

Rlabkey Users Guide

Rlabkey version 2.1.xxx LabKey Server version 16.1	Author: Peter Hussey info@labkey.com Document first release March 28, 2010 Document updated Apr 17 th , 2016
This User Guide supplements the help files on the individual functions of Rlabkey. It includes an overview of the integration between R and LabKey Server. It also covers specific topics that require interaction with the LabKey Server via the browser. Note that the screen shots of LabKey Server user pages may be different on the version of LabKey to which you are connecting.	

Working with R and LabKey Server together

Rlabkey is an interface between the R language and LabKey Server that has been designed to combine the strengths of LabKey Server and the R language platform. R has emerged as a leading open source analysis tool for bioinformatics. The combination of the R language, its packaging and distribution architecture, and an active international community of contributors have created a body of statistical and bioinformatics functionality that is widely used and constantly improving. LabKey Server is also an open-source tool for bioinformatics that helps researchers organize, analyze, and share their research data. LabKey Server is designed to manage the huge range of sizes and formats of data files generated by research instruments. LabKey Server also makes its data accessible to popular analysis tools, including R, SAS, and Excel. By facilitating both input and analysis, LabKey Server acts as a data hub for groups of researchers, whether they are in the same lab or collaborating across continents.

R was first integrated into LabKey Server as a reporting and visualization tool. "[R Reports](#)" are scripts run by LabKey Server. They are defined in the context of a specific grid view of data created through the Views -> Create -> R Report menu in the data grid. In an R Report, the current dataset shown in the grid is available to the script implicitly as a data frame named labkey.data. R Reports are commonly used to produce graphs or summary reports that are incorporated into web pages. LabKey then released the first version of the Rlabkey package (version "0.0") that was designed to enable R scripts running on a client machine to read and write data in LabKey. The second major release of the Rlabkey package (renamed version "2.1") added support for the interactive R user. The new functions in the Rlabkey package enable R users to explore and analyze the data stored in LabKey entirely from the R command prompt. Researchers and statisticians can also use Rlabkey to save their results back to LabKey Server, enabling tracking and aggregation of results over multiple runs. The combination of R Reports and the Rlabkey package makes R useful in many roles on a LabKey Server, as depicted in Figure 1.

For more information, see [The R project for Statistical Computing](http://www.r-project.org) (www.r-project.org) and [LabKey Software](http://www.labkey.com) (www.labkey.com).

