

Jun 07, 2024



## Mouse Habituation - Head Fixation on Disk

DOI

#### dx.doi.org/10.17504/protocols.io.j8nlkojmxv5r/v1

Avalon Amaya<sup>1</sup>, Jackie Swapp<sup>1</sup>

<sup>1</sup>Allen Institute for Neural Dynamics

Allen Institute for Neural D...



# Avalon Amaya

Allen Institute

# OPEN ACCESS



DOI: dx.doi.org/10.17504/protocols.io.j8nlkojmxv5r/v1

Protocol Citation: Avalon Amaya, Jackie Swapp 2024. Mouse Habituation - Head Fixation on Disk. protocols.io https://dx.doi.org/10.17504/protocols.io.j8nlkojmxv5r/v1

License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working We use this protocol and it's working

Created: January 25, 2024

Last Modified: June 07, 2024

Protocol Integer ID: 94173

Keywords: Rodent Head Fixation, Rodent Behavioral Habituation



### Abstract

This protocol describes a customized mouse habituation protocol for mice within experiments involving head fixation onto disks.

Mouse habituation is a common technique utilized for mice undergoing behavioral experimentation and/or head fixation. Habituation allows the mouse to acclimate to new environments, handlers, and handling techniques. This exposure can lower the mouses stress and can decrease the likelihood of mouse bites and/or jumps.

**Habituation timeline summary:** Mice are habituated for a total of 5 days, involving daily handling and increasing durations of head fixation in the behavior enclosure. Mice can be either water restricted or on Adlib water during habituation. In days 1 and 2 mice are habituated to handling, and on days 3-5 head fixing is introduced.

Utilize proper PPE when handling mice. Prioritize handling the mouse over the cage to minimize risk of mouse jumping/falling. Keep mouse cage covered when not attending to mouse. Follow IACUC and veterinary requirements for handling mice.

Mice under water restriction should be closely monitored for health concerns due to the effects of dehydration. All mice should receive 1.0mL of water daily. Follow IACUC and veterinary requirements for water restricting mice.

#### Guidelines

Only perform this procedure in accordance with IACUC and veterinary requirements.

#### Animal Behavior:

Utilize a 12-hour light cycle.

Mice should have access to ad-lib food and water until placed on diet/liquid restriction.

Single house mice on diet or water restriction.

Ensure cleanliness of experimental environment.



### Materials

### Materials:

Tool / Supply	Manufacturer / Supplier	Part Number
70% Ethanol (Diluted in house)	Sigma Aldrich	459836
Kimwipes	Kimberly - Clark Professional	S47299A
Liquid Reward	-	-
Nalgene® 125 ml PET Square Media Bottles, Sterile CP Lab Safety		342040-0125
1mL Luer-Lok Syringe	Becton Dickinson and Company	309628
25G 5/8 Needle *	Advantor	89134-134
1000mL Nalgene™ Straight-Sided Wide-Mouth Polycarbonate Jars	Fisher Scientific	21161000
Ohaus® Scout® Portable Balance	Sigma Aldrich	OH30253024
Mouse Igloo® and Fast Tracs, Certified	Bio-Serv	K3327
HEPA Replacement Filter for DC515	Dewalt	DC5151H
18V Cordless 1/2 Gallon Wet/Dry Portable Vac (Tool Only)	Dewalt	DC515B
Swiifer Sweeper	Uline	H-1960
Swiffer Sweeper Pads - Wet Cloths	Uline	S-13901
Water Bowl	Made in-house	0146-000-01
Cart Cover	Life Science Products	CC-34X19X38Z
Wheel Foam (MC® Solid Grip Black 18 in. x 4 ft.)	Kittrich	04F-187110-06
Precision Screwdriver Hex 1/16th	WIHA	084705263137

# Supplies can be substituted with their equivalent.

# **Equipment:**

Material	Part Number
Behavior Stage 1 (Behavior Platform, Behavior Training, Visual Behavior, Hotshoe Version 2) AND	0107-032-00
Behavior Stage 2 (Passive Behavior Shroud Assembly )	0312-300-00

# All equipment can be substituted with their equivalent.

# Key:

<sup>\* =</sup> Optional



AND = Including the tool/supply in row below.

### **Personal Protective Equipment (PPE):**

Suggested PPE		
Gloves		
Disposable lab coat		
Disposable face mask		
Scrubs		
Biohazard sharps disposal container		
Biohazard waste disposal container		

Utilize PPE in accordance with IACUC and veterinary requirements.

