

New Phytologist Supporting Information Figs S1-S18, Methods S1 and Tables S1-S7

Article title: The complete functional characterization of the terpene synthase family in tomato

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The following Supporting Information is available for this article:

Fig. S1 Annotated sequences of *TPS51* and *TPS52*.

Fig. S2 SDS-PAGE analysis of the purified recombinant His-tagged TPS proteins.

Fig. S3 Mass spectra of identified terpenes from Figs 3-5 and 10.

Fig. S4 GC-MS analysis of elemol and (+)-hedycaryol.

Fig. S5 GC-MS analysis of the products formed in planta by transiently co-expressing TPS with CPT genes in *Nicotiana benthamiana* leaves.

Fig. S6 Analysis of tomato terpene synthases with isoprene synthases from other plants.

Fig. S7 Sequence alignment of the proteins encoded by the functional tomato TPS-c genes and CPS genes from other plants.

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Fig. S14 Sequence alignment of the proteins encoded by the functional tomato TPS-e/f

genes.

Fig. S15 Sequence analysis of TPT homologs from tomato and other plants.

Fig. S16 GC-MS analysis of terpenes extracted from tomato leaves.

Fig. S17 Correlation analysis of terpene biosynthesis genes and volatile terpenes from tomato.

Fig. S18 Terpene biosynthesis gene clusters identified from tomato genome by plantiSMASH.

Table S1 Primers used in this study.

Table S2 A list of terpene compounds identified from Figs 3-5 and 10.

Table S3 A list of tomato terpene synthase genes, their characteristics, and the enzymes they encode.

Table S4 A list of tomato terpene synthase pseudogenes.

Table S5 A list of the *trans*-prenyltransferase genes in tomato.

Table S6 A list of terpene compounds produced by all the functional tomato terpene synthases.

Table S7 A list of *Arabidopsis* terpene synthase genes, their characteristics, and the enzymes they encode.

Methods S1 Details on experimental procedures.

Fig. S1 Annotated sequences of *TPS51* and *TPS52*. Exons are shaded in yellow, gaps in the reference genome (https://solgenomics.net/organism/Solanum_lycopersicum/genome; the 2017 release of version SL3.0) are underlined and filled by sequencing the cloned genomic DNA. Stop codons are shown in red. The obtained genomic sequences of *TPS51* and *TPS52* have been deposited to GenBank under accession numbers MN366354 and MN366355, respectively. The genomic sequences of other genes studied in this work can be found in the tomato genome database under loci listed in Tables S3-S5.

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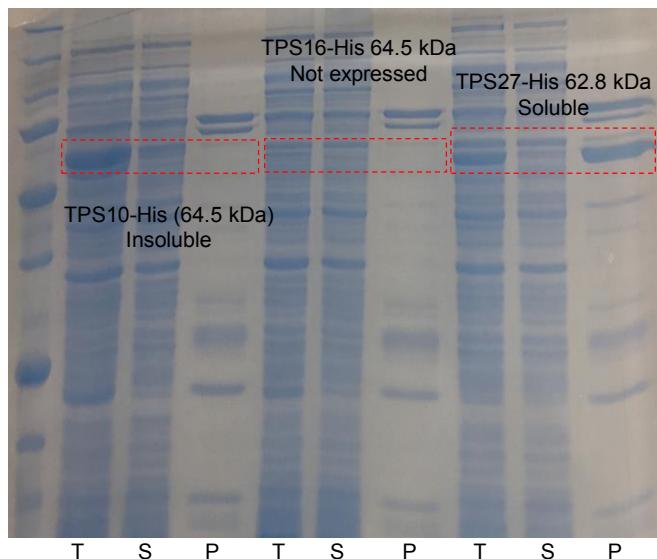
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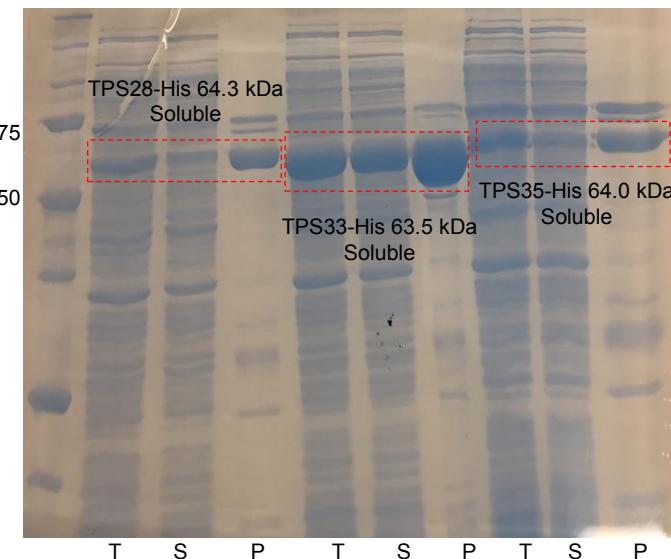
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Fig. S2 SDS-PAGE analysis of the purified recombinant His-tagged TPS proteins. T, total protein after IPTG induction; S, soluble fraction; P, purified fraction with Ni-NTA agarose as described in Methods S1.

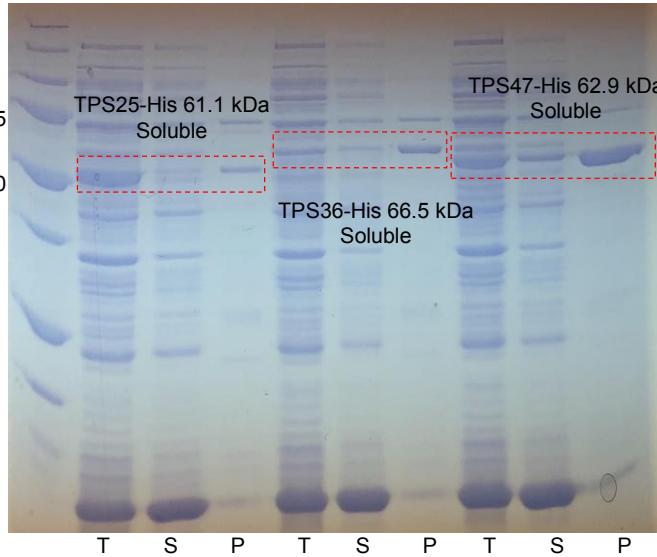
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Marker (kDa)



Marker (kDa)



Marker (kDa)

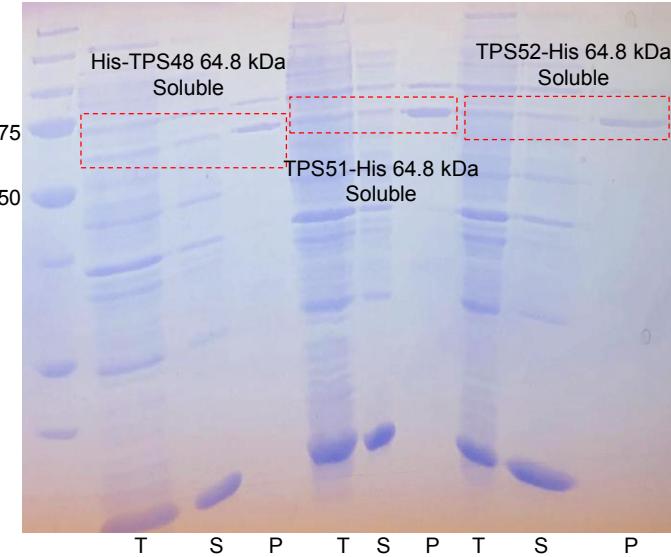
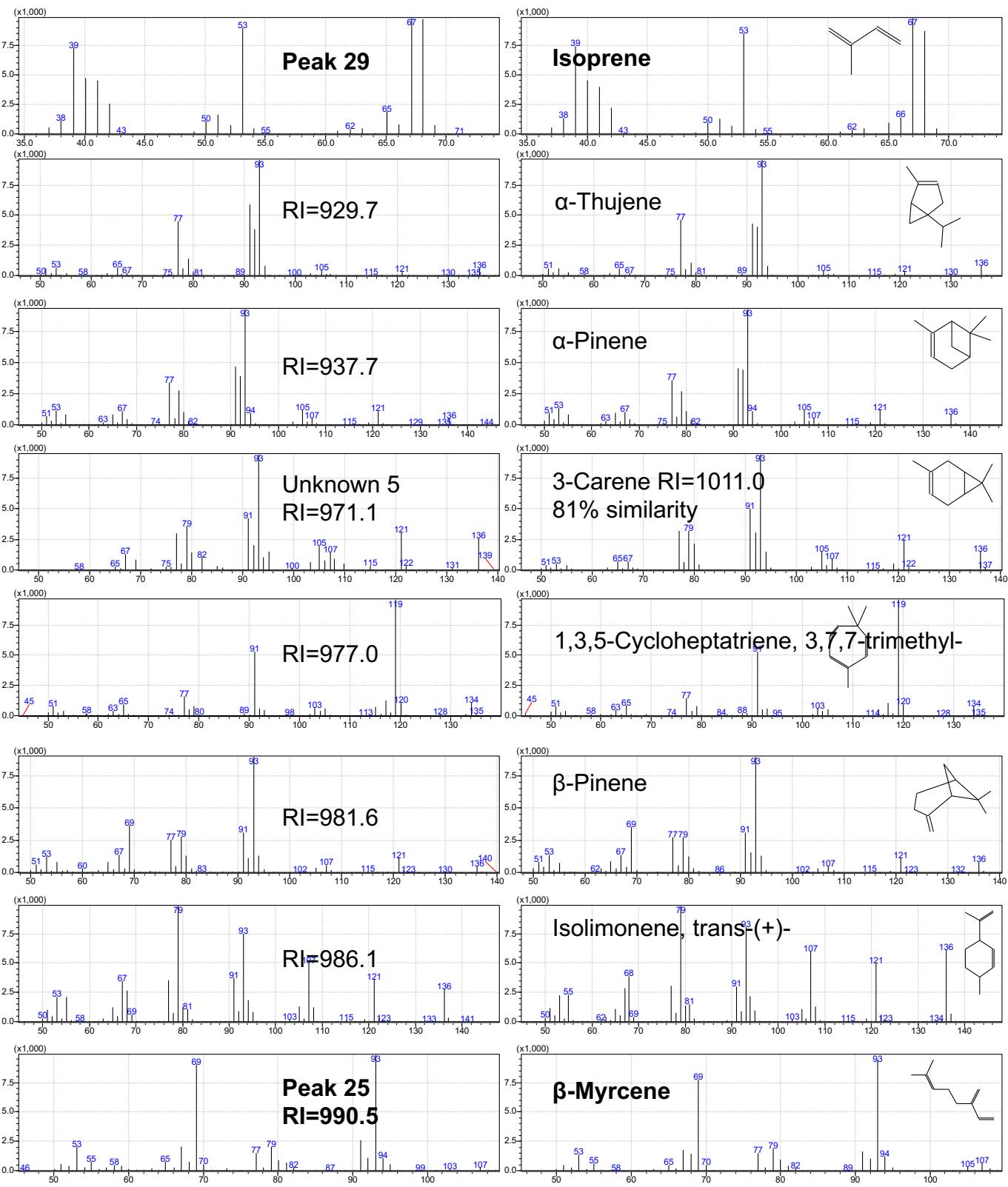
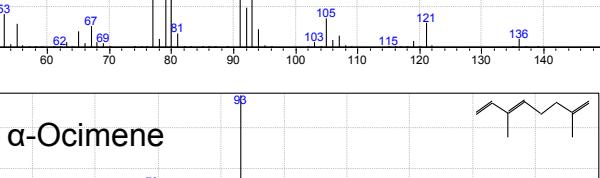
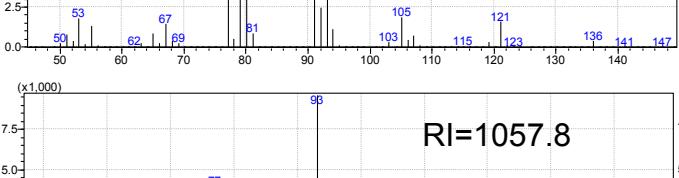
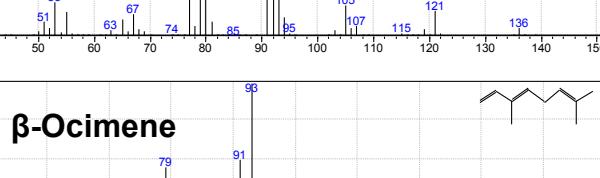
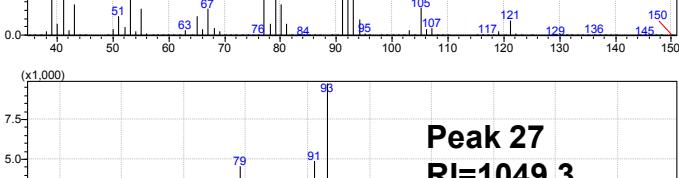
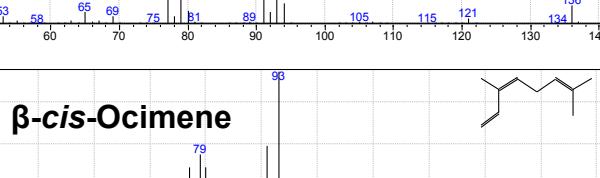
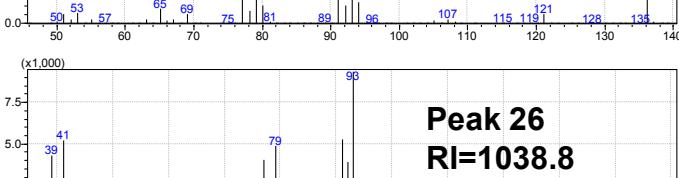
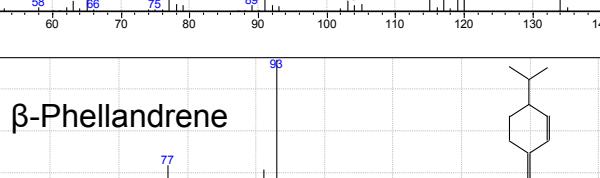
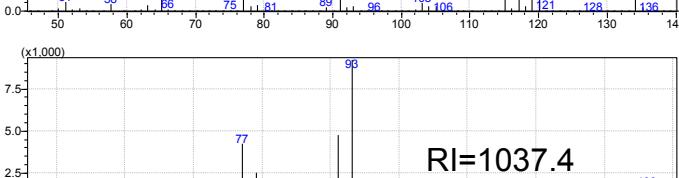
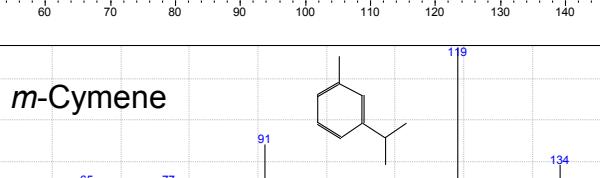
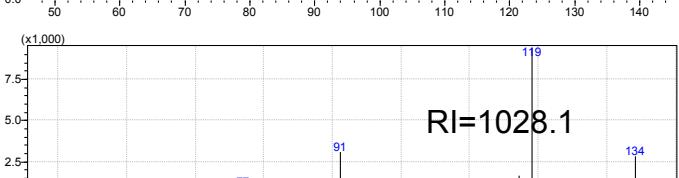
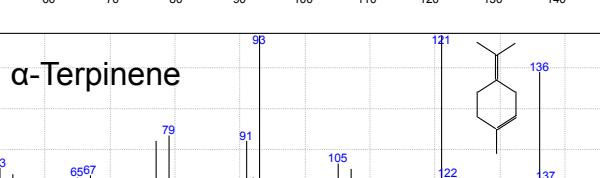
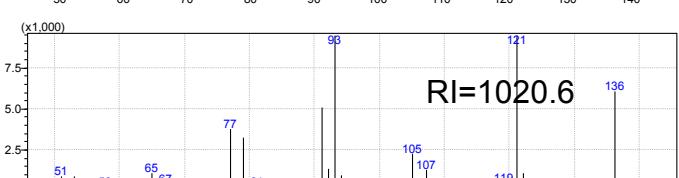
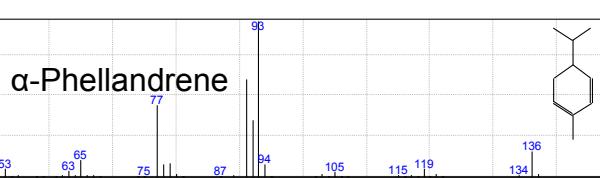
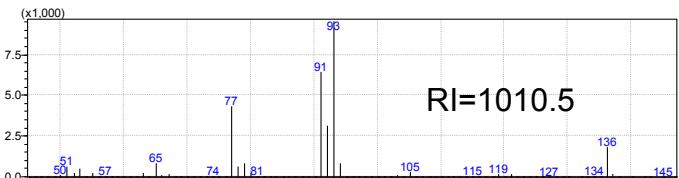
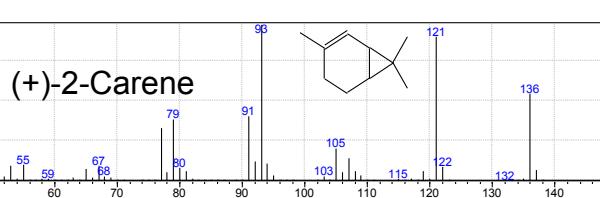
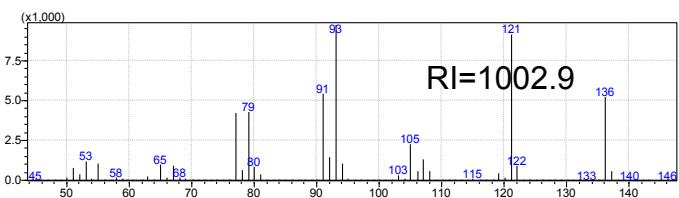
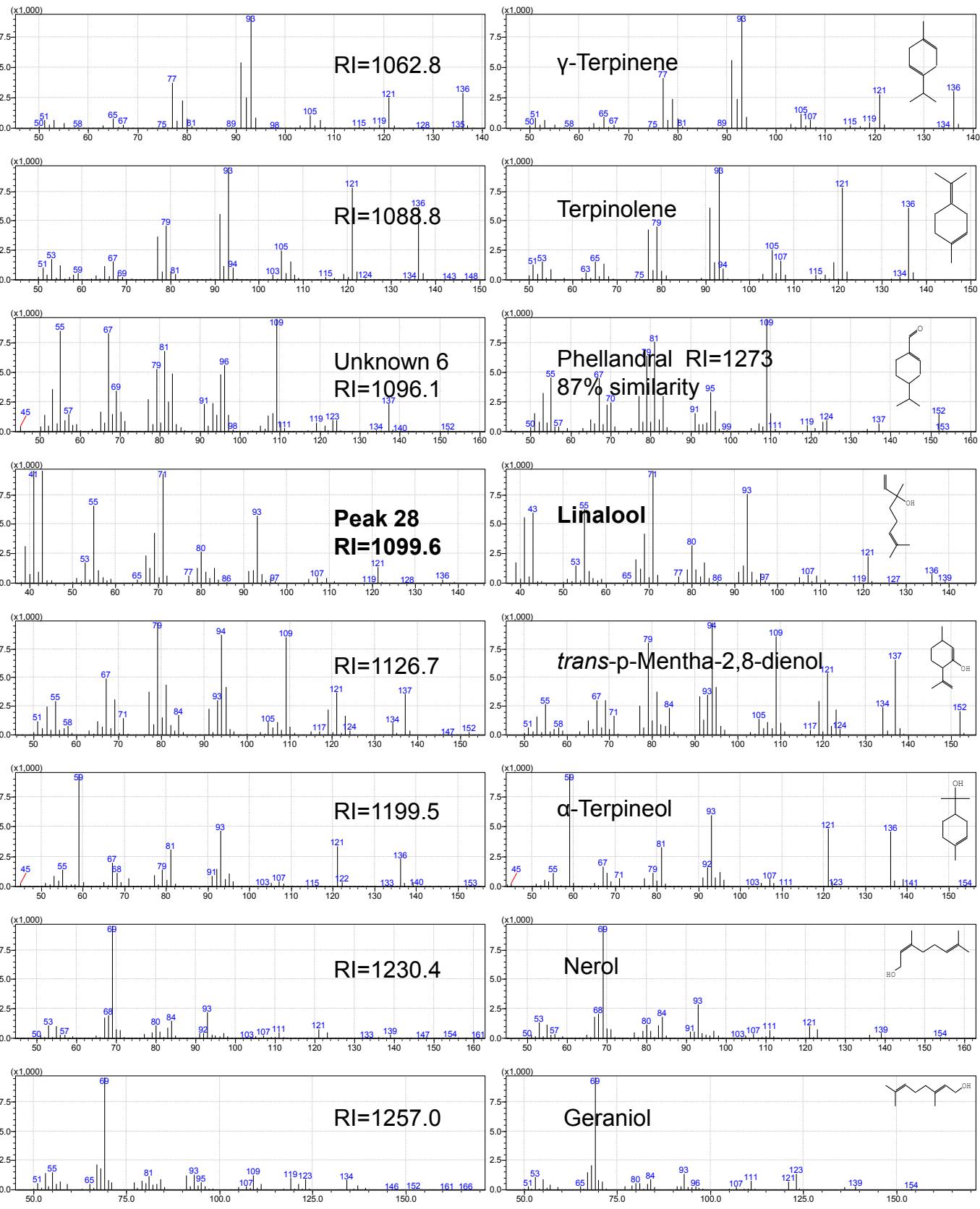
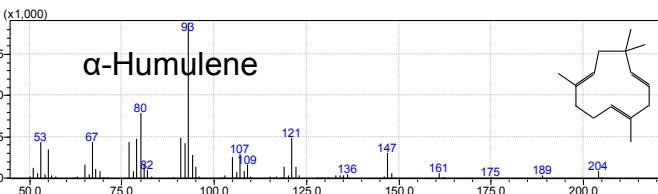
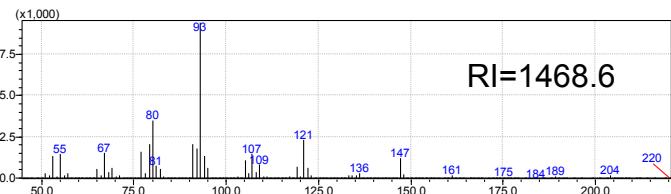
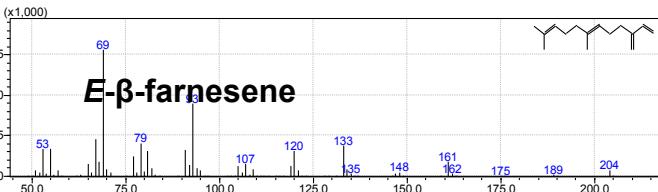
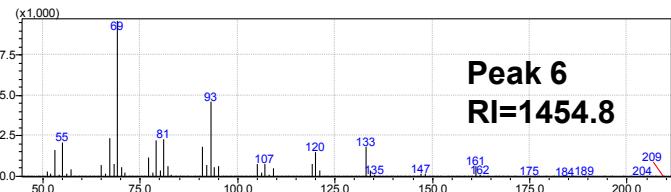
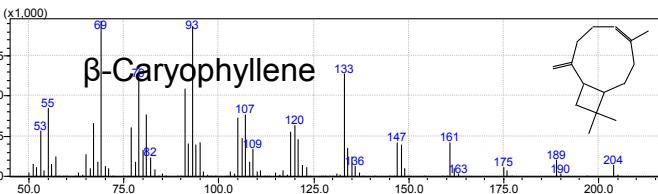
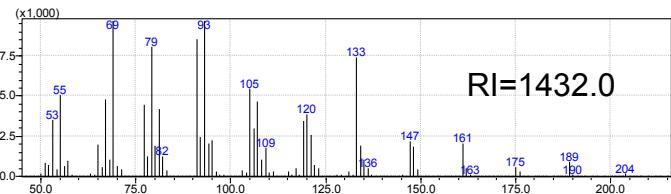
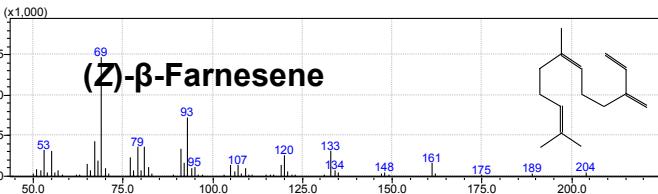
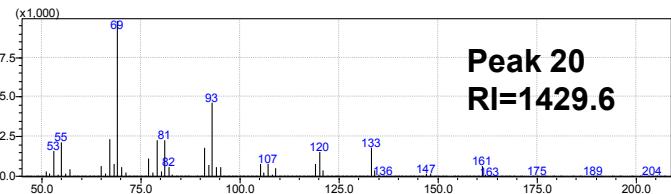
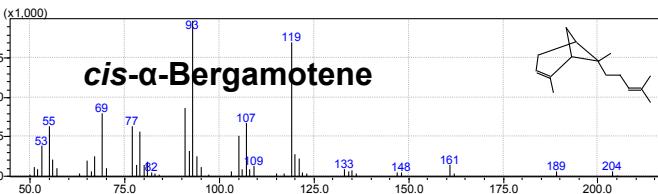
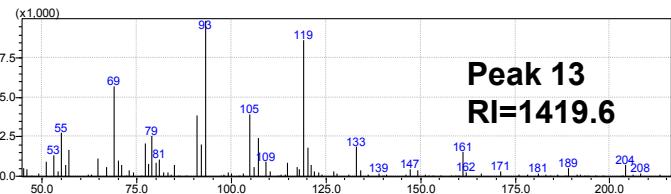
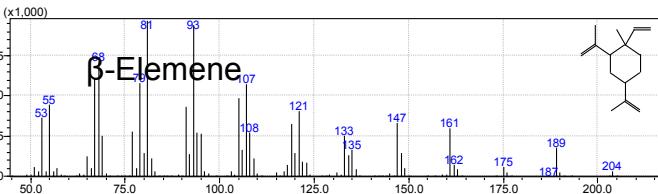
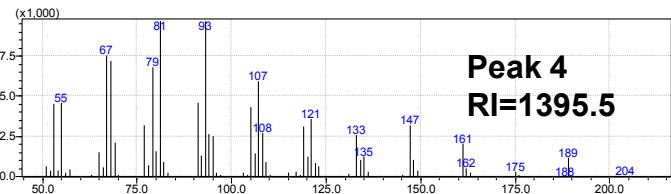
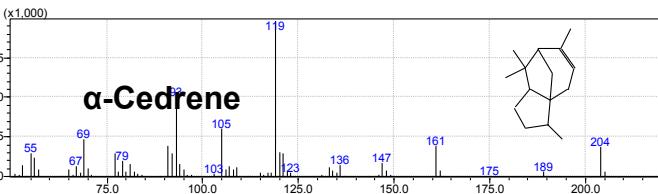
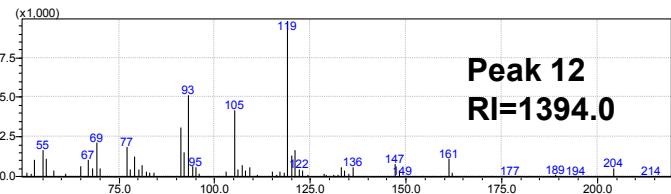
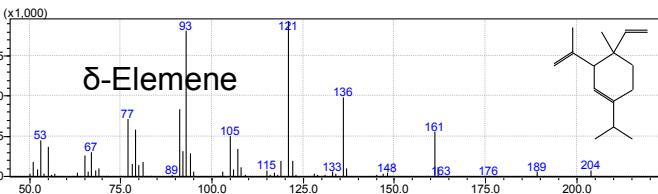
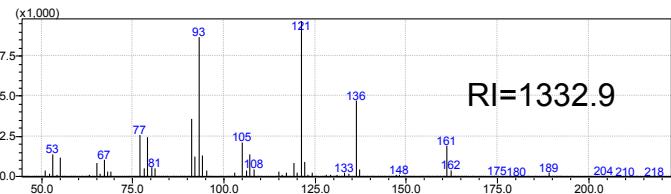


