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Protocol status: Working
 We use this protocol and it's working

Assembly and sterilization of EcoFAB 2.0 for plant growth experiments

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ABSTRACT

The following instructions are for the sterilization, and assembly of EcoFAB 2.0 for studying small plants and their microbiomes during growth.

IMAGE ATTRIBUTION

A Fabricated ecosystem (EcoFAB 2.0) is a standardized device for reproducible aseptic plant growth, sampling of the growth medium, and determination of plant-microbe interactions. It can accommodate small model plants such as *Brachypodium distachyon* and is compatible with automated systems found in most laboratories. Photo credit: Thor Swift/Berkeley Lab (CC BY-NC-ND 4.0).

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Materials

- 1 EcoFAB 2.0 parts supplied by LBNL (quantity per device)
 - A. EcoFAB Chamber, Clear Polycarbonate (1)
 - B. EcoFAB Base, Polycarbonate (1)
 - C. Gasket, Silicone Rubber (1)
 - D. Backing Plate, Polycarbonate with 20% Glass Fiber (1)

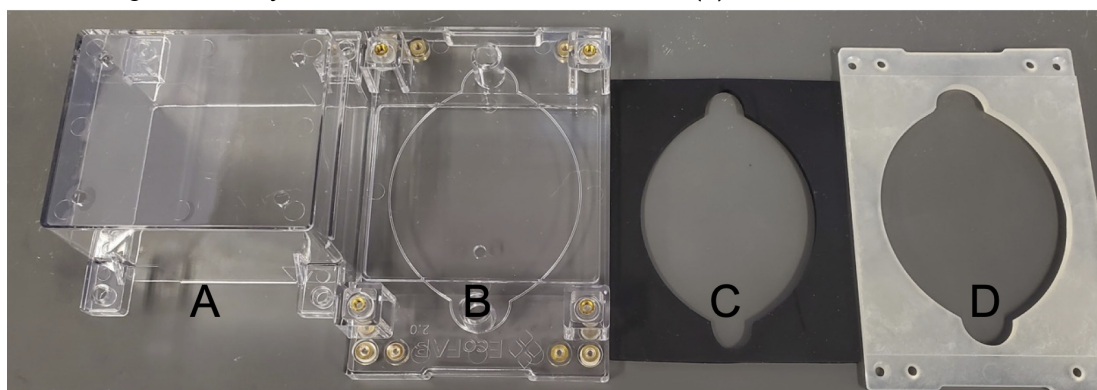


Fig. 1: The main EcoFAB 2.0 parts

Additional parts that may need to be purchased separately (quantity per device / supplier / supplier ID / quantity per package)

- E. 4" x 3" Glass slide (1 / Ted Pella / 260230-50 / 50)
- F. 3M Micropore Tape 1530-0 (a few pieces / FischerSci / 19-061655 / 24 rolls)
- G. Suba-Seal 8 mm septa (1 / Sigma Aldrich / #Z167231 / 100)
- H. 4/40 x 7/16" screws (4 / McMaster Carr / 91099A459 / 100)
- I. 4/40 x 1/4" screws (4 / McMaster Carr / 91099A155 / 100)
- J. 1/4" OD EPDM Rubber Washers (4 / McMaster Carr / 99186A111 / 100)

