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# HMW DNA extraction for diatoms

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1 Works for me

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[dx.doi.org/10.17504/protocols.io.qzudx6w](https://dx.doi.org/10.17504/protocols.io.qzudx6w)

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## ABSTRACT

Protocol for genomic DNA extraction from diatom cell culture without use of phenol/chloroform. Resulting DNA fragments were suitable for nanopore sequencing.

## DOI

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Jun 15, 2018

## LAST MODIFIED

Sep 06, 2021

## PROTOCOL INTEGER ID

13076

## MATERIALS TEXT

### MATERIALS

☒ [TAE \(TRIS-ACETATE-EDTA\) Buffer](#),

**50x Amresco Catalog #K915**

☒ [Ethyl alcohol, Pure 200 proof, for molecular biology](#) **Sigma**

**Aldrich Catalog #E7023**

☒ [RNase A \(10 mg/mL\)](#) **Thermo Fisher**

**Scientific Catalog #EN0531**

☒ [PBS](#) **Contributed by users**

☒ [Agarose](#) **Sigma**

☒ [Nuclei Lysis Solution](#),

**50ml Promega Catalog #A7941**

☒ [Protein Precipitation Solution](#),

**25ml Promega Catalog #A7951**

☒ [Isopropanol, molecular grade](#) **Sigma**

**Aldrich Catalog #I9516**

☒ [Tris-EDTA buffer, pH 8.0, molecular grade](#) **Sigma**

**Aldrich Catalog #93283**

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### Harvest cells

1. Harvest approximately  $7 \times 10^7$  -  $2 \times 10^8$  cells from mid-exponential phase by centrifugation (5000xg, 3 minutes)
2. Wash pelleted cells once with 1X PBS (5000xg, 3 minutes) and remove the supernatant

### Cell lysis and RNase treatment

2. 1. Resuspend cells in 600  $\mu$ L Nuclei Lysis Solution (Promega #A7941)
2. Freeze in dry ice or liquid nitrogen for 3 minutes and thaw in 65°C heat block or water bath for 3 minutes (or until completely thawed)
3. Repeat freeze/thaw cycle 3X
4. Heat to 65°C for 15 minutes
5. Let sample cool to room temperature and add 1.2  $\mu$ L of 10 mg/ml RNase A (ThermoFisher #EN0531)
6. Incubate for 15 minutes at 37°C in heat block or water bath
7. Let sample cool to room temperature



Be aware that rapid temperature changes can cause pressure to build up inside epi tubes, causing the lids to pop open. Use safe lock epi tubes or lid locks for the freeze/thaw step.

#### Protein precipitation

- 3
1. Add 250  $\mu$ L Protein Precipitation Solution and mix well (Promega # A7951)
2. Centrifuge (16,000xg) for 40 minutes at 4°C
3. Transfer supernatant to fresh tube

#### DNA precipitation

- 4
1. Precipitate DNA by adding 700  $\mu$ L 100% isopropanol, mix well by inversion
2. Centrifuge (16,000xg) for 30 minutes at room temperature
3. Wash DNA pellet with 600  $\mu$ L freshly prepared 70% molecular grade ethanol
4. Centrifuge (16,000xg) for 1 minute and remove all ethanol from the pellet (use quick spin and a smaller pipette tip if necessary to remove all residual ethanol)
5. Air dry DNA pellet for 10-15 minutes
6. Resuspend DNA pellet in ~25-50  $\mu$ L TE buffer, depending on pellet size and desired DNA concentration
7. Let DNA pellet resuspend in TE buffer at room temperature overnight (HMW gDNA takes longer to dissolve)
8. Determine DNA concentration by Nanodrop

After isopropanol precipitation, DNA pellet may appear as a translucent smear on the side of the tube. Pellet becomes more visible (white) following the ethanol wash step.

#### Gel electrophoresis

- 5
1. Run 100-200 ng DNA on a 0.8% agarose gel

Example data:



L = ladder (top band = 40kb)  
 1. *Phaeodactylum tricornutum*  
 2. *Thalassiosira pseudonana*  
 3. *Chaetoceros muelleri*



Species	Yield (ng/uL)	260/280	260/230
<i>Thalassiosira pseudonana</i>	1255.8	1.8	1.86
<i>Phaeodactylum tricornutum</i>	673.8	1.85	2.07

Chaetoceros muelleri	1066.5	1.89	1.49
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