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# Modified Frame-projected Independent Fiber Photometry (FIP) System\_Hardware

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## ABSTRACT

This is a step-by-step protocol to build a modified FIP (Frame-projected Independent FP) system. FIP was first implemented and reported by Kim et al. (2016). Their protocol is available [here](#).

We modified previous design so that it can be built mostly with Thorlabs products (still some products from other manufactures are required), which are off-the-shelf and easy to be purchased within a reasonable lead time. The system described in this protocol is designed to record signals from 1)GFP-based sensors, 2)mApple-based sensors, and 3) iso sbestic signals from up to 4 sites. When applied to measurements of other fluorescent protein-based sensors or spectrally shifted sensors, selection of optical filters and/or excitation light sources should be modified accordingly. Also, the selection of fiber patch cable should be based on the number of simultaneously recorded locations, and diameter/NA of fiber implants.

We thank Takeo Katsuki Ph.D., (Thorlabs Japan) and Sho Yagishita M.D., Ph.D., (U.Tokyo, Japan) for sharing their galvano-mirror-based photometry design described in Iino et al. (2020). TK also kindly provided CAD model used in this protocol.

## MATERIALS

### Thorlabs

product#	Description	#
Excitation		
M415F3	LED415nm	1
M470F3	LED470nm	1
M565F3	LED565nm	1
LEDD1B	t-cube LED driver(1200mA)	3
KPS201	PowerUnit for T-cube	3
FB410-10	410 bandpass	1

## OPEN ACCESS

**Protocol Citation:** Kenta M. Hagihara 2023. Modified Frame-projected Independent Fiber Photometry (FIP) System\_Hardware.

**protocols.io**

<https://protocols.io/view/modified-frame-projected-independent-fiber-photometry-cn96vh9e>

## MANUSCRIPT CITATION:

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**Protocol status:** Working  
We use this protocol and it's working

**Created:** Feb 11, 2023

**Last Modified:** Jul 10, 2023

**PROTOCOL integer ID:**  
76830

**Keywords:** fiber photometry, calcium imaging, neuromodulator imaging, CMOS-based photometry

product#	Description	#
FB470-10	470 bandpass	1
FB560-10	560 bandpass	1
M59L01	SMA-SMA 1000um 0.48NA fiber	3
KAD11F	collimator mount (kinematic)	2
AD15F	collimator mount	1
F220SM-532	collimator	2
F950SMA-A	HighNA collimator	1
General cage system (30mm cage)		
DFM1	kinematic filter cube	4
RS1P	post	4
CP33	cage plate	5
ER025	rod	16
ER2-P4	rod	2
ER4-P4	rod	1
SM1CP2	Externally SM1-Threaded End Cap	4
SM1L05	LensTube	4
SM1L03	LensTube	2
-	Breadboard with a proper size	1
Emission		
SM1A12	M25*0.75 Objective adaptor	1
AC254-080-A-ML	achromatic lens f=80	2
SM1A9	C-Mount/SMA adaptor	2
SM1AB2	C-Mount/SMA adaptor	2
SM1P1	Optic mount	2
Fine-tunable Fiber Mount		
CXYZ05	xyz fiber mount	1
SM05L03	lens tube	1

product#	Description	#
SM05FC	fiber adapter	1
SM05T2	capla for lens tube	1

Note, Thorlabs products can get obsolete routinely; replace them with corresponding newer products.

### Semrock

product#	Description
FF493/574-Di01-25x36	493/574 nm BrightLine® dual-edge standard epi-fluorescence dichroic beamsplitter
FF01-520/35-25	520/35 nm BrightLine® single-band bandpass filter
FF01-630/69-25	630/69 nm BrightLine® single-band bandpass filter
FF562-Di03-25x36	562 nm edge BrightLine® single-edge standard epi-fluorescence dichroic

### Nikon

product#	Description
MRD70170	CFI60 PLAN APOCHROMAT LAMBDA D 10X

### Edmund Optics

product#	Description
#69-899	500nm, 25.2 x 35.6mm, Dichroic Longpass Filter
#69-898	450nm, 25.2 x 35.6mm, Dichroic Longpass Filter

### Doric

Fiber patch cable should be selected based on application. We recommend "[Low-Autofluorescence Bundle Branching Fiber-optic Patch Cord](#)" from Doric for standard applications.