K. 3D Printed Seed Inserts (1 / LBNL - optional based on request)

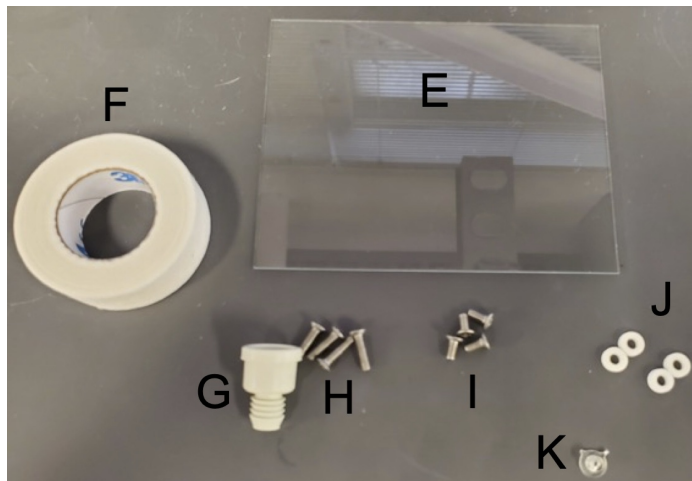


Fig. 2: Additional parts for EcoFAB 2.0 assembly

Consumables that may be required

L. Serological pipette

M. Syringes

N. 21 G needles

O. 0.22 μ m syringe filters (PES)

P. Sterilization pouches or aluminum foil to cover during sterilization.

Q. Manual or Willdone 008 electric screwdriver with Philips #1 head (Amazon, B08LPYPTV9)

R. Tweezers

S. Scissors

Note

We highly recommend utilizing an electric screwdriver (e.g., Willdone 008) to ensure uniform results and prevent the occurrence of cracks or leaks that may arise when employing a manual screwdriver.

Notes on materials

- 2
 1. The gaskets sometimes have an odor due to the manufacturing process. It is recommended to remove any plastic coatings from the gasket and wash it with alcohol (ethanol or isopropanol) and then purified water and then allow them to sit out in a well-ventilated area (e.g., biological safety cabinet or fume hood) overnight before use.
 2. The EcoFAB chamber, base, and backing plates can be reused; however, after repeated autoclaving, they may warp somewhat. We have not done extensive testing to determine how many times these parts can be reused. To clean, a laboratory-style dishwasher or soap should be suitable. Alcohol may streak the polycarbonate parts.
 3. The gasket will often fuse with the glass slide after experiments, and both parts will need to be

replaced.

4. One type of suba seal has been listed. There are other options available through Sigma Aldrich.
5. Seed inserts are not required. However, several have been designed for smaller seeds that are 3D printed out of biologically compatible resin that can be autoclaved.
6. Alternatively to glass slides, the design is also compatible with thinner materials like 0.55 mm thick gorilla glass. These can be obtained from specialty manufacturers.
7. Instead of covering the holes on top of the chamber with micropore tape. Breathe-Easy or Breathe-Easier membrane material can also be used (Diversified Biotech, #BMTM-1000 or BMTM-2000).
8. The EcoFAB Base will pick up fingerprints that are visible under the microscope, so wiping down the surface with a paper towel and wearing gloves is recommended.

