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Minimal Event Distance Aneuploidy Lineage Tree (MEDALT) inference based on single cell copy number profile

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1 Works for me dx.doi.org/10.17504/protocols.io.bfhpij5n

Fang Wang

ABSTRACT

This protocol describes two innovative algorithms:

- 1) A minimal event distance aneuploidy lineage tree (MEDALT) inference algorithm allows implementing genetically meaningful distances and is scalable to current single-cell datasets containing thousands of cells, and
- 2) A statistical routine, Lineage Speciation Analysis (LSA), enables prioritization of CNAs and genes that are non-randomly associated with the observed lineage expansion and thereby are potentially functionally important.

EXTERNAL LINK

<https://www.biorxiv.org/content/10.1101/2020.04.12.038281v1.full>

- 1 Install Python 2.7 and R 3.5
Download MEDALT tool from <https://github.com/KChen-lab/MEDALT.git>

**MEDALT 1.0**

by Fang Wang and Qihan Wang

Extract input dataset



Single cell copy number profile generated by single cell DNA seq



Single cell copy number profile inferred from single cell RNA se

2 Decompress gzipped files (MEDALT-1.0.tar.gz)



```
tar -zxvf MEDALT-1.0.tar.gz
cd MEDALT-1.0
```

```
#help document
python scTree.py -h
```

Usage: python scTree.py <-P path> <-I input> <-D datatype>

**Input integer copy number profile. Columns correspond to chromosomal position.
Rows correspond to cells.**

Options:

--version show program's version number and exit
-h, --help Show this help message and exit.
-P PATH, --Path=PATH Path to script
-I INPUT, --Input=INPUT
Input file
-G GENOME, --Genome=GENOME
Genome version hg19 or hg38
-O OUTPUT, --Output=OUTPUT
Output path
-D DATATYPE, --Datatype=DATATYPE
The type of input data. Either D (DNA-seq)
or R (RNA-seq).
-W WINDOWS, --Windows=WINDOWS
the number of genes you want to merge when
you input copy number profile inferred from
scRNA-seq. Default 30.
-R PERMUTATION, --Permutation=PERMUTATION
Whether reconstructed permuted tree (T) or
not (F). If not, permuted copy number
profile will be used to perform LSA. Default
value is F due to time cost.

3 Run the example data generated based on single cell DNA sequencing technology

 **scDNA.CNV.txt**



```
python scTree.py -P ./ -I ./example/scDNA.CNV.txt -D D -G hg19 -O
./example/outputDNA
```

Transfer data to segmental level
Inferring MEDALT.
MEDALT inference finish

MEDALT inference finish.

Performing LSA.

Loading required package: BiocGenerics

Loading required package: parallel

Attaching package: 'BiocGenerics'

The following objects are masked from 'package:parallel':

**clusterApply, clusterApplyLB, clusterCall, clusterEvalQ,
clusterExport, clusterMap, parApply, parCapply, parLapply,
parLapplyLB, parRapply, parSapply, parSapplyLB**

The following objects are masked from 'package:stats':

IQR, mad, sd, var, xtabs

The following objects are masked from 'package:base':

**anyDuplicated, append, as.data.frame, basename, cbind, colMeans,
colnames, colSums, dirname, do.call, duplicated, eval, evalq,
Filter, Find, get, grep, grepl, intersect, is.unsorted, lapply,
lengths, Map, mapply, match, mget, order, paste, pmax, pmax.int,
pmin, pmin.int, Position, rank, rbind, Reduce, rowMeans, rownames,
rowSums, sapply, setdiff, sort, table, tapply, union, unique,
unsplit, which, which.max, which.min**

Loading required package: S4Vectors

Loading required package: stats4

Attaching package: 'S4Vectors'

The following object is masked from 'package:base':

expand.grid

Loading required package: IRanges

Loading required package: GenomicRanges

Loading required package: GenomeInfoDb

Loading required package: Biostrings

Loading required package: XVector

Attaching package: 'Biostrings'

