Package 'EvidenceSynthesis'

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```
Type Package
Title Synthesizing Causal Evidence in a Distributed Research Network
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Maintainer Martijn Schuemie <schuemie@ohdsi.org>
Description Routines for combining causal effect estimates and study diagnostics across multi-
     ple data sites in a distributed study, without sharing patient-level data.
      Allows for normal and non-normal approximations of the data-
     site likelihood of the effect parameter.
SystemRequirements Java (>= 8)
Depends survival,
     R (>= 3.5.0)
Imports dplyr,
     ggplot2,
     ggdist,
     gridExtra,
     meta,
     EmpiricalCalibration,
     rJava,
     BeastJar (>= 10.5.1),
     Cyclops (>= 3.6.0),
     HDInterval,
     coda,
     rlang,
     methods
Suggests knitr,
     rmarkdown,
     testthat,
     sn,
     tidyr
License Apache License 2.0
URL https://ohdsi.github.io/EvidenceSynthesis/, https:
      //github.com/OHDSI/EvidenceSynthesis
BugReports https://github.com/OHDSI/EvidenceSynthesis/issues
VignetteBuilder knitr
```

2 Contents

RoxygenNote 7.3.2 Roxygen list(markdown = TRUE) Encoding UTF-8 LazyData true

Contents

approximateHierarchicalNormalPosterior
approximateLikelihood
approximateSimplePosterior
biasCorrectionInference
buildLabelReferences
computeBayesianMetaAnalysis
computeConfidenceInterval
computeFixedEffectMetaAnalysis
computeHierarchicalMetaAnalysis
constructDataModel
createApproximations
createSccsSimulationSettings
createSimulationSettings
customFunction
detectApproximationType
extractSourceSpecificEffects
fitBiasDistribution
generateBayesianHMAsettings
hermiteInterpolation
hmaLikelihoodList
likelihoodProfileLists
loadCyclopsLibraryForJava
ncLikelihoods
ooiLikelihoods
plotBiasCorrectionInference
plotBiasDistribution
plotCovariateBalances
plotEmpiricalNulls
plotLikelihoodFit
plotMcmcTrace
plotMetaAnalysisForest
plotPerDbMcmcTrace
plotPerDbPosterior
plotPosterior
plotPreparedPs
preparePsPlot
prepareSccsIntervalData
sequentialFitBiasDistribution
simulateMetaAnalysisWithNegativeControls
simulatePopulations
skewNormal
summarizeChain
supportsJava8

Index 43

approximateHierarchicalNormalPosterior

Approximate Bayesian posterior for hierarchical Normal model

Description

Approximate a Bayesian posterior from a set of Cyclops likelihood profiles under a hierarchical normal model using the Markov chain Monte Carlo engine BEAST.

Usage

```
approximateHierarchicalNormalPosterior(
   likelihoodProfiles,
   chainLength = 1100000,
   burnIn = 1e+05,
   subSampleFrequency = 100,
   effectPriorMean = 0,
   effectPriorSd = 0.5,
   nu0 = 1,
   sigma0 = 1,
   effectStartingValue = 0,
   precisionStartingValue = 1,
   seed = 1
)
```

Arguments

likelihoodProfiles

List of grid likelihoods profiled with Cyclops.

chainLength Number of MCMC iterations.

burnIn Number of MCMC iterations to consider as burn in.

subSampleFrequency

Subsample frequency for the MCMC.

effectPriorMean

Prior mean for global parameter

effectPriorSd Prior standard deviation for the global parameter

nu0 Prior "sample size" for precision (with precision ~ gamma(nu0/2, nu0*sigma0/2))

sigma0 Prior "variance" for precision (with precision ~ gamma(nu0/2, nu0*sigma0/2))

effectStartingValue

Initial value for global & local parameter

precisionStartingValue

Initial value for the precision

seed Seed for the random number generator.

Value

A data frame with the point estimates and 95% credible intervals for the global and local parameter, as well as the global precision. Attributes of the data frame contain the MCMC trace for diagnostics.

Examples

TBD

approximateLikelihood Approximate a likelihood function

Description

Approximate the likelihood function using a parametric (normal, skew-normal, or custom parametric), or grid approximation. The approximation does not reveal person-level information, and can therefore be shared among data sites. When counts are low, a normal approximation might not be appropriate.

Usage

```
approximateLikelihood(
  cyclopsFit,
  parameter = 1,
  approximation = "grid with gradients",
  bounds = c(log(0.1), log(10))
)
```

Arguments

Value

A vector of parameters of the likelihood approximation.

See Also

computeConfidenceInterval, computeFixedEffectMetaAnalysis, computeBayesianMetaAnalysis

```
# Simulate some data for this example:
populations <- simulatePopulations()

cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),
    data = populations[[1]],
    modelType = "cox"
)

cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)
approximation <- approximateLikelihood(cyclopsFit, "x")
approximation</pre>
```

```
# (Estimates in this example will vary due to the random simulation)
```

```
approximateSimplePosterior
```

Approximate simple Bayesian posterior

Description

Approximate a Bayesian posterior from a Cyclops likelihood profile and normal prior using the Markov chain Monte Carlo engine BEAST.

Usage

```
approximateSimplePosterior(
   likelihoodProfile,
   chainLength = 1100000,
   burnIn = 1e+05,
   subSampleFrequency = 100,
   priorMean = 0,
   priorSd = 0.5,
   startingValue = 0,
   seed = 1
)
```

Arguments

likelihoodProfile

Named vector containing grid likelihood data from Cyclops.

chainLength Number of MCMC iterations.

burnIn Number of MCMC iterations to consider as burn in.

subSampleFrequency

Subsample frequency for the MCMC.

priorMean Prior mean for the regression parameter

priorSd Prior standard deviation for the regression parameter

startingValue Initial state for regression parameter

seed Seed for the random number generator.

Value

A data frame with the point estimates and 95% credible intervals for the regression parameter. Attributes of the data frame contain the MCMC trace for diagnostics.

6 biasCorrectionInference

Examples

```
# Simulate some data for this example:
population <- simulatePopulations(createSimulationSettings(nSites = 1))[[1]]

# Fit a Cox regression at each data site, and approximate likelihood function:
cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),
    data = population,
    modelType = "cox"
)
cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)
likelihoodProfile <- approximateLikelihood(cyclopsFit, parameter = "x", approximation = "grid")

# Run MCMC
mcmcTraces <- approximateSimplePosterior(
    likelihoodProfile = likelihoodProfile,
    priorMean = 0, priorSd = 100
)

# Report posterior expectation
mean(mcmcTraces$theta)

# (Estimates in this example will vary due to the random simulation)</pre>
```

biasCorrectionInference

Bias Correction with Inference

Description

Perform Bayesian posterior inference regarding an outcome of interest with bias correction using negative control analysis. There is an option to not perform bias correction so that un-corrected results can be obtained.

Usage

