

Kevin M. Dean, Ph.D.

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PROFESSIONAL ACCOMPLISHMENTS

My research portfolio includes 39 peer-reviewed manuscripts that have been cited 2100 times (h-index of 23). I am a MPI on an NCI Cellular Cancer Biology Imaging Research Center (U54CA268072) grant, an NIGMS Biomedical Technology Development and Dissemination Center (RM1GM145399) grant, and a Harold C. Simmons Comprehensive Cancer Center Translational Research Pilot grant. My lab is highly collaborative, and we also receive funding as a co-investigator, collaborator, or as key personnel, on 5 grants, including a P30, R01, R21, R34, and S10. I am listed as an inventor on 3 patents, including one that is licensed to Intelligent Imaging Innovations, Inc (3i).

VISION

As a scientist, I am driven to develop and apply cutting-edge imaging technologies that provide molecular insight into incredibly complex biological processes and systems. My scientific training spans multiple disciplines, including biology, biochemistry, computer vision, fluorescence microscopy, and optical probes (including optogenetics, chemogenetics, and biosensors), and I have a strong history of leading and completing highly-technical, interdisciplinary, and collaborative research projects.

RESEARCH AND WORK ACTIVITIES

Tenure-Track Assistant Professor

2022 – Present

Lyda Hill Department of Bioinformatics and Cecil H. and Ida Green Center for Systems Biology.
UT Southwestern Medical Center, Dallas, TX, USA.

Advanced fluorescence imaging of tissues, single cell sequencing, and other single-cell approaches have provided incredible insight into the earliest events in the metastatic cascade, including the growth of the primary tumor and invasion of single cells or cell clusters into adjacent tissues. In contrast, the molecular and cellular processes that enable colonization of a remote tissue are poorly understood and data are scarce. The overarching goal of the Dean Lab is to identify the molecular mechanisms that enable cancer cells to colonize and populate a distant tissue. To achieve this, we are developing a series of self-driving microscopes that leverage cutting-edge computer vision routines and adaptively image diverse specimens, and advancing integrated multiplexing imaging approaches using a combination of hydrogel-based tissue clearing and directed evolution of labeling technologies. Member of the Biomedical Engineering and Molecular Biophysics Graduate Programs.

Research-Track Assistant Professor, and Director of the Microscopy Innovation Lab

2018 – 2022

Department of Cell Biology.

UT Southwestern Medical Center, Dallas, TX, USA.

The pace of microscopy technology development far outstrips the rate at which it can be commercialized and delivered to the greater biological community. The Microscopy Innovation Lab at UT Southwestern works to eliminate this delay by developing potentially transformative instrumentation that is tailored to address specific biological questions, and engaging in close collaborations with life scientists to gain molecular insight into physiological and pathophysiological phenomena. Towards these means, I work one-on-one with diverse biomedical scientists and physicians to develop custom imaging workflows that synergistically combine advances in optical probes, sample preparation, instrument design and operation, and computer vision-

based analyses of TB-scale datasets. Imaging systems include 1) two cleared tissue Axially Swept Light-Sheet Microscopes that achieve 300 and 650 nm isotropic resolution, respectively, throughout centimeter-scale optically cleared tissues, 2) a high-speed single-objective oblique plane light-sheet microscope with 200 and 450 nm lateral and axial resolution, respectively, 3) a hyperspectral TIRF system capable of imaging 6 genetically encoded fluorescent probes in living cells, 4) a 2 and 3-photon laser scanning confocal equipped with adaptive optics, extended depth of focusing, and rapid axial scanning for ultradeep intravital imaging, and 5) a mesoSPIM microscope for macro-scale imaging with 5 μm isotropic resolution. Furthermore, through collaborations with the Danuser and Fiolka labs, a 6) 2-photon micro-environmental selective plane illumination microscope with 300 nm isotropic resolution, 7) a live-cell 1-photon Axially Swept Light-Sheet Microscope with 350 nm isotropic resolution, 7) a Field Synthesis variant of Lattice Light-Sheet Microscopy, 8) and a multi-directional illumination light-sheet microscope for imaging of developing organisms. Research combines biology, biophysics, optics, and computer vision.

Founder 2019 – 2022
Discovery Imaging Systems, LLC, Dallas, TX, USA.
<https://www.discoveryimagingystems.com>

Established an LLC to provide customized and world-class imaging solutions to academic research centers. Business portfolio focused on new methods for imaging chemically transparent biological specimens with the cleared tissue variants of Axially Swept Light-Sheet Microscopy. Customer purchases third-party components, and Discovery Imaging Systems provides expert assembly on site and annual maintenance thereafter.

