

FEB 27, 2023

OPEN ACCESS

יוסם

dx.doi.org/10.17504/protocol s.io.bcy6ixze

Protocol Citation: Qiye Li, Qunfei Guo, Yang Zhou, Huishuang Tan, Terry Bertozzi, Yuanzhen Zhu, Ji Li, Stephen Donnellan, Guojie Zhang 2023. DNA extraction protocol for the eastern banjo frog using the Gentra Puregene Tissue Kit.

protocols.io https://dx.doi.org/10.17504/p rotocols.io.bcy6ixze

MANUSCRIPT CITATION:

Li Q, Guo Q, Zhou Y, et al. Protocols for "DNA extraction protocol for the eastern banjo frog using the Gentra Puregene Tissue Kit.". 2020, protocols.io. dx.doi.org/10.17504/protocol s.io.bcy6ixze.

License: This is an open access protocol distributed under the terms of the Creative Commons
Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

Created: Feb 26, 2020

© DNA extraction protocol for the eastern banjo frog using the Gentra Puregene Tissue Kit

In 2 collections

Qiye Li^{1,2}, Qunfei Guo^{1,3}, Yang Zhou¹, Huishuang Tan^{1,4}, Terry Bertozzi^{5,6}, Yuanzhen Zhu^{1,7}, Ji Li^{2,8}, Stephen Donnellan⁵, Guojie Zhang^{2,8,9,10}

¹BGI-Shenzhen, Shenzhen 518083, China;

²State Key Laboratory of Genetic Resources and Evolution, Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming 650223, China;

³College of Life Science and Technology, Huazhong University of Science and Technology, Wuhan 430074, China;

⁴Center for Informational Biology, University of Electronic Science and Technology of China, Chengdu 611731, China;

⁵South Australian Museum, North Terrace, Adelaide 5000, Australia; ⁶School of Biological Sciences, University of Adelaide, North Terrace, Adelaide 5005, Australia;

⁷School of Basic Medicine, Qingdao University, Qingdao 266071, China; ⁸China National Genebank, BGI-Shenzhen, Shenzhen 518120, China; ⁹Center for Excellence in Animal Evolution and Genetics, Chinese Academy

¹⁰Section for Ecology and Evolution, Department of Biology, University of Copenhagen, DK-2100 Copenhagen, Denmark



Qunfei Guo

of Sciences, 650223, Kunming, China;

ABSTRACT

Genomic DNA was extracted from the liver of an adult female <u>Limnodynastes</u> <u>dumerilii</u> using the Gentra Puregene Tissue Kit (QIAGEN, Hilden, Germany) with modifications as outlined in this protocol.

MATERIALS

MATERIALS

№ 2-propanol **Sigma Aldrich Catalog #19516**

🔀 Ethanol (undenatured) chem-supply Catalog #EA043

Last Modified: Feb 27, 2023

PROTOCOL integer ID:

33534

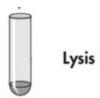
Keywords: DNA extraction, DNA purification, Gentra Puregene Tissue Kit, the eastern banjo frog

1 The following protocol is a modification of the protocol for "DNA purification from 5-10mg fresh or frozen solid tissue" using the Gentra Puregene Genomic DNA Purification Kit.

The amounts listed in this protocol are for a single extraction. Four replicate extractions were made using this protocol.

Approximately 30mg of frozen liver tissue was minced with a scapel blade on ice.

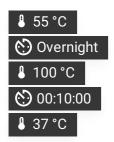
2



Dispense 900 μ l Cell Lysis Solution into a 1.5 ml microfuge tube and add the minced tissue from the previous step. Add 6 μ l of Proteinase K Solution (20mg/ml) and mix by inverting 25 times. Incubate at 55 °C overnight with agitation.

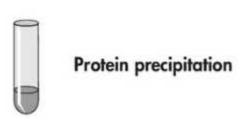
Heat RNase A Solution (12mg/ml) to 100 °C for 10 min. Add 1.5 μ l to the microfuge tube, mix by inverting 25 times and incubate at 37 °C for 60 min.

Cool to room temperature.



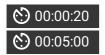


3

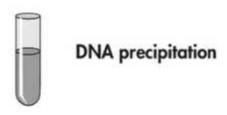


Add 300 µl Protein Precipitation Solution and vortex vigorously for 20s at high speed.

Centrifuge for 5min at 16000 x g.



4



Add 900 μ l cold 100% Isopropanol to a clean 2 ml microfuge tube and add the supernatent from the previous step by pouring carefully. Gently invert the tube 50 times to mix.

Spool the precipitated DNA onto a sterile glass rod.



Wash the spooled DNA by immersing in 300 μ l 70% ethanol and then place in a clean microfuge tube with 500 μ l 70% ethanol at -20 °C for 2h.



Note

This is the adapted step which is different from the manufacturer's instructions.

6



DNA hydration

Dry the DNA at room temperature for 1h by inverting the glass rod. Dissolve the DNA in 200 μ l of the recommended elution buffer (add more elution buffer if the DNA does not fully dissolve).



Note

This is the adapted step which is different from the manufacturer's instructions.