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TaqMan SNP genotyping protocol

PLOS One

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1 Works for me dx.doi.org/10.17504/protocols.io.bpapmidn

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ABSTRACT

The protocol is based on TaqMan SNP genotyping protocol

EXTERNAL LINK

<https://doi.org/10.1371/journal.pone.0243509>

THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Jin J, Robeson H, Fagan P, Orloff MS (2020) Association of *PARP1*-specific polymorphisms and haplotypes with non-small cell lung cancer subtypes. PLoS ONE 15(12): e0243509. doi: [10.1371/journal.pone.0243509](https://doi.org/10.1371/journal.pone.0243509)

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MANUSCRIPT CITATION please remember to cite the following publication along with this protocol

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44079

MATERIALS TEXT

TaqMan genotyping master mix
TaqMan genotyping SNP genotyping assays
Adhesive film

ABSTRACT




The protocol is based on TaqMan SNP genotyping protocol

BEFORE STARTING

- Extract and purify genomic DNA (gDNA)
- Quantify sample gDNA
- Prepare assays, DNA samples, and master mix

Dilute the predesigned and custom TaqMan SNP genotyping assays to 20X working stock solution




PCR reactions

- 1 384-well Fast (5- μ L reaction)
Prepare the reaction mix:
Combine the following components for the number of reactions required, plus 10% overage.
2X TaqMan Master Mix:  **2.5 μ l**
20X Assay Working Stock:  **0.25 μ l**
Total volume per well:  **2.75 μ l**
- 2 Vortex to mix
- 3 Centrifuge to bring the reaction mix to the bottom of the tube and remove the air bubbles
- 4 Prepare the reaction plate:
Dilute each DNA sample, positive and negative controls in Nuclease-free water. The final concentration should be at least 0.2 ng/ μ L
- 5 Add reaction mix to each well of the reaction plate
- 6 Seal the plate with adhesive film, then centrifuge to bring the reaction mix to the bottom of the well and eliminate air bubbles.
- 7 Remove film from the plate, then add the appropriate volume of sample or control to the wells

- 8 Seal the plate with adhesive film, then centrifuge to bring the reaction mix to the bottom of the well and eliminate air bubbles.

Perform PCR

10m

- 9 Place the plate in a Real-Time PCR instrument ABI 7900 HT. Use the thermal cycling conditions specified.
Polymerase activation:
Temp.  **95 °C** , Time 10mins , Cycles HOLD
Denaturation:
Temp.  **95 °C** , Time 15 seconds, Cycles 40
Annealing extension:
Temp.  **60 °C** , Time 1 min, Cycles 40

Post-PCR plate read and analyses

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