Postdoctoral Fellow, Fiolka and Danuser Laboratories 2014 – 2018
Lyda Hill Department of Bioinformatics
UT Southwestern Medical Center, Dallas, TX, USA.

Development and application of light-sheet microscopy and computer vision technologies to elucidate mechanisms of cell-extracellular matrix adhesions, Rho GTPase signaling, and cytoskeletal dynamics of breast cancer cells disseminating into 3D tissue-like environments. Ruth L. Kirschstein Postdoctoral Fellow. Research conducted under the guidance of Drs. Reto Fiolka and Gaudenz Danuser.

Director, BioFrontiers Advanced Imaging Resource 2013 – 2014
BioFrontiers Institute
University of Colorado, Boulder, CO, USA.

Established first campus-wide microscopy facility and expanded user base expanded to over 100 users from eight departments, two universities, and two biotechnology firms. Led a collaborative team in the building and development of novel microscopy and super-resolution instrumentation and techniques; executed independent research; provided facility strategic vision; organized analytical training for users; worked with researchers to develop customized protocols and assays; collaborated with faculty to compete for grants and contracts to support facility research activities and lead infrastructure grants; interfaced and worked closely with industrial users. Directly reported to the Chief Scientific Officer, Dr. Leslie Leinwand, and Dr. Tom Cech.

Graduate Research Assistant, Palmer Laboratory. 2007 – 2013
Department of Biochemistry
University of Colorado, Boulder, CO, USA.

Investigated the role of site-specific mutations in dark-state conversion and irreversible photobleaching for red-fluorescent proteins; developed high-throughput microfluidic cell-sorter capable of measuring fluorescence lifetime and the rate of photobleaching on single mammalian cells; directed-evolution of red-fluorescent proteins for improved photostability; collaborated with other lab members on the development and evaluation of optical biosensors. Research conducted under the guidance of Drs. Amy E. Palmer and Ralph Jimenez.

Undergraduate Research Assistant, Williamson Laboratory. 2003 – 2006

Department of Chemistry
Willamette University, Salem, OR, USA.

Development of semi-automated laser light scattering instrumentation and the measurement of highly accurate and precise binary liquid-liquid phase diagrams. Research conducted under the guidance of Dr. J. Charles Williamson.

EDUCATION

Ph.D., Biochemistry GPA: 3.9
University of Colorado, Boulder, CO, USA. 2007 - 2013
Dissertation: Fluorescent Proteins: Spectroscopic Studies, Microfluidic Analysis, and Generation of Improved Variants. Research conducted under the guidance of Drs. Amy E. Palmer and Ralph Jimenez.

B.A., Chemistry GPA: 3.64
Willamette University, Salem, OR, USA. December 2002 - 2006
Thesis: Laser Light Scattering Investigations of the Isobutyric Acid/Water Binary Liquid System. Research conducted under the guidance of Dr. J. Charles Williamson.

OTHER TRAINING

Marine Biological Laboratory, Woods Hole, MA. (2016)
Computational Image Analysis, Summer Course.
Directors: Lani Wu, Steven Altschuler, and Gaudenz Danuser

CURRENT GRANT SUPPORT

- 2023 – 2024 Next-Generation Histopathology Using Rapid Ultraviolet Photoacoustical Microscopy.
Harold C. Simmons Comprehensive Cancer Center, UTSW.
MPI
- 2023 – 2024 Illuminating the Molecular and Cellular Origins of Tissue Function in Health & Disease.
President's Research Council, UTSW.
PI
- 2021 – 2026 UTSW-UNC Center for Cell Signaling Analysis.
National Institute of General Medical Sciences, NIH.
RM1GM145399
MPI
- 2021 – 2026 Imaging Mechanisms of Metastatic Tumor Formation in Situ
National Cancer Institute, NIH.
U54CA268072
MPI

PREVIOUS GRANT SUPPORT

1. 2022 – 2023 Mechanisms of Cell Rearrangements During Germband Extension in *D. Melanogaster*.
National Institute of Child Health and Human Development, NIH.
R21HD105189
Collaborator
2. 2022 – 2023 Linking Function, Structure, and Molecular Identity of Lateral Habenula Neurons.
National Institute of Mental Health, NIH.

