

JUL 28, 2023

Cell surface biotinylation

rosanne.wouters¹, Peter Vangheluwe¹

¹KU Leuven



rosanne.wouters

ABSTRACT

The protocol describes cell surface biotinylation to identify plasma membrane localized proteins in cell culture via western blotting.

OPEN ACCESS



DOI:

dx.doi.org/10.17504/protocol s.io.x54v9d9o4g3e/v1

Protocol Citation: rosanne. wouters, Peter Vangheluwe 2023. Cell surface biotinylation. **protocols.io** https://dx.doi.org/10.17504/protocols.io.x54v9d9o4g3e/v1

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Protocol status: Working We use this protocol and it's working

Created: Jun 01, 2023

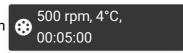
Last Modified: Jul 28, 2023

PROTOCOL integer ID:

82746

Keywords: ASAPCRN	Kevw	ords:	ASAPCRN
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- 1 grown cells until 70-80% confluency in 10cm dish
- 2 place cells on ice and wash with ice-cold PBS
- 3 incubate cells for 30 min c on ice with PBS containing 2.5 mg/ml Sulfo-NHS-SS-biotin (Pierce).
- 4 stop the biotinylation reaction by washing 3 times for 5min with quenching solution (0.5% BSA and 100 mM glycine in PBS).
- collect cells by scraping in PBS and centrifugation



- 6 lyse cells by resuspending cell pellet RIPA buffer supplemented with protease inhibitor and incubating for 30min on ice
- 7 clear cell lysate by centrifugation (20min, 14000 gavg, 4°C) and collect supernatant
- 8 isolate biotinylated proteins with immobilized Neutravidin beads

5m

- **8.1** wash neutravidin beads 2 times with RIPA buffer
- 8.2 add supernatant from step 7 to beads
- 8.3 incubate while rotating head-over-head © 02:00:00 at 4°
- **8.4** wash neutravidin beads 3 times with RIPA buffer
- 9 Input and bound proteins were processed for Western blotting with 4x LDS loading buffer.
- 9.1 remove all RIPA buffer from neutravidin beads
- 9.2 add 4x LDS loading buffer
- 9.3 denature proteins for 10min, 70°C

10 proceed with Western blotting