

# Introduction to Bioinformatics

JTMS-19

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**Key idea of probability models (repeated), multiple sequence alignment, final exam: first preview**

**What is this session about?**

One key idea of probability models is revisited. An algorithm for multiple sequence alignment is introduced. A first preview version of the final exam is discussed.

**How can you revise the material after the session?**

Read Durbin et al. chapters 6.1-6.4

*alternative reading:* Hütt/Dehnert chapter 3.2.2

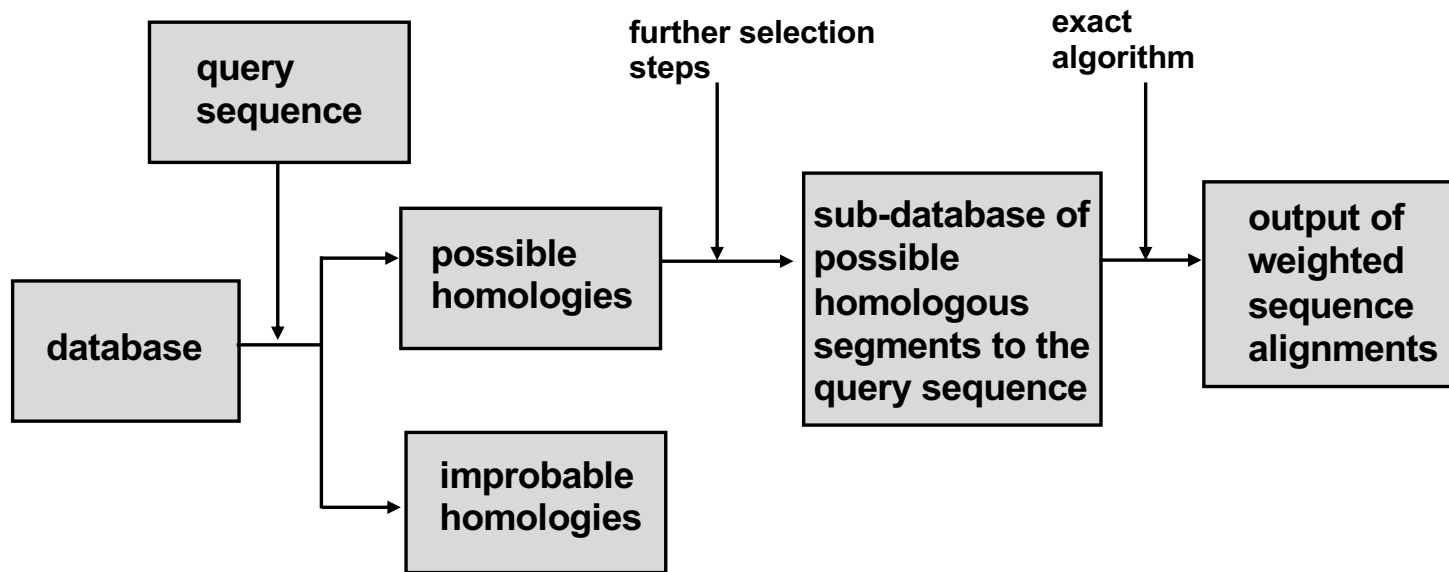
**... from the previous lecture**

$$F_{i,j} = \max \begin{cases} 0 \\ F_{i-1,j-1} + s(x_i, y_j) \\ F_{i-1,j} - d \\ F_{i,j-1} - d \end{cases}$$

KEFHN - GH  
 | | | |  
 KYFHKAGN

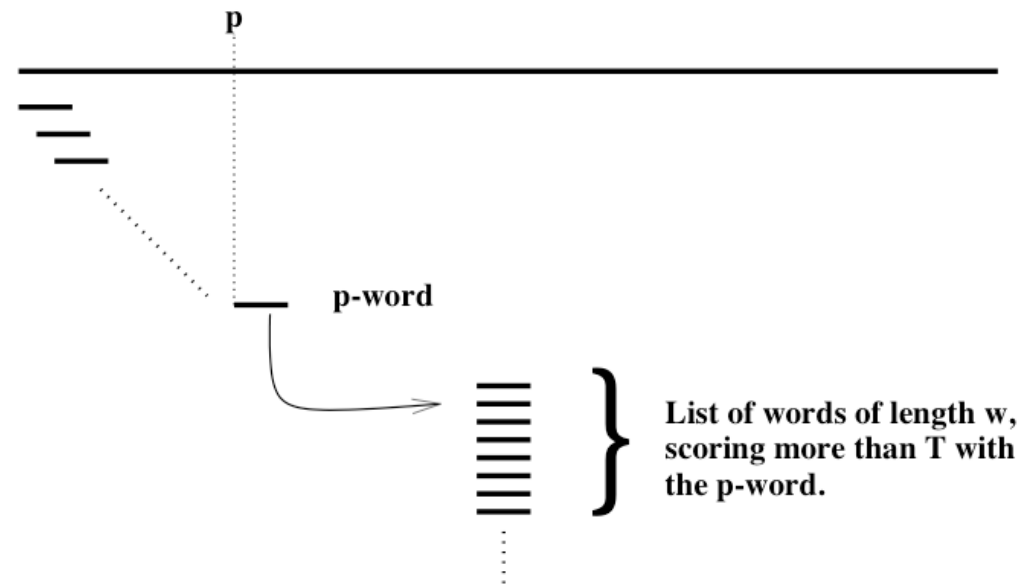
		K	Y	F	H	K	A	G	N	Q	H	S	P	T
	0	0	0	0	0	0	0	0	0	0	0	0	0	0
K	0	5	← 0	0	0	5	← 0	0	0	1	0	0	0	0
E	0	1	↖ 3	0	0	1	4	0	0	2	1	0	0	0
F	0	0	↖ 4	9	← 4	0	0	1	0	0	1	0	0	0
H	0	0	↖ 2	4	↑ 17	← 12	← 7	← 2	2	0	8	← 3	0	0
N	0	0	0	0	12	↑ 17	← 12	← 7	8	← 3	3	↑ 9	← 4	0
G	0	0	0	0	7	12	↑ 17	18	← 13	← 8	← 3	4	7	← 2
H	0	0	↖ 2	0	8	7	12	15	19	← 14	16	← 11	← 6	5
T	0	0	↖ 0	0	3	7	7	10	15	18	← 13	17	← 12	11

► Looking inside BLAST



► Looking inside BLAST

**A: For each position  $p$  of the query, find the list of words of length  $w$  scoring more than  $T$  when paired with the word starting at  $p$ :**



## ▶ Looking inside BLAST

query sequence: QLNFSAGW

(1) parameters

word length  $w = 2$

score threshold  $T = 8$

(2) determine all words of length  $w$  in the query sequence:

QL LN NF FS SA AG GW

(3) for each word determine a word list with an alignment score larger than (or equal to) the threshold  $T$ :

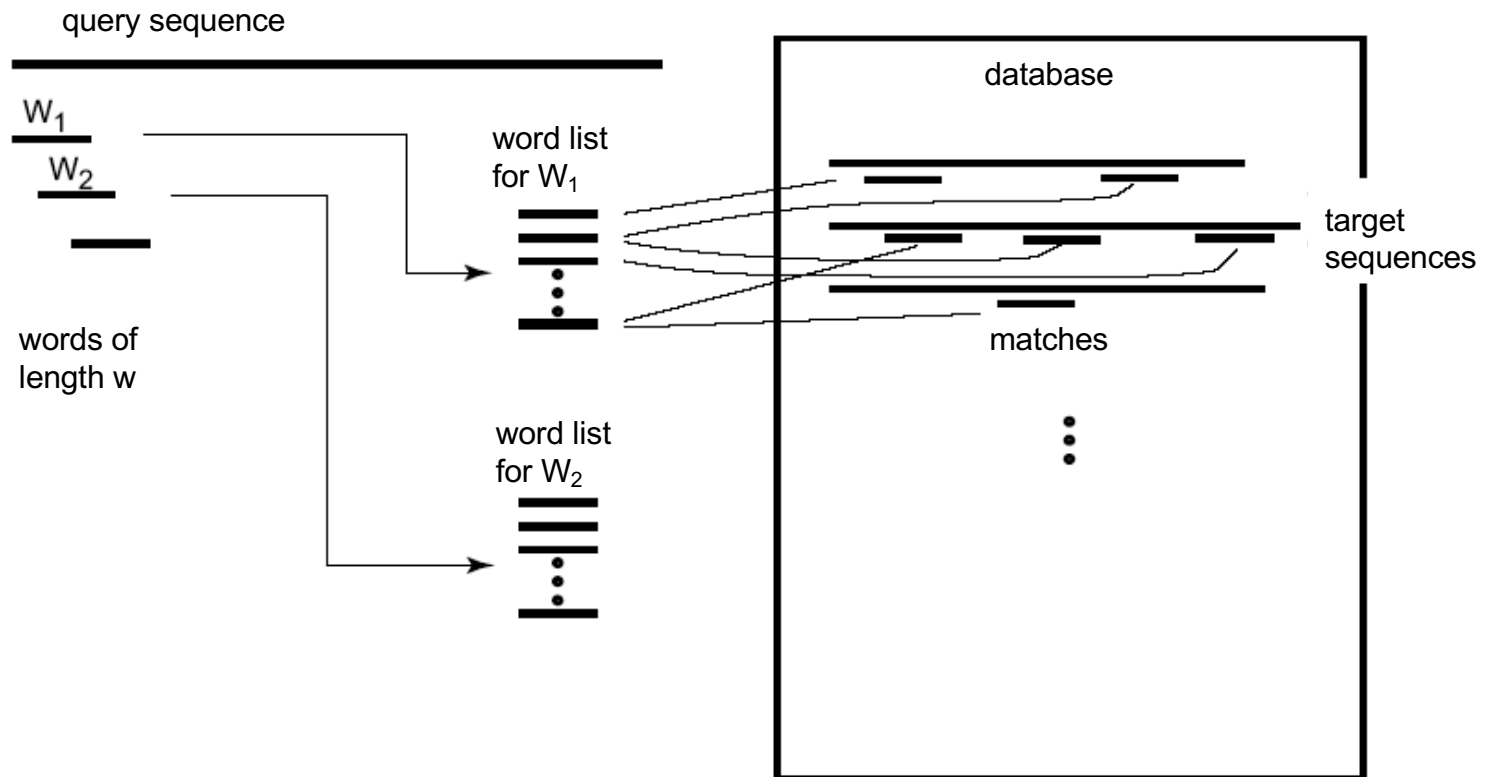
QL: QL=11, QM=9, HL=8, ZL=9

LN: LN=9, LB=8

NF: NF=12, AF=8, NY=8, DF=10, ...

