Package 'MapRtools'

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Title Tools for genetic mapping

Version 0.28

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Description Tools for genetic mapping

Depends R (>= 3.5.0)

License GPL-3

LazyData true

RoxygenNote 7.2.3

Encoding UTF-8

Imports ggplot2, scam, seriation, CVXR, Matrix, HMM, optimx

Suggests knitr, rmarkdown

VignetteBuilder knitr

R topics documented:

`EGQ` *Expected Genotype Quality*

Description

Expected Genotype Quality for Binomial Model

Usage

```
EGQ(depth, error, ploidy, prior)
```

Arguments

`depth`	read count
`error`	allelic error
`ploidy`	ploidy
`prior`	numeric vector of length ploidy + 1

Details

As defined in Matias et al. (2019), EGQ is the PHRED-scaled expected error of the genotype call, conditional on the true genotype. This function returns EGQ for the genotype most frequently miscalled, which is the balanced heterozygote (i.e., ploidy/2).

Value

numeric scalar

References

Matias et al. (2019) Plant Genome 12:190002. https://doi.org/10.3835/plantgenome2019.01.0002

`genetic_map` *Multi-point estimation of a genetic map*

Description

Multi-point estimation of a genetic map

Usage

```
genetic_map(x, LOD, n.point = 5)
```

Arguments

`x`	matrix of pairwise map distances (cM) between the marker-bins for one chromosome
`LOD`	matrix of LOD scores between marker-bins
`n.point`	Number of points used for estimation

Details

Uses LOD-score weighted least-squares regression method described by Stam (1993). Markers must be binned (e.g., using LDbin) for this function to work properly. Argument `n.point` controls how many pairwise distances are used in the linear regression. `n.point=2` means only adjacent bins; `n.point=3` means adjacent bins and bins with one intervening marker, etc. Marker names taken from the rownames attribute of `x`.

Value

data frame with columns marker,position (in cM)

`inverse_map_fn` *Inverse map function*

Description

Computes recombination frequency from map distance

Usage

```
inverse_map_fn(x, model)
```

Arguments

`x`	map distance (cM)
`model`	Either "Haldane" or "Kosambi"

Value

recombination frequency

`LDbin` *Create marker bins based on LD*

Description

Create marker bins based on LD

Usage

```
LDbin(geno, r2.thresh = 0.99)
```

Arguments

`geno`	matrix of haplotype dosages (markers x indiv)
`r2.thresh`	threshold for binning

Details

Bins are created based on hierarchical clustering with `hclust` and `method='single'`, using $1 - r^2$ as the dissimilarity metric. The argument `r2.thresh` controls the height for cutting the dendrogram to create the bins. The marker with the least missing data for each bin is chosen to represent it.

Value

List containing

bins data frame with two columns: marker,bin

geno genotype matrix for the bins

r2 r2 matrix for the bins

`LG` *Make linkage groups based on clustering*

Description

Make linkage groups based on clustering

Usage

```
LG(LODmat, thresh = seq(2, 20, by = 2))
```

Arguments

`LODmat`	matrix of LOD scores for the marker bins
`thresh`	numeric vector of thresholds for clusterings

Details

If `thresh` is a numeric vector with multiple LOD thresholds, the function returns a plot showing the number of markers per LG. If `thresh` is a single value, the function returns a data frame with the LG assignment for each marker. LGs are numbered from the largest to smallest group.

Value

Either a ggplot2 object or data frame of linkage groups (see Details)

LGtrim *Trim a linkage group based on genotype frequencies*

Description

Trim a linkage group based on genotype frequencies

Usage

```
LGtrim(geno, LODmat, thresh)
```

Arguments

geno	matrix of haplotype dosages (markers x samples)
LODmat	matrix of LOD scores for the markers
thresh	numeric vector of thresholds for clusterings

Details

This function should only be run on a single linkage group (to form the linkage groups, use LG. If thresh is a numeric vector with multiple LOD thresholds, the function returns a plot showing the impact of the threshold on genotype frequencies. If thresh is a single value, the function returns a vector of the marker names that are retained. The rownames of geno and LODmat must match.

