

# TODO title

## TODO

### **Abstract**

Through their interactions, their activities, and even their mere presence, organisms change the environment for themselves and others. This “niche construction” process becomes particularly interesting when it creates evolutionary feedback, whereby selective pressures are altered in response to environmental change. Here we consider how niche construction influences the evolution of cooperation, which has been a long-standing challenge to evolutionary theory. We simulate populations of individuals that cooperatively produce a public good that permits increased growth in a stressful environment and investigate how local- and global-scale niche construction affects the ability of these populations to resist invasion by non-producing cheats. We find that niche construction profoundly impacts the evolution of cooperation by creating new opportunities for adaptation. Cooperators are able to escape subversion by cheats as long as niche construction clears these paths of adaptation. This work provides a crucial step towards understanding how evolution occurs in complex environments like those found in nature.

## 20 Introduction

21 Cooperative behaviors are common across all branches of the tree of life. In-  
22 sects divide labor within their colonies, plants and soil bacteria exchange es-  
23 sential nutrients, birds care for others' young, and the trillions of cells in the  
24 human body restrain their growth and coordinate to provide vital functions.  
25 Each instance of cooperation presents an evolutionary challenge: How can in-  
26 dividuals that sacrifice their own well-being to help others avoid subversion by  
27 those that do not? Over time, we would expect these *defectors* to rise in abun-  
28 dance at the expense of others, eventually driving cooperators—and perhaps  
29 the entire population—to extinction.

30 Several factors can defer this potential *tragedy of the commons* (Hamilton,  
31 1964; Hardin, 1968; Nowak, 2006; West *et al.*, 2007b). For example, coopera-  
32 tors must benefit more from the cooperative act than others. This can occur  
33 when cooperators are clustered together in spatially-structured populations  
34 (Fletcher and Doebeli, 2009; Nadell *et al.*, 2010; Kuzdzal-Fick *et al.*, 2011) or  
35 when cooperators use communication (Brown and Johnstone, 2001; Darch *et*  
36 *al.*, 2012) or other cues (Sinervo *et al.*, 2006; Gardner and West, 2010; Veelders  
37 *et al.*, 2010) to cooperate conditionally with kin. Interestingly, cooperation can  
38 also be bolstered by genetic linkage with self-benefitting traits (Foster *et al.*,  
39 2004; Dandekar *et al.*, 2012; Asfahl *et al.*, 2015), setting the stage for an “adap-  
40 tive race” in which cooperators and defectors vie for the first highly-beneficial  
41 non-social adaptation (Waite and Shou, 2012; Morgan *et al.*, 2012).

42 Hankshaw and Kerr (2015) recently showed that in spatially-structured popu-

lations, cooperators can gain a substantial leg up on defectors in an adaptive race when the cooperative behavior increases local population density, thus increasing the likelihood of acquiring beneficial non-social mutations. By hitchhiking along with these adaptations, the cooperative trait can rapidly rise in abundance. Nevertheless, this advantage is fleeting. As soon as the opportunities for adaptation are exhausted, cooperators are once again at a disadvantage against defectors, particularly those that arise from within via mutations that disable cooperation. However, Hankshaw and Kerr (2015) demonstrated that cooperation can be maintained indefinitely when frequent environmental changes produce a steady stream of non-social adaptive opportunities. Although organisms typically find themselves in dynamic environments, change might not occur at a rate that provides sufficient adaptive opportunities to ensure long-term cooperator survival.

Here, we build upon the model presented in Hankshaw and Kerr (2015) to explore whether the selective feedbacks produced as populations modify their environment can act as an additional source of adaptive opportunities. As previously described, this model follows the evolution of cooperation in a metapopulation of populations connected by spatially-limited migration. Cooperators produce a public good that increases the local carrying capacity. Through mutation, individuals gain non-social adaptations that confer fitness benefits. These benefits are large enough so that an adapted cooperator is more fit than a less-adapted defector (note that for simplicity, we refer to all non-cooperators as “defectors”, regardless of their origin). Because of their larger sizes, cooperator populations are more likely to acquire these adaptations. However, coop-

erator populations remain susceptible to invasion by adapted defectors either immigrate from a nearby populaion or arise via social mutation. In our expanded version of this model, populations alter their local environment based on the the presence of different non-social adaptations. Frequency-dependent selection on these adaptations creates an eco-evolutionary feedback that increasingly favors the adaptations present in each population. We focus on two aspects in which the production of public goods is affected by these selective feedbacks. First, the creation of unique niches may diminish the ability of both cooperators and defectors to invade neighboring patches due to maladaptation. However, because cooperator populations are larger, the greater number of emigrants that they produce will allow them to “export” their niche at a higher rate than defectors. As a result, cooperators may be able to expand more quickly. Second, larger cooperator populations will experience more mutations, which better enables these populations to adapt to changing environments. If niche construction produces continual change, can the resulting adaptive opportunities maintain cooperation?

