



Dec 05, 2020

# SPOT1 revised protocol

Forked from [SPOT1 assay](#)Guanhua Xun<sup>1</sup>, Huimin Zhao<sup>1</sup>, [stlane2](#)<sup>1</sup><sup>1</sup>University of Illinois at Urbana-Champaign**1** *Works for me* [dx.doi.org/10.17504/protocols.io.bqfkmktkw](https://dx.doi.org/10.17504/protocols.io.bqfkmktkw)**SPOT**[stlane2](#)

DOI

[dx.doi.org/10.17504/protocols.io.bqfkmktkw](https://dx.doi.org/10.17504/protocols.io.bqfkmktkw)

## PROTOCOL CITATION

Guanhua Xun, Huimin Zhao, [stlane2](#) 2020. SPOT1 revised protocol. **protocols.io**  
<https://protocols.io/view/spot1-revised-protocol-bqfkmktkw>

## FORK NOTE

For use with the SPOT at-home device.

## FORK FROM

Forked from [SPOT1 assay](#), [stlane2](#)

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## CREATED

Dec 04, 2020

## LAST MODIFIED

Dec 05, 2020

## PROTOCOL INTEGER ID

45260

## MATERIALS TEXT

### STEP MATERIALS

|   |        |
|---|--------|
| <a href="#">☒ Nuclease-free Water - 25 ml</a> <b>New England</b>                    |        |
| <b>Biolabs Catalog #B1500S</b>  | Step 2 |
| <a href="#">☒ Bst 2.0 WarmStart DNA Polymerase - 8,000 units</a> <b>New England</b> |        |
| <b>Biolabs Catalog #M0538L</b>  | Step 2 |
| <a href="#">☒ Deoxynucleotide Solution Mix - 8 umol of each</a> <b>New England</b>  |        |
| <b>Biolabs Catalog #N0447S</b>  | Step 2 |
| <a href="#">☒ Non-CRISPR nuclease</a> <b>Contributed by</b>                         |        |
| <b>users Catalog #N/A</b>   | Step 2 |
| <a href="#">☒ Non-primer oligos</a> <b>Contributed by</b>                           |        |
| <b>users Catalog #N/A</b>   | Step 2 |
| <a href="#">☒ E gene primer mix</a> <b>Contributed by</b>                           |        |
| <b>users Catalog #N/A</b>   | Step 2 |
| <a href="#">☒ Isothermal Amplification Buffer - 6.0 ml</a> <b>New England</b>       |        |
| <b>Biolabs Catalog #B0537S</b>  | Step 2 |
| <a href="#">☒ N gene primer mix</a> <b>Contributed by</b>                           |        |
| <b>users Catalog #N/A</b>   | Step 2 |
| <a href="#">☒ Reporter Probe 2</a> <b>Contributed by</b>                            |        |
| <b>users Catalog #N/A</b>   | Step 2 |
| <a href="#">☒ WarmStart RTx Reverse Transcriptase - 250 rxns</a> <b>New England</b> |        |
| <b>Biolabs Catalog #M0380L</b>  | Step 2 |
| <a href="#">☒ Manganese(II) chloride tetrahydrate</a> <b>Sigma</b>                  |        |
| <b>Aldrich Catalog #M3634</b>   | Step 2 |
| <a href="#">☒ Magnesium Sulfate (MgSO4) Solution - 6.0 ml</a> <b>New England</b>    |        |
| <b>Biolabs Catalog #B1003S</b>  | Step 2 |
| <a href="#">☒ Reporter probe 1</a> <b>Contributed by</b>                            |        |
| <b>users Catalog #N/A</b>   | Step 2 |
| <a href="#">☒ Saliva sample</a> <b>Contributed by</b>                               |        |
| <b>users Catalog #N/A</b>   | Step 2 |
| <a href="#">☒ QuickExtract DNA Extraction</a>                                       |        |
| <b>Solution Lucigen Catalog #QE09050</b>  | Step 1 |

- 1 Using the first provided microcap, collect a saliva sample into capillary A, containing QuickExtract DNA Extraction Solution (Lucigen). Insert the capillary into the SPOT1 device and press the "Start" button to run the 5-minute pretreatment.

