

Dec 07, 2020

Protein Concentration Determination using Qubit

In 1 collection

Steven J Burgess¹

¹University of Illinois at Urbana-Champaign

In Development This protocol is published without a DOI.

Ag SynBio Lab UIUC



University of Illinois at Urbana-Champaign

ABSTRACT

Procedure for quantification of protein concentration. This version is adapted for use with samples that have been extracted in protein extraction buffer as part of processing soybean/cowpea samples, but the general outline is similar for other samples. See the manual for further details: https://assets.thermofisher.com/TFS-Assets/LSG/manuals/Qubit_Protein_Assay_UG.pdf

PROTOCOL CITATION

Steven J Burgess 2020. Protein Concentration Determination using Qubit. protocols.io https://protocols.io/view/protein-concentration-determination-using-qubit-bqhkmt4w

COLLECTIONS (i)

Immunoblot Analysis of Leaf Tissue

LICENSE

 $_{ extstyle }$ This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Dec 07, 2020

LAST MODIFIED

Dec 07, 2020

PROTOCOL INTEGER ID

45324

PARENT PROTOCOLS

Part of collection

Immunoblot Analysis of Leaf Tissue

MATERIALS TEXT

- Qubit® protein assay kit (Life Technologies; Q33211)
- Qubit® assay tubes (Life Technologies; Q32856)
- Qubit® 4 Fluorometer (Life Technologies; Q33238)

ABSTRACT

Procedure for quantification of protein concentration. This version is adapted for use with samples that have been extracted in protein extraction buffer as part of processing soybean/cowpea samples, but the general outline is similar for other samples. See the manual for further details: https://assets.thermofisher.com/TFS-Assets/LSG/manuals/Qubit_Protein_Assay_UG.pdf

Create Working Solution for Analysis

Create a working solution of Qubit assay buffer by diluting the reagent 1:200 in the provided buffer.

1

The final volume in each tube must be 200 μ L. Each standard tube requires 190 μ L of Qubit® working solution, and each sample tube requires anywhere from 180–199 μ L. Prepare sufficient Qubit® working solution to accommodate all standards and samples.

Create Sample Dilution for Analysis

15m 6s

2 Dilute sample 1:50 (\square 196 μ l dH₂0 + \square 4 μ l sample)

After taking into consideration the Qubit dilution factor (1:20; 10 μ L sample + 190 μ L qubit assay buffer) the sample being analyzed has been diluted 1:1000. This should give a value in the linear range for qubit (1.25-25 μ g/mL). This should also reduce the impact of SDS on the quantification, which must be >0.2% in the 10 μ L sample added (conc SDS in PEB is 2 %, so diluted 1:50 yields 0.04 %)

- 3 Add 190 μl of Qubit working solution to a fresh Qubit assay tube, one for each sample to be analyzed (including the three protein standards)
- 4 Add 110 μl of protein standard to the appropriate tube and mix by vortexing © 00:00:03 s.

Add $\Box 10 \mu I$ of diluted sample to the appropriate tube and mix by vortexing $\bigcirc 00:00:03$ s.

3s

3s

15m

6 Allow samples to incubate at room temperature for © 00:15:00

Measure protein sample concentration using the Qubit, following the instructions on the machine (i.e. start by measuring the standard curve).

Remember to adjust measured values according to the dilution factor applied. In this example, after taking into consideration the Qubit dilution factor (1:20; $\Box 10 \ \mu I$ sample + $\Box 190 \ \mu I$ Qubit assay buffer) the sample has been diluted 1:1000

Qubit[™] 4 Fluorometer, with WiFi Fluorometer

Invitrogen

Q33238