TODO title

₂ TODO

1

10

11

12

13

14

15

16

17

18

19

3 Abstract

Through their interactions, their activities, and even their mere presence, organisms change the environment for themselves and others. This "niche construction" process becomes particularly interesting when it creates evolutionary feedback, whereby selective pressures are altered in response to environmental change. Here we consider how niche construction influences the evolution of cooperation, which has been a longstanding challenge to evolutionary theory. We simulate populations of individuals that cooperatively produce a public good that permits increased growth in a stressful environment and investigate how local- and global-scale niche construction affects the ability of these populations to resist invasion by non-producing cheats. We find that niche construction profoundly impacts the evolution of cooperation by creating new opportunities for adaptation. Cooperators are able to escape subversion by cheats as long as niche construction clears these paths of adaptation. This work provides a crucial step towards understanding how evolution occurs in complex environments like those found in nature.

20 Introduction

Cooperative behaviors are common across all branches of the tree of life. Insects divide labor within their colonies, plants and soil bacteria exchange essential nutrients, birds care for others' young, and the trillions of cells in the human body restrain their growth and coordinate to provide vital functions. Each instance of cooperation presents an evolutionary challenge: How can individuals that sacrifice their own well-being to help others avoid subversion by those that do not? Over time, we would expect these defectors to rise in abundance at the expense of others, eventually driving cooperators—and perhaps the entire population—to extinction. Several factors can defer this potential tragedy of the commons (Hamilton, 1964; Hardin, 1968; Nowak, 2006; West et al., 2007b). For example, cooperators must benefit more from the cooperative act than others. This can occur when cooperators are clustered together in spatially-structured populations 33 (Fletcher and Doebeli, 2009; Nadell et al., 2010; Kuzdzal-Fick et al., 2011) or when cooperators use communication (Brown and Johnstone, 2001; Darch et al., 2012) or other cues (Sinervo et al., 2006; Gardner and West, 2010; Veelders et al., 2010) to cooperate conditionally with kin. Interestingly, cooperation can also be bolstered by genetic linkage with self-benefitting traits (Foster et al., 2004; Dandekar et al., 2012; Asfahl et al., 2015), setting the stage for an "adaptive race" in which cooperators and defectors vie for the first highly-beneficial non-social adaptation (Waite and Shou, 2012; Morgan et al., 2012).

42 Hankshaw and Kerr (2015) recently showed that in spatially-structured popu-

lations, cooperators can gain a substantial leg up on defectors in an adaptive race when the cooperative behavior increases local population density, thus increasing the likelihood of acquiring beneficial non-social mutations. By hitchhiking along with these adaptations, the cooperative trait can rapidly rise in abundance. Nevertheless, this advantage is fleeting. As soon as the opportunities for adaptation are exhausted, cooperators are once again at a disadvantage against defectors, particularly those that arise from within via mutations that disable cooperation. However, Hankshaw and Kerr (2015) demonstrated that cooperation can be maintained indefinitely when frequent environmental changes produce a steady stream of non-social adaptive opportunities. Although organisms typically find themselves in dynamic environments, change might not occur at a rate that provides sufficient adaptive opportunities to ensure long-term cooperator survival. In this work, we examine whether the changes in selection that arise as organisms modify their environment can create opportunities that allow cooperators to be maintained by hitchhiking. We expand upon the model presented in Hankshaw and Kerr (2015), by allowing populations to alter their local environment based on the presence of different non-social adaptations. Frequencydependent selection on these adaptations creates an eco-evolutionary feedback that increasingly favors the adaptations present in each population. We first examine how the intensity of these feedbacks affects the hitchhiking process. Because the production of public goods increases population density, populations containing cooperators will exert a greater influence on their environment. As a result, these environments will be more quickly brought to states where

67 fitness is higher.

