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# *Anthoceros agrestis* (hornwort) transformation v01

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Works for me

[dx.doi.org/10.17504/protocols.io.6izhcf6](https://dx.doi.org/10.17504/protocols.io.6izhcf6)

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## ABSTRACT

*Anthoceros agrestis* (hornwort) transformation

pre-print:

<https://biorxiv.org/cgi/content/short/2021.01.07.425778v1>

This protocol works better for the Oxford strain

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## PROTOCOL CITATION

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## CREATED

Aug 14, 2019

## LAST MODIFIED

Jan 10, 2021

## PROTOCOL INTEGER ID

26937

## GUIDELINES

Be patient...

## MATERIALS TEXT

### KNOP recipe:

#### Stock 1

25g/L KH<sub>2</sub>PO<sub>4</sub>

#### Stock 2

25g/L KCl

#### Stock 3

25g/L MgSO<sub>4</sub> 7H<sub>2</sub>O

#### Stock 4

100g/L Ca(NO<sub>3</sub>)<sub>2</sub> 4H<sub>2</sub>O

autoclave and store at RT or 4°C

### KNOP solid working solution:

In 600 mL of water add:

10ml Stock 1

10ml Stock 2

10ml Stock 3

10ml Stock 4

12.5mg FeSO<sub>4</sub>7H<sub>2</sub>O

pH to 5.8 with KOH

top up water to 1L after adjusting pH

add 7 gr of Gelzan - G1910 - CAS Number [71010-52-1](#)

#### **KNOP liquid working solution:**

In 600 mL of water add:

10ml Stock 1

10ml Stock 2

10ml Stock 3

10ml Stock 4

12.5mg FeSO<sub>4</sub>7H<sub>2</sub>O

20 gr of sucrose (2% w/v final concentration)

pH to 5.8 with KOH

top up water to 1L after adjusting pH

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Sterile disposable scalpels (#0501, Swann Morton)

Homogenizer (#727407, IKA Ultra-Turrax T25 S7 Homogenizer)

Dispensing tools (#10442743, IKA Dispensing Element)

100 µm cell strainer (#352360, CORNING),

6-well plate (#140675, ThermoFisher)

3',5'-dimethoxy-4'-hydroxyacetophenone (acetosyringone) (#115540050, Acros Organics, dissolved in dimethyl sulfoxide (DMSO) (#D8418, SIGMA))

Cefotaxime (#BIC0111, Apollo Scientific)

Hygromycin (#10687010, Invitrogen)

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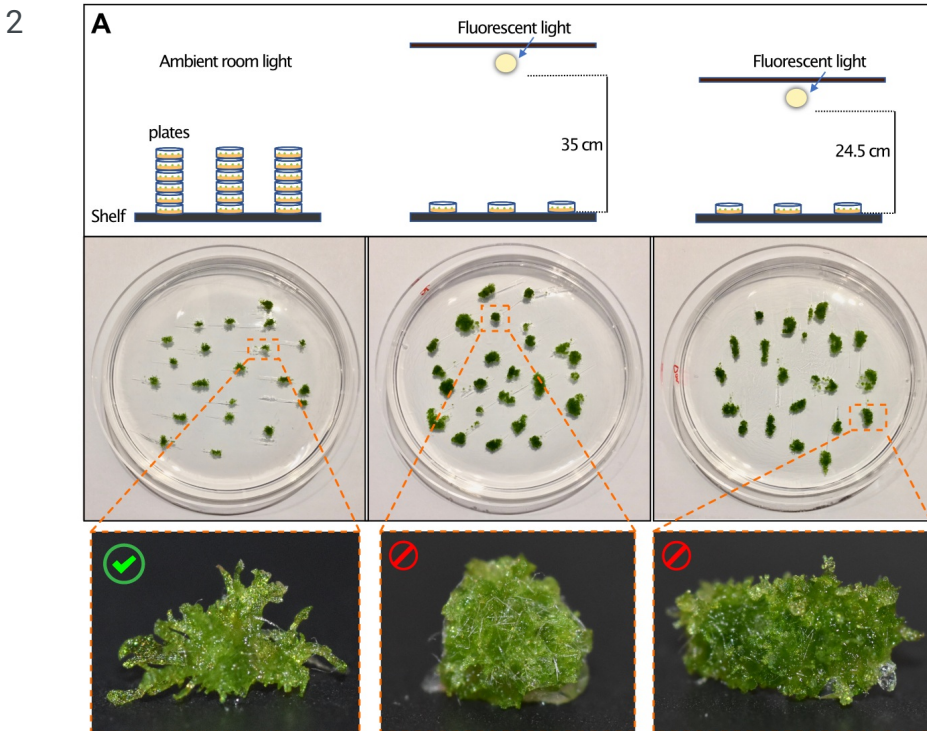
This protocol works better for the Oxford strain

