

Sample and Data Relationship Format for Proteomics (SDRF- Proteomics)

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Table of Contents

1. Status of this document	1
2. Abstract	2
3. Motivation	3
4. Quick Start	5
4.1. Minimal Example	5
4.2. Key Concepts	6
4.3. Format Requirements	6
4.4. Scope	6
4.5. Getting Started Steps	6
5. Validating SDRF Files	8
6. Specification structure	9
6.1. Versioning	9
6.2. Notational Conventions	10
6.3. Relationship to other specifications	10
7. SDRF-Proteomics specification	11
7.1. Format rules	11
7.2. Reserved words	12
7.3. SDRF file-level metadata	12
7.3.1. Template Inheritance and Versioning Rules	13
7.4. Table Column headers	14
7.5. Table Cell values	14
8. SDRF-Proteomics: Samples metadata	16
8.1. BioSamples database integration	17
8.2. Encoding sample technical and biological replicates	17
8.3. Pooled samples	19
8.4. Sample Metadata Guidelines	19
9. SDRF-Proteomics: data files metadata	20
9.1. CV Term Format for Data File Metadata	20
9.2. Sample Preparation and Fragmentation (MS-based only)	21
9.3. Proteomics data acquisition method	21
9.4. MS-Proteomics Template	22
10. Additional SDRF Rules	23
10.1. Column Cardinality	23
10.2. Row Uniqueness Requirements	23
11. Templates	25
11.1. What is a Template?	25
11.2. Template Layered Architecture	25
11.3. Template Inheritance	26
11.4. Specifying Templates in File Headers	27
11.5. Choosing Templates	28
11.6. Core Templates	28
11.7. Experiment-Type Templates	29
11.8. Extending a Template	30

11.8.1. When to Add Custom Columns	30
11.8.2. Rules for Custom Columns	30
11.8.3. Common Additional Columns	30
11.8.4. Example: Adding Study-Specific Columns.....	31
12. Factor Values (Study Variables)	32
12.1. Column Format	32
12.2. When to Use Factor Values	32
12.3. Rules	32
12.4. Example	32
13. Ontologies and Controlled Vocabularies	33
14. Examples of Annotated Datasets	35
15. Templates Under Discussion	36
15.1. MS-Imaging Datasets	36
15.2. Derived Samples (Patient-Derived Xenografts)	36
15.3. Turnover and TMT Datasets.....	36
15.4. Contributing New Templates	37
16. Intellectual Property Statement	38
17. Copyright Notice	39
18. How to cite	40
References	41

Chapter 1. Status of this document

This document provides information to the proteomics community about a proposed standard for sample metadata annotations in public repositories called Sample and Data Relationship Format (SDRF)-Proteomics. Distribution is unlimited.

Version v1.1.0 - 2025-01

Chapter 2. Abstract

The Human Proteome Organisation (HUPO) Proteomics Standards Initiative (PSI) defines community standards for data representation in proteomics to facilitate data comparison, exchange, and verification. This document presents a specification for the Sample and Data Relationship Format (SDRF-Proteomics).

Further detailed information, including any updates to this document, implementations, and examples is available at [SDRF GitHub Repository](#). The official PSI web page for the document is: [HUPO-PSI SDRF](#).

Chapter 3. Motivation

Many resources have emerged that provide raw or integrated proteomics data in the public domain. If these are valuable individually, their integration through re-analysis represents a huge asset for the community [1].

Unfortunately, proteomics experimental design and sample related information are often missing in public repositories or stored in very diverse ways and formats. For example:

- The [CPTAC Consortium](#) provides for every dataset a set of Excel files with the information on [each sample](#) including tumor size, origin, but also how every sample is related to a specific raw file (e.g. instrument configuration parameters).
- As a resource routinely re-analysing public datasets, [ProteomicsDB](#) captures for each sample in the database a minimum number of properties to describe the sample and the related experimental protocol such as tissue, digestion method and instrument.

Such heterogeneity often prevents data interpretation, reproducibility, and integration of data from different resources. For every proteomics dataset we propose to capture at least three levels of metadata:

- (i) dataset description
- (ii) the sample metadata and data files acquisition metadata.
- (iii) The relation between the sample and the data files. The experimental design.

The general description includes minimum information to describe the study overall: [title](#), [description](#), [date of publication](#), [type of experiment](#). In ProteomeXchange partners this metadata is captured at the dataset level, in other omics resources this is captured as IDF file format (e.g. MAGE-TAB). Currently, all ProteomeXchange partners mandate this information for each dataset. However, the information regarding the sample and its relation to the data files (**Figure 1**) is mostly missing [1].



Figure 1: SDRF-Proteomics file format stores the information of the sample and its relation to the data files in the dataset. The file format includes not only information about the sample but also about how the data was acquired and processed.

Here, we introduced the Sample and Data Relationship Format (SDRF-Proteomics) to capture the sample metadata and its relation to the data files for proteomics experiments. The SDRF-Proteomics format is a tab-delimited file format that describes the sample characteristics and the relationships between samples and data files included in a dataset.

This specification, which is a community effort, aims to provide a standard for the proteomics community to annotate the sample metadata and its relation to the data files.

Chapter 4. Quick Start

If you're new to SDRF-Proteomics, here's a minimal example to get you started. An SDRF file is a tab-separated file where each row represents a sample-to-data-file relationship.

4.1. Minimal Example

source	characteristic	characteristic	characteristic	characteristic	assay	technology	commodity	commodity	commodity	commodity	commodity	commodity	commodity	commodity	commodity	factor
name	actuator	actuator	actuator	actuator	name	type	men t[proto	men t[lab el]	men t[ins trument]	men t[cle ava	men t[fra ctio	men t[tec hnic al	men t[dat a file]	men t[replic ate]	value[di seas e]	
sample_1	hom o sapiens	liver	nor mal	1	run_1	protoeomic profiling by mass spectrometry	Data -dependent acquisition	label free sample	Q Exactive HF	NT= Trypsin;A C=M S:10 0125 1	1	1	sample_1.ra w	nor mal		
sample_2	hom o sapiens	liver	hepatocellular carcinoma	1	run_2	protoeomic profiling by mass spectrometry	Data -dependent acquisition	label free sample	Q Exactive HF	NT= Trypsin;A C=M S:10 0125 1	1	1	sample_2.ra w	hepatocellular carcinoma		

Blue: Sample metadata | Green: Data file metadata | Orange: Factor values

NOTE

This minimal example shows a **mass spectrometry-based** proteomics experiment. For detailed requirements see the [MS-Proteomics template](#). For **affinity-based proteomics** (Olink, SomaScan), different columns are required - see the [Affinity-Proteomics template](#).

4.2. Key Concepts

1. **Sample metadata** uses `characteristics[...]` columns (e.g., organism, disease)
2. **Data file metadata** uses `comment[...]` columns (e.g., instrument, label)
3. **Factor values** use `factor value[...]` columns to indicate variables under study
4. Each row links one sample to one data file

4.3. Format Requirements

The SDRF-Proteomics format has the following core requirements:

- The SDRF file is a **tab-delimited format** where each row corresponds to a relationship between a Sample and a Data file.
- Each column MUST correspond to an attribute/property of the Sample or the Data file.
- Each cell value MUST be the property value for the corresponding Sample or Data file.
- The file MUST start with columns describing sample properties (e.g., organism, disease), followed by data file properties (e.g., label, fraction identifier, data file).
- Unknown values MUST be handled using **reserved words**: `not available` (value is unknown), `not applicable` (property doesn't apply), or `pooled` (value is a mixture from multiple samples).

4.4. Scope

The SDRF-Proteomics format aims to capture the **sample metadata** and its **relationship with data files** (e.g., raw files from mass spectrometers).