R34NS121873

Collaborator

3. 2021 – 2026 UTSW Cancer Center Support Grant
National Cancer Institute, NIH.
P30CA142543
Key Personnel
4. 2020 – 2025 Prune Belly Syndrome: Mechanisms of Filamin A Mutations.
National Institute of Diabetes and Digest and Kidney Diseases, NIH.
R01DK127589
Co-Investigator
5. 2020 – 2024 Neuronal Signaling Mechanisms of Stress-Induced Anhedonia in the Lateral Habenula.
National Institute of Mental Health, NIH.
R01MH120131
Collaborator
6. 2019 – 2020 Laser for Ultra-Deep Intravital Imaging.
National Institute of General Medical Sciences, NIH.
S10OD026722
Key Personnel
7. 2016 – 2018 Symmetry breaking and polarization of cells in 3D environments.
National Institute of General Medical Sciences, NIH.
F32GM117793
Principal Investigator
8. 2011 – 2013 Integrative Graduate Education and Research Trainee (IGERT)
Computational, Optical, Sensing and Imaging.
Division Of Graduate Education, NSF
0801680
Trainee
9. 2009 – 2011 Interdisciplinary Predoctoral Training in Molecular Biophysics.
National Institute of General Medical Sciences, NIH.
T32GM065103
Trainee
10. 2006 – 2007 Sigma Xi Undergraduate Research Fellowship.
Trainee

PEER-REVIEWED PUBLICATIONS

1. Jinlong Lin et al, Mechanically sheared Axially Swept Light-Sheet Microscopy. *Submitted.*
2. Zach Marin, Xiaoding Wang et al, navigate: an open-source platform for smart light-sheet microscopy. *Submitted*
3. Benjamin Nanes et al, Keratin isoforms modulate motility signals during wound healing. *Submitted.*
4. Erik S. Welf et al, Worrying drives cell migration in mechanically unrestrained environments. *Submitted.*
5. Tadamoto Isogai, Kevin M. Dean et al, Direct Arp2/3-vinculin binding is essential for cell spreading, but only on compliant substrates and in 3D. *Submitted.*
6. Robert J. Ju et al, Compression-dependent microtubule reinforcement comprises a mechanostat which enables cells to navigate confined environments. *Accepted, Nat. Cell Biol.*
7. Pancheng Xie et al, Mammalian circadian clock proteins form dynamic interacting microbodies distinct from phase separation. *Proc. Nat. Acad. Sci. USA.* 2023 Dec; (120) e2318274120.

8. Philippe Roudot et al, u-track 3D: measuring and interrogating intracellular dynamics in three dimensions. *Cell Rep. Methods*. 2023 Dec; (3) 100655.
9. Mohamad I. Cheikh et al, A comprehensive model of Drosophila epithelium reveals the role of embryo geometry and cell topology in mechanical responses. *eLife*. 2023. Oct; (12) e85569.
10. Andrew D. Weems et al, Blebs Promote Cell Survival by Assembling Oncogenic Signaling Hubs. *Nature*. 2023 Mar; (615) 517-525.
11. Alec Bancroft et al, Discoidin Domain Receptor 2 regulates aberrant mesenchymal progenitor cell fate and matrix organization. *Sci. Advances*. 2022 Dec; (51) eabq6152.
12. Bingying Chen et al, Increasing the field-of-view in oblique plane microscopy via optical tiling. *Biomed. Opt. Exp.* 2022 Nov; (13) 5616-5627.
13. Bingying Chen et al, Resolution doubling in light-sheet microscopy via oblique plane structured illumination. *Nat. Methods* 2022 Nov; (11) 1419-1426.
14. Kevin M. Dean et al, Isotropic imaging across spatial scales with axially swept light-sheet microscopy. *Nat. Protoc.* 2022 Jul; (17) 2025-2053.
15. Sue Y. Kim et al, Particle Retracking Algorithm Capable of Quantifying Large, Local Matrix Deformation for Traction Force Microscopy. *PLoS ONE*. 2022 Jun; (17) e0268614.
16. Iliodora V Pop et al, Structure of long-range direct and indirect spinocerebellar pathways as well as local spinal circuits mediating proprioception. *Neurosci.* 2022 Jan; (42) 581-600.
17. Hao Liu et al, Heterozygous mutation of Vegfr3 decreases renal lymphatics but is dispensable for renal function. *J. Am. Soc. Neph.* 2021 Sep; (32) 3099-3113.
18. Bo-Jui Chang et al, Real-time multi-angle projection imaging of biological dynamics. *Nat. Methods*. 2021 Jun; (18) 829-834.
19. Sangyoon J. Han et al, Pre-complexation of talin and vinculin without tension is required for efficient nascent adhesion maturation. *eLife*. 2021 Mar; (10) e66151. *Co-corresponding author*.
20. Erik S. Welf et al, Actin-membrane release initiates cell protrusion. *Dev. Cell*. 2020 Dec; (55) 723-736.
21. Etai Sapoznik et al, A versatile Oblique Plane Microscope for large-scale and high-resolution imaging of subcellular dynamics. *eLife*. 2020 Nov; (9) e57681. *Co-corresponding author*.
22. Kyung-min Lee et al, Proline rich 11 (PRR11) overexpression amplifies PI3K signaling and promotes antiestrogen resistance in breast cancer. *Nat. Comm.* 2020 Oct; (11) 1-15.
23. Tonmoy Chakraborty, Bingying Chen et al, Converting Lateral Scanning Into Axial Focusing to Speed Up 3D Microscopy. *Light Sci. Appl.* 2020 Sep; (9) 00401-00409.
24. Bo-Jui Chang, Kevin M. Dean, and Reto Fiolka. Systematic and quantitative comparison of lattice and Gaussian light-sheets. *Opt. Express*. 2020 Aug; (28) 27052-27077.
25. Bingying Chen, Tonmoy Chakraborty et al, Extended depth of focus multiphoton microscopy via incoherent pulse splitting. *Biomed. Opt. Express*. 2020 Jun; (11) 3830-3842.
26. Tonmoy Chakraborty et al, Light-sheet microscopy of Cleared Tissues with Isotropic, Subcellular Resolution. *Nat. Methods*. 2019 Nov; (16), 1109-1113. *Co-corresponding author*.
27. Meghan K. Driscoll et al, Robust and automated detection of subcellular morphological motifs in 3D microscopy images. *Nat. Methods*. 2019 Sept; (16) 1037-1044.
28. Ashwathi S. Mohan et al, Enhanced dendritic actin network formation in lamellipodia drives proliferative signaling in growth-challenged Rac1^{P29S} melanoma cells. *Dev. Cell*. 2019 May; (49) 444-460.
29. Bo-Jui Chang, Mark Kittisopikul et al, Universal Light-Sheet Generation with Field Synthesis. *Nat. Methods*. 2019 Feb; (16) 235-238.
30. Kevin M. Dean and Reto Fiolka. Lossless Three-Dimensional Parallelization in Digitally Scanned Light-Sheet Fluorescence Microscopy. *Sci. Rep.* 2017 Aug; (7), 9332.