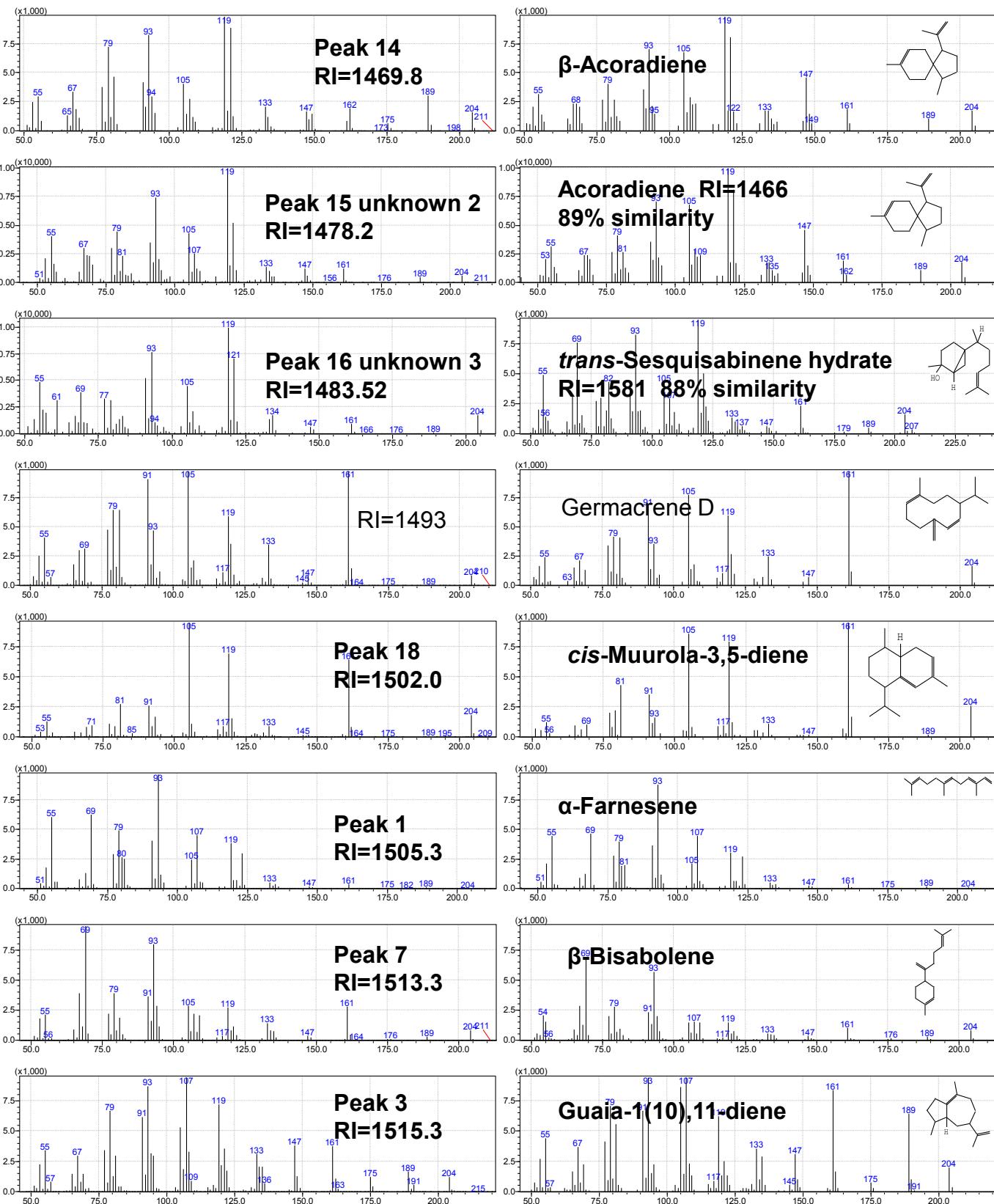
Fig. S3 Mass spectra of identified terpenes from Figs 3-5 and 10. The mass spectra of identified terpenes from NIST library are shown on the right. Compounds listed in order of elution from the Rx-5Sil MS column, the enzymatic products are shown in bold. Detailed information can be found in Table S2.

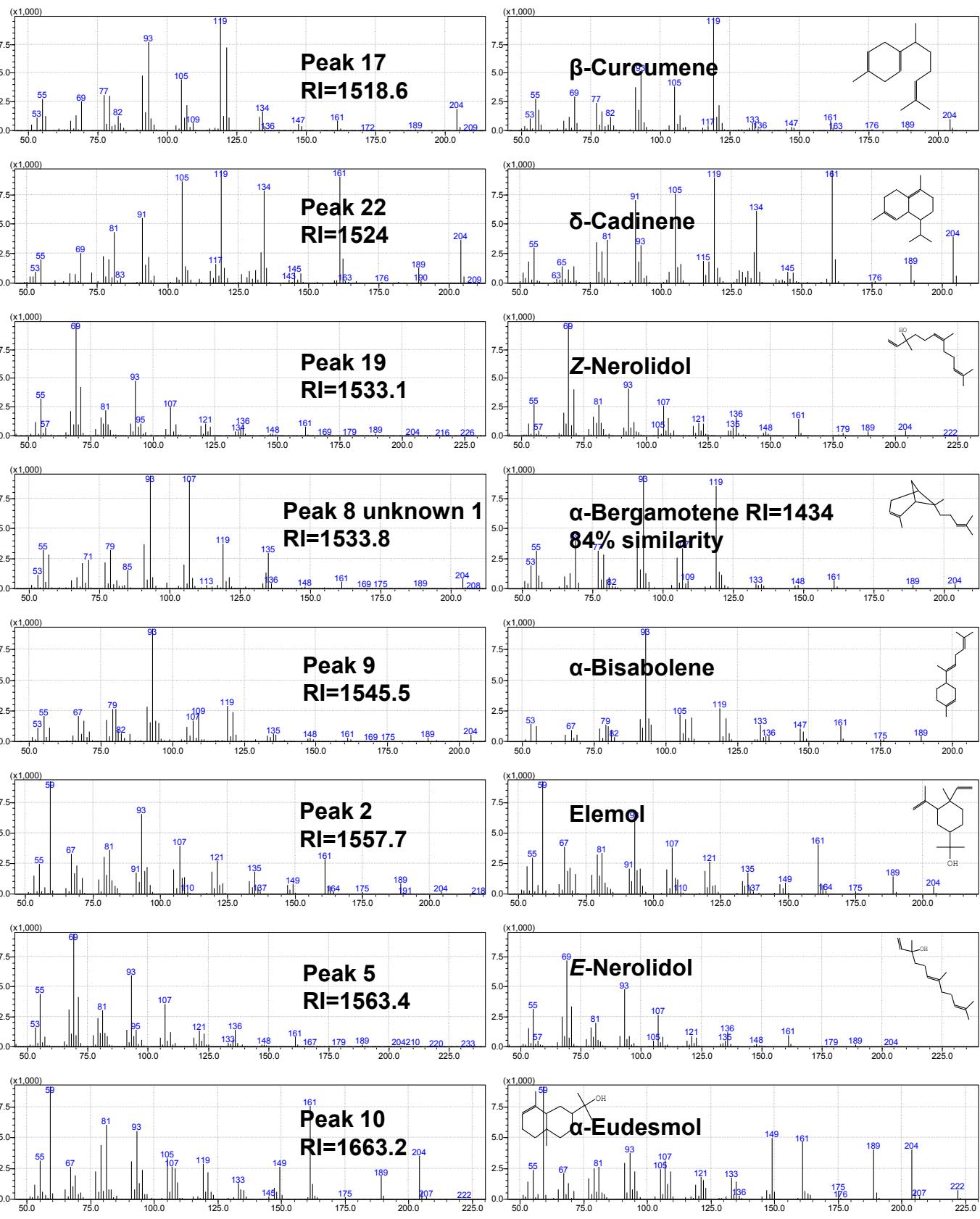












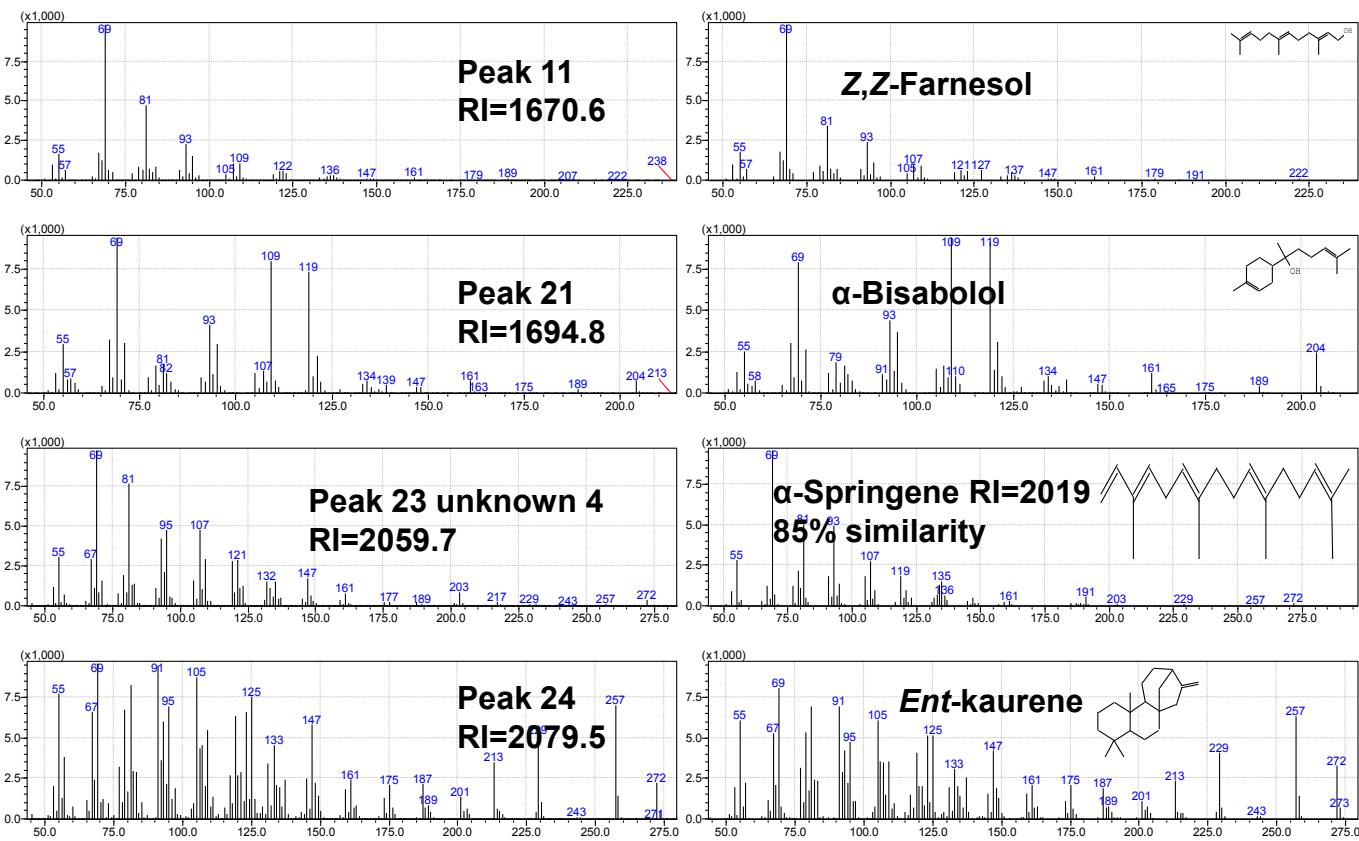


Fig. S4 GC-MS analysis of elemol and (+)-hedycaryol. Elemol is the thermal breakdown product of (+)-hedycaryol, which can be detected by lowering the injection port temperature.

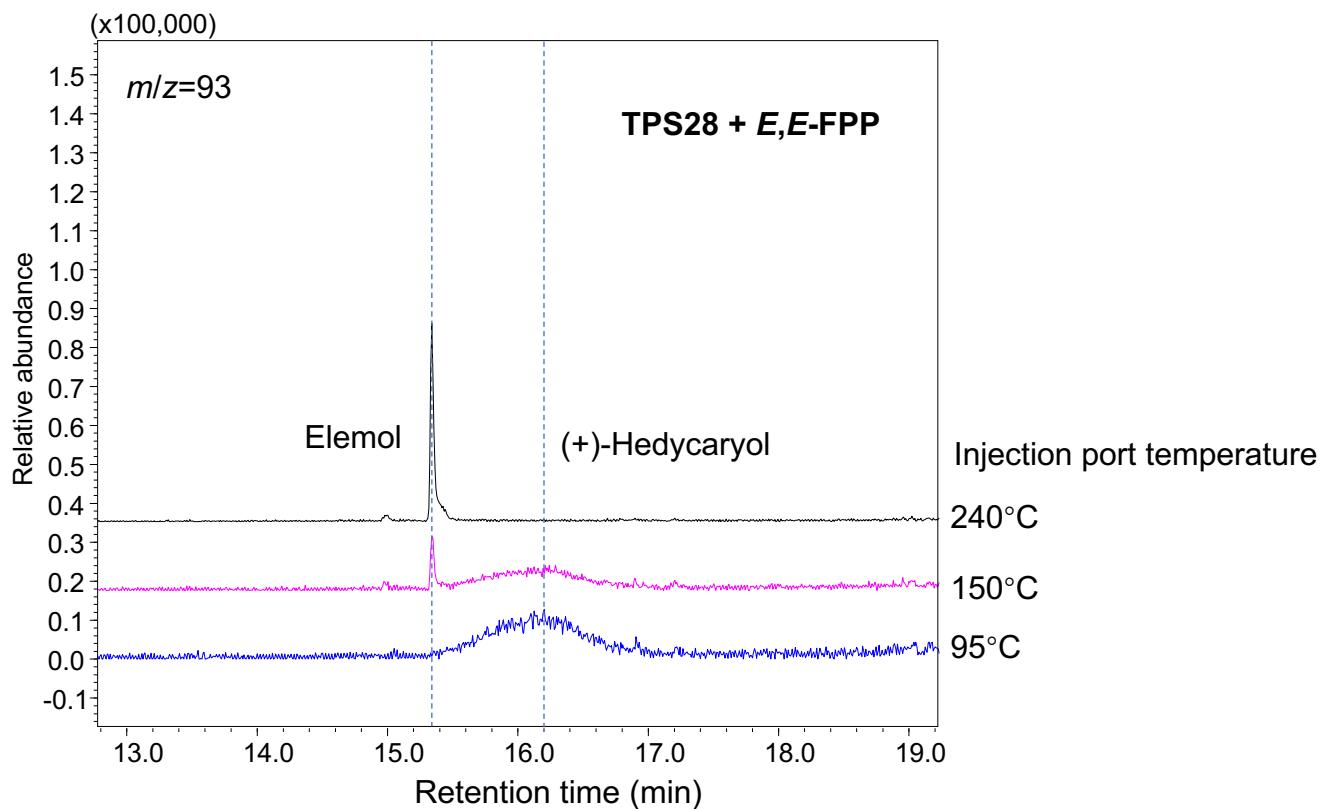


Fig. S5 GC-MS analysis of the products formed in planta by transiently co-expressing TPS with CPT genes in *Nicotiana benthamiana* leaves.

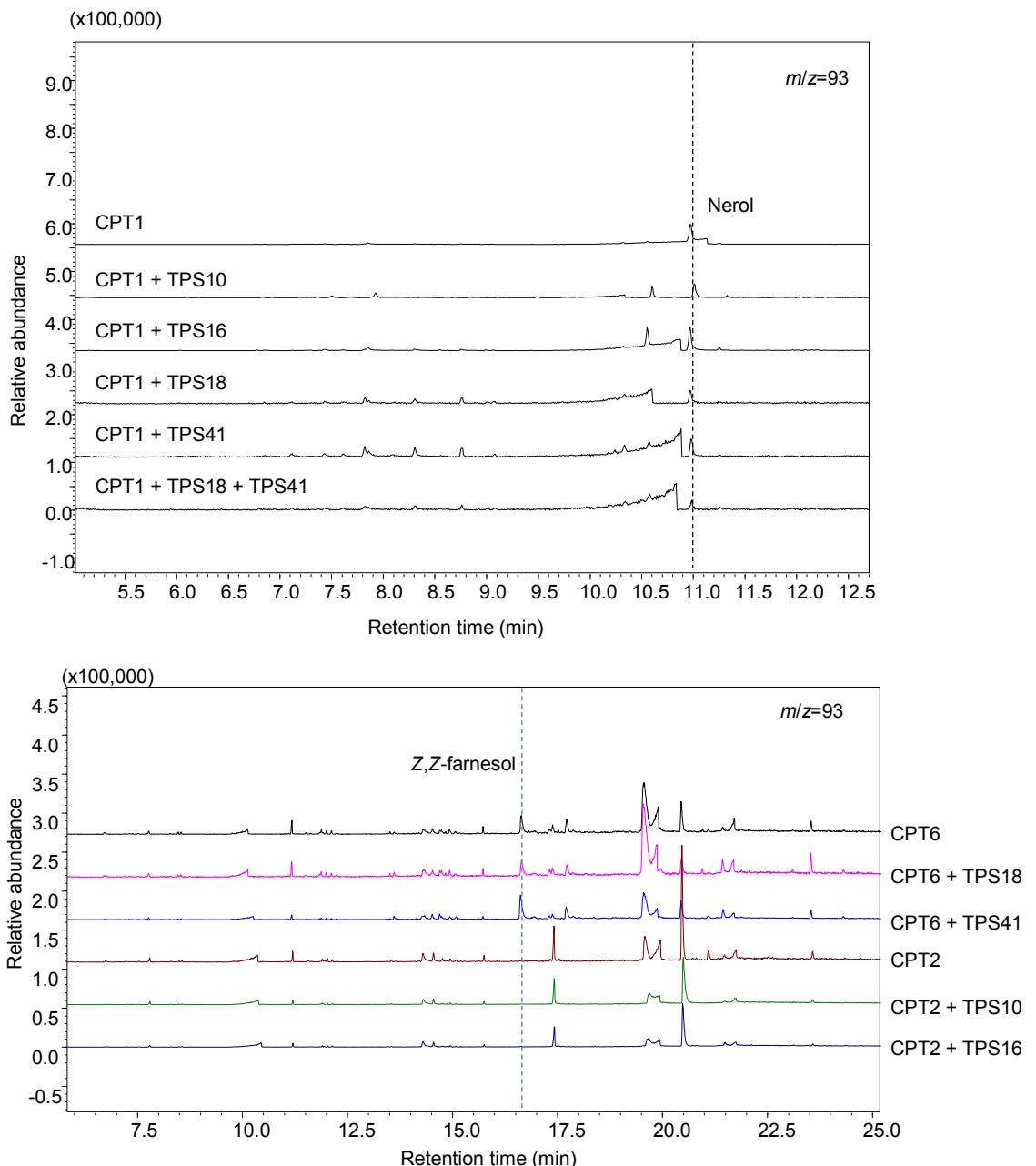


Fig. S6 Analysis of tomato terpene synthases with isoprene synthases from other plants. (a)

Phylogenetic analysis of 34 tomato TPSs with three functionally characterized isoprene synthases. The three TPSs clustered with known isoprene synthase are shown in bold and shaded in green. (b) Multiple sequence alignment tomato TPSs from TPS-a, TPS-b and TPS-g clades with isoprene synthases. Black shading indicates identical amino acid residues and grey shading are the conserved or semi-conserved substitutions observed and dashes indicate gaps. Amino acids belonging to the ‘isoprene score’ amino acids are shown in red, the calculated “isoprene score” is shown next to each TPS. Among the calculated tomato TPSs, TPS47 has the highest isoprene score and is shown in bold. PaIspS, isoprene synthase from *Populus alba* (AB198180); EgIspS, isoprene synthase from *Eucalyptus globulus* (BAF02831.1); PmIspS, isoprene synthase from *Pueraria montana* (AY316691.1).