IMPORTANT

SDRF-Proteomics does **not** aim to capture downstream analysis details, including: which samples were compared to which other samples, how samples are combined into study variables, or analysis parameters such as FDR thresholds or p-value cutoffs.

4.5. Getting Started Steps

1. Choose a **technology template** based on your **technology type**: `ms-proteomics` for *proteomic profiling by mass spectrometry*, `affinity-proteomics` for *protein expression profiling by antibody/aptamer array* (Olink, SomaScan)
2. Add the appropriate **sample template** for your organism (Human, Vertebrates, Invertebrates, Plants, or Cell-lines)
3. Fill in sample metadata (characteristics columns)
4. Fill in data file metadata (comment columns)
5. Add factor values for your experimental variables
6. Validate your file using `sdrf-pipelines`

For detailed guidance, continue reading the full specification below.

Chapter 5. Validating SDRF Files

The official validator for SDRF-Proteomics files is **sdrf-pipelines**, a Python tool that checks your SDRF file for errors and compliance with the specification.

Installation:

```
pip install sdrf-pipelines
```

Basic Validation:

```
# Validate an SDRF file
parse_sdrf validate-sdrf --sdrf_file your_file.sdrf.tsv

# Validate with a specific template
parse_sdrf validate-sdrf --sdrf_file your_file.sdrf.tsv --template human
```

For more information, visit: [sdrf-pipelines on GitHub](#)

Chapter 6. Specification structure

This document describes the main specification of SDRF-Proteomics, the structure of the specification (**Figure 2**), how to contribute, and extend the specification. SDRF-Proteomics uses a three-tier system for organizing metadata requirements:

- **The SDRF-Proteomics core specification:** This document contains the main specification, requirements and rules for the SDRF-Proteomics format. It also includes the notational conventions and the relationship to other specifications.
- **Templates:** All templates are organized in the `templates/` directory. This includes:
 - **Core templates** (organism-based): `human`, `vertebrates`, `invertebrates`, `plants`
 - **Specialized templates** (experiment-type): `cell-lines` (sample), `DDA acquisition` (data), `DIA acquisition` (data), `single-cell` (mixed), `affinity-proteomics` (data), `crosslinking` (data), `immunopeptidomics` (mixed), `metaproteomics` (mixed)

Each template has its own directory containing a detailed README.adoc with checklists and examples, plus a template file (`{name}-template.sdrf.tsv`) with column headers. See the [Templates](#) section.

[Logo] | *images/sdrf-guidelines-structure.png*

Figure 2: SDRF-Proteomics specification structure. The main specification defines the core rules and is extended by technology templates (ms-proteomics, affinity-proteomics), sample templates (human, vertebrates, etc.), and specialized experiment-type templates.

NOTE

The main specification is in the `sdrf-proteomics` directory. All templates are organized in `sdrf-proteomics/templates/`. Templates are extensions of the core specification and should follow all the rules and requirements in the main specification. Each template includes its own detailed documentation with checklists, examples, and annotation guidelines specific to that template type.

The official website for SDRF-Proteomics project is <https://github.com/bigbio/proteomics-metadata-standard>. New use cases, changes to the specification and examples can be added by using Pull requests or issues in GitHub to reach the bigbio team.

A set of examples and annotated projects from ProteomeXchange can be found here: <https://github.com/bigbio/proteomics-metadata-standard/tree/master/annotated-projects>

Multiple tools have been implemented to validate, annotate and convert SDRF-Proteomics files. The official validator of SDRF-Proteomics is sdrf-pipelines (Python - <https://github.com/bigbio/sdrf-pipelines>). This tool allows to validate an SDRF-Proteomics file. In addition, it allows converting SDRF to other popular pipelines and software configure files such as MaxQuant or OpenMS.

6.1. Versioning

The SDRF-Proteomics specification is versioned using the Semantic Versioning 2.0.0 (<https://semver.org/>) scheme. The version number is in the format MAJOR.MINOR.PATCH, where:

- MAJOR version is incremented for incompatible changes to the specification, when major changes are done to the specification.
- MINOR version is incremented for new features that are backward compatible with the previous version. Guidelines and templates are added or modified.
- PATCH version is incremented for bug fixes and minor changes that do not affect the specification or the templates. This includes typos, formatting changes, and other minor updates.

Every change in the specification should be done in GitHub using pull requests into the dev branch. The pull request should include a description of the changes and the reason for the changes. The pull request will be reviewed by the community and merged into the main branch when approved. After the merge, the version number will be updated according to the changes made, the release will be performed, and the Zenodo record will be updated.

NOTE

We added the prefix v to the version number to indicate that it is the version of the specification that was used to create the file. Examples: v1.1.0, v2.0.0, v3.0.0.

6.2. Notational Conventions

The key words “MUST”, “MUST NOT”, “REQUIRED”, “SHALL”, “SHALL NOT”, “SHOULD”, “SHOULD NOT”, “RECOMMEND/RECOMMENDED”, “MAY”, “COULD BE”, and “OPTIONAL” are to be interpreted as described in RFC 2119 (<https://www.rfc-editor.org/rfc/rfc2119>).

6.3. Relationship to other specifications

SDRF-Proteomics is fully compatible with the SDRF file format part of [MAGE-TAB](#). MAGE-TAB is the file format used to store metadata and sample information for transcriptomics experiments. When the ProteomeXchange project file is converted to idf file (project description in MAGE-TAB) and is combined with the SDRF-Proteomics a valid MAGE-TAB is obtained.

SDRF-Proteomics sample information can be embedded into mzTab metadata files. The sample metadata in mzTab contains properties as the columns in the SDRF-Proteomics and values as Sample cell values.

The SDRF-Proteomics aims to capture the sample metadata and its relationship with the data files (e.g. raw files from mass spectrometers). The SDRF-Proteomics do not aim to capture the downstream analysis part of the experimental design such as what samples should be compared, how they can be combined or parameters for the downstream analysis (FDR or p-values thresholds). The HUPO-PSI community will work in the future to include this information in other file formats such as mzTab or a new type of file format.

Chapter 7. SDRF-Proteomics specification

The SDRF-Proteomics file format describes the sample characteristics and the relationships between samples and data files. The file format is a tab-delimited one where each **ROW** corresponds to a relationship between a Sample and a Data file (in an ms proteomics experiment the data file containing the mass spectra), each **COLUMN** corresponds to an attribute/property of the Sample, the Data file, or the Factor values; and the value in each **CELL** is the specific value of the property for a given Sample/Data file/Factor value (**Figure 3**).

source name	character istics[organism]	character istics[...]	character istics[bio logical replicate]	assay name	technolo gy type	comment [...]	comment [data file]	factor value[...]
sample_1	homo sapiens	...	1	run_1	proteomic profiling by mass spectrometry	...	sample_1.raw	...
sample_2	homo sapiens	...	2	run_2	proteomic profiling by mass spectrometry	...	sample_2.raw	...

Blue: Sample metadata (characteristics) | Green: Data file metadata (comments) | Orange: Factor values

Figure 3: SDRF-Proteomics in a nutshell. Each **row** links a sample to a data file. **Columns** represent sample properties (characteristics), data file properties (comments), or experimental variables (factor values).

The SDRF-Proteomics format contains three main sections:

- The first section contains the [sample metadata](#).
- The second section contains the [data file metadata](#).
- The third section contains the [factor values](#) properties.

7.1. Format rules

- **Case sensitivity:** Text values are case-insensitive, but **column names are case-sensitive**. Use lowercase for all column names (e.g., `source_name`, `characteristics[organism]`, `comment[label]`). Incorrect casing like `Source Name` or `Characteristics[organism]` will cause validation failures.
- **Space sensitivity:** The SDRF is sensitive to spaces in column names (`sourcename` ↘ `source name`). Column names must include appropriate spaces (e.g., `source name`, not `sourcename`) but must NOT have a space before the bracket (e.g., `characteristics[organism]`, not `characteristics [organism]`).

