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Version 2

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# Amplicon clean-up using SPRI beads for RAPID nanopore kit RBK004 V.2

Forked from [Amplicon clean-up using SPRI beads](#)

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1 Works for me This protocol may be deleted by the owner

Coronavirus Method Development Community

Nikki Freed

## MATERIALS

NAME	CATALOG #	VENDOR
<a href="#">Agencourt AMPure XP beads</a>		

## STEPS MATERIALS

NAME	CATALOG #	VENDOR
<a href="#">Agencourt AMPure XP</a>	A63880	<a href="#">Beckman Coulter</a>
<a href="#">QuantiFluor(R) ONE dsDNA System, 100rxn</a>	E4871	<a href="#">Promega</a>

## MATERIALS TEXT

Freshly prepared 80% ethanol  
10 mM Tris-HCl pH 8.0 with 50 mM NaCl

## EQUIPMENT

NAME	CATALOG #	VENDOR
<a href="#">Quantus</a>	E6150	

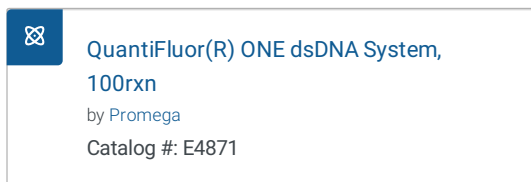
### Ampure XP bead clean up

- 1 Vortex SPRI beads thoroughly to ensure they are well resuspended, the solution should be a homogenous brown colour.

**Agencourt AMPure XP**  
by [Beckman Coulter](#)  
Catalog #: [A63880](#)

- 2 Add an equal volume (1:1) of SPRI beads to the sample tube and mix gently by either flicking or pipetting. For example add **50 µl** room temperature SPRI beads to a **50 µl** reaction.
- 3 Pulse centrifuge to collect all liquid at the bottom of the tube.
- 4 Incubate for **00:05:00** at room temperature.

- 5 Place on magnetic rack and incubate for 🕒 **00:02:00** or until the beads have pelleted and the supernatant is completely clear.
- 6 Carefully remove and discard the supernatant, being careful not to touch the bead pellet.
- 7 Add 🧴 **200 µl** of freshly prepared room-temperature **80 % volume** ethanol to the pellet.
- 8 Keeping the magnetic rack on the benchtop, rotate the bead-containing tube by 180°. Wait for the beads to migrate towards the magnet and re-form a pellet. Remove the ethanol using a pipette and discard.
- 9 🔄 and repeat ethanol wash.
- 10 Pulse centrifuge to collect all liquid at the bottom of the tube and carefully remove as much residual ethanol as possible using a P10 pipette.
- 11 With the tube lid open incubate for 🕒 **00:01:00** or until the pellet loses its shine (if the pellet dries completely it will crack and become difficult to resuspend).
- 12 Remove the tube from the magnetic rack. Resuspend pellet in 🧴 **10 µl** 10 mM Tris-HCl pH 8.0 with 50 mM NaCl, mix gently by flicking and incubate for 🕒 **00:02:00**.
- 13 Place on magnet and transfer sample to a clean 1.5mL Eppendorf tube ensuring no beads are transferred into this tube.
- 14 Quantify 🧴 **1 µl** product using the Quantus Fluorometer using the ONE dsDNA assay.





Quantus  
Fluorometer

Promega E6150 [↗](#)