

PGS Analyses

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```
library(magrittr)
library(dplyr)
library(readr)
library(forcats)
library(tidyr)
library(stringr)
library(ggplot2)
library(rethinking)
library(ggdist)
library(showtext)
font_add_google("Lato")
set_ulam_cmdstan(TRUE)
theme_set(theme_light(base_family = "Lato"))
showtext_auto()
histo_color = "#4C0099"
```

Prepare modeling data

Load

```
dat <- readr::read_csv("../data/chole_pgs.csv", col_types = "iiiddddlli")
skimr::skim_without_charts(dat)
```

Table 1: Data summary

Name	dat
Number of rows	179
Number of columns	10
Column type frequency:	
logical	1
numeric	9
Group variables	None

Variable type: logical

skim_variable	n_missing	complete_rate	mean	count
gb_hole	0	1	0.27	FAL: 131, TRU: 48

Variable type: numeric

skim_variable	n_missing	complete_rate	mean	sd	p0	p25	p50	p75	p100
videoid	0	1.00	90.00	51.82	1.00	45.50	90.00	134.50	179.00
surgid	0	1.00	9.61	7.07	1.00	4.00	8.00	17.00	23.00
pgs	0	1.00	2.46	1.21	1.00	1.00	2.00	3.00	5.00
time_until_1st_clip	0	1.00	15.53	12.42	1.07	5.57	13.00	21.15	60.92
time_cvs_attained	117	0.35	17.51	10.78	3.02	9.59	14.70	23.31	52.95
laparoscopic_duration	0	1.00	42.87	25.52	5.62	24.50	38.53	60.90	131.60
dissection_duration	0	1.00	16.08	13.20	1.07	6.15	13.00	21.05	63.22
gb_removal_duration	1	0.99	6.87	5.16	0.82	3.16	5.64	9.31	27.22
gb_holes	0	1.00	0.35	0.66	0.00	0.00	0.00	1.00	4.00

Keep surgeons with 5 or more cases

Arrange rows by surgeons number of cases (surgeons with most cases will be at the top, fewest cases at the bottom). Then replace the `videoid` with a new sequential integer id based on this order. This will allow us to drop surgeons with few cases and then still have a nice sequential id to join back on to during analysis of results:

```
dat <- dat %>%
  mutate(surgid = as.integer(fct_infreq(as.character(surgid)))) %>%
  arrange(surgid) %>%
  mutate(videoid = 1:nrow(.))
```

Keep surgeons with 5 or more cases:

```
dat <- dat %>%
  group_by(surgid) %>%
  filter(n() >= 5) %>%
  ungroup()
```

```
nsurgs <- max(dat$surgid)
nsurgs
```

```
## [1] 10
```

Prepare data for ulam/stan

Duration analyses

We performed analyses on the log scale given we were concerned about the magnitude of duration and that factors that increase duration tend to do so exponentially. For example, an inflamed gallbladder is harder to grasp, but also harder to dissect, and combining the two together, you have an exponential increase in time.

We also standardize our variables, as it assists with making weakly regularizing priors:

```
log_duration <- log(dat$laparoscopic_duration)
mean_log_duration <- mean(log_duration)
centered_log_duration <- log_duration - mean_log_duration
sd_log_duration <- sd(centered_log_duration)
std_log_duration <- centered_log_duration / sd_log_duration
```

Put list of data for modeling together. Note the `alpha`, which is our dirichlet prior (prior of 2 for PGS2, PGS3, PGS4, and PGS5):

```
ddat <- list(
  duration = std_log_duration,
  sid = dat$surgid,
  pgs = dat$pgs,
  alpha = rep(2, 4)
)
```

GB hole analyses

```
gdat <- list(
  hole = as.integer(dat$gb_hole),
  sid = dat$surgid,
  pgs = dat$pgs,
  alpha = rep(2, 4)
)
```

CVS attainment analyses

Analyses data is nearly ready, except for `dat$time_cvs_attained`.

We want to look at the binomial outcome, of whether or not CVS was achieved. To do so, just need to transform the data, so if it's NA then the CVS was never achieved.

```
cvs <- as.integer(!is.na(dat$time_cvs_attained))
```

Put the data together for stan:

```
cdat <- list(
  cvs = cvs,
  sid = dat$surgid,
  pgs = dat$pgs,
  alpha = rep(2, 4)
)
```

Utility functions

standard error

```
std_err <- function(xs) {
  sd(xs) / sqrt(length(xs))
}
```

De-standardize (back to original scale)

Used to move from the standardized/centered log scale to normal scale to facilitate analyses:

```
unstd <- function(x) {
  exp(x * sd_log_duration + mean_log_duration)
}
```

Find rows with high pareto k

```
high_k_rows <- function(results) {
  results %>%
```

```

    as_tibble(rownames = "videoid") %>%
    mutate(videoid = as.integer(videoid)) %>%
    inner_join(dat, by = "videoid") %>%
    filter(k >= 0.5)
}

```

samples from stan model

This is a thin wrapper around `rethinking` packages `extract.samples()` that returns a tibble with `janitor` fixing column names and a sample number given to each sample.

```

extract_samples <- function(..., seed = 1234) {
  set.seed(seed)
  extract.samples(...) %>%
  as_tibble() %>%
  # case = "none" to not mess up capitalization of our parameters
  # otherwise fixes brackets, commas, and other problem chars
  janitor::clean_names(case = "none") %>%
  mutate(sample_num = paste0("sample", 1:nrow()), .before = 1L)
}

```

tidy surgeons

Each sample has the info for one or more surgeons on each row. We want this info “tidy”, that is, one per row, so make a function to do that:

```

tidy_surgeons <- function(df) {
  df %>%
    pivot_longer(
      starts_with(c("a_", "bP")),
      names_to = c(".value", "surgeon"),
      names_sep = "_"
    )
}

```

tidy pgs

Each sample, when using pgs levels, has 5 tidy rows of info, 1 for each PGS. So need to pivot these longer:

```

tidy_pgs <- function(df) {
  df %>%
    pivot_longer(
      starts_with("PGS"),
      names_to = c(NA, "PGS"),
      names_pattern = "(PGS)([1-5])",
      names_transform = list("PGS" = as.integer),
      # to keep consistent with var name in model definition
      values_to = "sum_delta_j"
    )
}

```

sum deltas

We treat PGS as an ordered categorical predictor following McElreath’s strategy in “Statistical Rethinking”, Chapter 12, section 4. This assigns a proportion of the maximum value (PGS5) to each of the other PGS’s.

PGS1 is absorbed into the intercept.

The below function just calculates this proportion to make later calculations easier.

```
sum_deltas <- function(df) {  
  df %>% mutate(  
    PGS1 = 0,  
    PGS2 = delta_1,  
    PGS3 = PGS2 + delta_2,  
    PGS4 = 1 - delta_4,  
    PGS5 = 1  
  )  
}
```

Plot half eye

This is a wrapper over `stat_halfeye()` that will provide a fill for both the distribution and the scale with labels for percents. Color palette is purple gradients.

```
halfeye <- function(df, xcol, ycol) {  
  ggplot(df, aes(y = {{ ycol }}, x = {{ xcol }})) +  
    stat_halfeye(  
      aes(  
        fill = stat(  
          cut_cdf_qi(cdf, .width = c(0.5, 0.8, 0.95), labels = scales::percent_format())  
        )  
      ),  
      .width = c(0.5, 0.8, 0.95)  
    ) +  
    scale_fill_brewer(direction = -1, palette = "Purples", na.translate = FALSE) +  
    labs(fill = "Compatibility\nIntervals")  
}
```

Compatibility intervals

Quickly calculate a bunch of compatibility intervals and pretty format them:

```
ci_ints <- function(df, variable) {  
  summarise(df,  
    mean = mean({{ variable }}),  
    low_50 = quantile({{ variable }}, probs = 0.25),  
    high_50 = quantile({{ variable }}, probs = 0.75),  
    low_66 = quantile({{ variable }}, probs = 0.17),  
    high_66 = quantile({{ variable }}, probs = 0.83),  
    low_80 = quantile({{ variable }}, probs = 0.20),  
    high_80 = quantile({{ variable }}, probs = 0.80),  
    low_89 = quantile({{ variable }}, probs = 0.055),  
    high_89 = quantile({{ variable }}, probs = 0.945),  
    low_95 = quantile({{ variable }}, probs = 0.025),  
    high_95 = quantile({{ variable }}, probs = 0.975)  
  ) %>%  
  ungroup() %>%  
  mutate(across(where(is.numeric), ~ round(., 2))) %>%  
  unite(int_50, contains("_50"), sep = ", ") %>%  
  unite(int_66, contains("_66"), sep = ", ") %>%  
  unite(int_80, contains("_80"), sep = ", ") %>%
```

```

unite(int_89, contains("_89"), sep = ", ") %>%
unite(int_95, contains("_95"), sep = ", ")
}

```

Abbreviations in formulas

- sid: Deidentified surgeon id
- PGS: Parkland Grading Scale for gallbladder inflammation
- MVNormal: Multivariate normal distribution
- LKJCorr: Lewandowski, Kurowicka, and Joe Correlation Distribution
- CVS: Critical View of Safety

Duration Analysis

Priors determination

We will use weakly regularizing priors, that is, those that constrain parameters to those that are logically possible, while still allowing for some implausibly strong relationships if that is what the data determines.

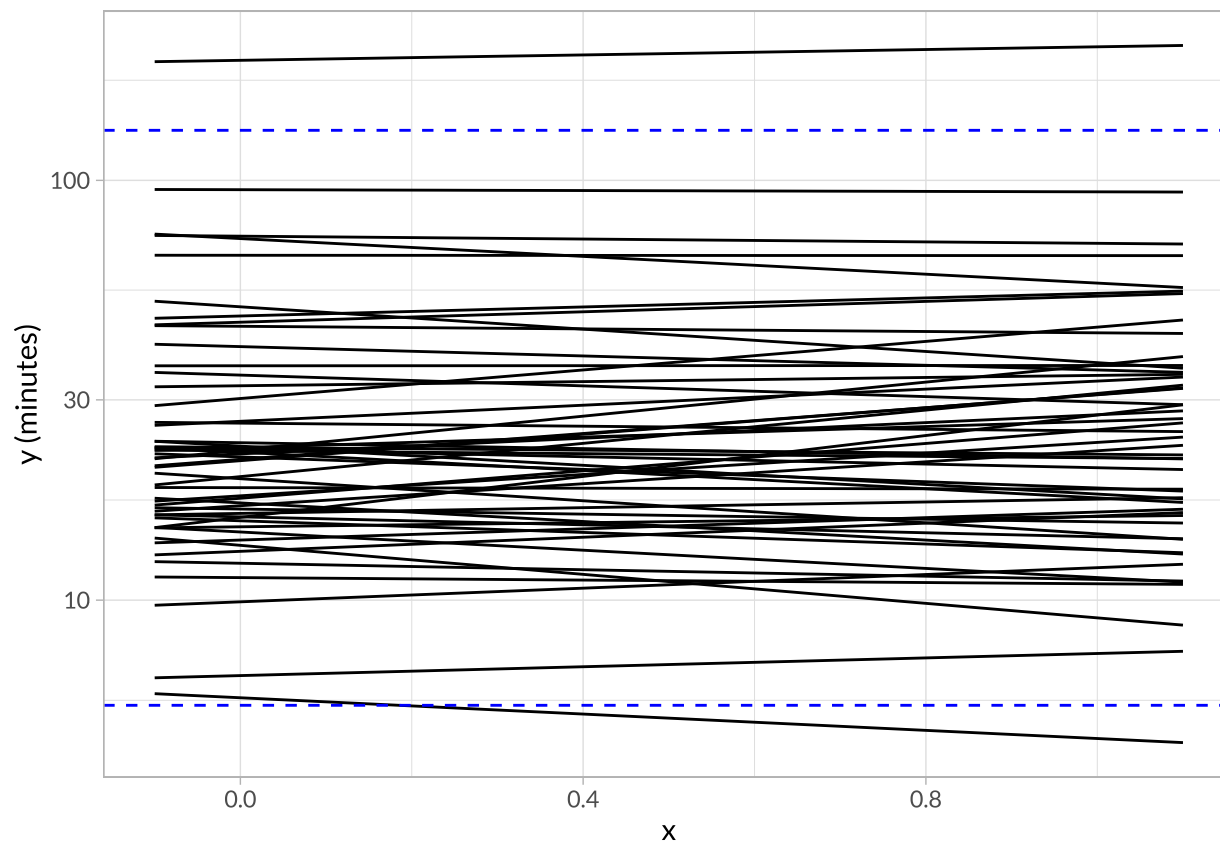
I will graph simulated values of the priors to help determine good ones to use:

Try a prior of $N(0, 1)$ for the intercept (a) and $N(0, 0.3)$ for the slope for PGS5, bp:

```

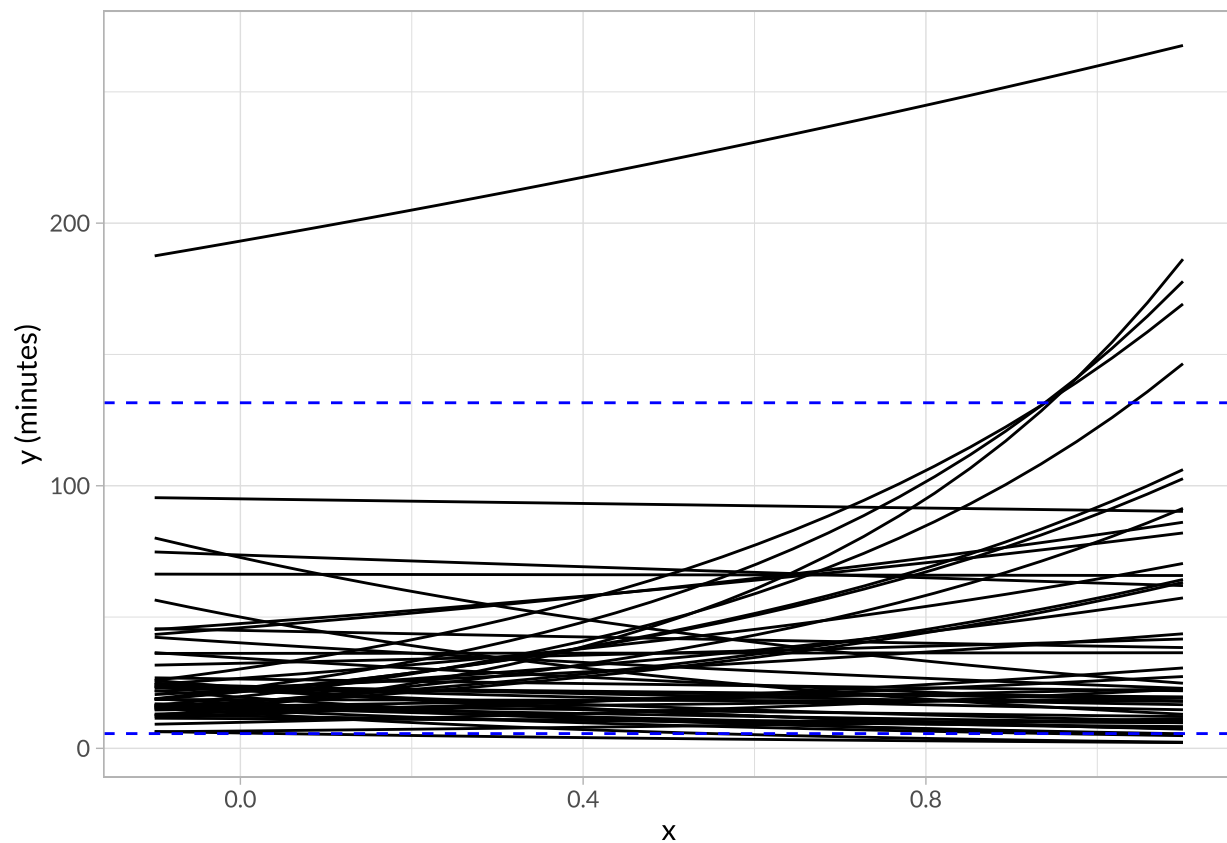
set.seed(1234)
max_dur <- max(ddat$duration)
min_dur <- min(ddat$duration)
tibble(
  sample_num = paste0("sample", 1:50),
  a = rnorm(50, mean = 0, sd = 1),
  bp = rnorm(50, mean = 0, sd = 0.3)
) %>%
  mutate(x = list(seq(from = -0.1, to = 1.1, length.out = 30))) %>%
  tidyr::unnest(x) %>%
  mutate(y = a + bp * x, og_y = unstd(y)) %>%
  ggplot(aes(x, og_y, group = sample_num)) +
  geom_line() +
  scale_y_log10() +
  labs(y = "y (minutes)") +
  geom_hline(yintercept = unstd(c(min_dur, max_dur)), linetype = 2, color = "blue")

```



The intercept prior looks acceptable. The slope prior is much too tight (remember slope is for a PGS5, so the maximum effect). The min/max of data never even reached. Let's try making it a bit bigger:

```
set.seed(1234)
max_dur <- max(ddat$duration)
min_dur <- min(ddat$duration)
tibble(
  sample_num = paste0("sample", 1:50),
  a = rnorm(50, mean = 0, sd = 1),
  bp = rnorm(50, mean = 0, sd = 1.2)
) %>%
  mutate(x = list(seq(from = -0.1, to = 1.1, length.out = 30))) %>%
  tidyr::unnest(x) %>%
  mutate(y = a + bp * x, og_y = unstd(y)) %>%
  ggplot(aes(x, og_y, group = sample_num)) +
  geom_line() +
  #scale_y_log10() +
  #labs(y = "y (log scale, minutes)") +
  labs(y = "y (minutes)") +
  geom_hline(yintercept = unstd(c(min_dur, max_dur)), linetype = 2, color = "blue")
```



The numbers are now constrained to mostly realistic values.

Other priors will be the usual weakly regularizing ones, including Dirichlet of 2, Exponential 1, and LKJCorr of 4.

Formula

Below is the centered version. The model given to Stan is the non-centered version that is mathematically equivalent but dramatically improves sampling.

$$\begin{aligned}
\log(\text{Duration}_i) &\sim \text{Normal}(\mu_i, \sigma) \\
\mu_i &= \alpha_{sid[i]} + \beta_{sid[i]} * \sum_{j=0}^{PGS_i-1} \delta_j \\
\begin{bmatrix} \alpha_{sid} \\ \beta_{sid} \end{bmatrix} &\sim \text{MVNormal}\left(\begin{bmatrix} \alpha \\ \beta \end{bmatrix}, \mathbf{S}\right) \\
\alpha &\sim \text{Normal}(0, 1) \\
\beta &\sim \text{Normal}(0, 1.2) \\
\delta &\sim \text{Dirichlet}(2) \\
\mathbf{S} &= \begin{pmatrix} \sigma_\alpha & 0 \\ 0 & \sigma_\beta \end{pmatrix} \mathbf{R} \begin{pmatrix} \sigma_\alpha & 0 \\ 0 & \sigma_\beta \end{pmatrix} \\
\mathbf{R} &= \begin{pmatrix} 1 & \rho \\ \rho & 1 \end{pmatrix} \\
\mathbf{R} &\sim \text{LKJCorr}(4) \\
\sigma, \sigma_\alpha, \sigma_\beta &\sim \text{Exponential}(1)
\end{aligned} \tag{1}$$

Code

```

set.seed(1234)
dur_mod <- ulam(
  alist(
    duration ~ normal(mu, sigma),
    mu <- a_bar + ab_sid[sid, 1] + (bP_bar + ab_sid[sid, 2]) * sum(delta_j[1:pgs]),
    transpar> matrix[sid, 2]:ab_sid <-
      compose_noncentered(sigma_sid, L_Rho_sid, z_sid),
    matrix[2, sid]:z_sid ~ normal(0, 1),
    a_bar ~ normal(0, 1),
    bP_bar ~ normal(0, 1.2),
    vector[2]:sigma_sid ~ exponential(1),
    sigma ~ exponential(1),
    cholesky_factor_corr[2]:L_Rho_sid ~ lkj_corr_cholesky(4),
    vector[5]:delta_j <-< append_row(0, delta),
    simplex[4]:delta ~ dirichlet(alpha),
    # compute correlation matrix from Cholesky matrix
    gq> matrix[2, 2]: Rho_sid <-< Chol_to_Corr(L_Rho_sid),
    # for our analysis sake, compute a[sid] and b[sid]
    gq> vector[sid]:a <-< a_bar + ab_sid[, 1],
    gq> vector[sid]:bP <-< bP_bar + ab_sid[, 2]
  ),
  data = ddat,
  cores = 4,
  chains = 4,
  iter = 5000,
  log_lik = TRUE
)

```

```

## This is cmdstanr version 0.3.0
## - Online documentation and vignettes at mc-stan.org/cmdstanr
## - CmdStan path set to: /home/thomas/.cmdstanr/cmdstan-2.26.1

```



```

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## Chain 3 finished in 4.2 seconds.

```

```
## Chain 2 Iteration: 4900 / 5000 [ 98%] (Sampling)
## Chain 4 Iteration: 5000 / 5000 [100%] (Sampling)
## Chain 4 finished in 4.3 seconds.
## Chain 2 Iteration: 5000 / 5000 [100%] (Sampling)
## Chain 2 finished in 4.4 seconds.
##
## All 4 chains finished successfully.
## Mean chain execution time: 4.3 seconds.
## Total execution time: 4.5 seconds.
```

Diagnostic Evaluation of Markov Chains

Rhat4 and effective sampling size