```
biasCorrectionInference(
   likelihoodProfiles,
   ncLikelihoodProfiles = NULL,
   biasDistributions = NULL,
   priorMean = 0,
   priorSd = 1,
   numsamps = 10000,
   thin = 10,
   doCorrection = TRUE,
   seed = 1,
   ...
)
```

Arguments

likelihoodProfiles

A list of grid profile likelihoods for the outcome of interest.

biasCorrectionInference 7

ncLikelihoodProfiles

Likelihood profiles for the negative control outcomes. Must be a list of lists of profile likelihoods; if there is only one analysis period, then this must be a length-1 list, with the first item as a list all outcome-wise profile likelihoods.

biasDistributions

Pre-saved bias distribution(s), formatted as the output from fitBiasDistribution() or sequentialFitBiasDistribution(). If NULL, then ncLikelihoodProfiles

must be provided.

priorMean Prior mean for the effect size (log rate ratio).

priorSd Prior standard deviation for the effect size (log rate ratio).

numsamps Total number of MCMC samples needed.

thin Thinning frequency: how many iterations before another sample is obtained?

doCorrection Whether or not to perform bias correction; default: TRUE.

seed Seed for the random number generator.

... Arguments to be passed to sequentialFitBiasDistribution() to fit bias dis-

tributions if biasDistributions is NULL.

Value

A dataframe with five columns, including posterior median and mean of log RR effect size estimates, 95% credible intervals (ci95Lb and ci95Ub), posterior probability that log RR > 0 (p1), and the period or group ID (Id).

It is accompanied by the following attributes:

- samplesCorrected: all MCMC samples for the bias corrected log RR effect size estimate.
- samplesRaw: all MCMC samples for log RR effect size estimate, without bias correction.
- $\bullet \ \ \ \text{biasDistributions: the learned empirical bias distribution from negative control analysis.}$
- summaryRaw: a summary dataframe (same format as in the main result) without bias correction.
- corrected: a logical flag indicating if bias correction has been performed; = TRUE if doCorrection = TRUE.

See Also

approximateSimplePosterior, fitBiasDistribution

```
# load example data
data("ncLikelihoods")
data("ooiLikelihoods")

# perform sequential analysis with bias correction, using the t model
# NOT RUN
# bbcResults = biasCorrectionInference(ooiLikelihoods,
# ncLikelihoodProfiles = ncLikelihoods,
# robust = TRUE,
# seed = 42)

# check out analysis summary
# bbcResults
```

buildLabelReferences Build a list of references that map likelihood names to integer labels for later use

Description

Build a list of references that map likelihood names to integer labels for later use

Usage

```
buildLabelReferences(data)
```

Arguments

data

The likelihood data. Can be a single approximation, approximations from multiple sites, or (adaptive) grid profile likelihoods.

Examples

```
data("likelihoodProfileLists")
refLabs <- buildLabelReferences(likelihoodProfileLists)</pre>
```

computeBayesianMetaAnalysis

Compute a Bayesian random-effects meta-analysis

Description

Compute a Bayesian meta-analysis using the Markov chain Monte Carlo (MCMC) engine BEAST. A normal and half-normal prior are used for the mu and tau parameters, respectively, with standard deviations as defined by the priorSd argument.

Usage

```
computeBayesianMetaAnalysis(
  data,
  chainLength = 1100000,
  burnIn = 1e+05,
  subSampleFrequency = 100,
  priorSd = c(2, 0.5),
  alpha = 0.05,
  robust = FALSE,
  df = 4,
  seed = 1,
  showProgressBar = TRUE
)
```

Arguments

data A data frame containing either normal, skew-normal, custom parametric, or grid

likelihood data, with one row per database.

chainLength Number of MCMC iterations.

burnIn Number of MCMC iterations to consider as burn in.

subSampleFrequency

Subsample frequency for the MCMC.

priorSd A two-dimensional vector with the standard deviation of the prior for mu and

tau, respectively.

alpha The alpha (expected type I error) used for the credible intervals.

robust Whether or not to use a t-distribution model; default: FALSE.

df Degrees of freedom for the t-model, only used if robust is TRUE.

seed The seed for the random number generator.

showProgressBar

Showing a progress bar for MCMC?

Value

A data frame with the point estimates and 95% credible intervals for the mu and tau parameters (the mean and standard deviation of the distribution from which the per-site effect sizes are drawn). Attributes of the data frame contain the MCMC trace and the detected approximation type.

See Also

approximate Likelihood, compute Fixed Effect Meta Analysis

```
# Simulate some data for this example:
populations <- simulatePopulations()</pre>
# Fit a Cox regression at each data site, and approximate likelihood function:
fitModelInDatabase <- function(population) {</pre>
  cyclopsData \leftarrow Cyclops::createCyclopsData(Surv(time, y) \sim x + strata(stratumId),
    data = population,
    modelType = "cox"
  cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)</pre>
 approximation <- approximateLikelihood(cyclopsFit, parameter = "x", approximation = "custom")</pre>
  return(approximation)
approximations <- lapply(populations, fitModelInDatabase)</pre>
approximations <- do.call("rbind", approximations)</pre>
# At study coordinating center, perform meta-analysis using per-site approximations:
estimate <- computeBayesianMetaAnalysis(approximations)</pre>
estimate
# (Estimates in this example will vary due to the random simulation)
```

computeConfidenceInterval

Compute the point estimate and confidence interval given a likelihood function approximation

Description

Compute the point estimate and confidence interval given a likelihood function approximation

Usage

```
computeConfidenceInterval(approximation, alpha = 0.05)
```

Arguments

approximation An approximation of the likelihood function as fitted using the approximateLikelihood() function.

alpha The alpha (expected type I error).

Details

Compute the point estimate and confidence interval given a likelihood function approximation.

Value

A data frame containing the point estimate, and upper and lower bound of the confidence interval.

Examples

```
# Simulate some data for this example:
populations <- simulatePopulations()

cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),
    data = populations[[1]],
    modelType = "cox"
)
cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)
approximation <- approximateLikelihood(cyclopsFit, "x")
computeConfidenceInterval(approximation)</pre>
```

```
{\tt computeFixedEffectMetaAnalysis}
```

Compute a fixed-effect meta-analysis

Description

Compute a fixed-effect meta-analysis using a choice of various likelihood approximations.

Usage

```
computeFixedEffectMetaAnalysis(data, alpha = 0.05)
```

Arguments

data A data frame containing either normal, skew-normal, custom parametric, or grid

likelihood data. One row per database.

alpha The alpha (expected type I error) used for the confidence intervals.

Value

The meta-analytic estimate, expressed as the point estimate hazard ratio (rr), its 95 percent confidence interval (lb, ub), as well as the log of the point estimate (logRr), and the standard error (seLogRr).

See Also

approximateLikelihood, computeBayesianMetaAnalysis

Examples

```
# Simulate some data for this example:
populations <- simulatePopulations()</pre>
# Fit a Cox regression at each data site, and approximate likelihood function:
fitModelInDatabase <- function(population) {</pre>
  cyclopsData \leftarrow Cyclops::createCyclopsData(Surv(time, y) \sim x + strata(stratumId),
    data = population,
    modelType = "cox"
  cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)</pre>
 approximation <- approximateLikelihood(cyclopsFit, parameter = "x", approximation = "custom")
  return(approximation)
}
approximations <- lapply(populations, fitModelInDatabase)</pre>
approximations <- do.call("rbind", approximations)</pre>
# At study coordinating center, perform meta-analysis using per-site approximations:
computeFixedEffectMetaAnalysis(approximations)
# (Estimates in this example will vary due to the random simulation)
```

computeHierarchicalMetaAnalysis

Compute a Bayesian random-effects hierarchical meta-analysis

Description

Compute a Bayesian hierarchical meta-analysis (two-level model) to learn the global effect with bias correction via negative control outcomes analysis. Bayesian inference is performed using the Markov chain Monte Carlo (MCMC) engine BEAST. Normal priors are used for the global effect, outcome-specific effects, and data-source-specific effects; a half normal prior is used for the standard deviation; a gamma prior is used for the precision parameters.

12 constructDataModel

Usage

