



JAN 31, 2024

MBP-Clu-tail purification from Escherichia coli cells

Andreas Bracher¹, F Ulrich Hartl¹

¹Department of Cellular Biochemistry, Max Planck Institute of Biochemistry



Patricia Yuste-Checa

ABSTRACT

This protocol details how to efficiently purify the fusion protein Maltose binding protein (MBP)-Clu-tail (204-238) from Escherichia coli.

ATTACHMENTS

[MBP-Clu\(204-238\)
purification
protocol_protocols.io.docx](#)

MATERIALS

Buffers

▪ Binding buffer:

A	B
Tris/HCl pH 7.4	20 mM
NaCl	200 mM
EDTA	1 mM

- Elution buffer: Binding buffer + **[M] 10 millimolar (mM)** maltose (final concentration)

OPEN ACCESS



DOI:

dx.doi.org/10.17504/protocols.io.6qpvr35xzvmk/v1

Protocol Citation: Andreas Bracher, F Ulrich Hartl 2024. MBP-Clu-tail purification from Escherichia coli cells.

protocols.io

<https://dx.doi.org/10.17504/protocols.io.6qpvr35xzvmk/v1>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: Jan 25, 2024

Last Modified: Jan 31, 2024

PROTOCOL integer ID: 94482

Keywords: ASAPCRN

Funders Acknowledgement:

Aligning Science Across

Parkinson's



Grant ID: ASAP-000282



His₆-Ubiquitin-GFP-Clu-tail expression and cell lysis

1 Express MBP-Clu-tail in E. coli BI21 (DE3) codon+RIL cells cultured in  1 L LB Medium containing  2 undetermined glucose at  37 °C with  1 millimolar (mM) IPTG during  02:00:00 . 2h

2 Centrifuge culture and keep pellet.



3 Re-suspend pellet in  25 mL volume of ice-chilled binding buffer, add Complete protease inhibitor cocktail (Roche) and  1 millimolar (mM) phenylmethylsulfonyl fluoride (PMSF).

4 Lyse cells by ultrasonication in ice bath (10 cycles of  00:00:30 ultrasonication with  00:01:30 intermittent cooling). 2m

5 Clear lysate by centrifugation at  22000 rpm in a JA25.50 rotor at  4 °C .

Amylose affinity chromatography

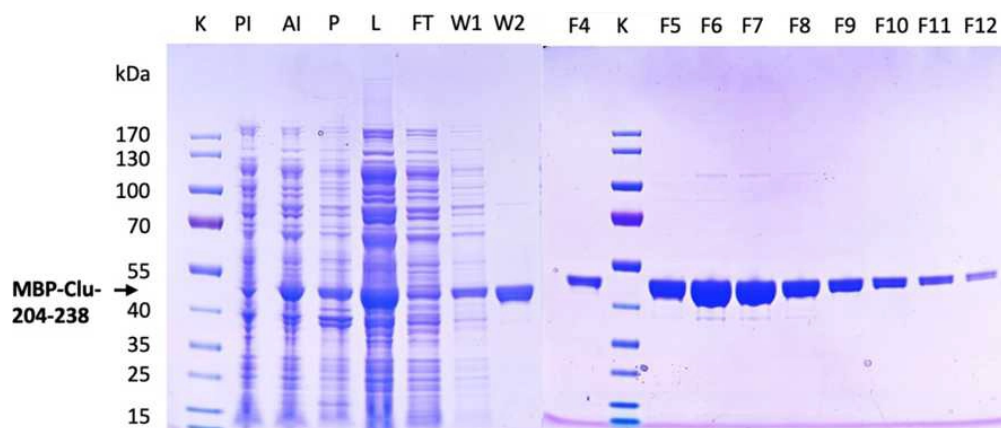
6 Load supernatant onto a 20 mL Amylose Resin (New England Biolabs) column previously equilibrated with binding buffer by gravity flow at 4 °C .

7 Wash the column with 12 CV of ice-chilled binding buffer.



8 Elute MBP-Clu-tail protein with 12x 3 mL of ice-chilled Elution buffer. Collect fractions of 3 mL volume. Store fractions On ice .

