



FEB 27, 2024

## OMS Atlas OCT Spatial Mapping - Limited

Forked from [OMS Atlas OCT Spatial Mapping](#)

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OMS Atlas



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### ABSTRACT

This protocol describes the procedure by which the OMS Atlas serially sections an OCT block, prepares the resulting slides and samples, and then distributes the specimens for downstream analysis.

### MATERIALS

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Superfrost Plus Microscope Slides **Fischer Scientific Catalog #12-550-15**

Additional equipment:

- Cryostat

### BEFORE START INSTRUCTIONS

Transfer OCT blocks to OHSU Knight Histopathology Shared Resource (HSR) for sectioning and processing.

OPEN ACCESS



DOI:

[dx.doi.org/10.17504/protocols.io.8epv5xy24g1b/v1](https://dx.doi.org/10.17504/protocols.io.8epv5xy24g1b/v1)

**Protocol Citation:** Brett Johnson, Danielle Galipeau, George Thomas 2024. OMS Atlas OCT Spatial Mapping - Limited. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.8epv5xy24g1b/v1>

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**Protocol status:** Working  
We use this protocol and it's working

**Created:** Feb 27, 2024


## Preparation

- 1 Verify the identity of the OCT block to be cut against written request for sectioning.
- 2 Remove OCT block from -80 °C freezer and acclimate to cryostat ( -20 °C ) for minimum of 03:00:00 .
- 3 Label all slides and cryotubes with a unique BEMS ID and Part#, corresponding to the written request and OCT spatial map (below).


| A     | B                      | C         | D                                       | E                 |
|-------|------------------------|-----------|---|-------------------|
| Part# | Description            | Thickness | Assay                                   | Recipient         |
| 1     | Superfrost Plus slide  | 5µm       | H&E                                     | OHSU, HSR         |
| 2     | Superfrost Plus slide  | 5µm       | Cyclic Immunofluorescence (Tumor Panel) | HMS, Alyce Chen   |
| 3     | Superfrost Plus slide  | 5µm       | Cyclic Immunofluorescence (Tumor Panel) | HMS, Alyce Chen   |
| 4     | Remainder of OCT block | NA        | Single Cell Indexing ATAC Sequencing    | OHSU, Andrew Adey |

## Sectioning

- 4 Affix OCT block to cryostat chuck.

- 5 Orient and face block to get adequate amount of core.  
*Note: Avoid excessive facing to reduce tissue loss.*
- 6 Set cryostat to 5 micron sections.  
*Note: All sections cut from here on should be sequential. The serial order, adjacency, and consistent orientation of the sections are all important factors. Please note any deviations from the protocol.*
- 7 Cut three sections at 5 microns (Part#1-3) and affix onto appropriately labeled slide according to OCT spatial map (step #3 above).
- 8 Place all slides and remaining OCT block in  -80 °C freezer.  
*Note: No slides are to be fixed under this protocol.*

## Processing

- 9 Perform hematoxylin and eosin (H&E) staining on slide labeled Part#1 (see OCT spatial map in step #3 above).
- 10 Deliver unstained slides (Part#2 and 3) and remainder OCT block (Part#4) to BioLibrary for distribution.  
*Note: Keep samples frozen at all times. Store at  -80 °C . Transfer/ship on dry ice.*