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Jerzy Pawlowski jp3900@nyu.edu

NYU Tandon School of Engineering

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Vector and Matrix Calculus

Let **v** and **w** be vectors, with $\mathbf{v} = \{v_i\}_{i=1}^{i=n}$, and let $\mathbb{1}$ be the unit vector, with $\mathbb{1} = \{1\}_{i=1}^{i=n}$.

Then the inner product of \mathbf{v} and \mathbf{w} can be written as $\mathbf{v}^T\mathbf{w} = \mathbf{w}^T\mathbf{v} = \sum_{i=1}^n v_i w_i$.

We can then express the sum of the elements of \mathbf{v} as the inner product: $\mathbf{v}^T \mathbb{1} = \mathbb{1}^T \mathbf{v} = \sum_{i=1}^n v_i$.

And the sum of squares of \mathbf{v} as the inner product: $\mathbf{v}^T\mathbf{v} = \sum_{i=1}^n v_i^2$.

Let \mathbb{A} be a matrix, with $\mathbb{A} = \{A_{ij}\}_{i,j=1}^{i,j=n}$.

Then the inner product of matrix \mathbb{A} with vectors \mathbf{v} and \mathbf{w} can be written as:

$$\mathbf{v}^T \mathbb{A} \mathbf{w} = \mathbf{w}^T \mathbb{A}^T \mathbf{v} = \sum_{i,j=1}^n A_{ij} v_i w_j$$

The derivative of a scalar variable with respect to a vector variable is a vector, for example:

$$\frac{d(\mathbf{v}^T \mathbb{1})}{d\mathbf{v}} = d_v[\mathbf{v}^T \mathbb{1}] = d_v[\mathbb{1}^T \mathbf{v}] = \mathbb{1}^T$$
$$d_v[\mathbf{v}^T \mathbf{w}] = d_v[\mathbf{w}^T \mathbf{v}] = \mathbf{w}^T$$
$$d_v[\mathbf{v}^T \mathbb{A} \mathbf{w}] = \mathbf{w}^T \mathbb{A}^T$$
$$d_v[\mathbf{v}^T \mathbb{A} \mathbf{v}] = \mathbf{v}^T \mathbb{A} + \mathbf{v}^T \mathbb{A}^T$$

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Eigenvectors and Eigenvalues of Matrices

The vector w is an eigenvector of the matrix \mathbb{A} , if it satisfies the eigenvalue equation:

$$\mathbb{A} w = \lambda w$$

Where λ is the eigenvalue corresponding to the eigenvector w.

The number of *eigenvalues* of a matrix is equal to its dimension.

Real symmetric matrices have real *eigenvalues*, and their *eigenvectors* are orthogonal to each other.

The eigenvectors can be normalized to 1.

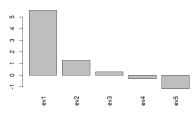
The eigenvectors form an orthonormal basis in which the matrix \mathbb{A} is diagonal.

The function eigen() calculates the eigenvectors and eigenvalues of numeric matrices.

An excellent interactive visualization of *eigenvectors* and *eigenvalues* is available here:

http://setosa.io/ev/eigenvectors-and-eigenvalues/

Eigenvalues of a real symmetric matrix



- > # Create a random real symmetric matrix
- > matrixv <- matrix(runif(25), nc=5)
- > matrixv <- matrixv + t(matrixv)
- > # Calculate the eigenvalues and eigenvectors
 > eigend <- eigen(matrixy)</pre>
- > eigenvec <- eigend\$vectors
- > eigenvec <- eigendavector > dim(eigenvec)
- > # Plot eigenvalues
- > barplot(eigend\$values, xlab="", ylab="", las=3,
- + names.arg=pasteO("ev", 1:NROW(eigend\$values)),
- + main="Eigenvalues of a real symmetric matrix")

Eigen Decomposition of Matrices

Real symmetric matrices have real *eigenvalues*, and their *eigenvectors* are orthogonal to each other.

The eigenvectors form an orthonormal basis in which the matrix $\mathbb A$ is diagonal:

$$\Sigma = \mathbb{O}^T \mathbb{A} \mathbb{O}$$

Where Σ is a diagonal matrix containing the eigenvalues of matrix \mathbb{A} , and \mathbb{O} is an orthogonal matrix of its eigenvectors, with $\mathbb{O}^T\mathbb{O}=\mathbb{1}$.

Any real symmetric matrix \mathbb{A} can be decomposed into a product of its eigenvalues and its eigenvectors (the eigen decomposition):

$$\mathbb{A} = \mathbb{O} \Sigma \mathbb{O}^T$$

The eigen decomposition expresses a matrix as the product of a rotation, followed by a scaling, followed by the inverse rotation.

- > # eigenvectors form an orthonormal basis
- > round(t(eigenvec) %*% eigenvec, digits=4)
- > # Diagonalize matrix using eigenvector matrix
- > round(t(eigenvec) %*% (matrixv %*% eigenvec), digits=4)
- > eigend\$values
- > # eigen decomposition of matrix by rotating the diagonal matrix
 > matrixe <- eigenvec %*% (eigend\$values * t(eigenvec))</pre>
- > # Create diagonal matrix of eigenvalues
- > # diagmat <- diag(eigend\$values)
- > # matrixe <- eigenvec %*% (diagmat %*% t(eigenvec))
 - > all.equal(matrixv, matrixe)

Orthogonal matrices represent rotations in hyperspace, and their inverse is equal to their transpose: $\mathbb{O}^{-1} = \mathbb{O}^T$

The diagonal matrix Σ represents a scaling (stretching) transformation proportional to the eigenvalues.

The %*% operator performs inner (scalar) multiplication of vectors and matrices.

Inner multiplication multiplies the rows of one matrix with the columns of another matrix, so that each pair produces a single number.

Positive Definite Matrices

Matrices with positive eigenvalues are called positive definite matrices.

Matrices with non-negative eigenvalues are called positive semi-definite matrices (some of their eigenvalues may be zero).

An example of *positive definite* matrices are the covariance matrices of linearly independent variables.

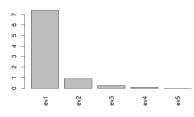
But the covariance matrices of linearly dependent variables have some *eigenvalues* equal to zero, in which case they are *singular*, and only *positive semi-definite*.

All covariance matrices are *positive semi-definite* and all *positive semi-definite* matrices are the covariance matrix of some multivariate distribution.

Matrices which have some *eigenvalues* equal to zero are called *singular* (degenerate) matrices.

For any real matrix \mathbb{A} , the matrix $\mathbb{A}^T \mathbb{A}$ is *positive* semi-definite.

Eigenvalues of positive semi-definite matrix



- > # Create a random positive semi-definite matrix
- > matrixv <- matrix(runif(25), nc=5)
 > matrixv <- t(matrixv) %*% matrixv</pre>
- > # Calculate the eigenvalues and eigenvectors
- > eigend <- eigen(matrixy)
- > eigend\$values
- > # Plot eigenvalues
- > barplot(eigend\$values, las=3, xlab="", ylab="",
- + names.arg=pasteO("ev", 1:NROW(eigend\$values)),
- + main="Eigenvalues of positive semi-definite matrix")

Singular Value Decomposition (SVD) of Matrices

The Singular Value Decomposition (SVD) is a generalization of the eigen decomposition of square matrices.

The SVD of a rectangular matrix $\mathbb A$ is defined as the factorization:

$$\mathbb{A}=\mathbb{U}\,\Sigma\,\mathbb{V}^{^{T}}$$

Where $\mathbb U$ and $\mathbb V$ are the left and right singular matrices, and Σ is a diagonal matrix of singular values.

If $\mathbb A$ has $\mathbb m$ rows and $\mathbb n$ columns and if $(\mathbb m > \mathbb n)$, then $\mathbb U$ is an $(\mathbb m \times \mathbb n)$ rectangular matrix, Σ is an $(\mathbb m \times \mathbb n)$ diagonal matrix, and $\mathbb V$ is an $(\mathbb m \times \mathbb n)$ orthogonal matrix, and if $(\mathbb m < \mathbb n)$ then the dimensions are: $(\mathbb m \times \mathbb m)$. $(\mathbb m \times \mathbb m)$. and $(\mathbb m \times \mathbb m)$.

The left $\mathbb U$ and right $\mathbb V$ singular matrices consist of columns of *orthonormal* vectors, so that $\mathbb U^T\mathbb U=\mathbb V^T\mathbb V=\mathbb 1.$

In the special case when $\mathbb A$ is a square matrix, then $\mathbb U=\mathbb V$, and the SVD reduces to the eigen decomposition.

The function svd() performs Singular Value Decomposition (SVD) of a rectangular matrix, and returns a list of three elements: the singular values, and the matrices of left-singular vectors and the right-singular vectors.

- \gt # Perform singular value decomposition
- > matrixv <- matrix(rnorm(50), nc=5)
 > sydec <- syd(matrixv)</pre>
- > svdec <- svd(matrixv)
 > # Recompose matrixv from SVD mat_rices
- > all.equal(matrixv, svdec\$u %*% (svdec\$d*t(svdec\$v)))
- > # Columns of U and V are orthonormal
- > round(t(svdec\$u) %*% svdec\$u, 4)
 > round(t(svdec\$v) %*% svdec\$v, 4)

The Left and Right Singular Matrices

The left $\mathbb U$ and right $\mathbb V$ singular matrices define rotation transformations into a coordinate system where the matrix $\mathbb A$ becomes diagonal:

$$\Sigma=\mathbb{U}^{T}\mathbb{A}\mathbb{V}$$

The columns of $\mathbb U$ and $\mathbb V$ are called the singular vectors, and they are only defined up to a reflection (change in sign), i.e. if vec is a singular vector, then so is -vec.

The left singular matrix $\mathbb U$ forms the $\it eigenvectors$ of the matrix $\mathbb A\mathbb A^T.$

The right singular matrix V forms the *eigenvectors* of the matrix A^TA .

```
> # Dimensions of left and right matrices
> nrows <- 6 ; ncols <- 4
> # Calculate left matrix
> leftmat <- matrix(runif(nrows^2), nc=nrows)
> eigend <- eigen(crossprod(leftmat))
> leftmat <- eigend$vectors[, 1:ncols]
> # Calculate right matrix and singular values
> rightmat <- matrix(runif(ncols^2), nc=ncols)
> eigend <- eigen(crossprod(rightmat))
> rightmat <- eigend$vectors
> singval <- sort(runif(ncols, min=1, max=5), decreasing=TRUE)
> # Compose rectangular matrix
> matrixv <- leftmat %*% (singval * t(rightmat))
> # Perform singular value decomposition
> sydec <- syd(matrixy)
> # Recompose matrixv from SVD
> all.equal(matrixv, svdec$u %*% (svdec$d*t(svdec$v)))
> # Compare SVD with matrixv components
> all.equal(abs(svdec$u), abs(leftmat))
> all.equal(abs(svdec$v), abs(rightmat))
> all.equal(svdec$d, singval)
> # Eigen decomposition of matrixv squared
> retsq <- matrixv %*% t(matrixv)
> eigend <- eigen(retsq)
> all.equal(eigend$values[1:ncols], singval^2)
> all.equal(abs(eigend$vectors[, 1:ncols]), abs(leftmat))
> # Eigen decomposition of matrixv squared
> retsq <- t(matrixv) %*% matrixv
> eigend <- eigen(retsq)
> all.equal(eigend$values, singval^2)
> all.equal(abs(eigend$vectors), abs(rightmat))
```

Inverse of Symmetric Square Matrices

The inverse of a square matrix $\mathbb A$ is defined as a square matrix $\mathbb A^{-1}$ that satisfies the equation:

$$\mathbb{A}^{-1}\mathbb{A}=\mathbb{A}\mathbb{A}^{-1}=\mathbb{1}$$

Where 1 is the identity matrix.

The inverse \mathbb{A}^{-1} of a *symmetric* square matrix \mathbb{A} can also be expressed as the product of the inverse of its *eigenvalues* (Σ) and its *eigenvectors* (\mathbb{O}):

$$\mathbb{A}^{-1} = \mathbb{O} \, \Sigma^{-1} \, \mathbb{O}^T$$

But *singular* (degenerate) matrices (which have some *eigenvalues* equal to zero) don't have an inverse.

The inverse of non-symmetric matrices can be calculated using Singular Value Decomposition (SVD).

The function solve() solves systems of linear equations, and also inverts square matrices.

> # Create a random positive semi-definite matrix
> matrixv <- matrix(runif(25), nc=5)
 matrixv <- t(matrixv) %*% matrixv
> # Calculate the inverse of matrixv
 invmat <- solve(a=matrixv)
> # Multiply inverse with matrix
> round(invmat %*% matrixv, 4)
> round(matrixv) %*% invmat, 4)
> # Calculate the eigenvalues and eigenvectors
 eigend <- eigen(matrixv)
> # eigenvec <- eigen(matrixv)
> # Calculate inverse from eigen decomposition
> # Calculate inverse from eigen decomposition
> inveigen <- eigenvec %*% (t(eigenvec) / eigend\$values)
> all.equal(invmat, inveigen)
> # Becompose diagonal matrix with inverse of eigenvalues

> # inveigen <- eigenvec %*% (diagmat %*% t(eigenvec))

> # diagmat <- diag(1/eigend\$values)

Generalized Inverse of Rectangular Matrices

The generalized inverse of an (m x n) rectangular matrix \mathbb{A} is defined as an $(n \times m)$ matrix \mathbb{A}^{-1} that satisfies the equation:

$$\mathbb{A}\mathbb{A}^{-1}\mathbb{A} = \mathbb{A}$$

The generalized inverse matrix \mathbb{A}^{-1} can be expressed as a product of the inverse of its singular values (Σ) and its left and right singular matrices (\mathbb{U} and \mathbb{V}):

$$\mathbb{A}^{-1} = \mathbb{V} \, \Sigma^{-1} \, \mathbb{U}^T$$

The generalized inverse \mathbb{A}^{-1} can also be expressed as the Moore-Penrose pseudo-inverse:

$$\mathbb{A}^{-1} = (\mathbb{A}^T \mathbb{A})^{-1} \mathbb{A}^T$$

In the case when the inverse matrix \mathbb{A}^{-1} exists, then the pseudo-inverse matrix simplifies to the inverse: $(\mathbb{A}^T \mathbb{A})^{-1} \mathbb{A}^T = \mathbb{A}^{-1} (\mathbb{A}^T)^{-1} \mathbb{A}^T = \mathbb{A}^{-1}$

The function MASS::ginv() calculates the generalized inverse of a matrix.

- > # Random rectangular matrix: nrows > ncols
- > nrows <- 6 ; ncols <- 4 > matrixv <- matrix(runif(nrows*ncols), nc=ncols)
- > # Calculate generalized inverse of matrixv
- > invmat <- MASS::ginv(matrixv)
- > round(invmat %*% matrixv, 4)
- > all.equal(matrixv, matrixv %*% invmat %*% matrixv)
- > # Random rectangular matrix: nrows < ncols > nrows <- 4 ; ncols <- 6
- > matrixv <- matrix(runif(nrows*ncols), nc=ncols)
- > # Calculate generalized inverse of matrixv
- > invmat <- MASS::ginv(matrixv)
- > all.equal(matrixv, matrixv %*% invmat %*% matrixv)
- > round(matrixv %*% invmat, 4) > round(invmat %*% matrixv, 4)
- > # Perform singular value decomposition
- > svdec <- svd(matrixv)
- > # Calculate generalized inverse from SVD
- > invsvd <- svdec\$v %*% (t(svdec\$u) / svdec\$d)
- > all.equal(invsvd, invmat)
- > # Calculate Moore-Penrose pseudo-inverse
- > invmp <- MASS::ginv(t(matrixv) %*% matrixv) %*% t(matrixv)
- > all.equal(invmp, invmat)

Regularized Inverse of Singular Matrices

Singular matrices have some singular values equal to zero, so they don't have an inverse matrix which satisfies the equation: $\mathbb{A}\mathbb{A}^{-1}\mathbb{A}=\mathbb{A}$

But if the singular values that are equal to zero are removed, then a regularized inverse for singular matrices can be specified by:

$$\mathbb{A}^{-1} = \mathbb{V}_n \, \Sigma_n^{-1} \, \mathbb{U}_n^T$$

Where \mathbb{U}_n , \mathbb{V}_n and Σ_n are the SVD matrices with the rows and columns corresponding to zero singular values removed.

- > # Create a random singular matrix
- > # More columns than rows: ncols > nrows
- > nrows <- 4 ; ncols <- 6
- > matrixv <- matrix(runif(nrows*ncols), nc=ncols)
- > matrixv <- t(matrixv) %*% matrixv
- > # Perform singular value decomposition
- > sydec <- syd(matrixy)
- > # Incorrect inverse from SVD because of zero singular values
- > invsvd <- svdec\$v %*% (t(svdec\$u) / svdec\$d)
- > # Inverse property doesn't hold
- > all.equal(matrixv, matrixv %*% invsvd %*% matrixv)

- > # Set tolerance for determining zero singular values
- > precv <- sqrt(.Machine\$double.eps) > # Check for zero singular values
- > round(svdec\$d, 12)
- > notzero <- (svdec\$d > (precv*svdec\$d[1]))
- > # Calculate regularized inverse from SVD
- > invsvd <- svdec\$v[, notzero] %*%
- (t(svdec\$u[, notzero]) / svdec\$d[notzero])
- > # Verify inverse property of matrixv
- > all.equal(matrixv, matrixv %*% invsvd %*% matrixv) > # Calculate regularized inverse using MASS::ginv()
- > invmat <- MASS::ginv(matrixv)
- > all.equal(invsvd, invmat)
- > # Calculate Moore-Penrose pseudo-inverse
- > invmp <- MASS::ginv(t(matrixv) %*% matrixv) %*% t(matrixv)
- > all.equal(invmp, invmat)

Diagonalizing the Inverse of Singular Matrices

The left-singular matrix $\mathbb U$ combined with the right-singular matrix $\mathbb V$ define a rotation transformation into a coordinate system where the matrix $\mathbb A$ becomes diagonal:

$$\Sigma = \mathbb{U}^T \mathbb{A} \mathbb{V}$$

The generalized inverse of singular matrices doesn't satisfy the equation: $\mathbb{A}^{-1}\mathbb{A}=\mathbb{A}\mathbb{A}^{-1}=\mathbb{I}$, but if it's rotated into the same coordinate system where \mathbb{A} is diagonal, then we have:

$$\mathbb{U}^{T}(\mathbb{A}^{-1}\mathbb{A})\,\mathbb{V}=\mathbb{1}_{n}$$

So that $\mathbb{A}^{-1}\mathbb{A}$ is diagonal in the same coordinate system where \mathbb{A} is diagonal.

- > # Diagonalize the unit matrix > unitmat <- matrixv %*% invmat
- > round(unitmat, 4)
- > round(matrixv %*% invmat, 4)
- > round(t(svdec\$u) %*% unitmat %*% svdec\$v, 4)

Solving Linear Equations Using solve()

A system of linear equations can be defined as:

$$\mathbb{A} x = b$$

Where \mathbb{A} is a matrix, b is a vector, and \mathbf{x} is the unknown vector.

The solution of the system of linear equations is equal to:

$$x = \mathbb{A}^{-1}b$$

Where \mathbb{A}^{-1} is the *inverse* of the matrix \mathbb{A} .

The function solve() solves systems of linear equations, and also inverts square matrices.

The %*% operator performs inner (scalar) multiplication of vectors and matrices.

Inner multiplication multiplies the rows of one matrix with the columns of another matrix, so that each pair produces a single number:

- > # Define a square matrix
- > matrixv <- matrix(c(1, 2, -1, 2), nc=2)
- > vectorv <- c(2, 1)
- > # Calculate the inverse of matrixv
- > invmat <- solve(a=matrixv)
- > invmat %*% matrixv
- > # Calculate solution using inverse of matrixv
- > solutionv <- invmat %*% vectorv
- > matrixv %*% solutionv
- > # Calculate solution of linear system
- > solutionv <- solve(a=matrixv, b=vectorv)
- > matrixv %*% solutionv

Fast Matrix Inverse Using C++

The Armadillo C++ functions can be several times faster than R functions - even those that are compiled from C++ code.

That's because the Armadillo C++ library calls routines optimized for fast numerical calculations.

The package RcppArmadillo allows calling from R the high-level Armadillo C++ linear algebra library.

The C++ Armadillo function arma::inv() calculates the matrix inverse several times faster than the function solve()

The function solve() calculates the matrix inverse several times faster than the function MASS::ginv().

// Rcpp header with information for C++ compiler

```
> # Create a random matrix
> matrixv <- matrix(rnorm(100), nc=10)
> # Calculate the matrix inverse using solve()
> invmatr <- solve(a=matrixv)
> round(invmatr %*% matrixv. 4)
> # Compile the C++ file using Rcpp
> Rcpp::sourceCpp(file="/Users/jerzy/Develop/Rcpp/test_fun.cpp")
> # Calculate the matrix inverse using C++
> invmat <- calc invmat(matrixv)
> all.equal(invmat, invmatr)
> # Compare the speed of RcppArmadillo with R code
> library(microbenchmark)
> summary(microbenchmark(
    ginv=MASS::ginv(matrixv),
    solve=solve(matrixv),
    cpp=calc_invmat(matrixv),
    times=10))[, c(1, 4, 5)]
```

```
// [[Rcpp::depends(RcppArmadillo)]]
#include <RcppArmadillo.h> // include RcppArmadillo header file
using namespace arma; // use Armadillo C++ namespace
// [[Rcpp::export]]
arma::mat calc_invmat(arma::mat& matrixv) {
  return arma::inv(matrixv);
} // end calc_invmat
```

Cholesky Decomposition

The Cholesky decomposition of a positive definite matrix A is defined as:

$$\mathbb{A}=\mathbb{L}^T\mathbb{L}$$

Where \mathbb{L} is an upper triangular matrix with positive diagonal elements.

The matrix \mathbb{L} can be considered the square root of \mathbb{A} .

The vast majority of random positive semi-definite matrices are also positive definite.

The function chol() calculates the Cholesky decomposition of a positive definite matrix.

The functions chol2inv() and chol() calculate the inverse of a positive definite matrix two times faster than solve()

```
> # Create large random positive semi-definite matrix
> matrixv <- matrix(runif(1e4), nc=100)
> matrixv <- t(matrixv) %*% matrixv
> # Calculate the eigen decomposition
> eigend <- eigen(matrixv)
> eigenval <- eigend$values
> eigenvec <- eigend$vectors
> # Set tolerance for determining zero singular values
> precv <- sqrt(.Machine$double.eps)
> # If needed convert to positive definite matrix
> notzero <- (eigenval > (precv*eigenval[1]))
> if (sum(!notzero) > 0) {
    eigenval[!notzero] <- 2*precv
    matrixv <- eigenvec %*% (eigenval * t(eigenvec))
+ } # end if
> # Calculate the Cholesky matrixv
> cholmat <- chol(matrixy)
> cholmat[1:5, 1:5]
> all.equal(matrixv, t(cholmat) %*% cholmat)
> # Calculate inverse from Cholesky
> invchol <- chol2inv(cholmat)
> all.equal(solve(matrixy), invchol)
> # Compare speed of Cholesky inversion
> library(microbenchmark)
> summary(microbenchmark(
```

times=10))[, c(1, 4, 5)] # end microbenchmark summary

solve=solve(matrixy). cholmat=chol2inv(chol(matrixv)),

Simulating Correlated Returns Using Cholesky Matrix

The *Cholesky* decomposition of a covariance matrix can be used to simulate correlated *Normal* returns following the given covariance matrix: $\mathbb{C} = \mathbb{L}^T \mathbb{L}$

Let $\mathbb R$ be a matrix with columns of *uncorrelated* returns following the *Standard Normal* distribution.

