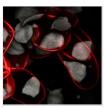


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Aug 10, 2022

# Anthoceros agrestis Bonn (hornwort) transformation v02

Eftychis Frangedakis<sup>1</sup>, Manuel Waller<sup>2</sup>

<sup>1</sup>University of Cambridge; <sup>2</sup>University of Zürich

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# Hornworts



**ABSTRACT** 

Anthoceros agrestis Bonn (hornwort) transformation v02

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MATERIALS TEXT

**KNOP** recipe:

Stock 1

25g/L KH<sub>2</sub>PO<sub>4</sub>

Stock 2

25g/L KCl

Stock 3

25g/L MgSO<sub>4</sub> 7H<sub>2</sub>O

Stock 4

100g/L Ca(NO<sub>3</sub>)<sub>2</sub> 4H<sub>2</sub>O

autoclave and store at RT or 4°C

# KNOP solid working solution:

In 600 mL of water add:

10ml Stock 1

10ml Stock 2

10ml Stock 3

10ml Stock 4

12.5mg FeSO<sub>4</sub>7H<sub>2</sub>O

pH to 5.8 with KOH

top up water to 1L after adjusting pH

add 7 gr of Gelzan - G1910 - CAS Number 71010-52-1

### KNOP liquid working solution:

In 600 mL of water add:

10ml Stock 1

10ml Stock 2

10ml Stock 3

10ml Stock 4

12.5mg FeSO<sub>4</sub>7H<sub>2</sub>O

10 gr of sucrose (1% w/v final concentration, 2% also fine)

40mM MES (very important)



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#### pH to 5.5 with KOH

top up water to 1L after adjusting pH

Filter sterile (do not autoclave), aliquot into 50mL falcon tubes and store at -20°C.

.....

Sterile disposable scalpels (#0501, Swann Morton)

Razor blades (#11904325, Fisher Scientific)

100 µm cell strainer (#352360, CORNING),

6-well plate (#140675, ThermoFisher)

3',5'-dimethoxy-4'-hydroxyacetophenone (acetosyringone) (#115540050, Acros Organics, dissolved in dimethyl sulfoxide (DMSO) (#D8418, SIGMA))

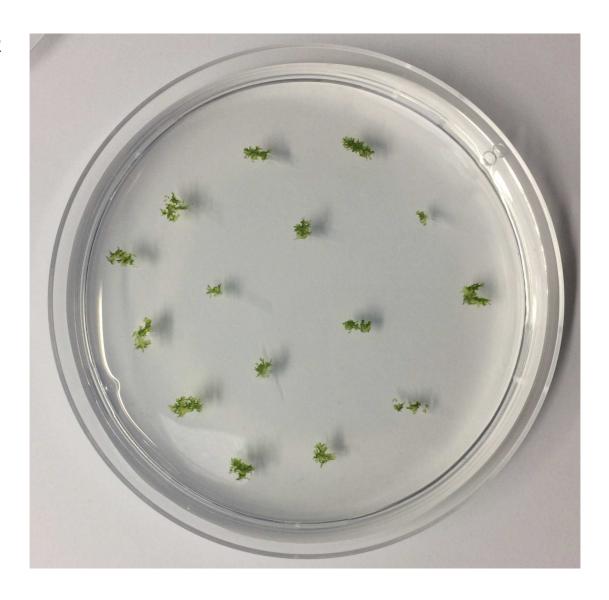
Cefotaxime (#BIC0111, Apollo Scientific)

Hygromycin (#10687010, Invitrogen)

Corning Disposable Vacuum Filter/Storage Systems (#430767)

# 1 1

**IMPORTANT**: The light intensity used to cultivate *A. agrestis* tissue is a very critical factor for successful transformation. Tissue should be grown under low light intensity (3-5  $\mu$ mol m<sup>-2</sup>s<sup>-1</sup>) and should have a morphology similar to the tissue in **Figure 1** 





**Figure 1:**Top: 4 week old *A. agrestis* Bonn thallus
Bottom: 7 week old *A. agrestis* Bonn thallus (this tissue is also good for transformation)

3 Axenic cultures of *A. agrestis* gametophytes can be routinely propagated by monthly subculturing as shown in **Figure 2**.

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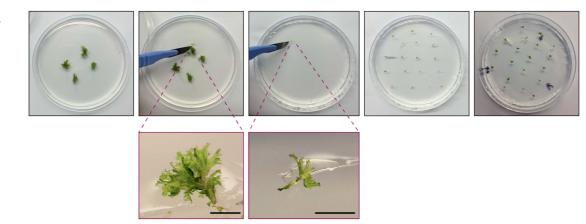
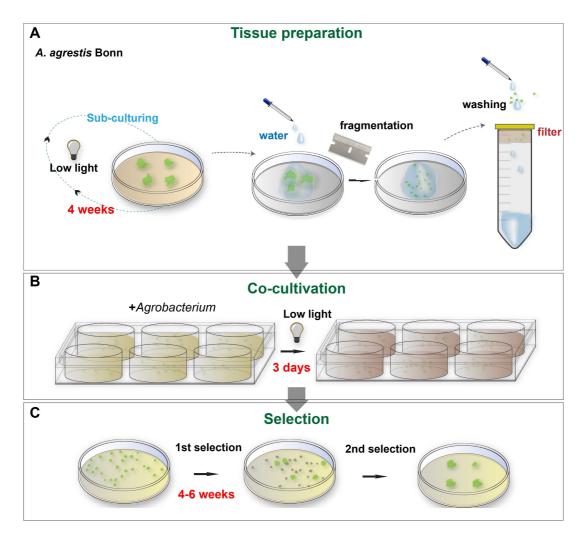


Figure 2: A. agrestis Bonn tissue culturing

For sub-culturing, a small piece of thallus tissue is cut using sterile disposable scalpels and placed on plates containing fresh growth medium. Scale bars: 2 mm. Petri dish dimensions: 92 x16 mm.

