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OT-2 Media dispensing and culture inoculation protocol

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1 Works for me

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dx.doi.org/10.17504/protocols.io.q26g7yb3kgwz/v1

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ABSTRACT

This protocol is meant to distribute reactive(s) into well plates with a single channel pipette, and then transfer culture samples from a source plate to these plates with a multi channel pipette.

This protocol uses a python script for an Opentrons 2 robot and a .csv file containing the required variables to set the number of samples, volumes of transfer, type of plates, etc... In our laboratory, this protocol has been used as part of the "High-throughput workflow for the genotypic characterization of transposon library variants" also available in protocols.io to prepare cultures plates to perform a subsequent counter-selection

DOI

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KEYWORDS

automation, opentrons, OT-2, media dispensing, culture inoculation

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OWNERSHIP HISTORY

Sep 27, 2022 | Lorea Alejaldre

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PROTOCOL INTEGER ID

70555

PARENT PROTOCOLS

In steps of

[High-throughput workflow for the genotypic characterization of transposon library variants](#)

GUIDELINES

This protocol was developed with python 3.9.7, OT App Software Version 6.1.0 and API level version 2.13. It has been tested with cultures from *Pseudomonas putida* KT2440 as part of the High-throughput workflow for the genotypic characterization of transposon library variants. LB and M9-citrate media with either kanamycin, gentamicin, streptomycin or ampicillin have been successfully run.

The maximum number of 96-well plates per run is 3 source plates to 3 final plates (using 4 tip racks and 1 tube rack).

MATERIALS TEXT

Software

- python 3.9.6
- python packages: pandas, logging, math, copy, numpy, opentrons
- OT App
- Excel

OT-2 Labware

- Opentrons Tipracks

Opentrons 96 Tip Rack 300 µL
Tip rack

Opentrons - [Link](#)

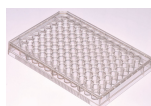
Opentrons 96 Filter Tip Rack 20 µL
Tip rack

Opentrons - [Link](#)

- 96- well plates

96-well plates, flat bottom, non treated
Cell culture plates

VWR 734-2781 [Link](#)



- Opentrons Falcon tubetrack

Opentrons 15 Tube Rack with Falcon 15 mL Conical OT Tube Rack

Opentrons - [Link](#)



- 15mL Falcon tubes

Falcon® Conical Centrifuge Tubes 15mL
Falcon Tube

Falcon 352096 [Link](#)

Equipment:

OT-2
Liquid handler

Opentrons OT-2

HEPA Module

Opentrons OT-2-HEPA [Link](#)

Single Channel Electronic Pipette (GEN2)
300uL
Opentrons Pipette
Opentrons - [Link](#)

8 Channel Electronic Pipette (GEN2) 20uL
Multi channel pipette
Opentrons - [Link](#)

SAFETY WARNINGS

It is important to use HEPA module to work in sterility

BEFORE STARTING

Note that the source and final 96-well plates will follow the same order (If a row of the source plates has empty wells the tips for the multichannel will aspirate in that well)

Files Preparation

1 Preparing Customized Template

Preparing the template (a .csv) with the specific variables for each experiment.

Here we attach one excel with several sheets:

1. Template to use in protocol
2. Explanation of each variable
3. Several examples

 [Template_Variables_PlateGeneration.xlsx](#)

1.1 Fill the template with the corresponding values

1.2 Store it as a comma-separated values file (csv) with the name *Variables-AntibioticPlatesCreation-OT.csv*



The file should be spelled exactly *Variables-SamplesMerging-OT.csv* or the python script won't work.

Make sure that the final csv is separated by commas, not by semicolons and that the variables have 2 quotation marks and not 6, in case they need quotation marks (these are added when going from excel to csv)

In Windows 10 to change excel separator from ";" to "," you should go to **Region Settings > Change date, time, or number formats > Additional settings > change List separator from ; to ,**

Other option is opening the exported .csv file in an editor and replace all ; to , and remove the extra quotation marks

2 Transferring csv to Robot

Transfer the *Variables-AntibioticPlatesCreation-OT.csv* to the directory */data/user_storage* of the OT system that we are going to use to perform the protocol.



Previous to transferring any file to the OT, we need to know the IP of the robot.

This can be obtained in the Networking section of the Device that we are going to use (In the OT-App > Devices > (three dots) Networking)

Here we present a summary of how to transfer the files in 3 OS: *Windows, Mac and Ubuntu* (applicable to most Linux)

Mac/Linux

We will use the command line with scp to transfer the file *Variables-PCRs-OT.csv* to the OT system.

