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Regulation of mitophagy by the NSL complex underlies genetic risk for Parkinson's disease: Drosophila Husbandry, Assays and Immunohistochemistry

Natalie J. Welsh¹, Alexander J. Whitworth¹

¹MRC Mitochondrial Biology Unit, University of Cambridge, Cambridge, UK



Benjamin O'Callaghan

ABSTRACT

This protocol describes Drosophila stocks and husbandry, Locomotor and lifespan assays and, Immunohistochemistry and sample preparation in order to assess the role of KAT8/KANSL1 in neuronal survival *in vivo*.

MATERIALS TEXT

Drosophila Strains

Bloomington *Drosophila* Stock Center (RRID:SCR_006457): *mof* RNAi lines, P{TRiP.JF01701} (RRID:BDSC_31401); and P{TRiP.HMS00537} (RRID:BDSC_58281); *nsl1* RNAi lines, P{TRiP.HMJ22458} (RRID:BDSC_58328); the pan-neuronal *nSyb-GAL4* driver (RRID:BDSC_51941); and dopaminergic neuron driver (TH-GAL4; RRID:BDSC_8848); and control (*lac2*) RNAi P{GD936}v51446) from the Vienna *Drosophila* Resource Center (RRID:SCR_013805).

Medium

Agar, cornmeal, molasses, propionic acid and yeast

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Drosophila stocks and husbandry

Raise flies under standard conditions in a humidified, temperature-controlled incubator with a 12h:12h light:dark cycle at \$\ 25 \circ \), on food consisting of agar, cornmeal, molasses, propionic acid and yeast.

Note

The following strains were obtained from the Bloomington *Drosophila* Stock Center (RRID:SCR_006457): *mof* RNAi lines, P{TRiP.JF01701} (RRID:BDSC_31401); and P{TRiP.HMS00537} (RRID:BDSC_58281); *nsl1* RNAi lines, P{TRiP.HMJ22458} (RRID:BDSC_58328); the pan-neuronal *nSyb-GAL4* driver (RRID:BDSC_51941); and dopaminergic neuron driver (TH-GAL4; RRID:BDSC_8848); and control (*lacZ*) RNAi P{GD936}v51446) from the Vienna *Drosophila* Resource Center (RRID:SCR_013805).

All experiments are conducted using male flies.

Locomotor Assay

- Perform the startle induced negative geotaxis (climbing) assay using a counter-current apparatus.
- Place 20-23 males into the first chamber, and tap to the bottom.
 Give 10 s to climb a 10 cm distance.
 Repeat this procedure five times (five chambers), and count the number of flies that remain in each chamber.
- 4 Normalize the weighted performance of several group of flies for each genotype to the maximum possible score and expressed as *Climbing index* as described previously in the manuscript by <u>Greene et al., 2003</u>.

Note

Reference:

Greene JC, Whitworth AJ, Kuo I, Andrews LA, Feany MB, Pallanck LJ. Mitochondrial pathology and

apoptotic muscle degeneration in Drosophila parkin mutants. *Proc Natl Acad Sci.* 2003;100(7):4078 LP - 4083. doi:10.1073/pnas.0737556100

Lifespan Assay

- For lifespan experiments, grow flies at low-density under identical conditions as described in Step 1

 5 go to step #1

 .
- 6 Collect progeny under very light anaesthesia and keep in tubes of approximately 20 males each, around 50-100 in total.
- 7 Transfer flies every 2-3 days to fresh medium and record the number of dead flies.
- 8 Calculate percent survival at the end of the experiment after correcting for any accidental loss.

Sample Preparation and Immunohistochemistry

3d 5h 30m

9 Dissect *drosophila* brains from aged flies and immunostain as previously described in the manuscript by Whitworth et al., 2005.

Note

Reference:

Whitworth AJ, Theodore DA, Greene JC, Beneš H, Wes PD, Pallanck LJ. Increased glutathione *S*-transferase activity rescues dopaminergic neuron loss in a *Drosophila* model of Parkinson's disease. *Proc Natl Acad Sci.* 2005;102(22):8024 LP - 8029. doi:10.1073/pnas.0501078102

 30m

Image brains with a Zeiss LMS 880 confocal. Count tyrosine hydroxylase-positive neurons under blinded

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conditions.