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LRRK2 microtubule sedimentation binding assay

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Assay to determine LRRK2 protein binding to microtubules.

Original assay by Andrea Dickey. Adapted by Mariusz Matyszewski for protocols.io.

Assay originally used in Snead, Matyszewski, Dickey et al. 2022

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Materials:

- Porcine brain tubulin purchased from Cytoskeleton, Inc.
- Purified LRRK2^{RCKW}

Buffers:

LRRK2 Binding Buffer:

- [M]20 millimolar (mM) HEPES pH 7.4
- [M]220 millimolar (mM) NaCl
- [M]0.5 millimolar (mM) TCEP
- [M]5 % volume glycerol
- [M]2.5 millimolar (mM) MgCl₂
- [M]20 micromolar (μM) GDP
- [M]20 micromolar (μM) Taxol

Microtubule preparation

25m

- 1 Polymerize tubulin at around [M]2.5 mg/mL for ⌚00:30:00 at 🌡 37 °C . Add Taxol for stabilization and incubate for another ⌚00:10:00 at 🌡 37 °C . 40m
- 2 Remove free tubulin by ultracentrifugation. 🌀108628 x g, 37°C, 00:15:00 through a [M]64 % volume glycerol cushion . 15m
- 3 Resuspend the resulting microtubule pellet in the **LRRK2 binding buffer**.

3.1 Determine microtubule concentration by running an SDS-PAGE with actin standards.

- 4 Incubate desired amount of LRRK2^{RCKW} protein ([M]200 nanomolar (nM) in our experiments) at 🌡 Room temperature for ⌚00:10:00 with varied concentrations of microtubules in the **LRRK2 binding buffer**. 10m

Assay can be modified to work with other proteins. We used the same assay to monitor LRRK1^{RCKW} binding.

- 5 Pellet the microtubules by ultracentrifugation. 🌀 **108628 x g, 25°C, 00:15:00** 15m
- 6 Quantify the depletion of LRRK2^{RCKW} by taking the supernatant and boiling for 🕒 **00:10:00** ^{10m} in SDS containing buffer for running a gel.
- 7 Samples were run on 4-12% polyacrylamide gels (NuPage, Invitrogen) and stained with SYPRO-Red Protein Gel Stain (ThermoFisher) for protein detection. Binding curves were fit in GraphPad Prism (9.2; GraphPad Software) with a nonlinear regression hyperbolic curve.