	-0.707	0.500	0.500				
Cdvsr10:Ld+	0.500	-0.354	-0.707	-0.354	-0.707	0.500	0.250	0.500			
Cdvsr20:Ld+	0.500	-0.354	-0.354	-0.707	-0.707	0.250	0.500	0.500	0.500		
Hstt:C10:L+	-0.354	0.500	0.500	0.250	0.500	-0.707	-0.354	-0.707	-0.707	-0.354	
Hstt:C20:L+	-0.354	0.500	0.250	0.500	0.500	-0.354	-0.707	-0.707	-0.354	-0.707	0.500
optimizer (nloptwrap) convergence code: 0 (OK)											
boundary (singular) fit: see help('isSingular')											

As expected the Run component is bound at zero, leading to a singular model. This results in a change in the estimate of the residual variance to 0.227. To allow for a negative estimate we will redo the analysis

with `Run` fixed, because with `lme4` (`lmerTest`) one cannot unconstrain the `Run` component to allow it to be negative. As Littell et al. (2006, p.150) say

if you do not set the negative variance component estimate to zero, but allow it to remain negative, you get better control over Type I error and, for cases of negative wholeplot error variance estimates, greater power. Therefore, this is the recommended procedure.

## Analyse with Reps fixed using `lm` to make the analysis equivalent to ANOVA

The function `lm` has to be used because there are no random terms; `lme4` cannot be used because it requires at least one random term.

```
m.lm <- lm(logitP ~ Run + Host*Cadavers*Ladybird,
             data=Ladybird.dat)
(aov.tab <- anova(m.lm))
```

Analysis of Variance Table

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Run	1	0.0677	0.0677	0.2944	0.58946
Host	1	13.5992	13.5992	59.1720	1.815e-10
Cadavers	2	17.0274	8.5137	37.0444	3.784e-11
Ladybird	1	11.0907	11.0907	48.2571	3.329e-09
Host:Cadavers	2	0.3078	0.1539	0.6695	0.51579
Host:Ladybird	1	0.2279	0.2279	0.9916	0.32341
Cadavers:Ladybird	2	1.7349	0.8675	3.7744	0.02867
Host:Cadavers:Ladybird	2	0.1999	0.1000	0.4350	0.64932
Residuals	59	13.5596	0.2298		

Now the `Run:Plant` variance estimate is equal to that for the `Residuals` `Mean Sq` for `Run:Plant` from the anova table.

## Obtain the marginality matrix for the fixed terms

The `pstructure` function from the `dae` package (Brien, 2025b) produce the marginality matrix for a formula as a side effect and we take advantage of that to obtain the matrix required here.

```
Ladybird.pstr <- pstructure(formula = ~ Host*Cadavers*Ladybird,
                           data = Ladybird.dat)
(HCL.marg <- marginality(Ladybird.pstr))
```

	Host	Cadavers	Host:Cadavers	Ladybird	Host:Ladybird	Cadavers:Ladybird	
Host	1	0	1	0	1		0
Cadavers	0	1	1	0	0		1
Host:Cadavers	0	0	1	0	0		0
Ladybird	0	0	0	1	1		1
Host:Ladybird	0	0	0	0	1		0
Cadavers:Ladybird	0	0	0	0	0		1
Host:Cadavers:Ladybird	0	0	0	0	0		0

```

Host                               1
Cadavers                          1
Host:Cadavers                     1
Ladybird                           1
Host:Ladybird                      1
Cadavers:Ladybird                  1
Host:Cadavers:Ladybird             1

```

This marginality matrix is interpreted by taking a row term and noting that it is marginal to any column term with a one in this row.

### Choose marginality-compliant model

```

chosen <- chooseModel(aov.tab, DF = "Df", denDF = 59, p.values = "Pr(>F)" ,
                        terms.marginality = HCL.marg)
(chosen$choose.summary)

```

```
#### Sequence of model investigations
```

	terms	DF	denDF	p	action
1	Host:Cadavers:Ladybird	2	59	0.6493	Nonsignificant
2	Cadavers:Ladybird	2	59	0.0287	Significant
3	Host:Ladybird	1	59	0.3234	Nonsignificant
4	Host:Cadavers	2	59	0.5158	Nonsignificant
5	Host	1	59	0.0000	Significant

```
(chosen$sig.terms)
```

```
[[1]]
[1] "Cadavers:Ladybird"
```

```
[[2]]
[1] "Host"
```

The `chooseModel` function produces a list with components `sig.terms`, a list with the terms in the marginality-compliant model, and `choose.summary`, a data.frame that details the tests performed in choosing the model. Note that `chooseModel` does not test the main effects for Cadavers or Ladybird, because these are marginal to the significant two-factor interaction Cadavers:Ladybird.

