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MP Biomedicals FastDNA™ SPIN Kit

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Protocol status: Working

We use this protocol and it's working

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Abstract

MP Biomedicals FastDNA™
SPIN Kit for feces

Materials

FASTDNA Spinkit for feces



MP Biomedicals FastDNA™ SPIN Kit for feces

- 1 In a 2 mL Lysing Matrix E tube, add 500 mg feces sample, 825 μ L Sodium Phosphate Buffer, and 275 μ L of PLS solution. Shake to mix. Vortex 10-15 seconds.
- 2 Centrifuge samples at 14,000 x g for 5 minutes and decant supernatant.
- 3 Add 978 μ L Sodium Phosphate Buffer and 122 μ L MT Buffer. Shake vigorously or vortex briefly to mix.
- 4 Homogenize samples in the FastPrep 24 instrument at setting 6.0m/s for 40 seconds.
- 5 Centrifuge samples at 14,000 x g for **15 minutes**. NOTE: Extending centrifugation to 15 minutes (did 15 instead of 5) can enhance elimination of excessive debris from large samples, or from cells with complex walls.
- 6 Transfer the supernatant to a clean 2.0 mL centrifuge tube.
- 7 Add 250 μ L of PPS solution, shake vigorously to mix, and incubate at 4°C for 10 minutes. Do not vortex! Centrifuge samples at 14,000 x g for 2 minutes.
- 8 While samples are centrifuging, add 1 mL of Binding Matrix Solution to a clean 15 mL conical tube (not supplied).
- 9 Transfer supernatant to the Binding Matrix Solution in the 15 mL conical. Shake gently by hand to mix, then place on a shaker/rocker for 3-5 minutes.
- 10 Centrifuge samples at 14,000 x g for 2 minutes. Decant the supernatant
- 11 Wash the binding mixture pellet by gently resuspending with 1 mL Wash Buffer #1.
- 12 The following step will require two spins. First, transfer approximately 600 μ L of the binding mixture to a SPIN Filter tube and centrifuge at 14,000 x g for 1 minute. Empty the catch tube. Add the remaining



- binding mixture to the SPIN Filter tube and centrifuge as before. Empty the catch tube again.
- 13 Add 500 μ L of prepared Wash Buffer #2 to the SPIN Filter tube and gently resuspend using the force of the liquid from the pipette tip to resuspend the pellet. Do not vortex. NOTE: Ensure that ethanol has been added to the Wash Buffer #2
 - 14 Centrifuge samples at 14,000 x g for 2 minute. Discard the flow-through.
 - 15 Centrifuge the sample again for 2 minutes to extract residual ethanol from the binding matrix and dry the sample.
 - 16 Transfer the SPIN Filter bucket to a clean 1.9 mL Catch Tube. Add 60-100 μ L TES. Flick the tube or stir the matrix with a pipette tip to resuspend the pellet. Do not vortex.
 - 17 Centrifuge samples at 14,000 x g for 2 minutes to elute purified DNA into the clean catch tube. Discard the SPIN filter. Store at -80°C until use.