

Seed ecology of European mesic meadows: the domestication of plant regeneration strategies

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**RESEARCH IN CONTEXT: Seed ecology of European mesic meadows:
the domestication of plant regeneration strategies**

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ABSTRACT (300 words)

- **Background and Aims** European mesic meadows are semi-natural open habitats of high biodiversity and an essential part of European landscapes. Their species-rich seed

mixes can be a source of plant material for ecological restoration, urban greening and rewilding. Here, we make a synthesis of the seed ecology of mesic meadows.

- **Methods** We combined our own experimental data with data obtained from databases to create a combined dataset containing 1,940 germination records of 104 plant species from 32 European countries. We performed a Bayesian meta-analysis of this dataset to test the seed germination response to environmental cues including scarification, stratification, temperature, alternating temperature and light. We also used multivariate ordination to check the relationship between seed traits (germination and morphology) and species ecological preferences, and to compare the seed ecology of mesic meadows with that of other herbaceous plant communities from the same area.

- **Key Results** The seed ecology of mesic meadows is characterized by (1) high seed germinability when compared to other herbaceous plant communities; (2) low correspondence between seed traits and species ecological preferences; and (3) a deep phylogenetic separation between the two major families, Poaceae and Fabaceae. Poaceae produce many light seeds which respond to gap-detecting germination cues (alternating temperatures and light); Fabaceae produce fewer heavy seeds, which need scarification to break their physical dormancy.

- **Conclusions** High germinability of meadow seeds will reduce their capacity to form persistent seed banks, resulting in dispersal limitations to passive regeneration. For centuries, human activities have shaped the regeneration of meadows, leading to a loss of seed dormancy and decoupling seeds from seasonal cycles, as has been found in many domesticated species. The same anthropic processes that have shaped semi-

natural mesic meadows have left them dependent on continued human intervention for their regeneration, highlighting the importance of active restoration via seed supply.

KEYWORDS

Arrhenatheretalia, *Arrhenatherion*, Asteraceae, Fabaceae, domestication, hay meadows, mesic grasslands, Poaceae, seed germination, seed morphology, semi-natural, species-rich meadows

INTRODUCTION

The mesic meadows of Europe are semi-natural open habitats that occupy clearings created by human intervention over a temperate wooded landscape (Poschlod *et al.* 2009; Hejerman *et al.* 2013), in sites with moderately fertile and well-drained soils (Mucina *et al.* 2016). Once, these were novel plant communities, with a composite flora made up of species from different ecological backgrounds; during historical times, their species composition further evolved following changes in human events and management practices (Chytrý 2012; Hejerman *et al.* 2013). Mesic meadows are thus the result of a process of habitat domestication that is characteristic of Europe's natural history (Flannery 2018). Nowadays, mesic meadows maintained by agricultural practices are an essential aspect of European landscapes, both real (Finck *et al.* 2002) and imagined (Judd and Judd 2017).

Traditional classification of mesic grasslands emphasises the separation between meadows used for hay making versus pastures maintained by grazing, but a recent revision at the European level showed that the main driver of variation in species composition is the

1 intensity rather than the type of management (Rodríguez-Rojo *et al.* 2017). Indeed, changes
2 in the intensity of management, including land abandonment and agricultural
3 intensification, are threatening the maintenance of mesic meadows in large parts of Europe
4 (Carboni *et al.* 2015). For this reason, the European Habitats Directive (92/43/EEC) lists as
5 habitats of conservation interest those species-rich meadows that are traditionally managed
6 by one or two annual cuts and light grazing.

7 One of the reasons for the high conservation interest of traditional mesic meadows is their
8 high species richness, and for this same reason they have been highlighted as a valuable
9 source of natural seed material to be used in ecological restoration, rewilding and urban
10 greening (Krautzer *et al.* 2013; Haslgrübler *et al.* 2014; Golińska *et al.* 2017). However, a
11 lack of knowledge about species germination traits has already been identified as a
12 bottleneck (Ladouceur *et al.* 2018) hampering the development of a competitive native seed
13 industry (De Vitis *et al.* 2017) in Europe. Understanding seed germination is part of the
14 practical scientific framework needed to tackle large-scale ecological restoration challenges
15 (Merritt and Dixon 2011) and to maintain *ex situ* collections of plant genetic resources (Li
16 and Pritchard 2009). When such a knowledge framework is available, ambitious regional
17 schemes of seed-based landscape restoration can be designed (Jiménez-Alfaro *et al.* 2020).

18 Mesic meadows are dominated by mesophilous grasses of the family Poaceae, which make
19 up most of their biomass and define the structure of the vegetation. The dispersal unit in
20 Poaceae is generally the floret (**Fig. 1a**), a composite structure made up of the caryopsis or
21 grain (a dry, indehiscent and monospermic fruit) and its surrounding bracts (i.e. modified
22 leaves): the lemma and the palea. Removal of the lemma and palea, as well as puncturing

the pericarp, can aid in seed germination (Probert *et al.* 1985). The dispersal unit often carries awns or hairs (**Fig. 1b**) that are assumed to aid dispersal, although it is not always the case that they do (Schonfeld 1983). Meadow Poaceae seeds have been reported as having physiological seed dormancy in various degrees (Sprague 1940; Dixon 1995; Baskin and Baskin 2014), but germinability is usually high even without treating the seeds with cold stratification (Grime *et al.* 1981; Schonfeld 1983; Williams 1983a; Bean 1984; Froud-Williams *et al.* 1984, 1986; Froud-Williams 1987; Dixon 1995; Pérez-Fernández and Rodríguez-Echeverría 2003; Pérez-Fernández *et al.* 2006; Stanisavljevic *et al.* 2011, 2015; Oliveira *et al.* 2012; Wille *et al.* 2013). Freshly harvested seeds are comparatively more dormant, but dormancy tends to disappear quickly in dry storage (Sprague 1940; Dixon 1995). Germination has been reported to occur at temperatures ranging from 5 to 30 °C (Grime *et al.* 1981; Williams 1983b; Pannangpetch and Bean 1984; Froud-Williams *et al.* 1986; Probert *et al.* 1986; Dixon 1995). Most Poaceae species have also been reported to germinate better in light than in darkness (Williams 1983b; c; Froud-Williams *et al.* 1984; Probert *et al.* 1985, 1986; Probert 1986; Thompson 1989; Dixon 1995) and even to be unable to germinate in darkness (Froud-Williams *et al.* 1986). Nonetheless, species of *Bromus* have been reported as germinating better in darkness (Thompson 1989) and *Cynosurus cristatus* as being indifferent to light/darkness (Williams 1983b; c). In *Poa trivialis*, germination is promoted by light but not by diurnal alternating temperatures (i.e. germination conditions where different temperatures are applied during the day and the night, in diurnal cycles) (Froud-Williams 1987), although alternating temperatures do encourage some germination in darkness (Froud-Williams *et al.* 1986). Wild Poaceae ecotypes usually have a germination response to alternating temperatures (Schonfeld 1983;

Williams 1983b; c; Pannangpetch and Bean 1984; Probert *et al.* 1985, 1986; Probert 1986; Thompson 1989), although this response is missing in some wild ecotypes and in the domesticated cultivars (Pannangpetch and Bean 1984), and some species such as *Lolium perenne* have been reported as insensitive to temperature alternation (Thompson 1977; Williams 1983c).

Next in abundance to Poaceae are the legumes of the family Fabaceae. Fabaceae species contribute to the nutritional value of meadow fodder, as thanks to their N-fixating capabilities they have high N contents (Reiné *et al.* 2020; Álvarez *et al.* 2021). The dispersal unit of most Fabaceae is the seed itself (**Fig. 1c**) but in some species dispersal units are more complex, including indehiscent monospermic fruits (e.g. *Onobrychis*, **Fig. 1d**) or indehiscent monospermic fruit fragments, i.e. lomentes (e.g. *Ornithopus*, **Fig. 1e**). Fabaceae are generally hard-seeded: they have a water-impermeable seed coat which needs to become permeable before germination can happen (i.e. physical seed dormancy) (Grime *et al.* 1981; Jones 1986; Ehrman and Cocks 1996; Kupferschmid *et al.* 2000; Baskin and Baskin 2014). In *Medicago*, seeds that have not reached full maturity can germinate before they become impermeable, but the completion of maturation imposes coat impermeability, and thereafter the seed must be scarified to allow water imbibition and germination (Gresta *et al.* 2007). Buried Fabaceae seeds can track the seasons, and in some species, germination seems to be promoted by cold stratification and alternating temperatures (Van Assche *et al.* 2003); some of these species have been described as having combinational dormancy (i.e. physical + physiological) (Van Assche and Vandeloos 2010). However, Fabaceae seeds have also been reported to germinate without any previous treatment (Marchiol *et al.*

2000; Nikolic *et al.* 2007; Kabouw *et al.* 2010; Oliveira *et al.* 2012), and to lose dormancy during storage (Van Assche and Vandeloek 2010). As in Poaceae, seeds of Fabaceae have been reported to germinate in high numbers across a range of temperatures from 5 to 25 °C (Grime *et al.* 1981; Gresta *et al.* 2007). Fabaceae seeds have been described as not responsive to light and capable of germinating in darkness (Silvertown 1980; Grime *et al.* 1981).

