Prototype QTL Strategy: Phenotype bp in Cross hyper

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1 Overview

This document analyzes trait bp for dataset hyper using the 1-D and 2-D Bayesian genome scan routines that build on Markov chain Monte Carlo (MCMC) samples from a posterior for the genetic architecture of a trait. Below the generic cross is actually the cross passed by the user in a call to qb.sweave. This entire document was created automatically by a function call in R. The function is not yet part of R/qtlbim. The actual call was

```
> library(qtlbim)
```

```
> qb.sweave(hyper, pheno.col = 1,
+ n.iter = 3000, n.draws = 64,
+ scan.type = "2logBF", hpd.level = 0.5,
+ threshold = c(upper = 2),
+ SweaveFile = "/tmp/Rinst1519441970/qtlbim/doc/hyperslide.Rnw",
+ SweaveExtra = "/tmp/Rinst1519441970/qtlbim/external/hyperpaperextra.Rnw",
+ PDFDir = "bpPDF",
+ remove.qb = TRUE)
```

At present, the threshold values are somewhat arbitrary, chosen for the hyper dataset to pick up apparently real QTL and previously detected epistasis.

This document automates a search for main and epistatic QTL. The main QTL positions are reliably estimated using qb.scanone. This pass also seems to capture the major chromosomes possibly involved in epistasis, although it does not provide very good estimates of positions of purely epistatic QTL within those chromosomes. Next we use qb.scantwo and to identify which pairs of QTL are epistatic, and to get initial estimates of their positions. We refine there positions with qb.slice. Along the way, we use generic summary and plot routines to view results.

Once we have reasonable estimates of QTL postions and effects, we use confirmatory ANOVA tools to refine the model. That is, we use R/qtl's simulation-based fitqtl followed by a stepwise backward fitting approach, using a new step.fitqtl, to confirm key QTL. That completes this automated analysis. It would be possible to add other, user-supplied refined analysis at the end of this document if desired.

2 Generating Samples

Here is a summary of the cross copy of the hyper object, followed by creation of 3000 MCMC samples.

> summary(cross)

Backcross

No. individuals: 250

No. phenotypes: 1

```
Percent phenotyped: 100

No. chromosomes: 19
    Autosomes: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19

Total markers: 170
    No. markers: 22 8 6 20 14 11 7 6 5 5 14 5 5 5 11 6 12 4 4
    Percent genotyped: 47.9
    Genotypes (%): AA:50.1 AB:49.9

> cross <- qb.genoprob(cross, step=2)
> cross.qb <- qb.mcmc(cross, pheno.col = pheno.col
+ genoupdate=TRUE, n.iter = 3000, mydir = "bpPDF")
```

3 1-D Scans

Now a 1-D scan picks out the major effects. We could uxe qb.scanone directly. Instead, we use qb.hpdone, which gives us a profile scan as well as a scan of genotypic means.

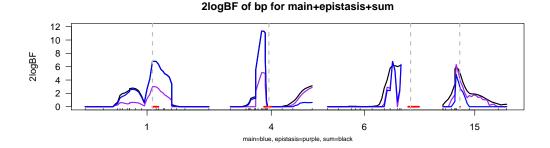
```
> hpd.level
[1] 0.5
> cross.hpd <- qb.hpdone(cross.qb, hpd.level)</pre>
> sum.one <- summary(cross.hpd)
> sum.one
     chr n.qtl pos lo.50% hi.50% 2logBF
       1 0.694 64.5
<NA>
                      64.5
                              69.9 6.796 103.604
                                                   99.073
       4 3.460 29.5
                      25.1
                              31.7 11.347 104.561 98.026
<NA>
<NA>
       6 1.107 59.0
                      56.8
                              66.7 6.179 99.606 102.924
     15 0.341 17.5
                      17.5
                              17.5 6.032 101.940 100.692
<NA>
```