```
computeHierarchicalMetaAnalysis(
  data,
  settings = generateBayesianHMAsettings(),
  alpha = 0.05,
  seed = 1,
  showProgressBar = TRUE
)
```

Arguments

data A data frame containing either normal, skew-normal, custom parametric, or grid

likelihood data, with one row per database.

settings Model settings list generated by generateBayesianHMAsettings()

alpha The alpha (expected type I error) used for the credible intervals.

seed Seed for the random number generator.

showProgressBar

Showing a progress bar for MCMC?

Value

A data frame with the point estimates, 95% credible intervals and sample standard errors for the (de-biased) global main effect, the average outcome effect, the average data source effect, and precision of random errors. Attributes of the data frame contain the MCMC trace and the detected approximation type.

See Also

approximateLikelihood, computeBayesianMetaAnalysis

Examples

```
data("hmaLikelihoodList")
estimates <- EvidenceSynthesis::computeHierarchicalMetaAnalysis(
  data = hmaLikelihoodList,
  seed = 666
)</pre>
```

constructDataModel

Construct DataModel objects from approximate likelihood or profile likelihood data

Description

Construct DataModel objects from approximate likelihood or profile likelihood data

Usage

```
constructDataModel(data, labelReferences = NULL)
```

createApproximations 13

Arguments

data

The likelihood data. Can be a single approximation, approximations from multiple sites, or (adaptive) grid profile likelihoods.

labelReferences

Optional parameter that provides a reference list that maps string names to integer indices; only applies to "grid" or "adaptive grip" type of data.

Examples

```
data("likelihoodProfileLists")
dataModel <- constructDataModel(likelihoodProfileLists[[1]])</pre>
```

createApproximations

Create likelihood approximations from individual-trajectory data

Description

Create likelihood approximations from individual-trajectory data

Usage

```
createApproximations(populations, approximation)
```

Arguments

populations Individual-level population data approximation Type of approximation method

createSccsSimulationSettings

Create SCCS simulation settings

Description

Create an object specifying a simulation for the Self-Controlled Case Series (SCCS).

Usage

```
createSccsSimulationSettings(
  nSites = 5,
  n = 10000,
  atRiskTimeFraction = 0.1,
  timePartitions = 24,
  timeCovariates = 5,
  timeEffectSize = log(2),
  minBackgroundRate = 0.001,
  maxBackgroundRate = 0.01,
  rateRatio = 2,
  randomEffectSd = 0
)
```

Arguments

nSites Number of database sites to simulate.

Number of subjects per site. Either a single number, or a vector of length nSites.

atRiskTimeFraction

Fraction of patient time when at risk (exposed). Either a single number, or a

vector of length nSites.

 $time Partitions \ \ Number \ of \ time \ partitions \ for \ seasonal \ covariates. \ Either \ a \ single \ number, \ or \ a$

vector of length nSites.

timeCovariates Number of covariates to represent seasonality. Either a single number, or a

vector of length nSites.

timeEffectSize Strength of the seasonality effect. Either a single number, or a vector of length

nSites.

minBackgroundRate

Minimum background outcome rate. Either a single number, or a vector of

length nSites.

maxBackgroundRate

Maximum background outcome rate. Either a single number, or a vector of

length nSites.

rateRatio The incidence rate ratio.

randomEffectSd Standard deviation of the log(hazardRatio). Fixed effect if equal to 0.

Value

An object of type simulationSccsSettings, to be used in the simulatePopulations() function.

See Also

simulatePopulations

```
settings <- createSccsSimulationSettings(nSites = 1, rateRatio = 2)
populations <- simulatePopulations(settings)

# Fit a SCCS regression for the simulated data site:
cyclopsData <- Cyclops::createCyclopsData(
    y ~ a + x1 + x2 + x3 + x4 + x5 + strata(stratumId) + offset(log(time)),
    data = populations[[1]],
    modelType = "cpr"
)
cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)
coef(cyclopsFit)

# (Estimates in this example will vary due to the random simulation)</pre>
```

```
createSimulationSettings
```

Create simulation settings

Description

Create an object specifying a simulation. Currently only Cox proportional hazard models are supported.

Usage

```
createSimulationSettings(
  nSites = 5,
  n = 10000,
  treatedFraction = 0.2,
  nStrata = 10,
  minBackgroundHazard = 2e-07,
  maxBackgroundHazard = 2e-05,
  hazardRatio = 2,
  randomEffectSd = 0,
  siteEffects = 0
)
```

Arguments

nSites Number of database sites to simulate.

Number of subjects per site. Either a single number, or a vector of length nSites.

treatedFraction

Fraction of subjects that is treated. Either a single number, or a vector of length

nSites.

nStrata Number of strata per site. Either a single number, or a vector of length nSites.

 $\verb|minBackgroundHaz| ard$

Minimum background hazard. Either a single number, or a vector of length

nSites.

 ${\tt maxBackgroundHazard}$

Maximum background hazard. Either a single number, or a vector of length

nSites.

hazardRatio Hazard ratio.

randomEffectSd Standard deviation of the log(hazardRatio). Fixed effect if equal to 0.

siteEffects Fixed site effects (if assuming varying site-specific effects). Same effects if 0.

Value

An object of type simulationSettings, to be used in the simulatePopulations() function.

See Also

simulatePopulations

16 customFunction

Examples

```
settings <- createSimulationSettings(nSites = 1, hazardRatio = 2)
populations <- simulatePopulations(settings)

# Fit a Cox regression for the simulated data site:
cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),
    data = populations[[1]],
    modelType = "cox"
)
cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)
coef(cyclopsFit)

# (Estimates in this example will vary due to the random simulation)</pre>
```

customFunction

A custom function to approximate a log likelihood function

Description

A custom function to approximate a log likelihood function

Usage

```
customFunction(x, mu, sigma, gamma)
```

Arguments

x The log(hazard ratio) for which to approximate the log likelihood.

mu The position parameter.
sigma The scale parameter.
gamma The skew parameter.

Details

A custom parametric function designed to approximate the shape of the Cox log likelihood function. When gamma = 0 this function is the normal distribution.

Value

The approximate log likelihood for the given x.