We need to perform the following line:

Passing Files to OT

```
scp -i [ot_key] [file] root@[IP_OT]:/data/user_storage
```

File passing from linux (our computer) to linux (OT raspberry)

Ubuntu



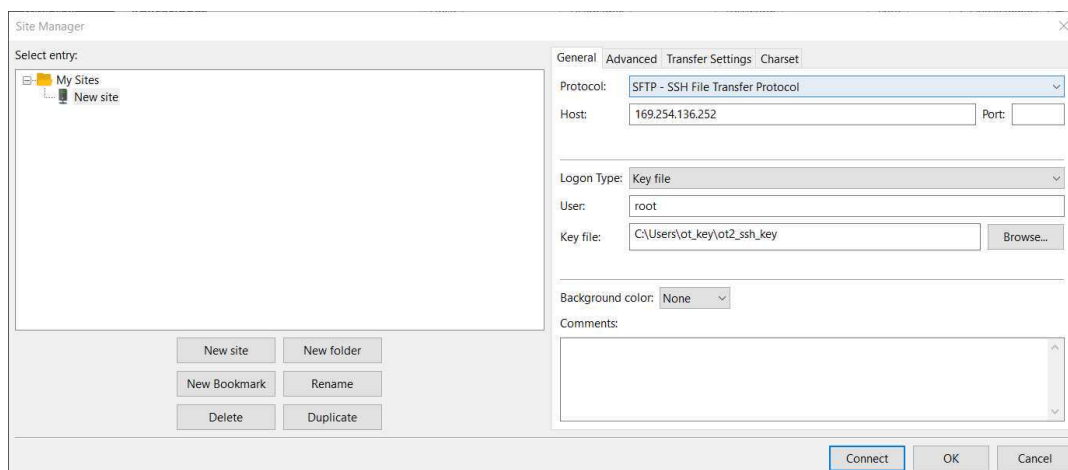
The ot key should have been previously generated, for more info visit <https://support.opentrons.com/s/article/Setting-up-SSH-access-to-your-OT-2>

Windows

There are several ways to pass files from a windows to a linux (for example with a virtual machine).

Here we will use FileZilla (<https://filezilla-project.org/download.php?type=client>).

Go to **File > Site Manager > New Site > Change Protocol to SFTP**. Then introduce in Host the OT IP, change the Logon Type to key file, change the user name to root and give the directory were the ot key is. Should look something like this



Example of setting the FileZilla to transfer files from windows (our computer) to linux (OT)

Then press connect and we will have a connection between our computer and the robot.

After this connection, we should be able to move out *Variables-SamplesMerging-OT.csv* (in our computer) to the directory */data/user_storage* in the robot.

Take in account that the IP of the robot could change, so it is possible that from time to time it will be needed to change the host in these connections.

The method described for the Windows system can be done as well in Ubuntu and Mac

3

Adding the custom labware

There is only a need to do this step when a labware that you are using is not OT official or your labware is not in the OT directory
/data/labware/v2/custom_definitions/custom_labware



This process/step is needed in the version 6.0.1 or below (when this protocol was developed).

According to their guide, this protocol should be possible only loading the labware in the OT-App (<https://support.opentrons.com/s/article/Using-labware-in-your-protocols>).

This bug has been reported and, hopefully, will be resolved as soon as possible. It is possible that you do not need to do this step if this error does not occur to you.

3.1 Creation of .json file

The description file can be obtained by describing the labware dimensions in <https://labware.opentrons.com/create/>

3.2 Creating description folder

We need to create for our custom labware a folder with the API name containing the description file (.json) called *1.json*

3.3 Transfer the description folder to the OT robot

We need to transfer the directory or directories created in Step 3.2

For that we need to introduce the following command (in Linux), for Windows process [go to step #2](#)

Transferring directory to OT

```
scp -i [ot_key] -r [directory_custom_labware]  
root@[IP_OT]:/data/labware/v2/custom_definitions/custom_be
```

Transferring the custom labware to OT

Linux

We do not need to execute this part every time the protocol is used, only when that labware is not included in the OT official labware and these directories are not in the robot that we are using

4 Simulating and getting the output (user instructions)

For more information in how to setup the ssh visit the official OT support blog (<https://support.opentrons.com/s/article/Setting-up-SSH-access-to-your-OT-2>)

4.1 Passing python script

To make sure that the protocol will work with the variables we have introduced we need to run the python script in the robot with the OT simulator.