- **Column order:** The SDRF columns follows some structure; first the sample metadata columns in [Chapter 8](#); then the data file metadata columns in [Chapter 9](#); followed by the factor values columns in [Chapter 12](#).
- **Extension:** The extension of the SDRF file SHOULD be **sdrf.tsv (preferred)** or .txt.

7.2. Reserved words

There are general scenarios where cell values cannot be provided with actual data. The following reserved words MUST be used in these cases:

- **not available:** In some cases, the column is mandatory in the format, but for some samples the corresponding value is unknown or could not be determined. In those cases, users SHOULD use **not available**.
- **not applicable:** In some cases, the column is mandatory, but for some samples the corresponding value or concept does not apply. In those cases, users SHOULD use **not applicable**.
- **anonymized:** In some cases, the value exists but has been intentionally redacted for privacy protection (e.g., in clinical studies with de-identified patient data). In those cases, users SHOULD use **anonymized**.
- **pooled:** In some cases, the sample is a pool of multiple samples (e.g., TMT reference channels), and the value cannot be represented as a single value. In those cases, users SHOULD use **pooled**.

Table 1. Reserved words for SDRF cell values

Term	Meaning	Example	Use Case
not available	Value exists but is unknown or could not be determined	characteristics[age] = not available	Patient age was not recorded in the study
not applicable	Value or concept does not apply to this sample	characteristics[age] = not applicable	Synthetic peptide library has no age
anonymized	Value exists but is redacted for privacy protection	characteristics[age] = anonymized	Clinical study with de-identified patient data
pooled	Value represents a mixture of multiple samples	characteristics[biological replicate] = pooled	TMT reference channel pooled from multiple replicates

7.3. SDRF file-level metadata

Since version 1.1.0, SDRF-Proteomics supports optional file-level metadata using header comments at the beginning of the file. These header comments provide information about the SDRF file itself, such as the format version, template used, and validation status. This approach is inspired by other omics formats such as VCF (Variant Call Format) file headers and is fully compatible with pandas and other tabular data processing tools.

Header comments MUST:

- Start with # (single hash) followed by a key-value pair
- Appear at the very beginning of the file, before the column header row
- Use the format `#key=value`

The following header fields are supported:

Key	Description	Example	Requirement	Ontology Term
<code>file_format</code>	Identifier for the file format	SDRF	RECO MMEN DED	PRIDE:0000831
<code>version</code>	SDRF-Proteomics specification version used	v1.1.0	RECO MMEN DED	NCIT:C25714
<code>template</code>	Template(s) used, comma-separated. Only list leaf templates; parent templates are implied by inheritance	human,crosslinking	OPTIONAL	PRIDE:0000832
<code>template_version</code>	Version(s) of the template(s). Use single value if all same; comma-separated only when versions differ	v1.1.0	OPTIONAL	PRIDE:0000833
<code>source</code>	Tool or origin that generated the file. Replaces per-row <code>comment[tool metadata]</code> column	lesSDRF v0.1.0	OPTIONAL	NCIT:C25683
<code>validation_hash</code>	Hash from validator certification	sha256:abc123...	OPTIONAL	PRIDE:0000834

Example of an SDRF file with header comments (simplified example showing only select columns; see [Chapter 11](#) for complete required columns):

```
#file_format=SDRF
#version=v1.1.0
#template=human
#template_version=v1.1.0
#source=lesSDRF v0.1.0
source name characteristics[organism]    characteristics[organism part]
characteristics[disease]    assay name comment[data file]
sample_1      homo sapiens    liver    normal    run_1    sample_1.raw
```

7.3.1. Template Inheritance and Versioning Rules

Template inheritance: Only list leaf templates in the `#template` field. Parent templates are implied:

- `crosslinking` inherits from `ms-proteomics` → use `#template=human,crosslinking` (not `ms-`)

- ```
proteomics,human,crosslinking)
• olink inherits from affinity-proteomics → use #template=human,olink (not affinity-
proteomics,olink,human)
```

### Version simplification:

- If all templates share the same version, use a single value: #template\_version=v1.1.0
- Only use comma-separated versions when templates have different versions

```
Same version for all templates - use single value
#template=human,crosslinking
#template_version=v1.1.0

Different versions - list each (matching template order)
#template=human,crosslinking
#template_version=v1.1.0,v1.0.0
```

**Source field:** The #source header replaces the per-row comment[tool metadata] column. Use it to record the tool or origin that generated the SDRF file (e.g., [lesSDRF v0.1.0](#), [PRIDE](#), [user-generated](#)).

**NOTE**

Header comments are OPTIONAL. SDRF files without header comments are still valid. When present, header comments provide valuable provenance information and enable tools to handle version-specific features appropriately. Header property names use underscores (e.g., `file_format`, `template_version`) rather than spaces to maintain consistency with the tab-delimited nature of SDRF files and avoid ambiguity when parsing.

## 7.4. Table Column headers

Depending on each section the column headers (property names) will be prefixed with the following prefixes:

- `characteristics`: Sample metadata (e.g. `characteristics[organism]`)
- `comment`: Data file metadata (e.g. `comment[data file]`)
- `factor value`: Factor values properties (e.g. `factor value[disease]`)

Each property name MUST be a valid ontology term or a valid controlled vocabulary term. Each section will have some specific order for column headers.

**NOTE**

A list of all controlled vocabularies and ontologies supported are in the [Chapter 13](#) section. On each section we also provide a list of properties that are supported.

## 7.5. Table Cell values

The value for each property, (e.g. characteristics, comment, factor value) corresponding to each sample or data file can be represented in multiple ways.

- Free Text (Human readable):** In the free text representation, the value is provided as text without Ontology support (e.g. colon or providing accession numbers). This is only RECOMMENDED when the text inserted in the table is the exact name of an ontology/CV term in EFO. If the term is not in EFO, other ontologies can be used.

| source name | characteristics[organism] |
|-------------|---------------------------|
| sample 1    | homo sapiens              |
| sample 2    | homo sapiens              |

- Ontology url (Computer readable):** Users can provide the corresponding URI (Uniform Resource Identifier) of the ontology/CV term as a value. This is recommended for enriched files where the user does not want to use intermediate tools to map from free text to ontology/CV terms.

| source name | characteristics[organism]                                                                                 |
|-------------|-----------------------------------------------------------------------------------------------------------|
| Sample 1    | <a href="http://purl.obolibrary.org/obo/NCBITaxon_9606">http://purl.obolibrary.org/obo/NCBITaxon_9606</a> |
| Sample 2    | <a href="http://purl.obolibrary.org/obo/NCBITaxon_9606">http://purl.obolibrary.org/obo/NCBITaxon_9606</a> |

- Key=value representation (Human and Computer readable):** The current representation aims to provide a mechanism to represent the complete information of the ontology/CV term including Accession, Name and other additional properties. In the key=value pair representation, the Value of the property is represented as an Object with multiple properties, where the key is one of the properties of the object and the value is the corresponding value for the particular key. An example of key value pairs is post-translational modification (see [Protein Modifications](#)):

```
NT=Glu->pyro-Glu;MT=fixed;PP=Anywhere;AC=Unimod:27;TA=E
```

## Chapter 8. SDRF-Proteomics: Samples metadata

The Sample metadata section provides information about the samples of origin and their characteristics. Each sample contains a *source name* (unique identifier) and a set of *characteristics* columns. The first column of the file should be the *source name* and the following columns should be the characteristics of the sample. For example, for any proteomics experiment (human, vertebrate, cell line), the following characteristics should be provided:

- **source name:** Unique sample name (it can be present multiple times if the same sample is used several times in the same dataset)
- **characteristics[organism]:** The organism of the Sample of origin. Values MUST come from [NCBI Taxonomy](#).
- **characteristics[organism part]:** The part of organism's anatomy or substance arising from an organism from which the biomaterial was derived (e.g., liver). Values SHOULD come from [UBERON](#) or [BTO](#).
- **characteristics[disease]:** The disease under study in the Sample. Values SHOULD come from [MONDO](#), [EFO](#), or [DOID](#). For healthy/control samples, use [normal](#) ([PATO:0000461](#)) - see [Disease Annotation Guidelines](#).
- **characteristics[cell type]:** A cell type is a distinct morphological or functional form of cell (e.g., epithelial, glial). Values SHOULD come from [Cell Ontology \(CL\)](#) or [BTO](#).

