

Methods for Solving the Dyadic Model Equations

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For dmm_3.1-1

Contents

1	Introduction	1
2	The dyadic model equations	3
3	Checking the dyadic model assumptions	5
3.1	The assumption of independence of columns of \mathbf{W}	5
3.2	The assumptions of independence of dyadic residuals	6
4	An example using the <i>warcolak</i> dataset	6
5	Discussion	10

1 Introduction

The function *dmm()* sets up equations which relate the observed covariance of pairs of individuals or dyads, to their expectation in terms of postulated genetic and environmental variance and covariance components.

These equations, termed *dyadic model equations* (DME's), can be solved directly to obtain estimates of variance and covariance components. The DME's are linear equations, and are exactly analogous to a set of multi-trait multiple regression equations.

The function *dmm()* therefore effectively turns variance component estimation into a regression problem. All of the statistical techniques for fitting a linear multiple regression are therefore available for solving the DME's. The function *dmm()* uses the *qr()* function by default, but can optionally use *lm()*, robust regression (*lmrob()*), principal component regression (*pls package*), or feasible generalised least squares (*fgls*).

Assumptions about dyadic residuals have to be considered in choosing a method to solve the DME's. The simplest assumption, that dyadic residuals are uncorrelated and equal in variance, leads to ordinary least squares (OLS). In *dmm()* the QR, lm, and pls options all use ordinary least squares. The robust regression (lmrob) option uses OLS after deleting dyads considered to be outliers. The fgls option is an implementation of generalised least squares (GLS), so it takes account of the known covariance structure of the dyadic residuals (see Section).

The type of variance component estimate that is obtained from *dmm()* depends on

1. The method (OLS or GLS) used to fit the fixed effects model, from which the residual variances to be partitioned into components are obtained
2. The method used to fit the dyadic or random effects model which does variance component estimation.

Table 1: Relationship between fitting methods for fixed and dyadic models and types of variance component estimate obtained

Method for fixed model	Method for dyadic model		
	Ordinary Least Squares qr or lm or pls	Generalised Least Squares fgls	Robust Regression lmrob
ols	MINQUE	REML	robust MINQUE
gls	BCML	REML	robust ML

The relationship between fitting methods and the type of variance component estimate obtained is summarized in Table 1

The types of variance component estimate are as follows

MINQUE minimum variance quadratic unbiased estimate defined by Rao ??

BCML bias corrected maximum likelihood estimate as derived by Anderson ??

REML restricted maximum likelihood estimate defined by

RMINQUE robust MINQUE estimate

RML robust ML estimate

The *dmm()* program can obtain any of the above estimates. By default *dmm()* uses ordinary least squares (OLS) for both the fixed model and the dyadic model. So the default variance component estimate type is MINQUE. If *gls* is specified for the fixed model *dmm()* does *ols* for the fixed model first, then does the *gls* iteration, and reports both *ols* and *gls* results. If *fgls* is specified for the dyadic model, *dmm* obtains REML variance component estimates, regardless of the method used for fixed effects. It is a property of REML estimates that they are independent of the method used to remove fixed effects. If *lmrob* is specified for the dyadic model, the corresponding robust estimates are obtained.

There are some limitations with *dmm()*. Robust regression can only be used for univariate models. The *fgls* option which leads to REML estimates is severely restricted by memory requirements. Obtaining REML estimates by doing generalised least squares on the dyadic model equations is an order N^4 problem where N is the number of individuals with data. In practice the *fgls* option can only handle about 200 individuals, so it is of academic interest only. If you want REML estimates use the *gremlin()* CRAN package (Wolak(2020) ?? All the other *dmm()* options are of order N^2 and they are useful up to about 10000 individuals.

Multivariate models require extra caution in interpretation. Multivariate analysis amounts to treating all the traits as if they were one trait and analysing the total variance. Multivariate MINQUE estimates will be identical to the

univariate MINQUE estimate for each trait. Multivariate BCML estimates will usually be similar to univariate BCML estimates for each trait, with minor deviations for traits that are highly correlated. Multivariate REML estimates will be quite different from univariate REML estimates, because using GLS on the dyadic model equations takes account of the correlated error structure of the dyadic model.

