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are credited **Protocol status: Working**

original author and source

We use this protocol and it's working

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GFP-sacB Characterization

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ABSTRACT

2023 NUS-Singapore iGEM team followed this protocol to characterise their New Composite Part "GFP-sacB" with IPTG and sucrose solution at various concentrations.

MATERIALS

- LB media
- M9 Media
- Correct Antibiotics
- IPTG Solution
- Sucrose Solution
- DI Water

SAFETY WARNINGS



- Proper lab PPE must be worn at all times.
- Since cells are used in this protocol, a Biosafety Cabinet (BSC) is required to ensure safety.



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PROTOCOL integer ID:

88978

Keywords: sacB, GFP, Negative Selection, Negative Selection Marker, Characterization

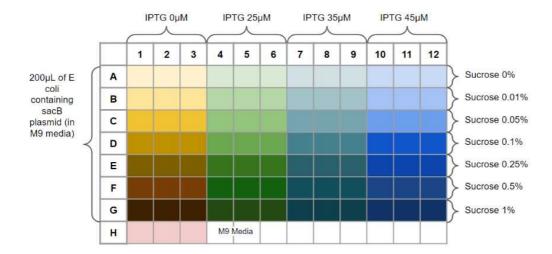
Cell Inoculation and Incubation (Day Before Characterization)

- 1 Inoculate cells with GFP-sacB gene from the cell stock.
- 2 Add \bot 5 mL of LB media and \bot 5 μ L of the appropriate antibiotic to a Falcon tube.
- 3 Incubate the Falcon tube at \$\ 37 \circ\$ in an incubator.

Sample Preparation (96-well Plate)

- 4 Decide the concentrations of IPTG and sucrose solutions for characterization.
- **5** Prepare new Falcon tube(s).
- Add \bot 5 mL of M9 media, \bot 5 μ L of the appropriate antibiotic, and \bot 100 μ L of cells cultured the previous day.
- 7 Add the required volume of IPTG to reach the desired concentration.

- 8 Incubate the cells for 02:00:00 at 37 °C
- 9 Prepare a sterile 96-well plate.
- Design an appropriate plate map, each sample (with a particular IPTG and sucrose concentration) must be repeated 3 times. Example of plate map:



One of the actual plate map used by the NUS-Singapore iGEM 2023 team when characterizing the GFP-sacB gene.

- Adjust sucrose concentration by adding DI water and sucrose solution to each well (final volume of $20~\mu L$) according to the plate map.

Characterization and Plate Reader Reading

- Place the 96-well plate into the plate reader. 14
- 15 Create a protocol in the plate reader's software according to the following setting:

A	В
Plate Type	96 WELL PLATE
- Set Temperature	Setpoint 37°C
	Preheat before moving to next step
Start Kinetic	Runtime 6:10:00 (HH:MM:SS), Interval 0:30:00, 13 Reads
- Shake	Orbital: Continuous
	Frequency: 282 cpm (3 mm)
Read 1	Absorbance Endpoint
	Full Plate
	Wavelengths: 600
	Read Speed: Normal, Delay: 100 msec, Measurements/Data Point: 8
Read 2	Fluorescence Endpoint
	Full Plate
	Excitation: 485, Emission: 528
	Optics: Top, Gain: 100
	Light Source: Xenon Flash, Lamp Energy: High
	Read Speed: Normal, Delay: 100 msec, Measurements/Data Point: 10
	Read Height: 7 mm
End Kinetic	

- 16 Start the continuous reading.
- 17 After the reading is complete, remove the 96-well plate from the plate reader.
- 18 Save the data as a CSV file for future analysis.
- 19 Discard the used 96-well plate in a biohazard bin.