

Version 4 ▼

Nov 21, 2020

© PBMC- 01b Isolation of human PBMC from Whole Blood V.4

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1 Works for me dx.doi.org/10.17504/protocols.io.bpxmmpk6

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ABSTRACT

Separation and purification of PBMC from FRESH BLOOD: list of published work using this protocol

Kustrimovic, N., Comi, C., Magistrelli, L., Rasini, E., Legnaro, M., Bombelli, R., Aleksic, I., Blandini, F., Minafra, B., Riboldazzi, G., Sturchio, A., Mauri, M., Bono, G., Marino, F., & Cosentino, M. (2018). Parkinson's disease patients have a complex phenotypic and functional Th1 bias: cross-sectional studies of CD4+ Th1/Th2/T17 and Treg in drug-naïve and drug-treated patients. Journal of neuroinflammation, 15(1), 205. https://doi.org/10.1186/s12974-018-1248-8

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Cosentino M., Ferrari M., Kustrimovic N., Rasini E., Marino F. (2015). Influence of dopamine receptor gene polymorphisms on circulating T lymphocytes: A pilot study in healthy subjects. Human immunology, 76, 10, 747-752. https://doi.org/10.1016/j.humimm.2015.09.032

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PROTOCOL CITATION

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KEYWORDS

PBMC, Fresh Blood, Neuroimmune-Pharmacology, Parkinson's Disease, Cell isolation, Primary cell culture

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OWNERSHIP HISTORY

Nov 21, 2020



Elisa Storelli Center for Research in Medical Pharmacology, University of Insubria

PROTOCOL INTEGER ID

44749

MATERIALS TEXT

MATERIALS

Healthcare Catalog #17144003-500 ml

(FBS) EuroClone Catalog #ECS0180L-500 ml

⊠ RPMI

1640 EuroClone Catalog #ECM 0495L- 500 ml

Aldrich Catalog #T8154- 100 ml

⊠ NaCl **Sigma**

Aldrich Catalog #S9625

⊠Na2HPO4*7H2O **Merck Serono**

GmbH Catalog #1.06574.1000

⊠NaH2P04 Merck Serono

GmbH Catalog #1.06346.0500

⊠NH4Cl Merck Serono

GmbH Catalog #1.01145.1000

⊠KHC03 Merck Serono

GmbH Catalog #1.04854.500

⊠EDTA **Sigma**

Aldrich Catalog #ED2SS

Acetic Acid 100% Sigma

Aldrich Catalog #A6283

Viti Catalog #not available

Instrumentation required:

- Laminar flow hood
- Autoclave

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Separation and purification of PBMC from FRESH BLOOD: list of published work using this protocol

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have a complex phenotypic and functional Th1 bias: cross-sectional studies of CD4+ Th1/Th2/T17 and Treg in drug-naïve and drug-treated patients. Journal of neuroinflammation, 15(1), 205. https://doi.org/10.1186/s12974-018-1248-8

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BEFORE STARTING

Add an equal volume of PBS 1X and mix well.

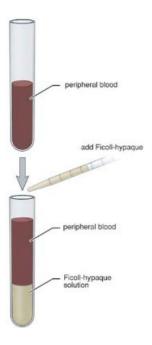
If you need to obtain **PBMC for cell culture**, make sure you are using **sterile PBS**, **culture medium**, **filtered Lysis Buffer and sterile plastic disposables as well**. Moreover, work under laminar flow hood when you are processing samples. Otherwise, use non-sterile solutions and plastic disposables, and process samples in cell isolation laboratory.

ALL REAGENTS USED IN THIS PROTOCOL MUST BE AT ROOM TEMPERATURE!

1	Put the needed amount of blood sampl into a 30 mL conical tube.	



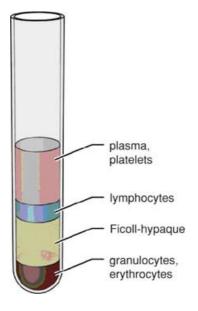
- 3 Place **□3 mL** of FICOLL in a **□15 mL** conical tube.
- Carefully layer **12 mL** of diluted blood on FICOLL with a glass Pasteur Pipette to a final volume of 15 ml as shown in the figure below.



5 Centrifuge samples **3400 x g, 00:40:00** at room temperature (RT) without break.

Allegra AVANTI 30
Centrifuge
Beckman Coulter Beckman Italy

After centrifugation, take out the tubes carefully to not disturb the mononuclear cell layer that appears as a white, cloudy band between the plasma and FICOLL as shown in the figure below.



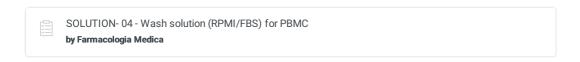
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7	\triangle
	Carefully with a glass Pasteur pipette transfer the mononuclear lymphocyte cell layer to another 15 ml conical tube.
8	Wash the isolated PBMC with PBS/FBS 2% to a final volume of □10 mL and centrifuge at ⊕600 x g, 00:10:00 at RT.
	SOLUTION- 05 - Wash solution (PBS/FBS) for PBMC by Elisa Storelli, Center for Research in Medical Pharmacology, University of Insubria
9	Remove supernatants, resuspend pellet in □1 mL of Lysis Buffer and add another □9 mL of Lysis Buffer. Immediately centrifuge tubes at ③300 x g, 00:10:00 at RT.
	SOLUTION- 06 - Lysis Buffer by Elisa Storelli, Center for Research in Medical Pharmacology, University of Insubria
10	Remove supernatant and resuspend pellet in ■10 mL of PBS/FBS 2% and centrifuge at 600 x g, 00:10:00 at RT.
	SOLUTION- 05 - Wash solution (PBS/FBS) for PBMC by Elisa Storelli, Center for Research in Medical Pharmacology, University of Insubria

Remove supernatant and resuspend the obtained pellet in **10 mL** of **RPMI/FBS 10%** for cell counting.

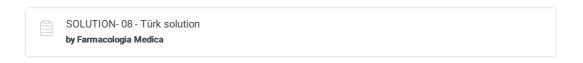
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12 For manual cell count use Türk solution for checking purity.

Follow protocol CELL COUNT- 02.

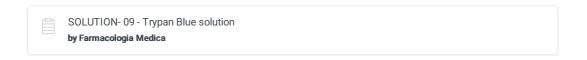


13



For automatic cell count with Cellometer machine use Trypan Blue.

Follow protocol CELL COUNT- 03.



14

If needed, check the purity of PBMC suspension by using morphological parameter of the flow cytometer.

For this test $0.5x10^6$ PBMC in 500 μl of PBS are enough.

BD FACS Celesta Flow Cytometer Becton Dickinson Milan Italy BD

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15 Expected results



VIABILITY - The expected viability by Trypan Blue should be ≥90 %.

PURITY - The PBMC suspension obtained should contain at least 80% of lymphocytes, 10-15% of monocytes and few contaminant PMN cells (\leq 5%) as confirmed by flow cytometry.

YIELD - The expected amount of PBMCs should be $\pm 28,5 \times 10^6$ starting from 25 ml of fresh blood.