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# Passaging of cells

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[dx.doi.org/10.17504/protocols.io.ewov1n3qkgr2/v1](https://dx.doi.org/10.17504/protocols.io.ewov1n3qkgr2/v1)

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General Lab procedure, Cell passage

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Warm up culture media, PBS and Trypsin in a **37 °C Waterbath**

- 1 Inspect cells for confluency, when 70-90 % confluency is reached cells are ready for passage
- 2 Discard media and wash once with PBS

- 3 Add Trypsin to fully cover the surface of the culture vessel
- 4 Incubate for 🕒 **00:05:00** at 🌡 **37 °C** 5m
  - 4.1 Lightly tap the culture vessel and check for cell detachment. If cells are still attached prolong incubation
- 5 Add 2/3 of culture media to 1/3 of trypsin and transfer cells into a Falcon tube
- 6 Centrifuge at 🌀 **300 rcf, Room temperature, 00:05:00** 5m
- 7 Discard supernatant and resubstitute with 🧴 **1 mL culture media**
- 8 Count cells and adjust media volume for required cell density
- 9 Plate cells