Example:

| source name    | characteristics[organism] | characteristics[organism part] | characteristics[disease] | characteristics[cell type] |
|----------------|---------------------------|--------------------------------|--------------------------|----------------------------|
| sample_treat   | homo sapiens              | liver                          | liver cancer             | not available              |
| sample_control | homo sapiens              | liver                          | liver cancer             | not available              |

### NOTE

Additional characteristics can be added depending on the type of the experiment and sample. The [SDRF-Proteomics templates](#) defines a set of templates and checklists of properties that should be provided depending on the proteomics experiment. In the core guidelines and templates, main document of SDRF-Proteomics, we explain the major sample properties for different experiments. However, SDRF-Proteomics can be extended using guidelines for specific experiments.

### IMPORTANT

Each characteristic name in the column header SHOULD be a CV term from the EFO ontology. For example, the header `characteristics[organism]` corresponds to the ontology term Organism. However the values could be from EFO or other ontologies. For example, we RECOMMEND to use MONDO for diseases because it has better coverage than EFO. For healthy samples, use [normal](#) ([PATO:0000461](#)) - see [Disease Annotation Guidelines](#).

### IMPORTANT

Multiple values (columns) for the same `characteristics` term are allowed in SDRF-Proteomics (see [Section 10.1](#)). However, it is RECOMMENDED not to use the same column in the same file. If you have multiple phenotypes, you can

specify what it refers to or use another more specific term, e.g., "immunophenotype".

## 8.1. BioSamples database integration

[BioSamples](#) provides persistent identifiers for biological samples that enable cross-database linking [5]. Use the optional *characteristics[biosample accession number]* column to link samples to BioSamples entries (EBI format: SAMEA\*, NCBI format: SAMN\*).

**Column:** `characteristics[biosample accession number]`

**Examples:** `SAMN12345678` (NCBI), `SAMEA12345678` (EBI)

Example usage:

| source name | characteristics[bio sample accession number] | characteristics[organism] | characteristics[organism part] | characteristics[disease] |
|-------------|----------------------------------------------|---------------------------|--------------------------------|--------------------------|
| sample_001  | SAMN12345678                                 | homo sapiens              | liver                          | liver cancer             |
| sample_002  | SAMN12345679                                 | homo sapiens              | liver                          | normal                   |
| sample_003  | SAMEA12345680                                | mus musculus              | brain                          | normal                   |

The BioSamples accession number enables connection to sample resources across databases, linking proteomics data with genomics, transcriptomics, and other omics datasets.

**NOTE**

BioSample accession numbers from NCBI follow the format `SAMNxxxxxxxxxx` and from EBI follow the format `SAMEAxxxxxxxxx`, where `x` represents digits. Either NCBI or EBI BioSample accession numbers can be used depending on where the sample is registered. The *characteristics[biosample accession number]* column is OPTIONAL, but when available, providing BioSample accession numbers is RECOMMENDED to enhance data integration and reusability. Users must first request BioSample accession numbers from the appropriate service (NCBI or EBI) before including them in their SDRF files.

## 8.2. Encoding sample technical and biological replicates

SDRF-Proteomics uses two columns to track replicates [4]:

- **characteristics[biological replicate]:** Identifies independent biological samples within each experimental condition (factor value group). Replicate numbers are assigned per condition, so if you have 2 cancer samples and 2 healthy samples, both groups would have biological replicates numbered 1 and 2.
- **comment[technical replicate]:** Identifies repeated measurements of the same sample (e.g., multiple LC-MS/MS injections)

**IMPORTANT**

Biological replicate numbering restarts for each experimental condition (factor value). For example, in a disease study with factor value[disease], the cancer samples would be numbered 1, 2, 3... and the normal samples would independently be numbered 1, 2, 3... This establishes the relationship between each sample and its experimental condition.

The following example shows 2 biological replicates, each with 2 fractions and 2 technical replicates:

| source name        | characteristics[biological replicate] | assay name | comment[label]    | comment[fraction identifier] | comment[technical replicate] | comment[data file] |
|--------------------|---------------------------------------|------------|-------------------|------------------------------|------------------------------|--------------------|
| patient_001_sample | 1                                     | run_01     | label free sample | 1                            | 1                            | P001_F1_TR 1.raw   |
| patient_001_sample | 1                                     | run_02     | label free sample | 2                            | 1                            | P001_F2_TR 1.raw   |
| patient_001_sample | 1                                     | run_03     | label free sample | 1                            | 2                            | P001_F1_TR 2.raw   |
| patient_001_sample | 1                                     | run_04     | label free sample | 2                            | 2                            | P001_F2_TR 2.raw   |
| patient_002_sample | 2                                     | run_05     | label free sample | 1                            | 1                            | P002_F1_TR 1.raw   |
| patient_002_sample | 2                                     | run_06     | label free sample | 2                            | 1                            | P002_F2_TR 1.raw   |
| patient_002_sample | 2                                     | run_07     | label free sample | 1                            | 2                            | P002_F1_TR 2.raw   |
| patient_002_sample | 2                                     | run_08     | label free sample | 2                            | 2                            | P002_F2_TR 2.raw   |

In this example:

- **Biological replicates:** `patient_001_sample` and `patient_002_sample` are different biological samples (different source names), annotated with `characteristics[biological replicate]` values 1 and 2
- **Technical replicates:** Each biological sample is measured twice (`comment[technical replicate] = 1 and 2`)
- **Fractions:** Each technical replicate has 2 fractions (`comment[fraction identifier] = 1 and 2`)

**IMPORTANT**

Both `characteristics[biological replicate]` and `comment[technical replicate]` columns are REQUIRED. When no replicates are performed in a study, set both columns to 1 (i.e., each sample is biological replicate 1 and technical replicate 1). For **pooled samples** (e.g., TMT reference channels), use `pooled` for biological replicate since these samples are mixtures of multiple replicates and assigning a specific replicate number would be misleading.

Some examples with explicit annotation of the biological replicates can be found here:

- <https://github.com/bigbio/proteomics-metadata-standard/blob/c3a56b076ef381280dfcb0140d2520126ace53ff/annotated-projects/PXD006401/PXD006401.sdrf.tsv>

### 8.3. Pooled samples

When multiple samples are pooled into one (e.g., TMT/iTRAQ reference channels for normalization), use the *characteristics[pooled sample]* column to indicate pooling status. Allowed values:

- **not pooled**: Regular individual samples
- **pooled**: Sample is pooled but individual sources are unknown
- **SN=sample1;SN=sample2;...:** Lists source names of pooled samples when known

Example:

| source name | characteristics[pooled sample] | characteristics[organism] | characteristics[age] | comment[label] | comment[data file] |
|-------------|--------------------------------|---------------------------|----------------------|----------------|--------------------|
| sample_1    | not pooled                     | homo sapiens              | 45Y                  | TMT126         | file01.raw         |
| sample_2    | not pooled                     | homo sapiens              | 52Y                  | TMT127N        | file01.raw         |
| pooled_ref  | SN=sample_1;SN=sample_2        | homo sapiens              | pooled               | TMT131C        | file01.raw         |

**TIP**

For pooled samples, use **pooled** for individual-specific fields (biological replicate, age, sex) to indicate a mixture rather than a single sample.