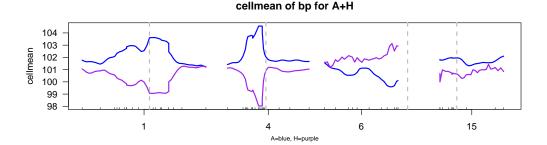
The qb.hpdone routine mostly automates the selection of peaks. We are still working on how to set reasonable thresholds, but for now a HPD level of 0.5 for the overall (sum) picks up the key features nicely for many phenotypes. The variable hpd.level was set in the call to qb.sweave that created this document. The new variables chrs and pos capture the chromosome numbers and main QTL positions, respectively.

```
> chrs <- as.vector(sum.one[, "chr"])
> pos <- sum.one[, "pos"]
> pos
<NA> <NA> <NA> <NA> <NA> 64.5 29.5 59.0 17.5
```

The following two figures highlight the selected chromosomes. In the top panel, the blue curve represents the 2logBF (twice log of Bayes factor) score for the main effect of a QTL at each locus, while the purple curve shows the score for any epistasis between a locus and any other loci. The black curve shows the combination of main and epistatic effects. The second panel shows the marginal means, which are roughly symmetric about the phenotype mean of 101.

```
> plot(cross.hpd)
```





4 2-D Scan

Now a look at a 2-D scan reveals the strength of epistasis. The summary suggests some other epistases, but some of this may be spurious [i.e. we will want to look further!]. We consider a subset of this summary above the upper threshold of 2.

```
> two <- qb.scantwo(cross.qb, chr = chrs, type = scan.type)
> sum.two <- summary(two, threshold = threshold, refine = TRUE)
> sum.two
      chr1 chr2 n.qtl l.pos1 l.pos2 lower u.pos1 u.pos2
                                                            upper
1:1
              1 0.362
                        43.7
                                77.6
                                      7.583 43.700 75.400
                                                            4.697
1:4
         1
              4 1.352
                        67.8
                                29.5 15.705 72.100 29.500
                                                            7.303
1:6
         1
              6 1.831
                        67.8
                                59.0 12.611 77.600 65.600
                                17.5 12.012 77.600 17.500
             15 1.145
                        67.8
                                                            5.794
1:15
         1
              4 0.298
                        29.5
                                74.3 11.820 2.029 30.600
                                                            4.756
4:4
         4
              6 1.561
                                66.7 14.884 74.300 59.000
4:6
                        29.5
                                                            7.728
4:15
             15 0.446
                        29.5
                                17.5 14.539 74.300 35.546
                                                            7.350
         6
              6 1.214
                                     7.442 27.300 66.700
                                                            4.756
6:6
                        61.2
                                66.7
6:15
         6
             15 1.080
                        59.0
                                17.5 12.779 59.000 17.500 12.751
                        17.5
                                27.5 8.125 17.500 25.500 7.234
15:15
        15
             15 0.105
```

Now let's extract the genetic architecture implied by this evidence for epistasis. The loci pairs that show epistasis are indexed to the vector of main QTL. In addition we see the pairs of chromosomes involved in epistasis. Some effort is made to merge nearby QTL estimated positions in qb.arch.

```
Epistatic pairs by qtl, chr, pos:
  qtla qtlb chra chrb posa posb
                   1 43.70 73.44
              1
                   4 73.44 29.87
3
    2
         7
                   6 73.44 61.86
              1
              1 15 73.44 19.10
    2
         8
                   4 2.03 29.87
5
    3
         4
              4
6
    5
         7
              4
                   6 74.30 61.86
7
     5
              4
         9
                 15 74.30 35.55
8
     6
         7
              6
                   6 27.30 61.86
    7
              6
                 15 61.86 19.10
Epistatic chromosomes by connected sets:
1,4,6,15
```

5 ANOVA Model Fit

Here we want to merge the 1-D chrs and pos with the 2-D epistatic pairs to determine the chromosomes and positions to include in an ANOVA fit. We equate QTL that are within, say 10cM of each other. After fitting a (very) full model, we use step.fitqtl, a newly written routine, to step-by-step reduce the model to key main effects and interactions, preserving hierarchy.

