Differences in Plasma Metabolites Related to Alzheimer's Disease, APOE- ϵ 4 status and Ethnicity

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Conflicts of interest: none

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Abstract

INTRODUCTION: We investigated metabolites in plasma to capture systemic

biochemical changes associated with Alzheimer's disease (AD).

METHODS: Metabolites in plasma were measured in 59 AD cases and 60 healthy

participants of African American (AA), Caribbean Hispanic (CH) and non-Hispanic white

(NHW) ancestry using untargeted liquid-chromatography based ultra-high resolution

mass spectrometry. Metabolite differences between AD and healthy, ethnic groups and

APOEε4 status were analyzed. Untargeted network analysis identified pathways

enriched in AD-associated metabolites.

RESULTS: 5,929 annotated metabolites were measured. PLS-DA analysis inferred that

AD clustered separately from healthy controls (AUC=0.9816); discriminating pathways

included glycerophospholipid, sphingolipid and non-essential amino acid (alanine,

aspartate, glutamate) metabolism. Metabolic features in AA clustered differently from

CH and NHW (AUC=0.9275), and differed between APOE:4-carriers and non-carriers

(AUC=0.9972).

DISCUSSION. Metabolites, specifically lipids, were associated with AD, APOE_E4 and

ethnic group. Metabolite profiling could identify perturbed AD pathways, but genetic and

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ancestral background need to be considered.

Introduction

The understanding of the pathogenesis of Alzheimer's disease (AD) is incomplete.

Genetic and molecular mechanisms have been proposed, but no single gene variant or

molecular mechanism can fully explain the complexity of this disorder. Thus, it is likely

that genetic variants affect downstream metabolic pathways. Finding systemic

between disease and healthy states could identify biological molecular changes

mechanisms, potentially leading to early diagnosis and therapeutic development and

possibly successful interventions.

Metabolomics is a method of deep molecular phenotyping that represents the

underlying biochemical and physiological layer of the genome, transcriptome and

proteome. Metabolomics provides a precise and comprehensive analysis of phenotypic

abnormalities in which the individual components are physiologically described. This

snap shot of the metabolic state of individuals is possible due to the advances in high-

resolution mass spectrometric platforms with high sensitivity and specificity¹ that can

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strengthen the study at the human population level. For this study, we conducted

untargeted metabolomics profile analyses in plasma to identify endogenous and

exogenous metabolites associated with clinical AD, and to identify differences by

racial/ethnic group and APOE genotype.

Methods

Participants. The Washington Heights, Inwood Columbia Aging Project (WHICAP) has

recruited a representative community based group of individuals aged 65 years and

older through a collaborative effort with the Centers for Medicare and Medicaid and through the use of marketing rolls. The composition of the cohort reflected the community surrounding Columbia University Medical Center and comprises of non-Hispanic whites (24%), African Americans (28%), Caribbean Hispanics (48%) and 67% of the cohort are women. After obtaining informed consent, participants are interviewed in either English or Spanish. During each assessment, participants received a standardized neuropsychological test battery and medical interview. Blood was drawn, bar-coded, and brought to the laboratory within two hours of collection for DNA extraction and storage of plasma and serum. All of the medical, neurological, psychiatric, and neuropsychological data collected were reviewed at a consensus conference by clinicians who are experienced in the diagnosis of AD and related dementias. Diagnosis of clinical AD was based on accepted criteria².

For this investigation, fresh plasma was obtained, processed and frozen within two hours from individuals who had been evaluated at least twice to ensure stability of the clinical diagnosis AD and the lack of dementia in healthy controls. These individuals were equally divided between the three ethnic groups: African American, Caribbean Hispanic and non-Hispanic white and between cases and controls (Table 1).

