



VERSION 2

JUL 03, 2023

OPEN ACCESS

DOI:

[dx.doi.org/10.17504/protocols.io.kxygx3mrwg8j/v2](https://dx.doi.org/10.17504/protocols.io.kxygx3mrwg8j/v2)

**Protocol Citation:** Annika Fendler 2023. DNA/RNA extraction from fresh-frozen tissue, AllPrep DNA/RNA/miRNA Universal Kit. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.kxygx3mrwg8j/v2> Version created by Annika Fendler

**License:** This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

**Protocol status:** Working Protocol has been established in house with minor changes from original protocol

**Created:** Jul 03, 2023

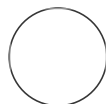
**Last Modified:** Jul 03, 2023

**PROTOCOL integer ID:** 84402

# DNA/RNA extraction from fresh-frozen tissue, AllPrep DNA/RNA/miRNA Universal Kit V.2

Annika Fendler<sup>1</sup>

<sup>1</sup>Charite



Annika Fendler

## ABSTRACT

Protocol for combined RNA and DNA extraction from fresh-frozen tissue using the AllPrep DNA/RNA/miRNA Universal Kit.

## MATERIALS

⊗ AllPrep DNA/RNA/miRNA Universal Kit (50) Qiagen Catalog #80224

⊗ Genomic DNA ScreenTape Agilent Technologies Catalog #5067-5365

⊗ Qubit™ dsDNA BR Assay Kit Thermo Fisher Scientific Catalog #Q32853

⊗ Qubit RNA BR Assay Kit Thermo Fisher Scientific Catalog #Q10211

⊗ RNA ScreenTape and Reagents Agilent Technologies

⊗ EB buffer Qiagen Catalog #19086

β-ME

EtOH

Isoprop

1.5 and 2 ml LoBind tubes

TissueLyser Beads 5 mm

## SAFETY WARNINGS



Dnase I stocks can be used 4 weeks after being thawed but should not be frozen again

**Keywords:** DNA, RNA, Fresh-frozen tissue, Qiagen AllPrep

## BEFORE START INSTRUCTIONS

### Preparations:

**FRN buffer:** Add 42 ml Isoprop to new bottle

**RPE buffer:** Add 44 ml EtOH to new bottle

**AW1 buffer:** Add 25 ml EtOH to new bottle

**AW2 buffer:** Add 30 ml EtOH


**DNase I stocks:** 550 µl RNase-free water to lyophilised DNase I, aliquot and store at -20°C for 9 months)

### Immediately

**DNase I:** 70 µl RDD + 10 µl DNase I per sample

**Proteinase K:** 60 µl AW1 + 20 µl Proteinase K per sample

## Tissue preparation

1 This protocol is for  Sample


Optional: Weigh tissue

Enter a complete list of samples used for each experiment below:

SampleID	Optionel Weigth (mg)	Comment


List of tissue sample used in experiment

2 Transfer tissue in  350 µL for up to 10 mg of tissue

 600 µL for 10 to 30 mg or stabilised tissue

RLT + β-ME in 2 ml DNA LoBind tube.

Make sure that the tissue does not defrost.

3 Add a 5mm bead to each tube and lyse tissue in TissueLyser for  00:02:00 @ 20 Hz

2m

## Equipment

**TissueLyser II**

NAME

Bead Mill

TYPE

QIAGEN








BRAND


85300

SKU

<https://www.qiagen.com/us/products/human-id-and-forensics/automation/tissuelyser-ii/#orderinginformation>



LINK



- 4 Spin down for  00:01:00 @  9000 x g 1m
- 5 Turn tube rack and lyse tissue for  00:02:00 @ 20 Hz 2m
- 6 Spin down for  00:01:00 @  9000 x g 1m
- 7 Add lysed product to DNA Mini Spin Column
- 8 Spin  00:00:30 @  max x g, repeat if any liquid remains on column 30s


9 Transfer column to a new collection tube and store tube at  4 °C until DNA extraction



10 Transfer flow-through to new 2 ml LoBind tube




## RNA extraction

11 Add  50 µL for up to 10 mg tissue  80 µL for 10 to 30 mg or stabilised tissue Proteinase K, mix by pipetting













12 Add  200 µL for up to 10 mg tissue  350 µL for 10 to 30 mg or stabilised tissue EtOH abs., mix by inverting, spin down liquid















13 Incubate  00:10:00 @  Room temperature 10m

14 Add  400 µL for up to 10 mg tissue  750 µL for 10 to 30 mg or stabilised tissue EtOH abs., mix by pipetting

15 Add  700 µL to RNeasy spin column and spin for  00:00:30 @  max x g and discard 30s  
flow-through

16 Repeat until all liquid passed through the column

- 17 Add  500 µL RPE
- 18 Spin  00:00:30 @  max x g and discard flow-through 30s
- 19 Add  80 µL DNase I working solution directly onto the membrane and incubate  00:15:00 at  Room temperature 15m
- 20 Add  500 µL FRN
- 21 Spin  00:00:30 @  max x g and *don't* discard flow-through 30s
- 22 Add flow-through again to column
- 23 Spin  00:00:30 @  max x g and discard flow-through 30s
- 24 Add  500 µL RPE

- 25 Spin  00:00:30 @  max x g and discard flow-through 30s
- 26 Add  500  $\mu$ L EtOH abs
- 27 Spin  00:02:00 @  max x g and place column in new collection tube 2m
- 28 Spin  00:02:00 @  max x g and place column in new 1.5 ml collection tube 2m
- 29 Add  30  $\mu$ L of RNase-free H<sub>2</sub>O
- 30 Incubate  00:01:00 @  Room temperature 1m
- 31 Spin down  00:01:00 @  8000 x g 1m
- 32 QC: Qubit  Qubit RNA BR Assay Kit Thermo Fisher Scientific Catalog #Q10211 and tape  
station  RNA ScreenTape and Reagents Agilent Technologies

32.1

Expected result

SampleID	Eluted Volume (µl)	Concentration (ng/ul)	Total mass (ng)	Storage Location

Qubit Results RNA

Expected result

Upload PDF from Tapestation here

DNA extraction11m

- 33

350 µL

 AW1 auf DNA Mini Spin Columnn geben
- 34

Spin

00:00:30

 @ 

max x g

 and discard flow-through

30s
- 35

80 µL

 Proteinase K working solution directly onto membrane
- 36













Incubate

00:05:00

 @ 

Room temperature


5m

- 37 Add  350  $\mu$ L AW1
- 38 Spin  00:00:30 @  max x g and discard flow-through 30s
- 39 Add  350  $\mu$ L AW2
- 40 Spin  00:02:00 @  max x g and place column in new 2 ml collection tube 2m
- 41 Spin  00:01:00 @  max x g and place column in new 1.5 ml collection tube 1m
- 42 Add  50  $\mu$ L  EB buffer Qiagen Catalog #19086 directly onto membrane
- 43 Incubate  00:01:00 @  Room temperature 1m
- 44 Spin down  00:01:00 @  8000 x g 1m




45

QC: Qubit DNA

 Qubit™ dsDNA BR Assay Kit Thermo Fisher Scientific Catalog #Q32853

and tape station

 Genomic DNA ScreenTape Agilent Technologies Catalog #5067-5365

45.1

Expected result

	Sample ID	Eluted Volume (µl)	Concentration (ng/µl)	Total Mass (ng)	Storage Location

Results from Qubit DNA

Note

Upload Tapestation results as pdf here