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Standard DAB Staining for Free-floating Fixed NHP Brain Tissue V.2

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ABSTRACT

This protocol details the procedure for immunohistochemical 3,3'-Diaminobenzidine (DAB) staining of free-floating fixed brain tissue sections using the avidin/biotin ABC complex.

This protocol has been tested with free-floating non-human primate (NHP) and rodent (mouse, rat) brain tissue that has been fixed (10% formalin or 4% paraformaldehyde), cryoprotected (sucrose or glycerol gradients), and cryo-sectioned + 20 µm -



GUIDELINES

When using 6 well tissue culture plates [Falcon, 353046] to react individual sections, you will need **2+ mL** solutions for *each* well plate.

When using circular staining nets [e.g., Brain Research Laboratories #4115] to react multiple series of sections, you will need **50 mL** solutions for **each**.



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Tissue

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Aligning Science Across

Parkinson's

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MATERIALS

Tissue:

Brain tissue sections (20 - 50 µm).

Materials/Equipment:

- Tissue culture plates or circular staining nets
- Orbital shaker
- Fume hood
- Nitrile Gloves
- Glass slides (charged or subbed)

Reagents:

- Phosphate-buffered saline (PBS)
- Hydrogen Peroxide: H₂O₂ (3% or 30%)
- Distilled water: dH₂O
- Primary Antibody
- Secondary Antibody (to match the host of the primary antibody)
- Normal Serum Blocking Solution (e.g., Normal Horse Serum, S-2000-20; Normal Goat Serum, S-1000-20)
- Vectastain Elite ABC Peroxidase Kit (Standard) (PK-6100) (Vector Laboratories)
- ABC-HRP Kit

Examples:

Vectastain ABC-HRP Kit, Peroxidase (Mouse IgG) (PK-4002, Vector Laboratories) Vectastain ABC-HRP Kit, Peroxidase (Rabbit IgG) (PK-4001, Vector Laboratories)

DAB Substrate Kit

Examples:

Peroxidase (HRP) with Nickel (3,3'-diaminobenzidine) (SK-4100) (Vector Laboratories) ImmPACT DAB (SK-4105)

SAFETY WARNINGS

Use appropriate care when using hydrogen peroxide (reactive, can cause skin/eye damage) and DAB (suspected carcinogen). Collect DAB solution for chemical waste disposal.

Part I (Day 1)

3h

- Bring tissue to Room temperature in buffer (e.g., Phosphate buffered saline, PBS) on an orbital shake 30m for 30 minutes. 00:30:00.
- Prepare Peroxide Solution $(0.3 3 \% H_2O_2)$ in dH_2O .

5m

E.g., for \triangle 10 mL 0.3% H₂O₂ use:

- Д 100 µL 30% H₂O₂
- Д 9900 µL dH₂O
- Prepare **Blocking Serum Solution** (e.g. Normal Horse, Normal Goat Serum) using a serum that matches the host of the secondary antibody (e.g. Normal Horse Serum for a Horse anti-Mouse secondary, Normal Goat Serum for a Goat anti-Rabbit secondary).

E.g., in 🔼 10 mL buffer (PBS) add:

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- 4 Prepare **Primary Antibody Solution** at the appropriate dilution in buffer (e.g., 1:1000 in PBS).

5m

Rinse in buffer (e.g. PBS) on a shaker at Room temperature : 3 x 3-5 minutes. 00:03:00

