

•



Jan 07, 2022

© Detection of Cryptosporidium in stool samples by Taq-Man qPCR

botchiesenyo 1

¹University of Ghana

2

dx.doi.org/10.17504/protocols.io.bwghpbt6

Ayi and Pawlowic Lab Collaborations

mcpawlowic

Detection of Cryptosporidium parvum and/or Cryptosporidium hominis from fecal samples by qPCR.

DOI

dx.doi.org/10.17504/protocols.io.bwghpbt6

botchiesenyo 2022. Detection of Cryptosporidium in stool samples by Taq-Man qPCR. **protocols.io**

https://dx.doi.org/10.17504/protocols.io.bwghpbt6

_____ protocol,

Jul 09, 2021

Jan 07, 2022

51433

Generating samples for a standard curve

To 100 mg of fecal material (known to not be infected with Cryptosporidium), spike in a known number of Cryptosporidium parvum.

25m

DNA Extraction 2h 30m

2 Follow protocol from Quick-DNA Fecal/Soil Microbe Miniprep Kit from Zymo Research to extract total genomic DNA from stool sample (both standard curve samples and experimental samples).

m protocols.io

1

Citation: botchiesenyo Detection of Cryptosporidium in stool samples by Taq-Man qPCR https://dx.doi.org/10.17504/protocols.io.bwghpbt6

Stool samples (0.1 g) should be thoroughly vortexed in lysis buffer and then subjected to five freeze-thaw cycles prior to DNA extraction.

qPCR

3 Combine the following for a single qPCR sample (9µl per sample). Scale accordingly for number of samples and technical replicates:

5 μl 10 x Luna Universal Probe qPCR Master Mix

 $0.25\,\mu l$ $10\,\mu M$ Forward Primer (ATGACGGGTAACGGGGAAT)

0.25 μl
10 μM Reverse Primer (CCAATTACAAAACCAAAAAGTCC)
1μl
1 μM Probe ([FAM]CGCGCCTGCTGCCTTCCTTAGATG[BHQ1])

2.5µl Ultrapure water

4 Load mastermix in 96-well PCR plate using a multichannel pipette.

1h

Add 1µl extracted DNA to corresponding wells.

Keep a detailed plate map of which wells contain standard curve DNA, sample DNA, positive control DNA, no DNA control.

- 5 Spin plate in a centrifuge at low speed to make sure the samples are at the bottom of the $\stackrel{5m}{\text{well}}$.
- 6 Run qPCR cycling as described for Luna Universal qPCR Mixture. Make sure qPCR machine is measuring FAM at the correct cycle:

6.1 95°C - 3 minutes

3m

6.2

57m

Repeat 40x:

95°C - 10 seconds

60°C - 30 seconds (record FAM in this step)

Calculating number of oocysts/g

2h

m protocols.io

2

7 Calculate delta Ct values for samples that contain the standard curve.

1h

1h

Using Microsoft excel (or a similar graphing program) plot the delta Ct values on the Y-axis and the number of oocysts/g on the x-axis.

Use the analysis tools to determine the linear equation of your standard curve samples.

8 Use the equation determined from your standard control samples, you can estimate the number of oocysts/g in your experimental samples.

Y = delta Ct value for experimental sample Solve for X = oocysts/g for your experimental sample