### Form formula for selected model

```

chosen.mod <- paste(unlist(chosen$sig.terms), collapse = " + ")
(chosen.mod <- as.formula(paste("~", chosen.mod)))

```

```
~Cadavers:Ladybird + Host
```

## Form predictions that conform to the chosen model

Use `emmeans` to get the predictions and associated statistics for the full model.

```
HCL.emm <- emmeans::emmeans(m1.lmer, specs = ~ Host:Cadavers:Ladybird)
HCL.preds <- summary(HCL.emm)
den.df <- min(HCL.preds$df)
HCL.vcov <- vcov(HCL.emm)
```

Setting the `specs` argument to `Host:Ladybird:Cadavers` requests predictions for all combinations of the three factors.

### Modify `HCL.preds` to be compatible with a `predictions.frame`

Basically, this is an exercise in renaming the columns in the `data.frame` containing the predictions.

```
names(HCL.preds)

[1] "Host"      "Cadavers" "Ladybird" "emmmean"   "SE"        "df"        "lower.CL"  "upper.CL"

HCL.preds <- as.predictions.frame(HCL.preds, predictions = "emmmean",
                                    se = "SE", interval.type = "CI",
                                    interval.names = c("lower.CL", "upper.CL"))
names(HCL.preds)

[1] "Host"                  "Cadavers"           "Ladybird"
[4] "predicted.value"       "standard.error"    "df"
[7] "lower.Confidence.limit" "upper.Confidence.limit" "est.status"
```

### Form an `alldiffs` object with predictions obtained with `emmeans`

```
HCL.diffs <- allDifferences(predictions = HCL.preds, classify = "Host:Ladybird:Cadavers",
                               vcov = HCL.vcov, tdf = den.df)
```

The functions `allDifferences` is used to form the `alldiffs.obj` that contains a `predictions` component, along with components related to pairwise comparisons. The `predictions` component contains upper and lower confidence limits produced by `emmeans`. The `tdf` is supplied so that it can be used to get the degrees of freedom for the *t*-value to be used in calculating the error intervals.

### Transform the prediction to conform to chosen model

The `linTransform` function is used to obtain estimated marginal means (EMMs) that conform to the chosen model. Because we would prefer error intervals based on  $\pm 0.5LSD$ , the `error.intervals` argument has been set to `"halfLeast"`, the `LSDtype` argument to `"factor.combination"` and the `LSDby` argument to `"Host"` so that the average LSD will be calculated for each Host. This necessary because, under the chosen model, the LSDs differ between Hosts. It results in `lower.halfLeastSignificant.limit` and `upper.halfLeastSignificant.limit` replacing the limits based on the confidence intervals in the `predictions` component of the resulting `alldiffs` object.

```

diffs <- linTransform(HCL.diffs, linear.transformation = ~Cadavers:Ladybird + Host,
                      error.intervals = "halfLeast",
                      LSDtype = "factor.combination", LSDby = "Host",
                      tables = "predictions")

```

Joining with 'by = join\_by(fac.comb)'  
 Joining with 'by = join\_by(Host)'

#### Predictions for transform(s) from Host:Ladybird:Cadavers

The original predictions, obtained as described below, have been linearly transformed to form estimated marginal means.

	Host	Ladybird	Cadavers	predicted.value	standard.error	df
1	bean	-	5	-1.6038338	0.1485977	47.2
2	bean	-	10	-1.1454308	0.1485977	47.2
3	bean	-	20	-0.7448097	0.1485977	47.2
4	bean	+	5	-1.0195475	0.1485977	47.2
5	bean	+	10	-0.5983440	0.1485977	47.2
6	bean	+	20	0.4786704	0.1485977	47.2
7	trefoil	-	5	-2.4730339	0.1485977	47.2
8	trefoil	-	10	-2.0146309	0.1485977	47.2
9	trefoil	-	20	-1.6140098	0.1485977	47.2
10	trefoil	+	5	-1.8887476	0.1485977	47.2
11	trefoil	+	10	-1.4675441	0.1485977	47.2
12	trefoil	+	20	-0.3905297	0.1485977	47.2
		upper.halfLeastSignificant.limit	lower.halfLeastSignificant.limit	est.status		
1		-1.4081535		Estimable	-1.7995140	
2		-0.9497506		Estimable	-1.3411111	
3		-0.5491295		Estimable	-0.9404900	
4		-0.8238673		Estimable	-1.2152278	
5		-0.4026637		Estimable	-0.7940242	
6		0.6743507		Estimable	0.2829901	
7		-2.2773537		Estimable	-2.6687142	
8		-1.8189507		Estimable	-2.2103112	
9		-1.4183296		Estimable	-1.8096901	
10		-1.6930674		Estimable	-2.0844279	
11		-1.2718638		Estimable	-1.6632243	
12		-0.1948495		Estimable	-0.5862100	

LSD values

```

minimum LSD = 0.3913605 0.3913605

mean LSD = 0.3913605 0.3913605

maximum LSD = 0.3913605 0.3913605

(sed range / mean sed = 2.45e-14 2.41e-14 )

```

The above LSD values can only be used to compare pairs of EMMs for the same Host.

