




Upload image

Jul 07, 2020

# Phenol-chloroform DNA purification

Tomasz Suchan<sup>1</sup><sup>1</sup>W. Szafer Institute of Botany, Polish Academy of Sciences**1** Works for me dx.doi.org/10.17504/protocols.io.re6d3he

Molecular Biogeography Group

 Tomasz Suchan  
W. Szafer Institute of Botany, Polish Academy of Sciences

## ABSTRACT

Because of the influence of salts on enzymatic reactions, Qiagen extractions can be purified before making RAD tags. First, it is necessary to perform a chloroform extraction (phenol / chloroform 50/50) by adding SDS to the aqueous phase (final conc. 0.1%), this eliminates contaminants introduced by the matrix of the column. Then, perform an ethanol precipitation followed by a wash with 70% ethanol. This is to eliminate the salt that was used to bind the nucleic acids to the column and also to remove EDTA (inhibitor!) which is 0.5 mM in the buffer AE. Eluting with water is nonsense as the pH is not optimal and thus yield is poor and the DNA is not buffered and there is always too much salt.

## DOI

[dx.doi.org/10.17504/protocols.io.re6d3he](https://dx.doi.org/10.17504/protocols.io.re6d3he)

## PROTOCOL CITATION

Tomasz Suchan 2020. Phenol-chloroform DNA purification. **protocols.io**  
[dx.doi.org/10.17504/protocols.io.re6d3he](https://dx.doi.org/10.17504/protocols.io.re6d3he)

## LICENSE

This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

## CREATED

Jul 02, 2018

## LAST MODIFIED

Jul 07, 2020

## PROTOCOL INTEGER ID


13502

## MATERIALS

NAME	CATALOG #	VENDOR
Phenol-chloroform-isoamyl alcohol 25:24:1 (PCI)	15593049	Invitrogen - Thermo Fisher
EtOH 100% at -20°C		
EtOH 70% at 4°C		
20% Sodium dodecyl sulfate (SDS)		
Na acetate 3M pH 4.8 or 5.2		

## Phenol-chloroform extraction

- 1 To 100ul eluted DNA, add 0.5 ul of 20% SDS (\*this may not be necessary as there are no cells) and 100 ul of phenol-chloroform.

- 2 Vortex well.
  - 3 Centrifuge at room temperature for 5 min, at full speed (14,000 rpm).
  - 4 Pipette the aqueous phase (upper phase, aprox 80 ul, it is better to leave some DNA than to pipette phenol) to a new labeled tube.
  - 5 Discard original tube.
- 

The tube contains phenol-chloroform.
- 6 Proceed to ethanol precipitation.

#### Ethanol precipitation

- 7 Add 1/10 volume Na acetate 3M pH 4.8 or 5.2 (ie 8ul for 80 ul DNA solution)  
add 2 volumes ETOH 100% (storage -20°C) = 176 ul  
  
Total volume: 264ul, possible with 264 ng of starting DNA - otherwise, you must add a carrier: tRNA, glycogen or linear acrylamide.
- 8 Vortex.
- 9 Put on dry ice for 30 min. or overnight at -20°C
- 10 Centrifuge at 4°C for 30 min at 14,000 rpm.
- 11 Discard the supernatant.
- 12 Wash with 500ul EtOH 70% (storage 4°C)
- 13 Centrifuge at 4°C for 5 min at 14,000 rpm.

- 14 Discard the supernatant
- 15 Quick spin
- 16 Pipette out the last drop of EtOH
- 17 Speed Vac for 3' or 5-7 min at room temperature.
- 18 Resuspend in 20ul of water or Tris 10 mM pH 7.5 or 8.0