Value

Either a ggplot2 object or a vector of marker names (see Details)

LL *Log-likelihood for mapping populations*

Description

Log-likelihood for mapping populations

Usage

```
LL(r, counts, pop.type)
```

Arguments

r	recombination frequency
counts	3x3 contingency table for haplotype dosages 0,1,2
pop.type	One of the following: "DH","BC","F2","S1r","RIL.self","RIL.sib"

Details

The argument counts can be constructed using the table function for two markers. Genotype coding must represent dosage of a founder haplotype. For BC populations, possible allele dosages are 0,1. For DH and RIL pops, it is 0,2. For F2 and S1 pops, it is 0,1,2. S1r is an S1 population with the 1 alleles in repulsion phase.

Value

log-likelihood

`map_fn` *Map functions*

Description

Computes cM map distance from recombination frequency

Usage

```
map_fn(r, model)
```

Arguments

`r`	recombination frequency
`model`	Either "Haldane" or "Kosambi"

Value

Map distance in cM

`MLEL` *Max Likelihood Estimation of Linkage*

Description

Max Likelihood Estimation of Linkage

Usage

```
MLEL(geno, pop.type, LOD, n.core = 1, adjacent = FALSE)
```

Arguments

`geno`	Matrix of haplotype dosages (markers x indiv)
`pop.type`	One of the following: "DH","BC","F2","S1","RIL.self","RIL.sib"
`LOD`	Logical, whether to return LOD (TRUE) or recomb freq (FALSE)
`n.core`	For parallel execution on multiple cores
`adjacent`	Logical, should calculation be done for all pairs (FALSE) or adjacent (TRUE) markers

Details

Can be used to estimate either the LOD score or recombination frequency, depending on the value of `LOD`. Genotype coding must represent dosage of a founder haplotype. For BC populations, possible allele dosages are 0,1. For DH and RIL pops, it is 0,2. For F2 and S1 pops, it is 0,1,2.

Value

If adjacent is FALSE, a matrix of recombination frequencies or LOD scores; otherwise, a three-column data frame with marker, the LOD or r value, and the phase ("c","r") with the previous marker

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order_markers *Order markers by solving the TSP*

Description

Order markers by solving the TSP

Usage

```
order_markers(x)
```

Arguments

x	distance matrix

Details

Uses R package seriation to minimize the distance between adjacent markers. For example, x could be a matrix of recombination frequencies or monotone decreasing transformation of LOD scores.

Value

a list containing

path optimized order as a vector of integers

distance sum of adjacent distances

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plot_coverage *Plot marker coverage of the genome*

Description

Plot marker coverage of the genome

Usage

```
plot_coverage(map, limits = NULL)
```

Arguments

map	data frame with columns chrom & position
limits	optional data frame with columns chrom & position, with the maximum length for each chromosome

Details

If `limits` not provided, then the maximum values in `map` are used.

Value

ggplot2 variable

`plot_genofreq` *Plot and filter markers based on genotype frequency vs position*

Description

Plot and filter markers based on genotype frequency vs position

Usage

```
plot_genofreq(geno, thresh = 0.1, span = 0.3)
```

Arguments

`geno`	haplotype dosage matrix (markers x indiv)
`thresh`	threshold for removing markers (see Details)
`span`	parameter to control degree of smoothing for spline (higher = less smooth)

Details

Genotypes should be coded 0,1,2. Markers are removed if their residual to the fitted spline exceeds `thresh`. Markers are assumed to be ordered. Function designed to be used for one chromosome.

Value

List containing

outliers character vector of marker names

plot ggplot2 variable

`plot_genoprob` *Plot genotype probabilities for one chromosome*

Description

Plot genotype probabilities for one chromosome

Usage

```
plot_genoprob(genoprob, map)
```

Arguments

genoprob	matrix (markers x genotypes) of probabilities for one individual
map	map data frame (markers,chrom,position)

Details

Names for the genotypes are taken from the colnames of genoprob.

Value

ggplot object

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plot_haplo *Graphical genotyping*

Description

Graphical genotyping

Usage

```
plot_haplo(geno, map)
```

Arguments

geno	genotype matrix (markers x indiv)
map	data frame with 3 columns (marker, chrom, position)

Details

Input matrix geno should have rownames attribute that matches marker names in the first column of map.

