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# Protocol for assembling SARS-Cov-2 runs with s-aligner

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**1** Works for me [dx.doi.org/10.17504/protocols.io.bsbvnan6](https://dx.doi.org/10.17504/protocols.io.bsbvnan6)

Coronavirus Method Development Community Contignant

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SUBMIT TO PLOS ONE

## ABSTRACT

This is a protocol for using Contignant s-aligner to de-novo assemble SARS-Cov-2 genomes. S-aligner outperforms common open-source software for de novo-assembly of viruses by 110% increased performance.

## DOI

[dx.doi.org/10.17504/protocols.io.bsbvnan6](https://dx.doi.org/10.17504/protocols.io.bsbvnan6)

## PROTOCOL CITATION

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<https://dx.doi.org/10.17504/protocols.io.bsbvnan6>

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## CREATED

Feb 11, 2021

## LAST MODIFIED

Mar 29, 2021

## PROTOCOL INTEGER ID

47189

## PARENT PROTOCOLS

In steps of

[untitled protocol](#)

## Indexing and fast-testing

1

Index the reads in the run. You can use as input a FASTA file (preferred) or a FASTQ. The file can also be compressed with gzip, having a .gz extension. The script will uncompress the file and pass it to FASTA format if it's a FASTQ file.

**`./index.sh your_run_id /mnt/c/your_run.fastq.gz`**

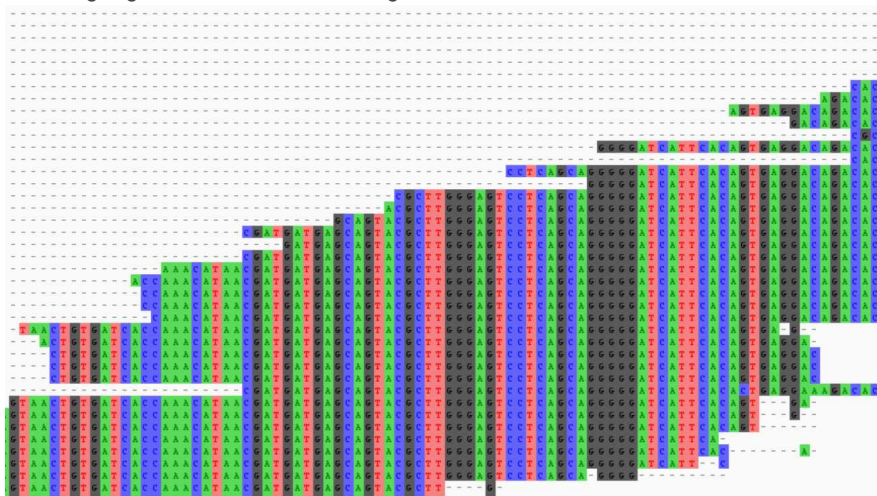
### Assemble

```
./assemble.sh your-run-id -output alignments 500 > results/your-run-id-500.fa
```

Assemble up to processing 500 reads



results/your-run-id-500.fa should have the largest contig bigger than 1.200bp and at least one of the files containing aligned reads to form a contig should look like that:



Step 2 includes a Step case.

**<1.200bp**

**>1200bp**

step case

**<1.200bp**

The largest contig in your-run-id-500.fa is shorter than 1200bp

Map your reads to a reference genome to see what's going wrong.

```
./map.sh your-run-id sequences/your-run-id/sra_data.part-71.fa reference.fa 4000  
> results/map-your-run-id.fa
```

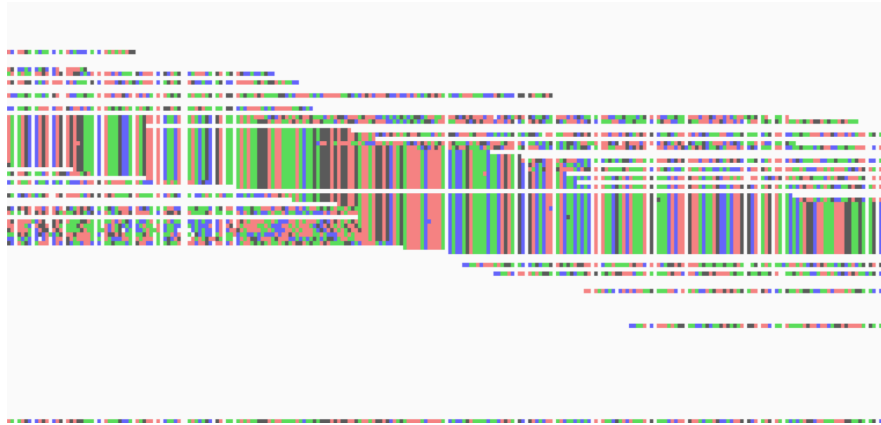
Step 3 includes a Step case.

**Data not trimmed**

Too many chimeras  
No overlapping  
Runs are short  
Contamination

step case

### Data not trimmed



If the data is not trimmed and has adaptors at the extremes of the runs it will look like the image above.

4 Trim your data and start again.

[go to step #1](#)