The last script version can be found at <https://github.com/Biocomputation-CBGP/OT2/tree/main/AntibioticPlatesGeneration> (name of this file is user's choice)

For that we can [go to step #2](#) but instead of passing the variables file, we pass the python script. In this case we can transfer it to the directory */data/user_storage* or we can directly pass it to the directory */root*

(recommended)

You can do this step only once and use always the same python script

The only time that you need to change it is when there is an update in github, then you should download the new script version and do this step

4.2 *Simulating*

We should connect to the robot via ssh

Connect to Linux based OT via ssh

ssh -i [ot_key] root@[Robot_IP]

Simulate the protocol so we can know if the protocol is going to work with the given variables

Linux

Then we would move to the directory where the python script is with the **cd** command

Finally we are going to perform the simulation of the opentrons

Simulate OT protocol

opentrons_simulate -o nothing -e [name_script].py > [name_instruction_file].txt 2>&1

Simulate the protocol so we can know if the protocol is going to work with the given variables

Linux

Do not worry if it takes a little bit of time

4.3

In case that you want to see the instruction file before the next step, you can see the instruction file with cat

Show content file cmd

cat [name_instruction_file].txt

Show in the command line the instruction file content

Linux

4.4

Retrieving output file (user instructions)

To retrieve the output from the OT system you can [go to step #2](#), replicate it but transferring the file from the OT to your computer

In this file we have 3 or more sections:

1. (Optional) **Warnings**
2. **General Information:** pieces of information that are extracted from variables in the variables file and are only remainders
3. **Deck Labware Position:** this information will be provided by the OT as well
4. **15mL Falcon Reactive Positions:** for each Falcon tuberack that is needed (with 15 positions) we have a table with the reactives and their respective volumes

It is recommended to have at least 10uL more of each reactive in each tube to make sure that the pipette does not take a bubble when aspirating

In the case that something went wrong, the variables are not consistent or there is not enough deck place to perform the experiments the instruction file will be shorter and will have a single section, **Errors**.

Run Protocol

5 Setting Labware

5.1 Wipe the surface of the deck with 70% ethanol to clean and disinfect the surfaces

For more information in how to clean the robot go to the following link
<https://support.opentrons.com/s/article/Cleaning-the-robot-s-surfaces>

5.2 Set the respective labwares in the slots as the OT App and the instruction file set



Make sure that the source and final A1 wells are at the left top (common mistake), and each labware fits appropriately.

5.3 Set the different reagents in their respective falcon tubetrack positions, as stated in the instruction file



The protocol is going to aspirate from the falcon at a specific height which is set with the volume of the falcon, so make sure that you don't have a falcon with more than 1mL more than the instruction file specific volume.

This way, the pipette will not end wet in the running of the protocol

6 Load script in OT-App

Now that we have made sure that the protocol is going to work we import the python script in the OT-App.



This whole step has been developed with version 6 of the OT-App

Indications may vary from version to version

6.1 Load the script

Protocols > Import > Drag the python script

The last script version can be found at <https://github.com/Biocomputation-CBGP/OT2/tree/main/AntibioticPlatesGeneration> (name of this file is user's choice)



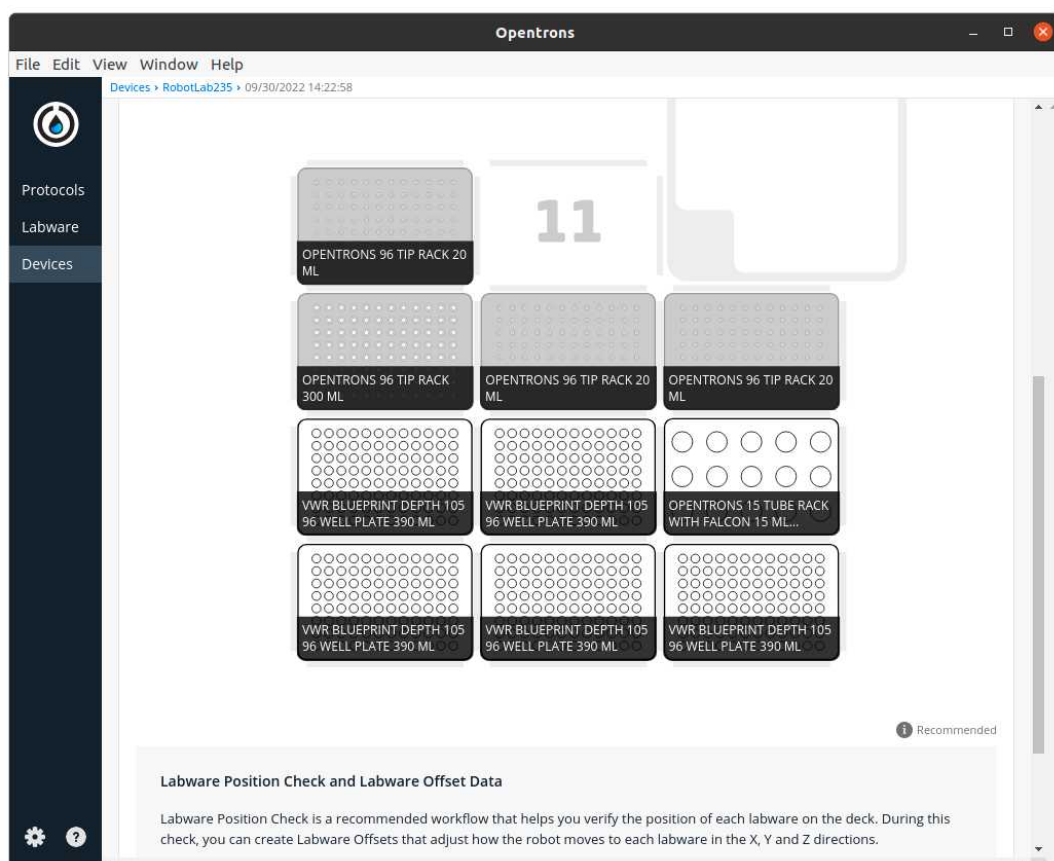
The App with version 6 analyzes your protocol before setting a robot to run, so the labware will not be shown before assigning the protocol to a specific robot.

