



Sample collection and eDNA extraction from Sterivex filter units V.2

COMMENTS 0

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VERSION 2

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WORKS FOR ME

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ABSTRACT

The following workflow covers several steps in the DNA analysis of environmental samples, from the water collection to the analysis back in the lab. The samples can be taken from several water systems (i.e. sea, lakes, rivers, streams) and collected in triplicate (1 L) in [Sterivex sterile filter units](#) (Merck, cat. no. SVGP01050). The DNA extraction protocol modifies the [Dneasy PowerWater Sterivex kit](#) (Qiagen, cat. no. 14600-50-nf).

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Sample collection

- 1 A portable peristaltic pump ([Vampire sampler](#); Buerkle) is used for sampling at an approximate 30-50 mL/min flow rate. The tubing inserted in the unit's head has a suction hose ([Marprene](#); 4.8 mm inner diameter), connected to a flexible [silicone hose](#). A suitable adapter connects both hoses.



- 2 A [Sterivex](#) filter unit (0.22 µm pore size) is attached to the suction hose by a stainless steel male [Luer-lock ring](#) hose barb (1/4"). We recommend using a [hose clip](#) (9-10 mm; Buerkle cat. no. BURK8678-000) to firmly tight the Luer-lock ring to the hose. Each filter can process up to 2L of water depending on the amount of the suspended material.



- 3 Once the water sample is filtered, remove the remaining liquid from the Sterivex unit using a 5 mL sterilized syringe or similar by pushing air.



- 4 Cap both ends of the Sterivex. If the units have a male tip outlet, Parafilm can be used to cap them. Be aware of potential leakage, and make sure the Sterivex is dried and well sealed.



- 5 Now that the samples are ready and sealed, do not forget to identify them accordingly. A good rule of thumb is to write down ID, date, replicate number, filtered volume and station on the filter. Do the same on the sample bags, especially if there are more filters per site or replicate. The samples are stored at **-20 °C** degrees.



DNA extraction

- 6 Prior to the experiment, prepare the following:
- Disinfect the surfaces and pipettes
 - Turn on waterbath and set it to 65°C
 - Turn on oven and set it to 90°C
 - Turn on heating block and set it to 37°C
 - Incubate MBL solution in waterbath
 - Attach vortex adapter to vortexer

Moreover, check if the Sterivex filter units are dry. If that is not the case, use a syringe to remove the remaining

water. In addition, seal the outlet tightly with Parafilm or replace the present Parafilm on the outlet with a new one, respectively.

Note: due to the limited space on the vortex adapter only 6 Sterivex filter units can be vortexed at the same time. However, in order to increase the number of samples to extract simultaneously, it is possible to prepare another 6 Sterivex filter units while the first batch is vortexing for 20min (step 8 and 9). Afterwards, the second batch of samples is vortexed. In the last 5 min of vortexing of the second batch, the first batch is incubated at 90°C in the oven (step 11). The following steps then allow to work on both batches simultaneously (work on one batch while the other is vortexing). Ultimately, the two batches can then be joined prior to step 15.

5m

- 7 Place the Sterivex vertically (with the inlet cap upward) and load Δ 900 μL of ST1B buffer. ST1B is stored at $\blacksquare 4^\circ\text{C}$. Be careful putting the pipette tip through the inlet (orange cap). Dispense the buffer slowly; a fraction of the volume can be lost.



8 Allow mixing in a vortex with a horizontal [adapter](#) (Qiagen, Cat. No. 13000-V1-5) for 00:05:00 at the minimum

5m

speed. Set the filters with the inlet facing out and check for potential leakage from the outlet sealed with Parafilm.



