



## GISTIC Documentation

**Description:** Genomic Identification of Significant Targets in Cancer  
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### Summary:

The GISTIC module identifies regions of the genome that are significantly amplified or deleted across a set of samples. Each aberration is assigned a G-score that considers the amplitude of the aberration as well as the frequency of its occurrence across samples. False Discovery Rate q-values are then calculated for the aberrant regions, and regions with q-values below a user-defined threshold are considered significant. For each significant region, a “peak region” is identified, which is the part of the aberrant region with greatest amplitude and frequency of alteration. In addition, a “wide peak” is determined using a leave-one-out algorithm to allow for errors in the boundaries in a single sample. The “wide peak” boundaries are more robust for identifying the most likely gene targets in the region. Each significantly aberrant region is also tested to determine whether it results primarily from broad events (longer than half a chromosome arm), focal events, or significant levels of both. The GISTIC module reports the genomic locations and calculated q-values for the aberrant regions. It identifies the samples that exhibit each significant amplification or deletion, and it lists genes found in each “wide peak” region.

### References:

- Beroukhim R, Getz G, et al. (2007). “Assessing the significance of chromosomal aberrations in cancer: Methodology and application to glioma.” *Proc Natl Acad Sci*, 104:20007-20012.

### Input Parameters

Name	Description
refgene file	The cytoband file to use in the analysis. Allowed values: {Human Hg18, Human Hg17, Human Hg16}. (Default=Human Hg16)
amplifications threshold	Threshold for copy number amplifications. Regions with a log2 ratio above this value are considered amplified. (Default=0.1)
deletions	Threshold for copy number deletions. Regions with a log2

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threshold	ratio below the negative of this value are considered deletions. (Default=0.1)
join segment size	Smallest number of markers to allow in segments from the segmented data. Segments that contain fewer than this number of markers are joined to the neighboring segment that is closest in copy number. (Default=4)
qv thresh	Threshold for q-values. Regions with q-values below this number are considered significant. (Default=0.25)
extension	Extension to append to all output files.
remove x	Flag indicating whether to remove data from the X-chromosome before analysis. Allowed values= {1,0}. (Default=1(yes))
seg file	The segmentation file contains the segmented data for all the samples identified by GLAD, CBS, or some other segmentation algorithm. (See GLAD file format in the Genepattern file formats documentation.) It is a six column, tab-delimited file with an optional first line identifying the columns. Positions are in base pair units.
markers file	The markers file identifies the marker names and positions of the markers in the original dataset (before segmentation). It is a three column, tab-delimited file with an optional header. If not already, markers are sorted by genomic position.
array list file	The array list file is an optional file identifying the subset of samples to be used in the analysis. It is a one column file with an optional header. The sample identifiers listed in the array list file must match the sample names given in the segmentation file.
cnv file	There are two options for the cnv file. The first option allows CNVs to be identified by marker name. The second option allows the CNVs to be identified by genomic location.

## Input Files

### 1. Segmentation File

### REQUIRED

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The segmentation file contains the segmented data for all the samples identified by GLAD, CBS, or some other segmentation algorithm. (See GLAD file format in the Genepattern file formats documentation.) It is a six column, tab-delimited file with an optional first line identifying the columns. Positions are in base pair units.

The column headers are:

- (1) *Sample* (sample name)
- (2) *Chromosome* (chromosome number)
- (3) *Start Position* (segment start position, in bases)
- (4) *End Position* (segment end position, in bases)
- (5) *Num markers* (number of markers in segment)
- (6) *Seg.CN* ( $\log_2()$  -1 of copy number)

[Example Segmentation File](#)

## 2. Markers File

### REQUIRED

The markers file identifies the marker names and positions of the markers in the original dataset (before segmentation). It is a three column, tab-delimited file with an optional header. The column headers are:

- (1) *Marker Name*
- (2) *Chromosome*
- (3) *Marker Position* (in bases)

[Example Markers File](#)

## 3. Array List File

### OPTIONAL

The array list file is an optional file identifying the subset of samples to be used in the analysis. It is a one column file with an optional header (*array*). The sample identifiers listed in the array list file must match the sample names given in the segmentation file.

