



Aug 19, 2021

Recombinant a-synuclein pre-formed fibril generation

In 1 collection

Alain Ndayisaba¹

¹Khurana Lab, Harvard Medical School, Brigham and Women's Hosptial



hendersa

ABSTRACT

This protocol details the generation of a-synuclein pre-formed fibrils.

ATTACHMENTS dh34biqa7.pdf

DOI

dx.doi.org/10.17504/protocols.io.bu53ny8n

PROTOCOL CITATION

Alain Ndayisaba 2021. Recombinant a-synuclein pre-formed fibril generation. **protocols.io** https://dx.doi.org/10.17504/protocols.io.bu53ny8n

COLLECTIONS (i)

Expression and purification of untagged asynuclein and recombinant a-synuclein pre-formed fibril generation

KEYWORDS

Recombinant a-synuclein pre-formed fibrils, Lyophilized monomeric synuclein

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

May 19, 2021

LAST MODIFIED

Aug 19, 2021

OWNERSHIP HISTORY

PROTOCOL INTEGER ID

50075

Citation: Alain Ndayisaba (08/19/2021). Recombinant a-synuclein pre-formed fibril generation. https://dx.doi.org/10.17504/protocols.io.bu53ny8n

Part of collection

Expression and purification of untagged asynuclein and recombinant a-synuclein pre-formed fibril generation

MATERIALS TEXT

Materials:

- Lyophilized monomeric synuclein
 Alternatively: aliquots of untagged alpha-synuclein in PBS, [M]0.5 mg/ml or [M]5 mg/ml
- Sterile dPBS
- 1.5 mL Protein LoBind tubes (Eppendorf)
- Parafilm

Recombinant a-synuclein pre-formed fibril generation 1w 0d 0h 30m	
1	Reconstitute $\blacksquare 1$ mg of lyophilized monomeric synuclein with $\blacksquare 100~\mu l$ cold sterile DPBS $\& On ice$ (do not pipet, close tube immediately).
2	Transfer tubes on wet ice to § 4 °C cold room and rotate on tube rotator for © 00:10:00 .
3	10m
	Centrifuge for © 00:10:00 at @15000 x g at § 4 °C .
4	Transfer supernatant to clean Protein-LoBind tube.
5	Prepare \blacksquare 5 μ l aliquots of serial dilutions (1:10, 1:25, 1:50, 1:100) and measure concentration with Nanodrop (A280 - MW=14,5 kDa; Extinction coefficient ϵ for human α -syn = 5,960 M ⁻¹ cm ⁻¹).
6	Dilute down to final concentration of [M]5 mg/ml , use $\Box 100~\mu l$ ($\Box 100~\mu l$ - $\Box 500~\mu l$) aliquots for reproducible results.
7	Take 1 μl - 12 μl aliquot of monomer and dilute to [M]1 ul/ml for electron microscopy, flash-freeze in dry ice – ethanol slurry.
8	Seal tubes with parafilm if no tube lock.



Place the tube in an orbital thermomixer with a heated lid for \circlearrowleft 168:00:00 at $\rat{8}$ 37 °C , shaking at $\rat{6}$ 1000 rpm .

NOTE: At the end of the 7 days, the contents of the tube should appear turbid. The thermomixer must have a lid to prevent condensation formation on the tube lids.



Gently flick the tube to resuspend a-syn fibrils.

- a. Fibrils for quality control steps can be stored at § Room temperature © Overnight.
- Prepare aliquots for QC assays: 11
 - 11.1 a. $\square 5 \mu I$ thioflavin T binding assay.
 - 11.2 b. $\mathbf{2}\mathbf{\mu}\mathbf{l}$ for TEM.
- 12 For long-term storage (12-18 months), flash-freeze fibrils in liquid nitrogen and store at 8-80 °C.

1w

10m