

Mar 31, 2025

## Modified Frame-projected Independent Fiber Photometry (FIP) System\_Triggering system

DOI

[dx.doi.org/10.17504/protocols.io.kxygx3e6wg8j/v1](https://dx.doi.org/10.17504/protocols.io.kxygx3e6wg8j/v1)



Kenta M. Hagihara<sup>1</sup>

<sup>1</sup>Allen Institute for Neural Dynamics

Allen Institute for Neural...



Kenta M. Hagihara

Allen Institute for Neural Dynamics

OPEN  ACCESS



DOI: [dx.doi.org/10.17504/protocols.io.kxygx3e6wg8j/v1](https://dx.doi.org/10.17504/protocols.io.kxygx3e6wg8j/v1)

**Protocol Citation:** Kenta M. Hagihara 2025. Modified Frame-projected Independent Fiber Photometry (FIP) System\_Triggering system. [protocols.io https://dx.doi.org/10.17504/protocols.io.kxygx3e6wg8j/v1](https://dx.doi.org/10.17504/protocols.io.kxygx3e6wg8j/v1)

### Manuscript citation:

Aggarwal, Abhi, Adrian Negrean, Yang Chen, Rishyashring Iyer, Daniel Reep, Anyi Liu, Anirudh Palutla, et al. "Glutamate Indicators with Increased Sensitivity and Tailored Deactivation Rates." bioRxiv, March 24, 2025. <https://doi.org/10.1101/2025.03.20.643984>.

**License:** This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

**Protocol status:** Working

We use this protocol and it's working

**Created:** July 06, 2023

**Last Modified:** March 31, 2025

**Protocol Integer ID:** 84608

**Keywords:** fiber photometry, calcium imaging, neuromodulator imaging, CMOS-based photometry

## Abstract

Frame-projected independent fiber photometry (FIP) is a method to measure fluorescent sensor signals through optical fibers implanted in living animals, using a camera to record video of the fiber faces. To precisely control timings of excitation and camera frame acquisition for FIP, and of other external stimulation apparatus (e.g. LEDs for photo-stimulation), we use a Teensy 4.1 microcontroller, which generates voltage pulses without relying on continuous communication with the operating system of the experimental computer for timing. This protocol is for the 3 excitation LED + 2 collection CMOS cameras design.

Triggering-controlling software (an Arduino sketch) and data aquisition software (a Bonsai workflow) can be found in an associated github repository.

## Guidelines

Teensy 4.1 only provides ~3.3V. If the device you want to trigger requires 5V (or more), the voltage needs to be up-regulated.

## Materials

### Equipment

Teensy 4.1	NAME
PJRC	BRAND
DEV-16771	SKU
<a href="https://www.pjrc.com/store/teensy41.html">https://www.pjrc.com/store/teensy41.html</a>	LINK

### Equipment

Terminal Block Breakout Board Module for Teensy 4.1, Screw Mount Version	NAME
Breakout board	TYPE
OONO	BRAND
D-1395P	SKU

## Software

[Arduino software](#)

[Teensy extension](#)

[AIND FIP DAQ Control github repo](#)

## Cables and connectors

USB2.0 cable (USB-A to Micro USB)

Jumper (or bare) wires

BNC cables

BNC sockets ([Amazon link for an example product](#))

BNC connector ([Amazon link for an example product](#))

[FLIR camera GPIO cable \(6-pin Hirose\)](#)

## Miscellaneous

Solder (we used: Lead free wire from Amerway)

Soldering tools (we used: Weller WX2)

3D-printer (we used: Stratasys Dimension 1200es and IC3D Virago 700, with ABS and PETG filament, respectively)

A computer that can run Arduino/Teensy software. We used Windows 10 PCs but Max/Linux would work too.

