



AUG 09, 2023

## Preparing whole cell samples for immunoblot analysis

Louise Uoselis<sup>1</sup>

<sup>1</sup>Lazarou Lab, WEHI



Louise Uoselis  
WEHI

### ABSTRACT

Protocol for preparation of HeLa cell lysates for immunoblot analysis.

OPEN  ACCESS



#### DOI:

[dx.doi.org/10.17504/protocols.io.e6nvwdj4dlmk/v1](https://dx.doi.org/10.17504/protocols.io.e6nvwdj4dlmk/v1)

**Protocol Citation:** Louise Uoselis 2023. Preparing whole cell samples for immunoblot analysis.

**protocols.io**

<https://dx.doi.org/10.17504/protocols.io.e6nvwdj4dlmk/v1>

**License:** This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited





**Protocol status:** Working  
We use this protocol and it's working

**Created:** Aug 09, 2023

**Last Modified:** Aug 09,  
2023

**PROTOCOL integer ID:**  
86214

**Keywords:** ASAPCRN

- 1 Add an appropriate volume of 1x LDS Sample Buffer (ThermoFisher) to each sample.
- 2 Boil each sample at  99 °C with shaking at maximum speed for  00:10:00 10m
- 3 Allow all samples to cool to  Room temperature, and quickly centrifuge the samples to collect all liquid in the bottom of the tube. Vortex each sample for ~3 seconds to ensure homogeneity.
- 4 Measure the concentration of each sample spectroscopically, blanking with the 1x LDS sample buffer and using an A280 measurement. Dilute samples with 1x LDS Sample buffer to a concentration of < 7 mg/mL if required.
- 5 Aliquot out the desired amount of each sample into a separate tube, and add 1x LDS Sample Buffer to each sample to make all samples in the same gel a standard volume.
- 6 Either freeze samples at  -20 °C until required, or load directly onto a SDS-PAGE gel.