## Explore the LSDs

To investigate the errors that would result from using the overall LSDs as opposed to the LSDs computed for each Host, the `exploreLSDs`, `pickLSDstatistics` and `plotLSDerrors` functions are used, firstly with the default value of "overall" for `LSDtype` and finally with the `LSDtype` set to "factor.combination" and `LSDby` to "Host".

The `exploreLSDs` function produces tables of statistics for the LSDs computed for the settings of the `LSDtype` and `LSDby`; the settings of these arguments does not have to match those used in producing the `alldiffs` object. For `LSDtype` set to "overall", a single LSD statistic is computed that is based on the standard errors of all pairwise differences. To ascertain the errors that arise from using this LSD value for determining, for all pairwise comparisons, whether a comparison is significant, `exploreLSDs` compares the results using the LSD value with the *p*-values in the `p.differences` component of the `alldiffs` object. For `LSDtype` set to "factor.combination" and `LSDby` to "Host", the LSD statistics are calculated from standard errors of the pairwise differences for each Host. Examination of the `sed` component of `diffs` reveals that there are only three different values for the standard errors of pairwise differences and, hence, only three unique values for the LSD. Of the statistics shown, there is no single value that will not result in errors.

Here are the tables of the numbers of false positive and negative error in using the values of the various LSD statistics for determining the significance of the 66 pairwise comparisons of the 12 predictions.

```
exploreLSDs(diffs, LSDtype = "overall")
```

```
#### Statistics calculated from LSD values
```

c	min	quant10	quant25	mean	median	quant75	quant90	max
1 66	0.2259521	0.3913605	0.3913605	0.4087627	0.3913605	0.4519042	0.4519042	0.4519042

```
#### False positives resulting from the use of various LSD statistics
```

c	min	quant10	quant25	mean	median	quant75	quant90	max	
false.pos	66	7	2	2	2	2	0	0	0

```
#### False negatives resulting from the use of various LSD statistics
```

c	min	quant10	quant25	mean	median	quant75	quant90	max	
false.neg	66	0	0	0	2	0	4	4	4

The function `findLSDminerrors` can be used to find a value of the LSD that minimizes the number of false positive (negative) errors i.e. declaring a pairwise difference to be significant (not significant) when it is not significant (significant). The function has an argument `false.pos.wt` that specifies how many false negatives are equivalent to one false positive; the default for `false.pos.wt` is 10, which is a conservative approach because it greatly favours false negatives over false positives.

```
(findLSDminerrors(diffs))
```

LSD	false.pos	false.neg	false.criterion	
overall	0.4486447	0	4	4

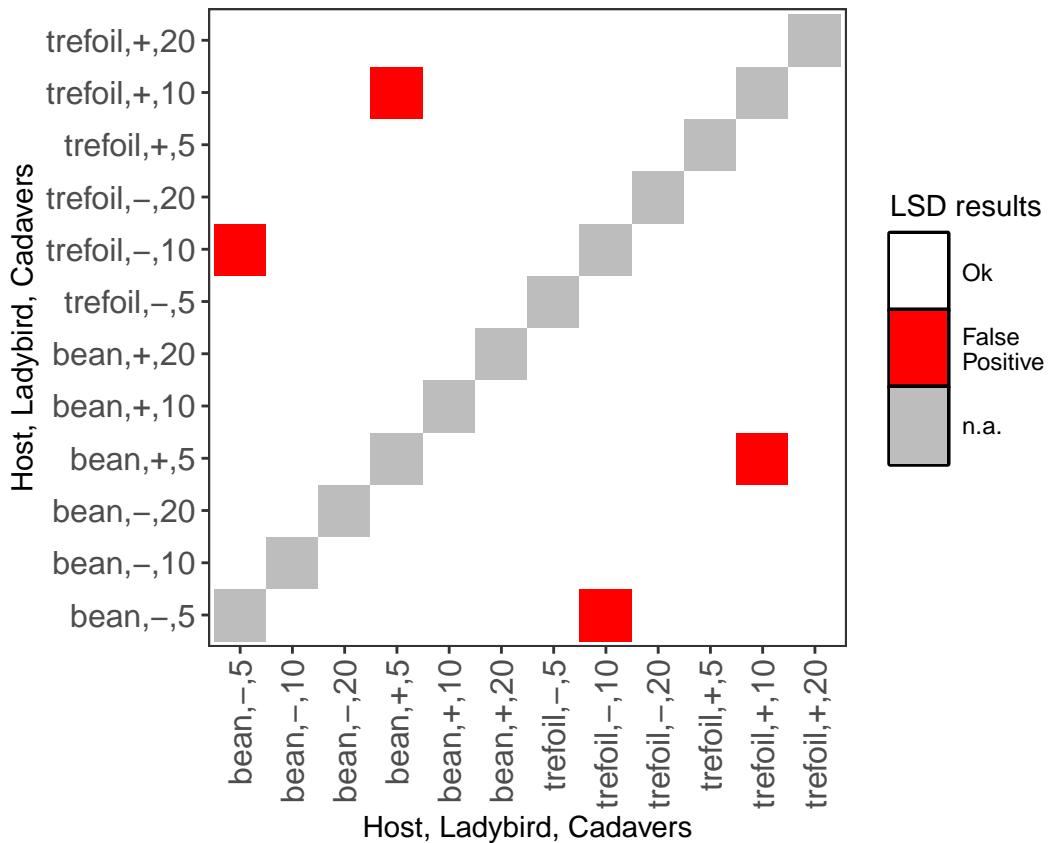
```
(minLSDpos <- findLSDminerrors(diffs, false.pos.wt = 1))
```

