



Jan 07, 2022

# Detection of *Cryptosporidium* in stool sample by PCR-RFLP

botchiesenyo<sup>1</sup><sup>1</sup>University of Ghana

2

[dx.doi.org/10.17504/protocols.io.bxaxpifn](https://dx.doi.org/10.17504/protocols.io.bxaxpifn)

Ayi and Pawlowic Lab Collaborations

mcpawlowic

Nested PCR-RFLP adapted from Nichols, R.A.B., Campbell, B.M. and Smith, H.V., 2003. Identification of *Cryptosporidium* spp. oocysts in United Kingdom noncarbonated natural mineral waters and drinking waters by using a modified nested PCR-restriction fragment length polymorphism assay. *Applied and environmental microbiology*, 69(7), p.4183.

DOI

[dx.doi.org/10.17504/protocols.io.bxaxpifn](https://dx.doi.org/10.17504/protocols.io.bxaxpifn)botchiesenyo 2022. Detection of *Cryptosporidium* in stool sample by PCR-RFLP.

protocols.io

<https://dx.doi.org/10.17504/protocols.io.bxaxpifn>

protocol ,

Aug 11, 2021

Jan 07, 2022

52279

- GoTaq MasterMix from Promega
- Primers

Nested PCR

12h 45m

1 Extract whole genomic DNA from stool samples.

3h

Positive control DNA for *Cryptosporidium parvum* may be obtained from ATCC. Alternatively you can extract genomic DNA from wild type *Cryptosporidium parvum* purchased from Bunchgrass Farms (Deary, Idaho, USA), Waterborne Inc (New Orleans, Louisiana, USA), or Sterling Labs (Tucson, Arizona, USA).

## 2 First round of PCR to amplify 18SrRNA

4h

- 2.1 Combine the following in a PCR tube: 30m
- 5 µl of DNA (from positive control, experimental sample, or water for negative control)
  - 0.1 µM Forward Primer ("out NDIAGF2": CAATTGGAGGGCAAGTCTGGTGCCAGC)
  - 0.1 µM Reverse Primer ("out NDIAGR2": CCTTCCTATGTCTGGACCTGGTGAGT)
  - 1x GoTaq Master Mix (Promega)
  - Ultrapure water to bring total volume to 25 microliters

- 2.2 Thermocycler program: 3h 30m
- 95 °C -5 minutes,
  - 30 cycles:
    - 94 °C -30 seconds
    - 58 °C - 1 minute
    - 72 °C - 30 seconds

## 3 Second round of PCR to amplify 18SrRNA

4h 30m

- 3.1 Combine the following in a PCR tube: 30m
- 1 µl of PCR product from Step 2
  - 0.1 µM Forward Primer ("DIAGF": AAGCTCGTAGTTGGATTTCTG)
  - 0.1 µM Reverse Primer ("DIAGR": TAAGGTGCTGAAGGAGTAAGG)
  - 1x GoTaq Master Mix (Promega)
  - Ultrapure water to bring total volume to 25 microliters

- 3.2 Thermocycler program: 4h
- 95 °C -5 minutes,
  - 45 cycles:
    - 94 °C -30 seconds
    - 58 °C - 1 minute
    - 50 °C - 1 minute

- 4 Run 7 µl of PCR product from Step 3 on a 2% agarose gel and visualise by staining with ethidium bromide. 45m

RFLP 4h 45m

- 5 Combine the following and incubate at 37 °C for 4 hours: 4h
- 5 µl PCR product from step 4 above
  - 20u Restriction enzyme (Ssp1, Vsp1, Ase1)
  - 1x final concentration Restriction enzyme digestion buffer (corresponds to enzyme used)
  - 1x final concentration BSA
- 6 Run digested PCR product from Step 5 on a 2% agarose gel and visualize by staining with ethidium bromide. 45m