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🌐 Preparation and transformation of chemically competent Escherichia coli

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ABSTRACT

Calcium chloride (CaCl₂) transformation is a laboratory technique in prokaryotic (bacterial) cell biology. The addition of calcium chloride to a cell suspension promotes the binding of plasmid DNA to lipopolysaccharides (LPS).

MATERIALS

LAF
Scale
Ice
Incubator
Centrifuge

OPEN ACCESS

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We use this protocol and it's working

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Preparation of Calcium chloride transformation buffer

1 In a sterile flask, add  200 mL distilled water

2 Measure  25 mL ( 1 Molarity (M)) Calcium chloride


Materials:

 Calcium chloride **Sigma-aldrich Catalog #C3881**



3 Add measured reagents and mix for  00:05:00

5m


4 Fill flask with distilled water to  250 mL


5 Filter sterilize solution with a filter (0.2 μ m) and store refrigerated ( 4 °C)

Preparation of chemically competent cells

6 Prepare a culture of *E. coli* on an LB agar plate. Pick a single colony, and inoculate in  500 mL LB broth in a  1000 mL flask.

Incubate at  18 °C with shaking at 100 rpm overnight, until OD₆₀₀ reaches 0.6














7 Aliquot entire culture volume into  50 mL canonical tubes

8 Place tubes on ice for  00:20:00

20m

9 Centrifuge tubes with  5000 rcf, 4°C, 00:10:00

10m

- 10 Discard supernatant and resuspend with  16 mL Calcium chloride transformation buffer
- 11 Incubate cells on ice for  00:10:00 10m
- 12 Centrifuge tubes with  5000 rcf, 4°C, 00:10:00 10m
- 13 Centrifuge tubes with  5000 rcf, 4°C, 00:10:00 . Meanwhile, prepare  50 mL glycerol- Calcium chloride transformation buffer by diluting  7.5 mL Glycerol in  42.5 mL Calcium chloride transformation buffer 10m
- Materials:
-  Glycerol **MP Biomedicals Catalog #194680**
- 14 Discard supernatant and resuspend with  8 mL Inoue transformation buffer. Pool into two tubes
- 15 Discard supernatant and resuspend with  10 mL DMSO-Inoue per tube
- 16 Incubate cells on ice for  00:30:00 30m
- 17 Aliquot  100 μ L cell suspension into sterile  500 μ L screw cap reaction tubes (Sarstedt #72.704.200).

Note

While creating aliquots, keep original tubes, and aliquots on ice, until snap-freeze take place

18 Snap freeze tubes in liquid nitrogen using a floating foam tube rack (Southern labware #HS2166)

19 Transfer aliquots storage box, and place in an ultra-low temperature freezer or vapor-phase nitrogen tank

Note

Store tubes in 50 mL canonical tubes, or similar containers

Transformation

1h 32m 30s

20 Quickly thaw a single reaction tube with  100 μL hyper-competent cells in Inoue-DMSO


21 Mix 1-5 μL plasmid (ligation product)

Note








Do not exceed 5% of the volume competent cells

Note

Use up-to  25 ng per  50 μL of competent cells

22 Incubate cells on ice for  00:30:00

30m

- 23 Heat-shock cells at  42 °C for  00:00:30 , followed by  00:02:00 at  4 °C 2m 30s
- 24 Add  500 µL prewarmed S. O. C. medium and incubate for  37 °C at 200 rpm for  01:00:00 1h
- 25 Add desired amount of suspension on LB plates with ampicillin (100 µg/mL) and incubate overnight