



## Supplementary Materials for

### **Cenozoic evolution of deep ocean temperature from clumped isotope thermometry**

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#### **The PDF file includes:**

Materials and Methods  
Figs. S1 to S7  
Tables S1 and S2  
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#### **Other Supplementary Material for this manuscript includes the following:**

Data S1 to S3

## Materials and Methods

### Material

#### 1. Newfoundland Margin

Large individual samples, for the most part core catchers, were taken from four different International Ocean Discovery Program (IODP) sites recovered by Expedition 342 on the Newfoundland margin in the North Atlantic (1406, 1407, 1409, and 1410; Table S1, Fig. S1), together covering the entire Cenozoic era (44). Age models for all Sites are based on detailed integrated bio-magneto-stratigraphies (44), updated to the CENOGRID timescale (2). Other supporting datasets that we show in Figures 1 and 2 (atmospheric CO<sub>2</sub> and previous estimates of deep ocean temperature and  $\delta^{18}\text{O}_{\text{sw}}$ ) are plotted on their published age models; minor resulting differences in age models are not relevant on the timescales investigated here.

For each sample, benthic foraminifera were picked from size fractions >150  $\mu\text{m}$ . A large variety of foraminifer species were used for analysis in order to achieve the necessary level of replication (Data S2), whereby aliquots for measurements were separated on the species or at least genus level where possible. Aliquots were cleaned by carefully opening foraminifera tests, followed by several iterations of ultrasonication using both deionized water and methanol, with subsequent rinses with deionized water and oven-drying at 50°C.

Scanning electron microscope images were obtained from a random selection of cleaned sample aliquots distributed over the whole study interval (Fig. S2).

#### 2. Walvis Ridge

For the Early Eocene, one additional clumped isotope temperature was obtained from Site 1263 (paleo-water depth ~1500 m) on the Walvis Ridge in the South Atlantic as an average of analyses from multiple individual samples spanning ~400 kyr (from 51.3 to 50.9 Ma). Foraminifer species *Nuttalides truempyi* and *Oridorsalis umbonatus* from the > 150  $\mu\text{m}$  size fraction were cleaned and analyzed at Utrecht University.

### Methods

#### 1. Clumped isotope thermometry

Clumped isotope thermometry takes advantage of the temperature dependence of isotopic ordering within molecules, with double substitution by heavy isotopes (e.g., <sup>18</sup>O and <sup>13</sup>C in carbonates) increasing with decreasing formation temperature (13). The excess abundance of such double substitution in carbonates compared to random ordering ( $\Delta_{47}$ ) is independent of the composition of the source water. The signal furthermore does not exhibit biological effects in foraminifera (46-50), appears to not be measurably affected by seawater pH within the range typical for the ocean (28, 30), and is largely robust to diagenetic overprinting in benthic foraminifera from typical ocean settings (19), if burial depth was sufficiently shallow (51). Using carbonate-based standardization

(52), calibrations can be reproduced in different laboratories, and foraminifera-based calibrations are indistinguishable from inorganically precipitated calcite (46, 47, 50). The comparatively large analytical uncertainty due to a small signal-to-noise ratio can be mitigated by extensive replication, where sample availability permits.

Clumped isotope measurements were performed between 2015 and 2020 on four different mass spectrometers in Bergen, Zürich, and Utrecht, all consisting of Thermo Fisher Scientific MAT253 or MAT253Plus instruments coupled to Thermo Fisher Scientific Kiel IV carbonate preparation devices. Most of the analyses ( $N = 936$ ) were performed on two different mass spectrometers in Bergen, with the exception of some North Atlantic sample aliquots measured in Zürich ( $N = 43$ ), and early Eocene samples from Walvis Ridge analyzed in Utrecht ( $N = 61$ ). The analytical approach was similar to that described by (53) and updated for the Bergen analyses as described by (49) and (46). The methodological details for the Utrecht lab are described in (18). In the Kiel device, sample aliquots are individually reacted with phosphoric acid at 70°C, and the sample gas is cryogenically separated from water and other trace gases. All Kiel devices were equipped with additional custom-built traps for organic contaminants (Porapak Q columns, held at -20°C; -45°C in the Utrecht lab) and sulfide components (Ag wool). All measurements were conducted in microvolume mode, and in the case of the Bergen and Utrecht instruments, the long-integration-dual-inlet method (LIDI; 54) was used. Masses 44-49 were monitored, and masses 44-47 were used for calculating  $\delta^{13}\text{C}$ ,  $\delta^{18}\text{O}$ , and  $\Delta_{47}$ . The raw data were corrected for pressure baseline effects based on daily peak scans (53) and further corrections for  $\Delta_{47}$  scale compression and transfer to the I-CDES scale (55) were made using carbonate standards ETH 1-3. The accepted values of these standards were derived by multiple laboratories (55) and reported for an acid digestion temperature of 90 °C. Our  $\Delta_{47}$  (I-CDES) results therefore also reflect an acid digestion temperature 90 °C (but note that by using carbonate standards, no acid fractionation factor needs to be applied). Standard aliquots were measured in roughly equal numbers as sample aliquots. In most cases, a moving window approach was used to base corrections on standard data from several adjacent days, with the number of standard measurements used for corrections depending on instrument stability. For the measurements performed in Bergen, data corrections were done with the Easotope software (56). All replicate-level data are archived in the Earthchem database (41, 42), including information regarding standard correction procedures. Sample-averaged data are reported as Data S1 and are archived in the Pangaea database (43).