```
precis(dur_mod, depth = 3)
```

##		mean	sd	5.5%	94.5%	n_eff
##	z_sid[1,1]	-2.384040295	0.64845693	-3.46047155	-1.3887519	4361.301
##	z_sid[1,2]	0.526393679	0.37258315	-0.05185201	1.1411565	3233.090
##	z_sid[1,3]	-0.196054682	0.34787152	-0.74475582	0.3629053	3283.831
##	z_sid[1,4]	0.743951570	0.40404040	0.11449700	1.4060161	3517.962
##	z_sid[1,5]	-0.121013920	0.38091790	-0.72746068	0.4919521	3743.610
##	z_sid[1,6]	-0.230778330	0.39064474	-0.86389002	0.3867039	4171.463
##	z_sid[1,7]	0.282479789	0.43019798	-0.39529988	0.9795907	4550.611
##	z_sid[1,8]	0.648260842	0.44872450	-0.04838142	1.3932141	4447.475
##	z_sid[1,9]	0.012495293	0.45714921	-0.71428326	0.7310225	4835.161
##	z_sid[1,10]	0.736366008	0.45693052	0.04007889	1.4858520	4238.050
##	z_sid[2,1]	-0.188884195	0.91739099	-1.64864895	1.2756814	11575.454
##	z_sid[2,2]	0.343427499	0.86451672	-1.07709830	1.6953510	9601.130
##	z_sid[2,3]	-0.103317120	0.88550879	-1.51337615	1.3156256	12563.761
##	z_sid[2,4]	0.366442393	0.91754956	-1.16928100	1.8118154	11928.553
##	z_sid[2,5]	-0.570701866	0.98628381	-2.10178525	1.0583522	10010.370
##	z_sid[2,6]	0.119371030	0.91779743	-1.32280940	1.5911848	13768.783
##	z_sid[2,7]	-0.029551626	0.95685800	-1.55359445	1.5153637	12634.192
##	z_sid[2,8]	-0.006542448	0.99020980	-1.60811240	1.5483340	13602.784
##	z_sid[2,9]	0.092697914	0.94128684	-1.42897290	1.5773290	12587.717
##	z_sid[2,10]	0.100754564	0.95920028	-1.45515890	1.6221550	13120.354
##	a_bar	0.078315842	0.24502259	-0.30479893	0.4641564	3025.395
##	bP_bar	0.525659390	0.18897720	0.23208460	0.8236595	8272.162
##	sigma_sid[1]	0.740994701	0.20399831	0.48811979	1.0992365	4714.022
##	sigma_sid[2]	0.230295577	0.19106328	0.01804434	0.5787149	5171.965
##	sigma	0.492707031	0.02953328	0.44816751	0.5413854	11622.999
##	L_Rho_sid[1,1]	1.000000000	0.00000000	1.00000000	1.0000000	NaN
##	L_Rho_sid[1,2]	0.000000000	0.00000000	0.00000000	0.0000000	NaN
##	L_Rho_sid[2,1]	0.060022547	0.31384906	-0.44226001	0.5565246	12951.443
##	L_Rho_sid[2,2]	0.944709241	0.07369466	0.80007415	0.9997231	3846.376
##	delta[1]	0.196725932	0.10723971	0.04812869	0.3827334	11843.450
##	delta[2]	0.258804358	0.12415547	0.08142931	0.4770108	11882.380
##	delta[3]	0.302800703	0.13683920	0.09792895	0.5322698	12468.829
##	delta[4]	0.241669012	0.12747727	0.05962633	0.4685847	13088.522
##	bP[1]	0.446410234	0.26111763	0.01871364	0.8462773	10784.322
##	bP[2]	0.620028919	0.20316337	0.31698957	0.9640038	10063.920
##	bP[3]	0.498760160	0.22936905	0.13005026	0.8536246	12062.550
##	bP[4]	0.639706544	0.25936135	0.26889784	1.0847010	9940.633

## bP[5]	0.347814625	0.34991123	-0.28751296	0.7852630	6497.632
## bP[6]	0.557224206	0.26923184	0.15688216	0.9945579	9822.049
## bP[7]	0.519682475	0.29183425	0.05966005	0.9573360	9073.491
## bP[8]	0.529620630	0.32998362	0.03607356	1.0077398	8659.421
## bP[9]	0.557250023	0.29183105	0.13178997	1.0110399	8494.040
## bP[10]	0.566535486	0.30895248	0.10406067	1.0517094	10190.231
## a[1]	-1.583054846	0.11740468	-1.76952275	-1.3955567	11001.569
## a[2]	0.444448537	0.12686933	0.23652517	0.6421926	10553.103
## a[3]	-0.059114236	0.10674821	-0.23049965	0.1137655	12680.428
## a[4]	0.597097399	0.14475824	0.35701904	0.8206876	10545.440
## a[5]	-0.006984282	0.16152566	-0.26077277	0.2542622	11058.199
## a[6]	-0.084064937	0.17829539	-0.36779120	0.1984511	13050.039
## a[7]	0.276363836	0.23111632	-0.08897297	0.6377499	11705.923
## a[8]	0.535090894	0.24211732	0.15167483	0.9162728	11661.807
## a[9]	0.087360977	0.26259003	-0.33214508	0.4956021	10718.759
## a[10]	0.596570234	0.23151126	0.22871259	0.9644537	13036.584
## Rho_sid[1,1]	1.000000000	0.00000000	1.00000000	1.0000000	NaN
## Rho_sid[1,2]	0.060022547	0.31384906	-0.44226001	0.5565246	12951.443
## Rho_sid[2,1]	0.060022547	0.31384906	-0.44226001	0.5565246	12951.443
## Rho_sid[2,2]	1.000000000	0.00000000	1.00000000	1.0000000	NaN
## ab_sid[1,1]	-1.661370708	0.25887463	-2.07702200	-1.2573134	3262.760
## ab_sid[1,2]	-0.079249148	0.23262053	-0.48389211	0.2463126	11377.196
## ab_sid[2,1]	0.366132700	0.25997624	-0.04014196	0.7845014	3350.392
## ab_sid[2,2]	0.094369531	0.19892059	-0.15844152	0.4576320	7184.016
## ab_sid[3,1]	-0.137430073	0.25730204	-0.54035649	0.2642420	3228.425
## ab_sid[3,2]	-0.026899230	0.20543773	-0.36813397	0.2818660	10592.281
## ab_sid[4,1]	0.518781563	0.27092806	0.08806932	0.9460820	3566.606
## ab_sid[4,2]	0.114047162	0.23811935	-0.16820897	0.5595589	8248.674
## ab_sid[5,1]	-0.085300116	0.27842923	-0.52605934	0.3578331	3606.977
## ab_sid[5,2]	-0.177844761	0.30259357	-0.76474788	0.1274416	6720.732
## ab_sid[6,1]	-0.162380778	0.28241711	-0.61705793	0.2837209	3843.780
## ab_sid[6,2]	0.031564815	0.22866791	-0.30383671	0.4197998	10104.012
## ab_sid[7,1]	0.198048005	0.30921125	-0.29264917	0.6896935	4362.857
## ab_sid[7,2]	-0.005976903	0.24165363	-0.39010508	0.3577238	9685.792
## ab_sid[8,1]	0.456775033	0.31616834	-0.03566903	0.9716484	4650.984
## ab_sid[8,2]	0.003961250	0.27642614	-0.39676551	0.4074769	9475.762
## ab_sid[9,1]	0.009045131	0.33133023	-0.51664892	0.5314908	4665.404
## ab_sid[9,2]	0.031590649	0.24464680	-0.30414098	0.4361623	9120.277
## ab_sid[10,1]	0.518254401	0.31394342	0.02934193	1.0175772	4378.663
## ab_sid[10,2]	0.040876102	0.25994092	-0.33061215	0.4617071	10584.545
##	Rhat4				
## z_sid[1,1]	1.0009547				
## z_sid[1,2]	1.0000485				
## z_sid[1,3]	0.9999519				
## z_sid[1,4]	1.0003597				
## z_sid[1,5]	1.0000571				
## z_sid[1,6]	0.9999865				
## z_sid[1,7]	0.9998480				
## z_sid[1,8]	1.0000571				
## z_sid[1,9]	0.9997443				
## z_sid[1,10]	1.0002478				
## z_sid[2,1]	0.9998427				
## z_sid[2,2]	0.9997749				
## z_sid[2,3]	0.9998908				

```

## z_sid[2,4]      0.9997793
## z_sid[2,5]      1.0002381
## z_sid[2,6]      0.9996765
## z_sid[2,7]      0.9997916
## z_sid[2,8]      1.0000862
## z_sid[2,9]      1.0000990
## z_sid[2,10]     0.9997940
## a_bar           1.0000781
## bP_bar          1.0000207
## sigma_sid[1]    1.0012668
## sigma_sid[2]    1.0007773
## sigma           0.9998212
## L_Rho_sid[1,1]   NaN
## L_Rho_sid[1,2]   NaN
## L_Rho_sid[2,1]   1.0002752
## L_Rho_sid[2,2]   1.0010717
## delta[1]         0.9998563
## delta[2]         0.9998311
## delta[3]         1.0000443
## delta[4]         0.9998103
## bP[1]            1.0000889
## bP[2]            0.9999250
## bP[3]            0.9999165
## bP[4]            0.9997010
## bP[5]            1.0005906
## bP[6]            0.9998104
## bP[7]            1.0001544
## bP[8]            1.0005158
## bP[9]            1.0001844
## bP[10]           1.0001504
## a[1]             0.9999226
## a[2]             0.9999620
## a[3]             1.0004889
## a[4]             0.9998053
## a[5]             1.0000256
## a[6]             0.9998460
## a[7]             1.0000033
## a[8]             1.0000659
## a[9]             0.9997889
## a[10]            1.0000340
## Rho_sid[1,1]     NaN
## Rho_sid[1,2]     1.0002752
## Rho_sid[2,1]     1.0002752
## Rho_sid[2,2]     NaN
## ab_sid[1,1]      1.0001677
## ab_sid[1,2]      1.0000533
## ab_sid[2,1]      0.9999082
## ab_sid[2,2]      1.0004184
## ab_sid[3,1]      1.0000298
## ab_sid[3,2]      0.9999735
## ab_sid[4,1]      0.9999604
## ab_sid[4,2]      0.9999812
## ab_sid[5,1]      1.0001224
## ab_sid[5,2]      1.0003042

```



```
## ab_sid[6,1]    1.0000758
## ab_sid[6,2]    0.9998335
## ab_sid[7,1]    1.0000718
## ab_sid[7,2]    1.0000265
## ab_sid[8,1]    0.9997919
## ab_sid[8,2]    1.0002419
## ab_sid[9,1]    0.9997953
## ab_sid[9,2]    1.0002789
## ab_sid[10,1]   0.9999335
## ab_sid[10,2]   1.0006268
```

All Rhat4 values are 1.

Each parameter also sampled well.

PSIS/WAIC

```
PSIS(dur_mod)
```

```
## Some Pareto k values are high (>0.5). Set pointwise=TRUE to inspect individual points.
```

```
##      PSIS      lppd  penalty  std_err
## 1 232.3422 -116.1711 13.90847 17.11978
```

```
WAIC(dur_mod)
```

```
##      WAIC      lppd  penalty  std_err
## 1 231.7685 -102.2626 13.62158 16.99175
```

The are some Pareto k values > 0.5. As long as < 0.7, not an issue. Are they?

```
PSIS(dur_mod, pointwise = TRUE) %>%
  high_k_rows()
```

```
## Some Pareto k values are high (>0.5). Set pointwise=TRUE to inspect individual points.
```

```
## # A tibble: 2 x 15
##   videoid PSIS  lppd penalty std_err      k surgid  pgs time_until_1st_c~
##   <int> <dbl> <dbl>   <dbl>   <dbl> <dbl> <int> <int>         <dbl>
## 1    146  5.93 -2.96   0.949   17.1 0.579      9     4           7.85
## 2    151  2.48 -1.24   0.327   17.1 0.563     10     2          12.6
## # ... with 6 more variables: time_cvs_attained <dbl>,
## #   laparoscopic_duration <dbl>, dissection_duration <dbl>,
## #   gb_removal_duration <dbl>, gb_hole <lgl>, gb_holes <int>
```

Only 2 rows total, and both have $k < 0.6$, so minimal issue with outliers.

Trace rank plot (trankplot)

```
trankplot(dur_mod)
```

Good mixing as shown with large amount of overlap.

Trace plot

```
traceplot(dur_mod)
```

Chains are stationary with a visible central tendency, have good mixing, and converge. Excellent.

Evaluate model results

List of the parameters I care about from the model:

1. Intercepts ($a[N]$ and average a_bar)
2. beta for pgs5 ($bP[N]$ and average bP_bar)
3. Correlation ($Rho_sid[1, 2]$)
4. sigmas (overall, for intercept, for beta)
5. deltas (for incremental effect of each PGS)

```
pars <- c(
  "a_bar",
  paste0("a[", 1:nsurgs, "]"),
  "bP_bar",
  paste0("bP[", 1:nsurgs, "]"),
  # sigma for entire model
  "sigma",
  # sigma_a
  "sigma_sid[1]",
  # sigma_b
  "sigma_sid[2]",
  paste0("Rho_sid[1,2]"),
  paste0("delta[", 1:4, "]")
)
```

Now time to extract samples from stan:

```
dur_samples <- extract_samples(dur_mod, pars = pars) %>%
  # change names for ease of remembering rather than having to
  # remember their position in the matrices
  rename(sigma_a = sigma_sid_1, sigma_bP = sigma_sid_2, rho = Rho_sid_1_2) %>%
  sum_deltas()
```

Now, let us generate a “new” unseen surgeon to see what the model will predict for a new surgeon, make the data tidy (one row per surgeon per PGS per sample), then calculate the outcome from the samples to see the model’s predictions.

```
tidy_dur_samples <- dur_samples %>%
  # generates a "new" unseen surgeon
  mutate(
    a_new = rnorm(nrow(.), a_bar, sigma_a),
    bP_new = rnorm(nrow(.), bP_bar, sigma_bP)
  ) %>%
  select(sample_num, starts_with(c("a_", "bP", "PGS")), sigma) %>%
  tidy_surgeons() %>%
  tidy_pgs() %>%
  mutate(mu = a + bP * sum_delta_j, mu_og = unstd(mu)) %>%
  select(sample_num, surgeon, PGS, mu, mu_og, sigma)
```

Plot results

Was duration correlated with starting time?

```
extract_samples(dur_mod, pars = c("Rho_sid[1,2]")) %>%
  rename(rho = Rho_sid_1_2) %>%
  ci_ints(rho) %>%
  knitr::kable()
```

```
)
caption = "Correlation between time to perform PGS1 case and the effect of incrementing PGS on laparoscopic duration"
)
```

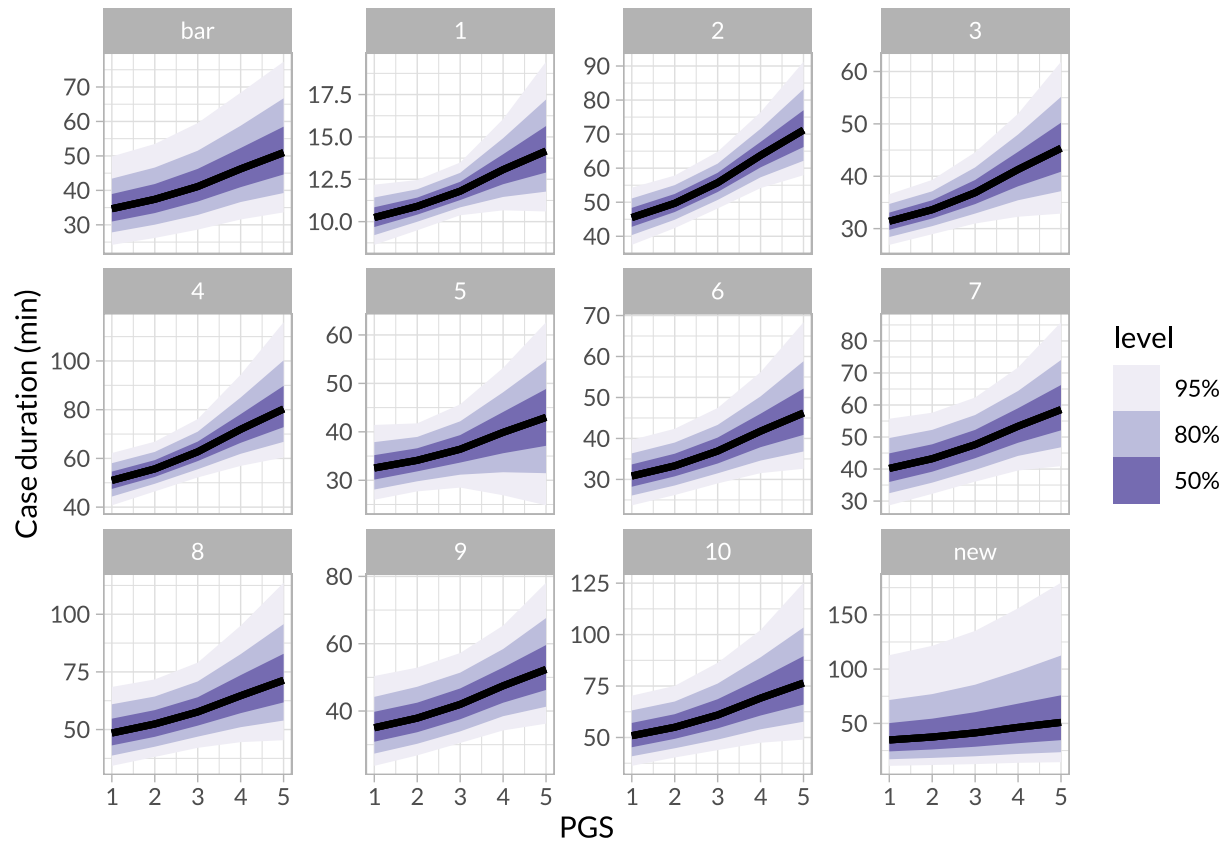
Table 4: Correlation between time to perform PGS1 case and the effect of incrementing PGS on laparoscopic duration

mean	int_50	int_66	int_80	int_89	int_95
0.06	-0.16, 0.29	-0.26, 0.37	-0.22, 0.34	-0.44, 0.56	-0.54, 0.65

For every surgeon

Visualization of the effects PGS has on each surgeon. Surgeon **bar** is the average surgeon, and surgeon **new** is for a future never seen surgeon.

```
tidy_dur_samples %>%
  mutate(surgeon = parse_factor(surgeon, levels = c("bar", as.character(1:nsurges), "new"))) %>%
  ggplot(aes(PGS, mu_og)) +
  stat_lineribbon() +
  facet_wrap(~ surgeon, scales = "free_y") +
  scale_fill_brewer(labels = c("95%", "80%", "50%"), palette = "Purples") +
  labs(y = "Case duration (min)")
```



Calculate deltas

Calculate the change from a PGS1 each PGS level has.

First, who is affected the most and the least by PGS? This will manifest by the smallest and the largest bP:

```
dur_bps <- precis(dur_mod, depth = 2, pars = paste0("bP[", 1:nsurgs, "]")) %>%
  as_tibble(rownames = "var")
# Least affected surgeon:
slice_min(dur_bps, mean)
```

```
## # A tibble: 1 x 7
##   var      mean      sd '5.5%' '94.5%' n_eff Rhat4
##   <chr> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl>
## 1 bP[5] 0.348 0.350 -0.288  0.785 6498.  1.00
```

```
# Most affected surgeon:
slice_max(dur_bps, mean)
```

```
## # A tibble: 1 x 7
##   var      mean      sd '5.5%' '94.5%' n_eff Rhat4
##   <chr> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl>
## 1 bP[4] 0.640 0.259  0.269   1.08 9941.  1.00
```

So the most affected surgeon is Surgeon “4” and the least affected surgeon is Surgeon “5”.

Now actually calculate the deltas:

```
dur_pgs_deltas <- tidy_dur_samples %>%
  # selecting our least, average, and most affected surgeons
  filter(surgeon %in% c("bar", "4", "5")) %>%
  mutate(
    surgeon = case_when(
      surgeon == "4" ~ "Most affected",
      surgeon == "5" ~ "Least affected",
      surgeon == "bar" ~ "Average"
    ),
    surgeon = parse_factor(
      surgeon,
      levels = c("Least affected", "Average", "Most affected")
    )
  ) %>%
  # ensure correct order, with PGS1 as first row in future group
  # will rely on this ordering to calculate delta
  arrange(sample_num, surgeon, PGS) %>%
  group_by(sample_num, surgeon) %>%
  mutate(pgs_delta = mu_og - mu_og[1]) %>%
  ungroup() %>%
  # needed for ggdist to work
  mutate(PGS = as.factor(PGS))
```

On average across all surgeons

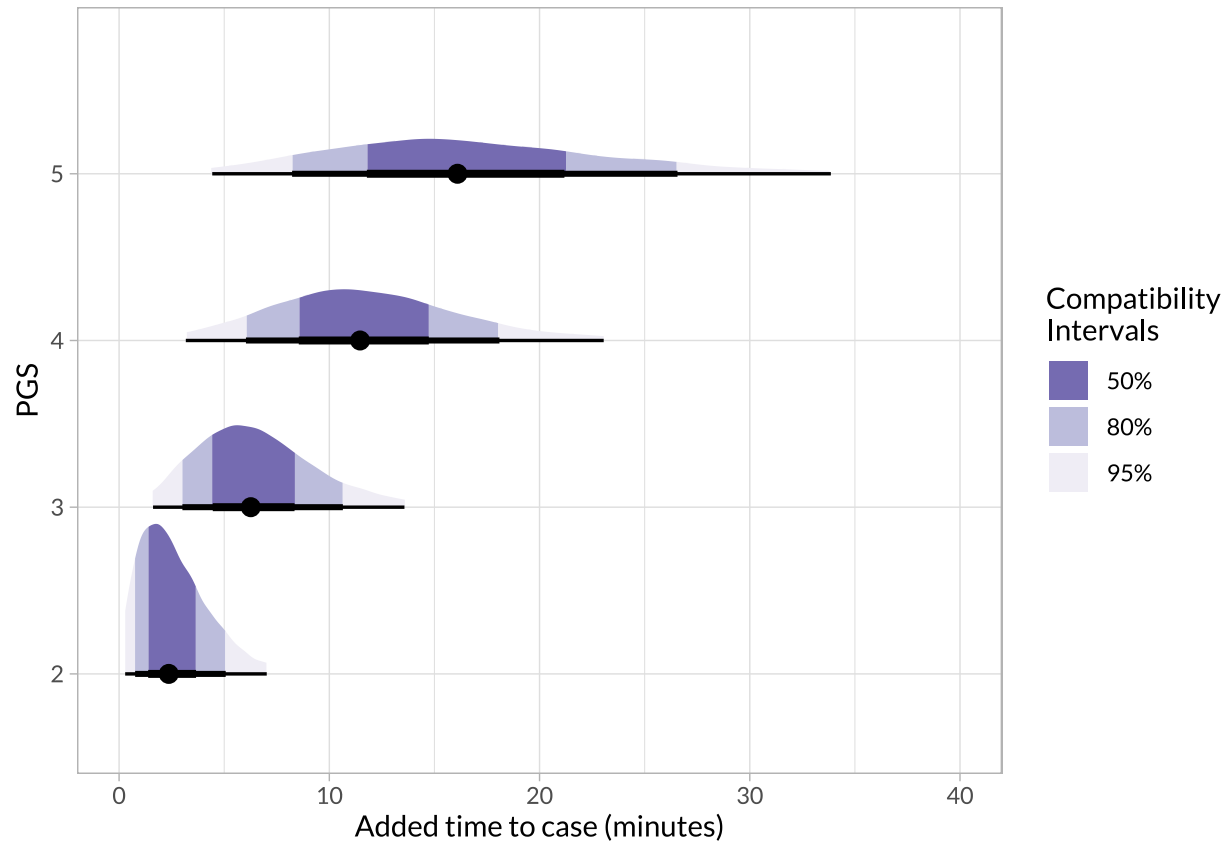
How was the average surgeon affected?

Then plot this:

```
dur_pgs_deltas_bar <- filter(dur_pgs_deltas, surgeon == "Average", PGS != "1")

dur_pgs_deltas_bar %>%
  halfeye(pgs_delta, PGS) +
  coord_cartesian(xlim = c(0, 40)) +
```

```
labs(
  x = "Added time to case (minutes)",
  y = "PGS"
)
```



```
ggsave("../output/pgs_duration.svg", width = 6, height = 6)
ggsave("../output/pgs_duration.pdf", width = 6, height = 6)
```

What are the numbers of the compatibility intervals:

```
dur_pgs_deltas_bar %>%
  group_by(PGS) %>%
  ci_ints(pgs_delta) %>%
  knitr::kable()
```

PGS	mean	int_50	int_66	int_80	int_89	int_95
2	2.70	1.38, 3.66	1.06, 4.3	1.18, 4.03	0.5, 5.9	0.29, 7.02
3	6.59	4.46, 8.35	3.76, 9.34	4.04, 8.93	2.38, 11.96	1.61, 13.57
4	11.87	8.55, 14.73	7.35, 16.24	7.82, 15.58	4.67, 20.37	3.17, 23.05
5	16.94	11.79, 21.18	10.06, 23.71	10.79, 22.64	6.49, 29.76	4.43, 33.86

All units are in minutes.

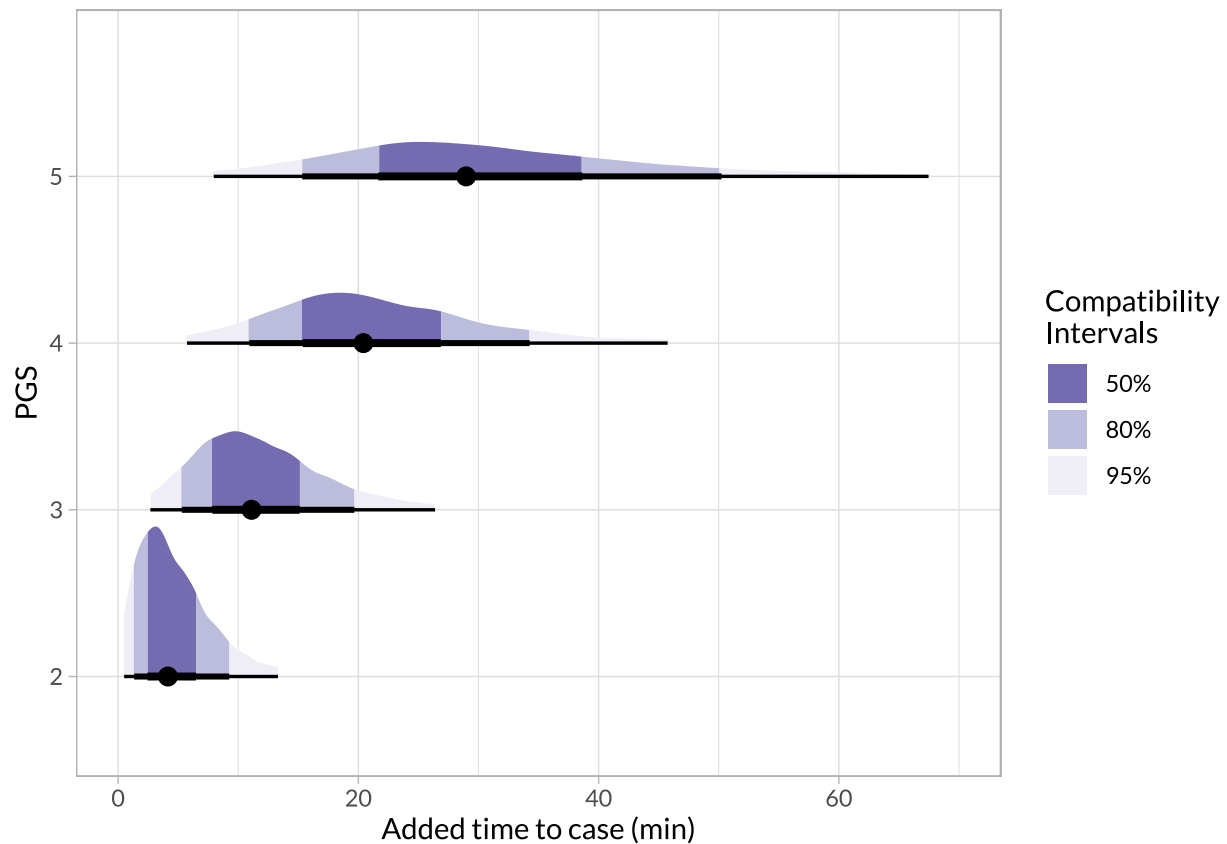
surgeon most affected

How much was the most affected surgeon, affected?

Then plot this:

