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# Bacterial Transformation - Mix & Go Competent Cells - CHEM 584

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THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Adapted from

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SAFETY WARNINGS

You should follow standard BSL-1 procedures when using this protocol.

BEFORE STARTING



Get an aliquot of the Mix &amp; Go competent cells from the -80 Freezer (ask a TA to get them for you)

- 1 Prewarm culture plates in a **37 °C** incubator for **00:30:00**.
- 2 Add **1 µl** to **5 µl** plasmid DNA to a **50 µl** aliquant of thawed competent cells (Mix & Go) on ice, mix gently for a few seconds (try to keep the added volume of DNA less than 5% of the total). Place tube on ice for **00:05:00**

minutes.



When selecting with Kanamycin, Tetracycline, etc., an outgrowth performed in SOC medium is required for efficient transformation. After the transformation mixture has incubated on ice for at least 5 min, add 4 volumes of SOC (e.g., 400 µl of SOC to 100 µl of the transformation mixture) and incubate for 1 hour at 37°C with gentle shaking at 200-300 rpm.

- 3 Spread  50 µl to  100 µl of the mixture onto a pre-warmed (37°C) LB plate containing Ampicillin (or other selection antibiotic).



Using chilled LB plates will decrease the transformation efficiency.

- 4 Incubate overnight at  37 °C in the bacterial incubator.



You can also incubate on the bench or in a drawer at room temperature for a longer time if this is convenient for scheduling. For example, leaving a plate on the bench for 2-3 days at room temperature usually generates nice sized colonies and you avoid having to come into the lab the next day (e.g., over the weekend).