As populations construct unique niches, they potentially decrease the threat of invasion from neighboring patches. This occurs when the traits that were advantageous in an immigrant's home niche are maladaptive elsewhere. Because environmental change is influenced solely by non-social phenotypes in 71 this model, this change of invasibility affects cooperators and defectors equally. Here again, however, populations containing a greater number of cooperators may have an advantage. The greater number of individuals that emigrate from these larger populations allow them to "export" their niche—and thus reduce the fitness of neighboring competitors—at a higher rate. We explore whether the range expansion that this process enables provides additional opportunities for cooperation to hitchhike. Finally, we demonstrate how *negative* niche construction, where populations change their environment in ways that reduce fitness, can further support cooperation. Even though the niche construction process creates selective feedbacks, we would expect the magnitude of these feedbacks to decrease as populations evolve. Once individuals can no longer gain adaptations that compensate for the costs of cooperation, they are then outcompeted by non-84 cooperators. However if populations construct their environment in a way 85 which decreases fitness, cooperation can still hitchhike when this change also

creates the opportunity to gain compensatory adaptations.

Materials and Methods

We build upon the model described in Hankshaw and Kerr (2015), in which cooperators and defectors compete and evolve in a spatially-structured metapopulation of populations. Each of these populations grows to carrying capacity,
mutates, and migrates to neighboring patches. During this process, populations adapt to their local environments. In our extended model, we allow
these individuals to modify their local environment, and these modifications
feed back to affect selection.

96 Model Description

Our simulated environment consists of N^2 patches arranged as an $N \times N$ 97 lattice (see Table 1 for model parameters and their values), where each patch 98 supports a population of zero or more individuals. Each individual in the population has a genotype, which is an ordered list of L+1 integers (loci). 100 The first L loci are adaptive loci, and are each occupied by a 0 or an integer 101 from the set $A = \{1, \dots, a_{max}\}$, where a_{max} is the number of potential alleles. 102 At each of these loci, the presence of a non-zero allele represents an adaptation 103 to the environment that confers fitness benefit δ . A binary allele at locus L+1104 determines whether or not that individual is a cooperator. Individuals with 105 allelic state 1 at this locus are cooperators, carrying a cost c, while individuals 106 with allelic state 0 are defectors. When $\delta \geq c$, an adapted cooperator recoups 107 the cost of cooperation. 108

Organisms also influence their environment, which can feed back to influence

selection. We model this as a form of frequency dependent selection. Specifically, the selective value of adaptive allele a at locus l increases with the 111 number of individuals in the population that have allele a-1 (modulo a_{max}) 112 at locus l-1 (modulo L). The slope of this increase is ϵ , which specifies the 113 intensity of niche construction. As a consequence of this form of frequency 114 dependence, genotypes with sequentially-increasing allelic states will tend to 115 evolve. Because mutations are random, as described later, each population 116 will evolve sequences that start with different allelic states. These different sequences represent the unique niches constructed by populations. Under this model, the fitness of an individual with genotype q in population P is:

$$W_g = z + \delta \sum_{l=1}^{L} I_A(a_{g,l}) + \epsilon \sum_{l=1}^{L} n(a_{g,l}) + ca_{g,L+1}$$
 (1)

where z is a baseline fitness, $a_{g,l}$ represents the allelic state of genotype g at locus l, L is the number of adaptive loci, and c is the cost of the cooperative allele. The function I_A indicates whether allelic state y is in A (i.e., it is non-zero). The function $n(a_{g,l})$ gives the number of individuals in the population with allelic state at the previous locus equal to one less than that at the focal locus $a_{g,l}$, or:

$$n(a_{g,l}) = \sum_{i \in P} I_{a_{g,l}} (1 + a_{\gamma(i),\{1 + (l - 2(\bmod L)\}}(\bmod a_{max}))$$
 (2)

Here, $I_x(y)$ indicates whether the allelic state y matches allelic state x (1) or not (0), and $\gamma(j)$ is the genotype of individual j. Note that the selective value

of an allele at the first locus is affected by the allelic state at locus L.

Cooperators produce a public good that is equally accessible to all members of the population. This public good increases the carrying capacity at that patch, allowing the population to reach greater density. This benefit increases linearly with the proportion of cooperators. Thus, if p is the proportion of cooperators in a population at the beginning of a growth cycle, then that population reaches the following size during the growth phase:

$$S(p) = S_{min} + p(S_{max} - S_{min}) \tag{3}$$

The function S(p) reflects the benefit of public good production. A population composed entirely of defectors reaches size S_{min} , while one composed entirely of cooperators reaches size S_{max} (with $S_{max} \geq S_{min}$). During growth, individuals compete for inclusion in the resulting population. The composition of population P with cooperator proportion p after growth is multinomial with parameters and S(p) and $\{\pi_1, \pi_2, \ldots, \pi_{|P|}\}$, where:

$$\pi_i = \frac{W_{\gamma(i)}}{\sum_{j \in P} W_{\gamma(j)}} \tag{4}$$

Here, $\gamma(i)$ is the genotype of individual i, and $W_{\gamma(i)}$ is its fitness (see Equation 1). π_i therefore reflects that an individual's ability to persist is proportional to its fitness relative to others'.

