# Package 'CellVizR'

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Type Package

Title Visualization and statistical analyses of single-cell data using manifold representations

**Description** The profiling of biological samples at the single-cell level, using either high-

dimensional cytometry or single-cell transcriptomics, is becoming more and more common.

Such generated data are usually analyzed using manifold algo-

rithms, such as UMAP, tSNE, or LargeVis, combined with cell clustering algorithms.

Nevertheless, this is still challenging for non-bioinformatician experts to easily han-

dle the whole pipeline of computational analyses with the purpose of answering specific biological questions.

CellVizR allows the statistical analysis and visualization of single-

cell data using manifold algorithms and clustering methods.

Especially, several key analysis steps are available to perform data importation, manifold generation, cell cluster identification, statistical analyses, cluster visualization, and quality controls of generated results.

CellVizR can import cell events from FCS, MTX or txt file formats using different transformation, down-sampling, and normalization approaches.

Manifold representations can be generated using the UMAP, tSNE or LargeVis algorithms to project cell events into a lower dimensionality space.

The identification of cell clusters can be done using multiple clustering algorithms, depending on user's assumptions.

The characteristics of cell clusters can be visualized using scatter plot, categori-

cal heatmap of marker expressions, or using parallel coordinates representations.

Cell clusters having abundances differently expressed between biological conditions can be identified using several statistical tests.

Statistical results can be visualized using volcano plots or heatmaps.

Unsupervised and supersized analysis approaches can be conducted by users to appreciate the homogeneity/heterogeneity of biological conditions, and to identify cell population biomarkers in a multivariate manner.

Additionally, CellVizR provides a workflow for asserting the quality of identified cell clusters using statistical approaches.

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Imports checkmate,

cluster, concaveman, cowplot, dbscan, 2 R topics documented:

dendextend,

	diptest,
	FactoMineR,
	flowCore,
	FNN,
	ggdendro,
	ggiraph,
	ggnewscale,
	ggplot2,
	ggpubr,
	ggrepel,
	ggridges,
	Gmedian,
	gridExtra,
	gtools,
	kohonen,
	MASS,
	methods,
	plyr,
	reshape,
	reshape2,
	rstatix,
	Rtsne,
	scales,
	Seurat,
	spade,
	stats,
	stringr,
	uwot,
	viridis
Sugg	ests knitr
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Vigno	etteBuilder knitr
biocV	iews Clustering, DataImport, FlowCytometry, Normalization, StatisticalMethod, Software, Vi-
	sualization
Roxy	genNote 7.2.1
Rto	opics documented:
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## Description

This function aims to attach meta-information to each biological sample.

Especially, the biological individual, the biological condition and the time point of each sample can be specified for subsequent analyses.

## Usage

assignMetadata(Celldata, metadata)

## **Arguments**

Celldata a Celldata object

metadata a data.frame containing contextual information about the biological samples.

This data.frame must have 3 columns specifying for each sample the associated individual (column named 'individual'), the biological condition (column

named 'condition') and the time point (column named 'timepoint')

#### Value

a S4 object of class 'Celldata'

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Celldata-class

Celldata class definition

## **Description**

The Celldata object is a S4 object containing all cytometry expressions.

#### **Slots**

samples a character vector containing the names of the biological samples

raw.markers a character vector containing the names of the raw markers

matrix.expression.r a data.frame containing the raw marker expressions of each cell

matrix.expression a data.frame containing the marker expressions of each cell

manifold a data.frame containing the manifold coordinates

manifold.params a list containing the parameters used for manifold creation

identify.clusters a vector containing the identifiers of cell clusters

identify.clusters.params a vector containing the parameters used for the identification of the cell clusters

concave.hulls a data.frame containing the coordinates of the cell cluster of the concave hulls for each cluster

matrix.cell.count a data.frame containing the number of cells associated to each cluster for each sample

matrix.abundance a data.frame containing the percentage of cells associated to each cluster for each sample

statistic a data.frame containing the statistics of cell clusters

metadata a data.frame containing the metadata associated to each sample

computeStatistics

Computes differential analysis statistics for cell clusters

#### **Description**

This function aims to compute the statistics of Differentially Abundant Clusters 'DAC'.

