COPAL Tool -- User Guide

Input file(s) details

The input data file(s):

- should be in csv or tsv text or excel format.
- should contain slices in separate columns, and proteins in rows.
- should contain a row of unique headers for each column
- should contain at least one column with identifiers for the proteins
- can contain any number of extra columns or excel sheets
- can contain multiple samples in one excel sheet/text file
- cannot contain information in rows below the protein migration data
- In the case of multiple files/sheets containing samples, protein identifiers should match, so the datasets can be combined.

Input instructions

Input frames

filename:

- Enter filename with file path, or select a file using the choose file button.
- Files can be either excel, .csv or .tsv files. Select the appropriate file format from the dropdown menu.
- In the case of an excel file: enter the name of the sheet containing the data in the sheetname entry. When using an text file, this entry will be ignored.

File details:

- Skip rows: in the case of more than one header row, skip all but one.
  (ie: 3 header rows → skip rows : 2). The 3rd header row will be used for the analysis, and should match the header names specified below.
- Protein identifier column: enter header of column containing protein identifiers

sample names and columns:

- In the first box, enter names for samples contained in this file/sheet
- In the first and last column boxes, enter first and last column header for the corresponding samples on the same line.
- Take care not to leave any unnecessary spaces, tabs or empty lines in these text boxes

Add another file:

- Use this option if samples from other excel sheets or excel/text files are to be added
- A new window will pop up

Proceed to output:

- If all input files and samples are specified, use the proceed to output button

Output frame

Job name: Enter preferred name of this analysis job, output folder and files will have this name

Output folder: Select folder location for output files

Data normalization:

- select type of normalization
- None: no normalization will be performed (only recommended if data has been normalized already)
- Using all Proteins: normalization will be based on complete data
- From column: A column in the input file with True/False specifies which proteins will be used for normalization. Enter column header in 'if from column' entry box
- From file: a separate plain text file containing a list of proteins is used for normalization. Enter file name and path in 'if from file' entry box (or use Choose file button). The file should contain one protein identifier on each line (that correspond with specified identifier column in input).

score analysis:

- check the box if hausdorff scores should be determined
- if checked, enter sample names in group 1 and 2 boxes, one sample name per line.
- Hausdorff scores will be determined between these groups
- Take care to not leave any unnecessary enters spaces or tabs in the entry boxes
- the hausdorff factor determines the protein abundance/molecular mass axes ratio of the plane in which hausdorff distances are calculated. Taking the default value of one will consider both dimensions equally. A higher value will weigh differences in protein abundance more heavily, where a value < 1 will weigh shifts in molecular mass more heavily.

Provide rank ordered protein list:

- This option can provide a ranked list of proteins with combined hausdorff scores. This list is in the .rnk format (which can be readily used in Gene Set Enrichment Analysis).
- If this option is checked, enter the header of the column containing protein identifiers to be used in rank ordered file in the entry box (for example, the "gene symbols" column if this is what is required for further analysis).

Back to start:

goes back to first frame, all data will have to be entered again. To be used in when a second dataset will be aligned, or when the input was wrong.

Save and run:

If all input is entered correctly, pressing save and run will start the complexome alignment process.

Status bar:

- Text box at the bottom will indicate the status of the analysis.
- If the analysis is complete, or if an error occurs, it will be displayed here.

Example walkthrough

This walkthrough shows how to run complexome profile alignment of the test data provided. In this example 9 samples spread over 3 input files will be aligned and scored. Normalisation of the samples is performed based on all proteins.

When you start the GUI the first input frame shows (figure 1-3). The first file containing a set of complexome profiles is specified here. Pressing the “add another file” button will create another input frame. Once the last input file is entered, press the “proceed to output” button. If all is well, pressing the “proceed to output” button will open the output details frame (figure 4). If the next frame is not opened, there is a problem with the input. In this case, consult the message at the bottom of the frame for more details.