...

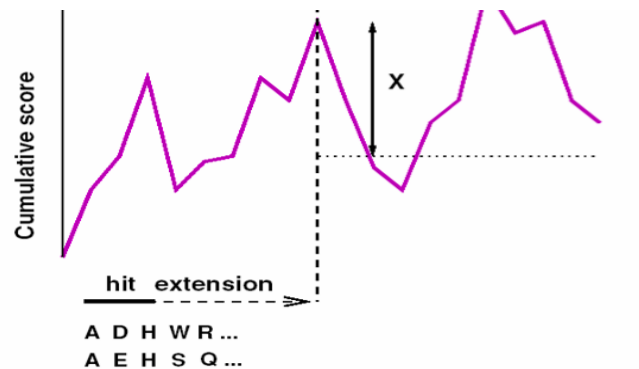
## ▶ Looking inside BLAST





► Looking inside BLAST

**C: For each word match («hit»), extend ungapped alignment in both directions. Stop when  $S$  decreases by more than  $X$  from the highest value reached by  $S$ .**



## ▶ Looking inside BLAST

▼ **Algorithm parameters**

**General Parameters**

<b>Max target sequences</b>	100	Select the maximum number of aligned sequences to display
<b>Short queries</b>	<input checked="" type="checkbox"/> Automatically adjust parameters for short input sequences	
<b>Expect threshold</b>	10	
<b>Word size</b>	3	

**Scoring Parameters**

<b>Matrix</b>	BLOSUM62	
<b>Gap Costs</b>	Existence: 11 Extension: 1	
<b>Compositional adjustments</b>	Composition-based statistics	

**Filters and Masking**

<b>Filter</b>	<input type="checkbox"/> Low complexity regions	
<b>Mask</b>	<input type="checkbox"/> Mask for lookup table only	
	<input type="checkbox"/> Mask lower case letters	

## ► Looking inside BLAST

### heuristic methods of sequence alignment

#### **FastA = fast Alignment**

D.J. Lipman and W.W.R. Pearson. Rapid and sensitive protein similarity searches. *Science*, 227:1435–1441, 1985.


W.R. Pearson and D.J. Lipman. Improved tools for biological sequence comparison. *Proc. Natl. Acad. Sci. USA*, 85:2444–2448, 1988.



#### **BLAST = Basic Local Alignment Search Tool**

S.F. Altschul, W. Gish, W. Miller, E.W. Myers, and D.J. Lipman. Basic local alignment search tool. *J. Mol. Biol.*, 215:403–410, 1990.

[a good introduction to these methods:  
Frédérique Galisson, The fasta and blast programs, 2002]

## ▶ Looking inside FastA

EMBL-EBI  EB-eye Search

Databases Tools EBI Groups Training Industry About Us Help [Site Index](#)  

- Help Index
- General Help
- Formats
- Gaps
- Matrix
- References
- FASTA Help
- MView Help
- VisualFASTA Help

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- View all FASTA's at EBI
- FASTA Programmatic Access


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- Database Information
  - UniProt
  - UniParc

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- Similar Applications
  - FASTA
  - BLAST
  - MPsrch
  - ScanPS
  - SSEARCH

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**FASTA Related Literature** 

Search for FASTA related literature in Medline... [more](#)

EBI > Tools > Similarity & Homology

### FASTA and SSEARCH - Protein Similarity Search

Provides sequence similarity searching against protein databases using the FASTA and SSEARCH programs. **SSEARCH** does a rigorous Smith-Waterman search for similarity between a query sequence and a database. **FASTA** can be very specific when identifying long regions of low similarity especially for highly diverged sequences. You can also conduct sequence similarity searching against [nucleotide databases](#) or complete [proteome/genome](#) databases using the [FASTA programs](#).

 [Download Software](#)

PROGRAM	DATABASES	RESULTS	SEARCH TITLE	YOUR EMAIL
FASTA	Protein UniProt Knowledgebase UniProtKB/Swiss-Prot UniProt Clusters 100% UniProt Clusters 100% (SEG filter)	interactive	Sequence	

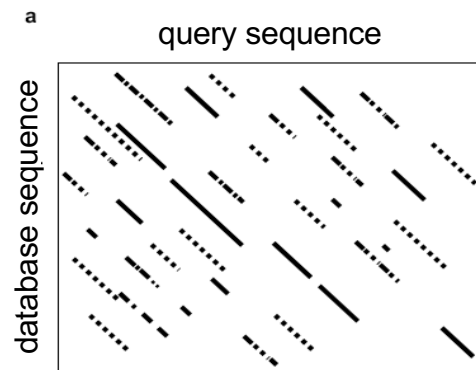
MATRIX	GAP OPEN	GAP EXTEND	KTUP	EXPECTATION UPPER VALUE	EXPECTATION LOWER VALUE
BLOSUM50	-10	-2	2	10.0	default

DNA STRAND	HISTOGRAM	MOLECULE TYPE
none	no	Protein

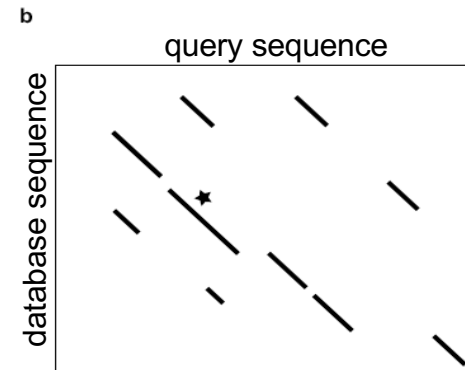
SCORES	ALIGNMENTS	SEQUENCE RANGE	DATABASE RANGE	FILTER	STATISTICAL ESTIMATES
50	50	START-END	START-END	none	Regress

Enter or Paste a  Sequence in any format:

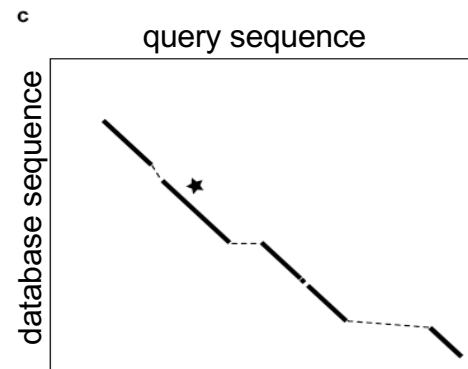
## ► Looking inside FastA



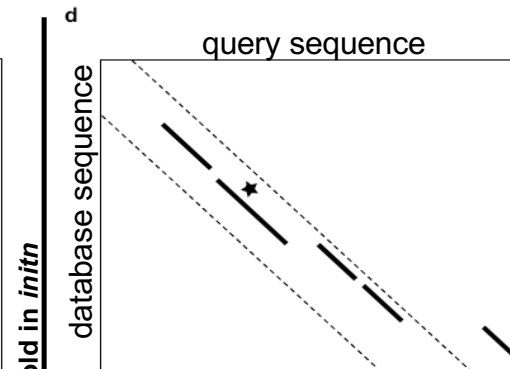
**Step 1:**  
Finding identical k-words



**Step 2:**  
Scoring of the regions with a PAM matrix;  
selection of highest scores (*init1*)



**Step 3:**  
Linking the segments  
with gaps (*initn* score)



**Step 4:**  
SW on a region of the plane; construction  
of the optimal alignment

## ► Looking inside FastA

remark on step 1

Query Sequence: WATSNANDCRICK

*ktup* = 1

1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
W	A	T	S	N	A	N	D	C	R	I	C	K		

Hashtable or  
lookup table:

A	C	D	I	K	N	R	S	T	W
2	9	8	11	13	5	10	4	3	1
6	12				7				

## ► Looking inside FastA

remark on step 1

# Target Sequence: BASEBALLANDCRICKET

Query Hashtable:

A	C	D	I	K	N	R	S	T	W
2	9	8	11	13	5	10	4	3	1
6	12				7				

Target  
Hashtable:

1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
B	A	S	E	B	A	L	L	A	N	D	C	R	I	C	K	E	T
	0	1			-4			-7	-5	-3	-3	-3	-3	-6	-3		-15
	4				0			-3	-3		0			-3			

Offset  
Table:

-15	-14	-13	-12	-11	-10	-9	-8	-7	-6	-5	-4	-3	-2	-1	0	1	2	3	4
1								1	1	1	1	6			3	1			1

			1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
			W	A	T	S	N	A	N	D	C	R	I	C	K		
1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
B	A	S	E	B	A	L	L	A	N	D	C	R	I	C	K	E	T

**... summary/repetition of probability models**





## Concepts from statistics

(probabilities, transition probabilities, Markov models)

### General idea:

Probability models can be seen as 'generators' of signal (e.g., DNA sequences) that can then be compared to real-life signals.

