# Package 'qPCRtools'

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Version 0.2.3

 ${\bf Title}\ \, {\bf Tools}\ \, {\bf for}\ \, {\bf qPCR}$ 

biological research. A crucial step in processing qPCR data is to calculate the amplification efficiency of genes to determine which method should be used to calculate expression level of genes. This Package can do it easily.  In addition to that, this package can calculate the expression level of genes based on three methods.
<pre>URL https://github.com/lixiang117423/qPCRtools</pre>
BugReports https://github.com/lixiang117423/qPCRtools/issues
License MIT + file LICENSE
Imports broom, data.table, dplyr, ggplot2, ggpmisc, ggthemes, kableExtra, magrittr, multcomp, readxl, reshape2, rstatix, sjmisc, stringr, tibble, tidyr, xlsx
RoxygenNote 7.2.1
Suggests knitr, rmarkdown
VignetteBuilder knitr
NeedsCompilation no
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CalCurve

CalCurve

Standard Curve Calculation.

# Description

The function can calculate the standard curve. At the same time, it can get the amplification efficiency of primer(s). Based on the amplification efficiency, we can know which method can be used to calculate the expression level.

# Arguments

cq.table The data frame of the position and Cq value.

concen.table The data frame of the position and concentration.

highest.concen The highest concentration for calculation.

lowest.concen The lowest concentration for calculation.

dilution Dilution factor of cDNA template. The default value is 4.

by.mean Calculation by mean Cq value or not. The default value is TRUE.

### Value

A list.

# Author(s)

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### Examples

```
df.1.path <- system.file("examples", "calsc.cq.txt", package = "qPCRtools")
df.2.path <- system.file("examples", "calsc.info.txt", package = "qPCRtools")
df.1 <- data.table::fread(df.1.path)
df.2 <- data.table::fread(df.2.path)
CalCurve(
    cq.table = df.1,
        concen.table = df.2,
        lowest.concen = 4,
        highest.concen = 4096,
        dilu = 4,
        by = "mean"
) -> p

p[["table"]]
p[["figure"]]
```

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CalExp2dCt	Calculate expression using standard curve.	
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### Description

Calculate expression using standard curve.

# Arguments

cq.table The data frame of the position and cq value.

design.table The data frame of the position and corresponding information.

ref.gene The name of reference gene.

#### Value

A list contain a table and a figure.

#### Author(s)

Xiang LI ¡lixiang117423@gmail.com;

#### Examples

CalExp2ddCt

Calculate expression using standard curve.

#### Description

Calculate expression using standard curve.

#### Arguments

cq.table The data frame of the position and cq value.

design.table The data frame of the position and corresponding information.

correction Correct expression value by reference gene.

ref.gene The name of reference gene.
ref.group The name of reference group.

stat.method Statistical method.

fig.type Output image type, 'box' represents 'boxplot', 'bar' represents 'barplot'.

fig.ncol Number of columes of figure.

CalExpCurve

#### Value

A list contain a table and a figure.

### Author(s)

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#### Examples

 ${\tt CalExpCurve}$ 

Calculate expression using standard curve.

### Description

Calculate expression using standard curve.

#### **Arguments**

cq.table The data frame of the position and Cq value.

 ${\tt design.table} \qquad {\tt The \ data \ frame \ of \ the \ position \ and \ corresponding \ information}.$ 

correction Correct expression value by reference gene.

ref.gene The name of reference gene.

stat.method Statistical method.

ref.group The name of reference group.

fig.type Output image type, 'box' represents 'boxplot', 'bar' represents 'barplot'.

fig.ncol Number of columes of figure.

# Value

A list contain a table and a figure.

### Author(s)

Xiang LI ¡lixiang117423@gmail.com;

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#### Examples

```
df1.path = system.file("examples", "cal.exp.curve.cq.txt", package = "qPCRtools")
df2.path = system.file("examples", "cal.expre.curve.sdc.txt", package = "qPCRtools")
df3.path = system.file("examples", "cal.exp.curve.design.txt", package = "qPCRtools")
cq.table = data.table::fread(df1.path)
curve.table = data.table::fread(df2.path)
design.table = data.table::fread(df3.path)
CalExpCurve(
   cq.table,
   curve.table,
   design.table,
   correction = TRUE,
   ref.gene = "OsUBQ",
   stat.method = "t.test",
   ref.group = "CK",
   fig.type = "box",
   fig.ncol = NULL) -> res
res[["table"]]
res[["figure"]]
```

CalExpRqPCR

Calculate expression using standard curve.

### Description

Calculate expression using standard curve.

# Arguments

cq.table The data frame of the position and cq value. design.table The data frame of the position and corresponding information. Correct expression value by reference gene. correction ref.gene The name of reference gene. ref.group The name of reference group. stat.method Statistical method. Output image type, 'box' represents 'boxplot', 'bar' represents 'barplot'. fig.type fig.ncol Number of columes of figure.

# Value

A list contain a table and a figure.

#### Author(s)

Xiang LI ¡lixiang117423@gmail.com;

6 CalRTable

#### Examples

CalRTable

Calculate RNA volume for reverse transcription.

#### Description

The first step of qPCR is usually the preparation of cDNA. We need to calculate the column of RNA for reverse transcription to cDNA. So, if we have the concentration of RNA, we can use the function 'CalRTable' to do that.

# Arguments

data A data frame contained the sample names and the concentration value.

The default unit of concentration is ng/uL.

template A data frame contained the information of reverse transcription. In this

data.frame there must be a column called 'all'.

RNA.weight RNA weight required for reverse transcription. Default is 1 ug.

#### Value

A list contain a table and a figure.

#### Author(s)

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#### Examples

```
df.1.path <- system.file("examples", "crtv.data.txt", package = "qPCRtools")
df.2.path <- system.file("examples", "crtv.template.txt", package = "qPCRtools")
df.1 <- data.table::fread(df.1.path)
df.2 <- data.table::fread(df.2.path)
result <- CalRTable(data = df.1, template = df.2, RNA.weight = 2)
head(result)</pre>
```

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```
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```