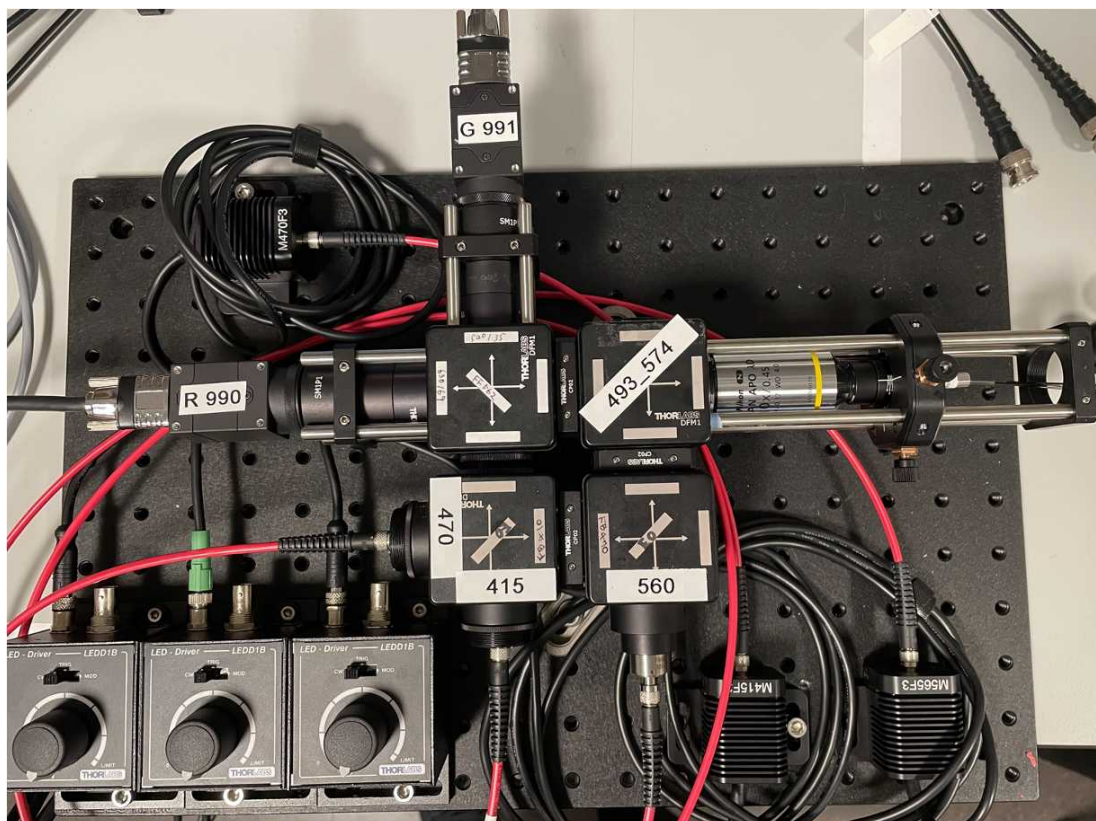
### FLIR-Teledyne

CMOS sensors should be selected based on required frame rate and sensitivity. We use "[BFS-U3-20S4M-C](#)", which is based on Sony IMX422 sensor, as default CMOSs.

