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© Expression and purification protocols of *Homo* sapiens E3-like ligase ATG12-ATG5/ATG16L1ß

Dorotea Fracchiolla¹

Team Hurley

¹Sascha Martens, University of Vienna, Max Perutz Labs - Vienna (Austria)



ABSTRACT

This protocol outlines the procedures for expression and purification of the human E3-like ligase ATG12–ATG5/ATG16L1ß (with or without GFP tag) of the ATG8 ubiquitin-like conjugation system in autophagy.

THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

A PI3K-WIPI2 positive feedback loop allosterically activates LC3 lipidation in autophagy 10.1083/jcb.201912098

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KEYWORDS

expression, purification, human E3-like ligase ATG12-ATG5/ATG16L1ß, recombinant protein

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IMAGE ATTRIBUTION

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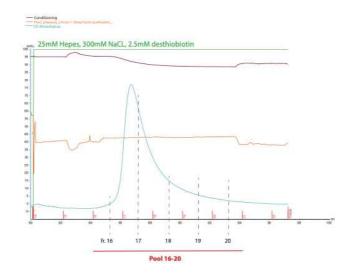
Feb 25, 2021 Emily Hasser University of Washington

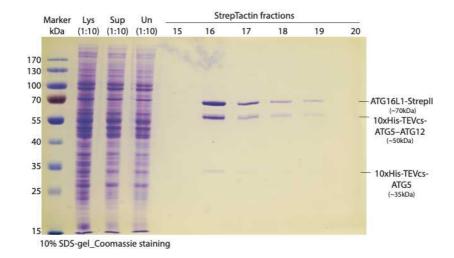
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GUIDELINES

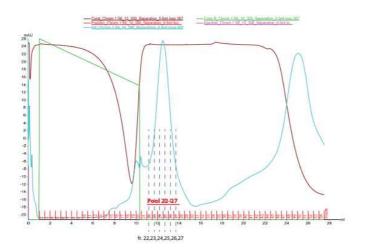
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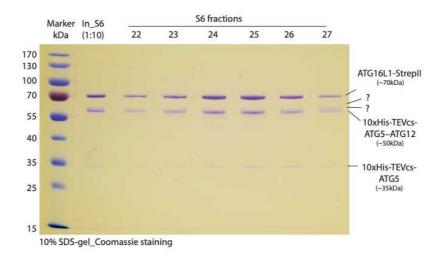




Chromatograph and Coomassie BB stained gel of Strep-tag affinity purification for ATG12-ATG5/ATG16L1ß.

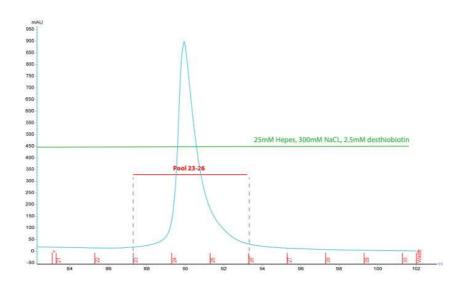
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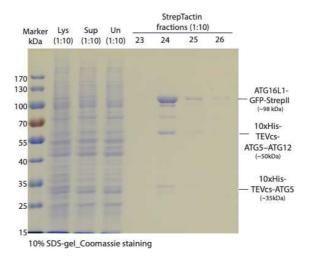




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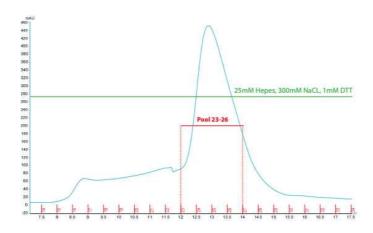
SMC1100_10xHis-TEVcs-ATG5-ATG12/ATG16L1-GFP-TEVcs_StrepII StrepTactin Affinity purification_17.12.18

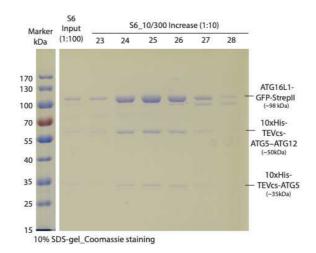




Chromatograph and Coomassie BB stained gel of Strep-tag affinity purification for ATG12-ATG5/ATG16L1ß-GFP.

Size Exclusion Chromatograhy_S6_10/300 Increase SMC1100_(10xHis-TEVcs-ATG5-ATG12/ATG16L1-GFP-TEVcs-StrepII)_17.12.'18





Chromatograph and Coomassie BB stained gel of Size Exclusion purification for ATG12-ATG5/ATG16L1ß-GFP.

Materials and Reagents

- Sf9 insect cells
- SF921 medium with antibiotics 100 IU/ml Penicillin and 100 μg/ml Streptomycin
- sterile cell culture hood
- 27°C shaker incubator
- sterile flasks and pipettes
- douncer 40 mL
- Virus coding for hATG7/hATG10/hATG12/10xHis-TEVcs-hATG5/10xHis-TEVcs-hATG16L1-TEVcs-StrepII (SMC1099, Addgene 169076) or hATG7/hATG10/hATG12/10xHis-TEVcs-hATG5/10xHis-TEVcs-hATG16L1-mGFP-TEVcsStrepII (SMC1100, Addgene 169077). Note: All the CDSs are codon-optimized for insect cell expression system (purchased from GenScript). Single subunits and tags were assembled into pLIB or pBIG1 vectors via classical restriction cloning or Gibson assembly strategy by Dorotea Fracchiolla. The final ATG12, 10xHis-TEVcs-ATG5, 10xHis-TEVcs-ATG16L1(±GFP)-TEVcs- StrepII, ATG7, ATG10 poli-cystronic gene constructs were cloned via Golden Gate approach by the Vienna BioCenter Core Facilities (VBCF) Protech Facility.