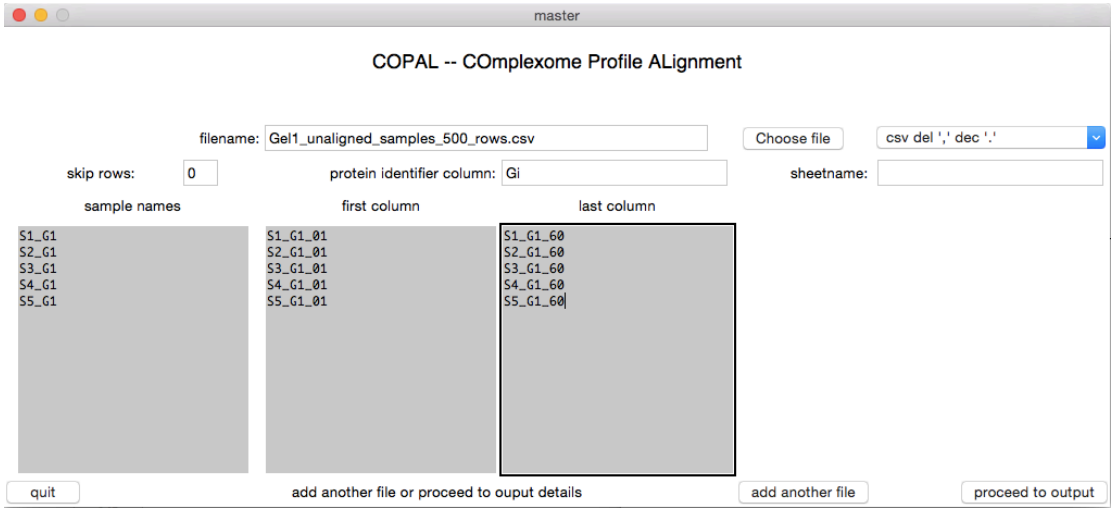

Figure 1: First input frame

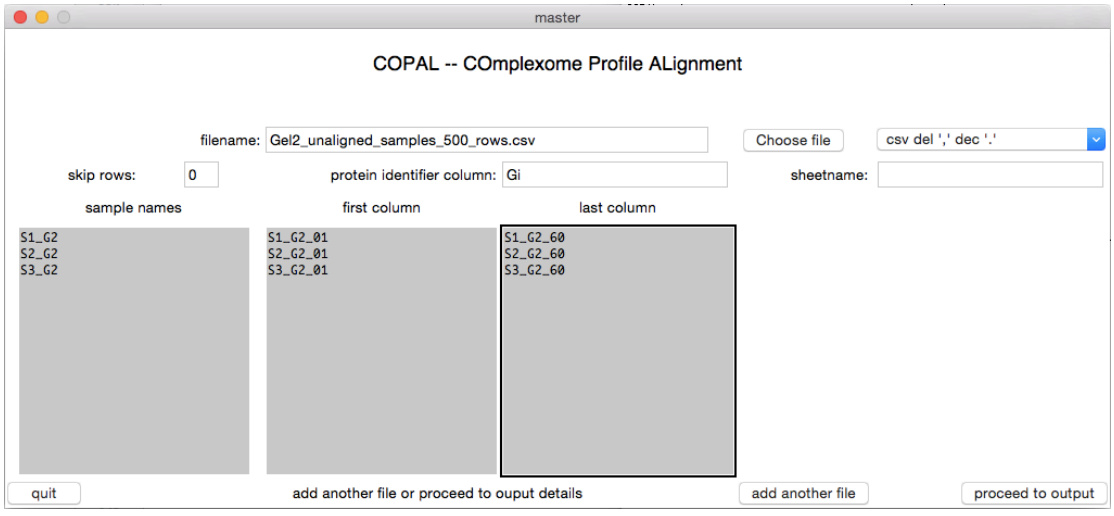

Figure 2: Second input frame

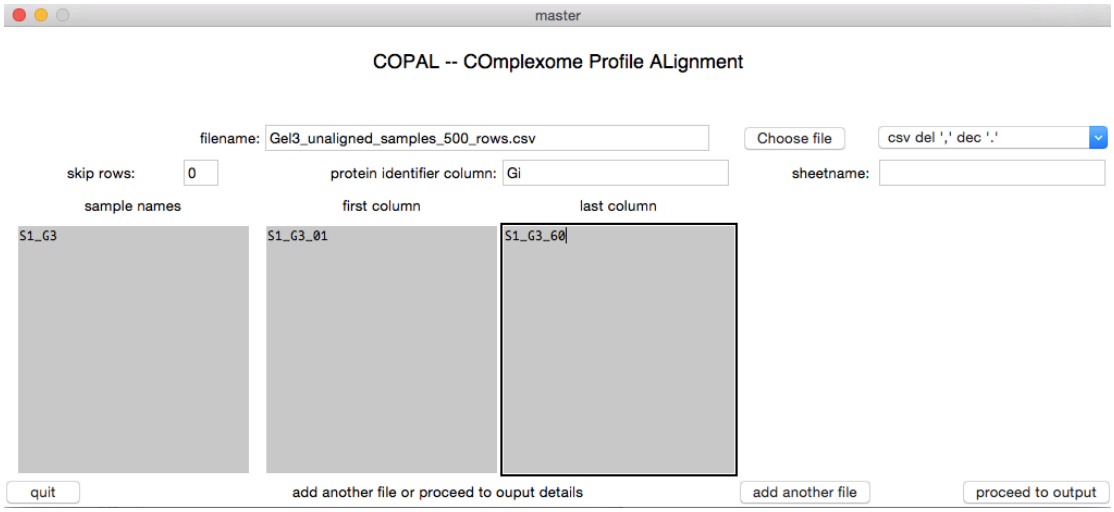

Figure 3: Third input frame

The output frames displays a list of all entered samples. Specify a name for the analysis job, and then specify a preferred output folder. Use the drop down menu to specify a type of normalization. Check the perform score analysis box if hausdorff effect sizes should be calculated after alignment. Enter sample names in the right groups for hausdorff effect sizes. If a rank ordered