Poaceae and Fabaceae are accompanied by a diversity of other families which, even if present in lower abundances, contribute to the high biodiversity of the system and increase the aesthetic value of the meadows as perceived by people (Southon *et al.* 2017; Chollet *et al.* 2018). They also add nutritional scope, being richer than Poaceae and Fabaceae in specific elements (Reiné *et al.* 2020; Álvarez *et al.* 2021). In many of these families (e.g. Asteraceae, Cyperaceae, Dipsacaceae, Lamiaceae, Polygonaceae, Ranunculaceae) the dispersal unit is the achene: dry, indehiscent and monospermic fruits. The morphology of these achenes is varied: cypselae with a hairy pappus in *Centaurea* (Asteraceae, **Fig. 1f**); beaked in *Ranunculus* (Ranunculaceae, **Fig. 1g**); hardened nutlets in *Prunella* (Lamiaceae, **Fig. 1h**); hairy and with an elaiosome in *Knautia* (Dipsacaceae, **Fig. 1i**); surrounded by a perigynium which aids in dispersal by water in *Carex* (Cyperaceae, **Fig. 1j**). In the genus *Sanguisorba* (Rosaceae), the dispersal unit is the urn-shaped receptacle containing one to three achenes (**Fig. 1k**). In Apiaceae, it is the mericarp (**Fig. 1l**), an indehiscent monospermic fragment of the fruit. In some other minor families, the dispersal unit is the seed itself, such as in Caryophyllaceae (**Fig. 1m**), Plantaginaceae (**Fig. 1n**), Juncaceae or the hemiparasitic species of *Rhinanthus* (Orobanchaceae) (**Fig. 1o**). In Asteraceae, high

1 germination without previous treatments has been reported in *Taraxacum officinale*
2 (Mezynski 1974; Washitani 1984; Noronha 1997; Benvenuti and Pardossi 2016; Masin *et*
3 *al.* 2017), *Hypochaeris radicata* (Oomes 1976; Benvenuti and Pardossi 2016) and *Achillea*
4 *millefolium* (Oomes 1976). *Taraxacum officinale* germinates between 5 and 30 °C (Masin
5 *et al.* 2017; RN1428; Washitani 1984) and has higher germination in light (Thompson
6 1989; Letchamo 1996; Noronha 1997) and in alternating temperatures (Mezynski 1974). In
7 *Stachys officinalis* (Lamiaceae), seeds need either cold stratification, light or alternating
8 temperatures to germinate (Wagner 2011; Kolodziejek *et al.* 2017). Underdeveloped
9 embryos that need to grow inside the seed before germination (i.e. morphological
10 dormancy) are widespread in Ranunculaceae and Apiaceae (Jauzein and Mansour 1992;
11 Baskin and Baskin 2014). *Ranunculus repens* (Ranunculaceae) germinates between 10 and
12 25 °C, but the germination percentages have been reported to be low (Harris 1998); the
13 same species has been reported to respond to alternating temperatures, which can promote
14 its germination even in darkness (Thompson and Grime 1983). In Polygonaceae, *Rumex*
15 *acetosa* can germinate immediately after dispersal and between 7 and 27 °C, while the
16 congeneric *Rumex acetosella* does not: this difference is due to the former being able to
17 germinate at constant temperatures in the darkness (Grime *et al.* 1981; Van Assche *et al.*
18 2002), while the latter has an absolute requirement for light (Van Assche *et al.* 2002). In
19 *Heracleum sphondylium* (Apiaceae), growth of the embryo only occurs below 10 °C, in
20 moist conditions (Jauzein and Mansour 1992). *Sanguisorba minor* (Rosaceae) increases its
21 germination after abrasion of the seeds with bleach (Tavşanoğlu *et al.* 2015; Benvenuti and
22 Pardossi 2016), although germination without previous treatment has also been reported
23 (Ludewig *et al.* 2014; Tavşanoğlu *et al.* 2015). Seeds of the hemiparasitic species

Rhinanthus angustifolius and *Rhinanthus minor* (Orobanchaceae) require relatively long periods of cold stratification to germinate (Ter Borg 2005) and can germinate in the dark (Marin *et al.* 2019).

Although a wealth of studies has accumulated, a synthesis of the seed ecology of European mesic meadows is still missing. In this article, we review for the first time this topic, combining newly generated data on seed morphology and germination with records from existing databases (Kleyer *et al.* 2008; Royal Botanic Gardens, Kew 2017; Carta *et al.* 2021; Fernández-Pascual 2021). The resulting dataset contains 1,940 germination records of 104 plant species from 32 European countries. We use this dataset to test the seed germination response to environmental cues including scarification, stratification, temperature, alternating temperature and light, applying Bayesian meta-analysis (Pappalardo *et al.* 2020). Further, using well-preserved meadows of the Iberian Peninsula as a study system, we analyse the covariation between seed traits and species environmental preferences, and compare the germination ecology of mesic meadows with that of other herbaceous plant communities from the same geographic area.

MATERIALS AND METHODS

Selection of mesic meadow species

To create a list of representative mesic meadow species for inclusion in our analysis, we used a dataset comprising 118 vegetation relevés (i.e. records of plants species co-occurring in sampling plots) from three Western European regions with well-maintained mesic meadows: 43 relevés from Northern Portugal, 25 from the Cantabrian Mountains of Spain

1 and 50 from the Pyrenees. These relevés contain a sample of mesic meadow diversity, as
2 they were recorded along a major stress gradient related to summer drought (Rodríguez-
3 Rojo *et al.* 2014): the Pyrenees and Cantabrian Mountains have a temperate macroclimate,
4 whereas Northern Portugal is transitional between the temperate and Mediterranean
5 macroclimates. Furthermore, meadows from the Pyrenees are closest to the European
6 optimum of mesic meadow vegetation while the Portuguese ones are in suboptimal areas at
7 the limit of the European distribution of temperate meadows (Rodríguez-Rojo *et al.* 2017).
8 Finally, the traditional management of meadows (i.e. mowing for haymaking once or twice
9 per year plus light grazing) is relatively well preserved in these three regions compared to
10 their European context (Prince *et al.* 2012; Guadilla-Sáez *et al.* 2019).

11 The studied meadows were maintained by traditional agricultural practices: mowing for
12 haymaking once or twice per year plus light grazing. Each vegetation sampling plot was
13 placed in a square area (25-100 m² area) situated in the central part of the meadow,
14 avoiding the margins. Vegetation sampling took place in 2016-2017, at the peak of plant
15 development, just before mowing. All vascular plant species in the plots were recorded and
16 given a cover value using the transformation of the Braun-Blanquet scale to coverage (+ =
17 0.1%, 1 = 5%, 2 = 17.5%, 3 = 37.5%, 4 = 62.5% and 5 = 87.5%). All plant names were
18 assigned following the nomenclature of Euro+Med (2006), which is used throughout this
19 article. As expected, the vegetation of the sampled meadows was dominated by Poaceae
20 and Fabaceae: these two families represented 47% and 17%, respectively, of the total plant
21 cover recorded in all the plots. Other 20 families were recorded, the largest of which was
22 Asteraceae; each of these other families represented less than 10% of the total cover.

Using all the relevés, we calculated the cumulative cover of each species in the entire area. To perform the calculation, first we standardized the cover values of the plots by dividing the cover of each species in each plot by the total plant cover in that plot. Then, for each species, we calculated its total cover in the dataset, by summing its standardized cover values from all the plots. Finally, we rescaled the values of all species to a 1-100 scale to obtain the cumulative cover values. From the resulting list of species, we removed 208 species with cumulative cover values below 2%, considering them to be transient species (Mariotte 2014) that might have been recorded by chance and may not represent the core mesic meadows flora. We used the remaining 116 species as the core list of meadows species to retrieve seed data for this article. Most of these species were hemicryptophytes (78%), with some therophytes (16%) and a few chamaephytes and geophytes (3% of each). The family with more species in the list was Poaceae (22%), with another 17% belonging to Fabaceae, 15% to Asteraceae, 8% to Apiaceae, and the rest of the families representing less than 5% each.

Species ecological requirements

We also used the relevés as a basis to characterize the preferences of the selected species for three environmental factors (cold, summer drought and soil reaction) which have been found to be major ecological drivers of mesic meadow plant diversity (Rodríguez-Rojo *et al.* 2014). For cold and drought, we used the coordinates of the plots to retrieve from CHELSA (Karger *et al.* 2017) the bioclimatic variables bio06 (minimum temperature of the coldest month) and bio14 (precipitation of the driest month). For soil reaction (pH), we took from each plot five soil samples from between 0 and 20 cm depth with a Dutch auger

and combined them to make a bulk soil horizon, which we subsequently air-dried, crumbled, finely crushed and sieved with a 2 mm screen, to finally measure the pH in H₂O with a glass electrode in a suspension of soil:water (1:2.5). With each of these three environmental variables (bio06, bio14, pH) measured at the plot level we calculated the species niche centroids (SNCs). The SNC for any given species and variable is the mean of the environmental variable in all the plots where the species occurs, weighted by species cover in each plot (Zelený 2018). The list of core species, with their cumulative covers and SNCs, is available at GitHub (see Data Availability Statement).

Seed morphology and germination dataset

From the vegetation plots described above, we collected dispersal units (hereafter called seeds) during the dispersal seasons of 2016, 2017 and 2018. Seed collection followed the methodology of ENSCONET (2009). To describe seed morphology, we acquired images of 100-seed samples of each species using a flatbed scanner (Brother LC985) with a resolution of 200 dpi and a scanning area of 1024 x 1024 pixels (Bacchetta *et al.* 2008). We distributed the seeds on the transparent glass of the scanner, in a 10 x 10 grid. For each sample, and without moving the seeds, we repeated the scan with black and white backgrounds. In the case of the black background, we covered samples with a black box to avoid interference from environmental light. For the white background, we used the scanner cover. We digitized the obtained images and stored them in JPEG format (Joint Photographic Experts Group). We processed the scanned images using *ImageJ*, an open-source image processing program designed for scientific multidimensional images (Schneider *et al.* 2012). The program calculates several biometric parameters for each seed

on the sample, and among those we chose seed length and width. Additionally, we retrieved species values of seed mass from the Seed Information Database (Royal Botanic Gardens, Kew 2017) and of seed number at the individual/ramet level from the LEDA database (Kleyer *et al.* 2008). The dataset with the length and width measures is available at GitHub (see Data Availability Statement).

We also germinated the collected seeds using three germination treatments to determine the germination response to temperatures that are representative of the study area: 14/4 °C representing the capacity of freshly-dispersed seeds to germinate at cool temperatures of spring and autumn, 22/12 °C as summer temperature, and 30/20 °C as sun-heated soil, e.g. soil exposed to sun after hay cutting. Additionally, we compared, for each of these temperature regimes, the germination of fresh seeds versus seeds subjected to a dormancy-breaking treatment. In the case of Fabaceae and other families that might present physical dormancy (Baskin and Baskin 2014), the treatment consisted in scarification by chipping the seed coat with a scalpel. For the rest of the families, we used gibberellic acid GA₃ (0.0645 mM) in darkness during 24h, as a treatment to remove potential physiological seed dormancy (Blandino *et al.* 2019). For each species and treatment, we sowed four Petri dishes with 25 seeds each. The germination substrate was 1% distilled water - agar. We sealed dishes with Parafilm to prevent desiccation. Trials took place in a germination chamber (KBW 400, Binder GmbH, Tuttlingen, Germany) with a 12/12-hour photoperiod (the light period corresponding to the higher temperature). Experiments lasted for four weeks, with germination scoring once per week. The germination criterion was 2 mm radicle emergence. After four weeks, we cut the seeds that failed to germinate and

1 examined them under a magnifying glass. We classified them as normal when the embryo
2 was visible and firm, empty when they lacked an embryo, and contaminated when they
3 were mouldy. We only considered normal seeds when calculating germination proportions
4 and conducting subsequent analyses.