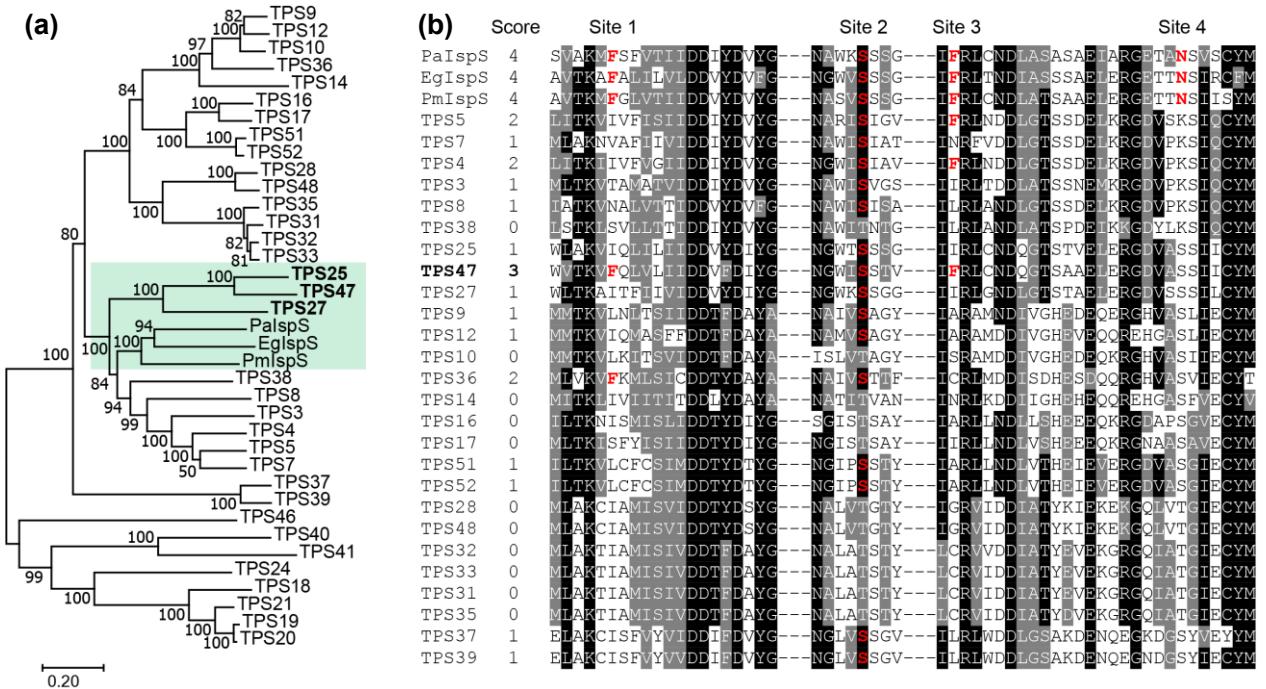


Fig. S7 Sequence alignment of the proteins encoded by the functional tomato TPS-c genes and CPS genes from other plants. Black shading indicates identical amino acid residues and grey shading are the conserved or semi-conserved substitutions observed and dashes indicate gaps. The conserved DxDD motif is underlined. The truncation sites for protein expression are indicated with triangle. AtCPS, copalyl diphosphate synthase from *Arabidopsis thaliana* (At4g02780); NtCPS2, 8-hydroxy-copalyl diphosphate from *Nicotiana tabacum* (HE588139.1); OsCPS1_{ent}, *ent*-copalyl diphosphate synthase 1 from *Oryza sativa* (Q6ET36.1); OsCPS2_{ent}, *ent*-copalyl diphosphate synthase 2 from *Oryza sativa* (Q6Z5I0.1); OsCPS3_{syn}, *syn*-copalyl diphosphate synthase 3 from *Oryza sativa* (Q0JF02.1); MtHPS, halimadienyl diphosphate synthase from *Mycobacterium tuberculosis* (NP_217894); SaCPS, copalyl diphosphate synthase from *Salinispora arenicola* (WP_012181499.1).

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TPS41	1	-----MLVTNSYRLSCHIHLRKLSPNFLLTQQAEEFKRKGHSRNWLQTEG---LPPISLNACPN
NtCPS2	1	-----MQVIITSSHFRFCHHLHQLKSLSAQAEFKKHGPRLWNLFQTEGSSLYKPVRLNCATs
OsCPS1ent	1	MIHLHSPPPTAPAAFGGAGSADWRRRRWSWSSSRAPVAKGGHLRPCVWRGGDDGGEDHHADGGGGGGAAWRARATTAGVSSSSTAGKLQANII
OsCPS2ent	1	-----MQMQVLTAASSLPRATLLRPAAAEPWRQSFQLQARPIQRPGIMTHCKAQLQ
OsCPS3syn	1	-----MPVFTASFCQCVTLFGQPASAADAQPLLQGQRPFHLHARRRRPCGPMIISKSPPY
MtHPS	1	-----
SaCPS	1	-----MSADL

TPS40	67	PVIKWDIAEEVDVETHTLEVY----DPS-SNEPDHIDAIRSMILGSMG-----DGEISVSAYDTAWVAMVKDVKG----TETPQFPSTIWEIANNQLADGSW
AtCPS	67	DLPLIHEWQQLQGEDAPQISVG----SNSNAFKEAVKSVTILRLNLT----DGEITISAYDTAWVALID---AG----DKTPAFPSAKWIAENQLSDGSW
TPS41	56	DASYFGNVSDLKSDHSNKYEE----KDIQMRNIDREEIKNMLSSMG----DGRSSVSFYSYDTAWVSFIEDNTNTINGTSKRFLFPSCIQWIIDDNQLDDGSW
NtCPS2	61	DASYLGNVNEYLESDHSKNSEEKDIQVSRТИQMKGTEEIKHMLNSME----DGRINLAYDTAWVSFIPNTTN--NGNDQRPMFPSCIQWIIDDNQLSDGSW
OsCPS1ent	101	HETPRITKWPNSRDLDDHQQNNEADEEADDELQFLVEOVRSMISSME----DGAIITASAYDTAWVALVPRIDC----EGGTQFQFPAAWRWIVGSQLADGSW
OsCPS2ent	53	GQETRERRQLDDDEHARPPQGGDDDVAASTSELPMIESIKSKLRAARNLSIGETTVSAYDTAWALVNRLDCGGE--RSPQFPEADWIARENQLPDSGW
OsCPS3syn	56	PASEETREWEADGQHEHTDELRT----ETTTTMDGIRTAIRSIG----EAEISISAYDTSLVALLKRLDGGD----GQFPSTIDWIVONQLPDSGW
MtHPS	1	-----METFRRLAKALAALGNGISSLAYDTAWVAKLGQFLDD-----ELSDLAIDLWICEROLPDSGW
SaCPS	6	GASA PAAVEPVPA-----RADTAESI VAELIRVP--AGQVSPSILYETARIVSLAPWLTG-----HAERVRHLITQSOPRDGGW

TPS40	154	GDNSIIPILVYDRVINTLACVIAIKS-----WNLHPDKILLGNSFMRNLSRIGDENAEHMPIGFEVAFPSLIEIAKKLGID-----FPYDSPVILQDIYASR
AtCPS	153	GDAYLFPSYHDRLINTLACVVAIKS-----WNLFPHOCNKKGITFFRENIGKLEDEDENDHMPIGFEVAFPSLIEIARGINID-----VPYDSPVILQDIYAKK
TPS41	147	GEELVFCITYDRLLNTLACVVATL-----WNTCLHKRNKGKMFIFIKENRERLEGGEVVMNTSGFFFVFPSSLDKAQQEHDIDDN-----IPYDAPVIFEDIYARR
NtCPS2	157	GEEIVFCIYDRLLNTLVCVIAITL-----WNTCLHKRNKGKMFIFIKENSLKETGEVNMNTSGFEIIVFPILLEKAQOLDID-----IPYDAPVIFEDIYARR
OsCPS1ent	194	GDEALAESAYDRVINTLACVVIAITR-----WSLHDQCKQGQIQLNLNWLRAAEEFDTTPMIGFEIAPFSLVEAARGLGID-----FPEYDHPAKLGIGIYANR
OsCPS2ent	150	GDAGMFITQDRLINTLCCVVAIAT-----WGVHEFQRARGIAYIQODNIWFLGEDDEBWWMMVGFEITFPVULKAKNLGID-----INYDDPAIQDIYAKR
OsCPS3syn	141	GDASFMMGDRINTSTLACVVIAIKS-----WNIHTDCKCERGIIIFIQENMWLAHEEDDWMLVGFELALAPSLLDMAKDIDID-----IPYDEPAKAIAYAER
MtHPS	56	GAEEPFCEYEDRLLSTLAAAMTSNKHRRRAAQVEKGLLAKNLTSGAFEGPQDOLDIKADTVGFELIAPTLNEAARLGIA-----JCHEESIIGLEVGR
SaCPS	77	GPPGEGMAVVPVMTSAATEALIAHTRAPAPAEPLIRATDAGLTVTRWLWSPRSPPDTPAIDIVPALAAAIRNLHVEADEPSALGHWRRAALARLPAGMDDR

TPS40	244	QLKLTRIPKDIMPKVPTTLLHSLEGMT-DLDWQKLLOQFCQT-DGGSFLFSPSSATAVLMOTQ---	DHNCILNYIKNAVHKFNGGVNPVYVVDLFEBHIWTVDR
AtCPS	243	ELKLTRIPKEIMPKPTTLLHSLEGMR-DLDWKLKLKLQSQDGSFLFSPSSATAFMOTR---	DSNCILEYIRNAVKRFNGGVNPVEVVDLFEBHIWTVDR
TPS41	238	EVKFTREPDKDLIHTIPTIVLFSLEGDR-DLDWQRLLKLQMEDGSFLTSPSSATAIVFMNTN---	DDKCFTEIQLNAVQKFNGGVPCSYAPIQARLIWAIDR
NtCPS2	247	EVKLTRIPKDVIITIPTTVLFSLEGDRDLDWQRLLKLQMPDGGSFLISPSTATAFAFMETN---	DEKCLAYIQNVVERKNG-GARQYPFDLVTRIWAIDR
OsCPS1ent	284	ELKLKRIPKDMMIIVPITSILHSLEGMP-GLDWQRLLKLQCSDGGSFLFSPSSATAVLMOTG---	DKKCFANVLDRIKEDGGVPNVYVVDLFEBHIWVWVDR
OsCPS2ent	240	QLKLAKXIPREALHARPTTLLHSLEGME-NLDWRLRLLQFKCPAGSLHSPPASAASAYALSETG---	DKELEYIETAINNEFGDGGAPCTYPVDNFDRWLSVDR
OsCPS3syn	231	ERKLAKRDPDVLISMPPTTLLHSLEGMV-DLDWKLKLRLCDGSFHCHSPSATAFOQTG---	DKCFKEVYDGFIVKVKFNGGVPCYIPLDLYVTRIWAIVDR
MtHPS	152	EOKLRKLGGSKINKHITAASFVELAGQDCVGMLDVNDLQETNGSVKYSPSASAFAHVKP--	CDKRALAMLISSIIQAGDGGAPAFYQAEIFFIVWSIWN
SaCPS	177	RRAAVHGLIGAGAAPEKLLHLALEVVG-SAAHGVGRVRPTRSRGIVGASPAAAWLGSPAGGHRRHPGASAYIERVVRQHDALAPCATPITVFERAWVAT	

TPS40	339	IQLRGISR---	YEELEIKEC	DYVSRW	WIN-KGIC	WARN	SPVQDIDDDT	TAMAFRL	RLRHG	YAVSA	DVF	KHFFSK	GFFC	VQGSN-QAVTGM	NLYRASHV		
AtCPS	338	IQLRGISR---	YEEEI	KECLDYV	HRYWTD-NGIC	WARMCSH	QDIDDDT	TAMAFRL	RQHG	YQVS	DVF	KNFKE	GFFC	VQGSN-QAVTGM	FNLYRASQ		
TPS41	333	IQLRGISY--	YFEEEIKD	LLEIYF	VFRYWD	KENGGFFS	ARNSNICEVDT	TCAIR	RLRLHGF	FDVSP	PVLHKFKD	GDEFFCLR	GESN-KSAT	VFMNLYRC	SQA		
NtCPS2	342	IQLRGISY--	YFAEFKEL	LNHVFRYD	EENGIF	SGRNSNNSDV	DDTCMAIR	RLRLHGF	YDVSP	PA	LNNF	KDGDQF	VCE	GEVD-GSP	THMENL	YRC	SQV
OsCPS1ent	379	LERLGISR--	YIQREIEQN	DYVN	RHTIE-DGIC	WARN	SNVKEV	DDTAMAFRL	RLRHG	YN	SPSVF	WKNDKF	GFFC	VQGST-QAVTGM	NLYRASQI		
OsCPS2ent	335	LRLRGISR--	YFTSEIEEY	FLAYARH	IISP-DGMSY	YGLCP	VKWDIDDDT	TAMAFRL	RLRHG	YN	SSSVF	NHF	KDGEYCF	QGS-SQ	LTAM	YNSYRASQI	
OsCPS3syn	326	LTRLGISR--	HFTSEIEE	DCLDYI	FRNWT-DGLA	HTKNCPV	KWDIDDDT	TAMAFRL	RLRHG	YQV	DPCV	LKF	KDGF	CFLH	GESNPSS-VTP	PMYNTYRASQ	
MtHPS	250	LSRTDIDLSDP	EIVRTYLPY	LHDV	QEHWVRGRGV	GTGNST	EDCDTTS	WADV	VLSKEGR	SPD	GAIVLQF	FE	DADWFRT	FHEVG-			
SaCPS	276	LARAGIAVT--	QAPD	LI	PGLADLT	TSV	IC-----	AGP	GPLP	LA	TTA	TLYALAH	GESV	DLECLWR	YTPDGFCT	WE	GEDG-----

TPS40	434	MFSGEK-ILENAKIFITSNYLREKFAQNQILDKWIITKDLPGEVGYALDVPWYASLPRLTRFFLEHYGGEDDVWIGKTLYRMPVUNNSLYLEI	LAKSDYNN	
AtCPS	433	AFFREE-IILKNAKEFSYNYLLEKREREELIDDKWIIMKDIPEGEIGEALEIPWYASLPRVETRFYIDQYGGENDVWIGKTLYRMPYVNNGYLEI	LAKQDYN	
TPS41	429	LFPGEI-ICEEAKNFTYNFLHQYLANNQSCKDKWVIAKDIPEGEIYALEFQWYASLPRVSRLYIDQYGADEIWISKTLYRMPDWVN	VNVLYEIAKLDYNR	
NtCPS2	438	LFPGEK-ILEEAKNFTYNFLQOQCIANNRCLDKWVIAKDIPEGEIYALEFQWYASLPRVARYIEQYGGADDIWIGKTLYRMPDVNNNVYLOQA	AKLQDYNR	
OsCPS1ent	474	SFPGED-ILQARARNFSYEFLREBFAQGTIDDKWIIISKDLPGEVCYTLDFPWYASLPRVEARTYI	GOYGGNDWVIGKTLYRMPIVVNNTAYLEIAKQDFNR	
OsCPS2ent	430	VFPGLDDGLEQIRAYCRAFLEERRATGNIMDKWVIAANGLPSEVEYALDFPWYASLPRVETRVYLBQYGASEDAWIGKGLYRMTLVNNNDLYL	AAKADFTN	
OsCPS3syn	422	KFPGDDGVGLGRAEVFCRSFLQDRGSNRMKDKWAIAKDIPGEVEYAMDYPWKASLPR	ETRLYLDQYGGSGDVWIGKVHMRMLFCNDLYLKA	AAKADFSN
MtHPS	334	-----	PSISTNVHVVLGALKQAGYDK	
SaCPS	352	-----	FSLSTNAHVLDVVGILTTDPGADRRH	

TPS40	533	CQAIHQFEWRRRKWVYECGLGFQ-----LSEKRLLVITYLGSASIFEAQRSTERMAWVKTAAIMDCVRSQFGSPQVAAAFLCEFAHYSSTALNSRYN	
AtCPS	532	CQAOHQOLEWDIIFQKWWYEEENRLSENG-----VRRSELLCECYLAAATIFESERSHERMWAKSSVIVKAISSEGESSSDRSFSQDFHEYIANARRSDHH	
TPS41	528	CQSQRHRFEWLIMQEWEFKGNFQKFG-----ISKKEVVLVSFLAASSIFEVEESRERLAWAKSICLCKMITSYINQEATWN-----SFLMEFKNYRDMSIKK	
NtCPS2	537	CQSQRHRFEWLIMQEWEFKCNFQKFG-----ISKKYLLIVSYFLAASIFEVESRERLAWAKSRIICKMITSYINQEATWN-----WTTRNSLLMEFKVSHDPTRKN	
OsCPS1ent	573	COAIHQELOCIQKWEIENGLEAFG-----MTPEDVIRRAYFLAACIFEPNPERASERLAWARVSVIANTISRHEYSDMSMKRMRERFMWSSLYEENGNVLG	
OsCPS2ent	530	FQRLSRLEWLSIKRWYIRNNLQAHG-----VTEQSVLRAYFLAANIFEPNRAAERLGWARTAIIAEAIASHLRQYSANGAADMGRTERLISGLASHDW	
OsCPS3syn	522	FQKECRVVELNGIIRRWRWYLSNLEKFGGT-----DPQTTLMTSYFLASANIIFEAANRAAERLGWAVALLADAVSSSHRRIGGPKNSTSNSLLELI	SLVPFDAYS
MtHPS	354	CHPVRVKVLEFTRSSKEPGRFCWRDKW-----HRSAYYTTAHLICAASNYDAAICSDAICWILNTQRPDGSWGHEDGQATAETAYCIQALAHWQRHSG-----	
SaCPS	379	VTAAPRLADALRQQQADGSWQDRWHASPYYATMCCALAGFP-----PGTAVTSLARAASIVDQTQRANGSGWGRKGTVEETAYAQVQLATVGRG-----	

TPS40	628	TEDR-----LVGVTLGTLNHLLSALLTHG-----RDHHYHLRHAENWLTVGEGEGEGE-----AELIIRTLNLC	
AtCPS	627	FNDRNMRLDRPGSVQASRLAGVLTQLNQMSFDLFMSHG-----DNLLLYLSWDWMEKWKLGYDGE-----LMVKMIIILMKN	
TPS41	620	SNET-----KEIIVLNNCQFLHQIKEYTDQDLG-----KDHQHQLHNVVEELENN-----TTCAEA-----AVLIVQTINLS	
NtCPS2	632	GNET-----KEIILVKNQRQFLRQLSETTFEDLG-----KDHQHQLQNWETWLFLIREEKNAQCQE-----TELLVRTINLS	
OsCPS1ent	668	LEGYAKDG-----ILARTCQLIDLSSQETPPVREGQKC-----HNLRCAWIEWMQQINMKDGRYDKGRVMHPGSCTVHNKETCLLIAQIVEICAGRI	
OsCPS2ent	625	RESK-----DSAARSLLYALDELIDLHAFGNASDSLREAQKWLMSWTNE-----SQGSTGG-----DTALLLVRTIEICS	
OsCPS3syn	619	G-----SLREAWKQWLMAWTAKESSQESIEG-----DTAILLVRRAIEIFG	
MtHPS	448	-----	
SaCPS	472	-----	

TPS40	692	SVHWISEEILLSHPTMOKILLEITNRVSHRIR-----LYKGHSEKQVGMLTFSEEIEGDMQIAELVLSHS DASEIDANIIDTFLTVAKSFFYSAVYCDRTIN	
AtCPS	705	NDLTN---FFTHTHVRDAEITNRCIPLRQ-----YLKARRNDEK-----EKTIKSMEMGKMEALASESDTFRDVSITFLDVRKAFFYFELICGD-HIQ	
TPS41	685	S-GHMTHDEILS-----KYTNKVCCHMIN-----EFQN-----DQICNSSKARDIELHMOALVKLVFSNTSSNN-----INQGIEDTYFKVVKTFYYTBHVSEETIN	
NtCPS2	700	G-GYMTHDEILFDADMENLNLNTNKVCGKLN-----ELCN-----DKVTGGSKNTNIELDMOALVKLVFGNTSSN-----INQDIDQFFAVVTFYYSHVSEEIN	
OsCPS1ent	759	EEAASMINNTEGSWFIQLASSICDSIHKAL-----LSCDTKKNETTINQIDKBIELGMQIAQYILPRVDDRRUNNKTQFLSIVKSCYYAANCSPHMID	
OsCPS2ent	691	GRHGSAEQSLKNSADMARLEQIASSMCSKLATKILAC-----NGGSMMDNVEGIDQEVDFVEMKELIQRVYGS-SNDVSSVT-----TFLDVVKSFCYVBHCSPETID	
OsCPS3syn	659	GRHVLTGQRPD-LWEISQLEQLTSSICCKISRRVLAENGESTEKVEEIDQQVDEMQILTRRVLQG-----CSANRLTEETFLHVVKSFCCYVBYCSPETID	
MtHPS	448	-----TSLSAQISRAGGWLSQHCEPPYAPLWIAKILYCSATVVKAAIL	
SaCPS	472	-----RPGAEAIRRGRAYLTEGTTAHDPGPLWHDKDLYRPAMIVRAAVV	

TPS40	789	FHIAKVLFERWV-----	
AtCPS	792	THISKVLFQKV-----	
TPS41	770	NHISKVLFQKA-----	
NtCPS2	792	FHISKVLFQKV-----	
OsCPS1ent	856	QHISEVIFEQWV-----	
OsCPS2ent	789	GHISKVLFEDWN-----	
OsCPS3syn	756	SHIDKVFQDWV-----	
MtHPS	491	SAJIRIVDESNQ-----	
SaCPS	518	AARHLAGAAGFATA	

Fig. S8 Predictions of subcellular localization for TPS and prenyltransferase proteins made by TargetP 1.1 (a) and ChloroP 1.1 (b).

(a) **### targetp v1.1 prediction results #####**
 Number of query sequences: 42
 Cleavage site predictions not included.
 Using PLANT networks.

Name	Len	cTP	mTP	SP	other	Loc	RC
TPS3	607	0.781	0.218	0.044	0.036	C	3
TPS4	590	0.756	0.205	0.040	0.055	C	3
TPS5	609	0.563	0.315	0.087	0.018	C	4
TPS7	592	0.499	0.155	0.233	0.054	C	4
TPS8	597	0.375	0.099	0.136	0.546	-	5
TPS9	548	0.050	0.182	0.180	0.693	-	3
TPS10	556	0.166	0.244	0.083	0.313	-	5
TPS12	548	0.053	0.301	0.139	0.461	-	5
TPS14	568	0.113	0.093	0.089	0.861	-	2
TPS16	553	0.075	0.144	0.059	0.858	-	2
TPS17	554	0.091	0.149	0.063	0.774	-	2
TPS18	773	0.024	0.900	0.009	0.207	M	2
TPS19	778	0.612	0.323	0.014	0.133	C	4
TPS20	778	0.612	0.323	0.014	0.133	C	4
TPS21	784	0.661	0.215	0.022	0.122	C	3
TPS24	820	0.930	0.035	0.075	0.070	C	1
TPS25	547	0.272	0.086	0.089	0.713	-	3
TPS27	554	0.214	0.166	0.081	0.576	-	4
TPS28	554	0.169	0.129	0.125	0.725	-	3
TPS31	555	0.243	0.057	0.145	0.727	-	3
TPS32	548	0.113	0.199	0.078	0.627	-	3
TPS33	552	0.171	0.123	0.193	0.514	-	4
TPS35	556	0.211	0.073	0.142	0.692	-	3
TPS36	591	0.548	0.061	0.144	0.293	C	4
TPS37	560	0.547	0.206	0.055	0.092	C	4
TPS38	546	0.157	0.177	0.046	0.816	-	2
TPS39	563	0.055	0.151	0.252	0.584	-	4
TPS40	800	0.921	0.093	0.038	0.044	C	1
TPS41	780	0.077	0.730	0.022	0.145	M	3
TPS46	821	0.231	0.055	0.112	0.891	-	2
TPS47	562	0.096	0.239	0.083	0.649	-	3
TPS48	560	0.327	0.105	0.160	0.528	-	4
TPS51	550	0.054	0.154	0.074	0.876	-	2
TPS52	551	0.038	0.359	0.059	0.720	-	4
FPPS1	342	0.027	0.053	0.378	0.864	-	3
SSU1	315	0.019	0.278	0.057	0.600	-	4
SSU2	334	0.366	0.190	0.170	0.100	C	5
GGPPS3	360	0.505	0.132	0.010	0.423	C	5
IPT1	372	0.158	0.866	0.013	0.029	M	2
IPT2	373	0.178	0.785	0.009	0.053	M	2
GGPPS2	363	0.322	0.082	0.028	0.170	C	5
GGPPS1	365	0.823	0.085	0.074	0.119	C	2
cutoff	0.000	0.000	0.000	0.000	0.000		

(b) **### chlorop v1.1 prediction results #####**
 Number of query sequences: 42

Name	Length	Score	cTP	CS-score	cTP-length
TPS3	607	0.564	Y	2.430	28
TPS4	590	0.550	Y	-0.472	38
TPS5	609	0.523	Y	-0.094	42
TPS7	592	0.538	Y	5.945	45
TPS8	597	0.443	-	-1.415	8
TPS9	548	0.437	-	1.897	2
TPS10	556	0.486	-	1.526	4
TPS12	548	0.446	-	3.181	38
TPS14	568	0.436	-	7.857	68
TPS16	553	0.428	-	7.547	9
TPS17	554	0.427	-	1.932	9
TPS18	773	0.454	-	0.863	28
TPS19	778	0.545	Y	8.925	36
TPS20	778	0.545	Y	8.925	36
TPS21	784	0.528	Y	5.963	36
TPS24	820	0.548	Y	4.184	53
TPS25	547	0.443	-	-0.394	8
TPS27	554	0.446	-	-0.085	33
TPS28	554	0.433	-	-2.312	16
TPS31	555	0.433	-	4.268	3
TPS32	548	0.438	-	-0.872	12
TPS33	552	0.442	-	6.280	3
TPS35	556	0.436	-	6.941	3
TPS36	591	0.485	-	3.693	41
TPS37	560	0.505	Y	8.499	30
TPS38	546	0.432	-	-2.033	13
TPS39	563	0.444	-	-0.156	86
TPS40	800	0.548	Y	2.548	46
TPS41	780	0.462	-	-0.630	26
TPS46	821	0.446	-	-1.743	13
TPS47	562	0.436	-	5.568	61
TPS48	560	0.469	-	0.624	42
TPS51	550	0.427	-	1.403	29
TPS52	551	0.428	-	2.759	34
FPPS1	342	0.432	-	1.534	12
SSU1	315	0.439	-	7.229	22
SSU2	334	0.512	Y	5.161	17
GGPPS3	360	0.567	Y	5.148	65
TPT1	372	0.506	Y	2.832	13
TPT2	373	0.507	Y	3.003	13
GGPPS2	363	0.528	Y	4.003	63
GGPPS1	365	0.553	Y	3.695	43

Fig. S9 Phylogenetic analysis of tomato TPT homologs and TPTs identified in other plants. The maximum likelihood phylogenetic tree was constructed using MEGA7 with default settings. The first two letters in each of the homologs are abbreviations for the species: Aa, *Artemisia annua*; Ag, *Abies grandis*; At, *Arabidopsis thaliana*; Am, *Antirrhinum majus*; Cb, *Clarkia breweri*; Cr, *Catharanthus roseus*; Hb, *Hevea brasiliensis*; Hl, *Humulus lupulus*; La, *Lupinus albus*; Mi, *Mangifera indica*; Mp, *Mentha piperita*; Nt, *Nicotiana tabacum*; Os, *Oryza sativa*; Pa, *Picea abies*; Sa, *Sinapis alba*; Vv, *Vitis vinifera*; Zm, *Zea mays*. The GenBank accession numbers of the proteins used in the phylogenetic analysis can be found in Methods S1. The tomato TPT homologs are shown in bold. TPT1, TPT2 and the four *Arabidopsis* GFPP synthases are shaded in red.

