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1 Works for me

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Invitrogen PureLink® Genomic DNA Mini Kit

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**BEFORE STARTING** 

Add 96-100% ethanol to PureLink® Genomic Wash Buffer 1 and PureLink® Genomic Wash Buffer 2 according to instructions on each label. Mix well.

Mark on the labels that ethanol is added.

Store both wash buffers with ethanol at room temperature.

### Preparing lysates: Mammalian cells, tissues

- Set a water bath or heat block at § 55 °C
- For adherent cells (≤5 × 10<sup>6</sup> cells): remove the growth medium from the culture plate and harvest cells by trypisinization or a method of choice.

For suspension cells (≤5 × 106 cells): harvest cells and centrifuge the cells at 250 × g for 5 minutes to pellet cells. Remove the growth medium.



Resuspend the cells from Step 2 in 200 µl PBS.

- 4 Add **□20 µl Proteinase K** (supplied with the kit, stored in § 4 °C) to the sample.
- 5

Add 20 µl RNase A (supplied with the kit, stored in § 4 °C) to the sample, mix well by brief vortexing, and incubate at room temperature for 2 minutes © 00:02:00.

6

Add 200 µl PureLink® Genomic Lysis/Binding Buffer and mix well by vortexing to obtain a homogenous solution.

7

Incubate at § 55 °C for 10 minutes © 00:10:00 to promote protein digestion.

8

Add  $\mathbf{200}\ \mu\text{l}$  96–100% ethanol to the lysate.

Mix well by vortexing for 5 seconds to yield a homogenous solution.

# Binding DNA

- 9 Remove a PureLink® Spin Column in a Collection Tube from the package.
- 10 Add the lysate (~640  $\mu L)$  to the PureLink® Spin Column.
- 11

Centrifuge the column at  $10,000 \times g$  for 1 minute at room temperature.

**(3)** 10000 x g, Room temperature 00:01:00

If you are processing >200 µL starting material such as blood, buccal swabs, or Oragene™ preserved saliva, you need to perform multiple loading of the lysate by transferring any remaining lysate to the same PureLink® Spin Column (above) and centrifuge at 10,000 × g for 1 minute.

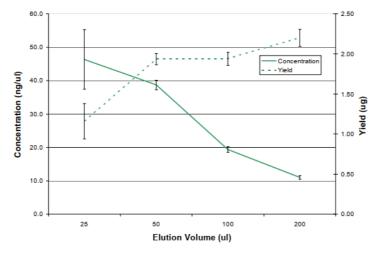
12 Discard the collection tube and place the spin column into a clean PureLink® Collection Tube supplied with the kit.

# Washing DNA

- 13 Add **3500 μl Wash Buffer 1 prepared with ethanol** to the column.
- 14 Centrifuge column at room temperature at 10,000 × g for 1 minute.
  - (3) 10000 x g, Room temperature 00:01:00
- 15 Discard the collection tube and place the spin column into a clean PureLink® collection tube supplied with the kit.
- 16 Add **Σ500 μl Wash Buffer 2 prepared with ethanol** to the column.
- 17 Centrifuge the column at maximum speed for 3 minutes at room temperature.
  - **\$\$16000 x g, Room temperature 00:03:00**
- 18 Discard collection tube.

# **Eluting DNA**

- 19 Place the spin column in a sterile 1.5-mL microcentrifuge tube.
- 20 Add 25–200 μL of PureLink® Genomic Elution Buffer to the column.



**Figure Legend:** Genomic DNA was purified from 100 μL blood samples with the PureLink® Genomic DNA Mini Kit using different elution volumes.

21



Incubate at room temperature for 1 minute.

§ Room temperature © 00:01:00

22



Centrifuge the column at maximum speed for 1 minute at room temperature.

**\$\$16000 x g, Room temperature 00:01:00** 



The tube now contains purified genomic DNA.

23



To recover more DNA, perform a second elution step using the same elution buffer volume as first elution in another sterile, 1.5-mL microcentrifuge tube.

24 Centrifuge the column at maximum speed for 1.5 minutes at room temperature.

**\$\$16000 x g, Room temperature 00:01:30** 

25 The tube contains purified DNA. Remove and discard the column.

# Storing DNA

- 26 Store the purified DNA at -20°C or use DNA for the desired downstream application.
- Por long-term storage, store the purified DNA in PureLink® Genomic Elution Buffer at −20°C as DNA stored in water is subject to acid hydrolysis.
- To avoid repeated freezing and thawing of DNA, store the purified DNA at  $4^{\circ}$ C for immediate use or aliquot the DNA and store at  $-20^{\circ}$ C for long-term storage.