BEFORE STARTING

Pray

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**IMPORTANT:** The light intensity used to cultivate *A. agrestis* tissue is a very critical factor for successful transformation. Tissue should be grown under low light intensity ( $3\text{--}5\ \mu\text{mol m}^{-2}\text{s}^{-1}$ ) and should have a morphology similar to the tissue in **Figure 1** bottom left image.



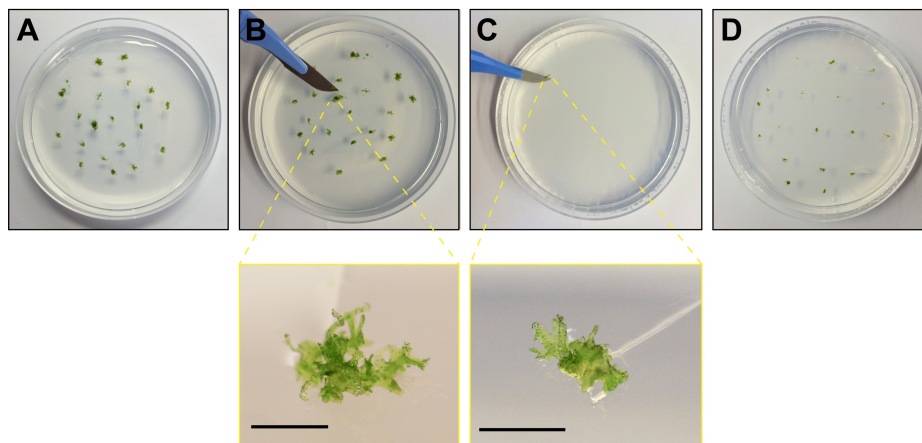
**Figure 1: Effect of light on *A. agrestis* growth**

Examples of *A. agrestis* tissue grown under different light regimes. From left to right: i) tissue morphology when plates were stacked in the laboratory under ambient room light ( $3\text{--}5\ \mu\text{mol m}^{-2}\text{s}^{-1}$ ), ii) tissue morphology under light supplemented by fluorescent tubes,  $35\ \mu\text{mol m}^{-2}\text{s}^{-1}$ , light intensity (PHILIPS, TL-D58W/835) and iii) tissue morphology under light supplemented by fluorescent tubes,  $20\ \mu\text{mol m}^{-2}\text{s}^{-1}$  light intensity (PHILIPS, TL-D 36W/840).

**Tissue similar to (i) is optimal for transformation. Use of tissue similar to (ii) and (iii) should be avoided.**

3 Axenic cultures of *A. agrestis* gametophytes can be routinely propagated by monthly sub-culturing as shown in **Figure 2**.

