

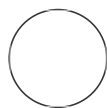


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Lentivirus preparation for neuronal transdifferentiation

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ABSTRACT

This is a protocol for the preparation of Lentiviruses.

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Protocol status: Working
We use this protocol and it's working

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Preparation of culture medium and plasmids

- 1
 1. HEK293T medium @4C lasts for 1 month. A total of 500 mL:
 - 435 ml DMEM (High glucose, GlutaMAX) (Thermo Fisher Scientific)
 - 50 ml Fetal bovine serum (FBS) (Thermo Fisher Scientific)
 - 5 ml Penicillin-Streptomycin (Thermo Fisher Scientific)
 - 5 ml HEPES (1M; Thermo Fisher Scientific)
 - 5 ml Sodium pyruvate (100 mM; Thermo Fisher Scientific)
 - Sterilized by filtration through a 0.22 µm filter
 2. Virus medium @4C lasts for 1 month. A total of 500 mL:
 - 470 ml DMEM (High glucose, GlutaMAX) (ThermoFisher Scientific)
 - 10 ml Fetal bovine serum (FBS) (Thermo Fisher Scientific)
 - 5 ml Penicillin-Streptomycin (Thermo Fisher Scientific)
 - 5 ml MEM NEAA (100X; Thermo Fisher Scientific)
 - 5 ml HEPES (1M; Thermo Fisher Scientific)
 - 5 ml Sodium pyruvate (100 mM; Thermo Fisher Scientific)
 - 0.5 ml beta-mercaptoethanol (Thermo Fisher Scientific)
 - Sterilized by filtration through a 0.22 µm filter
 3. OptiMEM+GlutaMAX (Thermo Fisher Scientific)
 4. Lipofectamine 2000 (Thermo Fisher Scientific)
 5. Plasmids:
 - Transfer: FUW-M2rtTA (Addgene plasmid# 20342), pTet-O-Ngn2-puro (Addgene plasmid# 52047), Tet-O-FUW-Ascl1 (Addgene plasmid# 27150), Tet-O-FUW-Brn2 (Addgene plasmid# 27151), Tet-O-FUW-Myt1l (Addgene plasmid# 27152)
 - Packaging: psPAX2 (Addgene plasmid# 12260)
 - Envelope: pMD2.G (Addgene plasmid# 12259)
 6. Millex syringe 0.22 & 0.45 µm filter

Generation of Lentiviruses

- 2 Seed 6×10^6 HEK293T cells in a 10-cm dish for each transfer plasmid and culture to reach 80% confluence next day.
- 3 Prepare DNA and Lipofectamine 2000 dilutions.
 - 5 µg Transfer plasmid
 - 4 µg Packaging plasmid
 - 2.5 µg Envelope plasmid
 - DNA diluted into 300 µL OptiMEM
 - 23 µL Lipofectamine 2000, diluted into 300 µL OptiMEM
- 4 Vortex DNA and Lipofectamine dilutions, centrifuge for 5 sec and incubate at room temperature for 5 mins.
- 5 Combine DNA and Lipofectamine dilutions, vortex, centrifuge and incubate the mixture for additional 15 mins.
- 6 Remove old HEK293T medium, rinse the cells with OptiMEM once and then add 5 mL OptiMEM to each dish.
- 7 Add a total of 600 µL of Lipofectamine/DNA mixture to the cells dropwise across the whole dish, gently shake in vertical × horizontal direction a few times and return the dishes to the incubator.
- 8 After 6-hr incubation, remove the transfection medium and add 9 mL of fresh Virus medium to each dish.
- 9 For additional 24-hr incubation, collect the viral particle-containing medium from each dish and store at 4°C.

- 10 After medium collection, add fresh 8 mL of pre-warm Virus medium to each dish.
- 11 After 24-hr incubation, collect the medium and pool it with the first collection.
- 12 The cells can now be bleached and disposed of accordingly.
- 13 Seal the pooled medium with parafilm and centrifuge in the swinging bucket at 400xG for 5 mins at 4°C to pellet cell debris.
- 14 Carefully take the cleared supernatant and pass through a 0.45 µm filter.
- 15 Viral supernatant is stored at 4°C until use and good for 2 weeks.