The correlated returns \mathbb{R}_c can be calculated from the uncorrelated returns \mathbb{R} by multiplying them by the Cholesky matrix \mathbb{L} :

$$\mathbb{R}_c = \mathbb{L}^T \mathbb{R}$$

- > # Calculate random covariance matrix
 > covmat <- matrix(runif(25), nc=5)</pre>
- > covmat <- t(covmat) %*% covmat
- > # Calculate the Cholesky matrix
- > cholmat <- chol(covmat)
- > cholmat
- > # Simulate random uncorrelated returns
- > nassets <- 5 > nrows <- 10000
- > retsp <- matrix(rnorm(nassets*nrows), nc=nassets)
- > # Calculate correlated returns by applying Cholesky
- > retscorr <- retsp %*% cholmat
- > # Calculate covariance matrix
- > covmat2 <- crossprod(retscorr) /(nrows-1)
- > all.equal(covmat, covmat2)

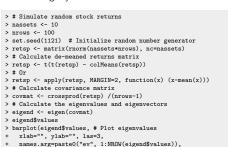
Eigenvalues of Singular Covariance Matrices

If \mathbb{R} is a matrix of returns (with zero mean) for a portfolio of k stocks (columns), over n time periods (rows), then the sample covariance matrix is equal to:

$$\mathbb{C} = \mathbb{R}^T \mathbb{R}/(n-1)$$

If the number of rows is less than the number of stocks, then the returns are collinear, and the sample covariance matrix is singular, with some eigenvalues equal to zero.

The function crossprod() performs inner (scalar) multiplication, exactly the same as the \%*% operator, but it is slightly faster.



90.0 0.04 0.02 00.0 15 20 > # Calculate the eigenvalues and eigenvectors

Smallest eigenvalue of covariance matrix

as function of number of returns

- > # as function of number of returns > ndata <- ((nassets/2):(2*nassets)) > eigenval <- sapply(ndata, function(x) { retsp <- retsp[1:x,] retsp <- apply(retsp, MARGIN=2, function(y) (y - mean(y))) covmat <- crossprod(retsp) / (x-1) min(eigen(covmat)\$values) + }) # end sapply > plot(y=eigenval, x=ndata, t="1", xlab="", ylab="", lwd=3, col="bl
 - main="Smallest eigenvalue of covariance matrix
 - as function of number of returns")

Regularized Inverse of Singular Covariance Matrices

The regularization technique allows calculating the inverse of singular covariance matrices while reducing the effects of statistical noise.

If the number of time periods of returns is less than the number of assets (columns), then the covariance matrix of returns is singular, and some of its eigenvalues are zero, so it doesn't have an inverse.

The regularized inverse \mathbb{C}_n^{-1} is calculated by removing the higher order eigenvalues that are almost zero, and keeping only the first n eigenvalues:

$$\mathbb{C}_n^{-1} = \mathbb{O}_n \, \Sigma_n^{-1} \, \mathbb{O}_n^T$$

Where Σ_n and \mathbb{O}_n are matrices with the higher order eigenvalues and eigenvectors removed.

The function MASS::ginv() calculates the regularized inverse of a matrix

- > # Create rectangular matrix with collinear columns
- > matrixv <- matrix(rnorm(10*8), nc=10) > # Calculate covariance matrix
- > covmat <- cov(matrixv)
- > # Calculate inverse of covmat error
- > invmat <- solve(covmat)
- > # Calculate regularized inverse of covmat
- > invmat <- MASS::ginv(covmat)
- > # Verify inverse property of matrixv
- > all.equal(covmat, covmat %*% invmat %*% covmat)
- > # Perform eigen decomposition > eigend <- eigen(covmat)
- > eigenvec <- eigend\$vectors
- > eigenval <- eigend\$values
- > # Set tolerance for determining zero singular values
- > precv <- sqrt(.Machine\$double.eps)
- > # Calculate regularized inverse matrix
- > notzero <- (eigenval > (precv * eigenval[1]))
- > invreg <- eigenvec[, notzero] %*%
- (t(eigenvec[, notzero]) / eigenval[notzero]) > # Verify that invmat is same as invreg
- > all.equal(invmat, invreg)

The Bias-Variance Tradeoff of the Regularized Inverse

Removing the very small higher order eigenvalues can also be used to reduce the propagation of statistical noise and improve the signal-to-noise ratio.

Removing a larger number of eigenvalues further reduces the noise, but it increases the bias of the covariance matrix.

This is an example of the bias-variance tradeoff.

Even though the *regularized* inverse \mathbb{C}_n^{-1} does not satisfy the matrix inverse property, its out-of-sample forecasts may be more accurate than those using the actual inverse matrix.

The parameter dimax specifies the number of eigenvalues used for calculating the *regularized* inverse of the covariance matrix of returns.

The optimal value of the parameter dimax can be determined using backtesting (cross-validation).

- > # Calculate regularized inverse matrix using cutoff > dimax <- 3
- > invmat <- eigenvec[, 1:dimax] %*%
 - (t(eigenvec[, 1:dimax]) / eigend\$values[1:dimax])
- > # Verify that invmat is same as invreg
- > all.equal(invmat, invreg)

> invmat <- solve(covshrink)

Shrinkage Estimator of Covariance Matrices

The estimates of the covariance matrix suffer from statistical noise, and those noise are magnified when the covariance matrix is inverted.

In the *shrinkage* technique the covariance matrix \mathbb{C}_s is estimated as a weighted sum of the sample covariance estimator \mathbb{C} plus a target matrix \mathbb{T} :

$$\mathbb{C}_s = (1 - \alpha) \, \mathbb{C} + \alpha \, \mathbb{T}$$

The target matrix \mathbb{T} represents an estimate of the covariance matrix subject to some constraint, such as that all the correlations are equal to each other.

The shrinkage intensity α determines the amount of shrinkage that is applied, with $\alpha=1$ representing a complete shrinkage towards the target matrix.

The *shrinkage* estimator reduces the estimate variance at the expense of increasing its bias (known as the *bias-variance tradeoff*).

```
> # Create a random covariance matrix
> set.seed(1121)
> matriv <- matrix(rnorm(5e2), nc=5)
> covant <- cov(matrixv)
> cornat <- cor(matrixv)
> stdev <- sqrt(diag(covant))
> # Calculate target matrix
> cornean <- mean(cornat[upper.tri(cormat)])
> targetmat <- matrix(cornean, nr=NROW(covmat), nc=NCOL(covmat))
> diag(targetmat) <- 1
> targetmat <- tictargetmat * stdev) * stdev)
> # Calculate shrinkage covariance matrix
> alpha <- 0.5
> covshrink <- (1-alpha)*covmat + alpha*targetmat
> # Calculate inverse matrix
```

Recursive Matrix Inverse

The inverse of a square matrix A can be calculated approximately using the recursive Schulz formula:

$$\mathbb{A}_{i+1}^{-1} \leftarrow 2\mathbb{A}_{i}^{-1} - \mathbb{A}_{i}^{-1}\mathbb{A}\mathbb{A}_{i}^{-1}$$

The Schulz formula requires a good initial value for the inverse matrix \mathbb{A}_1^{-1} or else the recursion diverges.

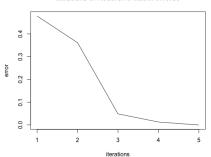
If the initial inverse matrix \mathbb{A}_1^{-1} is very close to the actual inverse \mathbb{A}^{-1} , then the Schulz formula produces a very good approximation with just a few iterations.

The Schulz formula is useful for updating the inverse when the matrix A changes only slightly. For example, for updating the inverse of the covariance matrix as it changes slowly over time.

The super-assignment operator "<<-" modifies variables in the enclosing environment in which the function was defined (lexical scoping).

- > # Create a random matrix
- > matrixv <- matrix(rnorm(100), nc=10) > # Calculate the inverse of matrixy
- > invmat <- solve(a=matrixv)
- > # Multiply inverse with matrix
- > round(invmat %*% matrixv, 4) > # Calculate the initial inverse
- > invmatr <- invmat + matrix(rnorm(100, sd=0.1), nc=10)
- > # Calculate the approximate recursive inverse of matrixv
- > invmatr <- (2*invmatr invmatr %*% matrixv %*% invmatr)
- > # Calculate the sum of the off-diagonal elements > sum((invmatr %*% matrixy)[upper.tri(matrixy)])

Iterations of Recursive Matrix Inverse



- > # Calculate the recursive inverse of matrixv in a loop
- > invmatr <- invmat + matrix(rnorm(100, sd=0.1), nc=10)
- > iterv <- sapply(1:5, function(x) {
- + # Calculate the recursive inverse of matrixy
- invmatr <<- (2*invmatr invmatr %*% matrixv %*% invmatr)
- + # Calculate the sum of the off-diagonal elements
- sum((invmatr %*% matrixv)[upper.tri(matrixv)])
- + }) # end sapply

Machine Learning

- > # Plot the iterations
- > plot(x=1:5, y=iterv, t="1", xlab="iterations", ylab="error",
 - main="Iterations of Recursive Matrix Inverse")

draft: Principal Components of Two Stocks

The scaled returns of XLP and VTI can be expressed as linear combinations of two orthogonal principal components:

The first principal component can be returns of *XLP* and *VTI* are highly correlated because they both share a common factor of market returns.

```
> retsp <- scale(na.omit(rutils::etfenv$returns
            [, as.character(formulav)[-1]]))
> crossprod(retsp) / NROW(retsp)
> w1 <- sqrt(0.5); w2 <- w1
> foo <- matrix(c(w1, w2, -w2, w1), nc=2)
> t(foo) %*% foo
> # bar <- retsp %*% t(solve(foo))
> (t(bar) %*% bar) / NROW(bar)
> covmat <- function(retsp, anglev=0) {
+ w1 <- cos(anglev)</p>
+ w2 <- sin(anglev)</p>
+ matrixv <- matrix(c(w1, -w2, w2, w1), nc=2)
+ pcav <- retsp %*% t(matrixv)
  (t(pcav) %*% pcav) / NROW(pcav)
+ } # end covmat
> bar <- covmat(retsp, anglev=pi/4)
> crossprod(retsp) / NROW(retsp)
> (t(bar) %*% bar) / NROW(bar)
> angles <- seq(0, pi/2, by=pi/24)
> covmat <- sapply(angles, function(angley)
   covmat(retsp, anglev=anglev)[1, 1])
> plot(x=angles, y=covmat, t="1")
> optiml <- optimize(
+ f=function(anglev)
```

> # Perform PCA for two stocks

```
> # Plot scatterplot of returns
> plot(formulav, data=vutils::etfenv$returns,
+ main="Regression XLP ~ VTI")
> # Add regression line
> abline(regmod, lwd=2, col="red")
```

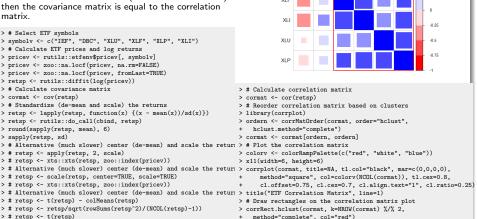
Covariance Matrix of ETF Returns

The covariance matrix \mathbb{C} , of the return matrix \mathbf{r} is given bv:

$$\mathbb{C} = \frac{\left(\mathbf{r} - \overline{\mathbf{r}}\right)^T \left(\mathbf{r} - \overline{\mathbf{r}}\right)}{n-1}$$

If the returns are standardized (de-meaned and scaled) then the covariance matrix is equal to the correlation matrix

```
> # Select ETF symbols
> symbolv <- c("IEF", "DBC", "XLU", "XLF", "XLP", "XLI")
> # Calculate ETF prices and log returns
> pricev <- rutils::etfenv$pricev[, symbolv]
> pricev <- zoo::na.locf(pricev, na.rm=FALSE)
> pricev <- zoo::na.locf(pricev, fromLast=TRUE)
> retsp <- rutils::diffit(log(pricev))
> # Calculate covariance matrix
> covmat <- cov(retsp)
> # Standardize (de-mean and scale) the returns
> retsp <- lapply(retsp, function(x) {(x - mean(x))/sd(x)})
> retsp <- rutils::do_call(cbind, retsp)
> round(sapply(retsp, mean), 6)
```



ETF Correlation Matrix

> # retsp <- retsp/sqrt(rowSums(retsp^2)/(NCOL(retsp)-1))

> # retsp <- xts::xts(retsp, zoo::index(pricev))

> # retsp <- scale(retsp, center=TRUE, scale=TRUE)

> # retsp <- xts::xts(retsp, zoo::index(pricev))

> sapply(retsp, sd)

> # retsp <- t(retsp)

> # retsp <- apply(retsp, 2, scale)

> # retsp <- t(retsp) - colMeans(retsp)

Machine Learning

Principal Component Vectors

Principal components are linear combinations of the k return vectors r::

$$\mathbf{pc}_{j} = \sum_{i=1}^{k} w_{ij} \, \mathbf{r}_{i}$$

Where \mathbf{w}_i is a vector of weights (loadings) of the principal component j, with $\mathbf{w}_{i}^{T}\mathbf{w}_{i}=1$.

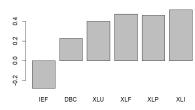
The weights \mathbf{w}_i are chosen to maximize the variance of the principal components, under the condition that they are orthogonal:

$$\mathbf{w}_{j} = \operatorname{arg\ max}\ \left\{\mathbf{pc}_{j}^{T}\ \mathbf{pc}_{j}\right\}$$

$$\mathbf{pc}_{i}^{T}\ \mathbf{pc}_{j} = 0\ (i \neq j)$$

- > # create initial vector of portfolio weights > nweights <- NROW(symboly) > weightv <- rep(1/sqrt(nweights), nweights) > names(weightv) <- symbolv > # Objective function equal to minus portfolio variance > objfun <- function(weightv, retsp) { retsp <- retsp %*% weightv
- -sum(retsp^2) + 1e7*(1 sum(weightv^2))^2 + } # end objfun
- > # Objective for equal weight portfolio > objfun(weightv, retsp)
- > # Compare speed of vector multiplication methods
- > summary(microbenchmark(
- transp=(t(retsp[, 1]) %*% retsp[, 1]),
- sumv=sum(retsp[, 1]^2),
- times=10))[, c(1, 4, 5)] Jerzy Pawlowski (NYU Tandon)

First Principal Component Weights



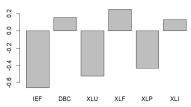
- > # Find weights with maximum variance
- > optiml <- optim(par=weightv, fn=objfun,
- retsp=retsp.
- method="L-BFGS-B".
- upper=rep(10.0, nweights),
- lower=rep(-10.0, nweights))
- > # Optimal weights and maximum variance
- > weights1 <- optiml\$par > -obifun(weights1, retsp)
- > # Plot first principal component weights
- > barplot(weights1, names.arg=names(weights1), xlab="", ylab="", main="First Principal Component Weights")

Higher Order Principal Components

The second *principal component* can be calculated by maximizing its variance, under the constraint that it must be orthogonal to the first *principal component*. Similarly, higher order *principal components* can be calculated by maximizing their variances, under the constraint that they must be orthogonal to all the previous *principal components*.

```
> # PC1 returns
> pc1 < drop(retsp %*% weights1)
> # Redefine objective function
> objfum <- function(weightv, retsp) {
    retsp <- retsp %*% weightv
    -sum(retsp^2) + le7*(1 - sum(weightv^2))^2 +
    1e7*(sum(weightal*weightv))^2 +
    1 # end objfum
> # Find second PC weights using parallel DEoptim
    optiml <- DEoptim: DEoptim(fn=objfum,
    upper=rep(10, NCOL(retsp)),
    1 lower=rep(10, NCOL(retsp)),
    retsp=retsp, control=list(parVar="weights1",
    trace=FALSE, itermax=1000, parallelType=1))
```

Second Principal Component Loadings



```
> # PC2 veights
> weights2 <- optiml%optim%bestmem
> names(veights2) <- colnames(retsp)
> sum(veights2"2)
> sum(veights2"2)
> # PC2 returns
> pc2 <- drop(retsp %*% weights2)
> # Plot second principal component loadings
```

> barplot(weights2, names.arg=names(weights2), xlab="", ylab="",
+ main="Second Principal Component Loadings")

Eigenvalues of the Covariance Matrix

The portfolio variance: $\mathbf{w}^T \mathbb{C} \mathbf{w}$ can be maximized under the quadratic weights constraint $\mathbf{w}^T \mathbf{w} = 1$, by maximizing the Lagrangian \mathcal{L} :

$$\mathcal{L} = \mathbf{w}^T \mathbb{C} \, \mathbf{w} \, - \, \lambda \, (\mathbf{w}^T \mathbf{w} - 1)$$

Where λ is a Lagrange multiplier.

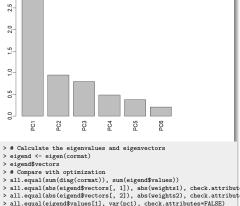
The maximum variance portfolio weights can be found by differentiating \mathcal{L} with respect to \mathbf{w} and setting it to zero:

$$\mathbb{C} \mathbf{w} = \lambda \mathbf{w}$$

This is the eigenvalue equation of the covariance matrix \mathbb{C} , with the optimal weights **w** forming an eigenvector, and λ is the eigenvalue corresponding to the eigenvector w.

The eigenvalues are the variances of the eigenvectors, and their sum is equal to the sum of the return variances:

$$\sum_{i=1}^{k} \lambda_i = \frac{1}{1-k} \sum_{i=1}^{k} \mathbf{r}_i^T \mathbf{r}_i$$



Principal Component Variances

- > all.equal(eigend\$values[2], var(pc2), check.attributes=FALSE)
- > # Eigenvalue equations
- > (cormat %*% weights1) / weights1 / var(pc1)
- > (cormat %*% weights2) / weights2 / var(pc2)
- > # Plot eigenvalues
- > barplot(eigend\$values, names.arg=paste0("PC", 1:nweights),
- + las=3, xlab="", ylab="", main="Principal Component Variances")

Principal Component Analysis Versus Eigen Decomposition

Principal Component Analysis (PCA) is equivalent to the eigen decomposition of either the correlation or the covariance matrix

If the input time series *are* scaled, then *PCA* is equivalent to the eigen decomposition of the *correlation matrix*.

If the input time series *are not* scaled, then *PCA* is equivalent to the eigen decomposition of the *covariance* matrix

Scaling the input time series improves the accuracy of the PCA dimension reduction, allowing a smaller number of principal components to more accurately capture the data contained in the input time series.

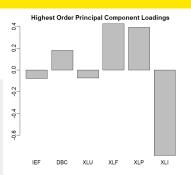
The number of *eigenvalues* is equal to the dimension of the covariance matrix.

- > # Calculate the eigen decomposition of the correlation matrix
 > eigend <- eigen(cormat)</pre>
- > # Perform PCA with scaling
- > pcad <- prcomp(retsp, scale=TRUE)
- > # Compare outputs
- > all.equal(eigend\$values, pcad\$sdev^2)
- > # Eigen decomposition of covariance matrix
- > eigend <- eigen(covmat)
- > # Perform PCA without scaling
- > pcad <- prcomp(retsp, scale=FALSE)
- > # Compare outputs
- > all.equal(eigend\$values, pcad\$sdev^2)
- > all.equal(abs(eigend\$vectors), abs(pcad\$rotation),
- check.attributes=FALSE)

Minimum Variance Portfolio

The highest order principal component, with the smallest eigenvalue, has the lowest possible variance, under the quadratic weights constraint: $\mathbf{w}^T\mathbf{w}=1$. So the highest order principal component is equal to the Minimum Variance Portfolio.

```
> # Redefine objective function to minimize variance
> objfun <- function(weightv, retsp) {
   retsp <- retsp %*% weightv
   sum(retsp^2) + 1e7*(1 - sum(weightv^2))^2
+ } # end objfun
> # Find highest order PC weights using parallel DEoptim
> optiml <- DEoptim::DEoptim(fn=objfun,
   upper=rep(10, NCOL(retsp)),
 lower=rep(-10, NCOL(retsp)),
 retsp=retsp, control=list(trace=FALSE,
     itermax=1000, parallelType=1))
> # PC6 weights and returns
> weights6 <- optiml$optim$bestmem
> names(weights6) <- colnames(retsp)
> sum(weights6^2)
> sum(weights1*weights6)
> # Compare with eigend vector
> weights6
```



- > # Plot highest order principal component loadings
- > x11(width=6, height=5)
- > par(mar=c(2.5, 2, 2, 3), oma=c(0, 0, 0, 0), mgp=c(2, 0.5, 0))
 > barplot(weights6, names.arg=names(weights2), xlab="", ylab="",
- + main="Highest Order Principal Component Loadings")

4□ > 4個 > 4 = > 4 = > = 900

> eigend\$vectors[, 6]
> # Calculate objective function
> objfun(weights6, retsp)
> objfun(eigend\$vectors[, 6], retsp)

Principal Component Analysis of ETF Returns

Principal Component Analysis (PCA) is a dimension reduction technique, that explains the returns of a large number of correlated time series as linear combinations of a smaller number of principal component time series.

The input time series are often scaled by their standard deviations, to improve the accuracy of *PCA dimension reduction*, so that more information is retained by the first few *principal component* time series.

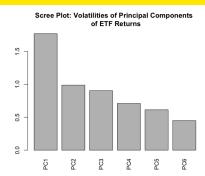
If the input time series are not scaled, then *PCA* analysis is equivalent to the *eigen decomposition* of the covariance matrix, and if they are scaled, then *PCA* analysis is equivalent to the *eigen decomposition* of the correlation matrix.

The function prcomp() performs *Principal Component Analysis* on a matrix of data (with the time series as columns), and returns the results as a list of class prcomp.

The prcomp() argument scale=TRUE specifies that the input time series should be scaled by their standard deviations.

The Kaiser-Guttman rule uses only principal components with variance greater than 1.

Another rule is to use the *principal components* with the largest standard deviations which sum up to 80% of the total variance of returns



A scree plot is a bar plot of the volatilities of the principal components.

- > # Perform principal component analysis PCA
- > pcad <- prcomp(retsp, scale=TRUE)
- > # Plot standard deviations of principal components
- > barplot(pcad\$sdev, names.arg=colnames(pcad\$rotation),
- + las=3, xlab="", ylab="",
- + main="Scree Plot: Volatilities of Principal Components \n of ET
 > # Calculate the number of principal components which sum up to at
- > pcavar <- pcad\$sdev^2
- > which(cumsum(pcavar)/sum(pcavar) > 0.8)[1]

Principal Component Loadings (Weights)

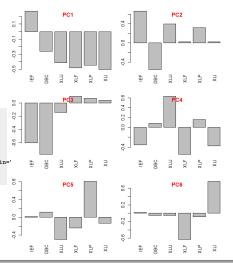
Principal component loadings are the weights of portfolios which have mutually orthogonal returns.

The principal component (PC) portfolios represent the different orthogonal modes of the return variance.

The PC portfolios typically consist of long or short positions of highly correlated groups of assets (clusters), so that they represent relative value portfolios.

> # Plot barplots with PCA loadings (weights) in multiple panels > pcad\$rotation

- > x11(width=6, height=7)
- > par(mfrow=c(nweights/2, 2))
- > par(mar=c(3, 2, 2, 1), oma=c(0, 0, 0, 0))
- > for (ordern in 1:nweights) {
- barplot(pcad\$rotation[, ordern], las=3, xlab="", ylab="", main=" title(paste0("PC", ordern), line=-1, col.main="red")
- # end for



Jerzy Pawlowski (NYU Tandon)

Principal Component Time Series

The time series of the *principal components* can be calculated by multiplying the loadings (weights) times the original data.

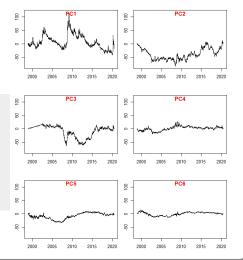
The *principal component* time series have mutually orthogonal returns.

Higher order principal components are gradually less volatile.

