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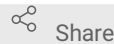
Primary cortical mixed culture

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ABSTRACT

To obtain cortical culture, pregnant mice were anesthetized, embryos were dissected, and cortex was collected in PBS. Tissues were incubated in 0.25% trypsin–EDTA at 37°C for 15 min. Trypsinized tissue was transferred into a high-glucose DMEM/F12 medium supplemented with 10% FBS. After centrifugation (1500rpm, 5 min), the pellet was resuspended. Cells were plated onto poly(L-lysine)-coated 24-well plates at 10⁶ cells per well and cultured in NB-A.

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KEYWORDS

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- 1 For primary cortical mixed culture - Use C57BL/6J mice at embryonic day 17

- 2 Anesthetized pregnant mice (1% sodium pentobarbital, 80mg/kg), dissect their embryos and collect the cortex.
(Separate and remove the soft membrane and blood vessels, rinse the cerebral cortex in PBS, and use the ophthalmic scissor to cut pieces of the cortex)

- 3 Collect the cortices in PBS in a 50 ml tube on ice
(The 50 ml tube contains 30 ml of PBS) ⚡ On ice 📦 5 mL) ⚡ On ice

- 4 Transfer the cortices to 15 ml tubes containing 1.5 ml trypsin–EDTA (0.25%) and incubate it at ^{15m}
⚡ 37 °C for ⌚ 00:15:00 Dissociate the cortices by triturating with a 10 mL serological pipette 10 – 15 times

- 5 Centrifuge the dissociated cortices (⚙️ 1500 rpm , ⌚ 00:05:00) and resuspend the pellet in ^{5m}10ml

[🔗 DMEM, high glucose Thermo Fisher](#)
Scientific Catalog #11965092
 medium supplemented with 10%
[🔗 Fetal Bovine Serum, certified, heat inactivated, United States Thermo](#)
Fisher Catalog #10082147

- 6 Seed the cells onto poly(L-lysine)-coated 24-well plates at a cell density of 1×10^6 cells/well containing specialized

[🔗 Neurobasal-A Medium Thermo Fisher](#)
Scientific Catalog #10888022 (NB-A)
 supplemented with 10%
[🔗 Fetal Bovine Serum, certified, heat inactivated, United States Thermo](#)
Fisher Catalog #10082147

- 7 After 5 days of culture, these cells are ready for further experiment