Final corrected  $\Delta_{47}$  values on the I-CDES scale were averaged per sample, before temperatures were calculated using the combined foraminifera-based calibration of (46), updated to the I-CDES scale by (57), that is based on three different calibration datasets measured in two different laboratories:

$$\Delta_{47}(I - CDES) = (0.0397 \pm 0.0011) * \frac{10^6}{T^2} + (0.1518 \pm 0.0128)$$

Thereby, temperature is given in °K. Choosing a different calibration has only a small effect on the results (Fig. S3), as long as calibrations are based on the same carbonate standardization

approach. We tested i) the travertine-based calibration of (58), as recalculated by (52), using  $\Delta_{47}$  from our samples standardized to the original values of the ETH standards published by (52), and ii) the combined inorganic-biogenic carbonate calibration of (59) covering a temperature range of more than 1000 °C. The differences between temperatures calculated with these equations and our preferred approach are -1.0 to 0.9 °C for (i) and -1.9 °C for (ii). For this study, we chose to use the foraminifera-based calibration (46), as it contains most foraminifera data, treats these in a consistent way (e.g., calculation of calcification temperatures), and has a larger data density in the interval of ocean temperatures compared to the travertine calibration. It furthermore avoids the influence of analytical uncertainty of sparse measurements on high-temperature samples and possible effects of the  $\Delta_{47}$  dependence on temperature deviating from a linear relationship with  $1/T^2$  over large temperature ranges (60).

All temperatures are plotted with 68% and 95% confidence intervals on the average temperatures, which combine the analytical uncertainty and the calibration uncertainty of (46; their Fig. A2), updated by (57) using a Monte Carlo approach that samples a random slope-intercept pair for the calibration and a random  $\Delta_{47}$  value from their respective probability distributions (5000 iterations). In addition, the number of replicates is indicated by different gray shading in order to optically give more weight to the better replicated analyses. LOESS smoothing was applied to the full North Atlantic temperature data set, taking into account the confidence levels of each data point. Assuming normal distribution of errors, we applied a Monte Carlo method to generate 10,000 realizations of the temperature time series. A LOESS curve (span = 0.2, degree = 1) was then fitted to each of the 10,000 realizations. These LOESS fits served as a basis for calculating 95% confidence intervals (2.5 and 97.5 percentiles; gray shading in Figures 1 and 2).

## 2. $\delta^{13}\text{C}$ , $\delta^{18}\text{O}$ , $\delta^{18}\text{O}_{\text{sw}}$

$\delta^{13}\text{C}$  and  $\delta^{18}\text{O}$  values were calibrated to the Vienna Pee Dee Belemnite (VPDB) scale with the ETH 1-4 carbonate standards that were also used for  $\Delta_{47}$  corrections in a similar moving window approach with standard results from several days. The clumped isotope standards have previously been calibrated to the VPDB scale using the international carbonate standards NBS-18, NBS-19, and LSVEC (52). In the case of  $\delta^{18}\text{O}$ , the standard correction also accounted for minor variations in  $\delta^{18}\text{O}$  scale compression.

$\delta^{18}\text{O}_{\text{sw}}$  was calculated from the  $\Delta_{47}$  temperatures and the  $\delta^{18}\text{O}_b$  values determined from *Cibicidoides spec.*, using a temperature-  $\delta^{18}\text{O}_b$  relationship derived for that genus (7; their equation 9). For early Eocene data from Walvis Ridge and Maud Rise, where *Cibicidoides* were not measured,  $\delta^{18}\text{O}_b$  from *Nuttalides* and *Oridorsalis* were first corrected for inter-species offsets from *Cibicidoides* using the relationships suggested by (9).

