

AUG 01, 2023

Environmental DNA (eDNA) Sample Shipping Protocol

Alexandria B

Meghan M. Shea¹, Boehm¹

¹Stanford University



Meghan M. Shea Stanford University

ABSTRACT

This is a protocol for mailing PCR amplicons generated from eDNA samples, specifically via the following earlier sampling & filtration, extraction, and PCR protocols, e.g.:

OPEN ACCESS



DOI:

dx.doi.org/10.17504/protocol s.io.dm6gp38jpvzp/v1

Protocol Citation: Meghan M. Shea, Alexandria B Boehm 2023. Environmental DNA (eDNA) Sample Shipping Protocol. protocols.io https://dx.doi.org/10.17504/protocols.io.dm6gp38jpvzp/v1

MANUSCRIPT CITATION: TBD

License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working We use this protocol and it's working

Protocol



NAME

Coastal Environmental DNA Sampling & Gravity Filtration Protocol

CREATED BY

Meghan M. Shea

PREVIEW

Protocol



NAME

DNA Extraction Protocol from Sterivex Filters

CREATED BY

Meghan M. Shea

PREVIEW

Created: Jun 19, 2023

Last Modified: Aug 01,

2023

PROTOCOL integer ID:

83661

Keywords: PCR amplicons, eDNA, shipping, environmental DNA

Protocol



NAME

Environmental DNA (eDNA) COI PCR Amplification and Gel Electrophoresis Protocol

CREATED BY

Meghan M. Shea

PREVIEW

While this protocol was designed for mailing to the <u>Georgia Genomics and</u> <u>Bioinformatics Core</u>, the general principles may be relevant to other shipping contexts.

Acknowledgements & Attributions:

Thanks to Bradley B. Tolar for guidance on successful shipping protocols.

IMAGE ATTRIBUTION

Meghan M. Shea

MATERIALS

General Laboratory Equipment:

| Equipment | Specific Model Used |
|--|-------------------------------------|
| 10% bleach solution in spray bottle | NA |
| >70% ethanol solution in spray bottle | NA |
| RNase Away solution in spray bottle | NA |
| 20 µL multichannel pipette with sterile tips | Various |
| UV Crosslinker | UVP CL-1000 Ultraviolet Crosslinker |
| PCR Tube Holders | Various |
| Ice bin | Various |
| Parafilm | Various |
| Kimwipes | Various |

Shipping Supplies:

| Material | Amount Needed | Source | Link | Approx. Cost |
|--|--|--|--|--------------------|
| 96-well plate | 1/96 samples | Applied Biosystems via ThermoFish er Scientific (4346906) | https://www.t hermofisher.c om/order/cat alog/product/ 4346906 | \$129/20 plates |
| Aluminum adhesive seal for 96-well plate | 1/plate | Unknown | NA | Unknown |
| Cardboard | Enough to cover both sides of each plate | Various | NA | Unknown |
| Ziploc bags | Various | Various | NA | Unknown |
| Bubble wrap | Various | Various | NA | Unknown |
| Hard container for holding plates | Various | Various | NA | Unknown |
| Styrofoam cooler with associated outer cardboard box | 1 | Reagents previously shipped on ice | NA | Recycled |

Preparing Samples for Shipping

1 Clean bench area with 10% bleach, 70% ethanol, and RNase Away

Note

Ideally, this should be a designated post-PCR bench area.

- 2 Wipe down 20 μ L multichannel pipette and several PCR tube holders with RNase away. Run in UV Crosslinker for 10 minutes on each side.
- Wipe down ice bin with 10% bleach, 70% ethanol, and RNase Away, fill with ice, and add PCR products stored in 8-strip PCR tubes

| 10 | Seal each plate in a new Ziploc bag and label accordingly |
|-----|---|
| 9 | Cut two pieces of cardboard that fit on either side of each plate; tape the cardboard around each plate to form a sandwich (to keep it from getting punctured while shipping) |
| 8 | Wrap each plate in Kimwipes to absorb any condensation that occurs during shipping |
| 7 | Parafilm around the edges of each plate |
| 6.1 | Press around the edges of each well to make sure they are sealed |
| 6 | Seal the plates carefully with aluminum adhesive seals |
| 5 | Being careful not to open PCR tubes directly over the 96-well plates, add 15 μ L of sample to each well using multichannel pipette, making sure you keep track of the orientation of how the plate was filled |
| 4 | Label all 96-well plates needed for storing samples accordingly |

- Seal hard container with tape and place in its own Ziploc bag; label accordingly
- 13 Place bagged box in a Styrofoam cooler that fits in an associated cardboard box and fill with ice packs
- 14 Tape Styrofoam cooler closed and secure it in the corresponding cardboard box

Note

If you need to include any order information or documents alongside the samples, don't forget to put them in the box here.

15 Store cardboard box at -20°C until ready to mail