Metabolomics acquisition and analyses. We acquired metabolites by conducting untargeted metabolomics from the plasma in 119 cases and controls from the WHICAP cohort using previously standardized method³. Untargeted LC-based ultra-high resolution mass spectrometry (LC-UHRMS) allowed deep phenotyping of the human

metabolome, providing measures of metabolites in most Kyoto Encyclopedia of Genes

and Genomes

(KEGG) human metabolic pathways⁴. After chromatographic separation using methods

previously described⁵, each sample was injected on each column (HILIC with positive

electrospray ionization (ESI) and C18 with negative ESI) three times, to obtain three

technical replicates per column. Data were processed through a computational pipeline

that leverages open source feature detection and peak alignment software, apLCMS⁶

and xMSanalyzer⁷, to create a feature table with the mass-to-charge (m/z) ratio,

retention time, and the abundance of each ion in each sample 6-8. The feature table was

used for statistical analyses followed by untargeted pathway analysis using the software

Mummichog⁸. The LC-UHRMS platform detects over 1,500 chemical signals that arise

from core nutrient metabolism, lipids, the microbiome, diet-derived chemicals,

pharmaceuticals and environmental contaminants, as well as over 100,000 untargeted

features⁹. For simplicity, we refer to the mass spectral features as metabolites. Results

described as annotations refer to level 4 and 5 confidence of feature identification by

criteria of Schymanski et al¹⁰. Results described as identifications refer to level 1

confidence (accurate mass, retention time and MS/MS fragmentation matching

authentic standards).

Metabolomics data analysis. Raw metabolite intensities were corrected for batch

effects using ComBat¹¹. Metabolites missing in over 30% of the samples were excluded

from further analysis. Metabolite values missing or of low quality were imputed to half

the value of the lowest level of detection for each metabolite¹². Intensities were quantile normalized, log-transformed and auto-scaled to normalize distributions¹³.

Principal component analyses (PCA) of normalized values were used to estimate stratification by ethnic/racial group, disease status and APOE ϵ 4. Partial Least Squares Discriminant analysis (PLS-DA) implemented in the mixOmics¹⁴ package was used to conduct supervised dimension reduction analysis to obtain a linear combination of the features that correlated with variability by ethnic/racial group, disease status and APOE ϵ 4 in the metabolomics data.

We tested association of individual metabolites with AD adjusting for age, sex, APOE- $\epsilon 4$ and ethnic/racial group. In addition, we tested the association of these metabolites with AD independently in each ethnic/racial group (defined by PCAs and ancestry analysis of GWAS data) to identify both ancestry specific and population-based metabolomics signatures in AD. Then, we tested association of $APOE \ \epsilon 4$ with metabolite intensities adjusting for age and sex. For association testing of independent metabolites, we declared nominal significance at P<0.05. Nominally significant metabolites were subsequently tested for pathway enrichment. Pathways enriched for metabolites associated with traits of interest were detected using module-enrichment analysis implemented in mummichog⁸. Statistical significance of enriched pathways were determined using a permutation test adjusting for multiple testing^{15,16}.

The project was reviewed and approved on 11/19/2019 by the Columbia University

Medical Center, Institutional Review Board (IRB-AAA09804).

Results

Untargeted metabolomics profiles were completed in 119 individuals that included 40

African Americans, 40 Caribbean Hispanics and 39 non-Hispanic whites from the

WHICAP cohort. Using the HILIC column with positive electrospray ionization (ESI), we

identified over 9,700 features, of which 5,929 were putatively annotated using

xMSannotator. The features did not show batch related variation and a total of 6375

features were present in at least 70% of the samples studied, of which 1,704

metabolites were annotated with xMSannotator confidence level 2 or 3. Using the C18

column with negative ESI, we measured over 6,700 features, of which 3759 were

present in at least 70% of the samples.

Metabolomic profiles of AD. We compared metabolites between AD and controls

adjusted for age, sex and ethnic/racial group and found several metabolic features

nominally associated with AD (uncorrected p<0.05) measured on both columns (figure

1A, Table S2) as well as unique metabolic profiles (Table 2).

While none of individual metabolites reached experiment-wide significance after

correcting for multiple testing, we identified several metabolites annotated with a

confidence level 2 by xmsannotator that were associated with AD at p<10e-03 (Table

2). In particular, we identified benzyl chloride, various adducts of benzenes and

toluenes, omega 3 fatty acids, ceramide and carnitines that are associated with

Alzheimer's Disease.

In addition, features from the HILIC positive data indicated that altered polyunsaturated

fatty acid biosynthesis, alanine, aspartate, glutamate, glycerophospholipid and

sphingolipid metabolism (figure 1C) were also associated with AD metabolome-wide.

Data from the C18 negative column implicated altered glycolysis and gluconeogenesis,

pyruvate and alanine and aspartate metabolism in cases versus controls of AD.