> # Calculate products of principal component time series

```
> round(t(pcad$x) %-%, pcad$x, 2)

# Calculate principal component time series from returns
> datev <- zoo::index(pricev)
> retspca <- xts::xts(retsp %-% pcad$rotation, order.by=datev)
> round(cov(retspca), 3)
> all.equal(coredata(retspca), pcad$x, check.attributes=FALSE)
> pcacum <- cumsum(retspca)
> # Plot principal component time series in multiple panels
> rangev <- range(pcacum)
> for (ordern in 1:nweights) {
+ plot.zoo(pcacum[, ordern], ylim=rangev, xlab="", ylab="")
+ title(pasteo("Pc", ordern), line=-1, col.main="red")
+ } # and for
```



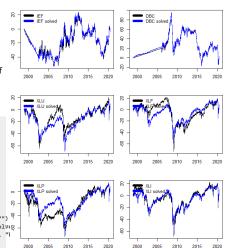
Dimension Reduction Using Principal Component Analysis

The original time series can be calculated exactly from the time series of all the *principal components*, by inverting the loadings matrix.

The original time series can be calculated approximately from just the first few *principal* components, which demonstrates that *PCA* is a form of dimension reduction.

The function solve() solves systems of linear equations, and also inverts square matrices.

```
> # Invert all the principal component time series
> retspca <- retsp %*% pcad$rotation
> solved <- retspca %*% solve(pcad$rotation)
> all.equal(coredata(retsp), solved)
> # Invert first 3 principal component time series
> solved <- retspca[, 1:3] %*% solve(pcad$rotation)[1:3, ]
> solved <- xts::xts(solved, datev)
> solved <- cumsum(solved)
> retc <- cumsum(retsp)
> # Plot the solved returns
> for (symbol in symboly) {
   plot.zoo(cbind(retc[, symbol], solved[, symbol]),
     plot.type="single", col=c("black", "blue"), xlab="", ylab="")
   legend(x="topleft", bty="n", legend=paste0(symbol, c("", " solv(@
     title=NULL, inset=0.0, cex=1.0, lwd=6, lty=1, col=c("black", "l
    # end for
```



Condition Number of Correlation Matrices

The condition number κ of a correlation matrix is equal to the ratio of its largest eigenvalue divided by the smallest:

$$\kappa = \frac{\lambda_{max}}{\lambda_{min}}$$

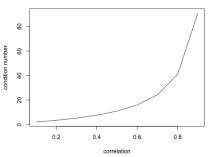
The condition number depends on the level of correlations. If correlations are small then the eigenvalues are close to 1 and the condition number is also close to 1. If the correlations are close to 1 then the condition number is large.

A large condition number indicates the presence of small eigenvalues, and a correlation matrix close to singular, with a poorly defined inverse matrix.

A very large condition number indicates that the correlation matrix is close to being singular.

> # Create a matrix with low correlation > ndata <- 10 > cormat <- matrix(rep(0.1, ndata^2), nc=ndata) > diag(cormat) <- rep(1, ndata) > # Calculate the condition number > eigend <- eigen(cormat) > eigenval <- eigend\$values > max(eigenval)/min(eigenval) > # Create a matrix with high correlation > cormat <- matrix(rep(0.9, ndata^2), nc=ndata) > diag(cormat) <- rep(1, ndata)





- > # Calculate the condition numbers as function correlation
- > corvec <- seq(0.1, 0.9, 0.1)
- > condvec <- sapply(corvec, function(corv) {
- cormat <- matrix(rep(corv, ndata^2), nc=ndata)
- diag(cormat) <- rep(1, ndata)
- eigend <- eigen(cormat)
- eigenval <- eigend\$values
- max(eigenval)/min(eigenval)
- + }) # end sapply
- > # Plot the condition numbers
- > plot(x=corvec, y=condvec, t="1",
- main="Condition Number as Function of Correlation",
- xlab="correlation", ylab="condition number")
- Machine Learning

> # Calculate the condition number

> eigend <- eigen(cormat)

> eigenval <- eigend\$values > max(eigenval)/min(eigenval)

Condition Number for Small Number of Observations

The condition number also depends on the number of observations

If the number of observations (rows of data) is small compared to the number of stocks (columns), then the condition number can be large, even if the returns are not correlated

That's because as the number of rows of data decreases, the returns become more collinear, and the sample correlation matrix becomes more singular, with some very small eigenvalues.

In practice, calculating the inverse correlation matrix of returns faces two challenges: not enough rows of data and correlated returns.

In both cases, the problem is that the columns of returns are close to collinear

9 condition number 2 9 20 60 ጸበ 100

Condition Number as Function of Number of Observations

```
> # Plot the condition numbers
> nstocks <- 10
> nrows <- 100
> set.seed(1121) # Initialize random number generator
> retsp <- matrix(rnorm(nstocks*nrows), nc=nstocks)
   Calculate the condition numbers as function of number of observations
> obsvec <- seg(20, nrows, 10)
> condvec <- sapply(obsvec, function(nobs) {
   cormat <- cor(retsp[1:nobs, ])
   eigend <- eigen(cormat)
```

> plot(x=obsvec, y=condvec, t="1", main="Condition Number as Function of Number of Observations", xlab="number of observations", ylab="condition number")

number of observations

eigenval <- eigend\$values max(eigenval)/min(eigenval)

+ }) # end sapply

Simulate uncorrelated stock returns

The Correlations of Stock Returns

Estimating the correlations of Stock returns is complicated because their date ranges may not overlap in time. Stocks may trade over different date ranges because of IPOs and corporate events (takeovers, mergers).

The function cor() calculates the correlation matrix of time series. The argument

use="pairwise.complete.obs" removes NA values from pairs of stock returns.

But removing NA values in pairs of stock returns can produce correlation matrices which are not positive semi-definite.

The reason is because the correlations are calculated over different time intervals for different pairs of stock returns.

- > # Load daily S&P500 log percentage stock returns
- > load(file="/Users/jerzy/Develop/lecture_slides/data/sp500_returns
 > # Calculate the number of NA values in returns
- > retsp <- returns100["2000/"]
- > colSums(is.na(retsp))
- > # Calculate the correlations ignoring NA values $\,$
- > cor(retsp\$DAL, retsp\$FOXA, use="pairwise.complete.obs")
 > cor(na.omit(retsp[, c("DAL", "FOXA")]))[2]
- > cormat <- cor(retsp, use="pairwise.complete.obs")
- > sum(is.na(cormat))
- > cormat[is.na(cormat)] <- 0
- > cormat[is.na(cormat)] <-

Principal Component Analysis of Stock Returns

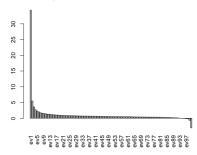
Removing NA values in pairs of stock returns can produce correlation matrices which are not positive semi-definite.

The function prcomp() produces an error when the correlation matrix is not positive semi-definite, so instead, eigen decomposition can be applied to perform Principal Component Analysis.

If some of the eigenvalues are negative, then the condition number is calculated using the eigenvalue with the smallest absolute value.

```
> # Perform principal component analysis PCA - produces error
> pcad <- proomp(retsp, scale=TRUE)
> # Calculate the eigen decomposition of the correlation matrix
> eigend <- eigen(cormat)
> # Calculate the eigenvalues and eigenvectors
> eigenval <- eigend$values
> eigenvec <- eigend$values
> # Calculate the number of negative eigenvalues
> sum (eigenval<0)
> # Calculate the condition number
```

Eigenvalues of Stock Correlation Matrix



- > # Plot the eigenvalues
 > barplot(eigenval, xlab="", ylab="", las=3,
- > barplot(eigenval, xlab="", ylab="", las=3,
 + names.arg=paste0("ev", 1:NROW(eigenval)),
- > max(eigenval/)min(abs(eigenval/) + names.arg=pasteu("ev", 1:NkUw(eigenval/),
 > # Calculate the number of eigenvalues which sum up to at least 8 + main="Eigenvalues of Stock Correlation Matrix")
 - main- Eigenvalues of Stock Cofferation Matrix)

> max(eigenval)/min(abs(eigenval))

> which(cumsum(eigenval)/sum(eigenval) > 0.8)[1]

Principal Component Analysis of Low and High Volatility Stocks

The returns of low volatility stocks have a higher correlation than those of high volatility stocks.

Low and high volatility stocks have different principal components.

The returns of low volatility stocks have a higher correlation so they can be explained by a smaller number of principal components, compared to high volatility stocks.

The correlation matrix of high volatility stocks may have a larger condition number than low volatility stocks

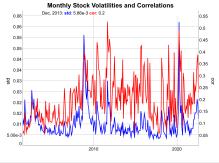
```
> # Calculate the stock volatilities
> stdevs <- sapply(retsp, var, na.rm=TRUE)
> # Calculate the returns of low and high volatility stocks
> nstocks <- NCOL(retsp)
> medianv <- median(stdevs)
> retlow <- retsp[, names(stdevs[stdevs<=medianv])]
> rethigh <- retsp[, names(stdevs[stdevs>medianv])]
> # Calculate the correlations of low volatility stocks
> cormat <- cor(retlow, use="pairwise.complete.obs")
> cormat[is.na(cormat)] <- 0
> # Calculate the mean correlations
> mean(cormat[upper.tri(cormat)])
> # Calculate the eigen decomposition of the correlation matrix
> eigend <- eigen(cormat)
> eigenval <- eigend$values
> # Calculate the number of negative eigenvalues
> sum(eigenval<0)
> # Calculate the number of eigenvalues which sum up to at least 80
> which(cumsum(eigenval)/sum(eigenval) > 0.8)[1]
> # Calculate the condition number
> max(eigenval)/min(abs(eigenval))
> # Calculate the correlations of high volatility stocks
> cormat <- cor(rethigh, use="pairwise.complete.obs")
> cormat[is.na(cormat)] <- 0
> # Calculate the mean correlations
> mean(cormat[upper.tri(cormat)])
> # Calculate the eigen decomposition of the correlation matrix
> eigend <- eigen(cormat)
> eigenval <- eigend$values
> # Calculate the number of negative eigenvalues
> sum(eigenval<0)
> # Calculate the number of eigenvalues which sum up to at least 80
> which(cumsum(eigenval)/sum(eigenval) > 0.8)[1]
> # Calculate the condition number
> max(eigenval)/min(abs(eigenval))
```

Correlations of Stock Returns in Intervals of High and Low Volatility

Correlations of stock returns are higher in time intervals with high volatility.

Stock returns have high correlations in time intervals with high volatility, and vice versa.

```
> # Subset (select) the stock returns after the start date of VTI
> retyti <- na.omit(rutils::etfenv$returns$VTI)
> colnames(retyti) <- "VTI"
> retsp <- returns[zoo::index(retvti)]
> datev <- zoo::index(retsp)
> retyti <- retyti[datev]
> nrows <- NROW(retsp)
> nstocks <- NCOL(retsp)
> head(retsp[, 1:5])
> # Calculate the monthly end points
> endd <- rutils::calc_endpoints(retvti, interval="months")
> retyti[head(endd)]
> retyti[tail(endd)]
> # Remove stub interval at the end
> endd <- endd[-NROW(endd)]
> npts <- NROW(endd)
> # Calculate the monthly stock volatilities and correlations
> stdcor <- sapply(2:npts, function(endp) {
   # cat("endp = ", endp, "\n")
   retsp <- retsp[endd[endp-1]:endd[endp]]
   cormat <- cor(retsp, use="pairwise.complete.obs")
   cormat[is.na(cormat)] <- 0
   c(std=sd(retvti[endd[endp-1]:endd[endp]]).
      cor=mean(cormat[upper.tri(cormat)]))
+ }) # end sapply
> stdcor <- t(stdcor)
```



```
> # Scatterplot of stock volatilities and correlations
> plot(x=stdcor[, "std"], y=stdcor[, "cor"],
+ xlab="volatility", ylab="correlation",
  main="Monthly Stock Volatilities and Correlations")
```

> # Plot stock volatilities and correlations > colnamev <- colnames(stdcor) > stdcor <- xts(stdcor, zoo::index(retvti[endd]))

> dvgraphs::dvgraph(stdcor.

- main="Monthly Stock Volatilities and Correlations") %>% dyAxis("y", label=colnamev[1], independentTicks=TRUE) %>% dyAxis("y2", label=colnamev[2], independentTicks=TRUE) %>%
- dySeries(name=colnamev[1], axis="y", label=colnamev[1], strokeW dySeries(name=colnamev[2], axis="y2", label=colnamev[2], stroke
- dvLegend(show="always", width=500)

> max(eigenval)/min(abs(eigenval))

Principal Component Analysis in Intervals of High and Low Volatility

Stock returns in time intervals with high volatility have high correlations and therefore require fewer eigenvalues to explain 80% of their total variance.

Stock returns in time intervals with low volatility have low correlations and therefore require more eigenvalues to explain 80% of their total variance.

```
> # Calculate the median VTI volatility
> medianv <- median(stdcor[, "std"])
> # Calculate the stock returns of low volatility intervals
> retlow <- lapply(2:npts, function(endp) {
   if (stdcor[endp-1, "std"] <= medianv)
      retsp[endd[endp-1]:endd[endp]]
+ }) # end lapply
> retlow <- rutils::do_call(rbind, retlow)
> # Calculate the stock returns of high volatility intervals
> rethigh <- lapply(2:npts, function(endp) {
   if (stdcor[endp-1, "std"] > medianv)
      retsp[endd[endp-1]:endd[endp]]
```

```
> # Calculate the correlations of low volatility intervals
> cormat <- cor(retlow, use="pairwise.complete.obs")
> cormat[is.na(cormat)] <- 0
> mean(cormat[upper.tri(cormat)])
> # Calculate the eigen decomposition of the correlation matrix
> eigend <- eigen(cormat)
> eigenval <- eigend$values
> sum(eigenval<0)
> # Calculate the number of eigenvalues which sum up to at least 80
> which(cumsum(eigenval)/sum(eigenval) > 0.8)[1]
> # Calculate the condition number
> max(eigenval)/min(abs(eigenval))
> # Calculate the correlations of high volatility intervals
> cormat <- cor(rethigh, use="pairwise.complete.obs")
> cormat[is.na(cormat)] <- 0
> mean(cormat[upper.tri(cormat)])
> # Calculate the eigen decomposition of the correlation matrix
> eigend <- eigen(cormat)
> eigenval <- eigend$values
> sum(eigenval<0)
> # Calculate the number of eigenvalues which sum up to at least 80
> which(cumsum(eigenval)/sum(eigenval) > 0.8)[1]
> # Calculate the condition number
```

> rethigh <- rutils::do_call(rbind, rethigh)

+ }) # end lapply

Trailing Correlations of Stock Returns

The trailing correlations of stock returns are typically higher with higher volatilities.

The function HighFreq::run_covar() calculates the trailing covariance of two time series using the decay factor λ .

$$\begin{split} & \bar{r}_{t}^{1} = \lambda \bar{r}_{t-1}^{1} + (1 - \lambda) r_{t}^{1} \\ & \bar{r}_{t}^{2} = \lambda \bar{r}_{t-1}^{2} + (1 - \lambda) r_{t}^{2} \\ & \operatorname{cov}_{t} = \lambda \operatorname{cov}_{t-1} + (1 - \lambda) (r_{t}^{1} - \bar{r}_{t}^{1}) (r_{t}^{2} - \bar{r}_{t}^{2}) \end{split}$$

- > # Calculate AAPL and XLK returns
- > retsp <- na.omit(cbind(returns\$AAPL, rutils::etfenv\$returns\$XLK)) > # Calculate the trailing correlations
- > lambda <- 0.99
- > covary <- HighFreq::run_covar(retsp, lambda)
- > correlv <- covarv[, 1, drop=FALSE]/sqrt(covarv[, 2]*covarv[, 3]) > volaapl <- sqrt(covarv[, 2])
- > # Scatterplot of trailing stock volatilities and correlations
- > endd <- rutils::calc_endpoints(retsp, interval="months")
- > plot(x=volaapl[endd], y=correlv[endd,], + xlab="volatility", ylab="correlation",
- main="Trailing Stock Volatilities and Correlations")
- > # Plot trailing stock volatilities and correlations
- > datav <- xts(cbind(volaapl, correlv), zoo::index(retsp))
- > colnames(datav) <- c("volatility", "correlation")
- > colnamev <- colnames(datav)
- > dygraphs::dygraph(datav[endd],
- main="AAPL Trailing Stock Volatility and Correlation") %>%
- dyAxis("y", label=colnamev[1], independentTicks=TRUE) %>%
- dvAxis("v2", label=colnamev[2], independentTicks=TRUE) %>%
- dySeries(name=colnamev[1], axis="y", label=colnamev[1], strokeWidth=2, col="blue") %>%

- AAPI Correlations With XI K - XI K -- correlation 0.4 2000 2010 2020
- > # Plot dygraph of XLK returns and AAPL correlations
- > datav <- cbind(cumsum(retsp\$XLK), correlv) > colnames(datay)[2] <- "correlation"
- > colnamev <- colnames(datav)
- > dygraphs::dygraph(datav[endd], main="AAPL Correlations With XLK") dyAxis("y", label=colnamev[1], independentTicks=TRUE) %>%
- dyAxis("y2", label=colnamev[2], independentTicks=TRUE) %>%
- dySeries(name=colnamev[1], axis="y", label=colnamev[1], strokeW dySeries(name=colnamev[2], axis="y2", label=colnamev[2], stroke
- dyLegend(show="always", width=500)

0.05

0.05

0.04

0.04

0.03

0.03

0.02

5.00e-3

- volatility - correlation

Portfolio Correlations

The average correlations of a stock portfolio are typically higher with higher volatilities.

```
> # Calculate portfolio returns
> retvti <- na.omit(rutils::etfenv$returns$VTI)
> colnames(retvti) <- "VTI"
> datev <- zoo::index(retvti)
> retsp <- returns100
> retsp[is.na(retsp)] <- 0
> retsp <- retsp[datev]
> nrows <- NROW(retsp)
> nstocks <- NCOL(retsp)
> head(retsp[, 1:5])
> # Calculate the average trailing portfolio correlations
> lambda <- 0.9
> correly <- sapply(retsp, function(retsp) {
   covary <- HighFreq::run_covar(cbind(retvti, retsp), lambda)
   covarv[, 1, drop=FALSE]/sqrt(covarv[, 2]*covarv[, 3])
+ }) # end sapply
> correlv[is.na(correlv)] <- 0
> correlp <- rowMeans(correly)
```

> # Scatterplot of trailing stock volatilities and correlations

> endd <- rutils::calc_endpoints(retvti, interval="months")

> plot(x=volvti[endd], y=correlp[endd], + xlab="volatility", ylab="correlation",

+ main="Trailing Stock Volatilities and Correlations")

> volvti <- sqrt(HighFreq::run_var(retvti, lambda))

dyAxis("y", label=colnamev[1], independentTicks=TRUE) %>% dvAxis("v2", label=colnamev[2], independentTicks=TRUE) %>%

dySeries(name=colnamev[1], axis="y", label=colnamev[1], strokeW

2010

> # Plot trailing stock volatilities and correlations > datay <- xts(cbind(volvti, correlp), datey)

> colnames(datav) <- c("volatility", "correlation")

> colnamev <- colnames(datav)

> dygraphs::dygraph(datav[endd],

Trailing Stock Volatilities and Correlations

dySeries(name=colnamev[2], axis="y2", label=colnamev[2], stroke

main="Trailing Stock Volatilities and Correlations") %>%

dvLegend(show="always", width=500)

4 D > 4 B > 4 B > 4 B >

2020

0.7

0.65

0.6

0.55

Vector and Matrix Calculus

Let **v** and **w** be vectors, with $\mathbf{v} = \{v_i\}_{i=1}^{i=n}$, and let $\mathbb{1}$ be the unit vector, with $\mathbb{1} = \{1\}_{i=1}^{i=n}$.

Then the inner product of \mathbf{v} and \mathbf{w} can be written as $\mathbf{v}^T\mathbf{w} = \mathbf{w}^T\mathbf{v} = \sum_{i=1}^n v_i w_i$.

We can then express the sum of the elements of \mathbf{v} as the inner product: $\mathbf{v}^T \mathbb{1} = \mathbb{1}^T \mathbf{v} = \sum_{i=1}^n v_i$.

And the sum of squares of \mathbf{v} as the inner product: $\mathbf{v}^T\mathbf{v} = \sum_{i=1}^n v_i^2$.

Let \mathbb{A} be a matrix, with $\mathbb{A} = \{A_{ij}\}_{i,j=1}^{i,j=n}$.

Then the inner product of matrix \mathbb{A} with vectors \mathbf{v} and \mathbf{w} can be written as:

$$\mathbf{v}^T \mathbb{A} \mathbf{w} = \mathbf{w}^T \mathbb{A}^T \mathbf{v} = \sum_{i,j=1}^n A_{ij} v_i w_j$$

The derivative of a scalar variable with respect to a vector variable is a vector, for example:

$$\begin{aligned} \frac{d(\mathbf{v}^T 1)}{d\mathbf{v}} &= d_v[\mathbf{v}^T 1] = d_v[1^T \mathbf{v}] = 1^T \\ d_v[\mathbf{v}^T \mathbf{w}] &= d_v[\mathbf{w}^T \mathbf{v}] = \mathbf{w}^T \\ d_v[\mathbf{v}^T A \mathbf{w}] &= \mathbf{w}^T A^T \\ d_v[\mathbf{v}^T A \mathbf{v}] &= \mathbf{v}^T A + \mathbf{v}^T A^T \end{aligned}$$

Formula Objects

Formulas in R are defined using the "~" operator followed by a series of terms separated by the "+" operator.

Formulas can be defined as separate objects. manipulated, and passed to functions.

The formula "z "x" means the response vector z is explained by the predictor x (also called the explanatory variable or independent variable).

The formula "z ~ x + y" represents a linear model: z = ax + bv + c.

The formula "z ~ x - 1" or "z ~ x + 0" represents a linear model with zero intercept: z = ax.

The function update() modifies existing formulas. The "." symbol represents either all the remaining

data, or the variable that was in this part of the formula.

```
> # Formula of linear model with zero intercept
> formulav <- z ~ x + y - 1
> formulay
> # Collapse vector of strings into single text string
> paste0("x", 1:5)
> paste(paste0("x", 1:5), collapse="+")
> # Create formula from text string
> formulay <- as.formula(
   # Coerce text strings to formula
   paste("z ~ ",
   paste(paste0("x", 1:5), collapse="+")
+ ) # end paste
+ ) # end as.formula
> class(formulay)
> formulay
> # Modify the formula using "update"
```

> update(formulav, log(.) ~ . + beta)

Simple Linear Regression

A Simple Linear Regression is a linear model between a response vector y and a single predictor x, defined by the formula:

$$y_i = \alpha + \beta x_i + \varepsilon_i$$

 α and β are the unknown regression coefficients.

 ε_i are the *residuals*, which are usually assumed to be standard normally distributed $\phi(0, \sigma_\varepsilon)$, independent, and stationary.

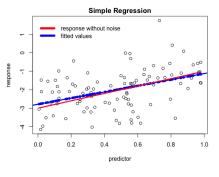
In the Ordinary Least Squares method (*OLS*), the regression parameters are estimated by minimizing the *Residual Sum of Squares* (*RSS*):

$$RSS = \sum_{i=1}^{n} \varepsilon_i^2 = \sum_{i=1}^{n} (y_i - \alpha - \beta x_i)^2$$

$$= (y - \alpha \mathbb{1} - \beta x)^{T} (y - \alpha \mathbb{1} - \beta x)$$

Where $\mathbb{1}$ is the unit vector, with $\mathbb{1}^T \mathbb{1} = n$ and $\mathbb{1}^T x = x^T \mathbb{1} = \sum_{i=1}^n x_i$

The data consists of n pairs of observations (x_i, y_i) of the response and predictor variables, with the index i ranging from 1 to n.



- > # Define explanatory (predv) variable
- > nrows <- 100
- > set.seed(1121) # Initialize random number generator
- > predv <- runif(nrows) > noisev <- rnorm(nrows)
- > # Response equals linear form plus random noise
- > respv <- (-3 + 2*predv + noisev)

The response vector and the predictor matrix don't have to be normally distributed.