DAC correspond to cell clusters having abundances statistically different between two biological conditions. The statistical test used for the comparisons can be defined by users. For each cluster, the p-value, log2 fold-change and effect size relative to the reference condition are computed. Statistical comparison can be performed in a paired and unpaired manner.

```
computeStatistics(
  Celldata,
  condition,
  ref.condition,
  test.statistics = c("wilcox.test", "t.test"),
  paired = FALSE
)
```

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#### **Arguments**

Celldata a Celldata object

condition a character value providing the name of the condition to be compared

ref.condition a character value providing the name of reference condition

test.statistics

a character value providing the type of statistical test to use. Possible values are:

'wilcoxon' or 't-test'

paired a boolean value indicating if a paired or unpaired comparison should be applied

#### Value

a S4 object of class 'Celldata'

createMetaclusters

Create metaclusters

#### **Description**

This function aims to gathered mutliple cell cluster to a large cell cluster

#### Usage

```
createMetaclusters(Celldata, clusters, metaclusters)
```

## **Arguments**

Celldata a Celldata object

clusters a character vector containing the identifiers of the clusters to use. By default, all

clusters are used

metaclusters a character value containing the name of the metacluster to create

#### Value

a Celldata object

export

Exports cell expression profiles to TSV or FCS files

## Description

Exports cell expression profiles from a Celldata object to a tab-separated or FCS files Cell expression profiles can be exported for a set of given samples and for a set of given cell clusters

```
export(Celldata, filename, clusters = NULL, samples = NULL)
## S4 method for signature 'Celldata'
export(Celldata, filename, clusters = NULL, samples = NULL)
```

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#### **Arguments**

Celldata a Celldata object

filename a character value providing the name of the output file

clusters a character vector containing the identifiers of the cell clusters to export. By

default, all clusters are extracted

samples a character vector containing the names of biological samples to export. By

default, all samples are extracted

#### Value

none

generateManifold

Generates a manifold of cell events

## **Description**

This function aims to generate a manifold representation for cell events stored in a Celldata object This function allows the use of several non-linear dimension reduction techniques such as UMAP, t-SNE or LargeVis The whole set of cell markers or specific cell markers can be used during the dimensionality reduction process

## Usage

```
generateManifold(
   Celldata,
   type = c("UMAP", "tSNE", "lvish"),
   markers = NULL,
   seed = 42,
   verbose = TRUE,
   ...
)
```

## **Arguments**

Celldata a Celldata object

type a character value specifying the type of manifold to compute. Possible values

are: 'UMAP' for Uniform Manifold Approximation and Projection, 'tSNE' for

t Stochastic Neighbor Embedding, and 'lvish' for LargeVis

markers a character vector providing the cell markers to use for the manifold generation seed a numeric value providing the random seed to use during stochastic operations

verbose a boolean value indicating if computational details must be displayed on the

console

... Other arguments passed on to methods

#### Value

a S4 object of class 'Celldata'

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getSamples	Filter to select samples
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#### **Description**

This function aims to select the samples of interest

## Usage

```
getSamples(Celldata, individual = NULL, condition = NULL, timepoint = NULL)
```

## Arguments

```
Celldata a Celldata object

individual a character vector containing the names of biological individual to use

condition a character vector containing the biological condition to use

timepoint a character vector containing the timepoint condition to use
```

#### Value

```
a ggplot2 object
```

identifyClusters

Identify cell cluster of having similar marker expressions

## Description

This function aims to identify cell clusters, which are groups of cells having similar expressions for selected markers, using different unsupervised clustering methods.

Several clustering method are available such as kmeans, kmedian, clara, DBSCAN, HDBSCAN and SOM. The cell clustering can be performed on the manifold representation or based on marker expression.

```
identifyClusters(
   Celldata,
   space = c("manifold", "markers"),
   markers = NULL,
   method = c("kmeans", "kmedian", "clara", "DBSCAN", "SOM"),
   concavity = 2,
   length.threshold = 0,
   seed = 42,
   ...
)
```