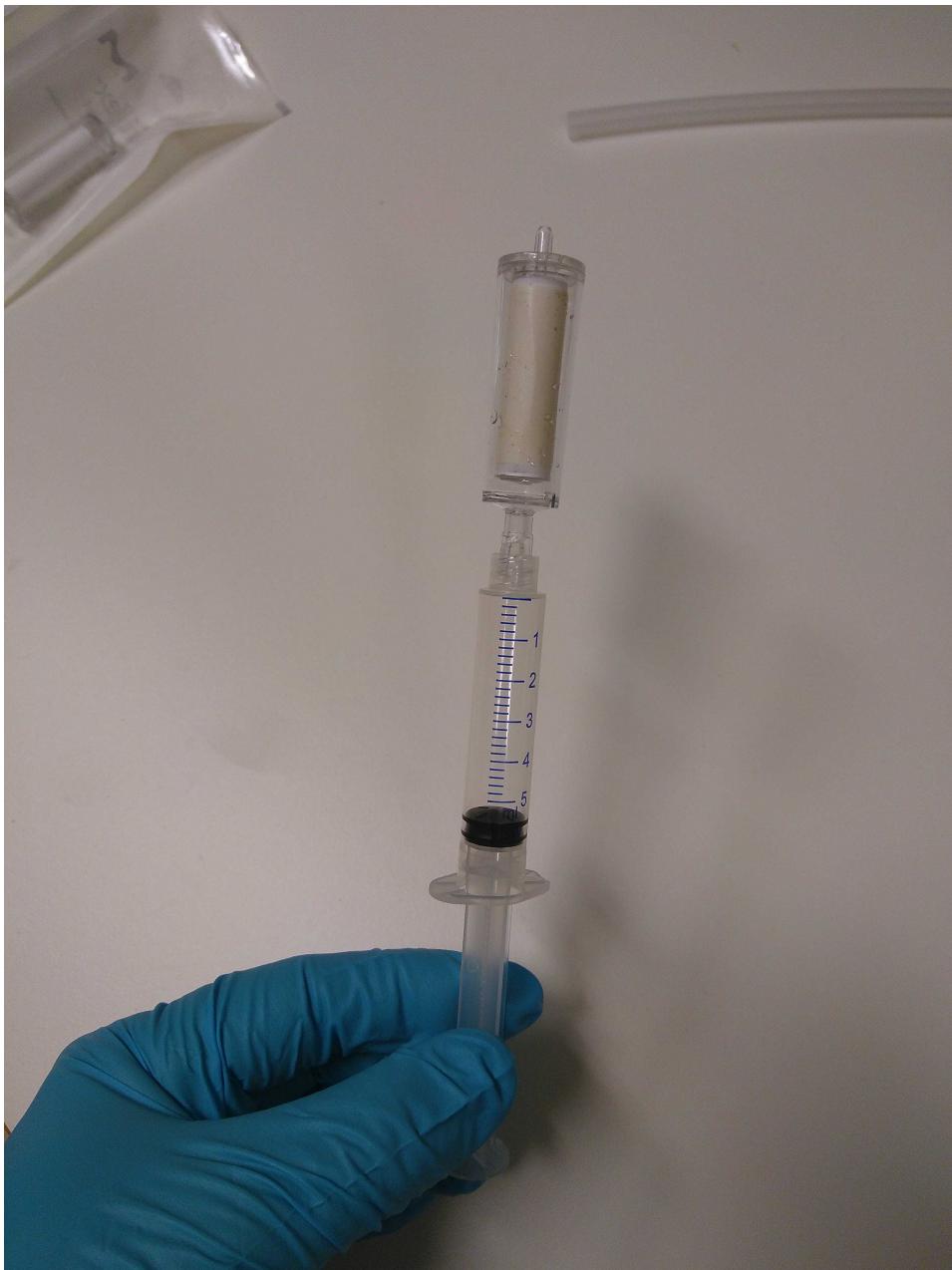
- 9 Rotate the Sterivex in 90 degrees and vortex for 00:05:00 at the minimum speed. Repeat the previous step 2 more times (4 times x 00:05:00 each filter).

15m

- 10 Add 900 µL of solution MBL. Dispense slowly; a fraction of the volume can be lost.
Before use, heat the MBL solution at 65°C for 00:10:00 as is suggested by the manufacturer.

10m

- 11 Incubate the filter units vertically with the inlet upward at $90\text{ }^{\circ}\text{C}$ for 00:05:00 in an oven. Before and after the incubation, check for any leakage in the Parafilm; replace it if needed. 5m
- 12 Let the filters cool down at room temperature for 00:02:00, and re-tight the caps and check the Parafilm. Then, vortex at maximum speed for 00:05:00. While mixing, check that the filters stay in place. If not, lower the speed. 7m
- 13 Transfer the lysate from the filter unit to a 3-5 mL syringe; push 1 mL of air into the filter while it is vertical, and then release the plunger. Continue pulling back until the lysate is recovered in the syringe. 6m



14 Pour the lysate into a **5 mL** Powerbeat tube and vortex horizontally for **00:05:00** at maximum speed.

5m



15 Centrifuge the tube at 4000 x g for 00:01:00 . 1m

16 Carefully transfer the lysate to a clean 2.2 mL collection tube. Then, add 1.6 µL of RNase (final concentration [M] 100 mg/mL). Incubate the samples at 37 °C x 00:30:00 in a heating block or a water bath. 35m