Individual metabolites that contributed to enrichment of these pathways are described in

Supplementary Material.

Metabolomic differences by ethnic/racial group. Within each ethnic/racial group, we

found that African Americans with AD had altered glycolysis and amino acid metabolism

as well as polyunsaturated fatty acid metabolism. Non-Hispanic whites with AD had

altered amino acid, fatty acid, and glycosphingolipid metabolism, and Caribbean

Hispanic cases had altered amino acid metabolism (figure 2). Restricting the analysis

to controls, we determined underlying metabolic differences that existed in participants

based on their ethnic/racial group was not driven by the presence of disease (figure 3A,

Table S1).

Compared with Caribbean Hispanics and non-Hispanic whites, partial least squares

discriminant analysis (PLS-DA) suggested features in African Americans clustered

differently (Figure 3), including altered amino acid metabolic pathways such as arginine

and proline metabolism, asparate and asparagine metabolism, and gylcine, serine,

alanine and threonine metabolism. Saturated fatty acid beta oxidation was also

identified based on variable importance in the projection (VIP) scores from the PLS-DA

(figure 3).

Metabolomic profiles associated with APOE genotype. We also investigated

metabolic differences driven by APOE-ε4. PLS-DA identified near complete separation

between APOE $\varepsilon 4$ carriers and non-carriers (Table S3). In contrast to pathways

identified by AD diagnosis, the presence of APOE-ε4 was associated with changes in

arachidonic acid metabolism, driven by features with putative matches to

octadecatrienoic acid and icosatrienoic acid (figure 4A and B), changes in the pentose

phosphate pathway, and in chondroitin sulfate. After adjusting for APOE-ε4, the

pathways associated with AD were largely the same (figure 1C).

Discussion

We compared untargeted metabolomics in patients with AD and healthy controls.

equally divided between African Americans, Caribbean Hispanics and non-Hispanic

whites. The goal of this investigation was to identify metabolites associated with

disease while adjusting for the strongest genetic risk factor, APOE and ethnic/racial

group. Therefore, we used a simple case-control design equally stratified by

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ethnic/racial group.

We found significant differences comparing exogenous and endogenous metabolites in patients with AD and healthy controls. Similar differences were observed in a comparison of metabolic profiling in brain tissue from the middle frontal and inferior temporal gyri in healthy controls, patients with AD or mild cognitive impairment (MCI) in the Baltimore Longitudinal Study¹⁷. An untargeted approach has also been used to correctly classify patients with mild cognitive impairment or AD when compared with controls with 97.7% accuracy¹⁸. However, metabolite analyses identified 22 putative biochemical pathways, including interlinked areas of metabolism (polyamine metabolism and L-arginine metabolism).

In this report, we identified several metabolites associated with AD that were altered in the fatty acid biosynthesis pathway. Interestingly, dysregulation of unsaturated fatty acid metabolism was previously reported in brain tissue from patients with AD compared to healthy controls¹⁷. The disruption in polyunsaturated fatty acid biosynthesis may have several downstream effects: inflammation, oxidative stress, cell maintenance or cell death. Several amino acid metabolism related pathways were also enriched among metabolites that were associated with risk of AD. Lower levels of glutamate and aspartate, and higher levels of glutamine have been observed in the temporal cortex, in postmortem samples from patients with AD¹⁹. A different cohort also showed elevations in plasma levels of glutamine in AD patients²⁰. We identified metabolites involved in dynorphin metabolism associated with AD, consistent with previous studies in postmortem samples from patients²¹. Expression of dynorphin, an endogenous opioid

peptide, increases with age and has been associated with cognitive impairment in rodent models²².