# Safety warnings



Personal Protective Equipment (PPE) should be used at all times while operating this protocol.

### Ethics statement

Research focused rodent behavior must be conducted according to internationally-accepted standards and should always have prior approval from an Institutional Animal Care and Use Committee (IACUC) or equivalent ethics committee(s).

This protocol has been approved by the Allen Institute Animal Care and Use Committee (IACUC).

PHS Assurance: D16-00781

AAALAC: Unit 1854

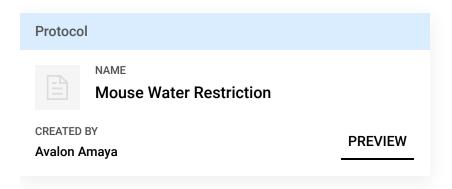


### Before start

Reference protocol below for specific start-up and take down procedures for behavioral training hardware and software:

Amaya, A. M., Swapp, J., Groblewski, P. A. (ND). Using behavioral training clusters. protocols.io. Unpublished. https://www.protocols.io/view/using-behavioral-training-clusters-c8vbzw2n

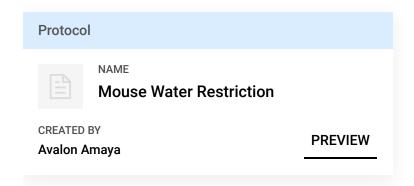
Reference water restriction protocol below for starting and maintaining a mouse on water restriction:





# Maintaining Water Restriction

Follow protocol below for maintaining a mouse on water restriction:



## Habituating a Mouse on a Disk

2 Day 1:

Handle mouse for 2-5 min.

2.1 Scoop/cup mouse out of cage. Cup/cover mouse with hands for around 30 seconds to 1 minute.

#### Note

This act of cuping/covering the mouse acts to "calm" the mouse down and to prevent mouse from immediately jumping out of hands.



Hands cupping mouse over cage.



- 2.2 Uncup/cover hands, and let mouse explore the environment of your open hand for another 1-4 minutes.
- 2.3 Weigh mouse and return back to cage.
- 2.4 Maintain mouse on water restriction if applicable (See above section "Maintaining Water Restriction").



3 Day 2:

Handle mouse for 2-5 min.

- 3.1 Repeat Day 1 handling steps go to step #2.3 - #2.4 .
- 3.2 Periodically touch the mouses headframe to habituate mouse to future head fixation.
- 3.3 Weigh and water mouse by following steps <u>■5</u> go to step #2.3 - #2.4 .
- 4 Day 3:

Handle mouse for 1-2 min, secure mouse to head fixation stage, and place mouse within designated behavioral training box for 5 minutes.

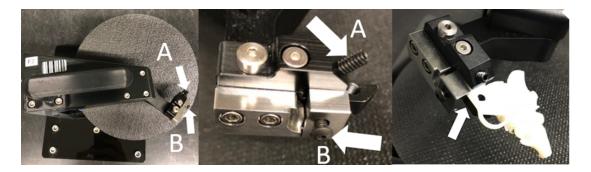
- 4.1 Cup or grab mouse out of cage by the tail.
- 4.2 Let mouse explore the environment of your hands for 1 minute. Periodically touch the mouses headframe to habituate mouse to head fixation.



Hands holding mouse head frame and tail over cage.

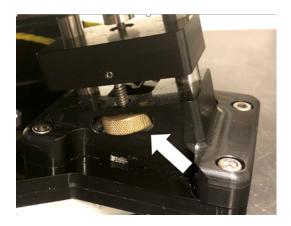


4.3 While grabbing the mouses headframe in one hand and the mouse's tail in the other. Secure mouse to disk head fixation stage.



Stage, screws, and model of attached headframe. A & B labels for securing points of contact in operational order.