[Example Array List File](#)

## 4. CNV File

### OPTIONAL

There are two options for the cnv file. The first option allows CNVs to be identified by marker name. The second option allows the CNVs to be identified by genomic location.

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Option #1: A two column, tab-delimited file with an optional header row. The marker names given in this file must match the marker names given in the markers\_file. The CNV identifiers are for user use and can be arbitrary. The column headers are:

- (1) *Marker Name*
- (2) *CNV Identifier*

Option #2: A 6 column, tab-delimited file with an optional header row. The 'CNV Identifier' is for user use and can be arbitrary. 'Narrow Region Start' and 'Narrow Region End' are also not used. The column headers are:

- (1) *CNV Identifier*
- (2) *Chromosome*
- (3) *Narrow Region Start*
- (4) *Narrow Region End*
- (5) *Wide Region Start*
- (6) *Wide Region End*

[Example CNV File](#)

## Output Files

### 1. All Lesions File (all\_lesions\_file.txt)

The all lesions file summarizes the results from the GISTIC run. It contains data about the significant regions of amplification and deletion as well as which samples are amplified or deleted in each of these regions. The identified regions are listed down the first column, and the samples are listed across the first row, starting in column 10.

#### *Region Data*

Columns 1-9 present the data about the significant regions as follows:

- (1) *Unique Name:* A name assigned to identify the region
- (2) *Descriptor:* The genomic descriptor of that region.
- (3) *Wide Peak Limits:* The "wide peak" boundaries most likely to contain the targeted genes. These are listed in genomic coordinates and marker (or probe) indices.
- (4) *Peak Limits:* The boundaries of the region of maximal amplification or deletion.
- (5) *Region Limits:* The boundaries of the entire significant region of amplification or deletion.
- (6) *q-values:* The q-value of the peak region.

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- (7) *Residual q-values*: The q-value of the peak region after removing (“peeling off”) amplifications or deletions that overlap other, more significant peak regions in the same chromosome.
- (8) *Broad or Focal*: Identifies whether the region reaches significance due primarily to broad events (called “broad”), focal events (called “focal”), or independently significant broad and focal events (called “both”).
- (9) *Amplitude Threshold*: Key giving the meaning of values in the subsequent columns associated with each sample.

## *Sample Data*

Each of the analyzed samples is represented in one of the columns following the lesion data (columns 10 through end). The data contained in these columns varies slightly by section of the file.

The first section can be identified by the key given in column 9 – it starts in row 2 and continues until the row that reads “Actual Log Value.” This section contains summarized data for each sample. A ‘0’ indicates that the copy number of the sample was not amplified or deleted beyond the threshold amount in that peak region. A ‘1’ indicates that the sample had low-level copy number aberrations (exceeding the low threshold indicated in column 9), and a ‘2’ indicates that the sample had high-level copy number aberrations (exceeding the high threshold indicated in column 9).

The second section can be identified the rows in which column 9 reads “Actual Log2 Ratio.” The second section exactly reproduces the first section, except that here the exact log2 ratios are provided rather than zeroes, ones, and twos.

The final section is similar to the first section, except that here only broad events are included. A 1 in the samples columns (columns 10+) indicates that the median copy number of the sample across the entire significant region exceeded the threshold given in column 9. That is, it indicates whether the sample had a geographically extended event, rather than a focal amplification or deletion covering little more than the peak region.

