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Preparation and use of 12-well plates for the rapid detection of terbinafine-resistant dermatophytes

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This protocol describes the steps necessary for the preparation and use of screening plates for the rapid detection of terbinafine resistant dermatophytes.

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- 1) When you take the medium out of the autoclave, be as quick as possible to add the terbinafine and transfer the medium to the plate wells. Since the medium contains agar, it will solidify fairly quickly.
- 2) Work in sterile conditions as much as possible. The medium is very easy to contaminate and can, if so, falsify the results.

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Terbinafine preparation

13m

Weigh 16.33 mg of

Terbinafine hydrochloride Merck Millipore

Sigma Catalog #T8826

and add to an erlenmeyer flask containing 5 mL of

Dimethyl sulfoxide Merck Millipore

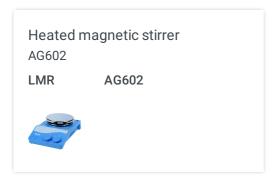
Sigma Catalog #D4540

. Mix without heating for 00:10:00 to dissolve. The final concentration of this terbinafine preparation is

3200 µg/mL . Do not forget to add a magnetic bar. Cover each flask with glass wool and aluminium foil.



2

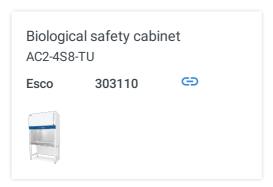


2 Autoclave this flask at § 121 °C for © 00:30:00





3 Under hood, transfer □1 mL of this preparation into 2 mL eppendorf tubes and store at 8 -80 °C for later use.



Medium preparation 4h 25m

4 Dissolve ■30 g of

Sigma Catalog #S3306

Medium preparation 4h 25m

8m

8m

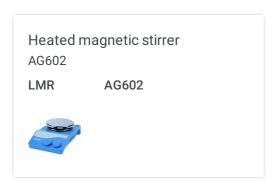
1 Sigma Catalog #S3306

in ■1 L of

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3

MilliQ water Contributed by users and let mix on the heated magnetic stirrer for
 00:05:00 (temperature and mixing speed knob at mid-step)





6 Pour **250 mL** of this medium into 4 erlenmeyer flasks and label each with final terbinafine concentrations (0.2 μg/mL for the first flask, 0.1 μg/mL for the second, 0.05 μg/mL for the third and 0 μg/mL for the last flask). Given the uncertainty of measurement, it is advisable to work from the highest concentration to the lowest. Do not forget to add a magnetic bar. Cover each flask with glass wool and aluminium foil.

4h

7 Autoclave these 4 flasks at 8 121 °C for © 00:30:00



Plates preparation 1h 11m

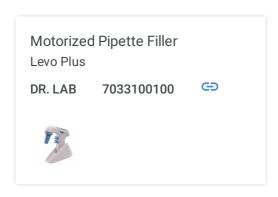
8 For the first flask (0.2 μ g/mL): add \Box 15.6 μ L of a 3200 μ g/mL terbinafine preparation (see

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4

section number 1). Let mix while heating (temperature and mixing speed knob at mid-step) for © **00:02:00** . Caution : do not leave for more than 2 minutes otherwise the medium will solidify.

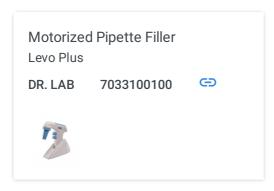
- 9 For the second flask (0.1 μg/mL): add ¬7.8 μL of a 3200 μg/mL terbinafine preparation (see section number 1). Let mix while heating (temperature and mixing speed knob at mid-step) for © 00:02:00 . Caution: do not leave for more than 2 minutes otherwise the medium will solidify.
- For the third flask (0.05 μg/mL): add 3.9 μL of a 3200 μg/mL terbinafine preparation (see section number 1). Let mix while heating (temperature and mixing speed knob at mid-step) for 00:02:00. Caution: do not leave for more than 2 minutes otherwise the medium will solidify.
- 11 For the last flask (0 μ g/mL): does not contain terbinafine.
- 12 In a 12-well plate (3 rows and 4 columns), the first well of each row does not contain terbinafine. It serves as a growth control. Using an automated pipettor, add **3 mL** of the medium that does not contains terbinafine (flask number 4) to all the first wells of the plate.

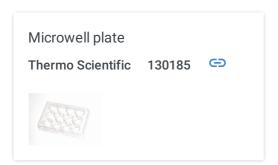


Microwell plate

Thermo Scientific 130185 🖘

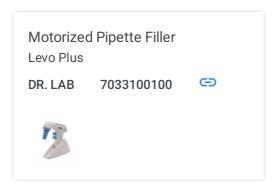
13 The second wells in each row contain a concentration of 0.05 μg/mL of terbinafine. Using an automated pipettor, add **3 mL** of the medium that contains **0.05 μg/mL** of terbinafine (flask number 3) to all the second wells of the plate.



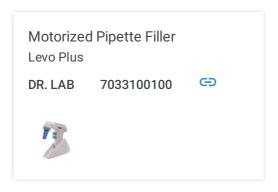


14 The third wells in each row contain a concentration of 0.1 μ g/mL of terbinafine. Using an automated pipettor, add $\square 3$ mL of the medium that contains $\square 0.1 \mu$ g/mL of terbinafine (flask number 2) to all the third wells of the plate.





15 The last wells in each row contain a concentration of 0.2 μg/mL of terbinafine. Using an automated pipettor, add **3 mL** of the medium that contains **0.2 μg/mL** of terbinafine (flask number 1) to all the last wells of the plate.





16 Cover the plates with their lids and let the medium solidify at room temperature. Label the plates with the corresponding terbinafine concentrations for each well and store at § 4 °C for future use.

Use of the plates

17 Using a sterile swab, prepare a suspension (sterile water) 0.5 McF of the dermatophyte to be



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studied.

- 18 Transfer 25μ L of this suspension to each of the 4 wells of the plate.
- Annotate the plate with the number of the studied strains and incubate at 30°C for 96h before reading the result.