Value

ggplot object

plot_LD *Plot LD vs distance*

Description

Plot LD vs distance

Usage

```
plot_LD(r2, map, max.pair = 10000, dof = 8)
```

Arguments

r2	squared correlation matrix
map	data frame with 3 columns (marker, chrom, position)
max.pair	maximum number of r2 pairs for the spline
dof	degrees of freedom for the spline

Details

A monotone decreasing, convex spline is fit using R package scam. The input matrix r2 should have rownames attribute that matches marker names in the first column of map.

Value

List containing

plot ggplot object

spline data frame with fitted values for the spline

plot_map *Plots data against map*

Description

Plots data against map

Usage

```
plot_map(data)
```

Arguments

data	data frame with 3 columns: chrom, position, y (the plotting variable)

Value

ggplot

plot_square *Plot square (dis)similarity matrix*

Description

Plot square (dis)similarity matrix

Usage

```
plot_square(data, lims = NULL)
```

Arguments

data	square matrix
lims	numeric 3-vector with the low,mid,high points for the colors

Details

Can be used to plot squared correlation, recomb frequency, LOD and more. By default, lims equals (0,median,max)

Value

ggplot2 variable

rabbit_diallel *Make RABBIT input files for diploid diallel population*

Description

Make RABBIT input files for diploid diallel population

Usage

```
rabbit_diallel(ped, geno, geno.founder, map, outstem)
```

Arguments

ped	data frame with pedigree (pop,parent1,parent2)
geno	list of genotype matrices (markers x indiv), one for each population in ped
geno.founder	matrix of genotype data for the founders (markers x indiv)
map	genetic map (marker,chromosome,position)
outstem	name for output files

Details

Populations must be numbered in ped corresponding to their position in geno. Founders are not included in ped. All genotype matrices must have identical markers. Genetic map position should be in cM. Genotypes need to be coded according to RABBIT format.

rabbit_read *Parse output from RABBIT MagicReconstruct*

Description

Parse output from RABBIT MagicReconstruct

Usage

```
rabbit_read(rabbit.file, ML.file = NULL, diaQTL.file = NULL)
```

Arguments

rabbit.file	name of RABBIT output file
ML.file	name of most likely genotype file to create
diaQTL.file	name of diaQTL genotype file to create

Details

Two different file formats can be created. The `ML.file` contains the most likely (i.e., posterior maximum) genotype for each individual at each marker. The `diaQTL.file` contains the full distribution of genotype probabilities in the format required by the diaQTL R package (`diaQTL.file`). The default value for each filename is NULL, which generates no file.

Value

data frame defining the genotypes

S1_haplo *Phase S1 parent and reconstruct progeny in terms of parental haplotypes*

Description

Phase S1 parent and reconstruct progeny in terms of parental haplotypes

Usage

```
S1_haplo(geno, r, error)
```

Arguments

geno	ordered genotype matrix (markers x indiv) for one chromosome
r	average recombination frequency to use for the HMM
error	average genotype error to use for the HMM

Details

It is assumed that only segregating markers are present. Progeny reconstruction occurs using an HMM with a uniform transition probability matrix, based on an average recombination frequency `r`, and a uniform model for the genotype `error`.

Value

List containing

parent two column matrix (rows = markers) with the haplotypes for the parent

progeny matrix with progeny reconstructed based on dosage of the second parental haplotype

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`S1_selection` *Signatures of selection in S1 populations*

Description

Signatures of selection in S1 populations

Usage

```
S1_selection(data, alpha = 0.05)
```

Arguments

`data`	data frame with columns: marker, chrom, position, AA, AB, BB. Columns 4-6 have count data.
`alpha`	significance level

Details

Genotypes must be coded based on the S1 parental haplotypes, not markers.

The null hypothesis is no selection, in which case the expected frequency of genotypes is (AA = 1/4, AB = 1/2, BB = 1/4). Two alternate hypotheses are tested for gametic selection: 1.selection in one sex, 2.selection in both sexes. Two models of zygotic selection are also tested: 1.selection against one homozygote, 2.selection against both homozygotes. The selection coefficient equals the sum of the absolute differences between the observed and expected frequencies. Positive values correspond to selection against A or AA, negative values for selection against B or BB. For zygotic2, positive (negative) values represent selection against (for) homozygotes.

P-values are computed based on the likelihood ratio test; in other words, the change in deviance is assumed to be chi-squared distributed under the null hypothesis.

Value

list with "plot" and "table" of results:

model name of best model

s selection coefficient

score -log10(p) value

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