



Feb 04, 2021

# Purification and quantification from PCR amplification protocol

# Rene Flores Clavo<sup>1</sup>, Nataly Ruiz Quinones<sup>1</sup>, Cristian Daniel Asmat Ortega<sup>2</sup>

<sup>1</sup>Universidade Estadual de Campinas; <sup>2</sup>Centro de Investigación en Innovación en Ciencias Activas Multidisciplinarias-CIICAM Rene Flores Clavo: Centro de Investigación e Innovación en Ciencias Activas Multidisciplinarias-CIICAM; Nataly Ruiz Quinones: Centro de Investigación e Innovación en Ciencias Activas Multidisciplinarias-CIICAM

1 Works for me

dx.doi.org/10.17504/protocols.io.brzsm76e

## **RENE FLORES**

Tech. support phone: +55 (19) 991640041 email: renefloresclavo@gmail.com Click here to message tech. support



Rene Flores Clavo

Universidade Estadual de Campinas, Centro de Investigación e...

DOI

dx.doi.org/10.17504/protocols.io.brzsm76e

PROTOCOL CITATION

Rene Flores Clavo, Nataly Ruiz Quinones, Cristian Daniel Asmat Ortega 2021. Purification and quantification from PCR amplification protocol. protocols.io

https://dx.doi.org/10.17504/protocols.io.brzsm76e

## **KEYWORDS**

null, Purification PCR amplification, visualization purification PCR amplification, GFX PCR DNA and Gel Band Purification Kit.

### LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Feb 01, 2021

LAST MODIFIED

Feb 04, 2021

PROTOCOL INTEGER ID

46866

02/04/2021

# GFX<sup>™</sup>PCR DNA and Gel Band Purification Kit

kit can be used to purify DNA and concentration of PCR products or DNA fragments ranging in size from 50 bp to 10 kb.

GFX<sup>™</sup> columns

collection tubes

Polypropylene tubes of 1,5 mL

Polypropylene tubes of 2,0 mL

Absorbent paper

Tips of 10 µL, 200 µL, 1000 µL

Permanent marker for labeling

Gloves

#### Reagents

Capture buffer

Washing buffer

Elution buffers (Tris-HCl and sterile water)

SYBER safe at 1:10000

Loading buffer dye (6X)

DNA Ladder 1 Kb (50 ng)  $0.5 \text{ ng/}\mu\text{L}$ 

 $\lambda$  phage (50 ng/ $\mu$ L)

TBE 1X (89mM Tris-borate, 89mM boric acid, 2mM EDTA)

## **Solutions**

DNA PCR amplification (25µL) Sterile deionized water

#### Other

Micropipette of 10  $\mu$ L, 200  $\mu$ L, 1000  $\mu$ L Analytical balance Freezer microcentrifugate

DISCLAIMER:

DISCLAIMER - FOR INFORMATIONAL PURPOSES ONLY: USE AT YOUR OWN RISK

The protocol content here is for informational purposes only and does not constitute legal, medical, clinical, or safety advice, or otherwise; content added to <u>protocols.io</u> is not peer reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol should not substitute for independent professional judgment, advice, diagnosis, or treatment. Any action you take or refrain from taking using or relying upon the information presented here is strictly at your own risk. You agree that neither the Company nor any of the authors, contributors, administrators, or anyone else associated with <u>protocols.io</u>, can be held responsible for your use of the information contained in or linked to this protocol or any of our Sites/Apps and Services.

# Purification steps

- 1 Prepare collection tubes for each sample with their corresponding columns
- 2 Add 500 μL of capture buffer on each column

Add 25 µL of PCR-amplified DNA

Citation: Rene Flores Clavo, Nataly Ruiz Quinones, Cristian Daniel Asmat Ortega (02/04/2021). Purification and quantification from PCR amplification protocol. <a href="https://dx.doi.org/10.17504/protocols.io.brzsm76e">https://dx.doi.org/10.17504/protocols.io.brzsm76e</a>

3	
4	Slowly mix the solution using a micropipette.
5	Centrifuge for 1 min at 16000 g
6	Keep the DNA in the column and discard the flow through on the tube
7	Add 500 μL of washing buffer on the column
8	Centrifuge for 1 min at 16000 g
9	Incubate for 1 min at room temperature
10	Transfer the column into a new tube
11	Add 10 µL of elution buffer
12	Incubate for 2 min at room temperature
13	Centrifuge for 2 min at 16000 g
14	Repeat step 11
15	Repeat step 12

Repeat step 13

17 Discard the columns and keep the tubes with purified DNA at 4 °C until visualization

# Quantification

- To visualize the amplified products, put 2  $\mu$ L of the PCR product with 2  $\mu$ L of loading buffer dye in agarose gel electrophoresis with 1X TBE stained with SYBER safe at 1:10000.
- 19 To compare, add 1  $\mu$ L of 1kb DNA ladder (final concentration 0,5 ng/ $\mu$ L) and observe a band of 1500 pb approximately.