For simplicity, we apply mutations after population growth. Mutations occur independently at each locus and cause the allelic state to change. Mutations

occur at each adaptive locus at rate μ_a , and cause a new allelic state to be chosen at random from the set $\{0\} \cup A$. At the binary cooperation locus, mutations occur at rate μ_c . These mutations flip the allelic state, causing cooperators to become defectors and vice versa. Therefore, the probability that genotype g mutates into genotype g' is given by:

$$\tau_{g \to g'} = \mu_a^{H_a(g, g')} (1 - \mu_a)^{\{L - H_a(g, g')\}} \mu_c^{H_c(g, g')} (1 - \mu_c)^{\{1 - H_c(g, g')\}}$$
 (5)

where $H_a(g, g')$ and $H_c(g, g')$ are the Hamming distances between genotypes gand g' at the cooperation locus and adaptive loci, respectively. The Hamming 152 distance is the number of loci at which allelic states differ (Hamming, 1950). 153 Because we define no inherent relationship among alleles, each of the $a_{max} + 1$ 154 allelic states is equally likely to arise via mutation at a given locus. 155 After mutation, individuals emigrate to an adjacent patch at rate m. The 156 destination patch is randomly chosen with uniform probability from the source 157 patch's Moore neighborhood, which is composed of the nearest 8 patches on the 158 lattice. Because the metapopulation lattice has boundaries, patches located 159 on an edge have smaller neighborhoods. 160 Metapopulations are initiated in a state that follows an environmental change. 161 First, populations are seeded at all patches with cooperator proportion p_0 and 162 grown to density $S(p_0)$. An environmental challenge is then introduced, which 163 subjects the population to a bottleneck. For each individual, the probability 164 of survival is μ_t , which represents the likelihood that a mutation occurs that 165

confers tolerance. Survivors are chosen by binomial sampling. Because indi-

viduals have not yet adapted to this new environment, the allelic state of each individual's genotype is set to 0 at each adaptive locus ($\forall i \in P, l \in \{1, ..., L\}$: $a_{\gamma(i),l} = 0$). Following initialization, simulations are run for T cycles, where each discrete cycle consists of growth, mutation, and migration. At the end of each cycle, populations are thinned to allow for growth in the next cycle. The individuals that remain are chosen by binomial sampling, where each individual persists with probability d, regardless of allelic state.

Source Code and Software Environment

The simulation software and configurations for the experiments reported are available online (Us, 2015). Simulations used Python 3.4.0, NumPy 1.9.1, Pandas 0.15.2 (McKinney, 2010), and NetworkX 1.9.1 (Hagberg *et al.*, 2008). Data analyses were performed with R 3.1.3 (R Core Team, 2015).

179 Results

Niche construction prolongs cooperator survival

"cooperator presence" greater in NC runs than with no-NC (GNH)

Negative Niche construction helps

Runs with wraparound persist indefinitely, while those without do not.

184 NC Prevents/Limits Defector invasion

NC Enables Cooperator Spread by "exporting" environ-

186 ment

187 Discussion

```
• summary of results
```

- similarities/differences from previous work
- Schwilk and Kerr (2002)
- -10.1073/pnas.0812644106
- public goods as niche construction
- future QS or other environmental sensing
- Facultative cooperation
- Rodrigues (2012)
- Dumas and Kümmerli (2010)
- Kümmerli and Brown (2010)
- Darch/Diggle
- QS?
- Environmental Sensing? (Koestler and Waters, 2014, Bernier et al. (2011))
- Negative Niche construction as a stragegy? would those that create this constant pressure (L=5, A=6) do better than those that do not (L=5, A=6)

A=5?