LSD	false.pos	false.neg	false.criterion
overall	0.3547448	2	0
			2

The smallest value of the LSD that results in the minimum weighted sum of the false positives and negatives is 0.4489466 when the weight is 10 and is 0.354983 when the weight is one. These values result in 4 false negatives and 2 false positives respectively.

To see which of the two pairwise comparisons will be falsely identified as being significant when the LSD value is set to minimum LSD, the LSDs stored in the `alldiffs` object need to be recalculated to be based on this LSD value. Then plot the errors or save the return values obtained using the function `plotLSDerrors`. The plot below shows that the two pairs whose differences are incorrectly identified as significant have the same level of the Ladybird factor, but differ in both of the levels for the Host and Cadaver factors.

```
diffs.overall <- recalcLSD(diffs, LSDtype = "supplied", LSDsupplied = minLSDpos["LSD"])
plotLSDerrors(diffs.overall)
```



This raises the question of whether the minimum LSD should be used. There are at least four alternatives: (i) use it without restriction, on the basis that it can be concluded that using it is unlikely to result in seriously flawed conclusions; (ii) use it with the restriction that it only be applied to pairwise comparisons that have the same Host or the same Cadaver treatment; (i11) investigate the use of the minimum LSD with only false negative errors; and (iv) rather than use an overall LSD value, use LSD values computed from the LSDs within each Host level.

Because `LSDtype` was set to "factor.combination" and `LSDby` to "Host" in forming the object `diffs`, the LSDs for alternative (iv) are stored in the `LSD` component of the object `diffs`. Printing out the LSD

component will summarize how those LSD values perform. Otherwise, the following call to `exploreLSDs` will display the properties of the LSDs for various LSD statistics:

```
exploreLSDs(diffs, LSDtype = "factor.combination", LSDby = "Host")
```

The following shows the contents of the LSD component of `diffs`:

```
(diffs$LSD)
```

	c	minLSD	meanLSD	maxLSD	assignedLSD	accuracyLSD	falsePos	falseNeg
bean	15	0.3913605	0.3913605	0.3913605	0.3913605	1.886492e-14	0	0
trefoil	15	0.3913605	0.3913605	0.3913605	0.3913605	1.843939e-14	0	0