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## Lesion Data

## Sample Data

Section 1

	Unique Na	Descriptor	Wide Peak Limits	Peak Limits	Region Lin	q values	Residual q	Broad or F	Amplitude Thre	AA_1	AA_2	AA_4	AA_5	AA_6	AA_7	AA_8
2	Amplificati	1q32.1	chr1:201017471-20	chr1:201512199-1	chr1:20082	6.07E-08	6.07E-08	focal	0: t<0.1; 1: 0.1	0	0	0	0	0	0	0
3	Amplificati	2p24.3	chr2:15719258-167	chr2:15830675-11	chr2:15830	0.23163	0.23163	focal	0: t<0.1; 1: 0.1	0	1	0	0	0	0	0
4	Amplificati	3q26.33	chr3:177090593-18	chr3:181261928-1	chr3:17709	0.043887	0.043887	focal	0: t<0.1; 1: 0.1	0	0	1	0	0	0	0
5	Amplificati	4q12	chr4:54505358-552	chr4:54603039-5	chr4:48833	3.74E-14	3.74E-14	focal	0: t<0.1; 1: 0.1	0	0	1	0	1	0	0
6	Amplificati	6p21.1	chr6:42094850-432	chr6:42664817-4	chr6:42664	0.13151	0.13151	focal	0: t<0.1; 1: 0.1	0	0	0	0	0	0	0
7	Amplificati	7p11.2	chr7:54640152-547	chr7:54709753-5	chr7:1-158	2.61E-79	2.61E-79	both	0: t<0.1; 1: 0.1	0	0	1	0	0	2	0
8	Amplificati	7q31.2	chr7:115842622-111	chr7:116102495-1	chr7:1-158	4.13E-24	9.48E-06	both	0: t<0.1; 1: 0.1	0	0	1	0	0	1	1
9	Amplificati	8q24.12	chr8:121983096-12	chr8:121997366-1	chr8:12198	0.048902	0.048902	broad	0: t<0.1; 1: 0.1	0	0	1	0	1	0	1
10	Deletion P	1p36.31	chr1:4257376-6053	chr1:5404535-60	chr1:1-240	8.69E-06	8.69E-06	focal	0: t>0.1; 1: 0.1	0	0	0	0	0	0	0
11	Deletion P	4q34.3	chr4:183322597-18	chr4:183555243-1	chr4:18355	0.21835	0.21835	focal	0: t>0.1; 1: 0.1	0	0	1	1	0	0	0
12	Deletion P	6q23.2	chr6:132978919-14	chr6:132978919-1	chr6:79415	0.000189	0.000189	broad	0: t>0.1; 1: 0.1	2	0	0	0	0	0	0
13	Amplificati	1q32.1	chr1:201017471-20	chr1:201512199-1	chr1:20082	6.07E-08	6.07E-08	focal	Actual Log2 R <sub>a</sub>	0.054818	0.042652	-0.29535	-0.00881	0	0.00089	0.005499
14	Amplificati	2p24.3	chr2:15719258-167	chr2:15830675-11	chr2:15830	0.23163	0.23163	focal	Actual Log2 R <sub>a</sub>	-0.00296	0.12565	-0.01509	-0.00581	0.03833	-0.00072	0.006214
15	Amplificati	3q26.33	chr3:177090593-18	chr3:181261928-1	chr3:17709	0.043887	0.043887	focal	Actual Log2 R <sub>a</sub>	-0.10861	-0.12928	0.17201	0.009299	0.013925	0.00367	-0.01703
16	Amplificati	4q12	chr4:54505358-552	chr4:54603039-5	chr4:48833	3.74E-14	3.74E-14	focal	Actual Log2 R <sub>a</sub>	0	0.067307	0.45864	-0.01232	0.25929	-0.02441	0
17	Amplificati	6p21.1	chr6:42094850-432	chr6:42664817-4	chr6:42664	0.13151	0.13151	focal	Actual Log2 R <sub>a</sub>	-0.03209	-0.01512	0.071373	-0.02192	0.025052	0.002768	0.002216
18	Amplificati	7p11.2	chr7:54640152-547	chr7:54709753-5	chr7:1-158	2.61E-79	2.61E-79	both	Actual Log2 R <sub>a</sub>	0.03151	0.079714	0.22638	-0.02749	0.014743	2.4949	-0.02561
19	Amplificati	7q31.2	chr7:115842622-111	chr7:116102495-1	chr7:1-158	4.13E-24	9.48E-06	both	Actual Log2 R <sub>a</sub>	0.03151	0.079714	0.22638	0.000868	0.014743	0.28996	0.38366
20	Amplificati	8q24.12	chr8:121983096-12	chr8:121997366-1	chr8:12198	0.048902	0.048902	broad	Actual Log2 R <sub>a</sub>	0.010292	-0.07417	0.11934	0.033819	0.24449	0.014686	0.39299
21	Deletion P	1p36.31	chr1:4257376-6053	chr1:5404535-60	chr1:1-240	8.69E-06	8.69E-06	focal	Actual Log2 R <sub>a</sub>	0.054818	-0.07143	0.1607	-0.07981	0	0.019109	0.005499
22	Deletion P	4q34.3	chr4:183322597-18	chr4:183555243-1	chr4:18355	0.21835	0.21835	focal	Actual Log2 R <sub>a</sub>	0	0.073541	-0.26992	-0.50535	-0.06473	-0.02441	0
23	Deletion P	6q23.2	chr6:132978919-14	chr6:132978919-1	chr6:79415	0.000189	0.000189	broad	Actual Log2 R <sub>a</sub>	-1.3294	0.056499	0.022594	-0.02325	-0.00897	-0.02443	-0.0073
24	Amplificati	7p	Amplitude values reBroad Event Corr	chr7:1-158	2.61E-79	2.61E-79	both	0: t<0.1; 1: t>0	0	0	0	1	0	0	1	0
25	Amplificati	7q	Amplitude values reBroad Event Corr	chr7:1-158	4.13E-24	9.48E-06	both	0: t<0.1; 1: t>0	0	0	1	0	0	1	0	0
26	Amplificati	8q	Amplitude values reBroad Event Corr	chr8:12198	0.048902	0.048902	broad	0: t<0.1; 1: t>0	0	0	1	0	1	0	1	1
27	Deletion P	6q	Amplitude values reBroad Event Corr	chr6:79415	0.000189	0.000189	broad	0: t>0.1; 1: t<0	0	0	0	0	0	0	0	0
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Section 2

