



Sep 08, 2020

# OnsiteGene 2 Protocol Saliva Direct

yliu<sup>1</sup><sup>1</sup>OnsiteGene Inc.**1** Works for me dx.doi.org/10.17504/protocols.io.bkuikwue

OnsiteGene 1 XPRIZE Rapid Covid Testing

yliu yliu

## ABSTRACT

This OnsiteGene protocol is designed for testing the normal saliva samples without nucleic acid extraction. A convenient saliva swab is used to collect about 50  $\mu$ L of saliva for the detection. The protocol uses the Star Array® Hi-Sense™ COVID-19 Molecular Testing Kit 1.0 in the one-step real-time RT-qPCR test assay to qualitatively detect RNA from SARS-CoV-2 virus in the human respiratory specimen. It combines the reverse transcription technology and real-time PCR method to provide accurate detection of the SARS-CoV-2 coronavirus. Along with the Star Array® SA™ Direct Extract Buffer, the protocol supports direct amplification without the need of specific RNA extraction equipment and kit. It significantly reduces the loss of RNA from the sample extraction and purification process, saves time and workload from sample preparation, and minimized the burdens on supply chain. The triplex fluorescence design of the kit simultaneously detects the N1 and Orf1ab genes of the virus and the human RNase P gene as an internal control to ensure the sample quality. This protocol uses the Star Array® XDive™ Superfast Real-Time PCR instrument to perform 40-cycle PCR in 8 minutes, and can test up to 16 samples or controls in each run. The total protocol time from sample collection to data interpretation is less than 11 minutes.

## DOI

[dx.doi.org/10.17504/protocols.io.bkuikwue](https://dx.doi.org/10.17504/protocols.io.bkuikwue)

## PROTOCOL CITATION

yliu 2020. OnsiteGene 2 Protocol Saliva Direct. **protocols.io**  
<https://dx.doi.org/10.17504/protocols.io.bkuikwue>

## KEYWORDS

XDive, superfast PCR, superfast RT-qPCR, fast PCR

## LICENSE

———— This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

## CREATED

Sep 04, 2020

## LAST MODIFIED

Sep 08, 2020

## PROTOCOL INTEGER ID

41578

## MATERIALS TEXT

Instruments and Consumables

Instrument and Consumable	Supplier	Catalog #
Real-Time PCR	OnsiteGene	XDIVE-16

Block heater	FOUR E'S SCIENTIFIC	TC0401005-UUSSAA
Mini centrifuge	Heathrow Scientific	HS120395
Vortex	Four E's Scientific	4ESS-MI0101002D
Reaction tube and cap	OnsiteGene	CM1010001
Pipette set	Microlit	RBOKit
Pipette tips	Extragene	TS-10-RS

#### Sample Collection and Extraction Kit

Reagent or Consumable	Supplier	Catalog #
Swab	Medcomma	MFS-03-01
1.5ml microcentrifuge tube	Biologix	88-915X
500uL PCR tube	Biologix	88-905X
Direct RNA release buffer	OnsiteGene	RG2010001
Storage solution	OnsiteGene	RG0000001

#### PCR Reaction Reagents

Reagent	Supplier	Catalog #
Hi-Sense COVID detection kit	OnsiteGene	KT1010001
External positive control	ZeptoMetrix	NATSARS(COV2)-ERC

#### Primer and Probe Sequences

Target Number	Target Gene	Sequence/antibody
1_F	N1	GACCCCAAATCAGCGAAAT
1_R	N1	TCTGGTTACTGCCAGTTGAATCTG
1_P	N1	ACCCCGCATTACGTTTGGTGGACC
2_F	ORF1ab	TATGTGGAAAGGTTATGGCTGTAG
2_R	ORF1ab	GATTGTGCATCAGCTGACTGAAG
2_P	ORF1ab	TTGTGATCAACTCCGCGAACCCATG
3_F	Rnase P	AGATTTGGACCTGCGAGCG
3_R	Rnase P	GAGCGGCTGTCTCCACAAGT
3_P	Rnase P	TTCTGACCTGAAGGCTCTGCGCG

#### SAFETY WARNINGS

**Most institutions will require samples potentially containing full-length SARS-CoV-2 RNA to be handled in a biosafety level 2 cabinet.** Please seek guidance from your local biosafety office on specific recommendations for working with samples which could contain live SARS-CoV-2 virus.