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#### **Arguments**

Celldata a Celldata object

space a character value containing the space of clustering method to use. Possible

values are: 'manifold' or 'markers'

markers a character vector providing the cell markers to use for the manifold generation

method a character value containing the name of the clustering method to use. Possible

values are: 'kmeans', 'kmedian', 'clara', 'DBSCAN' and 'SOM'

concavity a numeric value providing a relative measure of concavity for the computation

of the concave hulls (please refer to the function 'concaveman' of the 'concave-

man' package)

length.threshold

a numeric value providing a threshold of the segment length for the computation

of the concave hulls (please refer to the function 'concaveman' of the concave-

man package)

seed a numeric value providing the random seed to use during stochastic operations

... Other arguments passed on to methods

#### **Details**

For each identify cell cluster, the boundaries of cells belonging to this cluster are delineated using a concave hull

#### Value

a S4 object of class 'Celldata'

import

Imports of cell expression profiles from TSV or FCS files

#### **Description**

This function aims to import acquired cell events from cytometric profiling into a Celldata object.

Input files can be tab-separated or FCS files. Different transformations can be applied such as logicle, arcsinh or logarithmic. Importantly, a downsampling of cell events can be performed using uniformly-based or density-based random selections. Cell marker having technical or biological biaises can be excluded during the import.

```
import(
   files,
   filetype = "fcs",
   transform = c("logicle", "arcsinh", "logarithmic", "none"),
   d.method = c("none", "uniform", "density"),
   parameters.method = list(exclude.pctile = 0.01, target.pctile = 0.05, target.number =
        NULL, target.percent = 0.1),
   exclude.markers = NULL,
   seed = 42
)
```

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#### **Arguments**

files a character vector specifying the path of the tab-separated or FCS files to load filetype a character vector specifying the format of the loaded files. By default, FCS is used transform a character value containing the type of the transformation to apply. Possible values are: 'logicle', 'arcsinh', 'logarithmic' or 'none' d.method a character value containing the type of the downsampling to apply. Possible values are: 'none', 'uniform' or 'density' parameters.method a list value containing the parameters to use for downsampling exclude.markers a character vector providing the marker names to be excluded during the import

a numeric value providing the random seed to use during stochastic operations seed

#### Value

a S4 object of class 'Celldata'

importMTX	
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## **Description**

This function aims to import acquired cell events from single-cell transcriptomic profiling into a Celldata object.

#### Usage

```
importMTX(mtx = mtx, cells = cells, features = features)
```

## **Arguments**

a character vector specifying the path of the mtx file to load mtx cells a character vector specifying the format of the mtx file to load features a character vector specifying the format of the mtx file to load

```
a S4 object of class 'Celldata'
```

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performUpsampling

Performs the upsampling of downsampled events

## **Description**

This function aims to perform upsample downsampled events based on an existing Celldata object and existing cell events stored in tab-separated or FCS files

Importantly, the identification of cell clusters must have been performed prior to this operation

## Usage

```
performUpsampling(
   Celldata,
   files,
   transform = c("logicle", "arcsinh", "logarithmic", "none")
)
```

## **Arguments**

Celldata a Celldata object

files a character vector providing the path of the tab-separated or FCS files

transform a character value containing the type of the transformation to apply. Possible

values are: 'logicle', 'arcsinh', 'logarithmic' or 'none'

#### Value

a S4 object of class 'Celldata'

plot

Plots graphics for all Celldata objects

## **Description**

Generates a graphical representation for a Celldata object The displayed representation depends on the current analysis status of the Celldata object

- If the manifold has not been calculated, then the number of cells per sample will be displayed
- If the manifold has been calculated but not the clustering, then the manifold representation will be displayed
- If the manifold and clustering have been calculated, then a heatmap of marker expressions will be displayed

## Usage

```
## S4 method for signature 'Celldata,ANY'
plot(x)
```

#### **Arguments**

x a Celldata object

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#### Value

```
a ggplot2 object
```

plotBoxplot

Plots cell cluster abundances using a boxplot representation

## **Description**

This function aims to visualize and compare the cell cluster abundances for each biological condition using boxplot and jitter representations.

The abundance of a specific cell cluster or a set of cell clusters can be displayed. The representation can be resticted to a specific set of samples. Moreover boxplot can be constructed based on sample meta information. Statistic can be computed for all comparisons.