5 In addition to this experimental germination data, we retrieved seed germination records for
6 the list of study species from ENSCOBASE (Carta *et al.* 2021), the seed germination
7 database of the European Native Seed Conservation Network
8 (<http://enscobase.maich.gr/index.tml>); and the *SylvanSeeds* database of seed germination
9 records for the nemoral biome (Fernández-Pascual 2021). These new records included
10 additional records of species in our experimental dataset, plus records of new species absent
11 from our experimental dataset, and in all cases corresponded to seed lots originally
12 collected within Europe. The combined dataset, including our own experimental data and
13 the records from ENSCOBASE and *SylvanSeeds*, contained 104 species (i.e. 90% of the
14 core list of meadow species was covered) and 1,940 germination records (i.e. germination
15 proportions for a given seed lot of a species, recorded in a set of laboratory experimental
16 conditions) from 32 European countries. Overall, 131,747 seeds had been used in the
17 experiments. The range of experimental germination temperatures (weighted average of the
18 daily thermoperiod) which had been used in the experiments spanned from 2 to 31 °C, with
19 1,146 records of constant temperatures (i.e. experiments that used the same temperature
20 during all their duration) and 794 of alternating temperatures (i.e. experiments where
21 different temperatures were applied during the day and the night, in diurnal cycles). Seeds
22 had been exposed to light during some part of the diurnal cycle in 1,861 records or kept in

total darkness in 79 records. Experiments had been performed with unstratified seeds (i.e. not subjected to a previous dormancy-breaking incubation) in 1,765 records and with stratified seeds (i.e. subjected to previous incubation in dormancy-breaking conditions, including treatments of wet incubation under cold, warm and combinations of cold and warm conditions) in 175 records. Finally, there were 212 records where GA₃ had been applied, and 460 records where seeds had been scarified. The full germination dataset is available at GitHub (see Data Availability Statement).

Statistical analysis

We conducted all analyses in R (R Core Team 2020), and the code for analysis and creation of the figures and manuscript is available at GitHub (see Data Availability Statement).

To test the effect of germination treatments on seed germination proportions, we performed a meta-analysis (Pappalardo *et al.* 2020) of the germination dataset by fitting binomial generalized mixed models with Bayesian estimation (Markov Chain Monte Carlo generalized linear mixed models, MCMCglms) (Carter *et al.* 2021; Fernández-Pascual *et al.* 2021) using the R package *MCMCglmm* (Hadfield 2010). We fitted four models: (1) to the entire dataset; and separately for each of the three botanical groups of mesic meadows: (2) Poaceae, (3) Fabaceae, and (4) a third group including the rest of the families. To account for the effect of a shared phylogenetic history in species traits, models included as a random effect a reconstructed phylogenetic tree of the study species. We created the phylogeny using the R package *V.PhylMaker* (Jin and Qian 2019) which contains an updated mega-tree of the seed plants based on Smith & Brown (2018). We placed taxa absent from the mega-tree at the genus-level basal node. The phylogenetic tree is available

at GitHub (see Data Availability Statement). The response variable of the models was the proportion of germinated seeds. The fixed effects were the germination treatments (scarification, stratification, GA₃, temperature, alternating temperature and light). Random effects included the phylogenetic tree, species identity, seed lot and source of the data. In all models, variables were scaled so their contribution to the effect sizes could be compared. We used weakly informative priors in all models, with parameter-expanded priors for the random effects. Each model was run for 500,000 MCMC steps, with an initial burn-in phase of 50,000 and a thinning interval of 50 (De Villemereuil and Nakagawa 2014), resulting, on average, in 9,000 posterior distributions. From the resulting posterior distributions, we calculated mean parameter estimates and 95% Highest Posterior Density (HPD) and Credible Intervals (CI). We estimated the significance of model parameters by examining CIs, considering parameters with CIs overlapping with zero as non-significant. To estimate the phylogenetic signal in the models we used Pagels's lambda (λ) (Pagel 1999), estimated simultaneously with the models by calculating the mean of the posterior distribution and the 95% CI of λ as indicated by De Villemereuil et al. (2014). When $\lambda = 0$, related taxa are no more similar than expected by chance, while when $\lambda = 1$, the trait is evolving following a constant variance random walk or Brownian motion model; intermediate values of λ indicate a phylogenetic correlation in trait evolution that does not fully follow a Brownian motion model (Pagel 1999). The detailed output of the models is available at GitHub (see Data Availability Statement).

To check whether seed traits and plant ecological preferences were related, we did a Principal Component Analysis (PCA) of seed traits and species SNCs for cold, drought and

pH. We performed the PCA ordination at the species level, i.e. calculating a series of continuous seed traits for each species. We transformed the final germination proportions to create a continuous variable for the germination cues (i.e. stratification, scarification, temperature, alternating temperatures and light). To do so, for each cue and species, we calculated a weighted average of the cue levels (in the case of temperature, cue levels were the temperature treatments; for the other cues, the levels were 0 = absence and 1 = presence), weighting by the germination proportion at each level. This approach underrepresents the importance of the levels that had not been tested for a given species, but can serve as a proxy of the response to the germination cues when visualized across the whole dataset. We also included seed mass and seed number in the ordination (species average values). We left GA₃ out of the PCA because its ecological interpretation is subordinated to stratification (as both cues break physiological seed dormancy). We also left seed length and width out because these values were not available for enough species. We calculated the PCA with the package *FactoMineR* (Lê *et al.* 2008).

Finally, to compare the germination of mesic meadow species with that of other herbaceous communities, we retrieved data from previous works on the seed germination ecology of bogs and fens (Fernández-Pascual *et al.* 2013; Fernández-Pascual 2016), alpine and subalpine grasslands (Eduardo Fernández-Pascual and Jiménez-Alfaro *et al.* 2017), and coastal plant communities of rocky cliffs and sand dunes (Eduardo Fernández-Pascual and Pérez-Arcoiza *et al.* 2017). These additional germination records had been obtained using the same experimental methodology as the one employed for some of the germination experiments of this study: recently collected seeds, untreated for physiological dormancy

(but scarified in the cases of families known for having physical dormancy), had been subjected to three germination thermoperiods (14/4, 22/12, 30/20 °C). All seeds had been collected in the Cantabrian Mountains of Spain or the neighbouring coast. We combined these records with the records with matching experimental conditions from the meadows germination dataset, and performed a PCA of the resulting dataset. The germination records for the other plant communities are available at GitHub (see Data Availability Statement).

RESULTS

Seed morphology

Poaceae had lower values of seed mass and higher values of seed number, while Fabaceae had heavier but fewer seeds (**Fig. 2**). The other families covered the range of values showed by Poaceae and Fabaceae, but their median values were high for both traits: their median seed mass was close to that of the Fabaceae, while their median seed number was higher than that of the Poaceae (**Fig. 2**). Seed shape also showed a divergence between Poaceae and Fabaceae, with seeds being elongated in the former and round in the latter (**Fig. 2**). The other families covered the full range of variation, with both elongated and round seeds (**Fig. 2**).

Seed germination

When considering the full set of mesic meadow species, all six studied germination cues had a significant effect on final germination proportions (**Fig. 3**). The germination of meadow seeds was positively associated with scarification, stratification, GA₃, alternating

1 temperatures, and light. Average temperature had a negative effect, indicating a trend
2 towards higher germination at lower temperatures. Averaging the whole dataset, the highest
3 germination proportions were achieved at 20 °C. Between 0 °C and 20 °C, germination
4 proportions increased steadily with increasing temperatures. Above 20 °C, germination
5 declined more sharply, and meadow seeds rarely germinated at 30 °C.

6 Some differences became apparent when dividing the dataset in the three floristic groups
7 that compose mesic meadow vegetation. In Poaceae (**Fig. 3**), no effect was found for
8 scarification, GA₃ or temperature. Stratification had a negative effect on germination. The
9 major drivers of Poaceae germination appeared to be alternating temperatures and light,
10 with both having a positive effect. In Fabaceae (**Fig. 3**), the largest positive effect on
11 germination was produced by scarification, with no effect of stratification, GA₃ or light.
12 Temperature had a negative effect, and in fact Fabaceae species had higher germination
13 proportion at temperatures under 20 °C. Fabaceae seed germination also responded
14 negatively to alternating temperatures. Finally, in the remaining families (**Fig. 3**), the main
15 cues having a positive effect on germination were stratification, GA₃, alternating
16 temperatures and light. These species did not respond to scarification nor to average
17 temperature.

18 To describe the effects of the random factors, we will refer only to the full model that
19 included all species in the dataset. The strongest effect was that of the phylogeny (mean =
20 6.48, CI = 3.7 - 9.79), followed by the source of the data (mean = 2.9, CI = 1.21 - 4.98) and
21 the seed lot (mean = 1.28, CI = 0.94 - 1.61). The phylogenetic signal in the germination
22 responses was relatively high ($\lambda = 0.75$, CI = 0.65 - 0.84).

Seed traits and species ecological preferences

PCA indicated a clear separation between environmental preferences and seed traits, with each set of variables contributing to different axes (**Fig. 4**). The first PCA axis explained 29% of the variation and was related to environmental preferences. The variables with the largest contribution to this first axis were soil reaction (pH), winter cold (bio06) and summer rainfall (bio14). This horizontal axis separated (i, left) species with preferences for sites with warm winter temperatures from (ii, right) species with preferences for sites with high summer rainfall and less acidic soils. Axis 2 explained 17% of the variability, mostly related to seed traits. The main contributing variables were seed mass, seed number and the germination response to scarification. This axis separated (iii, bottom) Poaceae species that produce many seeds with a positive germination response to alternating temperatures from (iv, top) Fabaceae species that produce heavy seeds with a positive response to scarification.

Comparison with other habitats

Ordination resulted in a separation of mesic meadow species from plant species belonging to natural herbaceous communities of the same region (**Fig. 5**). Axis 1 explained 78% of the variance and was positively associated with high germination at all three temperature treatments. This horizontal axis separated (i, left) species with low germination across treatments from (ii, right) species with high germinability across treatments. The second axis explained 17% of the variation and separated (iii, bottom) species that responded more to the cool germination temperature from (iv, top) species that responded more to the warm germination treatment. Meadow species tended to be situated in the right sector of the

ordination, indicating high germinability; and at the center of the vertical axis, indicating a neutral response to temperature. On the other hand, species from the other communities were positioned at the left of the horizontal axis, indicating a lower germinability; and were more separated along the vertical axis, indicating a preference for either warmer (bogs and fens) or cooler (alpine grasslands, coastal communities) germination treatments.

DISCUSSION

Our meta-analysis of germination records showed that, overall, the seed ecology of mesic meadows is characterized by (1) high seed germinability when compared to other herbaceous plant communities; (2) low correspondence between seed trait variability and the natural environmental drivers of mesic meadow diversity; and (3) a deep phylogenetic separation between the two major families, Poaceae and Fabaceae.