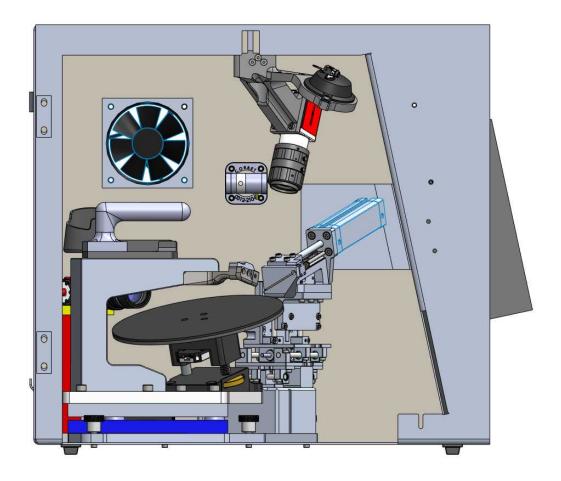
4.4 Adjust disk height to allow mice to run/walk at a natural and comfortable gait.



Knob for disk height adjustment

4.5 Place mouse secured to head fixation stage within designated behavioral training box for 5 minutes.





Computer model design of inside of behavior box with behavior stage.

- 4.6 Remove mouse from box and head fixation stage.
- 4.7 Weigh and water mouse by following steps go to step #2.3 - #2.4 .
- 5 Day 4:

Handle mouse for 1-2 min, secure mouse to head fixation stage, and place mouse within designated behavioral training box for 10 minutes.

- 5.1 Repeat Day 3 handling and head fixing steps go to step #4.1 - #4.4 .
- 5.2 Place mouse secured to head fixation stage within designated behavioral training box for 10 minutes.



- 5.3 Remove mouse from box and head fixation stage.
- Weigh and water mouse by following steps go to step #2.3 #2.4.

\*

6 Day 5 (Passive Behavior):

Handle mouse for 1-2 min, secure mouse to head fixation stage, and place mouse within designated behavioral training box for 10 minutes.

- 6.1 Repeat Day 4 steps <u>so to step #5</u> #5.4.
- 7 Day 5 (Active Behavior):

Handle mouse for 1-2 min, secure mouse to head fixation stage, place mouse within designated behavioral training box, and run 15 minute training session.

- 7.1 Open MouseDirector and load your username and the mouse ID onto its designated behavioral training box.
- 7.2 Repeat Day 3 steps go to step #4.1 #4.4
- 7.3 Use lickspout controller to find Mouse Offset.

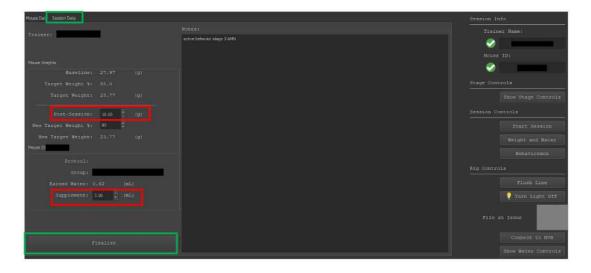
Mentioned in protocol *Using Behavioral Training Clusters*, section *Aligning Lickspout* 

Amaya, A. M., Swapp, J., Groblewski, P. A. (ND). Using behavioral training clusters. protocols.io. Unpublished. <a href="https://www.protocols.io/view/using-behavioral-training-clusters-c8vbzw2n">https://www.protocols.io/view/using-behavioral-training-clusters-c8vbzw2n</a>

- 7.4 Confirm script regimen and the mouse ID.
- 7.5 Press "start session" to begin auto train session.
- 7.6 Confirm that script has begun via camera view (typically static grating images).



- 7.7 Monitor mouse behavior. If not licking or if tongue is not able to make contact with lickspout, move lickspout closer to mouth.
- 7.8 Once session is completed, press the "safe" button on Mouse Director to retract the lickspout.
- 7.9 Remove mouse from box and head fixation stage.
- 7.10 Weigh mouse.
- 7.11 Return mouse to cage.
- 7.12 Click "Session Data" tab on Mouse Director.
- 7.13 Enter mouse weight in "Post-Session: " text box.



Session Data tab in Mouse Director.

Post-Session weight and Supplemental water calculation boxes outlined in red.

Session Data tab and Finalize button outlined in green.



#### Note

Mouse baseline weight and restriction percentage should already be associated with the mouse ID if water restricted through Waterlog. See water restriction protocol in above section.

7.14 Provide mouse with amount of supplemental water as written in "Supplemental:" box.

### Note

If the supplemental water text box displays 0.00mL, ensure mouse has received at least 1.0mL of water for the session.

7.15 Press the "Finalize" to save session data.

### Clean

- 8 Clean and disinfect behavior room.
- 8.1 Use 70% Ethanol spray and Kimwipes to clean weigh boats and surface areas.