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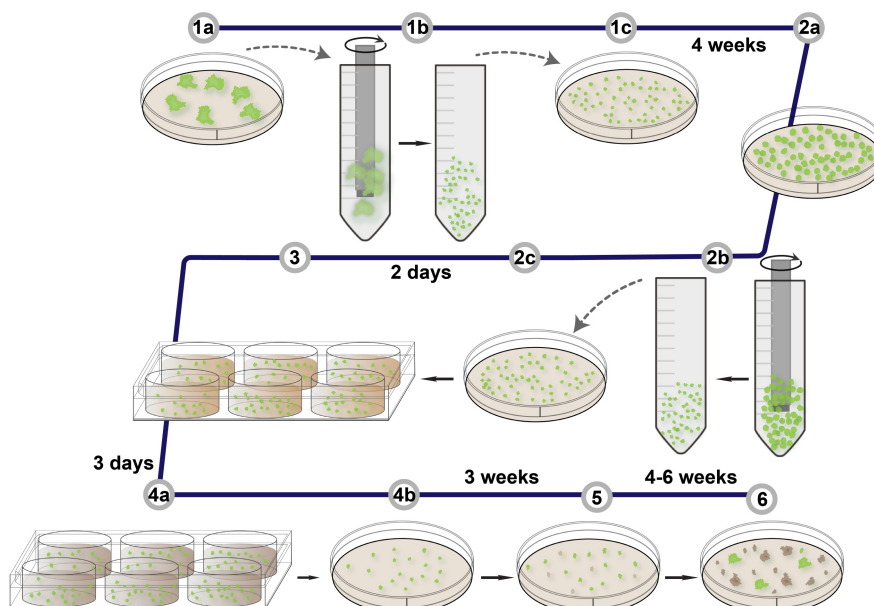


**Figure 2: *A. agrestis* growth and tissue culturing**

For sub-culturing, a small piece of thallus tissue is cut using sterile disposable scalpels and placed on plates containing fresh growth medium. Scale bars: 2 mm. Petri dish dimensions: 92 x16 mm.

Tissue similar to the bottom images is optimal for transformation

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**Figure 3: Transformation pipeline outline**

**1a-c:** Tissue is homogenized, transferred on growth medium, and placed under low light conditions. **2a-c:** After 4 weeks, the tissue is homogenized again and grown for two additional days. **3:** The tissue is co-cultivated with *Agrobacterium* for three days and then **4a-b:** spread on appropriate antibiotic-containing growth medium. **5:** After 3 weeks, the tissue is transferred again onto freshly prepared antibiotic-containing growth medium for a second round of selection. **6:** After approximately 4-8 weeks, putative transformants are visible.

## 6 Tissue preparation:

- Collect approximately 2 g of thallus tissue grown for 4 weeks under low light intensity (approximately 0.1 g of tissue per petri dish - 20 petri dishes in total) - **Figure 4.1**
- Split tissue into three parts, transfer each part into a 50 mL falcon tubes containing 15 mL of sterile water and homogenize using a homogenizer and corresponding dispensing tools (for 5 sec, lowest power 8000 rpm) - **Figure**

#### 4.2-5

- Transfer the homogenized tissue from one falcon tube into a 100 µm cell strainer, wash with 50 mL of sterile water and transfer onto two petri dishes (six plates in total) containing solid KNOP medium add 2-3 mL of sterile water and place at 21°C under 12 hours light and 12 hours dark, light intensity 3-5 µmol m<sup>-2</sup>s<sup>-1</sup> - **Figure 4.6-7**
- After 3-4 weeks transfer the tissue from the petri dishes into a falcon tube using sterile scalpels - **Figure 4.8-9**
- Re-homogenize in 15-20 mL of sterile water and filter using a cell strainer - **Figure 4.10-11**
- Spread out again onto 4 petri dishes with solid KNOP medium (if necessary, to facilitate spreading of the tissue, add 2 mL of sterile water to the petri dish) and allow to grow for 2 days at 21°C under continuous light (12 hours light and 12 hours dark also ok) (light intensity 30 µmol m<sup>-2</sup>s<sup>-1</sup>) - **Figure 4.12**

### 7 **Agrobacterium culture preparation:**

- Inoculate 5 mL LB media with a single *Agrobacterium* colony (AGL1 strain), 10 µg/mL rifampicin, 50 µg/mL carbenicillin and the plasmid-specific selection antibiotic.
- Incubate the preculture at 28°C for 2 days at 120 rpm.