```
dur_pgs_deltas_ma <- filter(dur_pgs_deltas, surgeon == "Most affected", PGS != "1")
```

```
dur_pgs_deltas_ma %>%
  halfeye(pgs_delta, PGS) +
  coord_cartesian(xlim = c(0, 70)) +
  labs(
    x = "Added time to case (min)"
  )
```



What are the numbers of the compatibility intervals:

```
dur_pgs_deltas_ma %>%
  group_by(PGS) %>%
  ci_ints(pgs_delta) %>%
  knitr::kable()
```

PGS	mean	int_50	int_66	int_80	int_89	int_95
2	4.86	2.44, 6.49	1.88, 7.7	2.09, 7.2	0.87, 10.96	0.5, 13.31
3	11.98	7.85, 15.1	6.71, 17.17	7.17, 16.29	4.12, 22.67	2.68, 26.39
4	21.77	15.38, 26.87	13.27, 29.88	14.16, 28.6	8.45, 38.9	5.74, 45.74
5	31.30	21.66, 38.64	18.73, 43.54	19.88, 41.48	12.03, 57.46	7.97, 67.46

All units are in minutes.

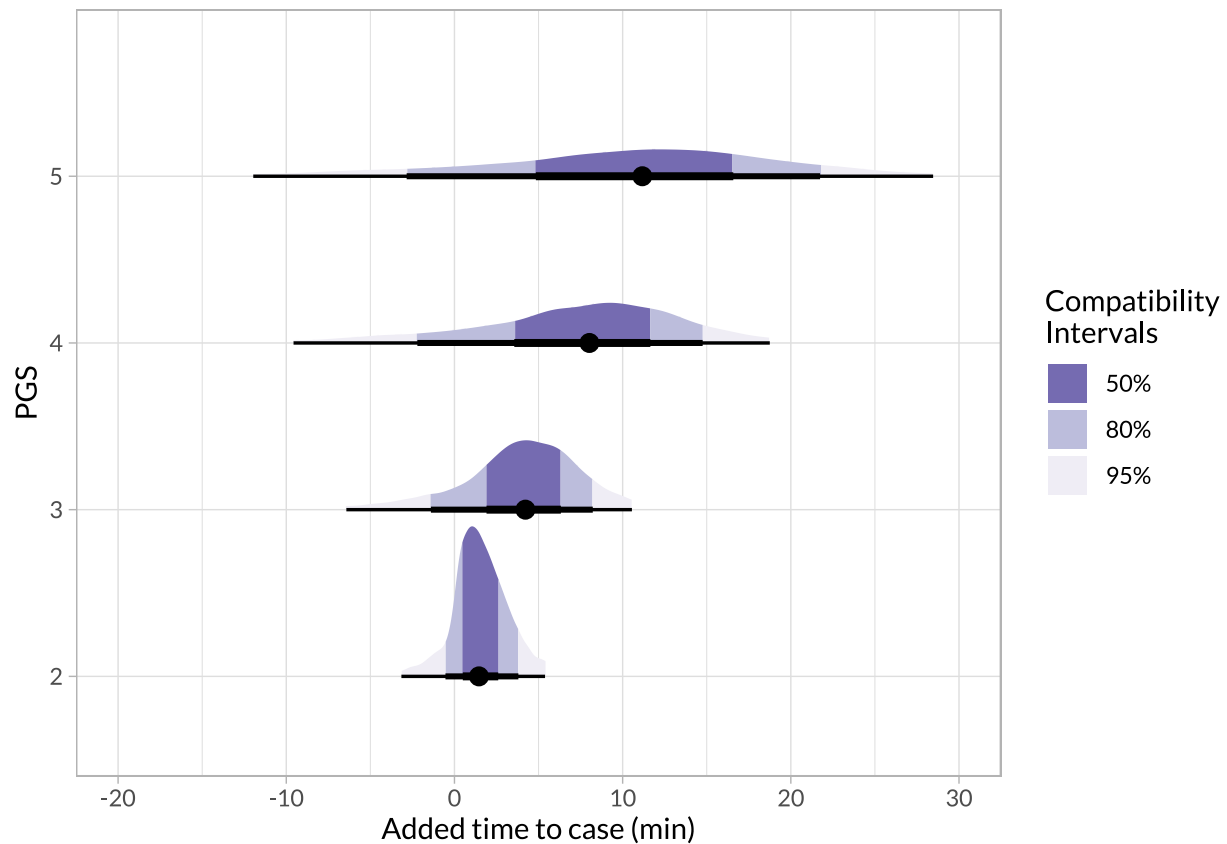
surgeon least affected

How much was the least affected surgeon, affected?

Then plot this:

```
dur_pgs_deltas_la <- filter(dur_pgs_deltas, surgeon == "Least affected", PGS != "1")
```

```
dur_pgs_deltas_la %>%
  halfeye(pgs_delta, PGS) +
  coord_cartesian(xlim = c(-20, 30)) +
  labs(
    x = "Added time to case (min)"
  )
```



What are the numbers of the compatibility intervals:

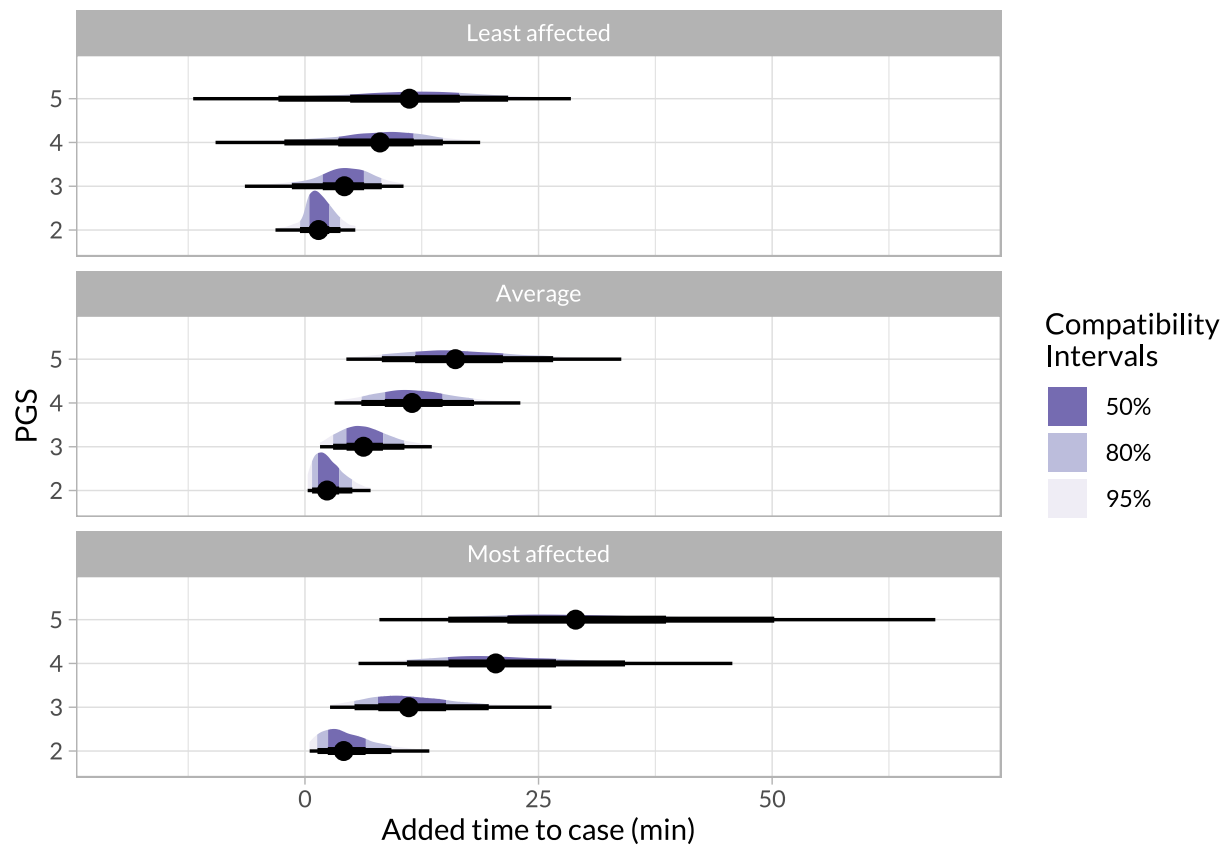
```
dur_pgs_deltas_la %>%
  group_by(PGS) %>%
  ci_ints(pgs_delta) %>%
  knitr::kable()
```

PGS	mean	int_50	int_66	int_80	int_89	int_95
2	1.46	0.5, 2.6	0.16, 3.13	0.3, 2.91	-1.55, 4.49	-3.15, 5.39
3	3.76	1.91, 6.33	0.64, 7.17	1.21, 6.83	-3.46, 9.34	-6.43, 10.55
4	7.07	3.56, 11.65	1.07, 13	2.12, 12.45	-5.4, 16.54	-9.57, 18.75
5	10.31	4.84, 16.58	1.44, 18.89	2.83, 17.89	-6.92, 24.43	-11.96, 28.46

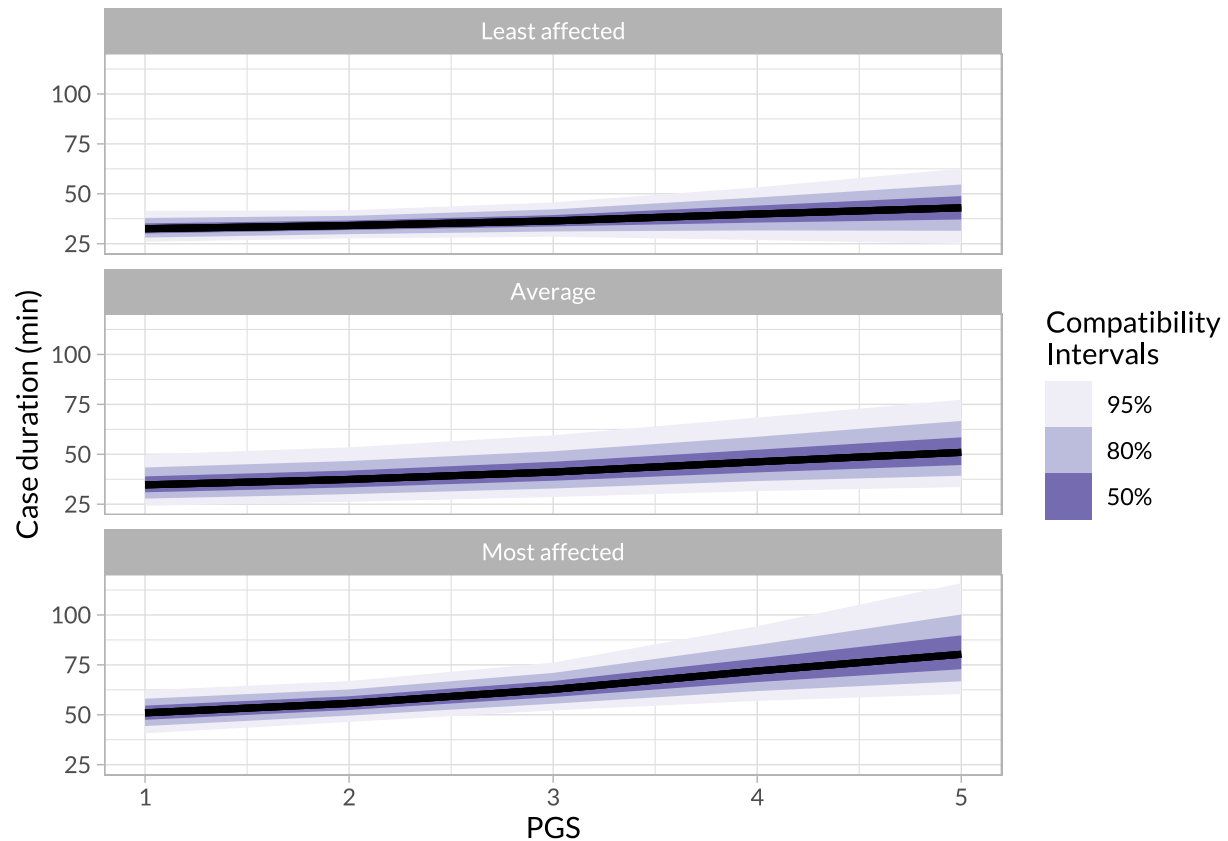
All units are in minutes.

all together now

```
dur_pgs_deltas %>%
  filter(PGS != "1") %>%
  halfeye(pgs_delta, PGS) +
  facet_wrap(~ surgeon, ncol = 1) +
  coord_cartesian(xlim = c(-20, 70)) +
  labs(
    x = "Added time to case (min)"
  )
```



```
dur_pgs_deltas %>%
  mutate(PGS = as.integer(PGS)) %>%
  ggplot(aes(PGS, mu_og)) +
  stat_lineribbon() +
  facet_wrap(~ surgeon, ncol = 1) +
  scale_fill_brewer(labels = c("95%", "80%", "50%"), palette = "Purples") +
  labs(y = "Case duration (min)", fill = "Compatibility\nIntervals")
```

Gallbladder Holes Analysis

We treated a gallbladder hole occurring during the removal of the gallbladder bed as a binomial outcome, that is, we cared if 1 or more holes occurred and treated those equal. We chose this because once a full-thickness perforation occurs, bile/stones spill. Adding further holes at this point is rather inconsequential, as what matters is going from no hole to a single hole.

Priors determination

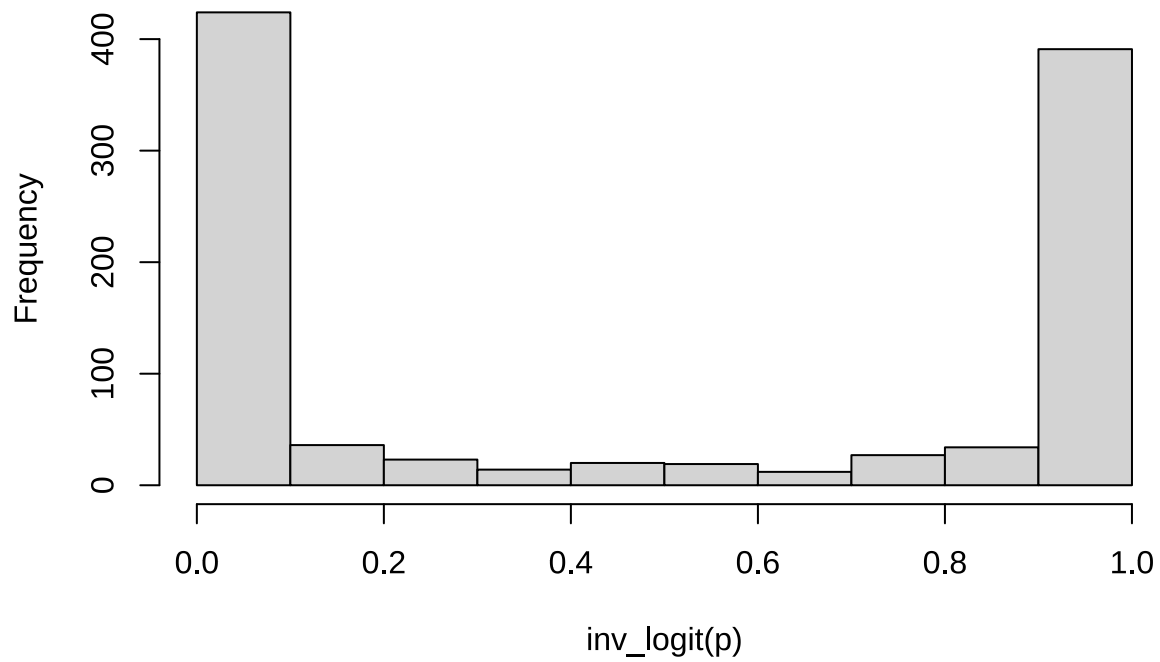
As before, we will use weakly regularizing priors.

Intercept

To see how `logit()` can mess up priors, look at a $N(0, 10)$ intercept prior:

```
p <- rnorm(1000, 0, 10)
hist(inv_logit(p))
```

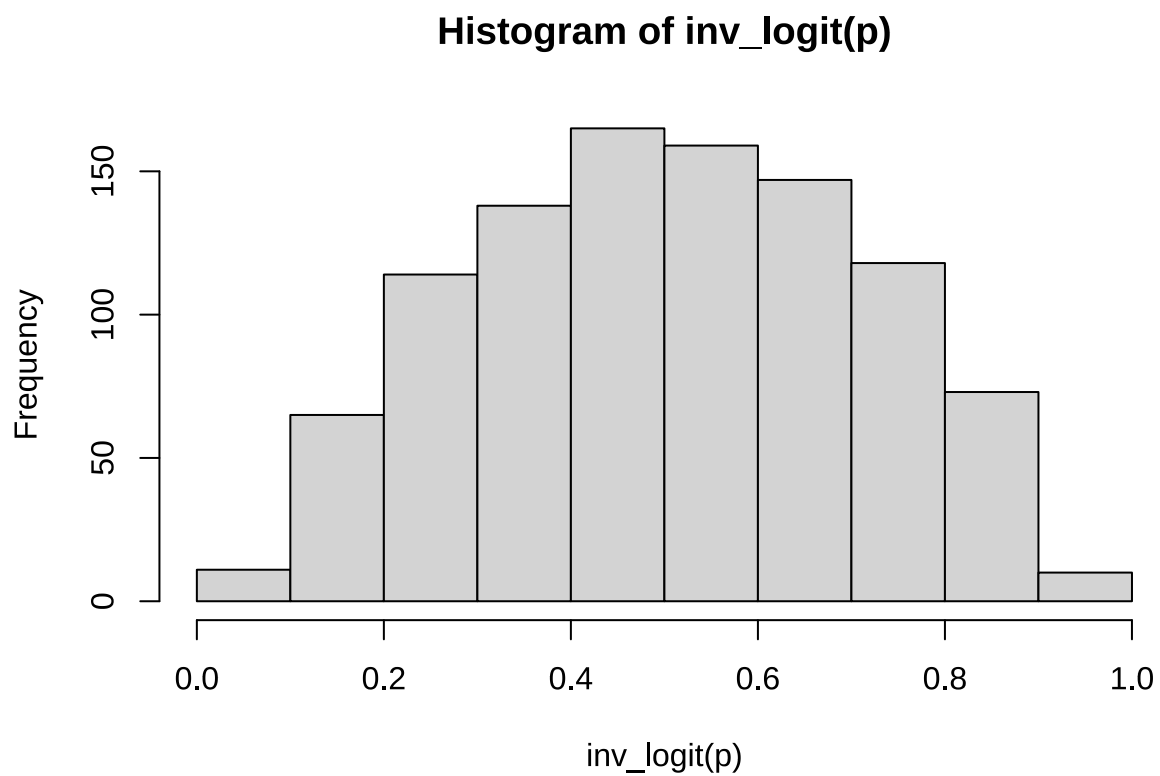
Histogram of $\text{inv_logit}(p)$



Nearly all the probability mass is at 0 or 1 for the prior, which is nonsensical.

Trying a $N(0, 1)$ prior:

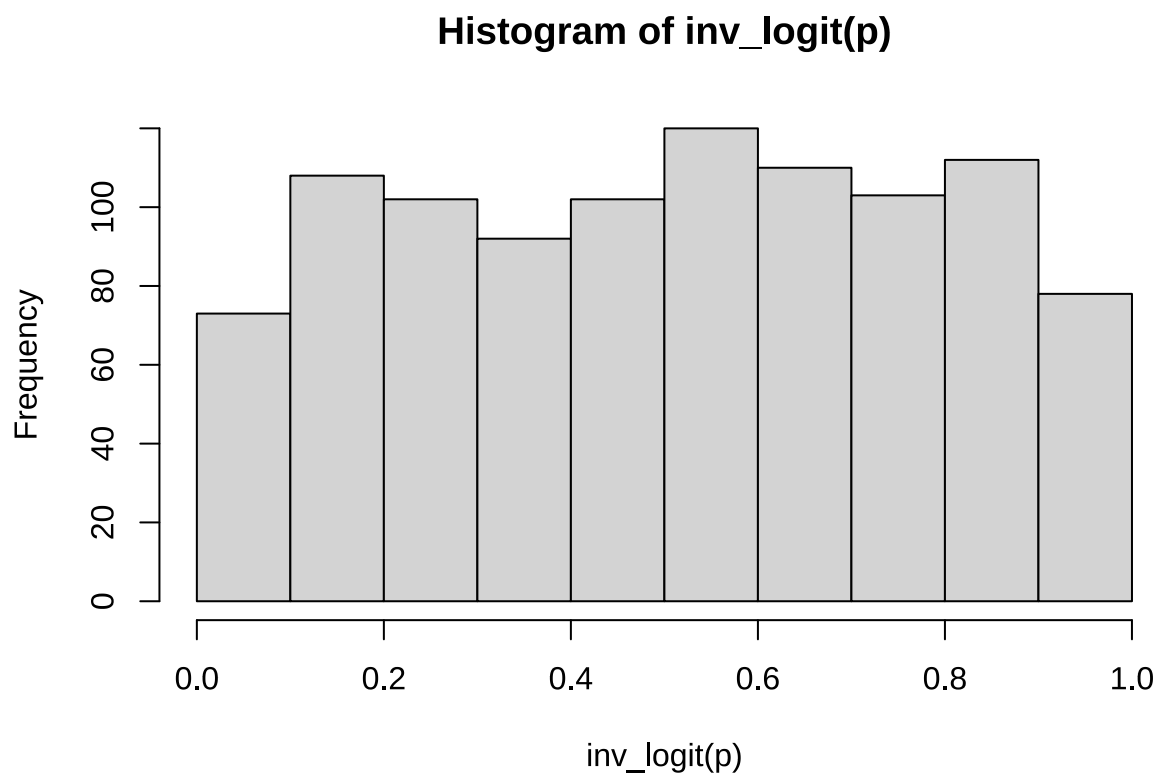
```
p <- rnorm(1000, 0, 1)
hist(inv_logit(p))
```



The bulk of the mass is too in the middle, that is, it is estimating the probability of a GB hole to be around 50%. a priori, that seems unlikely.

A $N(0, 1.5)$ is much more even and the one we will use:

```
p <- rnorm(1000, 0, 1.5)
hist(inv_logit(p))
```

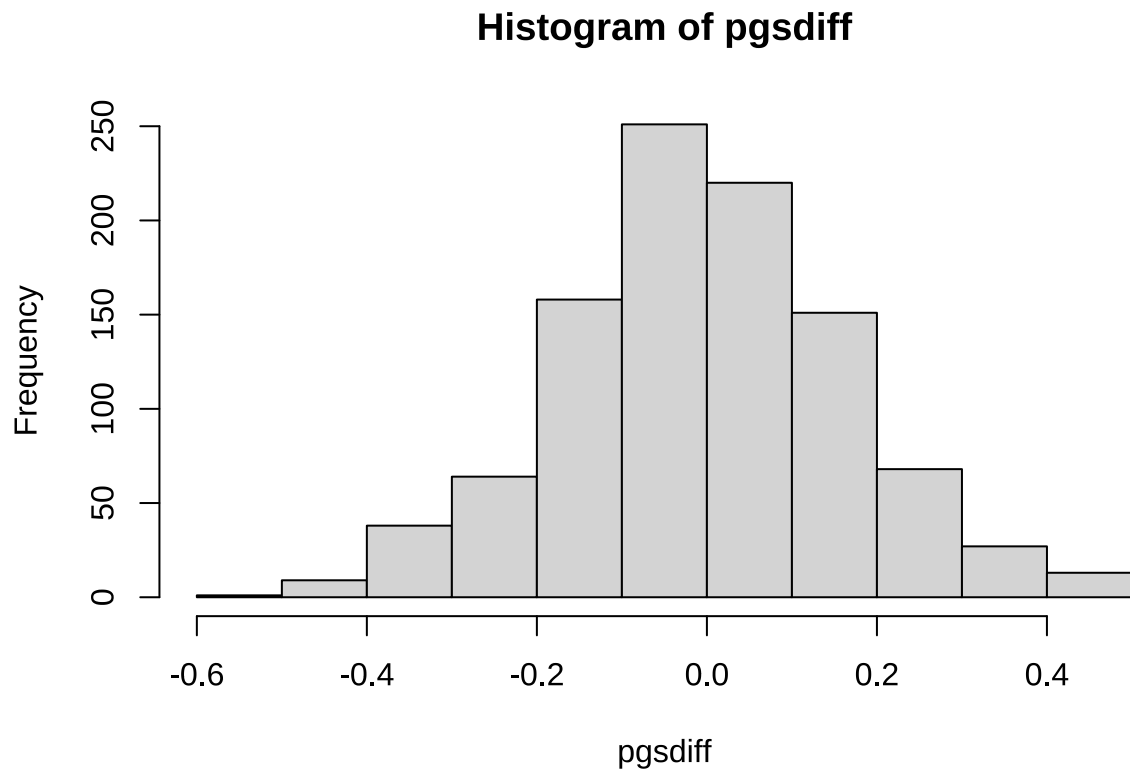


PGS coefficient

Remember, this is the effect of a PGS5. We will need to simulate the intercept then add the slope, followed by undoing the logit to see the probability:

A $N(0, 1)$

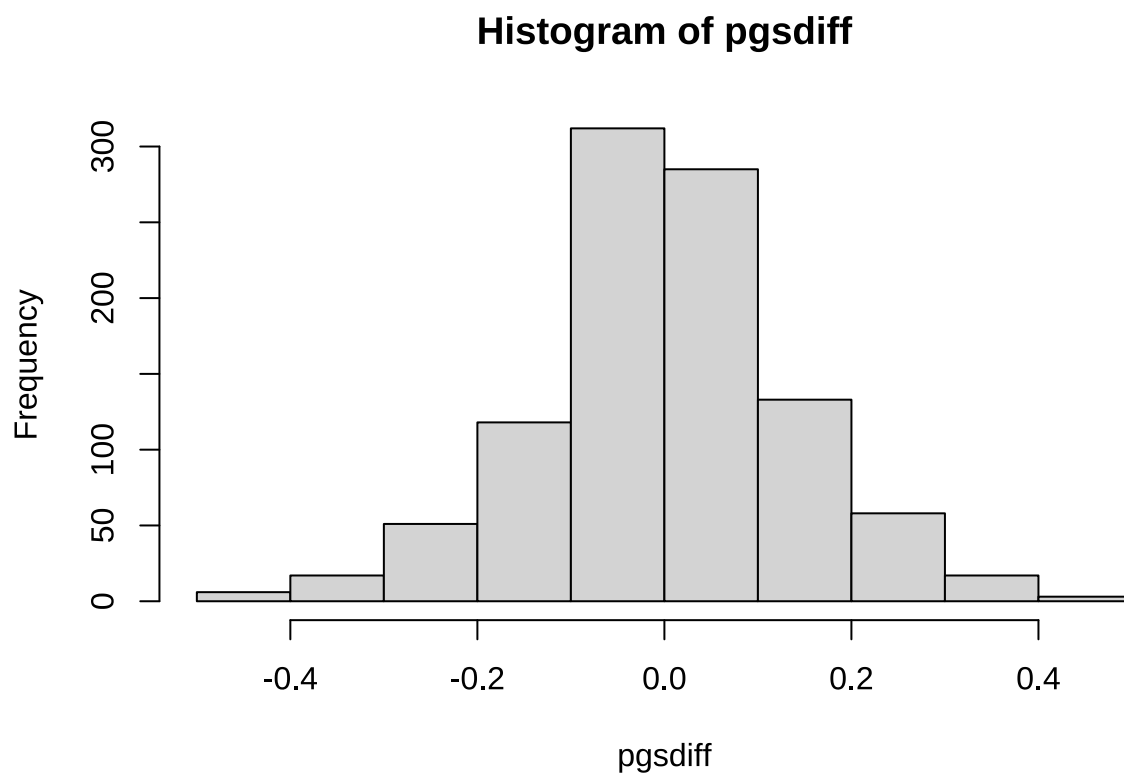
```
pint <- rnorm(1000, 0, 1.5)
ppgs <- rnorm(1000, 0, 1)
pgsdiff <- inv_logit(pint + ppgs) - inv_logit(pint)
hist(pgsdiff)
```



Is fairly reasonable (PGS5 does on average no change in getting a GB hole, but can change it by 60% probability in rare instances).

That seems a little too strong of an effect, so let's decrease with $N(0, 0.75)$:

```
pint <- rnorm(1000, 0, 1.5)
ppgs <- rnorm(1000, 0, 0.75)
pgsdiff <- inv_logit(pint + ppgs) - inv_logit(pint)
hist(pgsdiff)
```



other priors

Other priors will be the usual weakly regularizing ones, including Dirichlet of 2, Exponential 1, and LKJCorr of 4.

Formula

Below is the centered version. The model given to Stan is the non-centered version that is mathematically equivalent but dramatically improves sampling.

$$\begin{aligned}
Hole_i &\sim Bernoulli(p_i) \\
logit(p_i) &= \alpha_{sid[i]} + \beta_{sid[i]} * \sum_{j=0}^{PGS_i-1} \delta_j \\
\begin{bmatrix} \alpha_{sid} \\ \beta_{sid} \end{bmatrix} &\sim MVNormal\left(\begin{bmatrix} \alpha \\ \beta \end{bmatrix}, \mathbf{S}\right) \\
\alpha &\sim Normal(0, 1.5) \\
\beta &\sim Normal(0, 0.75) \\
\delta &\sim Dirichlet(2) \\
\mathbf{S} &= \begin{pmatrix} \sigma_\alpha & 0 \\ 0 & \sigma_\beta \end{pmatrix} \mathbf{R} \begin{pmatrix} \sigma_\alpha & 0 \\ 0 & \sigma_\beta \end{pmatrix} \\
\mathbf{R} &= \begin{pmatrix} 1 & \rho \\ \rho & 1 \end{pmatrix} \\
\mathbf{R} &\sim LKJCorr(4) \\
\sigma_\alpha, \sigma_\beta &\sim Exponential(1)
\end{aligned}
\tag{2}$$

```

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```



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```

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```

```

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## Chain 3 Iteration: 5000 / 5000 [100%] (Sampling)
## Chain 4 Iteration: 4900 / 5000 [ 98%] (Sampling)
## Chain 1 finished in 3.0 seconds.
## Chain 2 finished in 2.9 seconds.
## Chain 3 finished in 3.0 seconds.
## Chain 4 Iteration: 5000 / 5000 [100%] (Sampling)
## Chain 4 finished in 3.0 seconds.
##
## All 4 chains finished successfully.
## Mean chain execution time: 3.0 seconds.
## Total execution time: 3.2 seconds.

```

Diagnostic Evaluation of Markov Chains

Rhat4 and effective sampling size

```
precis(hole_mod, depth = 3)
```

##		mean	sd	5.5%	94.5%	n_eff
##	a_bar	-1.3826316500	0.31848139	-1.90273440	-0.8852383	8979.762
##	bP_bar	0.5860863112	0.56080178	-0.32735928	1.4632966	9197.078
##	delta[1]	0.1963108936	0.12032544	0.04255204	0.4174099	13942.335
##	delta[2]	0.2960796980	0.15259443	0.07844509	0.5607019	12859.262
##	delta[3]	0.2517295763	0.14119610	0.05996026	0.5003064	13981.086
##	delta[4]	0.2558798319	0.14226006	0.05865852	0.5109675	13325.849
##	z_sid[1,1]	-0.2680234545	0.84707966	-1.57988155	1.0960221	13389.775
##	z_sid[1,2]	0.4428785263	0.91494280	-1.08626245	1.8223225	8565.378
##	z_sid[1,3]	0.1404743506	0.86371689	-1.26426440	1.4953206	11537.128
##	z_sid[1,4]	-0.4406078745	0.89892164	-1.84642425	1.0187180	12285.393
##	z_sid[1,5]	0.0451315484	0.88610944	-1.38255100	1.4457191	16115.530
##	z_sid[1,6]	-0.3144259405	0.93568743	-1.78824695	1.1856433	13722.441
##	z_sid[1,7]	0.0026572858	0.93992277	-1.52948015	1.4780507	15456.629
##	z_sid[1,8]	0.1596764919	0.93939233	-1.35336765	1.6614847	14951.069
##	z_sid[1,9]	0.3262559676	0.98434474	-1.26689045	1.8541843	12670.376
##	z_sid[1,10]	-0.3311636644	0.98888466	-1.88631695	1.2736729	14990.080
##	z_sid[2,1]	-0.0137703779	0.86450538	-1.39452375	1.3764800	13618.729
##	z_sid[2,2]	0.7225369725	0.87893525	-0.76879505	2.0615121	8494.220
##	z_sid[2,3]	-0.5404047987	0.95252796	-2.03077895	1.0141886	11923.747
##	z_sid[2,4]	-0.1671835743	0.91256035	-1.61525055	1.2915584	15271.088
##	z_sid[2,5]	-0.0542666528	0.92549679	-1.54724060	1.4008160	15807.034
##	z_sid[2,6]	0.0080325677	0.92640357	-1.48253045	1.4824769	13544.957
##	z_sid[2,7]	0.0919204801	0.91020555	-1.39482765	1.5261582	14220.981
##	z_sid[2,8]	0.2811958137	0.96989427	-1.29406300	1.8349766	14744.338
##	z_sid[2,9]	0.5066919372	0.95003818	-1.05127485	1.9600038	12444.594
##	z_sid[2,10]	-0.1933510552	0.96749125	-1.73644495	1.3588837	16637.308
##	sigma_sid[1]	0.3678874557	0.29865146	0.03045718	0.9266509	5074.428
##	sigma_sid[2]	0.8103188851	0.64316838	0.06166090	1.9784393	5034.385
##	L_Rho_sid[1,1]	1.0000000000	0.00000000	1.00000000	1.0000000	NaN
##	L_Rho_sid[1,2]	0.0000000000	0.00000000	0.00000000	0.0000000	NaN
##	L_Rho_sid[2,1]	-0.0256299181	0.32498198	-0.53956611	0.4973029	11833.618
##	L_Rho_sid[2,2]	0.9424830887	0.07393621	0.79544456	0.9996901	5309.883
##	bP[1]	0.6166678177	0.82824939	-0.69834796	1.8805989	10343.783
##	bP[2]	1.2854067142	0.79149138	0.14505816	2.6461369	7764.960
##	bP[3]	-0.0008982751	1.19111342	-2.16518825	1.4472370	7306.758
##	bP[4]	0.4301040061	0.98189209	-1.25428810	1.7856643	9887.653
##	bP[5]	0.5298460929	1.00643362	-1.12598080	1.9328967	10097.770
##	bP[6]	0.6371909064	0.98080046	-0.89593169	2.0722067	10980.284
##	bP[7]	0.7126906019	0.92445199	-0.72701333	2.1021274	12428.390
##	bP[8]	0.9147067141	1.07995680	-0.57429527	2.6369995	10302.683
##	bP[9]	1.1660275711	1.03873357	-0.21625851	2.9779732	9469.926
##	bP[10]	0.3539717791	1.18717124	-1.67208630	1.8848414	9283.098
##	a[1]	-1.4998808111	0.38872381	-2.15612210	-0.9283229	11298.882
##	a[2]	-1.1847544972	0.41283226	-1.81172980	-0.4988825	9009.631
##	a[3]	-1.3202538583	0.36199746	-1.89868430	-0.7429484	12326.175
##	a[4]	-1.5954647520	0.47683464	-2.43332935	-0.9432220	7995.159
##	a[5]	-1.3618487657	0.41966448	-2.04731440	-0.7180963	12922.930
##	a[6]	-1.5434330775	0.50406777	-2.41351815	-0.8693691	9429.728

```

## a[7] -1.3818445936 0.47788656 -2.12114660 -0.6750021 11025.689
## a[8] -1.2960082544 0.48596934 -2.02055715 -0.5223752 11380.072
## a[9] -1.2083199819 0.53304410 -1.95187165 -0.3068774 9219.834
## a[10] -1.5645614981 0.56525156 -2.53296495 -0.8353756 8994.751
## Rho_sid[1,1] 1.0000000000 0.00000000 1.00000000 1.0000000 NaN
## Rho_sid[1,2] -0.0256299181 0.32498198 -0.53956611 0.4973029 11833.618
## Rho_sid[2,1] -0.0256299181 0.32498198 -0.53956611 0.4973029 11833.618
## Rho_sid[2,2] 1.0000000000 0.00000000 1.00000000 1.0000000 NaN
## ab_sid[1,1] -0.1172491578 0.32711263 -0.69330797 0.3241707 10354.589
## ab_sid[1,2] 0.0305814860 0.70615294 -1.01004720 1.1915695 10798.797
## ab_sid[2,1] 0.1978771722 0.35856443 -0.24741457 0.8657138 6931.912
## ab_sid[2,2] 0.6993203966 0.81335084 -0.19849025 2.2116013 5276.416
## ab_sid[3,1] 0.0623778177 0.32044568 -0.40059408 0.6243177 9933.756
## ab_sid[3,2] -0.5869845653 1.01341438 -2.49088730 0.4345075 7897.709
## ab_sid[4,1] -0.2128330683 0.40330567 -0.96309798 0.2370936 7909.462
## ab_sid[4,2] -0.1559823313 0.81901048 -1.52752265 0.9932486 11893.812
## ab_sid[5,1] 0.0207829060 0.34671726 -0.51554953 0.5833995 12662.388
## ab_sid[5,2] -0.0562402390 0.85953494 -1.42155190 1.1727073 10685.694
## ab_sid[6,1] -0.1608014557 0.42397913 -0.94423806 0.3492115 9204.745
## ab_sid[6,2] 0.0511046170 0.84817391 -1.20988545 1.4175699 11039.948
## ab_sid[7,1] 0.0007870842 0.39387131 -0.60889799 0.6047590 11938.298
## ab_sid[7,2] 0.1266042848 0.81509078 -1.01727775 1.4560045 9975.510
## ab_sid[8,1] 0.0866233637 0.40300116 -0.45738196 0.7835542 11373.189
## ab_sid[8,2] 0.3286203927 0.99651317 -0.85936993 2.0869241 8225.266
## ab_sid[9,1] 0.1743116289 0.45115193 -0.36216547 1.0119296 8484.258
## ab_sid[9,2] 0.5799412533 1.00453399 -0.48558835 2.4571680 7113.841
## ab_sid[10,1] -0.1819298869 0.48537782 -1.04371630 0.3621447 9063.103
## ab_sid[10,2] -0.2321145447 0.99522748 -1.87405685 1.0377873 10736.335
## Rhat4
## a_bar 0.9997749
## bP_bar 1.0001568
## delta[1] 0.9997256
## delta[2] 0.9999080
## delta[3] 0.9997599
## delta[4] 0.9999473
## z_sid[1,1] 0.9999703
## z_sid[1,2] 1.0002516
## z_sid[1,3] 0.9997692
## z_sid[1,4] 1.0000150
## z_sid[1,5] 0.9997936
## z_sid[1,6] 0.9999944
## z_sid[1,7] 0.9997764
## z_sid[1,8] 0.9998223
## z_sid[1,9] 0.9998046
## z_sid[1,10] 0.9998023
## z_sid[2,1] 1.0000344
## z_sid[2,2] 1.0003897
## z_sid[2,3] 0.9999215
## z_sid[2,4] 0.9998411
## z_sid[2,5] 0.9999645
## z_sid[2,6] 1.0000676
## z_sid[2,7] 0.9999433
## z_sid[2,8] 1.0000963
## z_sid[2,9] 0.9999750

```

```

## z_sid[2,10]      0.9996388
## sigma_sid[1]     1.0006535
## sigma_sid[2]     1.0008714
## L_Rho_sid[1,1]   NaN
## L_Rho_sid[1,2]   NaN
## L_Rho_sid[2,1]   0.9996744
## L_Rho_sid[2,2]   1.0001170
## bP[1]            1.0000179
## bP[2]            1.0003592
## bP[3]            1.0002064
## bP[4]            1.0002444
## bP[5]            1.0000277
## bP[6]            0.9999787
## bP[7]            0.9998517
## bP[8]            0.9999688
## bP[9]            1.0000127
## bP[10]           0.9999843
## a[1]             1.0000490
## a[2]             1.0002031
## a[3]             0.9998702
## a[4]             0.9999863
## a[5]             0.9999142
## a[6]             1.0003368
## a[7]             0.9999020
## a[8]             0.9999780
## a[9]             0.9999501
## a[10]            0.9997137
## Rho_sid[1,1]     NaN
## Rho_sid[1,2]     0.9996744
## Rho_sid[2,1]     0.9996744
## Rho_sid[2,2]     NaN
## ab_sid[1,1]      1.0001156
## ab_sid[1,2]      1.0002616
## ab_sid[2,1]      1.0003290
## ab_sid[2,2]      1.0005997
## ab_sid[3,1]      0.9998307
## ab_sid[3,2]      1.0002095
## ab_sid[4,1]      1.0000470
## ab_sid[4,2]      1.0001087
## ab_sid[5,1]      0.9997569
## ab_sid[5,2]      0.9998781
## ab_sid[6,1]      1.0003932
## ab_sid[6,2]      0.9997136
## ab_sid[7,1]      0.9999204
## ab_sid[7,2]      1.0002205
## ab_sid[8,1]      0.9999696
## ab_sid[8,2]      1.0000858
## ab_sid[9,1]      1.0000457
## ab_sid[9,2]      1.0003082
## ab_sid[10,1]     0.9998571
## ab_sid[10,2]     1.0000915

```

All Rhat4 values are 1.

Each parameter also sampled well.

PSIS/WAIC

```
PSIS(hole_mod)
```

```
## Some Pareto k values are high (>0.5). Set pointwise=TRUE to inspect individual points.
```

```
##      PSIS      lppd  penalty  std_err  
## 1 172.8493 -86.42464 6.148971 12.23386
```

```
WAIC(hole_mod)
```

```
##      WAIC      lppd  penalty  std_err  
## 1 172.6463 -80.27567 6.047468 12.17055
```

The are some Pareto k values > 0.5. As long as < 0.7, not an issue. Are they?

```
PSIS(hole_mod, pointwise = TRUE) %>%  
  high_k_rows()
```

```
## Some Pareto k values are high (>0.5). Set pointwise=TRUE to inspect individual points.
```

```
## # A tibble: 2 x 15  
##   videoid PSIS  lppd penalty std_err      k surgid  pgs time_until_1st_  
##   <int> <dbl> <dbl>   <dbl>   <dbl> <dbl> <int> <int>      <dbl>  
## 1     98 0.733 -0.367  0.0530   12.2 0.503     4     5      39.5  
## 2    130 3.23  -1.61   0.330   12.2 0.557     6     4      22.6  
## # ... with 6 more variables: time_cvs_attained <dbl>,  
## #   laparoscopic_duration <dbl>, dissection_duration <dbl>,  
## #   gb_removal_duration <dbl>, gb_hole <lgl>, gb_holes <int>
```

Only 2 rows total, and both have k < 0.6, so minimal issue with outliers.

Trace rank plot (trankplot)

```
trankplot(hole_mod)
```

Good mixing as shown with large amount of overlap.

Trace plot

```
traceplot(hole_mod)
```

Chains are stationary with a visible central tendency, have good mixing, and converge.

Evaluate model results

First, obtain tidy samples, as we did with the duration model. Additionally, transform the result to get the absolute probability of a hole and the OR of a hole for each PGS:

```
tidy_hole_samples <- extract_samples(  
  hole_mod,  
  pars = c(  
    "a_bar",  
    paste0("a[", 1:nsurgs, "]"),  
    "bP_bar",  
    paste0("bP[", 1:nsurgs, "]"),  
    # sigma_a  
    "sigma_sid[1]",
```

```

      # sigma_b
      "sigma_sid[2]",
      paste0("Rho_sid[1,2]"),
      paste0("delta[", 1:4, "]")
    )
  ) %>%
  tidy_surgeons() %>%
  sum_deltas() %>%
  tidy_pgs() %>%
  mutate(
    p = inv_logit(a + bP * sum_delta_j),
    bOR = exp(bP * sum_delta_j)
  ) %>%
  arrange(sample_num, surgeon, PGS) %>%
  group_by(surgeon, sample_num) %>%
  # calc change in probability for each sample
  # group by surgeon as well because each sample_num
  # has the info for all 10 surgeons and bar surgeon
  mutate(
    # change from one PGS level to the other
    incr_delta_p = p - lag(p),
    # change from PGS1 to that PGS level
    delta_p = p - p[1]
  ) %>%
  ungroup()

```

Key to know, is that:

`incr_delta_p` is the probability difference from one level to the next. It will be NA for a PGS1.

`delta_p` is probability difference from a certain PGS level to PGS1.

`p` is the probability of a hole at that level

`bOR` is the increased odds, compared to a PGS of 1, of a gb hole.