The following object is masked from 'package:base':

strsplit

Loading required package: BSgenome

Loading required package: rtracklayer

Loading required package: GenomicFeatures

Loading required package: AnnotationDbi

Loading required package: Biobase

Welcome to Bioconductor

welcome to bioconductor

**Vignettes contain introductory material; view with
'browseVignettes()'. To cite Bioconductor, see
'citation("Biobase")', and for packages 'citation("pkgname")'.**

Loading required package: VariantAnnotation

Loading required package: SummarizedExperiment

Loading required package: DelayedArray

Loading required package: matrixStats

Attaching package: 'matrixStats'

The following objects are masked from 'package:Biobase':

anyMissing, rowMedians

Loading required package: BiocParallel

Attaching package: 'DelayedArray'

The following objects are masked from 'package:matrixStats':

colMaxs, colMins, colRanges, rowMaxs, rowMins, rowRanges

The following object is masked from 'package:Biostrings':

type

The following objects are masked from 'package:base':

aperm, apply

Loading required package: Rsamtools

Attaching package: 'VariantAnnotation'

The following object is masked from 'package:base':

tabulate

Loading required package: GenomicAlignments

There were 20 warnings (use warnings() to see them)

Attaching package: 'igraph'

The following objects are masked from 'package:DelayedArray':

path, simplify

The following objects are masked from 'package:rtracklayer':

blocks, path

The following object is masked from 'package:Biostrings':

union

The following object is masked from 'package:GenomicRanges':

union

The following object is masked from 'package:IRanges':

union

The following object is masked from 'package:S4Vectors':

union

The following objects are masked from 'package:BiocGenerics':

normalize, path, union

The following objects are masked from 'package:stats':

decompose, spectrum

The following object is masked from 'package:base':

union

Warning message:

package 'igraph' was built under R version 3.5.2

Attaching package: 'DescTools'

The following object is masked from 'package:igraph':

%c%

Warning message:

package 'DescTools' was built under R version 3.5.2

[1] Visualization MEDALT!

null device

1

[1] LSA segmentation!

[1] Calculating CFL

[1] Calculating permutation CFL

[1] Estimate emperical p value

[1] Estimate parallel evolution

null device

1

Done!



R packages (igraph, HelloRanges and DescTools) are loaded.



Three text files are expected:

(1) CNV.tree.txt which is an rooted directed tree including three columns: parent node, child node and distance.

☐ **CNV.tree.txt**

(2) segmental.LSA.txt which includes broad CNAs significantly associated with lineage expansion.

