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## Qiagen All-Prep DNA/RNA Mini Kit V.2

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protocol.



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The AllPrep DNA/RNA Mini Kit is designed to purify genomic DNA and total RNA simultaneously from a single biological sample. Lysate is first passed through an AllPrep DNA spin column to selectively isolate DNA and then through an RNeasy spin column to selectively isolate RNA. Pure DNA and RNA are purified from the entire sample, in contrast to other procedures where either the biological sample or the purified total nucleic acids is divided into two before being processed separately. The kit is compatible with small amounts of a wide range of animal cells and tissues.

The AllPrep DNA/RNA Mini Kit allows the parallel processing of multiple samples in less than 40 minutes. Time-consuming and tedious methods, such as CsCl stepgradient ultracentrifugation and alcohol precipitation steps, or methods involving the use of toxic substances, such as phenol and/or chloroform, are replaced by the AllPrep DNA/RNA procedure.

Genomic DNA purified with the AllPrep DNA/RNA procedure has an average length of 15–30 kb depending on homogenization conditions. DNA of this length is particularly suitable for PCR, where complete denaturation of the template is important to achieve the highest amplification efficiency. The purified DNA is ready to use in any downstream application, including:

- Next-generation sequencing (NGS)
- PCR
- Southern, dot and slot blot analyses
- Comparative genome hybridization (CGH)
- Genotyping, SNP analysis

HB-2697-003\_HB\_AllPrep\_DNARNA \_Mini\_1120\_WW.pdf

George Testo 2022. Qiagen All-Prep DNA/RNA Mini Kit. **protocols.io** https://protocols.io/view/qiagen-all-prep-dna-rna-mini-kit-cajiscke George Testo

Updated steps to match what is shown

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#### **Protocols**

- Extraction List
- Reagent Calculations
- Buffer RLT Plus
- Buffer RPE
- Buffer AW1
- Buffer AW2

#### Reagents

- 1 x conical of prepared Buffer RLT Plus
- 1 x conical of **prepared** Buffer RPE
- 1 x conical of **prepared** Buffer AW1
- 1 x conical of **prepared** Buffer AW2

## **Supplies**

- Lysis matrix E tubes
- DNA + RNA spin columns
- 1.5mL + 2.5mL collection tubes
- 1 x biohazard trash bags
- 1 x biohazard trash holders
- 1 x black sharpie

### **Equipment**

- 1000uL (traditional) pipettes, tip boxes, & tips
- 200uL (traditional) pipettes, tip boxes, & tips
- 20uL (traditional) pipettes, tip boxes, & tips
- Conical fliprack

When working with chemicals, always wear a suitable lab coat, disposable gloves and protective goggles. For more information, please consult the appropriate safety data sheets (SDSs). These are available online in convenient and compact PDF format at www.qiagen.com/safety where you can find, view and print the SDS for each QIAGEN kit and kit component.

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1



## **Buffer RLT**

Add Good plus Qiagen Catalog #1053393 tubes.

to Lysing Matrix E

- 2 Place the entire tissue or a portion into Lysing Matrix E tubes for homogenization.
  - For Swab(s): pick up with tweezers and cut with surgical scissors sterilized in 2.1 a Germinator 500. Place the cut end of a single swab into the Lysing Matrix E tube.
  - 2.2 For Nasal Lavage(s): Thoroughly thaw the lavage and pipette up and down 6 times. Then transfer  $\Box 500 \mu L$  of the lavage to the Lysing Matrix E tube.

3

Homogenize the lysate in Buffer RLT Plus (do not use more than 30 mg of tissue).

- 3.1 Use a homogenizer designed for Lysing Matrix E tubes.
- 3.2 Use a disruption setting that works best for your tissue (i.e. fecal, lung, etc).

3m 4

Centrifuge the lysate for **© 00:03:00** at maximum speed. Carefully remove supernatant by pipetting and transfer it to the All-Prep DNA spin column placed in a 2mL collection tube.

30s 4.1 Close the lid gently, and centrifuge for © 00:00:30 at @10.000 rpm.

**Note:** Make sure that no liquid remains on the column membrane after centrifugation. If necessary, repeat the centrifugation until all liquid has passed through the membrane.

5

Place the All-Prep DNA spin column in a new 2mL collection tube, and store at room temperature (15-25C) or a 8 4 °C for later DNA purification in the following steps.

**Note:** Do not store the All-Prep DNA spin column at room temperature or at 4C for long periods of time. Do not freeze the column.

