



INSIGHT: a scalable isothermal NASBA-based platform for COVID-19 diagnosis

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1 Works for me [dx.doi.org/10.17504/protocols.io.bghsjt6e](https://doi.org/10.17504/protocols.io.bghsjt6e)

Coronavirus Method Development Community

Chenqu Suo

ABSTRACT

We present here INSIGHT (Isothermal NASBA-Sequencing based HIGH-throughput Test): a two-stage COVID-19 testing strategy, using a combination of an isothermal NASBA reaction and next generation sequencing. From commercially acquired human saliva with spiked-in viral RNA as input, the first stage employs isothermal amplification of viral RNA to give a rapid result in one to two hours, using either fluorescence detection or a dipstick readout, whilst simultaneously incorporating sample-specific barcodes into the amplification product. In the first stage, fluorescent viral RNA detection can be consistently achieved at 10-100 copies per 20 µl reaction. The second stage pools post-amplification barcoded products from multiple samples for scalable sequencing that could be centralised, to further improve the accuracy of the test in a massively parallel way. Our two-stage testing strategy is suitable for further development into a home-based or point-of-care assay, and is potentially scalable to population level.

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CREATED

May 17, 2020

LAST MODIFIED

Jun 03, 2020

MATERIALS

| NAME | CATALOG # | VENDOR |
|--------------------------------------|------------|--|
| QuickExtract DNA Extraction Solution | QE09050 | Lucigen |
| NASBA liquid kit | SKU: NWK-1 | Life Sciences Advanced Technologies Inc. |
| Tris (1 M) pH = 8 RNase free | AM9855G | Invitrogen - Thermo Fisher |
| Sodium Hydroxide | 71687 | Sigma-aldrich |
| 1M MgCl ₂ | AM9530G | Invitrogen - Thermo Fisher |
| 2M KCl | AM9640G | Invitrogen - Thermo Fisher |
| DTT | 43816 | Sigma-aldrich |
| DMSO | 276855 | Sigma-aldrich |
| dNTP set 100 mM | 10297018 | Invitrogen - Thermo Fisher |
| NTP set 100 mM | R0481 | Thermo Scientific |
| Bio-11-UTP (75 mM) | AM8451 | Invitrogen - Thermo Fisher |
| RNase H | M0297L | NEB |
| ProtoScript II reverse transcriptase | M0368S | NEB |
| T7 RNA polymerase | M0251L | NEB |
| BSA 20 mg/ml | B9000S | NEB |
| Direct-zol RNA Miniprep Plus | R2070 | Zymo Research |
| PCRD lateral flow assay | FG-FD51673 | Abingdon Health |
| Qubit RNA HS Assay Kit | Q32852 | Invitrogen - Thermo Fisher |
| PowerUp™ SYBR™ Green qPCR Master Mix | 15340939 | Applied Biosystems |

MATERIALS TEXT

Primers pair sequence:

| | |
|--|---|
| FWD primer | CCAGCAACTGTTTG TGGACCTA |
| REV primer with T7 handle | aattctaatacgactcact atagggagaaggACAC CTGTGCCTGTTAAA CCAT |
| FWD primer with 5-nt barcode and Illumina handle | tgactggagttcagacgt gtgctctccgatctnnnn nCCAGCAACTGTTT GTGGACCTA |
| REV primer with 5-nt barcode and T7 handle | aattctaatacgactcact atagggagaaggnnnnn ACACCTGTGCCTGT TAAACCAT |

Molecular beacon:

FAM-AUUGACAGUCUACUAAUUUGGUUAAAAACAAUGUGUCA-BHQ1dT-UUCAACUCAAUG-propyl

FAM labelled RNA capture oligo for PCR

FAM-AAAAGTCTACTAATTTGGTTAAAAACAAATGTGTCAATTTCAACTTC

SAFETY WARNINGS

***** IMPORTANT: This protocol has not been validated on patient samples and should not be used for clinical diagnosis without further validation and certification. *****

1 Lysis of saliva samples

Mix crude saliva at 1:1 ratio with QuickExtract DNA Extraction Solution. Incubate at 95 °C for 5 min to ensure complete lysis of virus and inactivation of proteinase K.

2 NASBA reaction

Take 1 µl from the product of Step 1 and add into the NASBA reaction mixture to make a total volume of 20 µl. Reaction mixture can either be prepared in-house or from the Life Sciences NASBA liquid kit (see tables below).