[☒ QuickExtract DNA Extraction](#)

**Solution Lucigen Catalog #QE09050**

SPOT1 Device  
Incubating fluorometer  
University of Illinois      N/A

🔥 95 °C

🕒 00:05:00

- 2 After pretreatment, remove capillary A from the SPOT1 device and use the second provided microcap to transfer a small volume of pretreated sample to capillary B, which contains the SPOT assay mastermix. Dispense the pretreated sample into **only the top layer of the capillary**, as shown in the diagram below. **Disturbing the wax dividing layer during sample transfer may lead to a failed reaction.**

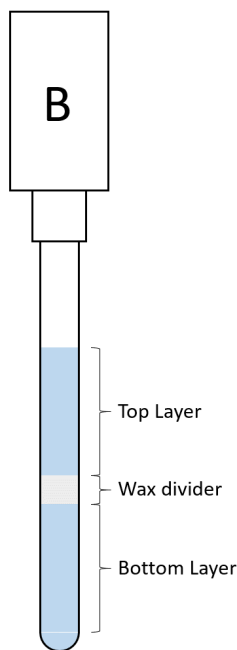


Diagram of capillary B layout. Pretreated saliva samples should be transferred into the top layer **only**. Disturbing the wax divider will result in a failed detection reaction.

**SPOT assay master mix:**

|                                    | Initial concentration | Final concentration | Amount (μL) |
|------------------------------------|-----------------------|---------------------|-------------|
| <b>Upper compartment</b>           |                       |                     |             |
| WarmStart® Bst 2.0                 | 8000 units/mL         | 160 units/mL        | 2           |
| WarmStart® RTx                     | 15,000 units/mL       | 150 units/mL        | 1           |
| Isothermal amplification buffer    | 10X                   | 0.5X                | 8           |
| dNTPs                              | 10 mM                 | 0.7 mM              | 5.6         |
| MgSO <sub>4</sub>                  | 100 mM                | 4 mM                | 3.2         |
| N gene primer mix                  | 10X                   | 0.25X               | 2           |
| E gene primer mix                  | 10X                   | 0.25X               | 2           |
| Saliva samples                     |                       |                     | 5           |
| Non-CRISPR nuclease                | 5 mg/mL or 55 μM      | 1.375 μM            | 2           |
| MnCl <sub>2</sub>                  | 50 mM                 | 0.5 mM              | 0.8         |
| Non-primer oligos (total 6 oligos) | 100 μM                | 625 nM              | 3           |
| Reporter probe 1                   | 100 μM                | 156.25 nM           | 0.125       |
| Reporter probe 2                   | 100 μM                | 312.5 nM            | 0.25        |
| Nuclease-free water                |                       |                     | 44.025      |
| <b>Total</b>                       |                       |                     | 80          |

[☒ Bst 2.0 WarmStart DNA Polymerase - 8,000 units](#) **New England**

**Biolabs Catalog #M0538L**

☒ 2 μl

[☒ WarmStart RTx Reverse Transcriptase - 250 rxns](#) **New England**

**Biolabs Catalog #M0380L**

☒ 1 μl

[☒ Isothermal Amplification Buffer - 6.0 ml](#) **New England**

**Biolabs Catalog #B0537S**

☒ 8 μl

[☒ Deoxynucleotide Solution Mix - 8 μmol of each](#) **New England**

**Biolabs Catalog #N0447S**

☒ 5.6 μl [M]0.7 Milimolar (mM)

[☒ Magnesium Sulfate \(MgSO<sub>4</sub>\) Solution - 6.0 ml](#) **New England**

**Biolabs Catalog #B1003S**

☒ 3.2 μl [M]4 Milimolar (mM)

[☒ N gene primer mix](#) **Contributed by**

**users Catalog #N/A**

[☒ E gene primer mix](#) **Contributed by**

**users Catalog #N/A**

[☒ Saliva sample](#) **Contributed by**

**users Catalog #N/A**

[☒ Nuclease-free Water - 25 ml](#) **New England**

**Biolabs Catalog #B1500S**

☒ Non-CRISPR nuclease Contributed by

users Catalog #N/A

☒ Manganese(II) chloride tetrahydrate Sigma

Aldrich Catalog #M3634

☒ 0.8 µl [M]0.5 Millimolar (mM)

☒ Non-primer oligos Contributed by

users Catalog #N/A

☒ Reporter probe 1 Contributed by

users Catalog #N/A

☒ Reporter Probe 2 Contributed by

users Catalog #N/A

- 3 Insert capillary B into the SPOT1 device and press the "Start" button to initiate the 35-minute detection reaction. 35m

SPOT1 device  
Incubating fluorometer  
University of Illinois N/A

☒ 63 °C ⌚ 00:30:00

☒ 98 °C ⌚ 00:05:00

- 4 Result ("Positive"/"Negative"/"Inconclusive") will be displayed on SPOT1 device LCD screen after completion of detection<sup>1m</sup> reaction and the 1-minute cooling period.