



Aug 17, 2020

An easy chromatographic method for purification of Immunoglobulin Y (IgY) using HiTrap™ Columns.

Angel A Justiz-Vaillant¹¹University of the West Indies St. Augustine**1** Works for me dx.doi.org/10.17504/protocols.io.bju7knzn[University of the West Indies](#) angel.vaillant@sta.uwi.eduAngel Justiz-Vaillant
University of the West Indies St. Augustine

DOI

dx.doi.org/10.17504/protocols.io.bju7knzn

PROTOCOL CITATION

Angel A Justiz-Vaillant 2020. An easy chromatographic method for purification of Immunoglobulin Y (IgY) using HiTrap™ Columns.. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.bju7knzn>

LICENSE

This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Aug 17, 2020

LAST MODIFIED

Aug 17, 2020

PROTOCOL INTEGER ID

40575

- 1 Fill the syringe or pump tubing with de-ionized water. Remove the stopper and connect HiTrap™ column to syringe (use the connector supplied).
- 2 Snap off tab on the column outlet.
- 3 Wash out the ethanol with 26 ml of de-ionized water.
- 4 Equilibrate column with 26 ml of binding buffer. The recommended flow rate is 5ml/min.
- 5 Apply the IgY sample using a syringe fitted to Luer connector or by pumping it onto the column.

- 6 For better results, use a flow rate of 0.5 to 5.1 ml/min during sample application.
- 7 Wash with at least 51 ml of binding buffer or no material remains in the effluent.
- 8 Maintain a flow rate of 5 to 11 ml/min for washing.
- 9 Elute with 51 ml of elution buffer using a one-step or using a linear gradient though larger volumes are often required to break the interaction.
- 10 After elution, regenerate the column by washing with 36 ml of wash buffer.
- 11 Re-equilibrate the column with 26 ml of binding buffer.
- 12 The column is now prepared for a new purification.