We found ceramide to be significantly associated with AD, and it has been suggested that ceramide promotes neuronal apoptosis, Aß accumulation²³ and has been seen at increased levels in patients with AD and co-morbid conditions²⁴. In addition, lower levels of plasma acylcarnitine were found in AD patients compared to controls. Plasma acylcarnitines predict altered fatty acid beta-oxidation, and ketogenesis and have been reported to be depressed in AD patients²⁵. Metabolic features also differed significantly in the three ethnic/racial groups, especially among African Americans, despite the individuals being similar in age and residing in the same geographical location. Very few studies have investigated overall metabolic differences by ethnicity/race. Investigators from the Women Health Initiative cohort identified significant differences in metabolite profiles between African Americas and individuals of European ancestry (https://www.ahajournals.org/doi/abs/10.1161/circ.139.suppl 1.MP55). Similar differences in amino acid and lipid metabolism were observed in a metabolic profiling study of serum metabolites in African American and European American patients with bladder cancer²⁶. Thus, genetic and metabolic factors could help to explain ethnic/racial and racial disparities in the risk factors associated with AD. Similar pathways emerge when we asses difference by AD or ethnic/racial group suggesting that metabolome profiling from plasma could be used to identify racial differences in disease.

Differences between APOE $\varepsilon 4$ carriers and non-carriers were apparent and remained when analyses were restricted to healthy individuals from all three ethnic/racial groups.

Interestingly, metabolites associated with AD were different from those associated with

APOE $\varepsilon 4$. Hexose phosphorylation and arachidonic acid metabolism pathways were

most enriched amongst metabolites that were associated with APOE $\varepsilon 4$. APOE $\varepsilon 4$

carriers converting to MCI/AD had higher arachidonic acid (AA)/docosahexaenoic acid

(DHA) ratios in phospholipids compared to cognitively normal ε4 and non-ε4 carriers²⁷.

Alterations in plasma arachidonic acid were observed in the brains of ε4 carrier mice

compared to non-carriers²⁷.

Taken together these results suggest that understanding metabolic heterogeneity in AD

pathogenesis may enable identification of biological mechanisms for specific subgroups

with the disease. Namely, those carrying an *APOE-ε4* allele and among those of African

or Hispanic ancestry. This study demonstrates the ability of untargeted metabolomics

to reveal biochemical differences in plasma based on ethnicity/race, the presence of

APOE $\varepsilon 4$, and AD, thus informing study design for optimal power of discovery in larger

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studies.

Acknowledgements

WHICAP. Data collection and sharing for this project was supported by the Washington Heights-Inwood Columbia Aging Project (WHICAP, PO1AG07232, R01AG037212, RF1AG054023 and R56AG063908) funded by the National Institute on Aging (NIA) and by the National Center for Advancing Translational Sciences, National Institutes of Health, through Grant Number UL1TR001873. The metabolomics work was supported by U2C ES030163 and R01 ES023839. This manuscript has been reviewed by WHICAP investigators for scientific content and consistency of data interpretation with previous WHICAP Study publications. We acknowledge the WHICAP study participants and the WHICAP research and support staff for their contributions to this study.

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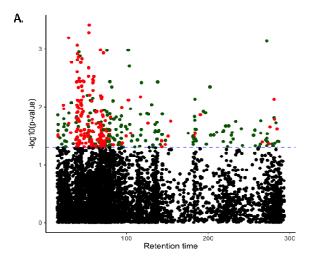
Figure 1A-D

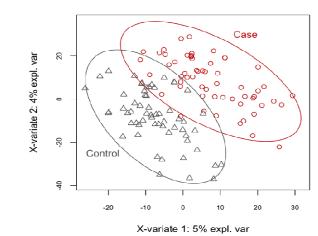
Metabolome wide association study in cases and controls of AD.

- A. A metabolome wide association study (MWAS) found 382 features altered in cases and controls at nominal significance of p<0.05 In red: features lower in cases, in green features higher in cases:
- B. A PLS-DA shows metabolomic patterns different between cases and controls (unadjusted for age and sex variables).
- C. Pathway analysis using mummichog shows most probable metabolic pathways altered between cases and controls.. We tested enrichment of pathways among metabolites that were nominally significant in association with AD (p<0.05) in models unadjusted and adjusted for APOE respectively. Displayed pathways are enriched at p<0.05, corrected for multiple testing.

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Pathways altered when in cases and controls with and without APOE adjustment

Size of bubble represents number of significant hits Enrichment is calculated as (Total Hits/Pathway size)

В.

