

Oct 26, 2020

3D Immunostaining for CLARITY-processed samples

Seth Currlin¹

¹University of Florida



dx.doi.org/10.17504/protocols.io.bnzdmf26

Human BioMolecular Atlas Program (HuBMAP) Method Development Community Tech. support email: Jeff.spraggins@vanderbilt.edu

Seth Currlin

ABSTRACT

This is a guide for immunostaining CLARITY-processed samples. dx.doi.org/10.17504/protocols.io.8jihuke

These steps are meant to be a guide for immunostaining large samples and should be optimized to suit your particular tissues and reagents. Large tissue volumes and dense tissue types will require longer incubation and wash times. The parameters suggested below are for a piece of human thymus tissue approximately 5 mm³ in size

A useful link: http://wiki.claritytechniques.org/index.php/Immunostaining

DO

dx.doi.org/10.17504/protocols.io.bnzdmf26

PROTOCOL CITATION

Seth Currlin 2020. 3D Immunostaining for CLARITY-processed samples. **protocols.io** https://dx.doi.org/10.17504/protocols.io.bnzdmf26

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Oct 26, 2020

LAST MODIFIED

Oct 26, 2020

PROTOCOL INTEGER ID

43781

MATERIALS TEXT

Solutions:

Wash buffer (PBST): 1x phosphate buffered saline (PBS), 0.5% Triton X-100, pH 7.4; at least 20x volume of tissue.

Blocking buffer: 1x PBS, 1% horse serum, 0.5% Triton-X 100, 0.05% Tween-20, pH 7.4; at least 10x volume of tissue. oAnother serum type should be used based on the secondary antibody.

Staining buffer: 1x PBS, 0.5% Triton X-100, 0.05% Tween-20, pH 7.4; at least 10x volume of tissue.

ABSTRACT

This is a guide for immunostaining CLARITY-processed samples. dx.doi.org/10.17504/protocols.io.8jihuke

 $\textbf{Citation:} \ \ \textbf{Seth Currlin (10/26/2020).} \ \ \textbf{3D Immunostaining for CLARITY-processed samples.} \ \ \underline{\textbf{https://dx.doi.org/10.17504/protocols.io.bnzdmf26}}$

These steps are meant to be a guide for immunostaining large samples and should be optimized to suit your particular tissues and reagents. Large tissue volumes and dense tissue types will require longer incubation and wash times. The parameters suggested below are for a piece of human thymus tissue approximately 5 mm³ in size.

A useful link: http://wiki.claritytechniques.org/index.php/Immunostaining

BEFORE STARTING

Ensure the tissue is sufficiently transparent for your imaging requirements. If the sample has not been successfully cleared then subsequent imaging will be difficult due to regions retaining light-obscuring lipids. Samples should be imaged prior to immunostaining to confirm adequate clearing, particularly among excitation/emission settings intended for later use.

Immunohistochemistry for large (5 mm³) tissue volumes 3w 5d		
1	Wash out residual clearing solution	3d
	Three washes in PBST, 24 hours each, 37C with gentle agitation.	
2	Nonspecific Epitope Blocking	3d
	Blocking buffer incubation: three days, 37C with gentle agitation.	
	Note: Serum type should be used based on the secondary antibody host species.	
3	Primary Antibody Incubation	1w
	1:100 dilution (~10 ug/mL) in Staining Buffer.	
	Primary antibody incubation: five days, 37C with gentle agitation; then two days, 4C with gentle agitation.	
4	Wash out Primary Antibody	3d
	Three washes, 24 hours each, at 37C with gentle agitation.	
5	Secondary Antibody Incubation	1w
	1:200 dilution (~5 ug/mL) in Staining Buffer.	
	Secondary antibody incubation: five days at 37C with gentle agitation; two days at 4C with gentle agitation.	
	Note: Protect samples from light.	
6	Wash out Secondary Antibody	3d
	Three washes, 24 hours each, at 37C with gentle agitation.	
	Note: Protect samples from light.	
7	Prepare for 3D Imaging or Storage	
	Prepare for imaging or store samples in washing buffer (PBST) at 4C.	
	Fac OD in a miner and a search than 1/1 day day and 1750 A/2 and a sale in branches	

For 3D imaging guide, see: https://dx.doi.org/10.17504/protocols.io.begajbse

i protocols.io 3 10/26/2020

 $\textbf{Citation:} \ \ \textbf{Seth Currlin (10/26/2020).} \ \ \textbf{3D Immunostaining for CLARITY-processed samples.} \\ \underline{\textbf{https://dx.doi.org/10.17504/protocols.io.bnzdmf26}} \\ \textbf{Citation:} \ \ \textbf{Seth Currlin (10/26/2020).} \ \ \textbf{3D Immunostaining for CLARITY-processed samples.} \\ \underline{\textbf{https://dx.doi.org/10.17504/protocols.io.bnzdmf26}} \\ \textbf{Citation:} \ \ \textbf{Seth Currlin (10/26/2020).} \ \ \textbf{3D Immunostaining for CLARITY-processed samples.} \\ \underline{\textbf{Mttps://dx.doi.org/10.17504/protocols.io.bnzdmf26}} \\ \textbf{Citation:} \ \ \textbf{Seth Currlin (10/26/2020).} \ \ \textbf{Seth Currlin (10/26/2020).} \\ \textbf{Seth Currlin (10/26/2020).} \ \ \textbf{Seth Currlin (10/26/2020).} \\ \textbf{Seth Currlin (10/26/2020).} \ \ \textbf{Seth Currlin (10/26/2020).} \\ \textbf{Seth Currlin (10/26/2020).} \ \ \textbf{Seth Currlin (10/26/2020).} \\ \textbf{Seth Currlin (10/26/2020$