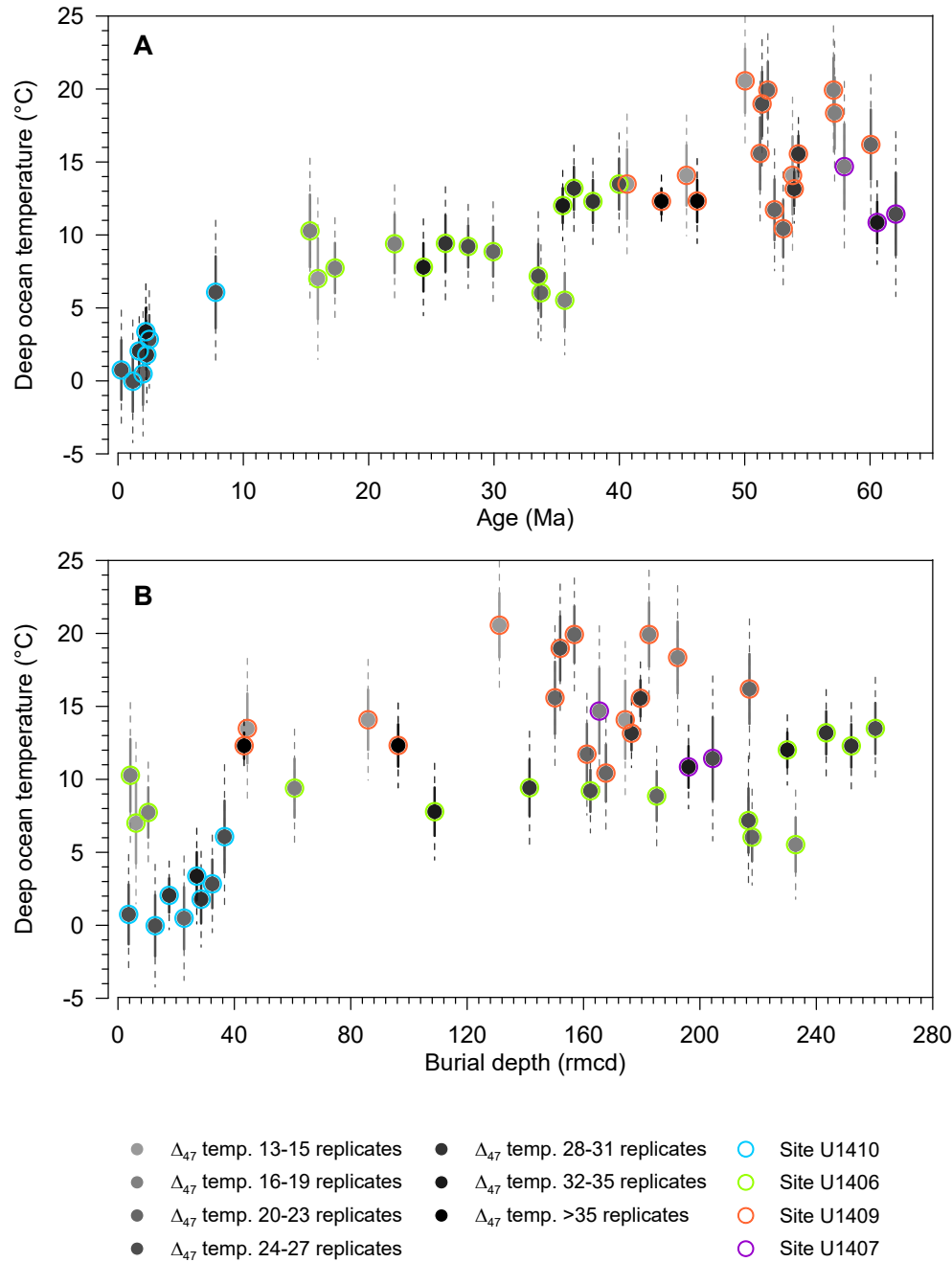
### 3. Deep Ocean pH

Boron isotope data from benthic foraminifera were generated from the North Atlantic (U1409) and Central Atlantic (ODP 1258 and 1260, 45-52 Ma) and combined with published data from the Central Atlantic (ODP 999 and 926; 40) – see Table S2 for details. These sites span paleo-depths from 2.5-3.6 km, which are today bathed by northern-sourced waters with similar properties (61), and we find consistent values between sites in our record. Analyses were made on mono-specific samples of the genus *Cibicidoides*: *C. wuellerstorfi* and *C. mundulus* in the Neogene and *C. eoceanus*, *C. havanensis*, and *C. velascoensis* in the Paleogene.

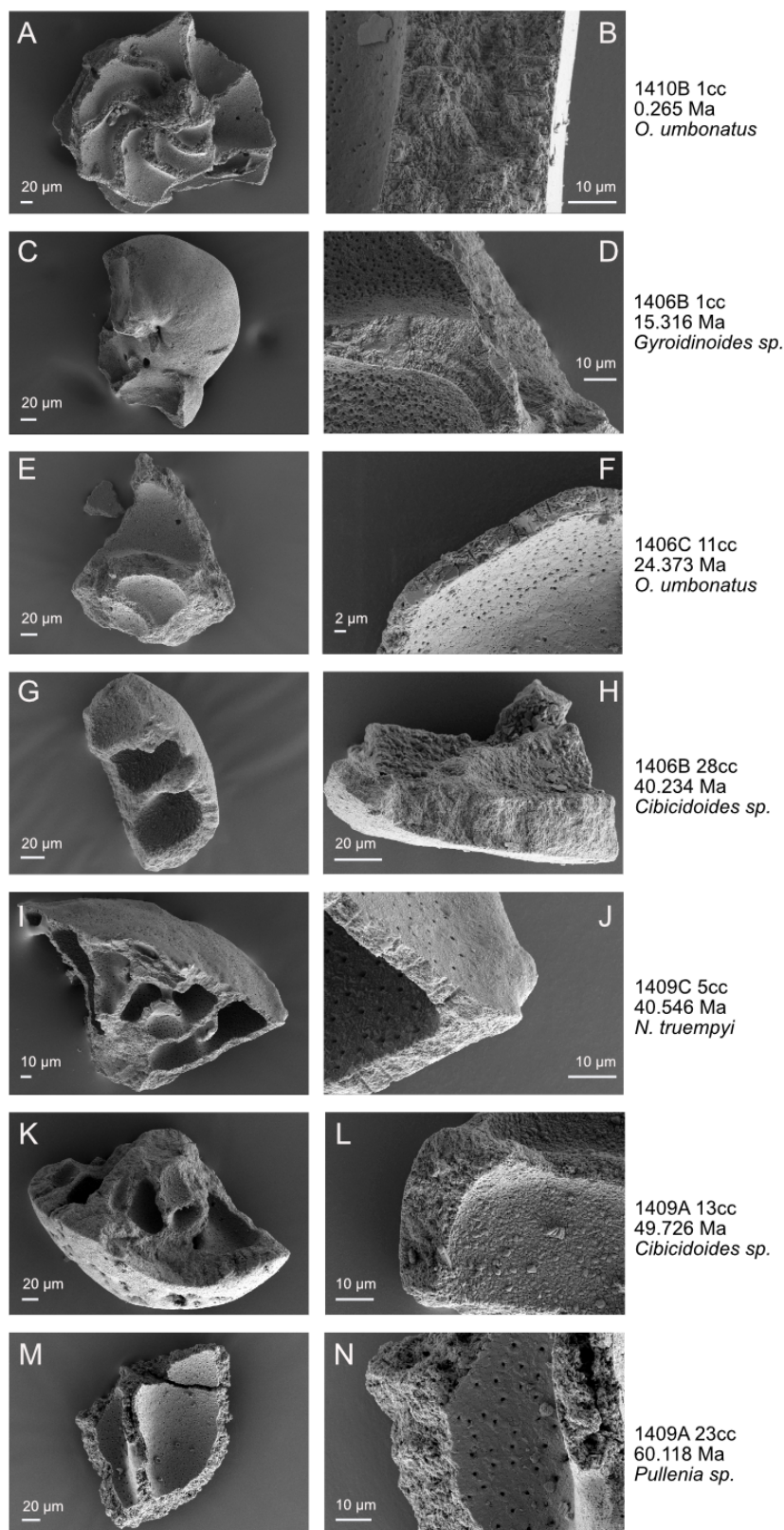
Analytical methods followed established protocols (e.g., 62-64). In brief, foraminifera were crushed, oxidatively cleaned with hydrogen peroxide, and dissolved following (62). A small split (<5 %) of the sample was analyzed for minor and trace element composition by ICPMS to assess boron content and sample cleanliness. Boron was then removed from the sample matrix, using ion exchange column chromatography with Amberlite 743, and analyzed by MC-ICPMS. Data were obtained from site U1409 in the St Andrews isotope Geochemistry (STAiG) laboratories at the University of St Andrews and from sites 1258 and 1260 in the Bristol Isotope Group (BIG) at the University of Bristol. Analyses in these laboratories follow very similar protocols, though with some minor differences (boron washout assisted using  $\text{NH}_3$  at BIG, HF at STAiG; and signal detection using  $10^{11} \Omega$  amplifiers at BIG,  $10^{13} \Omega$  amplifiers at STAiG). Cross calibration between these laboratories has been ensured by participation in interlaboratory comparison exercises (64-66) and use of identical primary and consistency standards, yielding consistent values (e.g. the pooled 2SD of boric acid consistency standards measured across both laboratories during these analytical sessions is 0.17 ‰, n=8). Uncertainties are reported at 2SD based on long-term reproducibility of samples of this size in each lab.