```
customFunction(x = 0:3, mu = 0, sigma = 1, gamma = 0)
```

detectApproximationType

Detect the type of likelihood approximation based on the data format

Description

Detect the type of likelihood approximation based on the data format

Usage

```
detectApproximationType(data, verbose = TRUE)
```

Arguments

data The approximation data. Can be a single approximation, or approximations

from multiple sites.

verbose Should the detected type be communicated to the user?

Value

A character vector with one of the following values: "normal", "custom", "skew normal", "pooled", "grid", or "adaptive grid".

Examples

```
detectApproximationType(data.frame(logRr = 1, seLogRr = 0.1))
```

${\tt extractSourceSpecificEffects}$

Compute source-specific biases and bias-corrected estimates from hierarchical meta analysis results

Description

Extract source-specific biases and obtain bias-corrected estimates for each data source, given the results from computeHierarchicalMetaAnalysis().

Usage

```
extractSourceSpecificEffects(estimates, alpha = 0.05)
```

Arguments

estimates A data frame as output from the computeHierarchicalMetaAnalysis() func-

tion.

alpha The alpha (expected type I error) used for the credible intervals.

18 fitBiasDistribution

Value

A data frame with point estimates, 95% credible intervals and sample standard errors for the effect size after bias correction within each data source.

See Also

computeHierarchicalMetaAnalysis

fitBiasDistribution Fit Bias Distribution

Description

Learn an empirical distribution on estimation bias by simultaneously analyzing a large set of negative control outcomes by a Bayesian hierarchical model through MCMC. Analysis is based on a list of extracted likelihood profiles.

Usage

```
fitBiasDistribution(
   likelihoodProfiles,
   priorSds = c(2, 0.5),
   numsamps = 10000,
   thin = 10,
   minNCs = 5,
   robust = FALSE,
   df = 4,
   seed = 1
)
```

Arguments

likelihoodProfiles

A list of grid profile likelihoods regarding negative controls.

priorSds A two-dimensional vector with the standard deviation of the prior for the average

bias and the sd/scale parameter, respectively.

numsamps Total number of MCMC samples needed.

thin Thinning frequency: how many iterations before another sample is obtained?

minNCs Minimum number of negative controls needed to fit a bias distribution; default

(also recommended): 5.

robust Whether or not to use a t-distribution model; default: FALSE.

df Degrees of freedom for the t-model, only used if robust is TRUE.

seed Seed for the random number generator.

Value

A dataframe with three columns and numsamps number of rows. Column mean includes MCMC samples for the average bias, scale for the sd/scale parameter, and bias for predictive samples of the bias.

See Also

computeBayesianMetaAnalysis

Examples

```
# load example data
data("ncLikelihoods")

# fit a bias distributions by analyzing a set of negative control outcomes
# for example, for the 5th analysis period, and using the t model
# NOT RUN
# biasDistribution = fitBiasDistribution(ncLikelihoods[[5]], robust = TRUE)
```

generateBayesianHMAsettings

Generate settings for the Bayesian random-effects hierarchical metaanalysis model

Description

This function generates a settings list for fitting a Bayesian hierarchical meta-analysis model. See computeHierarchicalMetaAnalysis() for more details.

Usage

```
generateBayesianHMAsettings(
  primaryEffectPriorStd = 1,
  secondaryEffectPriorStd = 1,
  globalExposureEffectPriorMean = c(0),
  globalExposureEffectPriorStd = c(2),
  primaryEffectPrecisionPrior = c(1, 1),
  secondaryEffectPrecisionPrior = c(1, 1),
  errorPrecisionPrior = c(1, 1),
  errorPrecisionStartValue = 1,
  includeSourceEffect = TRUE,
  includeExposureEffect = TRUE,
  exposureEffectCount = 1,
  separateExposurePrior = FALSE,
  chainLength = 1100000,
  burnIn = 1e+05,
  subSampleFrequency = 100
)
```

Arguments

```
primaryEffectPriorStd
```

Standard deviation for the average outcome effect.

secondaryEffectPriorStd

Standard deviation for the average data-source effect.

20 hermiteInterpolation

globalExposureEffectPriorMean

Prior mean for the global main exposure effect; can be a multiple entry vector if there are multiple outcomes of interest

globalExposureEffectPriorStd

Prior standard deviation for the global main exposure effect; can be a multiple entry vector if there are multiple outcomes of interest

primaryEffectPrecisionPrior

Shape and scale for the gamma prior of the precision term in the random effects model (normal) for individual outcome effects.

secondaryEffectPrecisionPrior

Shape and scale for the gamma prior of the precision term in the random effects model (normal) for individual data-source effects.

errorPrecisionPrior

Shape and scale for the gamma prior of the precision term in the normal model for random errors.

errorPrecisionStartValue

Initial value for the error distribution's precision term.

includeSourceEffect

Whether or not to consider the data-source-specific (secondary) random effects. Default is TRUE.

include Exposure Effect

Whether or not to estimate the main effect of interest. Default is TRUE.

exposureEffectCount

Number of main outcomes of interest to estimate effect for? Default = 1 separateExposurePrior

Use a separable prior on the main exposure effect? Default is FALSE.

chainLength Number of MCMC iterations.

burnIn Number of MCMC iterations to consider as burn in.

subSampleFrequency

Subsample ("thinning") frequency for the MCMC.

Value

A list with all the settings to use in the computeHierarchicalMetaAnalysis() function.

See Also

compute Hierarchical Meta Analysis

hermiteInterpolation Cubic Hermite interpolation using both values and gradients to approximate a log likelihood function

Description

Cubic Hermite interpolation using both values and gradients to approximate a log likelihood function

Usage

hermiteInterpolation(x, profile)

hmaLikelihoodList 21

Arguments

x The log(hazard ratio) for which to approximate the log likelihood.

profile A profile as created with approximateLikelihood() with approximation =

"grid with gradients". This is a data frame with 3 columns: point, value,

and derivative, sorted by point.

Details

Performs spline interpolation using cubic Hermite polynomials (Catmull et al. 1974) between the points specified in the profile. We use linear extrapolation outside the points.

Value

The approximate log likelihood for the given x.

References

Catmull, Edwin; Rom, Raphael (1974), "A class of local interpolating splines", in Barnhill, R. E.; Riesenfeld, R. F. (eds.), Computer Aided Geometric Design, New York: Academic Press, pp. 317–326

Examples

```
profile <- data.frame(point = c(1.1, 2.1), value = c(1, 1), derivative = c(0.1, -0.1)) hermiteInterpolation(x = 0:3, profile = profile)
```

hmaLikelihoodList

Example profile likelihoods for hierarchical meta analysis with bias correction

Description

A list that contains profile likelihoods for two negative control outcomes and a synthetic outcome of interest, across four data sources. Each element of the list contains profile likelihoods for one outcome, where each row provides profile likelihood values (over a grid) from one data source.

Usage

hmaLikelihoodList

Format

An objects of class list; the list contains 3 dataframes, where each dataframe includes four rows of likelihood function values corresponding to the points in the column names.

```
data("hmaLikelihoodList")
hmaLikEx <- hmaLikelihoodList[[1]]
plot(as.numeric(hmaLikEx[2, ]) ~ as.numeric(names(hmaLikEx)))</pre>
```

likelihoodProfileLists

A bigger example of profile likelihoods for hierarchical meta analysis with bias correction

Description

A list that contains profile likelihoods for 10 negative control outcomes and an outcome of interest, across data sources. Each element of the list contains a named list of profile likelihoods for one outcome, where each element is a data frame that provides likelihood values over a grid of parameter values, the element name corresponding to data source name.

Usage

likelihoodProfileLists

Format

An objects of class list; the list contains 11 named lists, each list for one outcome. Each list contains data frames that record profile likelihoods from different data sources. The first 10 list corresponds to 10 negative control outcomes, whereas the last list the outcome of interest.

Examples

```
data("likelihoodProfileLists")
exLP <- likelihoodProfileLists[[1]][[1]]
plot(value ~ point, data = exLP)</pre>
```

loadCyclopsLibraryForJava

Load the Cyclops dynamic C++ library for use in Java

Description

Load the Cyclops dynamic C++ library for Markov chain Monte Carlo (MCMC) engine BEAST sampling.

Usage