Niche construction and selective feedbacks Niche construction and other social interactions

207 Public Goods

TODO: merge this in with the "Cooperative Niche Construction" section

TEST

By their very nature, public goods benefit populations by making their environ-210 ment more hospitable. For example, bacteria produce extracellular products 211 that find soluble iron (Griffin et al., 2004), digest large proteins (Diggle et al., 212 2007; Darch et al., 2012), and reduce the risk of predation (Cosson et al., 2002), 213 among many others (West et al., 2007a). While many studies have explored 214 how the environment affects the evolution of cooperative bahviors, relatively 215 few have examined how those behaviors affect the environment and the result-216 ing change in evolutionary trajectories. Lehmann (2007) demonstrated analyti-217 cally that when niche construction act benefits future generations, cooperation 218 is favored due to reduced competition among kin. When rate-benefitting and 219 yield-benefitting altruistic acts co-evolve, Van Dyken and Wade (2012) showed that "reciprocal niche construction", where the selective feedbacks produced 221 by one act benefitted the other, can lead to increased selection for both traits. 222 While these studies have focused on the niche constructing effects of cooperation, we instead focus our attention here on how niche construction enables cooperators to escape defection by hitchhiking along with non-social traits.

226 Primacy/Recency

In our model, alterations to the environment were immediately echoed by changes in selection. However, decoupling the timescales on which these pro-228 cesses occur can have substantial effects (Laland et al., 1996). By integrating past allelic states into Equation 1, we can begin to explore how the cumulative 230 effects of niche construction affect the creation of non-social adaptive oppor-231 tunities and the benefits that they offer cooperation. Here, how these past 232 allelic states are integrated will play an important role. For example, when 233 the effects of earlier generations are weighted more heavily, the influence of 234 migration may be diminished. While this will reduce the threat of emigration 235 by defectors, cooperator populations will also be less effective at exporting 236 their niche. 237

238 Cooperative Niche Construction

While our focus for this work has been on the eco-evolutionary feedbacks created by non-social traits, it would also be interesting to explore how this system
is affected by the timescale at which carrying capacity at a given patch is increased by public goods. In natural settings, a multitude of factors including
protein durability (Brown, 2007; Kümmerli and Brown, 2010), diffusion (Allison, 2005; Driscoll and Pepper, 2010), and resource availability (Zhang and
Rainey, 2013; Ghoul et al., 2014) influence both the rate and the degree to
which public goods alter the environment (and thereby selection). Lehmann
(2007) demonstrated that a cooperative, niche constructing behavior can be fa-

vored when it only affected selection for future generations, thus reducing the potential for competition among contemporary kin. The evolutionary inertia that this creates, however, may ultimately work against cooperators. When public good accumulates in the environment, cooperators must reduce their investment in production to remain competitive (Kümmerli and Brown, 2010).

TODO: wrap up. Facultative cooperation requires sensing.

Host-Symbiont

In many instances of cooperation, the environment is itself a biological entity, which can produce additional evolutionary feedbacks. As the host population 256 changes, so too will selection on their symbiont populations. Here, evolution-257 ary outcomes depend greatly on the degree of shared interest between the host 258 and symbiont. For example, the cooperative production of virulence factors by 259 the human pathogen P. aeruqinosa in lung infections is harmful to those with 260 cystic fibrosis (Harrison, 2007). Conversely, cooperative light production by A. 261 fischeri is vital for the survival of its host, the Hawaiian bobtail squid (Ruby, 262 1996). It was recently argued that incorporating the effects of niche construc-263 tion is critical for improving our understanding of viral evolution (Hamblin et 264 al., 2014) and evolution in co-infecting parasites (Hafer and Milinski, 2015). 265 Incorporating host dynamics, co-evolution, and the feedbacks that they pro-266 duce into models is likely to be equally important for gaining an understanding of how cooperative behaviors evolve in these host-symbiont settings.

Acknowledgments

• TODO: Organizers?

• TODO: lab comments

We thank Anuraag Pakanati for assistance with simulations. This material is based upon work supported by the National Science Foundation Postdoctoral Research Fellowship in Biology under Grant No. 1309318 (to BDC) and under Cooperative Agreement No. DBI-0939454. Any opinions, findings, and conclusions or recommendations expressed in this material are those of the authors and do not necessarily reflect the views of the National Science Foundation. Computational resources were provided by an award from Google (to BDC and BK).