A fluorescence plate reader can be used to monitor the reaction in real-time, or as an endpoint assay.

| | vol. | stock conc. | conc. in RM |
|-----------------------|--------|-------------|------------------|
| sample | 1 µl | | |
| primers* | 1 µl | 0.5 µM each | 25 nM each |
| water +/- beacon | 3 µl | | 20 nM for beacon |
| buffer (NECB-24) | 6.7 µl | | |
| nucleotide (NECN-24) | 3.3 µl | | |
| enzyme mix (NEC-1-24) | 5 µl | | |
| total volume | 20 µl | | |

Life Sciences reaction mixture (RM)

* Primer sequence available in Materials.

| | vol. | stock conc. | conc. in RM |
|---------------------|-------|-------------|------------------|
| sample | 1 µl | | |
| primers* | 1 µl | 0.5 µM each | 25 nM each |
| water +/- beacon | 4 µl | | 20 nM for beacon |
| buffer with DMSO* | 5 µl | | |
| nucleotide mix* | 4 µl | | |
| enzyme mix* | 5 µl | | |
| total volume | 20 µl | | |

In-house reaction mixture (RM)

* For detailed mixture composition, see tables below. Primer sequence available in Materials.

| | vol. | stock conc. | conc. in RM |
|---------------------|----------|-------------|-------------|
| Tris-HCl pH 8.4* | 120 µl | 1 M | 40 mM |
| MgCl ₂ | 39.6 µl | 1 M | 13.2 mM |
| KCl | 112.5 µl | 2 M | 75 mM |
| DTT | 30 µl | 1 M | 10 mM |
| DMSO | 450 µl | 100% | 11% |
| water | 247.9 µl | | |
| total volume | 1000 µl | | |

Buffer with DMSO

*Tris-HCl pH 8.4 is made in-house by titrating Tris-HCl pH 8.0 with NaOH pellet and pH determined by pH meter.

| | vol. | stock conc. | conc. in RM |
|-------------------|----------|-------------|-------------|
| Tris-HCl pH 8.4 | 120 µl | 1 M | 40 mM |
| MgCl ₂ | 39.6 µl | 1 M | 13.2 mM |
| KCl | 112.5 µl | 2 M | 75 mM |
| DTT | 30 µl | 1 M | 10 mM |
| water | 697.9 µl | | |

| | | | |
|---------------------|---------|--|--|
| total volume | 1000 µl | | |
|---------------------|---------|--|--|

Buffer without DMSO

| | vol. | stock conc. | conc. in RM |
|---------------------|--------------|--------------------|--------------------|
| dNTP | 0.22 µl each | 100 mM | 1 mM each |
| NTP | 0.88 µl each | 100 mM | 4 mM each |
| total volume | 4.4 µl | | |

Nucleotide mix (incl. 10% excess)

| | vol. | stock conc. | conc. in RM |
|---------------------|-------------|--------------------|--------------------|
| diluted Rnase H | 0.17 µl | 500 U/ml | 3.75 U/ml |
| Photocript RT | 0.28 µl | 200000 U/ml | 2500 U/ml |
| T7 polymerase | 2.75 µl | 50000 U/ml | 6250 U/ml |
| BSA | 0.13 µl | 20 mg/ml | 0.12 mg/ml |
| buffer without DMSO | 1.78 µl | | |
| water | 0.40 µl | | |
| total volume | 5.5 µl | | |

Enzyme mix (incl. 10% excess)

| | vol. | stock conc. |
|---------------------|-------------|--------------------|
| Rnase H | 5 µl | 5000 U/ml |
| BSA (0.48mg/ml) | 1.2 µl | 20 mg/ml |
| buffer without DMSO | 16.67 µl | |
| water | 27.13 µl | |
| total volume | 50 µl | |

Diluted Rnase H

Step 2 includes a Step case.

(a) With denaturation step

(b) Completely isothermal

step case

(a) With denaturation step

Reaction mixture without the enzyme mix is incubated at 65 °C for 2 min followed by a 10-min incubation at 41 °C. Following that, 5 µl enzyme mix is added into the reaction and incubated at 41 °C for a further of 90-120 min.

3 Detection with lateral flow dipstick (if desired)

For detection with lateral flow assay, Bio-11-UTP can be added into the NASBA nucleotide mixture in Step 2 at a final concentration of 0.5 mM. At the end of the NASBA reaction, RNA is purified from the end-product using Direct-zol RNA Miniprep kit, and eluted with 30 µl of RNase free water. After purification, 4.2 µl of purified RNA is mixed with 1.8 µl of FAM labelled RNA capture oligo and 84 µl of PCRD extraction buffer. Take 75 µl of mix to the sample well of a PCRD test cassette. Results will be shown within 10 min.

4 Sequencing stage

To allow for pooled sequencing of NASBA reaction end products, barcode sequences are added upstream of each of the forward and reverse primers. In addition, an Illumina sequencing adaptor is added upstream of the forward primer barcode sequence as a universal PCR handle (see Materials and Reagents for the exact sequence).

Uniquely barcoded NASBA end products from different samples are pooled and purified. A one step PCR can be carried out at local sequencing centres using universal P5 and P7 primers. Here, if needed, another layer of indexing barcodes can be added to further increase the multiplexing capacity.