Buffers for Protein Purification

- Lysis buffer: 50mM Hepes pH=7.5, 300mM NaCl, Benzonase Nuclease (Sigma, use 1μl/50ml lysis buffer), 1mM
 Dithiothreitol (DTT), 1x Protease Inhibitor (EDTA-free CIP tablet, Roche), 300μl/1lt CIP (Sigma)
- Binding/Washing buffer: 50mM Hepes pH=7.5, 300mM NaCl, 1mM Dithiothreitol (filtered and degassed)
- Elution buffer: 50mM Hepes pH=7.5, 300mM NaCl, 2.5mM des-thiobiotin (Sigma), 1mM Dithiothreitol (DTT)(filtered and degassed)
- Size Exclusion Chromatography Buffer: 25mM Hepes pH=7.5, 300mM NaCl, 1mM Dithiothreitol (DTT) (filtered and degassed)

Note: all purification buffers are filtered and degassed. Reducing agent (DTT, Dithiothreitol) is added after degassing step.

Columns:

- StrepTactin 5ml column (GE Healthcare)
- S6 10/300 Increase (GE Healthcare)

Gels: 10% Poly-acrylamide SDS-gels

SAFETY WARNINGS

For hazard information and safety warnings, please refer to the SDS (Safety Data Sheet).

Infection/expression/harvest

- 1 Infect □1 L culture of Sf9 cells growing in Sf921 medium containing antibiotics Penicillin/Streptomycin at 1-1.5 mil/ml cells/volume at 99-100% viability in log phase with a volume of Virus 1 (V1), according to viral titer.
- 2 Monitor infection and harvest cells when viability goes to 97-98%.

Always check reporter gene fluorescence under the microscope to monitor the viral infection: when all alive cells are brightly fluorescent and only few dead -> harvest!

3

To harvest spin down the culture at @4000 rpm, 4°C, 00:15:00 in a Sorvall RC6+ centrifuge (Thermo Scientific).

4 Pour off the supernatant without disturbing the cell pellet.

5

Gently wash the pellet 1x in cold PBS buffer.

6



Centrifuge again and remove PBS.

7 Flash freeze cell pellet in liquid nitrogen and store at 8-80 °C for long storage.

Protein purification

45m

- 8 All steps are to be executed at 4°C or on ice.
 Re-suspend the cell pellet corresponding to 1 lt culture in □50 mL ice cold Lysis buffer; gently stir at 8 4°C avoiding bubbling until pellet dissolves.
- 9 Mechanically lyse the cells passing them through a pre-cooled 40ml vol. douncer for 3x (10x pestle A followed by 10x pestle B).

10

45m

Clear the lysate by spinning at 325000 rpm in a Ti45 Rotor for 00:45:00 at 4 °C using Beckman centrifuge.

- 11 Inject the supernatant at 8 4 °C onto a 5 ml StrepTactin column pre-equilibrated in **Binding buffer** at 1ml/min flow rate to allow protein binding.
- 12

Wash the column for 5CV (Column Volume) with **Wash buffer** at 2ml/min flow rate to remove unspecifically bound proteins.

- 13 Perform elution at 1ml/min flow rate with Elution buffer and collect corresponding fractions.
- 14 Check fractions on a SDS-PAGE gel (usually elution is very clean), pool and concentrate down those containing the proteins of interest at 8 4 °C using a 30kDa cutoff Amicon Filter (considering ATG12–ATG5 conjugate as lower MW limit) to 500 µl volume in a 5810R centrifuge (Eppendorf).
- Keep centrifugation steps short (© 00:05:00) to avoid protein local concentration/aggregation on the filter. Be very careful when pipetting at this step.

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- Inject 500 μl protein onto a S6_10/300 Increase column pre-equilibrated in Size Exclusion
 Chromatography buffer containing [M]25 Milimolar (mM) Hepes PH7.5, [M]300 Milimolar (mM) NaCl and [M]1 Milimolar (mM) DTT.
- 17 Check fractions on a SDS-PAGE gel, pool and concentrate down those containing the proteins of interest at 8 4 °C in a 30kDa cut-off Amicon Filter.
- 18 Keep centrifugation steps short to avoid protein precipitation.
- 19 👸

Measure final concentration with a spectrophotometer at A₂₈₀.

E3, Epsilon= $148280 \text{ M}^{-1} \text{ cm}^{-1}$; MW= 120,58 kDaE3-GFP, Epsilon= $170170 \text{ M}^{-1} \text{ cm}^{-1}$; MW= 148,05 kDa

A good concentration value when the protein does not aggregate is $20\mu M$.

Aliquot the protein, snap freeze it in liquid Nitrogen and store it at 8-80 °C. Protein yield is 1-3 mg/liter of culture. Protein activity is kept for at least 18 months.