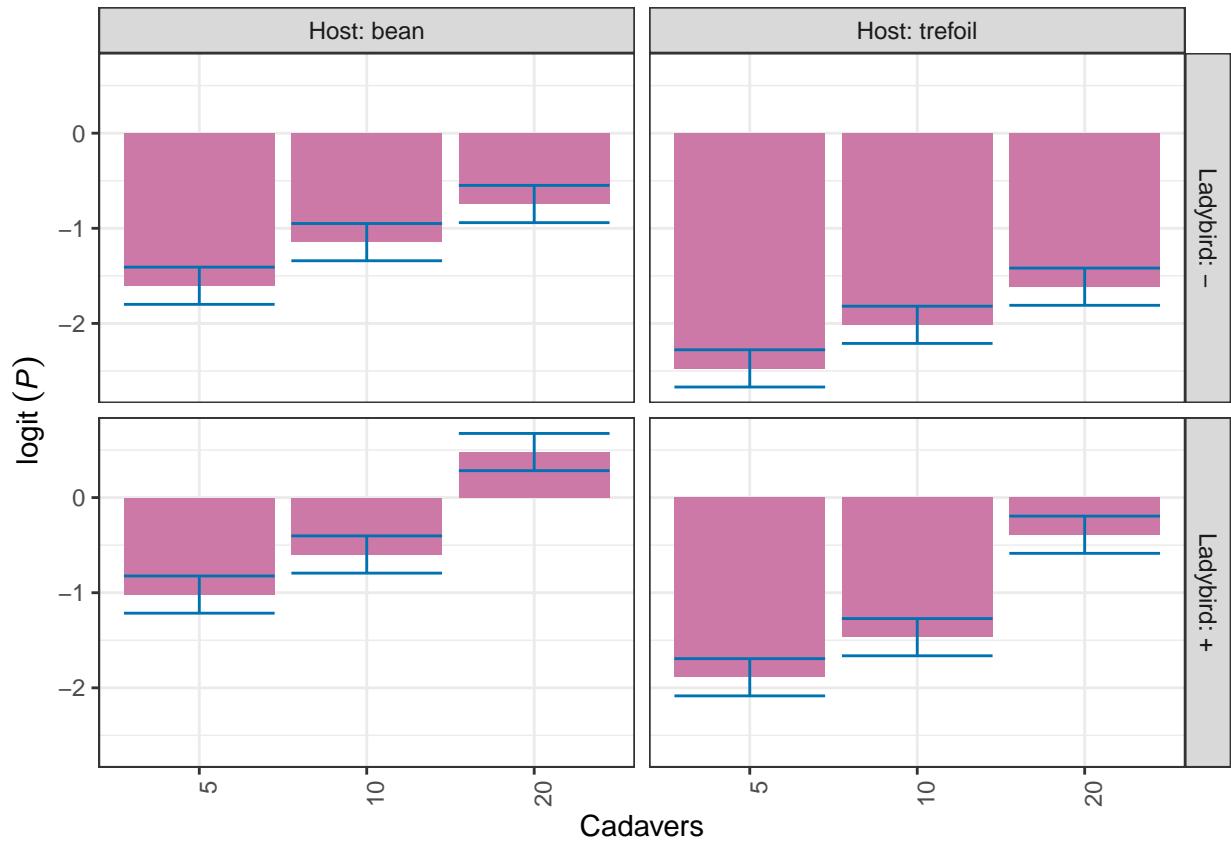
Because the minimum and maximum LSD values are equal, it follows that there is only one value of the LSD for all pairwise comparisons within each Host level and it happens that the values for the two Hosts are also equal. The table shows that zero false positive and negative errors will result from the use of the value of 0.39 for the 30 within-Host comparisons. Thus, using `findLSDminerrors(diffs, LSDtype = "factor.combinations", LSDby = "Host")` to select the minimum LSDs would only confirm what is already obvious.

## Plot the predictions

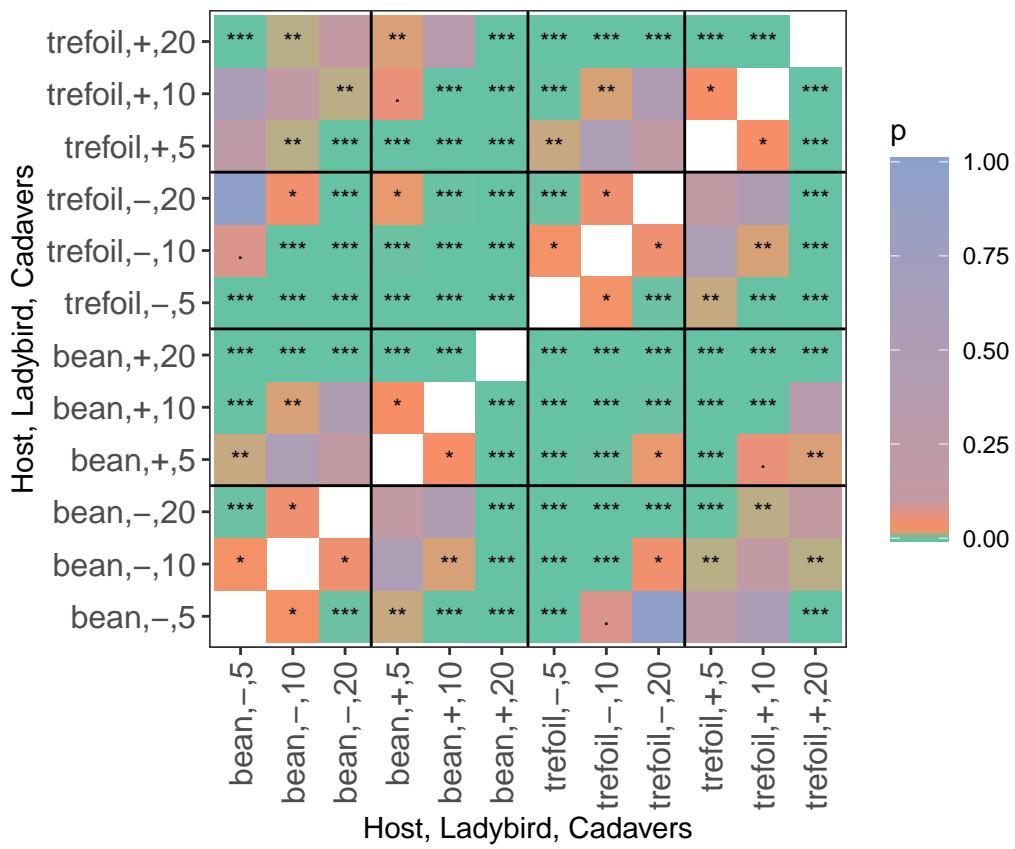
```
titl <- str2expression("logit~italic(P))")
names(titl) <- "logitP"
plotPredictions(diffs$predictions, y ="predicted.value",
                 y.title = titl,
                 classify = "Host:Ladybird:Cadavers",
                 error.intervals = "halfLeast", interval.annotate = FALSE,
                 ggplotFuncs = list(facet_grid(Ladybird ~ Host,
                                               labeller = label_both)))
```