Probability models contain parameters.

Each 'run' of a probability model will give you (generally speaking) a different output. Observing a large number of such outputs may allow you to extract the underlying parameters.

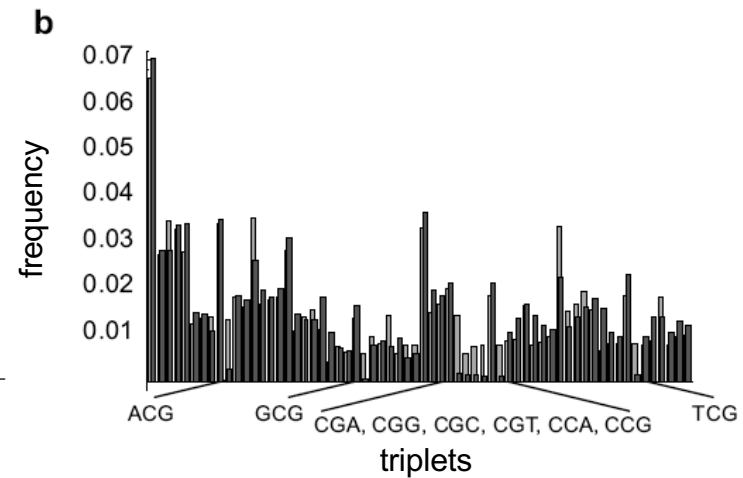
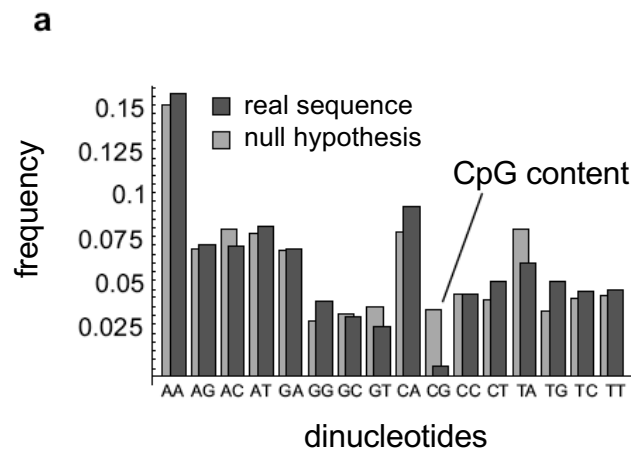
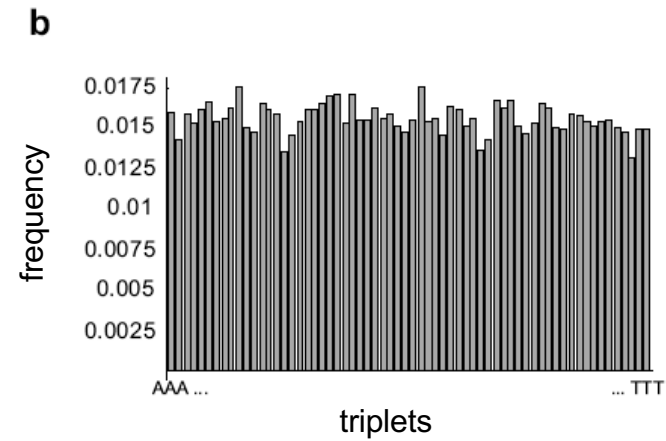
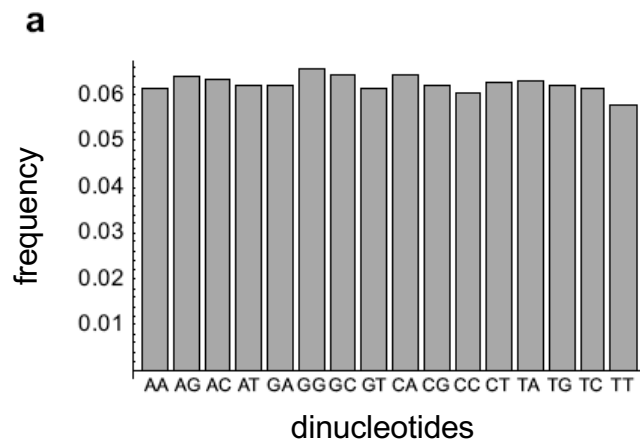
In the case of Markov models, these parameters are transition probabilities from one state (e.g., symbol in a sequence) to the next.

### Training:

Parameter estimation from data.

### Scoring:

What is the probability that a given model can produce a given sequence?





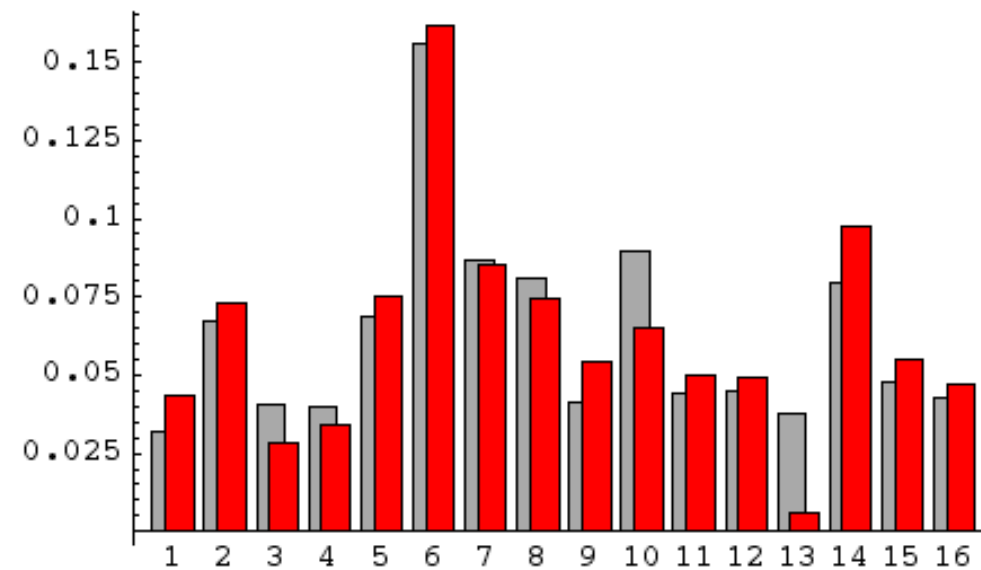
## DNA sequence from *Jurassic Park*

```
>JurassicPark DinoDNA from the book Jurassic Park
gcgttgctgg cgtttttcca taggtctcgc cccctgacg agcatcacia aaatcgacgc
ggtggcgaaa cccgacagga ctataaagat accaggcggt tccccctgga agctccctcg
tgttccgacc ctgccgetta cgggatacct gtccgccttt ctcccttcgg gaagcgtggc
tgctcacgct gtaggtatct cagttcgggt taggtcgttc gctccaagct gggctgtgtg
ccgttcagcc cgaccgctgc gccttatccg gtaactatcg tcttgagtcc aacccggtaa
agtaggacag gtgccggcag cgtctctgggt cattttcggc gaggaccgct ttcgctggag
atcggcctgt cgttgccggt attcggaatc ttgcacgccc tcgctcaagc ctctgctact
ccaaacgttt cggcgagaag caggccatta tcgccggcat ggccggccgac gcgctgggct
ggcgttcgag acgcgaggct ggatggcctt cccattatg attcttctcg ctccggcgag
cccgcgttgc aggccatgct gtccaggcag gtagatgacg accatcaggg acagcttcaa
cgctcttac cagcctaact tcgatcactg gaccgctgat cgtcacggcg atttatgccg
caagtcagag gtggcgaaac ccgacaagga ctataaagat accaggcggt tccccctgga
gcgctctcct gttccgaccc tgccgcttac cggatacctg tccgcctttc tcccttcggg
ctttctcatt gctcacgctg taggtatctc agttcgggtg aggtcgttcg ctccaagctg
acgaaccccc cgttcagccc gaccgctgcg ctttatccgg taactatcgt cttgagtcca
acacgactta acgggttggc atggattgta ggcgcgcgcc tataccttgt ctgcctcccc
gcggtgcatg gagccgggac acctcgacct gaatggaagc cggcggcacc tcgctaacgg
ccaagaattg gagccaatca attcttgcgg agaactgtga atgcgcaaac caacccttgg
ccatcgctgc cgccatctcc agcagccgca cgcggcgcat ctccggcagc gttgggtcct
gcgcatgata gtgctagcct gtcgttgagg acccggttag gctggcgggg ttgccttact
atgaatcacc gatacgcgag cgaacgtgaa gcgactgctg ctgcaaaacg tctgcgacct
atgaatggtc ttcggtttcc gtgtttcgta aagtctggaa acgcggaagt cagcgcctcg
```

Boguski, M.S.

