

# Package ‘sveval’

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**Title** SV evaluation

**Version** 1.2.1

**Description** Evaluate SV in a call set against a truth set using overlap-based approaches and sequence comparison for insertions.

**Depends** R (>= 3.4.4)

**License** MIT + file LICENSE

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GenomicRanges,  
IRanges,  
magrittr,  
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DelayedArray,  
Biostrings,  
parallel,  
testthat,  
ggplot2,  
shiny,  
DiagrammeR,  
DT

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sveval-package	<i>SV evaluation</i>
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**Description**

Evaluate SV in a call set against a truth set using overlap-based approaches and sequence comparison for insertions.

**Details**

Package:	sveval
Type:	Package
Version:	1.2.1
Date:	2019-02-28
License:	MIT

**Author(s)**

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**See Also**

<http://www.github.com/jmonlong/sveval>

**Examples**

```
## Not run:
eval = sveval01('calls.vcf', 'truth.vcf')
plot_prcurve(eval$curve)

# Comparing multiple methods
eval.1 = sveval01('calls1.vcf', 'truth.vcf')
eval.2 = sveval01('calls2.vcf', 'truth.vcf')
plot_prcurve(list(eval.1$curve, eval.2$curve), labels=c('method1', 'method2'))

## End(Not run)
```

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`filterSVs`*Filter SVs for size and regions of interest*

---

**Description**

Filter SVs for size and regions of interest

**Usage**

```
filterSVs(sv.gr, regions.gr = NULL, ol.prop = 0.5, min.size = 0,
          max.size = Inf)
```

**Arguments**

<code>sv.gr</code>	the input SVs (e.g. read from readSVvcf)
<code>regions.gr</code>	the regions of interest. Ignored if NULL (default).
<code>ol.prop</code>	minimum proportion of sv.gr that must overlap regions.gr. Default is 0.5
<code>min.size</code>	the minimum SV size to be considered. Default 0.
<code>max.size</code>	the maximum SV size to be considered. Default is Inf.

**Value**

a subset of sv.gr that overlaps regions.gr or in the specified size range.

**Author(s)**

Jean Monlong

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`findNocalls`*Find no-calls variants*

---

**Description**

Compare calls with a truth set and identifies which variants from the truth set specifically not called (genotype ./.).

**Usage**

```
findNocalls(calls.gr, truth.gr, max.ins.dist = 20, min.cov = 0.5,
            min.del.rol = 0.1, ins.seq.comp = FALSE, nb.cores = 1,
            sample.name = NULL, check.inv = FALSE)
```

**Arguments**

calls.gr	call set. A GRanges or the path to a VCF file.
truth.gr	truth set. A GRanges or the path to a VCF file.
max.ins.dist	maximum distance for insertions to be clustered. Default is 20.
min.cov	the minimum coverage to be considered a match. Default is 0.5
min.del.rol	minimum reciprocal overlap for deletions. Default is 0.1
ins.seq.comp	compare sequence instead of insertion sizes. Default is FALSE.
nb.cores	number of processors to use. Default is 1.
sample.name	the name of the sample to use if VCF files given as input. If NULL (default), use first sample.
check.inv	should the sequence of MNV be compared to identify inversions.

**Details**

Same overlapping strategy as in sveval01 although here no-calls are kept and there is no splitting by genotype.

**Value**

a data.frame with coordinates and variant ids from the truth set corresponding to no-calls.

**Author(s)**

Jean Monlong

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ivg\_sv

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*Interactive exploration of SVs in a variation graph*


---

**Description**

Opens a Shiny app with a dynamic table that contains input SVs. Clicking on a SV (row in the table) generates a simplified representation of the variation graph around this SV. The number of flanking nodes (context) can be increased if necessary, e.g. for large insertions. vg needs to be installed (<https://github.com/vgteam/vg>).

**Usage**

```
ivg_sv(svs, xg, ucsc.genome = "hg38")
```

**Arguments**

svs	either a GRanges with SVs (e.g. from readSVvcf) or the path to a VCF file.
xg	the path to the xg object of the variation graph.
ucsc.genome	the genome version for the UCSC Genome Browser automated link.