Plot results

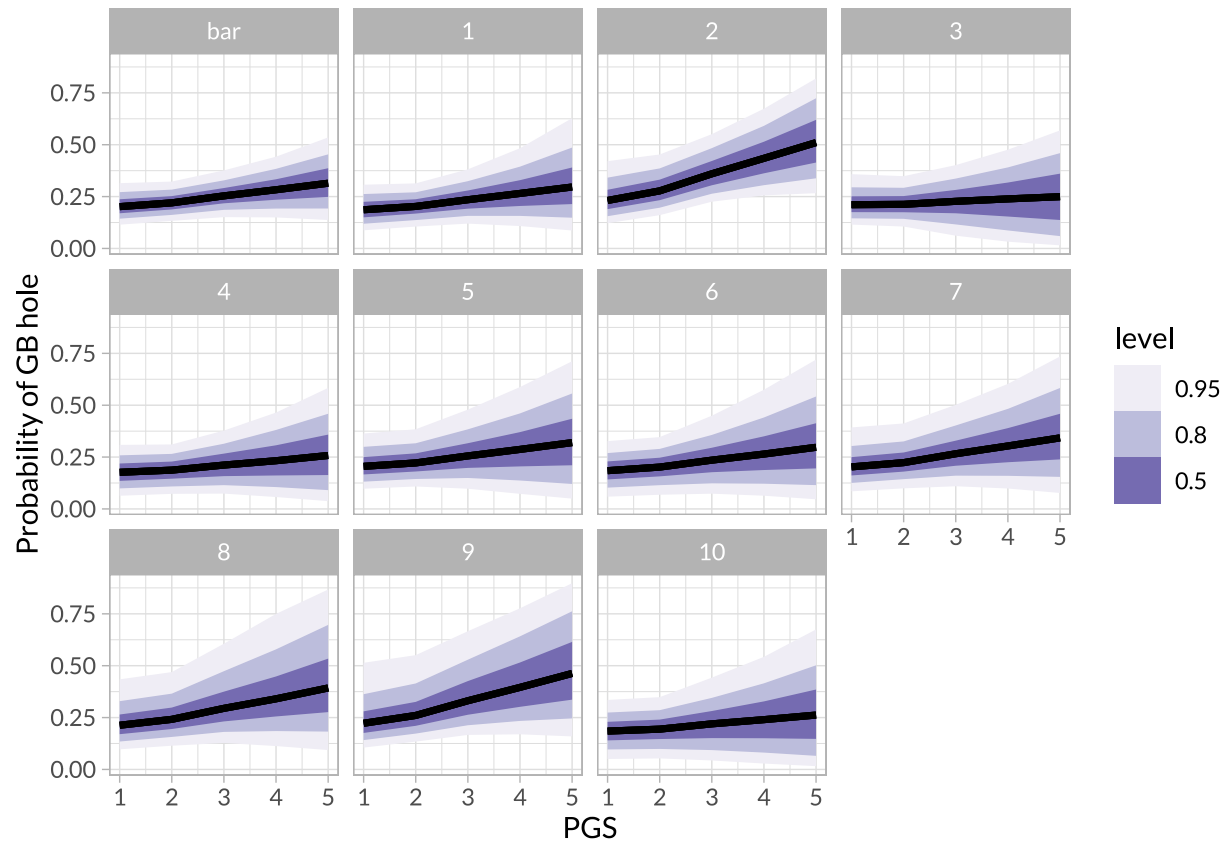
For all surgeons

Remember that “bar” is the average of all the surgeons:

```

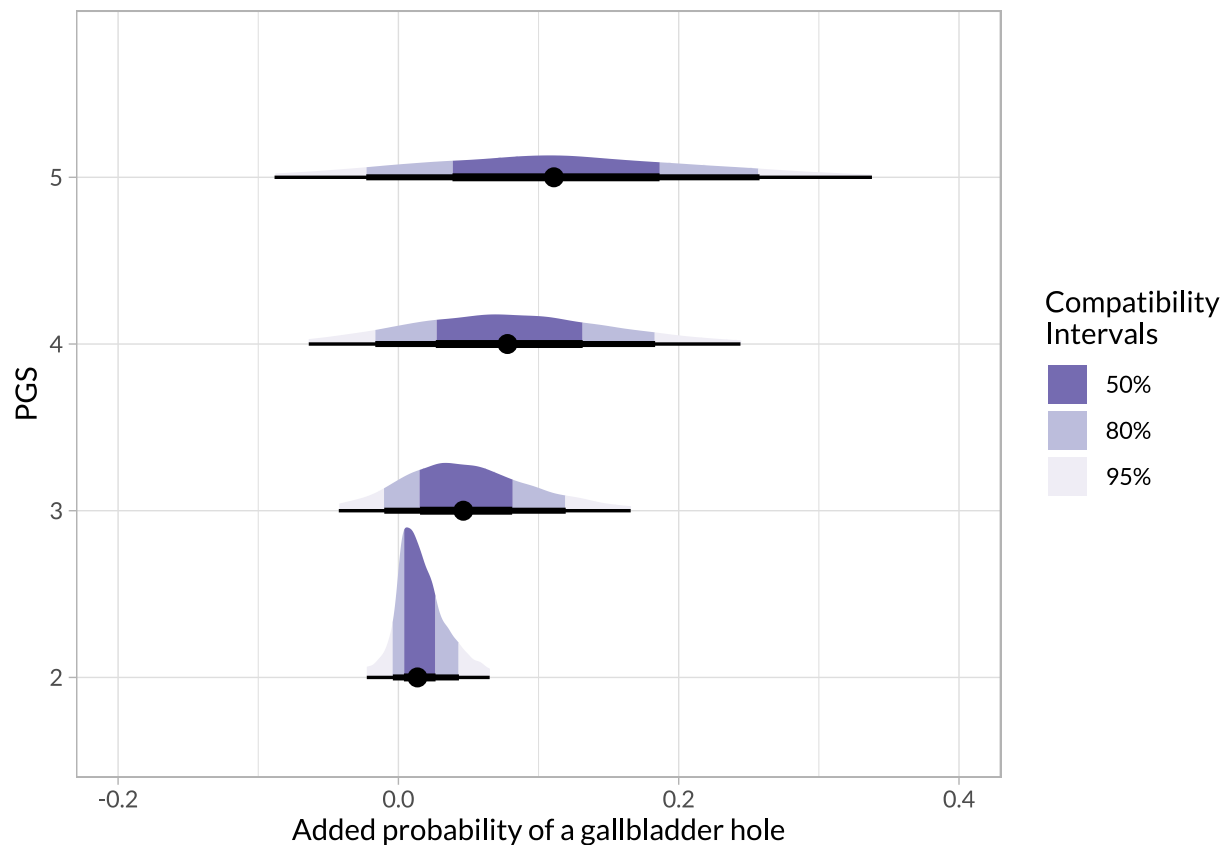
tidy_hole_samples %>%
  mutate(surgeon = parse_factor(surgeon, levels = c("bar", as.character(1:nsurgs)))) %>%
  ggplot(aes(PGS, p)) +
  stat_lineribbon() +
  facet_wrap(~ surgeon) +
  scale_fill_brewer(palette = "Purples") +
  labs(y = "Probability of GB hole")

```

On average across all surgeons

```
bar_gb <- tidy_hole_samples %>%
  filter(surgeon == "bar", PGS != 1) %>%
  mutate(PGS = as.factor(PGS))
bar_gb %>%
  halfeye(delta_p, PGS) +
  coord_cartesian(xlim = c(-0.2, 0.4)) +
  labs(
    y = "PGS",
    x = "Added probability of a gallbladder hole"
  )
```



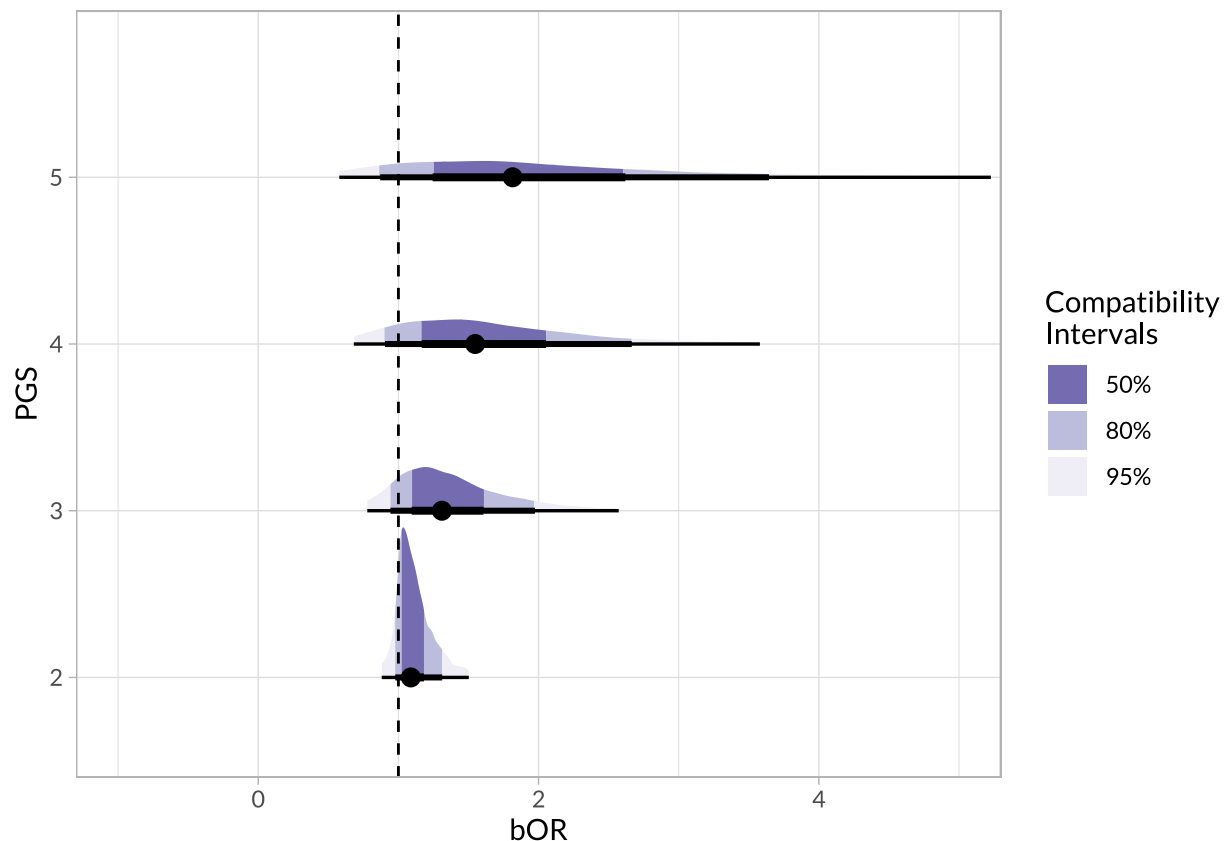
And numbers for this:

```
bar_gb %>%
  group_by(PGS) %>%
  ci_ints(delta_p)
```

```
## # A tibble: 4 x 7
##   PGS   mean int_50      int_66      int_80      int_89      int_95
##   <fct> <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1 2      0.02 0, 0.03    0, 0.03    0, 0.03    -0.01, 0.05 -0.02, 0.07
## 2 3      0.05 0.02, 0.08 0, 0.1      0.01, 0.09 -0.03, 0.14 -0.04, 0.17
## 3 4      0.08 0.03, 0.13 0.01, 0.15 0.01, 0.15 -0.04, 0.21 -0.06, 0.24
## 4 5      0.11 0.04, 0.19 0.01, 0.22 0.02, 0.21 -0.05, 0.3  -0.09, 0.34
```

And looking at the odd ratios:

```
bar_gb %>%
  halfeye(bOR, PGS) +
  coord_cartesian(xlim = c(-1, 5)) +
  geom_vline(xintercept = 1, linetype = 2)
```



Numbers for this:

```
bar_gb %>%
  group_by(PGS) %>%
  ci_ints(bOR)
```

```
## # A tibble: 4 x 7
##   PGS   mean int_50      int_66      int_80      int_89      int_95
##   <fct> <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1 2      1.12 1.02, 1.18 1.01, 1.24 1.01, 1.22 0.94, 1.4 0.88, 1.5
## 2 3      1.4  1.09, 1.61 1.02, 1.77 1.05, 1.7  0.86, 2.23 0.78, 2.57
## 3 4      1.7  1.17, 2.05 1.04, 2.3  1.09, 2.2  0.79, 3.07 0.68, 3.58
## 4 5      2.1  1.24, 2.62 1.06, 3.04 1.13, 2.85 0.72, 4.32 0.58, 5.23
```

For a surgeon most affected by PGS

```
hole_bps <- precis(hole_mod, depth = 2, pars = paste0("bP[", 1:nsurgs, "]")) %>%
  as_tibble(rownames = "var")
# Least affected surgeon:
slice_min(hole_bps, mean)
```

```
## # A tibble: 1 x 7
##   var      mean    sd '5.5%' '94.5%' n_eff Rhat4
##   <chr>    <dbl> <dbl> <dbl> <dbl> <dbl> <dbl>
## 1 bP[3] -0.000898 1.19 -2.17 1.45 7307. 1.00
```

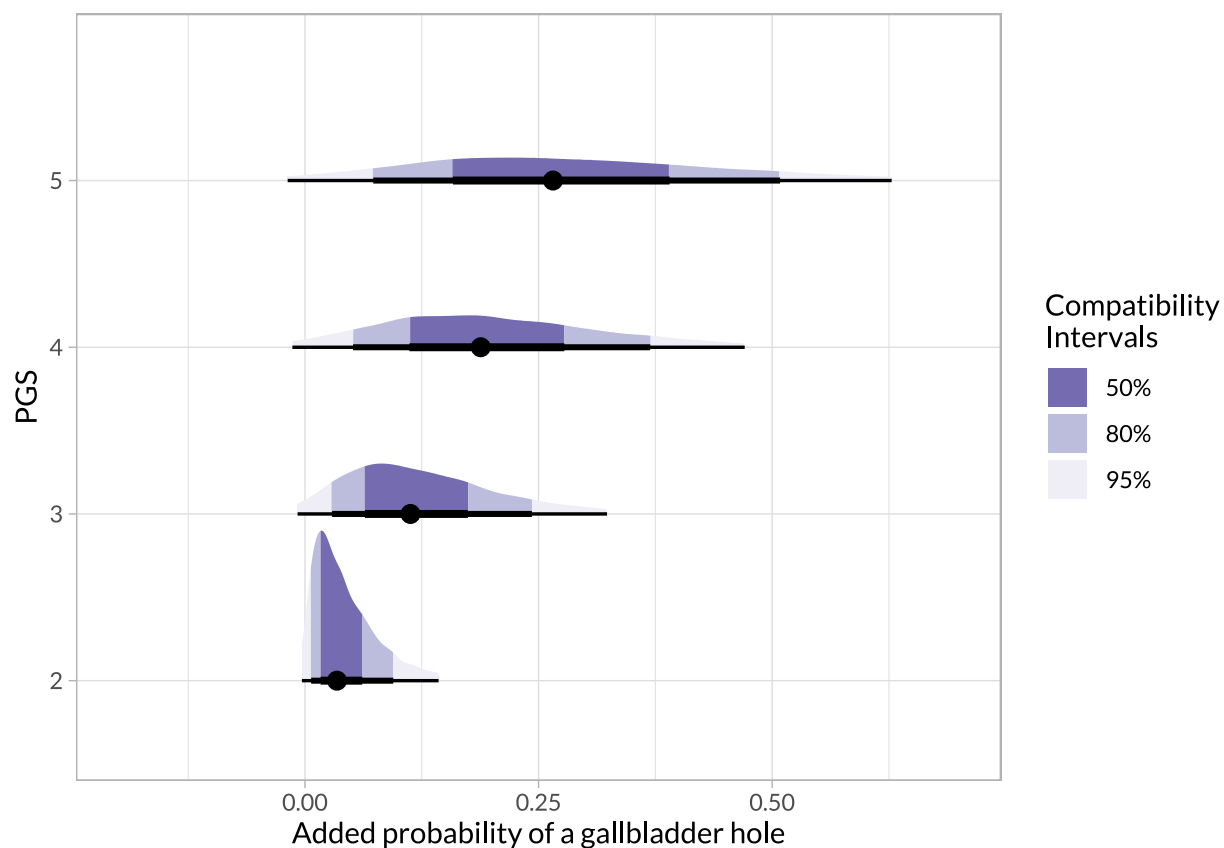
```
# Most affected surgeon:
slice_max(hole_bps, mean)
```

```
## # A tibble: 1 x 7
##   var      mean    sd '5.5%' '94.5%' n_eff Rhat4
##   <chr> <dbl> <dbl> <dbl> <dbl> <dbl> <dbl>
## 1 bP[2]  1.29 0.791 0.145   2.65 7765.  1.00
```

Surgeon 2 is most affected.

```
surg2_gb <- tidy_hole_samples %>%
  filter(surgeon == "2", PGS != 1) %>%
  mutate(PGS = as.factor(PGS))

surg2_gb %>%
  halfeye(delta_p, PGS) +
  coord_cartesian(xlim = c(-0.2, 0.7)) +
  labs(
    y = "PGS",
    x = "Added probability of a gallbladder hole"
  )
```



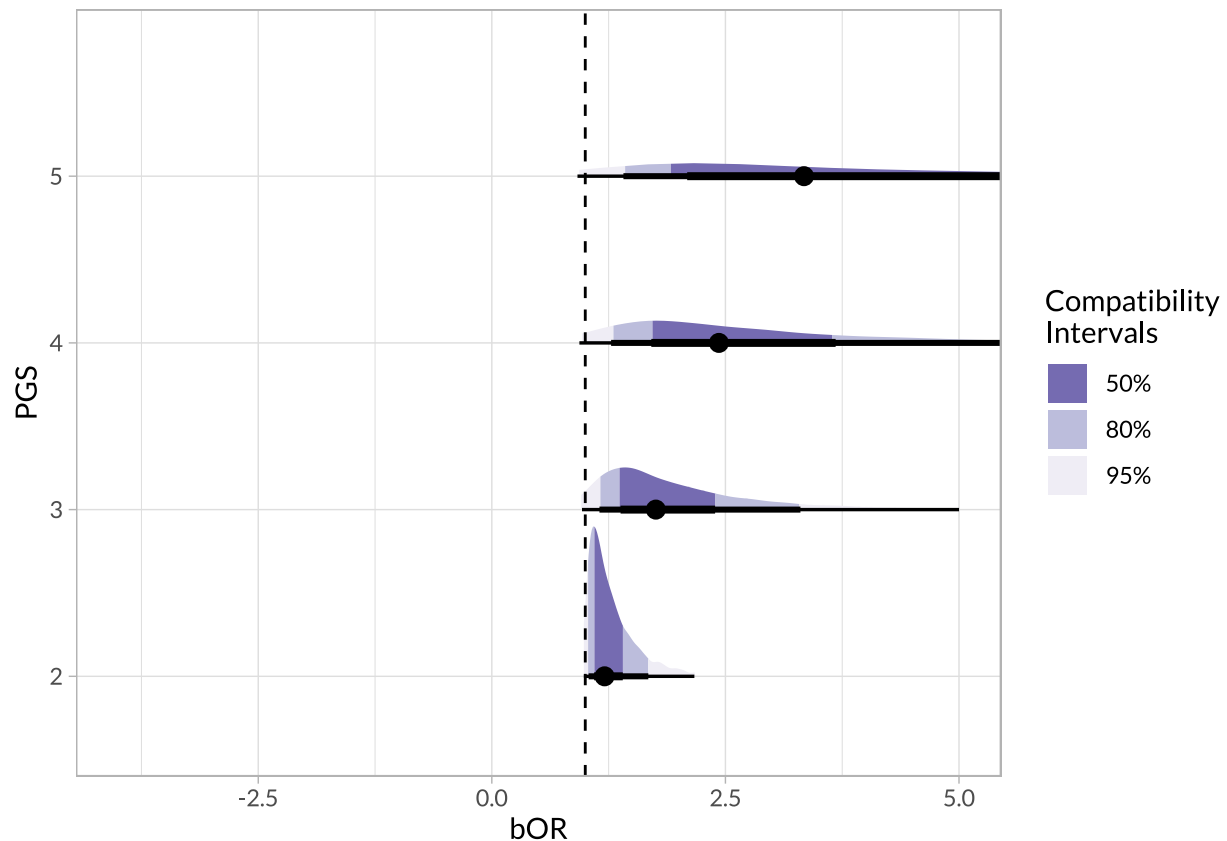
And numbers for this:

```
surg2_gb %>%
  group_by(PGS) %>%
  ci_ints(delta_p)
```

```
## # A tibble: 4 x 7
##   PGS   mean int_50      int_66      int_80      int_89      int_95
##   <fct> <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1 2      0.04 0.02, 0.06 0.01, 0.08 0.01, 0.07 0, 0.12    0, 0.14
## 2 3      0.13 0.06, 0.17 0.05, 0.2  0.05, 0.19 0.01, 0.28 -0.01, 0.32
## 3 4      0.2  0.11, 0.28 0.08, 0.32 0.1, 0.3   0.02, 0.42 -0.01, 0.47
## 4 5      0.28 0.16, 0.39 0.12, 0.44 0.14, 0.42 0.03, 0.57 -0.02, 0.63
```

And looking at odds ratios:

```
surg2_gb %>%
  halfeye(bOR, PGS) +
  coord_cartesian(xlim = c(-4, 5)) +
  geom_vline(xintercept = 1, linetype = 2)
```



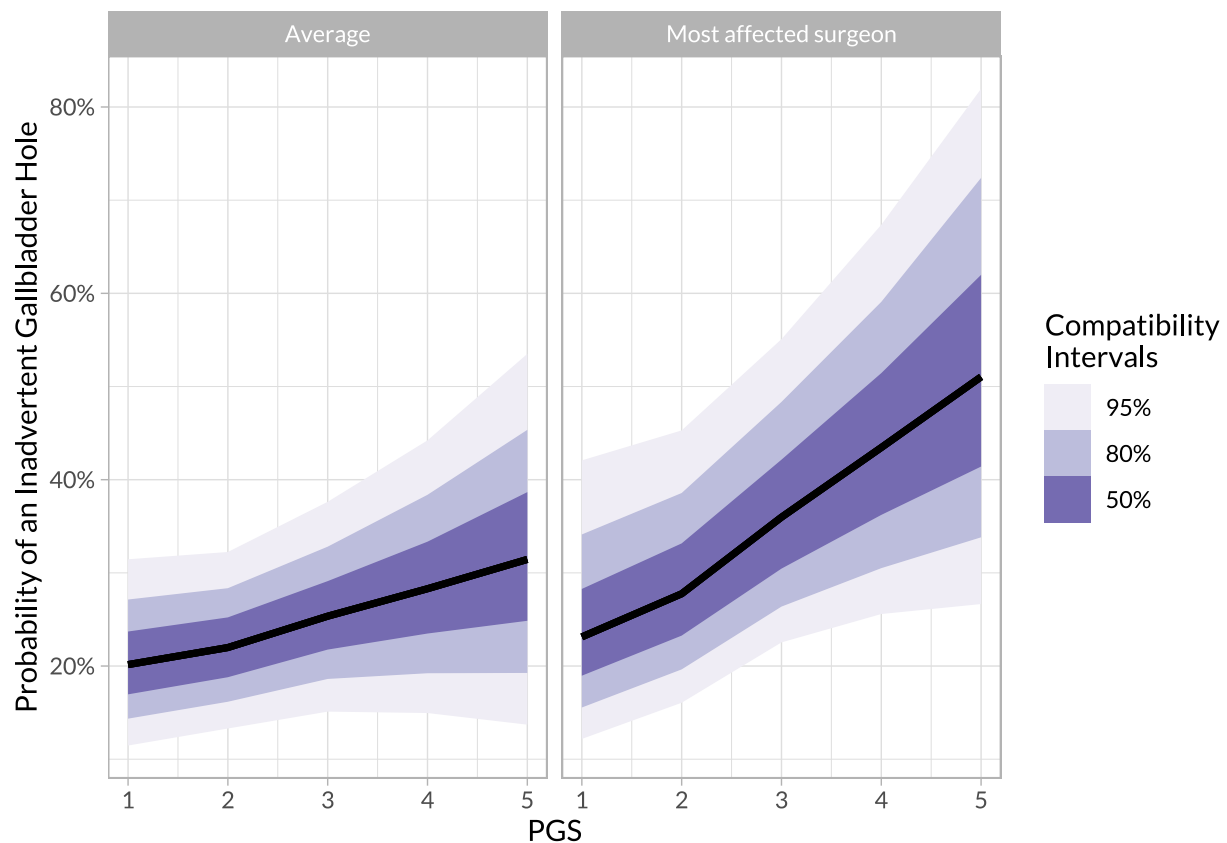
Numbers for this:

```
surg2_gb %>%
  group_by(PGS) %>%
  ci_ints(bOR)
```

```
## # A tibble: 4 x 7
##   PGS   mean int_50      int_66      int_80      int_89      int_95
##   <fct> <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1 2      1.3  1.09, 1.4  1.06, 1.52 1.08, 1.47 1.01, 1.87 0.99, 2.17
## 2 3      2.06 1.38, 2.39 1.26, 2.77 1.3, 2.61  1.06, 3.99 0.96, 5
## 3 4      3.09 1.71, 3.68 1.5, 4.47  1.58, 4.13 1.11, 7.06 0.94, 9.12
## 4 5      5.22 2.09, 5.83 1.74, 7.59 1.89, 6.81 1.16, 14.1 0.92, 20.45
```

And comparing to the average surgeon:

```
tidy_hole_samples %>%
  filter(surgeon %in% c("bar", "2")) %>%
  mutate(surgeon = if_else(surgeon == "bar", "Average", "Most affected surgeon")) %>%
  #mutate(surgeon = parse_factor(surgeon, levels = c("bar", as.character(1:nsurgs)))) %>%
  ggplot(aes(PGS, p)) +
  stat_lineribbon() +
  facet_wrap(~ surgeon) +
  scale_fill_brewer(labels = c("95%", "80%", "50%"), palette = "Purples") +
  scale_y_continuous(labels = scales::percent_format()) +
  labs(
    x = "PGS",
    y = "Probability of an Inadvertent Gallbladder Hole",
    fill = "Compatibility\nIntervals"
  )
)
```



```
ggsave("../output/pgs_hole.svg", width = 6, height = 4)
ggsave("../output/pgs_hole.pdf", width = 6, height = 4)
```

numbers for average surgeon:

```
tidy_hole_samples %>%
  filter(surgeon == "bar") %>%
  group_by(PGS) %>%
  ci_ints(p)
```

```
## # A tibble: 5 x 7
```

```
##      PGS mean int_50      int_66      int_80      int_89      int_95
##      <dbl> <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1      1 0.21 0.17, 0.24 0.16, 0.25 0.16, 0.25 0.13, 0.29 0.11, 0.31
## 2      2 0.22 0.19, 0.25 0.18, 0.27 0.18, 0.26 0.15, 0.3 0.13, 0.32
## 3      3 0.26 0.22, 0.29 0.2, 0.31 0.21, 0.3 0.17, 0.35 0.15, 0.38
## 4      4 0.290 0.23, 0.33 0.21, 0.36 0.22, 0.35 0.17, 0.41 0.15, 0.44
## 5      5 0.32 0.25, 0.39 0.22, 0.42 0.23, 0.4 0.17, 0.49 0.14, 0.53
```

numbers for most affected surgeon:

```
tidy_hole_samples %>%
  filter(surgeon == "2") %>%
  group_by(sample_num, PGS) %>%
  ci_ints(p)
```

