TEST - manual DNA Purification via magnetic beads V.2

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OPEN ACCESS

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We are still developing and optimizing this protocol

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ABSTRACT

manual purification of DNA/PCRs via magnetic beads

ATTACHMENTS

manual purification_CaJu.xlsx

MATERIALS

- NGS clean-up and size selection magnetic beads (Macherey-Nagel)
- 80% EtOH
- WFI (Ampuwa)

- magnetic rack
- tube rack
- fresh tubes

BEFORE START INSTRUCTIONS

bring magnetic beads to room temperature (at least 30min)

prepare magnetic beads

1 00:30:00 bring magnetic beads to room temperature (RT) 30m

2 vortex until homogeneous

Binding of DNA

3 Add beads

0.9 times the amount of PCR volume

4 00:05:00 incubate for at least 5min 5m

5 00:05:00 place tube in magnetic rack 5m

6 remove supernatant

1st wash

7 add ethanol

1.9 times the amount of PCR volume

vortex thoroughly

8 00:05:00 place in magnetic rack 5m

9 remove supernatant

2nd wash

10 add ethanol

1.9 times the amount of PCR volume

vortex thoroughly

11 00:05:00 place in magnetic rack 5m

12 remove supernatant

Airdry beads

13 00:10:00 keep lid open in magnetic rack 10m

14 remove residual supernatant

Elute DNA 10m

15 place tube in sample rack

16 add indicated volume of water and mix/vortex thoroughly 5m

00:05:00 incubate

17 00:05:00 place in magnetic rack 5m

18 transfer supernatant into labeled tube