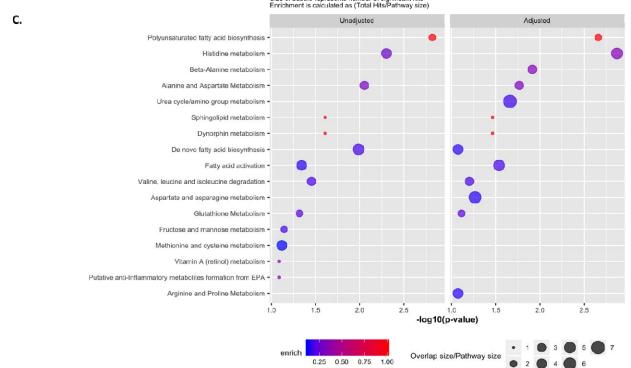
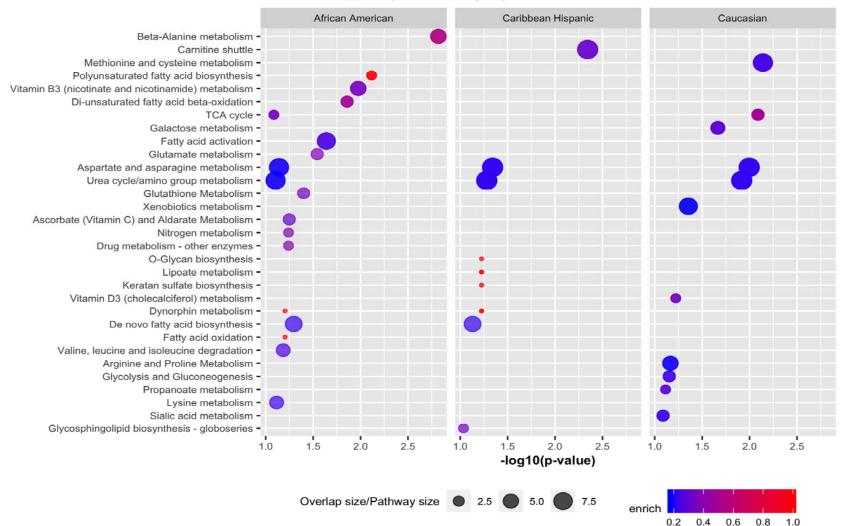


Figure 2. Ethnic differences. A bubble plot that shows pathways altered in controls of the three ethnic groups. Size of bubbles is proportional to overlap size of pathway and p-values are corrected for multiple testing.

Pathways altered in cases across different ethnicities

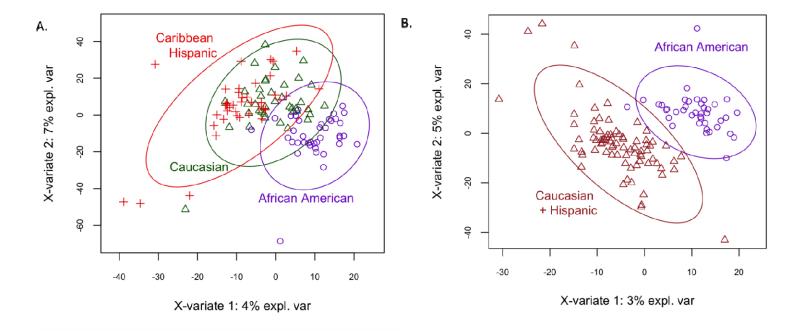
Size of bubble represents number of significant hits Enrichment is calculated as (Total Hits/Pathway size)

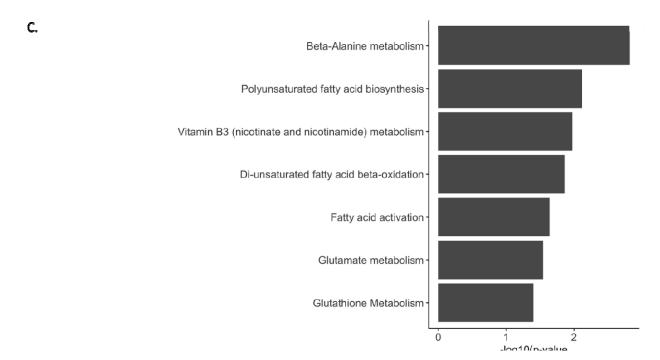


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Figure 3. African Americans v others

- A. A PLS-DA with all ethnic groups shows a African Americans differ from the Caucasian and Hispanic individuals.
- B. A PLS-DA the shows differences when comparing AAs with all others.
- C. Pathways analysis of features that are significantly different in cases and controls of African Americans.
- D. Distribution of top 3 features significantly different in cases and controls of African Americans in African American and other ethnic groups combined.