By their very nature, public goods benefit populations by making their environment more hospitable. For example, bacteria produce extracellular products that find soluble iron (Griffin *et al.*, 2004), digest large proteins (Diggle *et al.*, 2007; Darch *et al.*, 2012), and reduce the risk of predation (Cosson *et al.*, 2002), among many others (West *et al.*, 2007a). While many studies have explored how the environment affects the evolution of cooperative bahviors, relatively few have examined how those behaviors affect the environment and the resulting change in evolutionary trajectories. Lehmann (2007) demonstrated analyti-

91 cally that when niche construction act benefits future generations, cooperation  
92 is favored due to reduced competition among kin. When rate-benefitting and  
93 yield-benefitting altruistic acts co-evolve, Van Dyken and Wade (2012) showed  
94 that “reciprocal niche construction”, where the selective feedbacks produced  
95 by one act benefitted the other, can lead to increased selection for both traits.  
96 While these studies have focused on the niche constructing effects of cooper-  
97 ation, we instead focus our attention here on how niche construction enables  
98 cooperators to escape defection by hitchhiking along with non-social traits.

## 99 **Materials and Methods**

100 We build upon the model described in Hankshaw and Kerr (2015), in which co-  
101 operators and defectors compete and evolve in a spatially-structured metapop-  
102 ulation of populations. Each of these populations grows to carrying capacity,  
103 mutates, and migrates to neighboring patches. During this process, popula-  
104 tions adapt to their local environments. In our extended model, we allow  
105 the presence of these individuals to modify their local environment, and these  
106 modifications feed back to affect selection.

## 107 **Model Description**

108 Our simulated environment consists of  $N^2$  patches arranged as an  $N \times N$   
109 lattice (see [Table 1](#) for model parameters and their values), where each patch  
110 supports a population of zero or more individuals. Each individual in the

111 population has a genotype, which is an ordered list of  $L + 1$  integers (loci).  
 112 At the first locus, a binary allele determines whether or not that individual  
 113 is a cooperator. Individuals with allelic state 1 at this locus are cooperators,  
 114 carrying a cost  $c$ , while individuals with allelic state 0 are defectors. The  
 115 remaining  $L$  loci are *stress loci*, and are each occupied by a 0 or an integer  
 116 from the set  $A = \{1, \dots, a_{max}\}$ , where  $a_{max}$  is the number of potential alleles.  
 117 These alleles represent adaptations to the environment, and the number of loci  
 118 determines the number of adaptations that are possible. All non-zero alleles  
 119 confer fitness benefit  $\delta$ . When  $\delta \geq c$ , an adapted cooperator recoups the cost  
 120 of cooperation.

121 Organisms also influence their environment, which can feed back to influence  
 122 selection. We model this as a form of frequency dependent selection. Specif-  
 123 ically, the selective value of stress allele  $a$  at locus  $l$  increases with the pro-  
 124 portion of the population that has allele  $a - 1$  (modulo  $a_{max}$ ) at locus  $l - 1$   
 125 (and the first stress locus is affected by the last). The slope of this increase  
 126 is  $\epsilon$ , which specifies the intensity of niche construction. As a consequence of  
 127 this form of frequency dependence, genotypes with sequentially increasing al-  
 128 lelic states will tend to evolve. Because mutations are random, as described  
 129 later, each population will evolve sequences that start with different allelic  
 130 states. These different sequences represent the unique niches constructed by  
 131 populations. Under this model, the fitness of an individual with genotype  $g$  in  
 132 population  $P$  is:

$$W_g = z + ca_{g,1} + \delta \sum_{l=2}^{L+1} I_A(a_{g,l}) + \epsilon \sum_{l=2}^{L+1} n(a_{g,l}) \quad (1)$$

where  $z$  is a baseline fitness,  $c$  is the cost of the cooperative allele,  $a_{g,l}$  represents the allelic state of genotype  $g$  at locus  $l$ ,  $L$  is the number of stress loci, and  $\gamma(j)$  is the genotype of individual  $j$ .  $I_A$  indicates whether allelic state  $y$  is in  $A$  (i.e., it is non-zero). The function  $n(a_{g,l})$  gives the number of individuals in the population with allelic state at the previous locus equal to one less than that at the focal locus  $a_{g,l}$ , or:

$$n(a_{g,l}) = \sum_{h=1}^N I_{a_{g,l}}(1 + a_{h,l-1}(\text{mod } a_{\max})) \quad (2)$$

Here,  $I_x(y)$  indicates whether the allelic state  $y$  matches allelic state  $x$  (1) or not (0).