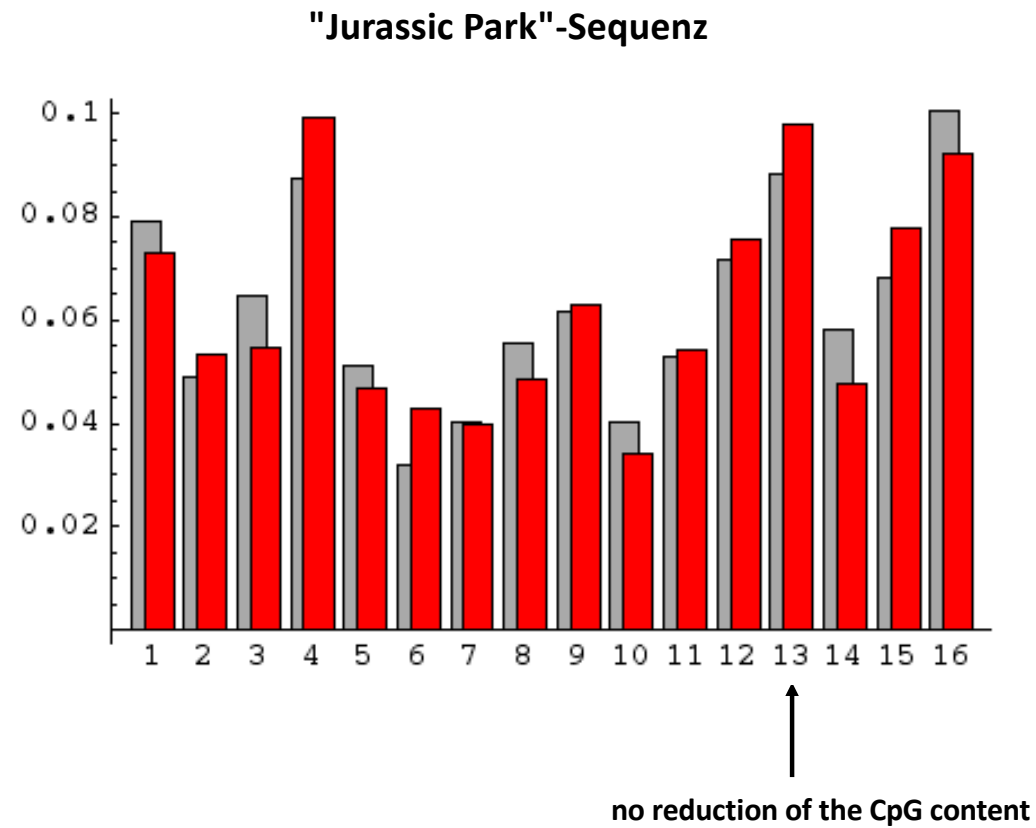
A Molecular Biologist Visits Jurassic Park. (1992) *BioTechniques* 12(5):668-669).

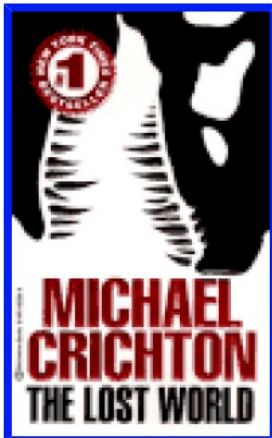
## elementary analysis of pair probabilities



reduction of the CpG content  
(lower probability for the dinucleotide "CG")

## elementary analysis of pair probabilities





## follow-up: "The Lost World"

```
>LostWorld DinoDNA from the book The Lost World
gaattccgga agcgagcaag agataagtcc tggcatcaga tacagttgga gataaggacg
gacgtgtggc agctcccgca gaggattcac tggaagtgca ttacctatcc catgggagcc
atggagttcg tggcgtggg ggggccggat gcgggtccc cactccgtt ccctgatgaa
gccggagcct tcctggggct gggggggggc gagaggacgg aggcgggggg gctgctggcc
tcctaccccc cctcaggccg cgtgtccctg gtgccgtggg cagacacggg tactttgggg
accccccagt ggggtgccgc cgcaccccaa atggagcccc cccactacct ggagctgctg
caaccccccc ggggcagccc cccccatccc tcctccgggc ccctactgcc actcagcagc
gggccccccac cctgcgaggc ccgtgagtgc gtcattggcca ggaagaactg cggagcgacg
gcaacgccgc tgtggcgccg ggacggcacc gggcattacc tgtgcaactg ggcctcagcc
tgccgggtct accaccgct caacggccag aaccgcccgc tcatccgcc caaaaagcgc
ctgcgggtga gtaagcgcgc aggcacagtg tgcagccacg agcgtgaaaa ctgccagaca
tccaccacca ctctgtggcg tcgcagcccc atgggggacc ccgtctgcaa caacattcac
gcctgcggcc tctactaaa actgcaccaa gtgaaccgcc ccctcacgat gcgcaaagac
ggaatccaaa ccgaaaccg caaagtttcc tccaagggtg aaaagcggcg cccccgggg
gggggaaacc cctccgccac cgcgggaggg ggcgtccta tggggggagg gggggacccc
tctatgcccc ccccgccgcc cccccggcc gccgcccccc ctcaaagcga cgctctgtac
gctctcggcc ccgtggtcct ttccggccat tttctgccct ttggaaactc cggagggttt
tttggggggg gggcgggggg ttacacggcc cccccggggc tgagcccgcg gatttaaata
ataactctga cgtgggcaag tgggccttgc tgagaagaca gtgtaacata ataatttgca
cctcggcaat tgcagagggt cgatctccac tttggacaca acagggtac tcggtaggac
cagataagca ctttgcctcc tggactgaaa aagaaaggat ttatctgttt gttcttgcct
gacaaatccc tgtgaaagg aaagtcgga cacagcaatc gattatttct cgcctgtgtg
aaattactgt gaattattga aatatatata tatatatata tatatctgta tagaacagcc
tcggaggcgg catggaccca gcgtagatca tgctggattt gtactgccgg aattc
```

follow-up: "The Lost World"

NCBI *nucleotide-nucleotide* **BLAST**

Nucleotide Protein Translations Retrieve results for an RID

[Search](#)

[Set subsequence](#) From:  To:

[Choose database](#)

Now: **BLAST!** or **Reset query** **Reset all**

### follow-up: "The Lost World"

```
>sp|P23770|GAT2_XENLA TRANSCRIPTION FACTOR XGATA-2 (GATA BINDING FACTOR-2)
  pir||C41602 transcription factor GATA-2 - African clawed frog
  gb|AAA49723.1| (M76564) GATA binding factor-2 [Xenopus laevis]
      Length = 452

Score = 193 bits (485), Expect = 4e-48
Identities = 92/124 (74%), Positives = 103/124 (82%)
Frame = +1

Query: 436 EARECVMARKNCGATATPLWRRDGTGHYLCN WASACGLYHRLNGQNRPLIRPKRLRVSK 615
      E RECV      NCGATATPLWRRDGTGHYLCN  ACGLYH++NGQNRPLI+PK+RL  ++
Sbjct: 263 EGRECV----NCGATATPLWRRDGTGHYLCN---ACGLYHKMNGQNRPLIKPKRRLSAAR 315

Query: 616 RAGTVCSHERENCQTSTTTTLWRRSPMGDPVCNNIHACGLYYKLHQVNRPLTMRKDG IQTR 795
      RAGT C+      NCQTSTTTTLWRR+  GDPVCN  ACGLYYKLH VNRPLTM+K+GIQTR
Sbjct: 316 RAGTCCA----NCQTSTTTTLWRRNANGDPVCN---ACGLYYKLHNVNRPLTMKKEGIQTR 368

Query: 796 NRKV 807
      NRK+
Sbjct: 369 NRKM 372
```



## Markov chains as a tool for studying CpG islands

+	A	C	G	T	-	A	C	G	T
A	0.180	0.274	0.426	0.120	A	0.300	0.205	0.285	0.210
C	0.171	0.368	0.274	0.188	C	0.322	0.298	0.078	0.302
G	0.161	0.339	0.375	0.125	G	0.248	0.246	0.298	0.208
T	0.079	0.355	0.384	0.182	T	0.177	0.239	0.292	0.292

$$S(x) = \log \left( \frac{P(x | \text{model } +)}{P(x | \text{model } -)} \right) = \log \left( \frac{P(B) \prod_{i=1}^L a_{x_{i-1}x_i}^+}{P(B) \prod_{i=1}^L a_{x_{i-1}x_i}^-} \right) = \sum_{i=1}^L \log \left( \frac{a_{x_{i-1}x_i}^+}{a_{x_{i-1}x_i}^-} \right) = \sum_{i=1}^L \beta_{x_{i-1}x_i}$$

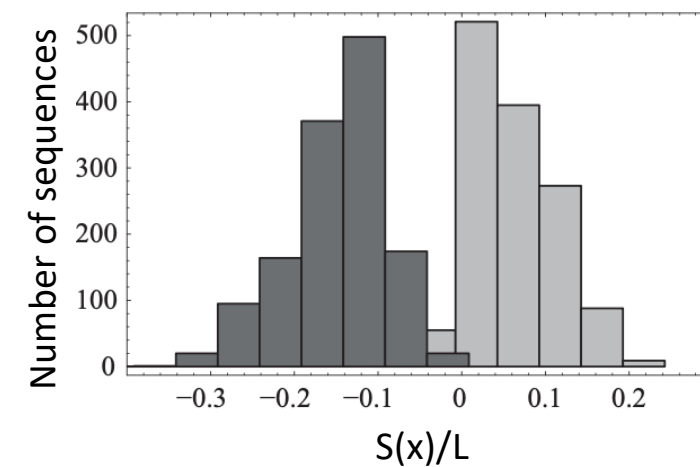
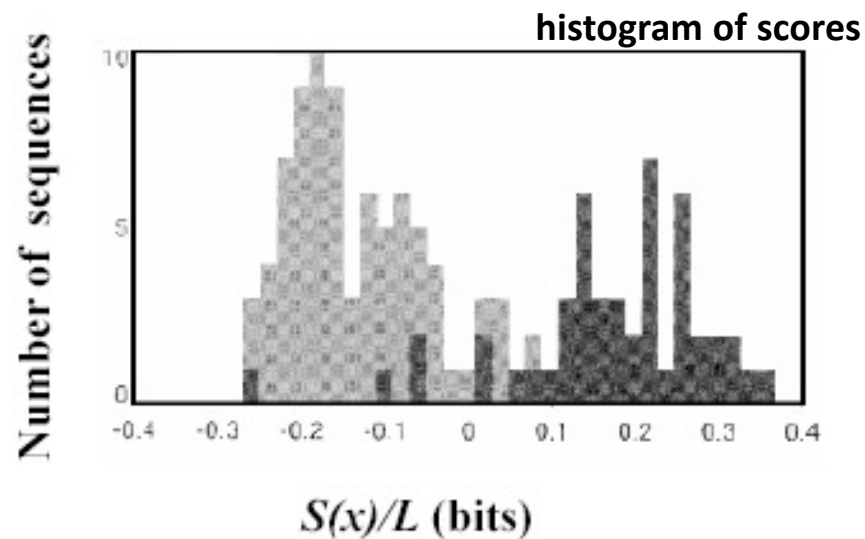
a number for each sequence  $x$   
 → histogram of score values  $S(x)$   
 for many sequences  $x$

a number for each dinucleotide  
 → table of "log-likelihoods"