Calculation of deep ocean pH as shown in Figure 2C follows the procedures in (67) and (24). We assume that these epifaunal *Cibicidoides* species record bottom water  $\delta^{11}\text{B}$  of borate, as seen in modern calibrations (62). To calculate the boric acid dissociation constant ( $K_B$ ) we interpolate the smoothed clumped isotope temperature record presented here (Fig. 1A), use salinity of 35, paleo water depths as shown in Table S2 (noting that depth and salinity have little influence on calculated pH), and we account for the (minor) influence of changing seawater  $[\text{Ca}^{2+}]$  and  $[\text{Mg}^{2+}]$  (e.g., 68) using MyAMI (69) as in (24). We use the compilation of  $\delta^{11}\text{B}_{\text{SW}}$  estimates from (24). The influence of  $\pm 0.5 \text{ ‰}$  in  $\delta^{11}\text{B}_{\text{SW}}$  on pH is shown with the shaded area in Figure 2C and the influence of analytical uncertainties in  $\delta^{11}\text{B}$  and  $\pm 2 \text{ °C}$  in temperature are shown by the 2SD error bars, based on the standard deviation of a 10,000 member Monte Carlo ensemble.

The deep ocean pH data are reported as Data S3 and archived in the Earthchem and Pangaea databases.

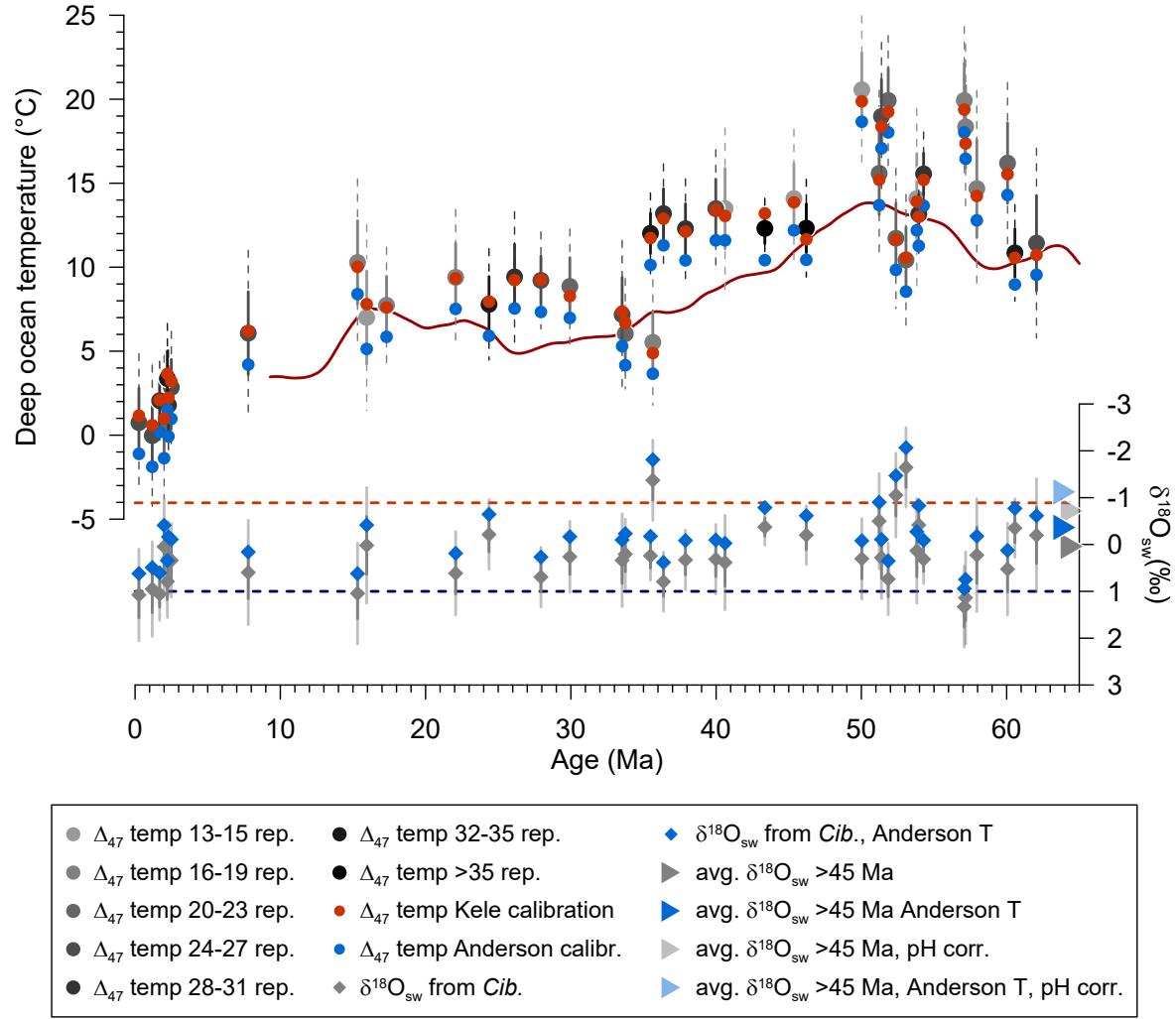