Solution of Linear Regression

The *OLS* solution for the *regression coefficients* is found by equating the *RSS* derivatives to zero:

$$RSS_{\alpha} = -2(y - \alpha \mathbb{1} - \beta x)^{T} \mathbb{1} = 0$$

$$RSS_{\beta} = -2(y - \alpha \mathbb{1} - \beta x)^{T} x = 0$$

The solution for α is given by:

$$\alpha = \bar{y} - \beta \bar{x}$$

The solution for β can be obtained by manipulating the equation for RSS_{β} as follows:

$$(y - (\bar{y} - \beta \bar{x})\mathbb{1} - \beta x)^{T}(x - \bar{x}\mathbb{1}) =$$

$$((y - \bar{y}\mathbb{1}) - \beta(x - \bar{x}\mathbb{1}))^{T}(x - \bar{x}\mathbb{1}) =$$

$$(\hat{v} - \beta \hat{x})^{T} \hat{x} = \hat{v}^{T} \hat{x} - \beta \hat{x}^{T} \hat{x} = 0$$

Where $\hat{x} = x - \bar{x}\mathbb{1}$ and $\hat{y} = y - \bar{y}\mathbb{1}$ are the de-meaned variables. Then β is given by:

$$\beta = \frac{\hat{\mathbf{y}}^T \hat{\mathbf{x}}}{\hat{\mathbf{x}}^T \hat{\mathbf{x}}} = \frac{\sigma_{\mathbf{y}}}{\sigma_{\mathbf{x}}} \rho_{\mathbf{x}\mathbf{y}}$$

 β is proportional to the correlation coefficient ρ_{xy} between the response and predictor variables.

If the response and predictor variables have zero mean, then $\alpha=0$ and $\beta=\frac{y^Tx}{T}$.

The residuals $\varepsilon = y - \alpha \mathbb{1} - \beta x$ have zero mean: $RSS_{\alpha} = -2\varepsilon^T \mathbb{1} = 0$.

The residuals ε are orthogonal to the predictor x: $RSS_{\beta} = -2\varepsilon^{T}x = 0$. The expected value of the RSS is equal to the degrees

The expected value of the RSS is equal to the degrees of freedom (n-2) times the variance σ_{ε}^2 of the residuals ε_i : $\mathbb{E}[RSS] = (n-2)\sigma_{\varepsilon}^2$.

- > # Calculate de-meaned predictor and response vectors
- > predzm <- predv mean(predv)
 > respzm <- respv mean(respv)</pre>
- > respzm <- respv mean(respv)
 > # Calculate the regression beta
- > betav <- cov(predv, respv)/var(predv)
- > # Calculate the regression alpha
- > alpha <- mean(respv) betav*mean(predv)
- > alpha <- mean(respv) betav*mean(predv)

check.attributes=FALSE)

[1] TRUE

Linear Regression Using Function lm()

Let the data generating process for the response variable be given as: $z=\alpha_{lat}+\beta_{lat}x+\varepsilon_{lat}$

Where α_{lat} and β_{lat} are latent (unknown) coefficients, and ε_{lat} is an unknown vector of random noise (error terms).

The error terms are the difference between the measured values of the response minus the (unknown) actual response values.

The function lm() fits a linear model into a set of data, and returns an object of class "lm", which is a list containing the results of fitting the model:

- call the model formula.
- coefficients the fitted model coefficients (α, β_j),
- residuals the model residuals (respv minus fitted values).

The regression *residuals* are not the same as the error terms, because the regression coefficients are not equal to the coefficients of the data generating process.

```
> # Specify regression formula
> formulav <- respv ~ predv
> regmod <- lm(formulav) # Perform regression
> class(regmod) # Regressions have class lm
[1] "1m"
> attributes(regmod)
$names
 [1] "coefficients"
                     "residuals"
                                      "effects"
                                                      "rank"
 [5] "fitted.values" "assign"
                                      "ar"
                                                      "df.residual"
 [9] "xlevels"
                      "call"
                                      "terms"
                                                      "model"
$class
[1] "]m"
> eval(regmod$call$formula) # Regression formula
respv ~ predv
> regmod$coeff
                # Regression coefficients
(Intercept)
                  predv
      -2.79
                   1.67
> all.equal(coef(regmod), c(alpha, betav),
```

The Fitted Values of Linear Regression

The fitted values y_{fit} are the estimates of the response vector obtained from the regression model:

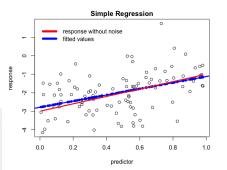
$$y_{fit} = \alpha + \beta x$$

The generic function plot() produces a scatterplot when it's called on the regression formula.

abline() plots a straight line corresponding to the regression coefficients, when it's called on the regression object.

```
> fittedv <- (alpha + betav*predv)
> all.equal(fittedv, regmod$fitted.values, check.attributes=FALSE)
> xi1(vidth+5, height+4) # Open xi1 for plotting
> # Set plot parameters to reduce whitespace around plot
> par(mar=c(5, 5, 2, 1), oma=c(0, 0, 0, 0))
> # Plot scatterplot using formula
> plot(formulav, xlab="predictor", ylab="response")
> title(main="Simple Regression", line=0.5)
> # Add regression line
> abline(regmod, lud=3, col="blue")
> # Plot fitted (predicted) response values
```

> points(x=predv, y=regmod\$fitted.values, pch=16, col="blue")



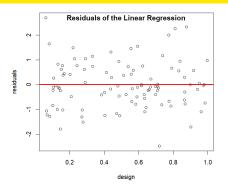
- > # Plot response without noise
 > lines(x=predv, y=(respv-noisev), col="red", lwd=3)
 > legend(x="topleft", # Add legend
 + lerend=c("response without noise". "fitted values").
- + title=NULL, inset=0.01, cex=1.0, lwd=6, y.intersp=0.5,
- + bty="n", lty=1, col=c("red", "blue"))

Linear Regression Residuals

The residuals ε_i of a linear regression are defined as the response vector minus the fitted values:

$$\varepsilon_i = y_i - y_{fit}$$

- > # Calculate the residuals
- > fittedy <- (alpha + betay*predy)
- > residv <- (respv fittedv)
- > all.equal(residv, regmod\$residuals, check.attributes=FALSE)
 [1] TRUE
- > # Residuals are orthogonal to the predictor
- > all.equal(sum(residv*predv), target=0)
- [1] TRUE
- > # Residuals are orthogonal to the fitted values
- > all.equal(sum(residv*fittedv), target=0)
- [1] TRUE
- > # Sum of residuals is equal to zero
- > all.equal(mean(residv), target=0)
- > all.equal(mean(residv), target=0,
- [1] TRUE



- > x11(width=6, height=5) # Open x11 for plotting
- > # Set plot parameters to reduce whitespace around plot
 > par(mar=c(5, 5, 1, 1), oma=c(0, 0, 0, 0))
- > par(mar=c(5, 5, 1, 1), oma=c(0, 0, 0, 0))
 > # Extract residuals
- > datav <- cbind(predv, regmod\$residuals)
- > datav <- cbind(predv, regmod\$residuais)
- > colnames(datav) <- c("predictor", "residuals")
 > # Plot residuals
- > # Plot residu
- > plot(datav)
- > title(main="Residuals of the Linear Regression", line=-1)
 > abline(h=0, lwd=3, col="red")
- > abline(n=0, iwd=3, col="red")

Standard Errors of Regression Coefficients

The *residuals* are the source of error in the regression model, producing uncertainty in the *response vector y* and in the regression coefficients: $y_i = \alpha + \beta x_i + \varepsilon_i$.

The standard errors of the regression coefficients are equal to their standard deviations, given the *residuals* as the source of error.

Since $\beta = \frac{\hat{y}^T \hat{x}}{\hat{x}^T \hat{x}}$, then its variance is equal to:

$$\sigma_{\beta}^{2} = \frac{1}{(n-2)} \frac{E[(\varepsilon^{T} \hat{x})^{2}]}{(\hat{x}^{T} \hat{x})^{2}} = \frac{1}{(n-2)} \frac{E[\varepsilon^{2}]}{\hat{x}^{T} \hat{x}} = \frac{\sigma_{\varepsilon}^{2}}{\hat{x}^{T} \hat{x}}$$

Since $\alpha = \bar{\mathbf{y}} - \beta \bar{\mathbf{x}}$, then its variance is equal to:

$$\sigma_{\alpha}^2 = \frac{\sigma_{\varepsilon}^2}{n} + \sigma_{\beta}^2 \bar{\mathbf{x}}^2 = \sigma_{\varepsilon}^2 (\frac{1}{n} + \frac{\bar{\mathbf{x}}^2}{\hat{\mathbf{x}}^T \hat{\mathbf{x}}})$$

- > # Degrees of freedom of residuals > degf <- regmod\$df.residual
- > # Standard deviation of residuals
- > residsd <- sqrt(sum(residv^2)/degf)
- > # Standard error of beta
- > betasd <- residsd/sqrt(sum(predzm^2))
- > # Standard error of alpha
- > alphasd <- residsd*sqrt(1/nrows + mean(predv)^2/sum(predzm^2))

Linear Regression Summary

The function summary.lm() produces a list of regression model diagnostic statistics:

- coefficients: matrix with estimated coefficients, their *t*-statistics, and *p*-values,
- r.squared: fraction of response variance explained by the model,
- adj.r.squared: r.squared adjusted for higher model complexity,
- fstatistic: ratio of variance explained by the model divided by unexplained variance,

The regression summary is a list, and its elements can be accessed individually.

```
> regsum <- summary(regmod) # Copy regression summary
> regsum # Print the summary to console
Call.
lm(formula = formulav)
Residuals:
   Min
           10 Median
-2.133 -0.649 0.106 0.590 3.321
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept)
              -2.787
                          0.196 -14.20 < 2e-16 ***
predv
               1 665
                          0.357
                                  4 67 9 8e-06 ***
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 '' 1
Residual standard error: 0.988 on 98 degrees of freedom
Multiple R-squared: 0.182.Adjusted R-squared: 0.173
```

F-statistic: 21.8 on 1 and 98 DF, p-value: 9.75e-06 > attributes(regsum)\$names # get summary elements

"terms"

"sigma"

[9] "adj.r.squared" "fstatistic"

"residuals"

"cov.unscaled"

"df"

[1] "call"

[5] "aliased"

"coefficients

"r.squared"

Regression Model Diagnostic Statistics

The *null hypothesis* for regression is that the coefficients are *zero*.

The *t*-statistic (*t*-value) is the ratio of the estimated value divided by its standard error.

The *p*-value is the probability of obtaining values exceeding the *t*-statistic assuming the null hypothese.

exceeding the \emph{t} -statistic, assuming the $\emph{null hypothesis}$ is true.

A small *p*-value means that the regression coefficients are very unlikely to be zero (given the data).

The key assumption in the formula for the standard error is that the *residuals* are normally distributed, independent, and stationary.

If they are not, then the standard error and the p-value may be much bigger than reported by summary.lm(), and therefore the regression may not be statistically significant.

Asset returns are very far from normal, so the small *p*-values shouldn't be automatically interpreted as meaning that the regression is statistically significant.

```
> regsum$coeff
            Estimate Std. Error t value Pr(>|t|)
(Intercept)
               -2.79
                          0.196
                                -14.20 1.61e-25
predv
                1 67
                          0.357
                                   4.67 9.75e-06
> # Standard errors
> regsum$coefficients[2, "Std, Error"]
Γ17 0.357
> all.equal(c(alphasd, betasd), regsum$coefficients[, "Std. Error"]
    check.attributes=FALSE)
[1] TRUE
> # R-squared
> regsum$r.squared
[1] 0.182
> regsum$adi.r.squared
[1] 0.173
> # F-statistic and ANOVA
> regsum$fstatistic
value numdf dendf
 21 8 1 0 98 0
> anova(regmod)
Analysis of Variance Table
Response: respv
          Df Sum Sq Mean Sq F value Pr(>F)
                               21.8 9.8e-06 ***
predv
               21.3
                      21.25
Residuals 98
               95.7
                       0.98
```

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Weak Regression

If the relationship between the response and predictor variables is weak compared to the error terms (noisev), then the regression will have low statistical significance.

- > # High noise compared to coefficient
 > respv <- (-3 + 2*predv + rnorm(nrows, sd=8))</pre>
- > regmod <- lm(formulav) # Perform regression
- > # Values of regression coefficients are not
- > # Statistically significant
- > summary(regmod)

Call:

lm(formula = formulav)

Residuals:

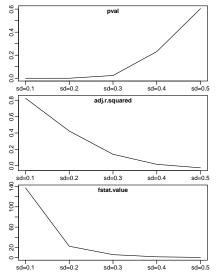
Min 1Q Median 3Q Max -16.430 -4.325 0.735 4.365 16.720

Coefficients:

Residual standard error: 7.25 on 98 degrees of freedom Multiple R-squared: 0.0043, Adjusted R-squared: -0.00586 F-statistic: 0.423 on 1 and 98 DF, p-value: 0.517

Influence of Noise on Regression

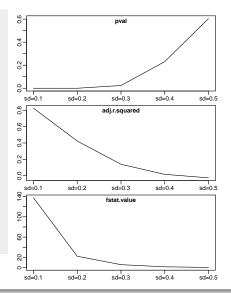
```
> regstats <- function(stdev) { # Noisy regression
    set.seed(1121) # initialize number generator
   Define explanatory (predv) and response variables
    predv <- rnorm(100, mean=2)
   respv <- (1 + 0.2*predv +
    rnorm(NROW(predy), sd=stdey))
   Specify regression formula
    formulay <- respy ~ predy
   Perform regression and get summary
    regsum <- summarv(lm(formulav))
 # Extract regression statistics
    with(regsum, c(pval=coefficients[2, 4],
     adj_rsquared=adj.r.squared,
     fstat=fstatistic[1]))
    # end regstats
> # Apply regstats() to vector of std dev values
> vecsd <- seg(from=0.1, to=0.5, bv=0.1)
> names(vecsd) <- paste0("sd=", vecsd)
> statsmat <- t(sapply(yecsd, regstats))
> # Plot in loop
> par(mfrow=c(NCOL(statsmat), 1))
> for (it in 1:NCOL(statsmat)) {
   plot(statsmat[, it], type="1",
  xaxt="n", xlab="", ylab="", main="")
    title(main=colnames(statsmat)[it], line=-1.0)
   axis(1, at=1:(NROW(statsmat)), labels=rownames(statsmat))
+ } # end for
```



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Influence of Noise on Regression Another Method

```
> regstats <- function(datav) { # get regression
+ # Perform regression and get summary
    colnamev <- colnames(datav)
    formulay <- paste(colnamev[2], colnamev[1], sep="~")
    regsum <- summarv(lm(formulav, data=datav))
 # Extract regression statistics
    with(regsum, c(pval=coefficients[2, 4],
     adj_rsquared=adj.r.squared,
     fstat=fstatistic[1]))
    # end regstats
   Apply regstats() to vector of std dev values
 vecsd <- seg(from=0.1, to=0.5, bv=0.1)
> names(vecsd) <- paste0("sd=", vecsd)
> statsmat <- t(sapply(vecsd, function(stdey) {
      set.seed(1121) # initialize number generator
 # Define explanatory (predy) and response variables
      predv <- rnorm(100, mean=2)
      respv <- (1 + 0.2*predv +
 rnorm(NROW(predy), sd=stdey))
      regstats(data.frame(predv. respv))
 # Plot in loop
 par(mfrow=c(NCOL(statsmat), 1))
> for (it in 1:NCOL(statsmat)) {
    plot(statsmat[, it], type="1",
  xaxt="n", xlab="", ylab="", main="")
   title(main=colnames(statsmat)[it], line=-1.0)
    axis(1, at=1:(NROW(statsmat)),
  labels=rownames(statsmat))
```



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+ } # end for

Linear Regression Diagnostic Plots

plot() produces diagnostic scatterplots for the residuals, when called on the regression object.

The diagnostic scatterplots allow for visual inspection to determine the quality of the regression fit.

- $^{\prime\prime}$ Residuals vs Fitted" is a scatterplot of the residuals vs. the predicted responses.
- "Scale-Location" is a scatterplot of the square root of the standardized residuals vs. the predicted responses.

The residuals should be randomly distributed around the horizontal line representing zero residual error.

A pattern in the residuals indicates that the model was not able to capture the relationship between the variables, or that the variables don't follow the statistical assumptions of the regression model.

- "Normal Q-Q" is the standard Q-Q plot, and the points should fall on the diagonal line, indicating that the residuals are normally distributed.
- "Residuals vs Leverage" is a scatterplot of the residuals vs. their leverage.

Leverage measures the amount by which the fitted values would change if the response values were shifted by a small amount.

Cook's distance measures the influence of a single observation on the fitted values, and is proportional to the sum of the squared differences between predictions made with all observations and predictions made without the observation.

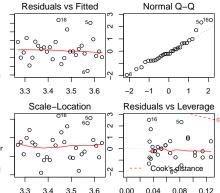
Points with large leverage, or a Cook's distance greater than 1 suggest the presence of an outlier or a poor model,

> par(mfrow=c(2, 2)) # Plot 2x2 panels

> plot(regmod) # Plot diagnostic scatterplots

> plot(regmod, which=2) # Plot just Q-Q

Im(reg_formula)



Durbin-Watson Test of Autocorrelation of Residuals

The *Durbin-Watson* test is designed to test the *null hypothesis* that the autocorrelations of regression *residuals* are equal to zero.

The test statistic is equal to:

$$DW = \frac{\sum_{i=2}^{n} (\varepsilon_i - \varepsilon_{i-1})^2}{\sum_{i=1}^{n} \varepsilon_i^2}$$

Where ε_i are the regression *residuals*.

The value of the *Durbin-Watson* statistic *DW* is close to zero for large positive autocorrelations, and close to four for large negative autocorrelations.

The ${\it DW}$ is close to two for autocorrelations close to zero.

The p-value for the reg_model regression is large, and we conclude that the null hypothesis is TRUE, and the regression residuals are uncorrelated.

> library(lmtest) # Load lmtest

> # Perform Durbin-Watson test

> lmtest::dwtest(regmod)

Durbin-Watson test

data: regmod DW = 2, p-value = 0.7

alternative hypothesis: true autocorrelation is greater than $\ensuremath{\text{0}}$

draft: Autocorrelated Time Series Regression

Filtering or smoothing a time series containing an error terms over overlapping periods introduces autocorrelations in the error terms of the time series.

Autocorrelations in the error terms introduces autocorrelations of the regression residuals, causing the Durbin-Watson test to fail.

Autocorrelations in the error terms introduce autocorrelations of the regression residuals, causing the Durbin-Watson test to fail

The failure of the Durbin-Watson test means that the standard errors and p-values calculated by the regression model are too small, and therefore the regression may not be statistically significant.

But the failure of the Durbin-Watson test doesn't reject the existence of a linear relationship between the response and predictor variables, it just puts it in doubt.

Links:

https://onlinecourses.science.psu.edu/stat510/node/72 http://stats.stackexchange.com/questions/6469/simple-linear-model-with-autocorrelated-errors-in-r

Regression of non-stationary time series creates *spurious* regressions.

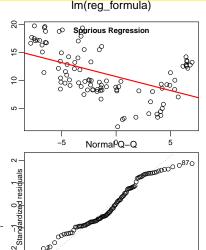
The *t*-statistics, *p*-values, and *R*-squared all indicate a statistically significant regression.

But the Durbin-Watson test shows residuals are

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Theoretical Quantiles

The Leverage for Univariate Regression

We can add an extra unit column to the *predictor* $matrix \mathbb{X}$ so that the univariate regression can be written in *homogeneous form* as:

$$v = X\beta + \varepsilon$$

With two regression coefficients: $\beta = (\alpha, \beta_1)$, and a predictor matrix $\mathbb X$ with two columns, with the first column equal to a unit vector.

After the second column of the *predictor matrix* $\mathbb X$ is de-meaned, its *covariance matrix* is given by:

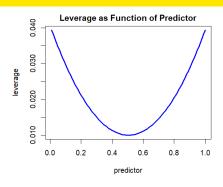
$$\mathbb{X}^{T}\mathbb{X} = \begin{pmatrix} n & 0 \\ 0 & \sum_{i=1}^{n} (x_{i} - \bar{x})^{2} \end{pmatrix}$$

And the *influence matrix* \mathbb{H} is given by:

$$\mathbb{H}_{ij} = [\mathbb{X}(\mathbb{X}^T\mathbb{X})^{-1}\mathbb{X}^T]_{ij} = \frac{1}{n} + \frac{(x_i - \bar{x})(x_j - \bar{x})}{\sum_{i=1}^n (x_i - \bar{x})^2}$$

The first term above is due to the influence of the regression intercept α , and the second term is due to the influence of the regression slope β_1 .

The diagonal elements of the *influence matrix* \mathbb{H}_{ii} form the *leverage vector*.



- > # Add unit column to the predictor matrix
- > predv <- cbind(rep(1, nrows), predv)
- > # Calculate generalized inverse of the predictor matrix
- > predinv <- MASS::ginv(predv)
- > # Calculate the influence matrix
- > infmat <- predv %*% predinv > # Plot the leverage vector
- > ordern <- order(predv[, 2])
- > plot(x=predv[ordern, 2], y=diag(infmat)[ordern],
- + type="1", lwd=3, col="blue",
- + xlab="predictor", ylab="leverage",
- + main="Leverage as Function of Predictor")

4 D > 4 A > 4 B > 4 B > B

Standard Deviations of Fitted Values

Covariance Matrix of Fitted Values in Univariate Regression

The fitted values y_{fit} can be considered to be random variables \hat{y}_{fit} :

$$\hat{y}_{fit} = \mathbb{H}\hat{y} = \mathbb{H}(y_{fit} + \hat{\varepsilon}) = y_{fit} + \mathbb{H}\hat{\varepsilon}$$

The covariance matrix of the fitted values \hat{y}_{fit} is:

$$\begin{split} \sigma_{\mathit{fit}}^2 &= \frac{\mathbb{E}\left[\mathbb{H}\hat{\varepsilon}(\mathbb{H}\hat{\varepsilon})^T\right]}{d_{\mathit{free}}} = \frac{\mathbb{E}\left[\mathbb{H}\,\hat{\varepsilon}\hat{\varepsilon}^T\,\mathbb{H}^T\right]}{d_{\mathit{free}}} = \\ \frac{\mathbb{H}\left[\mathbb{E}[\hat{\varepsilon}\hat{\varepsilon}^T]\,\mathbb{H}^T\right]}{d_{\mathit{free}}} &= \sigma_{\varepsilon}^2\,\mathbb{H} = \sigma_{\varepsilon}^2\,\mathbb{X}(\mathbb{X}^T\mathbb{X})^{-1}\mathbb{X}^T \end{split}$$

The square of the *influence matrix* $\mathbb H$ is equal to itself (it's idempotent): $\mathbb H \mathbb H^T = \mathbb H$.

The variance of the *fitted values* σ_{fit}^2 increases with the distance of the *predictors* from their mean values.

This is because the *fitted values* farther from their mean are more sensitive to the variance of the regression slope.

- > # Calculate the influence matrix > infmat <- predv %*% predinv > # The influence matrix is idempotent
- > all.equal(infmat, infmat %*% infmat)

0.10 12 1.3 14 1.5 17 1.8 > # Calculate covariance and standard deviations of fitted values > betay <- prediny %*% respy > fittedv <- drop(predv %*% betav) > residv <- drop(respv - fittedv) > degf <- (NROW(predv) - NCOL(predv)) > residsd <- sqrt(sum(residv^2)/degf) > fitcovar <- residsd*infmat > fitsd <- sgrt(diag(fitcovar)) > # Plot the standard deviations > fitsd <- cbind(fitted=fittedv, stdev=fitsd) > fitsd <- fitsd[order(fittedv),] > plot(fitsd, type="1", lwd=3, col="blue",

xlab="Fitted Value", ylab="Standard Deviation", main="Standard Deviations of Fitted Values\nin Univariate Re

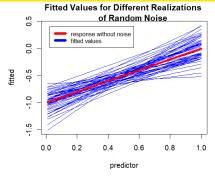
Fitted Values for Different Realizations of Random Noise

The fitted values are more volatile for *predictor* values that are further away from their mean, because those points have higher *leverage*.