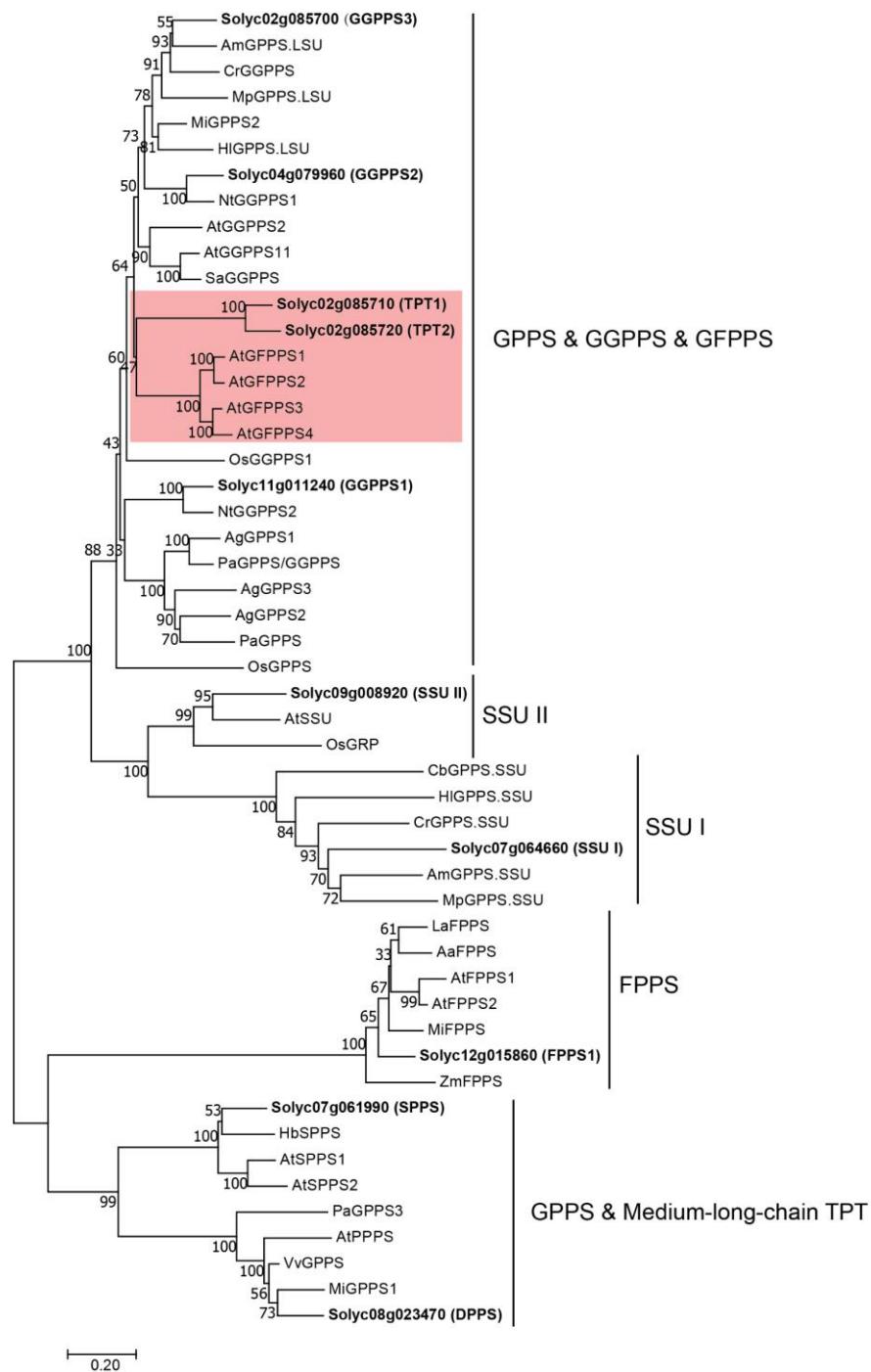


Fig. S10 GC-MS analysis of the products formed in planta by transiently co-expressing TPS with TPT genes in *Nicotiana benthamiana* leaves. Transit peptide truncated versions (“d”) of all tested genes were transiently expressed in tobacco cytosol, co-expression of *AtGFPPS2* with *AtTPS19* was used as a positive control to produce the sesterterpene (-)-retigeranin B (Shao *et al.*, 2017).

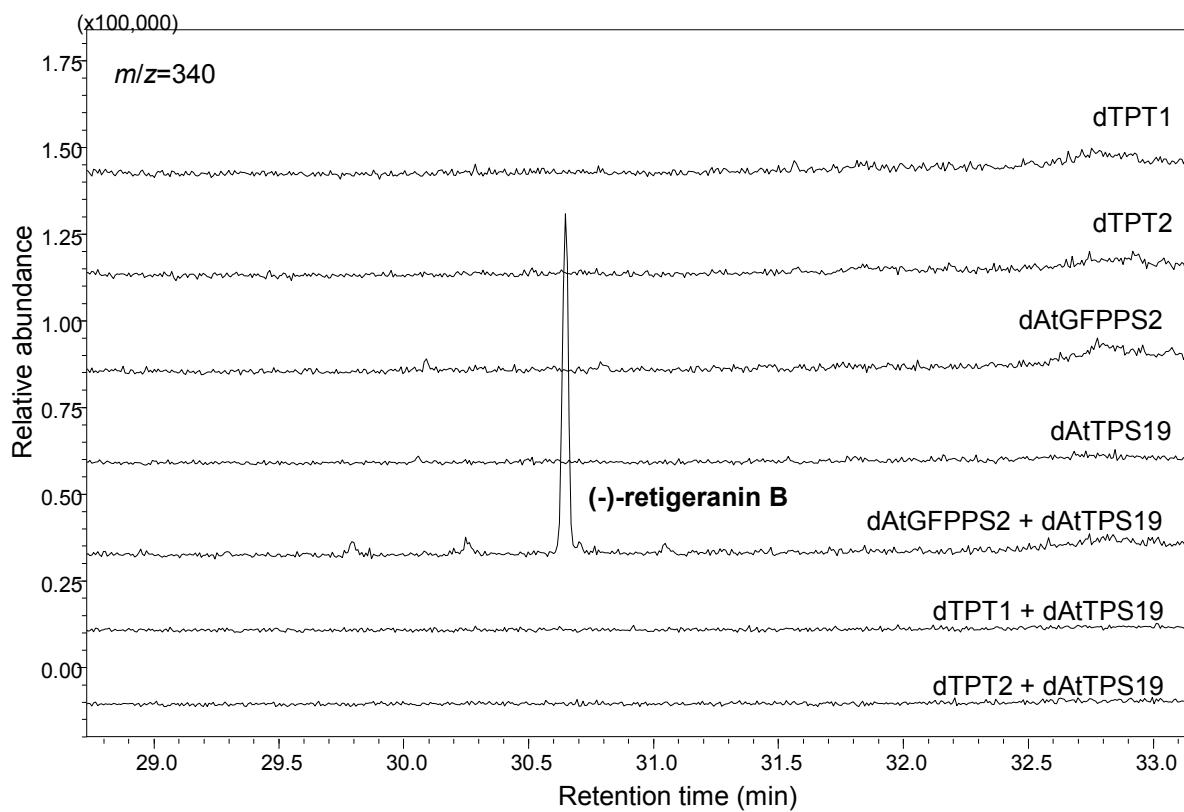


Fig. S11 Sequence alignment of five *Arabidopsis* sesterterpene synthases and tomato TPSs from TPS-a, TPS-b and TPS-g clades. (a) Key residues determining the substrate specificity of sesterterpene synthases are shown in red, the corresponding residues of tomato TPSs are shown in blue. Black shading indicates identical amino acid residues and grey shading are the conserved or semi-conserved substitutions observed and dashes indicate gaps. (b) Amino acids with large side chain (shaded in red) from tomato TPSs. (c) Amino acids with small side chain (shaded in yellow) from *Arabidopsis* sesterterpene synthases.

(a)	TPS5	323 F-SRDILVENVNMF W AVGALFEPQHS
	TPS7	305 F-SRDRILVED F WSVGLAFEPQHS
	TPS4	303 F-SRNRLIVEVNL F WAvgTNFEPQHS
	TPS3	315 F-ARDRLIVEAFQ W ITGMIFPQEN
	TPS8	286 F-ARDRLIVENE W TIGVNEIPQYG
	TPS38	259 F-VRDRLIVEGF W AVGFTPNPQFG
	TPS25	265 F-TRERLIVSEFF E AVGIASEGEHG
	TPS47	273 F-TDRDLIVECE E AGGIGSKEPQOG
	TPS27	269 F-SRDRIVESEL W SVGVAFEPQHS
	AtTPS30	317 C-VRDRIVEVTF P TLGLYFEFRFS
	AtTPS17	315 Y-IRDRIVEVVF G ALALYFEFRYS
	AtTPS25	316 Y-SIRERNVETYL G GLGVLFEEFRYS
	AtTPS18	317 Y-IRDRILVESSH L GGLGPYFEEPHYS
	AtTPS19	314 Y-IRDRSVESSH L GGLGPYFEPQYS
	TPS9	261 Y-ARDRLIVECYF W ILGVYFEEKYS
	TPS12	260 Y-ARDRLIVECYF W ILGVYFEEKYS
	TPS10	266 Y-ARDRLIVECYF W ILGVYFEEKYS
	TPS36	301 Y-ARDKLVEGYI T TLGVYFEPQYS
	TPS14	280 Y-ARDKLVBCYF W ATGVYFPQYK
	TPS16	264 Y-VRDRILABVYF W SLGVYFEPQYS
	TPS17	267 Y-ARDRLIVECYF W CLGTYFEPQYS
	TPS51	262 F-ARDRIVVLIYF W SLGVYFEEPHYS
	TPS52	263 F-ARDRIVVLIYF W SLGVYFEEPHYS
	TPS28	266 Y-VRDRMVECF F TVGVYFEPQYS
	TPS48	272 Y-VRDRMVECF F AVGVYFEPQYS
	TPS32	262 Y-ARDRAVEBCYF W TMGVYAEPOYS
	TPS33	266 Y-ARDRAVEBCYF W TMGVYAEPOYS
	TPS31	269 Y-ARDRAVEBCYF W TMGVYAEPOYS
	TPS35	270 Y-ARDRAVEBCYF W TMGVYAEPOYS
	TPS37	279 L-LRDQPLKWT T WPMTMLTDPKMS
	TPS39	280 L-LRDQPLKWT T WPMTMLTDPKMS

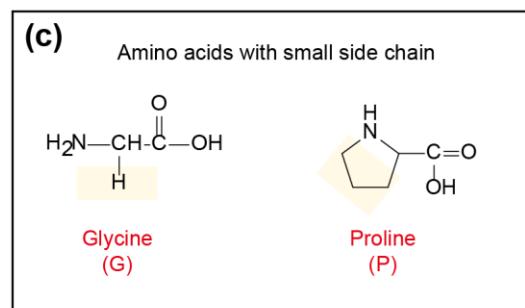
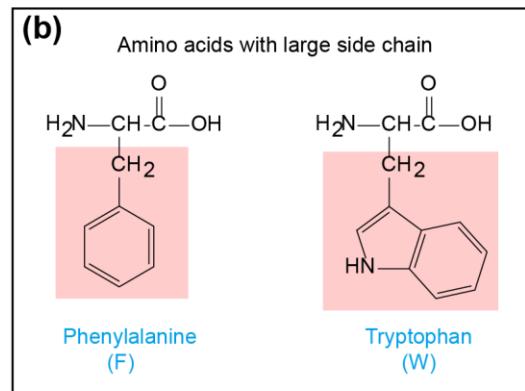
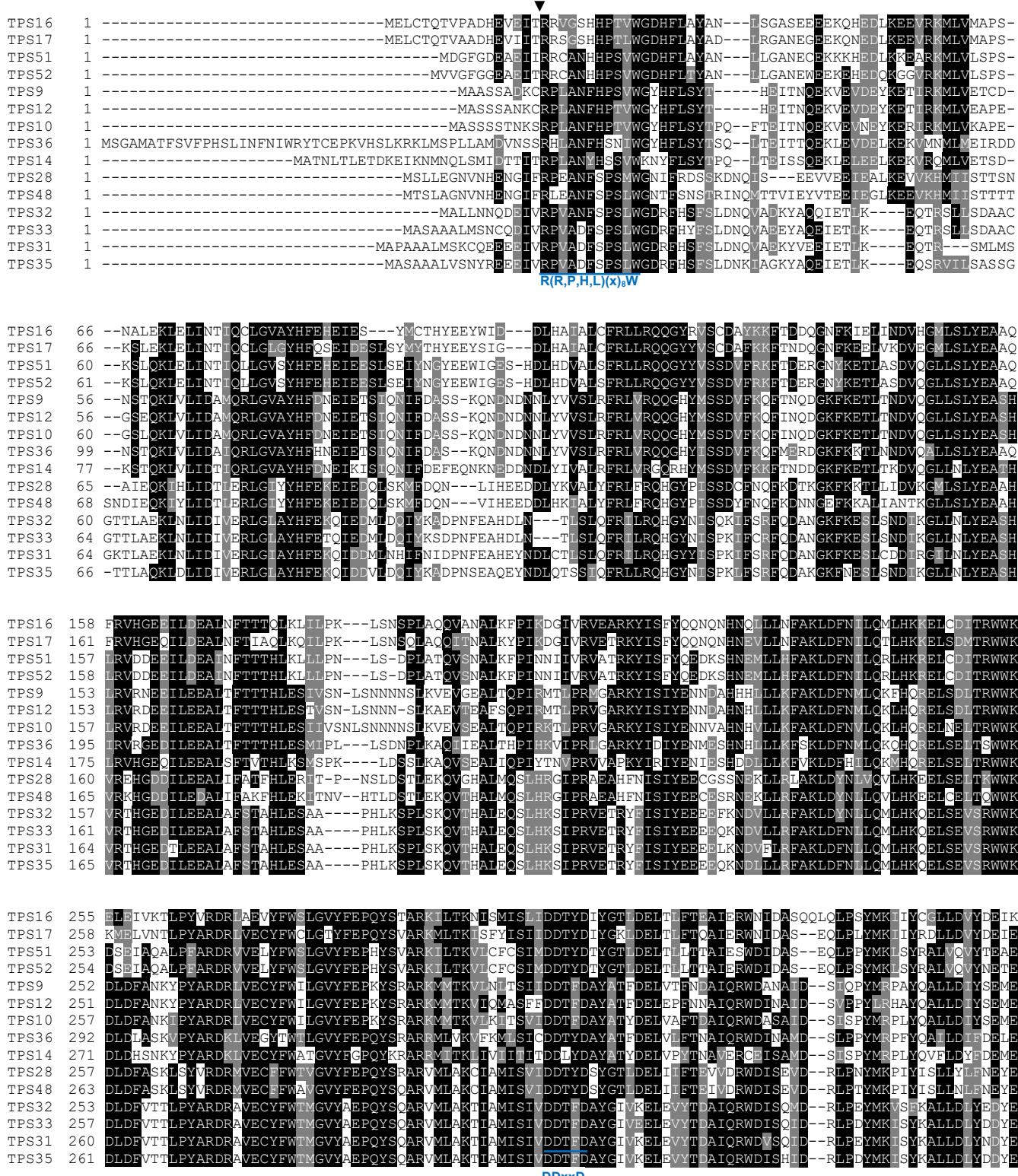


Fig. S12 Sequence alignment of the proteins encoded by the functional tomato TPS-a genes. Black shading indicates identical amino acid residues and grey shading are the conserved or semi-conserved substitutions observed and dashes indicate gaps. The conserved R(R,P,H,L)(x)8W and DDxxD motifs are underlined. The truncation sites for protein expression in *E. coli* are indicated with triangle.



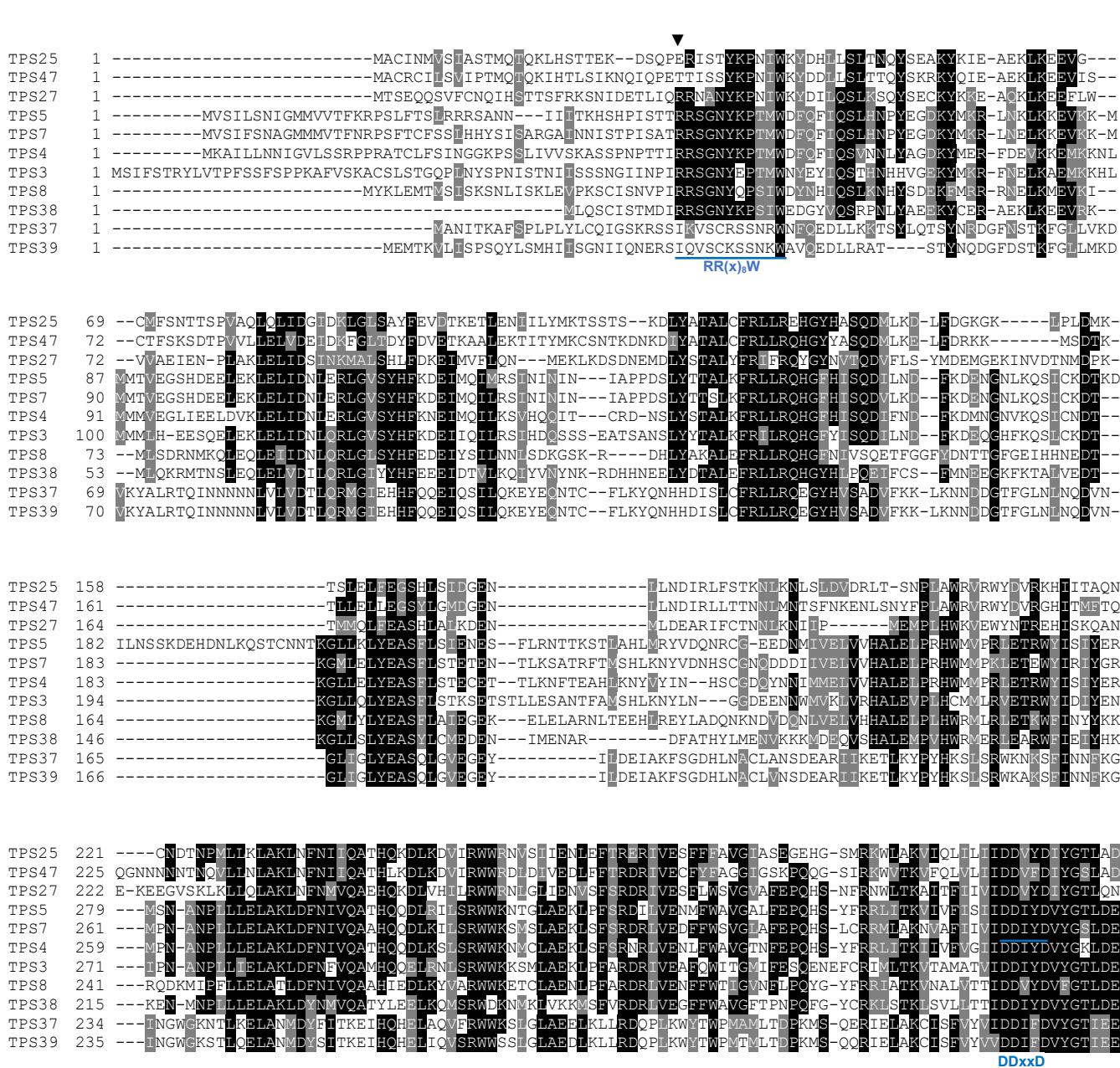
TPS16 355 KDLANE--NKSFLVNYNSIIEMKKMVMAFYQEEKWVY-GKTFPKMEEYIKSGISITSAYQVQVATTSWLGMGNVATKDFDWIVNEEPIILVASSIIARLINDL
 TPS17 356 KELANE--NKSFLVNYNSINEMKKVVRGYFQEEKWVY-GKKVPTMEQYMKNGISITSAYILLTTSWLAMGNVATKDAFDWVATEFFIVVASCYIIIRLINDL
 TPS51 351 RELENLGNKMTFRVKYSINEMKKLLRAYQEEKWVH-EKVVPTMEQYIKNGIPSSSTYLLLATTWSLGMGDVATKDAFDWISNEPTIILVLSIIARLINDL
 TPS52 352 KELENLGNKMTYRVKYSINEMKKLLRAYQEEKWVH-GKDVTMEQYIKNGIPSSSTYLLLATTWSLGMGDVATKDAFDWISNEPTIILVASSIIARLINDL
 TPS9 350 QVLSKEG--KLDRVVYAKNEMKKLVRAYFKEOTMLNDCDHIEPKYEEQENAIVSAGYMMISTTCLVGIEEFISHETFEWLMNESVIVRASALIARLINDL
 TPS12 349 QALAKEF--KSDRVVYAKYEMKKLVRAYFKEAOQLNNDNHIEPKYEEHNAMEVSAGYMMGATTCLVGIEEFISKETFEWMINELIVRASALIARLINDD
 TPS10 355 QVLSNEG--KLDRVVYKGKEMKKLARAYFKEAQMLN-AGYIPNCDEYIKNAIVSITTFMALGTTSLIGMEEFIFTKDIFEWITNEBESIILRASSTICRLMDD
 TPS36 390 BELTKEG--KSDRVVYAKNEMKKLARAYFKEAQMLN-AGYIPNCDEYIKNAIVSITTFMALGTTSLIGMEEFIFTKDIFEWITNEBESIILRASSTICRLMDD
 TPS14 369 BELTKEG--KAHYVYYAKVEMNKLIKSVAEMLK-NDIEPKCSEYIKRNATITVANQMILITCLIVAMEFISKETFEWMINESLIAPASSLINRUKDD
 TPS28 355 REINE--QDRFNGVNYYKEAMKEIVRSYNTIEAEWFT-EGKIPSFEYEINNALVTCFYLLPASLLGMESTS-KRTFDWMMKKPKILVASAIIIGRVIDDI
 TPS48 361 IEIELE--QDRFNGVWYVKEAMKEIVKSYNTIEAEWEL-EGKIPSFKEYIICNALVTFGYYLLPASLLGMKLAS-KRTFDWMMKKPKILVASAIIIGRVIDDI
 TPS32 351 KELSK--DGRSDVVQYAKERMKEIVRNYYVEAKWFI-EGYMPVSEYIISNALATSTYYLLTTSYLGVMKSAT-KEDFEWLAKNPKILEANVTLCRVVD
 TPS33 355 TELSK--DGRSDVVWYAKERMKEIVRNYYVEAKWFI-EGYMPVSEYIISNALATSTYYLLTTSYLGVMKSAT-KEDFEWLAKNPKILEANVTLCRVDD
 TPS31 358 TELSN--DGRSDVVWYAKERMKEIVRNYYVEAKWFI-EGYMPVSEYIISNALATSTYYLLTTSYLGVMKSAT-KEDFEWLAKNPKILEANVTLCRVDD
 TPS35 359 TELSK--DGRSDVVWYAKERMKEIVRNYYVEAKWFI-EGYMPVSEYIISNALATSTYYLLTTSYLGVMKSAN-KEDFEWLAKNPKILEANVTLCRVDD