'summarise()' has grouped output by 'sample_num'. You can override using the '.groups' argument.

```
## # A tibble: 50,000 x 8
##   sample_num PGS mean int_50      int_66      int_80      int_89      int_95
##   <chr>      <dbl> <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1 sample1      1 0.13 0.13, 0.~ 0.13, 0.~ 0.13, 0.~ 0.13, 0.~ 0.13, 0.~
## 2 sample1      2 0.23 0.23, 0.~ 0.23, 0.~ 0.23, 0.~ 0.23, 0.~ 0.23, 0.~
## 3 sample1      3 0.25 0.25, 0.~ 0.25, 0.~ 0.25, 0.~ 0.25, 0.~ 0.25, 0.~
## 4 sample1      4 0.3 0.3, 0.3 0.3, 0.3 0.3, 0.3 0.3, 0.3 0.3, 0.3
## 5 sample1      5 0.37 0.37, 0.~ 0.37, 0.~ 0.37, 0.~ 0.37, 0.~ 0.37, 0.~
## 6 sample10     1 0.14 0.14, 0.~ 0.14, 0.~ 0.14, 0.~ 0.14, 0.~ 0.14, 0.~
## 7 sample10     2 0.17 0.17, 0.~ 0.17, 0.~ 0.17, 0.~ 0.17, 0.~ 0.17, 0.~
## 8 sample10     3 0.26 0.26, 0.~ 0.26, 0.~ 0.26, 0.~ 0.26, 0.~ 0.26, 0.~
## 9 sample10     4 0.43 0.43, 0.~ 0.43, 0.~ 0.43, 0.~ 0.43, 0.~ 0.43, 0.~
## 10 sample10    5 0.47 0.47, 0.~ 0.47, 0.~ 0.47, 0.~ 0.47, 0.~ 0.47, 0.~
## # ... with 49,990 more rows
```

```
tidy_hole_samples %>%
  filter(surgeon == "2")
```

```
## # A tibble: 50,000 x 17
##   sample_num sigma_sid_1 sigma_sid_2 Rho_sid_1_2 delta_1 delta_2 delta_3
##   <chr>      <dbl>      <dbl>      <dbl>      <dbl>      <dbl>      <dbl>
## 1 sample1      0.350      0.774      0.0426     0.488     0.0862     0.209
## 2 sample1      0.350      0.774      0.0426     0.488     0.0862     0.209
## 3 sample1      0.350      0.774      0.0426     0.488     0.0862     0.209
## 4 sample1      0.350      0.774      0.0426     0.488     0.0862     0.209
## 5 sample1      0.350      0.774      0.0426     0.488     0.0862     0.209
## 6 sample10     0.176      1.10      0.111     0.118     0.325     0.459
## 7 sample10     0.176      1.10      0.111     0.118     0.325     0.459
## 8 sample10     0.176      1.10      0.111     0.118     0.325     0.459
## 9 sample10     0.176      1.10      0.111     0.118     0.325     0.459
## 10 sample10    0.176      1.10      0.111     0.118     0.325     0.459
## # ... with 49,990 more rows, and 10 more variables: delta_4 <dbl>,
## #   surgeon <chr>, a <dbl>, bP <dbl>, PGS <int>, sum_delta_j <dbl>,
## #   p <dbl>, bOR <dbl>, incr_delta_p <dbl>, delta_p <dbl>
```

Correlation of PGS1 and effect of incremental PGS

```
extract_samples(hole_mod, pars = c("Rho_sid[1,2]")) %>%
  rename(rho = Rho_sid_1_2) %>%
```

```
ci_ints(rho) %>%
knitr::kable(
  caption = "Correlation between GB hole in PGS1 case and the effect of incrementing PGS"
)
```

Table 8: Correlation between GB hole in PGS1 case and the effect of incrementing PGS

mean	int_50	int_66	int_80	int_89	int_95
-0.03	-0.27, 0.21	-0.35, 0.31	-0.32, 0.27	-0.54, 0.5	-0.63, 0.6

Critical View of Safety Attainment Analysis

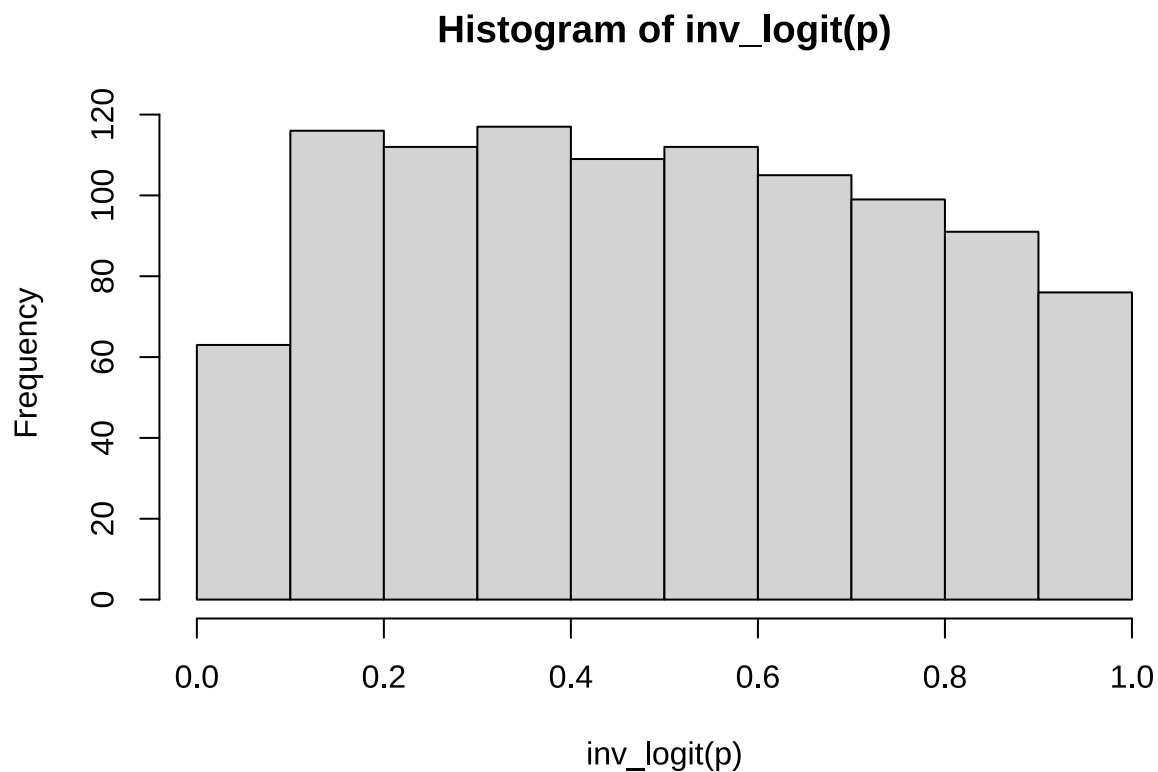
Priors determination

As before, we will use weakly regularizing priors.

Intercept

We will reuse the $N(0, 1.5)$ prior we established for GB hole:

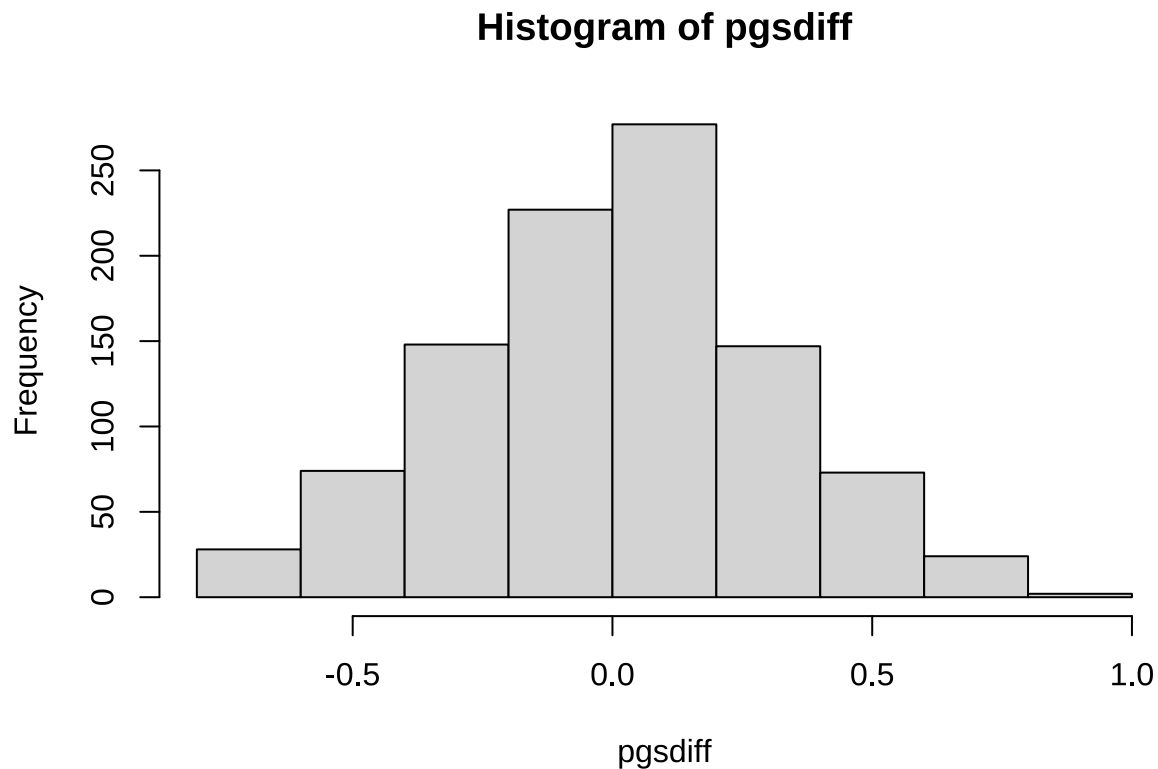
```
p <- rnorm(1000, 0, 1.5)
hist(inv_logit(p))
```



PGS coefficient

Remember, this is the effect of a PGS5. We will need to simulate the intercept then add the slope, followed by undoing the logit to see the probability:

```
pint <- rnorm(1000, 0, 1.5)
ppgs <- rnorm(1000, 0, 2)
pgsdiff <- inv_logit(pint + ppgs) - inv_logit(pint)
hist(pgsdiff)
```



This prior, $N(0, 2)$, is a wide and rather uninformative prior, but still fair. It allows for some surgeons to obtain CVS for a PGS1 but for PGS5 to never obtain it (due to technique preference). However, it still clusters most of the effects around zero, as we would, a priori, predict.

other priors

Other priors will be the usual weakly regularizing ones, including Dirichlet of 2, Exponential 1, and LKJCorr of 4.

Formula

Below is the centered version. The model given to Stan is the non-centered version that is mathematically equivalent but dramatically improves sampling.

$$\begin{aligned}
CVS_i &\sim \text{Bernoulli}(p_i) \\
\text{logit}(p_i) &= \alpha_{sid[i]} + \beta_{sid[i]} * \sum_{j=0}^{PGS_i-1} \delta_j \\
\begin{bmatrix} \alpha_{sid} \\ \beta_{sid} \end{bmatrix} &\sim \text{MVNormal}\left(\begin{bmatrix} \alpha \\ \beta \end{bmatrix}, \mathbf{S}\right) \\
\alpha &\sim \text{Normal}(0, 1.5) \\
\beta &\sim \text{Normal}(0, 2) \\
\delta &\sim \text{Dirichlet}(2) \\
\mathbf{S} &= \begin{pmatrix} \sigma_\alpha & 0 \\ 0 & \sigma_\beta \end{pmatrix} \mathbf{R} \begin{pmatrix} \sigma_\alpha & 0 \\ 0 & \sigma_\beta \end{pmatrix} \\
\mathbf{R} &= \begin{pmatrix} 1 & \rho \\ \rho & 1 \end{pmatrix} \\
\mathbf{R} &\sim \text{LKJCorr}(4) \\
\sigma_\alpha, \sigma_\beta &\sim \text{Exponential}(1)
\end{aligned}
\tag{3}$$

```

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```

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```

```

## Chain 4 Iteration: 3800 / 5000 [ 76%] (Sampling)
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## Chain 3 Iteration: 4700 / 5000 [ 94%] (Sampling)
## Chain 4 Iteration: 4600 / 5000 [ 92%] (Sampling)
## Chain 4 Iteration: 4700 / 5000 [ 94%] (Sampling)
## Chain 2 finished in 2.9 seconds.
## Chain 1 Iteration: 4900 / 5000 [ 98%] (Sampling)
## Chain 1 Iteration: 5000 / 5000 [100%] (Sampling)
## Chain 3 Iteration: 4800 / 5000 [ 96%] (Sampling)
## Chain 3 Iteration: 4900 / 5000 [ 98%] (Sampling)
## Chain 4 Iteration: 4800 / 5000 [ 96%] (Sampling)
## Chain 1 finished in 3.0 seconds.
## Chain 3 Iteration: 5000 / 5000 [100%] (Sampling)
## Chain 4 Iteration: 4900 / 5000 [ 98%] (Sampling)
## Chain 4 Iteration: 5000 / 5000 [100%] (Sampling)
## Chain 3 finished in 3.0 seconds.
## Chain 4 finished in 3.0 seconds.
##
## All 4 chains finished successfully.
## Mean chain execution time: 3.0 seconds.
## Total execution time: 3.2 seconds.

```

Diagnostic Evaluation of Markov Chains

Rhat4 and effective sampling size

```
precis(cvs_mod, depth = 3)
```

##		mean	sd	5.5%	94.5%	n_eff
##	a_bar	-0.448264277	0.61688817	-1.41086435	0.5275515	5526.114
##	bP_bar	-0.889484396	0.78108251	-2.11310205	0.3496151	9201.120
##	delta[1]	0.328473683	0.16433750	0.08260989	0.6057762	13496.905
##	delta[2]	0.220419204	0.13195595	0.04725604	0.4609032	14146.265
##	delta[3]	0.212912739	0.13240279	0.04365943	0.4563196	12076.815
##	delta[4]	0.238194371	0.13766828	0.05413588	0.4869947	13531.513
##	z_sid[1,1]	-1.839045735	0.65210868	-2.91555170	-0.8459421	9366.012
##	z_sid[1,2]	0.618261270	0.45749453	-0.06635933	1.3862307	6149.916
##	z_sid[1,3]	0.669119309	0.46456034	-0.03185255	1.4434717	5964.906
##	z_sid[1,4]	0.362315778	0.47940086	-0.37027781	1.1554911	7389.905
##	z_sid[1,5]	-1.380382002	0.67677304	-2.51579955	-0.3565313	11137.968
##	z_sid[1,6]	-0.302936852	0.57023348	-1.22991925	0.5780491	9718.295
##	z_sid[1,7]	0.582620856	0.60083384	-0.35222335	1.5378422	8832.196
##	z_sid[1,8]	0.157058844	0.63259009	-0.84518418	1.1522848	9723.647
##	z_sid[1,9]	0.564585927	0.65212853	-0.48601591	1.5798250	8548.197
##	z_sid[1,10]	0.153927964	0.60398823	-0.82178581	1.1014727	9986.034
##	z_sid[2,1]	-0.249554790	1.00763955	-1.86455245	1.3696495	13200.527
##	z_sid[2,2]	-0.065789320	0.88995102	-1.49860715	1.3800517	12929.078
##	z_sid[2,3]	-0.018175847	0.90890870	-1.48154160	1.4394117	13921.625
##	z_sid[2,4]	-0.151404376	0.94401335	-1.64393310	1.3765873	13931.730
##	z_sid[2,5]	-0.145643791	0.98203323	-1.71762960	1.4358294	14578.909
##	z_sid[2,6]	-0.090901718	0.97910390	-1.65641080	1.4929421	15407.370
##	z_sid[2,7]	0.199337105	0.98718006	-1.39282815	1.7434569	14347.200
##	z_sid[2,8]	0.124666472	0.97829507	-1.44821740	1.6918144	14862.009
##	z_sid[2,9]	0.345549564	1.02640615	-1.30792115	1.9698708	12629.499
##	z_sid[2,10]	-0.150004857	0.98051893	-1.72198380	1.4395869	14374.933
##	sigma_sid[1]	1.621404837	0.57342794	0.88822188	2.6459997	4731.374
##	sigma_sid[2]	0.659058739	0.62805927	0.03962365	1.8287401	5749.896
##	L_Rho_sid[1,1]	1.000000000	0.00000000	1.00000000	1.0000000	NaN
##	L_Rho_sid[1,2]	0.000000000	0.00000000	0.00000000	0.0000000	NaN
##	L_Rho_sid[2,1]	-0.001936964	0.33372487	-0.53620862	0.5380030	14076.456
##	L_Rho_sid[2,2]	0.939499448	0.07730767	0.78549857	0.9996702	4329.513
##	bP[1]	-1.150665657	1.29460826	-3.19523830	0.5955109	9564.878
##	bP[2]	-0.955404812	0.80787821	-2.23963440	0.2865176	12252.271
##	bP[3]	-0.896671158	0.89001770	-2.26516665	0.5080129	10467.681
##	bP[4]	-1.042222982	0.95837704	-2.57519565	0.3985956	9751.798
##	bP[5]	-1.055264904	1.23202517	-2.91185735	0.7117994	9737.563
##	bP[6]	-0.988572624	1.03040089	-2.57387165	0.5573008	9884.912
##	bP[7]	-0.688147867	1.09174104	-2.21805555	1.1006252	8313.455
##	bP[8]	-0.747784559	1.15793385	-2.35319760	1.0687032	8203.859
##	bP[9]	-0.516838955	1.24193269	-2.12125605	1.6121935	6285.792
##	bP[10]	-1.065238608	1.10530393	-2.78919965	0.5314901	9779.706
##	a[1]	-3.322331744	1.22642001	-5.46087965	-1.7082114	9547.034
##	a[2]	0.478285940	0.53969654	-0.35338086	1.3800427	12297.721
##	a[3]	0.553435894	0.46637565	-0.16874549	1.3219033	12143.054
##	a[4]	0.102501321	0.58940133	-0.81825293	1.0626413	11874.579
##	a[5]	-2.647260416	1.29108536	-4.93535810	-0.9683662	9198.918
##	a[6]	-0.905369345	0.79879007	-2.24964860	0.3317886	12565.145

```

## a[7]          0.463634188 0.92330483 -0.97912540 1.9642975 10672.536
## a[8]         -0.193327342 0.95623393 -1.70277165 1.3077560 11482.991
## a[9]          0.439898077 1.04467563 -1.20548275 2.0686177 8635.116
## a[10]         -0.196167550 0.89093668 -1.64701190 1.2158240 13280.297
## Rho_sid[1,1]  1.000000000 0.00000000 1.00000000 1.0000000 NaN
## Rho_sid[1,2] -0.001936964 0.33372487 -0.53620862 0.5380030 14076.456
## Rho_sid[2,1] -0.001936964 0.33372487 -0.53620862 0.5380030 14076.456
## Rho_sid[2,2]  1.000000000 0.00000000 1.00000000 1.0000000 NaN
## ab_sid[1,1]  -2.874067524 1.23309749 -4.97845385 -1.2337064 8000.900
## ab_sid[1,2]  -0.261181339 1.03882541 -1.98891005 0.9202769 8139.012
## ab_sid[2,1]   0.926550247 0.68617632 -0.10575894 2.0547187 6761.132
## ab_sid[2,2]  -0.065920439 0.62229765 -1.11103025 0.8080064 9042.591
## ab_sid[3,1]   1.001700163 0.68442639 -0.05153216 2.1029684 6368.713
## ab_sid[3,2]  -0.007186774 0.66777693 -1.01856935 0.9753703 10406.442
## ab_sid[4,1]   0.550765612 0.73280885 -0.58177873 1.7597441 7195.507
## ab_sid[4,2]  -0.152738645 0.70993754 -1.38853530 0.7410747 9046.928
## ab_sid[5,1]  -2.198996156 1.27711390 -4.46134320 -0.5011197 7855.024
## ab_sid[5,2]  -0.165780510 0.96096568 -1.69709050 0.9941787 8570.812
## ab_sid[6,1]  -0.457105066 0.87067648 -1.88403190 0.8954287 9200.391
## ab_sid[6,2]  -0.099088247 0.77991732 -1.32565025 0.9478919 10856.604
## ab_sid[7,1]   0.911898440 0.93759330 -0.52004717 2.4357948 8736.683
## ab_sid[7,2]   0.201336486 0.81568376 -0.78696891 1.6469500 8503.783
## ab_sid[8,1]   0.254936940 0.97391174 -1.28704465 1.7970104 9482.088
## ab_sid[8,2]   0.141699801 0.84800448 -0.86385854 1.4592503 8121.752
## ab_sid[9,1]   0.888162378 1.03249034 -0.70713162 2.5327493 7999.904
## ab_sid[9,2]   0.372645364 0.96846062 -0.60613709 2.1131399 6499.924
## ab_sid[10,1]  0.252096718 0.93648487 -1.24014530 1.7312969 9554.567
## ab_sid[10,2] -0.175754206 0.86016709 -1.56562500 0.8249945 7986.352
##
## Rhat4
## a_bar         0.9999567
## bP_bar        1.0007505
## delta[1]      1.0002862
## delta[2]      0.9998575
## delta[3]      1.0000244
## delta[4]      0.9998040
## z_sid[1,1]    0.9997372
## z_sid[1,2]    1.0000619
## z_sid[1,3]    0.9999466
## z_sid[1,4]    1.0000051
## z_sid[1,5]    0.9999790
## z_sid[1,6]    0.9999521
## z_sid[1,7]    0.9998447
## z_sid[1,8]    0.9997917
## z_sid[1,9]    0.9998741
## z_sid[1,10]   0.9999204
## z_sid[2,1]    0.9998742
## z_sid[2,2]    1.0001594
## z_sid[2,3]    0.9999579
## z_sid[2,4]    0.9998734
## z_sid[2,5]    0.9999513
## z_sid[2,6]    0.9997571
## z_sid[2,7]    1.0000343
## z_sid[2,8]    0.9997416
## z_sid[2,9]    0.9998339

```



```

## z_sid[2,10]      1.0002454
## sigma_sid[1]     1.0003029
## sigma_sid[2]     1.0001184
## L_Rho_sid[1,1]    NaN
## L_Rho_sid[1,2]    NaN
## L_Rho_sid[2,1]    1.0003038
## L_Rho_sid[2,2]    1.0003288
## bP[1]            1.0009580
## bP[2]            0.9999730
## bP[3]            1.0002129
## bP[4]            1.0002224
## bP[5]            1.0006391
## bP[6]            1.0011100
## bP[7]            1.0009017
## bP[8]            1.0003042
## bP[9]            1.0002106
## bP[10]           0.9999089
## a[1]             0.9998172
## a[2]             1.0002256
## a[3]             1.0001210
## a[4]             0.9998188
## a[5]             0.9999735
## a[6]             1.0004143
## a[7]             1.0004496
## a[8]             0.9999813
## a[9]             1.0002249
## a[10]            0.9998948
## Rho_sid[1,1]     NaN
## Rho_sid[1,2]     1.0003038
## Rho_sid[2,1]     1.0003038
## Rho_sid[2,2]     NaN
## ab_sid[1,1]      1.0000385
## ab_sid[1,2]      1.0010492
## ab_sid[2,1]      0.9997134
## ab_sid[2,2]      1.0000161
## ab_sid[3,1]      0.9999292
## ab_sid[3,2]      0.9999158
## ab_sid[4,1]      0.9998295
## ab_sid[4,2]      0.9998102
## ab_sid[5,1]      1.0001105
## ab_sid[5,2]      1.0004355
## ab_sid[6,1]      1.0000184
## ab_sid[6,2]      0.9999682
## ab_sid[7,1]      1.0000505
## ab_sid[7,2]      1.0004745
## ab_sid[8,1]      0.9997837
## ab_sid[8,2]      0.9999249
## ab_sid[9,1]      0.9999622
## ab_sid[9,2]      1.0004938
## ab_sid[10,1]     0.9997966
## ab_sid[10,2]     0.9998074

```

All Rhat4 values are 1.

Each parameter also sampled well.

PSIS/WAIC

```
PSIS(cvs_mod)
```

```
## Some Pareto k values are high (>0.5). Set pointwise=TRUE to inspect individual points.
```

```
##      PSIS      lppd  penalty  std_err  
## 1 161.6147 -80.80735 9.814636 9.838208
```

```
WAIC(cvs_mod)
```

```
##      WAIC      lppd  penalty  std_err  
## 1 161.2069 -70.99271 9.610717 9.756552
```

The are some Pareto k values > 0.5. As long as < 0.7, not an issue. Are they?

