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© cDNA synthesis using SuperScript™ IV

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1 Works for me

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ABSTRACT

The following protocol is intended as a downstream application for our

<u>Purification of RNA from a DNA/RNA Extract</u> protocol. This protocol describes how to synthesise a first-strand non-specific complementary DNA (cDNA) from a purified RNA extract using <u>SuperScript IV Reverse Transcriptase</u>. The second strand synthesis is usually not required for most downstream applications. This protocol is a simplified and condensed version of the full protocol provided by the manufacturer.

ATTACHMENTS

SSIV_First_Strand_Synthes is_System_UG.pdf

MATERIALS

NAME	CATALOG #	VENDOR
Bovine Serum Albumin (BSA)	B14	Thermo Fisher Scientific
SuperScript™ IV First-Strand Synthesis System	18091050	Thermo Fisher Scientific
RNaseOUT™ Recombinant Ribonuclease Inhibitor	10777019	Thermo Fisher Scientific
Ribonuclease H (RNase H)	18021071	Thermo Fisher Scientific
DNA Polymerase I (10 U/μL)	EP0041	Thermo Fisher Scientific
Random hexamers	N8080127	Thermo Scientific

STEPS MATERIALS

NAME	CATALOG #	VENDOR
Random hexamers	N8080127	Thermo Scientific
Nuclease-free autoclaved DEPC-treated water	T143.1	Carl Roth
USB Dithiothreitol (DTT) 0.1M Solution	707265ML	Thermo Fisher Scientific
RNaseOUT™ Recombinant Ribonuclease Inhibitor	10777019	Thermo Fisher Scientific
SuperScript™ IV First-Strand Synthesis System	18091050	Thermo Fisher Scientific
Bovine Serum Albumin (BSA)	B14	Thermo Fisher Scientific
Ribonuclease H (RNase H)	18021071	Thermo Fisher Scientific
DNA Polymerase I (10 U/μL)	EP0041	Thermo Fisher Scientific

BEFORE STARTING

Make sure your RNA is pure and contains no traces of DNA. A simple and very sensitive way to ensure that is to use the purified RNA as a template for a PCR reaction targeting a gene that should be present in the sample. A negative result indicates a lack of DNA template in the sample.

Primer annealing

1 Prepare the following mixture in a PCR tube:

- 1. $\square 1 \mu l$ to $\square 4 \mu l$ purified RNA ($\square 10 pg \square 5 \mu g$; usually $\square 200 ng$ for soil extract)
- 2. $\square 1 \mu l$ random hexamers (50 μM) or a gene-specific primer ($\square 2 \mu M$)

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3. 3. RNase free water



Nuclease-free autoclaved DEPCtreated water by Carl Roth Catalog #: T143.1 CAS Number: 7732-18-5

2 Mix gently and spin down the solution.

3 Incubate the mixture at &65 °C for &00:05:00 in a thermocycler and chill &0n ice (or in the cycler at > &4 °C) for at least &00:01:00.

Reverse transcription

- 4 Prepare the following mixture and add to each tube:
 - 1. **34 μl 5x Reaction buffer**
 - 2. **1 μl dNTP mix, 10 mM**
 - 3. **□1 μl 0.1M DTT**
 - 4. **□1 μl RNaseOUT™ (40 U/μl)** *
 - 5. **30.2 μl BSA (20 μg/μl)**
 - 6. □1 μl SuperScript™ IV RT (200 units/μl)
 - * Optional



 SuperScript™ IV First-Strand Synthesis
System
by Thermo Fisher Scientific
Catalog #: 18091050

RNaseOUT™ Recombinant
Ribonuclease Inhibitor
by Thermo Fisher Scientific
Catalog #: 10777019

Bovine Serum Albumin (BSA)
by Thermo Fisher Scientific
Catalog #: B14

- 5 Incubate the mixture in a thermocycler at § 23 °C for © 00:10:00 (only if using random hexamers, skip if using a specific primer) followed by § 50 °C for © 01:00:00 to © 03:00:00 and § 80 °C for © 00:10:00 . Chill § On ice .
- 6 For PCR templates > 1kb remove the RNA by adding $\Box 1 \mu I$ (2 units) of E. coli RNase H and incubate at 8 37 °C for $\odot 00:20:00$.
 - Ribonuclease H (RNase H)
 by Thermo Fisher Scientific
 Catalog #: 18021071

Optional: Second strand synthesis

- 7 Prepare the following mixture and add to each tube:
 - 1. **□1 μl DNA Polymerase I reaction buffer**
 - 2. **Q**0.75 μl DNA Polymerase I
 - 3. **Q0.2** μl RNase H
 - 4. 3.05 μl RNase-free water
 - 5. **3** μl Template cDNA
 - DNA Polymerase I (10 U/μL)
 by Thermo Fisher Scientific
 Catalog #: EP0041

- Ribonuclease H (RNase H)
 by Thermo Fisher Scientific
 Catalog #: 18021071
- Nuclease-free autoclaved DEPCtreated water
 by Carl Roth
 Catalog #: T143.1
 CAS Number: 7732-18-5
- 8 Incubate for at § 15 °C for © 02:00:00 followed by © 00:10:00 at § 75 °C for deactivation.
- 3. Purify the reaction through phenol/chloroform purification followed by ethanol precipitation or using a PCR purification kit.