TPS16 452 LSHEEEOKRGDAPSGVECYMKEYGVTKTEAHIKIRNTIENSWKDLYEYEFKVNGTHIPRVLIMCIINLARVIEFIYK-DEDAYTFPKNNLKDVYRLLID
 TPS17 453 VSHEEEOKRGNAASAVECYMNEYSVTKTEAHIKIRNTIENSWKDLNEEYFKVDMITIPRVLIMCIINLARVIEFIYK-DEDAYTFPSKNNLKDVISDLV
 TPS51 450 VTHEIEVERGDVASGLIECYMNEYGATKTEAYMETRKJIIENNWKVLRGOLKP-TTVPRVLIMFVNLNRVAEEFYK-DDDAYTFSTNNLKDIISILV
 TPS52 451 VTHEIEVERGDVASGLIECYMNEYGATKTEAYMETRKJIIENNWKVLRGOLKP-TTVPRVLIMFVNLNRVAEEFYK-DEDAYTFPSKNNLKDVISMLV
 TPS9 448 VGHEDEQERGHVASLIECYMKDYGASKQETYIKFLKEVTNAWDINKQFFRP-TEVPMFVIERVLNLRVADTYLYK-EKDITYTNAKGKLKNMINSILIE
 TPS12 447 VGHEVEQREHGHASLIECYMKDYGASKQETYIKFLKEVTNAWDINKQFFRP-TEVPMFVIERVLNLRVADTYLYK-EKDITYTNAKGKLKNMINSILIE
 TPS10 453 VGHEDEQERGHVASLIECYMKDYGASKQETYIKFLKEVTNAWDINKQFFRP-TEVPMFVIERALNFarVIDTLYQ-EVDGYTN SKGILKDLVNSLLIE
 TPS36 487 SDHESDQQRGHVASVIECYTKEYGASKQETYIKFLKEVKDAWKGINKAILRP-IEPVPIFVQILNIAFTMDTF-QDEEDGYTNNSNSKGDIVTILV
 TPS14 466 IGHEHEQBEHGASFWE CYVKEYRASKQETYVKEARRQIANAWKDINTDYLHA-TQVPTFVIIQALNLSPRLVDIQE--DPTDSQNFLLKDTIKIALLVD
 TPS28 451 ATYKIEKEKGQLVIGIECYMOENNLSVEAKASAQLSETAESAWKDLNKECIKTTTSNIPNEIILMRVVLNLRLLIDVYVYKNNQDGYSNPKNNVKSVIEALLVN
 TPS48 458 ATYKIEKEKGQLVIGIECYMEOBNLSVEAKASAQLSETAEAWKDLNKECIKSTDSPTEIILMRVVLNLRLLIDVYVYKNNQDGYSNPKNNVKSVIEALLVN
 TPS32 447 ATYEVKEKGRGQIAITGIECYMRDYGVSTOVAMDKEQMAETIAWKDVNEGILR--PTPVSTEIILTRILNLRLLIDVYVYKHNQDGYTHPEKVLKPHIIALLVD
 TPS33 451 ATYEVKEKGRGQIAITGIECYMRDYGVSTOVAMDKEQMAETIAWKDVNEGILR--PTPVSAKIITRILNLRLLIDVYVYKHNQDGYTHPEKVLKPHIIALLVD
 TPS31 454 ATYEVKEKGRGQIAITGIECYMRDYGVSTBEAMEKEEEAEIAWKDVNEGILR--PTPVSTEIILTRILNLRLLIDVYVYKHNQDGYTHPEKVLKPHIIALLVD
 TPS35 455 ATYDVEKGRGQIAITGIECYMRDYGVSTBEAMEKEEEAEIAWKDVNEGILR--PTPVSTEIILTRILNLRLLIDVYVYKHNQDGYTHPEKVLKPHIIALLVD

TPS16 551 PII----
 TPS17 552 PII----
 TPS51 547 PIITT----
 TPS52 548 PIKA----
 TPS9 545 SVKI----
 TPS12 545 SVEI----
 TPS10 550 SVKISIS-
 TPS36 585 SVTIIGRS-
 TPS14 561 SVNSTSCG
 TPS28 551 PIINM----
 TPS48 557 SIKL----
 TPS32 545 SIEI----
 TPS33 549 SVEI----
 TPS31 552 SIEI----
 TPS35 553 SIEI----

Fig. S13 Sequence alignment of the proteins encoded by the functional tomato TPS-b and TPS-g genes.

Black shading indicates identical amino acid residues and grey shading are the conserved or semi-conserved substitutions observed and dashes indicate gaps. The conserved RR(x)₈W and DDxxD motifs are underlined. The truncation sites for protein expression in *E. coli* are indicated with triangle.



Panel 4: Positions 221-331 (Continued)

Gene	Position	Sequence
TPS25	221	---CNDTNPMLIKLAKLNFNIQATHOKDLDKDVIRWWRNVIENLFTTRERIVESTFFFAGVIASEGEHG-SMRKWLAKVQLILLIDDVYDLYGTIAD
TPS47	225	QGNNNNNNTNOVQLINLAKLNFNIQATHOKDLDKDVIRWWRNVIEDLETFTRDRIVEOFYEAQGIGSKPQOG-SIRKWLTKVFLQVLIIDDVFDIYGSLAD
TPS27	222	E-KEEGVSKLKLLOLAKLNFNIQVAEHQDKLVHIIIRWWRNVIENVSFSRDRIVESLWSVGVAFEPQHS-NFRNWLTKAITFIIVIDDVYDLYGTIQLN
TPS5	279	---MSN-ANPLLLELAKLDFNIVQATHQODLRLISRWWKNTCLAEKLPEFSRDIILVBNMFWAVALFEPOHS-YFRRRLITKVIIFISIIDDYDLYGTIQLD
TPS7	261	--MPN-ANPLLLELAKLDFNIVQATHQODLKLISRWWKNTCLAEKLPEFSRDIILVBNMFWAVALFEPOHS-LCRMLIANKVAFIIVDDIYDLYGTIQLD
TPS4	259	--MPN-ANPLLLELAKLDFNIVQATHQODLKLISRWWKNTCLAEKLPEFSRDIILVBNMFWAVALFEPOHS-LCRMLIANKVAFIIVDDIYDLYGTIQLD
TPS3	271	--IPN-ANPLLLELAKLDFNIVQATHQODLKLISRWWKNTCLAEKLPEFSRDIILVBNMFWAVALFEPOHS-LCRMLIANKVAFIIVDDIYDLYGTIQLD
TPS8	241	--RQDKMIPFLLELATLDFNIVQATHQODLKLISRWWKNTCLAEKLPEFSRDIILVBNMFWAVALFEPOHS-YFRRRIATKVNALVTTIDDVYDLYGTIQLD
TPS38	215	--KEN-MNPLLLELAKLDFNIVQATHQODLKLISRWWKNTCLAEKLPEFSRDIILVBNMFWAVALFEPOHS-YFRRRIATKVNALVTTIDDVYDLYGTIQLD
TPS37	234	--INGWCKNTLKELANMDYITKEIHOHELAQFIRWWKSLCGLAEELKLIRDQPLKWTWPMALTDPKMS-QERIELAKCISEVYVVIDDIFDVYGTIEE
TPS39	235	--INGWCKSTLQELANMDYITKEIHOHELIQSRWSSLCLAEELKLIRDQPLKWTWPMALTDPKMS-QERIELAKCISEVYVVIDDIFDVYGTIEE

DDxxD

TPS25 414 HVLGLTN----ENLHLCK-NCQEIIYYTSLLIRLCNDQGTSIVELERGDVASSIIICYMH-QENVSEEDAREHTESTILNSWEKTN-YHFNRLSTSHRK
TPS47 424 YVIYGLTNNKITNESLDSSN-NFQEIIYHTSVIFRLCNDQGTSAAEELRGDVAASSIIICYMQ-QENVSEDVAREHIESIILDWSKKIN-YHFTNLMSHRE
TPS27 419 HVLLGLAODFS--QWDYFLE-NERDIKYSSLIIRGNDLGTTAELERGDVSSSIICYMR-KENVKEDVARKHIEEMVIETWKKMN-RHCFENSS---P
TPS5 471 HSLFLVTNQITKEALDSLTN--YPDIIRWSATIFRLNDDLGTSDELKRGDVSKSIQCYMN-EKGASEBAAIEHIEFLIQETWEAMN-TAQSKNSPLSET
TPS7 453 HAFIFLTNPITQFAESLNN--YPDIIRRCALINEFVDDLGTSDELKRGDVPKSIQCYMN-EKGVSEEBAREHINILIKEPEVMNKDQISKQVLFSEE
TPS4 451 HALFLVTDPITKEALDSLTN--YPDIIRCSATIFRLNDDLGTSDELKRGDVPKSIQCYMN-EKGVSEEBAREHIRFLIKEETWKFMNTAHHKEKSLFCET
TPS3 464 NAFFLVTNPITKEVLEYLFSNKYPDIIRWPAТИIIRLTDDLATSSNEVKRGDVPKSIQCYMK-ENGASEEEARKHINLMIKETWKMINTAQHDN-SLFCEK
TPS8 434 HAYFLVANPVNKEAHYLEN-NYHDITRCSALILRRIANDLGTSDELKRGDVPKSIQCYMN-EHQASEEBARQYIRRLISQTWKKLNEAWLAADPFPKI
TPS38 407 HAYFCITNPIKEDEIQLRN--HYPAIISPSYLRLANDLATSPDEIKNGDYLKSIIQCYMH-LSKSCEBNARYIKKLIDETWKKMNR-DILRDESLSKD
TPS37 427 HLYFYILGLCVS---SMHLQD--ISLMSTSIAKILRUWDDLGSAKDENQEKDGSSYVEYYMKENKDSSMLAREHVIKLIEDEWQLNKEFCLMQSOTRS
TPS39 428 HMFYLLGFGLNNQNQSIYLED--SSAMASSVATIIRLWDDLGSAKDENQEGNDGSYECYMKQQKNASIBLAREHVVKLIEDEWQLNKKHFNLMMNGSLGS

TPS25 506 IMKHVINEARMAHVMYLSGDCFGVQDGETQDQVIINLVQSHI-----
TPS47 521 IAKHVINIAARMGHVMYQFGDCFGVQDGKTRDQIIINLMEPIT-----
TPS27 511 LIKYIMNIARVTHFIYQONGDCFGVQDRETQOIISSLVQSHIPN-----
TPS5 567 FLEVAKNITKASHFMYLHSD-----VKSSISKILFEPIISNVAFALK-----
TPS7 550 FTKIVFNFSRTSHCVYQHGDCHGIQNSHITNRISKULFEPIII-----
TPS4 548 FVEIAKNTIATTAAHOMYLGKGDHSQIQTDVKNSISNULFHPIII-----
TPS3 562 FMGCAVNIAARTCQTLYQHGDCHCIQNYKIQNRISKULFEPIIT SMP-----
TPS8 532 FVTCAMNIALARMAQCQMYQHGDCHGNNNSTKHNIMAILFESWPIGHKHSSAEKDHSMVNYREKFMI
TPS38 503 FRRTSMNIALRIAQCQMYQHGDCFGIPDRETKDRILSLEFFQPIPT-----
TPS37 522 FSKASLNSARMVSIMYSYDDKQSIPILQEYIKSMLDGNL-----
TPS39 526 YSKASLNSARMVPLMYNDDKQSLHVLQEYINTLYDV-----

Fig. S14 Sequence alignment of the proteins encoded by the functional tomato TPS-e/f genes. Black shading indicates identical amino acid residues and grey shading are the conserved or semi-conserved substitutions observed and dashes indicate gaps. The truncation sites for protein expression are indicated with triangle.

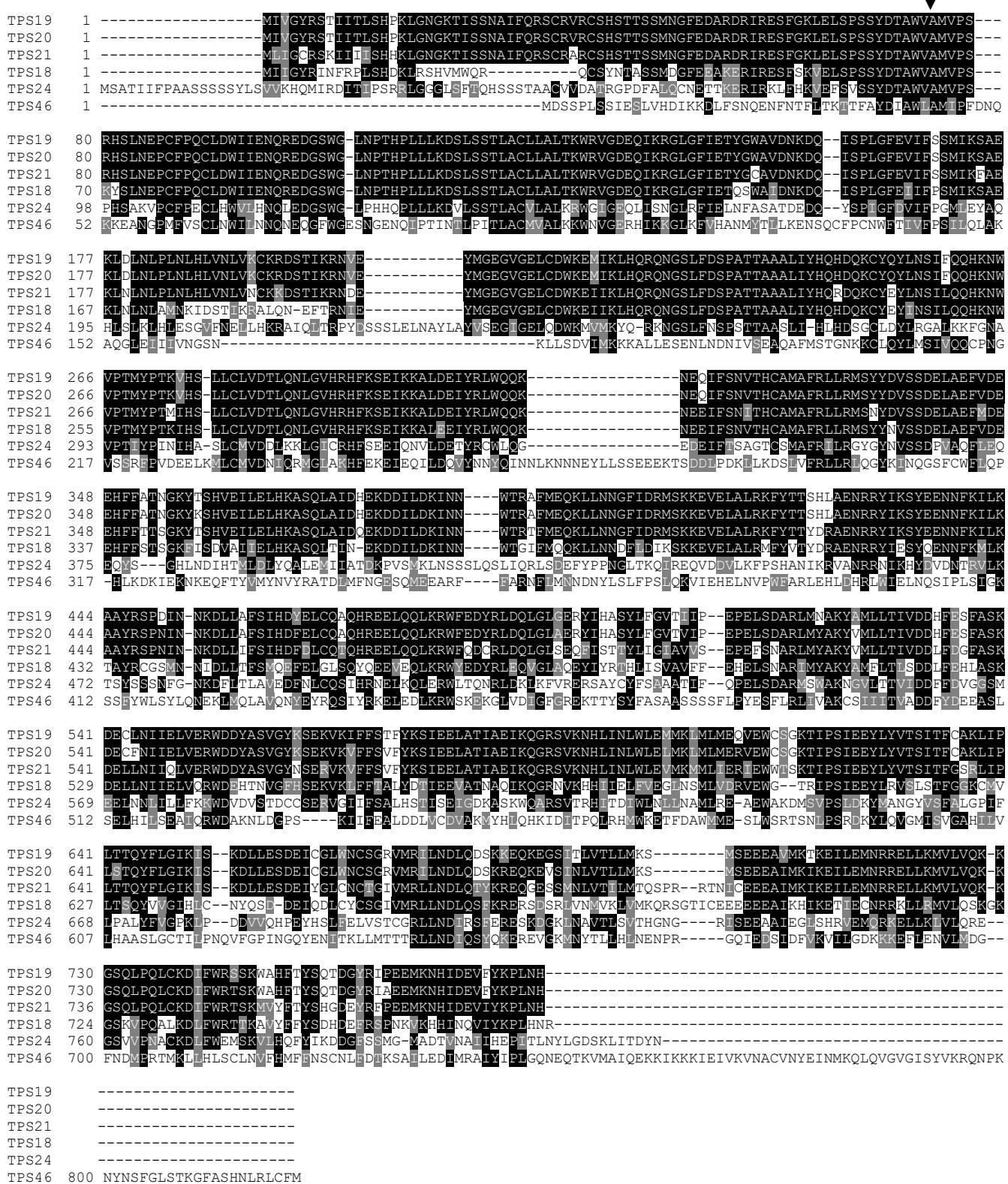
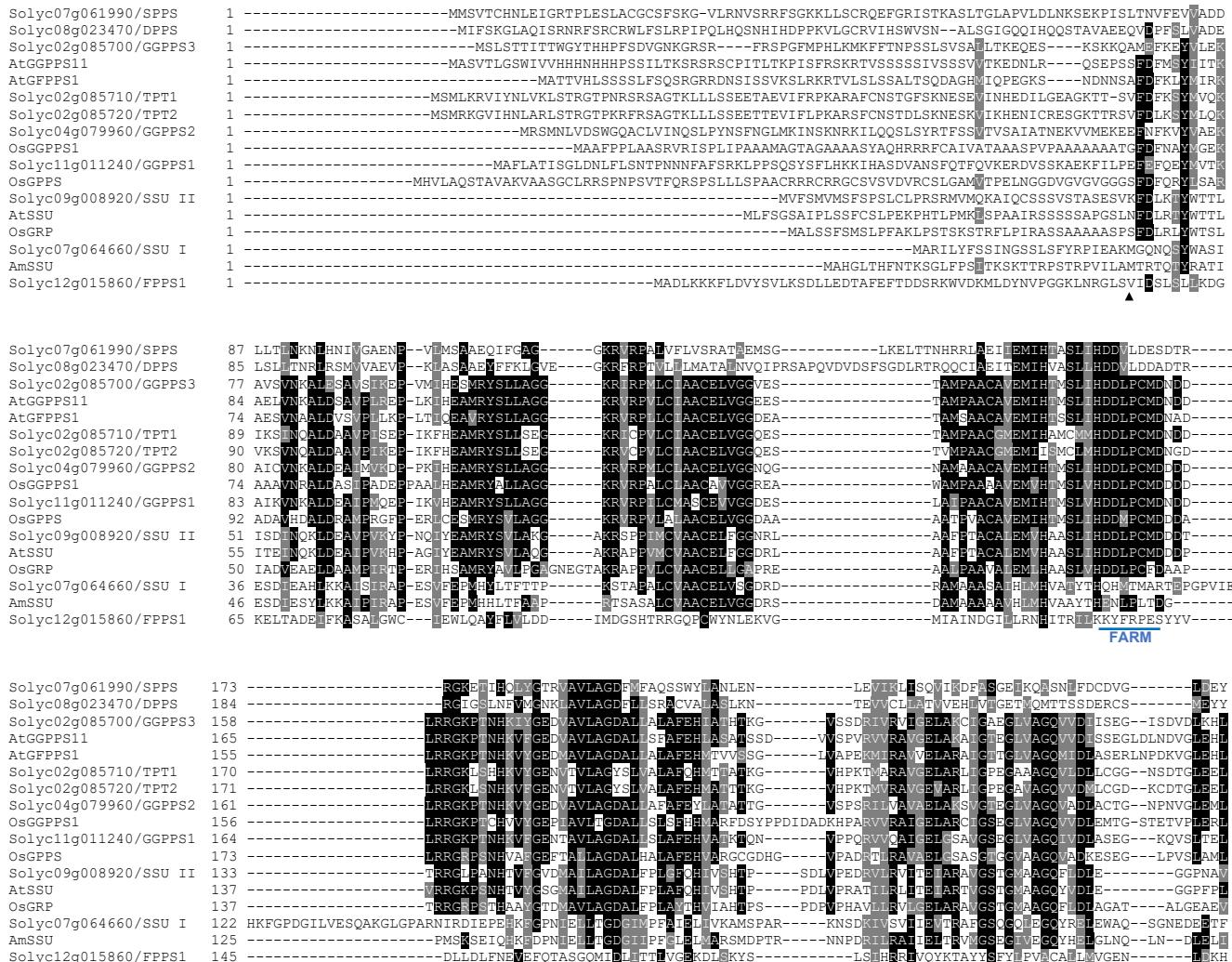


Fig. S15 Sequence analysis of TPT homologs from tomato and other plants. Black shading indicates identical amino acid residues and grey shading are the conserved or semi-conserved substitutions observed and dashes indicate gaps. The two aspartate-rich motifs (FARM, first aspartate-rich motif; SARM, second aspartate-rich motif) are underlined, the black arrow indicates the site of truncation of the N-termini for protein expression in this study. AtGGPPS11, geranylgeranyl diphosphate synthase 11 from *Arabidopsis thaliana* (At4g36810); AtGFPPS1, geranyl farnesyl diphosphate synthase 1 (At1g49530); OsGGPPS1, geranylgeranyl diphosphate synthase 1 from *Oryza sativa* (Os07g39270); OsGPPS, geranyl diphosphate synthase (Os01g14630); AtSSU, small subunit of geranyl(geranyl) diphosphate synthase (At4g38460); OsGRP, GGPPS recruiting protein (Os02g44780); AmSSU, small subunit of geranyl diphosphate synthase from *Antirrhinum majus* (AAS82859).



		LLKSYYTKASDIASTIKGKFIAFSEVGSDFISE	DQYGRNIGISFCIVDDILDFQCSAQAGLKGPA	SLAKGNITAVFELKEPNLRNIESEFHADGSLEA	EAINLVKS
SolyC07g061990/SPPS	243	LLKSYYTKASDIASTIKGKFIAFSEVGSDFISE	DQYGRNIGISFCIVDDILDFQCSAQAGLKGPA	SLAKGNITAVFELKEPNLRNIESEFHADGSLEA	EAINLVKS
SolyC08g023470/DPPS	254	MOKTYYTKASLISNSCKAALIAGHGSASFVSI	AFLYGRNIGISFCIVDDILDFQCSAQAGLKGPA	SLAKGNITAVFELKEPNLRNIESEFHADGSLEA	EAINLVKS
SolyC02g085700/GGPPS3	239	EFIHLHKTAAILLEASVILGAI	GCGGSDDEIEIRLKRKARCGILLFQVWDDILDV	TKSQQLGKTAGKDIAVDKTYPKLGIE	
AtGGPPS11	250	EFIHLHKTAAILLEASVILGAI	GCGGSDDEIEIRLKRKARCGILLFQVWDDILDV	TKSQQLGKTAGKDIAVDKTYPKLGIE	
AtGFPSS1	239	EFIHLHKTAAILLEASVILGAI	GCGGSDDEIEIRLKRKARCGILLFQVWDDILDV	TKSQQLGKTAGKDIAVDKTYPKLGIE	
SolyC02g085710/TPT1	251	EYIHLHKTAADFACAAAWVGAMIGGASEK	IINLKEPKSCIGLGLFQVWDDILDVTKSSE	ELGKTAGKDIAVDKTYPKLGIE	
SolyC02g085720/TPT2	252	MVIIHSRHTADFTEEAAAWVGAMIGGASEK	IINLKEPKSCIGLGLFQVWDDILDVTKSSE	ELGKTAGKDIAVDKTYPKLGIE	
SolyC04g079960/GGPPS2	242	EFIHLHKTAAILLEASVILGAI	GCGGSDDEIEIRLKRKARCGILLFQVWDDILDV	TKSSEELGKTAGKDIAVDKTYPKLGIE	
OsGGPPS1	245	EYIHLHKTAAILLEASVILGAI	GCGGSDDEIEIRLKRKARCGILLFQVWDDILDV	TKSSEELGKTAGKDIAVDKTYPKLGIE	
SolyC11g011240/GGPPS1	244	EYIHLHKTAAILLEASVILGAI	GCGGSDDEIEIRLKRKARCGILLFQVWDDILDV	TKSSEELGKTAGKDIAVDKTYPKLGIE	
OsGGPS	255	EYIHLHKTAAILLEASVILGAI	GCGGSDDEIEIRLKRKARCGILLFQVWDDILDV	TKSSEELGKTAGKDIAVDKTYPKLGIE	
SolyC09g008920/SSU II	210	DFVQEKKYGEPOS	AVCGAILGASDREIQMHRKYGRAVGL	HRVWDDILPARKTE	--NKTGCKKKKGKSYVSMYGP
AtSSU	214	SEFQKQFKFGMGCSA	WCAGLGLGQFQVWDIFDEDRK	--YLGCAEKG-	
OsGRP	219	MVKLTKEFGBMACCSA	ACAGMGLGGAGPDDEAALRKYGRIVGWL	YLVDIRSAE	-GNGKMRNSA
SolyC07g064660/SSU I	225	EYVCKKREGEI	HACGATCGAIALDNDREIEKLRLN	YGLVGLIQCIIIN	-GRNYNK-
AmSSU	205	EYVCKKREGEI	HACGATCGAIALGCGDEDKIEKLRLRFGLY	CVGLL	-KNRS
SolyC12g015860/FPPS1	215	WDVKKKILIDMGY	FYFQVQDDYDFCFEP	EVGLKIGTDIQDFKCS	TVVKAELCLNEEQKKLFENYGDNAACIAKI
ARM					
SolyC07g061990/SPPS	353	CGGIQRAQDIAK	KADLAMQNKLCLPSS-----	FQIALEEIVYVNLERIE	-----
SolyC08g023470/DPPS	364	SRGIQRFER	EARKHASLASAID	DSLPESDDEEVQRSSRAL	EDTHRITRTRK
SolyC02g085700/GGPPS3	321	-----	KSRDEAEPEINKEA	KACLVGFQK	-----
AtGGPPS11	332	-----	KSRDEAEKLNREB	ADQLGFDPSDVR	-----
AtGFPSS1	321	-----	KSRDEAEKLNREB	ADQLGFDPSDVR	-----
SolyC02g085710/TPT1	353	-----	KSRDEAQKUNKEA	KEQLVGFDPERS	-----
SolyC02g085720/TPT2	334	-----	KSRDEAQKULSKB	AEQCLVGFGRAPERKA	-----
SolyC04g079960/GGPPS2	324	-----	KSRDEAEKLNREB	ADQLGFDPSDVR	-----
OsGGPPS1	327	-----	KSRDEAEKULSDAB	BOLSGCFQDPT	-----
SolyC11g011240/GGPPS1	326	-----	KSRDEAEKULSDAB	BOLSGCFQDPT	-----
OsGGPS	337	-----	KSRDEA	AAQLLLAMAEEAED	LGFDAAERA-----
SolyC09g008920/SSU II	287	-----	KAVKVAEDILRACA	FEELDGLKYG	-----
AtSSU	279	-----	MMEAAPEEL	KEAKELLOVFINKY	EGG-DTLVFLYVYAHRRHFLLP
OsGRP	291	-----	RALGIVEELK	KAQAA	NEADRFRGDKY
SolyC07g064660/SSU I	279	-----	KMEKRVNELLRI	LAKEMLNTSF	KGK-----
AmSSU	257	-----	FGFRGRIKEL	KEKLAKEVKE	RSFGEGERI-----
SolyC12g015860/FPPS1	297	-----	IKL	PEVFILEY	KEKTSYKE
					TTTSIAAHF-SKAVQVLSLFLGKLYKRQK

Fig. S16 GC-MS analysis of terpenes extracted from tomato leaves. No significant differences between terpenes were observed from three extraction methods, data present in Fig. 10 was obtained by using MTBE dipping method as described in Methods S1. Terpene signals are identified by comparison of their mass spectra and retention indices with authentic standards and NIST libraries. Monoterpenes and sesquiterpenes are indicated with red and blue dotted lines, respectively. Non-terpene compounds are indicated with asterisks. Terpenes identified from left to right are as follows: α -thujene, α -pinene, unknown 5, 1,3,5-cycloheptatriene, 3,7,7-trimethyl-, β -pinene, isolimonene, β -myrcene, (+)-2-carene, α -phellandrene, α -terpinene, m-cymene, β -phellandrene, β -ocimene, α -ocimene, γ -terpinene, terpinolene, unknown 6, *trans*-*p*-mentha-2,8-dienol, α -terpineol, nerol, geraniol, δ -elemene (germacrene C), β -elemene (germacrene A), β -caryophyllene, α -humulene, germacrene D, cis-muurola-3,5-diene, α -farnesene, E-nerolidol. Mass spectra and retention indices of the terpene compounds can be found in Fig. S3 and Table S2, respectively.