### 8.4. Sample Metadata Guidelines

For detailed guidance on annotating sample metadata, refer to the following conventions documents:

- [Sample Metadata Guidelines](#) - Detailed guidelines for age, sex, disease, organism part, cell type, developmental stage, spiked-in samples, and other sample characteristics
- [Human Sample Metadata Guidelines](#) - Human-specific metadata including disease staging, treatment history, demographics, and lifestyle factors

# Chapter 9. SDRF-Proteomics: data files metadata

The connection between samples and data files is done using properties annotated with the `comment` prefix. All properties referring to a data file (e.g., MS run file) are annotated with the category `comment`. This differentiates data file properties from sample properties (characteristics).

## 9.1. CV Term Format for Data File Metadata

For data file metadata (`comment` columns) that reference ontology terms, use the structured format:  
`NT={term name};AC={accession}`

Examples: `NT=HCD;AC=PRIDE:0000590`, `NT=Orbitrap;AC=MS:1000484`

This format enables automated validation and software extraction from raw files. Sample metadata (characteristics) can use simple term names since they are typically human-annotated.

The following properties MUST be provided for each data file in **mass spectrometry-based proteomics** experiments. For **affinity-based proteomics** (Olink, SomaScan), see the [Affinity-Proteomics template](#) for different required columns.

| Column                                                   | Requirement | Description                                                               | Ontology                 |
|----------------------------------------------------------|-------------|---------------------------------------------------------------------------|--------------------------|
| <code>assay_name</code>                                  | REQUIRED    | Unique identifier for an MS run/data file                                 | Free text                |
| <code>technology_type</code>                             | REQUIRED    | Technology used to capture the data                                       | Fixed values             |
| <code>comment[proteomics data acquisition method]</code> | REQUIRED    | DDA, DIA, PRM, SRM                                                        | PRIDE:0000659            |
| <code>comment[label]</code>                              | REQUIRED    | Label applied to sample (or "label free sample")                          | PRIDE - Labels           |
| <code>comment[instrument]</code>                         | REQUIRED    | Mass spectrometer model                                                   | PSI-MS - Instruments     |
| <code>comment[cleavage agent details]</code>             | REQUIRED    | Enzyme information (use "not applicable" for top-down/undigested samples) | PSI-MS - Cleavage agents |
| <code>comment[fraction identifier]</code>                | REQUIRED    | Fraction number (1 if not fractionated)                                   | Integer                  |
| <code>comment[technical replicate]</code>                | REQUIRED    | Technical replicate number (1 if none)                                    | Integer                  |
| <code>comment[data file]</code>                          | REQUIRED    | Name of the raw file                                                      | Free text                |

Example:

| source name | assay name    | technology type                          | comment[proteomics data acquisition method] | comment[label]    | comment[instrument] | comment[data file] |
|-------------|---------------|------------------------------------------|---------------------------------------------|-------------------|---------------------|--------------------|
| sample_1    | sample1_ru_n1 | proteomic profiling by mass spectrometry | data-dependent acquisition                  | label free sample | Q Exactive HF       | sample1.raw        |

## 9.2. Sample Preparation and Fragmentation (MS-based only)

**NOTE**

This section applies to **mass spectrometry-based proteomics** experiments only. For affinity-based proteomics, these properties do not apply.

For detailed documentation of sample preparation and MS/MS fragmentation properties, see the [MS-Proteomics Template](#):

- **Sample preparation:** depletion, reduction reagent, alkylation reagent
- **Fractionation:** fractionation method (used with `comment[fraction identifier]`)
- **Fragmentation:** collision energy, dissociation method

## 9.3. Proteomics data acquisition method

Proteomics data acquisition method can happen in multiple ways: Data Dependent Acquisition (DDA), Data Independent Acquisition (DIA), and targeted approaches. The SDRF-Proteomics file format REQUIRES capturing the method used for the data acquisition in the `comment[proteomics data acquisition method]` column. The values MUST be children of the PRIDE ontology term [proteomics data acquisition method \(PRIDE:0000659\)](#). The following values are commonly used:

- data-dependent acquisition
- data-independent acquisition
  - diaPASEF
  - SWATH MS
- parallel reaction monitoring
- selected reaction monitoring

**IMPORTANT**

The `comment[proteomics data acquisition method]` column is REQUIRED for all mass spectrometry-based SDRF files. This field must be explicitly specified and cannot be omitted or assumed.

You can find an example of a DIA experiment in the following link: [DIA example](#)

**TIP**

For DIA experiments, additional properties like MS1 scan range can be captured. See [DIA Scan Window Limits](#) in the MS-Proteomics Template.

## 9.4. MS-Proteomics Template

For detailed guidance on data file metadata, refer to the conventions document:

- [MS-Proteomics Template](#) - Detailed guidelines for labels, instruments, modifications, cleavage agents, mass tolerances, RAW file URLs, and other data file properties

# Chapter 10. Additional SDRF Rules

## 10.1. Column Cardinality

Some columns can appear multiple times for the same sample. The cardinality rules are:

- **Single (1):** Column appears exactly once per sample (e.g., `characteristics[biological replicate]`)
- **Multiple (\*):** Column can appear multiple times (e.g., `comment[modification parameters]` can specify multiple post-translational modifications)

Example of multiple `comment[modification parameters]` columns:

| source name | characteristics[...] | comment[modification parameters]                         | comment[modification parameters]                      | ... |
|-------------|----------------------|----------------------------------------------------------|-------------------------------------------------------|-----|
| sample-1    | ...                  | NT=Carbamidomethyl;AC=UNIMOD:4;TA=C;MT=fixed;PP=Anywhere | NT=Oxidation;AC=UNIMOD:5;TA=M;MT=variable;PP=Anywhere | ... |

## 10.2. Row Uniqueness Requirements

SDRF files must satisfy specific uniqueness constraints to ensure data integrity and enable proper indexing by analysis tools.

**Error-level constraint (validation fails):** The combination of `source name + assay name + comment[label]` MUST be unique across all rows in the SDRF file. If two rows have identical values for all three columns, validation will fail with an error. This constraint ensures that each sample-run-label combination can be uniquely identified.

**Warning-level constraint (validation warns):** The combination of `source name + assay name` SHOULD be unique across all rows. Non-unique combinations will generate a warning during validation. This constraint helps identify potential issues where the same sample appears to have multiple entries for the same MS run without distinguishing labels.

**Assay name uniqueness:** Each distinct MS run/data file MUST have exactly one globally unique `assay name`, and no two different data files may share an assay name. To ensure uniqueness, it is RECOMMENDED to incorporate sample-specific information in assay names, such as sample IDs or replicate numbers (e.g., "sample1\_run1", "sample1\_run2", "patient001\_fraction01").

**NOTE** For multiplexed experiments (e.g., TMT, iTRAQ), multiple SDRF rows will share the same `assay name` because multiple samples are analyzed in a single MS run. In these cases, the `comment[label]` column distinguishes between different samples within the same run, and the combination of `source name + assay name + comment[label]` remains unique.