## Usage

```
plotBoxplot(
   Celldata,
   clusters,
   samples = NULL,
   observation = c("individual", "condition", "timepoint"),
   test.statistics = c("wilcox.test", "t.test"),
   paired = FALSE,
   hide.ns = TRUE
)
```

#### **Arguments**

Celldata a Celldata object

clusters a character vector containing the identifiers of the clusters to use

samples a character vector containing the names of biological samples to use. By default,

all samples are used

observation a character value containing the parameters to use

test.statistics

a character value providing the type of statistical test to use. Possible values are:

'wilcox.test' or 't.test'

paired a boolean value indicating if a paired or unpaired comparison should be applied

hide.ns a boolean value indicating if non significant p-value must be hidden

```
a ggplot2 object
```

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Plots the numbers of cells for each sample

## Description

This function aims to visualize the number of cells associated to each sample.

This representation displays the samples in the X-axis and the number of associated cells in the Y-axis. Several statistics can be computed and shown.

## Usage

```
plotCellCounts(
   Celldata,
   stats = c("min", "median", "mean", "q75", "max"),
   samples = NULL,
   sort = TRUE
)
```

## **Arguments**

Celldata	a Celldata object
stats	a character vector providing the statistics to display. Possible values are: 'min', 'median', 'mean', 'q75', 'max'
samples	a character vector containing the names of biological samples to use. By default, all samples are used
sort	a boolean value indicating if clusters must be sorted by the number associated sample

## **Details**

The following statistic can be computed:

- -'min' corresponds to the lowest number of cells within a data set
- -'median' corresponds to the number of cells separates the lower half from the upper half within data set
- -'mean' corresponds to the number of cells quantity shared within data set
- -'q75' corresponds to the number of cells separates the quantiles 75
- -'max' corresponds to the largest number of cells within a data set

```
a ggplot2 object
```

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plotClustersCounts

Plots the numbers of cells for each clusters

## **Description**

This function aims to visualize the number of cells associated to each clusters.

This representation displays the clusters in the X-axis and the total number of associated cells in the Y-axis.

## Usage

```
plotClustersCounts(
   Celldata,
   clusters = NULL,
   sort = TRUE,
   legend.max.samples = 10
)
```

### **Arguments**

Celldata a Celldata object

clusters a character vector containing the identifiers of the clusters to use. By default, all

clusters are used

sort a boolean value indicating if clusters must be sorted by the number associated

cluster

legend.max.samples

a numerical value specifying the maximal number of samples to display in the

legend

#### Value

a ggplot2 object

plotCombineHM

Plots a combined expression and statistic heatmaps

## Description

This function aims to combine the expression and statistic heatmaps.

## Usage

```
plotCombineHM(HM1, HM2)
```

## Arguments

HM1 a ggplot object containing the expression heatmap
HM2 a ggplot object containing the statistic heatmap

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#### Value

```
a ggplot2 object
```

plotCoordinates

Plots of phenotype of cell clusters using parallels coordinates

#### **Description**

This function aims to visualize the caracterisites of cell clusters using parallels coordinates. Each line in the plot corresponds to a biological sample for which marker/gene expression are indicated.

## Usage

```
plotCoordinates(
   Celldata,
   condition.samples = c("condition", "timepoint"),
   samples = NULL,
   clusters
)
```

## **Arguments**

Celldata a Celldata object condition.samples

a character vector containing the variable to be studied for the samples. Possible

values are: 'condition' or 'timepoint"

samples a character vector containing the names of biological samples to use. By default,

all samples are used

clusters a character vector containing the identifiers of the clusters to use

## Value

a ggplot2 object

plotDistogram

Plots of a distogram of marker co-expression

## **Description**

This function aims to visualize the pairwise co-expression between all markers using a distogram representation. Each tile corresponds to the co-expression between two markers and is gradient-colored based on the Pearson correlation

```
plotDistogram(Celldata, clusters = NULL)
```

plotHmAbundances 15

#### **Arguments**

Celldata a Celldata object

clusters a character vector containing the identifier of the cluster to use

#### **Details**

The Pearson correlation is computed based on the marker expressions

#### Value

```
a ggplot2 object
```

plotHmAbundances

Plots an heatmap of cell cluster abundances

## **Description**

This function aims to visualize the abundances of cell clusters using an heatmap representation

In such heatmap each column corresponds a cell cluster and he row corresponds the different samples. The heatmap can be restricted to specific cell clusters and samples. The levels of abundance of each sample in each cluster is represented using a color gradient scale Abundance values can be centered and reduced.