## Before start

Please see the accompanying protocol for the main FIP hardware design:

### Protocol



NAME

Modified Frame-projected Independent Fiber Photometry (FIP) System\_Hardware

CREATED BY

Kenta M. Hagihara

PREVIEW

## Preparing an enclosure

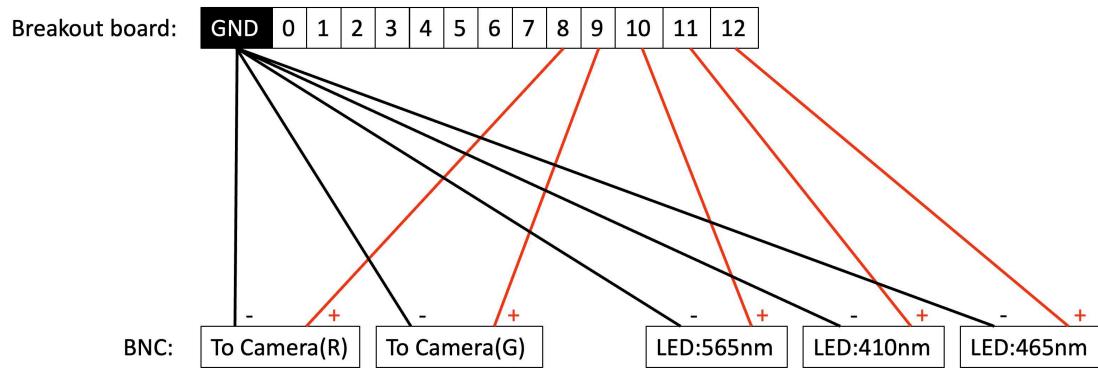
- As the Teensy 4.1 is a bare board, it should securely covered in an enclosure. The enclosure should be able to securely house Teensy 4.1, at least 5 BNC sockets, and optionally a small breadboard. Plastic 3D-printing (CAD design can be found here:  
 0294-100-01\_X01.step 246KB     0294-100-02\_X01.step 3MB ) would be sufficient and could be any materials and designs.

## Setting up an experimental PC and Teensy 4.1 device

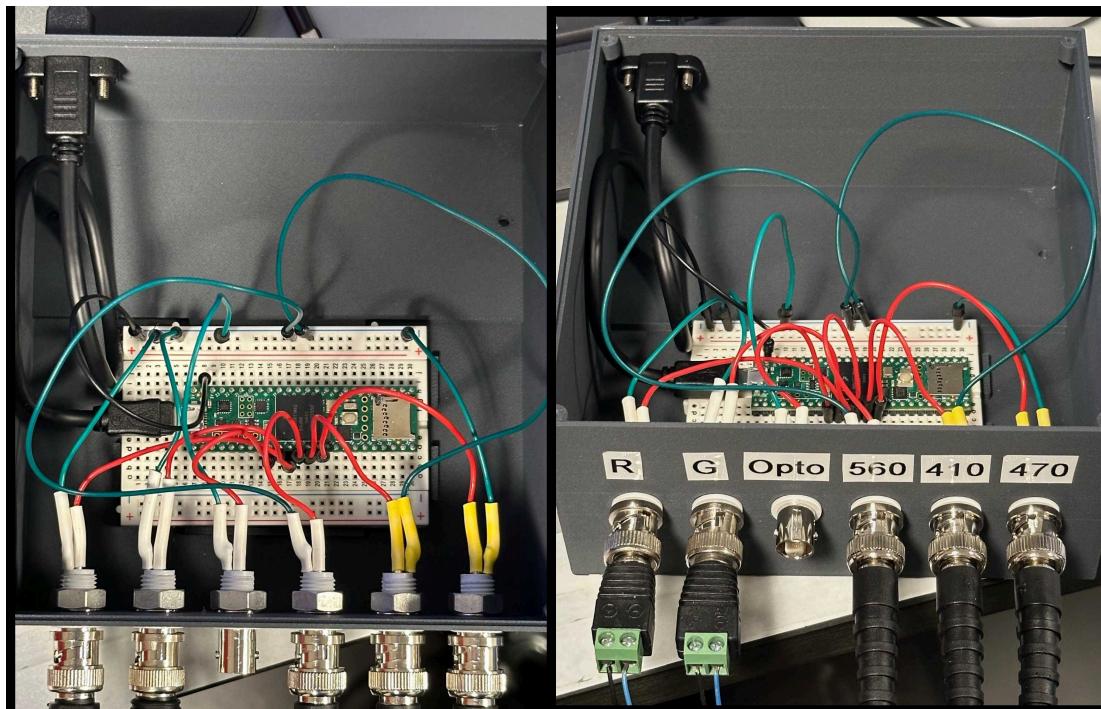
- Install Arduino app on your experimental PC from the following link: [Arduino software](#). 25m
- Add Teensy extension to Arduino app by following [the official instructions](#). 10m
- Download .ino sketch from the [github repo](#). 10m
- Connect Teensy 4.1 device to the experimental PC using a USB cable, set proper COM port in the Arduino app, and upload the .ino sketch to the device. How to find and set up the corresponding COM port can be found here: <https://support.arduino.cc/hc/en-us/articles/4406856349970-Select-board-and-port-in-Arduino-IDE>. 5m