31. Kevin M. Dean et al, Imaging Subcellular Dynamics with Fast and Light-Efficient Volumetrically Parallelized Microscopy. *Optica*. 2017 Feb; (4), 263-271.
32. Jun Chu et al, A Bright Cyan-Excitable Orange Fluorescent Protein for Dual-Emission Microscopy and Highly Sensitive Bioluminescence Imaging In Vivo. *Nat. Biotechnol.* 2016 May 30; (34), 760-767.
33. J. Charles Williamson et al. Determination of Liquid-Liquid Critical Point Composition Using 90° Laser Light Scattering. *Phys. Rev. E*. 2016 Apr 21; (93), 042610.
34. Kevin M. Dean, Philippe Roudot et al, Diagonally Scanned Light-Sheet Microscopy for Fast Volumetric Imaging of Adherent Cells. *Biophys. J.* 2016 Mar 29; (110), 1456-1465.
35. Erik S. Welf, Meghan K. Driscoll et al, Quantitative Multiscale Cell Imaging in Controlled 3D Microenvironments. *Dev. Cell*. 2016 Feb 22; (36), 462-475.
36. Kevin M. Dean et al. Deconvolution-Free Subcellular Imaging with Axially Swept Light Sheet Microscopy. *Biophys. J.* 2015 Jun 16; 108(12), 2807-2815.
37. Kevin M. Dean et al, High-Speed Multiparameter Photophysical Analyses of Fluorophore Libraries. *Anal. Chem.* 2015 Apr 21; 87(10), 5026-30.
38. Kevin M. Dean et al, Microfluidics-Based Selection of Red-Fluorescent Proteins with Decreased Rates of Photobleaching. *Int. Biol.* 2014 Nov 21; 7(2), 263-73.
39. Kevin M. Dean and Reto Fiolka. Uniform and Scalable Light-Sheets Generated by Extended Focusing. *Opt. Express*. 2014 Oct 16;22(21),26141-26152.
40. Kevin M. Dean and Amy E. Palmer. Advances in Fluorescence Labeling Strategies for Dynamic Cellular Imaging. *Nat. Chem. Biol.* 2014 May 16;10(7):512-23.
41. Yan Qin et al, Direct Comparison of a Genetically Encoded Sensor and Small Molecule Indicator: Implications for Quantification of Cytosolic Zn²⁺. *ACS Chem. Biol.* 2013 Aug 30;8(11):2366-71.
42. Lloyd M. Davis et al, Microfluidic Cell sorter for Use in Developing Red Fluorescent Proteins with Improved Photostability. *Lab Chip*. 2013 Jun 21;13(12):2320-7.
43. Kevin M. Dean, Yan Qin, and Amy E. Palmer. Visualizing Metal Ions in Cells: An Overview of Analytical Techniques, Approaches, and Probes. *Biochim. et Biophys. Acta*. 2012 Sep;1823(9):1406-15.
44. Jennifer L. Lubbeck et al, Microfluidic Flow Cytometer for Quantifying Photobleaching of Fluorescent Proteins in Cells. *Anal. Chem.* 2012 May 1;84(9):3929-37.
45. Kevin M. Dean et al, Analysis of Red-Fluorescent Proteins Provides Insight Into Dark-State Conversion and Photodegradation. *Biophys. J.* 2011 Aug 17;101(4):961-9.
46. Kevin M. Dean and J. Charles Williamson. The Stir-Settle Approach to Semiautomated Light Scattering Determination of Liquid-Liquid Coexistence Curves. *J. Chem. Eng. Data*. 2011 Jan 26;56(4):1433-7.
47. Kevin. M. Dean et al, The Accuracy of Liquid-Liquid Phase Transition Temperatures Determined from Semiautomated Light Scattering Measurements. *J. Chem. Phys.* 2010 Aug 21;133, 074506.