The higher *leverage* of points further away from the mean of the *predictor* is due to their greater sensitivity to changes in the slope of the regression.

The fitted values for different realizations of random noise can be calculated using the influence matrix.

- > # Calculate response without random noise for univariate regressic > # equal to weighted sum over columns of predictor.
- > betav <- c(-1, 1)
- > respv <- predv %*% betav
- > # Perform loop over different realizations of random noise
- > fittedv <- lapply(1:50, function(it) {
- + # Add random noise to response
- + respv <- respv + rnorm(nrows, sd=1.0)
- + # Calculate fitted values using influence matrix
- + infmat %*% respv
- + infmat %*% resp
- + }) # end lapply
- > fittedv <- rutils::do_call(cbind, fittedv)



- > # Plot fitted values
- > matplot(x=predv[. 2], v=fittedv.
- + type="l", lty="solid", lwd=1, col="blue",
- + xlab="predictor", ylab="fitted",
- + main="Fitted Values for Different Realizations
- + of Random Noise")
- > lines(x=predv[, 2], y=respv, col="red", lwd=4)
- > legend(x="topleft", # Add legend
 - legend=c("response without noise", "fitted values"),
- + title=NULL, inset=0.05, cex=0.8, lwd=6, y.intersp=0.5,
- + bty="n", lty=1, col=c("red", "blue"))

Predictions From Univariate Regression Models

The prediction y_{pred} from a regression model is equal to the *response value* corresponding to the *predictor* vector with the new data \mathbb{X}_{new} .

$$v_{pred} = X_{new} \beta$$

The variance σ_{pred}^2 of the predicted value is:

$$\begin{split} \sigma_{pred}^2 &= \frac{\mathbb{E}\left[\mathbb{X}_{new}\mathbb{X}_{inv}\hat{\varepsilon}\left(\mathbb{X}_{new}\mathbb{X}_{inv}\hat{\varepsilon}\right)^T\right]}{d_{free}} = \\ &\frac{\mathbb{E}\left[\mathbb{X}_{new}\mathbb{X}_{inv}\hat{\varepsilon}\hat{\varepsilon}^T\mathbb{X}_{inv}^T\mathbb{X}_{new}^T\right]}{d_{free}} = \sigma_{\varepsilon}^2\mathbb{X}_{new}\mathbb{X}_{inv}\mathbb{X}_{inv}^T\mathbb{X}_{new}^T = \\ \sigma_{\varepsilon}^2\mathbb{X}_{new}(\mathbb{X}^T\mathbb{X})^{-1}\mathbb{X}_{new}^T = \mathbb{X}_{new}\sigma_{\beta}^2\mathbb{X}_{new}^T \end{split}$$

The variance σ^2_{pred} of the predicted value is equal to the predictor vector multiplied by the covariance matrix of the regression coefficients $\sigma^2_{\rm R}$.

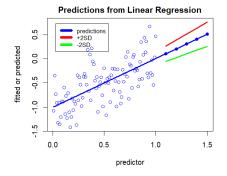
- > # Inverse of predictor matrix squared
 > predv2 <- MASS::ginv(crossprod(predv))
 > # Define new predictors
 > newdata <- (max(predv[, 2]) + 10*(1:5)/nrows)
 > # Calculate the predicted values and standard errors
 > predv <- cbind(rep(1, NROW(newdata)), newdata)
 > predsd <- sqrt(predv %*%, predv2 %*% t(predv))
 > predv <- cbind(
 + predictiom=drop(predv %*%, betav),
- + stdev=diag(residsd*predsd))
 > # Or: Perform loop over predv
- > # of. Felicim loop over predv
 > predv <- apply(predv, MARGIN=1, function(predv) {
 + # Calculate predicted values</pre>
- + prediction <- predv %*% betav
- + # Calculate standard deviation + predsd <- sqrt(t(predv) %*% predv2 %*% predv)
- + predictsd <- residsd*predsd
 + c(prediction=prediction, stdev=predictsd)</pre>
- + }) # end sapply > predv <- t(predv)

Confidence Intervals of Regression Predictions

The variables σ_{ε}^2 and σ_y^2 follow the *chi-squared* distribution with $d_{free} = (n-k-1)$ degrees of freedom, so the *predicted value* y_{pred} follows the *t-distribution*.

```
> # Prepare plot data
> xdata <- c(predv[, 2], newdata)
> xlim <- range(xdata)
> vdata <- c(fittedy, predy[, 1])
> # Calculate t-quantile
> tquant <- qt(pnorm(2), df=degf)
> predictlow <- predv[, 1]-tquant*predv[, 2]
> predicthigh <- predv[, 1]+tquant*predv[, 2]
> ylim <- range(c(respv, ydata, predictlow, predicthigh))
> # Plot the regression predictions
> plot(x=xdata, y=ydata, xlim=xlim, ylim=ylim,
      type="1", lwd=3, col="blue",
      xlab="predictor", ylab="fitted or predicted",
      main="Predictions from Linear Regression")
> points(x=predv[, 2], y=respv, col="blue")
> points(x=newdata, y=predv[, 1], pch=16, col="blue")
> lines(x=newdata, y=predicthigh, lwd=3, col="red")
> lines(x=newdata, y=predictlow, lwd=3, col="green")
> legend(x="topleft", # Add legend
        legend=c("predictions", "+2SD", "-2SD"),
        title=NULL, inset=0.05, cex=0.8, lwd=6, y.intersp=0.5,
```

bty="n", lty=1, col=c("blue", "red", "green"))



Predictions From Linear Regression Using Function 1m()

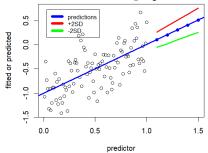
The function predict() is a *generic function* for forecasting based on a given model.

predict.lm() is the predict method for linear models (regressions) produced by the function lm().

```
> # Perform univariate regression
> predv <- predv[, 2]
> regmod <- lm(respv ~ predv)
> # Perform prediction from regression
> newdata <- data.frame(predv=newdata)
> predictlm <- predict(object=model,
   newdata=newdata, confl=1-2*(1-pnorm(2)),
    interval="confidence")
> predictlm <- as.data.frame(predictlm)
> all.equal(predictlm$fit, predv[, 1])
> all.equal(predictlm$lwr, predictlow)
> all.equal(predictlm$upr, predicthigh)
> plot(respy ~ predy.
      xlim=range(predv, newdata),
      ylim=range(respv, predictlm),
      xlab="predictor", ylab="fitted or predicted",
```

main="Predictions from lm() Regression")

Predictions from Im() Regression



- > abline(regmod, col="blue", lwd=3)
- > with(predictlm, {
- + points(x=newdata\$predv, y=fit, pch=16, col="blue")
- + lines(x=newdata\$predv, y=lwr, lwd=3, col="green")
- + lines(x=newdata\$predv, y=upr, lwd=3, col="red")
- + }) # end with
- > legend(x="topleft", # Add legend
- + legend=c("predictions", "+2SD", "-2SD"),
- + title=NULL, inset=0.05, cex=0.8, lwd=6, y.intersp=0.5,
- + bty="n", lty=1, col=c("blue", "red", "green"))

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Spurious Time Series Regression

Regression of non-stationary time series creates *spurious* regressions.

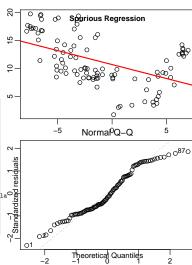
The *t*-statistics, *p*-values, and *R*-squared all indicate a statistically significant regression.

But the Durbin-Watson test shows residuals are autocorrelated, which invalidates the other tests.

The Q-Q plot also shows that residuals are *not* normally distributed.

```
> predv <- cumsum(rnorm(100)) # Unit root time series
         > respv <- cumsum(rnorm(100))
         > formulav <- respv ~ predv
         > regmod <- lm(formulay) # Perform regression
         > # Summary indicates statistically significant regression
         > regsum <- summary(regmod)
*** residuals are autocorrelated 
         > regsum$coeff
```

lm(reg_formula)



Multivariate Linear Regression

A multivariate linear regression model with k predictors x_j , is defined by the formula:

$$y_i = \alpha + \sum_{j=1}^k \beta_j x_{i,j} + \varepsilon_i$$

 α and β are the unknown regression coefficients, with α a scalar and β a vector of length k.

The *residuals* ε_i are assumed to be normally distributed $\phi(0, \sigma_{\varepsilon})$, independent, and stationary.

The data consists of *n* observations, with each observation containing *k predictors* and one *response* value.

The response vector y, the predictor vectors x_j , and the residuals ε are vectors of length n.

The k predictors x_j form the columns of the (n, k)-dimensional predictor matrix \mathbb{X} .

The *multivariate regression* model can be written in vector notation as:

$$y = \alpha + \mathbb{X}\beta + \varepsilon = y_{fit} + \varepsilon$$
$$v_{fit} = \alpha + \mathbb{X}\beta$$

Where y_{fit} are the *fitted values* of the model.

- > # Define predictor matrix
- > nrows <- 100 > ncols <- 5
- > set.seed(1121) # initialize random number generator
- > predv <- matrix(runif(nrows*ncols), ncol=ncols)
- > # Add column names
- > colnames(predv) <- paste0("pred", 1:ncols)
- > # Define the predictor weights
- > weightv <- runif(3:(ncols+2), min=(-1), max=1)
- > # Response equals weighted predictor plus random noise
- > noisev <- rnorm(nrows, sd=2)
- > respv <- (1 + predv %*% weightv + noisev)

Solution of *Multivariate Regression*

The Residual Sum of Squares (RSS) is defined as the sum of the squared residuals:

$$RSS = \varepsilon^{T} \varepsilon = (y - y_{fit})^{T} (y - y_{fit}) = (y - \alpha + \mathbb{X}\beta)^{T} (y - \alpha + \mathbb{X}\beta)$$

The *OLS* solution for the regression coefficients is found by equating the *RSS* derivatives to zero:

$$RSS_{\alpha} = -2(y - \alpha - \mathbb{X}\beta)^{T} \mathbb{1} = 0$$

$$RSS_{\beta} = -2(y - \alpha - \mathbb{X}\beta)^{T} \mathbb{X} = 0$$

The solutions for α and β are given by:

$$\begin{split} \alpha &= \bar{y} - \bar{\mathbb{X}}\beta \\ RSS_{\beta} &= -2(\hat{y} - \hat{\mathbb{X}}\beta)^T \hat{\mathbb{X}} = 0 \\ \hat{\mathbb{X}}^T \hat{y} - \hat{\mathbb{X}}^T \hat{\mathbb{X}}\beta &= 0 \\ \beta &= (\hat{\mathbb{X}}^T \hat{\mathbb{X}})^{-1} \hat{\mathbb{X}}^T \hat{y} &= \hat{\mathbb{X}}^{inv} \hat{y} \end{split}$$

Where \bar{y} and $\bar{\mathbb{X}}$ are the column means, and $\hat{\mathbb{X}} = \mathbb{X} - \bar{\mathbb{X}}$ and $\hat{y} = y - \bar{y} = \hat{\mathbb{X}}\beta + \varepsilon$ are the de-meaned variables.

The matrix $\hat{\mathbb{X}}^{inv}$ is the generalized inverse of the de-meaned *predictor matrix* $\hat{\mathbb{X}}$.

The matrix $\mathbb{C} = \hat{\mathbb{X}}^T \hat{\mathbb{X}}/(n-1)$ is the covariance matrix of the matrix \mathbb{X} , and it's invertible only if the columns of \mathbb{X} are linearly independent.

- > # Perform multivariate regression using lm()
- > regmod <- lm(respv ~ predv)
- > # Solve multivariate regression using matrix algebra
- > # Calculate de-meaned predictor matrix and response vector
 > predzm <- t(t(predv) colMeans(predv))</pre>
- > # predv <- apply(predv, 2, function(x) (x-mean(x)))
- > respzm <- respv mean(respv)
- > # Calculate the regression coefficients
- > betav <- drop(MASS::ginv(predzm) %*% respzm)
- > # Calculate the regression alpha
- > alpha <- mean(respv) sum(colSums(predv)*betav)/nrows
- > # Compare with coefficients from lm()
- > all.equal(coef(regmod), c(alpha, betav), check.attributes=FALSE)
 [1] TRUE
- > # Compare with actual coefficients
- > all.equal(c(-1, weightv), c(alpha, betav), check.attributes=FALSE
- [1] "Mean relative difference: 1.42"

Multivariate Regression in Homogeneous Form

We can add an extra unit column to the *predictor* $matrix \ \mathbb{X}$ to represent the intercept term, and express the *linear regression* formula in *homogeneous form*:

$$y = X\beta + \varepsilon$$

Where the regression coefficients β now contain the intercept α : $\beta = (\alpha, \beta_1, \dots, \beta_k)$, and the predictor matrix $\mathbb X$ has k+1 columns and n rows.

The OLS solution for the β coefficients is found by equating the RSS derivative to zero:

$$RSS_{\beta} = -2(y - \mathbb{X}\beta)^{T} \mathbb{X} = 0$$

$$\mathbb{X}^{T} y - \mathbb{X}^{T} \mathbb{X}\beta = 0$$

$$\beta = (\mathbb{X}^{T} \mathbb{X})^{-1} \mathbb{X}^{T} y = \mathbb{X}_{inv} y$$

The matrix $\mathbb{X}_{inv} = (\mathbb{X}^T \mathbb{X})^{-1} \mathbb{X}^T$ is the generalized inverse of the *predictor matrix* \mathbb{X} .

The coefficients β can be interpreted as the projections of the *response vector y* onto the columns of the *predictor matrix* \mathbb{X} .

The predictor matrix $\mathbb X$ maps the regression coefficients β into the response vector $\mathbf y$.

The generalized inverse of the *predictor matrix* \mathbb{X}_{inv} maps the *response vector y* into the *regression coefficients* β .

- > predv <- cbind(rep(1, NROW(predv)), predv)
 > ncols <- NCOL(predv)
 > # Add column name
 > colnames(predv)[i] <- "intercept"
 > # Calculate generalized inverse of the predictor matrix
- > predinv <- MASS::ginv(predv)
 > # Calculate the regression coefficients

> # Add intercept column to predictor matrix

- > betav <- predinv %*% respv
- > # Perform multivariate regression without intercept term > regmod <- lm(respv ~ predv - 1)
- > all.equal(drop(betav), coef(regmod), check.attributes=FALSE)
 [1] TRUE

The Residuals of Multivariate Regression

The *multivariate regression* model can be written in vector notation as:

$$y = X\beta + \varepsilon = y_{fit} + \varepsilon$$
$$v_{fit} = X\beta$$

Where y_{fit} are the *fitted values* of the model.

The residuals are equal to the response vector minus the fitted values: $\varepsilon = y - y_{fit}$.

The residuals ε are orthogonal to the columns of the predictor matrix $\mathbb X$ (the predictors):

$$\varepsilon^{T} \mathbb{X} = (y - \mathbb{X}(\mathbb{X}^{T} \mathbb{X})^{-1} \mathbb{X}^{T} y)^{T} \mathbb{X} = y^{T} \mathbb{X} - y^{T} \mathbb{X}(\mathbb{X}^{T} \mathbb{X})^{-1} \mathbb{X}^{T} \mathbb{X} = y^{T} \mathbb{X} - y^{T} \mathbb{X} = 0$$

Therefore the *residuals* are also orthogonal to the *fitted* values: $\varepsilon^T y_{fit} = \varepsilon^T \mathbb{X} \beta = 0$.

Since the first column of the *predictor matrix* $\mathbb X$ is a unit vector, the *residuals* ε have zero mean: $\varepsilon^T\mathbb 1=0$.

- > # Calculate fitted values from regression coefficients > fittedv <- drop(predv %*% betav)
- > all.equal(fittedv, regmod\$fitted.values, check.attributes=FALSE)
- [1] TRUE
 > # Calculate the residuals
- > residv <- drop(respv fittedv)
- > all.equal(residv, regmod\$residuals, check.attributes=FALSE)
- [1] TRUE
 > # Residuals are orthogonal to predictor columns (predvs)
- > sapply(residv %*% predv, all.equal, target=0)
- [1] TRUE TRUE TRUE TRUE TRUE TRUE
- > # Residuals are orthogonal to the fitted values
- > all.equal(sum(residv*fittedv), target=0)
 [1] TRUE
- > # Sum of residuals is equal to zero
- > all.equal(sum(residv), target=0)
- [1] TRUE

The Influence Matrix of Multivariate Regression

The vector $y_{fit} = \mathbb{X}\beta$ are the fitted values corresponding to the response vector y:

$$y_{fit} = \mathbb{X}\beta = \mathbb{X}(\mathbb{X}^T\mathbb{X})^{-1}\mathbb{X}^Ty = \mathbb{X}\mathbb{X}_{inv}y = \mathbb{H}y$$

Where $\mathbb{H} = \mathbb{X}\mathbb{X}_{inv} = \mathbb{X}(\mathbb{X}^T\mathbb{X})^{-1}\mathbb{X}^T$ is the influence matrix (or hat matrix), which maps the response vector y into the fitted values y_{fit} .

The influence matrix \mathbb{H} is a projection matrix, and it measures the changes in the fitted values y_{fit} due to changes in the response vector y.

$$\mathbb{H}_{ij} = \frac{\partial y_i^{nt}}{\partial y_j}$$

The square of the *influence matrix* \mathbb{H} is equal to itself (it's idempotent): $\mathbb{H} \mathbb{H}^T = \mathbb{H}$.

- > # Calculate the influence matrix
- > infmat <- predv %*% predinv
- > # The influence matrix is idempotent
- > all.equal(infmat, infmat %*% infmat)
 [1] TRUE
- > # Calculate fitted values using influence matrix
- > fittedv <- drop(infmat %*% respv)
 > all.equal(fittedv, regmod\$fitted.values, check.attributes=FALSE)
- [1] TRUE
- > # Calculate fitted values from regression coefficients > fittedv <- drop(predv %*% betav)
- > iltedv <- drop(predv /*/, betav)
 > all.equal(fittedv, regmod\$fitted.values, check.attributes=FALSE)
 [1] TRUE

Multivariate Regression With de-Meaned Variables

The *multivariate regression* model can be written in vector notation as:

$$y = \alpha + \mathbb{X}\beta + \varepsilon$$

The intercept α can be substituted with its solution: $\alpha = \bar{y} - \bar{\mathbb{X}}\beta$ to obtain the regression model with de-meaned response and predictor matrix:

$$y = \bar{y} - \bar{\mathbb{X}}\beta + \mathbb{X}\beta$$
$$\hat{y} = \hat{\mathbb{X}}\beta + \varepsilon$$

The regression model with a de-meaned *predictor* matrix produces the same fitted values (only shifted by their mean) and residuals as the original regression model, so it's equivalent to it. has the same influence matrix, and

But the de-meaned regression model has a different influence matrix, which maps the de-meaned response vector \hat{y} into the de-meaned fitted values \hat{y}_{fit} .

> # Calculate zero mean fitted values > predzm <- t(t(predv) - colMeans(predv)) > fitted_zm <- drop(predzm %*% betav) > all.equal(fitted_zm, regmod\$fitted.values - mean(respv), check.attributes=FALSE) [1] TRUE > # Calculate the residuals > respzm <- respv - mean(respv) > residv <- drop(respzm - fitted_zm) > all.equal(residv, regmod\$residuals, check.attributes=FALSE) [1] TRUE > # Calculate the influence matrix > influence_zm <- predzm %*% MASS::ginv(predzm) > # Compare the fitted values > all.equal(fitted_zm, drop(influence_zm %*% respzm), check.attributes=FALSE)

[1] TRUE

Multivariate Regression for Orthogonal Predictors

The generalized inverse can be written as:

$$\mathbb{X}_{\textit{inv}} = (\mathbb{X}^{\textit{T}}\mathbb{X})^{-1}\mathbb{X}^{\textit{T}} = \mathbb{C}^{-1}\mathbb{X}^{\textit{T}}$$

Where $\mathbb{C}=\mathbb{X}^T\mathbb{X}$ is the matrix of inner products of the predictors \mathbb{X} .

If the predictors are orthogonal $(x_i \cdot x_j = 0 \text{ for } i \neq j,$ and $x_i \cdot x_i = \sigma_i^2)$ then the squared predictor matrix $\mathbb C$ is diagonal:

$$\mathbb{C} = \begin{pmatrix} \sigma_1^2 & 0 & \cdots & 0 \\ 0 & \sigma_2^2 & \cdots & 0 \\ \vdots & \vdots & \ddots & \vdots \\ 0 & 0 & \cdots & \sigma_n^2 \end{pmatrix}$$

And the inverse of the squared predictor matrix \mathbb{C}^{-1} is also diagonal, so the *regression coefficients* can then be written simply as:

$$\beta_i = \frac{x_i \cdot y}{\sigma_i^2}$$

Where $x_i \cdot y$ are the inner products of the predictors x_i times the *response vector* y.

Conversely, if the predictors are *collinear* then their squared predictor matrix is *singular* and the regression is also singular. Predictors are *collinear* if there's a linear combination that is constant.

> # Perform PCA of the predictors > pcad <- prcomp(predv, center=FALSE, scale=FALSE) > # Calculate the PCA predictors > predpca <- predv %*% pcad\$rotation > # Principal components are orthogonal to each other > round(t(predpca) %*% predpca, 2) > # Calculate the PCA regression coefficients using lm() > regmod <- lm(respv ~ predpca - 1) > summary(regmod) > regmod\$coefficients > # Calculate the PCA regression coefficients directly > colSums(predpca*drop(respv))/colSums(predpca^2) > # Create almost collinear predictors > predc <- predv > predc[, 1] <- (predc[, 1]/1e3 + predc[, 2]) > # Calculate the PCA predictors > pcad <- prcomp(predc, center=FALSE, scale=FALSE) > predpca <- predc %*% pcad\$rotation > round(t(predpca) %*% predpca, 6) > # Calculate the PCA regression coefficients

> # Calculate the PCA regression coefficients directly

> colSums(predpca*drop(respv))/colSums(predpca^2)

> drop(MASS::ginv(predpca) %*% respv)

Omitted Variable Bias

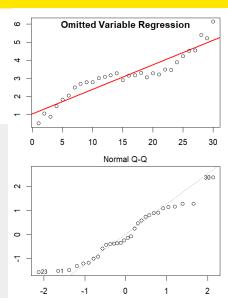
Omitted Variable Bias occurs in a regression model that omits important predictors.

The parameter estimates are biased, even though the *t*-statistics, *p*-values, and *R*-squared all indicate a statistically significant regression.

But the Durbin-Watson test shows that the residuals are autocorrelated, which means that the regression coefficients may not be statistically significant (different from zero).

```
> library(lmtest) # Load lmtest
> # Define predictor matrix
> predv <- 1:30
> omity <- sin(0.2*1:30)
> # Response depends on both predictors
> respv <- 0.2*predv + omitv + 0.2*rnorm(30)
> # Mis-specified regression only one predictor
> model_ovb <- lm(respv ~ predv)
> regsum <- summary(regmod_ovb)
> regsum$coeff
> regsum$r.squared
> # Durbin-Watson test shows residuals are autocorrelated
> lmtest::dwtest(regmod_ovb)
> # Plot the regression diagnostic plots
> x11(width=5, height=7)
> par(mfrow=c(2,1)) # Set plot panels
> par(mar=c(3, 2, 1, 1), oma=c(1, 0, 0, 0))
> plot(respv ~ predv)
> abline(regmod_ovb, lwd=2, col="red")
> title(main="Omitted Variable Regression", line=-1)
```

> plot(regmod ovb, which=2, ask=FALSE) # Plot just Q-Q



Regression Coefficients as Random Variables

The residuals $\hat{\varepsilon}$ can be considered to be random variables, with expected value equal to zero $\mathbb{E}[\hat{\varepsilon}]=0,$ and variance equal to $\sigma_{\varepsilon}^2.$

The variance of the *residuals* is equal to the expected value of the squared *residuals* divided by the number of *degrees of freedom*:

$$\sigma_{arepsilon}^2 = rac{\mathbb{E}[arepsilon^T arepsilon]}{d_{free}}$$

Where $d_{free} = (n - k)$ is the number of degrees of freedom of the residuals, equal to the number of observations n, minus the number of predictors k (including the intercept term).