## Usage

```
plotHmAbundances(
   Celldata,
   clusters = NULL,
   samples = NULL,
   saturation = 2.5,
   rescale = FALSE
)
```

#### **Arguments**

Celldata a Celldata object

clusters a character vector containing the identifiers of the clusters to use. By default, all

clusters are used

samples a character vector containing the names of biological samples to use. By default,

all samples are used

saturation a numeric value providing the saturation threshold of cell cluster abundances rescale a boolean specifying if cell cluster abundances must be centered and reduced

```
a ggplot2 object
```

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plotHmExpressions

Plots an heatmap of cell marker expressions

## **Description**

This function aims to visualize the cell marker expressions for selected markers and clusters

The mean of median marker expressions is computed for each cluster, and marker expressions displayed using a categorical heatmap (5 categories are defined by default) The range expression of each cell marker is discretized into several categories between bounds of marker expressions To hierarchical clustering, shown using dendrogramm, can be computed on both marker and cluster levels

#### Usage

```
plotHmExpressions(
   Celldata,
   markers = NULL,
   clusters = NULL,
   method.hclust = c("ward.D", "ward.D2", "single", "complete", "average", "mcquitty",
        "median", "centroid"),
   nb.cat = 5,
   seed = 42
)
```

#### **Arguments**

Celldata a Celldata object

markers a character vector providing the marker names to use. By default, all markers

are used

clusters a character vector containing the identifiers of the clusters to use. By default, all

clusters are used

method.hclust a character value providing the agglomeration method to be use. Possible values

are: 'ward.D', 'ward.D2', 'single', 'complete', 'average', 'mcquitty', 'median'

or 'centroid' (please refer to the function 'hclust' of the 'stats' package)

nb.cat a numeric specifying the number of categories to use

seed a numeric value providing the random seed to use during stochastic operations

```
a ggplot2 object
```

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plotHmStatistics

Plots an heatmap of a statistical analysis results

## **Description**

This function aims to visualize the results of differential cell clusters analysis

This representation displays statistical information for each cell cluster for a given comparison of samples Different statistics can be visualized, such as the p-value, the log2(fold-change), and effect size

## Usage

```
plotHmStatistics(
   Celldata,
   clusters = NULL,
   statistics = c("pvalue", "lfc", "effsize"),
   saturation = 3
)
```

## **Arguments**

Celldata a Celldata object

clusters a character vector containing the identifiers of the clusters to use. By default, all

clusters are used

statistics a character value providing the name of the statistic to display. Possible values

are: 'pvalue' for p-value, 'lfc' for log2 fold change or 'eff' for effect size

saturation a numeric value providing the saturation value for statistics to display

#### Value

a ggplot2 object

plotLDA

Plots a LDA representation based cell cluster abundances

## **Description**

This function aims to represent a Linear Discriminant Analysis representation based on cell cluster abundances

```
plotLDA(
   Celldata,
   levels = c("predictions", "coefficients"),
   ref.condition,
   condition,
   clusters = NULL
)
```

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#### **Arguments**

Celldata a Celldata object

levels a character value containing the variable to be displayed. Possible values are:

'clusters' or 'samples'

ref.condition a character value providing the name of reference condition

condition a character value providing the name of the condition to be compared

clusters a character vector containing the identifiers of the clusters to use. By default, all

clusters are used

#### Value

XX

plotManifold

Plots a representation of a computed manifold

## **Description**

This function aims to visualize a computed manifold representation for given analysis.

This representation can be used on a Celldata object for which a manifold analysis has been performed.

If a cell clustering has been performed, then the clusters are delineated using concave hulls. Additionnally, the manifold can be colored based on the local cell density or marker expressions. It is possible to centred ans reduce the values of expressions.

