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Protocol for variant calling in SARS-Cov-2 enabling long indel detection

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ABSTRACT

Protocol for SARS-Cov-2 variant calling departing from raw reads in a run. First, we will de-novo assemble the genome using s-aligner, then we will use standard software for variant calling: BWA, samtools, and freebayes. The advantage of this protocol is that it also finds long indels if they exist. Other protocols based on reference mapping will often miss these.

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PROTOCOL CITATION

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protocols.io

https://dx.doi.org/10.17504/protocols.io.btrunm6w

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Assemble the genome

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1.1

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Index the reads in the run. You can use as input a FASTA file (preferred) or a FASTQ. The file can also be compressed with gzip, having a .gz extension. The script will uncompress the file and pass it to FASTA format if it's a FASTQ file.

./index.sh your run id /mnt/c/your run.fastq.gz

1.2

Assemble

./assemble.sh your-run-id -output alignments 500 > results/your-run-id-500.fa
Assemble up to processing 500 reads



Step 1.2 includes a Step case.

<1.200bp

>1200bp

step case

<1.200bp

The largest contig in your-run-id-500.fa is shorter than 1200bp

1.3

 Map your reads to a reference genome to see what's going wrong.

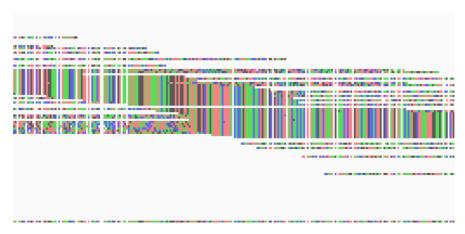
./map.sh your-run-id sequences/your-run-id/sra_data.part-71.fa reference.fa 4000 > results/map-your-run-id.fa

Step 1.3 includes a Step case.

Data not trimmed
Too many chimeras
No overlapping
Runs are short
Contamination

step case

Data not trimmed



If the data is not trimmed and has adaptors at the extremes of the runs it will look like the image above.

1.4 Trim your data and start again.

Ó

2 Call variants. sh to find variants. The script will call BWA, samtools, varscan and freebayes to generate a VCF file with the resulting variants.

./scripts/variants.sh RUN-ID ./results/RUN-ID-xxxx.fa PATH_TO_SARS_COV_2_REFERENCE_FASTA