The response vector y can also be considered to be a random variable \hat{y} , equal to the sum of the deterministic fitted values y_{fit} plus the random residuals $\hat{\varepsilon}$:

$$\hat{\mathbf{y}} = \mathbb{X}\boldsymbol{\beta} + \hat{\boldsymbol{\varepsilon}} = \mathbf{y}_{\mathrm{fit}} + \hat{\boldsymbol{\varepsilon}}$$

The regression coefficients β can also be considered to be random variables $\hat{\beta}$:

$$\hat{\beta} = \mathbb{X}_{inv} \hat{y} = \mathbb{X}_{inv} (y_{fit} + \hat{\varepsilon}) = (\mathbb{X}^T \mathbb{X})^{-1} \mathbb{X}^T (\mathbb{X}\beta + \hat{\varepsilon}) = \beta + \mathbb{X}_{inv} \hat{\varepsilon}$$

Where β is equal to the expected value of $\hat{\beta}$:

$$\beta = \mathbb{E}[\hat{\beta}] = \mathbb{X}_{inv} y_{fit} = \mathbb{X}_{inv} y.$$

- > # Regression model summary
- > regsum <- summary(regmod)
- > # Degrees of freedom of residuals
- > nrows <- NROW(predv)
- > ncols <- NCOL(predv)
- > degf <- (nrows ncols)
- > all.equal(degf, regsum\$df[2])
- [1] TRUE
- > # Variance of residuals
- > residsd <- sum(residv^2)/degf

> # Inverse of predictor matrix squared

Covariance Matrix of the Regression Coefficients

The covariance matrix of the regression coefficients $\hat{\beta}$ is given by:

$$\begin{split} \sigma_{\beta}^2 &= \frac{\mathbb{E}[(\hat{\beta} - \beta)(\hat{\beta} - \beta)^T]}{d_{free}} = \\ &\frac{\mathbb{E}[\mathbb{X}_{inv} \hat{\varepsilon}(\mathbb{X}_{inv} \hat{\varepsilon})^T]}{d_{free}} &= \frac{\mathbb{E}[\mathbb{X}_{inv} \hat{\varepsilon} \hat{\varepsilon}^T \mathbb{X}_{inv}^T]}{d_{free}} = \\ &\frac{(\mathbb{X}^T \mathbb{X})^{-1} \mathbb{X}^T \mathbb{E}[\hat{\varepsilon} \hat{\varepsilon}^T] \mathbb{X}(\mathbb{X}^T \mathbb{X})^{-1}}{d_{free}} = \\ (\mathbb{X}^T \mathbb{X})^{-1} \mathbb{X}^T \sigma_{\varepsilon}^2 \mathbb{1} \mathbb{X}(\mathbb{X}^T \mathbb{X})^{-1} = \sigma_{\varepsilon}^2 (\mathbb{X}^T \mathbb{X})^{-1} \end{split}$$

Where the expected values of the squared residuals are proportional to the diagonal unit matrix 1:

$$\frac{\mathbb{E}[\hat{\varepsilon}\hat{\varepsilon}^T]}{d_{free}} = \sigma_{\varepsilon}^2 \mathbb{1}$$

If the predictors are close to being *collinear*, then the squared predictor matrix becomes singular, and the covariance of their regression coefficients becomes very large.

The matrix $\mathbb{X}_{inv} = (\mathbb{X}^T \mathbb{X})^{-1} \mathbb{X}^T$ is the generalized inverse of the *predictor matrix* \mathbb{X} .

```
> predv2 <- MASS::ginv(crossprod(predv))
> # predv2 <- t(predv) /* /* predv
> # Variance of residuals
> residsd <- sum(residv '2)/degf
> # Calculate covariance matrix of betas
> betacovar <- residsd*predv2
> # Round(betacovar, 3)
```

- > betasd <- sqrt(diag(betacovar))
- > all.equal(betasd, regsum\$coeff[, 2], check.attributes=FALSE) [1] TRUE
- > # Calculate t-values of betas
- > betatvals <- drop(betav)/betasd
 > all.equal(betatvals, regsum\$coeff[, 3], check.attributes=FALSE)
- [1] TRUE
- > # Calculate two-sided p-values of betas > betapvals <- 2*pt(-abs(betatvals), df=degf)
- > all.equal(betapvals, regsum\$coeff[, 4], check.attributes=FALSE)
 [1] TRUE
- > # The square of the generalized inverse is equal > # to the inverse of the square
- > all.equal(MASS::ginv(crossprod(predv)), predinv %*% t(predinv))
 [1] TRUE

Covariance Matrix of the Fitted Values

The fitted values yfit can also be considered to be random variables \hat{v}_{fit} , because the regression coefficients $\hat{\beta}$ are random variables:

$$\hat{y}_{fit} = \mathbb{X}\hat{\beta} = \mathbb{X}(\beta + \mathbb{X}_{inv}\hat{\varepsilon}) = y_{fit} + \mathbb{X}\mathbb{X}_{inv}\hat{\varepsilon}.$$

The covariance matrix of the fitted values σ_{fit}^2 is:

$$\sigma_{\mathit{fit}}^2 = \frac{\mathbb{E}[\mathbb{X}\mathbb{X}_{\mathit{inv}}\hat{\varepsilon}\left(\mathbb{X}\mathbb{X}_{\mathit{inv}}\hat{\varepsilon}\right)^T]}{d_{\mathit{free}}} = \frac{\mathbb{E}[\mathbb{H}\,\hat{\varepsilon}\hat{\varepsilon}^T\mathbb{H}^T]}{d_{\mathit{free}}} =$$

$$\frac{\mathbb{H} \ \mathbb{E}[\hat{\varepsilon}\hat{\varepsilon}^T] \ \mathbb{H}^T}{d_{\text{free}}} = \sigma_{\varepsilon}^2 \ \mathbb{H} = \sigma_{\varepsilon}^2 \ \mathbb{X}(\mathbb{X}^T\mathbb{X})^{-1}\mathbb{X}^T$$

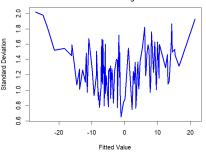
The square of the influence matrix $\mathbb H$ is equal to itself (it's idempotent): $\mathbb{H} \mathbb{H}^T = \mathbb{H}$.

The variance of the *fitted values* σ_{fit}^2 increases with the distance of the predictors from their mean values.

This is because the fitted values farther from their mean are more sensitive to the variance of the regression slope.

- > # Calculate the influence matrix
- > infmat <- predv %*% predinv
- > # The influence matrix is idempotent
- > all.equal(infmat, infmat %*% infmat)

Standard Deviations of Fitted Values in Multivariate Regression



- > # Calculate covariance and standard deviations of fitted values
- > fit.covar <- residsd*infmat
- > fitsd <- sqrt(diag(fitcovar)) > # Sort the standard deviations
- > fitsd <- cbind(fitted=fittedv, stdev=fitsd)
 - > fitsd <- fitsd[order(fittedv),]
 - > # Plot the standard deviations
 - > plot(fitsd, type="1", lwd=3, col="blue",
- xlab="Fitted Value", ylab="Standard Deviation",
- main="Standard Deviations of Fitted Values\nin Multivariate

Standard Errors of Time Series Regression

Bootstrapping the regression of asset returns shows that the actual standard errors can be over twice as large as those reported by the function lm().

This is because the function lm() assumes that the data is normally distributed, while in reality asset returns have very large skewness and kurtosis.

```
> # Load time series of ETF percentage returns
> retsp <- rutils::etfenv$returns[, c("XLF", "XLE")]
> retsp <- na.omit(retsp)
> nrows <- NROW(retsp)
> head(retsp)
> # Define regression formula
> formulav <- paste(colnames(retsp)[1],
    paste(colnames(retsp)[-1], collapse="+"),
    sep=" ~ ")
> # Standard regression
> regmod <- lm(formulav, data=retsp)
> regsum <- summary(regmod)
> # Bootstrap of regression
> set.seed(1121) # initialize random number generator
> bootd <- sapply(1:100, function(x) {
    samplev <- sample.int(nrows, replace=TRUE)
    regmod <- lm(formulav, data=retsp[samplev, ])
    regmod$coefficients
+ }) # end sapply
> # Means and standard errors from regression
> regsum$coefficients
> # Means and standard errors from bootstrap
> dim(bootd)
> t(apply(bootd, MARGIN=1,
+ function(x) c(mean=mean(x), stderror=sd(x))))
```

Predictions From Multivariate Regression Models

The prediction y_{pred} from a regression model is equal to the *response value* corresponding to the *predictor* vector with the new data \mathbb{X}_{new} :

$$y_{pred} = \mathbb{X}_{new} \beta$$

The prediction is a random variable \hat{y}_{pred} , because the regression coefficients $\hat{\beta}$ are random variables:

$$\hat{y}_{pred} = \mathbb{X}_{new} \hat{\beta} = \mathbb{X}_{new} (\beta + \mathbb{X}_{inv} \hat{\varepsilon}) = y_{pred} + \mathbb{X}_{new} \mathbb{X}_{inv} \hat{\varepsilon}$$

The variance σ_{pred}^2 of the predicted value is:

$$\sigma_{pred}^{2} = \frac{\mathbb{E}\left[\mathbb{X}_{new} \mathbb{X}_{inv} \hat{\varepsilon} \left(\mathbb{X}_{new} \mathbb{X}_{inv} \hat{\varepsilon}\right)^{I}\right]}{d_{free}} = \frac{\mathbb{E}\left[\mathbb{X}_{new} \mathbb{X}_{inv} \hat{\varepsilon} \hat{\varepsilon}^{T} \mathbb{X}_{inv}^{T} \mathbb{X}_{new}^{T}\right]}{d_{free}} = \sigma_{\varepsilon}^{2} \mathbb{X}_{new} \mathbb{X}_{inv} \mathbb{X}_{inv}^{T} \mathbb{X}_{new}^{T} = \sigma_{\varepsilon}^{2} \mathbb{X}_{new} \mathbb{X}_{inv} \mathbb{X}_{inv}^{T} \mathbb{X}_{new}^{T} = \sigma_{\varepsilon}^{2} \mathbb{X}_{new} \mathbb{X}_{new}^{T} \mathbb{X}_{new}^{T} = \sigma_{\varepsilon}^{2} \mathbb{X}_{new}^{T} \mathbb{X}_{new}^{T} = \sigma_{\varepsilon}^{2} \mathbb{X}_{new}^{T} \mathbb{X}_{new}^{T} \mathbb{X}_{new}^{T} = \sigma_{\varepsilon}^{2} \mathbb{X}_{new}^{T} \mathbb{X}_{new}^{T} \mathbb{X}_{new}^{T} = \sigma_{\varepsilon}^{2} = \sigma_{\varepsilon}^{2} \mathbb{X}_{new}^{T} = \sigma_{\varepsilon}^{2} = \sigma_{\varepsilon}^{2} = \sigma_{\varepsilon}^$$

The variance σ_{pred}^2 of the predicted value is equal to the predictor vector multiplied by the covariance matrix of the regression coefficients σ_{β}^2 .

> set.seed(1121)
> newdata <- data.frame(matrix(c(1, rnorm(5)), nr=1))
> colnamev <- colnames(predv)
> colnames(newdata) <- colnamev
> newdatav <- as.matrix(newdata)

> predsd <- drop(sqrt(newdatav %*% betacovar %*% t(newdatav)))

> # New data predictor is a data frame or row vector

> prediction <- drop(newdatav %*% betav)

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Predictions From *Multivariate Regression* Using lm()

The function predict() is a *generic function* for forecasting based on a given model.

predict.lm() is the predict method for linear models (regressions) produced by the function lm().

In order for predict.lm() to work properly, the multivariate regression must be specified using a formula.

> # Calculate t-quantile
> tquant <- qt(pnorm(2), df=degf)</pre>

> # Create formula from text string

- > predicthigh <- (prediction + tquant*predsd)
- > predictlow <- (prediction tquant*predsd)
 > # Compare with matrix calculations
- > all.equal(predictlm[1, "fit"], prediction)
- > all.equal(predictlm[1, "lwr"], predictlow)
- > all.equal(predictlm[1, "upr"], predicthigh)

Total Sum of Squares and Explained Sum of Squares

The Total Sum of Squares (TSS) and the Explained Sum of Squares (ESS) are defined as:

$$TSS = (y - \bar{y})^{T}(y - \bar{y})$$

$$ESS = (y_{fit} - \bar{y})^{T}(y_{fit} - \bar{y})$$

$$RSS = (y - y_{fit})^{T}(y - y_{fit})$$

Since the residuals $\varepsilon = v - v_{fit}$ are orthogonal to the

fitted values y_{fit}, they are also orthogonal to the fitted excess values $(y_{fit} - \bar{y})$:

$$(y - y_{fit})^T (y_{fit} - \bar{y}) = 0$$

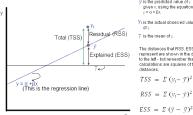
Therefore the *TSS* can be expressed as the sum of the ESS plus the RSS:

$$TSS = ESS + RSS$$

It also follows that the RSS and the ESS follow independent *chi-squared* distributions with (n - k) and (k-1) degrees of freedom.

The degrees of freedom of the Total Sum of Squares is equal to the sum of the RSS plus the ESS:

$$d_{free}^{TSS} = (n-k) + (k-1) = n-1.$$



given x, using the equation Ville the actual observed value The distances that RSS, ESS and TSS represent are shown in the diagram calculations are squares of these

$$RSS = \Sigma (y_t - \hat{y})^2$$

$$ESS = \Sigma (\hat{y} - \hat{y})^2$$

R-squared of Multivariate Regression

The *R-squared* is the fraction of the *Explained Sum of Squares* (*ESS*) divided by the *Total Sum of Squares* (*TSS*):

$$R^2 = \frac{ESS}{TSS} = 1 - \frac{RSS}{TSS}$$

The *R*-squared is a measure of the model goodness of fit, with *R*-squared close to 1 for models fitting the data very well, and *R*-squared close to 0 for poorly fitting models.

The *R-squared* is equal to the squared correlation between the response and the *fitted values*:

$$\rho_{yy_{fit}} = \frac{\left(y_{fit} - \bar{y}\right)^{T} \left(y - \bar{y}\right)}{\sqrt{TSS \cdot ESS}} = \frac{\left(y_{fit} - \bar{y}\right)^{T} \left(y_{fit} - \bar{y}\right)}{\sqrt{TSS \cdot ESS}} = \sqrt{\frac{ESS}{TSS}}$$

- > # Set regression attribute for intercept
- > attributes(regmod\$terms)\$intercept <- 1
- > # Regression summary
- > regsum <- summary(regmod)
- > # Regression R-squared
- > rsquared <- ess/tss
- > all.equal(rsquared, regsum\$r.squared)
- [1] TRUE
- > # Correlation between response and fitted values
- > cor_fitted <- drop(cor(respv, fittedv))
- > # Squared correlation between response and fitted values
- > all.equal(cor_fitted^2, rsquared)
- [1] TRUE

Adjusted R-squared of Multivariate Regression

The weakness of *R-squared* is that it increases with the number of predictors (even for predictors which are purely random), so it may provide an inflated measure of the quality of a model with many predictors.

This is remedied by using the *residual variance* $\left(\sigma_{\varepsilon}^{2} = \frac{RSS}{d_{free}}\right)$ instead of the *RSS*, and the *response variance* $\left(\sigma_{v}^{2} = \frac{TSS}{B-1}\right)$ instead of the *TSS*.

The adjusted R-squared is equal to 1 minus the fraction of the residual variance divided by the response variance:

$$R_{adj}^2 = 1 - rac{\sigma_{arepsilon}^2}{\sigma_y^2} = 1 - rac{RSS/d_{free}}{TSS/(n-1)}$$

Where $d_{free} = (n - k)$ is the number of degrees of freedom of the residuals.

The adjusted R-squared is always smaller than the R-squared.

The performance of two different models can be compared by comparing their adjusted R-squared, since the model with the larger adjusted R-squared has a smaller residual variance, so it's better able to explain the response.

```
> nrous <- NROW(predv)
> ncols <- NCOL(predv)
> # Degrees of freedom of residuals
> degf <- (nrows - ncols)
> # Adjusted R-squared
> rsqadj <- (1-sum(residv^2)/degf/var(respv))
> # Compare adjusted R-squared from lm()
> all.equal(drop(rsqadj), regsum$adj.r.squared)
[11 TRUE
```

Fisher's F-distribution

Let χ^2_m and χ^2_n be independent random variables following chi-squared distributions with m and n degrees of freedom.

Then the F-statistic random variable:

$$F = \frac{\chi_m^2/m}{\chi_n^2/n}$$

Follows the F-distribution with m and n degrees of freedom, with the probability density function:

$$P(F) = \frac{\Gamma((m+n)/2)m^{m/2}n^{n/2}}{\Gamma(m/2)\Gamma(n/2)} \frac{F^{m/2-1}}{(n+mF)^{(m+n)/2}}$$

The F-distribution depends on the F-statistic F and also on the degrees of freedom, m and n.

The function ${\tt df}$ () calculates the probability density of the F-distribution.

- F-Distributions

 Gegrees of freedom
 Gegrees of free
- > # Add title
- > title(main="F-Distributions", line=0.5)
- > # Add legend
 > labely <- paste("df", degf, sep="=")</pre>
- > labelv <- paste("df", degf, sep="=")
- > legend("topright", inset=0.05, title="degrees of freedom",
 - y.intersp=0.5, bty="n", labelv, cex=0.8, lwd=2, lty=1, col

[9] 0.001888 0.000793 0.000329

The F-test For the Variance Ratio

Let x and y be independent standard *Normal* variables, and let $\sigma_x^2 = \frac{1}{m-1} \sum_{i=1}^m (x_i - \bar{x})^2$ and

$$\sigma_y^2 = \frac{1}{n-1} \sum_{i=1}^n (y_i - \bar{y})^2$$
 be their sample variances.

The ratio $F = \sigma_x^2/\sigma_y^2$ of the sample variances follows the *F*-distribution with m and n degrees of freedom.

The *F*-test tests the *null hypothesis* that the *F*-statistic *F* is not significantly greater than 1 (the variance σ_x^2 is not significantly greater than σ_y^2).

A large value of the *F-statistic F* indicates that the variances are unlikely to be equal.

The function pf(q) returns the cumulative probability of the *F-distribution*, i.e. the cumulative probability that the *F-statistic F* is less than the quantile q.

This *F-test* is very sensitive to the assumption of the normality of the variables.

```
> sigmax <- var(rnorm(nrows))
> sigmay <- var(rnorm(nrows))
> fratio <- sigmax/sigmay
> # Cumulative probability for q = fratio
> pf(fratio, nrows-1, nrows-1)
[i] 0.0642
> # p-value for fratios
> -pf(fratios) /- nrows-1, nrows-1)
```

[1] 0.500000 0.318150 0.182964 0.096784 0.047876 0.022467 0.010123

The *F-statistic* for Linear Regression

The performance of two different regression models can be compared by directly comparing their *Residual Sum of Squares* (*RSS*), since the model with a smaller *RSS* is better able to explain the *response*.

Let the restricted model have p_1 parameters with $df_1 = n - p_1$ degrees of freedom, and the unrestricted model have p_2 parameters with $df_2 = n - p_2$ degrees of freedom, with $p_1 > p_2$.

Then the F-statistic F, defined as the ratio of the scaled Residual Sum of Squares:

$$F = \frac{(RSS_1 - RSS_2)/(df_1 - df_2)}{RSS_2/df_2}$$

Follows the F-distribution with (p_2-p_1) and $(n-p_2)$ degrees of freedom (assuming that the residuals are normally distributed).

If the restricted model only has one parameter (the constant intercept term), then $df_1 = n - 1$, and its fitted values are equal to the average of the response: $y_i^{fit} = \overline{y}$, so RSS₁ is equal to the TSS:

 $RSS_1 = TSS = (y - \bar{y})^2$, so its Explained Sum of Squares is equal to zero: $ESS_1 = TSS - RSS_1 = 0$.

Let the *unrestricted* multivariate regression model be defined as:

$$y = X\beta + \varepsilon$$

Where y is the response, $\mathbb X$ is the predictor matrix (with k predictors, including the intercept term), and β are the k regression coefficients.

So the *unrestricted* model has k parameters ($p_2 = k$), and $RSS_2 = RSS$ and $ESS_2 = ESS$, and then the F-statistic can be written as:

$$F = \frac{ESS/(k-1)}{RSS/(n-k)}$$

> # p-value of F-statistic

[1] 0.00757

The F-test for Linear Regression

The Residual Sum of Squares RSS = $\varepsilon^T \varepsilon$ and the Explained Sum of Squares ESS = $(y_{\rm fit} - \bar{y})^T (y_{\rm fit} - \bar{y})$ follow independent *chi-squared* distributions with (n-k) and (k-1) degrees of freedom.

Then the *F*-statistic, equal to the ratio of the *ESS* divided by *RSS*:

$$F = \frac{ESS/(k-1)}{RSS/(n-k)}$$

Follows the *F-distribution* with (k-1) and (n-k) degrees of freedom (assuming that the *residuals* are normally distributed).

```
> # F-statistic from lm()
> regsum$fstatistic
value numdf dendf
3.37 5.00 94.00
> # Degrees of freedom of residuals
> degf <- (nrows - ncols)
> # F-statistic from ESS and RSS
> fstat <- (ess/(ncols-1))/(rss/degf)
> all.equal(fstat, regsum$fstatistic[1], check.attributes=FALSE)
```

> 1-pf(q=fstat, df1=(ncols-1), df2=(nrows-ncols))

Regularized Inverse of Rectangular Matrices

The SVD of a rectangular matrix $\mathbb A$ is defined as the factorization:

$$\mathbb{A} = \mathbb{U}\Sigma\mathbb{V}^T$$

Where $\mathbb U$ and $\mathbb V$ are the singular matrices, and Σ is a diagonal matrix of singular values.

The generalized inverse matrix \mathbb{A}^{-1} satisfies the inverse equation: $\mathbb{A}\mathbb{A}^{-1}\mathbb{A}=\mathbb{A}$, and it can be expressed as a product of the SVD matrices as follows:

$$\mathbb{A}^{-1} = \mathbb{V} \, \Sigma^{-1} \, \mathbb{U}^{\mathsf{T}}$$

If any of the *singular values* are zero then the *generalized inverse* does not exist.

The *regularized inverse* is obtained by removing very small *singular values*:

$$\mathbb{A}^{-1} = \mathbb{V}_n \, \Sigma_n^{-1} \, \mathbb{U}_n^T$$

Where \mathbb{U}_n , \mathbb{V}_n and Σ_n are the *SVD* matrices without very small *singular values*.