MINQUE, BCML, and REML estimates will agree for simple balanced data sets. Robust regression estimates will differ. For complex pedigrees, MINQUE and BCML estimates of variance components will be estimates of the variances existing in the population of related individuals from which the data were obtained. REML estimates will be adjusted to estimate what the variances would have been if the individuals were unrelated. This is because using GLS on the dyadic model equations allows for the covariance structure of the dyadic errors. The dyadic error covariances come from two sources, the covariance structure of products, and the relationship matrix between individuals.

Because of the above, REML estimates need to be interpreted differently from MINQUE and BCML. If you want the conventional genetic parameters which are adjusted to a hypothetical population of unrelated individuals, use REML. If you want to know what the selection responses might be in the population at hand, use MINQUE or BCML.

In many of the datasets available to quantitative geneticists, the (co)variance components which we would like to estimate are partially confounded, sometimes to the point where they are not separably estimable. This is particularly so in dealing with nonadditive genetic components. The function *dmm()* offers an experimental approach (*pls()* option) which allows partially confounded components to be estimated by constraining some components, using principal component regression. This is still ordinary least squares, but it is constrained to a certain subspace of the full variance component space.

A related issue is omitted variable bias. If you leave a variance component which is large and significant out of the fitted dyadic model, its variance does not necessarily go into dyadic error, but will be spread unpredictably among the components which are fitted, resulting in biased estimates. It is therefore always important to include all significant variances in the dyadic model. There will be problems if all significant variances are not separable with the data at hand. The *pls* option may help with this, by allowing a fit of a combination of variance components, rather than fitting them individually.

2 The dyadic model equations

The dyadic model is presented in Section 6.2.2 of the document *dmmOverview.pdf*[1]. It results in a set of equations (the DME's) which are given in matrix form as equation 12, which is reproduced below

$$\Psi = W\Gamma + \Delta \tag{1}$$

It is important to understand each of the matrix components of these equations, so we elaborate as follows

First, the following variables set the size of the problem and the sizes of the above matrices

n number of individuals with data

m number of individuals in pedigree

l number of traits

k number of fixed effects

c number of variance components to be estimated

Now we explain each matrix

Ψ $n^2 \times l^2$ matrix of dyadic covariances for each pair of individuals (row) and each traitpair (col). Each covariance needs to be appropriately adjusted for fixed effects. The columns of **Ψ** become the dependent variables in a multi-trait multiple regression.

W $n^2 \times c$ matrix containing the coefficients of the dyadic model equations, which become the independent variables of a multiple regression. Each column of **W** has the form $Vec(MZ_cR_cZ'_cM')$ where Vec is an operator that vectorizes a matrix, **M** is a matrix from the fixed effect model such that $Y - X\hat{\alpha} = MY$, **Z_c** is an incidence matrix relating individuals with data to individuals in the pedigree, and **R_c** is a relationship matrix relevant to component c . Note that relationship matrices are used, not their inverse.

Γ $c \times l^2$ matrix of (co)variance component parameters to be estimated, which become the partial regression coefficients of a multiple regression.

Δ $n^2 \times l^2$ matrix of dyadic model residuals. Note the variance of these is not the individual environmental variance component - that has to be explicitly fitted as one of the columns of matrix **W** and appears as one row of matrix **Γ** . The **Δ** matrix elements are the extent to which each dyadic covariance in matrix **Ψ** deviates from its expectation. The (co)variances of the elements of **Δ** enter into the standard errors of variance component estimates, in the same way that the (co)variances of residuals enter into the standard errors of any regression coefficient estimates.

Matrices **Ψ** and **W** have n^2 rows, so we are looking at solving n^2 equations in c unknowns. This is an overdetermined system. We can use least squares to obtain an approximate solution.

3 Checking the dyadic model assumptions

Before attempting a regression fit of model (1), it is worth looking at how well the data conform to the assumptions made in using least squares to fit a multiple regression model. The critical assumptions are

- the independent variables (columns of \mathbf{W}) are uncorrelated
- the residuals (columns of $\mathbf{\Delta}$) are uncorrelated with each other and with the independent variates

A least squares fit does not involve assumptions regarding the distribution of residuals, but this does become involved when using residuals to obtain standard errors of parameter estimates.

3.1 The assumption of independence of columns of \mathbf{W}

The correlations among independent variables (columns of \mathbf{W}) are returned by *dmm()* in the attribute *dme.correl* of the returned object. These are pairwise correlations between the columns of the \mathbf{W} matrix. They are not quite the same thing as correlations between relationship matrix elements, because the columns of \mathbf{W} also involve \mathbf{M} and \mathbf{Z} matrices.