We found that alternating temperatures and light are the two most influential factors triggering germination, especially in species from the family Poaceae and the minor families, with the exception of Fabaceae. Alternating temperatures and light are usually considered to be micro-environmental cues that indicate, at a fine scale, the existence of safe sites for regeneration (Jumpponen *et al.* 1999). The diurnal alternation of temperature decreases with burial depth in the soil, and the alternation is also reduced by vegetation cover (Thompson 1977; Van Assche and Vanlerberghe 1989; Saatkamp *et al.* 2011). Thus, a positive germination response to alternating temperatures could detect the depth of seed burial and prevent the germination of seeds that are buried too deep for seedlings to survive before emerging from the ground (Bond *et al.* 1999). Perhaps more importantly in mesic

1 meadows, alternating temperatures could also indicate that the vegetation cover has been
2 diminished by either mowing or grazing, and therefore mark an appropriate time for
3 seedling regeneration, when the competition by the established plants is lessened. Similar
4 functions can be ascribed to the germination response to light, as physiologically-active
5 light in significant amounts only travels the uppermost millimetres of soil (Tester and
6 Morris [1987](#)) and the quality of light will be affected by vegetation cover (Jankowska-
7 Blaszcuk and Daws [2007](#)). Therefore, the germination response to alternating
8 temperatures and light indicates conditions that are found immediately after mowing for
9 haymaking, a predictable perturbation that occurs approximately at the same time every
10 year, and which forces meadow plants to establish, grow and reproduce in the time frame
11 determined by this perturbation (Grime [2006](#); Klimešová *et al.* [2010](#)). Theoretically, these
12 germination traits would also promote the formation of a transient soil seed bank (Williams
13 [1983b](#); Venn and Morgan [2010](#)), but it is worth mentioning that Poaceae, which show the
14 germination response to alternating temperatures and light, also have the more elongated
15 seed shape, a trait that is thought to reduce the capacity of seeds to enter the soil seed bank
16 in temperate meadows (Thompson *et al.* [1993](#); Funes *et al.* [1999](#)).

17 The strong effect of scarification was specifically related to the hard-seeded Fabaceae. In
18 this family, physical dormancy as a result of an impermeable seed coat works as a
19 mechanism to detect seasonal cycles of temperature and humidity (Van Assche *et al.* [2003](#)).
20 It has been also proposed that physical dormancy can avoid seed predation, by preventing
21 the emission of olfactory cues that are elicited by seed imbibition and that can be perceived
22 by seed predators (Paulsen *et al.* [2013](#)). In the case of this study, Fabaceae seeds are

1 amongst the heaviest, and they clearly follow a separate regeneration strategy from that of
2 the other families. While other groups rely on alternating temperatures and light as gap-
3 detecting mechanisms, Fabaceae seeds do not respond to these factors, and they are
4 regulated instead by scarification, while they also respond to cooler temperatures than the
5 rest of the families. This suggests that Fabaceae seeds would tend to germinate when
6 temperatures are cooler, and thus before or after the summer haymaking season. Their
7 larger size, and the related larger reserves, could allow Fabaceae seedlings to emerge from
8 greater depths (Bond *et al.* 1999), e.g. when the meadow grass is still high, before mowing.
9 Furthermore, larger seed size also increases seed survival in cattle dung (Peco *et al.* 2006)
10 and thus improves the capacity of being dispersed by animal depositions (Traba *et al.*
11 2003).

12 The germination response to average temperatures and stratification is understood to detect
13 cues related to seasonal cycling and macroclimatic variation (Finch-Savage and Leubner-
14 Metzger 2006; Carta *et al.* 2021). Apart from the response to cool temperatures in
15 Fabaceae, the rest of the families did not show a response to average temperatures, further
16 highlighting their reliance on micro-habitat cueing. Moreover, the dominant family Poaceae
17 showed a negative response to cold stratification. The less frequent families, however,
18 showed a divergence from the Poaceae strategy in their positive response to stratification.
19 These non-dominant species appear to require a period of cold stratification, indicating that
20 overwintering has occurred, before they can germinate (Baskin and Baskin 2014). These
21 minor families are also the only group of species that showed a positive response to GA₃, a

1 phytohormone which can work as a substitute of cold stratification to overcome
2 physiological dormancy (Bewley *et al.* 2013).

3 The strong phylogenetic signal in germination responses highlights the phylogenetic
4 clustering of regeneration strategies that we have described so far: (1) the dominant family
5 Poaceae, showing a lack of response to seasonal cues (average temperature and cold
6 stratification), and relying instead on large amounts of propagules and on detecting micro-
7 niche cues (alternating temperatures, light) that can be associated to the yearly perturbation
8 of mowing, (2) the second most-dominant family, Fabaceae, which do not respond to the
9 micro-niche cues, and regulate their germination timing through scarification and cooler
10 germination temperatures, possibly because their larger size and reserves allow their seeds
11 to decouple their emergence timing from the mowing disturbance, and finally (3) the minor
12 families that respond to micro-cues in a similar way than Poaceae, but which are
13 differentiated from them by showing a positive response to cold stratification and G₃,
14 indicating that they rely on physiological seed dormancy to track seasonal cycles. This third
15 strategy would allow to fine-tune their germination to the micro-environmental conditions
16 plus the inter-annual climatic variation. Phylogenetic clustering is also apparent regarding
17 seed shape, mass and number, with Poaceae producing many elongated and light seeds, and
18 Fabaceae producing fewer, rounder and heavier seeds. The shape of Poaceae seeds could
19 make them particularly successful in attaching to hay and dispersing with it, and Poaceae
20 seeds tend to be overrepresented in seed mixes obtained via haymaking (Scotton *et al.*
21 2009; Haslgrübler *et al.* 2014). Asteraceae and Apiaceae, some of the most abundant
22 among the minor families, also tend to have shapes resembling those of Poaceae.

1 As we have seen, the regeneration strategy of high germinability appears to be general in
2 meadow species, unrelated to regional environmental gradients, and to be more responsive
3 to the predictable perturbation that is yearly mowing. In an abandoned meadow of the
4 Swiss Prealps, resuming mowing promoted germination and emergence, although natural
5 regeneration was limited due to a lack of seed dispersal to the site (Kupferschmid *et al.*
6 2000). Mowing also promoted autumn germination in a dry grassland of northern Germany
7 (Kahmen and Poschlod 2008). These results agree with the lack of dormancy and the high
8 germinability of meadows seeds that we have found, and which has also been highlighted
9 by previous studies. For example, when comparing populations of *Poa trivialis*, grassland
10 seeds were less dormant than seeds collected in an arable field (Froud-Williams *et al.*
11 1986). Ten Brink *et al.* (2013), when comparing congeneric herbaceous species from open
12 and forest habitats, found that species from open habitats needed less cold stratification.
13 Similarly, seeds collected from a hay meadow germinated better when left untreated, rather
14 than when being exposed to dormancy-breaking treatments (Haslgrübler *et al.* 2014). These
15 high-germinability strategy has the practical consequence of greatly limiting the long-term
16 seed bank persistence of mesic meadow species. Rather, the soil seed bank has been
17 reported as being transient in hay meadows and related grasslands (Milberg 1992;
18 McDonald 1993; Hutchings and Booth 1996). In Poland, the soil bank of a hay meadow
19 was dominated by arable and weedy forbs, with low representation of Poaceae and
20 Fabaceae (Janicka 2017).

21 Overall, our results suggest that mesic meadows are quite sensitive to land use change.
22 Their inability to form persistent seed banks leads to dispersal limitations to passive natural

regeneration (Kupferschmid *et al.* 2000), highlighting the importance of active actions of meadow restoration via seed supply. For centuries, human activities have shaped the regeneration of hay meadows, leading to a loss of seed dormancy and decoupling seeds from seasonal cycles, as has been found in many domesticated species (Dürr *et al.* 2015). The same anthropic processes that have shaped semi-natural mesic meadows have left them dependent on continued human intervention for their regeneration. Understanding the germination requirements of the different plant groups that coexist in mesic meadows can help to manage, conserve and restore their biological diversity.

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AUTHOR CONTRIBUTIONS

All authors contributed data. E.F.P. conceived the study and performed the analyses. E.F.P. wrote the manuscript with help from A.C. All authors revised the manuscript and approved the final version.

DATA AVAILABILITY

The original data, R code for the analysis and creation of the manuscript can be accessed at the GitHub repository <https://github.com/efernandezpascual/meadows>. Upon publication, a version of record of the repository will be deposited in Zenodo.

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FIGURES

Figure 1: Diversity of dispersal units in plant species from mesic meadows: (a) Floret of Cynosurus cristatus (Poaceae); (b) floret of Arrhenatherum elatius (Poaceae); (c) seed of Lathyrus pratensis (Fabaceae); (d) legume of Onobrychis viciifolia (Fabaceae); (e) loment fragment of Ornithopus perpusillus (Fabaceae); (f) achene with pappus of Centaurea scabiosa (Asteraceae); (g) achene of Ranunculus acris (Ranunculaceae); (h) nutlet of Prunella grandiflora (Lamiaceae); (i) achene of Knautia nevadensis (Dipsacaceae); (j) perigynium of Carex binervis (Cyperaceae); (k) receptacle of Sanguisorba minor (Rosaceae); (l) mericarp of Carum verticillatum (Apiaceae); (m) seed of Cerastium fontanum (Caryophyllaceae); (n) seed of Plantago lanceolata (Plantaginaceae); (o) seed of Rhinanthus angustifolius (Orobanchaceae).