PATENTS

1. 13,360,706 – Optically Integrated Microfluidic Cytometer for High Throughput Screening of Photophysical Properties of Cells or Particles.
2. 62,155,980 – Uniform and Scalable Light-Sheets Generated by Extended Focusing.
3. 16,093,561 – Light-Sheet Microscope with Parallelized 3D Image Acquisition.

INVITED TALKS

1. Light Microscopy Australia. Melbourne, VIC, Australia, 2024.
2. Microscopy & Microanalysis. Minneapolis, MN, USA, 2023.
3. EMBL-Janelia Bioimaging Seminar Series. Virtual. 2023.
4. MC2 Image Analysis Working Group. Virtual. 2023.
5. Willamette University. Salem, OR, USA. 2023.
6. Photonics Media Webinar Series. Virtual. 2022.
7. CLEO: Laser Science to Photonic Applications Technical Conference. San Jose, CA, USA. 2022.
8. Light-Sheet Conference and Workshop, MBL. Woods Hole, MA, USA. 2022.
9. IQ Health Science & Engineering, Michigan State University. Lansing, MI, USA. 2021.
10. Photonics Media Webinar Series. Virtual. 2020.
11. Imaging ONE WORLD Series. Virtual. 2020.
12. Next Generation Microscopy Workshop. Cambridge, U.K. 2019.
13. Cell Biology Seminar Series, Denver University, Denver, CO, USA. 2017.
14. Andor Academy, University of Colorado, Boulder, CO, USA. 2014.
15. American Chemical Society, Orlando, FL, USA. 2019.

INVITED WORKSHOPS

1. M2PC Workshop. Pacific Northwest National Labs. Virtual. 2021.
2. Software for Microscopy. Janelia Farm, Howard Hughes Medical Institute. Ashburn, VA. 2020.

OTHER TALKS & PRESENTATIONS

1. Kevin M. Dean. American Society for Cell Biology (ASCB). Virtual. 2021.
2. Kevin M. Dean. Society for Neuroscience (SfN). Virtual. 2021.
3. Kevin M. Dean. EMBO & EMBL Symposium: Seeing is Believing - Imaging the Processes of Life. Virtual. 2021.
4. Doug P. Shepherd, Alfred Millett-Sikking, Andrew G. York, Reto P. Fiolka, Kevin M. Dean. Focus On Microscopy. Virtual. 2021.
5. Kevin M. Dean, Denise Ramirez, Reto Fiolka, Hu Zhao, and Kate Luby-Phelps. Arizona Imaging and Microanalysis Conference. Virtual. 2021.
6. Tonmoy Chakraborty, Reto Fiolka, and Kevin M. Dean. The Network of European Bioimage Analysts (NEUBIAS). Bordeaux, France. 2020.
7. Kevin M. Dean. Imaging Mouse Development. Janelia Farm, Howard Hughes Medical Institute. Ashburn, VA, USA. 2020.
8. Alfred Millett-Sikking, Andrew G. York, Reto Fiolka, Kevin M. Dean. Focus On Microscopy. Osaka, Japan. *Cancelled*. 2020.
9. Kevin M. Dean. The Academy of Medicine, Engineering & Science of Texas (TAMEST). Dallas, TX, USA. 2020.
10. Tonmoy Chakraborty, Reto Fiolka, and Kevin M. Dean. EMBL Seeing is Believing. Heidelberg, Germany. 2019.
11. Kevin M. Dean. Frontiers and Challenges in Laser-Based Biological Microscopy. Telluride, CO, USA. 2019.

