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Crystallization of Zika NS5 RdRp

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ASAP Discovery



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External link: https://asapdiscovery.org/outputs/target-enabling-packages/#ASAP-ZIKA-NS5-RDRP

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Protocol status: Working
We use this protocol and it's

working

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Disclaimer

The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

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Abstract

The main aim of this work was to identify small molecules that bind Zika NS5 RdRp (catalytic RNA-dependent RNA polymerase domain) through X-ray fragment-based screening. The Zika NS5 RDRP domain was cloned, expressed, purified, and crystallised. Suitable crystals for fragment screening were produced and optimised allowing an extensive fragment campaign to be performed. A native high-resolution structure was determined at 1.8Å and formed the basis for the fragment campaign.

Materials

SwissCl 3 lens crystallization plates https://swissci.com/product/3-lens-crystallisation-plate/ Codes: Midi: UVXPO-3LENS 3W96T-PS 3W96T-UVP

Morpheus HT-96 single reagent 250mL Catalog # MDSR-47-250-2-10

Purified Zika NS5 polymerase protein ([M] 5 mg/mL) in [M] 20 millimolar (mM) HEPES https://doi.org/10.1016/j.j. [M] 300 millimolar (mM) NaCl, 2.5% Glycerol, [M] 10 micromolar (µM) ZnCl₂, [M] 2 millimolar (mM) TCEP

306-903 residues- construct 2A1 (6 Hist sumo tag)



Safety warnings

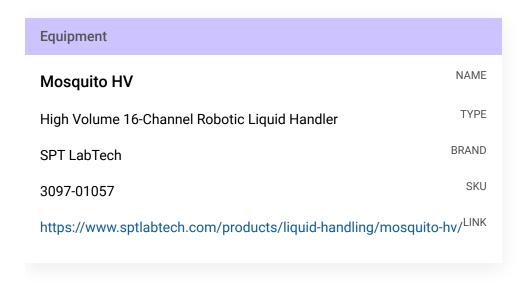


• Follow all handling warning for the chemicals used in the crystalllisation screen composition.



Equipment needed

<u>Formulatrix Rock Imager</u> (or incubator of choice) **SPT mosquito**



P100 8 multi-channel pipette

SwissCI 3 lens plate

Crystallization experiment

1d

2 Protein and buffer requirements:

> ∆ 3.264 mL Crystallization screen

3 **Crystallisation screen composition:**

Morpheous I E10 condition

[M] 0.12 Molarity (M) Ethylene glycols [M] 0.1 Molarity (M) Buffer system 3 PH 8.5 30 % v/v Precipitant Mix 2

Stock solutions used:

Morpheous I E10 condition



Note

The crystallisation screen can be stored in a duran bottle.

For long term storage keep the Crystallisation screen in the fridge at 4°C.

4 Dispense 🚨 34 µL Crystallisation screen into SwissCl 3 lens plate reservoir wells using a 100 μl multi-channel pipette.

Sample to each lens using the SPT mosquito.

Dispense 🚨 50 undetermined Crystallisation screen to each lens using the SPT mosquito.

Drop ratio: 2:1 ratio (100 nl Sample : 50 nl reservoir solution)

Final drop volume: 150 nl

5 Incubate at \$\mathbb{L}\$ 20 °C for \(\frac{1}{20} \) 24:00:00 h in Formulatrix Rock Imager.

Imaging Schedule: The first images are taken after 12 h and the imaging schedule follows a Fibonacci sequence of days for further collections.

6 Crystal form after ~24 h. 1d



Expected result

The crystals reach their maximum size after 96 h and the precipitant has gone.

Crystals grown inconsistently to 2 sizes, one being half the size described below, but both sizes achieve the same results

Morphology: typically plates.

Size: $\sim 250 \, \mu m$ in length and $\sim 60 \, \mu m$ in width, depth of the crystals is $\sim 2 \, \mu m$

Appearance: glass shard. Average resolution: 2.0 Å Space group: P4₃2₁2 **Unit cell:** 79 Å, 79 Å, 210 Å

90.00°, 90.00°, 90.00°



An example of a drop containing Zika NS5 RdRp ploymerase crystals.



Data collection at Synchrotron

7 Diamond Light Source

> **Unattended Data Collection (UDC) Data Collection Temperature:** 100K **Detector:** DECTRIS EIGER2 X 9M

Beamline: 104-1

Wavelength: 0.9212 Å **Resolution (Å):** 1.78 **Beam Size (µm):** 60 X 50 Number of images: 3600

Oscillation: 0.10° **Exposure (s):** 0.0020 Transmission (%): 100 Flux (ph/s): 9.50e+11

Protocol references

N/A