### Sample Collection Procedure

1m

- 1 Saliva sample should be collected with the assistance of a healthcare worker or technician.
- 2 Before collection, clean hands using alcohol-based sanitizer or soap and water (no fragrances) and wear appropriate PPE (at minimum, gloves and a mask).
  - 2.1 Back away as far as possible from the sample donor during the whole process. While preparing collection materials, direct the sample donor to begin pooling saliva in their mouth.

2.2 This protocol is intended for the collection of the normal saliva that naturally pools into the mouth. No coughing or sniffing prior to sample collection is required. Ideally, water should be avoided 10 minutes prior to collection. Other drinks, food, and nasal sprays should be avoided half an hour before sample collection.

3 Ensure all collection materials are labelled with the correct identifying information.

4 Remove the lid of the collection tube containing 450  $\mu$ L sample storage solution.

5s

5 Take the saliva swab out from the package and direct the sample donor to soak the swab pad in the mouth with saliva for 15 seconds.

30s

6 Place the soaked swab into the storage solution in the tube. Remove stick through the opening and leave swab pad in the tube. Discard stick and secure the lid.

10s

6.1 Sterilize the tube surface with 70% ethanol or a disinfecting wipe, and place the sample in a secondary container or an appropriately labeled biohazard bag.

6.2 Dispose of gloves, and register the sample collection information (including date and time).

7 Transfer the sample at room temperature for sample processing. The virus RNA remains stable at room temperature for 3-5 days.

### Real-Time PCR Procedure

9m 45s

8 Vortex each sample collection tube for 5 seconds at 3000-5000 RPM.

5s

8.1 This step can be substituted by tapping or shaking each sample collection tube to mix the sample.

9 Spin down the tube for 15 seconds using centrifuge.

15s

9.1 The purpose is to: 1) Remove liquid from the cap to avoid the contamination when cap is removed; 2) Spin down solid particles to the bottom of the tube and allow supernatant to be acquired.

10 Transfer 20  $\mu$ L supernatant of the sample storage solution to a tube containing 20  $\mu$ L SA<sup>TM</sup> direct RNA release buffer. Secure the lid of the tube.

5s

- 11 Heat the tube with the sample for 1 minute at 95°C on a heating block. 1m
- 12 Transfer 7.5 µL viral RNA sample solution (this sample can be stored at -20°C for 14 days) and 7.5 µL OnsiteGene Hi-5<sup>5s</sup> Sense™ COVID-19 detection kit reaction mix (with primers and probes) into a superfast RT-qPCR reaction tube. Secure the lid of the tube.
- 13 Spin down the tube for 15 seconds using centrifuge or spinner. 15s
- 14 Load up to 16 superfast RT-qPCR reaction tubes onto the OnsiteGene XDive Superfast Real-Time PCT instrument and run the following conditions: 8m

Step	Temperature	Time
1	50°C	30 sec
2	95°C	0 sec
3	60°C	2 sec
4	Read Fluorescence intensity and overhead	(10 sec)
5	Repeat steps 2-4 for 40 cycles.	

a) XDive takes 8 minutes to complete 40 thermal cycles with fluorescence imaging.

b) Run external positive control and external negative control twice per day using 5 µL of positive control and no-template control (NTC - water). The control tests are recommended to be performed at the beginning and the end of the work/day.

- 15 15. Report results per the following interpretation criteria:

Results	IC CT	SARS-Cov-2 Targets		Interpretation
		ORF1ab CT	N1 CT	
Negative	< 35	≥ 40	≥ 40	Indicate the absence of SARS-Cov-2 RNA
Positive	any value	< 40	any value *	Indicate the presence of SARS-Cov-2 RNA
		any value *	≤ 40	
Invalid	≥ 35	≥ 40	≥ 40	Inability to conclusively determine presence or absence of SARS-Cov-2 RNA. This may be due to 1) Internal Control failure; or 2) failure to detect sufficient specimen volume. The sample needs to be retested.

\* In the case of one SARS-Cov-2 target positive (CT < 40) and one SARS-Cov-2 target negative (CT ≥ 40), the result is suggestive of: 1) a sample at concentrations near or below the limit of detection of the test, 2) a mutation in one of the target regions, or 3) other factors.