



VERSION 2  
SEP 28, 2023

OPEN ACCESS



**DOI:**  
[dx.doi.org/10.17504/protocols.io.81wgbxnk3lpk/v2](https://dx.doi.org/10.17504/protocols.io.81wgbxnk3lpk/v2)

**Protocol Citation:** NUS iGEM 2023. DNA Isolation (Gel Clean-up). **protocols.io** <https://dx.doi.org/10.17504/protocols.io.81wgbxnk3lpk/v2> Version created by NUS iGEM

**License:** This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

**Protocol status:** Working  
We use this protocol and it's working

**Created:** Sep 28, 2023

## DNA Isolation (Gel Clean-up) V.2

NUS iGEM<sup>1</sup>

<sup>1</sup>National University of Singapore

NUS iGEM 2023



NUS iGEM  
National University of Singapore

### ABSTRACT

2023 NUS-Singapore iGEM team followed this protocol to isolate the DNA fragments from the agarose gel after the gel electrophoresis.

### PROTOCOL MATERIALS

Buffer QG Qiagen Catalog #19063 Step 5

Buffer PE Qiagen Catalog #19065 In [3 steps](#)

### SAFETY WARNINGS

- Proper lab PPE must be worn at all times.
- When using the LED transilluminator, wear eyewear that is designed to block blue light to protect the eyes.

**Last Modified:** Sep 28, 2023





**PROTOCOL integer ID:**  
88520




**Keywords:** DNA Isolation,  
DNA, Gel, Gel  
Electrophoresis, Buffer QG,  
Buffer PE






- 1 Prepare and label an Eppendorf tube.
- 2 Place the agarose gel onto the LED Transilluminator (blue light) to observe the DNA band(s).

#### Safety information


- Wear protective eyewear to protect the eyes from blue light.
- Turn off the LED transilluminator immediately when it is not in use.

- 3 Cut out the target DNA band from the agarose gel.
- 4 Put the gel piece into the Eppendorf tube.
- 5 Add  450 µL of  Buffer QG Qiagen Catalog #19063 into the Eppendorf tube.
- 6 Heat the Eppendorf tube at  55 °C for  00:20:00 in the Thermo-Shaker. 20m

- 7 Add  150 µL of 100% isopropanol (IPA) into the Eppendorf tube and shake the tube to mix the solution well.
- 8 Transfer the whole solution into a QIAquick Spin Column (purple tube with a maximum volume of  750 µL ).
- 9 Centrifuge it for  13 rpm, 00:01:00 . 1m
- 10 Discard the flow-through and place the QIAquick column back into the same tube.
- 11 Add  700 µL of  Buffer PE QIAGEN Catalog #19065 into the QIAquick column.
- 12 Centrifuge it for  13 rpm, 00:01:00 .
- 13 Discard the flow-through and place the QIAquick column back into the same tube.
- 14 Add  700 µL of  Buffer PE QIAGEN Catalog #19065 again into the QIAquick column.

- 15 Centrifuge it for  13 rpm, 00:01:00 .
- 16 Discard the flow-through and place the QIAquick column back into the same tube.
- 17 Centrifuge the emptied QIAquick column at  13 rpm, 00:01:00 to remove residual  Buffer PE Qiagen Catalog #19065 . 1m
- 18 Transfer the QIAquick column into the newly labelled Eppendorf tube.
- 19 Add  30  $\mu$ L of DI water into the QIAquick column.
- 20 Centrifuge the tube at  13 rpm, 00:01:00 , ensuring that the direction of the Eppendorf tube's cap is the same as the direction of spinning to avoid breaking. 1m
- 21 Discard the QIAquick column, the solution left in the Eppendorf tube contains the DNA fragment of interest.
- 22 Use the Nanodrop to measure and record the purity and concentration of the DNA fragment.

Equipment	
<b>NanoDrop™ One/OneC Microvolume UV-Vis Spectrophotometer</b>	NAME
UV-Vis Spectrophotometer	TYPE
Thermo Scientific	BRAND
ND-ONE-W	SKU

**23** Keep the isolated DNA fragment in the  Room temperature .