

SEP 26, 2023

## OPEN ACCESS



#### DOI:

dx.doi.org/10.17504/protocol s.io.81wgbxnnqlpk/v1

**Protocol Citation:** Jian You Wang, Lamis Berqdar, Salim Al-Babili 2023. Protocol for abscisic acid (ABA) extraction from plant seeds.

#### protocols.io

https://dx.doi.org/10.17504/protocols.io.81wgbxnnqlpk/v1

License: This is an open access protocol distributed under the terms of the Creative Commons
Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

**Protocol status:** Working We use this protocol and it's working

Created: Sep 22, 2023

# Protocol for abscisic acid (ABA) extraction from plant seeds

Jian You Salim Al-Wang<sup>1,2</sup>, Lamis Berqdar<sup>1</sup>, Babili<sup>1,2</sup>

<sup>1</sup>The BioActives Lab, Center for Desert Agriculture, King Abdullah University of Science and Technology, Saudi Arabia;

<sup>2</sup>The Plant Science Program, Biological and Environmental Science and Engineering Division, King Abdullah University of Science and Technology (KAUST), Saudi Arabia

Salim Al-Babili: \*Correspondence: salim.babili@kaust.edu.sa;

#### **KAUST**



lamis.bergdar

#### **DISCLAIMER**

The authors declare no competing interests.

#### **ABSTRACT**

The plant hormone abscisic acid (ABA) regulates seed dormancy and germination. Here, we present a protocol for ABA extraction from plant seeds. We describe necessary steps required for preparation and extraction, followed by liquid chromatography-mass spectrometry (LC-MS/MS) analysis for ABA quantification.

**ATTACHMENTS** 

851-294.pdf

Last Modified: Sep 26,

2023

GUIDELINES

### PROTOCOL integer ID:

88357

**Keywords:** abscisic acid, plant seeds, LC-MS/MS, plant hormone

#### **Author Contributions**

S.A.-B. and J.Y.W. conceived the project. J.Y.W., and L.B. conducted experiments. L.B., J.Y.W., and S. A.-B. wrote, reviewed, and edited the protocol.

#### **Competing interests**

The authors declare no competing interests.

#### **MATERIALS**

#### List of equipment

- Calibrated balance.
- Grinder (Mixer Mill MM 400, RETSCH®),
- Sonicator (Branson 5510R-DTH Ultrasonic machine),
- ultra-centrifuge (Centrifuge 5424 R, EppendorfTM),
- speedVac (Concentrator Plus, EppendorfTM),
- 0.2 um filter (Non-Sterile Syringe Filter 13 mm; WhatmanTM UnifloTM),
- 0.1 mL Micro-insert clear glass tube (fisher scientific),
- 1.5 mL LCMS vial.

#### BEFORE START INSTRUCTIONS

#### To prepare before to start:

## Sample preparation

2h 42m

1m

- Grind 7-8 seeds in a Safe-Lock 2.0 mL Eppendorf tube with 3-4 beads for 00:01:00 frequency 25-26 Hz.
- Weight 4 100 mg fine powder of ground seeds.

Add 1 mL of Standard solution in each tube. 15m Sonicate the samples for 00:15:00 4 Centrifuge for 3 14000 rpm, 00:05:00 , then transfer the supernatant to a new 2 mL Eppendorf tube. Add another **A** 1 mL of Standard solution. 7 15m Sonicate (5) 00:15:00 Centrifuge for (14000 rpm, 00:05:00) 8 5m 9 Transfer the supernatant ( $\mathbb{Z}_2$  mL) to a 2 mL Eppendorf tube. 10 Dry the supernatant under vacuum by speedVac for 6002:00:00 2h

1m

- Filter the re-suspended solution through a 1- 0.22 μm filter into 0.1 mL micro-insert 29x5.7 mm clear glass tubes with inserted vials.
- **14** Tap the bottle to remove any bubbles.

## Sample quantification

7m

- ABA quantification is performed by LC-MS/MS using a UHPLC-Triple-Stage Quadrupole Mass Spectrometer (Thermo Scientific Altis) machine.
- Chromatographic separation is achieved on the Hypersil GOLD C18 Selectivity HPLC Columns (150 × 4.6 mm; 3 μm; fisher scientific) with mobile phases consisting of water (A) and acetonitrile (B), both containing 0.1% formic acid, and the following linear gradient (flow rate, 0.5 mL/min): 0–10 min, 15%–100 % B, follow it by washing with 100 % B for 00:05:00 and equilibration with 15 % B for 00:02:00.

7m

- 17 Inject  $\underline{L}$  10  $\mu L$  of sample, maintain the column temperature at  $\underline{l}$  35 °C for each run.
- 18 Set the MS parameters of Thermo ScientificTM AltisTM as follows:
  - negative mode,
  - ion source of H-ESI,

- ion spray voltage of 3000 V,
- sheath gas of 40 arbitrary units,
- aux gas of 15 arbitrary units,
- sweep gas of 0 arbitrary units,
- ion transfer tube gas temperature of \$\mathbb{g}\$ 350 °C
- vaporizer temperature of \$\mathbb{I}\$ 350 °C
- collision energy of 20 eV,
- CID gas of 2 mTorr, and
- full width at half maximum (FWHM) 0.4 Da of Q1/Q3 mass.

#### Note

The characteristic Multiple Reaction Monitoring (MRM) transitions (precursor ion  $\rightarrow$  product ion) were the characteristic MRM transitions (precursor ion  $\rightarrow$  product ion) were 263.2  $\rightarrow$  153.1 for ABA; 269.2  $\rightarrow$  159.1 for D6-ABA.