



FEB 16, 2024

# DAB Staining for GFP on Free-floating Fixed NHP Brain Tissue

Forked from [Standard DAB Staining for Free-floating Fixed NHP Brain Tissue](#)

Aydin Alikaya<sup>1</sup>, William Stauffer<sup>1</sup>

<sup>1</sup>Department of Neurobiology, University of Pittsburgh, Pittsburgh, PA, USA



Andreea C. Bostan

## ABSTRACT

This protocol details the procedure for immunohistochemical 3,3'-Diaminobenzidine (DAB) staining of free-floating fixed brain tissue sections using the avidin/biotin ABC complex to detect Green Fluorescence Protein (GFP).

This protocol has been tested with free-floating non-human primate (NHP) brain tissue that has been fixed with 4% paraformaldehyde, cryoprotected with sucrose gradients, and cryo-sectioned  $\pm 50 \mu\text{m}$ .

## GUIDELINES

When using 6 well tissue culture plates [Falcon, 353046] to react individual sections, you will need **2+ mL** solutions for **each** well plate.

When using circular staining nets [e.g., Brain Research Laboratories #4115] to react multiple series of sections, you will need **50 mL** solutions for **each** staining net.

OPEN ACCESS



DOI:

[dx.doi.org/10.17504/protocols.io.eq2lyjy4mlx9/v1](https://dx.doi.org/10.17504/protocols.io.eq2lyjy4mlx9/v1)

**Protocol Citation:** Aydin Alikaya, William Stauffer 2024. DAB Staining for GFP on Free-floating Fixed NHP Brain Tissue .

protocols.io

<https://dx.doi.org/10.17504/protocols.io.eq2lyjy4mlx9/v1>

**License:** This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

**Protocol status:** Working

We use this protocol and it's working

**Created:** Feb 07, 2024

Last Modified: Feb 16, 2024

PROTOCOL integer ID: 94871

**Keywords:** ASAPCRN, Immunostaining, DAB, NHP Brain Tissue

**Funders Acknowledgement:**  
Aligning Science Across Parkinson's  
Grant ID: ASAP-020519

MATERIALS

**Tissue:**  
NHP brain tissue sections (50 µm).

- Materials/Equipment:**
- Tissue culture plates or circular staining nets
  - Orbital shaker
  - Fume hood
  - Nitrile Gloves
  - Glass slides (charged or subbed)




- Reagents:**
- Phosphate-buffered saline (PBS)
  - Hydrogen Peroxide: H<sub>2</sub>O<sub>2</sub> (30%)
  - Distilled water: dH<sub>2</sub>O
  - Primary Antibody: Thermo Fisher, Molecular Probes Cat# A11122, RRID: AB\_221569
  - Vectastain ABC-HRP Kit, Peroxidase (Rabbit IgG) (PK-4001, Vector Laboratories)
  - Peroxidase (HRP) with Nickel (3,3'-diaminobenzidine) (SK-4100) (Vector Laboratories)















SAFETY WARNINGS

⚠ Use appropriate care when using hydrogen peroxide (reactive, can cause skin/eye damage) and DAB (suspected carcinogen). Collect DAB solution for chemical waste disposal.

Part I (Day 1)

3h

- 1 Bring tissue to  Room temperature in Phosphate Buffered Saline (**PBS**, pH 7.2-7.4) on an orbital shake  30m for 30 minutes.  00:30:00

- 2 Prepare **Peroxide Solution (0.3 % H<sub>2</sub>O<sub>2</sub>)** in **dH<sub>2</sub>O**. 5m  
 For  10 mL **0.3% H<sub>2</sub>O<sub>2</sub>** use:
  -  100 µL **30% H<sub>2</sub>O<sub>2</sub>**
  -  9900 µL **dH<sub>2</sub>O**
  
- 3 Prepare **Normal Goat Serum Blocking Solution** in **PBS**. 5m  
 To  10 mL **PBS** add:
  -  150 µL **Normal Goat Serum** (= 3 drops of serum from Vectastain ABC-HRP Kit, Peroxidase Rabbit IgG PK-4001)
  
- 4 Prepare **Primary Antibody Solution (rabbit anti-GFP)** at **1:10000** dilution in **PBS**: 5m
  -  1 µL **rabbit anti-GFP** (Thermo Fisher, Molecular Probes Cat# A11122, RRID:AB\_221569)
  -  9999 µL **PBS**.
  