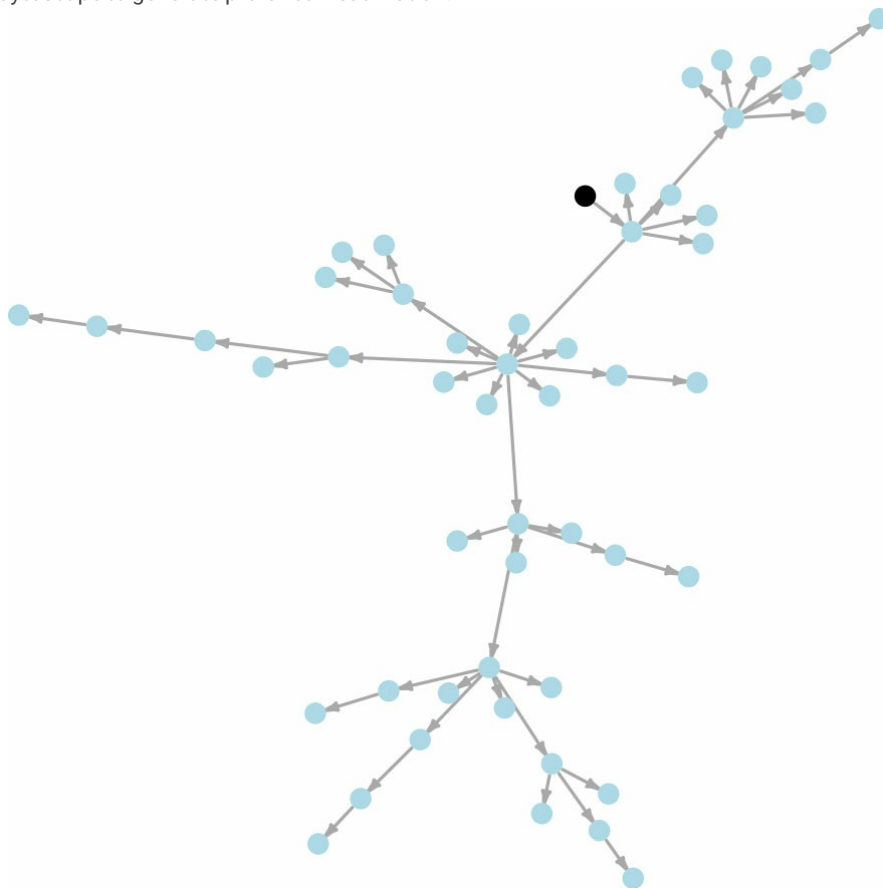
☐ **segmental.LSA.txt**

(3) gene.LSA.txt which includes focal (gene) CNAs significantly associated with lineage expansion.

☐ **gene.LSA.txt**

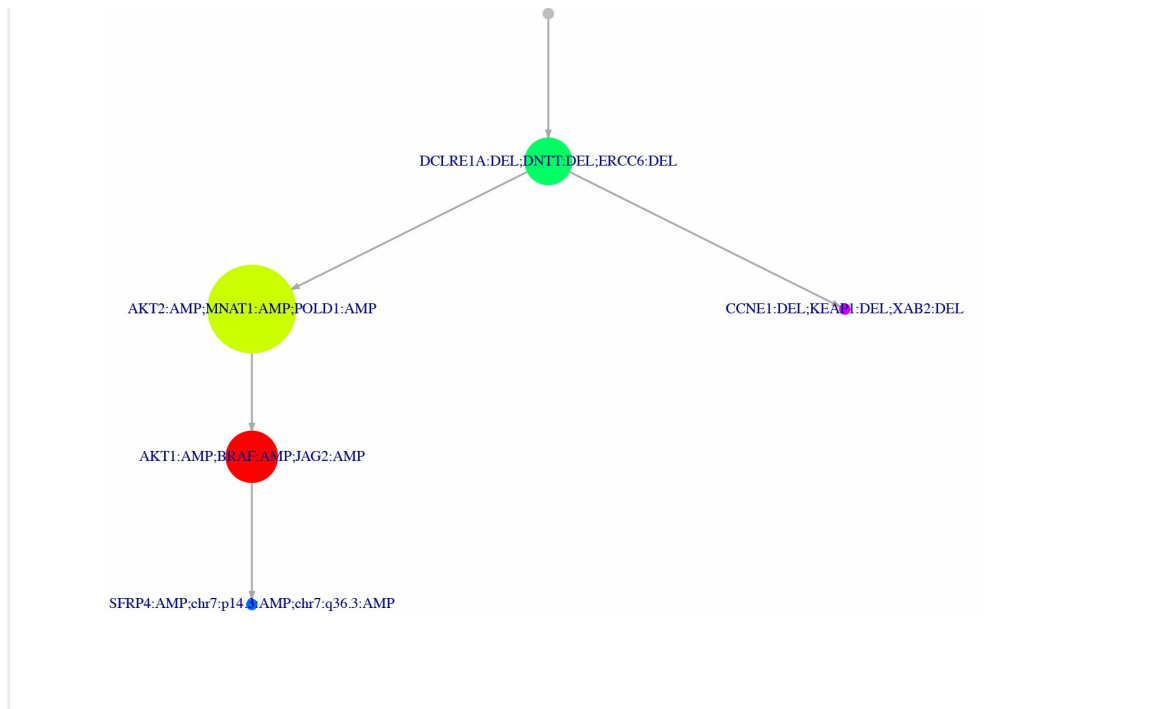
Two figures are also expected:

(1) singlecell.tree.pdf which is a visualization of MEDALT by igraph. You also can input CNV.tree.txt into Cytoscape to generate preferred visualization.



