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15-minute DNA extraction from beetle legs for PCR barcoding

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1 Works for me dx.doi.org/10.17504/protocols.io.bmj4k4qw

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ABSTRACT

This method allows for quick extraction of DNA from beetle specimens for routine PCR amplification of barcoding genes such as COI (cytochrome oxidase subunit 1) for beetle ID. It is likely to also be effective for other high-copy genes such as other mitochondrial genes or ribosomal genes. It may not yield enough DNA to amplify large fragments or low-copy genes, and is certainly not appropriate for next-generation sequencing. However, for quick ID of beetle vouchers (for example, for beetles from which fungal cultures were isolated in a mycological study) it's hard to beat a ~15 minute DNA extraction.

This protocol is part of the Bark Beetle Mycobiome (BBM) Research Coordination Network. For more information on the BBM international network: Hulcr J, Barnes I, De Beer ZW, Duong TA, Gazis R, Johnson AJ, Jusino MA, Kasson MT, Li Y, Lynch S, Mayers C, Musvuugwa T, Roets F, Seltmann KC, Six D, Vanderpool D, & Villari C. 2020. Bark beetle mycobiome: collaboratively defined research priorities on a widespread insect-fungus symbiosis. Symbiosis 81: 101–113 https://doi.org/10.1007/s13199-020-00686-9.

THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Mayers CG, Harrington TC, Mcnew DL, Roeper RA, Biedermann PH, Masuya H, Bateman CC. Four mycangium types and four genera of ambrosia fungi suggest a complex history of fungus farming in the ambrosia beetle tribe Xyloterini. Mycologia. 2020 Jun 18:1-34.

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GUIDELINES

The manufacurer's protocol recommends **200 µl** of PrepMan per extraction, but we found good results with **50 µl** volumes, which quadruples the number of extractions possible from a single bottle of PrepMan and results in relatively inexpensive DNA extractions.

MATERIALS TEXT

MATERIALS

PrepMan™ Ultra Sample Preparation Reagent **Thermo**

Fisher Catalog #4318930

ABSTRACT

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1 Use a pipette to transfer **50 µl** of PrepMan Ultra Sample Preparation Reagent into a small microcentrifuge tube. Label the tube appropriately. 1m

If possible, this should be done in a sterile environment and away from PCR equiptment to avoid contaminating the PrepMan reagent with unwanted DNA.

2 Using sterile forceps, remove one leg (or multiple) from the beetle and transfer to the PrepMan reagant. 1m

Fresh, dried, and ethanol-pickled beetle specimens have all yielded usable results with varying success rates.

3 Using a sterile transfer needle (or other tool), thoroughly crush the leg(s) against the inner wall of the microcentrifuge tube. 2m

4 Incubate the microcentrifuge tube at **100 °C** for **00:10:00** . 10m

5 *Optional:* Centrifuge the microcentrifuge tube briefly and pipette the supernatant to a new microcentrifuge tube. Label appropriately. 2m

6 The liquid in the tube is now ready to be used as template for PCR. It can be stored at **-20 °C** .

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Typically we used

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