15m

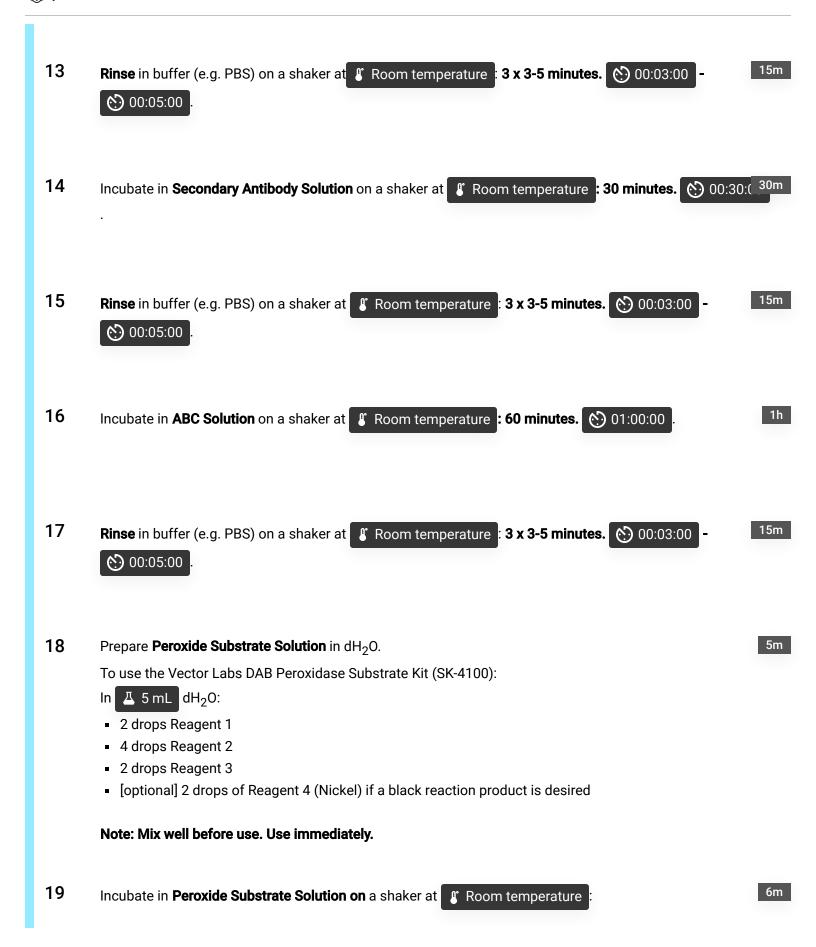
© 00:05:00

6 Quench endogenous peroxide in **Peroxide Solution (0.3 - 3 % H₂O₂)** on a shaker at 8 Room temperature 30 - 60 minutes. 8 00:30:00 -01:00:00 7 15m Rinse in buffer (e.g. PBS) on a shaker at Room temperature: 3 x 3-5 minutes. (5) 00:03:00 -00:05:00 8 Incubate in **Blocking Serum Solution** on a shaker at RT: 1 hour. 1h DO NOT RINSE after blocking serum. 9 20h Incubate in **Primary Antibody Solution** on a shaker at **4** °C Overnight, or longer (20 - 72 hours depending on the antibody). 4h Part II (Day 2) 10 30m Bring tissue (in the **Primary Antibody Solution**) to Room temperature on a shaker (30 - 60 minutes). **(?)** 00:30:00 **(:)** 01:00:00 11 5m Prepare **ABC Solution** in buffer (e.g. PBS) (at least 30 minutes before use). (5) 00:30:00 12 5m Prepare **Secondary Antibody Solution** (1:200) in buffer (e.g. PBS). △ 10 mL buffer add: Δ 150 μL (= 3 drops of normal serum from a Vector Labs kit) of normal serum (matched to the host of your secondary antibody)

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(matched to the host of your primary Antibody)

Δ 50 μL (= 1 drop secondary antibody from a Vector Labs kit) of biotinylated secondary antibody



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Note: Watch the tissue closely to avoid high background staining.

- Rinse in buffer (e.g. PBS) on a shaker at Room temperature : 3 x 3-5 minutes. 00:03:00 15m
- Mount tissue on glass slides (subbed or charged) in 1:8 buffer in dH₂O and let air dry.
- Rinse slides with dH₂O and let air dry (preferably in a hood).
- 23 Coverslip clean and dry slides with Cytoseal 60 (Thermo Fisher #830-16).

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