12. Tonmoy Chakraborty, Bo-Jui Chang, Kevin M. Dean, and Reto Fiolka. Focus On Microscopy. London, U.K. 2018.
13. Kevin M. Dean, Tadamoto Isogai, Philippe Roudot, Erik S. Welf, Meghan K. Driscoll, Reto Fiolka, and Gaudenz Danuser. Focus On Microscopy. Singapore, Republic of Singapore. 2018.
14. Kevin M. Dean, Erik S. Welf, Philippe Roudot, Meghan K. Driscoll, Reto Fiolka, and Gaudenz Danuser. Frontiers in Imaging Science. Janelia Farm, Howard Hughes Medical Institute. Ashburn, VA, USA. 2017.
15. Kevin M. Dean, Erik S. Welf, Philippe Roudot, Meghan K. Driscoll, Reto Fiolka, and Gaudenz Danuser. Biophysical Society. New Orleans, LA, USA. 2017.
16. Kevin M. Dean, Erik S. Welf, Philippe Roudot, Meghan K. Driscoll, Reto Fiolka, and Gaudenz Danuser. Quantitative BioImaging 2017. College Station, TX, USA. 2017.
17. Kevin M. Dean et al, Microscope Technologies for Quantitative Imaging in 3D Microenvironments. *SPIE, Biomedical Optics and Medical Imaging*. 2016.
18. Kevin M. Dean and Reto Fiolka. Isotropic 3D Imaging over Large Volumes with Axially Swept Light-Sheet Microscopy. *inFocus Magazine, Royal Microscopical Society*. 2016.
19. Kevin M. Dean, Erik S. Welf, Philippe Roudot, Meghan K. Driscoll, Reto Fiolka, and Gaudenz Danuser. Gordon Research Conference - Signaling by Adhesion Receptors. Bates College, Lewiston, Maine, USA. 2016.
20. Kevin M. Dean, Philippe Roudot, Erik S. Welf, Meghan K. Driscoll, Reto Fiolka, and Gaudenz Danuser. EMBO & EMBL Symposium: Seeing is Believing - Imaging the Processes of Life. EMBL, Heidelberg, Germany. 2015.
21. Kevin M. Dean, Jennifer L. Lubbeck, Premashis Manna, Lloyd M. Davis, Ralph Jimenez, Amy E. Palmer. EMBO & EMBL Symposium: Seeing is Believing - Imaging the Processes of Life. EMBL, Heidelberg, Germany. 2013.
22. Kevin M. Dean, Jennifer L. Lubbeck, Lloyd M. Davis, Ralph Jimenez, Amy E. Palmer. Signaling and Cellular Regulation Symposium. University of Colorado, Boulder, CO, USA. 2013.
23. Kevin M. Dean, Jennifer L. Lubbeck, Lloyd M. Davis, Ralph Jimenez, Amy E. Palmer. Computational, Optical, Sensing and Imaging. University of Colorado, Boulder, CO, USA. 2013.
24. Kevin M. Dean, Jennifer L. Lubbeck, Lloyd M. Davis, Ralph Jimenez, Amy E. Palmer. Computational Optical, Sensing and Imaging IAB Meeting. Breckenridge, CO, USA. 2012.
25. Kevin M. Dean, Jennifer L. Lubbeck, Lloyd M. Davis, Ralph Jimenez, Amy E. Palmer. Colorado Photonics Industry Association Annual Meeting. Boulder, CO, USA. 2011.
26. Kevin M. Dean, Jennifer L. Lubbeck, Lloyd M. Davis, Ralph Jimenez, Amy E. Palmer. Molecular and Cellular Biophysics Symposium at the University of Denver. Denver, CO, USA. 2011.
27. Kevin M. Dean, Jennifer L. Lubbeck, Lloyd M. Davis, Ralph Jimenez, Amy E. Palmer. Multiphoton Imaging: The Next 6×10^{23} Femtoseconds, Janelia Farm, Howard Hughes Medical Institute. Ashburn, VA, USA. 2010.
28. Kevin M. Dean, Jennifer L. Lubbeck, Hairong Ma, Ralph Jimenez, Amy E. Palmer. New Optical Methods in Cell Physiology, Society of General Physiologists. Wood's Hole, MA, USA. 2010.
29. Kevin M. Dean, Jennifer L. Lubbeck, Hairong Ma, Ralph Jimenez, Amy E. Palmer. Novel Approaches to Bioimaging II, Janelia Farm, Howard Hughes Medical Institute. Ashburn, VA, USA. 2010.
30. Kevin M. Dean, Joseph C. Williamson, Student Scholarship Recognition Day, Salem, OR, USA. 2006.
31. Kevin M. Dean, Joseph C. Williamson, Northwest Undergraduate Science Research Conference at Oregon Health & Science University, Portland, OR, USA. 2006.
32. Kevin M. Dean, Joseph C. Williamson, Student Scholarship Recognition Day, Salem, OR, USA. 2005.