- 5 **Rinse** in PBS on a shaker at  Room temperature : **3 x 5 minutes.**  00:05:00 15m
  
- 6 Quench endogenous peroxide in **Peroxide Solution (0.3 H<sub>2</sub>O<sub>2</sub>)** on a shaker at  Room temperature : **30 minutes.**  00:30:00 30m
  
- 7 **Rinse** in PBS on a shaker at  Room temperature : **3 x 5 minutes.**  00:05:00 15m
  
- 8 Incubate in **Normal Goat Serum Blocking Solution** on a shaker at RT: **1 hour.**  01:00:00 1h  
  
**DO NOT RINSE after blocking serum.**

9 Incubate in **Primary Antibody Solution** on a shaker at 4 °C Overnight . 12h

## Part II (Day 2)

4h

10 Bring tissue (in the **Primary Antibody Solution**) to Room temperature on a shaker (**30 minutes**). 30m  
00:30:00

11 Prepare **ABC Solution** in PBS (**at least 30 minutes before use**). 00:30:00 5m  
To 10 mL PBS add:  

- **2 drops A** from Vectastain ABC-HRP Kit, Peroxidase Rabbit IgG PK-4001.
- **2 drops B** from Vectastain ABC-HRP Kit, Peroxidase Rabbit IgG PK-4001.





12 Prepare **Secondary Antibody Solution (1:200)** in PBS. 5m  
To 10 mL PBS add:  

- 150 µL (= 3 drops from Vectastain ABC-HRP Kit, Peroxidase Rabbit IgG PK-4001) of **Normal Goat Serum**.
- 50 µL (= 1 drop from Vectastain ABC-HRP Kit, Peroxidase Rabbit IgG PK-4001) **biotinylated goat anti-rabbit IgG secondary antibody**.


13 **Rinse** in **PBS** on a shaker at Room temperature : **3 x 5 minutes**. 00:05:00 15m

14 Incubate in **Secondary Antibody Solution** on a shaker at Room temperature : **30 minutes**. 00:30:00 30m






15 **Rinse** in **PBS** on a shaker at Room temperature : **3 x 5 minutes**. 00:05:00 15m

- 16 Incubate in **ABC Solution** on a shaker at  Room temperature : **60 minutes.**  01:00:00 . 1h
- 17 **Rinse** in **PBS** on a shaker at  Room temperature : **3 x 5 minutes.**  00:05:00 15m
- 18 Prepare **DAB Peroxide Substrate Solution** in **dH<sub>2</sub>O**. 5m

To use the Vector Labs DAB Peroxidase Substrate Kit (SK-4100):

In  5 mL **dH<sub>2</sub>O**:

  - **2 drops Reagent 1**
  - **4 drops Reagent 2**
  - **2 drops Reagent 3**
  - [optional] 2 drops of Reagent 4 (Nickel) if a black reaction product is desired

**Note: Mix well before use. Use immediately.**
- 19 Incubate in **DAB Peroxide Substrate Solution** on a shaker at  Room temperature :  00:03:00 -  00:06:00 . 6m
- 20 **Rinse** in **PBS** on a shaker at  Room temperature : **3 x 5 minutes.**  00:05:00 15m
- 21 Mount tissue on charged glass slides in **1:8 PBS** in **dH<sub>2</sub>O** and let air dry.

- 22 Rinse slides with **dH<sub>2</sub>O** and let air dry (preferably in a hood).
- 23 Coverslip clean and dry slides with Cytoseal 60 (Thermo Fisher #830-16).