**Fig. S1: North Atlantic IODP Expedition 342 Sites used for the different parts of the record (A) and clumped isotope temperatures versus burial depth (B).** Note the close reproducibility of temperatures at 40 Ma, with overlapping data from two sites with very different burial depths in each (1406B core 28 at 260 m vs. 1409C core 5 at 44 m). See also the respective SEM images of foraminifer tests from each of these two samples in Figure S2. Burial depth is given in revised meter composite depth (rmcd; 44). We do not observe a consistent relationship between clumped isotope temperatures and burial depth (panel B).



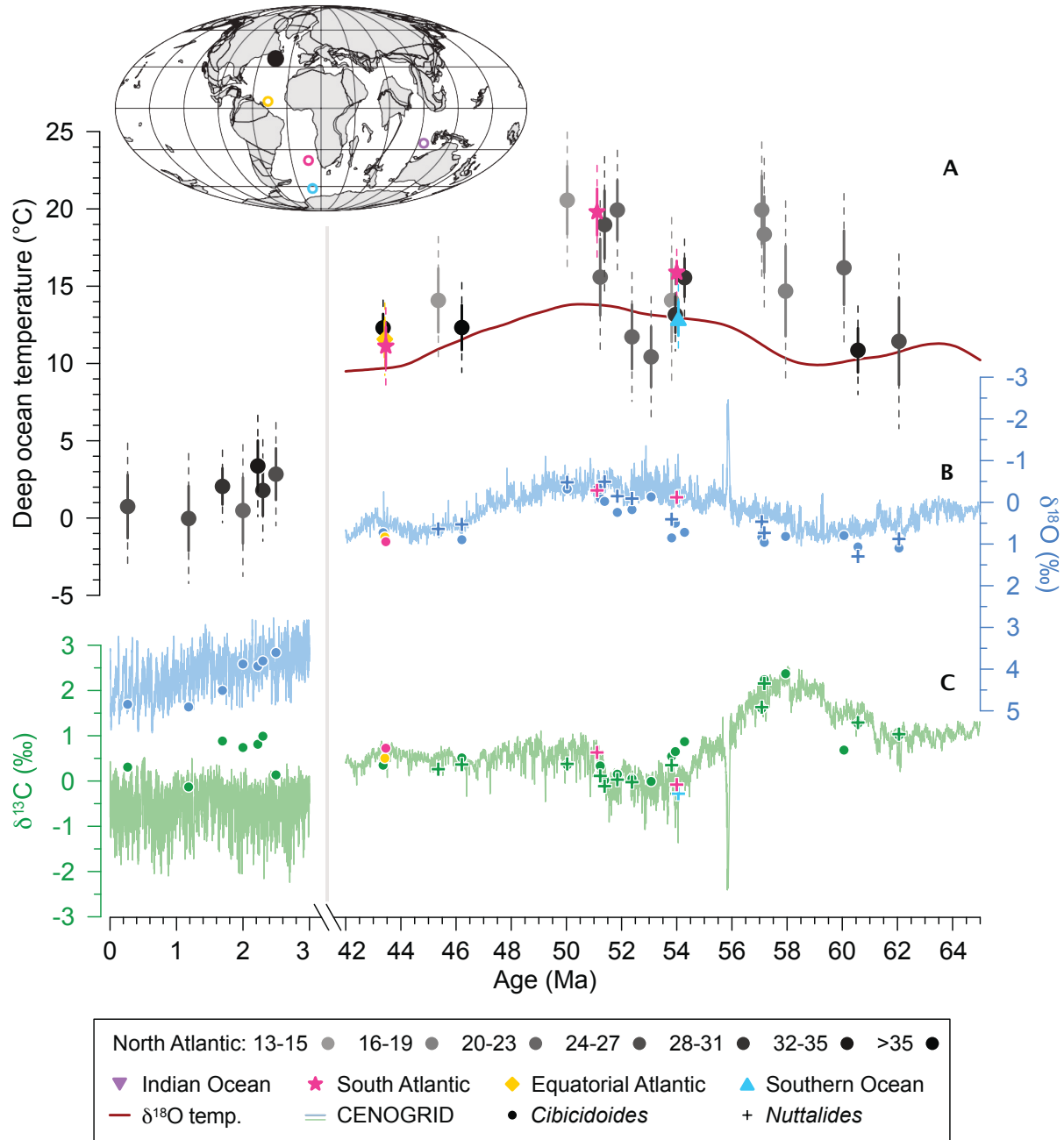
**Fig. S2. SEM images of benthic foraminifera analyzed for clumped isotopes spanning the Cenozoic record.**

Preservation state is generally good to excellent (clearly visible microstructures such as pores and layered structure of wall cross-sections) but shows the expected deterioration (addition of blocky calcite overgrowth) with depth (increasing core number X in "Xcc"). Note for example the different extent of overgrowth on two samples for ~40 Ma stemming from different sites and burial depths (G, H vs. I, J). The sample from 1406B 28cc shows blocky calcite overgrowth on the inside of the shell fragment (H), which is not observed on the fragment from 1409C 5cc (J). These samples yielded nonetheless indistinguishable temperatures.

