





• What's the first command you should run in a python/data analysis project any after starting a terminal?

conda install ...



pip install ...



conda activate ...



jupyter lab





• What characterizes a bio-image analysis workflow that is reproducible?

It outputs numbers



It is fully automated



It is well documented



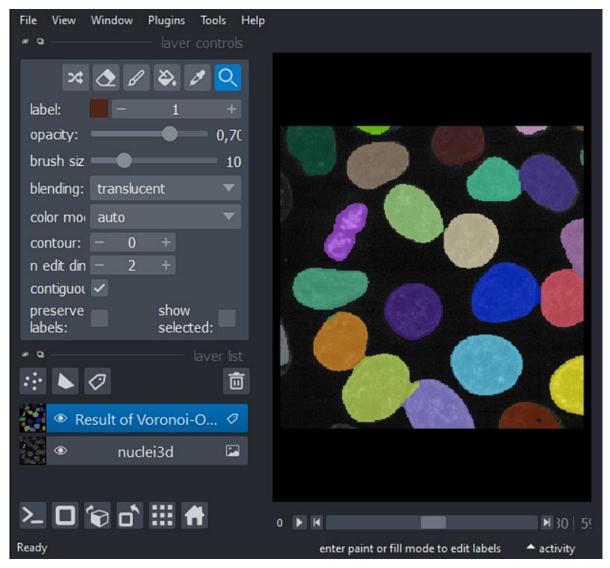
It visualizes results

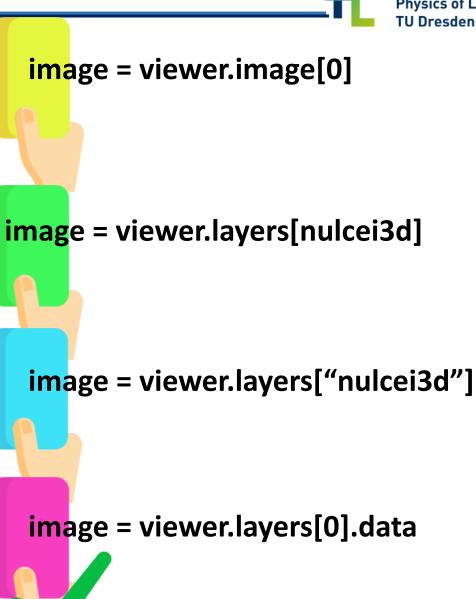


Napari



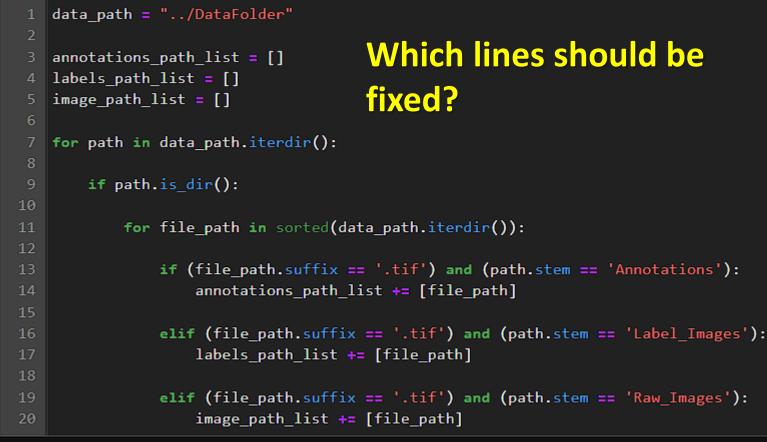
How do I get the image from napari?





Folder Structures

- DataFolder
 - Annotations
 - image_01.tif
 - image_02.tif
 - image_03.tif
 - Label_Images
 - image_01.tif
 - image 02.tif
 - image_03.tif
 - Raw_Images
 - image_01.tif
 - image_02.tif
 - image_03.tif





1 and 19

11 and 19







In Python, images can often be read by providing a path to a "imread" function. The path can be a relative path or an absolute path.

```
from skimage.io import imread

image = imread('../../data/blobs.tif')

Relative path to image

Absolute path to image

Absolute path to image
```

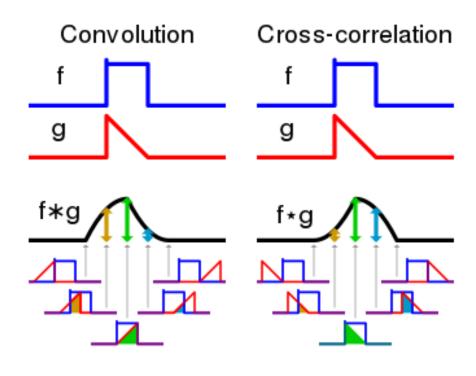
Backslash ('\') is a special character. In Windows, this may lead to errors.

```
Replace backslashes ('\') by forward slashes ('/')

from skimage.io import imread
  image = imread("C:/Users/mazo260d/Quantitative_Image_Analysis_with_Python_2022/data/blobs.tif")
```

Correlation vs Convolution





https://github.com/scipy/scipy/blob/1023d9207fdc1430a8ba196f1a1616ac3c264acf/scipy/ndimage/filters.py

Cross-Correlation

$$F \circ I(x) = \sum_{i=-N}^{N} F(i)I(x+i)$$

Convolution

$$F * I(x) = \sum_{i=-N}^{N} F(i)I(x-i)$$

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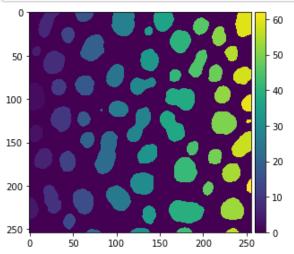


- Diversity is cool.
- Also community needs differ.
- Example: matplotlibs's imshow was not developed specifically for the life-sciences. cle.imshow was.

