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Minimum inhibitory concentration of butanol for *E. coli* KJK01

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This protocol helps you determine the minimum inhibitory concentration (MIC) of butanol for *E. coli* KJK01. However, this technique can be extended to determine the MIC of any metabolite for any strain by making adjustments to the concentration of the metabolite in the starting wells.

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<https://dx.doi.org/10.17504/protocols.io.byz8px9w>*E. coli*, Minimum inhibitory concentration, butanol

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1 Take a 96 wellled plate.

Media condition preparation

- 2 Calculate the amount of butanol that you require in the starting well. Wilbanks 2017 paper suggests that butanol shows strong toxic effects below 10 g/L and entirely inhibits growth at 15 g/L. We should expect our MIC to fall in this range. Hence, we can start with an initial concentration of 60 mg/ml.

Wilbanks B, Trinh CT (2017). Comprehensive characterization of toxicity of fermentative metabolites on microbial growth..

Biotechnology for biofuels.

<https://doi.org/10.1186/s13068-017-0952-4>

2.1 Take **185.2 μ L of LB** and pour it in wells B2 to D2.

- 3 Pour **100 μ L of LB** of LB in all wells from B3 to B11 and the same for rows C and D.

- 4 Take **14.6 μ L of Butanol** and pour it wells B2 to D2 and mix well.

- 5 Take **100 μ L of sample from well B2** and pour it in well B3. Mix well.

- 6 Take **100 μ L of sample from well B3** and pour it in well B4. Mix well. Repeat this for the subsequent wells till you reach well B10.

- 7 Take **100 μ L of sample from well B10** and discard in the well B12.

Do not add the 100 μ L sample into well B11 as it is supposed to serve as our control (i.e. culture without butanol).

8 Repeat steps 5 to 7 for rows C and

Inoculation

- 9 Take  **0.2 µL secondary overnight culture of wild type strain** and inoculate wells B2 to B11.
- 10 Take  **0.2 µL secondary overnight culture of uninduced KJK01 strain** and inoculate wells C2 to C11.
- 11 Take  **0.2 µL secondary overnight culture of induced KJK01 strain** and inoculate wells D2 to D11.
- 12 Fill the border wells with milliQ.

Measurement

- 13 Cover the 96 well plate with foil and place it in the incubator at 37°C.
- 14 Take a reading of the plate using any plate reader every hour and place it back in the incubator. You may run this for 24 hrs.
- 15 Obtain a growth curve for each strain and concentration. Identify the least concentration for which the growth curve shows a sharp drop.