BB512/BB612 - Week VI

```
suppressPackageStartupMessages(library(Biobase))
suppressPackageStartupMessages(library(bladderbatch))
```

Data

We are going to use data from the paper Evaluating gene expression in C57BL/6J and DBA/2J mouse striatum using RNA-Seq and microarrays. that is a comparative RNA-seq analysis of different mouse strains.

```
con <- url("http://bowtie-bio.sourceforge.net/recount/ExpressionSets/bottomly_eset.RData")
load(file = con)
close(con)

pdata <- pData(bottomly.eset)
edata <- as.matrix(exprs(bottomly.eset))
fdata <- fData(bottomly.eset)</pre>
```

Transform the expression data and remove lowly expressed genes:

```
edata <- log2(as.matrix(edata) + 1)
edata <- edata[rowMeans(edata) > 10, ]
head(pdata)
```

```
sample.id num.tech.reps strain experiment.number lane.number
##
## SRX033480 SRX033480 1 C57BL/6J
                                                           6
                                                                      1
## SRX033488 SRX033488
                                1 C57BL/6J
## SRX033481 SRX033481
                                1 C57BL/6J
                                                           6
                                                                      2
                                                           7
## SRX033489 SRX033489
                                 1 C57BL/6J
                                                                       2
                                                           6
## SRX033482 SRX033482
                                 1 C57BL/6J
                                                                       3
## SRX033490 SRX033490
                                 1 C57BL/6J
                                                                       3
```

```
table(pdata$strain)
```

```
## ## C57BL/6J DBA/2J
## 10 11
```

Calculate p-values parametrically

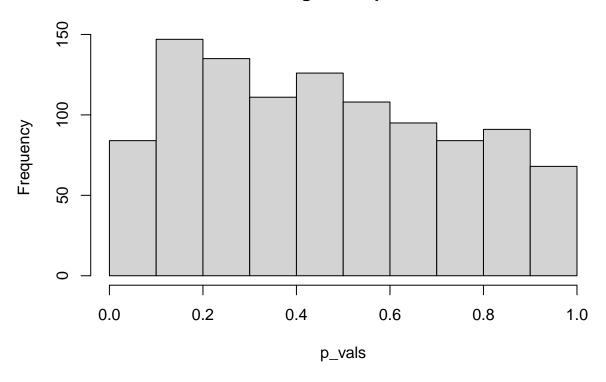
t-test

```
idxA <- which(colnames(edata) %in% pdata$sample.id[pdata$strain == "C57BL/6J"])
idxB <- which(colnames(edata) %in% pdata$sample.id[pdata$strain == "DBA/2J"])
raw_p_vals <- apply(edata, 1, function(x) t.test(x[idxA], x[idxB])$p.value)
p_vals <- sort(raw_p_vals)
head(p_vals, 10)</pre>
```

ENSMUSG00000033585 ENSMUSG00000070003 ENSMUSG000000022212 ENSMUSG00000044708

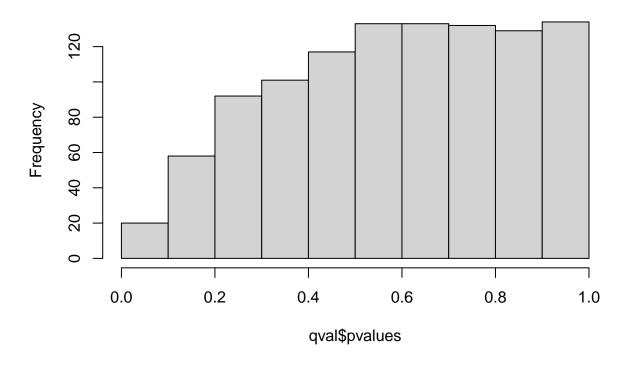
```
0.00019311
                               0.00348277
                                                   0.00573420
                                                                      0.00738418
##
## ENSMUSG00000024883 ENSMUSG00000036473 ENSMUSG00000038020 ENSMUSG00000074045
##
           0.00927920
                               0.01023011
                                                  0.01119852
                                                                      0.01435736
## ENSMUSG00000032340 ENSMUSG00000041297
           0.01560463
                               0.01826414
p_adj_vals <- p.adjust(p_vals, method = "fdr")</pre>
head(p_adj_vals, 10)
## ENSMUSG00000033585 ENSMUSG00000070003 ENSMUSG000000022212 ENSMUSG00000044708
              0.20257
                                  0.84267
                                                      0.84267
                                                                         0.84267
## ENSMUSG00000024883 ENSMUSG00000036473 ENSMUSG000000038020 ENSMUSG00000074045
##
              0.84267
                                  0.84267
                                                      0.84267
                                                                         0.84267
## ENSMUSG00000032340 ENSMUSG00000041297
##
              0.84267
                                  0.84267
hist(p_vals)
```