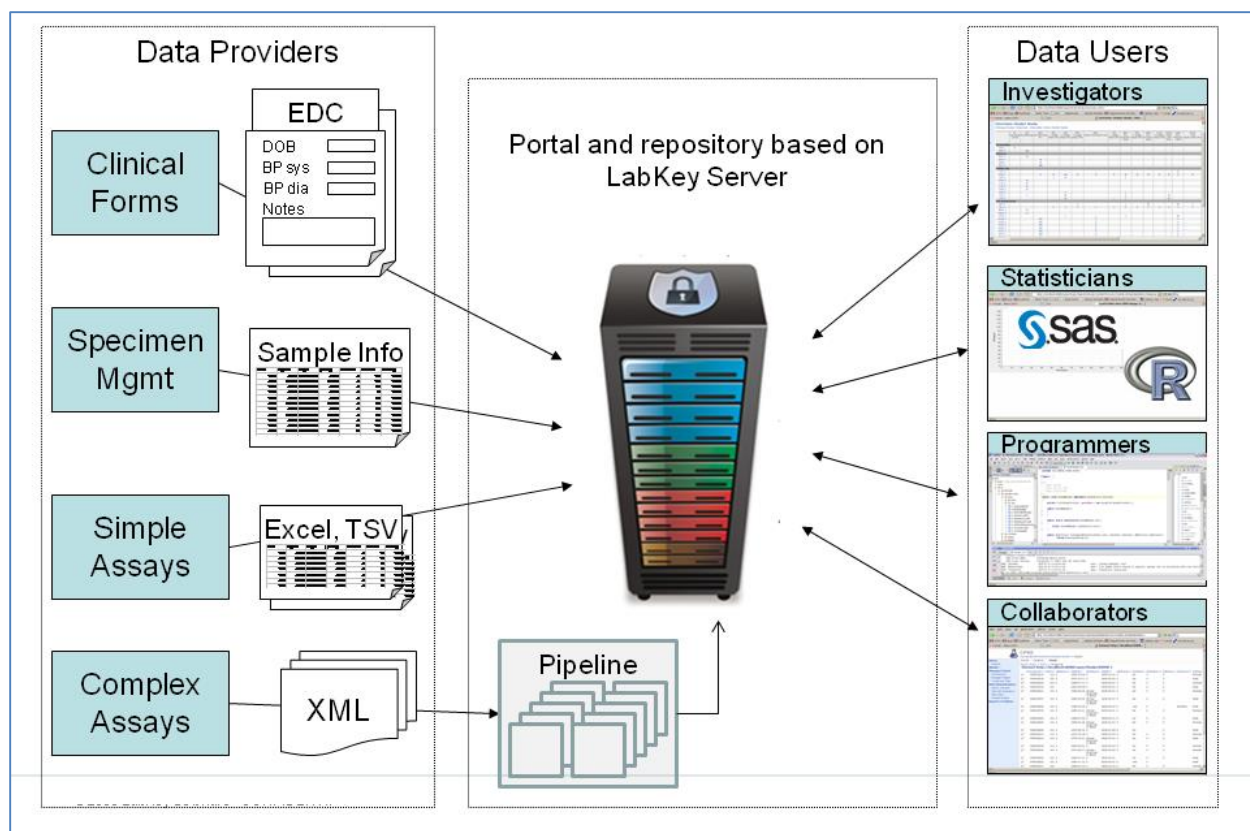


FIGURE 1.

Overview of the Rlabkey API

There are two basic types of functions available in Rlabkey. The "stateless" functions have names that begin with the prefix "labkey." The parameters of these stateless functions contain all the information required to read or update their target data. The `labkey.SelectRows` function is a good example:

```
rows <- labkey.selectRows(baseUrl="http://localhost:8080/labkey",
                          folderPath="/apisamples",
                          schemaName="lists",
                          queryName="AllTypes")
```

The parameters to `labkey.SelectRows` specify the server address, the project and folders within the server that are the context of the rows to select, plus the schema name and query name of the data to retrieve. Additional parameters allow specification of column lists, row filters and sort order of the returned data frame. Almost all of the parameters to the stateless functions are specified as strings.

In contrast, the session-based functions in Rlabkey version 2.1 are designed to be used in sets. A simple example that retrieves the same result as the function above is the following:

```
session <- getSession(baseUrl="http://localhost:8080/labkey",
                     folderPath="/apisamples")
schema <- getSchema(session, "lists")
rows <- getRows(session, schema$AllTypes)
```

There are two primary advantages to the session-style functions for an interactive R user:

- **Efficiency:** The user need only specify the baseUrl and folderPath once, and it is saved in the session object where it can be used to do multiple selects or updates against one or more data sets. The function names are shorter to save typing. Also, the arguments to the session-based are usually object-valued, allowing the R editor to provide statement completion choices as the user types.
- **Discovery:** The object returned by getSchema is a list of *all available* query objects within that schema on the server. These query objects in turn are a list of their fields and field attributes. These schema objects allow the R user to discover what is available on the server. Field attributes for "lookup" fields connect to other queries, effectively allowing the user to specify joins without knowing SQL.

All of the Rlabkey APIs can be used in an R Report script as well as on client machines.

User logins and passwords

To access most data on LabKey Server, the user must be logged in to LabKey and have the appropriate permissions to the data. (The exception is when the anonymous "Guest" user has been given access permission. See [security topics](#) for details.) Rlabkey connects using login information stored in a netrc file. The netrc file contains configuration and login information for the File Transfer Protocol client (ftp) and other programs such as CURL.

On a UNIX system this file should be named .netrc (dot netrc) and on Windows it should be named _netrc (underscore netrc). The file should be located in the user's home directory and the permissions on the file should be unreadable for everybody except the owner.

To create the _netrc on a Windows machine, first create an environment variable called 'HOME' that is set to your home directory (for example: c:\Users\<User-Name> on recent Windows releases) or any directory you want to use. In that directory, create a text file named _netrc.

