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# Relative quantification of mRNA transcript levels by qPCR

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This method describes isolation of RNA from cultured cells, generation of cDNA, and relative quantification of transcript levels by qPCR.

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### Solutions to prepare:

#### DMEM solution:

FBS	10%
Penicillin	100 U/ml
Streptomycin	100 mg/ml
L-glutamine	2 mM

### Cell culture and treatments

3d

- 1 Culture the HeLa-M cells at **37 °C** in 5% CO<sub>2</sub> and DMEM containing 10% FBS, **100 U/ml** penicillin, **100 mg/mL** streptomycin, and **2 Milimolar (mM)** L-glutamine (all from Gibco).
- 2 For any given experiment, plate the cells at such density so as to be approximately 90% confluent at the time of lysis.
- 3 For experiments using siRNA, transfect 60 pmols of the indicated siRNA using **6 µL** Lipofectamine RNAiMax (ThermoFisher) in Opti-MEM (Gibco) per well according to manufacturer protocol. Lyse the cells **72:00:00** after siRNA transfection.

3d

### Cell lysis, RNA purification, and qPCR

- 4 Aspirate media from cells and rinse cells with PBS **On ice**.
- 5 Isolate RNA using RNeasy Micro Plus kit (Qiagen) according to manufacturer's protocol.
- 6 Generate cDNA from **1 µg** purified RNA using iScript cDNA synthesis Kit (Bio-Rad)

according to manufacturer's protocol.

- 7 Dilute the iScript reaction to a total of **400  $\mu\text{L}$**  Sterile Water (American Bio).



Combine **10  $\mu\text{L}$**  SYBR Green Master Mix (BioRad) with **6.78  $\mu\text{L}$**  Sterile Water (American Bio) per sample.



Combine **16.78  $\mu\text{L}$**  diluted SYBR Green Master Mix with **0.61  $\mu\text{L}$**  each of **10 Micromolar ( $\mu\text{M}$ )** forward and reverse primers per sample. Pipette this mixture into wells of 96-well qPCR plate. Perform at least two technical replicates for each sample.



Pipette **2  $\mu\text{L}$**  of diluted RNA from step 7 in well with SYBR Green Master Mix.

- 11 Cover plate with Optical Adhesive Covers (Applied Biosystems).



Spin down plate in table top centrifuge.



Run qPCR in CFX96 Real-Time System (BioRad) using the following protocol:

A	B	C
95 °C	3 min	Repeat 39x
95 °C	10 sec	
55 °C	10 sec	
72 °C	30 sec	
95 °C	10 sec	
65 °C	5 sec	
95 °C	5 sec	

### Data analysis

- 14 Subtract the housekeeping gene (b-actin) mean threshold cycle (Ct) values from transcript of interest mean Ct values to calculate  $\Delta C_t$ .
- 15 Subtract the  $\Delta C_t$  of the control sample from each sample  $\Delta C_t$  to calculate the  $\Delta\Delta C_t$  value.
- 16 Calculate relative expression using the  $2^{-\Delta\Delta C_t}$  method.