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# Freezing of mouse embryonic fibroblasts (MEFs) for hPSC cultures

In 1 collection

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#### **ABSTRACT**

This protocol describes the freezing of mouse embryonic fibroblasts (MEFs), which can later be used as feeder cells for human pluripotent stem cell (hPSC) culture.

#### **General notes**

- 1. This protocol can be used to freeze MEFs before or after inactivation.
- 2. Throughout this protocol, the term hPSC is used to collectively refer to both hiPSCs and hESCs. All described procedures have been tested and work equally well for hiPSCs and hESCs.
- 3. MEFs were obtained as described in Manipulating the Mouse Embryo: A Laboratory Manual, Third Edition (ISBN: 0879695919)

Andras Nagy, Marina Gertsenstein, Kristina Vintersten, & Richard Behringer. Manipulating the Mouse Embryo: A Laboratory Manual, 3rd ed.. Cold Spring Harbor Laboratory Press.

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PROTOCOL CITATION

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#### COLLECTIONS (1)

Maintenance and inactivation of mouse embryonic fibroblasts (MEFs) as feeder cells for human pluripotent stem cell culture

**KEYWORDS** 

**ASAPCRN** 

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PARENT PROTOCOLS

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Maintenance and inactivation of mouse embryonic fibroblasts (MEFs) as feeder cells for human pluripotent stem cell culture

#### **MATERIALS TEXT**

Item	Vendor	Catalog #
DMEM	Corning	10-013-CV
FB Essence	Avantor	10803-034
FBS	Gibco	10437028
200mM L-Glutamine	Sigma	G8540
Penicillin & Streptomycin	Gibco	15140-122
MEM Non-Essential Amino Acids	Gibco	11140-050
DMSO	Thermo Fisher	BP231-100
0.25% Trypsin/EDTA (Trypsin)	Gibco	25200-056
DPBS w/o Ca & Mg (DPBS)	Corning	MT21031CV
50ml centrifuge tubes	Corning	1495949A
Nunc 1.8 ml cryovials	Thermo Fisher	377267
10ml serological pipet	Corning	7200574
Styrofoam microtube freezer box	Labnet	R8000

## 1 Wash the plates twice with DPBS

2 Add Trypsin (5 ml for 15-cm plate) and incubate for **© 00:05:00** (37°C; 5% CO2)

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### 3 Add 10 ml MEF medium to neutralize the Trypsin and collect the solution into a conical tube.

## 3.1 MEF medium

A	В
DMEM	435 ml
FB Essence/FBS*	75 ml
200mM L-Glutamine	5 ml
Penicillin & Streptomycin (100x)	5 ml
MEM Non-Essential Amino Acids	5 ml

<sup>\*</sup>We have successfully used either FB Essence or FBS and have not observed an obvious difference. Final volume: 500ml



- 4 Centrifuge the cell suspension at **3250 x g, Room temperature, 00:10:00**
- 5 Discard supernatant, and re-suspend the cells in MEF Freezing Medium I at a concentration 2X the desired final freezing concentration (Freezing concentration will vary with MEF usage.)

## 5.1 MEF Freezing Medium I

Α	В
FB Essence/FBS*	5 ml
MEF media	5 ml

<sup>\*</sup>We have successfully used either FB Essence or FBS and have not observed an obvious difference. Final volume: 10ml

6 While swirling, slowly and drop-wise, add an equal volume of MEF Freezing Medium II to the cell suspension.

## 6.1 MEF Freezing Medium II

Α	В
FB Essence/FBS*	8 ml
DMSO	2 ml

<sup>\*</sup>We have successfully used either FB Essence or FBS and have not observed an obvious difference. Final volume: 10ml

- Dispense cell mixture into pre-labeled cryovials (10x10<sup>6</sup>) including type of cells, passage number, date frozen, and who handled cells.
- 8 Place cryovials into Styrofoam microtube freezer box or NALGENE™ Cryo 1°C Freezing Container pre-filled with 250 ml room temperature isopropanol. (This ensures that a -1°C/min rate of cooling is achieved, which is critical to cell viability.)
- 9 Freeze at © Overnight § -80 °C
- 10 For long term storage, store cryovials in liquid nitrogen (-196°C).

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