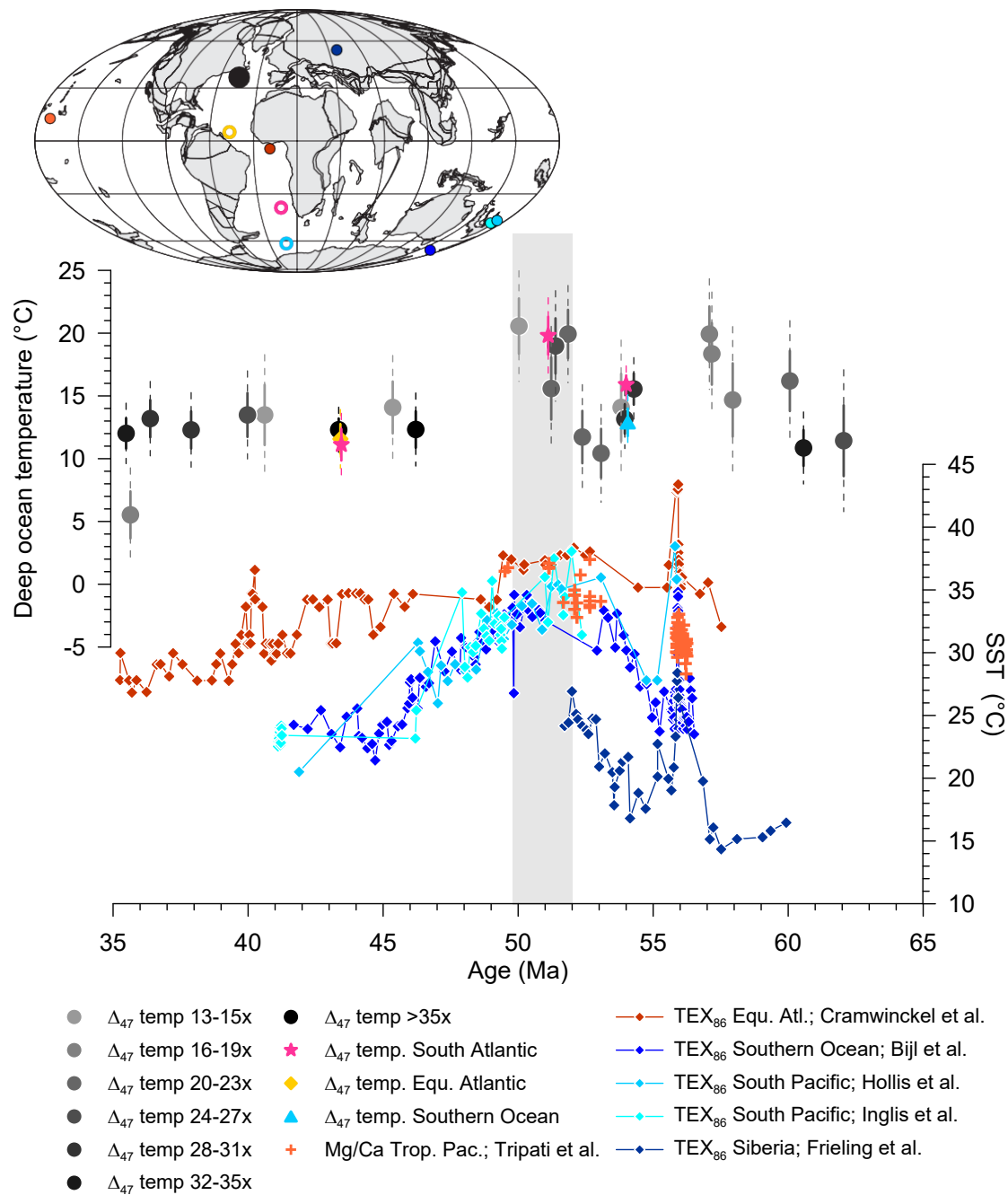


**Fig. S3. Influence of different clumped isotope calibration on reconstructed temperatures and  $\delta^{18}\text{O}_{\text{sw}}$ .** The calibration used in this study is based on a compilation of planktic and benthic foraminifera from surface sediments (46), updated to the I-CDES scale by (57). The gray symbols in the top and bottom panels represent the North Atlantic data calculated with this calibration and are the same as shown in Figs. 1A and 2A and in Fig. 2D, respectively. The results match well to the temperatures obtained when using a calibration based on travertines and tufas covering a larger temperature range (“Kele calibration” (58), recalculated by (52)) with differences of -1.0 to 0.9 °C. The differences to this calibration include the effects of slightly different standardization (CDES versus I-CDES scale). A recent calibration covering a temperature range of 1000°C and including previous calibration data including most, but not all, of the above (59), on the other hand yields temperatures slightly colder (by 1.9 °C), but the general observations remain unchanged. With this (“Anderson”) calibration,  $\delta^{18}\text{O}_{\text{sw}}$  is 0.4 ‰ heavier (blue symbols in bottom panel). Average  $\delta^{18}\text{O}_{\text{sw}}$  for >45 Ma (arrows to the right) with and without a pH correction become -1.13 ‰ and -0.37 ‰, respectively.

