



Danielle Gutierrez¹, Jamie Allen¹, Zach Jenkins¹, Jeff Spraggins¹

¹Vanderbilt University

Works for me

dx.doi.org/10.17504/protocols.io.bs7hnhj6

Mar 10, 2021

VU Biomolecular Multimodal Imaging Center

Human BioMolecular Atlas Program (HuBMAP) Method Development Community

Vanderbilt University

SUBMIT TO PLOS ONE

ABSTRACT

Description of settings used to acquire LC-MS/MS data from label-free proteomic samples.

DOI

dx.doi.org/10.17504/protocols.io.bs7hnhj6

PROTOCOL CITATION

Danielle Gutierrez, Jamie Allen, Zach Jenkins, Jeff Spraggins 2021. LC-MS/MS Label-Free Proteomic Data Acquisition. protocols.io

https://dx.doi.org/10.17504/protocols.io.bs7hnhj6

Version created by Danielle Gutierrez

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Mar 10, 2021

LAST MODIFIED

Mar 10, 2021

OWNERSHIP HISTORY

Vanderbilt University Mar 10, 2021 Jamie Allen

PROTOCOL INTEGER ID

48073

DISCLAIMER:

DISCLAIMER - FOR INFORMATIONAL PURPOSES ONLY; USE AT YOUR OWN RISK

The protocol content here is for informational purposes only and does not constitute legal, medical, clinical, or safety advice, or otherwise; content added to protocols.io is not peer reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol should not substitute for independent professional judgment, advice, diagnosis, or treatment. Any action you take or refrain from taking using or relying upon the information presented here is strictly at your own risk. You agree that neither the Company nor any of the authors, contributors, administrators, or anyone else associated with protocols.io, can be held responsible for your use of the

mprotocols.io

03/10/2021

Citation: Danielle Gutierrez, Jamie Allen, Zach Jenkins, Jeff Spraggins (03/10/2021). LC-MS/MS Label-Free Proteomic Data Acquisition. https://dx.doi.org/10.17504/protocols.io.bs7hnhj6

information contained in or linked to this protocol or any of our Sites/Apps and Services.

- 1 Label-free proteomic samples were analyzed on a Thermo Scientific Orbitrap Fusion Tribrid mass spectrometer in line with an EvoSep One HPLC system.
- 2 Samples,1 μL at a 1 μg/μL, were loaded onto EvoTips and diluted with 19 μL of mobile phase A (100% water, 0.1% formic acid). Samples were processed on tip according to the manufacturer's recommended protocol. EvoTips were placed onto the EvoSep One, and samples were eluted from the EvoTips onto a Dr. Maisch C18 AQ, 100 micron inner diameter, 15 cm long column with 3 micron particles (EV1074) and analyzed using the 30 sample per day method settings.
- 3 Peptides were separated at a flow rate of 500nL/min over a 44 minute gradient according to the 30 samples/day settings. Mobile phase A was 100% water, 0.1% formic acid, and mobile phase B was 100% acetonitrile, 0.1% formic acid.
- 4 Eluted peptides were ionized via positive mode nanoelectrospray ionization (nESI) using a Nanospray Flex ion source (Thermo Fisher Scientific).
- 5 The mass spectrometer was operated using a top 12 data-dependent acquisition mode.
- Fourier transform mass spectra (FTMS) were collected using 60,000 resolving power, an automated gain control (AGC) target of $1e^6$, and a maximum injection time of 50 ms over the mass range of 375-1500 m/z.
- Precursor ions were filtered using monoisotopic precursor selection of peptide ions with charge states ranging from 2 to 6. Previously interrogated precursor ions were excluded using a 30 s dynamic window (± 10 ppm).
- Precursor ions for tandem mass spectrometry (MS/MS) analysis were isolated using a 1.2 m/z quadrupole mass filter window and then fragmented in the ion-routing multipole via higher energy dissociation (HCD) using a normalized collision energy of 35%.
- 9 Ion trap fragmentation spectra were acquired using an AGC target of 10,000 and maximum injection time of 60 ms, and 120 *m/z* was set for the first scan mass to enable detection of the lysine residue fragmented ion.