Histogram of p_vals



Adjusting for variables with edge

Histogram of qval\$pvalues



Using moderated statistics with limma

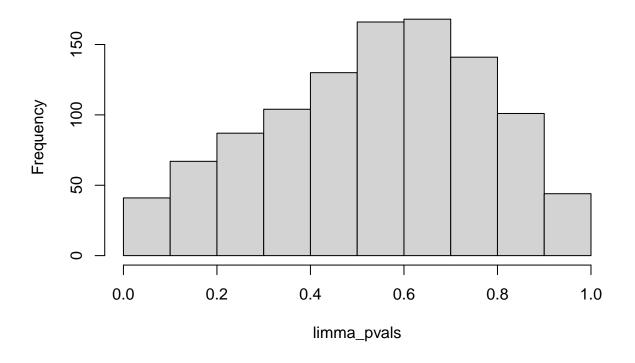
```
library(limma)

##
## Attaching package: 'limma'
## The following object is masked from 'package:BiocGenerics':
##
## plotMA

mod <- model.matrix(~ pdata$strain + pdata$lane.number)
fit_limma <- lmFit(edata, mod)
ebayes_limma <- eBayes(fit_limma)
limma_pvals <- topTable(ebayes_limma, number = Inf)$P.Value

## Removing intercept from test coefficients
hist(limma_pvals)</pre>
```

Histogram of limma_pvals



Calculating empirical permutation p-values

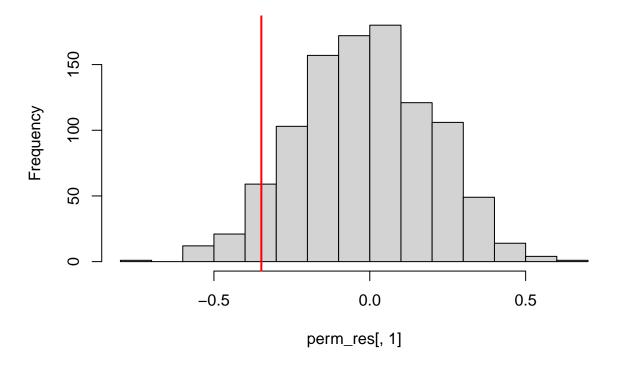
```
idxA <- which(colnames(edata) %in% pdata$sample.id[pdata$strain == "C57BL/6J"])
idxB <- which(colnames(edata) %in% pdata$sample.id[pdata$strain == "DBA/2J"])

actual_diffs <- apply(edata, 1, function(x) mean(x[idxA]) - mean(x[idxB]))

B <- 1000
perm_res <- c()
set.seed(123)
for (i in seq_len(B)) {
    cur_idxA <- sample(seq_len(ncol(edata)), 10)
    cur_idxB <- setdiff(seq_len(ncol(edata)), cur_idxA)
    cur_diffs <- apply(edata, 1, function(x) mean(x[cur_idxA]) - mean(x[cur_idxB]))
    perm_res <- rbind(perm_res, cur_diffs)
}

hist(perm_res[, 1])
abline(v = actual_diffs[1], col = "red", lwd = 2)</pre>
```

Histogram of perm_res[, 1]

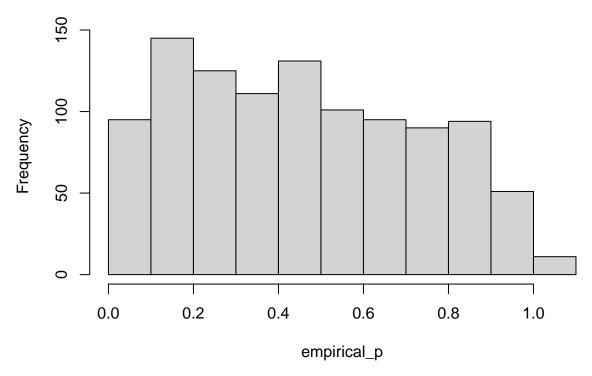


```
# empirical p value
2 * sum(perm_res[, 1] <= actual_diffs[1]) / B

## [1] 0.114

empirical_p <- c()
for (j in 1:ncol(perm_res)) {
    if (actual_diffs[j] < 0) {
        empirical_p <- c(empirical_p, 2 * sum(perm_res[, j] <= actual_diffs[j]) / B)
    } else {
        empirical_p <- c(empirical_p, 2 * sum(perm_res[, j] >= actual_diffs[j]) / B)
    }
}
hist(empirical_p)
```

Histogram of empirical_p



```
plot(raw_p_vals, empirical_p)
abline(a = 0, b = 1, col = "red", lwd = 3)
```