6.2 Select Robot to Perform Script

Click in the protocol > Run Protocol > Choose the OT where the file Variables-AntibioticPlatesCreation-OT.csv is > Proceed To Setup

After clicking in Proceed to Setup you should obtain the setup of the labware as instructed in the Deck Labware Positions section of the instructions file.

The Labware Setup tab should look similar to the following image



Example of Labware Setup for this protocol

It is recommended that you perform a labware position check.

You can do it with test plates before cleaning the surface, that way you reduce the probability of contamination (using the test plates and labware) and pipetting errors (position check).

7 Run Protocol in OT

7.1 Make sure the needed calibrations are done

7.2 Labware position check is performed (if needed)

7.3 Start Run

The procedure that the robot is going to do is mainly divided in 2 parts:

1. Distributing of antibiotic or reactivities in respective plates
2. Transferring samples to corresponding plates



Make sure that the wells of the source plates are at least filled with 25µl so the pipettes can aspirate a sample



As many final plates as antibiotics per source plates introduced in the variables .csv file

We are going to obtain 1 plate per antibiotic and source plate, there will be no combining of source plates samples

The script is going to be the same so we can load it only once to the OT-App.

To re-run the protocol make sure that the filled template is in the OT system and [go to step #6.2](#)

After-Running

8 Retrieve labware from the OT

Example

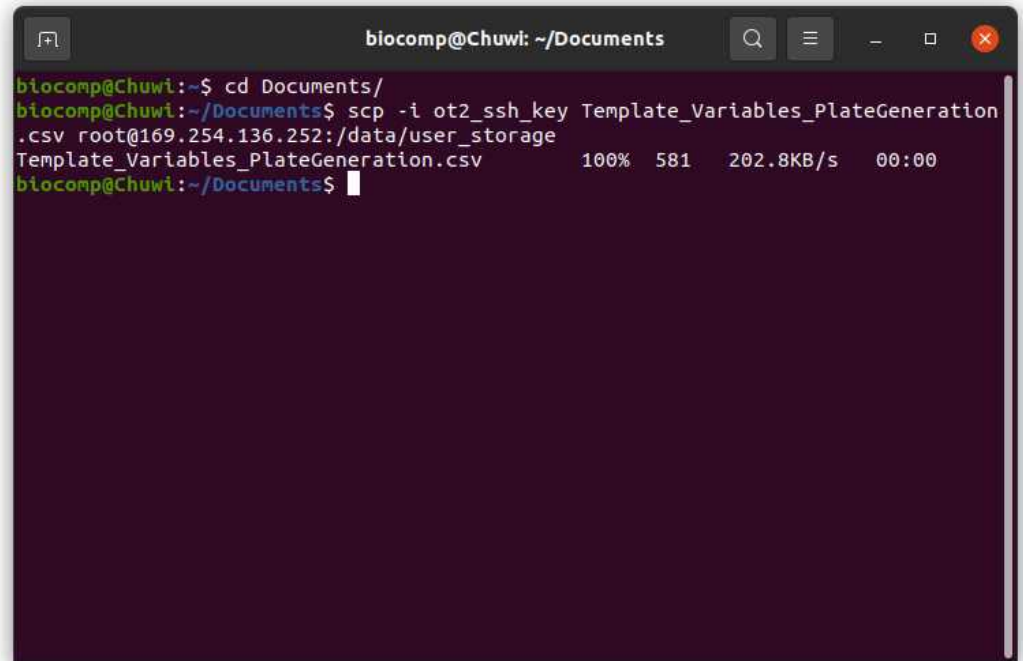
9 We want to transfer samples from 3 source plates to different antibiotic plates, one per antibiotic and source plate (Example 2 in *Template_Variables_PlateGeneration.xlsx*)