The following three lines must be included in the .netrc or _netrc file either separated by white space (spaces, tabs, or newlines) or commas.

```
machine <remote-machine-name>
login <user-email>
password <user-password>
```

One example would be:

```
machine localhost
login peter@labkey.com
password mypass
```

Another example would be:

```
machine atlas.scharp.org login vobench@fhcrc.org password mypass
```

Multiple such blocks can exist in one file. Also, a netrc file is not used by functions in an R Report script.

Using Rlabkey to save analysis results

You must define an assay on the LabKey Server in order to use the saveResults function. The following steps go through the basic steps. (See the [Assays](#) topic on LabKey.org for more details.)

1. In R, write out a sample result table as a tab-separated text file without NA indicators, quotes or row names

```
write.table(topTable, "c:/temp/limmaResults.txt",  
            sep="\t", na="", quote=FALSE, row.names=FALSE)
```

2. Login to LabKey Server via a browser
3. If you don't already have a project to work in, define a new project where the results will be stored. Choose the Folder type of "Assay "

LabKey Software Foundation
Open Source Software for Scientists

1. Create Project
2. Users / Permissions
3. Project Settings

Create Project

Name:
miRNA Analysis

Folder Type:
☒ Assay
☐ Collaboration
☐ Dataspace
☐ Flow
☐ MS1
☐ MS2
☐ Microarray
☐ Panorama
☐ Study
☐ Custom
☐ Create From Template Folder

NEXT CANCEL

FIGURE 2

Leave the user/permission settings and project settings at their defaults for now.

4. On the home page (also called the "Dashboard") of the project in which the assay results will be stored, you will see an Assay List. On this Assay List, click the New Assay Design button.

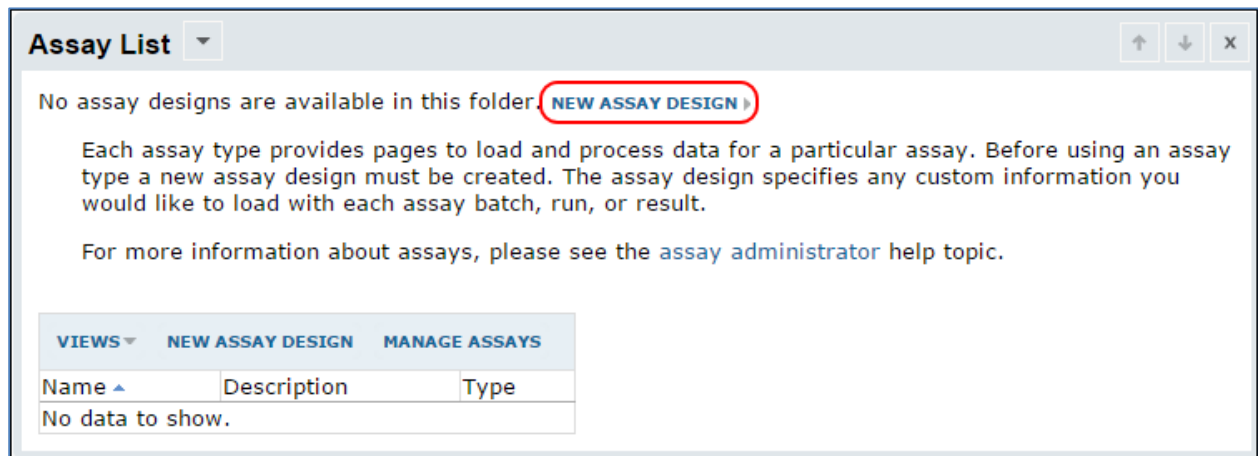


FIGURE 3

Then select the "General" assay type from the list and click Next:

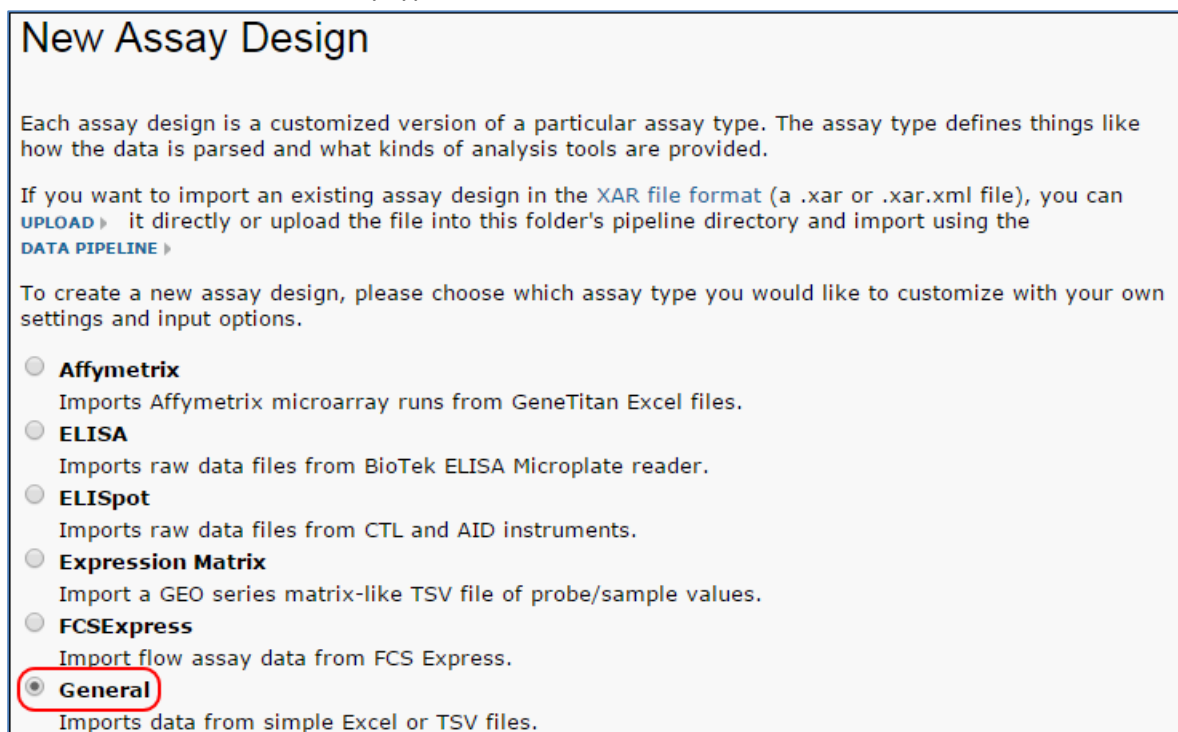


FIGURE 4

5. You now are presented with the General Assay Designer page.

General Assay Designer

SAVE & CLOSE **SAVE** **CANCEL**

Assay Properties

Name (Required)

Description

Auto-copy Data? ☐

Auto-copy Target?

Transform Scripts? **ADD SCRIPT**

Save Script Data? ☐

Editable Runs? ☐

Editable Results? ☐

Import in Background? ☐

Batch Fields

The user is prompted for batch properties once for each set of runs they import. The batch is a convenience to let users set properties that seldom change in one place and import many runs using them. This is the first step of the import process.

	Name	Label	Type
<input type="checkbox"/>	ParticipantVisitResolve	Participant Visit Resolve	xt (String)
<input type="checkbox"/>	TargetStudy	Target Study	xt (String)

ADD FIELD **IMPORT FIELDS** **EXPORT FIELDS** **INFER FIELDS FROM FILE**

Run Fields

The user is prompted to enter run level properties for each file they import. This is the second step of the import process.

No fields have been defined.

ADD FIELD **IMPORT FIELDS** **EXPORT FIELDS** **INFER FIELDS FROM FILE**

Data Fields

The user is prompted to enter data values for row of data associated with a run, typically done as uploading a file. This is part of the second step of the upload process.