The function `plotPredictions` uses `ggplot` to produce the plot and the `ggplotFuncs` argument allows the addition of `ggplot` functions to modify the plot. In this case, the `facet.grid` function is respecified to include prepender functions that modify the labels of the facets to include the factor names. Note the statement in the legend of Figure 1 that restricts the use of the error bars to determining the significance of differences for the pairwise comparisons of EMMs for the same Host.

```
plotPvalues(diffs, factors.per.grid = 1, show.sig = TRUE)
```



**Figure 1.** Estimated marginal means (EMMs) for  $\text{logit}(P)$ , where  $P$  is the proportion of live aphids that were infected, for two Hosts, two Ladybird levels and three Cadaver levels. Error bars are an EMM  $\pm$  half-LSD (5%). The two EMMs for the same Host are significantly different ( $p \leq 0.05$ ) if their error bars do not overlap.



**Figure 2.** The  $p$ -values for each of the pairwise comparisons of the estimated marginal means for  $\text{logit}(P)$ , where  $P$  is the proportion of live aphids that were infected, for two Hosts, two Ladybird levels and three Cadaver levels

```
options(width = 90)
diffs$differences
```

	bean,-,5	bean,-,10	bean,-,20	bean,+,5	bean,+,10	bean,+,20
bean,-,5	0.00000000	-0.4584030	-0.8590241	-0.5842863	-1.0054898	-2.0825042
bean,-,10	0.45840297	0.0000000	-0.4006211	-0.1258833	-0.5470869	-1.6241012
bean,-,20	0.85902408	0.4006211	0.0000000	0.2747378	-0.1464657	-1.2234801
bean,+,5	0.58428627	0.1258833	-0.2747378	0.0000000	-0.4212036	-1.4982179
bean,+,10	1.00548982	0.5470869	0.1464657	0.4212036	0.0000000	-1.0770144
bean,+,20	2.08250420	1.6241012	1.2234801	1.4982179	1.0770144	0.0000000
trefoil,-,5	-0.86920012	-1.3276031	-1.7282242	-1.4534864	-1.8746899	-2.9517043
trefoil,-,10	-0.41079715	-0.8692001	-1.2698212	-0.9950834	-1.4162870	-2.4933014
trefoil,-,20	-0.01017604	-0.4685790	-0.8692001	-0.5944623	-1.0156659	-2.0926802
trefoil,+,5	-0.28491385	-0.7433168	-1.1439379	-0.8692001	-1.2904037	-2.3674180
trefoil,+,10	0.13628970	-0.3221133	-0.7227344	-0.4479966	-0.8692001	-1.9462145
trefoil,+,20	1.21330408	0.7549011	0.3542800	0.6290178	0.2078143	-0.8692001
	trefoil,-,5	trefoil,-,10	trefoil,-,20	trefoil,+,5	trefoil,+,10	trefoil,+,20
bean,-,5	0.8692001	0.4107972	0.01017604	0.2849139	-0.1362897	-1.2133041
bean,-,10	1.3276031	0.8692001	0.46857901	0.7433168	0.3221133	-0.7549011
bean,-,20	1.7282242	1.2698212	0.86920012	1.1439379	0.7227344	-0.3542800
bean,+,5	1.4534864	0.9950834	0.59446231	0.8692001	0.4479966	-0.6290178
bean,+,10	1.8746899	1.4162870	1.01566586	1.2904037	0.8692001	-0.2078143
bean,+,20	2.9517043	2.4933014	2.09268024	2.3674180	1.9462145	0.8692001
trefoil,-,5	0.0000000	-0.4584030	-0.85902408	-0.5842863	-1.0054898	-2.0825042
trefoil,-,10	0.4584030	0.0000000	-0.40062111	-0.1258833	-0.5470869	-1.6241012
trefoil,-,20	0.8590241	0.4006211	0.0000000	0.2747378	-0.1464657	-1.2234801
trefoil,+,5	0.5842863	0.1258833	-0.27473781	0.0000000	-0.4212036	-1.4982179
trefoil,+,10	1.0054898	0.5470869	0.14646574	0.4212036	0.0000000	-1.0770144
trefoil,+,20	2.0825042	1.6241012	1.22348012	1.4982179	1.0770144	0.0000000

```
options(width = 90)
print(diffs$sed)
```

