



Removal of Melanin V.2

Jason E Stajich¹

¹UC Riverside

Version 2 ▾

Aug 22, 2020

1

Works for me

dx.doi.org/10.17504/protocols.io.bj6zkrf6

stajichlab



Jason Stajich
UC Riverside

ABSTRACT

Principle is that CTAB is charging the anionic nucleotides whereby neutral polysaccharides/ melanins are remaining in supernatant. This method also uses urea with the idea that the presence of urea helps to solubilize hydrophobic compounds that would otherwise potentially interact with the hydrophobic core of the CTAB micelles.

EXTERNAL LINK

<http://onlinelibrary.wiley.com/doi/10.1111/j.1600-0749.2004.00155.x/abstract>

DOI

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PROTOCOL CITATION

Jason E Stajich 2020. Removal of Melanin. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.bj6zkrf6>



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CREATED

Aug 22, 2020

LAST MODIFIED

Aug 22, 2020

PROTOCOL INTEGER ID

40889

STEPS MATERIALS

NAME	CATALOG #	VENDOR
NaCl	53014	Sigma Aldrich
Water		
Guanidine hydrochloride	G3272-1KG	Sigma Aldrich
70% ethanol		Fisher Scientific

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1

Add

 Water

to ~  **100 μ l** -  **200 μ l** DNA/RNA solution until a volume of  **400 μ l** is reached.

2 Add  **130 μ l** [M] **5 Molarity (M)**

 NaCl
by Sigma Aldrich
Catalog #: 53014

3 Add  **1.6 mL** mL of CTAB-Urea solution

  CTAB-Urea buffer
by Jason Stajich

[PREVIEW](#) [RUN](#)

3.1 50 mM Tris-HCl, pH 7.0

3.2 1% CTAB

3.3 4M Urea

3.4 1 mM EDTA

- 4 Mix samples (by hand).
- 5 Incubate overnight at 4°C
 Overnight
- 6 Centrifuge for **00:15:00** at max speed at 4°C
- 7 Remove the solution. Be very careful in this step!
- 8 Resuspended in **400 μL** **7 Molarity (M)**

Guanidine hydrochloride
by Sigma Aldrich
Catalog #: **G3272-1KG**
CAS Number: 50-01-1

- 9 Add 2 **2 μL** **VOLUMES** Vol of EtOH (100%)
- 10 Incubate on ice for
 01:00:00
- 11 **15300 x g, 4°C 00:15:00**

- 12 Wash with

70% ethanol
by Fisher Scientific

- 13 Wash again with

70% ethanol
by Fisher Scientific

14  **20800 x g, Room temperature 00:10:00 , (or maximum speed)**

15 Remove the supernatant.

16 Dry pellet

17 Resuspend in TE buffer