

Oct 03, 2024

## Immunocytochemistry

DOI

**[dx.doi.org/10.17504/protocols.io.5jyl82ym8l2w/v1](https://dx.doi.org/10.17504/protocols.io.5jyl82ym8l2w/v1)**

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**Protocol Citation:** Michael Henderson, Naman Vatsa 2024. Immunocytochemistry. **protocols.io**  
**<https://dx.doi.org/10.17504/protocols.io.5jyl82ym8l2w/v1>**

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**Protocol status:** Working

**We use this protocol and it's working**

**Created:** October 02, 2024

**Last Modified:** October 03, 2024

**Protocol Integer ID:** 108928

**Funders Acknowledgement:**  
**Aligning Science Across**  
**Parkinson's**

## Abstract

This is a protocol for immunocytochemistry for 24 and 96 well cell culture plates.

## Materials

A	B	C	D
Vendor	Catalog#	Qty	Description
RPI	A30075-1 00.0	1	Bovine Serum Albumin
Electron Microscopy Services	15713	10 X 10ml	20% Paraformaldehyde



## Preparation

- 1 Prepare 1x phosphate buffer saline (PBS)
- 2 Prepare fixation solution: 4% paraformaldehyde/4% sucrose in PBS

### Note

can be stores at 4°C for up to 1 month

- 3 Prewarm the fixative solution to 37 degrees Celsius
- 4 Prepare blocking buffer: 3% Bovine Serum Albumin (BSA) in PBS
  - Seal the conical with parafilm to prevent evaporation and store at 4 degrees Celsius
- 5 Prepare 10% stock of Triton-X-100 in PBS
- 6 Prepare permeabilization buffer: 0.3% TX-100 in 3% BSA in PBS

## Staining

- 7 Thoroughly aspirate media from each well without disturbing cells
- 8 Add pre-warmed fixative solution to the wells and incubate at room temperature for 15 minutes. (0.5 mL/well for 24-well plate, 100µL/well for 96-well plate).
- 9 Rinse 5X with PBS. This and subsequent washes can be done with the plate washer for 96-well plates.
- 10 Permeabilize neurons with a permeabilization buffer for 15 minutes at room temperature.



- 11 Rinse 3X with PBS.
- 12 Block neurons for a further 50 minutes with a blocking buffer at room temperature.
- 13 Dilute primary antibodies in blocking buffer.
- 14 Incubate cells with primary antibody for 2 hours at room temperature or overnight at 4°C.
- 15 Rinse 5X with PBS.
- 16 Dilute Alexa Fluor conjugated secondary antibodies corresponding to primary antibodies in blocking buffer in 1:500 dilution
- 17 Incubate cells in secondary antibodies for 1 hour at room temperature.

**Note**

Make sure that the secondary antibodies will not recognize each other or crossreact with the same primary antibodies!

- 18 Rinse 5X with PBS.
- 19 Mount coverslips onto glass slides with Prolong Gold mounting media. 96-well should be incubated with 100 µL DAPI 1:10,000 in PBS.
- 20 Seal the 96-well plate with a black seal.
- 21 Visualize staining on a fluorescent microscope.