



FEB 29, 2024

Western blot for tissue extract

María Sanchiz Calvo¹, eduard.bentea¹, Veerle Baekelandt¹

¹KU Leuven. Neurobiology and Gene therapy lab

ASAP Collaborative Research Network



María Sanchiz Calvo

ABSTRACT

Protocol for the detection of proteins by Western blot from tissue extract

OPEN ACCESS



DOI:

dx.doi.org/10.17504/protocols.io.yxmvm38q9l3p/v1

Protocol Citation: María Sanchiz Calvo, eduard.bentea, Veerle Baekelandt 2024. Western blot for tissue extract.

protocols.io

<https://dx.doi.org/10.17504/protocols.io.yxmvm38q9l3p/v1>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: Feb 29, 2024

Last Modified: Feb 29, 2024

MATERIALS

PROTOCOL integer ID: 95960

Recipe for 100 mL 1x RIPA buffer :

Keywords: ASAPCRN

- o 5 mL Tris pH 7.4
- o 1 mL Triton x100
- o 1 g Deoxycholate
- o 3 mL 5M NaCl
- o 200 µL 0.5M EDTA (pH 8.0)
- o 1 mL 10% SDS

Add water until 100 mL (MilliQ water)

Add protease and phosphatase inhibitors (Pi + PPI) right before use

Reagents used during Western blotting :

- o 4X Laemmli buffer
- o PageRuler™ Plus Prestained Protein Ladder
- o 4–15% Criterion™ Tris-HCl Protein Gel
- o Trans-Blot® Turbo™ PVDF Membrane
- o 5% milk powder dissolved in PBS-T 0.1% (PBS + 0.1% Triton-X100)
- o Goat anti-rabbit HRP secondary antibody (Dako)
- o ECL Prime chemiluminescence kit (GE Healthcare)

Recipe for 10.2 mL 4x Laemmli buffer (0.24M Tris pH 6.8, 7.27% SDS, 40% Glycerol, 10% b-Mercaptoethanol, 0.01% Bromophenol blue):

- 2.4 ml 1M Tris
pH6.8
- 0.8g SDS
- 4ml 100% glycerol
- 2.8ml dH2O
- 1ml b-mercaptoethanol
- 0.01% Bromophenol
blue



BEFORE START INSTRUCTIONS


Perform protein extraction from snap-frozen brain tissue:







- weigh tissue
- add RIPA buffer (see Materials) : 10 X of the weight (40 mg = 400 µL)
- homogenize samples using sample homogenizer
- sonicate samples at 4 degrees C, 3 times 15 seconds (keep the samples on ice between each sonication)
- centrifuge samples at 6000 g for 10 minutes at 4 degrees C
- collect supernatant and measure protein concentration
- aliquot protein extracts and store at -20 degrees




Day 1



- 1 Prepare sample for western blot 10m

 15 µg of protein in  12 µL PBS



Add  4 µL Laemmli buffer


Boil at  98 °C for  00:10:00
- 2 Load samples, together with  7 µL of mass marker (PageRuler™ Plus Prestained Protein Ladder) on a on 4–15% Criterion™ Tris-HCl Protein Gel.
- 3 Run the gel for  00:10:00 at 80V 10m
- 4 Run the gel for  00:45:00 at 150V 45m
- 5 Transfer the proteins on PVDF membrane (Trans-Blot® Turbo™ PVDF Membrane) using Trans-Blot Turbo Transfer System (Bio-Rad), using the pre-programmed protocol STANDARD SD:  00:30:00 , up to 1.0 A, 25 V. 30m

6 Block membranes for  01:00:00 using 5% milk dissolved in PBS-T 0,1% at  Room temperature 

7 Incubate membranes  Overnight in primary antibody in 5% milk PBS-T 0,1% at  4 °C




Day 2



8 Wash membranes with PBS-T 0,1% for 10 minutes, 3 times at T  Room temperature 

Wash PBS-T  00:10:00

Wash PBS-T  00:10:00

Wash PBS-T  00:10:00

9 Incubate membranes for  01:00:00 horseradish peroxidase-conjugated secondary antibody (Dako) diluted 1/10000 in 5% milk PBS-T 0,1% at  Room temperature 

10 Wash membranes with PBS-T 0,1% for 10 minutes, 3 times at T  Room temperature 

Wash PBS-T  00:10:00

Wash PBS-T  00:10:00

Wash PBS-T  00:10:00

11 Develop membranes using ECL Prime (GE Healthcare)