## Total RNA Purification

4m 45s

Add 1 volume (~1x) of 70% Ethanol to the flowthrough from step 5 and mix well by pipetting. Do not centrifuge. Proceed immediately to step 7.

If some lysate was lost during homogenization, adjust the volume of ethanol.

**Note:** Precipitates may be visible after addition of ethanol, but this does not affect the procedure. For maximum RNA yields from live, use 50% ethanol instead of 70%.

- Transfer up to □700 μL of the sample, including any precipitate that may have formed, to a RNeasy spin column placed in a 2mL collection tube. Close the lid gently, and centrifuge for © 00:00:15 at ⊚10.000 rpm . Discard the flowthrough.
  - 7.1 Reuse the collection tube in step 7.

**Note:** If the sample volume exceeds 700uL, centrifuge successive aliquots in the same RNeasy spin column. Discard the flow-through after each centrifugation.

15s





## **8** Buffer

Add 700 µL of RW1 Qiagen Catalog #1053394 to the RNeasy spin column.

Close the lid gently, and centrifuge for **© 00:00:15** at **® 10.000 rpm** to wash the spin column membrane. Discard the flow-through.

9



**8** Buffer

Add 500 µL of RPE Qiagen Catalog #1018013

to the RNeasy spin column.

Close the lid gently, and centrifuge for **© 00:00:15** at **®10.000 rpm** to wash the spin column membrane. Discard the flow-through.

9.1 Reuse the collection tube in step 9.

> Note: Buffer RPE is supplied as a concentrate. Ensure that ethanol is added to Buffer RPE before use.

10



2m

**Buffer** 

Add 500 µL of RPE Qiagen Catalog #1018013 to the RNeasy spin column.

Close the lid gently, and centrifuge for **© 00:02:00** at **® 10.000 rpm** to wash the spin column membrane.

Note: The long centrifugation dries the spin column membrane, ensuring that no ethanol is carried over during RNA elution. Residual ethanol may interfere with downstream reactions or usage of resultant RNAs and or DNAs.

Note: After centrifugation, carefully remove the RNeasy spin column from the collection

tube so that the column does not contact the flow-through; carryover ethanol may occur.

Place the RNeasy spin column in a new 2mL collection tube and discard the old collection tube with the flowthrough. Centrifuge at full speed for © 00:01:00.

**Note:** Perform this step to eliminate any possible carryover of Buffer RPE, or if residual flow-through remains on the outside of the RNeasy spin column after step 10.

Place the RNeasy spin column in a new 1.5ml collection tube. Add □30-50 μL of 

⊠RNAse-free Water Contributed by users directly to the spin column membrane. Close the lid gently, and centrifuge for ⑤ 00:01:00 at ⑥10.000 rpm to elute the RNA.

# 12.1

Genomic DNA Purification 2m 15s

13

15s

**⊠** Buffer

Add **■500 µL of** AW1 **Qiagen Catalog #19081** 

to the All-Prep DNA spin

column from step 5. Close the lid gently, and centrifuge for  $\circlearrowleft 00:00:15$  at \$10.000 rpm . Discard the flow-through.

13.1 Reuse the spin column in step 13.

Note: Buffer AW1 is supplied as a concentrate. Ensure that ethanol is



14



2m

## **Buffer**

Add  $\blacksquare 500~\mu L$  of AW2 Qiagen Catalog #19072 to the All-Prep DNA spin column. Close the lid gently, and centrifuge for © 00:02:00 at full speed to wash the spin column membrane.

**Note:** Buffer AW2 is supplied as a concentrate. Ensure that ethanol is added to Buffer AW2 before use.

**Note:** The long centrifugation dries the spin column membrane, ensuring that no ethanol is carried over during RNA elution. Residual ethanol may interfere with downstream reactions or usage of resultant RNAs and or DNAs.

**Note:** After centrifugation, carefully remove the All-Prep DNA spin column from the collection tube. If the column contacts the flow-through, empty the collection tube and centrifuge the spin column again for © 00:01:00 at full speed.

15 Place the All-Prep DNA spin column in a new 1.5mL collection tube. Add  $\,\Box$ 50  $\,\mu$ L of

2m

**⊠** Buffer

the lid. Incubate at room temperature ( § 15-25 °C ) for © 00:01:00 , and then centrifuge for © 00:01:00 at © 10.000 rpm to elute the DNA.

**Note:** To achieve a higher DNA concentration, elute with 2 x  $\blacksquare$  50  $\mu$ L Buffer EB. The final DNA yield, however, may be reduced.