## Connecting the Teensy 4.1 device to hardware to be controlled

- Place the Teensy 4.1 device on a breakout board (or a generic electric breadboard). 5m
- Attach the breakout board and BNC(female) sockets to the enclosure. 10m
- Solder wires between the breakout board and the BNC sockets according to this diagram for 3 excitation LEDs and 2 CMOS sensors configuration. 30m



**Fig1. Diagram for connections**



Red wires (hot), green wires (ground)

- 9 Connect the device to hardware (cameras and LEDs) to be controlled using BNC cables.

10m

Protocol



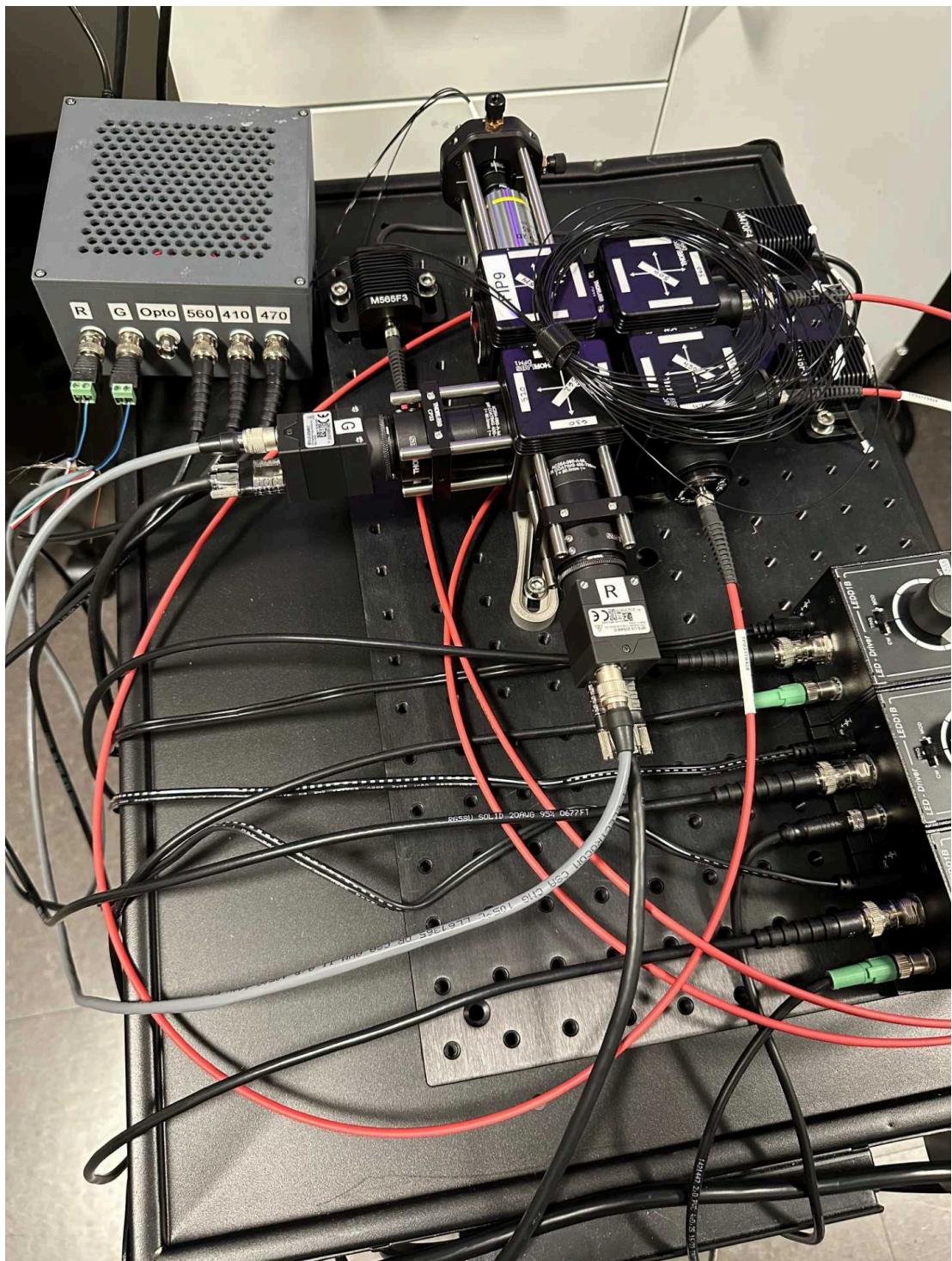
NAME

**Modified Frame-projected Independent Fiber Photometry  
(FIP) System\_Hardware**

CREATED BY

Kenta M. Haghara

**PREVIEW**



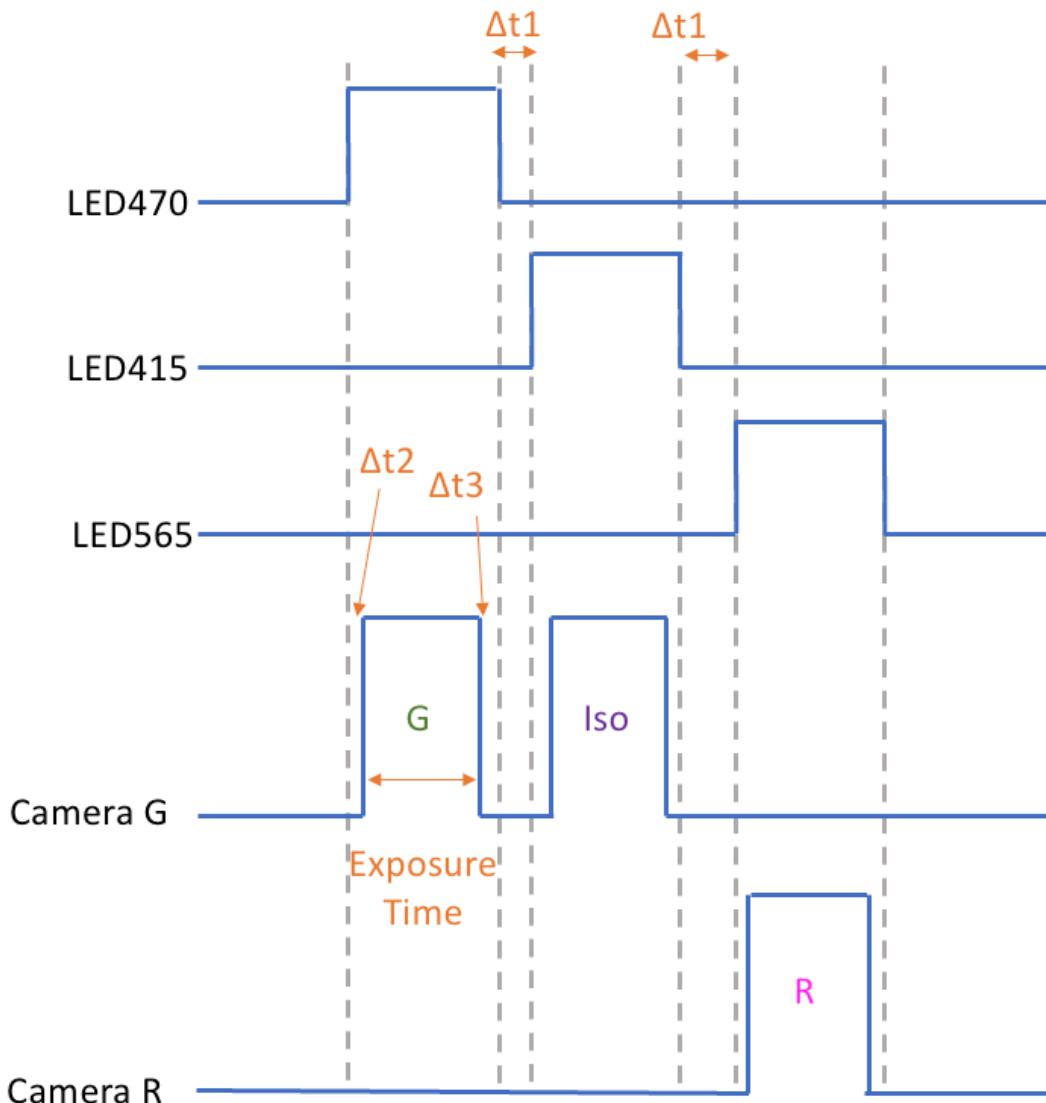
All BNC ports connected to corresponding devices of the main hardware

