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# Monosynaptic Rabies Tracing

Forked from [Mouse Stereotaxic Surgery](#)

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## ABSTRACT

This protocol describes the steps for performing stereotaxic surgery in mice. It is applicable to intracranial injections (e.g. virus, drug) and placement of implants (e.g. optical fibers, electrode arrays) into targeted regions of mouse brains.

OPEN ACCESS



DOI:

[dx.doi.org/10.17504/protocols.io.3byl4qp9jvo5/v1](https://dx.doi.org/10.17504/protocols.io.3byl4qp9jvo5/v1)

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<https://dx.doi.org/10.17504/protocols.io.3byl4qp9jvo5/v1>

**MANUSCRIPT CITATION:**  
Jonathan S Schor, Isabelle Gonzalez Montalvo, Perry WE Spratt, Rea J Brakaj, Jasmine A Stansil, Emily L Twedell, Kevin J Bender, Alexandra B Nelson (2022) Therapeutic deep brain stimulation disrupts movement-related subthalamic nucleus activity in Parkinsonian mice eLife 11:e75253

<https://doi.org/10.7554/eLife.75253>

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**Protocol status:** Working  
We use this protocol and it's working

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- 1 Rabies injections were performed in a BSL-2 surgical suite following the protocol described in [dx.doi.org/10.17504/protocols.io.n2bvj6qynlk5/v1](https://dx.doi.org/10.17504/protocols.io.n2bvj6qynlk5/v1)
  - 1.1 For modified, G-deleted rabies viruses, first inject a Cre-dependent helper virus (AAV-DIO-sTpEpB-GFP) to restrict expression of the EnvA receptor (TVA) and rabies glycoprotein necessary for subsequent rabies virus infection.
  - 1.2 2 weeks later, inject the modified, G-deleted rabies virus (EnvA-G-deleted-rabies-mCherry) into the same coordinates as the helper virus.
  - 1.3 For a detailed protocol regarding subsequent perfusion, sectioning, staining, imaging, and analysis with MBF NeuroInfo software, see Eastwood, et al., 2019

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6570587/>