Figures Figures

Figure 1

Figure 1A

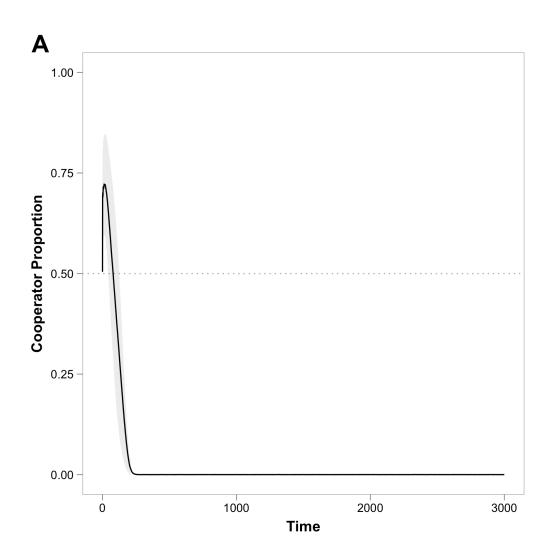


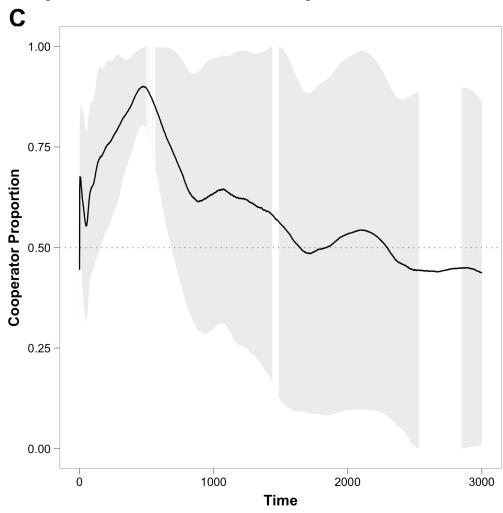
Figure 1: Proportion of cooperators over time when there are no opportunities for non-social adaptation

²⁸³ Figure 1B

²⁸⁴ Proportion of cooperators over time with non-social adaptation

Figure 1C

286 Proportion of cooperators over time with non-social adaptation and niche con-



struction

²⁸⁸ {#fig:fig1c}

Figure 2

- 290 Plot showing integral for baseline values, double adaptive benefit (no NC,
- epsilon=0), no negative NC (L=5, A=5), Extreme negative niche construction
- 292 (L=1, A=6)

Figure 3

 294 Mean fitness over time for the treatments shown in Figure 2

Figure 4

296 Cooperators invade from single population

Figure 5

298 Defectors are kept at bay

Figure 6

- 300 Figure 6A Effect of Migration Rate (m)
- Figure 6B Effect of Public Good Benefit (Smax-Smin)

Tables

Table 1: Model parameters and their value

Parameter	Description	Base Value
$\overline{N^2}$	Number of metapopulation sites	625
L	Number of adaptive loci	5
a_{max}	Number of alleles	6
δ	Fitness benefit, nonzero alleles	0.3
c	Production cost	0.1
ϵ	Fitness benefit, sequential alleles	0.00015
z	Baseline fitness	1
S_{min}	Minimum population size	800
S_{max}	Maximum population size	2000
μ_c	Mutation rate (cooperation)	10^{-5}
μ_a	Mutation rate (adaptation)	10^{-5}
m	Migration rate	0.05
p_0	Initial cooperator proportion	0.5
μ_t	Mutation rate (tolerance to new stress)	10^{-5}
T	Number of simulation cycles	1000
d	Population dilution factor	0.1