## Markov chains as a tool for studying CpG islands

$\beta(\log_2)$	A	C	G	T
A	-0.740	0.419	0.580	-0.803
C	-0.913	0.302	1.812	-0.0685
G	-0.624	0.461	0.331	-0.730
T	-1.169	0.573	0.393	-0.679

table of  
"log-likelihoods"

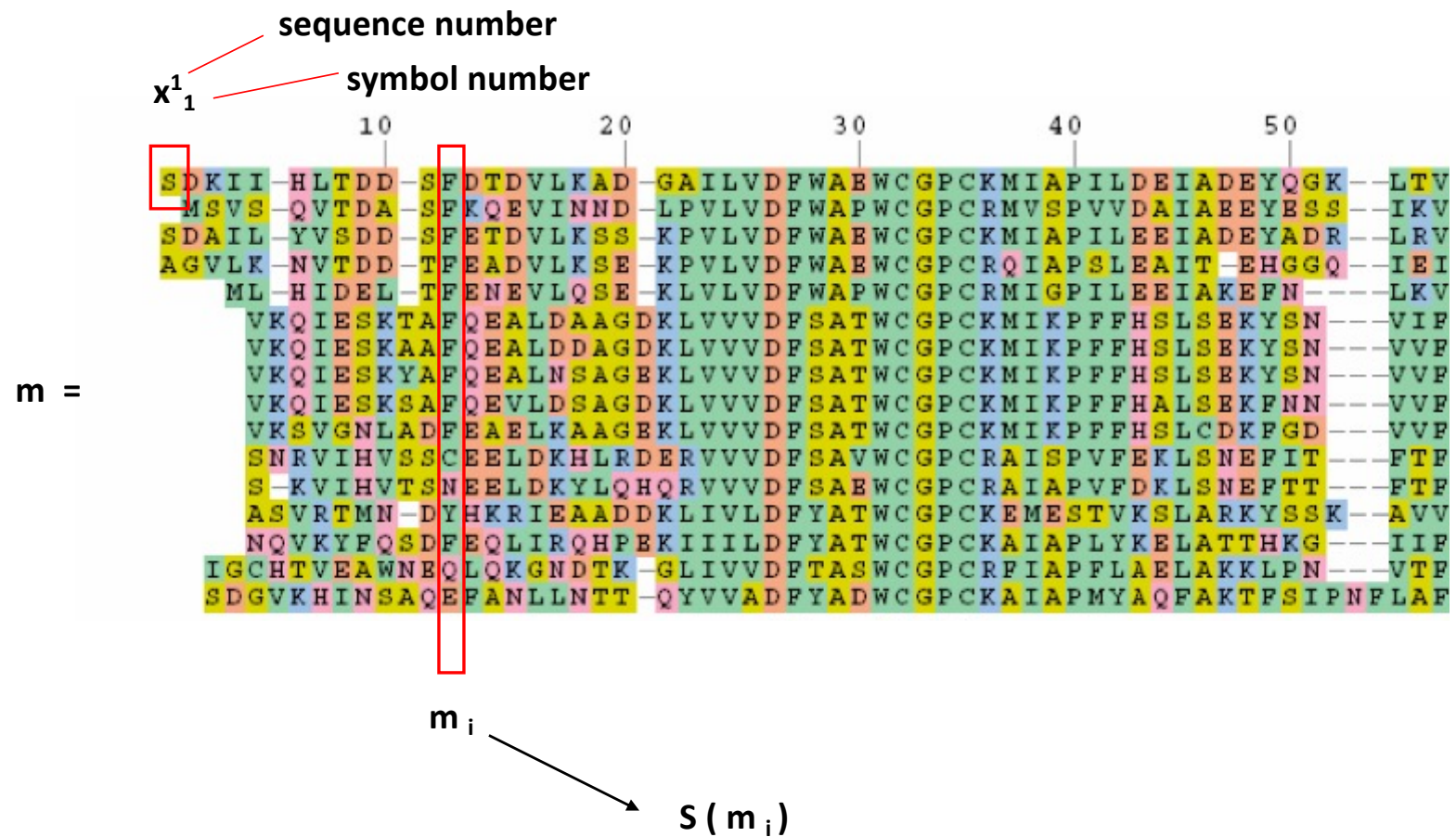


## Multiple sequence alignment

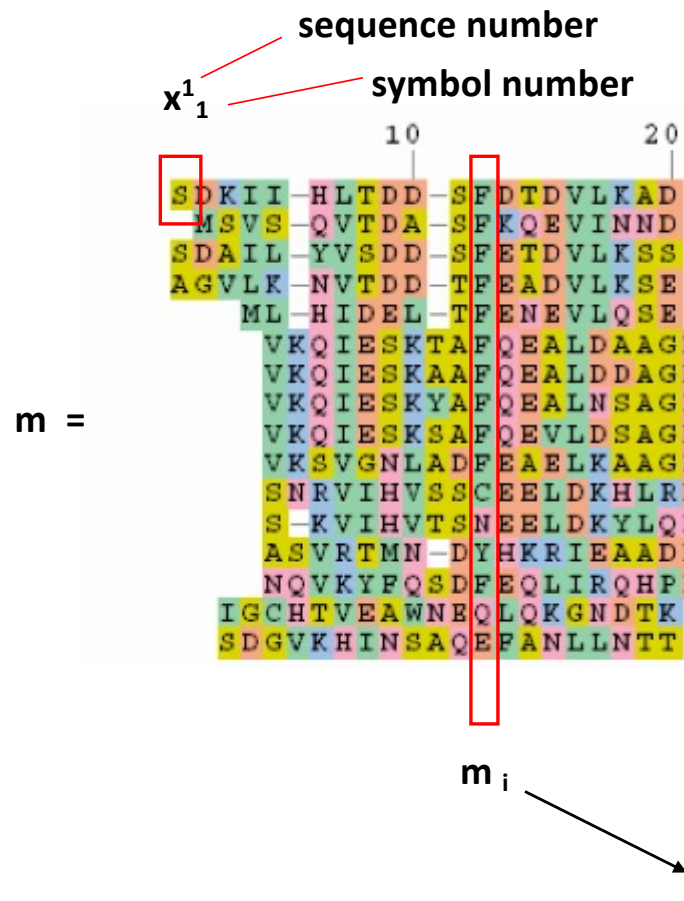
*Escherichia coli*  
*Porphyra purpurea*  
*Thiobacillus ferrooxidans*  
*Streptomyces clavuligerus*  
*Cyanidioschyzon merolae*  
 Human  
 Rhesus monkey  
 Sheep  
 Rabbit  
 Chicken  
*Dictyostelium discoideum*  
*Dictyostelium discoideum*  
*Drosophila melanogaster*  
*Caenorhabditis elegans*  
*Ricinus communis*  
*Neurospora crassa*

*Escherichia coli*  
*Porphyra purpurea*  
*Thiobacillus ferrooxidans*  
*Streptomyces clavuligerus*  
*Cyanidioschyzon merolae*  
 Human  
 Rhesus monkey  
 Sheep  
 Rabbit  
 Chicken  
*Dictyostelium discoideum*  
*Dictyostelium discoideum*  
*Drosophila melanogaster*  
*Caenorhabditis elegans*  
*Ricinus communis*  
*Neurospora crassa*

[illegible]



Sum of pairs (SP) score



$$S(m_i) = \sum_{j < k} s(m_i^j, m_i^k)$$

$$s(a, -) = s(-, a) = -d, \quad s(-, -) = 0$$

$$\alpha_{i_1, i_2, \dots, i_N} = \max \begin{cases} G_0 \\ G_1 \\ G_2 \\ \vdots \\ G_{N-1} \end{cases},$$

$$G_0 = \alpha_{i_1-1, i_2-1, i_3-1, \dots, i_N-1} + S(x_{i_1}^1, x_{i_2}^2, x_{i_3}^3, \dots, x_{i_N}^N)$$

$$G_1 = \begin{cases} \alpha_{i_1, i_2-1, i_3-1, \dots, i_N-1} + S(-, x_{i_2}^2, x_{i_3}^3, \dots, x_{i_N}^N) \\ \alpha_{i_1-1, i_2, i_3-1, \dots, i_N-1} + S(x_{i_1}^1, -, x_{i_3}^3, \dots, x_{i_N}^N) \\ \vdots \\ \alpha_{i_1-1, i_2-1, i_3-1, \dots, i_N} + S(x_{i_1}^1, x_{i_2}^2, x_{i_3}^3, \dots, -) \end{cases}$$

$$G_2 = \begin{cases} \alpha_{i_1, i_2, i_3-1, \dots, i_N-1} + S(-, -, x_{i_3}^3, \dots, x_{i_N}^N) \\ \alpha_{i_1-1, i_2, i_3, \dots, i_N-1} + S(x_{i_1}^1, -, -, \dots, x_{i_N}^N) \\ \vdots \end{cases}$$



