

APR 06, 2023

OPEN ACCESS

DOI:

dx.doi.org/10.17504/protocol s.io.36wgqjerovk5/v1

Protocol Citation: Hannah M. Alt, David G. Brandner, Gregg T. Beckham, Kelsey J. Ramirez 2023. Lignin Reductive Catalytic Fractionation (RCF) Monomers Analysis by Gas Chromatography Flame Ionization Detection (GC-FID). protocols.io

https://dx.doi.org/10.17504/protocols.io.36wgqjerovk5/v1

License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working We use this protocol and it's working

Created: Feb 07, 2023

Last Modified: Apr 06, 2023

PROTOCOL integer ID: 76618

Lignin Reductive Catalytic Fractionation (RCF)
Monomers Analysis by Gas Chromatography Flame
Ionization Detection (GC-FID)

Hannah M. Alt¹, David G. Brandner¹, Gregg T. Beckham¹, Kelsey J. Ramirez¹

¹Renewable Resources and Enabling Sciences Center, National Renewable Energy Laboratory

NREL

Tech. support email: stefan.haugen@nrel.gov



Hannah M. Alt

ABSTRACT

A gas chromatography with flame ionization detection (GC-FID) method was developed to quantify reductive catalytic fractionation (RCF) monomers.

GUIDELINES

NOTICE

This work was authored by the National Renewable Energy Laboratory, operated by Alliance for Sustainable Energy, LLC, for the U.S. Department of Energy (DOE) under Contract No. DE-AC36-08G028308. Funding provided by U.S. Department of Energy Office of Energy Efficiency and Renewable Energy Bioenergy Technologies Office. The views expressed herein do not necessarily represent the views of the DOE or the U.S. Government.

MATERIALS

Woody RCF Standards

- Phenol Contributed by users
- **⊠** Guaiacol **Contributed by users**
- **⋈** 4-Methylguaiacol **Contributed by users**
- Syringol Contributed by users
- **⋈** 4-Propylguaiacol **Contributed by users**

Keywords: Reductive Catalytic Fractionation, Gas Chromatography

- **⋈** Isoeugenol **Contributed by users**
- Methylparaben Contributed by users
- **⋈** 4-Ethylsyringol **Contributed by users**

- Propenylsyringol Contributed by users
- **⋈** 4-Propanolsyringol **Contributed by users**

Non-Woody RCF Standards

- Methyl-3-(4-hydroxyphenyl) propionate Contributed by users
- Methyl-3-(4-hydroxy-3-methoxyphenyl) propionate Contributed by users
- Methylcoumarate Contributed by users
- Methylferulate Contributed by users

Internal Standard

X Tri-tert-butylbenzene Contributed by users

Reagents

Methanol P212121 Catalog #PA-33900HPLCCS4L

Equipment

Equipment	
Gas Chromatograph	NAME
8890 GC System	TYPE
Agilent	BRAND
8890 GC System	SKU
https://www.agilent.com/en/product/gas-chromatography/gc- systems/8890-gc-system	LINK

Equipment		
HP5-MS	NAME	
GC Column	TYPE	
Agilent	BRAND	
19091S-433UI	SKU	
https://www.agilent.com/store/productDetail.jsp?catalogId=19091S-433UI LINK		

Equipment		
Inlet Liner		NAME
Agilent		BRAND
5183-4711		SKU
https://www.agilent.com/store/productDetail.jsp?catalogId=518	3-4711	LINK
Inlet liner, split, single taper, glass wool, deactivated	SPECIFI	CATIONS

SAFETY WARNINGS

All chemicals used for this procedure are hazardous. Read the Safety Data Sheet (SDS) for all chemicals and follow all applicable chemical handling and waste disposal procedures. Manufacturer specific SDS information can be found by following the CAS numbers of compounds in 'Materials' list.

BEFORE START INSTRUCTIONS

All solvents, analytes, and chemicals used in this protocol are listed in the 'Materials' section. They are excluded from in-line referencing to keep steps clear and concise.

