



Nov 02, 2022

# Macherey-Nagel Nucleospin 96 Food protocol for bee pollen

Lauren Ponisio<sup>1</sup>, Jocelyn Zorn<sup>1</sup><sup>1</sup>University of Oregon1 *Works for me* Share[dx.doi.org/10.17504/protocols.io.kxygxpbro18j/v1](https://dx.doi.org/10.17504/protocols.io.kxygxpbro18j/v1)**FFAR** **Ponisiolab** Jocelynz

## ABSTRACT

Macherey-Nagel Nucleospin 96 Food protocol for bee pollen

DOI

[dx.doi.org/10.17504/protocols.io.kxygxpbro18j/v1](https://dx.doi.org/10.17504/protocols.io.kxygxpbro18j/v1)

## PROTOCOL CITATION

Lauren Ponisio, Jocelyn Zorn 2022. Macherey-Nagel Nucleospin 96 Food protocol for bee pollen. **protocols.io**  
<https://dx.doi.org/10.17504/protocols.io.kxygxpbro18j/v1>

## FUNDERS ACKNOWLEDGEMENT

Foundation for Food and Agricultural Research  
Grant ID: CA18-SS-0000000009

## LICENSE

————— This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

## CREATED

Sep 03, 2021

## LAST MODIFIED

Nov 02, 2022

- 1 UV sterilize supplies for 2 96 well plates worth of extractions: 4 50mL centrifuge tubes, 2 15mL centrifuge tubes, zirconia beads, 2 96 deep well plates and clear strip caps, 2 s-blocks, 2 96 well elution plates, 14 1000uL tip boxes, 2 200uL tip boxes, 2 10uL tip boxes, 6 reagent troughs, and 6 96 well microplates

- 2 [Lysis Buffer CF Macherey](#)

Aliquot [Nagel Catalog #740946](#) into sterile centrifuge tube and warm in **65 °C** water bath. You will need to aliquot **30 mL** per 1/2 plate (48 samples)

- 3 [Proteinase K Macherey](#)

Add **540 µL** [Nagel Catalog #740506](#) to each warmed **30 mL**

[Lysis Buffer CF Macherey](#)  
[Nagel Catalog #740946](#)

aliquot and invert gently to mix

- 4 [Lysis Buffer CF Macherey](#)

Add **560 µL** [Nagel Catalog #740946](#) +

[Proteinase K Macherey](#)  
[Nagel Catalog #740506](#)

solution to each sample

Each sample will receive **550 µL**

[Lysis Buffer CF Macherey](#)  
[Nagel Catalog #740946](#)

[Proteinase K Macherey](#)  
and **10 µL** [Nagel Catalog #740506](#)

- 5 Place in tissue lyser and run at **10 Hz** for **00:01:00**

1m

Make sure pollen ball is removed from bee leg after lysing. If still attached, repeat this step, increasing Hz if needed

TissueLyser II  
Bead Mill

QIAGEN 85300 [🔗](#)

## 6 Centrifuge 🌀 **3220 x g, 00:01:00**

1m

Eppendorf™ 5810R Centrifuge  
Centrifuge

Eppendorf 02-262-8187 [🔗](#)

## 7 Using sterile tweezers, remove bee leg, rinse with 200 proof ethanol, and place leg into labeled sterile microcentrifuge tube. Sterilize tweezers between each use with flame. Leave leg tubes open in fume hood until remaining ethanol has evaporated, then store at 🌡 **-80 °C**

If extracting pollen not attached to bee leg, skip steps 5-7

## 8 Add ~ 📄 **100 µL** zirconia beads to each pollen sample

## 9 Place in tissue lyser and run at 📄 **24 Hz** for 🕒 **00:01:30** , rotate plates 180 degrees, and lyse again at 📄 **24 Hz** for 🕒 **00:01:30**

TissueLyser II

Bead Mill

QIAGEN

85300



10 Place in **65 °C** water bath for **00:30:00**

30m

Incubation time may be increased up to overnight if extraction of DNA from pollen during lysis was not sufficient

11 Centrifuge **3220 x g, 00:35:00**

35m

12 Transfer **300 µL** of supernatant into 96 deep well plate

Samples may be stored at **-20 °C** after this step

13 Add **300 µL** (or equal volume)

**Binding Buffer C4 Macherey**

**Nagel Catalog #740366.250**

and **300 µL** (or equal volume) 200 proof ethanol

**Binding Buffer C4 Macherey**

**Nagel Catalog #740366.250**

combined ahead of time. Check note on reagent bottle. If already combined, add

**Binding Buffer C4 Macherey**

**600 µL** **Nagel Catalog #740366.250**

and ethanol may be

/EtOH

 Binding Buffer C4 Macherey

solution. **Nagel Catalog #740366.250**

with EtOH

added may be stored at room temperature for up to 1 month



 Binding Buffer C4 Macherey

**Nagel Catalog #740366.250**

contains

guanidine salt - do not mix with bleach

14 Vortex samples until thoroughly combined

15 Centrifuge  **1500 x g** for  **00:00:30**




30s

Do not centrifuge at a higher g-force or for a longer duration - this will precipitate out DNA

16 Place food binding plate onto s-block and transfer sample to food binding plate. Seal with gas-permeable foil

17 Centrifuge  **3220 x g, 00:09:00** , discard flowthrough

9m

18 While centrifuging, aliquot out  **12 mL** per plate of  **Elution Buffer CE Macherey Nagel** and place in  **70 °C** water bath

19

 Wash Buffer CQW MachereyAdd  500 µL Nagel Catalog #740313.125

and seal

 Wash Buffer CQW Macherey

Nagel Catalog #740313.125

contains

guanidine salt - do not mix with bleach

20

Centrifuge  3220 x g, 00:04:00 , discard flowthrough

4m

21

 Wash Buffer C5 MachereyAdd  900 µL Nagel Catalog #740931

, do not seal

22

Centrifuge  3220 x g, 00:20:00 , discard flowthrough

20m

23

Incubate binding plate  37 °C for  00:30:00

30m

24

Place binding plate over elution block. Add  100 µL pre-heated Elution Buffer CE Macherey Nagel

directly onto center of each binding plate filter membrane

25

Incubate room temperature  00:05:00

5m

26

Centrifuge  3220 x g, 00:04:00

4m

27 Aliquot entirety of product into 3 96 well microplates per DNA plate

28 Store at  $-20^{\circ}\text{C}$  (short term) to  $-80^{\circ}\text{C}$  (long term)