References

- Allison, S.D. 2005. Cheaters, diffusion and nutrients constrain decomposition
- by microbial enzymes in spatially structured environments. Ecology Letters,
- **8**: 626–635.
- Asfahl, K.L., Walsh, J., Gilbert, K. and Schuster, M. 2015. Non-social adap-
- tation defers a tragedy of the commons in Pseudomonas aeruginosa quorum
- sensing. The ISME Journal, doi: 10.1038/ismej.2014.259.
- Bernier, S.P., Ha, D.-G., Khan, W., Merritt, J.H.M. and O'Toole, G.A. 2011.
- Modulation of Pseudomonas aeruginosa surface-associated group behaviors by
- individual amino acids through c-di-GMP signaling. Research in Microbiology,
- 313 **162**: 680–688.
- Brown, F., Sam P. AND Taddei. 2007. The durability of public goods changes
- the dynamics and nature of social dilemmas. PLoS ONE, 2: e593. Public
- 316 Library of Science.
- Brown, S.P. and Johnstone, R.A. 2001. Cooperation in the dark: Signalling
- and collective action in quorum-sensing bacteria. Proceedings of the Royal
- 319 Society of London B: Biological Sciences, 268: 961–965.
- Cosson, P., Zulianello, L., Join-Lambert, O., Faurisson, F., Gebbie, L. and
- Benghezal, M. et al. 2002. Pseudomonas aeruginosa virulence analyzed in a
- dictyostelium discoideum host system. Journal of Bacteriology, 184: 3027-
- 323 3033.
- Dandekar, A.A., Chugani, S. and Greenberg, E.P. 2012. Bacterial quorum

- sensing and metabolic incentives to cooperate. Science, 338: 264–266.
- Darch, S.E., West, S.A., Winzer, K. and Diggle, S.P. 2012. Density-dependent
- fitness benefits in quorum-sensing bacterial populations. Proceedings of the
- National Academy of Sciences, 109: 8259–8263.
- Diggle, S.P., Griffin, A.S., Campbell, G.S. and West, S.A. 2007. Cooperation
- and conflict in quorum-sensing bacterial populations. *Nature*, **450**: 411–414.
- Driscoll, W.W. and Pepper, J.W. 2010. Theory for the evolution of diffusible
- ³³² external goods. *Evolution*, **64**: 2682–2687.
- Fletcher, J.A. and Doebeli, M. 2009. A simple and general explanation for the
- evolution of altruism. Proceedings of the Royal Society B: Biological Sciences,
- **276**: 13–19.
- Foster, K., Shaulsky, G., Strassmann, J., Queller, D. and Thompson, C. 2004.
- Pleiotropy as a mechanism to stabilize cooperation. *Nature*, **431**: 693–696.
- Nature Publishing Group.
- Gardner, A. and West, S.A. 2010. Greenbeards. Evolution, 64: 25–38.
- Ghoul, M., West, S.A., Diggle, S.P. and Griffin, A.S. 2014. An experimental
- test of whether cheating is context dependent. Journal of Evolutionary Biology,
- **27**: 551–556.
- Griffin, A.S., West, S.A. and Buckling, A. 2004. Cooperation and competition
- in pathogenic bacteria. Nature, 430: 1024–1027.
- Hafer, N. and Milinski, M. 2015. When parasites disagree: Evidence
- for parasite-induced sabotage of host manipulation. Evolution, doi:

- 347 10.1111/evo.12612.
- Hagberg, A.A., Schult, D.A. and Swart, P.J. 2008. Exploring network struc-
- ture, dynamics, and function using NetworkX. In: Proceedings of the 7th
- Python in Science Conference (SciPy2008), pp. 11–15.
- Hamblin, S.R., White, P.A. and Tanaka, M.M. 2014. Viral niche construction
- alters hosts and ecosystems at multiple scales. Trends in Ecology & Evolution,
- **29**: 594–599.
- Hamilton, W.D. 1964. The genetical evolution of social behaviour I & II.
- Journal of Theoretical Biology, 7: 1–52.
- Hamming, R.W. 1950. Error detecting and error correcting codes. Bell System
- 357 Technical Journal, **29**: 147–160.
- Hankshaw, S. and Kerr, B. 2015. Thumbs up!: Hitching a ride on the evolu-
- tionary superhighway. *Unknown Journal*, **0**: 0–0.
- 360 Hardin, G. 1968. The tragedy of the commons. *Science*, **162**: 1243–1248.
- Harrison, F. 2007. Microbial ecology of the cystic fibrosis lung. *Microbiology*,
- 362 **153**: 917–923.
- 363 Koestler, B.J. and Waters, C.M. 2014. Bile acids and bicarbonate inversely
- regulate intracellular cyclic di-GMP in Vibrio cholerae. Infection and Immu-
- 365 nity, **82**: 3002–3014.
- Kuzdzal-Fick, J.J., Fox, S.A., Strassmann, J.E. and Queller, D.C. 2011. High
- relatedness is necessary and sufficient to maintain multicellularity in Dic-
- ³⁶⁸ tyostelium. *Science*, **334**: 1548–1551.