Figure 2: Morphology of mesic meadow seeds. The two panels on the left show the probability densities of species values for seed mass and seed number, log-transformed for ease of visualization. The three horizontal lines within the probability densities represent the first quartile, the median and the third quartile of the values. The panel on the right shows values of seed length and width obtained by image analysis, with each point being a

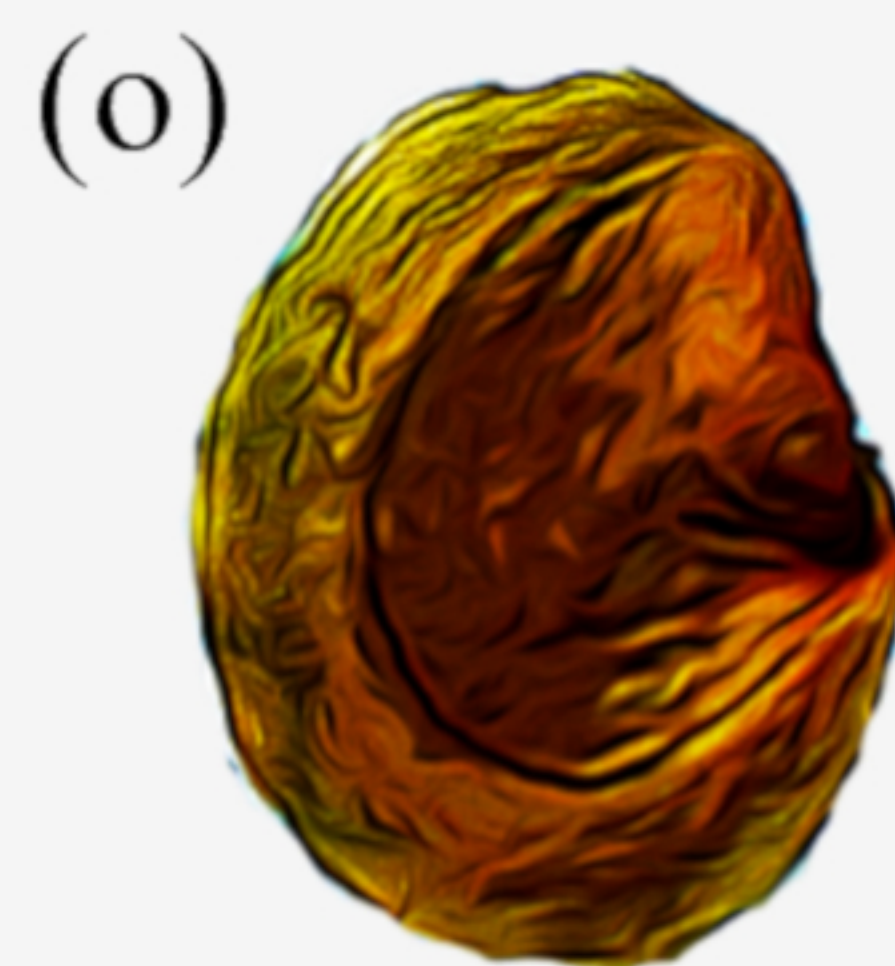
