



Sep 13, 2022

# Immunofluorescence on paraffin sections

[miquel.vila](#)<sup>1</sup><sup>1</sup>Vall d'Hebron Research Institute1 *Works for me* Share[dx.doi.org/10.17504/protocols.io.rm7vzbn62vx1/v1](https://dx.doi.org/10.17504/protocols.io.rm7vzbn62vx1/v1) [joan.compte](#)

## ABSTRACT

Immunofluorescence protocol on paraffin-embedded rat brain sections

## DOI

[dx.doi.org/10.17504/protocols.io.rm7vzbn62vx1/v1](https://dx.doi.org/10.17504/protocols.io.rm7vzbn62vx1/v1)

## PROTOCOL CITATION

miquel.vila 2022. Immunofluorescence on paraffin sections. **protocols.io**  
<https://protocols.io/view/immunofluorescence-on-paraffin-sections-cgietube>



## LICENSE

————— This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

## CREATED

Sep 13, 2022

## LAST MODIFIED

Sep 13, 2022

## PROTOCOL INTEGER ID

69926

## MATERIALS TEXT

### Reagents :

- TBS 10X : Tris base 121.1g + NaCl 90g in 1L H<sub>2</sub>O.pH 7.4.
- TBS 1X-Triton 0,5%
- Xilen
- Ethanol : 100%, 95%, 70%
- Unmasking buffer epitopes : Citrate solution 10mM pH6.0
- Blocking Buffer : TBS 1X + 5% NGS
- 1st Ab : Diluted in1X PBS +2%NGS
- 2nd Ab : Diluted in1X PBS +2%NGS
- Endogenous peroxidase blocking solution : TBS 1x + 3% H<sub>2</sub>O<sub>2</sub>(30%) + 10% methanol

### 1. Deparaffinization and hydratation :

- 1 Put the slides 30 min in the incubator at 60°C
- 2 Wash 3x3 min in xylene
- 3 Wash 1x10 min in ethanol 100%
- 4 Wash 1x10 min in ethanol 95%
- 5 Wash 1x5 min in ethanol 70%
- 6 Wash 2x5 min in PBS 1X

**IMPORTANT:** The buffer used for immunofluorescence is PBS, not TBS as for immunohistochemistry and the endogenous peroxidase blocking step is not required.

### Antigen retrieval

- 7 Add 200 mL/section of citrate buffer 10mM, pH 6

7.1 Put sections in a boiling water bath for 20 min

8 Let sections cool down for 20 min at room temperature (RT)

#### Washing

9 Wash 3x5 min in PBS 1X-Triton (0.5%)

#### Blocking

10 Put wet paper in a black box for immunohistochemistry

11 Gently dry and circle sections with the hydrophobic pen ImmEdge Vector H 4000 without touching the tissue

12 Add 200 uL/section of blocking solution (5% NGS (in PBS 1X) (200uL/section) + 0.1% Triton (in PBS 1X)) 1h at RT

#### 1ary antibody

13 Gently wipe off the water of the slides

14 Put 200ul/section of 1ary antibody in 2% NGS + 0.1% Triton overnight at 4°C (cold room)

#### Washing

15 Wash 3x5min in PBS 1X-Triton (0.5%)

## 2ary antibody

16 Gently wipe off the water of the slides

17 Put 200ul/section of 2ary antibody in 2% NGS + 0.1% Triton + DAPI for 1 h at RT

Alexa goat anti-mouse 488 (green color) 1:500 or 1: 1000, Alexa goat anti-rabbit 594 (red color) and 1:5000 DAPI (nuclei staining in blue color)

## Washing

18 Wash 3x5min in PBS 1X

## Mounting

19 Mount sections with fluorescent mounting medium. Put the coverslip (washed with ethanol previously) on the slide. Remove bubbles

## Storage

20 Let dry the slides on the tray in the hood for 5 minutes and store at 4°C **asap**