



Aug 12, 2021

Single-cell ATAC sequencing

Klaus H. Kaestner Lab¹, Suzanne Shapira¹¹University of Pennsylvania Perelman School of Medicine

Suzanne Shapira: HIRN-HPAP correspondence: sshapira@penmedicine.upenn.edu

1 Works for me

 Sharedx.doi.org/10.17504/protocols.io.bwddpa26

Human Islet Research Network



Lili Liang

ABSTRACT

Interrogating cell-type specific chromatin accessibility can reveal cis-regulatory elements linked to downstream gene expression patterns. Assay for Transposase-Accessible Chromatin using Sequencing (ATAC-Seq) is a technique that assays genome-wide chromatin accessibility and can reveal nucleosome positioning patterns, map enhancer and promoter regions, or reveal transcription factor binding sites. Single-cell Assay for Transposase-Accessible Chromatin using Sequencing (scATAC-seq) adapts the bulk ATAC-sequencing protocol to separate individual nuclei and assay chromatin state with single-cell resolution. This technique allows for resolution of gene accessibility patterns within specific pancreatic cell types such as α -cells, β -cells, and acinar cells.

Date submitted: August 13, 2020

DOI

dx.doi.org/10.17504/protocols.io.bwddpa26

EXTERNAL LINK

<https://hpap.pmacs.upenn.edu/explore/workflow/islet-molecular-phenotyping-studies?protocol=5>

PROTOCOL CITATION

Klaus H. Kaestner Lab, Suzanne Shapira 2021. Single-cell ATAC sequencing. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.bwddpa26>



KEYWORDS

null, Single-cell ATAC sequencing, HPAP, HIRN

LICENSE

This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited









CREATED

Jul 06, 2021

LAST MODIFIED

Aug 12, 2021

Steps in pre-processing

- 1 1. Transfer handpicked islets (approximately 5,000 IEQs) into  **15 mL** conical tube.
2. Add  **10 mL** of 1xPBS w/o Ca²⁺, Mg²⁺ ([Rockland, MB-008](#)). Centrifuge for 2 min at RT, 180 xg. Aspirate the supernatant.
3. Add  **1 mL** of warm ( **37 °C**) 0.05% Trypsin ([Invitrogen, 25300054](#)) to the islets. Pipette up and down with p1000.
4. Incubate at  **37 °C** for 9 min, or until cells are in single cells. Pipette up and down at t=7 min, 4 min, 2 min, 0 min.
5. Stop the trypsin reaction by adding  **1 mL** of 100% FBS ([Hyclone, SH3091003](#)) to the dissociated islets and pass cells through BD FACs tube with strainer top ([Corning 352235](#))
6. Use  **1 mL** of 100% FBS to rinse the tube and pass through the strainer.
7. Transfer cells to  **15 mL** conical. Centrifuge 4 min, 400 xg.
8. Remove the supernatant and wash cells with PBS with 10% FBS. Centrifuge for 4 min, 400 xg.
9. Wash the cells with PBS with 10% FBS and centrifuge for 4 min, 400 xg. Remove the supernatant.
10. Count cells using a countess chamber.
11. For the scATACseq, do the final resuspension in 0.04% BSA in PBS as is consistent with the instructions for using fresh cell in the Nuclei Isolation Protocol.

Links to kits used in post-processing

- 2 1. For nuclei isolation the protocol used is: [Nuclei Isolation for Single Cell ATAC Sequencing](#). We target a 5000 nuclei recovery for this protocol.
2. Samples are processed for scATAC seq using [Chromium Single Cell ATAC Reagent Kits](#).