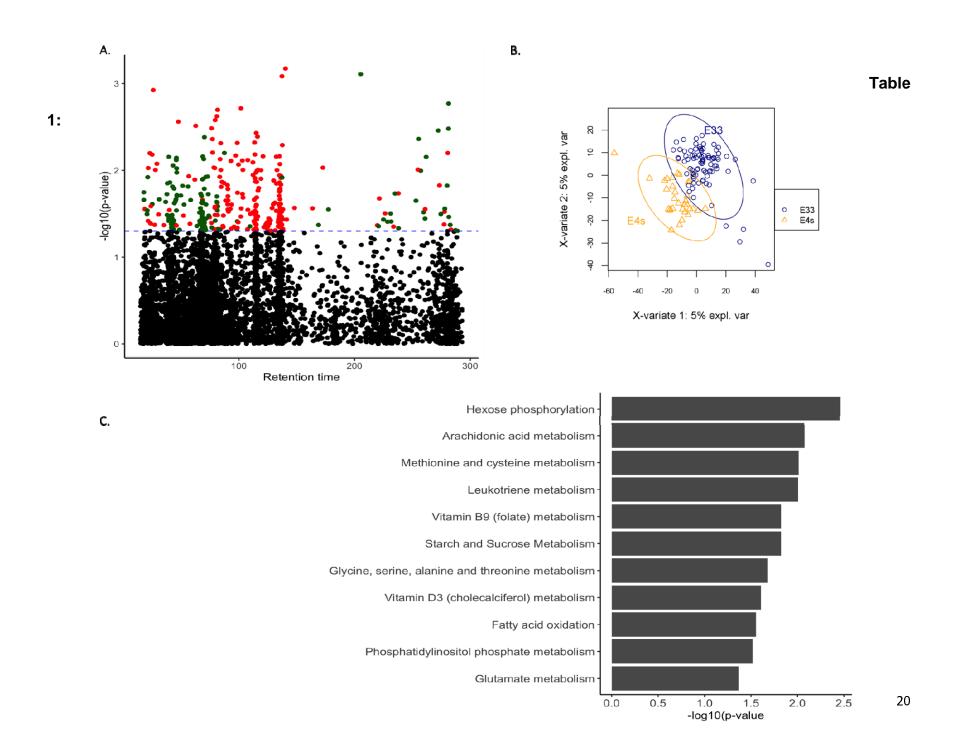




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Figure 4. APOE status and metabolic differences.

- A. MWAS shows features associated with APOE status.
- B. PLS-DA showing separation between APOE ε4 carriers and non-carriers
- C. Metabolic pathways enriched in metabolites that are associated with APOE $\epsilon 4$ status at a nominal significance of p<0.05.



Demographic characteristics of cases and controls analyzed in the study

	Cases	Controls
Caucasians (N)	19	20
Age	89.21301	93.00000
Sex (% female)	79%	60%
APOE allele frequency (%)	13.2	2.5
African Americans (N)	20	20
Age	86.38012	91.05000
Sex (% female)	80%	90%
APOE allele frequency (%)	30	17.5
Hispanics (N)	20	20
Age	83.80178	91.55000
Sex (% female)	85%	90%
APOE allele frequency (%)	15	10

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Table 2. Individual metabolites associated with AD status and annotated with a confidence score>2.

mz	time	BETA	Р	chemical_ID	Match Category	Theoretical mz	delta_ppm	Name	Formula	Monoisotopic Mass	
127.03	53.80	-0.82	5.23E-04	HMDB59882	Unique	127.03	7.08	Benzyl chloride	C7H7CI	126.02	
		HMDB59877		Propylbenzene	C9H12	120.09					
	21.10 271.70 0.79 7		7.31E-04	HMDB59819	Multiple				o-Ethyltoluene		
				HMDB13733						124-Trimethylbenzene	
121.10		0.79		HMDB40458		121.10	21.10 0.17	124- Tris(methylene)cyclohexane			
		7.512 01	HMDB59901	TTGTT.		0.27	Hemimellitene	C9H12	120.09		
		HMDB41924 HMDB59848 HMDB34029		Mesitylene		¥Va⊪e					
				HMDB59848				m-Ethyltoluene		i i	
				HMDB34029							
594.58	68.20	-0.72	1.04E-03	HMDB04951	Unique	594.58	0.62	Ceramide (d18:120:0)	C38H75NO3	593.57	
291.20	41.60	0.72	1.12E-03	HMDB31098	Multiple	291.20	1.27	(9Z11E13E15Z)-4-Oxo- 9111315- octadecatetraenoic acid	C18H26O3	290.19	
				HMDB13288				Nonanoylcarnitine			
302.23	41.10	-0.79	1.38E-03	HMDB06320	Multiple	302.23	3 0.93	26 Dimethylheptanoyl carnitine	C16H31NO4	301.23	