While the samples are incubating, prepare the following:

- Incubate MR solution in the waterbath at 65°C

- Prepare the VacValve vacuum system and connect it to the vacuum pump for later use

17 Add $\text{300 } \mu\text{L}$ IRS solution to the tube and vortex. Then incubate at 4°C for $00:05:00$.

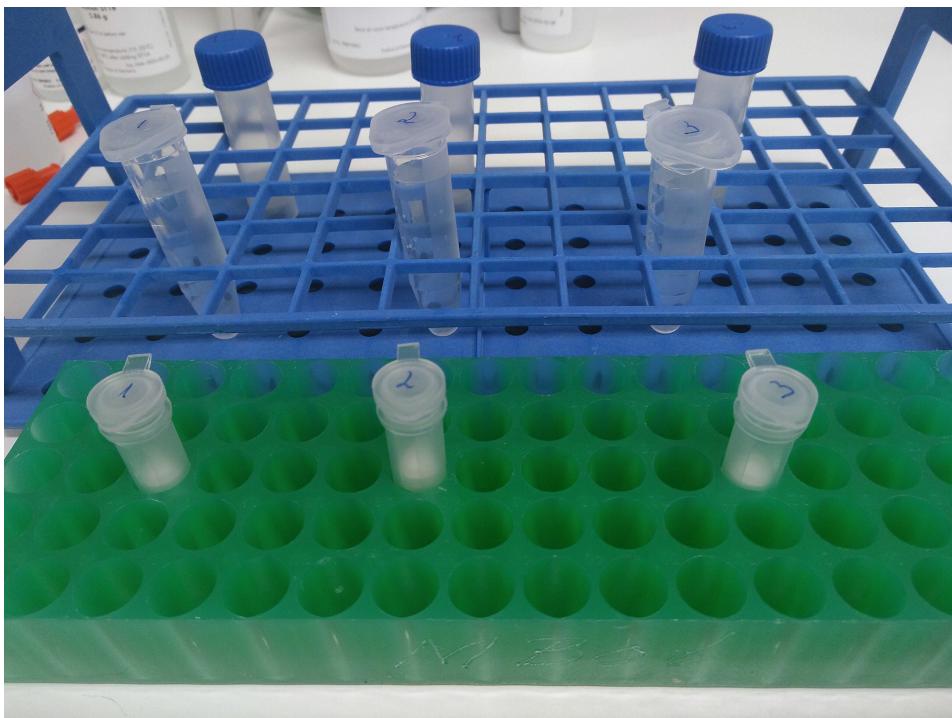
7m

18 Centrifuge the tube at $13000 \times g$ for $00:01:00$

1m

19 Transfer the supernatant to a 5 mL collection tube, avoiding the pellet, and add 3 mL of MR solution and vortex. Heat the MR solution at 65°C for $00:10:00$ before using it.