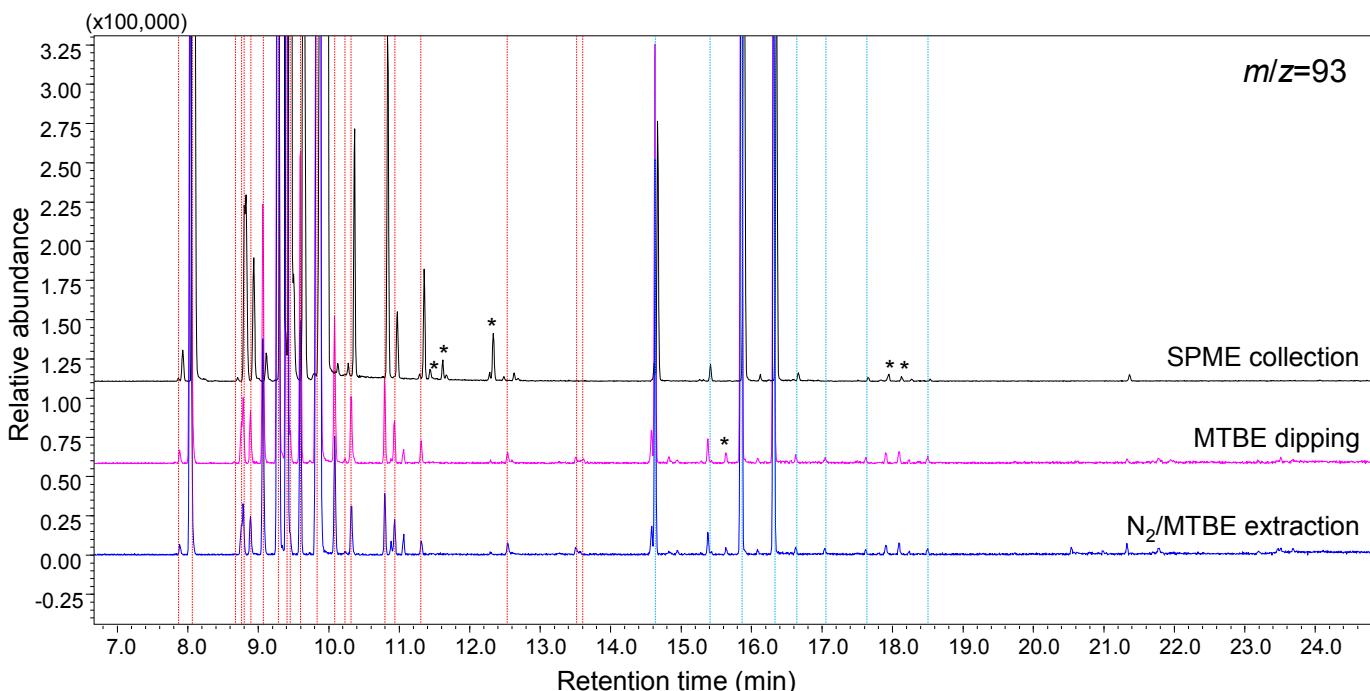


Fig. S17 Correlation analysis of terpene biosynthesis genes and volatile terpenes from tomato.

The Spearman correlation coefficients were calculated using gene expression data and terpene profiling data as indicated in Fig. 9 and Fig. 10, respectively. Genes characterized in this work are shown in bold. The hierarchical clustering was performed using average linkage method by Cluster 3.0.

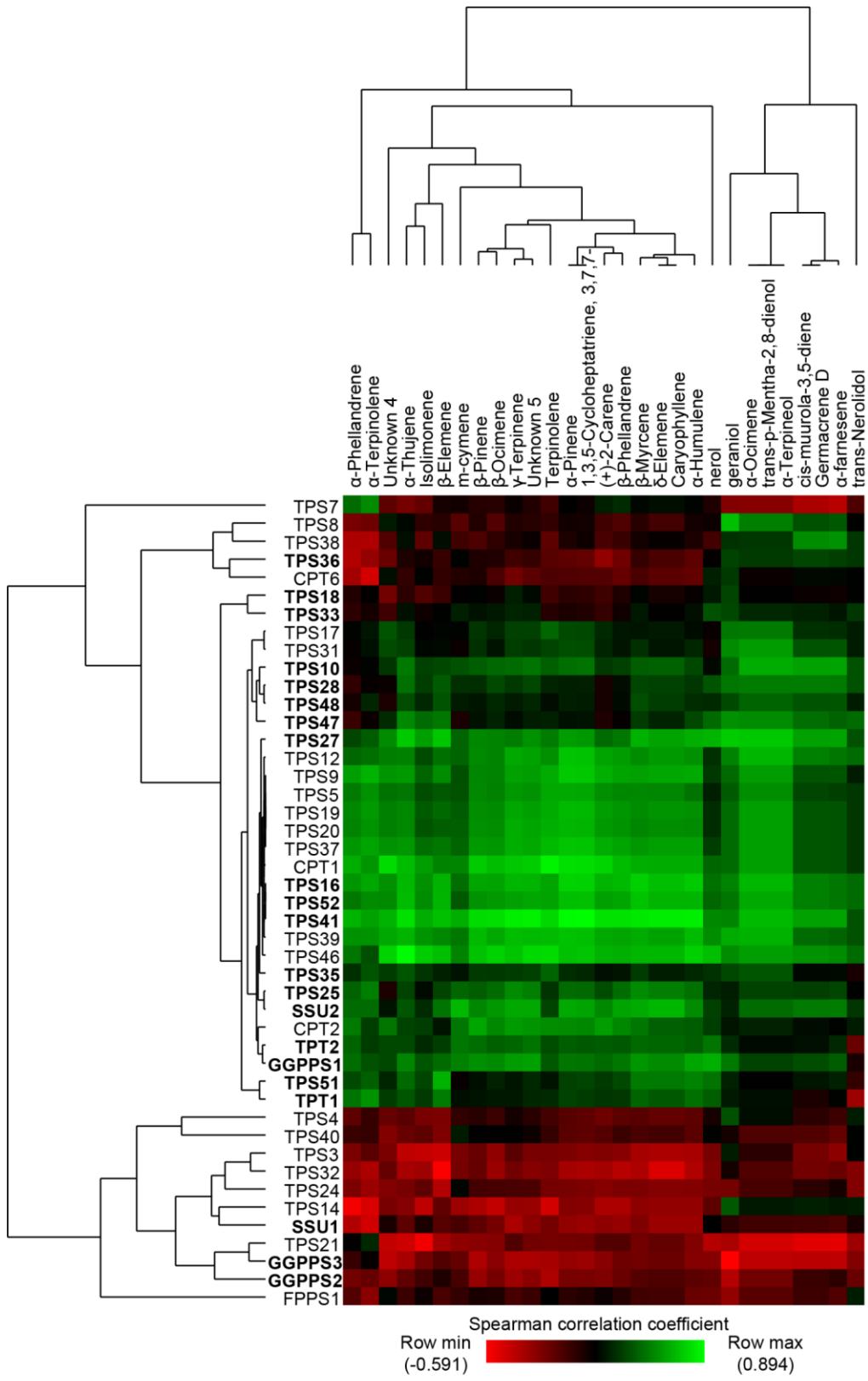
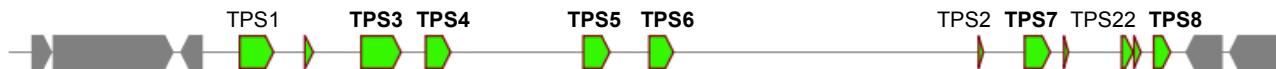


Fig. S18 Terpene biosynthesis gene clusters identified from tomato genome by plantiSMASH.
 The functional TPS genes are shown in bold. The eight clusters (7, 10, 17, 18, 26, 36, 42 and 43; numbered in the analysis of plant secondary metabolite clusters for *Solanum lycopersicum*) are obtained from plantiSMASH website (http://plantismash.secondarymetabolites.org/precalc/precalc/Solanum_lycopersicum/).

SL2.50ch01 - Gene Cluster 7. Type = terpene. Location: 93839435 - 93979803 nt. Click on genes for more information.
 Show pHMM detection rules used



SL2.50ch02 - Gene Cluster 36. Type = terpene. Location: 44246088 - 44308948 nt. Click on genes for more information.
 Show pHMM detection rules used



SL2.50ch06 - Gene Cluster 42. Type = terpene. Location: 37803946 - 38001175 nt. Click on genes for more information.
 Show pHMM detection rules used



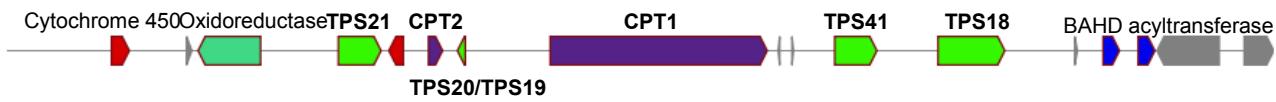
SL2.50ch06 - Gene Cluster 43. Type = terpene. Location: 38130615 - 38191506 nt. Click on genes for more information.
 Show pHMM detection rules used



SL2.50ch07 - Gene Cluster 17. Type = lignan-terpene. Location: 60595725 - 60709207 nt. Click on genes for more information.
 Show pHMM detection rules used



SL2.50ch08 - Gene Cluster 26. Type = terpene. Location: 464459 - 617526 nt. Click on genes for more information.
 Show pHMM detection rules used



SL2.50ch09 - Gene Cluster 10. Type = saccharide-terpene. Location: 71529135 - 71576652 nt. Click on genes for more information.
 Show pHMM detection rules used



SL2.50ch12 - Gene Cluster 18. Type = saccharide-terpene. Location: 836959 - 1084067 nt. Click on genes for more information.
 Show pHMM detection rules used



Legend:

Cytochrome 450	Terpene synthase	Glycosyltransferase	BAHD acyltransferase	Epimerase
Dioxygenase	Amino oxidase	Aminotransferase	(Other) Biosynthetic Genes	Other Genes
biosynthetic genes	transport-related genes	regulatory genes	other genes	

Table S1 Primers used in this work.

Primer name ¹	Sequence (5'-3') ²
<i>For cloning of genomic DNA and sequencing</i>	
TPS47-F	CACACTTGATGTACCTCCAATGTAC
TPS47-R	TTATGTAATAGGCTCCATCAAGTTGATTAGG
TPS48-F	GTATTATCACAATATAGCAACAATGAC
TPS48-R	TTACAGCTTGATGGAGTTGACGAGTAAAGC
TPS49-F	ATGGAATACTTAAACAAATATCC
TPS49-R	TAATTGAGACAATATGCATTGAG
TPS50-F	ATGGAGATTAACAATATGAATGTTC
TPS50-R	GTCACATTACATACATAAATACATAATG
TPS51-F	TGTTGGTTCTTTCTTCACC
TPS51-R	TTCATCATTCACTAATAATTCTTG
TPS52-F	ATGGTCGTTGGATTGGTGGTGAAGC
TPS52-R	CAAGCTTTAATCTTGACTATATTC
TPS53-F	CTACTGGGTTCAATTCTAAGTAATTATTACAC
TPS53-R	CTCAAACCTTAATCATTACTCTACCTC
<i>For cloning in pET32b and creating His-tagged recombinant protein</i>	
TPS10-opti-NdeIF	GCTAAT <u>CCATATGGCATCCTCCAGTAGTACG</u>
TPS10-opti-XhoIR-	CCG <u>CTCGAGACTGATGGAAATTTCACGCTC</u>
TPS16-opti-NdeIF	GCTAAT <u>CCATATGCCCGCGTGGGTCCCACAC</u>
TPS16-opti-XhoIR-	CCG <u>CTCGAGGATAATTGGTCGATAAGGATACG</u>
TPS25-NdeIF	GCTAAT <u>CCATATGGAAAGGATTCTACCTATAAACCAAC</u>
TPS25-NotIR-	TTCAC <u>CGGGCCGCTATAATAGACTGTACCAAGTTATAAG</u>
TPS27-NdeIF	GCTAAT <u>CCATATGCGACGAAATGCTAATTACAAGCC</u>
TPS27-XhoIR-	CCG <u>CTCGAGATTGAGAGGAAGTGATTGAACC</u>
TPS47-opti-NdeIF	GCTAAT <u>CCATATGACTACGATTCTACCTATAAGCCC</u>
TPS47-opti-XhoIR-	CCG <u>CTCGAGAGTAATTGGCTCCATAAGATTAATTAAAATC</u>
TPS51-opti-NdeIF	GCTAAT <u>CCATATGGACGGCTCGGGATGAGG</u>
TPS51-opti-XhoIR-	CCG <u>CTCGAGGGTTGTGATAGGATCGATTAAG</u>
TPS52-opti-NdeIF	GCTAAT <u>CCATATGGTGGTAGGATTGGCGCG</u>
TPS52-opti-XhoIR-	CCG <u>CTCGAGTGCCTGATGGTCGATAAGTACC</u>
dGGPPS1-NdeIF	GCTAAT <u>CCATATGCCTGAGTTGAGTTCAAGAAC</u>
GGPPS1-XhoIR-	CCG <u>CTCGAGATTCTGTCGATTGCAATATAAC</u>
dGGPPS2-NdeIF	GCTAAT <u>CCATATGGAAAAAGAAGAACATTAAATTTC</u>
GGPPS2-XhoIR-	CCG <u>CTCGAGATTGACGATTAGCAATGTAATC</u>
dGGPPS3-NdeIF	GCTAAT <u>CCATATGGAGTTAAAGAACATCGTTCTG</u>
GGPPS3-XhoIR-	CCG <u>CTCGAGATTCTCTGTAAGCAATATAATTGC</u>
dTPT1-XbaIF	GCT <u>CTAGATGAGTGTCTTGATTCAAAAGTTAC</u>
TPT1-EcoRIR-	CGGAATT <u>CCATTGGCCGATGAAGAACG</u>

dTPT2-XbaIF	GCT <u>CTAGATGAGTGTCTTGATTGAAAAGTTAC</u>
TPT2-EcoRIR-	CGGA <u>ATTCCATTGGCCGATGAAGAAG</u>
<i>To create non-tagged protein in pET32b and co-express with pET28a-SSU I/SSU II</i>	
GGPPS1-XhoIR+ (with dGGPPS1-NdeIF)	CCG <u>CTCGAGTCAATTCTGTCGATTGCAATATAAAC</u>
GGPPS2-XhoIR+ (with dGGPPS2-NdeIF)	CCG <u>CTCGAGTTAATTGACGATTAGCAATGTAATC</u>
GGPPS3-XhoIR+ (with dGGPPS3-NdeIF)	CCG <u>CTCGAGTTAATTCTCTGTAAGCAATATAATTGC</u>
<i>For cloning in pET28a and creating His-tagged recombinant protein</i>	
TPS28-NcoIF	CATGCC <u>CATGGTTCGCCAGAGGCTAATTCTCTC</u>
TPS28-XhoIR-	CCG <u>CTCGAGCATATTGATAGGATTACCGAGTAAAG</u>
TPS33-NcoIF	CATGCC <u>CATGGTTCGCCCTGTTGCTGATTCTCTCC</u>
TPS33-XhoIR-	CCG <u>CTCGAGAATCTAACACAGAACATCACCAACAAGGC</u>
TPS35-NcoIF	CATGCC <u>CATGGGCCCTGTTGCTGACTTCTCTCC</u>
TPS35-XhoIR-	CCG <u>CTCGAGAAATTCAATAGAGTCCACCAACAAACGC</u>
TPS36-NheIF	TTCAC <u>GCTAGCATGGATGTTAACTCTTCACGTC</u>
TPS36-NotIR-	TTCAC <u>GCGGCCGCTTATGATCGCTTATAGTAAC</u>
TPS48-NcoIF	CATGCC <u>CATGGACCCTAGAGGCTAATTCTCCTCTCAG</u>
TPS48-XhoIR-	CCG <u>CTCGAGCAGCTTGTGGAGTTGACGAGTAAAG</u>
dSSUI-NdeIF	GCTAAT <u>CCATATGGGCCAAATCAATCTTATTG</u>
SSUI-EcoRIR-	CGGA <u>ATTCAAGACTCTCAACAAAGACTAGC</u>
dSSUII-NdeIF	GCTAAT <u>CCATATGACAGCATCTGAGTCCGTCAAG</u>
SSUII-EcoRIR-	CGGA <u>ATTCACTTGGCCATCAATGCTAAAC</u>
<i>For cloning in pEZS-NL and subcellular localization analysis</i>	
TPS3-XhoIF	CCG <u>CTCGAGATGTCAATATTCTCCACAAGATATTAG</u>
TPS3-BamHIR-	CGGG <u>ATCCGACGCCATAAGGATTAATTATTCCATTG</u>
TPS4-EcoRIF	CGGA <u>ATTCACTGAAGGCAATTATTGAACACATAGG</u>
TPS4-BamHIR-	CGGG <u>ATCCGCATAGTAGGTTGTAATTCTGAACG</u>
TPS5-XhoIF	CCG <u>CTCGAGATGGTTCAATTGAGTAACATAGGG</u>
TPS5-BamHIR-	CGGG <u>ATCCGAGGCTTGTAAATTCCCTGAACGCC</u>
TPS7-EcoRIF	CGGA <u>ATTCACTGGTTCAATTTCAGTAACGCAGGG</u>
TPS7-BamHIR-	CGGG <u>ATCCGAGGCTTGTAAATTCCCCGAACGCC</u>
TPS8-XhoIF	CCG <u>CTCGAGATGTATAAACTGGAAATGACAATGTC</u>
TPS8-BamHIR-	CGGG <u>ATCCCGTTCTCGTCTGAATAATGATTCTCAGTG</u>
TPS9-XhoIF	CCG <u>CTCGAGATGGCTGCTTCTGCTGATAAGTGT</u>
TPS9-BamHIR-	CGGG <u>ATCCGCAA AAA TGT TTT GAA TGG ATG TTT C</u>
TPS10-EcoRIF	CGGA <u>ATTCACTGGCTTCTTCTTCTTACTAATAAG</u>
TPS10-BamHIR-	CGGG <u>ATCCGCGCGTCGATCAACACAAAGCTTTG</u>
TPS12-XhoIF	CCG <u>CTCGAGATGGCTTCTTCTGCTAATAAGTGT</u>

TPS12-BamHIR-	CGGGATCCCGAAAAATGTTGAATGGATGTTTC
TPS16-EcoRIF	CGGAATTCATGGAGCTGTGCACACAAACCGTTC
TPS16-BamHIR-	CGGGATCCCGTAACTCTCAATCTCATGTTCAAAATG
TPS17-XhoIF	CCGCTCGAGATGGAGTTGTGCACACAAACCGTTGC
TPS17-BamHIR-	CGGGATCCCGGATTCAATCTCACTTGAAAATG
TPS18-EcoRIF	CGGAATTCATGATAATTGGCTATAGAATCAATTTC
TPS18-BamHIR-	CGGGATCCAAGGTAGGGTTAGTCCCCAAGATC
TPS19-XhoIF	CCGCTCGAGATGATAGTTGGCTATAGAAGCACAATC
TPS19-BamHIR-	CGGGATCCCGAGAAAGTGAGTCCTTAGAAGCAATGG
TPS20-XhoIF	CCGCTCGAGATGATAGTTGGCTATAGAAGCACAATC
TPS20-BamHIR-	CGGGATCCCGAGAAAGTGAGTCCTTAGAAGCAATGG
TPS21-XhoIF	CCGCTCGAGATGCTAATTGGCTGTAGAAGCAAAATC
TPS21-BamHIR-	CGGGATCCCGAGAAAGTGAGTCCTTAGAAGCAATGG
TPS24-XhoIF	CCGCTCGAGATGTCTGCTACCATTATTTCTG
TPS24-BamHIR-	CGGGATCCCAGTCTCAAGTTGATTATGCAACACCC
TPS25-EcoRIF	CGGAATTCATGGCTTGCTAAACATGGTGT
TPS25-BamHIR-	CGGGATCCTTGTATCACACCTCAAAGTAAGCAC
TPS27-EcoRIF	CGGAATTGACAAGTGAGCAGCAATCTG
TPS27-BamHIR-	CGGGATCCCGTAGAACACCATGATTCCTTGTC
TPS28-EcoRIF	CGGAATTGAGTTATTGGAAGGAAATGTAAACC
TPS28-BamHIR-	CGGGATCCCGCTTGCTTAGTTGATCTCAATCTCC
TPS31-XhoIF	CCGCTCGAGATGGCCCCAGCTGCTGCATTGATGAG
TPS31-BamHIR-	CGGGATCCCGTCAACATGTCGCTATTGTTTTTC
TPS32-XhoIF	CCGCTCGAGATGCCCTACTTAACAACCAAGATGAG
TPS32-BamHIR-	CGGGATCCCGCCAACATGTCCTCTATTGTTTTTC
TPS33-EcoRIF	CGGAATTGAGCTCAGCTGCTGCATTGATG
TPS33-BamHIR-	CGGGATCCCGGTCTCTATTGTGTCTAAAATG
TPS35-EcoRIF	CGGAATTGAGCTCAGCTGCTGCATTGGTG
TPS35-BamHIR-	CGGGATCCCGTTGAAATTGATCCAACACATCG
TPS37-XhoIF	CCGCTCGAGATGGCAAATATTACTAAAGCTTCTCCC
TPS37-BamHIR-	CGGGATCCCG GCGATTAGCTCGTTGAAGATAAGAGG
TPS38-XhoIF	CCGCTCGAGATGCTCAAAGTTGCATATCAACTATG
TPS38-BamHIR-	CGGGATCCGAATTCTAAAGCTGTATCATACAACTC
TPS39-XhoIF	CCGCTCGAGATGGAATGACTAAAGCTTGATTCCC
TPS39-BamHIR-	CGGGATCCCGTACGTGGAGGTTGCTCTGAGAAGATC
TPS40-XhoIF	CCGCTCGAGATGTCGATCTCAGCTTCTTTAAG
TPS40-BamHIR-	CGGGATCCCGCATCGAATAGCATCGATATGGTC
TPS41-SalIF	ACGCGTCGACATGCTGTAATTACATCCAATTAC
TPS41-BamHIR-	CGGGATCCAACGATGAATATCCTTCTTCTACTTG
TPS46-XhoIF	CCGCTCGAGATGGATTCTCACCATGCTTCTATTG

