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Jul 23, 2020

Perfusion

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1 Works for me This protocol is published without a DOI.

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PROTOCOL CITATION

George Lab

Molly Brennan, Olivier George 2020. Perfusion. **protocols.io** https://protocols.io/view/perfusion-bivzke76

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CREATED

Jul 22, 2020

LAST MODIFIED

Jul 23, 2020

PROTOCOL INTEGER ID

39577

1 WHOLE ANIMAL FIXATION VIA TRANSCARDIAL PERFUSION

List of Stuff Needed for Perfusion

- -4% Formaldehyde
- -0.9% Hep-saline
- -Bottles & pump
- -Surgical tool (scissors, clamps, bone rongeurs, spatula)
- -Vials
- -timer

2 Preparation

- -Switch waste flow to deposit into chemical waste bucket
- -Prepare perfusion pumps with 0.9% saline and 4% paraformaldehyde, check flow
- -Test system; check for flow and air bubbles

3 Anesthesia

- -Inject Clorohydrate 35 % into the left lower ventral abdomen
- -Wait approximately 5 min for anesthetic to take effect
- -Pinch the tip of the tail forcefully to test for reaction
- -Add additional amount of anesthesia if necessary

4 Perfusion Rates:

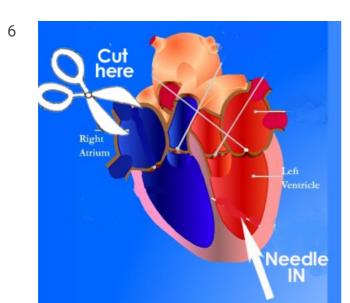
Koob Lab-Pump runs in RPMS

-Prefixative rinse -saline- 3 minutes @ max 90 RPM(i.e. ~24mls/min)

- -Formaldehyde- 5 minutes @ max 90 RPM(i.e. ~24mls/min)
- -Continue Form- 15 minutes @ 65 RPM(i.e. ~16mls/min)

5 Surgery and Fixation (do under hood or safe place)

- -Place animal on the downdraft, make a "V" shape above the genital
- -Lift skin upwards firmly using forceps, pinch a small hole into the abdomen, then cut along the "V" line
- -Continue to cut upwards to expose the diaphragm, and then cut through the diaphragm
- -Cut through the rib cage to expose the thoracic cavity
- -Clamp open excess tissue for better access of the heart
- -While holding the middle of the heart steady with forceps, make a cut slightly right to the apex of the heart (left ventricle), using one tip of a sharp scissors, then insert both tips of small scissors to make a bigger hole
- -Insert catheter needle through the cut, up to the aorta. Clamp the entrance



- -Make a cut in the dark red-colored right atrium,
- -Circulate 0.9% saline, make sure fluid is flowing freely, until the limbs of the rat are discolored (3 min)
- -Turn off saline pump, and circulate 4% paraformaldehyde (approximately 250 ml) until neck is stiffened (3+15 min)

7 Brain Removal

- -Use a pair of scissors to cut connective tissue under the skin, reaching in between the eyes
- -Cut skin open, exposing muscles on top of skull to the eyes
- -Use scalpel, cut muscle connecting to the skull bone on both sides
- -Carefully place bone cutter against the inside of the skull, and make cuts following the scalpel incision
- -Lift the occipital, parietal, and frontal bones using forceps, fully exposing the brain
- -Cut membrane on top and scoop brain out of the skull; cut optical nerves and connective tissue
- -Store brain in 4% fixative for 1 day.

8 Sucrose Treatment

- -Take the brain out of the fixative, dab dry on utility towel
- -Place brain in 20% sucrose solution (in distilled water) until it sinks to the bottom of the vial
- -Place brain in 30% sucrose solution (in distilled water) for another day until it sinks to the bottom of the vial
- -Freeze brains before cutting in dry ice.