CONFERENCE PAPERS

1. Zach Marin et al, Autonomous Multiscale Axially Swept Light-Sheet Microscopy. Microscopy & Microanalysis, 2023.
2. Hazel Borges et al, Quantitative Cleared Tissue Imaging. Microscopy & Microanalysis, 2023.
3. Kevin M. Dean et al, Microfluidic Flow Cytometer for Multiparameter Screening of Fluorophore Photophysics. IEEE, 2014.

CONFERENCE ORGANIZING

2023	CLEO: Laser Science to Photonic Applications Technical Conference.
2022	Biomedical Applications Program. OSA Biophotonics Congress.
2022	BRAIN Technical Program Committee. OSA Biophotonics Congress.

STUDY SECTION

2023	National Institutes of Health, MOSAIC K99/R00 Special Emphasis Panel Meeting.
2023	National Institutes of Health, Neurotech. and Vision Review Special Emphasis Panel.
2022	National Institutes of Health, Neurotech. and Vision Review Special Emphasis Panel.
2022	Netherlands Organization for Scientific Research (NWO, the Dutch Research Council).
2019	Netherlands Organization for Scientific Research (NWO, the Dutch Research Council).

ADVISORY PANELS

2021 – Present	Beckman Center for Advanced Light Sheet Microscopy and Data Science, University of Colorado BioFrontiers Institute.
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TEACHING EXPERIENCE

2024 – Present	Lead Instructor, Introduction to Python, UTSW.
2024 – Present	Instructor, Optical Imaging and Probe Engineering, UTSW.
2023 – Present	Lead Instructor, Introduction to Python Software Development on GitHub, UTSW.
2022 – Present	Discussion Leader, Responsible Conduct of Research, UTSW.
2021 – Present	Assistant Instructor, Computational Image Analysis, UTSW.
2015 – 2019	Assistant Instructor, Optical Microscopy, UTSW.
2013 – 2014	Laboratory Assistant, Light Microscopy, University of Colorado.
2007 – 2008	Teaching Assistant, Introductory Chemistry, University of Colorado.
2006 – 2007	Tutor, Introductory Chemistry and Physical Chemistry, Willamette University.

INSTITUTIONAL COMMITTEES

2022 – Present	Molecular Biophysics Work-In-Progress Evaluator.
2022 – Present	UT Southwestern Committee on Improving Research Staff Recruitment and Hiring.
2022 – Present	Lyda Hill Dept. of Bioinformatics and BME-Computational Biology Retreat.
2022 – Present	UT Southwestern Graduate School Admissions Committee.
2021 – Present	Dept. of Biophysics and Lyda Hill Dept. of Bioinformatics Faculty Search.

2021 – Present Faculty Senate at UT Southwestern.
2021 – Present UT Southwestern Medical School Admissions Committee.

SCIENTIFIC MENTORSHIP

High-School Students

2021 – 2022 Samir Mamtani, Undergraduate Student, University of Wisconsin.
2021 – 2022 Sampath Rapuri, Undergraduate Student, Johns Hopkins University.
2021 – 2022 Renil Gupta, High-School Student, St. Marks H.S.
2019 – 2019 Ryan Hammond, Undergraduate Student, Carnegie Mellon University.

Undergraduate Students

2023 – Present Cammy Kutter, Undergraduate Student, UCLA.
2023 – Present Sai Bommisetty, Undergraduate Student, UT Dallas.
2023 – Present Joy Camposagrado, Undergraduate Student, UT Dallas.
2023 – Present James Harrigan, Undergraduate Student, UT Dallas.
2023 – Present Rayyan Sappayani, Undergraduate Student, UT Dallas.
2023 – Present Abhijeet Kulkarni, Undergraduate Student, UT Dallas.
2023 – Present Beshar Nusair, Undergraduate Student, UT Dallas.
2023 – 2023 Evan Wylie, Undergraduate Student, Willamette University.
2012 – 2013 Jordan Gringauz, Legal Associate, Finnegan.
2011 – 2013 Derek Gann, Senior Devices Engineer, Heirloom.
2010 – 2011 Jennifer Binder, Graduate Student, Arizona State University.

Graduate Students

2023 – Present Seweryn Galecki, M.S., Fulbright Graduate Student.
2023 – 2023 Calvin Jones, Graduate Research Assistant, Boise State University.
2011 – 2013 Pramashis Manna, Assistant Professor, Ohio State University.

Postdoctoral Fellows

2024 – Present Derek Abbott, Ph.D., UT Southwestern Medical Center.
2023 – Present Qionghua Shen, Ph.D., UT Southwestern Medical Center.
2023 – Present Dushyant Mehra, Ph.D., UT Southwestern Medical Center.
2023 – Present John Haug, Ph.D., UT Southwestern Medical Center.
2022 – 2023 Zach Marin, Ph.D., Postdoctoral Fellow, Max Perutz Labs Vienna.
2018 – 2019 Qiuyan Shao, Principal Data Scientist at Capital One.