We will use a computer with an Ubuntu system connected via USB to the robot

9.1 Excel temple filled and exported to csv with the name *Variables-*

 **Variables-AntibioticPlatesCreation-OT.csv**

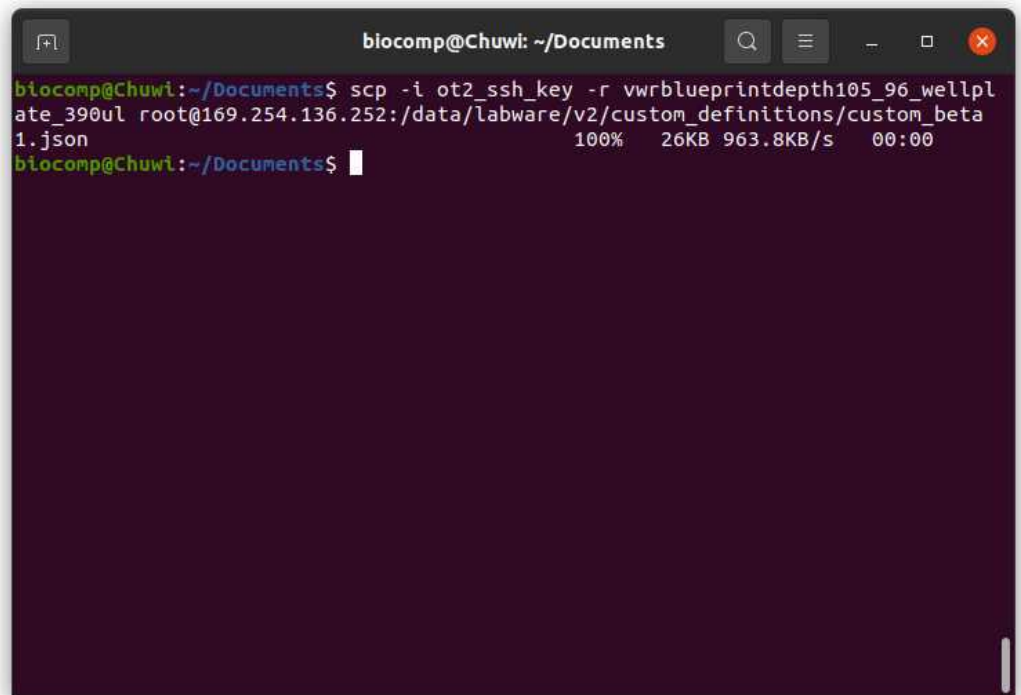
9.2 Export the csv to /data/user_storage



```
biocomp@Chuwi: ~/Documents
biocomp@Chuwi:~$ cd Documents/
biocomp@Chuwi:~/Documents$ scp -i ot2_ssh_key Template_Variables_PlateGeneration
.csv root@169.254.136.252:/data/user_storage
Template_Variables_PlateGeneration.csv      100% 581 202.8KB/s 00:00
biocomp@Chuwi:~/Documents$
```

cmd window with scp commands to transfer the .csv from our computer to the OT

9.3 We have never used a 96-well tissue plate labware with the API name vwrblueprintdepth105_96_wellplate_390ul, so we must pass the directory regarding this labware to the robot



```
biocomp@Chuwi: ~/Documents
biocomp@Chuwi:~/Documents$ scp -i ot2_ssh_key -r vwrblueprintdepth105_96_wellplate_390ul root@169.254.136.252:/data/labware/v2/custom_definitions/custom_beta1.json
100% 26KB 963.8KB/s 00:00
biocomp@Chuwi:~/Documents$
```

cmd window with scp commands to transfer the custom labware directory from our computer to the OT