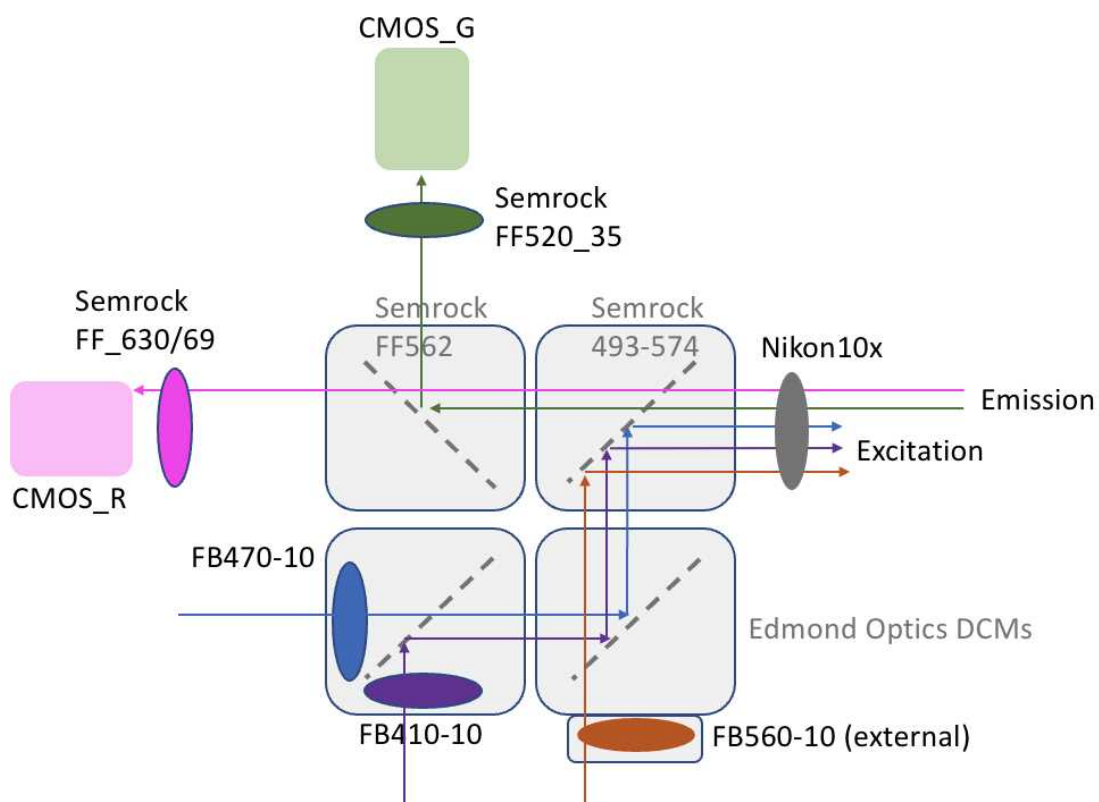
## SAFETY WARNINGS

- ! Although resultant FIP setup will be a fairly optically closed system, in particular during building, experimenter(s) could be exposed to stray light from LEDs. In general, inappropriate use of any Fiber-Coupled LEDs may result in permanent eye damage. To prevent injury, use the LEDs in accordance with the International Standard "Photobiological Safety of Lamps & Lamp Systems" IEC 62471. The LEDs used in this system fall under RG2 - Moderate Risk Group in accordance to the standard IEC 62471:2006.

