



VERSION 4

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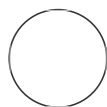
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Total Starch Enzymatic Digestion V.4

Lynn Doran¹, Amanda P. De Souza¹

¹Realizing Increased Photosynthetic Efficiency (RIPE)

UIUC Long Lab



Lynn Doran

Realizing Increased Photosynthetic Efficiency (RIPE)





ABSTRACT

Enzymatic digestion of total soluble starch to glucose in plant tissue extracts for preparation for quantification via the GOD-POD Method (NZYtech).

MATERIALS





Reagents

- [α-amylase \(Bacillus licheniformis\) enzyme, 3000 U/mL](#)
- [Amyloglucosidase \(Aspergillus niger\) enzyme, 3260 U/mL](#)
- MOPS Buffer 10mM, pH 6.5

 2.09 g of MOPS per  1 L of water ( 0.4598 g for  220 mL water), pH adjusted to 6.5 using NaOH.

Store refrigerated for up to 2 months.

- Acetate Buffer 100mM , pH 4.5

 6 mL of acetic acid per  1 L of water ( 1.5 mL per  250 mL water), pH adjusted to 6.5 using NaOH.

Store refrigerated for up to 2 months.

- Ice

Materials

- Pipette tips

Equipment

- Graduated cylinder
- Water bath
- Floating tube holder
- Single channel pipette
- Ice bucket

Protocol status: Working
We use this protocol and it's working

BEFORE START INSTRUCTIONS

Extract and dry total starch pellet from plant tissue per [Extraction of Non-Structural Carbohydrates \(Total Soluble Sugars + Starch\) in Plant Tissues](#).

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- 1
 - Prepare fresh daily 120 U/mL α -amylase (*Bacillus licheniformis*) in MOPS buffer. 1 mL per sample will be needed. Initial concentration of α -amylase is 3000 U/mL. Use $C_1V_1 = C_2V_2$ to calculate the volume of α -amylase and MOPS buffer to use.

Note


α -amylase comes at 3000 U/mL concentration in a 20 mL bottle.

To prepare 250 mL of 120 U/mL α -amylase in MOPS buffer, dilute 10 mL α -amylase (3000 U/mL) into 240 mL MOPS buffer.

- 2
 - Prepare fresh daily 30 U/mL amyloglucosidase (*Aspergillus niger*) in acetate buffer. 1 mL per sample will be needed. Initial concentration of amyloglucosidase is 3260 U/mL. Use $C_1V_1 = C_2V_2$ to calculate the volume of amyloglucosidase and acetate buffer to use.

Note

To prepare 250 mL of 30 U/mL amyloglucosidase in acetate buffer, dilute 2.3 mL amyloglucosidase (3260 U/mL) into 247.7 mL acetate buffer.


- 3 Heat the water bath to  75 °C .

- 4 Add  500 μ L of α -amylase in MOPS buffer (120 U/mL) to each sample tube.

5 Vortex to suspend all solids. Flicking the tube may help dislodge the dried starch pellet.

6 Incubate for  00:30:00 at  75 °C in the water bath.


30m

7 Add another  500 μ L of α -amylase in MOPS buffer (120 U/mL) to each sample tube.

8 Vortex to suspend all solids.


9 Incubate for  00:30:00 at  75 °C in the water bath.

30m






10 Cool down the water bath to  50 °C, tubes can be stored at room temp on the counter while water bath cools.

Note

Leaving the lid off, especially if it is a shaking water bath, will help cool. For faster cooling, remove some water from the water bath and refill with cool distilled water. Do not use tap water as mineral buildup or heated chlorine could damage the water bath.

11 Add  500 μ L amyloglucosidase in acetate buffer (30 U/mL) to each sample tube.

12 Vortex to suspend all solids.

- 13** Incubate for  00:30:00 at  50 °C in the water bath. 30m
- 14** Add another  500 µL amyloglucosidase in acetate buffer (30 U/mL) to each sample tube.
- 15** Vortex to suspend all solids.
- 16** Incubate for  00:30:00 at  50 °C in the water bath. 30m
- 17** Place the tubes in ice to stop the reaction, until cool to the touch.
- 18** Proceed to total starch (as glucose) quantification by NZYtech GOD-POD method or store the samples at -20°C up to one month.