The following uses R/qtl tools calc.genoprob, sim.geno and makeqtl, plus the new step.fitqtl, which calls fitqtl multiple times.

```
> cross.sub <- subset(cross, chr = cross.arch$qtl$chr)</pre>
> n.draws
[1] 64
> cross.sub <- sim.geno(cross.sub, n.draws = n.draws, step = 2,
      error = 0.01)
> qtl <- makeqtl(cross.sub, cross.arch$qtl$chr, cross.arch$qtl$pos)
  Now we run stepwise backward elimination, preserving hierarchy.
> cross.step <- step.fitqtl(cross.sub, qtl, pheno.col, cross.arch)</pre>
  drop
                        LOD
1 Chr1073.44:Chr6061.86 0.0678 0.592
2 Chr4@2.03:Chr4@29.87 0.1030 0.508
3 Chr6@27.3:Chr6@61.86 0.2930 0.263
4 Chr6@27.3
                        0.2640 0.287
5 Chr4@2.03
                        0.2910 0.262
6 Chr1073.44:Chr15019.1 0.4520 0.161
7 Chr1@43.7:Chr1@73.44 0.4110 0.181
> summary(cross.step$fit)
Full model result
       df
                SS
                          MS
                                   LOD
                                           %var Pvalue(Chi2) Pvalue(F)
Model 11 7533.98 684.90732 30.17357 42.63969
Error 238 10134.96 42.58385
Total 249 17668.94
```

Drop one QTL at a time ANOVA table:

	df	Type III SS	LOD	%var	${\tt F} \ {\tt value}$	Pvalue(F)
Chr1@43.7	1	358.110	1.885	2.027	8.410	0.004082 **
Chr1073.44	2	875.544	4.498	4.955	10.280	5.22e-05 ***
Chr4@29.87	2	2481.225	11.888	14.043	29.133	4.81e-12 ***
Chr4074.3	3	1318.237	6.638	7.461	10.319	2.07e-06 ***
Chr6@61.86	3	1928.498	9.456	10.915	15.096	5.02e-09 ***
Chr15@19.1	2	1377.604	6.919	7.797	16.175	2.59e-07 ***
Chr15@35.55	2	811.959	4.184	4.595	9.534	0.000104 ***
Chr1@73.44:Chr4@29.87	1	199.916	1.060	1.131	4.695	0.031250 *
Chr4074.3:Chr6061.86	1	300.820	1.588	1.703	7.064	0.008397 **
Chr4@74.3:Chr15@35.55	1	775.841	4.004	4.391	18.219	2.84e-05 ***
Chr6@61.86:Chr15@19.1	1	1216.722	6.155	6.886	28.572	2.11e-07 ***

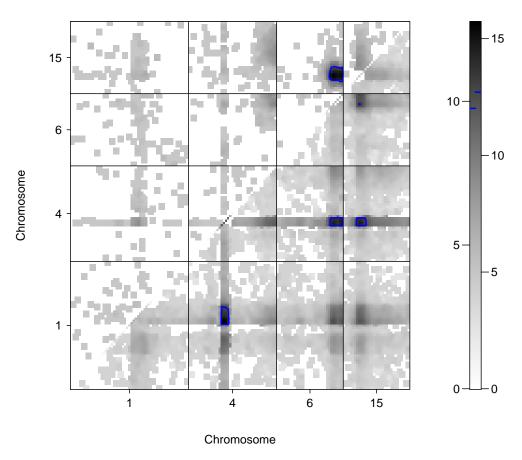
The final model may be more complicated than a model found 'by hand' using standard R/qtl tools. Hopefully that model is a subset of the automatically found model.