**Value**

Starts a Shiny app in a web browser.

**Author(s)**

Jean Monlong

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plot_prcurve	<i>Create precision-recall graphs</i>
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**Description**

Create a precision/recall curve using metrics computed by the sveval01 function. The sveval01 function returns a list containing a "curve" data.frame with the evaluation metrics for different quality thresholds.

**Usage**

```
plot_prcurve(eval, labels = NULL)
```

**Arguments**

eval	a data.frame, a list of data.frames, or a vector with one or several paths to files with "curve" information.
labels	the labels to use for each input (when multiple inputs are used). Ignored is NULL (default).

**Details**

If the input is a data.frame (or list of data.frames) it should be the "curve" element of the list returned by the sveval01 function. If the input is a character (or a vector of characters), they are considered to be file names and the data will be read from these files.

If multiple inputs are given, either using a list of data.frames or a vectors with several filenames, one curve per input will be created. This is to be used to quickly compare several methods. The "labels" parameters can be used to specify a label for each input to use for the graphs.

**Value**

list of ggplot graph objects

**Author(s)**

Jean Monlong

## Examples

```
## Not run:
eval = sveval01('calls.vcf', 'truth.vcf')
plot_prcurve(eval$curve)

# Comparing multiple methods
eval.1 = sveval01('calls1.vcf', 'truth.vcf')
eval.2 = sveval01('calls2.vcf', 'truth.vcf')
plot_prcurve(list(eval.1$curve, eval.2$curve), labels=c('method1', 'method2'))

# Or if the results were previously written in files
plot_prcurve(c('methods1-prcurve.tsv', 'methods2-prcurve.tsv'), labels=c('method1', 'method2'))

## End(Not run)
```

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readSVvcf

*Read SVs from a VCF file*


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## Description

Read a VCF file that contains SVs and create a GRanges with relevant information, e.g. SV size or genotype quality.

## Usage

```
readSVvcf(vcf.file, keep.ins.seq = FALSE, sample.name = NULL,
  qual.field = c("GQ", "QUAL"), check.inv = FALSE, keep.ids = FALSE,
  nocalls = FALSE, right.trim = TRUE)
```

## Arguments

vcf.file	the path to the VCF file
keep.ins.seq	should it keep the inserted sequence? Default is FALSE.
sample.name	the name of the sample to use. If NULL (default), use first sample.
qual.field	fields to use as quality. Will be tried in order.
check.inv	should the sequence of MNV be compared to identify inversions.
keep.ids	keep variant ids? Default is FALSE.
nocalls	if TRUE returns no-calls only (genotype ./). Default FALSE.
right.trim	if TRUE (default) the REF/ALT sequences are right-trimmed after splitting up multi-ALT variants.

## Details

By default, the quality information is taken from the QUAL field. If all values are NA or 0, the function will try other fields as specified in the "qual.field" vector. Fields can be from the INFO or FORMAT fields.

**Value**

a GRanges object with relevant information.

**Author(s)**

Jean Monlong

**Examples**

```
## Not run:
calls.gr = readSVvcf('calls.vcf')

## End(Not run)
```

---

svevalOl

*SV evaluation based on overlap and variant size*


---

**Description**

SV evaluation based on overlap and variant size

**Usage**

```
svevalOl(calls.gr, truth.gr, max.ins.dist = 20, min.cov = 0.5,
  min.del.rol = 0.1, ins.seq.comp = FALSE, nb.cores = 1,
  min.size = 50, max.size = Inf, bed.regions = NULL,
  bed.regions.ol = 0.5, qual.field = c("QUAL", "GQ"),
  sample.name = NULL, outfile = NULL, out.bed.prefix = NULL,
  qual.ths = c(0, 2, 3, 4, 5, 7, 10, 12, 14, 21, 27, 35, 45, 50, 60, 75,
  90, 99, 110, 133, 167, 180, 250, 350, 450, 550, 650),
  qual.quantiles = seq(0, 1, 0.1), check.inv = FALSE,
  geno.eval = FALSE, stitch.hets = FALSE, stitch.dist = 20,
  merge.hets = FALSE, merge.rol = 0.8)
```