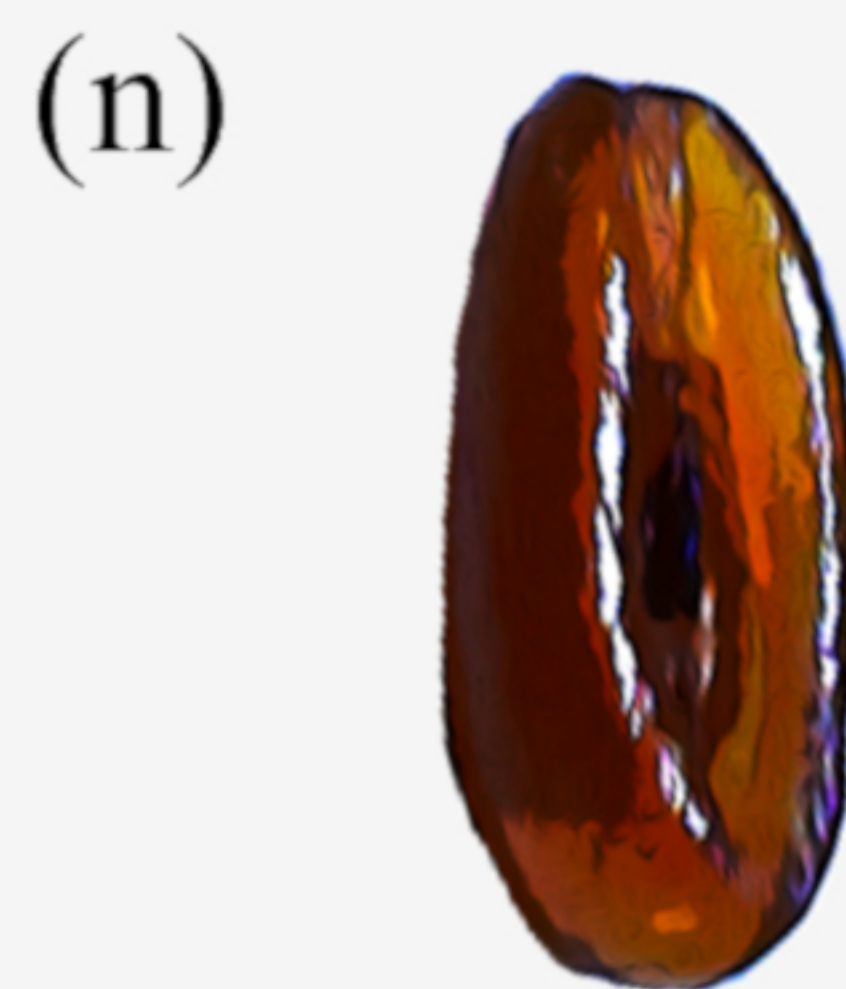
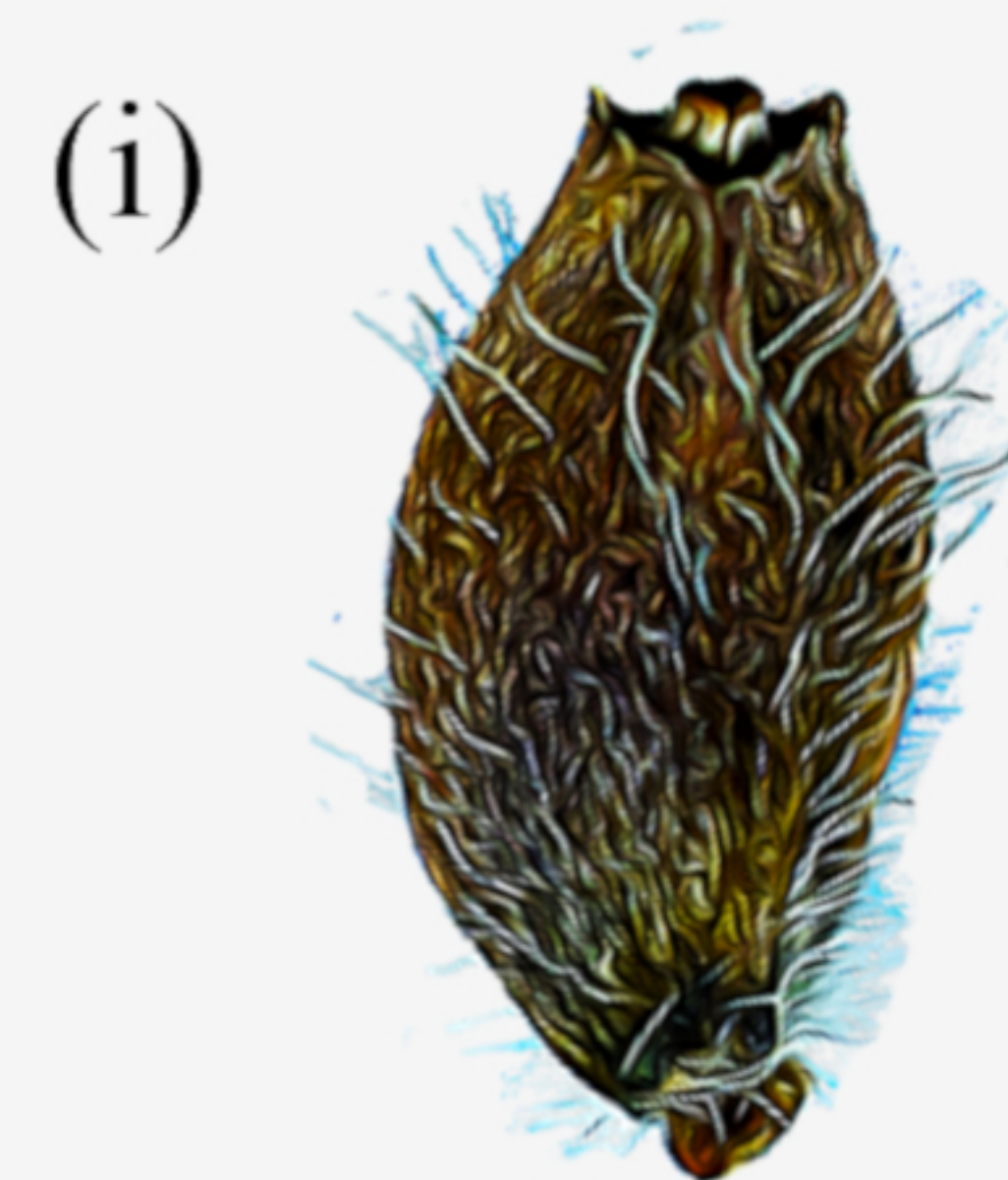
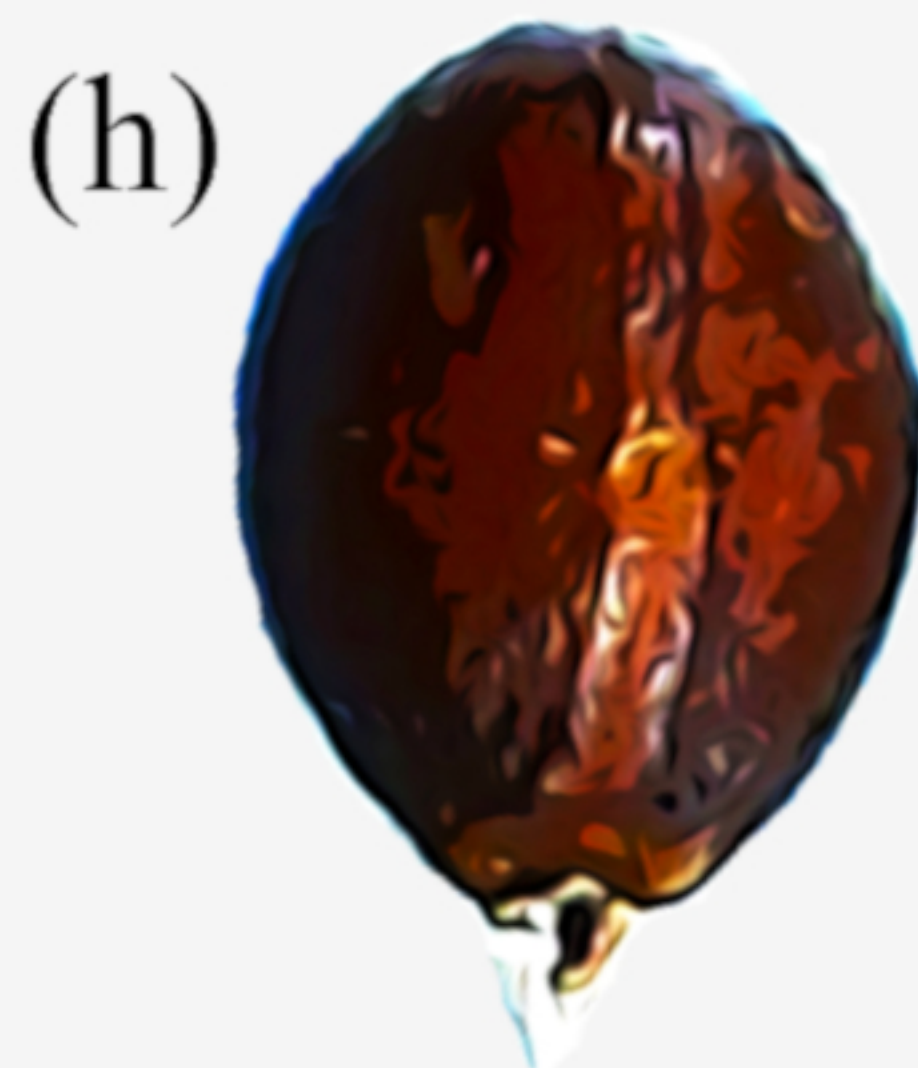
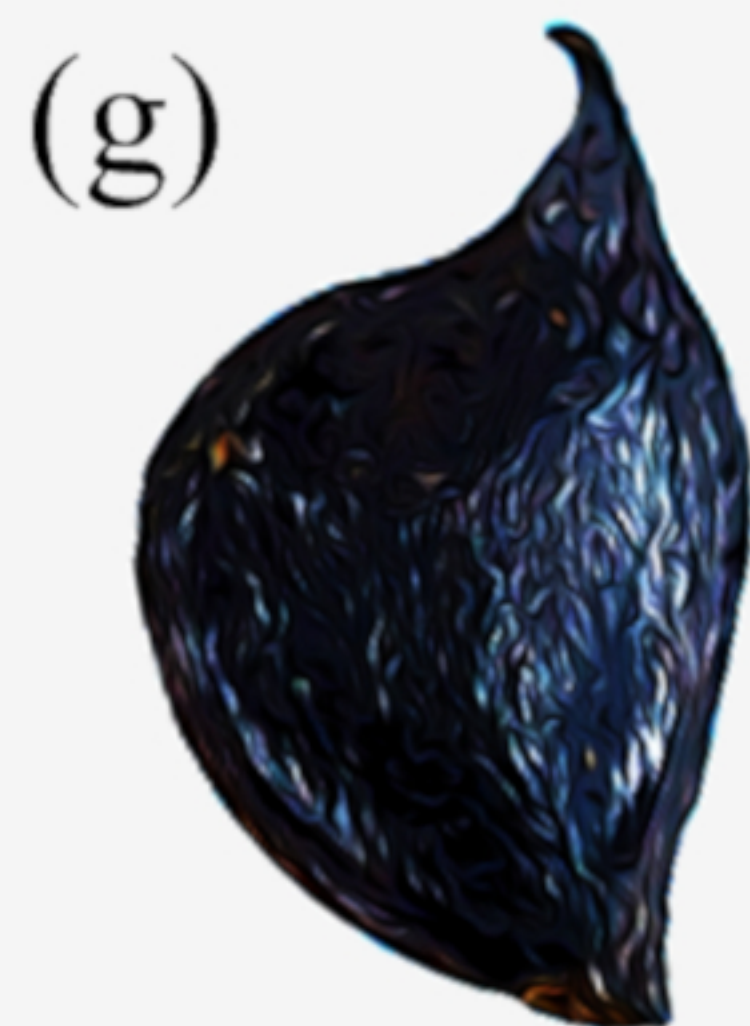
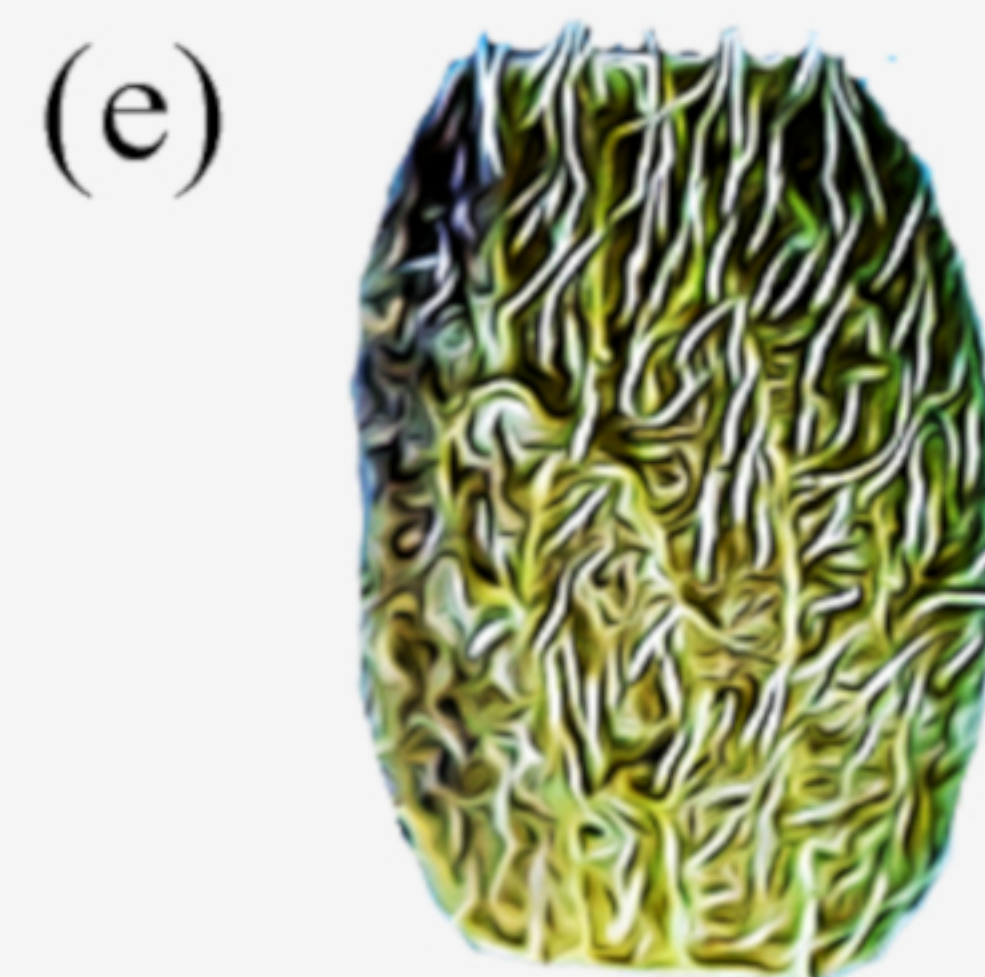
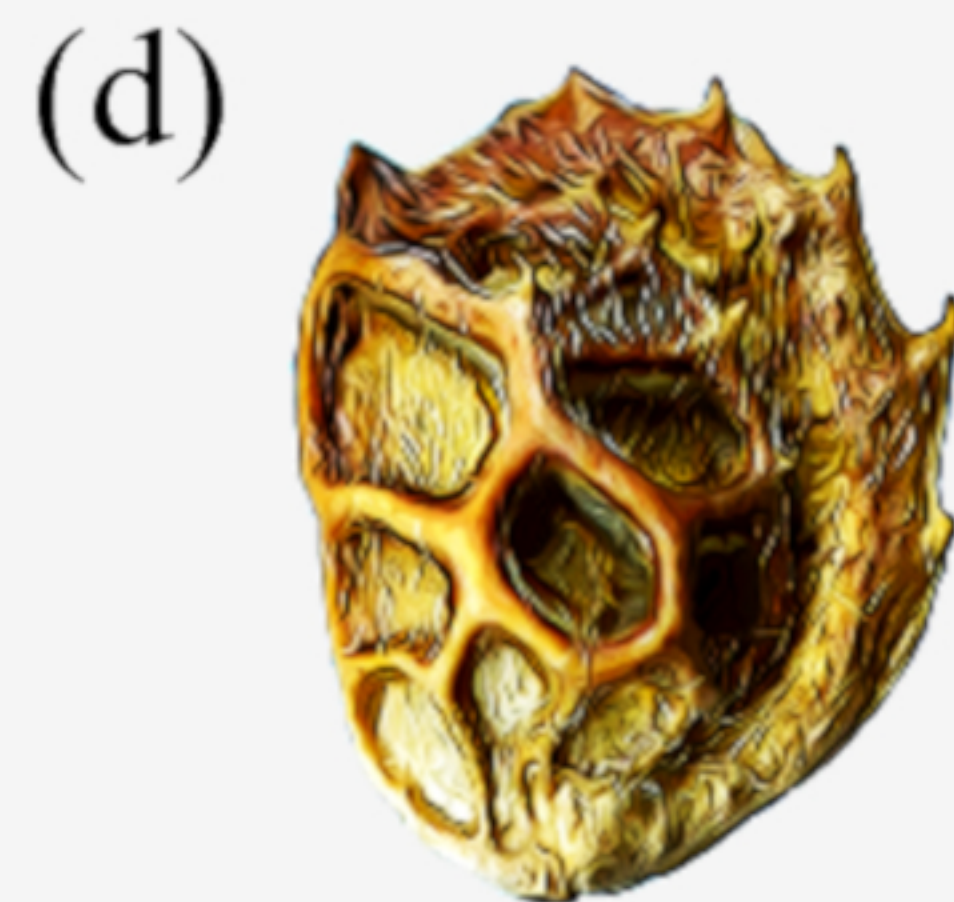
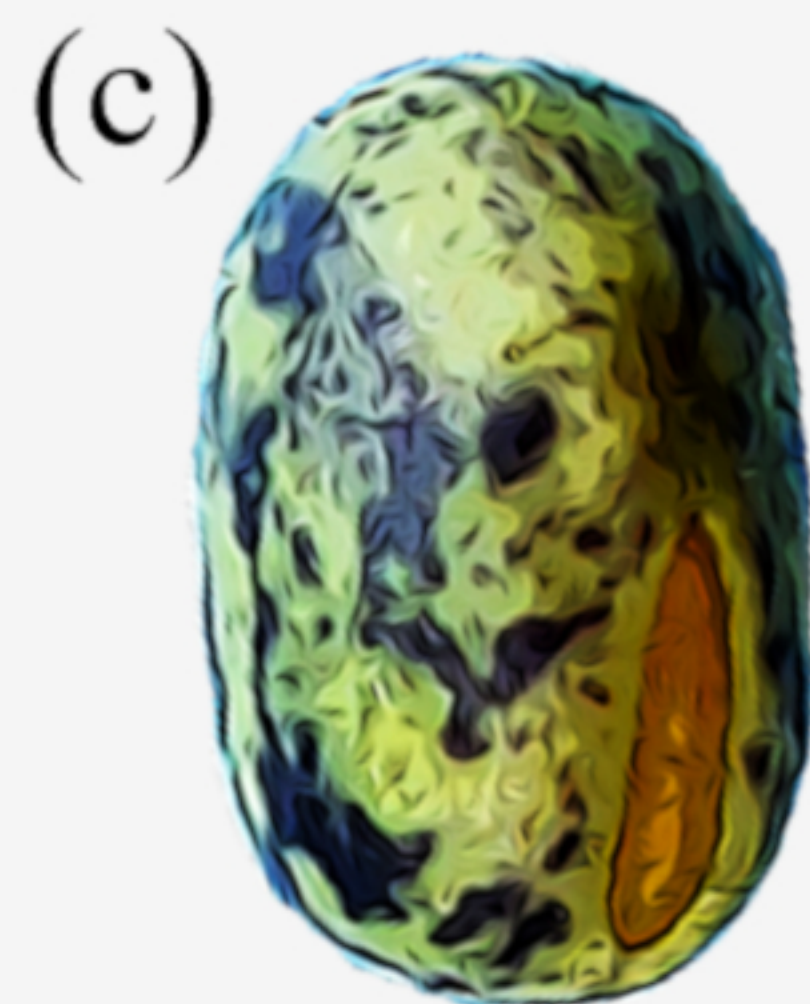
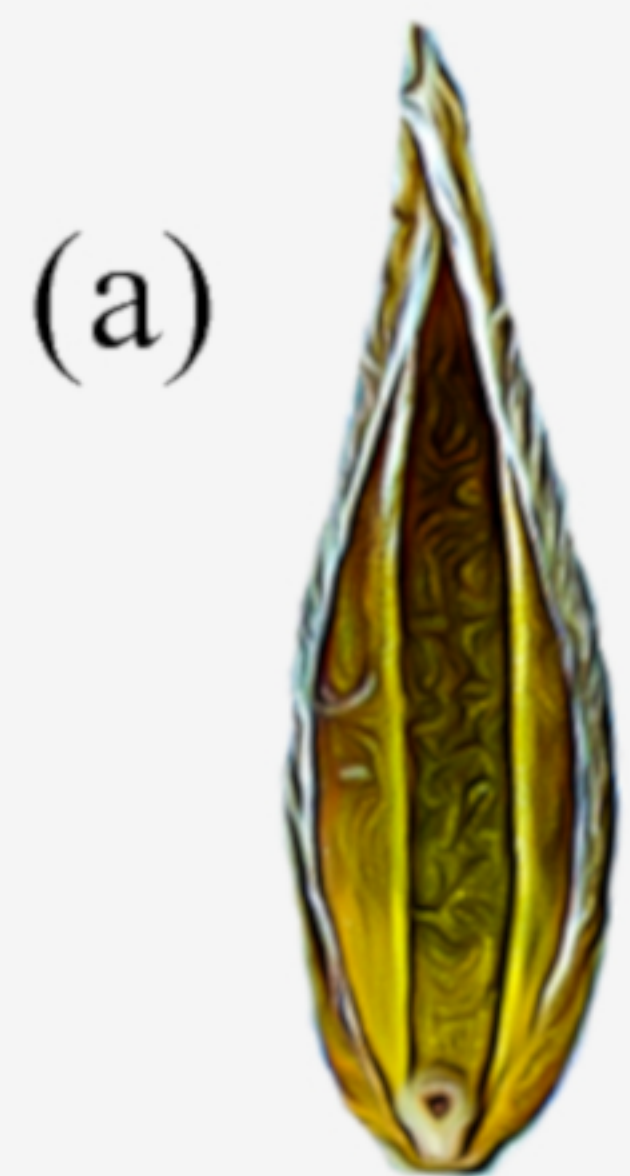
1 seed. In all cases, data is divided between the grasses (*Poaceae*), legumes (*Fabaceae*) and
2 the other plant families.