- 9.4 Pass the script that I have downloaded from <https://github.com/Biocomputation-CBGP/OT2/tree/main/AntibioticPlatesGeneration> (I named it AntibioticPlateGeneration-example.py), connect to OT computer, simulate, exit OT system, store the output in *instructions_AntibioticPlateGeneration_Example2.txt* and import it to the computer

```

biocomp@Chuw1: ~/Documents
biocomp@Chuw1:~/Documents$ scp -i ot2_ssh_key AntibioticPlateGeneration-example.py root@169.254.136.252:/root
AntibioticPlateGeneration-example.py
biocomp@Chuw1:~/Documents$ ssh -i ot2_ssh_key root@169.254.136.252

      00000 00000
      0000 0000 0000 00000 00000
      000 00 000 000000 00000
      00 000000 000 000000 00
      00 -00000 0 #00 00
      000 0 0 000 00
      000 00-00 000 00
      0000 0000 00 00000
      00000000000000000000
      (000000)
      ## 000000#

# opentrons_simulate -o nothing -e AntibioticPlateGeneration-example.py > instructions_AntibioticPlateGeneration_Example2.txt 2>&1
# exit
Connection to 169.254.136.252 closed.
biocomp@Chuw1:~/Documents$ scp -i ot2_ssh_key root@169.254.136.252:/root/instructions_AntibioticPlateGeneration_Example2.txt .
instructions_AntibioticPlateGeneration_Example2.txt
biocomp@Chuw1:~/Documents$

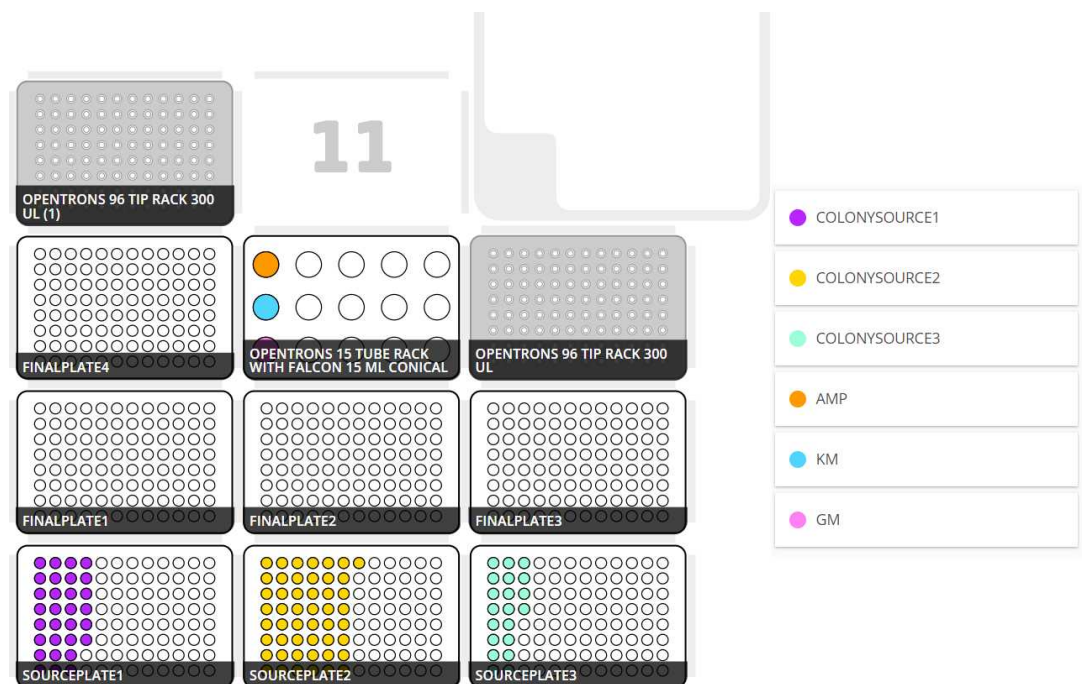
```

cmd window with scp commands to transfer the python script from our computer to the OT

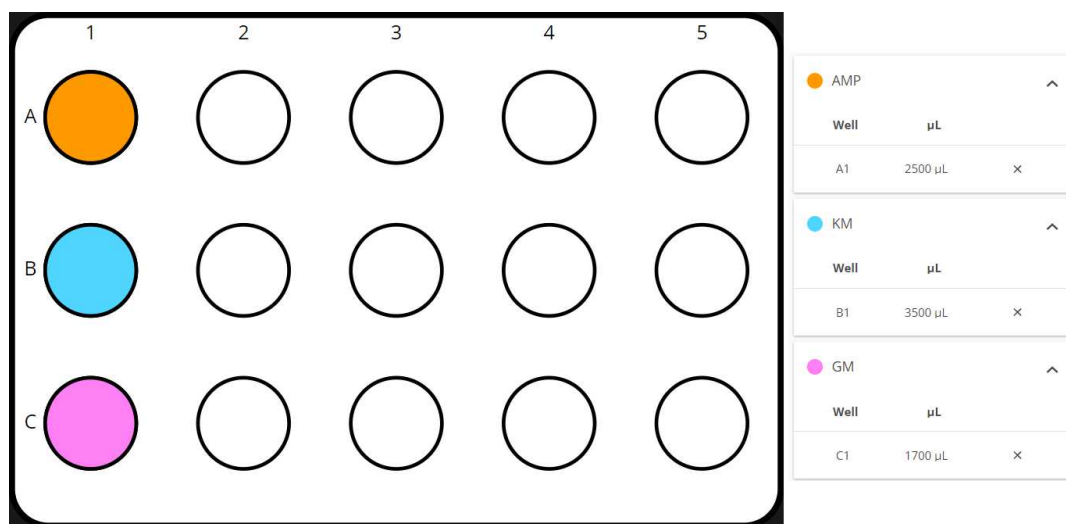


9.5 Prepare all reactivities and labwares

Initial Labware Setup and Coldblocks setup should look like the following picture