	Name	Label	Type
<input type="checkbox"/>	SpecimenID	Specimen ID	xt (String)
<input type="checkbox"/>	ParticipantID	Participant ID	object/Participant (Strir
<input type="checkbox"/>	VisitID	Visit ID	mber (Double)
<input type="checkbox"/>	Date	Date	teTime

ADD FIELD **IMPORT FIELDS** **EXPORT FIELDS** **INFER FIELDS FROM FILE**

Figure 5

This page is broken into four sections, namely the properties of the assay definition itself, metadata properties that apply to a "batch" of one or more runs, metadata properties of individual runs, and then the data properties that are the rows of the data frames (result sets) to be uploaded. "Metadata" refers to property values describing the results as a whole, including any necessary information about how they were calculated that is not recorded in the data frame itself.

In our example we will be assuming a single run per batch, and we will record two example properties recording the versions of the R packages used, just for illustration.

- a. Give the assay a name and a description. Avoid spaces or special characters in the name.
- b. In the Batch Fields section:
 - i. Remove the pre-defined Batch properties by clicking the X next to each
 - ii. Press Add field to add text fields named LimmaVersion and RlabkeyVersion.

General Assay Designer

SAVE & CLOSE SAVE CANCEL

Assay Properties

Name (Required)

Description

Auto-copy Data? ☐

Auto-copy Target?

Transform Scripts?

Save Script Data? ☐

Editable Runs? ☐

Editable Results? ☐

Import in Background? ☐

Batch Fields

The user is prompted for batch properties once for each set of runs they import. The batch is a convenience to let users set properties that seldom change in one place and import many runs using them. This is the first step of the import process.

	Name	Label	Type	
<input type="checkbox"/>	LimmaVersion		kt (String)	<input type="button" value="Display"/>
<input type="checkbox"/>	RlabkeyVersion		kt (String)	<input type="button" value="Format"/>

Description

Figure 6

- c. In the Data Fields section:
 - i. Click the Infer Fields from File button (and click OK if you are prompted to delete existing fields).
 - ii. In the Infer Fields dialog click the Browse button and navigate to the file written out in step 1. Then press Submit. The bottom part of the page should now look like

Run Fields

The user is prompted to enter run level properties for each file they import. This is the second step of the import process.

Name	Label	Type
ProbeName		String
GeneName		String
logFC		Double
t		Double
P_Value	P.Value	Double
adj_P_Val	adj.P.Val	Double
B		Double

ADD FIELD IMPORT FIELDS EXPORT FIELDS INFER FIELDS FROM FILE

Display Format Validators Reporting Advanced

Description

URL ?

Shown In Display Modes

Grid Insert Update Details

Figure 7

If the column names of your data frame contained characters that aren't allowed in the Data Field names of an assay, they will be replaced by underscores as shown above. If the field names and types inferred by LabKey Server look correct, press Save & Close. Your newly defined assay should now appear in the Assay List:

Assay List

VIEWS NEW ASSAY DESIGN MANAGE ASSAYS 1 - 5 of 5

Name	Description	Type
LimmaTopTable	The assay stores the calculated results of the .miRNATwoGroupDifferentialExpression.	General
NAb High/Single Assay		TZM-bl Neutralization (NAb), High-throughput (Single Plate Dilution)
PhysicalExam		General

Figure 8

- You are now ready to save results sets from R. The highlighted names in the code block below correspond to the names you defined in the preceding steps. Note that there is a built-in property of Batch Name that you will likely want to define. In this code the batch name will be "Batch <timestamp>". Also note that the topTable rows being saved are only those which have a B value greater than 0.

```
S <- getSession(baseUrl="http://localhost:8080/labkey", folderPath="/miRNA Analysis")

## get the versions of the currently loaded limma and Rlabkey libraries,
## taken from installed.packages()
>ipdf <- data.frame(installed.packages(), stringsAsFactors=FALSE)
>bprops <- list(LimmaVersion=ipdf["limma", "Version"],
               RlabkeyVersion=ipdf["Rlabkey", "Version"])

## format as a list of lists
bpl <- list(name=paste("Batch ", as.character(date())), properties = bprops)
```



```
## call saveResults
>assaybatch <- saveResults(s, assayName="LimmaTopTable",
                           resultDataFrame=topTable[topTable$B>0, ],batchPropertyList= bpl)
```

After saving two assay results in this way, clicking on the name of your assay in the Assay List will bring up the Runs view listing the datasets you have saved. Click on the Assay Id link to see the results of a single run, or the "view results" link to see the data across multiple runs. These data can also be queried and retrieved from R. In this example, they will belong to a data set named "LimmaTopTable Data"