## Software parameter setting

30m

- 10 Based on acquisition frame rate and the CMOS cameras to be used, set the acquisition parameters in the .ino sketch.

30m



**Fig2. Scheme of temporal multiplexing of excitation and data acquisition**

When acquisition is 60Hz (20Hz for each channel) with sufficiently fast CMOS sensors (e.g. FLIR, BFS-U3-20S4M-C), an example parameter set could be:

$$\text{ExpT} = 15650 \mu\text{s}$$

$$\Delta t_1 = 666 \mu\text{s}$$

$$\Delta t_2 = 300 \mu\text{s}$$

$$\Delta t_3 = 50 \mu\text{s}$$

$$\text{ExpT} + \Delta t_1 + \Delta t_2 + \Delta t_3 = 16666 \mu\text{s} (= 1/60 \text{ s})$$

For better signal,  $\Delta t_1/\Delta t_2/\Delta t_3$  should be minimized to make the exposure time longer. If they are too short, however, the camera will not be able to unload the data (when  $\Delta t_1$  is

too short) or LED instability at the onsets (when  $\Delta t_2$  is too short) would lead to periodic noise generation. Based on excitation light sources and CMOS models, optimize these parameters empirically.

## Protocol references

Kim, C., Yang, S., Pichamoorthy, N. *et al.* Simultaneous fast measurement of circuit dynamics at multiple sites across the mammalian brain. *Nat Methods* **13**, 325–328 (2016). <https://doi.org/10.1038/nmeth.3770>

TeensyDuino .ino sketch can be found at [AIND github repo](#).

CAD design can be found here: [!\[\]\(4cafc60cd39da821525d7c6589540296\_img.jpg\) 0294-100-01\\_X01.step 246KB](#) [!\[\]\(775cbf51955011dd735a723560100a76\_img.jpg\) 0294-100-02\\_X01.step 3MB](#)

## Acknowledgements

We thank Joon Jung (Allen Institute, SIPE) for designing and 3D-printing the enclosure.