10m

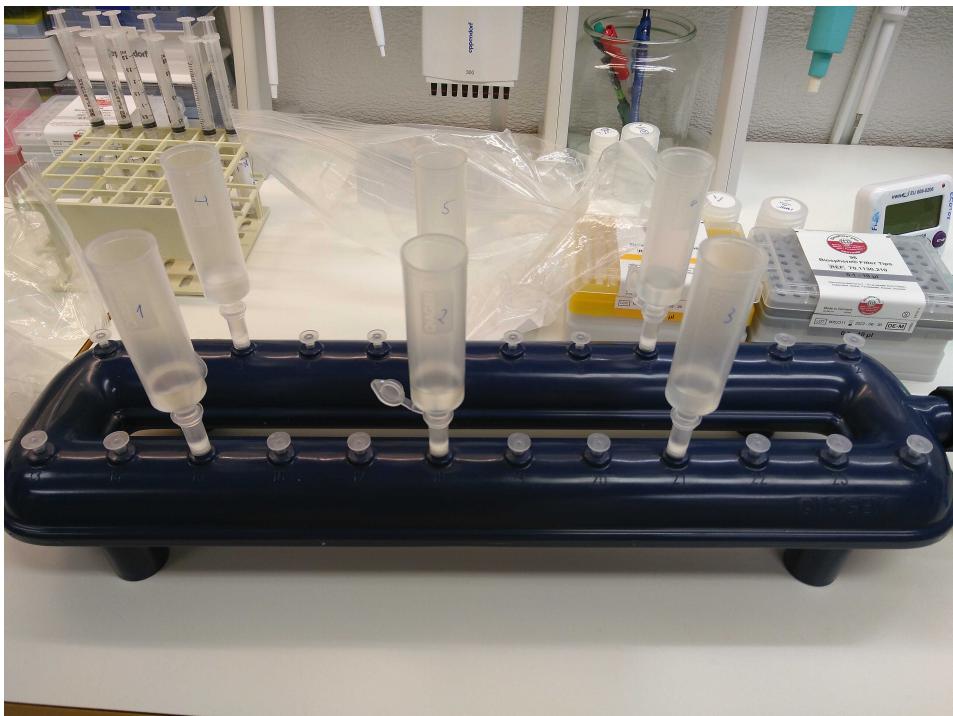


20 A [VacValve vacuum system](#) and a vacuum pump are used for the following steps.



- 21 Load the supernatant (4.5 mL) into a tube extender and MB spin column. Filtrate at low pressure. The column is attached to the [VacValve vacuum system](#) using an extra adapter to avoid contamination.

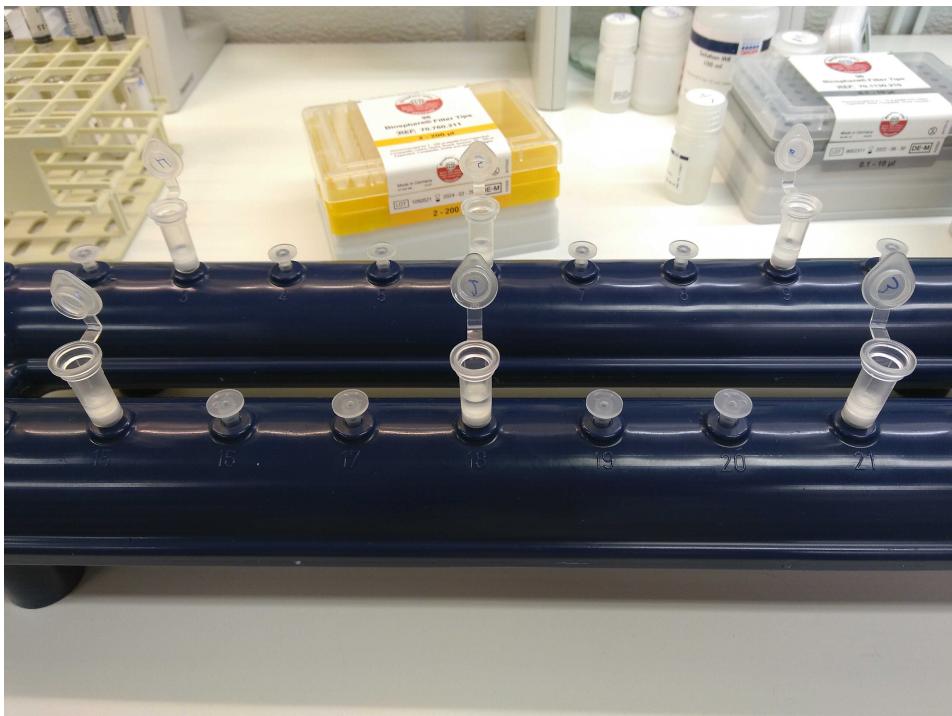
10m



- 22 Once the lysate passes through, the column extender is carefully removed. Next, wash the spin column with

5m

800 μ L of ethanol. Keep the columns open.



23 Shake the PW solution sufficiently prior to use. Then, wash the spin column with 800 μ L of PW solution.
Keep the columns open.
Allow the membrane to dry by keeping the vacuum pump running for 1 min.

5m

24 Wash the spin column again by adding 800 μ L of ethanol. Allow the membrane to dry as before.
Afterwards, close the lids, turn off the vacuum pump and open one port to get rid of the vacuum.

5m

25 Place the spin column in a clean collection tube and dry it out by centrifugation at 13000 x g for
 00:02:00

2m

26 Transfer the spin column to a new clean tube (1.5 mL safe-lock collection tube), and add 50-100 μ L of solution EB (or DNA-free grade water). Centrifuge at 13000 x g for 00:01:00 .
Transfer the eluate back onto the column at centrifuge a second time at 13000 x g for 00:01:00 .

