Total nucleic acid extraction - Maxwell(R) HT Environmental TNA Kit, custom (Promega) V.4

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In Development Share

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ABSTRACT

Total nucleic acid extraction from wastewater using Maxwell(R) HT Environmental TNA Kit, custom (Promega)

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PARENT PROTOCOLS

In steps of

Concentration of viruses from sewage using HA filters

GUIDELINES

When work is completed, remove equipment and supplies from the cabinet. Wipe the work area with 10% bleach, let stand for 10 min, rinse with water, then with 70% ethanol, and finally with RNAase AWAY.

MATERIALS TEXT

MATERIALS
•Ethanol USP/ACS or molecular biology grade (100%)
•Molecular biology grade water
•**Isopropanol** molecular biology grade (100%)
•**6x** KingFisher **96-well plates** (Cat. no.: 95040460)
•**1x** KingFisher **96-tip comb well plate**(Cat. no.: 97002534)
•**Screw cap** microcentrifuge tubes

Mini-Beadbeater-16
high-energy cell disrupter
BioSpec **607**
1 speed

Kingfisher Flex
Automated Extraction System
ThermoFisher **5400630**

Promega_Maxwell_HT_RNA_Wastewater_V1.bdz

DISCLAIMER:

BEFORE STARTING

1. Clean the working area and all equipment: wipe down with 10% bleach and let dry. Wipe down with 70% ethanol and let dry. Then, wipe down using RNase AWAY and let dry.

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2. Prepare the 50% ethanol solution (it must be fresh!)

3. Prepare the 6 purification plates:
- **Wash 1 plate**: Add 100 µl of 50% ethanol and 900 µl of wash buffer (WBA) to each well required for purification.
- **Wash 2 plate** (same as plate Wash 1): Add 100 µl of 50% ethanol and 900 µl of wash buffer (WBA) to each well required for purification.
- **Ethanol Wash plate**: Add 450 µl of 50% ethanol to each well required for purification.
- **Elution plate**: Add 100 µl of 25 mM Tris-HCl (pH 8.0) to each well required for purification.
- **Tip plate**: Place KingFisher 96-tip comb into an empty KingFisher 96-well plate. *While opening the 96-tip comb plate, pay attention to not touch the tips.*
- **Lysis and Bind plate**: Add 35 µL of Resin to each well required for purification (v*ortex the bottle at max speed before use)*. Add 50 µl of Alkaline Protease Solution custom (APA) to each well required for purification. Add 250 µl of cell lysis solution (CLD) to each well required for purification. Add 400 µl of Isopropanol (100%) to each well required for purification.

Total nucleic acid extraction 2h

1 5m

For **HA filter** extraction, let the sample thaw on ice and go to **step 2**.

For **BCoV/BRSV** extraction (in duplicate), add 5 µL of BCoV/BRSV solution to the 2-mL tube containing 250 µL CTAB. Vortex for 15 seconds (speed 7 out of 10) and flash freeze the tube. Go to step 4.

For **Direct extraction**, add 150 µL of wastewater to the 2-mL tube containing 100 µL CTAB. Vortex for 15 seconds (speed 7 out of 10) and flash freeze the tube. Go to step 4.

2

For **HA filter** extraction, place the 2-mL tubes in the bead beater.

Mini-Beadbeater-16
high-energy cell disrupter
BioSpec **607**
1 speed

2.1 2m 30s

Bead beat for **00:02:30**

Start the bead beating when the beads start to be loose in the tubes.

2.2 5m

Cooldown the samples on ice for **00:05:00** .

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2.3 Repeat Steps 9.1 and 9.2 once .

7m 30s

3 Centrifuge at maximum speed for 1 min at room temperature. **150000 rpm, Room temperature , 00:01:00**

1m

4 For **HA filter** extraction, transfer 125-250 µL of supernatant to the **Lysis/Bind plate**.
For **BCoV/BRSV/Direct extraction**, transfer all supernatant to the **Lysis/Bind plate**.

10m

The default volume transferred is 250 µL. However, for WWTPs with "dirty" influents, we only transfer 125 µL.

5 Start the protocol Promega_Maxwell_HT_RNA_Wastewater_V1.bdz on the KingFisher Flex **01:14:00**

1h 14m

Kingfisher Flex
Automated Extraction System
ThermoFisher **5400630**

6 Transfer the purified sample from the **Elution plate** to the **microcentrifuge tubes**.

10m

The DNA/RNA is now ready for downstream applications. RNA extract may be stored in RNase-free water at -80ºC for 1 year.

RT-ddPCR

7 Quantification by Droplet Digital PCR (ddPCR)

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