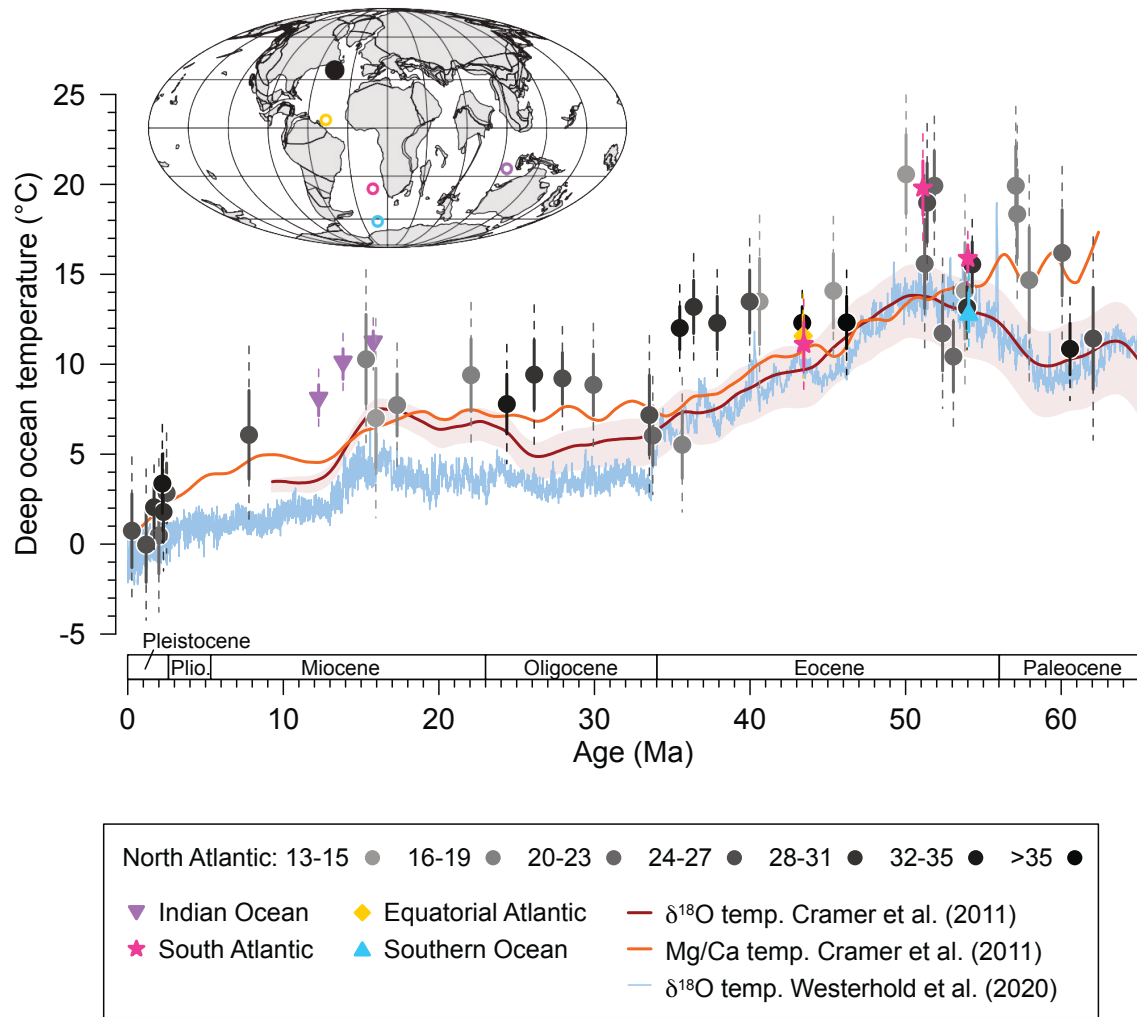




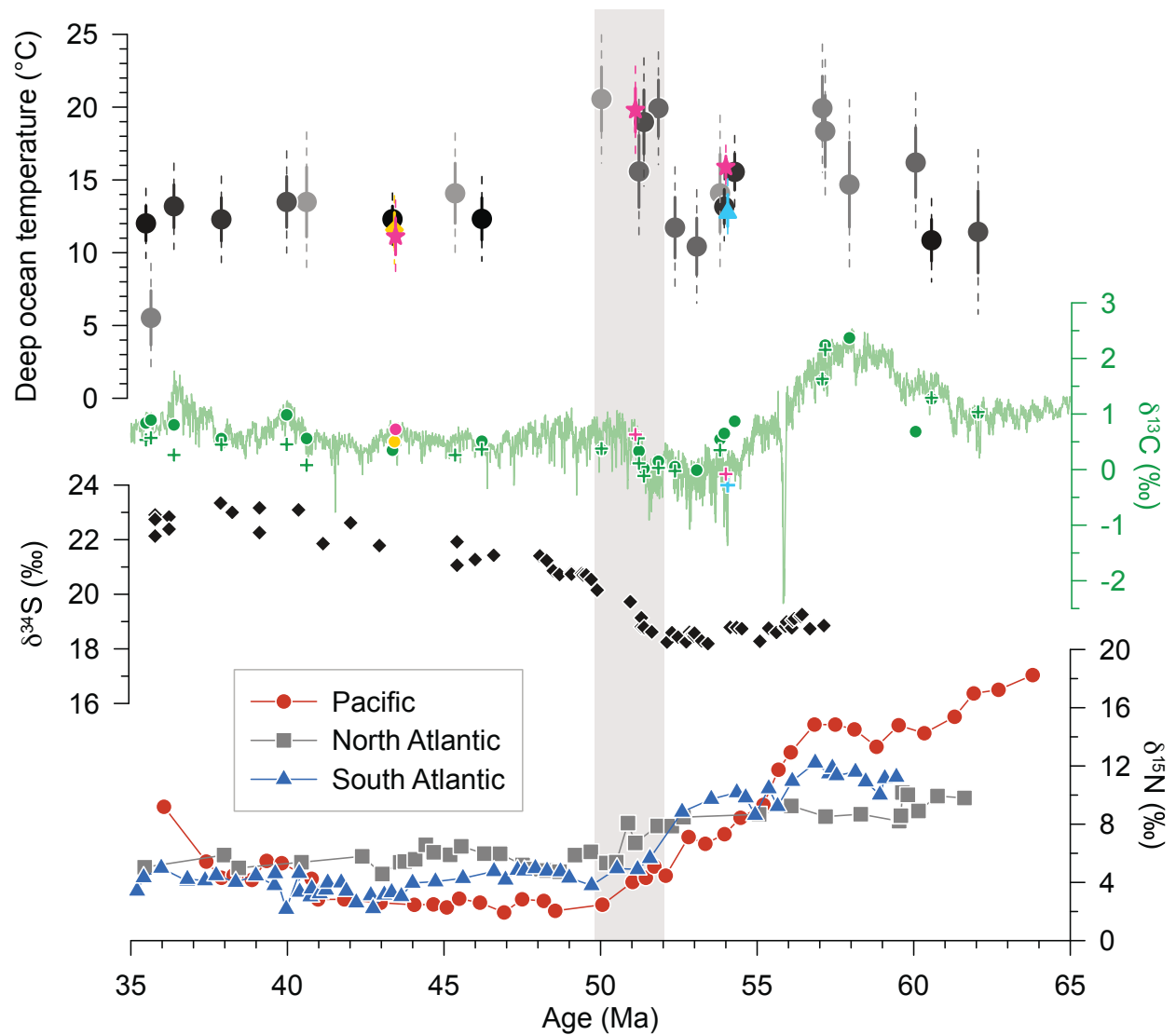
**Fig. S4. Close-up of the early and late Cenozoic intervals of Fig. 1.** As explained in the caption for Fig. 1, some of the apparent offsets in  $\delta^{18}\text{O}_b$  and  $\delta^{13}\text{C}$  between our data and CENOGRID (2) are due to adjustments for inter-site/basin offsets in the CENOGRID data set. For example, the South Atlantic data used for most of the Paleocene and early Eocene sections of the CENOGRID record have been adjusted by  $-0.2\text{‰}$  (2), but such a correction was not applied to our data for lack of constraints.



**Fig. S5. Comparison of deep ocean temperatures for the early Cenozoic with the most detailed sea surface temperature (SST) records currently available.** Most SST records that span a significant time period in the Early Cenozoic with sufficient resolution are based on organic biomarkers (TEX<sub>86</sub> proxy). The SST data shown here (70-78) were obtained from a recent compilation of early Cenozoic temperature data (8) and age models were updated to the CENOGRID timescale for direct comparison to our data. The gray bar highlights the early Eocene interval of elevated Atlantic deep ocean temperatures. The locations of the SST sites are indicated on the inset map by closed small circles.



**Fig. S6. Comparison of clumped isotope temperatures to estimates of deep ocean temperature based on  $\delta^{18}\text{O}$  and Mg/Ca.** Red line with uncertainty band: temperature estimate based on  $\delta^{18}\text{O}_b$  and reconstructed sea level (6); Orange line: Mg/Ca-based temperature record (6), in the version using Equation 7b of (6) for temperature calculation. This reconstruction is based on a global compilation of selected Mg/Ca records, but in the Paleocene to middle Eocene is based on sparse data predominantly from the Pacific Ocean. Light blue