5m

27 Voila! The DNA is in the collection tube, ready for further processing.
If DNA quality and yield is not checked immediately, store the samples at -20°C.

30m

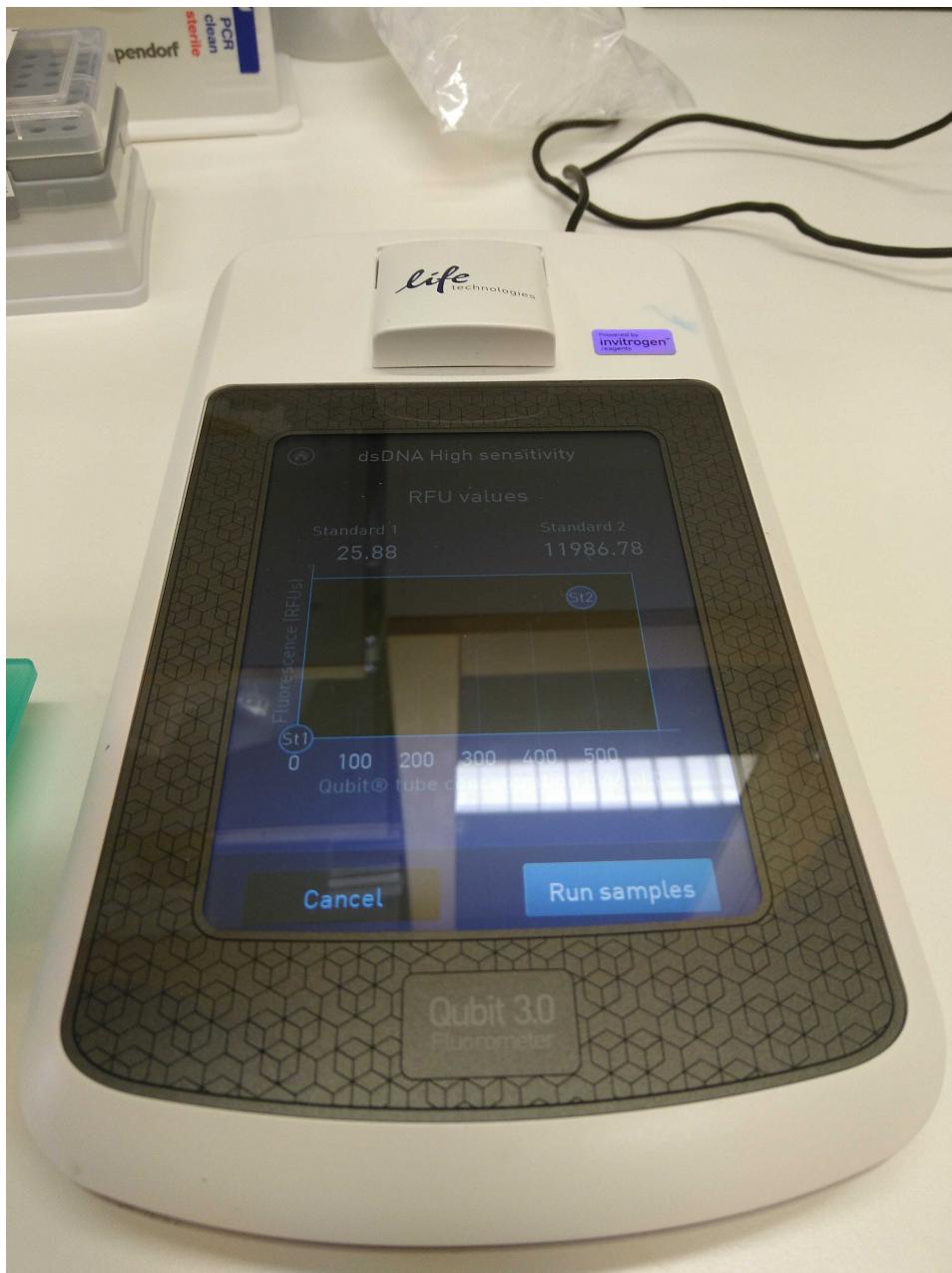
DNA quality check

10m

- 28 The DNA purity is checked with a Nanodrop. $\text{1 } \mu\text{L}$ of blank or sample is used for the measurement, respectively.

20m

- 29 The DNA concentration is quantified by Qubit. The preparation of standards and samples is following the Qubit instructions.



30 After measuring DNA quality and yield, store the DNA samples at -20°C for later use.