References

- 1. Yoshida M, Hatano N, Nishiumi S, et al. Diagnosis of gastroenterological diseases by metabolome analysis using gas chromatography-mass spectrometry. *J Gastroenterol*. 2012;47(1):9-20.
- 2. McKhann GM, Knopman DS, Chertkow H, et al. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's & dementia : the journal of the Alzheimer's Association*. 2011;7(3):263-269.
- 3. Soltow QA, Strobel FH, Mansfield KG, Wachtman L, Park Y, Jones DP. High-performance metabolic profiling with dual chromatography-Fourier-transform mass spectrometry (DC-FTMS) for study of the exposome. *Metabolomics : Official journal of the Metabolomic Society.* 2013;9(1 Suppl):S132-S143.
- 4. Jones DP, Park Y, Ziegler TR. Nutritional metabolomics: progress in addressing complexity in diet and health. *Annual review of nutrition*. 2012;32:183-202.
- 5. Liu KH, Walker Dl, Uppal K, et al. High-Resolution Metabolomics Assessment of Military Personnel: Evaluating Analytical Strategies for Chemical Detection. *J Occup Environ Med.* 2016;58(8 Suppl 1):S53-61.
- 6. Yu T, Park Y, Johnson JM, Jones DP. apLCMS--adaptive processing of high-resolution LC/MS data. *Bioinformatics*. 2009;25(15):1930-1936.
- 7. Uppal K, Soltow QA, Strobel FH, et al. xMSanalyzer: automated pipeline for improved feature detection and downstream analysis of large-scale, non-targeted metabolomics data. *BMC Bioinformatics*. 2013;14:15.
- 8. Li S, Park Y, Duraisingham S, et al. Predicting network activity from high throughput metabolomics. *PLoS Comput Biol.* 2013;9(7):e1003123.
- 9. Johnson JM, Yu T, Strobel FH, Jones DP. A practical approach to detect unique metabolic patterns for personalized medicine. *Analyst.* 2010;135(11):2864-2870.
- 10. Schymanski EL, Jeon J, Gulde R, et al. Identifying small molecules via high resolution mass spectrometry: communicating confidence. *Environ Sci Technol.* 2014;48(4):2097-2098.
- 11. Johnson WE, Li C, Rabinovic A. Adjusting batch effects in microarray expression data using empirical Bayes methods. *Biostatistics*. 2007;8(1):118-127.
- 12. Menni C, Fauman E, Erte I, et al. Biomarkers for type 2 diabetes and impaired fasting glucose using a nontargeted metabolomics approach. *Diabetes*. 2013;62(12):4270-4276.
- van den Berg RA, Hoefsloot HC, Westerhuis JA, Smilde AK, van der Werf MJ. Centering, scaling, and transformations: improving the biological information content of metabolomics data. *BMC Genomics*. 2006;7:142.
- 14. Rohart F, Gautier B, Singh A, Le Cao KA. mixOmics: An R package for 'omics feature selection and multiple data integration. *PLoS Comput Biol.* 2017;13(11):e1005752.
- 15. Berriz GF, King OD, Bryant B, Sander C, Roth FP. Characterizing gene sets with FuncAssociate. *Bioinformatics*. 2003;19(18):2502-2504.
- 16. Hosack DA, Dennis G, Jr., Sherman BT, Lane HC, Lempicki RA. Identifying biological themes within lists of genes with EASE. *Genome Biol.* 2003;4(10):R70.
- 17. Snowden SG, Ebshiana AA, Hye A, et al. Association between fatty acid metabolism in the brain and Alzheimer disease neuropathology and cognitive performance: A nontargeted metabolomic study. *PLoS Med.* 2017;14(3):e1002266.
- 18. Graham SF, Chevallier OP, Elliott CT, et al. Untargeted metabolomic analysis of human plasma indicates differentially affected polyamine and L-arginine metabolism in mild cognitive impairment subjects converting to Alzheimer's disease. *PloS one*. 2015;10(3):e0119452.