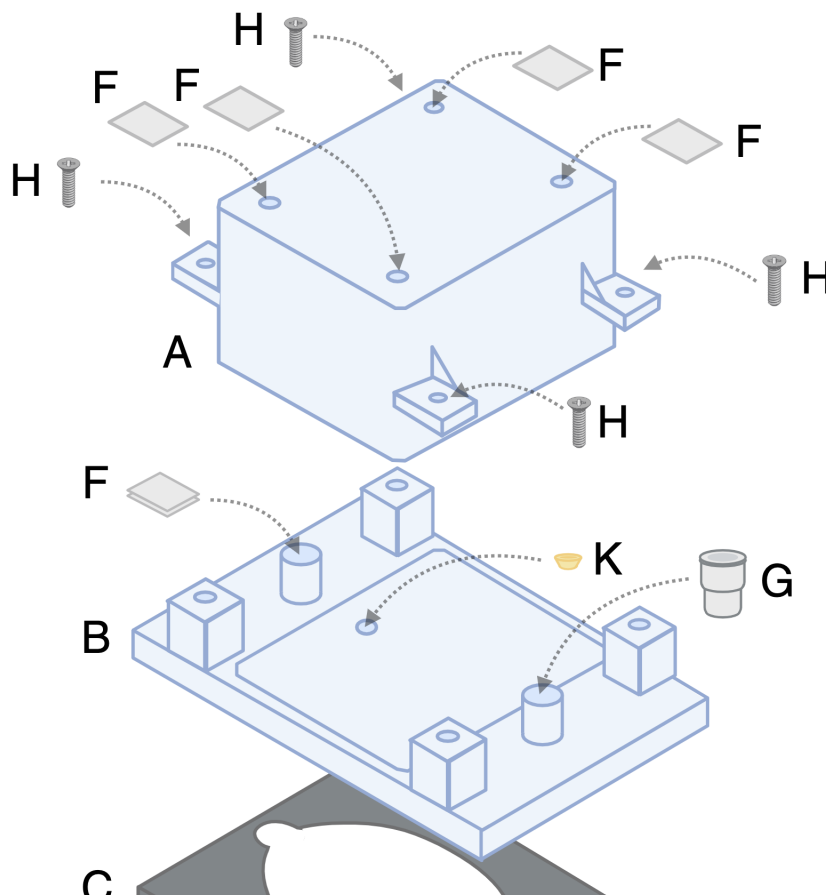
Sterilization

- 3
 1. Most of the parts for the EcoFAB 2.0 can be sterilized by autoclaving. For most uses it is recommended to partially assemble the EcoFABs, autoclave, and then finish assembly, add media and seedlings within a sterile environment (e.g., biological safety cabinet).
 2. The one part that can't be autoclaved is the micropore tape. Misting with 70% ethanol or UV may be used, but typically isn't necessary if handled carefully.

Assembly

4

EcoFAB 2.0 Typical Assembly



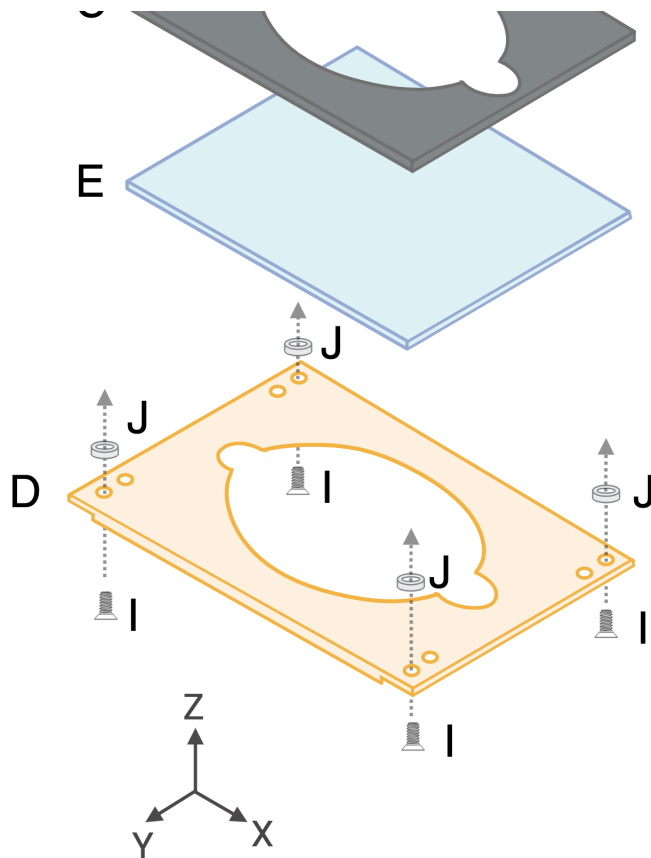


Fig. 3: Typical assembly of EcoFAB 2.0 consists of A. EcoFAB Chamber, B. EcoFAB Base, C. Gasket, D. Backing Plate, E. Glass slide, F. Micropore Tape, G. Suba-Seal septa, H. 7/16" screws, I. 1/4" screws, J. Rubber Washers, and K. Seed Insert. Figure created with Biorender.com

- 5 Place the EcoFAB base upside down on a clean surface and lay the gasket down between the threaded inserts. If the gasket has matte and glossy surfaces, place the matte side up and facing the glass so the slide can be adjusted after autoclaving if needed.