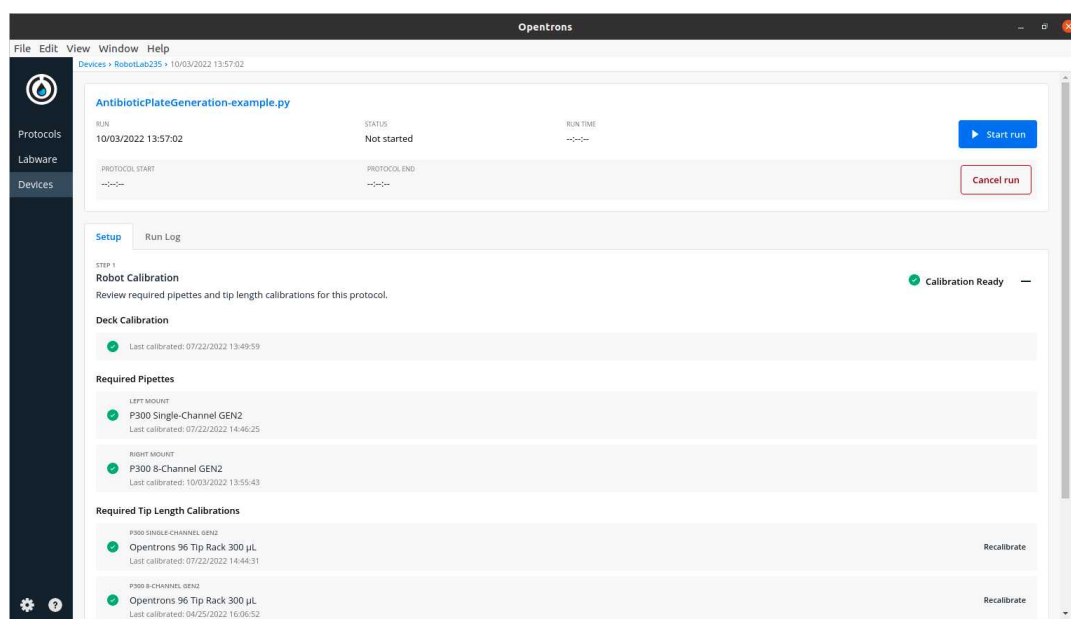
Deck sketch with labware and reactivities legend



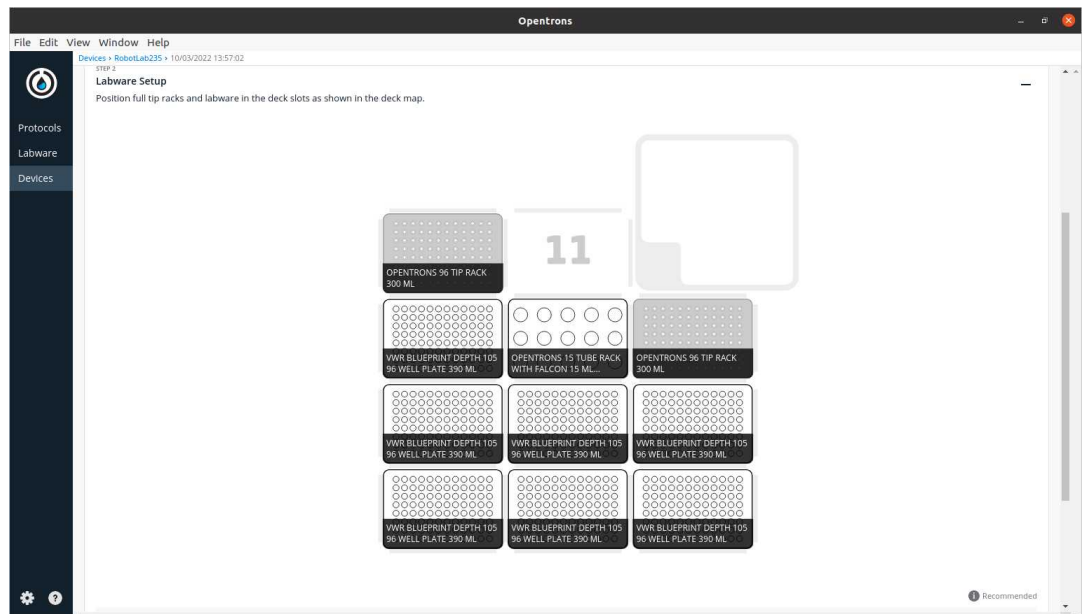
Falcon Tuberack with reactives (positions and volumes)