**step 1**

$\{S_1, S_2, S_3, S_4, S_5\}$



$S_1$      $S_1$     ...     $S_4$   
 $S_2$      $S_3$              $S_5$

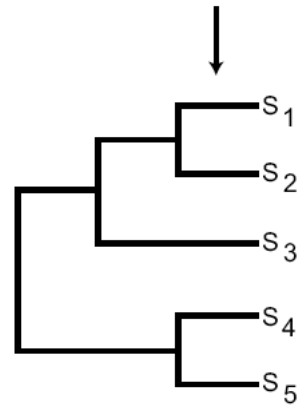
**step 2**



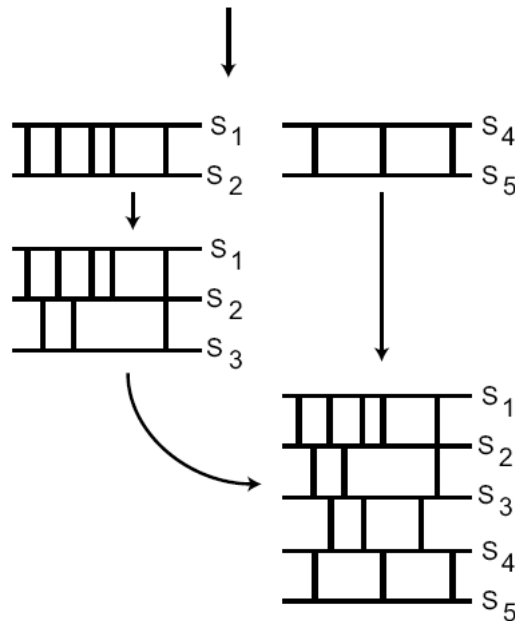
	$S_1$	$S_2$	$S_3$	$S_4$	$S_5$
$S_1$	0	$a_{12}$	$a_{13}$	$a_{14}$	$a_{15}$
$S_2$	$a_{21}$	0	$a_{23}$	$a_{24}$	$a_{25}$
$S_3$	$a_{31}$	$a_{32}$	0	$a_{34}$	$a_{35}$
$S_4$	$a_{41}$	$a_{42}$	$a_{43}$	0	$a_{45}$
$S_5$	$a_{51}$	$a_{52}$	$a_{53}$	$a_{54}$	0



step 3



step 4



**a**

```

KEFHN - G - - H - - T
|  |  |  |  |  |  |
KYFHKAGNQHSPT

```

S=11

**b**

```

KEFH - - - NGHT
|  |  |  |  |  |
KYFHKAGNGHT

```

S=27

**c**

```

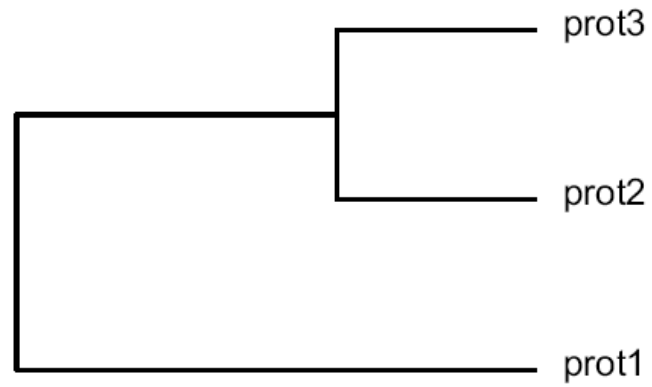
KYFHKAGNQHSPT
| | | | | | | | | |
KYFHKAGNGH - - T

```

S=48

$$d = -\log \frac{S - S_{rand}}{S_{max} - S_{rand}}$$

	prot1	prot2	prot3
prot1	0	1.30429	0.75107
prot2	1.30429	0	0.393446
prot3	0.75107	0.393446	0



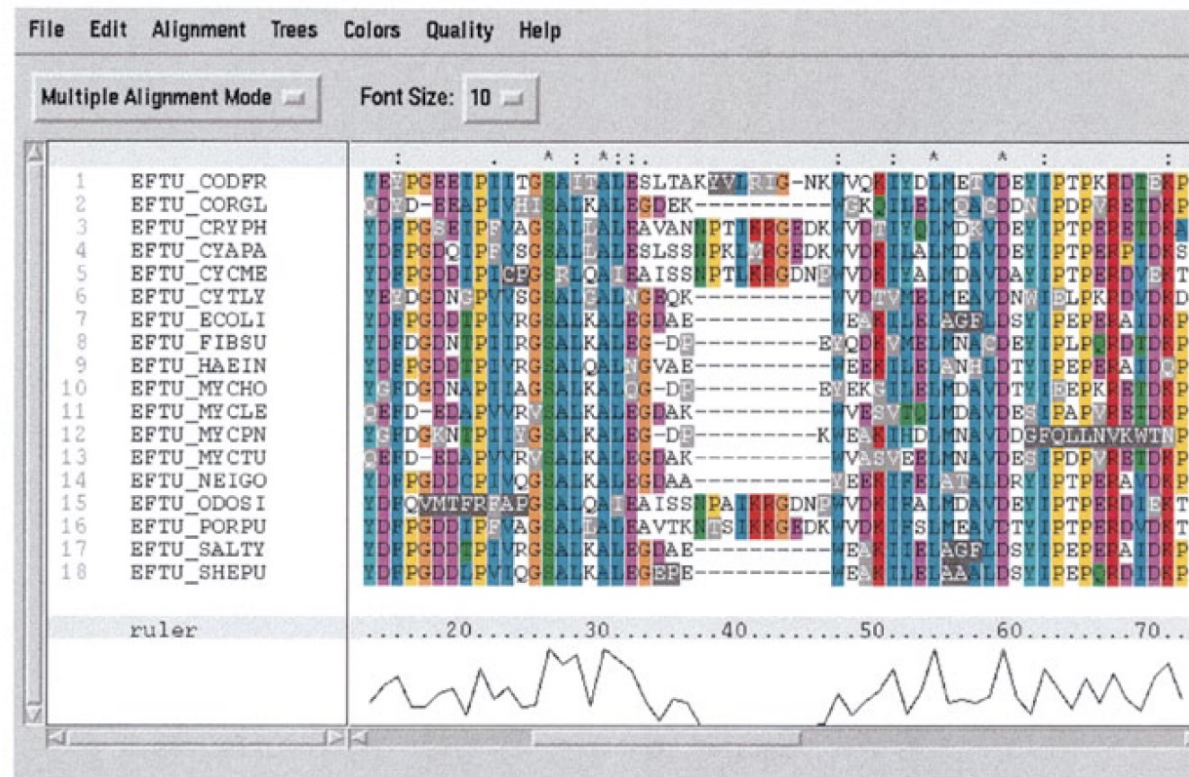
prot2	KYFHKAGNQHSPT
prot3	KYFHKAGNGH - - T
prot1	KEFH - - - NGH - - T

## The CLUSTAL\_X windows interface: flexible strategies for multiple sequence alignment aided by quality analysis tools

Julie D. Thompson, Toby J. Gibson<sup>1</sup>, Frédéric Plewniak, François Jeanmougin\* and Desmond G. Higgins<sup>2</sup>

### ABSTRACT

CLUSTAL X is a new windows interface for the widely-used progressive multiple sequence alignment program CLUSTAL W. The new system is easy to use, providing an integrated system for performing multiple sequence and profile alignments and analysing the results. CLUSTAL X displays the sequence alignment in a window on the screen. A versatile sequence colouring scheme allows the user to highlight conserved features in the alignment. Pull-down menus provide all the options required for traditional multiple sequence and profile alignment. New features include: the ability to cut-and-paste sequences to change the order of the alignment, selection of a subset of the sequences to be realigned, and selection of a sub-range of the alignment to be realigned and inserted back into the original alignment. Alignment quality analysis can be performed and low-scoring segments or exceptional residues can be highlighted. Quality analysis and realignment of selected residue ranges provide the user with a powerful tool to improve and refine difficult alignments and to trap errors in input sequences. CLUSTAL X has been compiled on SUN Solaris, IRIX5.3 on Silicon Graphics, Digital UNIX on DECstations, Microsoft Windows (32 bit) for PCs, Linux ELF for x86 PCs, and Macintosh PowerMac.



**Figure 1.** The CLUSTAL X window in multiple alignment mode. An alignment of some EFTU proteins is displayed. Low-scoring segments are highlighted using a white character on a black background. Exceptional residues are shown as a white character on a grey background. The quality analysis reveals two anomalously low scoring regions, ruler positions 16–25 in EFTU\_ODOSI and 61–71 in EFTU\_MYCPN. These were found to be caused by frameshift errors. Two more sequences (EFTU\_RICPR and EFTU\_SPIPL), not shown here, have 4-residue sequencing errors in this region which CLUSTAL X will also highlight.

# RESEARCH ARTICLE

## Crystal Structure of the Potassium Channel KirBac1.1 in the Closed State

Anling Kuo,<sup>1</sup> Jacqueline M. Gulbis,<sup>2</sup> Jennifer F. Antcliff,<sup>3</sup> Tahmina Rahman,<sup>1</sup> Edward D. Lowe,<sup>1</sup> Jochen Zimmer,<sup>1</sup> Jonathan Cuthbertson,<sup>1</sup> Frances M. Ashcroft,<sup>3</sup> Takayuki Ezaki,<sup>4</sup> Declan A. Doyle<sup>1\*</sup>

K<sup>+</sup> channels are involved in a wide range of physiological processes, such as propagation of the action potential, cardiac function, K<sup>+</sup> reabsorption in the kidney, and hormone regulation (1, 2). This diversity is possible because many different signals can open or close K<sup>+</sup> channels, a process known as gating. The signals are received by domains attached to the pore-forming subunit.

