



NOV 09, 2022

SHARE

WORKS FOR ME 1

Scanning electron microscopy protocol

DOI

dx.doi.org/10.17504/protocols.io.j8nlkk796l5r/v1Rene Flores Clavo¹, Francisco Breno S. Teófilo¹¹Universidade Estadual de CampinasRene Flores Clavo: CENTRO DE INVESTIGACIÓN E INNOVACIÓN EN CIENCIAS ACTIVAS MULTIDISCIPLINARIAS
Francisco Breno S. Teófilo: Electron Microscopy Laboratory - University of Campinas

RENE FLORES

Protocol Electronic Microscopic



Rene Flores Clavo

Universidade Estadual de Campinas, Centro de Investigación e...

COMMENTS 0

ABSTRACT

This protocol briefly summarizes the basic steps of a scanning electron microscopy processing. The methods adopted for fixation, post-fixation, dehydration, drying in a critical point chamber, sputter coating, and the visualization and acquisition of images are described here.

DOI

dx.doi.org/10.17504/protocols.io.j8nlkk796l5r/v1

PROTOCOL CITATION

Rene Flores Clavo, Francisco Breno S. Teófilo 2022. Scanning electron microscopy protocol .
protocols.io
<https://dx.doi.org/10.17504/protocols.io.j8nlkk796l5r/v1>

KEYWORDS

Scanning electron, microscopy protocol, bacterial identification's, LBME, CIICAM

LICENSE

This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Aug 16, 2022

LAST MODIFIED

Nov 09, 2022

PROTOCOL INTEGER ID

68749

MATERIALS TEXT

1. Sample;
2. Glutaraldehyde, 2,5%;
3. Sodium cacodylate buffer (pH 7.3) 0.1 M;
4. Osmium tetroxide, 1.0%;
5. Acetone;
6. CO₂;
7. Aluminum stubs;
8. Balzers CPD-030 Critical Point Dryer;
9. Balzers SCD 050 Sputter-Coater;
10. Scanning Electron Microscope JEOL JSM 5800LV, at 10 kV;
11. SemAfore 5.21 software.

MATERIAL SELECTION

- 1 1 mm² samples were selected in the colony.

FIXATION

- 2 Samples were fixed in a solution of **2.5 % volume** **Glutaraldehyde 25% Aqueous Solution 10 x 10 ml ampoules Electron Microscopy Sciences Catalog #16220** and **0.1 Molarity (M)** **Sodium cacodylate trihydrate Merck Millipore Sigma Catalog #C0250** buffer (**pH 7.3**), at **Room temperature** for **04:00:00**.

WASHING AFTER FIXATION

- 3 Samples was then rinsed three times in [M] 0.1 Molarity (M) [S] Sodium cacodylate trihydrate Merck Millipore Sigma Catalog #C0250 (pH 7.3), at [T] Room temperature, for [D] 00:20:00 for each rinse. 20m

POST-FIXATION

- 4 Samples was then post-fixed with [M] 1 % volume osmium tetroxide in [M] 0.1 Molarity (M) [S] Sodium cacodylate trihydrate Merck Millipore Sigma Catalog #C0250 buffer, (pH 7.3), at [T] Room temperature, for [D] 01:00:00, protected from ambient light. 1h

WASHING AFTER POST-FIXATION

- 5 Samples was then rinsed three times in sodium [M] 0.1 Molarity (M) [S] Sodium cacodylate trihydrate Merck Millipore Sigma Catalog #C0250 buffer (pH 7.3), at [T] Room temperature, for [D] 00:20:00 for each rinse. 20m

DEHYDRATION

- 6 Samples were dehydrated with ascending acetone series, for [D] 00:20:00 minutes each step - [M] 30 % volume, [M] 50 % volume, [M] 70 % volume, [M] 90 % volume, [M] 100 % volume (last one step, for three times). 20m

CRITICAL POINT DRYING

- 7 Samples was then dried using the critical point method with CO₂.

MOUNTING ON THE STUBS

- 8 Samples was then placed on aluminum stubs.

SPUTTER-COATER

- 9 Samples was then coated with a layer of 30–40 nm gold using a Balzers SCD 050 sputter-coater.

OBSERVATIONS AND IMAGE ACQUISITION

- 10 Observations and photomicrograph acquisitions were obtained using a JEOL JSM 5800LV at 10 kV with SemAfore 5.21 software.