### 8 **Co-cultivation:**

- Centrifuge 5 mL of 2 d *Agrobacterium* culture (OD: 2.5 - 2.7) for 7 min at 1800 xg.
- Remove supernatant and re-suspend in 5 mL liquid KNOP plus 2% (w/v) sucrose and 100 µM acetosyringone.
- Incubate the culture with shaking (120 rpm) at 28°C for 5 hours.
- Wash the regenerating thallus using a 100 µm cell strainer.
- Transfer ½ of the tissue from one plate into a single well of a 6-well plate containing 4 mL of liquid KNOP medium supplemented with 2% (w/v) sucrose - **Figure 4.13**
- Add 80 µL of *Agrobacterium* culture and 100 µM (final concentration) acetosyringone to the medium.
- Co-cultivate the tissue and *Agrobacterium* for 3 days with shaking at 110 rpm at 21°C with only ambient light from the room (1-3 µmol m<sup>-2</sup>s<sup>-1</sup>) - **Figure 4.14**

### 9

#### **1st selection:**

- Using a sterile plastic pipette transfer the tissue from a single well into a 100 µm cell strainer, drain and then transfer on growth media containing the appropriate antibiotic. To facilitate spreading of the tissue, add 2 mL of sterile water to the petri dish - **Figure 4.15-17**
- Plate the tissue on solid KNOP plates supplemented with 100 µg/mL cefotaxime and 10 µg/mL Hygromycin and grow at 21°C under 12 hours light and 12 hours dark, 35 µmol m<sup>-2</sup>s<sup>-1</sup>

## 10 2nd selection:

- After 3 weeks transfer the tissue to fresh growth media containing 100 µg/mL cefotaxime and 10 µg/mL Hygromycin. To facilitate spreading of tissue on the petri dish add 2 mL of sterile water. Grow at 21°C under 12 hours light and 12 hours dark, 35 µmol m<sup>-2</sup>s<sup>-1</sup> - **Figure 4.18**
- After 6-8 weeks successful transformants are visible on the petri dish (successful transformants can be identified using a microscope after 4 weeks selection based on rhizoid production) - **Figure 4.19 and Figure 5 & 6.**

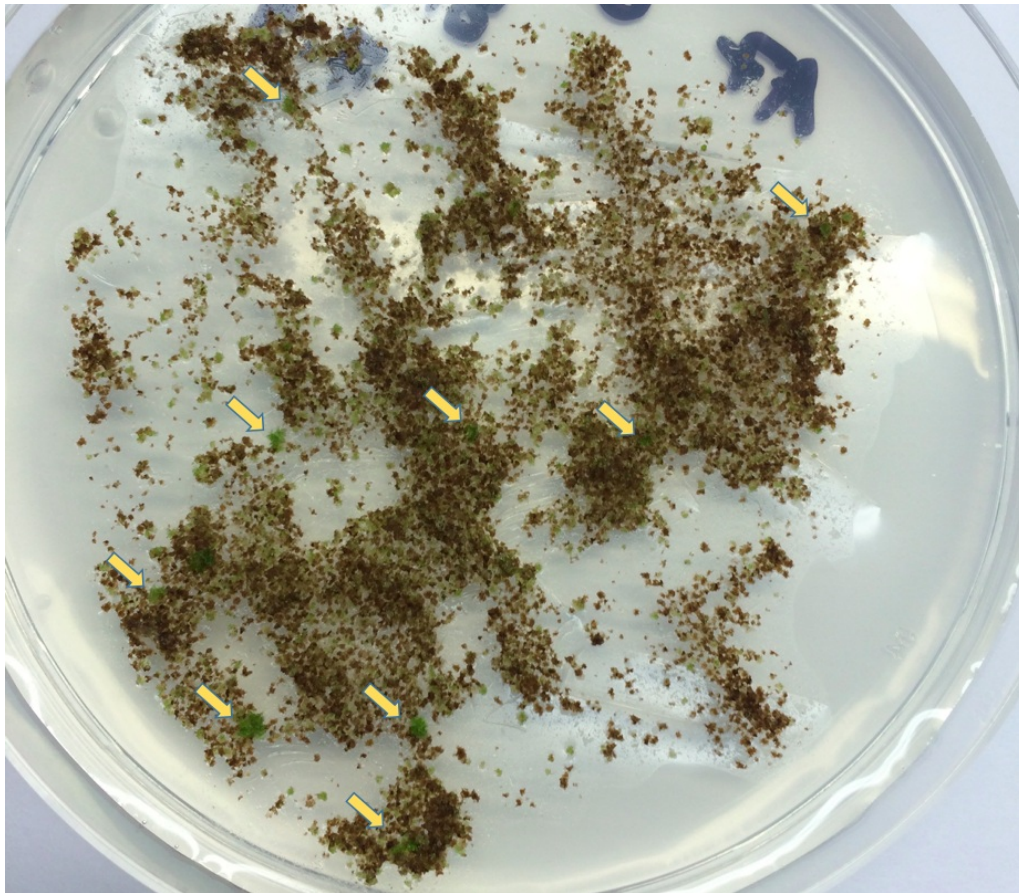
## 11 3rd selection:

To eliminate false positives, transfer surviving tissue fragments again on 100 µg/mL cefotaxime and 10 µg/mL Hygromycin containing growth media. Grow at 21°C under 12 hours light and 12 hours dark, 3-35 µmol m<sup>-2</sup>s<sup>-1</sup> - **Figure 7.**



Scale bars: 2 mm.

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**Figure 5: Example of plate with tissue after two months of selection.**

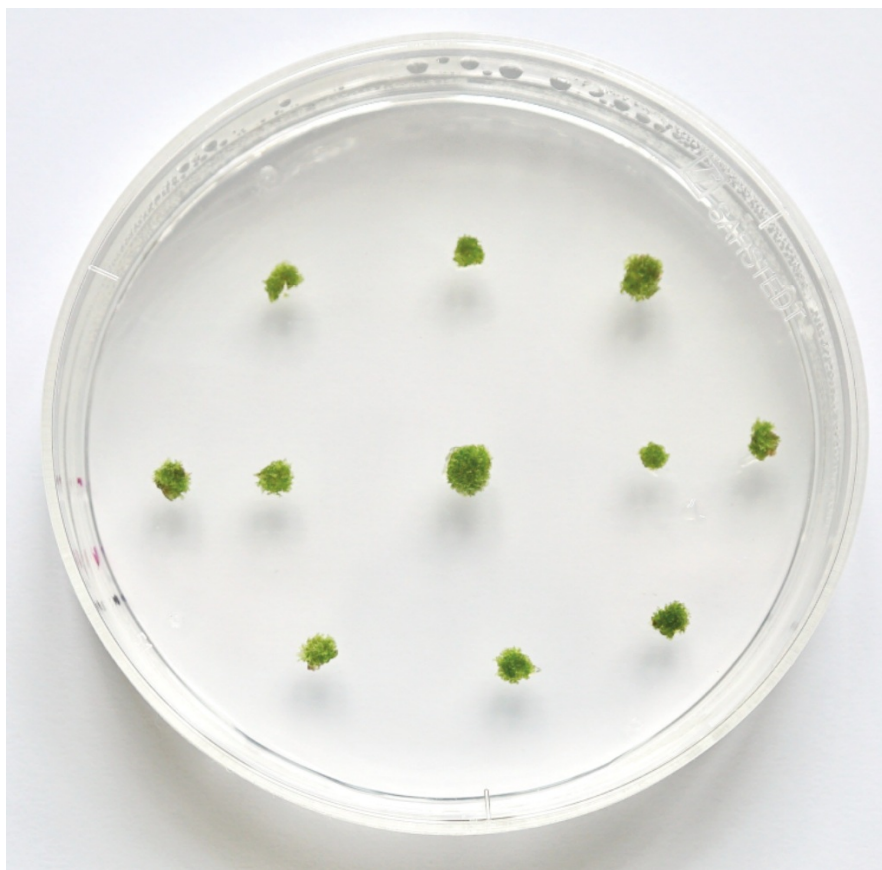
Transformants are indicated with arrows  
Petri dish dimensions: 92 x16 mm.

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**Figure 6: Transgenic regenerating thallus fragment.**  
Rhizoid indicated with an arrow head.





**Figure 7: Tissue after 3rd selection**

Petri dish dimensions: 92 x16 mm.