Section 3

## 2. Amplification Genes File (Amp\_genes.txt)

The amp genes file contains one column for each amplification identified in the GISTIC analysis. The first four rows are:

- (1) *cytoband*
- (2) *q-value*
- (3) *residual q-value*
- (4) *wide peak boundaries*

These rows identify the lesion in the same way as the all lesions file.

The remaining rows list the genes contained in each wide peak. For peaks that contain no genes, the nearest gene is listed in brackets.

## 3. Deletion Genes File (Del\_genes.txt)

The del genes file contains one column for each deletion identified in the GISTIC analysis. The file format for the del genes file is identical to the format for the amp genes file.

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	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	
1	cytoband	1q32.1	2p24.3	3q26.33	4q12	6p21.1	7p11.2	7q31.2	8q24.12	12p13.32	12p12.1	12q14.1	12q14.3	12q15	17q24.1	19p1
2	q value	6.07E-08	0.23163	0.043887	3.74E-14	0.13151	2.61E-79	4.13E-24	0.048902	0.001345	0.010735	0.036891	0.066517	2.57E-05	0.066517	0.066517
3	residual q value	6.07E-08	0.23163	0.043887	3.74E-14	0.13151	2.61E-79	9.48E-06	0.048902	0.001345	0.1772	0.098942	0.23163	2.57E-05	0.066518	0.066518
4	wide peak boundaries	chr1:20101	chr2:15715	chr3:17705	chr4:54505	chr6:42094	chr7:54640	chr7:11584	chr8:12196	chr12:2815	chr12:2404	chr12:5582	chr12:6513	chr12:6737	chr17:5295	chr1
5	genes in wide peak	KISS1	DDX1	ACTL6A	PDGFRA	GUCA1A	[EGFR]	CAPZA2	[SNTB1]	CCND2	BCAT1	CDK4	GRIP1	CPM	CA4	ACP
6		MDM4	MYCN	NDUFB5	CHIC2	GUCA1B		CFTR		FGF6	ITPR2	CYP27B1		MDM2	CD79B	AES
7		PIK3C2B	NAG	PIK3CA	GSH2	MEA1		MET		FOXM1	KRAS	DDIT3		SLC35E3	CLTC	AMH
8		REN	FAM49A	SOX2	LOC40217	PEX6		WNT2		KCNA1	LRMP	B4GALNT1		NUP107	COX11	ASN

## 4. Gistic Scores File (scores.gistic.txt)

The scores file lists the q-values [presented as  $-\log_{10}(q)$ ], G-scores, average amplitudes among aberrant samples, and frequency of aberration, across the genome for both amplifications and deletions. The scores file is viewable with the Genepattern SNPViewer module.