Cooperators produce a public good that is equally accessible to all members of the population. This public good increases the carrying capacity at that patch, allowing the population to reach greater density. This benefit increases linearly with the proportion of cooperators. Thus, if  $p$  is the proportion of cooperators in a population at the beginning of a growth cycle, then that population reaches the following size during the growth phase:

$$S(p) = S_{\min} + p(S_{\max} - S_{\min}) \quad (3)$$

The function  $S(p)$  reflects the benefit of public good production. A population

148 composed entirely of defectors reaches size  $S_{min}$ , while one composed entirely  
 149 of cooperators reaches size  $S_{max}$  (with  $S_{max} \geq S_{min}$ ). During growth, indi-  
 150 viduals compete for inclusion in the resulting population. The composition of  
 151 population  $P$  with cooperator proportion  $p$  after growth is multinomial with  
 152 parameters and  $S(p)$  and  $\{\pi_1, \pi_2, \dots, \pi_{|P|}\}$ , where:

$$\pi_i = \frac{W_{\gamma(i)}}{\sum_{j \in P} W_{\gamma(j)}} \quad (4)$$

153 Here,  $\gamma(i)$  is the genotype of individual  $i$ , and  $W_{\gamma(i)}$  is its fitness (see Equation  
 154 1).  $\pi_i$  is proportional to  $W_{\gamma(i)}$  (and  $\sum_{i \in P} \pi_i = 1$ ).

155 For simplicity, we apply mutations after population growth. Mutations occur  
 156 independently at each locus and cause the allelic state to change. At the binary  
 157 cooperation locus, mutations occur at rate  $\mu_c$ . These mutations flip the allelic  
 158 state, causing cooperators to become defectors and vice versa. Mutations  
 159 occur at each stress locus at rate  $\mu_s$ , and cause a new allelic state to be chosen  
 160 at random from the set  $\{0\} \cup A$ . Therefore, the probability that genotype  $g$   
 161 mutates into genotype  $g'$  is given by:

$$\tau_{g \rightarrow g'} = \mu_c^{H_c(g, g')} (1 - \mu_c)^{\{1 - H_c(g, g')\}} \mu_s^{H_s(g, g')} (1 - \mu_s)^{\{L - H_s(g, g')\}} \quad (5)$$

162 where  $H_c(g, g')$  and  $H_s(g, g')$  are the Hamming distances between genotypes  
 163  $g$  and  $g'$  at the cooperation locus and stress loci, respectively. The Hamming  
 164 distance is the number of loci at which allelic states differ (Hamming, 1950).  
 165 Because we define no inherent relationship among alleles, each of the  $a_{max} + 1$



166 allelic states is equally likely to arise via mutation at a given locus.  
 167 After mutation, individuals emigrate to an adjacent patch at rate  $m$ . The  
 168 destination patch is randomly chosen with uniform probability from the source  
 169 patch's Moore neighborhood, which is composed of the nearest 8 patches on the  
 170 lattice. Because the metapopulation lattice has boundaries, patches located  
 171 on an edge have smaller neighborhoods.  
 172 Metapopulations are initiated in a state that follows the onset of an environ-  
 173 mental stress. First, populations are seeded at all patches with cooperator  
 174 proportion  $p_0$  and grown to density  $S(p_0)$ . Stress is then introduced, which  
 175 subjects the population to a bottleneck. For each individual, the probability of  
 176 survival is  $\mu_t$ , which represents the likelihood that a mutation occurs that con-  
 177 fers tolerance. Survivors are chosen by binomial sampling. Because individuals  
 178 have not yet adapted to this new stress, the allelic state of each individual's  
 179 genotype is set to 0 at each stress locus ( $\forall i \in P, l \in \{2, \dots, L+1\} : a_{\gamma(i),l} = 0$ ).  
 180 Following initialization, simulations are run for  $T$  cycles, where each discrete  
 181 cycle consists of growth, mutation, and migration. At the end of each cycle,  
 182 populations are thinned to allow for growth in the next cycle. The individuals  
 183 that remain are chosen by binomial sampling, where each individual persists  
 184 with probability  $d$ , regardless of allelic state.