```
loadCyclopsLibraryForJava(
  file = system.file("libs", "Cyclops.so", package = "Cyclops")
)
```

Arguments

file

The full system path to the Cyclops. so library

ncLikelihoods 23

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ncLikelihoods	E

Example profile likelihoods for negative control outcomes

Description

A list that contain profile likelihoods a large set of negative control outcomes. They are extracted from a real-world observational healthcare database, with the likelihoods profiled using adaptive grids using the Cyclops package.

Usage

ncLikelihoods

Format

An object of class list containing 12 lists, where each list includes several dataframes ith column point and value for adaptive grid profile likelihoods.

References

Schuemie et al. (2022). Vaccine safety surveillance using routinely collected healthcare data—an empirical evaluation of epidemiological designs. Frontiers in Pharmacology.

Examples

```
data("ncLikelihoods")
ncLikEx <- ncLikelihoods[["5"]][[1]]
plot(value ~ point, data = ncLikEx)</pre>
```

ooiLikelihoods

Example profile likelihoods for a synthetic outcome of interest

Description

A list that contain profile likelihoods for a synthetic outcome of interest. They are extracted from a real-world observational healthcare database, with the likelihoods profiled using adaptive grids using the Cyclops package.

Usage

ooiLikelihoods

Format

An objects of class list; the list contains 12 lists, where each list includes several dataframes with column point and value for adaptive grid profile likelihoods.

References

Schuemie et al. (2022). Vaccine safety surveillance using routinely collected healthcare data—an empirical evaluation of epidemiological designs. Frontiers in Pharmacology.

Examples

```
data("ooiLikelihoods")
ooiLikEx <- ooiLikelihoods[["5"]][[1]]
plot(value ~ point, data = ooiLikEx)</pre>
```

plotBiasCorrectionInference

Plot bias correction inference

Description

Plot bias correction inference

Usage

```
plotBiasCorrectionInference(
  bbcResult,
  type = "raw",
  ids = bbcResult$Id,
  limits = c(-3, 3),
  logScale = FALSE,
  numericId = TRUE,
  fileName = NULL
)
```

Arguments

bbcResult	A (sequential) analysis object generated by the biasCorrectionInference() function.
type	The type of plot. Must be one of c("corrected", "raw", "compare").
ids	IDs of the periods/groups to plot result for; default is all IDs.
limits	The limits on log RR for plotting.
logScale	Whether or not to show bias in log-RR; default FALSE (shown in RR).
numericId	Whether or not to treat Id as a numeric variable; default: TRUE.
fileName	Name of the file where the plot should be saved, for example 'plot.png'. See the function ggplot2::ggsave in the ggplot2 package for supported file formats.

Details

Plot empirical bias distributions learned from analyzing negative controls.

plotBiasDistribution 25

Value

A ggplot object. Use the ggplot2::ggsave function to save to file.

See Also

biasCorrectionInference

Examples

```
# Perform sequential analysis using Bayesian bias correction for this example:
data("ncLikelihoods")
data("ooiLikelihoods")
# bbcSequential = biasCorrectionInference(ooiLikelihoods, ncLikelihoodProfiles = ncLikelihoods)
# Plot it
# NOT RUN
# plotBiasCorrectionInference(bbcSequential, type = "corrected")
```

plotBiasDistribution Plot bias distributions

Description

Plot bias distributions

Usage

```
plotBiasDistribution(
  biasDist,
  limits = c(-2, 2),
  logScale = FALSE,
  numericId = TRUE,
  fileName = NULL
)
```

Arguments

biasDist	A bias distribution object generated by the fitBiasDistribution() or sequentialFitBiasDistribution.
limits	The lower and upper limits in log-RR to plot.
logScale	Whether or not to show bias in log-RR; default FALSE (shown in RR).
numericId	(For sequential or group case only) whether or not to treat Id as a numeric variable; default: TRUE.
fileName	Name of the file where the plot should be saved, for example 'plot.png'. See the function ggplot2::ggsave in the ggplot2 package for supported file formats.

Details

Plot empirical bias distributions learned from analyzing negative controls.

plotCovariateBalances

Value

26

A ggplot object. Use the ggplot2::ggsave function to save to file.

See Also

fitBiasDistribution, sequentialFitBiasDistribution

Examples

```
# Fit a bias distribution for this example:
data("ncLikelihoods")
# NOT RUN
# singleBiasDist = fitBiasDistribution(ncLikelihoods[[5]], seed = 1)
# Plot it
# NOT RUN
# plotBiasDistribution(singleBiasDist)
```

plotCovariateBalances Plot covariate balances

Description

Plots the covariate balance before and after matching for multiple data sources.

Usage

```
plotCovariateBalances(
  balances,
  labels,
  threshold = 0,
  beforeLabel = "Before matching",
  afterLabel = "After matching",
  fileName = NULL
)
```

Arguments

ba	lances A	A list of covariate balance	objects as created	d using the comp	uteCovariateBalance()	
----	----------	-----------------------------	--------------------	------------------	-----------------------	--

function in the CohortMethod package. Each balance object is expected to be a data frame with at least these two columns: beforeMatchingStdDiff and

 $after {\tt MatchingStdDiff}.$

labels A vector containing the labels for the various sources.

threshold Show a threshold value for the standardized difference.

beforeLabel Label for before matching / stratification / trimming.

afterLabel Label for after matching / stratification / trimming.

fileName Name of the file where the plot should be saved, for example 'plot.png'. See the

function ggplot2::ggsave for supported file formats.

plotEmpiricalNulls 27

Details

Creates a plot showing the covariate balance before and after matching. Balance distributions are displayed as box plots combined with scatterplots.

Value

A Ggplot object. Use the ggplot2::ggsave.

Examples

```
# Some example data:
balance1 <- data.frame(
   beforeMatchingStdDiff = rnorm(1000, 0.1, 0.1),
   afterMatchingStdDiff = rnorm(1000, 0.001)
)
balance2 <- data.frame(
   beforeMatchingStdDiff = rnorm(1000, 0.2, 0.1),
   afterMatchingStdDiff = rnorm(1000, 0.05)
)
balance3 <- data.frame(
   beforeMatchingStdDiff = rnorm(1000, 0, 0.1),
   afterMatchingStdDiff = rnorm(1000, 0, 0.03)
)
plotCovariateBalances(
   balances = list(balance1, balance2, balance3),
   labels = c("Site A", "Site B", "Site C")
)</pre>
```

plotEmpiricalNulls

Plot empirical null distributions

Description

Plot the empirical null distribution for multiple data sources.

Usage

```
plotEmpiricalNulls(
  logRr,
  seLogRr,
  labels,
  xLabel = "Relative risk",
  limits = c(0.1, 10),
  showCis = TRUE,
  fileName = NULL
)
```

28 plotLikelihoodFit

Arguments

logRr	A numeric vector of effect estimates for the negative controls on the log scale.
seLogRr	The standard error of the log of the effect estimates. Hint: often the standard error = $(\log(\text{lower bound 95 percent confidence interval}) - \log(\text{effect estimate}))/\text{qnorm}(0.025)$.
labels	A vector containing the labels for the various sources. Should be of equal length as logRr and seLogRr.
xLabel	The label on the x-axis: the name of the effect estimate.
limits	The limits of the effect size axis.
showCis	Show the 95 percent confidence intervals on the null distribution and distribution parameter estimates?
fileName	Name of the file where the plot should be saved, for example 'plot.png'. See the function ggplot2::ggsave() for supported file formats.

Details

Creates a plot showing the empirical null distributions. Distributions are shown as mean plus minus one standard deviation, as well as a distribution plot.

Value

A Ggplot object. Use the ggplot2::ggsave() function to save to file.

See Also

EmpiricalCalibration::fitNull, EmpiricalCalibration::fitMcmcNull

Examples