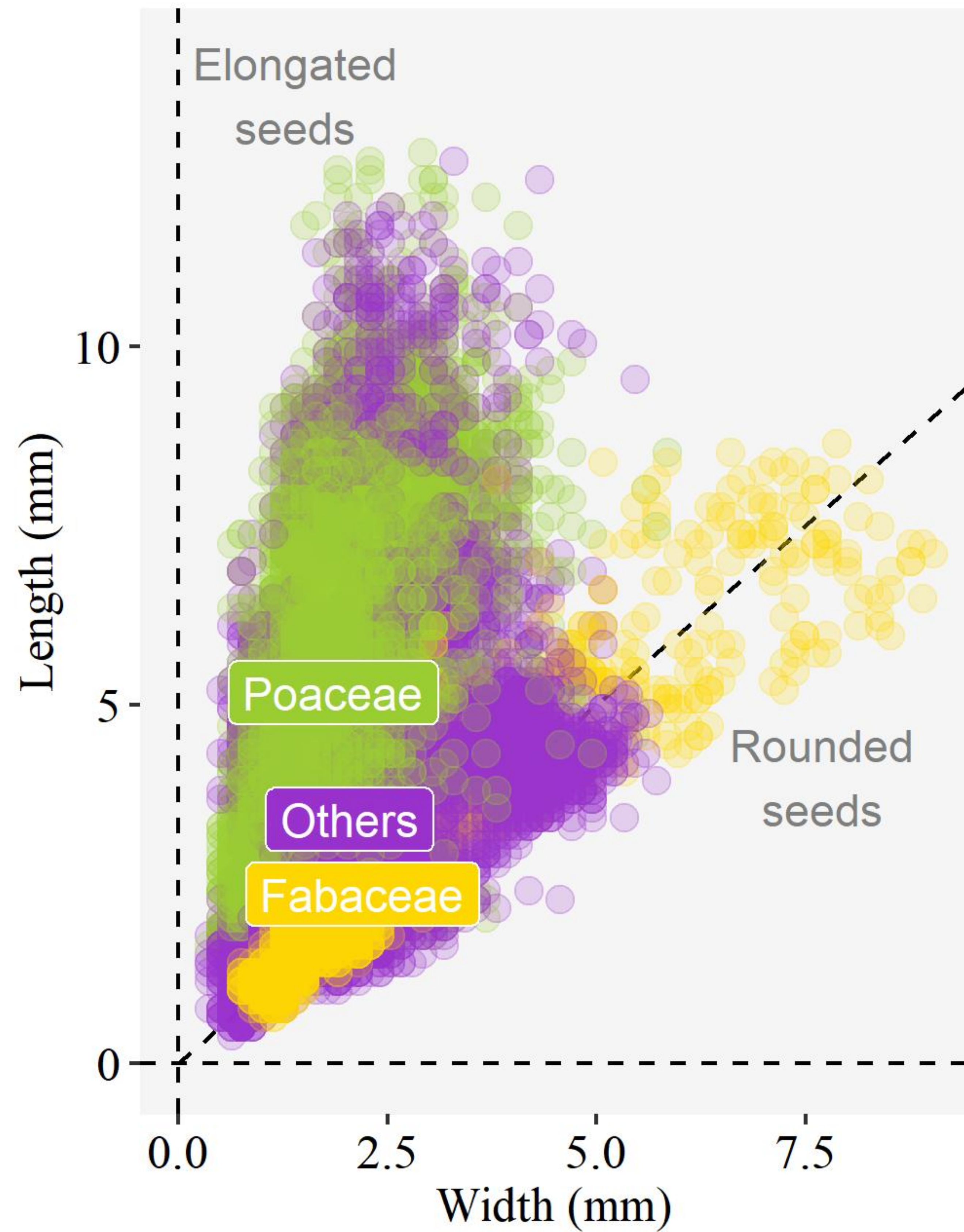
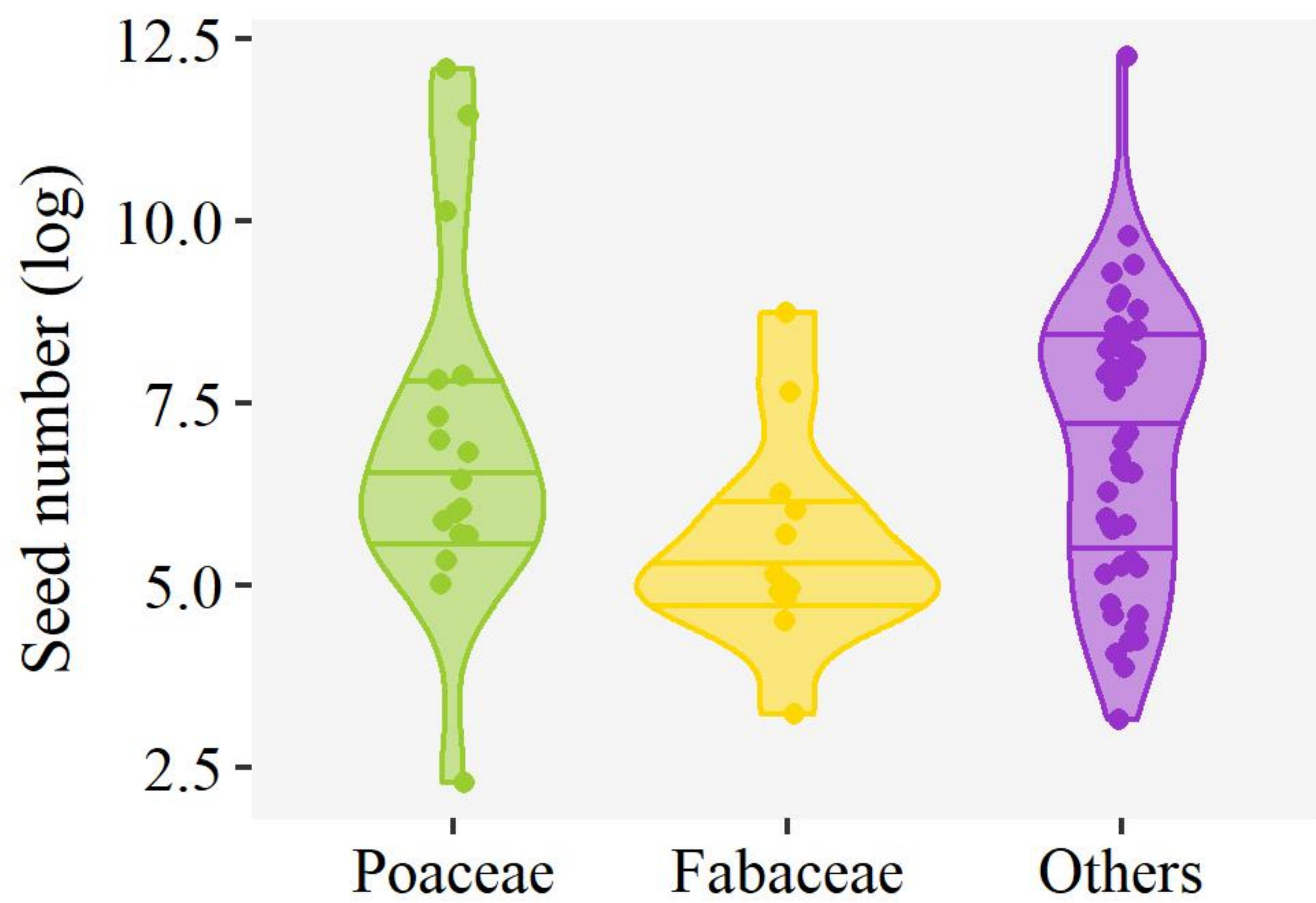
3
4 *Figure 3: Germination cues in mesic meadows. Effect of germination cues simulated in the*
5 *laboratory over the final germination proportions of mesic meadow seeds. Dots indicate*
6 *the posterior mean of the effect size for each cue, and whiskers the 95% credible interval of*
7 *the effect size. The line of zero-effect is shown: when a credible interval overlaps with the*
8 *zero-effect line, the effect can be regarded as non-significant. In separate panels, the figure*
9 *shows the results of a general model including data for all species, plus specific models for*
10 *the three main botanical groups of mesic meadows: grasses (*Poaceae*), legumes*
11 *(*Fabaceae*) and the other families.*