	bean,-,5	bean,-,10	bean,-,20	bean,+,5	bean,+,10	bean,+,20	trefoil,-,5
bean,-,5	NA	0.1945600	0.1945600	0.1945600	0.1945600	0.1945600	0.1123293
bean,-,10	0.1945600	NA	0.1945600	0.1945600	0.1945600	0.1945600	0.2246586
bean,-,20	0.1945600	0.1945600	NA	0.1945600	0.1945600	0.1945600	0.2246586
bean,+,5	0.1945600	0.1945600	0.1945600	NA	0.1945600	0.1945600	0.2246586
bean,+,10	0.1945600	0.1945600	0.1945600	0.1945600	NA	0.1945600	0.2246586
bean,+,20	0.1945600	0.1945600	0.1945600	0.1945600	0.1945600	NA	0.2246586
trefoil,-,5	0.1123293	0.2246586	0.2246586	0.2246586	0.2246586	0.2246586	NA
trefoil,-,10	0.2246586	0.1123293	0.2246586	0.2246586	0.2246586	0.2246586	0.1945600
trefoil,-,20	0.2246586	0.2246586	0.1123293	0.2246586	0.2246586	0.2246586	0.1945600
trefoil,+,5	0.2246586	0.2246586	0.2246586	0.1123293	0.2246586	0.2246586	0.1945600
trefoil,+,10	0.2246586	0.2246586	0.2246586	0.2246586	0.1123293	0.2246586	0.1945600
trefoil,+,20	0.2246586	0.2246586	0.2246586	0.2246586	0.2246586	0.1123293	0.1945600
	trefoil,-,10	trefoil,-,20	trefoil,+,5	trefoil,+,10	trefoil,+,20		
bean,-,5	0.2246586	0.2246586	0.2246586	0.2246586	0.2246586		
bean,-,10	0.1123293	0.2246586	0.2246586	0.2246586	0.2246586		
bean,-,20	0.2246586	0.1123293	0.2246586	0.2246586	0.2246586		
bean,+,5	0.2246586	0.2246586	0.1123293	0.2246586	0.2246586		
bean,+,10	0.2246586	0.2246586	0.2246586	0.1123293	0.2246586		
bean,+,20	0.2246586	0.2246586	0.2246586	0.2246586	0.1123293		
trefoil,-,5	0.1945600	0.1945600	0.1945600	0.1945600	0.1945600		
trefoil,-,10	NA	0.1945600	0.1945600	0.1945600	0.1945600		
trefoil,-,20	0.1945600	NA	0.1945600	0.1945600	0.1945600		
trefoil,+,5	0.1945600	0.1945600	NA	0.1945600	0.1945600		
trefoil,+,10	0.1945600	0.1945600	0.1945600	NA	0.1945600		
trefoil,+,20	0.1945600	0.1945600	0.1945600	0.1945600	NA		

## Perform the analysis with just the selected model fitted

The model with nonsignificant fixed terms dropped is obtained in order to compare it with the fit when they are retained and the estimated marginal means for the chosen model are obtained.

```
m.sig.lm <- lm(logitP ~ Run + Cadavers*Ladybird + Host,
                 data=Ladybird.dat)
(aov.tab <- anova(m.sig.lm))
```

Analysis of Variance Table

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
Run	1	0.0677	0.0677	0.3029	0.58398
Cadavers	2	17.0274	8.5137	38.1160	1.255e-11
Ladybird	1	11.0907	11.0907	49.6531	1.542e-09
Host	1	13.5992	13.5992	60.8836	7.179e-11
Cadavers:Ladybird	2	1.7349	0.8675	3.8836	0.02559
Residuals	64	14.2952	0.2234		

```
HCL.emm <- emmeans::emmeans(m.sig.lm, specs = ~ Host:Cadavers:Ladybird)
HCL.preds <- summary(HCL.emm)
den.df <- min(HCL.preds$df)
HCL.vcov <- vcov(HCL.emm)
HCL.preds <- as.predictions.frame(HCL.preds, predictions = "emmmean",
                                    se = "SE", interval.type = "CI",
                                    interval.names = c("lower.CL", "upper.CL"))
diffs.red <- allDifferences(predictions = HCL.preds, classify = "Host:Ladybird:Cadavers",
                               vcov = HCL.vcov, tdf = den.df)
diffs.red <- redoErrorIntervals(diffs, error.intervals = "halfLeast",
                                 LSDtype = "factor.combination", LSDby = "Host")
```

