

AUG 29, 2023

Qiagen RNEasy PowerMicrobiome RNA extraction kit

Michael Dan

Siemon¹, Christelle Schang et al², Jessica Pardy³,

Dilan Justin

Richard Gibson³, Joseph³, Donovan³,

christopher.degroot³

¹Univeristy of Western Ontario; ²Monash University;

³University of Western Ontario



Michael Dan Siemon Western University

DISCLAIMER

DISCLAIMER - FOR INFORMATIONAL PURPOSES ONLY; USE AT YOUR OWN RISK

The protocol content here is for informational purposes only and does not constitute legal, medical, clinical, or safety advice, or otherwise; content added to protocols.io is not peer reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol should not substitute for independent professional judgment, advice, diagnosis, or treatment. Any action you take or refrain from taking using or relying upon the information presented here is strictly at your own risk. You agree that neither the Company nor any of the authors, contributors, administrators, or anyone else associated with protocols.io, can be held responsible for your use of the information contained in or linked to this protocol or any of our Sites/Apps and Services.

OPEN ACCESS



DOI:

dx.doi.org/10.17504/protocol s.io.q26g7p181gwz/v1

Protocol Citation: Michael Dan Siemon, Christelle Schang et al, Jessica Pardy, Richard Gibson, Dilan Joseph, Justin Donovan, christopher.degroot 2023. Qiagen RNEasy PowerMicrobiome RNA extraction kit. protocols.io https://dx.doi.org/10.17504/protocols.io.q26g7p181gwz/v1

License: This is an open access protocol distributed under the terms of the Creative Commons
Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

ABSTRACT

The samples were processed using the Qiagen RNeasy PowerMicrobiome kit with the modifications described by Schang et al., 2021. As a substitute for vortexing described in the kit protocol, bead-beating was used. 100ul of phenol-chloroform-isoamyl alcohol was added to the bead beating tubes before the addition of the membranes. Bead-beating was conducted 4x for 30s at 4 m/s. After pelletizing the samples with centrifugation, the supernatant was processed through spin columns. Spin columns were incubated with DNase Digestion Solution for 15 minutes to isolate only RNA. The RNA was eluted from the spin columns by passing 50 uL of DEPC water through twice.

Protocol status: Working This protocol was used for RNA extraction of wastewater samples in April 2023. It was replaced by the Qiagen PowerFecal Pro kit.

IMAGE ATTRIBUTION

Michael Dan Siemon on behalf of ImPaKT Lab, University of Western Ontario

MATERIALS

Qiagen RNEasy PowerMicrobiome RNA extraction kit

Created: Aug 28, 2023

Last Modified: Aug 29,

2023

7055	L Integer ID:	
	Qiagen RNEasy PowerMicrobiome RNA	52m
1	Thoroughly mix wastewater sample then aliquot 40 mL into 50 mL Falcon tube. Centrifuge at 4500 x g for 20 min. Decant supernatant, assume 280 µl pellet.	20m
2	Add 100 µl phenol-chloroform-isoamyl alcohol to a PowerBead Bead Tube, Glass 0.1 mm. Place 0.25 g stool or biosolid sample into the Bead Tube.	
3	Add 650 μl PM1 and 6.5 μl β-mercaptoethanol to the PowerBead Tube	
4	As a substitute for vortexing described in the kit protocol, bead-beating was used. Bead-beating was conducted 4x for 30s at 4 m/s.	4m
5	After pelletizing the samples with centrifugation (13,000 x g for 1 min), mix the supernatant with 650 µl each of Solution PM3 and Solution PM4.	1m

6

Process the solution through MB RNA Spin Column by centrifugation (13,000 x g for 1 min).

Discard flow-through and repeat until all the solution has been processed.

3m

7 Shake solution PM5, add 650 µl to the Spin Column and centrifuge (13,000 x g for 1 min). 8 Conduct a drying step by centrifuging at 13,000 x g for 1 min to remove residual wash. 9 15m Incubate spin columns with DNase Digestion Solution for 15 minutes at room temperature to isolate only RNA. 10 Add 400 µl Solution PM7 and centrifuge at 13,000 x g for 1 min. 11 Discard flow-through. Add 650 µl Solution PM5. Centrifuge at 13,000 x g for 1 min. 12 Discard flow-through. Add 650 µl Solution PM4. Centrifuge at 13,000 x g for 1 min. 13 Conduct a drying step by centrifuging at 13,000 x g for 2 min. 2m 14 Elute RNA from the spin columns into 1.5 ml eppendorf tubes by passing 50 uL of DEPC water 2m through twice. Centrifuge each elution 13,000 x g for 1 min.