

Johor Straits Sampling Checklist

Date: _____

Item or Sample Type	Preparations in the Lab – qty is per trip	What to do in the Field	Back in the Lab
Autosampler	<ul style="list-style-type: none"> - 10x sterivex (4 per site, +2 extra) - 4x 10ml syringes - RNA Later - Blue tack - Toilet roll 	<ul style="list-style-type: none"> - Run autosampler for 10 minutes with local water - Collect each sterivex sample for 15 minutes, then remove water by pushing air using syringe - Fill each sterivex sample with 2ml RNA Later using pipette - Put sterivex samples in Dryshipper. 	<p>Store sterivex in -80C</p> <p>Rinse Autosampler with tap water for 10 minutes</p> <p>[Once a week, rinse the autosampler with 5L 10% bleach, then 10 mins with tap water.]</p>
Nutrients	<ul style="list-style-type: none"> - 5x 15ml tubes, acid washed (1x acid, 3x RO water), dry (2 per site, +1 extra) 	<ul style="list-style-type: none"> - Use mid-stream sterivex filtrate. - Rinse each tube 3 times with sample before collection - Place in Dryshipper 	Store tubes in -20C.
FCM Samples	<ul style="list-style-type: none"> - 10 cryovials (4 per site, +2 extra) - Pre-label with sample ID, location, date, sample type - Pre-aliquot 20ul of 25% glutaraldehyde into each vial - Store in 4C fridge until the morning of sampling - Bring for sampling inside cooler + ice pack 	<ul style="list-style-type: none"> - Add 980ul of water sample inside each vial, accordingly to each sample type - Fix samples in the cooler for 15-30 minutes - Flash freeze in Dryshipper - For each sample, do 2 for 150um pre-filtrate & 2 for 0.22um filtrate (total 4 per site). 	Store cryovials in -80C.
Chlorophyll GF/F	<ul style="list-style-type: none"> - 4x GF/F (2 per site) - 8x 50ml syringe (4 per site) - filter holder - Forceps - Aluminium foil <p>For each site, do 1 sample for completely unfiltered water, 1 for 150um pre-filtrate</p>	<ul style="list-style-type: none"> - Place GF/F in filter holder using forceps - Gently, filter 50ml of sample through the GFF, then rinse filter with 50ml of DI water - Push air out of syringe to dry filter - Fold in 2 (sample side in), place in aluminium foil - Put in Dryshipper - Rinse filter holder with DI water. 	Store GFF at -20C.
Fluorescence	<ul style="list-style-type: none"> - Aquaflash + glass cuvette - Kimwipes - 6x 50ml tubes (3 per site, 1 each for completely unfiltered, 150um pre-filtered, 0.22um filtered) 	<ul style="list-style-type: none"> - Aliquot 2ml of sample into glass cuvette - Measure & record. - Do for completely unfiltered, 150um pre-filtered & 0.22um filtered sample 	
Secchi disk	<ul style="list-style-type: none"> - Bring Secchi disk 	<ul style="list-style-type: none"> - Lower disk until it just disappears. Record depth. - Pull disk until it just appears. Record depth. - Repeat 5 times 	<ul style="list-style-type: none"> - Average the 10 numbers, that is the Secchi Depth
Cooler & Dryshipper & other consumables	<ul style="list-style-type: none"> - Put ice packs in -80 - Prepare cooler - Prepare Dryshipper - Prepare sampling materials - At least 1l of DI Water - 1000ul pipette & tips - Gloves, 70% EtOH, tissue - Salinity & temperature probe - Sampling notebook, pen, sharpies 	<ul style="list-style-type: none"> - Record date, sampling location, sampling time, weather, sampling personnel - Record measurements: <ul style="list-style-type: none"> o Fluorescence & yield o Secchi depth o Salinity & Temperature o Number of samples of each kind taken - Record any other observations 	<ul style="list-style-type: none"> - Thaw/re-freeze ice packs - Clean cooler - Remove all samples to proper storage places - Dispose remaining DI Water - Fully charge the Dryshipper when not cold

Completely unfiltered = from bypass filter, turn red knob

150um pre-filtered = from the outhouse, the one that spews water very far

0.22um filtered = from sterivex