Joining with ‘by = join\_by(fac.comb)’  
Joining with ‘by = join\_by(Host)’

```
options(width = 90)
print(diffs.red$sed)
```

	bean,-,5	bean,-,10	bean,-,20	bean,+,5	bean,+,10	bean,+,20	trefoil,-,5
bean,-,5	NA	0.1945600	0.1945600	0.1945600	0.1945600	0.1945600	0.1123293
bean,-,10	0.1945600	NA	0.1945600	0.1945600	0.1945600	0.1945600	0.2246586
bean,-,20	0.1945600	0.1945600	NA	0.1945600	0.1945600	0.1945600	0.2246586
bean,+,5	0.1945600	0.1945600	0.1945600	NA	0.1945600	0.1945600	0.2246586
bean,+,10	0.1945600	0.1945600	0.1945600	0.1945600	NA	0.1945600	0.2246586
bean,+,20	0.1945600	0.1945600	0.1945600	0.1945600	0.1945600	NA	0.2246586
trefoil,-,5	0.1123293	0.2246586	0.2246586	0.2246586	0.2246586	0.2246586	NA
trefoil,-,10	0.2246586	0.1123293	0.2246586	0.2246586	0.2246586	0.2246586	0.1945600
trefoil,-,20	0.2246586	0.2246586	0.1123293	0.2246586	0.2246586	0.2246586	0.1945600
trefoil,+,5	0.2246586	0.2246586	0.2246586	0.1123293	0.2246586	0.2246586	0.1945600
trefoil,+,10	0.2246586	0.2246586	0.2246586	0.2246586	0.1123293	0.2246586	0.1945600
trefoil,+,20	0.2246586	0.2246586	0.2246586	0.2246586	0.2246586	0.1123293	0.1945600
trefoil,-,10	trefoil,-,20	trefoil,+,5	trefoil,+,10	trefoil,+,20			

bean,-,5	0.2246586	0.2246586	0.2246586	0.2246586	0.2246586
bean,-,10	0.1123293	0.2246586	0.2246586	0.2246586	0.2246586
bean,-,20	0.2246586	0.1123293	0.2246586	0.2246586	0.2246586
bean,+,5	0.2246586	0.2246586	0.1123293	0.2246586	0.2246586
bean,+,10	0.2246586	0.2246586	0.2246586	0.1123293	0.2246586
bean,+,20	0.2246586	0.2246586	0.2246586	0.2246586	0.1123293
trefoil,-,5	0.1945600	0.1945600	0.1945600	0.1945600	0.1945600
trefoil,-,10	NA	0.1945600	0.1945600	0.1945600	0.1945600
trefoil,-,20	0.1945600	NA	0.1945600	0.1945600	0.1945600
trefoil,+,5	0.1945600	0.1945600	NA	0.1945600	0.1945600
trefoil,+,10	0.1945600	0.1945600	0.1945600	NA	0.1945600
trefoil,+,20	0.1945600	0.1945600	0.1945600	0.1945600	NA

## References

- Brien, C. J. (2025a) **asremlPlus**: *Augments ASReml-R in fitting mixed models and packages generally in exploring prediction differences*. Version 4.4.55. <https://cran.r-project.org/package=asremlPlus/> or <http://chris.brien.name/rpackages/>.
- Brien, C. J. (2025b) **dae**: *Functions useful in the design and ANOVA of experiments*. Version 3.2.33. <https://cran.r-project.org/package=dae/> or <http://chris.brien.name/rpackages/>.
- Kuznetsova, A., Brockhoff, P. B. and Christensen, R. H. B. (2017) **lmerTest** Package: Tests in Linear Mixed Effects Models. *Journal of Statistical Software*, **82**, 1–26.
- Lenth, R. V., Banfa, B., Bolker, B., Buerkner, P., Giné-Vázquez, I., Herve, M. J., Jung, M., Love, J., Miguez, F., Piaskowski, J., Riebl, H., & Singmann, H. (2025) **emmeans**: *Estimated Marginal Means, aka Least-Squares Means*. Version 1.11.0. <https://cran.r-project.org/package=emmeans/>.
- Littell, R. C., Milliken, G. A., Stroup, W. W., Wolfinger, R. D., & Schabenberger, O. (2006). *SAS for Mixed Model.* (2nd ed.). Cary, N.C.: SAS Press.
- R Core Team (2025) **R**: *A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. <https://www.r-project.org>.
- Snee, R. D. (1981). Graphical Display and Assessment of Means. *Biometrics*, **37**, 835–836.
- Welham, S. J., Gezan, S. A., Clark, S. J., & Mead, A. (2014). *Statistical Methods in Biology: Design and Analysis of Experiments and Regression*. Boca Raton: Chapman and Hall/CRC.