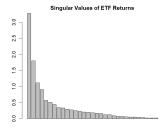
The regularized inverse satisfies the inverse equation only approximately (it has *bias*), but it's often used in machine learning because it has lower *variance* than the exact inverse

> # Calculate ETF returns

> retsp <- na.omit(rutils::etfenv\$returns)
> # Perform singular value decomposition

> svdec <- svd(retsp)

> barplot(svdec\$d, main="Singular Values of ETF Returns")



- > # Calculate generalized inverse from SVD > invmat <- svdec\$v %*% (t(svdec\$u) / svdec\$d)
- > # Verify inverse property of inverse
- > all.equal(zoo::coredata(retsp), retsp %*% invmat %*% retsp)
 > # Calculate regularized inverse from SVD
- > # Calculate regularized inverse from SV!
 > dimax <- 1:3</pre>
- > invreg <- svdec\$v[, dimax] %*%
 + (t(svdec\$u[, dimax]) / svdec\$d[dimax])</pre>
- > # Calculate regularized inverse using RcppArmadillo
- > invcpp <- HighFreq::calc_inv(retsp, dimax=3)
 > all.equal(invreg, invcpp, check.attributes=FALSE)
- > all.equal(invreg, invcpp, check.attributes=FALSE)
 > # Calculate regularized inverse from Moore-Penrose pseudo-inverse
- > retsq <- t(retsp) %*% retsp > eigend <- eigen(retsq)
- > inv2 <- eigend\$vectors[, dimax] %*%
 + (t(eigend\$vectors[, dimax]) / eigend\$values[dimax])</pre>
- > invmp <- inv2 %*% t(retsp)

> all.equal(invreg, invmp, check.attributes=FALSE)

Linear Transformation of the Predictor Matrix

A *multivariate* linear regression model can be transformed by replacing its *predictors* x_j with their own linear combinations.

This is equivalent to multiplying the *predictor matrix* \mathbb{X} by a transformation matrix \mathbb{W} :

$$X_{trans} = X W$$

The transformed predictor matrix \mathbb{X}_{trans} produces the same influence matrix \mathbb{H} as the original predictor matrix \mathbb{X} .

$$\begin{aligned} &\mathbb{H}_{trans} = \mathbb{X}_{trans} (\mathbb{X}_{trans}^{T} \mathbb{X}_{trans})^{-1} \mathbb{X}_{trans}^{T} = \\ &\mathbb{X} \mathbb{W} (\mathbb{W}^{T} \mathbb{X}^{T} \mathbb{X} \mathbb{W})^{-1} \mathbb{W}^{T} \mathbb{X}^{T} = \\ &\mathbb{X} \mathbb{W} \mathbb{W}^{-1} (\mathbb{X}^{T} \mathbb{X})^{-1} \mathbb{W}^{T-1} \mathbb{W}^{T} \mathbb{X}^{T} = \\ &\mathbb{X} (\mathbb{X}^{T} \mathbb{X})^{-1} \mathbb{X}^{T} = \mathbb{H} \end{aligned}$$

Since the *influence matrix* $\mathbb H$ is the same, the transformed regression model produces the same *fitted values* and *residuals* as the original regression model, so it's equivalent to it.

- > # Define transformation matrix
- > matv <- matrix(runif(ncols^2, min=(-1), max=1), ncol=ncols)
- > # Calculate linear combinations of predictor columns > predt <- predv %*% matv
- > # Calculate the influence matrix of the transformed predictor
- > influencet <- predt %*% MASS::ginv(predt)
- > # Compare the influence matrices
- > all.equal(infmat, influencet)
- [1] TRUE

Principal Component Regression

In Principal Component Regression (PCR), the predictor matrix $\mathbb X$ is multiplied by the PCA rotation matrix $\mathbb W$:

$$X_{pca} = XW$$

So that the principal component vectors form the columns of the new predictor matrix.

Since the new *PCR* predictors x_i^{pca} are orthogonal, the regression coefficients are simply:

$$\beta_i = \frac{x_i^{pca} \cdot y}{\sigma_i^2}$$

Where $x_i^{pca} \cdot y$ are the inner products of the *PCR* predictors x_i^{pca} times the *response vector y*, and $\sigma_i^2 = x_i^{pca} \cdot x_i^{pca}$ are the inner products (sum of squares) of the predictors x_i^{pca} .

- > # Perform PCA of the predictors
- > pcad <- prcomp(predv, center=FALSE, scale=FALSE)
- > # Calculate the PCA predictors
- > predpca <- predv %*% pcad\$rotation
 > # Principal components are orthogonal to each other
- > round(t(predpca) %*% predpca, 2)
- > # Calculate the PCA influence matrix
- > infmat <- predv %*% MASS::ginv(predv)
- > infpca <- predpca %*% MASS::ginv(predpca)
- > all.equal(infmat, infpca)
- > # Calculate the regression coefficients
 > coeffy <- drop(MASS::ginv(predy) %*% respy)</pre>
- > # Transform the collinear regression coefficients to the PCA
- > drop(coeffv %*% pcad\$rotation)
- > # Calculate the PCA regression coefficients
 > drop(MASS::ginv(predpca) %*% respv)
- > # Calculate the PCA regression coefficients directly
- > # Calculate the PCA regression coefficients directly
 > colSums(predpca*drop(respv))/colSums(predpca^2)

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> # Create almost collinear predictors

Dimension Reduction Using Principal Component Regression

If the predictor columns are *collinear* then some of the *PCR* predictor squares are zero $\sigma_i^2 = 0$, and the associated regression coefficients are infinite (indeterminate) and should be discarded.

The regression can also become *singular* if the number of rows of the predictor is too small, or is even less than the number of its columns.

The regression can be *regularized* by removing the infinite or very large *PCR* regression coefficients, and transforming the coefficients back to the original predictor coordinates.

This is called *dimension reduction* - excluding the principal components with very small squares.

Dimension reduction can also be applied to reduce model overfitting by reducing the number of effective predictors.

```
> predc <- predv
> predc[, 1] <- (predc[, 1]/1e3 + predc[, 2])
> # Calculate the collinear regression coefficients
> coeffv <- drop(MASS::ginv(predc) %*% respv)
> coeffv
> # Calculate the PCA predictors
> pcad <- prcomp(predc, center=FALSE, scale=FALSE)
> predpca <- predc %*% pcad$rotation
> round(t(predpca) %*% predpca, 6)
```

- > # Transform the collinear regression coefficients to the PCA > drop(coeffv %*% pcad\$rotation)
- > # Calculate the PCA regression coefficients
- > coeffpca <- drop(MASS::ginv(predpca) %*% respv)
 > # Calculate the PCA regression coefficients directly
- > colSums(predpca*drop(respv))/colSums(predpca^2)
- > # Transform the PCA regression coefficients to the original coord > drop(coeffpca %*% MASS::ginv(pcad\$rotation))
 - > coeffv
 > # Calculate the regression coefficients after dimension reduction
 > npca <- NROW(coeffpca)</pre>
- > drop(coeffpca[-npca] %*% MASS::ginv(pcad\$rotation)[-npca,])
- > # Compare with the collinear regression coefficients
 - > coeffv
 - > # Calculate the original regression coefficients
 - > drop(MASS::ginv(predv) %*% respv)

The Logistic Function

The *logistic* function expresses the probability of a numerical variable ranging over the whole interval of real numbers:

$$p(x) = \frac{1}{1 + \exp(-\lambda x)}$$

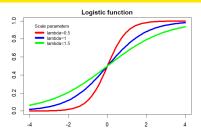
Where λ is the scale (dispersion) parameter.

The *logistic* function is often used as an activation function in neural networks, and logistic regression can be viewed as a perceptron (single neuron network).

The *logistic* function can be inverted to obtain the *Odds Ratio* (the ratio of probabilities for favorable to unfavorable outcomes):

$$\frac{p(x)}{1-p(x)}=\exp(\lambda x)$$

The function plogis() gives the cumulative probability of the *Logistic* distribution,



Performing Logistic Regression Using the Function glm()

Logistic regression (logit) is used when the response are discrete variables (like factors or integers), when linear regression can't be applied.

The function glm() fits generalized linear models, including *logistic* regressions.

The parameter family=binomial(logit) specifies a binomial distribution of residuals in the *logistic* regression model.

The Mann-Whitney test null hypothesis is that the two samples, x_i and y_i , were obtained from probability distributions with the same median (location).

The function wilcox.test() with parameter paired=FALSE (the default) calculates the Mann-Whitney test statistic and its p-value.

```
> set.seed(1121) # Reset random number generator
> # Simulate overlapping scores data
> sample1 <- runif(100, max=0.6)
> sample2 <- runif(100, min=0.4)
> # Perform Mann-Whitney test for data location
> wilcox.test(sample1, sample2)
> # Combine scores and add categorical variable
> predv <- c(cample1, sample2)
> respv <- c(clogical(100), !logical(100))
> # Perform logit regression
> logmod <- glm(respv ~ predv, family=binomial(logit))
> class(logmod)
> summary(logmod)
```



Category Densities and Logistic Function

```
> ordern <- order(predv)
 plot(x=predv[ordern], y=logmod$fitted.values[ordern],
       main="Category Densities and Logistic Function".
       type="1", lwd=4, col="orange", xlab="predictor", vlab="densi
> densityv <- density(predv[response])
> densitvv$v <- densitvv$v/max(densitvv$v)
> lines(densityv, col="red")
> polygon(c(min(densityv$x), densityv$x, max(densityv$x)), c(min(densityv$x)
> densityv <- density(predv[!response])
> densityv$y <- densityv$y/max(densityv$y)
> lines(densityv, col="blue")
> polygon(c(min(densityv$x), densityv$x, max(densityv$x)), c(min(densityv$x)
> # Add legend
> legend(x="top", cex=1.0, bty="n", lty=c(1, NA, NA),
+ lwd=c(6, NA, NA), pch=c(NA, 15, 15), y.intersp=0.5,
+ legend=c("logistic fit", "TRUE", "FALSE"),
+ col=c("orange", "red", "blue"),
```

+ text.col=c("black", "red", "blue"))

The Likelihood Function of the Binomial Distribution

Let b be a binomial random variable, which either has the value b=1 with probability p, or b=0 with probability (1-p).

Then b follows the binomial distribution:

$$f(b) = b p + (1 - b) (1 - p)$$

The log-likelihood function $\mathcal{L}(p|b)$ of the probability p given the value b is obtained from the logarithms of the binomial probabilities:

$$\mathcal{L}(p|b) = b \log(p) + (1-b) \log(1-p)$$

The *log-likelihood* function measures how *likely* are the distribution parameters, given the observed values.

The Likelihood Function of the Logistic Model

Let b; be binomial random variables, with probabilities p_i that depend on the numerical variables s_i through the logistic function:

$$p_i = \frac{1}{1 + \exp(-\lambda_0 - \lambda_1 s_i)}$$

Let's assume that the b; and s; values are known (observed), and we want to find the parameters λ_0 and λ_1 that best fit the observations.

The log-likelihood function \mathcal{L} is equal to the sum of the individual log-likelihoods:

$$\mathcal{L}(\lambda_0, \lambda_1 | b_i) = \sum_{i=1}^n b_i \log(p_i) + (1-b_i) \log(1-p_i)$$

The log-likelihood function measures how likely are the distribution parameters, given the observed values.

- > # Specify predictor matrix
- > predv=cbind(intercept=rep(1, NROW(respv)), predv) > # Likelihood function of the logistic model
- > likefun <- function(coeff, respv, predv) {
- probs <- plogis(drop(predv %*% coeff)) -sum(respv*log(probs) + (1-respv)*log((1-probs)))
- + } # end likefun
- > # Run likelihood function > coeff <- c(1, 1)
- > likefun(coeff, respv, predv)

Multi-dimensional Optimization Using optim()

The function optim() performs multi-dimensional optimization.

The argument fn is the objective function to be minimized

The argument of fn that is to be optimized, must be a vector argument. The argument par is the initial vector argument value.

optim() accepts additional parameters bound to the dots "..." argument, and passes them to the fn objective function.

The arguments lower and upper specify the search range for the variables of the objective function fn.

method="L-BFGS-B" specifies the quasi-Newton gradient optimization method.

optim() returns a list containing the location of the minimum and the objective function value.

The gradient methods used by optim() can only find the local minimum, not the global minimum.

```
> # Rastrigin function with vector argument for optimization
> rastrigin <- function(vectorv, param=25) {
   sum(vectorv^2 - param*cos(vectorv))
+ } # end rastrigin
> vectorv <- c(pi/6, pi/6)
> rastrigin(vectorv=vectorv)
> # Draw 3d surface plot of Rastrigin function
> options(rgl.useNULL=TRUE); library(rgl)
> rgl::persp3d(
+ x=Vectorize(function(x, y) rastrigin(vectorv=c(x, y))),
+ xlim=c(-10, 10), ylim=c(-10, 10),
   col="green", axes=FALSE, zlab="", main="rastrigin")
> # Render the 3d surface plot of function
> rgl::rglwidget(elementId="plot3drgl", width=400, height=400)
> # Optimize with respect to vector argument
> optiml <- optim(par=vectorv, fn=rastrigin,
          method="L-BFGS-B".
         upper=c(4*pi, 4*pi),
          lower=c(pi/2, pi/2),
          param=1)
> # Optimal parameters and value
> optiml$par
> optiml$value
> rastrigin(optiml$par, param=1)
```

Maximum Likelihood Calibration of the Logistic Model

The logistic model depends on the unknown parameters λ_0 and λ_1 , which can be calibrated by maximizing the likelihood function.

The function optim() with the argument hessian=TRUE returns the Hessian matrix.

The Hessian is a matrix of the second-order partial derivatives of the likelihood function with respect to the optimization parameters:

$$H = \frac{\partial^2 \mathcal{L}}{\partial \lambda^2}$$

The Hessian matrix measures the convexity of the likelihood surface - it's large if the likelihood surface is highly convex, and it's small if the likelihood surface is flat.

If the likelihood surface is highly convex, then the coefficients can be determined with greater precision, so their standard errors are small. If the likelihood surface is flat, then the coefficients have large standard errors.

The inverse of the Hessian matrix provides the standard errors of the logistic parameters: $\sigma_{SF} = \sqrt{H^{-1}}$.

upper=c(20, 20), # Upper constraint

lower=c(-20, -20), # Lower constraint

- + hessian=TRUE)
 > # Optimal logistic parameters
- > optiml\$par
- > unname(logmod\$coefficients)
 > # Standard errors of parameters
- > sqrt(diag(solve(optiml\$hessian)))
- > regsum <- summary(logmod)
 > regsum\$coefficients[, 2]

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Package ISLR With Datasets for Machine Learning

The package *ISLR* contains datasets used in the book *Introduction to Statistical Learning* by Gareth James, Daniela Witten, Trevor Hastie, and Robert Tibshirani.

The book introduces machine learning techniques using R, and it's a must for advanced finance applications.

```
> library(ISLR) # Load package ISLR
> # get documentation for package tseries
> packageDescription("ISLR") # get short description
> help(package="ISLR") # Load help page
> library(ISLR) # Load package ISLR
> data(package="ISLR") # list all datasets in ISLR
> ls("package:ISLR") # list all objects in ISLR
> detach("package:ISLR") # Remove ISLR from search path
```

The Default Dataset

The data frame Default in the package *ISLR* contains credit default data.

The Default data frame contains two columns of categorical data (factors): default and student, and two columns of numerical data: balance and income.

The columns default and student contain factor data, and they can be converted to Boolean values, with TRUE if default == "Yes" and student == "Yes", and FALSE otherwise.

This avoids implicit coercion by the function glm().

- > # Coerce the student and default columns into Boolean
- > Default <- ISLR::Default
- > Default\$default <- (Default\$default == "Yes")
- > Default\$student <- (Default\$student == "Yes")
 > colnames(Default)[1:2] <- c("default", "student")
- > attach(Default) # Attach Default to search path
- > # Explore credit default data
- > summary(Default)

default	student	balance	income
Mode :logical	Mode :logical	Min. : 0	Min. : 772
FALSE:9667	FALSE:7056	1st Qu.: 482	1st Qu.:21340
TRUE :333	TRUE :2944	Median: 824	Median :34553
		Mean : 835	Mean :33517
		3rd Qu.:1166	3rd Qu.:43808
		May .2654	May .7355/

- > sapply(Default, class)
- default student balance income
- "logical" "logical" "numeric" "numeric"
- > dim(Default)
- [1] 10000 4
- > head(Default)

	deraurt	Student	Dalance	THCOME	
1	FALSE	FALSE	730	44362	
2	FALSE	TRUE	817	12106	
3	FALSE	FALSE	1074	31767	
4	FALSE	FALSE	529	35704	
5	FALSE	FALSE	786	38463	
c	CALCE	TRITE	020	7/02	

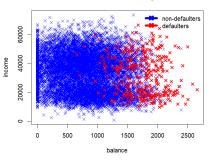
The Dependence of default on The balance and income

The columns student, balance, and income can be used as *predictors* to predict the default column.

The scatterplot of income versus balance shows that the balance column is able to separate the data points of default = TRUE from default = FALSE.

But there is very little difference in income between the default = TRUE versus default = FALSE data points.

Default Dataset from Package ISLR



Boxplots of the Default Dataset

A Box Plot (box-and-whisker plot) is a graphical display of a distribution of data:

The box represents the upper and lower quartiles. The vertical lines (whiskers) represent values beyond the quartiles,

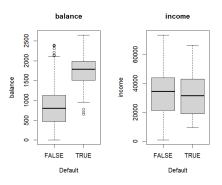
Open circles represent values beyond the nominal range (outliers).

The function boxplot() plots a box-and-whisker plot for a distribution of data

boxplot() has two methods: one for formula objects (involving categorical variables), and another for data frames

The Mann-Whitney test shows that the balance column provides a strong separation between defaulters and non-defaulters, but the income column doesn't.

- > # Perform Mann-Whitney test for the location of the balances > wilcox.test(balance[default], balance[!default])
- > # Perform Mann-Whitney test for the location of the incomes
- > wilcox.test(income[default], income[!default])



- > x11(width=6, height=5)
- > # Set 2 plot panels
- > par(mfrow=c(1,2)) > # Balance boxplot
- > boxplot(formula=balance ~ default.
- col="lightgrev", main="balance", xlab="Default")
- > # Income boxplot
- > boxplot(formula=income ~ default.
- col="lightgrey", main="income", xlab="Default")

Modeling Credit Defaults Using Logistic Regression

The balance column can be used to calculate the probability of default using *logistic* regression.

The residuals are the differences between the actual response values (0 and 1), and the calculated probabilities of default.

The residuals are not normally distributed, so the data is fitted using the *maximum likelihood* method, instead of least squares.

> # Fit logistic regression model

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```
> logmod <- glm(default ~ balance, family=binomial(logit))
> class(logmod)
[1] "glm" "lm"
> summary(logmod)
Call:
glm(formula = default ~ balance, family = binomial(logit))
Deviance Residuals:
  Min
           10 Median
                                   May
-2.270 -0.146 -0.059 -0.022
                                3.759
Coefficients:
             Estimate Std. Error z value Pr(>|z|)
(Intercept) -10.65133
                        0.36116
                                   -29 5
                                          <20-16 ***
balance
             0.00550
                        0.00022
                                   24.9
                                         <2e-16 ***
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' '1
(Dispersion parameter for binomial family taken to be 1)
   Null deviance: 2920.6 on 9999 degrees of freedom
```

Residual deviance: 1596.5 on 9998 degrees of freedom

```
> x11(width=6, height=5)

> par(mar=c(4, 4, 2, 2), oma=c(0, 0, 0, 0), mgp=c(2.5, 1, 0))

> plot(x=balance, y=default,
```

- main="Logistic Regression of Credit Defaults",
- + col="orange", xlab="credit balance", ylab="defaults")
 > ordern <- order(balance)
- > lines(x=balance[ordern], y=logmod\$fitted.values[ordern], col="blu
- > legend(x="topleft", inset=0.1, bty="n", lwd=6, y.intersp=0.5,

99 / 118

- + legend=c("defaults", "logit fitted values"),
- + col=c("orange", "blue"), lty=c(NA, 1), pch=c(1, NA))

Modeling Cumulative Defaults Using Logistic Regression

The function glm() can model a *logistic* regression using either a Boolean response variable, or using a response variable specified as a frequency.

In the second case, the response variable should be defined as a two-column matrix, with the cumulative frequency of success (TRUE) and a cumulative frequency of failure (FALSE).

These two different ways of specifying the *logistic* regression are related, but they are not equivalent, because they have different error terms.

```
> # Calculate cumulative defaults

> sumd <- sum(default)

> defaultv <- sapply(balance, function(balv) {

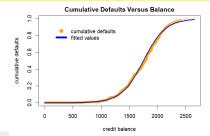
+ sum(default[balance <= balv])

+ }) # end sapply

> # Perform logit regression

> logmod <- glm(cbind(defaultv, sumd-defaultv) - balance,

+ family=binomial(logiti)
```



- > plot(x=balance, y=defaultv/sumd, col="orange", lwd=1,
- + main="Cumulative Defaults Versus Balance",
 + xlab="credit balance", ylab="cumulative defaults")
- > ordern <- order(balance)
- > ordern <- order(balance)
 > lines(x=balance[ordern], y=logmod\$fitted.values[ordern],
- + col="blue", lwd=3)
 - + COI="DIUE", IWG=3)
- > legend(x="topleft", inset=0.1, bty="n", y.intersp=0.5,
- + legend=c("cumulative defaults", "fitted values"),
- + col=c("orange", "blue"), ltv=c(NA, 1), pch=c(1, NA), lwd=6)

> summary(logmod)

> # Fit multifactor logistic regression model

Multifactor Logistic Regression

Logistic regression calculates the probability of categorical variables, from the Odds Ratio of continuous predictors:

$$p = \frac{1}{1 + \exp(-\lambda_0 - \sum_{i=1}^n \lambda_i x_i)}$$

The *generic* function summary() produces a list of regression model summary and diagnostic statistics:

- coefficients: matrix with estimated coefficients, their z-values, and p-values,
- Null deviance: measures the differences between the response values and the probabilities calculated using only the intercept,
- Residual deviance: measures the differences between the response values and the model probabilities,

The balance and student columns are statistically significant, but the income column is not.

```
> colnamev <- colnames(Default)
> formulav <- as.formula(paste(colnamev[1],
   paste(colnamev[-1], collapse="+"), sep=" ~ "))
> formulay
default " student + balance + income
> logmod <- glm(formulav, data=Default, family=binomial(logit))
> summary(logmod)
Call:
glm(formula = formulav, family = binomial(logit), data = Default)
Deviance Residuals:
   Min 10 Median 30
-2.469 -0.142 -0.056 -0.020 3.738
Coefficients:
            Estimate Std. Error z value Pr(>|z|)
(Intercept) -1.09e+01 4.92e-01 -22.08
                                        <2e-16 ***
studentTRUE -6.47e-01 2.36e-01 -2.74
                                        0.0062 **
balance
            5.74e-03 2.32e-04 24.74
                                         <2e-16 ***
income
            3 03e-06 8 20e-06
                                  0.37
                                        0 7115
Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
(Dispersion parameter for binomial family taken to be 1)
```

Null deviance: 2920.6 on 9999 degrees of freedom Residual deviance: 1571.5 on 9996 degrees of freedom

Number of Fisher Scoring iterations: 8

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Cumulative defaults of

students and non-students

Confounding Variables in Multifactor Logistic Regression

The student column alone can be used to calculate the probability of default using single-factor *logistic* regression.

But the coefficient from the single-factor regression is positive (indicating that students are more likely to default), while the coefficient from the multifactor regression is negative (indicating that students are less likely to default).

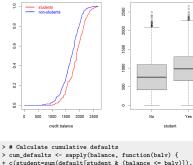
The reason that students are more likely to default is because they have higher credit balances than non-students - which is what the single-factor regression shows.

But students are less likely to default than non-students that have the same credit balance - which is what the multifactor model shows.

The student column is a confounding variable since it's correlated with the balance column.

That's why the multifactor regression coefficient for student is negative, while the single factor coefficient for student is positive.

- > # Fit single-factor logistic model with student as predictor
 > glm_student <- glm(default ~ student, family=binomial(logit))</pre>
- > summary(glm_student)
- > # Multifactor coefficient is negative
- > logmod\$coefficients
- > # Single-factor coefficient is positive
- > glm_student\$coefficients



```
non student=sum(default[!student & (balance <= balv)]))
+ }) # end sapply
> total defaults <- c(student=sum(student & default).
        student=sum(!student & default))
> cum defaults <- t(cum defaults / total defaults)
> # Plot cumulative defaults
> par(mfrow=c(1,2)) # Set plot panels
> ordern <- order(balance)
> plot(x=balance[ordern], v=cum defaults[ordern, 1],
       col="red", t="1", lwd=2, xlab="credit balance", vlab="",
       main="Cumulative defaults of\n students and non-students")
> lines(x=balance[ordern], v=cum defaults[ordern, 2], col="blue", 1
> legend(x="topleft", btv="n", v.intersp=0.5.
+ legend=c("students", "non-students"),
   col=c("red", "blue"), text.col=c("red", "blue"), lwd=3)
> # Balance boxplot for student factor
```

halance

draft: Modeling Credit Defaults Using Student Status

The student column can be used to calculate the probability of default using *logistic* regression.

Persons who are students are more likely to default because students have higher credit balances.