TPS46-BamHIR-	CGGGATCCCGTACCATGCATGCAAGAGTAATAGGGAG
TPS47-XhoIF	CCG <u>CTCGAG</u> ATGGCTTGCAGATGCATATTATCC
TPS47-BamHIR-	CGGGATCCCGTGGAGTATCACTCTTGAAACGTG
TPS48-XhoIF	CCG <u>CTCGAG</u> ATGACTTCACTGGCTGGAAATG
TPS48-BamHIR-	CGGGATCCC CGTCATCTGATTAATCCGAGTAG
TPS51-XhoIF	CCG <u>CTCGAG</u> ATGGATGGATTGGT GATGAGGCAGAG
TPS51-BamHIR-	CGGGATCCC GTTATAGATTTCACTCAATGATT CTC
TPS52-XhoIF	CCG <u>CTCGAG</u> ATGGCTGTTGGATTGGTGAAGCGGAG
TPS52-BamHIR-	CGGGATCCC GATTATAGATTTCACTCAATGATT CTC
FPPS1-XhoIF	CCG <u>CTCGAG</u> ATGGCTGATCTGAAGAAGAAATTTTG
FPPS1-BamHIR-	CGGGATCCC GCTCTGCCTCTATAAAATCTTCCAAG
GGPPS1-EcoRIF	CGGAATT CATGGCATTTTAGCTACCATTCTGG
GGPPS1-BamHIR-	CGGGATCCC ATTCTGTCGATTGCAATATAACTAG
GGPPS2-XhoIF	CCG <u>CTCGAG</u> ATGAGATCTATGAACCTGTTGATT
GGPPS2-SalIR-	ACGCG <u>TGACT</u> GATTTGACGATTAGCAATGTAATCTGC
GGPPS3-EcoRIF	CGGAATT CATGAGTCTTCAACACAATTACAAC
GGPPS3-BamHIR-	CGGGATCCC GATTCTCTCTGTAAGCAATATAATTGC
SSU I-EcoRIF	CGGAATT CATGGCAAGAATT TATATT TT CCTCC
SSU I-BamHIR-	CGGGATCCC GAGACTCTCAACAAAGACTAGCAATTG
SSU II-EcoRIF	CGGAATT CATGGTTTCTCCATGGT GATGAGC
SSU II-BamHIR-	CGGGATCCC GGACTTGGCCATCAATGCTAAACC
TPT1-EcoRIF	CGGAATT CATGGCCCATACTAAGTCAAATAGG
TPT1-BamHIR-	CGGGATCCC ATTTGCCGATGAAGAACGAAATC
TPT2-EcoRIF	CGGAATT CATGTCTATGCGAAAAGGTGTAATC
TPT2-BamHIR-	CGGGATCCC ATTTGCCGATGAAGAACGAAATC
<i>For cloning in pEAQHT and tobacco transient expression</i>	
TPS10-NruIF	CTAG <u>TCGCGA</u> ATGGCTTCTTCTTCTACTAATAAG
TPS10-XmaIR	TCCCC <u>CCGGT</u> TATTGATATT TGACAGATTCAATCAG
TPS16-NruIF	CTAG <u>TCGCGA</u> ATGGAGCTGTGCACACAAACCG
TPS16-XhoIR	CCG <u>CTCGAG</u> CTATATAATAGGATCAATAAGTATCC
TPS18-NruIF	CTAG <u>TCGCGA</u> TGATAATTGGCTATAGAATCAATTTC
dTPS18-NruIF	CTAG <u>TCGCGA</u> TGGTCCCTCAAAATATTCACTAAATG
TPS18-XmaIR	TCCCC <u>CCGGT</u> CATCGATTATGGAGTGGTTGTAATGAC
TPS41-NruIF	CTAG <u>TCGCGA</u> ATGCTGGTAATTACATCCAATTAC
dTPS41-NruIF	CTAG <u>TCGCGA</u> ATGAGGAATTGAGAGAACGAGATA
TPS41-XmaIR	TCCCC <u>CCGGGG</u> CTTCTGAAAAAGAACTTGG
CPT1-NruIF	CTAG <u>TCGCGA</u> ATGAGTTCTTGGTTCTCAATGTTG
CPT1-XmaIR	TCCCC <u>CCGGG</u> ATATGTGTGTCACCAAAACGTCTATG
CPT2-NruIF	CTAG <u>TCGCGA</u> ACTCTCAATAGTGTCTCAAC
dCPT2-NruIF	CTAG <u>TCGCGA</u> GTGCAAATTCTAAAGCTGCCTCAACATGTC

CPT2-XmaIR	TCCCCCCC <u>GGGT</u> CAATATGTGTGTCCACCAAAACGTC
CPT6-NruIF	CTAG <u>TCGCGA</u> TGA <u>ACT</u> CTTGT <u>TTGAGGGAGGCC</u>
CPT6-XmaIR	TCCCCCCC <u>GGGT</u> CAATATGTGTGTCCACCAAAACGTC <u>TATG</u>
CPS-NruIF	CTAG <u>TCGCGA</u> AT <u>GTCGAT</u> CTCAG <u>CTTCTTTTAAG</u>
dCPS-NruIF	CTAG <u>TCGCGA</u> TGG <u>GGTTCGTTGCCGGT</u> TATCAAGTGG
CPS-XmaIR	TCCCCCCC <u>GGGAA</u> CTACT <u>CTTCAA</u> ATAGCA <u>CTTGGC</u>
KS-NruIF	CTAG <u>TCGCGA</u> AT <u>GTCGAT</u> CT <u>ACCATT</u> AT <u>TTTCTGCC</u>
dKS-NruIF	CTAG <u>TCGCGA</u> T <u>GCGAGGACCGG</u> AT <u>TTGCTCTCAATG</u>
KS-XmaIR	TCCCCCCC <u>GGGATT</u> GTAA <u>TCAGTA</u> ATTAG <u>TTTGAATCCC</u>
GGPS1-NruIF	CTAG <u>TCGCGA</u> AT <u>GAGTCTTCAACA</u> ACAATT <u>ACAC</u>
dGGPS1-NruIF	CTAG <u>TCGCGA</u> T <u>GGAGTTAAAGA</u> ATAC <u>GTCTTGAAAAG</u>
GGPS1-XmaIR	TCCCCCCC <u>GGGATT</u> CT <u>CTGTAA</u> AG <u>CAATAATTG</u>
dTPT1-AgeIF	CCG <u>ACCGGT</u> AT <u>GACAG</u> TT <u>TTCCAAGA</u> AT <u>GAAAG</u>
TPT1-XmaIR	TCCCCCCC <u>GGGTTT</u> GC <u>GATGAAGAACGAAATC</u>
dTPT2-AgeIF	CCG <u>ACCGGT</u> AT <u>GACAG</u> AT <u>CTTCCAAGA</u> AT <u>GAATC</u>
TPT2-XmaIR	TCCCCCCC <u>GGGTTT</u> GC <u>GATGAAGAACGAAATC</u>
dAtTPS19-NruIF	CTAG <u>TCGCGA</u> T <u>GCGTCCTCGACGT</u> ACT <u>TTTC</u>
AtTPS19-XmaIR	TCCCCCCC <u>GGGT</u> GA <u>AAGGGG</u> TAT <u>GGGTGGACGTAC</u>
dAtGFPS2-NruIF	CTAG <u>TCGCGA</u> AT <u>GATTCCACCAGAGGGAAAATGCAATG</u>
AtGFPPS2-XmaIR	TCCCCCCC <u>GGGTTG</u> T <u>GTCTGCAAGCGATGTAG</u>

For qRT-PCR

TPS3-QF	GTTCATGGGTGTGCAGTAAAT
TPS3-QR	TAGGGCATGGAAATAGTGTATGG
TPS4-QF	TGCTGTTCAAAGATGGATACA
TPS4-QR	GTAGGGTAGGACGTTGATGTTG
TPS5-QF	GAGGTGACGTTCCAATCAATAC
TPS5-QR	GCTTCCC <u>ATGTCTCCTGTATC</u>
TPS7-QF	AGCAGCAC <u>CCAACAAGA</u>
TPS7-QR	GCTAAT <u>CCCAC</u> TGACCAGAAA
TPS8-QF	ACCTAGCAAGAAT <u>GGCACAA</u>
TPS8-QR	CCTAGAGGAAC <u>GGATTCAAAGAG</u>
TPS9-QF	ATCCAGAGAT <u>GGGATGCTAATG</u>
TPS9-QR	CACGGTCC <u>AGTTACCTTCTT</u>
TPS10-QF	GGTGGGAG <u>CAAGGAAATACATATC</u>
TPS10-QR	AAGTAAC <u>ACTCAACCAGTCTGTCC</u>
TPS12-QF	GAGACAG <u>GGTTGGT</u> GAGTGT <u>TA</u>
TPS12-QR	CATCAAAGAAG <u>GGAGGCCATTG</u>
TPS14-QF	GATTGTT <u>GGGGCGAATT</u> TAT
TPS14-QR	GCTCT <u>CTTGTGTTCGTGTTC</u>
TPS16-QF	CAACGACGT <u>GCATGGAATGTTAAG</u>

TPS16-QR	TTCCTTGCTTCTACCCCTACAATAC
TPS17-QF	CCACCAATAGTTGTTGCTTCTTG
TPS17-QR	GCTTCTTCCTTGTAAACGCTATATT
TPS18-QF	GTAATAGAAGAAAGTTGCTAAGGATGG
TPS18-QR	ATGACTTGATTGATATGATGCTTGAC
TPS19-QF	TCTTGAACCTCACAAAGCATCAC
TPS19-QR	CAAAGCAAGTTCCACCTCCTCT
TPS20-QF	TTCCCACATGTATCCAACAAAGG
TPS20-QR	TGTTGCCATAGCCTGTATATTTCATC
TPS21-QF	AACTCAAGAGGTGGTTCAAGATTG
TPS21-QR	CTCAGGTTCAGACACAAACAGCTATG
TPS24-QF	AGTTTCACCCAGCATTCCAGTTC
TPS24-QR	CAACCCAGGCAGTATCGTATGAAG
TPS25-QF	ATGTTTCAGAGGATGTAGCTCG
TPS25-QR	GACCCCAAATCCATCTCCAG
TPS27-QF	CATGTTCTTCTGGTTGGCTCAAG
TPS27-QR	GATGAAGACACGTACCTCTTCC
TPS28-QF	CCATGAAGAACAGATGATCTATAACAGG
TPS28-QR	CTAACATGTGCTGCTTCATACAAAC
TPS31-QF	GAAGTATCAAGGTGGTGGAAAGA
TPS31-QR	GCCTGAGAGTATTGAGGTTAG
TPS32-QF	TTTGGACGATGGGAGTGTATG
TPS32-QR	CGGTGTAAACCTCCAGTTCTT
TPS33-QF	TAGGGATAGAGCAGTGGAGTGTAC
TPS33-QR	ACCTCTGTATGGCATCGGTATAAAC
TPS35-QF	TCATGGAGAACGATATTTGGAAGAGG
TPS35-QR	TCGTAGATAGAGATGAAGTATCGTGT
TPS36-QF	GGTAAATCAGACCGCGTCTAC
TPS36-QR	GTTGTTCCAAGCGCCATAAAGGTTAG
TPS37-QF	GGGCTTGTAGTTCTGGAGTT
TPS37-QR	GCTATGGAAGTGGACATGAGTG
TPS38-QF	GCTTCCTAGCTCTTCAACTC
TPS38-QR	GTTAGCCCAGTGTGGCTTAGTT
TPS39-QF	CTGGAGTTCATGTGGCTTAGTT
TPS39-QR	ATCATCCCAAAGGCGAAGG
TPS40-QF	GGTAGGAGAACGGAGAACGGAGG
TPS40-QR	GGAGTCGGTGAGAAACTCTATTGG
TPS41-QF	CACTTGGATATGTGATTGTTGATTG
TPS41-QR	GCATGTGACAGACTTATTGGTGTAC
TPS46-QF	AAGGTGGATGCAAAGAACATC

TPS46-QR	CATTTGGCAACATCACCA
TPS47-QF	TTGGCCTCACTGATTACTCGACG
TPS47-QR	CCATGCCAAATATGACCCTTCC
TPS48-QF	ATGGAAGGATTGAACAAAGAGTGC
TPS48-QR	AAAGCTTCGATTACAGCTTCACG
TPS51-QF	ATGGATGGATTGGTGATGAGGCAGAG
TPS51-QR	GATTGGAAGGAGAAAGCACTAGC
TPS52-QF	ATGGTCGTTGGATTGGTGGT
TPS52-QR	GATTGGAAGGAGAAAGCACTAGC
FPPS1-QF	CGC CGG ATT GTC CAG TAT AAA AC
FPPS1-QR	GGT CAG CAA AGC AGT CCA GAT AAT C
GGPPS1-QF	CAGCGGGTAAGGACCTAATAAC
GGPPS1-QR	TGCCTTGATAGTCGAAGTAG
GGPPS2-QF	CGCTACTTGCTTCGCATTC
GGPPS2-QR	CCAGCTACTAACCTTCAGTTC
GGPPS3-QF	GGGATGCACCTCTGCATTAG
GGPPS3-QR	CTACAACCTGACCAGCTACAA
SSU I-QF	GGACAGCTAGAAGGCCAATATC
SSU I-QR	GCTCCACATGCATGAATTCC
SSU II-QF	TCGAGTCCTCCGGGTTATTA
SSU II-QR	CACTCACCCATTCACCATATTTC
TPT1-QF	GCGTCCGAGAAGGAGATTAATAG
TPT1-QR	GCTCAGAGGATTAGTCACATCA
TPT2-QF	TTCGTGCTGTTGGAGAAGTAG
TPT2-QR	GCTCTCCAATCCAGTATCACA
CPT1-QF	CAAAGGGCAACAAAGGACTTCACC
CPT1-QR	CAGGATTGGACACTGCTTCAAG
CPT2-QF	GAACAAAGAACTTGAAAGCAAGTGT
CPT2-QR	CGTCTATGCCCTTGTGAAAGTTAAG
CPT6-QF	TACCACCAAGTGTGCTAAATTCCC
CPT6-QR	ATGAGAGCTTCTCCAAAGTCAG
Ubi-QF	CAGCTCGCTTCTCAGTTCC
Ubi-QR	AGCTCGACAAACAGACTGGAA

¹⁺, has a stop codon in the sequence; -, does not contain a stop codon in the sequence. d, for truncating the N-terminal transit peptide. opti, codon optimized version.

²Restriction sites are underlined.

Table S2 A list of terpene compounds identified from Figs 3-5 and 10.

Identified terpenes ¹	MS similarity to NIST library	Identification method	Retention indices ²	Reference retention indices ³	Source of reference
29. Isoprene	99%	b, c	n.c.	n.r.	Standard
α -Thujene	97%	a, b, c	929.7	929.7	Mandarin oil
α -Pinene	98%	a, b, c	937.7	937.7	Standard
Unknown 5	81% to 3-carene	-	971.1	3-carene RI=1011.0	Standard
1,3,5-Cycloheptatriene, 3,7,7-trimethyl	97%	a, b	977.0	972.6	NIST 2014
β -Pinene	97%	a, b, c	981.6	981.6	Standard
Isolimonene	95%	a, b, c	986.1	986.1	Standard
25. β-Myrcene	93%	a, b, c	990.5	990.5	Standard
(+)-2-Carene	96%	a, b, c	1002.9	1002.9	Standard
α -Phellandrene	97%	a, b, c	1010.5	1010.5	Standard
α -Terpinene	96%	a, b, c	1020.6	1020.6	Standard
<i>m</i> -cymene	97%	a, b, c	1028.1	1028.1	Standard
β -Phellandrene	95%	a, b, c	1037.4	1037.4	SIPHS1 (Schilmiller <i>et al.</i> , 2009)
26. β-<i>cis</i>-Ocimene	94%	a, b, c	1038.8	1038.8	Standard

27. β-trans-Ocimene	99%	a, b, c	1049.3	1049.3	Standard
α -Ocimene	90%	a, b	1057.8	1056	Jordán <i>et al.</i> (2013)
γ -Terpinene	98%	a, b, c	1062.8	1062.8	Standard
Terpinolene	95%	a, b, c	1088.8	1088.8	Standard
Unknown 6	87% to phellandral	-	1096.1	Phellandral RI=1273	Babushok <i>et al.</i> (2011)
28. Linalool	93%	a, b, c	1099.6	1099.6	Standard
<i>trans</i> -p-Mentha-2,8-dienol	95%	a, b	1126.7	1127.5	Babushok <i>et al.</i> (2011)
α -Terpineol	93%	a, b, c	1199.5	1199.5	Standard
nerol	95%	a, b, c	1230.4	1230.4	Standard
geraniol	90%	a, b, c	1257.0	1257.0	Standard
δ -Elemene	95%	a, b	1332.9	1333.8	Zizovic <i>et al.</i> (2007)
12. α-Cedrene	92%	a, b, c	1394.0	1394.0	Standard
4. β-Elemene	96%	a, b, c	1395.5	1395.5	Standard
13. <i>cis</i>-α-Bergamotene	90%	a, b	1419.6	1419.5	NIST 2014
20. (<i>Z</i>)-β-Farnesene	94%	a, b, c	1429.6	1429.6	Standard
β -Caryophyllene	95%	a, b, c	1432.0	1432.0	Standard
6. (<i>E</i>)-β-Farnesene	96%	a, b, c	1454.8	1454.8	Standard
α -Humulene	95%	a, b, c	1468.6	1468.6	Standard

14. β-Acoradiene	91%	a, b	1469.8	1466	Babushok <i>et al.</i> (2011)
15. Unknown 2	89% to β -acoradiene	-	1478.2	β -Acoradiene RI=1466	Babushok <i>et al.</i> (2011)
16. Unknown 3	88% to trans-Sesquisabinene hydrate	-	1483.5	trans-Sesquisabinene hydrate RI=1581	Babushok <i>et al.</i> (2011)
Germacrene D	90%	a, b, c	1493.0	1493.0	Ginger oil
18. <i>cis</i>-muurola-3,5-diene	93% to <i>cis</i> -muurola-3,5-diene 92% to α -cubebene	a, b	1494.0	n.r. α -cubebene RI=1351	Babushok <i>et al.</i> (2011)
1. (<i>E,E</i>)-α-Farnesene	97%	a, b, c	1505.3	1505.3	Standard
7. β-Bisabolene	94%	a, b, c	1513.3	1513.3	Ginger oil
3. Guaia-1(10),11-diene	94%	a, b	1515.3	1515	Novak <i>et al.</i> (2010)
17. β-Curcumene	93%	a, b	1518.6	1516	Kahriman <i>et al.</i> (2011)
22. δ-Cadinene	92%	a, b	1523.1	1524	Novak <i>et al.</i> (2010)
19. Z-Nerolidol	94%	a, b, c	1533.1	1533.1	Standard
8. Unknown 1	84% to α -Bergamotene	-	1533.8	α -Bergamotene RI=1434	Babushok <i>et al.</i> (2011)
9. α-Bisabolene	90%	a, b	1545.5	1541	Babushok <i>et al.</i> (2011)
2. Elemol	96%	a, b	1557.7	1557	Pinheiro <i>et al.</i> (2013)
5. <i>E</i>-Nerolidol	95%	a, b, c	1563.4	1563.4	Standard

10. α-Eudesmol	90%	a, b	1663.2	1664	Watanabe <i>et al.</i> (2005)
11. (Z,Z)-Farnesol	95%	a, b, c	1670.6	1670.6	SICPT6 (Akhtar <i>et al.</i> , 2013)
21. α-Bisabolol	93%	a, b, c	1694.8	1694.8	Standard
23. Unknown 4	85% to α -springene	-	2059.7	α -springene RI=2019	Setzer <i>et al.</i> (2006)
24. ent-Kaurene	95%	a, b, c	2079.5	2079.5	SITPS40 (Falara <i>et al.</i> , 2011)

¹Compounds listed in order of elution from the Rxi-5Sil MS column, the enzymatic terpene products are shown in bold. ²Retention indices relative to C₇-C₄₀ n-alkanes on the Rxi-5Sil MS column. ³Reference retention indices were obtained using the similar nonpolar column.

a, identification based on retention index; b, identification based on comparison with mass spectral data; c, identification based on comparison with authentic compounds; n.c. not calculated; n.r. no reference available from the similar column.

Akhtar TA, Matsuba Y, Schauvinhold I, Yu G, Lees HA, Klein SE, Pichersky E. 2013. The tomato *cis*-prenyltransferase gene family. *Plant J* **73**(4): 640-652.

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Table S3 A list of tomato terpene synthase genes, their characteristics, and the enzymes they encode.

Chromosome	Locus	TPS	TPS-clade	No. of Exons	Full-length cDNA obtained	Protein size (aa)	Subcellular localization	Substrate	Main product
1	Solyc01g101170	31	a	6	Yes	555	Cytosol ^c	<i>E,E</i> -FPP	Viridiflorene ^a
	Solyc01g101180	32	a	6	Yes	548	Cytosol ^c	<i>E,E</i> -FPP <i>Z,Z</i> -FPP	Viridiflorene ^b Unidentified ^b
	Solyc01g101190	33	a	6	Yes	552	Cytosol^c	<i>E,E</i>-FPP <i>Z,Z</i>-FPP	Guaia-1(10),11-diene^c β-Acoradiene^c
	Solyc01g101210	35	a	6	Yes	556	Cytosol^c	<i>E,E</i>-FPP <i>Z,Z</i>-FPP	Guaia-1(10),11-diene^c (<i>Z,Z</i>)-Farnesol^c
	Solyc01g105870	3	b	7	Yes	607	Plastid ^c	GPP	Camphene, tricyclene ^b
	Solyc01g105880	4 (MTS2)	b	7	Yes	590	Plastid ^c	GPP	β-Phellandrene ^d
	Solyc01g105890	5 (MTS1)	b	8	Yes	609	Plastid ^c	GPP	Linalool ^d
	Solyc01g105920	7	b	7	Yes	592	Plastid ^c	GPP	β-Myrcene, limonene ^b
	Solyc01g105960	8	b	7	Yes	597	Cytosol ^c	GPP NPP	1,8-Cineole ^b 1,8-Cineole ^b
2	Solyc02g079840	38	b	7	Yes	546	Cytosol ^c	<i>E,E</i> -FPP	α-Bergamotene ^b
	Solyc02g079890	25	b	7	Yes	547	Cytosol^c	GPP	β-Ocimene^c
	Solyc02g079910	27	b	7	Yes	554	Cytosol^c	<i>E,E</i>-FPP	α-Farnesene^c
	Solyc00g154480								
3	Solyc03g006550	46	e/f	11	Yes	821	Cytosol ^c	GGPP	Geranylinalool ^e
	Solyc03g007730	47	b	7	Yes	562	Cytosol^c	DMAPP	Isoprene^c
4	Solyc04g051620	48	a	8	Yes	560	Cytosol^c	<i>E,E</i>-FPP <i>Z,Z</i>-FPP	Hedycaryol (Elemol)^c Unidentified^c
	Solyc04g054380	28	a	7	Yes	554	Cytosol^c	<i>E,E</i>-FPP <i>Z,Z</i>-FPP	Hedycaryol (Elemol)^c Unidentified^c
6	Solyc06g059885	9 (Sst1)	a	7	Yes	548	Cytosol ^c	<i>E,E</i> -FPP	Germacrene ^f

	Solyc06g059910	10	a	7	Yes	556	Cytosol^c	Z,Z-FPP	α-Bisabolol^c
	Solyc06g059920								
	Solyc06g059930	12 (CAHS)	a	7	Yes	548	Cytosol ^c	<i>E,E</i> -FPP	β -Caryophyllene, α -humulene ^g
	Solyc06g060180	36	a	7	Yes	591	Mitochondria^b	Z,Z-FPP	<i>cis</i>-Muurola-3,5-diene^c
	Solyc06g084240	40 (CPS1)	c	15	Yes	800	Plastid ^c	GGPP	Copaly diphosphate ^h
7	Solyc07g008680	16	a	7	Yes	553	Cytosol^c	<i>E,E</i> -FPP	δ -Cadinene ^c
	Solyc07g008690								
	Solyc07g052120	51	a	7	Yes	550	Cytosol^c	<i>E,E</i> -FPP	<i>E</i> -Nerolidol ^c
	Solyc07g052130							Z,Z-FPP	α -Bisabolol ^c
	Solyc07g052140	52	a	7	Yes	551	Cytosol^c	<i>E,E</i> -FPP	<i>E</i> -Nerolidol ^c
	Solyc07g052150							Z,Z-FPP	α -Bisabolol ^c
	Solyc07g066670	24 (KS)	e/f	15	Yes	820	Plastid ^c	CPP	<i>ent</i> -Kaurene ^b
	Solyc07g066675								
8	Solyc08g005640	21	e/f	14	Yes	784	Plastid ^c	NNPP	Lycosantalene ⁱ
	Solyc08g005665	20 (PHS1)	e/f	14	Yes	778	Plastid ^c	NPP	β -Phellandrene ^j
	Solyc08g005670	19	e/f	14	Yes	778	Plastid ^c	NPP	β -Myrcene, β -ocimene ⁱ
	Solyc08g005710	41	c	15	Yes	780	Mitochondria^c	GGPP	Copaly diphosphate^c
	Solyc08g005720	18	e/f	14	Yes	773	Mitochondria^c	NNPP	Unidentified^c
9	Solyc09g092470	14	a	7	Yes	568	Cytosol ^b	<i>E,E</i> -FPP	β -Bisabolene ^b
								Z,Z-FPP	α -Bisabolene ^b
10	Solyc10g005390	39	g	7	Yes	563	Cytosol ^c	GPP	Linalool ^b
	Solyc10g005410	37	g	7	Yes	560	Cytosol ^c	<i>E,E</i> -FPP	<i>E</i> -Nerolidol ^b
12	Solyc12g006570	17	a	7	Yes	554	Cytosol ^c	<i>E,E</i> -FPP	Valencene ^a

^aBleeker *et al.* (2011). ^bFalara *et al.* (2011). ^cThis work. ^dvan Schie *et al.* (2007). ^eFalare *et al.* (2014). ^fColby *et al.* (1998). ^gSchilmiller *et al.* (2010). ^hBensen and Zeevaart, (1990). ⁱMatsuba *et al.* (2013). ^jSchilmiller *et al.* (2009).

