

Oct 13, 2020

DNA-Extraction-ClostridiaProject

Tobias Wandt¹

¹Universität Leipzig

Works for me

This protocol is published without a DOI.

Tobias Wandt

ABSTRACT

Protocol used for DNA-Extraction of Clostridia in my project.

DNA-Kit used is GenFind V3 by Beckmann/Coulter.

Some steps used from:

Sim JH, Anikst V, Lohith A, Pourmand N, Banaei N. Optimized Protocol for Simple Extraction of High-Quality Genomic DNA from Clostridium difficile for Whole-Genome Sequencing. J Clin Microbiol. 2015;53(7):2329-2331. doi:10.1128/JCM.00956-15

PROTOCOL CITATION

Tobias Wandt 2020. DNA-Extraction-Clostridia Project. protocols.io https://protocols.io/view/dna-extraction-clostridiaproject-bnb9mar6

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Oct 12, 2020

LAST MODIFIED

Oct 13, 2020

PROTOCOL INTEGER ID

43105

ABSTRACT

Protocol used for DNA-Extraction of Clostridia in my project.

DNA-Kit used is GenFind V3 by Beckmann/Coulter.

Some steps used from:

Sim JH, Anikst V, Lohith A, Pourmand N, Banaei N. Optimized Protocol for Simple Extraction of High-Quality Genomic DNA from Clostridium difficile for Whole-Genome Sequencing. J Clin Microbiol. 2015;53(7):2329-2331. doi:10.1128/JCM.00956-15

Growing and Preparing Clostridia 2d

Inoculate Clostridia in 10ml TSB-Broth, incubating at least 48h in anaerobe atmosphere at 37°C

2d

■10 mL TSB broth (§ 48:00:00 at 37°C anaerobic

Take 1ml of Clostridia TSB-Broth and centrifuge for 15min at 3,000g, after that remove supernatant

15m

■1 mL Clostridia TSB broth © 00:15:00 centrifuge at 3000g

- 3 Add 500ul Lysis-Buffer(LBB) and 30ul Proteinase K, tipmixing or vortexing to resuspend the pellet
 - ■500 µl LBB ■30 µl Proteinase K
- 4 Incubate at 57°C for 1h while continously shaking at ~500/min

© 01:00:00 at 57°C + shaking 500/min

Add 300ul vortexed Black Beads(BBB) and tipmix(at least 10 times), incubate for 5 min at room temperature, place on a magnetic tube stand until no or few beads are left in suspension ~10-15min, then carefully remove supernatant without disturbing the beads-formation

1h

5m

■300 µl Black Beads © 00:05:00 at room temperature © 00:15:00 magnetic tube stand

6 Add 800ul WBB and tipmix(at least 10 times), place on a magnetic tube stand until no or few beads are left in suspension ~5min, remove supernatant

■800 µl WBB © 00:05:00 magnetic tube stand

- 7 ogo to step #6 for a total of 2 WBB washes
- 8 Add 800ul WBC and tipmix (at least 10 times), than add 600ul WBC and tipmix, place on a magnetic tube stand until no or few beads left in suspension \sim 5min, remove supernatant

□1400 µl WBC © 00:05:00 magnetic tube stand

- 9 go to step #8 for a total of 2 WBC washes
- Add 100ul distilled water, tipmix 10 times, incubate for 2min at roomtemp, tipmix 10 times again and place in a 4°C ^{2m} fridge overnight

□100 µl distilled water \bigcirc 00:02:00 room temperature \bigcirc 0 vernight at 4°C

- 11 Tipmix 10times and place on a magnetic tube stand until no beads are left in suspension
- 12 Take the supernatant with your DNA and put it in a fresh tube