#### Usage

```
plotManifold(
   Celldata,
   markers = "density",
   samples = NULL,
   scale = FALSE,
   quant.low = 0.05,
   quant.high = 0.95
)
```

## **Arguments**

Celldata a Celldata object

markers a character value providing the name of the marker to use for the colouring. By

default, cells are colored based on their local density

samples a character vector containing the names of biological samples to use

scale a boolean value specifing if expression calue must be rescaled

quant.low a numeric value providing the number of first quantile quant.high a numeric value providing the number of last quantile

#### Value

a ggplot2 object

plotMarkerDensity 19

plotMarkerDensity

Plots of phenotype of identified cell clusters

#### **Description**

This function aims to visualize xxx

#### Usage

```
plotMarkerDensity(
   Celldata,
   clusters,
   quant.low = 0.05,
   quant.high = 0.95,
   dip.th = 0.01
)
```

## **Arguments**

```
Celldata a Celldata object

clusters a character vector containing the identifier of the cluster to use

quant.low a numeric value providing the number of first quantile

quant.high a numeric value providing the number of last quantile

dip.th a numeric value specifing the p-value threshold of the Hartigan's dip test
```

#### Value

a ggplot2 object

plotMDS

Plots a MDS representation based on cell cluster abundances

## **Description**

This function aims to visualize the similarities between samples or clusters based on their abundances, using a Multidimensional Scaling representation. Each dot represents a sample or a cluster and the distances between the dots are proportional to the Euclidean distance between these objects. The representation can be restricted to specific cell clusters and samples. In addition, it is possible to choose the levels displayed, clusters or samples.

```
plotMDS(
   Celldata,
   matrix = c("abundance", "expression"),
   levels = c("clusters", "samples"),
   condition.samples = c("condition", "timepoint"),
   clusters = NULL,
   samples = NULL,
   plot.text = TRUE
)
```

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#### **Arguments**

Celldata a Celldata object

matrix a character vector containing the matrix to be studied. Possible values are:

'abundance' or 'expression'

levels a character value containing the variable to be displayed. Possible values are:

'clusters' or 'samples'

condition.samples

a character vector containing the variable to be studied for the samples. Possible

values are: 'condition' or 'timepoint"

clusters a character vector containing the identifiers of the clusters to use. By default, all

clusters are used

samples a character vector containing the names of biological samples to use. By default,

all samples are used

plot.text a boolean value specifing if adds text directly at the plot

#### Value

a ggplot2 object

plotPCA

Plots a PCA representation based cell cluster abundances

## Description

This function aims to represent a Principal Component Analysis representation based on cell cluster abundances. In such representation, clusters or samples are positioned based on computed principal components. The representation can be displayed based on specific principal components. The representation can be restricted to specific cell clusters and samples. In addition, it is possible to choose the levels displayed, clusters or samples.

## Usage

```
plotPCA(
   Celldata,
   levels = c("both", "clusters", "samples"),
   clusters = NULL,
   samples = NULL,
   components = c(1, 2),
   condition.samples = c("condition", "timepoint"),
   cor.radius.th = 0.6,
   plot.text = TRUE
)
```

## Arguments

Celldata a Celldata object

levels a character value containing the variable to be displayed. Possible values are:

'both', 'clusters' or 'samples'

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clusters a character vector containing the identifier of the cluster to use. By default, all

clusters are used

samples a character vector containing the names of biological samples to use. By default,

all samples are used

components a numeric vector providing the components to display

condition.samples

a character vector containing the variable to be studied for the samples. Possible

values are: 'condition' or 'timepoint"

cor.radius.th a numeric value specifing the radius of the correlation plot radius

plot.text a boolean value specifing if adds text directly at the plot

## Value

a ggplot2 object

plotScatter	Plots of a scatter plot of marker co-expression

## Description

This function aims to visualize co-expression between two markers using a scatter representation

## Usage

```
plotScatter(Celldata, marker1, marker2, samples = NULL, clusters = NULL)
```

## **Arguments**

Celldata	a Celldata object
marker1	a character value specifying the first marker to be visualised
marker2	a character value specifying the second marker to be visualised
samples	a character vector containing the names of biological samples to use. By default, all samples are used
clusters	a character vector containing the identifiers of the clusters to use. By default, all clusters are used

```
a ggplot2 object
```

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plotVolcano	Plots of a volcano plot of statistical analysis	
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## Description

This function aims to visualize the results of a differentially abundant a analysis using a Volcano plot In such representation, each in dot corresponds to a cell cluster and dots are positioned in two dimensional space where the X-axis represents the log2(fold-change) and the Y-axis represents the -log10 of the p-value. Un horizontal line is displayed accordingly to the p-value threshold and to vertical lines are displayed accordingly to the fold-change threshold.