- Kümmerli, R. and Brown, S.P. 2010. Molecular and regulatory properties of
- a public good shape the evolution of cooperation. Proceedings of the National
- 371 Academy of Sciences, **107**: 18921–18926.
- Laland, K.N., Odling-Smee, F.J. and Feldman, M.W. 1996. The evolutionary
- consequences of niche construction: A theoretical investigation using two-locus
- theory. Journal of Evolutionary Biology, 9: 293–316.
- Lehmann, L. 2007. The evolution of trans-generational altruism: Kin selec-
- tion meets niche construction. Journal of Evolutionary Biology, 20: 181–189.
- 377 Blackwell Publishing Ltd.
- McKinney, W. 2010. Data structures for statistical computing in Python. In:
- Proceedings of the 9th Python in Science Conference (S. van der Walt and J.
- 380 Millman, eds), pp. 51–56.
- Morgan, A.D., Quigley, B.J.Z., Brown, S.P. and Buckling, A. 2012. Selection
- on non-social traits limits the invasion of social cheats. *Ecology Letters*, 15:
- 383 841–846.
- Nadell, C.D., Foster, K.R. and Xavier, J.B. 2010. Emergence of spatial struc-
- ture in cell groups and the evolution of cooperation. PLoS Computational
- зв6 *Biology*, **6**: e1000716.
- Nowak, M.A. 2006. Five rules for the evolution of cooperation. Science, 314:
- зва 1560-1563.
- R Core Team. 2015. R: A language and environment for statistical computing.
- ³⁹⁰ Vienna, Austria: R Foundation for Statistical Computing.

- Ruby, E.G. 1996. Lessons from a cooperative, bacterial-animal association:
- The Vibrio fischeri–Euprymna scolopes light organ symbiosis. Annual Review
- оf Microbiology, **50**: 591–624.
- 394 Schwilk, D.W. and Kerr, B. 2002. Genetic niche-hiking: An alternative expla-
- nation for the evolution of flammability. Oikos, 99: 431–442.
- Sinervo, B., Chaine, A., Clobert, J., Calsbeek, R., Hazard, L. and Lancaster,
- Let al. 2006. Self-recognition, color signals, and cycles of greenbeard mutu-
- ³⁹⁸ alism and altruism. Proceedings of the National Academy of Sciences, 103:
- 399 7372-7377.
- 400 Us. 2015. Name for data and code.
- Van Dyken, J.D. and Wade, M.J. 2012. Origins of altruism diversity II: Run-
- away coevolution of altruistic strategies via "reciprocal niche construction".
- 403 Evolution, **66**: 2498–2513.
- Veelders, M., Brückner, S., Ott, D., Unverzagt, C., Mösch, H.-U. and Essen,
- 405 L.-O. 2010. Structural basis of flocculin-mediated social behavior in yeast.
- 406 Proceedings of the National Academy of Sciences, 107: 22511–22516.
- Waite, A.J. and Shou, W. 2012. Adaptation to a new environment allows coop-
- erators to purge cheaters stochastically. Proceedings of the National Academy
- of Sciences, **109**: 19079–19086.
- West, S.A., Diggle, S.P., Buckling, A., Gardner, A. and Griffin, A.S. 2007a.
- 411 The social lives of microbes. Annual Review of Ecology, Evolution, and Sys-
- 412 tematics, **38**: 53–77.

- West, S.A., Griffin, A.S. and Gardner, A. 2007b. Evolutionary explanations
- for cooperation. Current Biology, 17: R661–R672.
- ⁴¹⁵ Zhang, X.-X. and Rainey, P.B. 2013. Exploring the sociobiology of pyoverdin-
- producing Pseudomonas. Evolution, 67: 3161–3174.