



## Nov 23, 2020

# Recombineering

#### Elizabeth Fozo<sup>1</sup>

<sup>1</sup>In-house protocol

1 Works for me

This protocol is published without a DOI.

Eadewunm

#### PROTOCOL CITATION

Elizabeth Fozo 2020. Recombineering. **protocols.io** https://protocols.io/view/recombineering-bpzhmp36

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Nov 23, 2020

LAST MODIFIED

Nov 23, 2020

PROTOCOL INTEGER ID

44809

DISCLAIMER:

DISCLAIMER: THIS IS A WORK IN PROGRESS. IT IS FOR INFORMATIONAL PURPOSES ONLY; USE AT YOUR OWN RISK

The protocol content here is for informational purposes only and does not constitute legal, medical, clinical, or safety advice, or otherwise; content added to <a href="protocols.io">protocols.io</a> is not peer-reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol should not substitute for independent professional judgment, advice, diagnosis, or treatment. Any action you take or refrain from taking using or relying upon the information presented here is strictly at your own risk. You agree that neither the Company nor any of the authors, contributors, administrators, or anyone else associated with <a href="protocols.io">protocols.io</a>, can be held responsible for your use of the information contained in or linked to this protocol or any of our Sites/Apps and Services.

### BEFORE STARTING

Template: if you are using pKD4 (kan cassette), you can use 200 ng of DNA for transformation. Plate on Kan 20 ug/mL plates (lower than normal). Pick to Kan 30 plates to verify really resistance.

Dr. Fozo has not had an issue obtaining transformants on Kan 30 initially: only problematic if a weak RBS is driving Kan.

Be careful not to overgrow cells: we have seen a decrease in efficiency if the cells start approaching an OD 600 of 0.7 before you induce at 42.

If you are not using Kan 20 plates, use more DNA, and rescue for 2 hours.

Storing transformation mix: some people find that leaving the mix on the bench overnight increases the number of positive transformants when they plate out the next day.

Tem	nl	lat	-
16111	μı	ıaı	יכ

- 1 Prepare a PCR product. Typically, 200-600 ng/transformation. See note below. If using a plasmid template, may have to digest using DpnI or another enzyme to remove the additional plasmid
- 2 Following purification, combine PCR products and ethanol precipitate. Resuspend products in water

I usually combine 2-4 PCR products into 5 ul and use 2.5 ul for a first recombineering attempt.

#### Strains

- 3 Grow up 25 ml of a strain of choice at 30 or 32 degrees (do NOT use DY330. Use NM400, NM1100, or some such derivative).
- 4 When OD600 approximately 0.4, transfer to 42 degrees shaking water bath for 15 20 minutes.
- 5 After 15-20 minutes, place the flask in an icy slurry and swirl. Keep in icy slurry for about 2 minutes.
- 6 Spin at 4 degrees, 10 minutes, 4150 rpm.
- 7 Wash cells with 1 ml cold 10% glycerol at 4 degrees, three times
- 8 Resuspend cells in **100 μl** cold 10% glycerol.
- 9 Electroporate: use **50 μl** of cells + DNA. Immediately after electroporation, add 950 ul of SOC. Shake at 30 degrees for 1 hour. Plate 1/10 onto one plate, and concentrate the remaining 900 ul and plate to a second plate.