- 19. Gueli MC, Taibi G. Alzheimer's disease: amino acid levels and brain metabolic status. *Neurol Sci.* 2013;34(9):1575-1579.
- 20. Niedzwiecki M, Walker D, Howell CJ, et al. High-resolution metabolomic profiling of Alzheimer's disease in plasma. *Ann Clin Transl Neurol*. 2019:In Press.
- 21. Yakovleva T, Marinova Z, Kuzmin A, et al. Dysregulation of dynorphins in Alzheimer disease. *Neurobiol Aging*. 2007;28(11):1700-1708.
- 22. Menard C, Herzog H, Schwarzer C, Quirion R. Possible role of dynorphins in Alzheimer's disease and age-related cognitive deficits. *Neurodegener Dis.* 2014;13(2-3):82-85.
- Jazvinscak Jembrek M, Hof PR, Simic G. Ceramides in Alzheimer's Disease: Key Mediators of Neuronal Apoptosis Induced by Oxidative Stress and Abeta Accumulation. *Oxid Med Cell Longev.* 2015;2015:346783.
- Filippov V, Song MA, Zhang K, et al. Increased ceramide in brains with Alzheimer's and other neurodegenerative diseases. *J Alzheimers Dis.* 2012;29(3):537-547.
- 25. Ciavardelli D, Piras F, Consalvo A, et al. Medium-chain plasma acylcarnitines, ketone levels, cognition, and gray matter volumes in healthy elderly, mildly cognitively impaired, or Alzheimer's disease subjects. *Neurobiol Aging*. 2016;43:1-12.
- 26. Vantaku V, Donepudi SR, Piyarathna DWB, et al. Large-scale profiling of serum metabolites in African American and European American patients with bladder cancer reveals metabolic pathways associated with patient survival. *Cancer*. 2019;125(6):921-932.
- 27. Abdullah L, Evans JE, Emmerich T, et al. APOE epsilon4 specific imbalance of arachidonic acid and docosahexaenoic acid in serum phospholipids identifies individuals with preclinical Mild Cognitive Impairment/Alzheimer's Disease. *Aging (Albany NY)*. 2017;9(3):964-985.

S1. PLS-DA performance in separating three ethnic groups using all 119 individuals (60 AD cases and 59 controls).

Component 1					
	AUC	p-value			
AFRICAN AMERICAN vs Other(s)	0.9275	2.95 E-14			
EUROPEAN vs Other(s)	0.5478	3.99E-01			
HISPANIC vs Other(s)	0.8804	1.36E-11			
Component 2					
	AUC	p-value			
AFRICAN AMERICAN vs Other(s)	0.9816	0.00E+00			
EUROPEAN vs Other(s)	0.6904	7.71E-04			
HISPANIC vs Other(s)	0.8994	1.25 E-12			
Component 3					
	AUC	p-value			
AFRICAN AMERICAN vs Other(s)	0.9972	0.00E+00			
EUROPEAN vs Other(s)	0.9804	0.00E+00			
HISPANIC vs Other(s)	0.9601	2.22E-16			

S2. PLS-DA performance in separating 60 AD cases from 59 age-matched healthy controls.

Component 1					
	AUC	p-value			
AD vs HEALTHY	0.8842	4.89E-13			
Component 2					
	AUC	p-value			
AD vs HEALTHY	0.9907	0			
Component 3					
	AUC	p-value			
AD vs HEALTHY	1	0			

S3. PLS-DA performance in separating APOE4 carriers from non-carriers

Component 1				
	AUC	p-value		
E33 vs E4s	0.919	5 2.56E-12		
Component 2				
	AUC	p-value		
E33 vs E4s	0.99	5 2.22E-16		
Component 3				
	AUC	p-value		
E33 vs E4s		1 0		