```
# Some example data:
site1 <- EmpiricalCalibration::simulateControls(n = 50, mean = 0, sd = 0.1, trueLogRr = 0)
site1$label <- "Site 1"
site2 <- EmpiricalCalibration::simulateControls(n = 50, mean = 0.1, sd = 0.2, trueLogRr = 0)
site2$label <- "Site 2"
site3 <- EmpiricalCalibration::simulateControls(n = 50, mean = 0.15, sd = 0.25, trueLogRr = 0)
site3$label <- "Site 3"
site3 <- rbind(site1, site2, site3)

plotEmpiricalNulls(logRr = sites$logRr, seLogRr = sites$seLogRr, labels = sites$label)</pre>
```

 ${\tt plotLikelihoodFit} \qquad \textit{Plot the likelihood approximation}$

Description

Plot the likelihood approximation

plotLikelihoodFit 29

Usage

```
plotLikelihoodFit(
  approximation,
  cyclopsFit,
  parameter = "x",
  logScale = TRUE,
  xLabel = "Hazard Ratio",
  limits = c(0.1, 10),
  fileName = NULL
)
```

Arguments

approximation An approximation of the likelihood function as fitted using the approximateLikelihood() function. cyclopsFit A model fitted using the Cyclops::fitCyclopsModel() function. The parameter in the cyclopsFit object to profile. parameter logScale Show the y-axis on the log scale? xLabel The title of the x-axis. limits The limits on the x-axis. fileName Name of the file where the plot should be saved, for example 'plot.png'. See the function ggplot2::ggsave in the ggplot2 package for supported file formats.

Details

Plots the (log) likelihood and the approximation of the likelihood. Allows for reviewing the approximation.

Value

A Ggplot object. Use the ggplot2::ggsave function to save to file.

```
# Simulate a single database population:
population <- simulatePopulations(createSimulationSettings(nSites = 1))[[1]]

# Approximate the likelihood:
cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),
    data = population,
    modelType = "cox"
)
cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)
approximation <- approximateLikelihood(cyclopsFit, parameter = "x", approximation = "custom")
plotLikelihoodFit(approximation, cyclopsFit, parameter = "x")</pre>
```

30 plotMcmcTrace

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Plot MCMC trace

Description

Plot MCMC trace

Usage

```
plotMcmcTrace(
   estimate,
   showEstimate = TRUE,
   dataCutoff = 0.01,
   fileName = NULL
)
```

Arguments

An object as generated using the computeBayesianMetaAnalysis() function.

ShowEstimate Show the parameter estimates (mode) and 95 percent confidence intervals?

This fraction of the data at both tails will be removed.

Name of the file where the plot should be saved, for example 'plot.png'. See the function ggplot2::ggsave in the ggplot2 package for supported file formats.

Details

Plot the samples of the posterior distribution of the mu and tau parameters. Samples are taken using Markov-chain Monte Carlo (MCMC).

Value

A Ggplot object. Use the ggplot2::ggsave function to save to file.

See Also

computeBayesianMetaAnalysis

```
# Simulate some data for this example:
populations <- simulatePopulations()

# Fit a Cox regression at each data site, and approximate likelihood function:
fitModelInDatabase <- function(population) {
   cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),
        data = population,
        modelType = "cox"
   )
   cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)
   approximation <- approximateLikelihood(cyclopsFit, parameter = "x", approximation = "custom")
   return(approximation)
}</pre>
```

```
approximations <- lapply(populations, fitModelInDatabase)
approximations <- do.call("rbind", approximations)

# At study coordinating center, perform meta-analysis using per-site approximations:
estimate <- computeBayesianMetaAnalysis(approximations)
plotMcmcTrace(estimate)</pre>
```

```
plotMetaAnalysisForest
```

Create a forest plot

Description

Creates a forest plot of effect size estimates, including the summary estimate.

Usage

```
plotMetaAnalysisForest(
  data,
  labels,
  estimate,
  xLabel = "Relative risk",
  summaryLabel = "Summary",
  limits = c(0.1, 10),
  alpha = 0.05,
  showLikelihood = TRUE,
  fileName = NULL
)
```

Arguments

data	A data frame containing either normal, skew-normal, custom parametric, or grid
	likelihood data. One row per database.

labels A vector of labels for the data sources.

estimate The meta-analytic estimate as created using either ['computeFixedEffectMeta-

Analysis()] or [computeBayesianMetaAnalysis()'] function.

xLabel The label on the x-axis: the name of the effect estimate.

summaryLabel The label for the meta-analytic estimate.

limits The limits of the effect size axis.

alpha The alpha (expected type I error).

showLikelihood Show the likelihood curve for each estimate?

fileName Name of the file where the plot should be saved, for example 'plot.png'. See the

function ggplot2::ggsave ifor supported file formats.

Details

Creates a forest plot of effect size estimates, including a meta-analysis estimate.

Value

A Ggplot object. Use the ggplot2::ggsave function to save to file.

Examples

```
# Simulate some data for this example:
populations <- simulatePopulations()</pre>
labels <- paste("Data site", LETTERS[1:length(populations)])</pre>
# Fit a Cox regression at each data site, and approximate likelihood function:
fitModelInDatabase <- function(population) {</pre>
  cyclopsData \leftarrow Cyclops::createCyclopsData(Surv(time, y) \sim x + strata(stratumId),
    data = population,
    modelType = "cox"
 cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)</pre>
 approximation <- approximateLikelihood(cyclopsFit, parameter = "x", approximation = "custom")</pre>
 return(approximation)
approximations <- lapply(populations, fitModelInDatabase)</pre>
approximations <- do.call("rbind", approximations)</pre>
# At study coordinating center, perform meta-analysis using per-site approximations:
estimate <- computeBayesianMetaAnalysis(approximations)</pre>
plotMetaAnalysisForest(approximations, labels, estimate)
# (Estimates in this example will vary due to the random simulation)
```

plotPerDbMcmcTrace

Plot MCMC trace for individual databases

Description

Plot MCMC trace for individual databases

Usage

```
plotPerDbMcmcTrace(
  estimate,
  showEstimate = TRUE,
  dataCutoff = 0.01,
  fileName = NULL
)
```

Arguments

An object as generated using the computeBayesianMetaAnalysis() function.

ShowEstimate Show the parameter estimates (mode) and 95 percent confidence intervals?

This fraction of the data at both tails will be removed.

Name of the file where the plot should be saved, for example 'plot.png'. See the

function ggplot2::ggsave in the ggplot2 package for supported file formats.

plotPerDbPosterior 33

Details

Plot the samples of the posterior distribution of the theta parameter (the estimated log hazard ratio) at each site. Samples are taken using Markov-chain Monte Carlo (MCMC).

Value

A Ggplot object. Use the ggplot2::ggsave function to save to file.

See Also

compute Bayesian Meta Analysis

Examples