The \mathbf{Z} matrix simply selects a subset of the relationship matrix corresponding to those individuals which have observations. The \mathbf{M} matrix comes from the fixed effects model, and consists of a set of weights which adjust for the degrees of freedom involved in each of the fixed effects applicable to each individual. So the columns of the \mathbf{W} matrix are a weighted subset of the elements of the appropriate relationship matrix. Their correlations are therefore not the same as relationship matrix element correlations.

In the case of a dataset with mean only, and all individuals in the pedigree with data, such as the *warcolak* dataset, the subset is all of the relationship matrix and the weights in \mathbf{M} are all equal, so the correlations of the columns of \mathbf{W} are exactly the same as the correlations of the elements of the relationship matrices, in this special case.

In regression analysis it is generally considered that if there are collinearities among the independent variates amounting to correlations greater than around 0.5 then the estimates of regression coefficients are suspect. Translating this to our dyadic model, the variance component estimates are not likely to achieve a realistic separation if the columns of \mathbf{W} have correlations exceeding 0.5. The option of using principal component regression (*dmeopt="pcr"*) has been developed as an experimental approach to dealing with serious collinearities among the components.

If we want to plot these correlations (eg as scatterplots) we need argument *dmekeep=T* in calling *dmm()*. The default for *dmm()* is not to save the DME's in its return object. This can be overridden with argument *dmekeep=T*. This results in 2 attributes *dme.psi* and *dme.wmat* being added to the returned object. Caution, this can result in a very large returned object. The attribute *dme.wmat*

is the W matrix, and its columns can be plotted with the standard `plot()` routine.

3.2 The assumptions of independence of dyadic residuals

To check the dyadic residuals, we can use the S3 `plot()` method included in the *dmm* package. This will output histograms, qqnorm plots, and scatterplots of residuals against fitted and observed values. Plots tend to be more informative for datasets with smaller numbers of individuals.

Dyadic residuals are usually not far from normally distributed, but may be leptokurtic and slightly skewed to the right.

If dyadic residuals are correlated with fitted values of the components, then there is something wrong with the model, probably some extra component should be fitted.

One can expect dyadic residuals to be correlated with observed dyadic covariances. It is normal for the fitted components in a dyadic model to explain only a small fraction of the total variation, and this of course leads to the observed covariances being highly correlated with residuals.

The question of patterns of correlation among the dyadic residuals themselves is a seriously difficult area. The covariances (or correlations) among the dyadic residuals for one trait form an $n^2 \times n^2$ matrix - ie n^4 elements. Too many to compute and cant be stored in R. This issue is discussed in the document *dmmOverview.pdf* [1] in relation to why *dmm()* is not able to do REML estimates. REML estimates require that the covariance (or correlation) matrix of the dyadic residuals be constructed and used to compute a GLS rather than an OLS solution to the DME's. That is not computationally feasible, and neither is an examination of the residual covariance/correlation matrix.

The covariance structure of dyadic residuals is actually known. It is derived in Searle et al (1992) [2] on pages 407-413. It involves fourth moments of the observations.

4 An example using the *warcolak* dataset

Using the *warcolak* dataset from the *nadiv* package (Wolak(2014) [3], and just analysing Trait2, we first setup the data file making all the appropriate relationship matrices, then run the usual analysis fitting 4 variance components, using the default "qr" option for solving the DME's, and setting options to save the DME's and the fit object.

```
> library(dmm)
> data(warcolak)
> warcolak.df <- warcolak.convert(warcolak)
> warcolak.mdf.univ <- mdf(warcolak.df,pedcols=c(1:3),factorcols=4,ycols=c(5:6),
                           sexcode=c(0,1),relmat=c("E","A","D","S"),keep=T)
.....
```

```
> warcolak.fit.t2 <- dmm(warcolak.mdf.univ, Trait2 ~ 1,
  components=c("VarE(I)", "VarG(Ia)", "VarG(Id)", "VarGs(Ia)"),
  relmat = "withdf", dmekeep=T, dmekeepfit=T)
.....
> summary(warcolak.fit.t2)
.....
Components partitioned by DME from residual var/covariance after OLS-b fit:
```

	Traitpair	Estimate	StdErr	CI95lo	CI95hi
VarE(I)	Trait2:Trait2	0.270	0.0369	0.198	0.342
VarG(Ia)	Trait2:Trait2	0.313	0.0134	0.287	0.339
VarG(Id)	Trait2:Trait2	0.327	0.0380	0.252	0.401
VarGs(Ia)	Trait2:Trait2	0.146	0.0130	0.121	0.172
VarP(I)	Trait2:Trait2	1.056	0.0142	1.028	1.084

These results are as for Trait 2 in the bivariate analysis reported in `dm-Overview.pdf` [1]. The only difference is that we have samed some additional information.