file with hausdorff effect sizes should be included in the output, check the "provide ranked protein list for GSEA box". Enter the name of the column to be added as protein identifier to the rank ordered file. To start the analysis, click save and run. This saves all input parameters and starts the analysis. The message box at the bottom displays the progress, or displays an error message is there is a problem. When the analysis is finished, a message will display the location of the folder containing the analysis results.

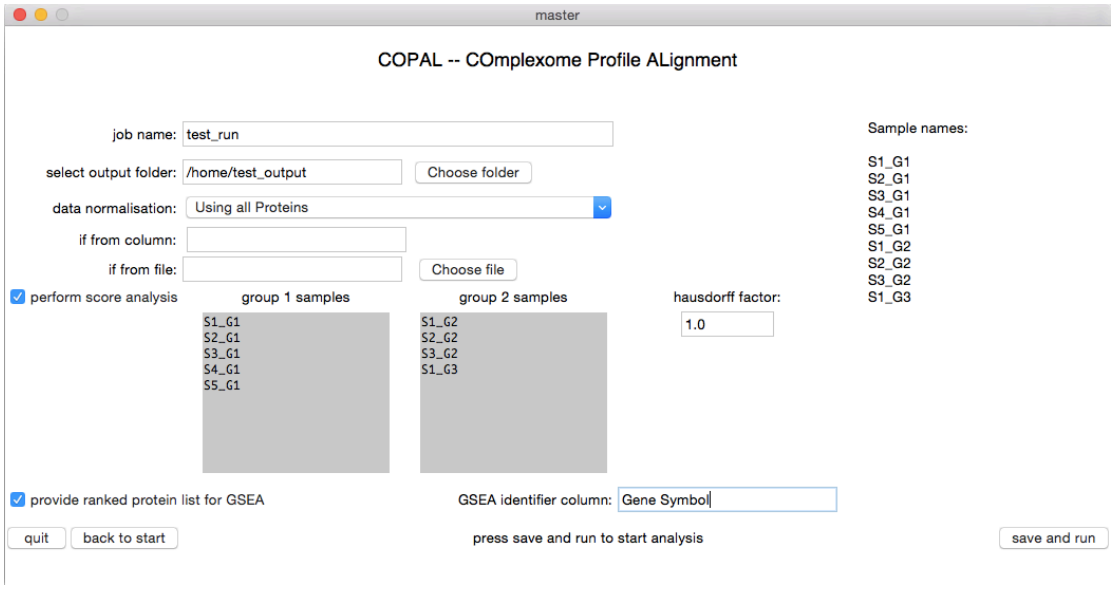

Figure 4: Output details frame