

### VERSION 1

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# OPEN ACCESS

#### יוסם

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**Protocol status:** Working We use this protocol and it's working

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Single-moclecule Immunofluorescence Tissue Staining Protocol for Oligomer Imaging V.1

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**ABSTRACT** 

This protocol details about immunofluorescence staining for oligomer imaging.

**ATTACHMENTS** 

kb3ib25np.pdf

#### **GUIDELINES**

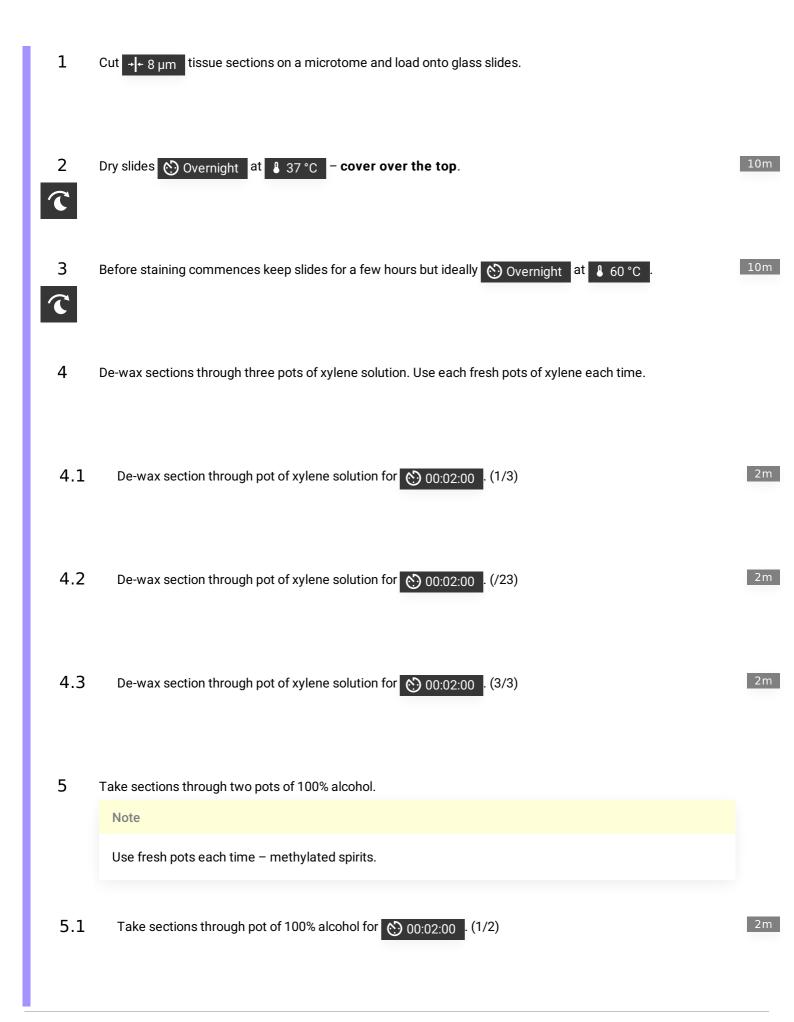
- Use only clean bottles, flasks, magnetic stirrers, tweezers, weighing spatulas, measuring cylinders – everything should be cleaned, dried and covered if left on the side before next use.
- Everything should be handled with clean tweezers gloves should not touch the samples, solutions and ideally anything placed into the solutions where the slides are.

**MATERIALS** 

#### **Materials and Reagents**

- Microtome
- Glass slides
- Xylene solution
- 100% alcohol
- Methanol
- Hydrogen peroxide (H<sub>2</sub>0<sub>2</sub>) solution
- Citrate buffer pH6
- Milli Q water
- Pressure cooker
- PBS
- Goat Serum 10%
- AlexaFluor antibody
- 0.1% Sudan black solution
- Vectashield
- Overslip

## Immunofluorescences staining protocol for oligomer imaging



10 Wash in PBS with fresh buffer (at least filtered if not cell culture grade).

Apply primary antibody for 50 01:00:00 at 8 Room temperature

10.1 Wash 00:05:00 in PBS clean squirty bottle with fresh buffer. (1/3)

9



Mount section with Vectashield and coverslip (Plasma cleaned slides).

16 Take for imaging.