Tissue similar to the bottom images is optimal for transformation



**Figure 3: Transformation method outline**. A) Thallus tissue is routinely propagated on a monthly basis under low light. 4-6 week old tissue is fragmented with the aid of a razor blade, transferred to a cell strainer, and washed thoroughly with sterile water. B) The tissue is then co-cultivated with *Agrobacterium* for three days (under low light) and C) spread on antibiotic-containing growth medium. After approximately 4-6 weeks, putative transformants are visible. A final round of selection is recommended to eliminate false-positive transformants.

### 6 Tissue preparation:

- Collect approximately 1 g of thallus tissue grown for 4-6 weeks under low light intensity (approximately 0.1 g of tissue per petri dish - 10 petri dishes in total). Figure 1 and Figure 4.1
- Transfer the tissue into an empty petri dish, add sterile water until the tissue is covered and fragment using a razor blade (it takes approximately 5 mins, similar to Video 1). Figure 4.2-3
- Transfer the tissue from the petri dish into a cell strainer positioned on a falcon tube using sterile scalpels and wash the tissue using ~100 ml of sterile water or until the flow through

# 7 Agrobacterium culture preparation:

- Inoculate 5 mL LB media with 3-4 Agrobacterium colonies (AGL1:15 μg/mL rifampicin, 50 μg/mL carbenicillin) (GV3101:50 μg/mL rifampicin, 25 μg/mL gentamicin) and the plasmid-specific selection antibiotic.
- Incubate the preculture at 28°C for 2 days at 110 rpm.
- Centrifuge 5 mL of 2 d Agrobacterium culture (no need to measure OD) for 7 min at 2000 xq.
- Remove supernatant and re-suspend in 5 mL liquid KNOP plus 1% (w/v) sucrose and 100  $\mu$ M acetosyringone.
- Incubate the culture with shaking (120 rpm) at 28°C for 3-5 hours.

### 8 Co-cultivation:

- Transferred the fragmented thallus tissue into a 6-well plate (transfer 1/6 of the 1 g tissue into a single well) with 5 mL of liquid KNOP medium supplemented with 1% (w/v) sucrose and 30-40 mM MES (VERY IMPORTANT), pH 5.5, 80 μL of Agrobacterium culture and acetosyringone at final concentration of 100 μM. Figure 4.7
- Co-cultivate the tissue with the *Agrobacterium* for 3 days on a shaker at 110 rpm, with only ambient light.
- Using a sterile plastic pipette transfer the tissue of one well into a cell strainer, drain and then transfer on growth media containing the appropriate antibiotic (onto 1 petri dish from one well). To facilitate spreading of the tissue, 1-2 mL of sterile water is added to the petri dish. Figure 4.8-11
- After 4-6 weeks successful transformants are visible on the petri dish (successful transformants can be identified using a dissecting scope after 4 weeks selection (sometimes as early as 2 weeks) based on rhizoid production and/or fluorescence if such a marker is present on the construct). Figure 4.12
- The emergence of rhizoids is an indication of successful transformation (yellow arrow: transformed thallus fragment, blue arrow: dying thallus fragment). Figure 5

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# 2nd selection (optional):

■ To eliminate false positives, after 4 weeks transfer the tissue to fresh growth media containing 100  $\mu$ g/mL cefotaxime and 10  $\mu$ g/mL Hygromycin. To facilitate spreading of tissue on the petri dish add 2 mL of sterile water. Grow at 21°C under 12 hours light and 12 hours dark, 35  $\mu$ mol m<sup>-2</sup>s<sup>-1</sup>

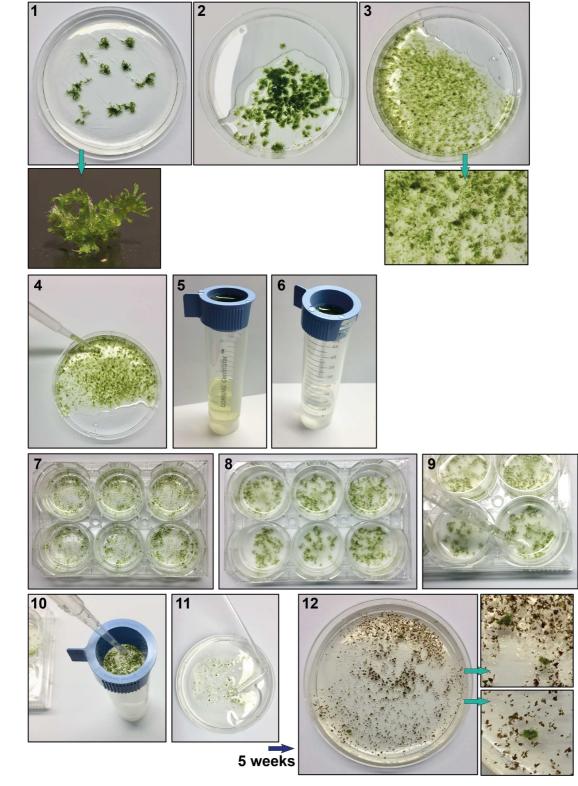


Figure 4



**Figure 5**: The emergence of rhizoids is an indication of successful transformation (yellow arrows: transformed thallus fragment, blue arrow: dying thallus fragment).

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Example of plate with successful transformants 8 weeks after co-cultivation.

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Video 1, Example of tissue fragmentation for *A. agrestis* Bonn.