Example of valid multiplexed experiment:

| <b>source name</b> | ... | <b>assay name</b> | <b>comment[label]</b> | ... | <b>comment[data file]</b> |
|--------------------|-----|-------------------|-----------------------|-----|---------------------------|
| sample_A           | ... | TMT_batch1_ru n1  | TMT126                | ... | batch1_run1.ra w          |
| sample_B           | ... | TMT_batch1_ru n1  | TMT127N               | ... | batch1_run1.ra w          |
| sample_C           | ... | TMT_batch1_ru n1  | TMT127C               | ... | batch1_run1.ra w          |
| sample_D           | ... | TMT_batch1_ru n1  | TMT128N               | ... | batch1_run1.ra w          |

In this example, all four rows share the same `assay name` and `comment[data file]` because they represent different samples multiplexed in a single MS run. The combination of `source name + assay name + comment[label]` is unique for each row.

# Chapter 11. Templates

## 11.1. What is a Template?

A **template** in SDRF-Proteomics is a predefined set of metadata columns (both required and recommended) that ensures consistent and complete annotation for a specific type of experiment or sample. Templates serve the same purpose as **metadata checklists**, **minimum information standards** (like MIAPE), or **validation schemas** in other data standards—they define what information must be captured to make a dataset FAIR (Findable, Accessible, Interoperable, Reusable).

Each template includes both **sample metadata** (characteristics columns describing the biological sample) and **data file metadata** (comment columns describing the MS data files)—everything needed to create a complete, validated SDRF file.

## 11.2. Template Layered Architecture

Templates in SDRF-Proteomics follow a **layered architecture** that separates concerns:

| Layer             | Purpose                                      | Templates                                                                                                                                                   | Valid Alone?                                   |
|-------------------|----------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------|------------------------------------------------|
| <b>BASE</b>       | Construction artifact with shared columns    | <a href="#">base</a>                                                                                                                                        | No - never use directly                        |
| <b>TECHNOLOGY</b> | Minimum valid templates for data acquisition | <a href="#">ms-proteomics</a> , <a href="#">affinity-proteomics</a>                                                                                         | <b>Yes</b> - these are the minimum valid SDRFs |
| <b>SAMPLE</b>     | Organism-specific sample metadata            | <a href="#">human</a> , <a href="#">vertebrates</a> , <a href="#">invertebrates</a> , <a href="#">plants</a>                                                | No - must combine with TECHNOLOGY              |
| <b>EXPERIMENT</b> | Experiment-specific columns                  | <a href="#">cell-lines</a> , <a href="#">single-cell</a> , <a href="#">crosslinking</a> , <a href="#">dda-acquisition</a> , <a href="#">dia-acquisition</a> | No - must combine with TECHNOLOGY              |

**TIP**

The **base** template is an internal construction artifact that defines shared columns inherited by all other templates. It should never be used directly by users. This specification document itself serves as the documentation for base template columns - all columns defined in the **sample metadata** and **data file metadata** sections are inherited by technology and sample templates.

### Key concepts:

- **Technology templates** ([ms-proteomics](#) or [affinity-proteomics](#)) are **required** - you must use exactly one
- **Sample templates** ([human](#), [vertebrates](#), etc.) are **recommended** but optional
- **Experiment templates** provide additional methodology-specific columns
- Templates can be **extended**: add custom columns beyond the template requirements for study-specific metadata

When combining templates, include all required columns from each applicable template. The validator will check compliance with all specified templates.

### 11.3. Template Inheritance

Templates form a layered inheritance hierarchy. Child templates inherit all columns and validators from parent templates and can add new columns or strengthen requirements.

```

base (CONSTRUCTION ARTIFACT - not valid alone)
 - Shared columns: source name, organism, organism part,
 - biological replicate, assay name, technology type,
 - instrument, technical replicate, data file
 -
 -> ms-proteomics (MINIMUM VALID for MS proteomics)
 - Adds: acquisition method, cleavage agent, label, fraction identifier
 -
 -> dda-acquisition (extends ms-proteomics with DDA-specific columns)
 -> dia-acquisition (extends ms-proteomics with DIA-specific columns)
 -> crosslinking (extends ms-proteomics with XL-MS columns)
 -> single-cell (extends ms-proteomics with SCP columns)
 -> immunopeptidomics (extends ms-proteomics with MHC-MS columns)
 -
 -> affinity-proteomics (MINIMUM VALID for affinity proteomics)
 - Adds: instrument, panel name, quantification unit, sample type
 -
 -> olink (extends affinity-proteomics with Olink-specific columns)
 -> somascan (extends affinity-proteomics with SomaScan-specific columns)

Sample templates (combine with TECHNOLOGY templates):
 -> human (adds: disease, age, sex, ancestry)
 -> vertebrates (adds: disease, developmental stage, strain)
 -> invertebrates (adds: disease, developmental stage, genotype)
 -> plants (adds: disease, developmental stage, growth conditions)

Experiment templates (organism-agnostic):
 -> cell-lines (adds: cell line name, Cellosaurus ID, passage number)

```

#### Inheritance rules:

1. Child templates inherit all columns from parent templates
2. Child templates may add new columns
3. Child templates may promote `optional` → `recommended` → `required`
4. Child templates **must not** weaken requirements (e.g., `required` → `optional`)
5. Multiple templates can be combined (e.g., `ms-proteomics + human + crosslinking`)

#### Column requirement levels in templates:

Each column in a template has one of three requirement levels:

- **REQUIRED:** The column **MUST** appear in the SDRF file. Validation will fail if this column is missing. Template header files (.sdrf.tsv) always include these columns.

- **RECOMMENDED:** The column **SHOULD** appear in the SDRF file when applicable to the experiment. These columns are included in template header files and validation will generate warnings if missing.
- **OPTIONAL:** The column **MAY** be included if relevant to the study. These columns are NOT included in template header files but are documented in the template checklist. Users can add them as needed.

**NOTE**

When a column is marked as RECOMMENDED or OPTIONAL, it means users should evaluate whether the column applies to their specific experiment. For example, `characteristics[cell line]` is RECOMMENDED in the immunopeptidomics template because cell lines are commonly used, but tissue samples may use `not applicable`.

**TIP**

**For developers and maintainers:** For detailed information on YAML template structure, validators, and LinkML schema, see the [Template Definitions Guidelines](#).

## 11.4. Specifying Templates in File Headers

When declaring templates in the `#template` header field, follow these rules:

### Rule 1: Only list leaf templates - parent templates are implied

Since child templates inherit from parents, you only need to list the most specific (leaf) template. The parent is automatically implied:

```
CORRECT: crosslinking inherits from ms-proteomics
#template=crosslinking

INCORRECT: don't list parent when child is present
#template=ms-proteomics,crosslinking
```

### Rule 2: List templates from different branches

When combining templates from different branches of the hierarchy (e.g., organism + experiment), list both:

```
Human sample with crosslinking experiment
- human is from the SAMPLE branch
- crosslinking is from the EXPERIMENT branch (inherits ms-proteomics)
#template=human,crosslinking
```

### Rule 3: Version simplification

Use a single version if all templates share the same version. Only use comma-separated versions when templates have different versions:

```
All templates at v1.1.0 - use single value
#template=human,crosslinking
#template_version=v1.1.0
```

```
Different versions - list each (matching template order)
#template=human,crosslinking
#template_version=v1.1.0,v1.0.0
```

### Common examples:

| Experiment Type                  | Templates                                | Header                                     |
|----------------------------------|------------------------------------------|--------------------------------------------|
| Human MS proteomics              | human (inherits ms-proteomics)           | #template=human                            |
| Mouse MS proteomics              | vertebrates (inherits ms-proteomics)     | #template=vertebrates                      |
| Non-organism-specific MS         | ms-proteomics only                       | #template=ms-proteomics                    |
| Human crosslinking               | human + crosslinking                     | #template=human,crosslinking               |
| Recombinant protein crosslinking | crosslinking only (no organism template) | #template=crosslinking                     |
| Human Olink                      | human + olink                            | #template=human,olink                      |
| Human DIA with cell lines        | human + dia-acquisition + cell-lines     | #template=human,dia-acquisition,cell-lines |