## 185 Source Code and Software Environment

186 The simulation software and configurations for the experiments reported are  
 187 available online (Us, 2015). Simulations used Python 3.4.0, NumPy 1.9.1,

188 Pandas 0.15.2 (McKinney, 2010), NetworkX 1.9.1 (Hagberg *et al.*, 2008). Data  
189 analyses were performed with R 3.1.3 (R Core Team, 2015).

## 190 Results

191 results...

## 192 Discussion

- 193 • summary of results
- 194 • similarities/differences from previous work
  - 195 – Schwilk and Kerr (2002)
- 196 • future primacy/recency
  - 197 – Laland et al. (1996)
  - 198 – Lehmann (2007)
- 199 • public goods as niche construction
- 200 • Host symbiont - many instances of cooperation occur among pathogens.
- 201 • future QS or other environmental sensing
- 202 • Facultative cooperation
  - 203 – Rodrigues (2012)
  - 204 – Dumas and Kümmerli (2010)
  - 205 – Kümmerli and Brown (2010)

206           – Darch/Diggle

207           – QS?

208           – Environmental Sensing?

209   Niche construction and selective feedbacks

210   Niche construction and other social interactions # Acknowledgments

211       • PRFB

212       • BEACON

213       • Google

214       • Organizers?

215 **Figures**

Table 1: Model parameters and their value

Parameter	Description	Base Value
$L$	Number of Stress Loci	5
$c$	Production Cost	0.1
$a_{max}$	Number of alleles	6
$\delta$	Fitness benefit, nonzero alleles	0.3
$\epsilon$	Fitness benefit, sequential alleles	TODO
$z$	Baseline fitness	1
$S_{min}$	Minimum Population Size	800
$S_{max}$	Maximum Population Size	2000
$\mu_c$	Mutation Rate (Cooperation)	$10^{-5}$
$\mu_s$	Mutation Rate (Stress)	$10^{-5}$
$N^2$	Number of Metapopulation Sites	625
$m$	Migration Rate	0.05
$p_0$	Initial Cooperator Proportion	0.5
$\mu_t$	Mutation Rate (Tolerance to New Stress)	$10^{-5}$
$T$	Number of Simulation Cycles	1000
$d$	Population Dilution Factor	0.1

## References

- Asfahl, K.L., Walsh, J., Gilbert, K. and Schuster, M. 2015. Non-social adaptation defers a tragedy of the commons in *Pseudomonas aeruginosa* quorum sensing. *The ISME Journal*, doi: [10.1038/ismej.2014.259](https://doi.org/10.1038/ismej.2014.259).
- Brown, S.P. and Johnstone, R.A. 2001. Cooperation in the dark: Signalling and collective action in quorum-sensing bacteria. *Proceedings of the Royal Society of London B: Biological Sciences*, **268**: 961–965.
- Cosson, P., Zulianello, L., Join-Lambert, O., Faurisson, F., Gebbie, L. and Benghezal, M.*et al.* 2002. *Pseudomonas aeruginosa* virulence analyzed in a dictyostelium discoideum host system. *Journal of Bacteriology*, **184**: 3027–3033.
- Dandekar, A.A., Chugani, S. and Greenberg, E.P. 2012. Bacterial quorum sensing and metabolic incentives to cooperate. *Science*, **338**: 264–266.
- Darch, S.E., West, S.A., Winzer, K. and Diggle, S.P. 2012. Density-dependent fitness benefits in quorum-sensing bacterial populations. *Proceedings of the National Academy of Sciences*, **109**: 8259–8263.
- Diggle, S.P., Griffin, A.S., Campbell, G.S. and West, S.A. 2007. Cooperation and conflict in quorum-sensing bacterial populations. *Nature*, **450**: 411–414.
- Fletcher, J.A. and Doebeli, M. 2009. A simple and general explanation for the evolution of altruism. *Proceedings of the Royal Society B: Biological Sciences*, **276**: 13–19.
- Foster, K., Shaulsky, G., Strassmann, J., Queller, D. and Thompson, C. 2004.

