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Epifluorescence-Microscopy-VLPs

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FOOD Micro UCPH



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ABSTRACT

Epifluorescence Microscopy for Viral-like Particles
-Virome Studies-

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ABSTRACT

Epifluorescence Microscopy for Viral-like Particles
-Virome Studies-

Reagents

- SYBR Gold
- Glycerol
- PBS: (0.05M Na₂HPO₄, 0.85%(wt/vol) NaCl (pH 7.5)
- 0.02µm filtered-autoclaved MilliQwater

Equipment

- 0.02µm filter whatman Al₂O₃ (Whatman: 6809-6002)
- Microscope glass slides
- Glass coverslips

- Falcon tubes
- Petri dishes
- Wipes
- Swinnex Filter holder (Millipore: SX0002500)
- 5ml syringe

Reagents Setup

- **50% glycerol/50% PBS (1:1) (Anti-fading/mounting Solution)**

Sterilized through 0.02µm membrane, autoclaved and stored in falcon tubes at -20°C.

- **SYBR Gold solution (1000x)**

975µl of 0.02µm filter-autoclaved MilliQwater

25 µl of SYBR Gold 1000x (prepared)

Prepare solution fresh-before use

Protocol

Filter Preparation

1. Filter holders (Swinnex) must be autoclaved or immersed in 70% EtOH overnight.
2. Subsequently, place the 0.02µm anodisc on top of the filter, close the lid and the filter is ready to use (**REMEMBER** to use filter forceps and wear gloves to avoid contamination).

Dilution of phages

3. Dilute 5 µl of CsCl gradient sample in 2.5 ml of 0.02µm filtered-autoclaved MilliQwater.
4. Using a 5 ml syringe, filter the diluted samples. When filtrating, be gentle with the pressure applied to avoid damage of the viral-like particles.
5. Carefully, remove anodisc (from holder) using filter forceps and place it on a clean Kimwipe to blot the back of the Anodisc filter.
6. Anodisc should be dried, by placing it on a new Kimwipe in a dark box or bench drawer. When the filter is dried, it will not be translucent (10-15 min).

Preparation of SYBR Gold for staining (while filters are drying)

7. Enumerate petri dishes according to your samples/treatments.
8. Dispense 100µl droplet of the SYBR Gold solution (1:38) on a Petri Dish.
9. Once anodiscs are dried, place one of them onto a 100 µl droplet and stain for 25-30 min on **DARK** conditions, e.g. in a dark box or bench drawer.
10. After staining, carefully pick up the anodiscs and repeat drying procedures from steps 6 and 7.
11. Thereafter, dried anodiscs are mounted on labeled glass slides. Add 15 µl of mounting medium (ANTI-FADING solution) onto the glass to facilitate the filter positioning. Subsequently, 15 µl of mounting medium (ANTI-FADING solution) are also deposited on top of the anodisc to facilitate attachment of the glass coverslip (Noble & Fuhrman, 1998).
12. Sample is ready to be observed though the FM.