## 11.5. Choosing Templates

First, determine your proteomics technology and select the appropriate technology template:

| Template                   | Use For                                                        | Documentation      | Valid Alone?               |
|----------------------------|----------------------------------------------------------------|--------------------|----------------------------|
| <b>MS-Proteomics</b>       | All mass spectrometry-based proteomics (DDA, DIA, SRM/MRM/PRM) | Full documentation | Yes - minimum for MS       |
| <b>Affinity-Proteomics</b> | Affinity-based proteomics (Olink, SomaScan)                    | Full documentation | Yes - minimum for affinity |

## 11.6. Core Templates

Add the appropriate sample template based on your sample organism:

| Template           | Use For                                  | Key Additional Columns                           | Documentation      |
|--------------------|------------------------------------------|--------------------------------------------------|--------------------|
| <b>Human</b>       | Human clinical samples                   | disease, age, sex, ancestry category, individual | Full documentation |
| <b>Vertebrates</b> | Mouse, rat, zebrafish, other vertebrates | disease, developmental stage, strain/breed, sex  | Full documentation |

| Template      | Use For                          | Key Additional Columns                                                   | Documentation                      |
|---------------|----------------------------------|--------------------------------------------------------------------------|------------------------------------|
| Invertebrates | Drosophila, C. elegans, insects  | disease, developmental stage, strain/breed, genotype                     | <a href="#">Full documentation</a> |
| Plants        | Arabidopsis, crops, other plants | disease, developmental stage, strain/breed, growth conditions, treatment | <a href="#">Full documentation</a> |

For detailed explanations of each column, see [sample metadata](#) for sample properties and [data file metadata](#) for data file properties.

**NOTE**

The [characteristics\[cell type\]](#) column is RECOMMENDED across all templates when the cell type is known or can be determined. Use [not available](#) if the cell type cannot be determined (e.g., whole tissue samples, mixed cell populations). For cell line experiments, use the cell-lines template which provides more specific guidance.

## 11.7. Experiment-Type Templates

In addition to sample templates, SDRF-Proteomics provides specialized experiment-type templates. These templates extend the core templates with methodology-specific columns.

- **DDA Acquisition:** Data-dependent acquisition experiments. Includes columns for dissociation method, collision energy, fractionation method, modification parameters, and mass tolerances.
- **DIA Acquisition:** Data-independent acquisition experiments. Includes columns for scan window limits, isolation window width, DIA method, and spectral library information.
- **Cell Lines:** Experiments using cell line samples. Includes Cellosaurus integration for cell line identification.
- **Single-Cell Proteomics:** Single-cell proteomics experiments. Includes columns for cell isolation method, carrier proteome, and single-cell identifiers.
- **Immunopeptidomics:** MHC peptide immunopeptidomics. Includes columns for MHC class, HLA typing, and enrichment methods.
- **Crosslinking MS:** Cross-linking mass spectrometry experiments. Includes columns for crosslinking reagents and enrichment methods.
- **Metaproteomics:** Environmental and microbiome proteomics. Includes columns for environmental sample type and geographic location.
- **Affinity Proteomics:** Affinity-based proteomics (Olink, SomaScan). Includes columns specific to these platforms.

The template files can be downloaded from the [templates](#) folder.

## 11.8. Extending a Template

Templates define the minimum required and recommended columns for a given experiment type. However, SDRF-Proteomics is designed to be **extensible**:

1. **Add experiment-type templates:** If your experiment type has a specialized template (DDA, DIA, cell-lines, etc.), add it to your base configuration. See [Experiment-Type Templates](#) above.
2. **Add custom columns:** You can add any additional columns to capture study-specific metadata beyond what templates define. This section provides guidance on adding custom columns.

### 11.8.1. When to Add Custom Columns

Add custom columns when your experiment requires metadata that is:

- Not covered by existing templates but important for data interpretation
- Specific to your experimental design or technology
- Required for cross-study integration or data reuse
- Needed to comply with domain-specific standards (e.g., clinical trials, environmental studies)

### 11.8.2. Rules for Custom Columns

#### 1. Use appropriate column prefixes:

- `characteristics[...]` - For sample-related metadata (properties of the biological material)
- `comment[...]` - For technical or protocol-related metadata (data acquisition, processing)
- `factor value[...]` - For experimental variables (see [Chapter 12](#))

#### 2. Follow naming conventions:

- Use lowercase for column names inside brackets
- Use descriptive, specific names (e.g., `characteristics[tumor grade]` not `characteristics[grade]`)
- Use ontology terms when they exist (e.g., `characteristics[patient bmi]` maps to EFO term)

#### 3. Use controlled vocabularies when available:

- Reference existing ontologies (EFO, MONDO, UBERON, etc.) for values
- Prefer standardized terms over free text when possible
- For new terms, follow the key=value format if ontology reference is needed

### 11.8.3. Common Additional Columns

Beyond template requirements, many commonly used columns can enhance your SDRF file. For a comprehensive list of additional columns for sample metadata (treatment, time point, dose, BMI, smoking status, etc.) and data metadata (sample preparation, enrichment method, etc.), see:

- [Common Additional Columns in the Sample Metadata Guidelines](#)

- [SDRF Terms Reference](#) for a comprehensive list of commonly used terms

#### 11.8.4. Example: Adding Study-Specific Columns

For a drug treatment time-course study, you might add columns beyond the template requirements:

| source name      | characteristics[organism] | characteristics[disease] | characteristics[treatment] | characteristics[time point] | characteristics[doze] | ... | factor value[treatment] | factor value[time point] |
|------------------|---------------------------|--------------------------|----------------------------|-----------------------------|-----------------------|-----|-------------------------|--------------------------|
| sample_ctrl_0h   | homo sapiens              | normal                   | vehicle control            | 0h                          | not applicable        | ... | vehicle control         | 0h                       |
| sample_d rug_24h | homo sapiens              | normal                   | dexamethasone              | 24h                         | 100 nM                | ... | dexamethasone           | 24h                      |

**TIP** When adding custom columns, check the [SDRF Terms Reference](#) and [EBI Ontology Lookup Service \(OLS\)](#) to find existing terms that match your metadata needs. Using standardized terms improves data interoperability.

# Chapter 12. Factor Values (Study Variables)

Factor values identify the experimental variables being studied - the conditions you want to compare in your analysis. They highlight which sample characteristics are the focus of your experiment.

## 12.1. Column Format

```
factor value[{variable name}]
```

## 12.2. When to Use Factor Values

Use factor values to indicate:

- The primary variable(s) under investigation
- Conditions being compared (e.g., disease vs. normal, treated vs. untreated)
- Variables that define experimental groups

**NOTE**

Use `normal` (not "control") in the disease field for healthy samples. "Control" is an experimental design concept, not a disease state. See [Disease Annotation Guidelines](#) for details.

## 12.3. Rules

- Factor value columns SHOULD appear after all characteristics and comment columns
- Multiple factor values can be used when studying multiple variables
- The value in a factor value column typically mirrors a characteristics column value

## 12.4. Example

In an experiment comparing tumor vs. normal tissue across different cancer stages:

| source name     | ... | characteristi cs[disease] | characteristi cs[disease staging] | ... | factor value[diseas e] | factor value[diseas e staging] |
|-----------------|-----|---------------------------|-----------------------------------|-----|------------------------|--------------------------------|
| tumor_sample_1  | ... | breast carcinoma          | stage II                          | ... | breast carcinoma       | stage II                       |
| normal_sample_1 | ... | normal                    | not applicable                    | ... | normal                 | not applicable                 |
| tumor_sample_2  | ... | breast carcinoma          | stage III                         | ... | breast carcinoma       | stage III                      |

In this example, both `disease` and `disease staging` are factor values because the experiment aims to compare expression differences between disease states and across cancer stages.

