

AUG 24, 2023

ABSTRACT

This is a routine protocol for extracting DNA from various fungi. This extraction method is suitable for follow-up molecular work such as PCR amplification.

OPEN ACCESS

MATERIALS

Sterile micropestles, isopropyl alcohol, ethyl alcohol, cell lysis buffer, protein precipitation buffer, elution buffer, metal scraper.

PROTOCOL MATERIALS





Protocol Citation: Angie Macias, Matthew T Kasson, Brian Lovett 2023. General Fungal DNA Extraction. protocols.io

https://protocols.io/view/gen ral-fungal-dna-extractioncy5gxy3wVersion created by Brian Lovett

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Protocol status: Working We use this protocol and it's working

Created: Aug 24, 2023

Last Modified: Aug 24,

2023

Before you begin

- 1 Turn on hot water bath, set to 65 °C
- 2 Pull two Eppendorf A 1.5 mL centrifuge tubes per sample.
- **2.1** Label both sets of tubes with (short) sample names.
- 2.2 Label one tube set for each sample with an "I" for

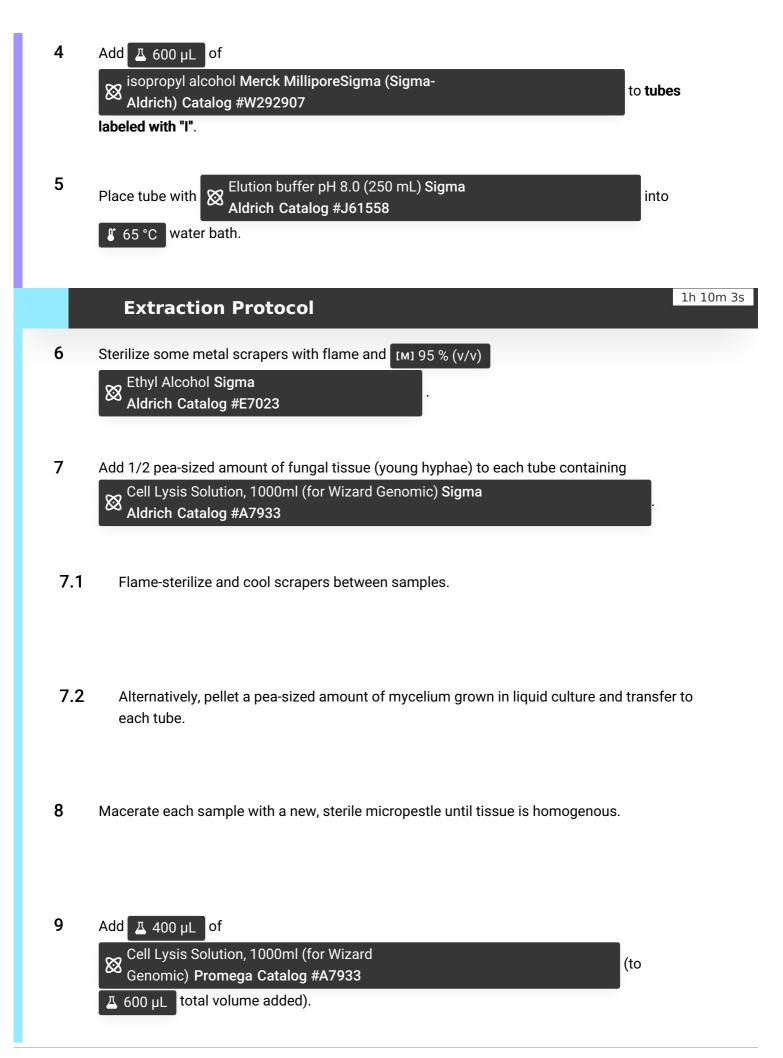


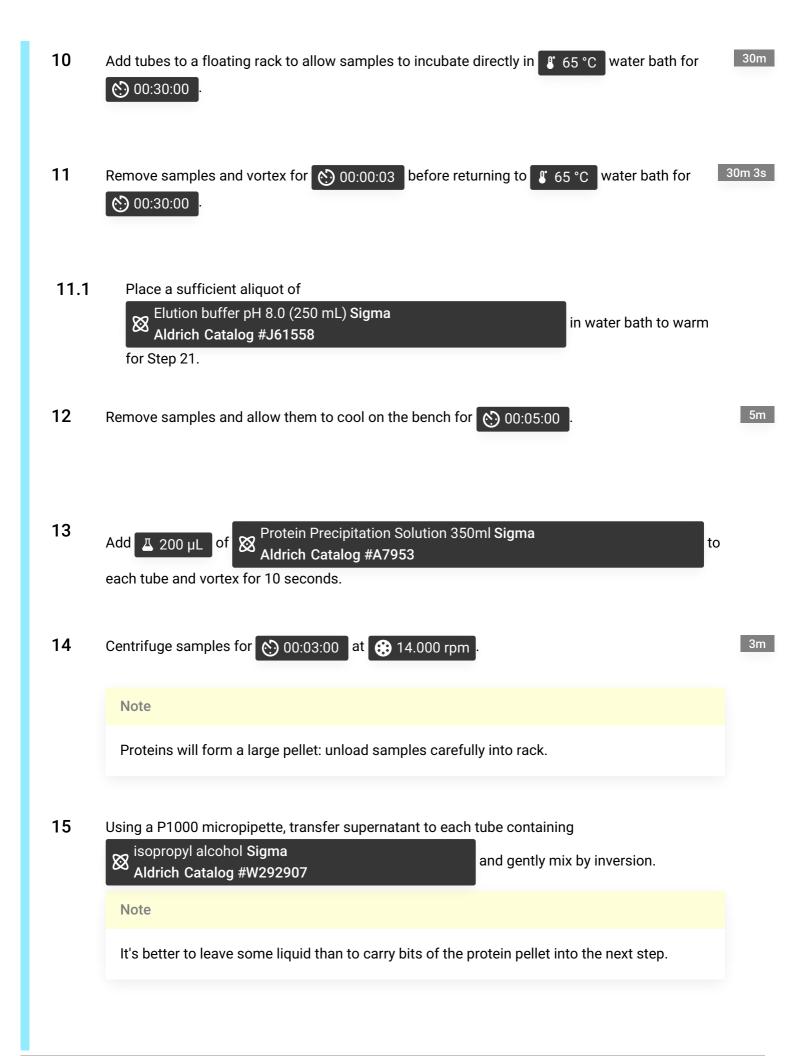


Sketch of "I"-labeled tubes (drawing from Angie Macias).

3 Add Δ 200 μL of









17 Carefully pour off the supernatant into waste container.

Note

Be careful to not lose your white DNA pellet!

- Add Add tube and mix gently by inversion.

 Ethyl Alcohol Sigma

 Aldrich Catalog #E7023

 tube and mix gently by inversion.
- 19 Centrifuge for (5) 00:01:00 at (3) 14.000 rpm

20 Repeat Step 16.

21 Open and invert tubes onto a clean paper towel.

Note

A tube rack can be placed on the tube lids to secure inverted tubes onto the paper towel.

- Add Δ 100 μL of warmed
 - Elution buffer pH 8.0 (250 mL) Sigma to each tube.

 Aldrich Catalog #J61558
- Store fully-labeled tubes in a box (not a tube rack) in the -20 °C freezer.