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## Plasmid Extraction (Plasmid Isolation)

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**Protocol status:** Working  
We use this protocol and it's working

**Created:** Sep 27, 2023

**Last Modified:** Sep 28, 2023

### ABSTRACT

2023 NUS-Singapore iGEM team followed this protocol to isolate the plasmid from the cells.

### PROTOCOL MATERIALS

⊗ Buffer PE Qiagen Catalog #19065	In <a href="#">2 steps</a>
⊗ Buffer P1 Qiagen Catalog #19051	Step 5
⊗ RNase A Qiagen Catalog #19101	Step 5
⊗ Buffer N3 Qiagen Catalog #19064	Step 9
⊗ PB buffer Qiagen Catalog #19066	Step 15
⊗ Buffer P2 Qiagen Catalog #19052	Step 7

### SAFETY WARNINGS





- ⚠ Proper lab PPE must be worn at all times.

### BEFORE START INSTRUCTIONS

Glycerol stock ( 900 µL of cell stock in 300 µL of 100% glycerol solution) may be prepared and keep it in the -80 °C fridge before the plasmid extraction procedure.






**Keywords:** Plasmid  
Extraction, Plasmid, Buffer  
P1, Buffer P2, Buffer N3,  
Buffer PB, Buffer PE, Plasmid  
Isolation, Isolation






## Cell Culture

- 1 Incubate the cells containing the plasmid of interest in a Falcon tube with  5 mL of LB media and  5 µL of the appropriate antibiotics  Overnight at  37 °C before starting the plasmid extraction.

## Plasmid Extraction

5m

- 2 Take out the Falcon tube with the cultured cells from the incubator.
- 3 Centrifuge the Falcon tube at  5000 rpm, 4°C, 00:05:00 . 5m
- 4 Discard the supernatant in the Falcon tube and keep the cell pellet.
- 5 Add  250 µL of  Buffer P1 Qiagen Catalog #19051 (with  RNase A Qiagen Catalog #19101 added, kept in the  4 °C fridge) into the Falcon tube and resuspend the cell pellet.
- 6 Transfer the whole solution into a Eppendorf tube.

- 7 Add  250  $\mu$ L of  Buffer P2 Qiagen Catalog #19052 into the Eppendorf tube to lyse the cells.
- 8 Shake the Eppendorf tube for mixing.
- 9 Add 350 $\mu$ L of  Buffer N3 Qiagen Catalog #19064 into the Eppendorf tube.
- 10 Shake the Eppendorf tube for mixing, a cloudy solution shall be observed.
- 11 Centrifuge the Eppendorf tube at  13 rpm, 00:10:00 . 10m
- 12 Transfer the supernatant into a Mini Prep tube and discard the cell debris.
- 13 Centrifuge it at  13 rpm, 00:01:00 . 1m
- 14 Discard the flow-through and place the Mini Prep tube back into the same tube.

15 Add  500 µL of  PB buffer Qiagen Catalog #19066 into the Mini Prep tube.



16 Centrifuge it at  13 rpm, 00:01:00 .

17 Discard the flow-through and place the Mini Prep tube back into the same tube.



18 Add  700 µL of  Buffer PE Qiagen Catalog #19065 into the Mini Prep tube.

19 Centrifuge it at  13 rpm, 00:01:00 .

20 Discard the flow-through and place the Mini Prep tube back into the same tube.

21 Centrifuge it at  13 rpm, 00:01:00 to remove the  Buffer PE Qiagen Catalog #19065 residual.

22 Transfer the Mini Prep column into the newly labelled Eppendorf tube.

- 23 Add  50 µL of DI water into the Eppendorf tube.
- 24 Centrifuge it at  13 rpm, 00:01:00 , ensuring that the direction of the Eppendorf tube's cap is the same as the direction of spinning to avoid breaking.
- 25 Discard the Mini Prep column, the solution left in the Eppendorf tube is the plasmid.
- 26 Use NanoDrop

#### Equipment

NanoDrop™ One/OneC Microvolume UV-Vis Spectrophotometer	NAME
UV-Vis Spectrophotometer	TYPE
Thermo Scientific	BRAND
ND-ONE-W	SKU

to measure the concentration (under "dsDNA" mode) and the purity of the plasmid.

- 27 Store the isolated plasmid at room temperature.