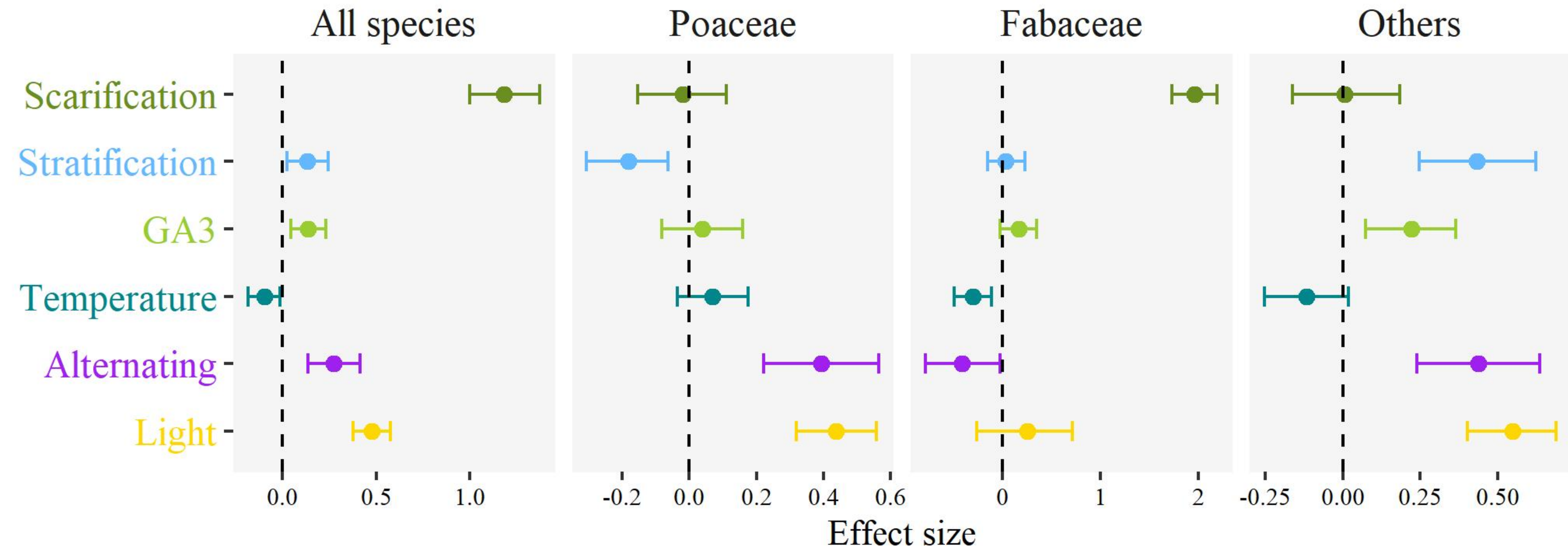
12
13 *Figure 4: Environment and seed traits are separate axis of variation in mesic meadows.*
14 *Principal Component Analysis ordination of mesic meadow species considering their*
15 *environmental preferences and their seed traits. Each point is a species, coloured by the*
16 *three main botanical groups of mesic meadows: grasses (*Poaceae*), legumes (*Fabaceae*)*
17 *and the other families. Labels indicate the contribution of the variables to the axes: grey-*
18 *background labels for environmental preferences, and white-background labels for seed*
19 *traits. Environmental preferences were calculated as species niche centroids (SNCs) for the*
20 *minimal temperature of the coldest month (bio06), precipitation of the driest month*
21 *(bio14), and soil pH. To calculate the SNCs, a vegetation dataset of mesic meadows of the*

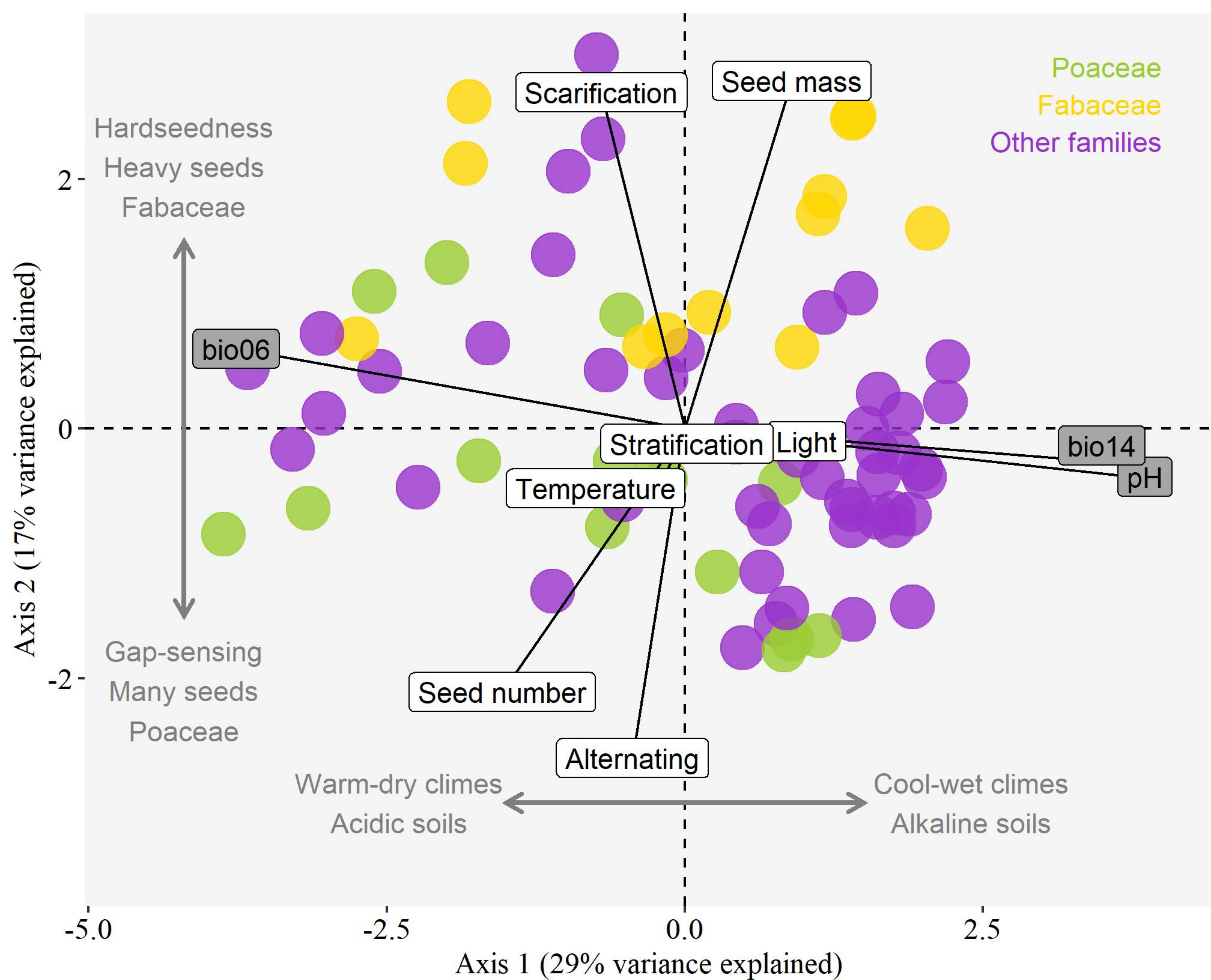
1 *Iberian Peninsula was used. The seed traits are seed mass and the germination relative*
2 *indices for the response to scarification, stratification, average germination temperature*
3 *(temperature), alternating temperature (alternating) and light. All environmental*
4 *preferences aligned to the first axis, while seed traits aligned to the second axis, showing*
5 *that environment and seed traits are separate axis of variation in mesic meadows.*
6 *Stratification and average germination temperature showed very low variation, in*
7 *accordance with their small effect on Figure 2.*

8
9 *Figure 5: Higher germinability in mesic meadows compared to other herbaceous plant*
10 *communities. Principal Component Analysis ordination of species responses to three seed*
11 *germination treatments. Species are grouped by their habitat, with points showing the*
12 *centroid for each habitat group, and rays linking the centroid with the position of each*
13 *species in the group. Labels indicate the contribution of the germination treatments to the*
14 *axes. All species had been collected in herbaceous vegetation types of the Cantabrian*
15 *Mountains of Spain and the neighbouring coast. All seeds were untreated, except for*
16 *scarification, which was applied routinely to all botanical families presenting physical*
17 *dormancy. Germination treatments consisted in 12/12 h periods with a warmer phase in*
18 *light and a cooler phase in darkness. The position of mesic meadows compared to the other*
19 *habitats indicates that their species tended to have higher germinability even if untreated*
20 *(i.e. less seed dormancy) and were more neutral in their thermal requirements.*









● Alpine grasslands ● Bogs and fens ● Coastal cliffs and dunes ● Mesic meadows

