



NOV 27, 2023

OPEN ACCESS



DOI:
dx.doi.org/10.17504/protocols.io.36wgq3yz5lk5/v1

Protocol Citation: jillian.seiler 2023. Open Field Test.
protocols.io
<https://dx.doi.org/10.17504/protocols.io.36wgq3yz5lk5/v1>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working
 We use this protocol and it's working

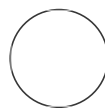
Created: Nov 22, 2023

Last Modified: Nov 27, 2023

🌐 Open Field Test

jillian.seiler¹

¹Northwestern University



jillian.seiler

ABSTRACT

The Open Field Test is a measure of rodent anxiety-like behavior. The rodent's natural "anxious" tendency is to spend time in the outer borders of the chamber, close to the walls; this is called thigmotaxis. A rodent exhibiting decreased anxiety will spend more time in the center of the chamber, where it is unprotected by the walls. If only measuring anxiety-like behavior, measure time spent in the outside of the field and time spent in the center of the field (in this case, "center" refers to the 33cmx33cm square shape in the center of the field). You can also use the Open Field Test to look at locomotion (distance traveled and velocity), as well as exploratory behaviors (rearing).

This protocol is adapted from Tye, et al., 2013 (doi: 10.1038/nature11740) and was originally designed to be used with optogenetic stimulation. If you are performing an Open Field Test without stimulation, you can decrease the time to 10 minutes.

ATTACHMENTS

[n433b9szf.pdf](#)

MATERIALS

Materials:

- Large open field (opaque white; 55 cm x 55 cm)
- Ethovision
- Two USB cameras of the same make and model
- 70% Ethanol and paper towels

Optional Materials:

- Prizmatix Blue LED
- Patch cord
- Ceramic slip
- Extra cage

Procedure

- 1 Open Ethovision and apply necessary settings.

Note

Duration of the test and External Hardware will vary depending on whether or not you'll be using optogenetic stimulation.

- 2 If not done already, swab Open Field with 70% Ethanol and let dry.

- 3 Grasp mouse lightly by the middle of the tail and place in center of the Open Field.

Note

If utilizing optogenetic stimulation, scruff mouse and apply patch cord to implant with help of ceramic slip. Let mouse recover from scruffing in empty cage for 2-5 minutes, then place mouse in the center of the Open Field.

- 4 Press "Start" in Acquisition window of Ethovision.

- 5 When the animal has finished the test, gently remove it from the Open Field and return it to the homepage.

Note

If utilizing optogenetic stimulation, gently remove animal from Open Field and carefully remove patch cord and ceramic slip from mouse's implant. You do not need to scruff the animal for patch cord removal.

- 6 Clean the Open Field with 70% Ethanol and paper towels, and continue with next animal.

Analysis

- 7 Compare time in center and time in thigmo between groups, or, if using optogenetic stimulation, compare time in center and time in thigmo across 3 minute periods.
- 8 Assess freezing behavior (time spent frozen and instances of freezing).