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Algae Agar Preparation

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ABSTRACT

This protocol details the making of algae concentrate agar for the growth of bacteria from Douglas Lake.

Protocol for class project in **Microbes in the Wild: Advanced Environmental Microbiology Lab (EEB 447)** course at University of Michigan Biological Station and on campus.

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Protocol status: Working We use this protocol and it's working

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- 1. During the peak of a cyanobacterial bloom, bring a 100µm mesh out into the water.
- 2. Collect as many colonies as possible using the mesh and collect them into three sterile 50 ml falcon tubes.

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- 1. Sterilize a conventional blender with 10% bleach. Rinse with 1-2L MilliQ water
- 2. Pour the contents of the three 50 ml falcon tubes into the blender. Blend on high for 2 minutes.
- 3. Add the contents of the blender into a sterile 1L round glass media bottle.
- 4. Add 15 grams of agar to the media bottle.
- 5. Bring up the mixture to 1L with MilliQ water
- 6. Steam autoclave at 121°C for 30 minutes on Liquid Cycle 2
- 7. Pour agar into sterile petri dishes and allow to set. Store plates at 4°C.