

FEB 16, 2024

## OPEN ACCESS



DOI:

dx.doi.org/10.17504/protocols.io. 3byl4qoxrvo5/v1

Protocol Citation: eduard.bente a, María Sanchiz Calvo, Veerle Baekelandt 2024. Immunofluorescence staining, vibratome sections. protocols.io https://dx.doi.org/10.17504/protoc ols.io.3byl4qoxrvo5/v1

License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

**Protocol status:** Working We use this protocol and it's working

Created: Feb 16, 2024

## • Immunofluorescence staining, vibratome sections

eduard.bentea<sup>1</sup>, María Sanchiz Calvo<sup>1</sup>, Veerle Baekelandt<sup>1</sup>

<sup>1</sup>Katholieke Universiteit Leuven



eduard.bentea

#### **ABSTRACT**

Protocol for performing immunofluorescence staining on free-floating vibratome cut brain sections from rats or mice.

#### **MATERIALS**

#### Antigen retrieval solution (10 mM citrate buffer pH 6.0)

Prepare stock solutions:

Solution A: 0.1 M citric acid solution Solution B: 0.1 M sodium citrate solution

Prepare working solution:

Add 9 mL of Stock solution A and 41 mL of Stock solution B to 400 mL of AD Adjust pH to 6.0 Fill until 500 mL with AD

any medium, provided the origin author and source are credited

Oct 16 2024

### protocols.io

Last Modified: Feb 16, 2024

PROTOCOL integer ID: 95341

Keywords: ASAPCRN

Funders Acknowledgement: ASAP (Aligning Science Across Parkinson's)

# Day 1 1 Transfer sections to be stained to a new 24-well plate filled with 1X PBS. All incubations are with Δ 500 μL solution per well unless stated otherwise. 2 1X PBS rinse. 3 Antigen retrieval step (using oven). 3.1 30m Incubate sections in antigen retrieval solution (10 mM citrate buffer pH 6.0; see recipe in Materials) at 80°C for (5) 00:30:00 3.2 20m Place the well plate in the same buffer on ice for 00:20:00

4

1X PBS rinse.

#### protocols.io

Wash sections in 1X PBS for 00:05:00 at room temperature on wobbler.

5m

6 Wash sections in 1X PBS for 00:05:00 at room temperature on wobbler.

5m

- 7 Incubate sections with 250 µL blocking solution (PBS-T + 10% donkey serum) per well for 31:00:00 at room temperature on wobbler. (PBS-T = PBS + 0.1% Tergitol)
- 1h

- 8 Dilute primary antibodies at the required concentration in PBS-T + 10% donkey serum.
- 9 Incubate sections with A 250 µL primary antibodies overnight at room temperature.

## Day 2

211

- 10 1X PBS-T rinse.
- Wash sections in 1X PBS-T for 00:05:00 at room temperature on wobbler.

5m

12 Wash sections in 1X PBS-T for © 00:05:00 at room temperature on wobbler.

5m

- 13 Dilute secondary antibodies at the required concentration in PBS-T.
- Incubate sections with  $\triangle$  250  $\mu$ L secondary antibodies for  $\bigcirc$  02:00:00 at room temperature (in the dark).
- 15 1X PBS rinse.
- Wash sections in 1X PBS for 00:05:00 at room temperature on wobbler.

5m

2h

Wash sections in 1X PBS for 00:05:00 at room temperature on wobbler.

5m

- 18 Briefly rinse sections in 1/2 PBS + 1/2 AD and allow to dry.
- **19** Mount with Mowiol.



Oct 16 2024