



Dec 24, 2021

Chelex DNA extraction Protocol

Sarah Chang¹, Michael Russello¹

¹University of British Columbia

1



dx.doi.org/10.17504/protocols.io.byvhpw36

Sarah Chang

Laboratory protocol for DNA extraction from animal tissue with Chelex.

DOI

dx.doi.org/10.17504/protocols.io.byvhpw36

<https://doi.org/10.1371/journal.pone.0261966>

Sarah Chang, Michael Russello 2021. Chelex DNA extraction Protocol.

protocols.io

<https://dx.doi.org/10.17504/protocols.io.byvhpw36>



protocol

Chang SL, Ward HGM, Russello MA (2021) Genotyping-in-Thousands by sequencing panel development and application to inform kokanee salmon (*Oncorhynchus nerka*) fisheries management at multiple scales. PLoS ONE 16(12): e0261966. doi: [10.1371/journal.pone.0261966](https://doi.org/10.1371/journal.pone.0261966)

protocol ,

Oct 07, 2021

Dec 24, 2021

53897

- 1 Make up enough 10% Chelex for entire plate (195 µL /individual + slop) (ie. 1 g Chelex: 9 mL dH2O)

- 2 Place a magnetic stir bar in the jar containing your 10% Chelex solution. The Chelex powder is very heavy and needs to be constantly stirred in order to maintain consistency.
- 3 Pipette **195 μ L** of Chelex into each well of a 96-well plate.
- 4 Add approximately 1mm² piece of tissue (usually operculum punch or adipose fin clip) in each well. Be careful when doing this, as it is easy to mess up.
- 5 Add **5 μ L** of proteinase K to each well using a repeater pipette. This should always be the last step, as proteinase K will digest itself at room temperature with no substrate.
- 6 Cover plate and incubate in the thermocycler under the following conditions: 60°C for 120 min (tissue digestion) 95°C for 15 min (heat kill for proteinase K) This protocol should be saved in most of the thermocyclers (just scroll through). Sometimes this is not sufficient for complete digestion, so a second addition of 5 μ L of proteinase K followed by a repeat of the incubation procedure may be necessary. Alternatively, you could increase the digestion period from 120 min to 240 min. This is what I usually do and seems to work quite well.
- 7 Once digestion is complete, vortex briefly to break up any remaining pieces of tissue, then vortex at 1500 rpm for 2 min to collect Chelex and any other remaining solids on the bottom of the well.
- 8 Pipette off the supernatant and add to a new 96 well. This will be your undiluted DNA stock that will be stored for future use. Store at -20°C in chimney well plates with caps.
- 9 Make up a 1:10 dilution plate to be used for the assays. Usually **200 μ L** is sufficient to run all assays. Add **180 μ L** of dH₂O to each well, and then add **20 μ L** of your undiluted stock. Vortex and spin to finish the procedure. The 1:10 dilution plate can be stored in the fridge to avoid multiple freeze/thaw cycles.