

Size selection (Purification)

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ABSTRACT

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Protocol status: Working We use this protocol and it's working

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1 Prepare \bot 20 μ L sample and \bot 9 μ L magnetic beads in 1.5 mL eppendorf tube.

A	В	С	D

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Component	Volume	Proportion	Note
Sample	20 μΙ	1x	
Magnetic beads	9 μΙ	0.45x	BeaverBeads™ DNA Select Isolation

2	Mix sample and beads gently by flicking then flash spin the tube. Put on the regular rack for
	5mins (5) 00:05:00 .

5m

Note

DON'T put the tube on magnetic rack during waiting in this step.

- 3 Transfer the tube to the magnetic rack. After most of the magnetic beads attach to the wall, remove the supernant.
- 4 Add Δ 300 μL 75% ethanol, flip whole magnetic rack around. Wait for 3 mins 00:03:00 and remove the supernant.

3m

- 5 Repeat step 4.
- Quick spin the tube, and put on the magnetic rack. Remove superfluous solution with 10 μ l pipette.

Note

Caution: DO NOT let the beads crack!

7 Add \perp 10 μ L elution buffer. Mix gently by flicking and flash spin the tube. Put on the regular rack for 10mins 00:10:00.

10m

Note

DON'T put the tube on magnetic rack during waiting in this step.

- 8 Transfer the tube to the magnetic rack. After most of the magnetic beads attach to the wall, collect the supernant to 200 µl PCR tube or 8-strip PCR tube.
- 9 Ready for 2' PCR.