Each node represents a cell, each edge represents a kinship between two cells, arrows point towards younger cells, and the root represents a normal diploid cell.

(2) LSA.tree.pdf which is a visualization of identified CNAs by igraph.



We run the example data only through permuting copy number profile instead of reconstructing tree based on permuted copy number profile. The setting can be changed via -R T.

4 Run the example data inferred using inferCNV based on single cell RNA sequencing technology

☐ **scRNA.CNV.txt**



```
python scTree.py -P ./ -I ./example/scRNA.CNV.txt -D R -G hg19 -O
./example/outputRNA
```

Transfer data to segmental level

The number of genes which are merger into the bin is default value 30. If you want change it please specify the value through -W

Inferring MEDALT.

MEDALT inference finish.

Performing LSA.

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The following object is masked from ‘package:Biostrings’:

union

The following object is masked from ‘package:GenomicRanges’:

union

The following object is masked from ‘package:IRanges’:

union

The following object is masked from 'package:S4Vectors':

union

The following objects are masked from 'package:BiocGenerics':

normalize, path, union

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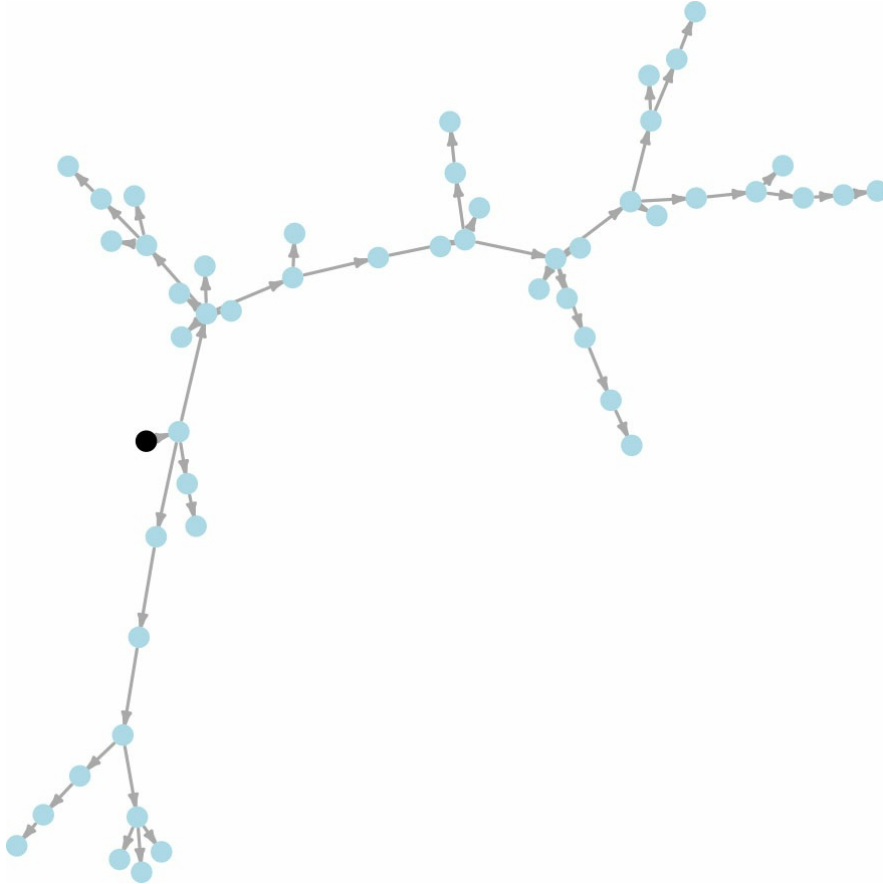
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