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Cell migration assay or Transwell assay

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1 Works for me dx.doi.org/10.17504/protocols.io.bgd9js96

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1 Cells are washed in cold PBS, detached with 5mM EDTA, centrifuged at 1200 g for 7 min and resuspended in serum-free medium.

2 Transwell chambers (Corning Costar) are prepared filling with 600 µl of complete medium the lower chamber.

3 The desired amount of cells is plated in 100 µl of serum-free medium in the upper insert (8.0 µm pore size) of each Transwell chamber and incubated at 37°C.

4 After 24 hours at 37°C, fill with 600 µl of 5mM EDTA an empty well beside each used Transwell chamber.

5 Eliminate DMEM from the upper insert and move it inside an EDTA well.

6 Collect DMEM from the lower chamber and put some EDTA inside, to detach cells possibly fallen from the upper insert.

7 Detach cells migrated on the underside of the upper insert using a plastic scraper and collecting them in the EDTA well.

8 Mix in one tube cells collected from the lower chamber and scraped from the insert and centrifuge at 1200 g for 7 min, to count the migrated cells.

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