TPS gene numbers are adopted from Falara *et al.* (2011), terpene synthases characterized in this work are shown in bold.

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- Colby SM, Crock J, Dowdle-Rizzo B, Lemaux PG, Croteau R.** 1998. Germacrene C synthase from *Lycopersicon esculentum* cv. VFNT cherry tomato: cDNA isolation, characterization, and bacterial expression of the multiple product sesquiterpene cyclase. *Proceedings of the National Academy of Sciences, USA* **95**(5): 2216-2221.
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Table S4 A list of tomato terpene synthase pseudogenes.

Chromosome	Locus	TPS	No. of Exons	Type of mutation	Putative translated protein (aa)
1	Between Solyc01g101190 and Solyc01g101210	34	(-)3	Deletion of exons	No in frame start codon was found
	Solyc01g101220	30	(-)3(-)1	Deletion of exons	No in frame start codon was found
	Solyc01g105850 Solyc01g105860	1	7	Aberrant splicing, incorrect 3' splice sequence at the end of intron 4, cloned cDNAs skip exon 5 and part of exon 6	400(-)
	Solyc01g105900	6	7	Deletion of about 80nt in exon 3 to cause frame shift	165(-)
	Solyc01g105910	2	3(-)	Deletion of exons and stop codons in frame	131(-)
	Solyc01g105930 Solyc01g105940 Solyc01g105950	22	7	Large insertion in exon 2 and numerous stop codons in frame	52(-)
2	Solyc02g079900 Solyc02g079895	26	(-)4	Deletion of exons and mutation of 3' splice site	No in frame start codon was found
3	Solyc03g112175	49	(-)2	Only the last two exons were found.	(-)92
4	Between Solyc04g005100 and Solyc04g005120	50	2(-)2	Deletion of exons	51(-)
5	Solyc05g026590 Solyc05g026600	43	7	Insertion of an extra T in first exon to cause frame shift	30(-)
6	Between Solyc06g059885 and Solyc06g059880	23	(-)1	Deletion of exons	No in frame start codon was found
	Part of Solyc06g059930	11	2(-)2(-)	Deletion of exons	50(-)
	Solyc06g060010 Solyc06g060013	13	7	Insertion in exon 2 to cause frame shift	156(-)
7	Solyc07g051940	15	7	The end of the first exon has a deletion including	29(-).

				the splice site, so the intron is not spliced correctly, cloned cDNA has stop codon in frame	
9	Solyc09g065230	44	(-3(-)	Deletion of exons	No in frame start codon was found
10	Solyc10g005420	42	(-4(-)	Deletion of exons to cause frame shift	15(-)
	Solyc10g052745	53	(-2	Deletion of exons	No in frame start codon was found
12	Solyc12g019240	29	(-1(g)2(g)	Deletion of exons	No in frame start codon was found

TPS gene numbers and type of mutation are adopted from Falara *et al.* (2011), the new terpene synthase pseudogenes identified in this work are shown in bold.

Table S5 A list of the *trans*-prenyltransferase genes in tomato.

TPT	Locus	No. of exons	Predicted product ¹	Main Product	Subcellular Localization
FPPS1	Solyc12g015860	12	C15	FPP ^a	Cytosol ^b
GGPPS1	Solyc11g011240	1	C20	GGPP ^{b, c}	Plastid ^b
GGPPS2	Solyc04g079960	1	C20	GGPP ^{b, c}	Plastid ^b
SPPS	Solyc07g061990	6	C20	SPP ^d	Plastid ^d
DPPS	Solyc08g023470	12	≥C30	DPP ^d	Cytosol ^d
GGPPS3	Solyc02g085700	1	C20	GGPP ^b	Plastid ^b
TPT1	Solyc02g085710	1	C20	n.d. ^b	Mitochondria ^b
TPT2	Solyc02g085720	1	C20	n.d. ^b	Mitochondria ^b
SSU I	Solyc07g064660	1	C20	n.d. ^b	Plastid ^b
SSU II	Solyc09g008920	2	C20	n.d. ^b	Plastid ^b

¹Prediction of *trans*-prenyltransferase activity was made by the well-established “three floors” model of TPT product chain-length determination mechanism (Wang *et al.*, 2016).

^aGaffe *et al.* (2000). ^bThis work. ^cAment *et al.* (2006). ^dJones *et al.* (2013). n.d., not detected. Enzymes characterized in this work are shown in bold.

Ament K, Van Schie CC, Bouwmeester HJ, Haring MA, Schuurink RC. 2006. Induction of a leaf specific geranylgeranyl pyrophosphate synthase and emission of (*E,E*)-4,8,12-trimethyltrideca-1,3,7,11-tetraene in tomato are dependent on both jasmonic acid and salicylic acid signaling pathways. *Planta* **224**(5): 1197-1208.

Gaffe J, Bru JP, Causse M, Vidal A, Stamitti-Bert L, Carde JP, Gallusci P. 2000. *LeFPS1*, a tomato farnesyl pyrophosphate gene highly expressed during early fruit development. *Plant Physiology* **123**(4): 1351-1362.

Jones MO, Perez-Fons L, Robertson FP, Bramley PM, Fraser PD. 2013. Functional characterization of long-chain prenyl diphosphate synthases from tomato. *Biochemical Journal* **449**(3): 729-740.

Wang CY, Chen QW, Fan DJ, Li JX, Wang GD, Zhang P. 2016. Structural analyses of short-chain prenyltransferases identify an evolutionarily conserved GFPPS clade in Brassicaceae plants. *Molecular Plant* **9**(2): 195-204.

Table S6 A list of terpene compounds produced by all the functional tomato terpene synthases.

TPS	Substrate	Products	Reference
3	GPP	Camphene , tricyclene, β -Myrcene, limonene, α -pinene, sabinene, β -pinene, unidentified	Falara <i>et al.</i> (2011)
4	GPP	β-Phellandrene , β -myrcene, sabinene	Van Schie <i>et al.</i> (2007)
5	GPP <i>E,E-FPP</i>	Linalool <i>E</i>-Nerolidol	Van Schie <i>et al.</i> (2007)
7	GPP	β-Myrcene , limonene, α -pinene, β -pinene, linalool, sabinene	Falara <i>et al.</i> (2011)
8	GPP NPP	1,8-Cineole , α -pinene 1,8-Cineole , α -pinene	Falara <i>et al.</i> (2011)
9	GPP <i>E,E-FPP</i> <i>Z,Z-FPP</i>	β-Myrcene , limonene, α -pinene, (+)-2-carene, α -terpinene, terpinolene Germacrene C , germacrene A, germacrene B, germacrene D Germacrene C , α -humulene, β -bisabolene, germacrene A, germacrene B	Bleeker <i>et al.</i> (2011) Colby <i>et al.</i> (1998) Bleeker <i>et al.</i> (2011)
10	Z,Z-FPP	α-Bisabolol	This work
12	GPP <i>E,E-FPP</i> <i>Z,Z-FPP</i>	β-Myrcene , limonene, terpinolene, <i>E</i> - β -ocimene β-Caryophyllene , α -humulene γ -Curcumene, β-bisabolene , <i>Z</i> - γ -bisabolene	Bleeker <i>et al.</i> (2011) Schilmiller <i>et al.</i> (2010)
14	<i>E,E-FPP</i> <i>Z,Z-FPP</i>	β-Bisabolene , <i>E</i> -nerolidol, <i>E</i> - γ -bisabolene, <i>Z</i> - γ -bisabolene, <i>E</i> - α -bisabolene <i>E</i>-α-Bisabolene , β -bisabolene, <i>E</i> - γ -bisabolene, <i>Z</i> - γ -bisabolene,	Falara <i>et al.</i> (2011)
16	<i>E,E-FPP</i>	δ-Cadinene	This work
17	GPP <i>E,E-FPP</i> <i>Z,Z-FPP</i>	β-Myrcene , (+)-2-carene, limonene, terpinolene, <i>E</i> - β -ocimene, <i>Z</i> - β -ocimene, γ -terpinene, linalool Valencene , γ -gurjunene, <i>E</i> - β -farnesene, α -farnesene Z-γ-bisabolene , <i>E</i> - α -bergamotene, <i>Z</i> - α -bergamotene, <i>E</i> - γ -bisabolene, β -bisabolene	Bleeker <i>et al.</i> (2011)
18	NNPP	Unidentified	This work
19	NPP	β-Ocimene , β -myrcene	Matsuba <i>et al.</i> (2013)
20	NPP	β-Phellandrene , (+)-2-carene, limonene, α -phellandrene, α -terpinene	Schilmiller <i>et al.</i> (2009)
21	NNPP	Lycosantalene	Matsuba <i>et al.</i> (2013)
24	CPP	<i>Ent</i>-Kaurene	Falara <i>et al.</i> (2011)
25	GPP	<i>E</i>-β-ocimene , <i>Z</i> - β -ocimene, linalool, β -Myrcene	This work
27	<i>E,E-FPP</i>	α-Farnesene	This work
28	<i>E,E-FPP</i> <i>Z,Z-FPP</i>	Elemol (hedyacryol) Unidentified , β -bisabolene, <i>E</i> - α -bisabolene, α -eudesmol, <i>Z,Z</i> -farnesol	This work
31	<i>E,E-FPP</i>	Viridiflorene	Bleeker <i>et al.</i> (2011)
32	<i>E,E-FPP</i> <i>Z,Z-FPP</i>	Viridiflorene Unidentified	Falara <i>et al.</i> (2011)
33	<i>E,E-FPP</i> <i>Z,Z-FPP</i>	Guaia-1(10),11-diene β -Acoradiene, β -curcumene, α -cedrene, <i>Z</i> - α -bergamotene, unidentified	This work
35	<i>E,E-FPP</i> <i>Z,Z-FPP</i>	Guaia-1(10),11-diene , β -elemene (germacrene A) <i>Z,Z</i>-Farnesol , β -acoradiene	This work
36	<i>Z,Z-FPP</i>	<i>cis</i>-Muurola-3,5-diene , β -acoradiene	This work

37	GPP <i>E,E-FPP</i>	Linalool <i>E-Nerolidol</i>	Falara <i>et al.</i> (2011)
38	<i>E,E-FPP</i>	<i>E-α-Bergamotene</i>	Falara <i>et al.</i> (2011)
39	GPP <i>E,E-FPP</i>	Linalool <i>E-Nerolidol</i>	Falara <i>et al.</i> (2011)
40	GGPP	Copaly diphosphate	Bensen and Zeevaart, (1990)
41	GGPP	Copaly diphosphate	This work
46	GGPP	Geranyl linalool	Falara <i>et al.</i> (2014)
47	DMAPP	Isoprene	This work
48	<i>E,E-FPP</i> <i>Z,Z-FPP</i>	Elemol (hedycaryol) Unidentified	This work
51	<i>E,E-FPP</i> <i>Z,Z-FPP</i>	<i>E-Nerolidol, E-β-farnesene</i> α-Bisabolol, Z-nerolidol, <i>E-α-bisabolene, β-bisabolene, Z-β-farnesene</i>	This work
52	<i>E,E-FPP</i> <i>Z,Z-FPP</i>	<i>E-Nerolidol, E-β-farnesene</i> α-Bisabolol, Z-nerolidol, <i>E-α-bisabolene, β-bisabolene, Z-β-farnesene</i>	This work

The main product of each TPS enzyme using the corresponding substrate is shown in bold. The preferred substrates are shown in bold, considering their subcellular localizations. Structures of terpene compounds can be found in Fig. 11. TPS enzymes characterized in this work are shown in bold. Elemol is the thermal breakdown product of (+)-hedycaryol (Hattan *et al.*, 2016); β -elemene and δ -elemene are the thermal rearrangement of germacrene A and germacrene C, respectively (Colby *et al.*, 1998). References can be found in Table S3.

Table S7 A list of *Arabidopsis* terpene synthase genes, their characteristics, and the enzymes they encode.

Chromosome	AGI No.	TPS	TPS-clade	No. of Exons	Protein size (aa)	Subcellular localization	Substrate	Main product	Reference
1	At1g31950	29	a	7	607	M/P	Unknown	-	-
	At1g33750	22	a	6	603	M/P	<i>E,E</i> -FPP	(<i>E</i>)- β -Farnesene, α -farnesene	Wang <i>et al.</i> (2016)
	At1g48800	28	a	7	603	M/P	Unknown	-	-
	At1g61120	4	e/f	12	877	<u>C</u>	GGPP	Geranylinalool	Herde <i>et al.</i> (2008)
	At1g61680	14	g	7	569	<u>P</u>	GPP	Linalool	Chen <i>et al.</i> (2003)
	At1g66020	26	a	7	598	M/P	GGPP	Unknown diterpenes	Wang <i>et al.</i> (2016)
	At1g70080	6	a	7	611	M/P	GGPP	Dolabelladienol	Wang <i>et al.</i> (2016)
	At1g79460-KS1	32	e/f	14	785	<u>P</u>	<i>Ent</i> -CPP	<i>Ent</i> -kaurene	Yamaguchi <i>et al.</i> (1998); Helliwell <i>et al.</i> (2001)
2	At2g23230	5	a	7	606	M/P	Unknown	-	-
	At2g24210	10	b	7	591	<u>P</u>	GPP	(<i>E</i>)- β -ocimene, myrcene	Bohlmann <i>et al.</i> (2000)
3	At3g14490	17	a	7	601	P	GFPP	Sesquiterpene blend	Huang <i>et al.</i> (2017)
	At3g14520	18	a	7	605	<u>P</u>	GFPP	(+)-thalianatriene	Shao <i>et al.</i> (2017); Chen <i>et al.</i> (2019)
	At3g14540	19	a	7	602	<u>P</u>	GFPP	(-)-retigeranin B	Shao <i>et al.</i> (2017); Chen <i>et al.</i> (2019)
	At3g25810	24	b	7	598	P	GPP	Monoterpene blend	Chen <i>et al.</i> (2003)
	At3g25820	27	b	7	600	<u>P</u>	GPP	1,8-Cineole	Chen <i>et al.</i> (2004); Roos <i>et al.</i> (2015)
	At3g25830	23	b	7	600	<u>P</u>	GPP	1,8-Cineole	Chen <i>et al.</i> (2004)
	At3g29110	16	a	7	595	P	Unknown	-	-
	At3g29190	15	a	7	601	P	Unknown	-	-
	At3g29410	25	a	7	603	<u>P</u>	GFPP	(-)-ent-quianulatene, (-)-variculatriene A	Huang <i>et al.</i> (2017); Chen <i>et al.</i> (2019)
4	At4g02780-CPS	31	c	15	802	<u>P</u>	GGPP	(+)-Astallatene	Huang <i>et al.</i> (2017); Chen <i>et al.</i> (2019)
									Sun and Kamly, 1997; Helliwell <i>et al.</i> (2001)

	At4g13280	12	a	8	524	C	<i>E,E</i> -FPP	(Z)- γ -bisabolene	Ro <i>et al.</i> (2006)
	At4g13300	13	a	7	554	C	<i>E,E</i> -FPP	(Z)- γ -bisabolene	Ro <i>et al.</i> (2006)
	At4g15870	1	a	7	598	P	Unknown	-	-
	At4g16730-Ws	2	b	7	589	<u>P</u>	GPP <i>E,E</i> -FPP	(E)- β -ocimene, myrcene (E,E)- α -farnesene	Huang <i>et al.</i> (2010)
	At4g16740	3	b	7	565	<u>P</u>	GPP <i>E,E</i> -FPP	(E)- β -ocimene, myrcene (E,E)- α -farnesene	Fäldt <i>et al.</i> (2003)
	At4g20210	8	a	7	600	<u>P</u>	GGPP	Rhizathalene A	Vaughan <i>et al.</i> (2013)
	At4g20230	9	a	7	607	M/P	GGPP	Unidentified diterpene	Wang <i>et al.</i> (2016)
	At4g20200	7	a	7	603	P	Unknown	-	-
5	At5g23960	21	a	7	547	C	<i>E,E</i> -FPP	(E)- β -caryophyllene	Chen <i>et al.</i> (2003)
	At5g44630	11	a	7	557	C	<i>E,E</i> -FPP	Several sesquiterpenes	Tholl <i>et al.</i> (2005)
	At5g48110-Cvi	20	a	8	592	<u>P</u>	GGPP	Dolathaliatriene, dolabelladienol	Wang <i>et al.</i> (2016)

TPS gene numbers are adopted from Aubourg *et al.* (2002). Underlined letters indicate the experimentally proved subcellular localizations and the other localizations were predicted by the TargetP and ChloroP programs. Ws, *Arabidopsis thaliana* Wassilewskija ecotype; Cvi, *A. thaliana* Cape Verde Islands ecotype. P, plastid; M, mitochondria; C, cytosol.

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1 **Methods S1 Details on experimental procedures.**

2 **Plant growth conditions**

3 Tomato seeds were germinated on moistened filter paper. After germination, the
4 seedlings were transplanted and grown in soil in a controlled growth room maintained
5 under 16 h of light ($300 \mu\text{E m}^{-2}\text{s}^{-1}$; Valoya LED grow lights, Helsinki, Finland) at 28°C
6 and 8 h of dark at 20°C . *Nicotiana benthamiana* and *Arabidopsis thaliana* (Col-0
7 ecotype) seeds were sown in potting soil, after germination the seedlings were transferred
8 and grown in soil in a growth chamber under controlled conditions of light intensity of
9 $200 \mu\text{E m}^{-2}\text{s}^{-1}$ (mixed cool-white and incandescent bulbs) for 16 h light period, and
10 day/night temperature of $25/20^\circ\text{C}$.

11

12 **RNA isolation, cDNA synthesis and quantitative real-time PCR**

13 Total RNA was isolated using the E.Z.N.A Plant RNA Kit (Omega Bio-tek, Norcross,
14 GA, USA) and incubated with DNase I (Promega, Madison, WI, USA) to avoid genomic
15 DNA contamination according to the manufacturer's protocol. One microgram of total
16 RNA was used to synthesize cDNA by using the High Capacity cDNA Reverse
17 Transcription Kit (Thermo Fisher, Waltham, MA, USA). For quantitative real-time PCR
18 (RT-qPCR), 1 μL of the 10-fold diluted cDNA and gene-specific primers were applied to
19 10 μL SYBR Green reactions (PowerUp SYBR Green Master Mix, Thermo Fisher),
20 which were run on a Real-Time PCR System (StepOnePlus, Thermo Fisher) with the
21 following cycles: 95°C for 10 min, followed by 40 cycles of 95°C for 15 s and 60°C for 1
22 min, and a melting curve analysis. Gene expression values were calculated according to
23 the previously described comparative C_T method (Schmittgen & Livak, 2008). For
24 normalizing gene expressions, *Ubiquitin* (*Solyc11g071260*) was used as a control. Three
25 biological replicates were used for each point with three or more technical replicates.
26 Primers used are listed in Table S1.

27

28 **Protein expression in *Escherichia coli* and purification**

29 For enzymatic assays, TPS genes without their fragments corresponding to putative
30 transit peptides were cloned upstream and in-frame of the (His)₆-tag sequences in
31 pET32b to express TPS-His recombinant proteins. Restriction sites and primers used are
32 detailed in Table S1. The resulting recombinant plasmids were sequence verified and
33 transformed into *E. coli* Rosetta2(DE3)pLysS cells (Novagen, Burlington, MA, USA).

34 Single positive colonies were used to inoculate 4 mL LB medium containing 34 ng μL^{-1}
35 chloramphenicol and 100 ng μL^{-1} ampicillin. One mL of the overnight culture was
36 transferred to 100 mL of the same medium and continued to grow at 37°C to reach an
37 OD₆₀₀ of approximately 0.3. Protein expression was induced by adding isopropyl β -
38 thiogalactopyranoside (IPTG) to a final concentration of 1 mM. After 16 h incubation at
39 16°C, cells were harvested by centrifugation and stored at -80°C until use. The
40 recombinant His-tagged proteins were affinity purified using nickel-nitrilotriacetic acid
41 (Ni-NTA) agarose (Qiagen, Hilden, Germany) according to the manufacturer's manuals.
42 Protein concentration was determined using the Bradford method (Bradford, 1976) with
43 BSA as a standard.

44

45 **GC-MS analysis of terpenes**

46 The analysis of terpene products (C₁₀-C₂₅) was performed as described previously (Falara
47 et al., 2011). One microliter of sample was auto-injected onto a Shimadzu (Kyoto, Japan)
48 QP-2010 SE GC-MS system equipped with the Rxi-5Sil column (30 m length, 0.25 mm
49 i.d., and 0.25 μm film thickness; Restek, Bellefonte, PA, USA). The injection port was
50 set to 240°C in splitless mode, and the flow rate of the He carrier gas was set at 1.4 mL
51 min⁻¹. The GC oven temperature was programmed as follows: 45°C for 2 min, 10°C min⁻¹
52 up to 250°C and hold for 3 min, 20°C min⁻¹ up to 320°C and hold for 5 min. The

53 analysis of prenyltransferase assay products was performed using an EC-WAX column
54 (30 m length, 0.32 mm i.d., and 0.25 μ m film thickness; Grace, Columbia, MD, USA).
55 The injection port was set to 220°C in splitless mode, and the flow rate of the He carrier
56 gas was set at 1.5 mL min⁻¹. The GC oven temperature was programmed as follows: 45°C
57 for 2 min, 10°C min⁻¹ up to 260°C and hold for 5 min. Interface temperature was 280°C
58 and ion source temperature was set to 260°C. The detector was activated after a 4.5-min
59 solvent delay. Mass spectra were obtained using the scan mode (total ion count, 45-450
60 mz^{-1}). The Kováts retention indices (Babushok *et al.*, 2011) of each terpene compounds
61 were determined based on a comparison with (C₇-C₄₀) n-alkanes. Products were
62 identified using the mass spectra database NIST11 and their mass spectra and retention
63 indices were compared with authentic compounds or the literature (Table S2).

64

65 Analysis of isoprene was performed by directly injecting the SPME fiber into the
66 Shimadzu QP-2010 SE GC-MS system equipped with the Rxi-5Sil column (30 m length,
67 0.25 mm i.d., and 0.25 μ m film thickness; Restek). The injection port was set to 220°C in
68 splitless mode, and the flow rate of the He carrier gas was set at 1.0 mL min⁻¹. The GC
69 oven temperature was programmed as follows: 40°C for 3 min, 10°C min⁻¹ up to 100°C
70 and hold for 3 min, 20°C min⁻¹ up to 260°C and hold for 3 min. Isoprene was identified
71 by comparison with the authentic standard.

72

73 **Correlation analysis of tomato terpene synthase genes and terpene volatiles**

74 The Spearman correlation coefficients were calculated by GraphPad Prism 6 using gene
75 expression data and terpene profiling data as indicated in Fig. 9 and Fig. 10, respectively.
76 Based on the Spearman correlation coefficient, the hierarchical clustering was performed
77 using average linkage method by Cluster 3.0 and a heatmap was built using TreeView
78 3.0.

79

80 **Accession numbers**

81 The GenBank accession numbers of the proteins used in the phylogenetic analysis (Fig.
82 S9) are as follows: AgGPPS1, AF513111; AgGPPS2, AF513112; AgGPPS3, AF513113;
83 PaGPPS, EU432047; PaGPPS/GGPPS, GQ369788; PaGPPS3, EU432048; MiGPPS1,
84 JN035297; MiGPPS2, JN035298; MiFPPS, JN035296; AmGPPS.LSU, AAS82860;
85 AmGPPS.SSU, AAS82859; CbGPPS.SSU, AY534745; CrGGPPS, JX417183;
86 CrGPPS.SSU, JX417184; HlGPPS.LSU, FJ455407; HlGPPS.SSU, ACQ90681;
87 MpGPPS.LSU, AF182828; MpGPPS.SSU, AF182827; VvGPPS, AAR08151; LaFPPS,
88 U15777; AaFPPS, U36376; ZmFPPS, L39789; NtGGPPS1, GQ911583; NtGGPPS2,
89 GQ911584; SaGGPPS, CAA67330; HbSPPS, DQ437520; OsGGPPS1, Os07g39270;
90 OsGPPS, Os01g14630; OsGRP; Os02g44780. AGI numbers for *A. thaliana* TPTs are as
91 follows: AtPPPS, At2g434630; AtFPPS1, At5g47770; AtFPPS2, At4g17190; AtSPPS1,
92 At1g78510; AtSPPS2, At1g17050; AtGGPPS11, At4g36810; AtGGPPS2, At2g18620;
93 AtSSU, At4g38460. Tomato TPT homologs are listed in Table S5.

94

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