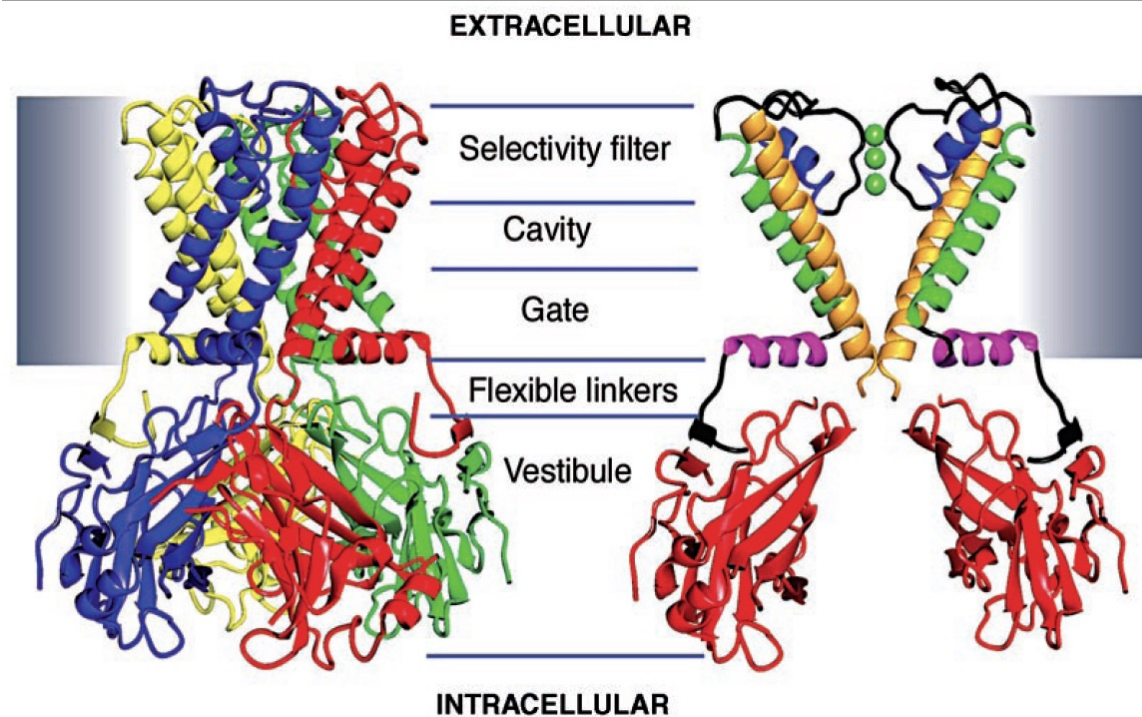
We present a complete K<sup>+</sup> channel structure that shows the nature of the physical link coupling domains that receive gating signals to the transmembrane helices.

20 JUNE 2003 VOL 300 SCIENCE [www.sciencemag.org](http://www.sciencemag.org)

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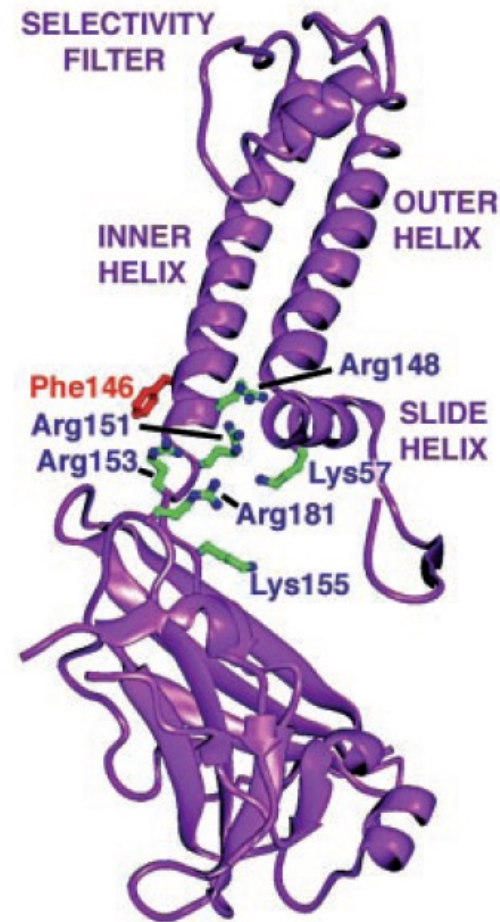




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Rahman,<sup>1</sup> Edward D. Lowe,<sup>1</sup> Jochen Zimmer,<sup>1</sup> Jonathan  
Cuthbertson,<sup>1</sup> Frances M. Ashcroft,<sup>2</sup> Takayuki Ezaki,<sup>4</sup>  
Declan A. Doyle<sup>1\*</sup>

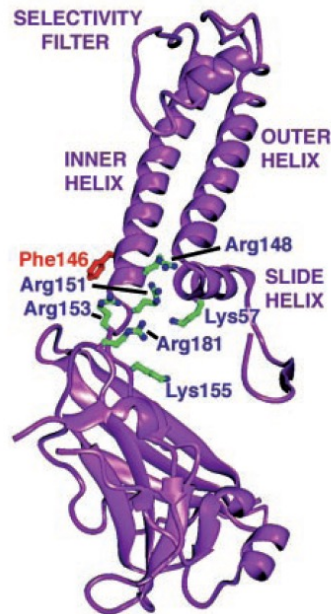




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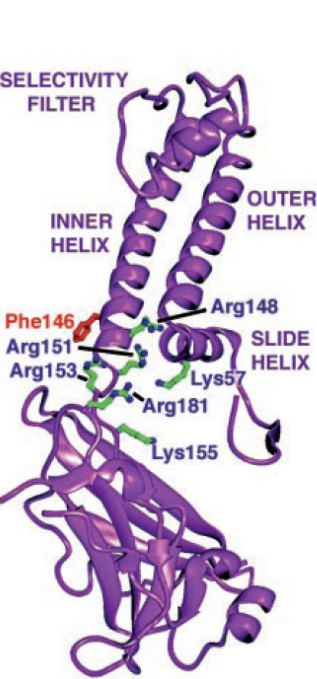