```
# Simulate some data for this example:
populations <- simulatePopulations()</pre>
# Fit a Cox regression at each data site, and approximate likelihood function:
fitModelInDatabase <- function(population) {</pre>
  cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),</pre>
    data = population,
    modelType = "cox"
 cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)</pre>
 approximation <- approximateLikelihood(cyclopsFit, parameter = "x", approximation = "custom")
  return(approximation)
}
approximations <- lapply(populations, fitModelInDatabase)</pre>
approximations <- do.call("rbind", approximations)</pre>
# At study coordinating center, perform meta-analysis using per-site approximations:
estimate <- computeBayesianMetaAnalysis(approximations)</pre>
plotPerDbMcmcTrace(estimate)
```

 ${\tt plotPerDbPosterior}$

Plot posterior density per database

Description

Plot posterior density per database

Usage

```
plotPerDbPosterior(
  estimate,
  showEstimate = TRUE,
  dataCutoff = 0.01,
  fileName = NULL
)
```

34 plotPosterior

Arguments

estimate An object as generated using the computeBayesianMetaAnalysis() function. showEstimate Show the parameter estimates (mode) and 95 percent confidence intervals? This fraction of the data at both tails will be removed.

Name of the file where the plot should be saved, for example 'plot.png'. See the function ggplot2::ggsave in the ggplot2 package for supported file formats.

Details

Plot the density of the posterior distribution of the theta parameter (the estimated log hazard ratio) at each site.

Value

A Ggplot object. Use the ggplot2::ggsave function to save to file.

Examples

```
# Simulate some data for this example:
populations <- simulatePopulations()</pre>
# Fit a Cox regression at each data site, and approximate likelihood function:
fitModelInDatabase <- function(population) {</pre>
  cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),</pre>
    data = population,
    modelType = "cox"
  )
 cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)</pre>
 approximation <- approximateLikelihood(cyclopsFit, parameter = "x", approximation = "custom")</pre>
 return(approximation)
approximations <- lapply(populations, fitModelInDatabase)</pre>
approximations <- do.call("rbind", approximations)</pre>
# At study coordinating center, perform meta-analysis using per-site approximations:
estimate <- computeBayesianMetaAnalysis(approximations)</pre>
plotPerDbPosterior(estimate)
```

plotPosterior

Plot posterior density

Description

Plot posterior density

Usage

```
plotPosterior(
   estimate,
   showEstimate = TRUE,
   dataCutoff = 0.01,
   fileName = NULL
)
```

plotPreparedPs 35

Arguments

An object as generated using the computeBayesianMetaAnalysis() function.

ShowEstimate Show the parameter estimates (mode) and 95 percent confidence intervals?

This fraction of the data at both tails will be removed.

Name of the file where the plot should be saved, for example 'plot.png'. See the function ggplot2::ggsave in the ggplot2 package for supported file formats.

Details

Plot the density of the posterior distribution of the mu and tau parameters.

Value

A Ggplot object. Use the ggplot2::ggsave function to save to file.

See Also

computeBayesianMetaAnalysis

Examples

```
# Simulate some data for this example:
populations <- simulatePopulations()</pre>
# Fit a Cox regression at each data site, and approximate likelihood function:
fitModelInDatabase <- function(population) {</pre>
  cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),</pre>
    data = population,
    modelType = "cox"
  )
 cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)</pre>
 approximation <- approximateLikelihood(cyclopsFit, parameter = "x", approximation = "custom")</pre>
  return(approximation)
approximations <- lapply(populations, fitModelInDatabase)</pre>
approximations <- do.call("rbind", approximations)</pre>
# At study coordinating center, perform meta-analysis using per-site approximations:
estimate <- computeBayesianMetaAnalysis(approximations)</pre>
plotPosterior(estimate)
```

plotPreparedPs

Plot the propensity score distribution

Description

Plot the propensity score distribution

36 plotPreparedPs

Usage

```
plotPreparedPs(
   preparedPsPlots,
   labels,
   treatmentLabel = "Target",
   comparatorLabel = "Comparator",
   fileName = NULL
)
```

Arguments

preparedPsPlots

list of prepared propensity score data as created by the preparePsPlot() func-

tion.

labels A vector containing the labels for the various sources.

treatmentLabel A label to us for the treated cohort.

comparatorLabel

A label to us for the comparator cohort.

fileName

Name of the file where the plot should be saved, for example 'plot.png'. See the function ggplot2::ggsave for supported file formats.

Value

A ggplot object. Use the ggplot2::ggsave function to save to file in a different format.

See Also

preparePsPlot

```
# Simulate some data for this example:
treatment <- rep(0:1, each = 100)
propensityScore <- c(rnorm(100, mean = 0.4, sd = 0.25), rnorm(100, mean = 0.6, sd = 0.25))
data <- data.frame(treatment = treatment, propensityScore = propensityScore)
data <- data[data$propensityScore > 0 & data$propensityScore < 1, ]
preparedPlot <- preparePsPlot(data)

# Just reusing the same data three times for demonstration purposes:
preparedPsPlots <- list(preparedPlot, preparedPlot, preparedPlot)
labels <- c("Data site A", "Data site B", "Data site C")

plotPreparedPs(preparedPsPlots, labels)</pre>
```

preparePsPlot 37

preparePsPlot	Prepare to plot the propensity score distribution

Description

Prepare to plot the propensity (or preference) score distribution. It computes the distribution, so the output does not contain person-level data.

Usage

```
preparePsPlot(data, unfilteredData = NULL, scale = "preference")
```

Arguments

data	A data frame with at least the two columns described below
unfilteredData	To be used when computing preference scores on data from which subjects have already been removed, e.g. through trimming and/or matching. This data frame should have the same structure as data.
scale	The scale of the graph. Two scales are supported: scale = 'propensity' or scale = 'preference'. The preference score scale is defined by Walker et al. (2013).

Details

The data frame should have a least the following two columns:

• **treatment** (integer): Column indicating whether the person is in the treated (1) or comparator (0) group. - **propensityScore** (numeric): Propensity score.

Value

A data frame describing the propensity score (or preference score) distribution at 100 equally-spaced points.

References

Walker AM, Patrick AR, Lauer MS, Hornbrook MC, Marin MG, Platt R, Roger VL, Stang P, and Schneeweiss S. (2013) A tool for assessing the feasibility of comparative effectiveness research, Comparative Effective Research, 3, 11-20

See Also

```
plotPreparedPs
```

```
# Simulate some data for this example:
treatment <- rep(0:1, each = 100)
propensityScore <- c(rnorm(100, mean = 0.4, sd = 0.25), rnorm(100, mean = 0.6, sd = 0.25))
data <- data.frame(treatment = treatment, propensityScore = propensityScore)
data <- data[data$propensityScore > 0 & data$propensityScore < 1, ]</pre>
```

preparedPlot <- preparePsPlot(data)</pre>

prepareSccsIntervalData

Prepare SCCS interval data for pooled analysis

Description

Prepare SCCS interval data for pooled analysis

Usage

prepareSccsIntervalData(sccsIntervalData, covariateId)

Arguments

sccsIntervalData

An object of type SccsIntervalData as created using the createSccsIntervalData

 $function\ in\ the\ OHDSI\ Self Controlled Case Series\ package.$

covariateId

The ID of the covariate of interest, for which the estimate will be synthesized.

All other covariates will be considered nuisance variables.

Value

A tibble that can be used in the computeBayesianMetaAnalysis() function.

sequentialFitBiasDistribution

Fit Bias Distribution Sequentially or in Groups

Description

Learn empirical bias distributions sequentially or in groups; for each sequential step or analysis group, bias distributions is learned by by simultaneously analyzing a large set of negative control outcomes by a Bayesian hierarchical model through MCMC.

Usage