LimmaTableTop Runs

MANAGE ASSAY DESIGN

VIEW BATCHES

VIEW RUNS

VIEW RESULTS

VIEW COPY-TO-STUDY HISTORY

VIEWS

CHARTS

EXPORT

PRINT

PAGE SIZE

DELETE

SHOW RESULTS

COPY TO STUDY

IMPORT DATA

RE-IMPORT RUN

REPLACED FILTER

<input type="checkbox"/>	Flag	Assay Id	Created	Created By	Run Groups	Batch	Rlabkey Version
<input type="checkbox"/>		LimmaTableTop-2015-02-16-1048-2.tmp	2015-02-16 10:48	steveh		2015-02-16 batch	2.1.126
<input type="checkbox"/>		LimmaTableTop-2015-02-16-1048-1.tmp	2015-02-16 10:48	steveh		2015-02-16 batch	2.1.126
<input type="checkbox"/>		LimmaTableTop-2015-02-16-1048.tmp	2015-02-16 10:48	steveh		2015-02-16 batch	2.1.126

FIGURE 9

Organizing multiple assay results

You may want to create separate folders within a project to hold results for different users. Segregating results into user-specific folders can be useful for security restrictions (not all users are allowed to view other user's data), to allow QC review before consolidation, or for simple organizational reasons. For consistency across subfolders, all users should use the same Assay, defined at the project level as described above. To refer to this definition, users must be given read permissions at the project level.

To create subfolders, go to Admin -> Folder -> Management, select your project, and click Create Subfolder:

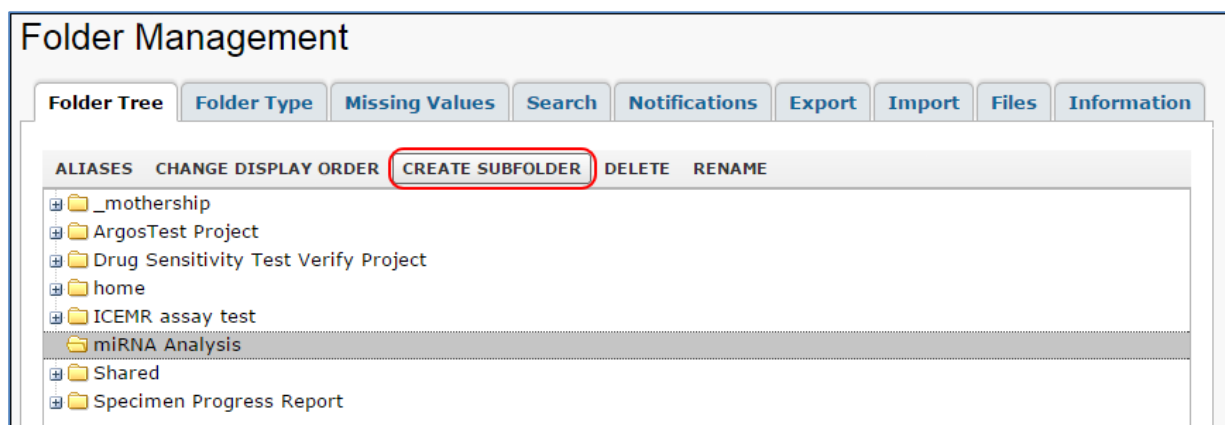


Figure 10

To save results to a folder named "johnd" within the project described in the previous section, the only difference in the R code is to use the full path to the subfolder:

```
S <- getSession(baseUrl="http://localhost:8080/labkey",  
               folderPath="/miRNA Analysis/johnd")
```

Troubleshooting

When querying data, if you encounter errors similar to the following, you must update your version of Rlabkey to the most recent version:

```
Error in function (type, msg, asError = TRUE) :  
error:1408F10B:SSL routines:SSL3_GET_RECORD:wrong version number
```

Or

```
Error in function (type, msg, asError = TRUE) :  
error:1411809D:SSL routines:SSL_CHECK_SERVERHELLO_TLSEXT:tls invalid  
ecpointformat list
```