The correlations among columns of \mathbf{W} are, of course, also the same

```
> warcolak.fit.t2$dme.correl
      VarE(I)  VarG(Ia)  VarG(Id)  VarGs(Ia)
VarE(I)  1.0000000  0.4856317  0.9190589  0.4081477
VarG(Ia)  0.4856317  1.0000000  0.6255694  0.8266557
VarG(Id)  0.9190589  0.6255694  1.0000000  0.5254541
VarGs(Ia) 0.4081477  0.8266557  0.5254541  1.0000000
>
```

Two of these correlations are somewhat larger than the nominal 0.5 mentioned above. We need to look and see if the multiple regression has been able to separate these 4 components properly. To that end we will redo the analysis using `dmeopt="pcr"` instead of the default `"qr"`. Using the principal component regression option requires at least two runs - the first retaining all 4 principal components, and then one or more reruns omitting the least important principal components. The first run is as follows

```
> warcolak.fitpcr1 <- dmm(warcolak.mdf.univ, Trait2 ~ 1,
  components=c("VarE(I)", "VarG(Ia)", "VarG(Id)", "VarGs(Ia)"),
  relmat = "withdf", dmekeep=T, dmekeepfit=T, dmeopt="pcr")
.....
DME substep:
PCR option on dyadic model equations:
Data: X dimension: 29160000 4
Y dimension: 29160000 1
Fit method: svdpc
Number of components considered: 4
```



```

VALIDATION: RMSEP
Cross-validated using 10 random segments.
      (Intercept) 1 comps 2 comps 3 comps 4 comps
CV           1.023   1.022   1.022   1.022   1.022
adjCV        1.023   1.022   1.022   1.022   1.022

```

```

TRAINING: % variance explained
      1 comps 2 comps 3 comps 4 comps
X      79.46086 92.89539 99.28012 100.00000
evec   0.02946 0.03157 0.03158 0.03158

```

DME substep completed:

OLS-b step completed:

```

>
> loadings(warcolak.fitpcr1$dme.fit)

```

Loadings:

```

      Comp 1 Comp 2 Comp 3 Comp 4
'VarE(I)'   0.209 0.663 0.192 -0.692
'VarG(Ia)'   0.699      -0.705
'VarG(Id)'   0.275 0.630 0.111 0.718
'VarGs(Ia)'  0.626 -0.393 0.674

```

```

      Comp 1 Comp 2 Comp 3 Comp 4
SS loadings      1.00 1.00 1.00 1.00
Proportion Var   0.25 0.25 0.25 0.25
Cumulative Var   0.25 0.50 0.75 1.00
>

```

What we need from this at the moment is the % of variance explained by various numbers of principal components. Obviously using all 4 components explains 100% of the variation, but what is interesting is that 3 components explain 99% and 2 components 92%. This is a signal that we should try regressing the observations on 3 principal components of the columns of \mathbf{W} , and see how it affects the estimates and their standard errors. We do that with a rerun

```

> warcolak.fitpcr2 <- dmm(warcolak.mdf.univ, Trait2 ~ 1,
      components=c("VarE(I)", "VarG(Ia)", "VarG(Id)", "VarGs(Ia)"),
      relmat = "withdf", dmeopt="pcr", ncomp=3)
.....