Fig. 4: Upside-down EcoFAB 2.0 base with silicone gasket

- 6 Place the glass slide down on top of the gasket. Make sure that the slide doesn't overlap with the threaded inserts and covers the cutout in the gasket.



Fig. 5: Glass slide placed on the silicone gasket

- 7 Place 4 washers on the four outer threaded inserts. Without the washers screwing in, the backing plate will need to be done very carefully, or the slide will crack, or the device will leak. Afterward, place the backing plate on top of everything with the protruding side facing up. Note: the backing plate is not reversible. Usage in the other direction is for instances where a thinner slide is used. In that case, the 4 internal threaded inserts will be used. You can tell this based on the indents in the backing plate screw taps.
- 8 Take the 4 shorter screws ($\frac{1}{4}$ " long) and lightly thread them into the four outer ports. If the devices will be sterilized, do not over-tighten the screws enough to distort the washers, or the slide is likely to crack during autoclaving. The screws should be just tight enough so that the backing plate doesn't fall off when it is turned over.

Note

If using the Willdone 008 electric screwdriver for the attachment of bottom plate screws, set the torque to 4.

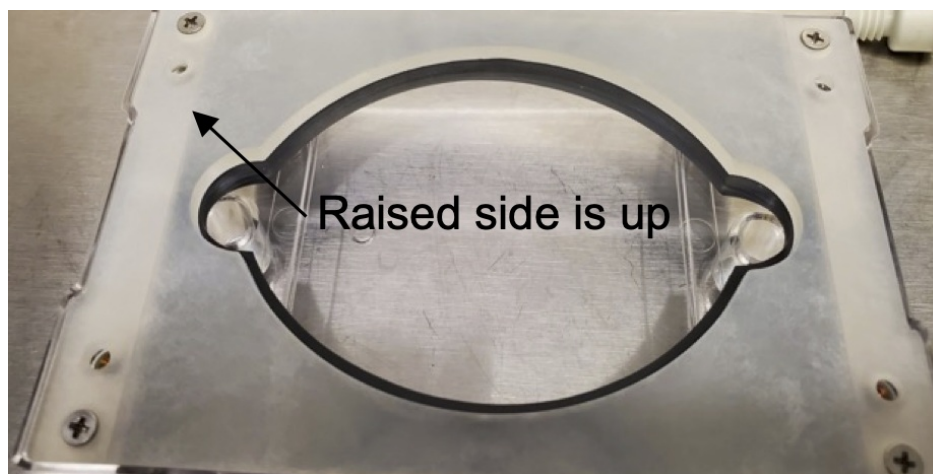


Fig. 6: Backing plate screwed to the base with the raised side facing away from the glass slide

- 9 Place the septa and seed insert (if being used) into the EcoFAB and place the chamber on top.