9.6 Load to OT-App the **AntibioticPlateGeneration-example.py** > **Run** > **Select robot** in which we are going to run the protocol

The window and tabs of the OT-App should look like the following pictures



Calibration Tab OT with Example2 variables



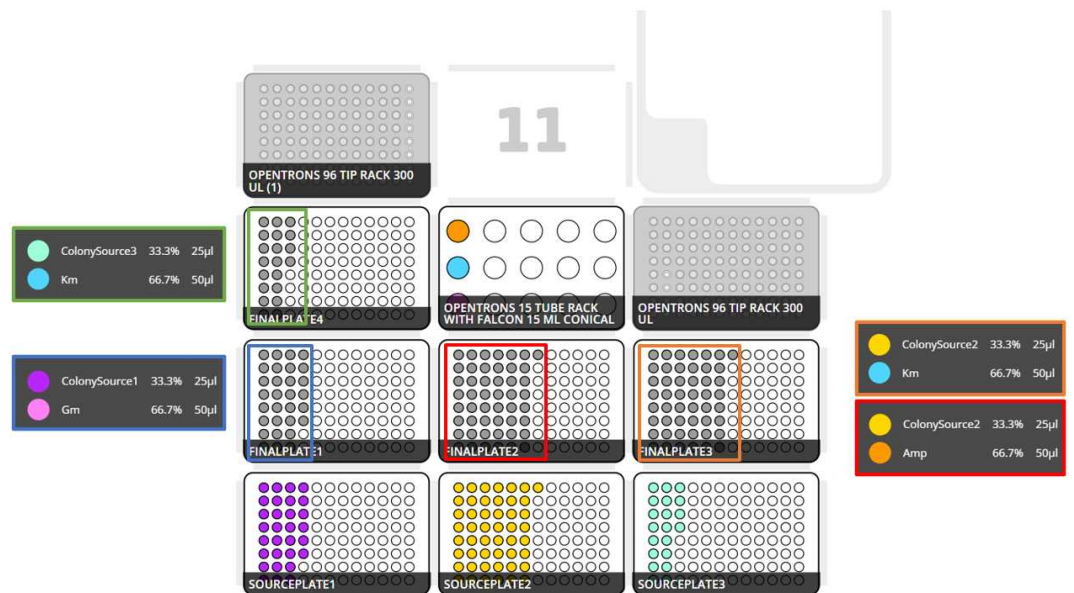
Labware Setup Tab OT with Example2 variables

9.7 Turn HEPA module on and disinfect OT-2

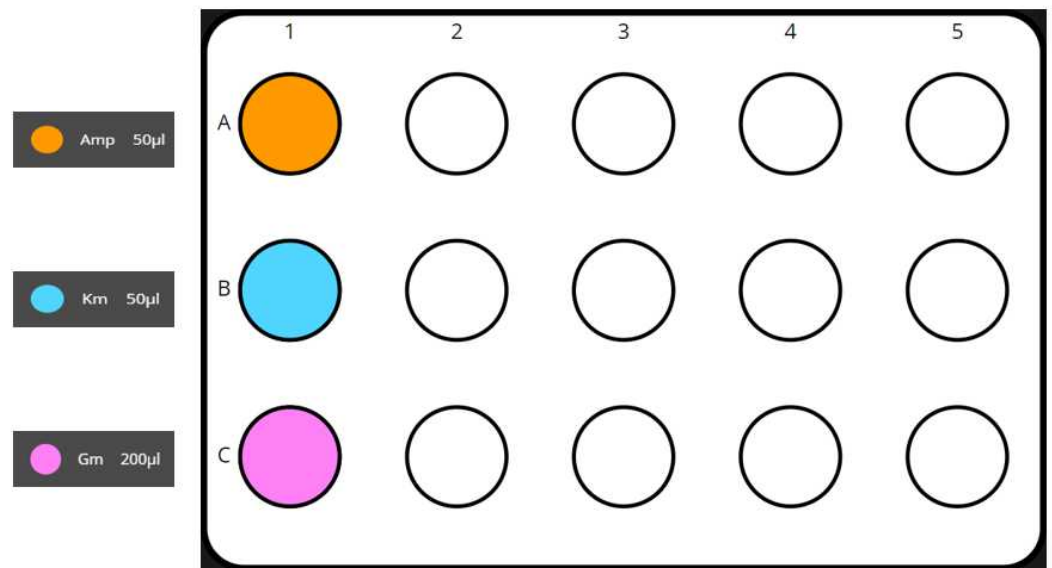
9.8 Load protocol labware as instructed in Step 9.6

9.9 Run protocol

The final results should look like the following pictures



Deck reactivities positions and volumes at the end of the run



Falcon tubetrack reactive volumes at the end of the protocol

9.10 Retrieve labwares from the OT