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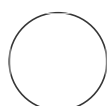
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Protocol status: Working
 We use this protocol and it's working

🌐 EdU Staining Protocol in juvenile *F. hepatica*

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Paul McCusker

ABSTRACT

This protocol describes the process of labelling and whole mount staining proliferative cells in *in vitro* juvenile *Fasciola hepatica* with the thymidine analogue 5-ethynyl-2'-deoxyuridine (EdU). The protocol is based on the *Schistosoma mansoni* EdU staining protocol developed by the Collins and Newmark groups. A full copy of that protocol is available here -

http://collinslab.org/PDF/Whole_mount_EdU_Smansoni.pdf.

GUIDELINES

Fixation

While flat fixing is necessary for worms grown for more than three weeks *in vitro* younger worms may be suitable for free fixing in 4%FA.

MATERIALS

CS50 - 50%

⊗ Chicken Serum, New Zealand Origin Thermo Fisher Catalog #16110082 and

50% ⊗ RPMI 1640 Medium, no phenol red Thermo Fisher Catalog #11835105

with x1

⊗ Antibiotic Antimycotic Solution (100x), Stabilized Merck MilliporeSigma (Sigma-Aldrich) Catalog #A5955

PBSTx - 1X PBS + 0.3%

⊗ Triton X-100 Merck MilliporeSigma (Sigma-Aldrich) Catalog #T8787-50ML

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85723

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Funders

Acknowledgement:

LIVER FLUKE MOTOR
FUNCTION AND PARASITE
CONTROL: EXPLOITING A
'TARGET VALIDATION
TOOLBOX' AS A DRUG
SCREEN-INTERFACE FOR
FLUKICIDE DISCOVERY

Grant ID: BB/K009583/1


Probing in vivo parasite
biology in vitro

Grant ID: NC/N001486/1








Exploiting stem cell biology
for liver fluke control

Grant ID: BB/T002727/1

4%FA -  500 μ L

 Formaldehyde solution Merck MilliporeSigma (Sigma-
Aldrich) Catalog #F8775-25ML


with  4 mL PBSTx

4%PFA - Add  8 g Paraformaldehyde to  100 mL millipore water, stir/heat in
a flow hood for 1 h between  55 °C and  60 °C on hotplate (**NB. must NOT
go above 60°C; if it does, start again**). Following 1 h heating turn off heat and add
NaOH dropwise until solution goes clear before adding  100 mL 2X PBS. and
mix. Store at  4 °C for 2 weeks or  -20 °C in aliquots for 6 months.

EdU Detection Solution


	A	B	C	D	E
Reagent		200 (μ L)	1 ml (μ L)	5ml (ml)	10ml (ml)
PBS		158	789	3.945	7.89
100mM Copper Protectant in Water (Store 4oC)		2	10	0.05	0.1
10mM AlexaFluor (488 or 545)/6-FAM azide		0.2	1	0.005	0.01
500mM Ascorbic Acid (Make fresh – 88 mg in 1 ml PBS)		40	200	1	2

Copper protectant -

 Copper(II) sulfate Merck MilliporeSigma (Sigma-
Aldrich) Catalog #451657

Alexafluor 488 -

 Azide-fluor 488 Merck MilliporeSigma (Sigma-
Aldrich) Catalog #760765

6-FAM azide -  6-Fam Azide Metabion

DAPI solution - 1:1000 of  1 mg/mL stock in PBSTx

PROTOCOL MATERIALS



Azide-fluor 488 Merck MilliporeSigma (Sigma-Aldrich) Catalog #760765

Materials



6-Fam Azide Metabion Materials



Formaldehyde solution Merck MilliporeSigma (Sigma-Aldrich) Catalog #F8775-25ML

Materials



VECTASHIELD Mounting Medium Vector Laboratories Catalog #H-1000

In 2 steps



Triton X-100 Merck MilliporeSigma (Sigma-Aldrich) Catalog #T8787-50ML

Materials



RPMI 1640 Medium, no phenol red Thermo Fisher Catalog #11835105

Materials



Antibiotic Antimycotic Solution (100×), Stabilized Merck MilliporeSigma (Sigma-Aldrich) Catalog #A5955

Materials



Copper(II) sulfate Merck MilliporeSigma (Sigma-Aldrich) Catalog #451657

Materials



Chicken Serum, New Zealand Origin Thermo Fisher Catalog #16110082

Materials

SAFETY WARNINGS


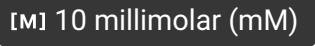
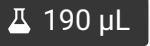


EdU Mutagen

As a thymidine analogue EdU is known for germ cell mutagenicity and reproductive toxicity. Particular care should be taken when disposing of the chemical and around pregnant mothers.

In vitro addition of EdU to juvenile *F. hepatica*


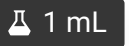
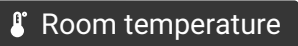

1d

- 1 Juvenile liver fluke are cultured in 50% Chicken serum (CS50) as described in McCusker et al. (2016).
- 2  10 μL of  10 millimolar (mM) 5-Ethynyl-2'-deoxyuridine (EdU) is mixed with  190 μL pre-warmed CS50 and added to worms for between 6 and 24 h.

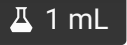
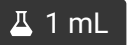
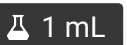

Fixation


STEP CASE

Stain immediately following EdU incubation  15 steps


- 3 Add  20 μL **4% FA/4% PFA** to a petri dish.
- 4 Pipette worms in into **4% FA/4% PFA** and immediately flatten with coverslip and weight (e.g. half full 15 mL falcon tube) for 10 min.
- 5 Add **4% FA/4% PFA** to float coverslip. Lift coverslip and move worms into 1.5 mL eppendorf with  1 mL **4% FA/4% PFA**. Place tube on a rotator for further 10 min (4%FA)/2 h (4%PFA) at  Room temperature **OR** O/N at  4 °C .

OPTIONAL Storage Step

- 6 Wash in  1 mL **PBSTx** for 10 min.
- 7 Wash in  1 mL 1:1 **PBSTx:Methanol** for 10 min.
- 8 Store in  1 mL methanol at  -20 °C until required.

9 Rehydrate in  1 mL 50:50 **PBSTx**:Methanol for 10 min.

Staining


10 x2 washes in  1 mL **PBSTx** for 10 min each.




11 Incubate in  1 mL fresh **Proteinase K Solution** for 15 min at  Room temperature .

12 Postfix in  1 mL **4%FA** for 10 min.

13 Wash in  1 mL **PBSTx** for 10 min.

14 Incubate in **EdU Detection Solution** for 30 min at  Room temperature . **NB: cover with tinfoil from this point on.**

15 x2 washes in  1 mL **PBSTx** for 5 min each

16 Incubate in  1 mL **DAPI Solution** for 20 min at  Room temperature or O/N at  4 °C .

17

Rinse in **PBSTx** and mount in



VECTASHIELD Mounting
Medium Metabion Catalog #H-1000