```
sequentialFitBiasDistribution(LikelihoodProfileList, ...)
```

Arguments

LikelihoodProfileList

A list of lists, each of which is a set of grid profile likelihoods regarding negative controls, indexed by analysis period ID for sequential analyses or group ID for group analyses.

... Arguments passed to the fitBiasDistribution() function.

Value

A (long) dataframe with four columns. Column mean includes MCMC samples for the average bias, scale for the sd/scale parameter, bias for predictive samples of the bias, and Id for the period ID or group ID.

See Also

fitBiasDistribution, computeBayesianMetaAnalysis

Examples

```
# load example data
data("ncLikelihoods")

# fit bias distributions over analysis periods
# NOT RUN
# biasDistributions = sequentialFitBiasDistribution(ncLikelihoods, seed = 42)
```

 ${\it simulate Meta Analysis With Negative Controls}$

Simulate survival data across a federated data network, with negative control outcomes as well.

Description

A function to simulate patient-level survival data for a hypothetical exposure, with simulated bias and data-source-specific random effects. Patient-level data for negative control outcomes are simulated as well to reflect systematic error.

Usage

```
simulateMetaAnalysisWithNegativeControls(
  meanExposureEffect = log(2),
  meanBias = 0.5,
  biasStd = 0.2,
  meanSiteEffect = 0,
   siteEffectStd = 0.1,
  mNegativeControls = 10,
  nSites = 10,
  sitePop = 2000,
  seed = 42,
  ...
)
```

Arguments

meanExposureEffect

Average exposure effect; has to be on the log-scale

meanBias Average bias for the bias distribution

biasStd Standard deviation for the bias distribution

40 simulatePopulations

meanSiteEffect Average of the data-source-specific effects (typically should be zero) siteEffectStd Standard deviation for data-source-specific effects mNegativeControls

Number of negative control outcomes

nSites Number of data sources

sitePop Population size per data source

seed Random seed

... Arguments that will be passed to other functions

See Also

computeHierarchicalMetaAnalysis

simulatePopulations S

Simulate survival data for multiple databases

Description

Simulate survival data for multiple databases

Usage

```
simulatePopulations(settings = createSimulationSettings())
```

Arguments

settings

Either an object of type simulationSettings, created by the createSimulationSettings() function or an object of type sccsSimulationSettings as created by the createSccsSimulationSetunction.

Value

A object of class simulation, which is a list of population data frames. Depending on the type of simulation, the data frames have different columns: Cox simulations will have the columns rowId, stratumId, x, time, and y. SCCS simulations will have the columns stratumId, a, x1...xN, time, and y.

```
settings <- createSimulationSettings(nSites = 1, hazardRatio = 2)
populations <- simulatePopulations(settings)

# Fit a Cox regression for the simulated data site:
cyclopsData <- Cyclops::createCyclopsData(Surv(time, y) ~ x + strata(stratumId),
    data = populations[[1]],
    modelType = "cox"
)
cyclopsFit <- Cyclops::fitCyclopsModel(cyclopsData)
coef(cyclopsFit)

# (Estimates in this example will vary due to the random simulation)</pre>
```

skewNormal 41

skewNormal	The skew normal function to approximate a log likelihood function

Description

The skew normal function to approximate a log likelihood function

Usage

```
skewNormal(x, mu, sigma, alpha)
```

Arguments

x The log(hazard ratio) for which to approximate the log likelihood.

mu The position parameter.
sigma The scale parameter.
alpha The skew parameter.

Details

The skew normal function. When alpha = 0 this function is the normal distribution.

Value

The approximate log likelihood for the given x.

References

Azzalini, A. (2013). The Skew-Normal and Related Families. Institute of Mathematical Statistics Monographs. Cambridge University Press.

Examples

```
skewNormal(x = 0:3, mu = 0, sigma = 1, alpha = 0)
```

summarizeChain Utility function to summarize MCMC samples (posterior mean, median, HDI, std, etc.)

Description

Utility function to summarize MCMC samples (posterior mean, median, HDI, std, etc.)

Usage

```
summarizeChain(chain, alpha = 0.05)
```

Arguments

chain A vector of posterior samples from MCMC.

alpha Alpha level for the credible interval.

42 supportsJava8

supportsJava8

Determine if Java virtual machine supports Java

Description

Tests Java virtual machine (JVM) java.version system property to check if version >= 8.

Usage

```
supportsJava8()
```

Value

Returns TRUE if JVM supports Java >= 8.

Examples

supportsJava8()

Index

* datasets	hermiteInterpolation, 20
hmaLikelihoodList, 21	hmaLikelihoodList, 21
likelihoodProfileLists, 22	
ncLikelihoods, 23	likelihoodProfileLists, 22
ooiLikelihoods, 23	loadCyclopsLibraryForJava, 22
approximateHierarchicalNormalPosterior,	ncLikelihoods, 23
approximateLikelihood, 4, 9, 11, 12	ooiLikelihoods, 23
approximateLikelihood(), 10, 29	plotBiasCorrectionInference, 24
approximateSimplePosterior, 5, 7	plotBiasDistribution, 25
biasCorrectionInference, 6, 25	plotCovariateBalances, 26
biasCorrectionInference(), 24	plotEmpiricalNulls, 27
buildLabelReferences, 8	plotLikelihoodFit, 28
bullulabelikerer enees, o	plotMcmcTrace, 30
computeBayesianMetaAnalysis, 4, 8, 11, 12,	plotMetaAnalysisForest, 31
19, 30, 33, 35, 39	plotPerDbMcmcTrace, 32
computeBayesianMetaAnalysis(), 30, 32,	plotPerDbPosterior, 33
34, 35	plotPosterior, 34
computeConfidenceInterval, 4, 10	plotPreparedPs, 35, 37
computeFixedEffectMetaAnalysis, 4, 9, 10	preparePsPlot, 36,37
computeHierarchicalMetaAnalysis, 11, 18,	<pre>preparePsPlot(), 36</pre>
20, 40	prepareSccsIntervalData,38
<pre>constructDataModel, 12</pre>	
createApproximations, 13	sequentialFitBiasDistribution, 26, 38
createSccsSimulationSettings, 13	sequentialFitBiasDistribution(), 7, 25
<pre>createSccsSimulationSettings(), 40</pre>	simulateMetaAnalysisWithNegativeControls,
createSimulationSettings, 15	39
<pre>createSimulationSettings(), 40</pre>	simulatePopulations, 14, 15, 40
customFunction, 16	simulatePopulations(), 14, 15
Cyclops::fitCyclopsModel(), 4, 29	skewNormal, 41
	summarizeChain, 41
<pre>detectApproximationType, 17</pre>	supportsJava8, 42
EmpiricalCalibration::fitMcmcNull, 28	
EmpiricalCalibration::fitNull, 28	
extractSourceSpecificEffects, 17	
fitBiasDistribution, 7, 18, 26, 39	
fitBiasDistribution(), 7, 25, 38	
${\tt generateBayesianHMAsettings}, 19$	
ggplot2::ggsave, 24–27, 29–36	
ggplot2::ggsave(), 28	