```

```

> summary(warcolak.fitpcr2)
Call:
summary.dmm(object = warcolak.fitpcr2)

```

Coefficients fitted by OLS for fixed effects:

```

      Trait Estimate StdErr CI95lo CI95hi
1 Trait2   -0.063 0.0138  -0.09 -0.036

```

Components partitioned by DME from residual var/covariance after OLS-b fit:

```

      Traitpair Estimate StdErr CI95lo CI95hi
VarE(I)   Trait2:Trait2    0.286 0.0127  0.262  0.311
VarG(Ia)  Trait2:Trait2    0.315 0.0182  0.279  0.351
VarG(Id)  Trait2:Trait2    0.310 0.0118  0.286  0.333
VarGs(Ia) Trait2:Trait2    0.146 0.0205  0.106  0.186
VarP(I)   Trait2:Trait2    1.057 0.0244  1.010  1.105

```

>

So comparing these estimates with those from the "qr" fit (which are the same as obtained with "pcr" with all 4 principal components), we find that omitting one principal component has not changed the estimated components substantially, and has reduced the standard errors of the two constrained components. By omitting the fourth principal component we have in effect set it to zero, which amounts to constraining the components estimates to be on the plane defined by

$$-0.692 \times VarE(I) + 0.718 \times VarG(Id) = 0 \quad (2)$$

We get this equation from the *loadings* which were given at the end of the run with all four principal components above. If we substitute the estimates of VarE(I) and VarG(Id) from the 3 component run into the above equation we find that they do indeed fall on the constraint plane. So we still get estimates of all 4 components, but two of them are constrained to be in a ratio 0.692/0.718, or approximately equal.

To my mind, that is a more satisfactory analysis than the unconstrained result from a "qr" fit. By regressing on the principal components of columns of \mathbf{W} instead of on the columns themselves we have avoided violating the assumption of independence, and by applying one constraint we have improved the standard errors.

One can go on and try omitting two principal components. This leads to two constraint equations, so the variance components would be constrained to lie on the intersection of two planes. We shall not do it here. We have done enough to demonstrate the method.

We can view the dyadic residuals using the S3 *plot()* method included in the *dmm* package. We do this for the "qr" fit as follows

```

> postscript(onefile=F, horizontal=F)
> plot(warcolak.fit.t2)
.....
> dev.off()

```

Five plots will be saved on five separate files. It is probably better to use *png()* rather than *postscript()*. The resulting plots are shown in Figure 1 to Figure 5. We can see that the histogram is more peaked than a normal distribution (Figure 1), that the qqplot is sigmoidal (Figure 2), that the residuals are not associated with fitted values (Figure 3), that the observed values are strongly correlated with the residuals (Figure 4), and that the fitted values and observed values are not strongly correlated (Figure 5). There is some discussion of these aspects in the next section.

5 Discussion

There is nothing special about the dyadic model used by *dmm()*. Quantitative genetics has always been about covariances between relatives and measures of relationship, that is about pairs of individuals. It is just not usually called a dyadic model, but that is what it is. The term is common in the social sciences where interactions between pairs of individuals are analysed with a dyadic model.

What is important is what the dyadic model allows us to do, not the terminology. By turning variance component estimation into a regression problem, a dyadic model opens the door to using the wide range of established regression techniques for variance component estimation. That includes techniques for dealing with collinearities among the independent variables, and these could be quite useful in quantitative genetic applications where the variance components which we wish to estimate are often partially confounded, as in the example above. There is a full presentation on the use of principal components regression in *dmmOverview.pdf* [1] Section 7.4. There are some issues, the interface to the "pcr" option via the *pls* package is clumsy and its use is seriously memory intensive. Some further work is indicated.

The dyadic residuals (Δ) are usually large and highly correlated with the observed dyadic covariances (Ψ), as in Figure 4. This is because the covariance for each dyad is obtained from only one replicate pair of observations. The R^2 for a dyadic model is tiny - only 3 percent of the variance in the case of the *warcolak* example above. This highlights the central problem of quantitative genetic analysis - it is trying to extract information from a system with a signal to noise ratio of 0.03. Modelling variances is much more demanding than modelling observations. It does not matter whether you do it by maximizing a likelihood, fitting a regression, or doing an AOV. The problem is intrinsic - *dmm()* just makes it obvious by attacking the problem directly.

References

- [1] Jackson, N. (2015) An Overview of the R package *dmm*. From <http://cran.r-project.org/package=dmm> Or <https://github.com/cran/dmm>

- [2] Searle, S.R., Casella, G., and McCulloch, C.E. (1992) *Variance Components*. John Wiley and Sons, New York.
- [3] Wolak, M.E. (2014) nadiv: an R package to create relatedness matrices for estimating non-additive genetic variances in animal models. *Methods in Ecology and Evolution* 3:792-796.