



Oct 01, 2021

NGM Agar

In 1 collection

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Adrien Assie: I am not the author of this protocol. This is a standard protocol for C. elegans maintenance available on worm book.

1 Works for me

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dx.doi.org/10.17504/protocols.io.zy7f7zn

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DISCLAIMER

This is a standard protocol for C. elegans maintenance available on wormbook.

ABSTRACT

NGM agar recipe from wormbook

DOI

dx.doi.org/10.17504/protocols.io.zy7f7zn

EXTERNAL LINK

http://www.wormbook.org/chapters/www_strainmaintain/strainmaintain.html

PROTOCOL CITATION

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<https://dx.doi.org/10.17504/protocols.io.zy7f7zn>

MANUSCRIPT CITATION please remember to cite the following publication along with this protocol

Brenner, S. (1974). Genetics 77, 71.

COLLECTIONS ⓘ



Samuel Lab Media and Buffers

KEYWORDS

C. elegans, Growing media, Caenorhabditis, nematode

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PARENT PROTOCOLS

In steps of








[Caenorhabditis elegans strain freezing procedure](#)





Part of collection

[Samuel Lab Media and Buffers](#)

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- 1 Start with  **975 mL water**
- 2  **3.0 g** NaCl
- 3  **2.5 g** Peptone
- 4  **17 g** Agar
- 5 Autoclave with stir bar
- 6 Cool to  **55 °C** (faster in water bath)
- 7 Then add the following while stirring on heat plate (DO NOT OVERHEAT)
- 8  **0.5 mL** of **[M] 1 Molarity (M)** CaCl₂ (sterile)
- 9  **1 mL** of 5 mg/mL Cholesterol (dissolved in ethanol)

- 10  1 mL of [M]1 Molarity (M) MgSO₄ (sterile)
- 11  25 mL of [M]1 Molarity (M) Potassium Phosphate Buffer, pH 6.0 (sterile)
- 12 Pour into petri dishes using sterile technique
 - 60 mm dishes = About  7 mL per plate
 - 100 mm dishes = About  20 mL per plate