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◆ Total Lipid Extraction from Baker's yeast (Saccharomyces cerevisiae) V.2

Israel Olayide¹, Monica Rieth²

¹St. Louis University Dept. of Chemistry; ²Southern Illinois University-Edwardsville

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This protocol outlines a method for extracting total lipids from Baker's yeast, *Saccharomyces cerevisiae.* It has been adapted from Roy et al., *J. Lipid Res.* **2018.** doi: 10.1194/jlr.M088559.

Extraction of lipids from yeast_lab website_v.3.pdf

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All culture ODs should be normalized for comparative analyses prior to the extraction. Weighed Eppendorf tubes to be used before getting started. Extracted lipids can be stored at -20 degC



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chloroform and methanol should be used in a chemical hood

| Day 1-3 | 3d |
|---------|----|

- 1 Sterilize the inoculating loop by dipping it in the ethanol and heating it for 30 second in the flame
 - 1.1 Allow loop to cool the loop at least for 10 seconds
- 2 Streak plate from prepared glycerol stock (-80 $^{\circ}\text{C})$ and spread on YPD agar plate
- 3 Allow plate to incubate for 2-3 days at 25-30 °C until colonies are 1-2mm in diameter

Day 4 1d

- 4 Inoculate 5 mL YPD with yeast colony from plate
- 5 Grow overnight at 30°C with shaking at 250 rpm

- 6 Check OD₆₀₀ at UV-Vis spectrophotometer
- 7 Harvest the whole cells when OD₆₀₀ is 1.5
 Centrifuge at 1000 x g for 2 minutes or until clear to pellet the yeast
- 8 Pour off the supernatant without disturbing cell pellet

Wash the cells three times by resuspending pellets in 1ml PBS, centrifuge for at 500 x g for 3-5

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| 9 | mins. |
|----|--|
| 10 | After washing three times with PBS, resuspend in 1ml PBS and transfer this mixture to a 15 ml conical centrifuge tube. |
| 11 | Add 3.75 mL of chloroform/methanol (1:2) to the 1 mL of resuspended cells Add 1.25 mL chloroform and 1.25 mL sterile water subsequently. |
| 12 | Vortex vigorously for 5 mins, and centrifuge at 150 x g for 5 mins at room temperature. |
| 13 | After centrifugation, a two-phase system is obtained: aqueous top phase and organic bottom phase which contains the lipids. |
| 14 | Carefully remove the top aqueous layer and the middle insoluble layers (precipitated proteins). |
| 15 | The organic bottom layer is dried using a speed vacuum or dried under a steady stream of nitrogen. |
| 16 | Record the weight of the dried sample by pre-weighing an Eppendorf tube or equivalent before or after drying the lipid. |