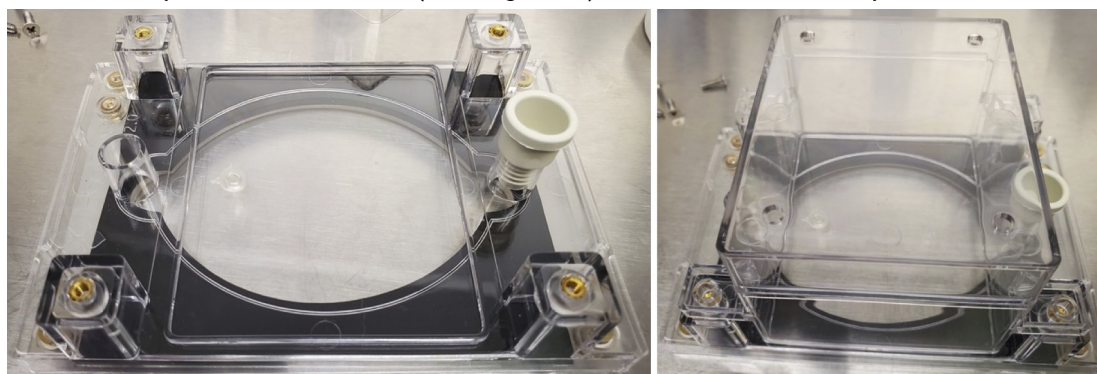


Fig. 7: Left: Assembled base. Right: top chamber placed on the base

- 10 Place the partially assembled EcoFAB in an autoclave bag or wrap it tightly in foil and autoclave it to sterilize under standard conditions.



Fig. 8: EcoFAB 2.0 in an autoclave bag

- 11** After sterilization, unpack the EcoFABs inside a biological safety cabinet or another sterile environment. Remove the chamber and turn the EcoFAB upside down to tighten the bottom screws. If any screws fall out during sterilization, make sure the washers are aligned with the screws and the slide and gasket with the ports. The screws should be tightened until the washers distort and begin to flatten.

Note

If using the Willdone 008 electric screwdriver to tighten the bottom plate screws, set the torque to 7.



Fig. 9: EcoFAB 2.0 with screws fully tightened

- 12** Place micropore tape pieces over the 4 top chamber vents (or other breathable covering like Breathe-Easy membranes). Place two pieces of micropore tape over the rear sampling port (near the seed insert) to allow air exchange during filling.



Fig. 10: Micropore tape covering the chamber vents and the sampling port

- 13** To fill the EcoFABs, prop the EcoFABs on an angle and fill from the bottom port either with a serological pipet with the septa removed or through the septa with a syringe and needle. Approximately 10 ml of liquid will fill the root chamber. Using sterile tweezers, place a seedling in the seed port/holder.
see

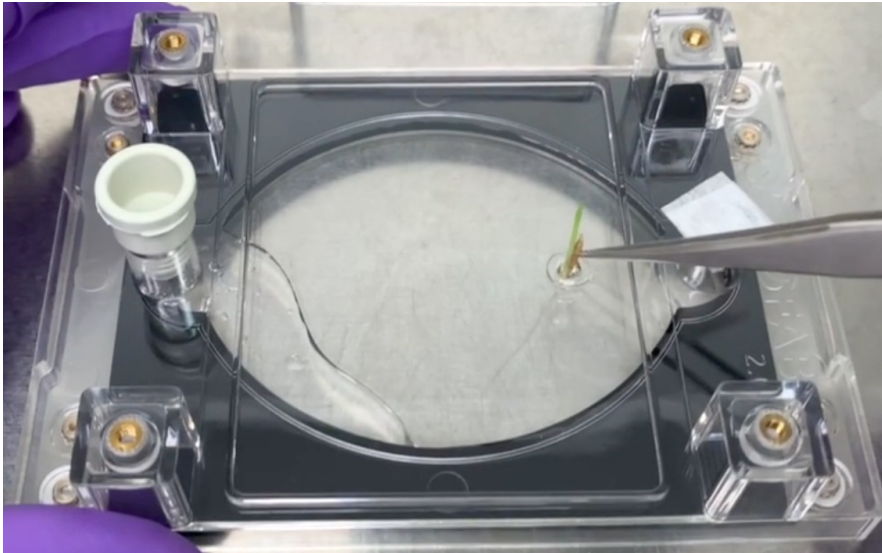


Fig. 11: Transfer of 3-day old *Brachypodium* seedling into EcoFAB filled with hydroponic medium

- 14** Screw down the top chamber and place the EcoFAB 2.0 in a growth chamber.

Note

If using the Willdone 008 electric screwdriver to attach the top chamber, set the torque to 4-5



Fig. 12: Assembled EcoFAB 2.0 with *B. distachyon* seedling