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# PNGase F Protocol (Denaturing Conditions) V.2

New England Biolabs<sup>1</sup>

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[dx.doi.org/10.17504/protocols.io.bikhkct6](https://doi.org/10.17504/protocols.io.bikhkct6)

**New England Biolabs (NEB)**

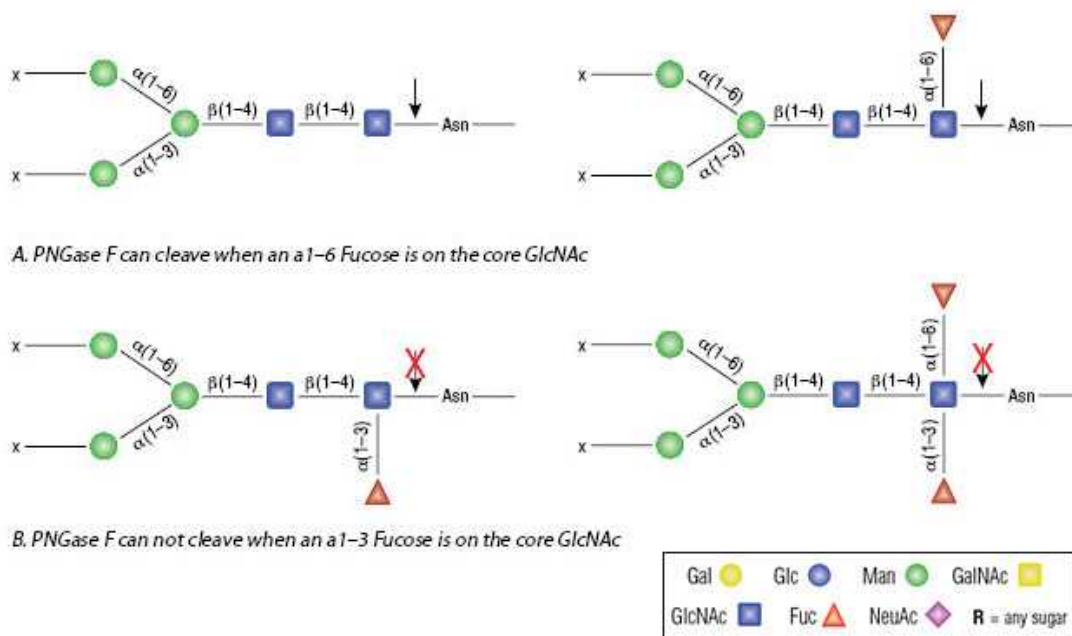
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PNGase F is the most effective enzymatic method for removing almost all *N*-linked oligosaccharides from glycoproteins. PNGase F is an amidase, which cleaves between the innermost GlcNAc and asparagine residues of high mannose, hybrid, and complex oligosaccharides.

This is a generic PNGase F protocol for **denaturing reaction conditions**. It is appropriate for both [P0704](#) and [P0708](#).



DOI

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<https://www.neb.com/protocols/2014/07/31/pngase-f-protocol>

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**protocols.io**

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Liz Brydon



Essentials of Glycobiology, glycoprotein, PNGase F is inhibited, deglycosylation, PNGase F denaturing reaction conditions, PNGase F non-denaturing reaction conditions, pngase f

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- If using [P0704/P0708](#), we recommend limiting PNGase F to 1/10 (or less) of the total reaction volume to keep the final glycerol concentration equal to (or less than) 5%.
- For unit conversion between different suppliers, please reference the [Glycobiology Unit Conversion Chart](#) page.

### Biology Unit Conversion Chart

Reagent companies differ in how a unit of enzyme is defined. This chart can be used to help determine how a unit of enzyme from one company compares to a unit of enzyme from NEB. All enzymes were assayed using NEB's assay protocols as a means of normalization (NEB Assay).

Enzyme	Company	Selling Conc. (U/ml)	Units/Vial	μl/Vial	NEB Assay (U/ml)	NEB Assay Units /Vial	μl Conversion (1 NEB μl = x Company μls)
PNGase F	NEB (NEB #P0704/P0705)	500,000	15,000	30	500,000	15,000	1
	Prozyme (GKE-5006A)	2.5	0.1	40	150,000	6,000	3.3
	Prozyme (GKE-5020B, Ultra)	10	0.4	40	500,000	20,000	1
	QA Bio (E-PNG01)	5	0.3	60	200,000	12,000	2.5
	Sigma (P7367)	500	50	50	90,000	4,500	5.5

### MATERIALS

[PNGase F \(native\) - 75,000 units New England](#)

**Biolabs Catalog #P0704L**

[PNGase F \(native\) - 15,000 units New England](#)

**Biolabs Catalog #P0704S**

[PNGase F Recombinant - 75,000 units New England](#)

**Biolabs Catalog #P0708L**

[PNGase F Recombinant - 15,000 units New England](#)

**Biolabs Catalog #P0708S**

Please refer to the Safety Data Sheets (SDS) for health and environmental hazards.

Reactions may be scaled-up linearly to accommodate larger amounts of glycoprotein and larger reaction volumes. Optimal incubation times may vary for particular substrates. Typical reaction conditions are as follows:

## Denaturing Reaction Conditions:

1 

Combine **1 µg** - **20 µg** glycoprotein ,  
**1 µL Glycoprotein Denaturing Buffer (10X)** and H<sub>2</sub>O (if necessary) to make a **10 µL**  
total reaction volume.

2 

Denature glycoprotein by heating reaction at **100 °C** for **00:10:00** .

3 

Chill denatured glycoprotein **On ice** and centrifuge **00:00:10** .

4 

Make a **total reaction volume** of **20 µL** by adding: **2 µL GlycoBuffer 2 (10X)** ,  
**2 µL 10% NP-40** and **6 µL H<sub>2</sub>O** .

*PNGase F is inhibited by SDS, therefore it is essential to have NP-40 in the reaction mixture under denaturing conditions. Failure to include NP-40 into the denaturing protocol will result in loss of enzymatic activity.*

5 

Add **1 µL PNGase F** and mix gently.

6 

Incubate reaction at **37 °C** for **01:00:00** .

7 

Analyze by method of choice.

*Note: The simplest method of assessing the extent of deglycosylation is by mobility shifts on SDS-PAGE gels.*