```
PSIS(cvs_mod, pointwise = TRUE) %>%  
  high_k_rows()
```

```
## Some Pareto k values are high (>0.5). Set pointwise=TRUE to inspect individual points.
```

```
## # A tibble: 1 x 15  
##   videoid PSIS  lppd penalty std_err      k surgid  pgs time_until_1st_c~  
##   <int> <dbl> <dbl>   <dbl>   <dbl> <dbl> <int> <int>          <dbl>  
## 1     98  3.07 -1.54   0.345    9.84 0.506      4     5          39.5  
## # ... with 6 more variables: time_cvs_attained <dbl>,  
## #   laparoscopic_duration <dbl>, dissection_duration <dbl>,  
## #   gb_removal_duration <dbl>, gb_hole <lgl>, gb_holes <int>
```

Only 1 row with a k of 0.506, so minimal issue with outliers.

Trace rank plot (trankplot)

```
trankplot(cvs_mod)
```

Good mixing as shown with large amount of overlap.

Trace plot

```
traceplot(cvs_mod)
```

Chains are stationary with a visible central tendency, have good mixing, and converge.

Evaluate model results

First, obtain tidy samples, as we did with the duration and GB hole model. Additionally, transform the result to get the absolute probability of CVS and the OR of a obtained CVS for each PGS:

```
tidy_cvs_samples <- extract_samples(  
  cvs_mod,  
  pars = c(  
    "a_bar",  
    paste0("a[", 1:nsurgs, "]"),  
    "bP_bar",  
    paste0("bP[", 1:nsurgs, "]"),  
    # sigma_a  
    "sigma_sid[1]",  
    # sigma_b
```

```

      "sigma_sid[2]",
      paste0("Rho_sid[1,2]"),
      paste0("delta[", 1:4, "]")
    )
  ) %>%
  tidy_surgeons() %>%
  sum_deltas() %>%
  tidy_pgs() %>%
  mutate(
    p = inv_logit(a + bP * sum_delta_j),
    bOR = exp(bP * sum_delta_j)
  ) %>%
  arrange(sample_num, surgeon, PGS) %>%
  group_by(surgeon, sample_num) %>%
  # calc change in probability for each sample
  # group by surgeon as well because each sample_num
  # has the info for all 10 surgeons and bar surgeon
  mutate(
    # change from one PGS level to the other
    incr_delta_p = p - lag(p),
    # change from PGS1 to that PGS level
    delta_p = p - p[1]
  ) %>%
  ungroup()

```

Key to know, is that:

`incr_delta_p` is the probability difference from one level to the next. It will be NA for a PGS1.

`delta_p` is probability difference from a certain PGS level to PGS1.

`p` is the probability of a cvs at that PGS level

`bOR` is the increased odds, compared to a PGS of 1, of obtaining CVS

Plot results

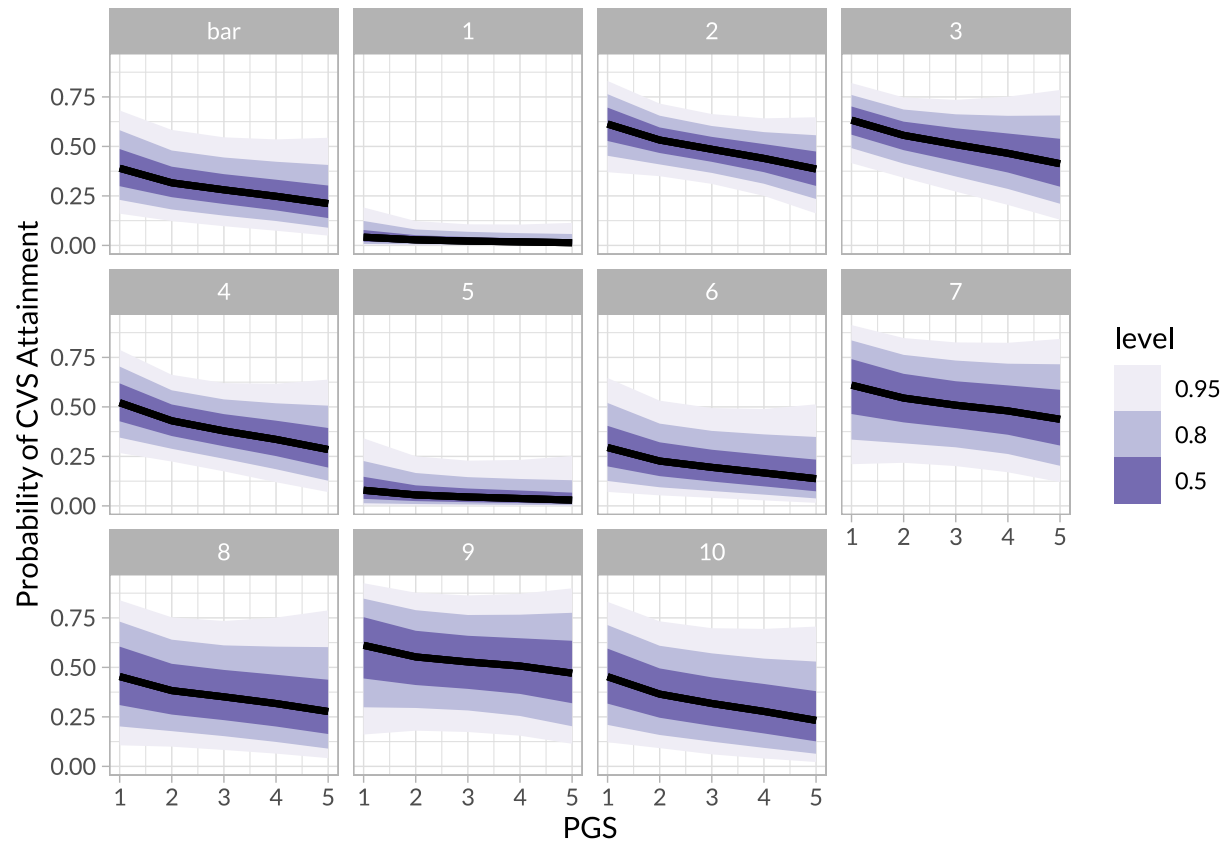
For all surgeons

Remember that “bar” is the average of all the surgeons:

```

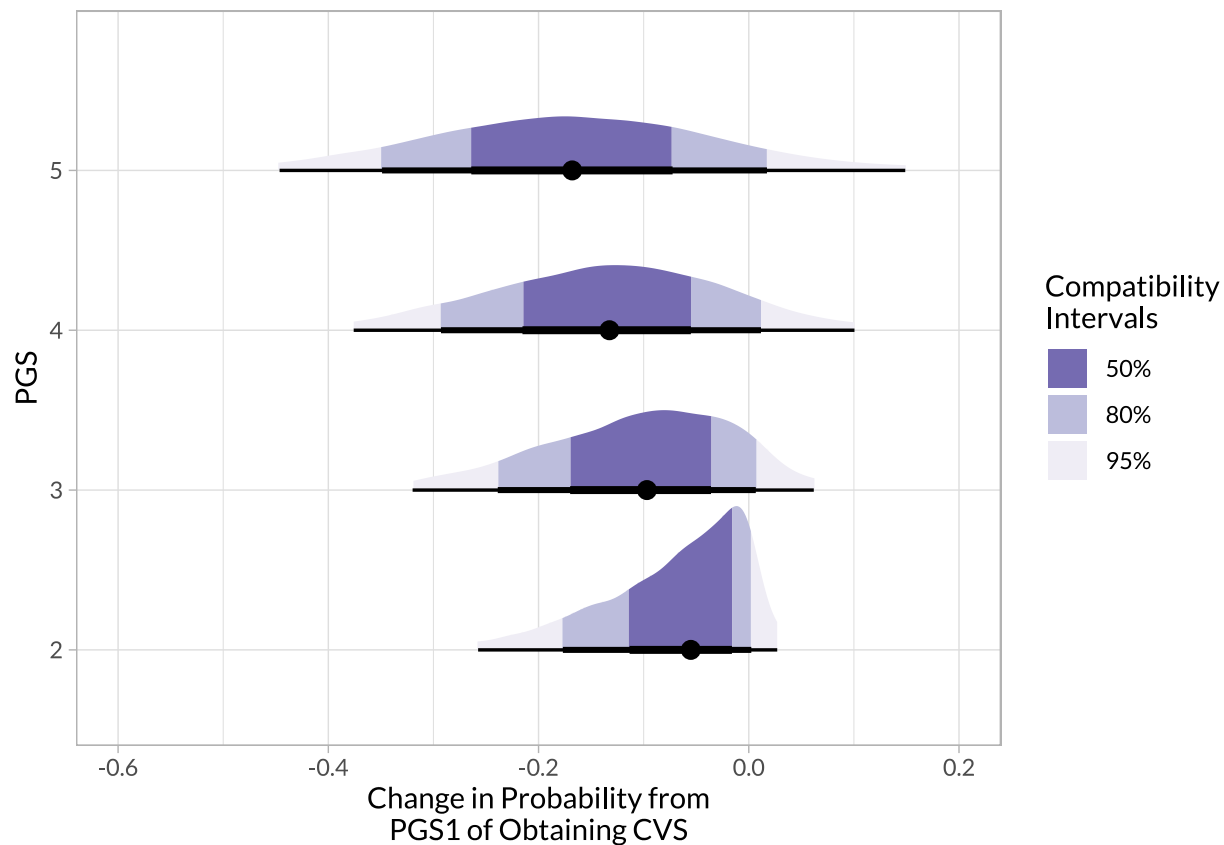
tidy_cvs_samples %>%
  mutate(surgeon = parse_factor(surgeon, levels = c("bar", as.character(1:nsurgs)))) %>%
  ggplot(aes(PGS, p)) +
  stat_lineribbon() +
  facet_wrap(~ surgeon) +
  scale_fill_brewer(palette = "Purples") +
  labs(y = "Probability of CVS Attainment")

```



On average across all surgeons

```
bar_cvs <- tidy_cvs_samples %>%
  filter(surgeon == "bar", PGS != 1) %>%
  mutate(PGS = as.factor(PGS))
bar_cvs %>%
  halfeye(delta_p, PGS) +
  coord_cartesian(xlim = c(-0.6, 0.2)) +
  labs(
    y = "PGS",
    x = "Change in Probability from\nPGS1 of Obtaining CVS"
  )
```



And numbers for this:

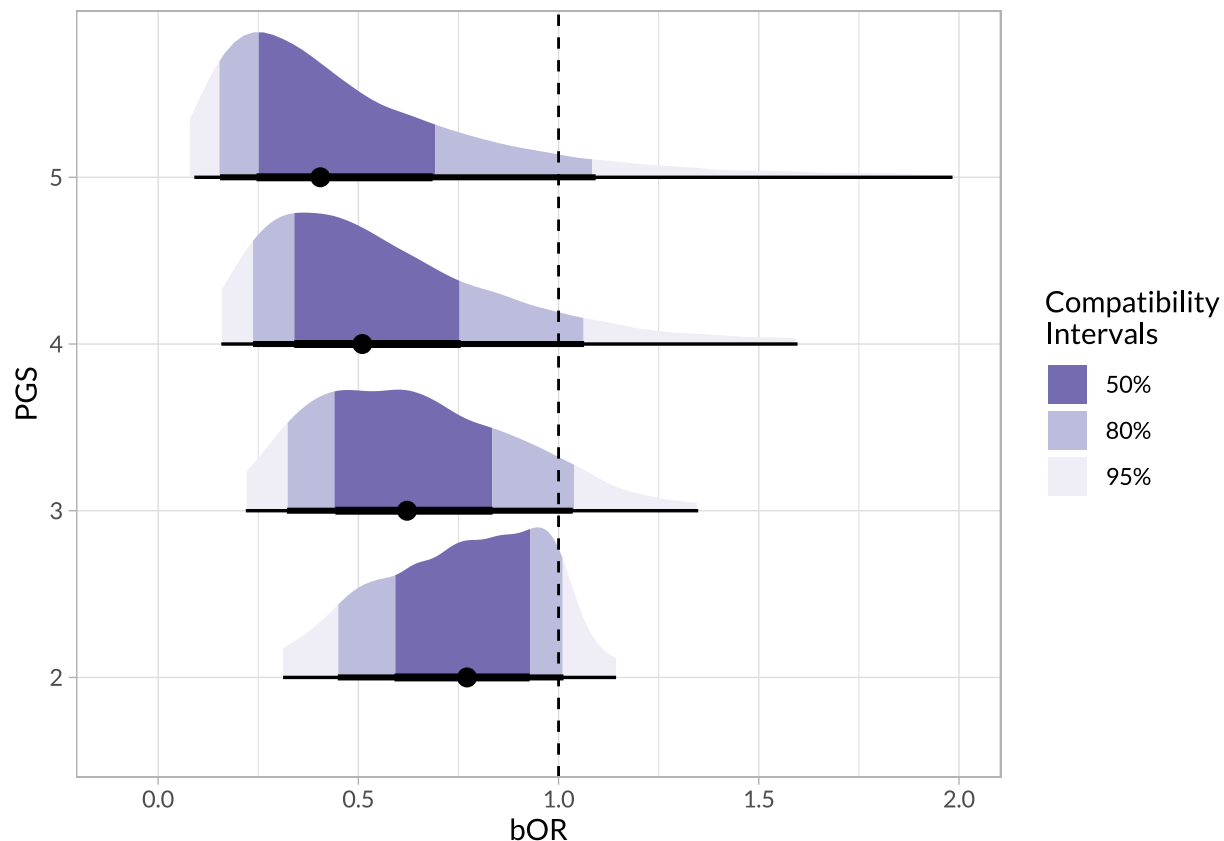
```
bar_cvs %>%
  group_by(PGS) %>%
  ci_ints(delta_p)
```

A tibble: 4 x 7

##	PGS	mean	int_50	int_66	int_80	int_89	int_95
##	<fct>	<dbl>	<chr>	<chr>	<chr>	<chr>	<chr>
## 1	2	-0.07	-0.11, -0.02	-0.14, -0.01	-0.13, -0.01	-0.21, 0.~	-0.26, 0.~
## 2	3	-0.11	-0.17, -0.04	-0.2, -0.01	-0.19, -0.02	-0.28, 0.~	-0.32, 0.~
## 3	4	-0.14	-0.22, -0.06	-0.25, -0.02	-0.24, -0.04	-0.33, 0.~	-0.38, 0.1
## 4	5	-0.17	-0.26, -0.07	-0.3, -0.03	-0.29, -0.05	-0.39, 0.~	-0.45, 0.~

And looking at the odd ratios:

```
bar_cvs %>%
  halfeye(bOR, PGS) +
  coord_cartesian(xlim = c(-0.1, 2)) +
  geom_vline(xintercept = 1, linetype = 2)
```



Numbers for this:

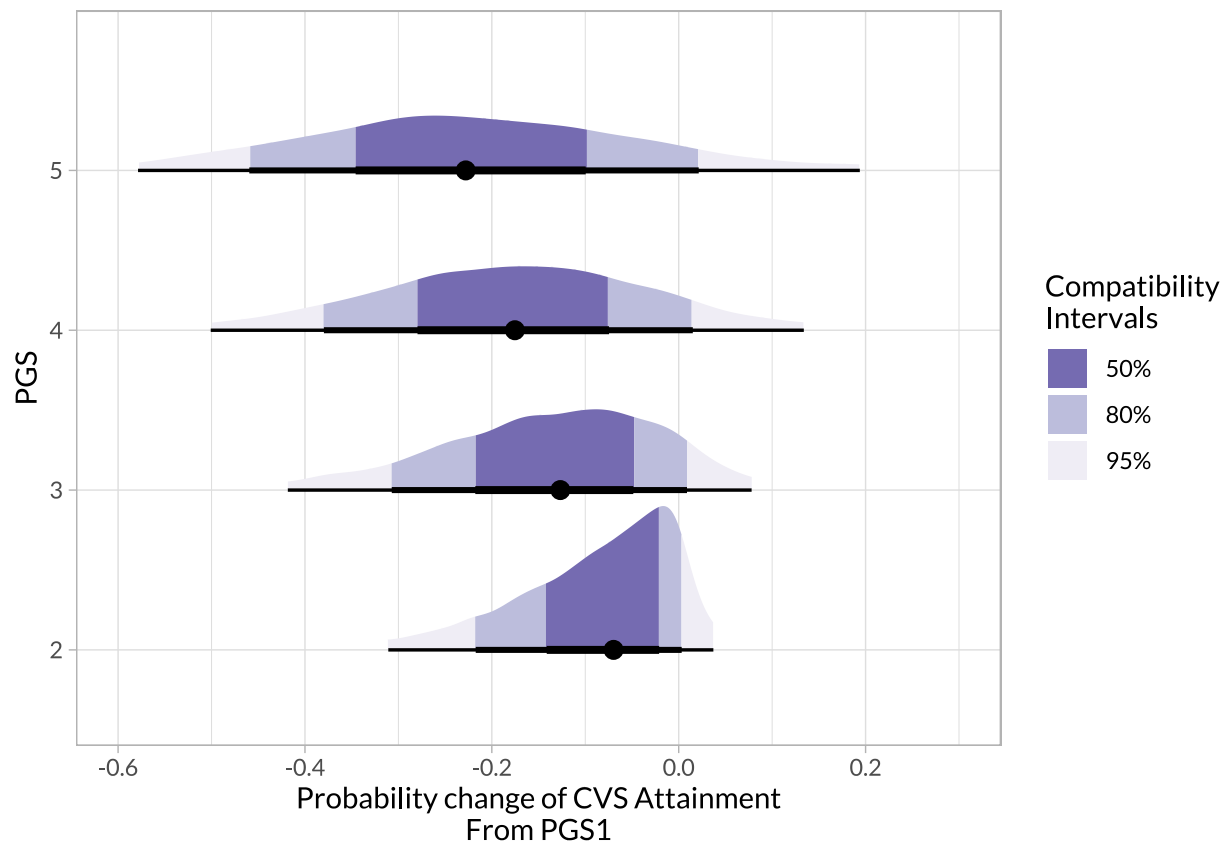
```
bar_cvs %>%
  group_by(PGS) %>%
  ci_ints(bOR)
```

```
## # A tibble: 4 x 7
##   PGS   mean int_50      int_66      int_80      int_89      int_95
##   <fct> <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1 2      0.75 0.59, 0.93 0.52, 0.97 0.55, 0.95 0.38, 1.06 0.31, 1.14
## 2 3      0.66 0.44, 0.84 0.38, 0.93 0.41, 0.89 0.27, 1.15 0.22, 1.35
## 3 4      0.61 0.34, 0.76 0.29, 0.88 0.31, 0.83 0.2, 1.28 0.16, 1.6
## 4 5      0.570 0.25, 0.69 0.2, 0.85 0.22, 0.78 0.12, 1.42 0.09, 1.98
```

For a surgeon particularly affected by PGS

```
surg4_cvs <- tidy_cvs_samples %>%
  filter(surgeon == "4", PGS != 1) %>%
  mutate(PGS = as.factor(PGS))

surg4_cvs %>%
  halfeye(delta_p, PGS) +
  coord_cartesian(xlim = c(-0.6, 0.3)) +
  labs(
    y = "PGS",
    x = "Probability change of CVS Attainment\nFrom PGS1"
  )
```



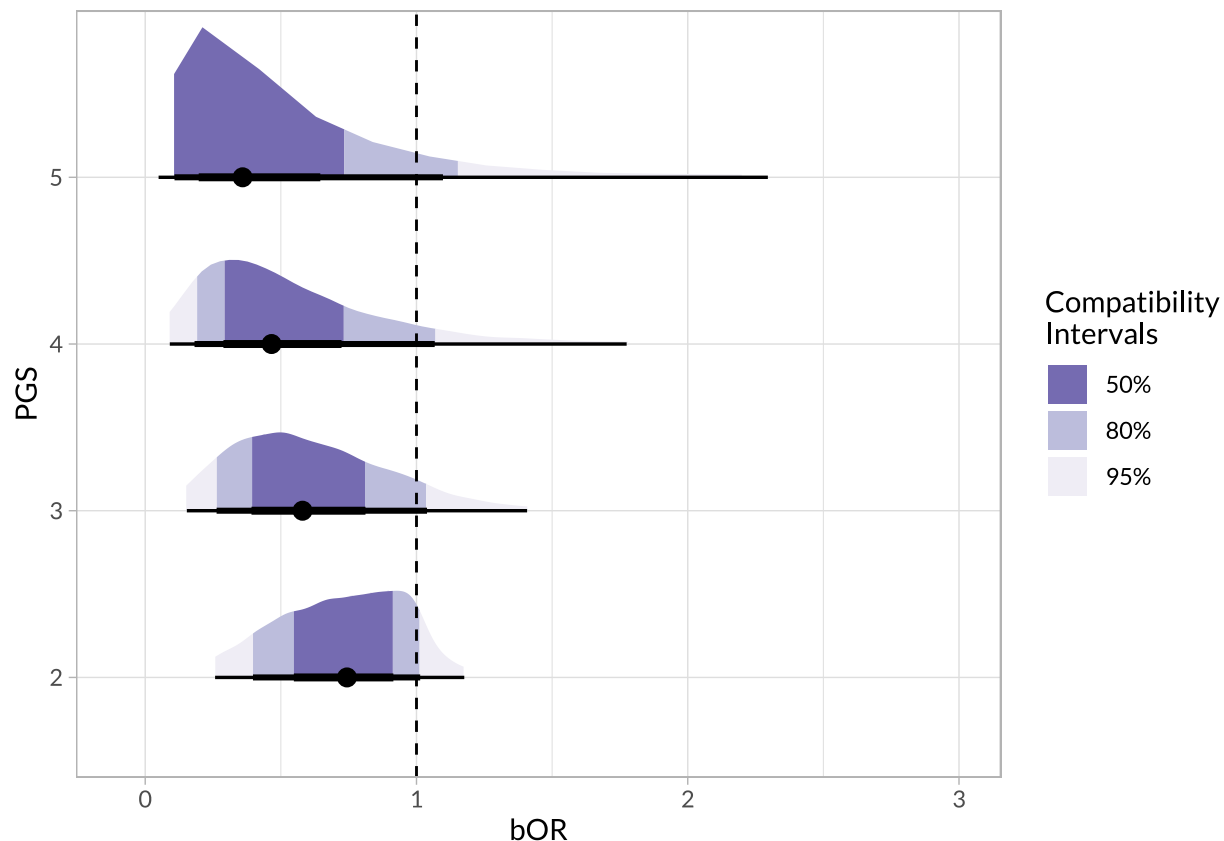
And numbers for this:

```
surg4_cvs %>%
  group_by(PGS) %>%
  ci_ints(delta_p)
```

```
## # A tibble: 4 x 7
##   PGS   mean int_50      int_66      int_80      int_89      int_95
##   <fct> <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1 2     -0.09 -0.14, -0.02 -0.17, -0.01 -0.16, -0.01 -0.26, 0.~ -0.31, 0.~
## 2 3     -0.14 -0.22, -0.05 -0.26, -0.02 -0.24, -0.03 -0.36, 0.~ -0.42, 0.~
## 3 4     -0.18 -0.28, -0.07 -0.33, -0.03 -0.31, -0.05 -0.43, 0.~ -0.5, 0.13
## 4 5     -0.22 -0.35, -0.1 -0.4, -0.04 -0.38, -0.07 -0.51, 0.~ -0.58, 0.~
```

And looking at odds ratios:

```
surg4_cvs %>%
  halfeye(bOR, PGS) +
  coord_cartesian(xlim = c(-0.1, 3)) +
  geom_vline(xintercept = 1, linetype = 2)
```



Numbers for this:

```
surg4_cvs %>%
  group_by(PGS) %>%
  ci_ints(bOR)
```

```
## # A tibble: 4 x 7
##   PGS   mean int_50      int_66      int_80      int_89      int_95
##   <fct> <dbl> <chr>      <chr>      <chr>      <chr>      <chr>
## 1 2      0.73 0.55, 0.91 0.48, 0.97 0.51, 0.95 0.32, 1.07 0.26, 1.18
## 2 3      0.63 0.39, 0.81 0.33, 0.92 0.35, 0.88 0.21, 1.18 0.15, 1.41
## 3 4      0.580 0.29, 0.72 0.23, 0.87 0.25, 0.81 0.14, 1.34 0.09, 1.77
## 4 5      0.580 0.2, 0.65 0.15, 0.83 0.17, 0.75 0.08, 1.49 0.05, 2.3
```

Average and surgeon most affected by PGS

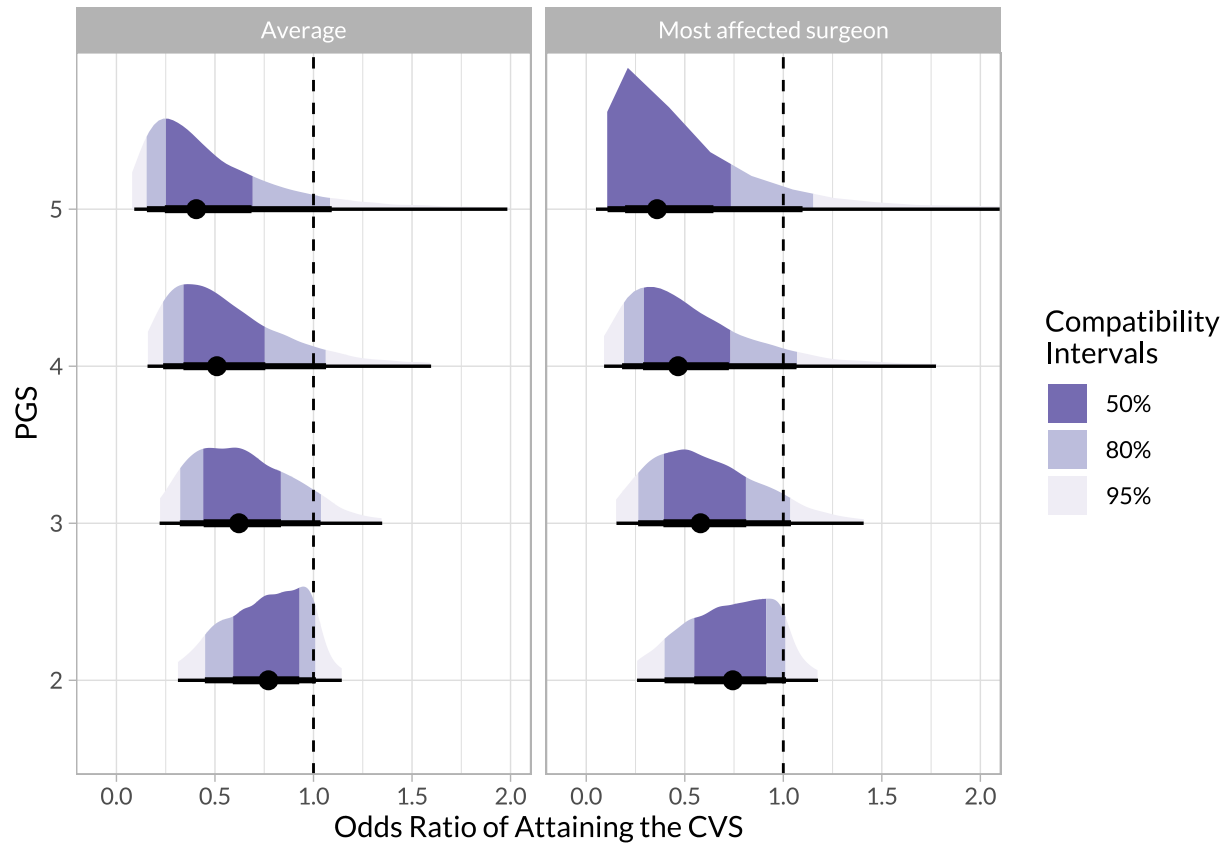
```
tidy_cvs_samples %>%
  filter(surgeon %in% c("bar", "4"), PGS != 1) %>%
  mutate(
    surgeon = if_else(surgeon == "bar", "Average", "Most affected surgeon"),
    PGS = as.factor(PGS)
  ) %>%
  halfeye(bOR, PGS) +
  facet_wrap(~ surgeon) +
  coord_cartesian(xlim = c(-0.1, 2)) +
  geom_vline(xintercept = 1, linetype = 2) +
  labs(
```



```

x = "Odds Ratio of Attaining the CVS",
y = "PGS"
)

```



```

ggsave("../output/pgs_cvs.svg", width = 6, height = 4)
ggsave("../output/pgs_cvs.pdf", width = 6, height = 4)

```

Correlation of PGS1 and effect of incremental PGS on CVS

```

extract_samples(cvs_mod, pars = c("Rho_sid[1,2]")) %>%
  rename(rho = Rho_sid_1_2) %>%
  ci_ints(rho) %>%
  knitr::kable(
    caption = "Correlation between attaining CVS in PGS1 case and the effect of incrementing PGS"
  )

```

Table 9: Correlation between attaining CVS in PGS1 case and the effect of incrementing PGS

mean	int_50	int_66	int_80	int_89	int_95
0	-0.24, 0.24	-0.34, 0.34	-0.3, 0.3	-0.54, 0.54	-0.63, 0.62

Summarise inputs/outcomes

Numbers

Number of surgeons:

```
nsurgs
```

```
## [1] 10
```

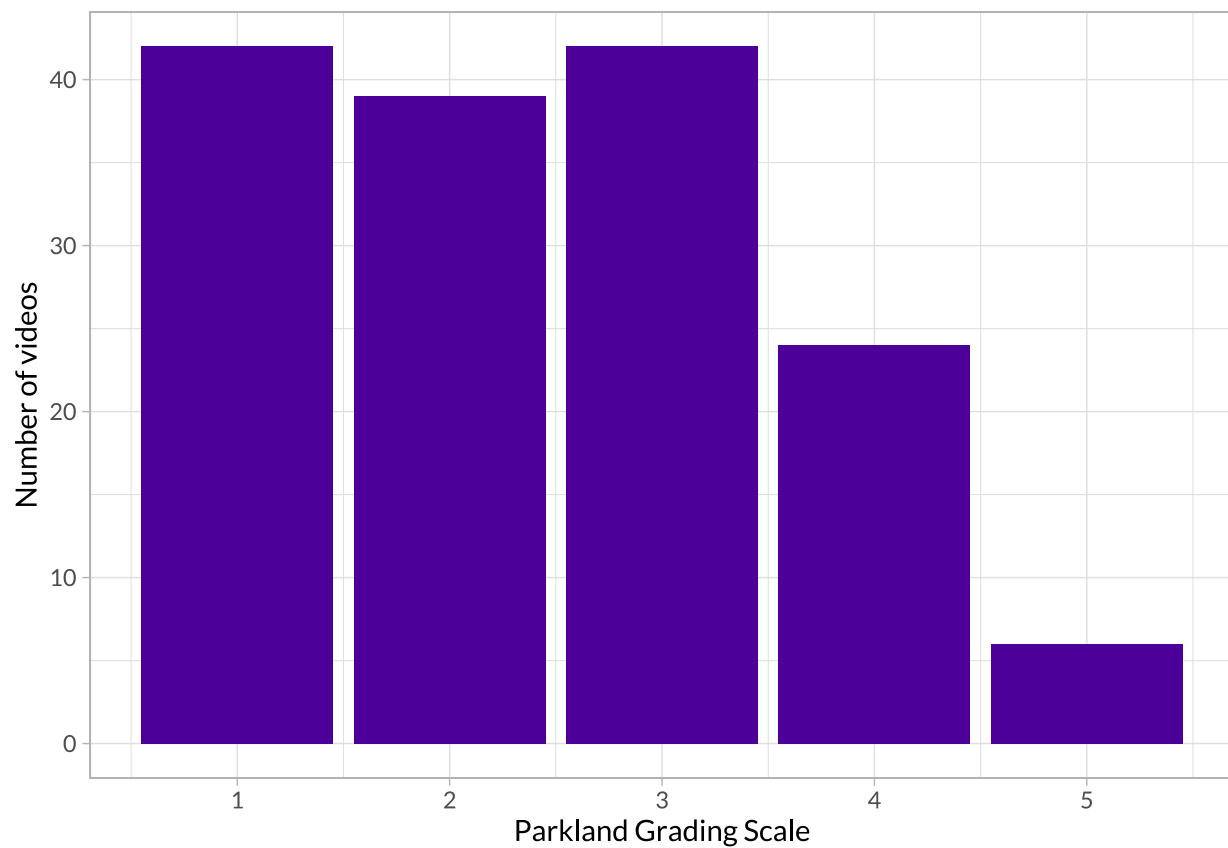
Number of videos with surgeons ≥ 5 cases:

```
nrow(dat)
```

```
## [1] 153
```

PGS

```
dat %>%  
  ggplot(aes(pgs)) +  
  geom_bar(fill = histo_color) +  
  labs(  
    x = "Parkland Grading Scale",  
    y = "Number of videos"  
  )
```



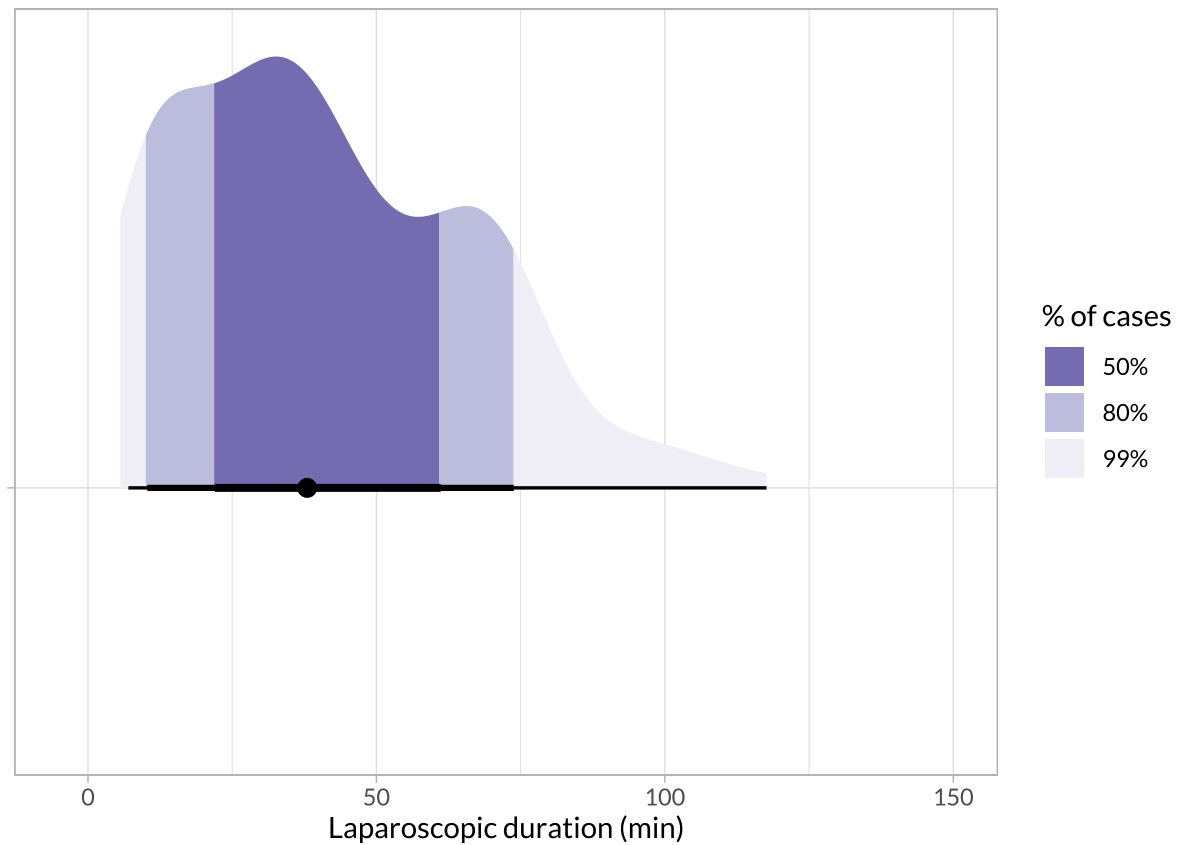
```
ggsave("../output/pgs_distribution.svg", width = 6, height = 6)  
ggsave("../output/pgs_distribution.pdf", width = 6, height = 6)
```

```
dat %>%
  count(pgs) %>%
  mutate(prop = n / sum(n)) %>%
  mutate(cum.sum = cumsum(prop))
```

```
## # A tibble: 5 x 4
##   pgs      n  prop cum.sum
##   <int> <int> <dbl> <dbl>
## 1     1    42 0.275  0.275
## 2     2    39 0.255  0.529
## 3     3    42 0.275  0.804
## 4     4    24 0.157  0.961
## 5     5     6 0.0392  1
```

Laparoscopic duration

```
dat %>%
  ggplot(aes(y = "", x = laparoscopic_duration)) +
  stat_halfeye(
    aes(
      fill = stat(
        cut_cdf_qi(cdf, .width = c(0.5, 0.8, 0.99), labels = scales::percent_format())
      )
    ),
    .width = c(0.5, 0.8, 0.99)
  ) +
  scale_fill_brewer(direction = -1, palette = "Purples", na.translate = FALSE) +
  labs(fill = "% of cases", y = "", x = "Laparoscopic duration (min)") +
  coord_cartesian(xlim = c(-5, 150))
```



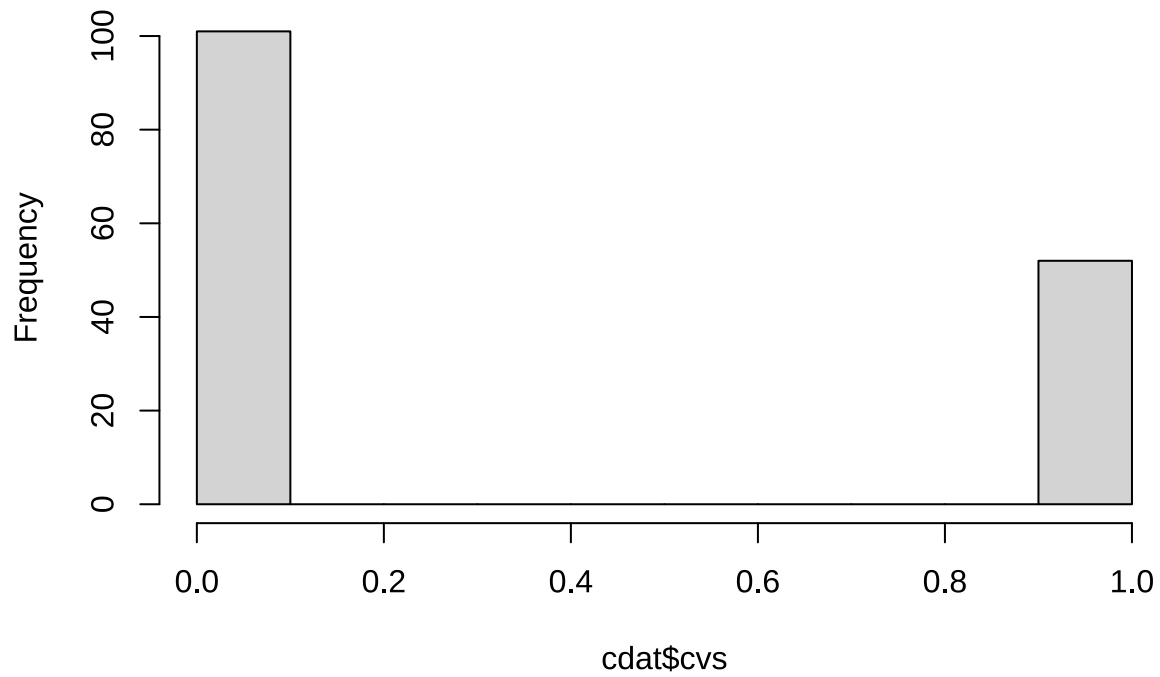
```
summary(dat$laparoscopic_duration)
```

```
##      Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
##    5.617  22.017   38.000   41.296  61.133  131.600
```

CVS number of cases with attainment

```
hist(cdat$cvs)
```

Histogram of cdat\$cvcs



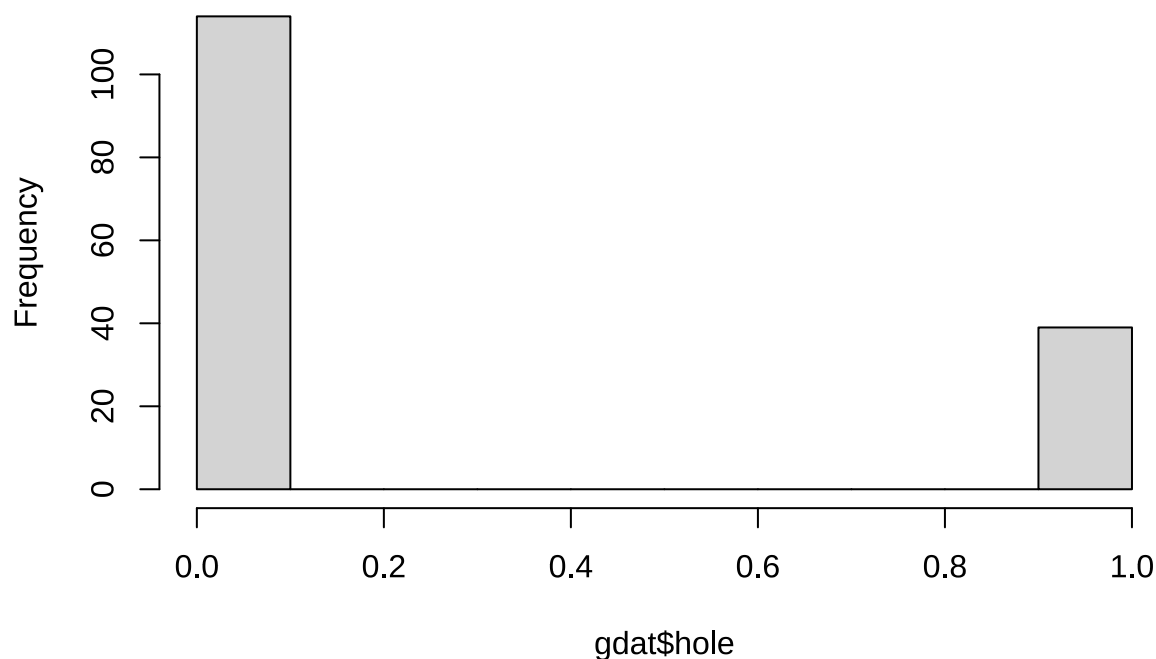
```
mean(cdat$cvcs)
```

```
## [1] 0.3398693
```

GB holes

```
hist(gdat$hole)
```

Histogram of gdat\$hole



```
mean(gdat$hole)
```

```
## [1] 0.254902
```

Environment

```
devtools::session_info()
```

```
## - Session info -----
## setting  value
## version  R version 4.0.5 (2021-03-31)
## os       Fedora 34 (Workstation Edition)
## system   x86_64, linux-gnu
## ui       X11
## language (EN)
## collate  en_US.UTF-8
## ctype    en_US.UTF-8
## tz       America/New_York
## date     2021-08-29
##
## - Packages -----
## package      * version  date      lib
## abind         1.4-5    2016-07-21 [1]
## assertthat    0.2.1    2019-03-21 [1]
## backports     1.2.1    2020-12-09 [1]
## base64enc     0.1-3    2015-07-28 [1]
```

## blob	1.2.1	2020-01-20	[1]
## callr	3.6.0	2021-03-28	[1]
## checkmate	2.0.0	2020-02-06	[1]
## cli	2.3.1	2021-02-23	[1]
## cmdstanr	* 0.3.0	2021-03-29	[1]
## coda	0.19-4	2020-09-30	[1]
## codetools	0.2-18	2020-11-04	[2]
## colorspace	2.0-0	2020-11-11	[1]
## crayon	1.4.1	2021-02-08	[1]
## curl	4.3	2019-12-02	[1]
## data.table	1.13.2	2020-10-19	[1]
## DBI	1.1.0	2019-12-15	[1]
## desc	1.3.0	2021-03-05	[1]
## devtools	2.3.2	2020-09-18	[1]
## digest	* 0.6.27	2020-10-24	[1]
## distributional	0.2.2	2021-02-02	[1]
## dplyr	* 1.0.5	2021-03-05	[1]
## ellipsis	0.3.1	2020-05-15	[1]
## evaluate	0.14	2019-05-28	[1]
## fansi	0.4.2	2021-01-15	[1]
## farver	2.1.0	2021-02-28	[1]
## forcats	* 0.5.1	2021-01-27	[1]
## fs	1.5.0	2020-07-31	[1]
## gdtools	* 0.2.3	2021-01-06	[1]
## generics	0.1.0	2020-10-31	[1]
## ggdist	* 2.4.0	2021-01-04	[1]
## ggplot2	* 3.3.3	2020-12-30	[1]
## glue	1.4.2	2020-08-27	[1]
## gridExtra	2.3	2017-09-09	[1]
## gtable	0.3.0	2019-03-25	[1]
## highr	0.8	2019-03-20	[1]
## hms	0.5.3	2020-01-08	[1]
## htmltools	0.5.1.1	2021-01-22	[1]
## inline	0.3.17	2020-12-01	[1]
## janitor	2.0.1	2020-04-12	[1]
## jsonlite	1.7.2	2020-12-09	[1]
## knitr	1.30	2020-09-22	[1]
## labeling	0.4.2	2020-10-20	[1]
## lattice	0.20-41	2020-04-02	[2]
## lifecycle	1.0.0	2021-02-15	[1]
## loo	2.4.1	2020-12-09	[1]
## lubridate	1.7.9	2020-06-08	[1]
## magrittr	* 2.0.1	2020-11-17	[1]
## MASS	7.3-53.1	2021-02-12	[2]
## matrixStats	0.58.0	2021-01-29	[1]
## memoise	1.1.0	2017-04-21	[1]
## munsell	0.5.0	2018-06-12	[1]
## mvtnorm	1.1-1	2020-06-09	[1]
## nvimcom	* 0.9-102	2021-05-17	[1]
## pillar	1.5.1	2021-03-05	[1]
## pkgbuild	1.2.0	2020-12-15	[1]
## pkgconfig	2.0.3	2019-09-22	[1]
## pkgload	1.1.0	2020-05-29	[1]
## posterior	0.1.4	2021-03-29	[1]

```

## prettyunits      1.1.1      2020-01-24 [1]
## processx         3.5.0      2021-03-23 [1]
## ps               1.6.0      2021-02-28 [1]
## purrr            0.3.4      2020-04-17 [1]
## R6                2.5.0      2020-10-28 [1]
## RColorBrewer      1.1-2      2014-12-07 [1]
## Rcpp              1.0.6      2021-01-15 [1]
## RcppParallel      5.0.3      2021-02-24 [1]
## readr             * 1.4.0      2020-10-05 [1]
## remotes           2.2.0      2020-07-21 [1]
## repr              1.1.0      2020-01-28 [1]
## rethinking        * 2.13      2020-10-24 [1]
## rlang              0.4.10     2020-12-30 [1]
## rmarkdown         * 2.5       2020-10-21 [1]
## rprojroot         2.0.2      2020-11-15 [1]
## rstan             * 2.21.2     2020-07-27 [1]
## rstudioapi        0.11      2020-02-07 [1]
## scales            1.1.1      2020-05-11 [1]
## sessioninfo       1.1.1      2018-11-05 [1]
## shape             1.4.5      2020-09-13 [1]
## showtext          * 0.9-2     2021-01-10 [1]
## showtextdb        * 3.0       2020-06-04 [1]
## skimr             2.1.2      2020-07-06 [1]
## snakecase         0.11.0     2019-05-25 [1]
## StanHeaders       * 2.21.0-7   2020-12-17 [1]
## stringi           1.5.3      2020-09-09 [1]
## stringr           * 1.4.0      2019-02-10 [1]
## svglite           1.2.3.2    2020-07-07 [1]
## sysfonts          * 0.8.3      2021-01-10 [1]
## systemfonts       0.3.2      2020-09-29 [1]
## tensorA           0.36.2     2020-11-19 [1]
## testthat          3.0.3      2021-06-16 [1]
## tibble            3.1.0      2021-02-25 [1]
## tidyr             * 1.1.3      2021-03-03 [1]
## tidyselect        1.1.0      2020-05-11 [1]
## usethis           1.6.3      2020-09-17 [1]
## utf8              1.2.1      2021-03-12 [1]
## V8                3.4.0      2020-11-04 [1]
## vctrs             0.3.7      2021-03-29 [1]
## withr             2.4.1      2021-01-26 [1]
## xfun              0.18       2020-09-29 [1]
## yaml              2.2.1      2020-02-01 [1]
## source
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## local
## CRAN (R 4.0.3)
## CRAN (R 4.0.5)

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##
## [1] /home/thomas/R/x86_64-redhat-linux-gnu-library/4.0
## [2] /usr/lib64/R/library
## [3] /usr/share/R/library

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