line: CENOGRID  $\delta^{18}\text{O}_b$ -based deep ocean temperature (2) calculated with the approach suggested by Hansen (79), which accounts for the varying contribution of ice sheet size by defining three different equations for different time intervals (0-3.6 Ma, 3.6-34 Ma, and prior to 34 Ma). Gray and colored symbols: Clumped isotope temperatures with uncertainties as in Figs. 1 and 2. Previous estimates are plotted on their published age models.



**Figure S7. Comparison of the deep ocean temperatures to proxy records of biogeochemical processes for the Palocene-Eocene.**  $\delta^{13}\text{C}$  data are as in Fig. 1C,  $\delta^{34}\text{S}$  data are from multiple sites (34), and  $\delta^{15}\text{N}$  data are from three different sites (35) as indicated in the legend.

**Table S1.** IODP Sites from Newfoundland margin studied, with their respective water depths below sea level at present (44) and at 50 Ma (45).

Site	Present location	Water depth (m) present	Water depth (m) 50 Ma
U1407	41°25.5'N, 49°48.8'W	3074	2600
U1410	41°19.7'N, 49°10.2'W	3387	2950
U1409	41°17.7'N, 49°14.0'W	3500	3050
U1406	40°21.0'N, 51°39.0'W	3813	3300

**Table S2.** Site information for benthic boron isotope samples with approximate paleo water depths for the respective age intervals.

Site	Present location	Age range of benthic $\delta^{11}\text{B}$ data	Paleo water depth (m)
U1409	41°17.7'N, 49°14.0'W	53-59 Ma	2500
1258	9°26.0'N, 54°44.0'W	47-52 Ma	3200
1260	9°16.0'N, 54°33.0'W	45-46 Ma	2500
926	3°43.1'N, 42°54.5'W	5-23 Ma	3600
999	12°44.6'N, 78°44.4'W	0-3 Ma	2800

**Data S1. (separate file)**

Clumped isotope data averaged per sample

**Data S2. (separate file)**

Numbers of  $\Delta_{47}$  aliquots analyzed per species/genus for each sample

**Data S3. (separate file)**

Deep ocean pH data from boron isotopes in benthic foraminifera

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