

Mar 19, 2021

Primary Lung Fibroblasts (PLF) Plating/Freezing Protocol

PennLBI¹

¹Penn Lung Biology Institute

Works for me

This protocol is published without a DOI.

LungMap2 Consortium Tech. support email: lungmap2dcc@gmail.com Click here to message tech. support

Stephanieloos

SUBMIT TO PLOS ONE

ABSTRACT

Primary Lung Fibroblasts (PLF) Plating/Freezing Protocol

90 mL DMEM NUTRIENT MIX F12 (Invitrogen/Cell Center, cat# 11320033)

10 mL 10% FBS (Invitrogen, cat#10437-028)

1 mL ANTIBIOTIC ANTIMYCOTIC-Anti-Anti 100x (Invitrogen/Cell Center, cat#15240096)

Freeze Media

40 ml PLF Media

5 mL 10% FBS (Invitrogen, cat# 10437-028)

5 mL DMSO (Sigma, cat#D2650-100ml)

Nunc™ EasYFlask™ Cell Culture Flasks, T75, filter (hermo/ Life Tech, cat#156499)

TRYPSIN 0.05% EDTA (Invitrogen/Cell Center, cat#25300054)

CryoVials (Neta Sci, cat#430659)

PROTOCOL CITATION

PennLBI 2021. Primary Lung Fibroblasts (PLF) Plating/Freezing Protocol. protocols.io https://protocols.io/view/primary-lung-fibroblasts-plf-plating-freezing-prot-bthenj3e

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Mar 19, 2021

LAST MODIFIED

Mar 19, 2021

PROTOCOL INTEGER ID

48390

DISCLAIMER:

DISCLAIMER - FOR INFORMATIONAL PURPOSES ONLY; USE AT YOUR OWN RISK

The protocol content here is for informational purposes only and does not constitute legal, medical, clinical, or safety advice, or otherwise; content added to protocols.io is not peer reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol should not substitute for independent professional judgment, advice, diagnosis, or treatment. Any action you take or refrain from taking using or relying upon the

information presented here is strictly at your own risk. You agree that neither the Company nor any of the authors, contributors, administrators, or anyone else associated with <u>protocols.io</u>, can be held responsible for your use of the information contained in or linked to this protocol or any of our Sites/Apps and Services.

| Ρ | lating | and | Freezin | aPLF |
|---|---------|-----|------------|-------|
| | iatilig | and | I I CCZIII | gi Li |

| ating | ating and FreezingPLF | | | | |
|-------|--|--|--|--|--|
| 1 | In TC hood label 1-2 T75 flasks with lung ID, date, disease or NI (normal), initials, and number of cells. | | | | |
| 2 | Obtain single cell suspension from lung, count cells, and keep on ice. | | | | |
| 3 | Spin down 10-12 million cells in 4C centrifuge for 5 min at 300xg(rcf). | | | | |
| 4 | Using a 10mL serological pipette (slow settings), resuspend cells with 10 mL of cold PLF media in T-75 flaks/s and put in incubator. | | | | |
| | 4.1 a. It is important to mix the cells evenly | | | | |
| 5 | After ~24 hours, change PLF media on the cells. | | | | |
| 6 | Change PLF media 2-3 times a week until cells are relatively confluent. They grow very patchy, so it is sometimes hard to tell, but this usually takes about 3 weeks. | | | | |
| 7 | Once confluent, split the cells out into more flasks. Usually 1:6 is OK for the first split, but depending on confluency, this might change. | | | | |
| | 7.1 a. If you started with 2 plates of cells, can split them each out 1:12 | | | | |
| 8 | Once those flaks are confluent (usually 3-4 days later), split 1:4 so you end up with 24 flasks total. | | | | |
| 9 | Once the 24 flasks are confluent (usually 3-4 days later) freeze down cells by trypsinizing the flasks with 0.05% trypsin and resuspending cells in 1 mL of Freeze Media/vial. | | | | |
| | 9 1 a. It is easiest to label cryo vials, with lung ID, disease/NI, date, initials, before trypsinizing flasks. | | | | |

m protocols.io 03/19/2021

There will be a total of 50 vials (resuspend pellet in 50 mL of Freeze media).

b.

9.2

- 9.3 c. There is approx. 1 million cells per vial. Can count cells if you want ,but is not necessary for freezing.
- 10 10. Put vials in a Mr. Freezy and put in -80 for at least 24 hours.
- 11. Transfer vials to liquid nitrogen tank and update google sheetàHuman Samples-à liquid nitrogen tank