

•



Aug 04, 2021

© E. coli CRISPR Barcoding

Leanne Hobbs¹

¹University of Newcastle-upon-Tyne

1	Works for me	≪ €	Share

This protocol is published without a DOI.

Leanne Hobbs

ABSTRACT

This is the protocol for barcoding *E. coli* using CRISPR for CellRepo

EXTERNAL LINK

https://identifiers.org/cellrepo:59

PROTOCOL CITATION

Leanne Hobbs 2021. E. coli CRISPR Barcoding. **protocols.io** https://protocols.io/view/e-coli-crispr-barcoding-bw5jpg4n

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Aug 04, 2021

LAST MODIFIED

Aug 04, 2021

PROTOCOL INTEGER ID

52107

Day 1

Start an overnight culture (37 °C) by inoculating LB medium from a single colony. **© 00:00:00**

Day 2

- 2 Prepare competent cells following your favorite protocol.
- 3 Transform E. coli cells with plasmid pREDCas9 and plate the cells at 30 $^{\circ}$ C in LB supplemented with 50 μ g/mL spectinomycin.

Day 3

4 Start an overnight culture at 30 °C in LB/Spec from a single colony.

Jay 4	
5	Next morning refresh the culture with LB/Spec and grow the cells until OD600 reaches 0.1.
6	Add IPTG to a final concentration of 2 mM and grow the cells to an OD600 = 0.6 (recombination proteins are being expressed at this point).
7	Frost cells on ice for 20 minutes and electrocompetent cells are then prepared by washing bacteria with ice-cold milliQ water after spinning aliquots 10 minutes at 5000 rpm in a 4 °C centrifuge. (Heat shock transformation also could be performed)
8	After two washes, resuspend cells in the residual milliQ water and electroporate with pEC-CRISPR2-BC.
9	Add 950 μ l of fresh LB without antibiotics and resuspend cells. Incubate cells 1 hour at 30 $^{\circ}$ C.
10	Then, spread 100 μ L of bacterial culture on LB plates supplemented with Spec (50 μ g/mL) /Amp (100 μ g/mL).
Day 5	Check colonies for barcode presence by colony-PCR.
12	Inoculate a positive clone in 2 mL of LB/Spec/Ara (30 mM) (sgRNA targeting pUC origin in pEC-CRISPR2-BC is expressed).
13	Grow for 4-6 hours.
14	Plate on LB/Spec.
15	Check some colonies for Ampicillin resistance by restreaking them on LB/Amp.
Day 7	Take a sensitive clone and restreak it on LB plates at 37°C.
Day 8	
17	Check again by colony PCR the presence of the barcode and restreak on LB/Spec and LB/Amp plates to double check

plasmid curing.