

# Package ‘qPCRtools’

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**Version** 0.1.1

**Title** Tools for qPCR

**Description** A set of tools for qPCR data process.

**URL** <https://github.com/lixiang117423/qPCRtools>

**BugReports** <https://github.com/lixiang117423/qPCRtools/issues>

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**Imports** broom, data.table, dplyr, ggplot2, ggpmisc, ggthemes,  
magrittr, multcomp, readxl, reshape2, rstatix, sjmisc, stringr,  
tibble, xlsx

**RoxygenNote** 7.2.0

**NeedsCompilation** no

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CalCurve	<i>Standard Curve Calculation.</i>
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## Description

Standard Curve Calculation.

## Arguments

cq.table	The data frame of the position and Cq value.
concen.table	The data frame of the position and concentration.
highest.concen	The highest concentration.
lowest.concen	The lowest concentration.
dilution	Dilution factor of cDNA template.
by.mean	Calculation by mean Cq value or not.

**Value**

A list.

**Author(s)**

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**Examples**

```
df.1.path <- system.file("examples", "calsc.cq.txt", package = "qPCRtools")
df.2.path <- system.file("examples", "calsc.info.txt", package = "qPCRtools")
df.1 <- data.table::fread(df.1.path)
df.2 <- data.table::fread(df.2.path)
CalCurve(
  cq.table = df.1,
  concen.table = df.2,
  lowest.concen = 4,
  highest.concen = 4096,
  dilu = 4,
  by = "mean"
) -> p

p[["table"]]
p[["figure"]]
```

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CalExpCurve

*Calculate expression using standard curve.*

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**Description**

Calculate expression using standard curve.

**Arguments**

<code>cq.table</code>	The data frame of the position and Cq value.
<code>design.table</code>	The data frame of the position and corresponding information.
<code>correction</code>	Correct expression value by reference gene.
<code>ref.gene</code>	The name of reference gene.
<code>stat.method</code>	Statistical method.
<code>ref.group</code>	The name of reference group.
<code>fig.type</code>	Calculation by mean Cq value or not.
<code>fig.ncol</code>	Number of columes of figure.

**Author(s)**

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**Examples**

```
df1.path = system.file("examples", "cal.exp.curve.cq.txt", package = "qPCRtools")
df2.path = system.file("examples", "cal.expre.curve.sdc.txt", package = "qPCRtools")
df3.path = system.file("examples", "cal.exp.curve.design.txt", package = "qPCRtools")

cq.table = data.table::fread(df1.path)
curve.table = data.table::fread(df2.path)
design.table = data.table::fread(df3.path)

CalExpCurve(
  cq.table,
  curve.table,
  design.table,
  correction = TRUE,
  ref.gene = "OsUBQ",
  stat.method = "t.test",
  ref.group = "CK",
  fig.type = "box",
  fig.ncol = NULL) -> res

res[["table"]]
res[["figure"]]
```

CalRTable

*Calculate volume.***Description**

Calculate RNA and other reagent volume required for reverse transcription.

**Arguments**

<b>data</b>	A data.frame contained the sample names and the concentration value. The default unit of concentration is ng/uL.
<b>template</b>	A data.frame contained the information of reverse transcription.
<b>RNA.weight</b>	RNA weight required for reverse transcription. Default is 1 ug.

**Value**

A data frame.

**Author(s)**

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**Examples**

```
df.1.path <- system.file("examples", "crtv.data.txt", package = "qPCRtools")
df.2.path <- system.file("examples", "crtv.template.txt", package = "qPCRtools")
df.1 <- data.table::fread(df.1.path)
df.2 <- data.table::fread(df.2.path)
result <- CalRTable(data = df.1, template = df.2, RNA.weight = 2)
head(result)
```

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