239 Pleiotropy as a mechanism to stabilize cooperation. *Nature*, **431**: 693–696.  
240 Nature Publishing Group.

241 Gardner, A. and West, S.A. 2010. Greenbeards. *Evolution*, **64**: 25–38.

242 Griffin, A.S., West, S.A. and Buckling, A. 2004. Cooperation and competition  
243 in pathogenic bacteria. *Nature*, **430**: 1024–1027.

244 Hagberg, A.A., Schult, D.A. and Swart, P.J. 2008. Exploring network struc-  
245 ture, dynamics, and function using NetworkX. In: *Proceedings of the 7th*  
246 *Python in Science Conference (SciPy2008)*, pp. 11–15.

247 Hamilton, W.D. 1964. The genetical evolution of social behaviour I & II.  
248 *Journal of Theoretical Biology*, **7**: 1–52.

249 Hamming, R.W. 1950. Error detecting and error correcting codes. *Bell System*  
250 *Technical Journal*, **29**: 147–160.

251 Hankshaw, S. and Kerr, B. 2015. Thumbs up!: Hitching a ride on the evolu-  
252 tionary superhighway. *Unknown Journal*, **0**: 0–0.

253 Hardin, G. 1968. The tragedy of the commons. *Science*, **162**: 1243–1248.

254 Kuzdzal-Fick, J.J., Fox, S.A., Strassmann, J.E. and Queller, D.C. 2011. High  
255 relatedness is necessary and sufficient to maintain multicellularity in Dic-  
256 tyostelium. *Science*, **334**: 1548–1551.

257 Laland, K.N., Odling-Smee, F.J. and Feldman, M.W. 1996. The evolutionary  
258 consequences of niche construction: A theoretical investigation using two-locus  
259 theory. *Journal of Evolutionary Biology*, **9**: 293–316.

260 Lehmann, L. 2007. The evolution of trans-generational altruism: Kin selec-

tion meets niche construction. *Journal of Evolutionary Biology*, **20**: 181–189.

Blackwell Publishing Ltd.

McKinney, W. 2010. Data structures for statistical computing in Python. In: *Proceedings of the 9th Python in Science Conference* (S. van der Walt and J. Millman, eds), pp. 51–56.

Morgan, A.D., Quigley, B.J.Z., Brown, S.P. and Buckling, A. 2012. Selection on non-social traits limits the invasion of social cheats. *Ecology Letters*, **15**: 841–846.

Nadell, C.D., Foster, K.R. and Xavier, J.B. 2010. Emergence of spatial structure in cell groups and the evolution of cooperation. *PLoS Computational Biology*, **6**: e1000716.

Nowak, M.A. 2006. Five rules for the evolution of cooperation. *Science*, **314**: 1560–1563.

R Core Team. 2015. *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing.

Schwilk, D.W. and Kerr, B. 2002. Genetic niche-hiking: An alternative explanation for the evolution of flammability. *Oikos*, **99**: 431–442.

Sinervo, B., Chaine, A., Clobert, J., Calsbeek, R., Hazard, L. and Lancaster, L.*et al.* 2006. Self-recognition, color signals, and cycles of greenbeard mutualism and altruism. *Proceedings of the National Academy of Sciences*, **103**: 7372–7377.

Us. 2015. Name for data and code.



- 283 Van Dyken, J.D. and Wade, M.J. 2012. Origins of altruism diversity II: Run-  
284 away coevolution of altruistic strategies via “reciprocal niche construction”.  
285 *Evolution*, **66**: 2498–2513.
- 286 Veelders, M., Brückner, S., Ott, D., Unverzagt, C., Mösch, H.-U. and Essen,  
287 L.-O. 2010. Structural basis of flocculin-mediated social behavior in yeast.  
288 *Proceedings of the National Academy of Sciences*, **107**: 22511–22516.
- 289 Waite, A.J. and Shou, W. 2012. Adaptation to a new environment allows coop-  
290 erators to purge cheaters stochastically. *Proceedings of the National Academy*  
291 *of Sciences*, **109**: 19079–19086.
- 292 West, S.A., Diggle, S.P., Buckling, A., Gardner, A. and Griffin, A.S. 2007a.  
293 The social lives of microbes. *Annual Review of Ecology, Evolution, and Sys-*  
294 *tematics*, **38**: 53–77.
- 295 West, S.A., Griffin, A.S. and Gardner, A. 2007b. Evolutionary explanations  
296 for cooperation. *Current Biology*, **17**: R661–R672.