



VERSION 1
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OPEN  ACCESS



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Protocol status: Working
We use this protocol and it's working

Created: Nov 29, 2023

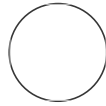
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PROTOCOL integer ID:
91568

Tissue processing and freezing after surgery V.1

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ABSTRACT

The aim of this protocol is to document the processing of fresh tissue after surgery.

The collected tissue must also be documented in the files which are linked down below

The protocol describes in detail how to process the resulting tissue from following protocols:

Protocol



NAME

Gewebesammlung Frischgewebe Zystektomie

CREATED BY

Bettina Ergün

PREVIEW

Protocol



NAME

Gewebesammlung Frischgewebe Prostatektomie

CREATED BY

Bettina Ergün

PREVIEW

MATERIALS

Transport medium

MEM : Prozent: 98 %

Zellshield : Prozent: 2 %

MEM mit Earle's Salzen, mit 2,2 g/l NaHCO₃, mit stabilem Glutamin

Hersteller/Lieferant:Biosell

Katalog #:BS.FG0325

ZellShield

Hersteller/Lieferant:Minerva BioLabs

Katalog #:13-0050

Freezing medium

FKS : Prozent: 90 %

DMSO : Prozent: 10 %

FBS Superior stabil

Hersteller/Lieferant:Biosell

Katalog #:FBS. S 0615

sterile forceps, scapels and petri dishes



1 ml Cryotubes

Sample processing in the lab

1

1h

Fresh tissue should be processed on the same day if possible, but can be stored at 4°C for a maximum of overnight.

To reduce pathogens, the tissue should be incubated for  01:00:00 at  Room temperature

2

Gewebe#	Organ

3

Note

Comments/remarks on the tissue/processing:

4

The tissue is divided in the lab into pieces with an edge length of 1-2 mm.

Mix the fragments of a tumor piece so that each aliquot is as representative as possible of the tumor area.

5

16 (decision from June 29, 2023; previously: 8) fragments are stored in 1 ml of freezing medium in a cryotube.

If there are enough fragments for more than 4 cryotubes, more fragments can be combined in one tube. A maximum of 5 cryotubes of each entity are frozen.

Cryo tubes should be labelled as follows and documented in the N2 binder:

Tissue ID, T1/T2 resp. N1/N2...

No. of Pieces

Date

6

Freeze samples in Mr. Frosty overnight.

Documentation

7

Store patient data and enter the corresponding liquid number:

https://charitede.sharepoint.com/:x:/r/sites/UroForsch/_layouts/15/doc2.aspx?sourcedoc=%7B1282209D-5A7C-4998-A577-145386116E54%7D&file=Biobank_Gewebe_ab01012019.xlsx&action=default&mobileredirect=true

8

Enter the corresponding tissue number in the liquid collection list:

https://charitede.sharepoint.com/:x:/r/sites/UroForsch/_layouts/15/Doc.aspx?sourcedoc=%7BCE8CD395-BFB0-4B2C-8220-37FB802726A7%7D&file=Biobanking-lfdListe_ab190311.xlsx&action=default&mobileredirect=true

9

Please add pictures of:

- *the origin of the sample in the organ (if available)*
- *each tissue piece that has been taken before cutting the fragments. The picture should include the tissue ID (in the picture)*

- 
- *the documentation sheet*