6 2-D Epistasis Plots

Should there be any evidence for epistasis that is confirmed by ANOVA, it can be useful to view 2-D plots similar to scantwo, but now using the marginal 2logBF. Here is the reduced, final genetic architecture:

```
> cross.arch <- cross.step$arch
> cross.arch
main QTL loci:
chr 1.0 1.00 4.00 4.0 6.00 15.0 15.00
pos 43.7 73.44 29.87 74.3 61.86 19.1 35.55
Epistatic pairs by qtl, chr, pos:
  q1 q2 chra chrb posa posb
                4 73.44 29.87
           1
  5 7
           4
                6 74.30 61.86
  5 9
           4
               15 74.30 35.55
  7 8
           6
               15 61.86 19.10
Epistatic chromosomes by connected sets:
1,4,6,15
Here are the plots by clique (if any).
> for(i in names(cross.arch$chr.by.set))
    plot(two, chr = cross.arch$chr.by.set[[i]], smooth = 3,
      col = "gray", contour = 3)
```

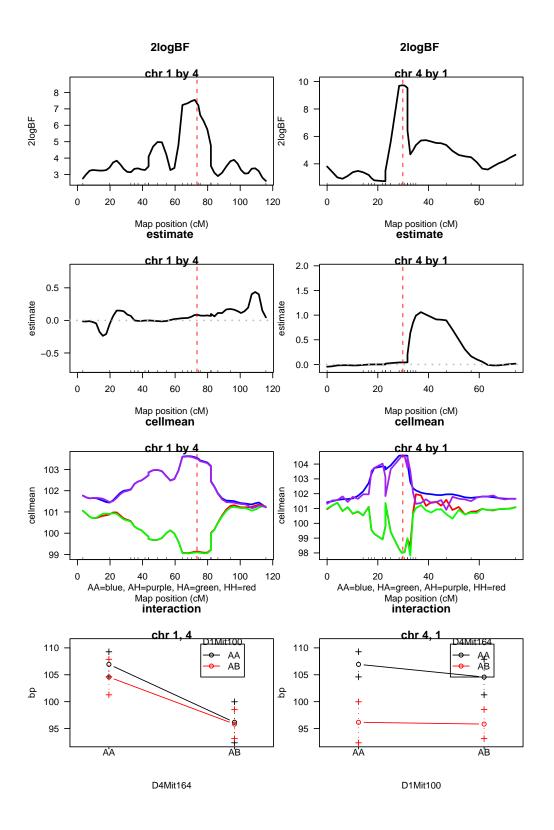


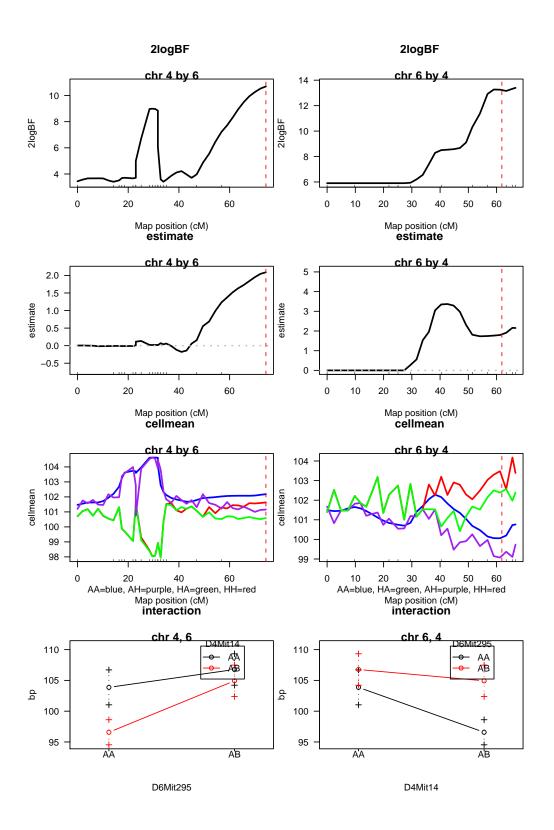


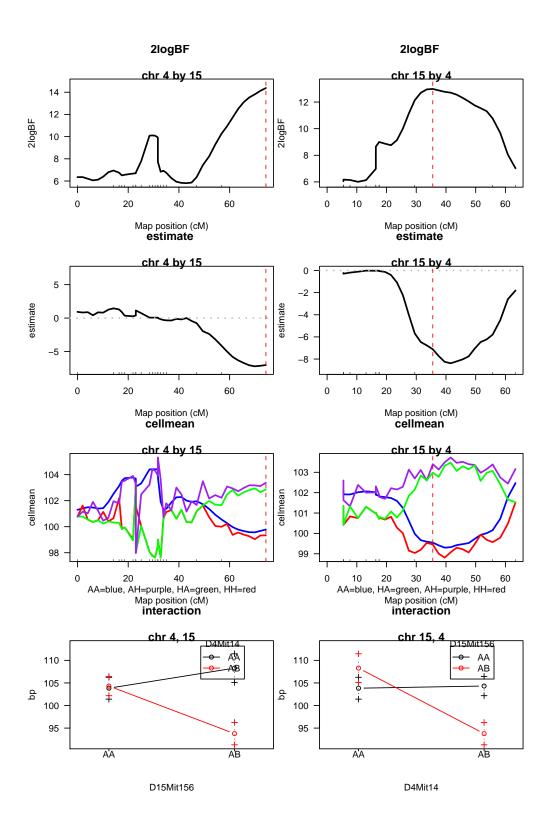
7 1-D Epistasis Slices

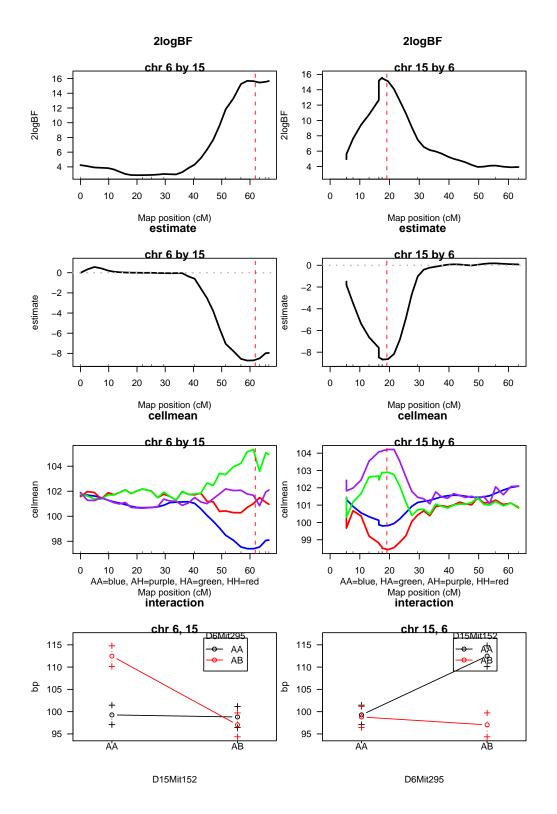
We then examine 1-D slices through the 2-D surface for epistatic pairs in the reduced model, to focus on epistasis for those identified pairs. We show 1-D slices of the LOD and the epistatic effects. In addition, we show interaction plots at the nearest markers.

```
> if(!is.null(cross.arch$pair.by.chr)) {
+ for(i in seq(nrow(cross.arch$pair.by.chr$chr))) {
+ chri <- cross.arch$pair.by.chr$chr[i,]
+ posi <- cross.arch$pair.by.chr$pos[i,]
+ plot(qb.slicetwo(cross.qb, chri, posi, scan.type), byrow = FALSE)
+ }
+}</pre>
```









8 User Customized Section

We know from previous work that there are main QTLs on chromosomes 1 and 4, and epistatic pairs involving 6 and 15, and 7 and 15. Here we pick the nested model that contains these QTL.

Now we do a formal comparison of this reduced model with the fuller model we automatically uncovered. It appears that the fuller model is a much better fit.

```
> anova(cross.step, cross.step2)
```

9 Cleaning Up

That completes the template. Now the penultimate task is to remove the objects created by R/qtlbim, if this is desired by the user.

Finally, externally rename file hyperslide.tex to bp.tex and run pdflatex twice on it. Use a free Acrobat reader to view.

```
> file.rename("hyperslide.tex", "bp.tex")
> invisible(system("pdflatex bp.tex",intern=TRUE))
> invisible(system("pdflatex bp.tex",intern=TRUE))
```