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# Sephadex/sephacryl purification of AFLP products

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Works for me

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GUIDELINES

It is the best to gather two full 96x plates (or at least one 96x plate).

All the volumes are given for two plates.

Never leave dry wells in a purification plate (membrane deterioration).

#### MATERIALS

NAME	CATALOG #	VENDOR
EDTA		
NaCl	53014	Sigma Aldrich
MultiScreen HTS DV 0.65µm Clear Non-sterile plates	MSDVN6550	Millipore
Sephadex G-50 Superfine	17 0041 01	Ge Healthcare
Sephacryl S-200	17 0584 01]	Ge Healthcare
Twin.Tec PCR Plate 96 semi-skirted, colourless wells, 25 pcs	0030128575	Eppendorf

Tris

**BEFORE STARTING** 

Solutions to prepare:

NaCl 0.5 M:

5.84 g NaCl in 200 ml ddH20

#### EDTA-Na2 100 mM pH 8.0:

Dissolve3.72 g EDTA-Na2 in 90 ml ddH20 Adjust pH to 8.0 Fill up with ddH20 to 100 ml

## TRIS-HCI 1 M pH 7.0:

Dissolve 12.114 g Tris in 80 ml dH20. Adjust pH to 7.0 with the appropriate volume of concentrated HCl. Bring final volume to 100 ml with deionized water. Stock at  $4^{\circ}$ C

#### 5% Sephadex G50:

2 ml NaCl 0.5 M
2 ml EDTA-Na2 100 mM pH 8.0
2 ml Tris-HCl 1 M pH 7.0
Add ddH20 up to 200 ml
Add 10 g Sephadex G 50 – mix [attention: it will never become clear]
Add 100 µl Triton X-100 (final conc. 0.05%; attention: viscous solution – cut tip end and pipet slowly!)
Stock at 4°C!
This amount serves ca. 5 purification cycles of two 96x plates each.

### Preparation of 96x plates

- 1 Put the Sephacryl bottle on the shaker for 5-10 minutes, adjust speed to 4-5 to make it liquid. For the last minute add also the Sephadex mixture bottle and decrease the speed to 3-4.
- Prepare a 1:1 (v/v) solution of pre-prepared 5% Sephadex G50 and Sephacryl S200 (35 ml of each) and mix well.
- Pipet 300 μl of the mixture to each well of the Millipore purification plate.NOTE: Use the Matrix electronic pipet >> Fill at 1250 μl, Disp at 300 μl).
- 4 Place the Millipore plates on trash plates.
- 5 Centrifugate 1 min at 2800 rpm. Purification of AFLP products

## Purification of AFLP products

- 6 Match and place the prepared purification plates (Millipore plates with fresh Sephadex/Sephacryl layer prepared as described above) on new Eppendorf plates.
  - describe the contents of the Eppendorf plate beforehand.
  - double-check carefully the orientation of the two plates!
  - fix the two plates additionally with small pieces of tape on both sides.
- Pipet the whole volume of the selective PCR product onto the centre of Sephadex/Sephacryl membranes without touching the membranes (using multichannel pipette).

 $\textbf{Citation:} \ \ \textbf{Michal Ronikier, Tomasz Suchan (07/07/2020).} \ \ \textbf{Sephadex/sephacryl purification of AFLP products.} \ \ \underline{\textbf{https://dx.doi.org/10.17504/protocols.io.rq8d5zw}}$ 

- 8 Centrifugate 2 min at 2800 rpm.
- 9 Remove the Millipore plate and use the filtrated PCR product in the Eppendorf plate for subsequent dilution of the selective PCR product.
- Prepare the mix of 10  $\mu$ l of Hi-Di formamide and 0.1  $\mu$ l of Gene Scan-500 ROX size standard per sample and pipet 1-1.5  $\mu$ l of the purified and diluted PCR product.



Millipore plates can be reused up to three times. Leave the Sephadex resin to dry in ambient temperature – it will contract and become easy to remove. Then, plates can be loaded with a new Sephadex/Sephacryl mixture.