## System Overview

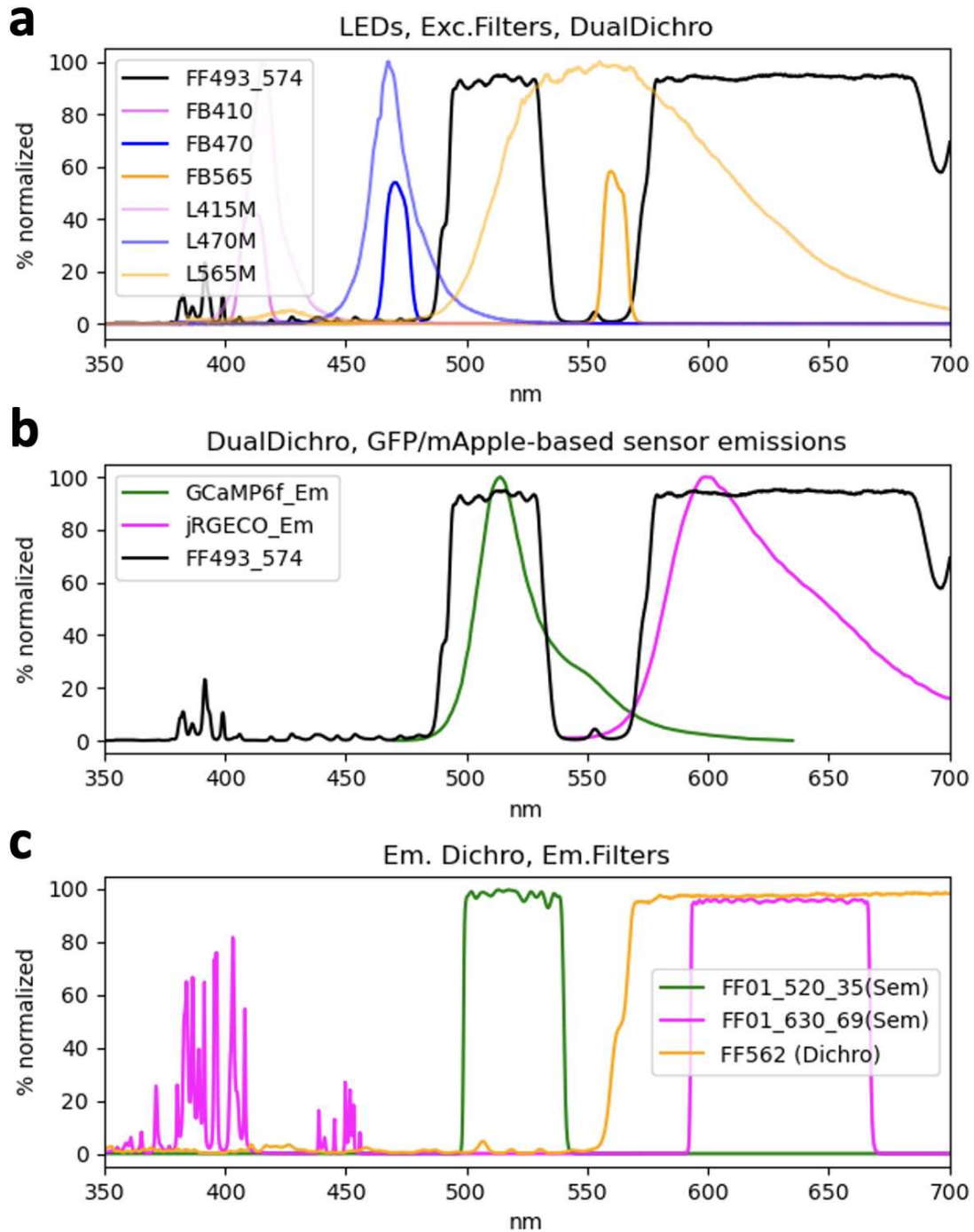


**Fig. 1. System Overview**



**Fig. 2. Design diagram**

See the **Materials** section for products used in the setup.



**Fig. 3. Spectral features of LEDs, optical filters, and genetic sensor emission signals.**

**a**, Normalized intensity of excitation LEDs (L415M, L470M, L565M), and transmission of excitation band-pass filters (FB410, FB470, FB560) and the dual dichroic filter (FF493\_574).

**b**, Transmission of the dual dichroic filter (duplicated from **a**) separating emission spectrum from excitation spectrum (**Fig. 1**), and normalized intensity of GCaMP6f and jRGECO1a. Note, this design would be suited for most cpGFP- and cpmApple-based sensors.

**c**, Transmission of the dichroic filter separating green and red emission signals (FF562), and emission band-pass filters (FF01\_520\_35 for green and FF01\_630\_69 for red)

## Assembling optical parts

2h

- 2 Assembling the cubes 30m  
(This step-by-step part will have corresponding CAD models later.)
- 3 Adding optical filters, collimators, and the objective lens 30m
- 4 Assembling the fiber positioning module 15m
- 5 Assembling the CMOS modules 30m
- 6 Connecting excitation light sources with fibers 15m

## Alignment

30m

- 7 Adjusting the fiber location so that fiber end makes a crisp image on CMOSs
- 8 Adjusting angle of collimators so that excitation light coming from fiber patch cables would get roughly uniform. 30m