**Arguments**

<code>calls.gr</code>	call set. A GRanges or the path to a VCF file.
<code>truth.gr</code>	truth set. A GRanges or the path to a VCF file.
<code>max.ins.dist</code>	maximum distance for insertions to be clustered. Default is 20.
<code>min.cov</code>	the minimum coverage to be considered a match. Default is 0.5
<code>min.del.rol</code>	minimum reciprocal overlap for deletions. Default is 0.1
<code>ins.seq.comp</code>	compare sequence instead of insertion sizes. Default is FALSE.
<code>nb.cores</code>	number of processors to use. Default is 1.
<code>min.size</code>	the minimum SV size to be considered. Default 50.
<code>max.size</code>	the maximum SV size to be considered. Default is Inf.

<code>bed.regions</code>	If non-NULL, a GRanges object or path to a BED file (no headers) with regions of interest.
<code>bed.regions.ol</code>	minimum proportion of <code>sv.gr</code> that must overlap <code>regions.gr</code> . Default is 0.5
<code>qual.field</code>	fields to use as quality. Will be tried in order.
<code>sample.name</code>	the name of the sample to use if VCF files given as input. If NULL (default), use first sample.
<code>outfile</code>	the TSV file to output the results. If NULL (default), returns a data.frame.
<code>out.bed.prefix</code>	prefix for the output BED files. If NULL (default), no BED output.
<code>qual.ths</code>	the QUAL thresholds for the PR curve. If NULL, will use quantiles. see <code>qual.quantiles</code> .
<code>qual.quantiles</code>	the QUAL quantiles for the PR curve, if <code>qual.ths</code> is NULL. Default is (0, .1, ..., .9, 1).
<code>check.inv</code>	should the sequence of MNV be compared to identify inversions.
<code>geno.eval</code>	should het/hom be evaluated separately (genotype evaluation). Default FALSE.
<code>stitch.hets</code>	should clustered hets be stitched together before genotype evaluation. Default is FALSE.
<code>stitch.dist</code>	the maximum distance to stitch hets during genotype evaluation.
<code>merge.hets</code>	should similar hets be merged into hets before genotype evaluation. Default is FALSE.
<code>merge.rol</code>	the minimum reciprocal overlap to merge hets before genotype evaluation.

**Value**

a list with

<code>eval</code>	a data.frame with TP, FP and FN for each SV type when including all variants
<code>curve</code>	a data.frame with TP, FP and FN for each SV type when using different quality thresholds
<code>svs</code>	a list of GRanges object with FP, TP and FN for each SV type (when using <code>QUAL&gt;=0</code> threshold).

**Author(s)**

Jean Monlong

**Examples**

```
## Not run:
## From VCF files
eval = sveval01('calls.vcf', 'truth.vcf')

## From GRanges
calls.gr = readSVvcf('calls.vcf')
truth.gr = readSVvcf('truth.vcf')
eval = sveval01(calls.gr, truth.gr)
```



```
## Genotype evaluation
eval = sveval01(calls.gr, truth.gr, geno.eval=TRUE, merge.hets=TRUE, stitch.hets=TRUE)

## End(Not run)
```

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**svOverlap***Overlap and annotate SV sets with coverage metrics*

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**Description**

Overlap and annotate SV sets with coverage metrics

**Usage**

```
svOverlap(query, subject, max.ins.dist = 20, min.cov = 0.5,
  min.del.rol = 0.1, ins.seq.comp = FALSE, nb.cores = 1)
```

**Arguments**

query	a GRanges object with SVs
subject	another GRanges object with SVs
max.ins.dist	maximum distance for insertions to be clustered. Default is 20.
min.cov	the minimum coverage to be considered a match. Default is 0.5
min.del.rol	minimum reciprocal overlap for deletions. Default is 0.1
ins.seq.comp	compare sequence instead of insertion sizes. Default is FALSE.
nb.cores	number of processors to use. Default is 1.

**Value**

a list with:

query	the query GRanges object annotated
subject	the subject GRanges object annotated

**Author(s)**

Jean Monlong

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