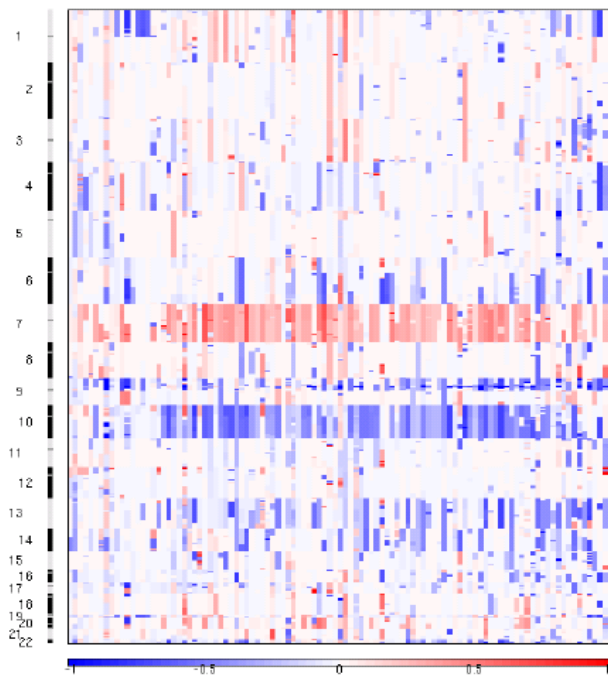
scores.gistic.txt								
	A	B	C	D	E	F	G	H
1	Type	Chromosome	Start	End	$-\log_{10}(q\text{-value})$	G-score	average amplitude	frequency
2	Amp	1	328296	80416443	0	0	0	C
3	Amp	1	80446996	85204863	0	0.003071	0.322479	0.009524
4	Amp	1	85205164	85230288	0.001931	0.013	0.682484	0.01904E
5	Amp	1	85260386	94097075	0	0.003071	0.322479	0.009524
6	Amp	1	94157911	1.09E+08	0	0	0	C
7	Amp	1	1.09E+08	1.09E+08	0	0.008671	0.910463	0.009524
8	Amp	1	1.09E+08	1.43E+08	0	0	0	C
9	Amp	1	1.44E+08	1.6E+08	0	0.006143	0.645044	0.009524

## 5. Segmented Copy Number (segmented\_copy\_number.xx.pdf)

The segmented copy number is a pdf file containing a colormap image of the segmented copy number profiles in the input data.

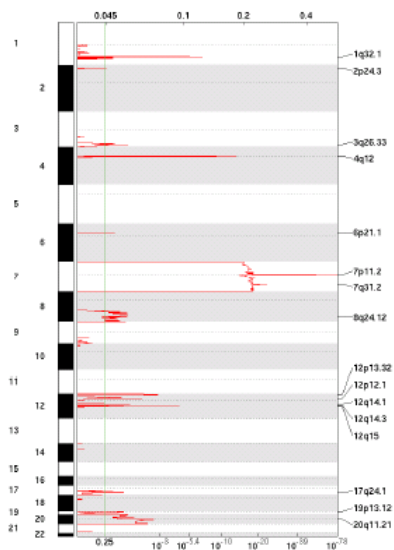


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## 7. Amplification GISTIC plot (amplification.xx.pdf)

The amplification pdf is a plot of the G-scores (top) and q-values (bottom) with respect to amplifications for all markers over the entire region analyzed.



## 8. Deletion GISTIC plot (deletion.xx.pdf)



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The deletion pdf is a plot of the G-scores (top) and q-values (bottom) with respect to deletions for all markers over the entire region analyzed.

