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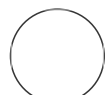
VU TIS Multimodal Molecular Imaging Pipeline for KPMP Biopsy Interrogation

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KPM

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ABSTRACT

This protocol provides an overview of the Multimodal Molecular Imaging pipeline used by the Vanderbilt University Tissue Interrogation Site (VU TIS) as part of the Kidney Precision Medicine Project (KPMP). Individual protocols are contextualized within our larger workflow.

Protocol Citation: Katerina V Djambazova, Lukasz Migas, Martin Dufresne, Jamie Allen, N. Heath Patterson, Léonore Tideman, Ellie Pingry, Madeline E. Colley, Angela R.S. Kruse, Melissa Farrow, Raf Van De Plas, Jeff Spraggins 2023. VU TIS Multimodal Molecular Imaging Pipeline for KPMP Biopsy Interrogation .

protocols.io
<https://protocols.io/view/vu-tis-multimodal-molecular-imaging-pipeline-for-k-cpcyvixw>

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Protocol status: Working
We use this protocol and it's working

Created: Feb 13, 2023

Last Modified: Jul 07, 2023

PROTOCOL integer ID:
76920

Tissue Screening and Assessment

1 Asses Kidney Tissue Sections

Sample Processing Go/No-Go Criteria

A	B
Go/No-Go Criteria	Reasoning
Biopsy must be LN2 core	Other storage conditions can induce analyte suppression effect in MALDI IMS analysis
Biopsy size must be > 3mm	Smaller tissue sections are difficult to handle, section, and mount
Time to preservation must be < 10 min	Prolonged exposure prior to preservation can cause metabolite/lipid degradation

On-Site Tissue Integrity Considerations

A	B
Tissue QC	Reasoning / Data Flags
Are there any glomeruli in the biopsy section?	Ensures proper coverage of biological features
Visible cracks/folds on the tissue surface	This can lower FTU segmentation performance, due to non-segmentable features
Visible freezing artifacts on the tissue surface	Freezing artifacts can induce analyte delocalization, jeopardizing MALDI IMS spatial integrity

Sample Interrogation Assays

2 **Autofluorescence Microscopy (AF)** is collected on all tissue sections prior to IMS

AF QC Protocol: [AFQC Protocol](#)

AF protocol: [AF Microscopy](#)

CITATION

Patterson NH, Tuck M, Van de Plas R, Caprioli RM (2018). Advanced Registration and Analysis of MALDI Imaging Mass Spectrometry Measurements through Autofluorescence Microscopy..

LINK

<https://doi.org/10.1021/acs.analchem.8b02884>

3 **MALDI Imaging Mass Spectrometry (IMS)**

Matrix Deposition Protocol (Sublimation): [Tissue Washing and Matrix deposition Protocol](#)

High Spatial Resolution MALDI IMS Data Acquisition: [MALDI IMS Protocol](#)

4 **LC-MS/MS Lipidomics**

LC-MS/MS is performed on a single serial biopsy section.

Protocol: [LC-MS/MS Protocol](#)

5 Post-IMS Autofluorescence Microscopy

Protocol: [Post-IMS AF](#)

6 PAS Staining

Matrix removal and Staining: [PAS Staining Protocol](#)

Data Pre-Processing and Analysis

7 Image registration - AF, PAS and IMS

Protocol: [Image Registration](#)

CITATION

Patterson NH, Tuck M, Van de Plas R, Caprioli RM (2018). Advanced Registration and Analysis of MALDI Imaging Mass Spectrometry Measurements through Autofluorescence Microscopy..

LINK

<https://doi.org/10.1021/acs.analchem.8b02884>

8 Data Pre-Processing

Protocol: [MALDI IMS Data Pre-Processing](#)

9 Functional Tissue Unit (FTU) Segmentation

Protocol: [FTU Segmentation](#)

CITATION

Patterson NH, Neumann EK, Sharman K, Allen J, Harris R, Fogo AB, De Caestecker M, Caprioli RM, Van de Plas R, Spraggins JM (2021). Autofluorescence microscopy as a label-free tool for renal histology and glomerular segmentation. bioRxiv.

LINK

<https://doi.org/10.1101/2021.07.16.452703>

10 Analyte Annotation

Tentative lipid annotation from the MALDI IMS data

Protocol: [Tentative Lipid Annotation](#)