```
> # Fit logistic regression model
```

> summary(logmod)

```
> x11(width=6, height=5)
> par(mfrow=c(1,2)) # Set plot panels
> # Balance boxplot
> boxplot(formula=balance ~ default,
   col="lightgrey", main="balance", xlab="Default")
> # Plot data points for non-students
> x11(width=6, height=5)
> xlim <- range(balance); ylim <- range(income)
> plot(income ~ balance,
       main="Default Dataset from Package ISLR",
       xlim=xlim, ylim=ylim, pch=4, col="blue",
       data=Default[!student, ])
> # Plot data points for students
> points(income ~ balance, pch=4, lwd=2, col="red",
+ data=Default[student, ])
> # Add legend
> legend(x="topright", bty="n", y.intersp=0.5,
+ legend=c("non-students", "students"),
+ col=c("blue", "red"), ltv=1, lwd=6, pch=4)
```

> logmod <- glm(default ~ student, family=binomial(logit))

Forecasting Credit Defaults using Logistic Regression

The function predict() is a generic function for forecasting based on a given model.

The method predict.glm() produces forecasts for a generalized linear (glm) model, in the form of numeric probabilities, not the Boolean response variable.

The Boolean forecasts are obtained by comparing the forecast probabilities with a discrimination threshold.

Let the null hypothesis be that the subject will not default: default = FALSE.

If the forecast probability is less than the discrimination threshold, then the forecast is that the subject will not default and that the null hypothesis is TRUE.

The in-sample forecasts are just the fitted values of the glm model.

```
> # Perform in-sample forecast from logistic regression model
> fcast <- predict(logmod, type="response")
> all.equal(logmod$fitted.values, fcast)
[1] TRUE
> # Define discrimination threshold value
> threshold <- 0.7
> # Calculate confusion matrix in-sample
> table(actual=!default, forecast=(fcast < threshold))
       forecast
actual FALSE TRUE
  FALSE
           57 276
           12 9655
  TRUE
> # Fit logistic regression over training data
> set.seed(1121) # Reset random number generator
> nrows <- NROW(Default)
> samplev <- sample.int(n=nrows, size=nrows/2)
> trainset <- Default[samplev, ]
> logmod <- glm(formulav, data=trainset, family=binomial(logit))
> # Forecast over test data out-of-sample
> testset <- Default[-samplev, ]
> fcast <- predict(logmod, newdata=testset, type="response")
> # Calculate confusion matrix out-of-sample
> table(actual=!testset$default, forecast=(fcast < threshold))
       forecast
```

actual FALSE TRUE FALSE

TRUE

29 132

9 4830

Γ17 9

Forecasting Errors

A binary classification model categorizes cases based on its forecasts whether the *null hypothesis* is TRUE or FALSE.

Let the *null hypothesis* be that the subject will not default: default = FALSE.

A *positive* result corresponds to rejecting the null hypothesis, while a *negative* result corresponds to accepting the null hypothesis.

The forecasts are subject to two different types of errors: *type I* and *type II* errors.

A *type I* error is the incorrect rejection of a TRUE *null hypothesis* (i.e. a "false positive"), when there is no default but it's classified as a default.

A type II error is the incorrect acceptance of a FALSE null hypothesis (i.e. a "false negative"), when there is a default but it's classified as no default.

```
> # Calculate confusion matrix out-of-sample

> confmat <- table(actual=!testset$default,

+ forecast+cfcast < threshold))

> confmat

forecast

actual FALSE TRUE

FALSE 29 132

TRUE 9 4830

> # Calculate FALSE positive (type I error)

> sum(!testset$default.k.(fcast.> threshold))
```

The Confusion Matrix of a Binary Classification Model

The confusion matrix summarizes the performance of a classification model on a set of test data for which the actual values of the *null hypothesis* are known.

	Null is FALSE	recast Null is TRUE
Actual Null is FALSE	True Positive (sensitivity)	False Negative (type II error)
Null is TRUE	False Positive (type I error)	True Negative (specificity)

- > # Calculate FALSE positive and FALSE negative rates
 > confmat <- confmat / rowSums(confmat)</pre>
- > c(typeI=confmat[2, 1], typeII=confmat[1, 2])
- typeI typeII
- 0.00186 0.81988
- > detach(Default)

Let the *null hypothesis* be that the subject will not default: default = FALSE.

The true positive rate (known as the sensitivity) is the fraction of FALSE null hypothesis cases that are correctly classified as FALSE.

The false negative rate is the fraction of FALSE null hypothesis cases that are incorrectly classified as TRUE (type II error).

The sum of the *true positive* plus the *false negative* rate is equal to 1.

The true negative rate (known as the specificity) is the fraction of TRUE null hypothesis cases that are correctly classified as TRUE.

The false positive rate is the fraction of TRUE null hypothesis cases that are incorrectly classified as FALSE (type I error).

The sum of the *true negative* plus the *false positive* rate is equal to 1.

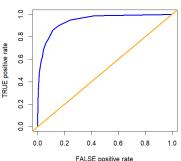
Receiver Operating Characteristic (ROC) Curve

The *ROC curve* is the plot of the *true positive* rate, as a function of the *false positive* rate, and illustrates the performance of a binary classifier.

The area under the *ROC curve* (AUC) is a measure of the performance of a binary classification model.

```
> # Confusion matrix as function of threshold
> confun <- function(actualy, fcast, threshold) {
      confmat <- table(actualy, (fcast < threshold))
      confmat <- confmat / rowSums(confmat)
      c(typeI=confmat[2, 1], typeII=confmat[1, 2])
    } # end confun
> confun(!testset$default, fcast, threshold=threshold)
> # Define vector of discrimination thresholds
> threshv <- seq(0.05, 0.95, by=0.05)^2
> # Calculate error rates
> error_rates <- sapply(threshy, confun,
   actualv=!testset$default, fcast=fcast) # end sapply
> error_rates <- t(error_rates)
> rownames(error_rates) <- threshv
> error_rates <- rbind(c(1, 0), error_rates)
> error_rates <- rbind(error_rates, c(0, 1))
> # Calculate area under ROC curve (AUC)
> truepos <- (1 - error_rates[, "typeII"])
> truepos <- (truepos + rutils::lagit(truepos))/2
> falsepos <- rutils::diffit(error_rates[, "typeI"])
```

ROC Curve for Defaults



```
> # Plot ROC Curve for Defaults
> x11(width=5, height=5)
> plot(x=error_rates[, "typeI"], y=1-error_rates[, "typeII"],
+ xlab="FALSE positive rate", ylab="TRUE positive rate",
+ main="ROC Curve for Defaults", type="1", lwd=3, col="blue")
> abline(a=0.0, b=1.0, lwd=3, col="oranse")
```

> abs(sum(truepos*falsepos))

> library(HighFreq)

Reading TAQ Data From .csv Files

Trade and Quote (TAQ) data stored in .csv files can be very large, so it's better to read it using the function data.table::fread() which is much faster than the function read.csv().

Each *trade* or *quote* contributes a *tick* (row) of data, and the number of ticks can be very large (hundred of thousands per day, or more).

The function strptime() coerces character strings representing the date and time into POSIX1t date-time objects.

The argument format="%H:%M:%OS" allows the parsing of fractional seconds, for example "15:59:59.989847074".

The function as.POSIXct() coerces objects into POSIXct date-time objects, with a numeric value representing the moment of time in seconds.

```
> # Read TAQ trade data from csv file
> tag <- data.table::fread(file="/Users/jerzy/Develop/data/xlk_tick
> # Inspect the TAQ data
> tag
> class(tag)
> colnames(tag)
> sapply(taq, class)
> symbol <- tag$SYM ROOT[1]
> # Create date-time index
> datev <- paste(tag$DATE, tag$TIME M)
> # Coerce date-time index to POSIX1t
> datev <- strptime(datev, "%Y%m%d %H:%M:%OS")
> class(datev)
> # Display more significant digits
> # options("digits")
> options(digits=20, digits.secs=10)
> last(datev)
> unclass(last(datev))
> as.numeric(last(datev))
> # Coerce date-time index to POSIXct
> datev <- as.POSIXct(datev)
> class(datev)
> last(datev)
> unclass(last(datev))
> as.numeric(last(datev))
> # Calculate the number of ticks per second
> nsecs <- as.numeric(last(datev)) - as.numeric(first(datev))
> NROW(taq)/(6.5*3600)
> # Select TAQ data columns
> tag <- tag[, .(price=PRICE, volume=SIZE)]
> # Add date-time index
> tag <- cbind(index=datev, tag)
```

Microstructure Noise in High Frequency Data

High frequency data contains microstructure noise in the form of price jumps and the bid-ask bounce.

Price jumps are single ticks with prices far away from the average.

Price iumps are often caused by data collection errors. but sometimes they represent actual very large lot trades

The bid-ask bounce is the bouncing of traded prices between the bid and ask prices.

The bid-ask bounce creates an illusion of rapidly changing pricey, while in fact the mid price is constant.



XLK Trade Ticks for 2020-03-16

- > # save(xtsv, file="/Users/jerzv/Develop/data/xlk tick trades2020
- > dygraphs::dygraph(xtsv\$XLK.Close,
- main="XLK Trade Ticks for 2020-03-16")
- > # Plot in v11 window
- > x11(width=6, height=5)
- > quantmod::chart Series(x=xtsv\$XLK.Close.
- name="XLK Trade Ticks for 2020-03-16")

XLK Trade Ticks for 2020-03-16 (after Hampel filter)

Removing Microstructure Noise From High Frequency Data

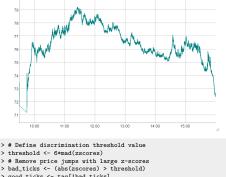
Microstructure noise can be removed from high frequency data by using a Hampel filter.

The z-scores are equal to the prices minus the median pricey, divided by the median absolute deviation (MAD) of prices:

$$z_i = \frac{p_i - \mathsf{median}(\mathbf{p})}{\mathsf{MAD}}$$

If the z-score exceeds the threshold value then it's classified as an outlier (jump in pricev).

```
> # Calculate centered Hampel filter to remove price jumps
> look_back <- 111
> half_back <- look_back %/% 2
> medianv <- roll::roll_median(tag$price, width=look_back)
> # medianv <- TTR::runMedian(tag$price, n=look_back)
> medianv <- rutils::lagit(medianv, lagg=(-half_back), pad_zeros=Fa
> madv <- HighFreq::roll_var(matrix(taq$price), look_back=look_back
> # madv <- TTR::runMAD(taq$price, n=look_back)
> madv <- rutils::lagit(madv, lagg=(-half_back), pad_zeros=FALSE)
> # Calculate Z-scores
> zscores <- (taq$price - medianv)/madv
> zscores[is.na(zscores)] <- 0
> zscores[!is.finite(zscores)] <- 0
> sum(is.na(zscores))
> sum(!is.finite(zscores))
> range(zscores); mad(zscores)
> hist(zscores, breaks=2000, xlim=c(-5*mad(zscores), 5*mad(zscores)
```



- > # Define discrimination threshold value

- > bad_ticks <- (abs(zscores) > threshold)
- > good_ticks <- taq[!bad_ticks]
- > # Calculate number of price jumps
- > sum(bad_ticks)/NROW(zscores)
- > # Coerce trade prices to xts
- > xtsv <- xts::xts(good_ticks[, .(price, volume)], good_ticks\$index
- > colnames(xtsv) <- c("XLK.Close", "XLK.Volume")
- > # Plot dygraph of the clean lots
- > dygraphs::dygraph(xtsv\$XLK.Close,
- main="XLK Trade Ticks for 2020-03-16 (Hampel filtered)") > # Plot the large lots
- > x11(width=6, height=5)
- > quantmod::chart_Series(x=xtsv\$XLK.Close,
- name="XLK Trade Ticks for 2020-03-16 (Hampel filtered)")

Classifying Data Outliers Using the Hampel Filter

The data points whose absolute z-scores exceed a threshold value are classified as outliers.

This procedure is a *classifier*, which classifies the prices as either good or bad data points.

If the bad data points are not labeled, then we can add jumps to the data to test the performance of the classifier

Let the null hypothesis be that the given price is a good data point.

A positive result corresponds to rejecting the null hypothesis, while a negative result corresponds to accepting the null hypothesis.

The classifications are subject to two different types of errors: type I and type II errors.

A type I error is the incorrect rejection of a TRUE null hypothesis (i.e. a "false positive"), when good data is classified as had

A type II error is the incorrect acceptance of a FALSE null hypothesis (i.e. a "false negative"), when bad data is classified as good.

- > # Define discrimination threshold value
- > threshold <- 6*mad(zscores)
- > # Calculate number of prices classified as bad data > isbad <- (abs(zscores) > threshold)
- > sum(isbad)
- > # Add 200 random price jumps into prices
- > set.seed(1121)
- > nbad <- 200 > isjump <- logical(NROW(closep))
- > isjump[sample(x=NROW(isjump), size=nbad)] <- TRUE
- > closep[isjump] <- closep[isjump]* sample(c(0.95, 1.05), size=nbad, replace=TRUE)
- > # Plot the prices and medians
- > dygraphs::dygraph(cbind(closep, mediany), main="VTI median") %>% dyOptions(colors=c("black", "red"))
- > # Calculate time series of z-scores
- > medianv <- roll::roll_median(closep, width=look_back)
- > # medianv <- TTR::runMedian(closep, n=look_back)
- > madv <- HighFreq::roll_var(closep, look_back=look_back, method="neth
- > # madv <- TTR::runMAD(closep, n=look_back) > zscores <- (closep - medianv)/madv
- > zscores[1:look back,] <- 0
- > # Calculate number of prices classified as bad data
- > isbad <- (abs(zscores) > threshold)
- > sum(isbad)

Eassass.

Confusion Matrix of a Binary Classification Model

A binary classification model categorizes cases based on its forecasts whether the null hypothesis is TRUE or FALSE.

The confusion matrix summarizes the performance of a classification model on a set of test data for which the actual values of the null hypothesis are known.

	Null is FALSE Null is TRUE		
Actual Null is FALSE	True Positive (sensitivity)	False Negative (type II error)	
Null is TRUE False Positive (type I error)		True Negative (specificity)	

Let the null hypothesis be that the given price is a good data point.

The true positive rate (known as the sensitivity) is the fraction of FALSE null hypothesis cases that are correctly classified as FALSE.

The false negative rate is the fraction of FALSE null hypothesis cases that are incorrectly classified as TRUE (type II error).

The sum of the true positive plus the false negative rate is equal to 1.

The true negative rate (known as the specificity) is the fraction of TRUE null hypothesis cases that are correctly classified as TRUE

The false positive rate is the fraction of TRUE null hypothesis cases that are incorrectly classified as FALSE (type I error).

The sum of the true negative plus the false positive rate is equal to 1.

- > # Calculate confusion matrix
- > table(actual=!isjump, forecast=!isbad) > sum(isbad)
- > # FALSE positive (type I error)
- > sum(!isjump & isbad)
- > # FALSE negative (type II error)
- > sum(isjump & !isbad)

Receiver Operating Characteristic (ROC) Curve

The *ROC curve* is the plot of the *true positive* rate, as a function of the *false positive* rate, and illustrates the performance of a binary classifier.

The area under the $ROC\ curve\ (AUC)$ measures the classification ability of a binary classifier.

```
> # Confusion matrix as function of threshold
> confun <- function(actualy, zscores, threshold) {
      confmat <- table(!actualv, !(abs(zscores) > threshold))
      confmat <- confmat / rowSums(confmat)
      c(typeI=confmat[2, 1], typeII=confmat[1, 2])
    } # end confun
> confun(isjump, zscores, threshold=threshold)
> # Define vector of discrimination thresholds
> threshv <- seq(from=0.2, to=5.0, by=0.2)
> # Calculate error rates
> error_rates <- sapply(threshy, confun,
   actualv=isjump, zscores=zscores) # end sapply
> error_rates <- t(error_rates)
> rownames(error_rates) <- threshv
> error_rates <- rbind(c(1, 0), error_rates)
> error_rates <- rbind(error_rates, c(0, 1))
> # Calculate area under ROC curve (AUC)
> truepos <- (1 - error_rates[, "typeII"])
> truepos <- (truepos + rutils::lagit(truepos))/2
> falsepos <- rutils::diffit(error_rates[, "typeI"])
> abs(sum(truepos*falsepos))
```

ROC Curve for Hampel Classifier

```
> # Plot ROC curve for Hampel classifier
> x11(width=6, height=5)
> plot(x=error_rates[, "typeII"], y=1-error_rates[, "typeII"],
+ xlab="FALSE positive rate", ylab="RUE positive rate",
+ xlim=c(0, 1), ylim=c(0, 1), ylim=c(0, 1),
+ main="ROC Curve for Hampel Classifier",
+ type="1", lwd=3, col="blue")
> abline(a=0.0, b=1.0, lwd=3, col="orange")
```

draft: Receiver Operating Characteristic (ROC) Curve

The performance of the Hampel noise classification model depends on the length of the look-back time interval. a binary. area under the ROC curve (AUC) is a measure of a binary. The optimal threshold value can be determined using cross-validation.

The ROC curve is the plot of the true positive rate, as a function of the false positive rate, and illustrates the performance of a binary classifier.

The area under the *ROC curve* (AUC) measures the classification ability of a binary classifier.

> # Confusion matrix as function of threshold

```
> confun <- function(actualy, zscores, threshold) {
      confmat <- table(!actualv, !(abs(zscores) > threshold))
      confmat <- confmat / rowSums(confmat)
      c(typeI=confmat[2, 1], typeII=confmat[1, 2])
   } # end confun
> confun(isjump, zscores, threshold=threshold)
   Define vector of discrimination thresholds
> threshv <- seq(from=0.2, to=5.0, by=0.2)
> # Calculate error rates
> error_rates <- sapply(threshv, confun,
    actualv=isjump, zscores=zscores) # end sapply
> error rates <- t(error rates)
> rownames(error rates) <- threshy
> error rates <- rbind(c(1, 0), error rates)
> error rates <- rbind(error rates, c(0, 1))
> # Calculate area under ROC curve (AUC)
> truepos <- (1 - error rates[, "typeII"])
> truepos <- (truepos + rutils::lagit(truepos))/2
> falsepos <- rutils::diffit(error rates[, "typeI"])
```

ROC Curve for Hampel Classifier

```
> # Plot ROC curve for Hampel classifier
> x11(vidth=6, height=5)
    plot(xe=ror_rates[, "typeI"], y=1-error_rates[, "typeII"],
+ xlab="FALSE positive rate", ylab="TRUE positive rate",
+ xlim=(0, 1), ylim=(0, 1),
+ main="ROC Curve for Hampel Classifier",
+ type="1", lwd=3, col="blue")
> ablin(a=0, 0, b=1.0, lwd=3, col="orange")
```

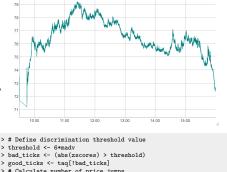
> abs(sum(truepos*falsepos))

Improved Microstructure Noise Filtering

The filtering of microstructure noise can be improved by calculating the median prices over an interval of only 3 data points.

If the *z-score* exceeds the *threshold value* then it's classified as an *outlier* (jump in pricev).

```
> # Calculate centered Hampel filter over 3 data points
> medianv <- roll::roll_median(taq$price, width=3)
> medianv (!:2] <- taq$price[1:2]
> medianv <- rutils::lagit(medianv, lagg=-1, pad_zeros=FALSE)
> madv <- HighFreq::roll_var(matrix(taq$price), look_back=3, method:
> madv <- rutils::lagit(madv, lagg=-1, pad_zeros=FALSE)
> # Calculate Z-scores
> zscores <- ifelse(madv > 0, (taq$price - medianv)/madv, 0)
> range(zscores); mad(zscores)
> madv <- mad(zscores[abs(zscores)>0])
> hist(zscores, breaks=2000, xlimsc(-5*madv, 5*madv))
```



XLK Trade Ticks for 2020-03-16 (after Hampel filter)

```
> # Define discrimination threshold value
> threshold <- 6*madov
> bad_ticks <- (abs(zscores) > threshold)
> good_ticks <- taq[!bad_ticks]
> # Calculate number of price jumps
> sum(bad_ticks)/NROW(zscores)
> # Coerce trade prices to xts
> xtov <- xts::xts(good_ticks[, (price, volume)], good_ticks$index
> colnames(xtsv) <- ("XLK.Close", "XLK.Volume")
> # Plot dygraph of the clean lots
> dygraphs::dygraph(xtsv$XLK.Close,
+ main="XLK Trade Ticks for 2020-03-16 (Hampel filtered)")
> # Plot the large lots
> x11(vidth=6, height=5)
> quantmod::chart_Series(x=xtsv$XLK.Close,
+ name="XLK Trade Ticks for 2020-03-16 (Hampel filtered)")
```

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draft: Classification Using K-Nearest Neighbor (KNN) Algorithm

The K-nearest neighbor (KNN) algorithm is a supervised learning classification technique.

Normalizing numeric data

function predict() is a *generic function* for forecasting based on a given model.

The method predict.glm() produces forecasts for a generalized linear model, in the form of probabilities for the Boolean response variable.

The Boolean forecasts are obtained by comparing the forecast probabilities with a discrimination threshold.

The null hypothesis is that default = FALSE.

A positive result corresponds to rejecting the null hypothesis, while a negative result corresponds to accepting the null hypothesis.

The forecasts are subject to two different types of errors: *type I* and *type II* errors.

A type I error is the incorrect rejection of a TRUE null hypothesis (i.e. a "false positive"), when good data is classified as bad.

A type II error is the incorrect acceptance of a FALSE null hypothesis (i.e. a "false negative"), when bad data is classified as good.

draft: Data Science

Data Science is very important to quantitative finance.

Below is an example of a simulation of the path of *Brownian Motion* crossing a barrier level.

- Data is never clean.
- 2. You will spend most of your time cleaning and preparing data.
- 3. 95% of tasks do not require deep learning.
- 4. In 90% of cases generalized linear regression will do the trick.
- 5. Big Data is just a tool.
- You should embrace the Bayesian approach.
 No one cares how you did it.
- 7. No one cares now you did it
- 8. Academia and business are two different worlds.
- 9. Presentation is key be a master of Power Point.
- 10. All models are false, but some are useful.
- 11. There is no fully automated Data Science. You need to get your hands dirty.

draft: Machine Learning

What is Machine Learning? What is Machine Learning? Machine Learning (ML) studies statistical models which can identify patterns in the data and make predictions. ML is closely related to statistics, but with an emphasis on prediction. ML models are divided into supervised learning or unsupervised learning. Supervised learning models require a training set to calibrate the model parameters. Examples of supervised learning models are linear regression, decision trees, support vector machines (SVM), and neural networks. Unsupervised learning models don't require a training set. Examples of unsupervised learning models are clustering models, like principal component analysis (PCA) and k-nearest neighbors (KNN). ML models are also divided into classification and regression models. An example of a regression model is linear regression. An example of a classification model is logistic regression. ML uses several techniques to calibrate models and improve prediction. First, ML uses cross-validation (backtesting) to determine the optimal model meta-parameters. Second, ML uses estimator shrinkage to achieve a better tradeoff between their bias and variance

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