### Usage

```
plotVolcano(Celldata, comparison, th.pv = 1.3, th.fc = 1.5, plot.text = TRUE)
```

## **Arguments**

Celldata a Celldata object

comparison a character value containing the comparison to study
th.pv a numeric value containing the p-value threshold to use
th.fc a numeric value containing the fold-change threshold to use
plot.text a boolean value specifing if adds text directly at the plot

## Value

a ggplot2 object

print

Prints information for a given Celldata object

## **Description**

Prints a preview for a Celldata object

## Usage

```
## S4 method for signature 'Celldata'
print(x)
```

## Arguments

x a Celldata object

#### Value

none

QCMarkerNames 23

QCMarkerNames	Verifies the consistency of the marker names within cell event files	

## **Description**

This function aims to check the consistency of marker names across multiple tab-separated or FCS files.

Additionally, the number of cells associated to each sample is displayed.

#### Usage

```
QCMarkerNames(files)
```

#### **Arguments**

files

a character vector specifying the path of the tab-separated or FCS files to check

#### Value

a data.frame containing the marker names and the associated number of cells for each sample (rownames = samples and colnames = markers)

QCMarkerRanges	Verifies the consistency of marker expressions integrity within cell
	event files

## **Description**

This function aims to check the consistency of marker expressions ranges across multiple tabseparated or FCS files.

The marker expressions ranges are calculated based on the user-defined quantiles.

## Usage

```
QCMarkerRanges(files, probs = c(0.05, 0.95))
```

## **Arguments**

files a character vector specifying the path of the FCS files to verified

probs a numerical vector providing the quantiles used to define marker expressions

ranges

### Value

a list containing two data.frame for the lower and upper marker expression ranges (rownames = samples and colnames = markers)

24 QCUniformClusters

OCSmallClusters	Computes the percentage	e of cell clusters	s with low nur	nher of cells

## Description

This function aims to compute and show cell clusters having a number of associated cells lower than a specific threshold

## Usage

```
QCSmallClusters(Celldata, th.size = 50, plot.device = TRUE)
```

## **Arguments**

Celldata a Celldata object

th.size a numeric value providing the minimum number of cells needed for a cluster to be considered a small cluster

plot.device a boolean value specifying a results representation must be displayed

### Value

a numerical value corresponding to the percentage of cell cluster with low number of cells

QCUniformClusters Computes the percentage of clusters with uniform phenotypes
-------------------------------------------------------------------------------

## Description

This function aims to identify and show cell clusters having a uniform phenotype

A uniform cluster corresponds to a cluster that have a unimodal expression and a low spread of expression for all its markers

```
QCUniformClusters(
   Celldata,
   uniform.test = c("both", "uniform", "IQR"),
   th.pvalue = 0.05,
   th.IQR = 2,
   plot.device = TRUE
)
```

renameMarkers 25

#### **Arguments**

Celldata a Celldata object

uniform.test a character providing the name of test assessment to perform. Possible value are : 'both', 'uniform', 'IQR'

th.pvalue a numeric value providing the p-value threshold of the Hartigan's dip test (unimodal if pvalue > th.pvalue)

th.IQR a numeric value providing the IQR (interquartile range) threshold to assume a distribution as uniform

plot.device a boolean value specifying if result representation must be displayed

## **Details**

-'uniform' corresponds to the verification of the unimodal distribution of markers with a Hartigans test

-'IQR' corresponds to the verification of the distribution of markers so that they are not below the IQR threshold (interquantile range)

-'both' corresponds to the combination of the two parameters : uniform and IQR

#### Value

a numerical value corresponding to the percentage of cell cluster with unimodal expression and a low spread

renameMarkers Renames markers within a Celldata object	
--------------------------------------------------------	--

## **Description**

This function aims to rename cell markers stored within a Celldata object.

This function is interesting to remove the names of the fluorochromes or metals recorded during the acquisition process.

## Usage

```
renameMarkers(Celldata, marker.names)
```

#### **Arguments**

Celldata a Celldata object

marker.names a character vector providing the new marker names to use

#### Value

a S4 object of class 'Celldata'

26 show

show

Prints information for a Celldata objects

## Description

Shows a preview for a Celldata object

## Usage

```
## S4 method for signature 'Celldata'
show(object)
```

## Arguments

object

a Celldata object

## Value

none

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```