9 Analyze eluted fractions by SDS-PAGE and Coomassie blue staining.



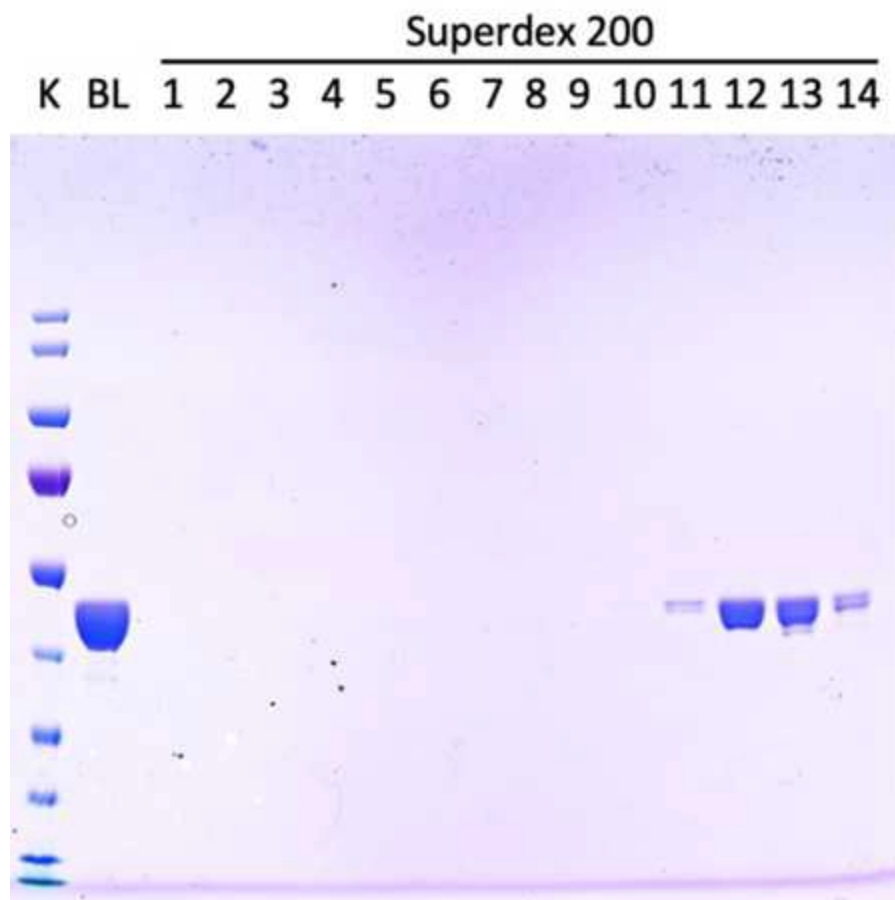
10 Pool fractions containing MBP-Clu-tail.

11 Concentrate pool to less than 10 mL volume by ultrafiltration using 10 kDa cut-off spin concentrator at 4 °C .

Size exclusion chromatography

12 Apply concentrate on a HiLoad 26/600 Superdex-200 (Cytiva 28-9893-36) column equilibrated with PBS. Develop the column at 4 °C and collect 10 mL fractions.

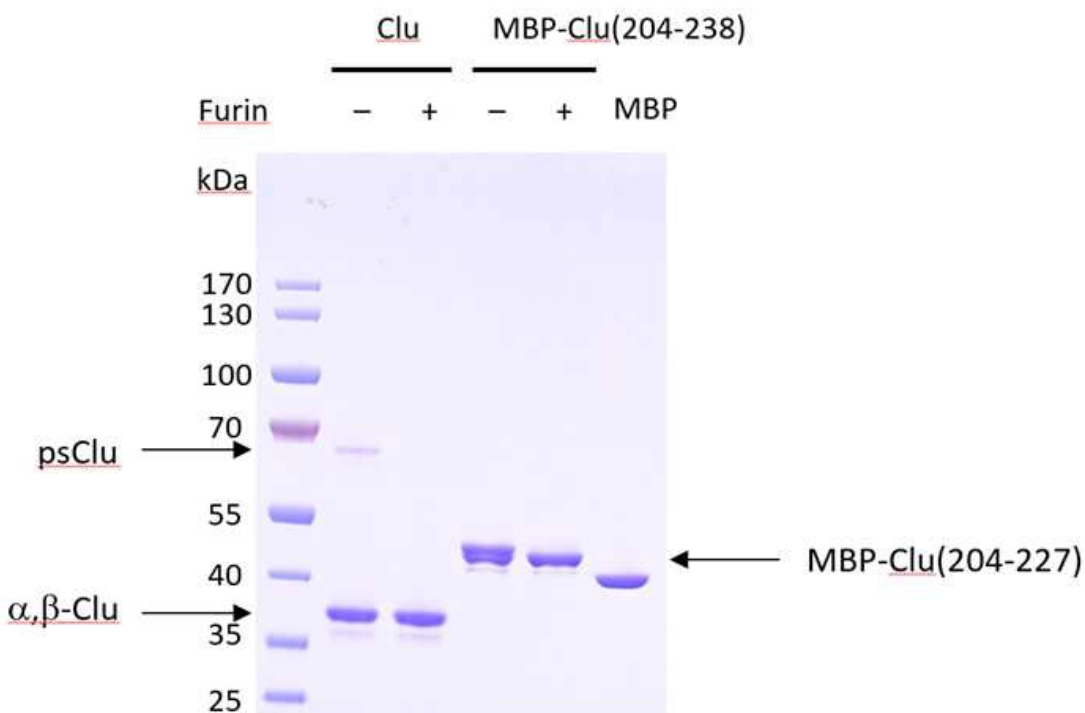
13 Analyze eluted fractions by SDS-PAGE and Coomassie blue staining.



14 Merge fractions with MBP-Clu-tail peak. Concentrate to 1.5 mL volume by ultrafiltration using 10 kDa cut-off spin concentrator at 4 °C, aliquot and flash-freeze purified MBP-Clu-tail in liquid nitrogen for storage at -70 °C.


Note

MBP-Clu-tail appears as a double band. In contrast to the lower band, the upper band is sensitive to cleavage by furin, suggesting that a protease from *E. coli* partially cleaves close to the furin site in MBP-Clu-tail.



Note

Concentrations were determined by absorbance at 280 nm using absorbance coefficients of $66,350 \text{ M}^{-1} \text{ cm}^{-1}$ or $1.645 \text{ L g}^{-1} \text{ cm}^{-1}$ for MBP-Clu-tail.

Approximate yield: From  1 L of culture around  15 mg of pure MBP-Clu-tail were obtained.