Staff Scientists

2023 – Present Samantha Calvache, Research Technician II, UT Southwestern Medical Center.
2022 – Present Xiaoding 'Annie' Wang, M.S., Software Engineer, UT Southwestern Medical Center.
2022 – 2023 Melissa Glidewell, Territory Manager, 89 North.
2021 – Present Jinlong Lin, Research Technician I, UT Southwestern Medical Center.
2021 – Present Hazel Borges, M.S., Research Assistant I, UT Southwestern Medical Center.
2021 – 2023 Dax Collision, Software Engineer, Atmos Energy.
2019 – 2021 Evgenia Azarova, Graduate Student, Johns Hopkins University.
2018 – 2020 Saumya Vora, M.S., Regulatory Affairs Specialist, Medtronic.
2017 – 2019 Justin Cillay, M.D., Resident Physician, University of Colorado.
2012 – 2014 Pia Friis, Laboratory Technician in Clinical Microbiomics.

TRAINEE GRANT SUPPORT

2023 Hazel Borges, Carl Storm Underrepresented Minority (CSURM) Fellowship.

DISSERTATION COMMITTEES

2020 – Present Harshida Pancholi, Roberts Lab, Dept. of Neuroscience, UTSW.
2020 – 2022 Byron Weiss, Danuser Lab, Lyda Hill Dept. of Bioinformatics, UTSW.

DISTINCTIONS

2023 Recipient, Distinguished Researcher Award from the UTSW President's Research Council
2022 Inductee, Weed High School Hall of Fame.
2018 Dean's Discretionary Award, UT Southwestern Medical Center.
2018 Runner-Up, UTSW Brown-Goldstein Excellence in Postdoctoral Research.
2016 Nominee, AAAS/Science Program for Excellence in Science.
2015 Finalist, 2015 Educational Award, Edmund Optics.
2011 Certificate of Achievement, Colorado Photonics Industry Association (CPIA).
2007 Excellence in Graduate Teaching.
2007 ESPN & CoSIDA District 8 Academic All-American.
2007 Inducted into National Football Foundation Hampshire Honor Society.
2007 Co-Recipient, Henry Booth Outstanding Senior Male Athlete Award.
2006 Willamette University Bill Trenbeath Award.
2006 ESPN & CoSIDA District 8 Academic All-American.
2006 Sigma Alpha Epsilon National Scholar.
2006 Willamette University Athlete of The Month.
2006 Northwest Conference Athlete of the Week.
2006 Willamette University Football Team Captain.
2005 Invited to Speak at the Murdock's NW Undergraduate Research Meeting.
2005 Willamette University Football Team Captain.
2005 Peterson Family Scholarship for Chemistry.
2005 Inducted into the Mortar Board National Honor Society.
2004 Sigma Alpha Epsilon National Scholar.
2004 Inducted into the National Society of Collegiate Scholars (NSCS).
2004 Selected for Science Collaborative Research Program.
2002 Haynes McHale Award.
2002 Neal Wade Award - Weed High School Outstanding Male Athlete of the Year.
2002 Inducted into California Scholarship Federation.

LEADERSHIP POSITIONS

2007 – 2013 Sigma Alpha Epsilon - University of Colorado Advisor and Regional Director
2007 Coast 2 Coast 4 Kalan Memorial Bike Ride - Philanthropy Coordinator and Participant
2005 – 2006 Willamette University Football Team Captain

PHILANTHROPY & COMMUNITY SERVICE

2016 – 2022 Neonatal Support Staff, Parkland Neonatal Intensive Care Unit

2015 – 2020	Science Advocate, Science Policy, Education, and Communication club (SPEaC)
2006	Participant and Organizer, Coast to Coast for Kalan Memorial Bicycle Ride
2004 – 2005	Participant, United Way Day of Caring
2004 – 2005	Participant, Willamette University Bearcat Day Student Athlete Youth Outreach
2002	Participant, Habitat for Humanity

REFeree FOR JOURNALS

- Cell
- Nature Methods
- Nature Protocols
- Nature Biotechnology
- Nature Scientific Data
- Scientific Reports
- Journal of Neuroscience Methods
- Optics Express
- Biomedical Optics Express
- Journal of Biophotonics
- Biophysical Journal
- PLoS One
- EMBO Molecular Systems Biology
- eLife
- International Journal of Biochemistry and Cell Biology
- Laser & Photonics Reviews

PROFESSIONAL MEMBERSHIPS

American Society for Cell Biology
American Chemical Society
Biophysical Society
Society for Neuroscience