# Chapter 13. Ontologies and Controlled Vocabularies

SDRF-Proteomics uses ontologies and controlled vocabularies (CVs) to standardize metadata values. The following ontologies are supported:

| Category                                                           | Ontology/CV                                         | Description                                                   | Notes                  |
|--------------------------------------------------------------------|-----------------------------------------------------|---------------------------------------------------------------|------------------------|
| <b>General Purpose</b>                                             |                                                     |                                                               |                        |
| General                                                            | <a href="#">Experimental Factor Ontology (EFO)</a>  | General experimental metadata                                 |                        |
| General                                                            | <a href="#">PATO</a>                                | Phenotype and Trait Ontology                                  |                        |
| General                                                            | <a href="#">NCI Thesaurus (NCIT)</a>                | Biomedical terminology                                        |                        |
| General                                                            | <a href="#">PRIDE Controlled Vocabulary</a>         | Proteomics-specific terms                                     |                        |
| <b>Organism and Taxonomy</b>                                       |                                                     |                                                               |                        |
| Taxonomy                                                           | <a href="#">NCBI Taxonomy (NCBITaxon)</a>           | Organism classification                                       |                        |
| <b>Anatomy and Cell Types</b>                                      |                                                     |                                                               |                        |
| Anatomy                                                            | <a href="#">UBERON</a>                              | Cross-species anatomy ontology                                |                        |
| Cell Type                                                          | <a href="#">Cell Ontology (CL)</a>                  | Cell type classification                                      |                        |
| Anatomy                                                            | <a href="#">BRENDA Tissue Ontology (BTO)</a>        | Tissues and cell lines                                        |                        |
| Anatomy                                                            | <a href="#">Plant Ontology (PO)</a>                 | Plant anatomy and development                                 | For plant samples      |
| Anatomy                                                            | <a href="#">FlyBase Anatomy (FBbt)</a>              | Drosophila anatomy                                            | For Drosophila samples |
| Anatomy                                                            | <a href="#">WormBase Anatomy (WBbt)</a>             | C. elegans anatomy                                            | For C. elegans samples |
| Anatomy                                                            | <a href="#">Zebrafish Anatomy (ZFA)</a>             | Zebrafish anatomy and development                             | For zebrafish samples  |
| <b>Disease (see <a href="#">Disease Annotation Guidelines</a>)</b> |                                                     |                                                               |                        |
| Disease                                                            | <a href="#">Mondo Disease Ontology (MONDO)</a>      | Unified disease ontology                                      | RECOMMENDED            |
| Disease                                                            | <a href="#">Experimental Factor Ontology (EFO)</a>  | Disease terms from EFO                                        |                        |
| Healthy samples                                                    | <a href="#">Phenotype And Trait Ontology (PATO)</a> | Use <a href="#">normal</a> (PATO:0000461) for healthy samples |                        |
| <b>Cell Lines</b>                                                  |                                                     |                                                               |                        |
| Cell Lines                                                         | <a href="#">Cellosaurus</a>                         | Cell line knowledge resource                                  | RECOMMENDED            |

| Category                                | Ontology/CV                                         | Description                              | Notes              |
|-----------------------------------------|-----------------------------------------------------|------------------------------------------|--------------------|
| Cell Lines                              | <a href="#">Cell Line Ontology (CLO)</a>            | Cell line ontology                       |                    |
| <b>Mass Spectrometry and Proteomics</b> |                                                     |                                          |                    |
| MS/Proteomics                           | <a href="#">PSI Mass Spectrometry CV (PSI-MS)</a>   | Instruments, methods, parameters         |                    |
| Modifications                           | <a href="#">Unimod</a>                              | Protein modifications database           |                    |
| Modifications                           | <a href="#">PSI-MOD CV</a>                          | Protein modifications ontology           |                    |
| <b>Other</b>                            |                                                     |                                          |                    |
| Chemistry                               | <a href="#">ChEBI</a>                               | Chemical Entities of Biological Interest |                    |
| Environment                             | <a href="#">Environment Ontology (ENVO)</a>         | Environmental sample classification      | For metaproteomics |
| Ancestry                                | <a href="#">Human Ancestry Ontology (HANCESTRO)</a> | Human ancestry categories                | For human samples  |

## Chapter 14. Examples of Annotated Datasets

The following table provides links to example SDRF files for different experiment types. Click "View in Explorer" to open the SDRF file in the interactive viewer.

| Experiment Type   | Dataset   | Description                              | View                             | Source                 |
|-------------------|-----------|------------------------------------------|----------------------------------|------------------------|
| Label-free        | PXD008934 | Human proteome label-free quantification | <a href="#">View in Explorer</a> | <a href="#">GitHub</a> |
| TMT               | PXD017710 | TMT-labeled quantitative proteomics      | <a href="#">View in Explorer</a> | <a href="#">GitHub</a> |
| SILAC             | PXD000612 | SILAC-based quantification               | <a href="#">View in Explorer</a> | <a href="#">GitHub</a> |
| DIA               | PXD018830 | Data-independent acquisition             | <a href="#">View in Explorer</a> | <a href="#">GitHub</a> |
| Phosphoproteomics | PXD000759 | PTM enrichment study                     | <a href="#">View in Explorer</a> | <a href="#">GitHub</a> |
| Cell lines        | PXD001819 | Cell line proteomics                     | <a href="#">View in Explorer</a> | <a href="#">GitHub</a> |

**TIP**

Use the [SDRF Explorer](#) to browse all {total\_datasets}+ annotated datasets with filtering, statistics, and interactive viewing.

A comprehensive collection of annotated projects is available at: [Annotated Projects Repository](#)

# Chapter 15. Templates Under Discussion

This section describes templates that are actively under discussion. If you want to discuss a particular template, please create an [issue on GitHub](#).

## 15.1. MS-Imaging Datasets

Discussions about MS-Imaging representation can be found in [issue #451](#). Key considerations for MSI datasets:

### Metadata that may not apply to MSI:

- **Reduction and Alkylation:** For most MSI studies this would be "not applicable"
- **Fraction identifier:** Never used in MSI - use "not applicable"
- **MS2/Database search terms:** Not mandatory, can be ignored for MSI data

### MSI-specific considerations:

- **File type:** imzML is a composite format (imzML + ibd files). Current practice references only the imzML file.
- **Multiple samples per run:** Multiple tissues can be placed on one slide. Samples are distinguished by position, requiring additional position identifiers or coordinates.

### Potential additional MSI metadata columns:

- **Origin of cleavage agent:** Important for calibration/QC (bovine vs. porcine trypsin)
- **Matrix:** For MALDI experiments, matrix composition helps with peak removal
- **Ionization source:** Customized sources may not match standard instrument terms
- **Analyte:** MSI can measure phospholipids, N-glycans, and other analytes beyond peptides

## 15.2. Derived Samples (Patient-Derived Xenografts)

A prototype for derived samples is documented in the [\[derived-samples\]](#) section. Outstanding issues include:

- Full examples with multiple species and mixture types are needed
- Capturing relationships between samples (e.g., in xenografts, how to link the human tumor sample to the mouse host)

## 15.3. Turnover and TMT Datasets

Turnover experiments may contain multiple labels - for example, TMT for quantification combined with SILAC for turnover/differential expression. See [PXD017710](#) for an example that was originally annotated as two independent datasets.

## 15.4. Contributing New Templates

To contribute to template development or propose a new template:

1. Open an [issue on GitHub](#) describing the experiment type
2. Discuss requirements with the community
3. Submit a pull request following the established template structure

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## Chapter 18. How to cite

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