	Slide helix	Outer helix			
KirBac1.1	-----REVIAVGMPPASVWRLFYWALKVSNPVPFASLAVLVVNNLTPALLYQLGDAPT	89			
Human_Kir1.1	-----FGNVRAQSRPIFFVDIWTTLVDLKNRYKMTIPITAFGLSWFFPGLLWYAVAYIH	106			
Human_Kir2.1	-----QFINVGEKGGQRYLADITTTCDVDIRNRWMLVIFCLAFVLWSLFGGCVFVLLIALH	110			
Human_Kir3.1	-----QHGNLGSSTSKYLSDLFTTLVDLKNRWNLFIPLTYTVAWLFMASMMWVIATYR	109			
Human_Kir3.4	-----HHGNVQETRYLSDLFTTLVDLKNRWNLVFTMVYTVTWLFFGPIWVLLIAYIR	115			
Human_Kir4.1	-----RMEHIAADKRFYLDLWTTTFIDMKNRYKLLPSATFAGTWLFGVWVYLVAVAH	93			
Human_Kir5.1	-----YFKHIFGEWGSYVVDITFTLVDTKWRHMFVIFSLSYLSWLIFGSGVWVLIAPFH	99			
Human_Kir6.2	-----AHKNIREQGRFLQDVFTTLVDLKNWPHLTLLITMSFLCSWLLFAMAWVLIAPAH	97			
Human_Kir7.1	-----QMDGAQGLALYLRDAGILMDWRWRMMLVFSASFVWVHLVFAVLVLAEMN	82			
KcsA	-----MAPMLSGLLARLVKLLLGRHGSALHWAAGATVLLVIVLLAGSLAVLAERG	53			
MthK	-----MVLVIEIIRKHLPRVLKV-PATRILLVLAVIITYGTAGPHEITEGE	44			
Shaker	-----GLQILGRTLKASMRRLGLLIFLFGVLLFSSAVYFAEAGSE	422			
	Turret	Pore helix	Filter	Inner helix	
KirBac1.1	AN-----	90	110	130	
Human_Kir1.1	K-DLPEPHPSANHTPCVENINGLTSAPLFSLEITQVTIGYGRFVTEQCATAIPLLIQGS	164			
Human_Kir2.1	G-DLDAS--KGKACVSEVNSFTAAFLFSLEITQVTIGYGRFVTEQCATAIPLLIQGS	165			
Human_Kir3.1	G-DLNKAH-VGNYTPCVANVNFPSAPLFFIETETATIGYGRYITDKCPGIIILFLFS	166			
Human_Kir3.4	G-DLDHVG-DQEWIPCVENLSGFVSAPLFSIETETATIGYGRVITEKCPGIIILLLVQA	172			
Human_Kir4.1	G-DLLEDPPANHTPCVQVHTLTGAPLFSLESQTIGYGRYISEECPALIVLLIAQL	151			
Human_Kir5.1	G-DLLND--PDITPCVDNVHSFTGAPLFSLEITQVTIGYGRFVTEECVAVMLVLQS	154			
Human_Kir6.2	G-DLAPS--EGTAEPVTSIHSFSSAPLFSIEVQVTIGYGRMVTEECPLAILSLIVQN	153			
Human_Kir7.1	GDLELDHDAPPENHTICVKYITSFTAAFSFSLTQLTIGYGTMPFSGDCPSAIIALLAQM	142			
KcsA	AP-----GAQLITYPRALWWSVETATTIGYGDLYP-VTLWGRCAVVVM	83			
MthK	-----SWTVSLYWTFTIATIGYGDYSP-STPLGMVFVETLLI	80			
Shaker	N-----SFFKSLIPDAFWWAVVMTTIGYGDMTF-VGVNGKIVGSLCA	463			
	Inner helix				
KirBac1.1	FVGMGSIALSTGLVFARFARPRAK---IMFARHAIVRFPNGRMVLMVRAANARQNVIAEA	188			
Human_Kir1.1	ILGVIIINSFMCAGAILAKISRPKKRAKTTITFSKNAVISKRGGKCLLIRVANLRKSLIIGS	224			
Human_Kir2.1	IVGCIIDAFIIGAVMAKMAKPKKRNETLVFSHNAVIAMRDGGKCLLMWRVGNLRKSHLVSA	225			
Human_Kir3.1	ILGSIIVDAFLICCMFVKMSQPKKRAETLMFSEHAVISMRRDGGKCLLMFVRVGNLRSHMVSA	226			
Human_Kir3.4	ILGSIIVDAFLICCMFVKMSQPKKRAETLMFSEHAVISMRRDGGKCLLMFVRVGNLRSHMVSA	232			
Human_Kir4.1	VLITILELPIGTTFIAKIAKPKKRAETIRFSQHAVASHNGKPCMLIRVANMRKSLIIGC	211			
Human_Kir5.1	ILSCIINTFIIGAADAKMATARKRAQTIRFSYPALIGMRDGGKCLLMWRIGDFRPNHVRG	214			
Human_Kir6.2	IVGLMINAIMLGCIFMKTAQAHRAETLIFSKHAVIALRHGRLCFMLRVGDLRKSIMIIISA	213			
Human_Kir7.1	LLGLMLEAFITGAFVAKIARPKNRAFRIETDTAVVAHMDGKPNLIFQVANTRPSPLTSV	202			
KcsA	VAGITSFGLVTAALATFVGRQERRGH-----	124			
MthK	VLGIGTFFAVAVERLLEFLINREQMK-----	105			
Shaker	TAGVLTIALPVFVIVSNFNFYFHRETD-----	490			
KirBac1.1	190	210	230		
Human_Kir1.1	RAKMRIMRREHSSEG---YSLMKTHDLKLVNRNEHPIFLLGNMMHVIDESSPLFGETPE	244			
Human_Kir2.1	HIYGLKLLKTTVTPEGETIILDOININFPVVDAGNENLFFISPLTIYHVIDHNSPPFHMA-A	283			
Human_Kir3.1	HVRAQLLKSRTISEGEYIPLDQIDINVGFDSDGIDRFLVSPITIVHEIDEDSPPLYDLRSQ	285			
Human_Kir3.4	QIRCKLLKSRTPEGEFLPLDQLELDVGFSTGADQLFLVSPITICHVIDAKSPPLYDLRSQ	286			
Human_Kir4.1	SIRAKLLKSRTKEGEFLPLNQDINVGFDGDDRLFLVSPITISHEINEKSPPLYDLRSQA	292			
Human_Kir5.1	QVTGKLLQTHQTEGENIRLNQVNVTFQVDTASDSPFLILPLTFYHVVDETSPDKDLP-L	270			
Human_Kir6.2	TVRAQLLRYTDESEG-RMTMAFKDKLVND---QIILVTPVTIVHEIDHESPLYALDRK	269			
Human_Kir7.1	TIHQVVRKTTSPGEVVPVLIQVDIPMENGVGNGNSLPLVAPLIIYHVIDANSPLYDLAPS	273			
	RVSAYLYQER---ENGLYQTSVDFPHLDGISDECPFFIFPLTYHISITPSSPLATLLQH	259			
KirBac1.1	250	270	290		
Human_Kir1.1	SLAE-GRAMLLVMIEGSDETTAQVMQARHAWEHDDIRWHRYVDLMSDVD-GMTHIDYTR	302			
Human_Kir2.1	ETLLQQDPFELVFLDQTVESATCQVRTSYVPEVVLWGYRFAPIVSKTEKGYRVVDPHN	343			
Human_Kir3.1	DIIDN-ADPEIVVILGGMVEATMETQCSRSSYLANELWGRHRYEPVLFEEK-HYKVDYSR	343			
Human_Kir3.4	SMQT-EQPEIVVILGIVETTGMTQCARSTYDEVLWGRHFPFVVISLEE-GPFKYVYSQ	344			
Human_Kir4.1	QLHQ-EFEVNVVILGEMVEATGMTQCARSSYMDTEVLWGRHFTPVVLTLEK-GPYEVDYNT	350			
Human_Kir5.1	RSGE-GDFELVLLSGTVESTSATCQVRTSYLPEELLWGYEFTPAISLSASGKYIADFSL	329			
Human_Kir6.2	AVAK-DNPEILVTFIYTGDSGTSHQSRSSYVPREILWGRHFNVDVLEVKR-KYYKVNCLQ	327			
Human_Kir7.1	DLHHHQDLLEIIVILEGVVETTGITTQARTSYLADEILWGRFVFPVIAEED-GRYSVDYSK	332			
	ENPS--HFELVVFLSAMQEGTGETICQRTSYLPSEIMLHRCFASLLTRGSKGEYQIKMEN	317			
KirBac1.1	FNDTEPV	309			
Human_Kir1.1	FSKTVEV	350			
Human_Kir2.1	PHKTYEV	350			
Human_Kir3.1	PHATFEV	351			
Human_Kir3.4	FHDYET	357			
Human_Kir4.1	FDQVVKV	336			
Human_Kir5.1	FEQSVVEV	334			
Human_Kir6.2	FGNTIKV	339			
Human_Kir7.1	FDKTVPE	324			

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KirBac1.1  
Human\_Kir1.1  
Human\_Kir2.1  
Human\_Kir3.1  
Human\_Kir3.4  
Human\_Kir4.1  
Human\_Kir5.1  
Human\_Kir6.2  
Human\_Kir7.1  
KcsA  
MthK  
Shaker

KirBac1.1  
Human\_Kir1.1  
Human\_Kir2.1  
Human\_Kir3.1  
Human\_Kir3.4  
Human\_Kir4.1  
Human\_Kir5.1  
Human\_Kir6.2  
Human\_Kir7.1  
KcsA  
MthK  
Shaker

Slide helix		Outer helix	
	50	70	
-----	REVIA	YGMPAS	VWRDLYYWALKVSWPVFFASLAVLFVNNNTLFALLYQLGDAP I 89
-----	FGNVEA	QSRFIF	FFVDIWTTVLDLKWRYKMTIFITAFSGSWFFFGLLWYAVAYIH 106
-----	QFINV	GKGRYL	ADIFTTCVDIRWRWMLVIFCLAFVLSWLFPGCVFWLIALLH 110
-----	QHGNL	GSETSR	YLSDLFTTLVDLKWWRNLFIFILTYTVAWLFMASMWWVIAYTR 109
-----	HHGNV	QETRYL	SDLFTTLVDLKWRFNLLVFTMVYTVTWLFFGFIWWLIAYIR 115
-----	RMEHI	ADKRFL	YLYKDLWTTTFIDMQWRYKLLLSATFAGTWFLFGVVWYLVAVAH 93
-----	YFKHI	FGWGSY	VVDIFTTLVDTKWRHMFVIFSLSYILSWLIFGSVFWLIAPFH 99
-----	AHKNI	REQGRF	LQDVFTTLVDLKWPHLLIFTMSFLCSWLLFAMAWWLIAPAH 97
-----	QMDGA	QRLAYL	RDAGWILMDMRWRWMLVFSASFVHVHLVFAVLWYVLAEMN 82
-----	MAPML	SGLLAR	LVLKLLGRHGSALHWRAGAATVLLVIVLLAGSYLAVLAERG 53
-----	MVLVI	EIRKHL	PRVLKV-PATRILLVLAVIITYGTAGFHEIEGE 44
-----	GLQIL	GRTLKAS	MRELGLLIEFFLFIGVVLFSASVYFAEAGSE 422
Turret		Pore helix	
		Filter	
		Inner helix	
90	AN-----	QSPPGF	VGAFFFSVETLATVGYGDMHP--QTVYAHAIATLEI 131
	K--DLPE	FHPSAN	HTPCVENINGLTSAFLFSLETQVTIGYGFRCVTEQCATAIFLLIFQS 164
	G--DLDA	S---KEG	KACVSEVNSFTAFLFSIETQTTIGYGFRCVTDECPPIAVFMVVFQS 165
	G--DLNKA	H-VGNY	TPCVANVYNFPFAFLFFIETEATIGYGYRYITDKCPEGIILFLFQS 166
	G--DLDHV	G-DQEW	IPCVENLSGFVSAFLFSIETETTIGYGF RVITEKCEPIIILLVQA 172
	G--DLLEL	DPPAN	HTPCVVQVHTLTGAFLFSLESQTTIGYGF RYISEECPLAIVLLIAQL 151
	G--DLLND	---PD	ITPCVDNVHSFTGAFLFSLETQTTIGYGYRCVTEECVAVLMVILQS 154
	G--DLAPS	---EG	TAEPCVTSIHSFSAFLFSIEVQVTIGFGGRMVTEECPLAILSLIVQN 153
	GDLELD	HDAPP	NHTICVKYITSFTAAFSFSLETQLTIGYGT MFPSGDCPSAIALLAIQM 142
	AP-----	GAQLI	TYPRALWWSVETATTVGYGDLYP--VTLWGRCVAVVVM 83
	-----	SWTV	SLYWTFVTIATVGYGDYSP--STPLGMYFTVTLI 80
	N-----	SFFKSI	PDAFWWAVVTMTTVGYGDMTP--VG VWGKIVGSLCA 463