Internal Standard Preparation

- This analysis uses 1,3,5-tri-tert-butylbenzene (TTB) as an internal standard. Prepare adequate volume to allow for addition of 1μ L of TTB per 100 μ L of sample or standard volume.
- 2 Create a 10,000 μ g/mL internal standard working solution by weight of TTB using methanol as the diluent.

Preparation of Standards

By weight, create individual 20,000 μ g/mL stock solutions of all monomers (listed in Materials section) and use methanol as the diluent.

Note

If compounds have not previously been analyzed on the column specified in this method, it is necessary to analyze each monomer separately (at a concentration of approximately 100 $\mu g/mL$) to determine retention time for each before analyzing a mixture containing all compounds.

- 4 Combine the stock solutions to create a 1000 μ g/mL mixed standard working solution in methanol.
- 4.1 For example, to prepare a 10 mL mixed standard working solution of woody RCF analytes (distinguished in Materials section), add 500 μ L of each of the 20,000 μ g/mL stock solutions (14 analytes) and add 3000 μ L methanol.
- Using the mixed standard working solution at 1000 μ g/mL, create a calibration curve from 10 μ g/mL to 1000 μ g/mL with a minimum of five calibration points using methanol as the diluent.

Add 1 μ L of 10,000 μ g/mL TTB internal standard working solution per every 100 μ L of calibration standard.

Example: $10 \mu L$ into $1000 \mu L$ or $5 \mu L$ into $500 \mu L$

6.1

Calibration	Concentration	Volume of Mixed Standard	Volume of	Volume of Internal
Level	(µg/mL)	Working Solution (μL)	Methanol (μL)	Standard (µL)
9	1000	1000	0	10
8	750	750	250	10
7	500	500	500	10
6	250	250	750	10
5	100	100	900	10
4	75	75	925	10
3	50	50	950	10
2	25	25	975	10
1	10	10	990	10

Example Calibration

Sample Preparation

7 Oil Samples:

- 1. Weigh approximately 5-10 mg of oil into a GC vial and record weight.
- 2. Add a known volume of methanol to each vial containing sample (for example 1 mL).
- 3. Add 1 μ L of 10,000 μ g/mL TTB for every 100 μ L of methanol added to the sample vial. Example: 10 μ L into 1000 μ L or 5 μ L into 500 μ L

Liquid Samples:

- 1. Aliquot a known volume of sample into a GC vial (for example 1 mL)
- 2. Add 1 μ L of 10,000 μ g/mL TTB for every 100 μ L of methanol added to the sample vial. Example: 10 μ L into 1000 μ L or 5 μ L into 500 μ L

GC-FID Analysis

Analyze samples using an 8890 Agilent Gas Chromatograph (GC) or equivalent equipped with a flame ionization detector (FID) per the method parameters below:

8.1 Method Parameters

Split/Splitless Inlet Parameters:

Inlet Temperature: 280 °C Injection Volume: 1 µL

Split Ratio: 2:1

Inlet Liner: split, single taper, glass wool, deactivated (see Materials)

Syringe: P/N 5181-8809 Wash Solvent: Methanol

Column:

HP5-MS (see Materials) Carrier Gas: Helium

Flow Rate: 1 mL/min (constant flow)

Oven Parameters:

Maximum Oven Temperature: 280 °C

	Rate	Value	Hold Time	Run Time
2	°C/min	°C	min	min
Initial		70	2	2
Ramp	10	280	2	25

Detector Parameters:

Detector: FID

Detector Temperature: 300 °C

Air Flow: 400 mL/min H2 Fuel Flow: 40 mL/min Makeup Flow: 10 mL/min

Analytical Quality Control

9 Several strategies are utilized when performing this analysis to ensure instrument stability and reproducibility.

9.1 Calibration Curves

All compounds must have a correlation coefficient (r^2) of 0.995 or greater using a linear calibration fit.

9.2 Calibration Verification Standards (CVS)

A calibration verification standard (CVS) is a standard from the calibration curve that is reanalyzed every 20 or fewer samples to ensure instrument drift remains within the determined acceptance criteria. Acceptable CVS recoveries for this analysis are within 15% of the expected amount. Acceptance criteria may differ between instruments and should be determined experimentally.