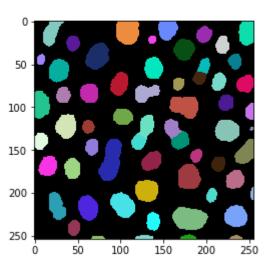
```
# labeling
labels = cle.connected_components_labeling_box(binary)
# visualize results
imshow(labels)

<matplotlib.image.AxesImage at 0x174b43f8610>

0
```



```
labels = cle.voronoi_otsu_labeling(image, spot_sigma=3.5, outline_sigma=1)
cle.imshow(labels, labels=True)
```

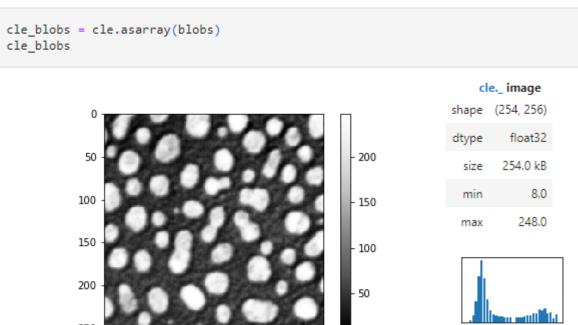


https://haesleinhuepf.github.io/BioImageAnalysisNotebooks/15 gpu acceleration/clesperanto.html



 Hint: The tools we develop (such as clesperanto) can be modified by us. Let us know what you need and we make it happen!

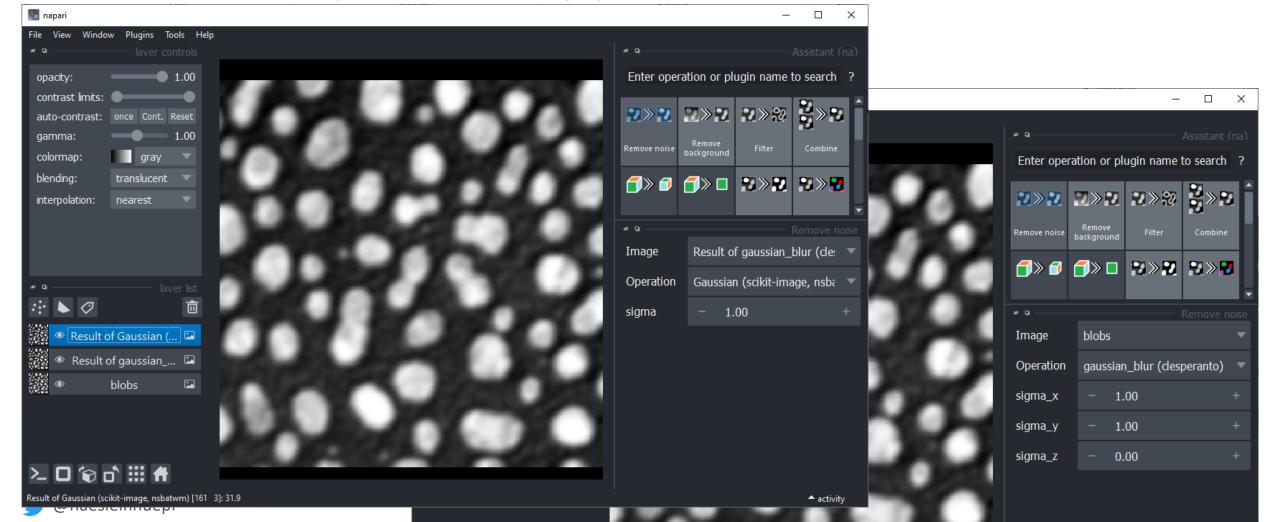
```
blobs = imread("../../data/blobs.tif")
blobs
array([[ 40, 32, 24, ..., 216, 200, 200],
        56, 40, 24, ..., 232, 216, 216],
             48, 24, ..., 240, 232, 232],
             80, 80, ..., 48, 48, 48],
       [ 96, 88, 80, ..., 48, 48, 48]], dtype=uint8)
cle.imshow(cle blobs, colormap='jet', colorbar=True)
                                      200
                                      150
                                      100
                                      - 50
```



https://github.com/clEsperanto/pyclesperanto_prototype/blob/master/demo/interoperability/jupyter.ipynb



- Example Gaussian blur:
 - Some are fast, some are slow.
 - Some run on all computers, some need a graphics card.





- Read the documentation of the different variants and make your choice. Try to stick to similar operations between projects.
- Also: Generate notebooks

```
[]: from skimage.io import imread
import pyclesperanto_prototype as cle # version 0.19.4
import napari_segment_blobs_and_things_with_membranes as nsbatwm # version 0.3.3
```

Loading 'blobs'

gaussian blur

```
[ ]: image1_gb = cle.gaussian_blur(image0_b, None, 1.0, 1.0, 0.0)
image1_gb
```

gaussian blur

```
[]: image2_G = nsbatwm.gaussian_blur(image1_gb, 1.0)
image2_G
```