#### Note

Make sure to remove all fecal matter from behavior boxes.

- 8.2 Wipe lickspouts with alcohol wipes.
- 8.3 Sweep floors.
- 8.4 Mop floors once a week at the end of the day once all mice have left the room.



Note

Make sure there are no animals present in room. Mop when room has sufficient time to air

8.5 Vacuum behavior boxes once a month at the end of the day once all mice have left the room.

Note

Use HVAC certified vacuum. Make sure no animals are present in room.

# Habituating a Mouse Quick Guide

9

#### Active Habituation

	М	1	W	Th	E
Week 1			Weigh	Weigh	Weigh Set baseline weight Water restrict
Week 2	Handle 2-5 minutes	Handle 2-5 minutes	Handle 1-2 minutes, headfix, place stage in box for 5 minutes	Handle 1-2 minutes, headfix, place stage in box for 10 minutes	Handle 1-2 minutes, headfix, 15 minute autoreward script
Week 3	Training day 1	Training day 2	Training day 3	Training day 4	Training day 5

Passive Habituation

	М	Т	W	Th	F
Week 1	Handle 2-5 minutes	Handle 2-5 minutes	Handle 1-2 minutes, headfix, place stage in box for 5 minutes	Handle 1-2 minutes, headfix, place stage in box for 10 minutes	Handle 1-2 minutes, headfix, place stage in box for 10 minutes
Week 3	Training day 1	Training day 2	Training day 3	Training day 4	Training day 5

#### **Quick Guide:**

Typical first 2-3 week schedule of an experimental behavior mouse.

The intended use for quick guide is to reference once familiarized with habituating a mouse.



### Protocol references

Groblewski, P. A., Ollerenshaw, D. R., Kiggins, J. T., Garrett, E., Mochizuki C., Casal, L., Cross, S., Mace, K., Swapp J., Manavi, S., Williams, D., Mihalas, S., Olsen, S. R. (2020). Characterization of learning, motivation, and visual perception in five transgenic mouse lines expressing GCaMP in distinct cell populations. Frontiers in Behavioral Neuroscience. https://doi.org/10.3389/fnbeh.2020.00104

Schoorlemmer, G. H., Evered, M. D. (2002). Reduced feeding during water deprivation depends on hydration of the gut. Am J Physiol Regul Integr Comp Physiol. Nov;283(5):R1061-9. doi: 10.1152/ajpregu.00236.2002. PMID: 12376399.

Rowland, N., E. (2007). Food or fluid restriction in common laboratory animals: balancing welfare considerations with scientific inquiry. Comparative Medicine 57(2). American Association for Laboratory Animal Science. https://www.ingentaconnect.com/content/aalas/cm/2007/00000057/00000002/art00001

Guo, Z. V., Hires, S. A., Li, N., O'Connor, D. H., Komiyama, T., Ophir, E., et al. (2014). Procedures for behavioral experiments in head-fixed mice. PLoS ONE 9(2): e88678. https://doi.org/10.1371/journal.pone.0088678

Barkus, C., Bergmann, C., Branco, T., Carandini, M., Chadderton, P. T., Galiñanes, G. L., Gilmour, G., Huber, H., Huxter, J. R., et. al. Refinements to rodent head fixation and fluid/food control for neuroscience. Journal of Neuroscience Methods. (2022). https://doi.org/10.1016/j.jneumeth.2022.109705.

Gouveia K, Hurst J.L. Reducing Mouse Anxiety during Handling: Effect of Experience with Handling Tunnels. PLoS ONE 8(6): e66401 (2013). doi:10.1371/journal.pone.0066401

Gouveia, K., Hurst, J. Optimising reliability of mouse performance in behavioural testing: the major role of non-aversive handling. Sci Rep 7, 44999 (2017). https://doi.org/10.1038/srep44999

Gouveia, K., Hurst, J.L. Improving the practicality of using non-aversive handling methods to reduce background stress and anxiety in laboratory mice. Sci Rep 9, 20305 (2019). https://doi.org/10.1038/s41598-019-56860-7

Henderson, L.J., Dani, B., Serrano, E.M.N. et al. Benefits of tunnel handling persist after repeated restraint, injection and anaesthesia. Sci Rep 10, 14562 (2020). https://doi.org/10.1038/s41598-020-71476-y

Juczewski, K., Koussa, J.A., Kesner, A.J. et al. Stress and behavioral correlates in the head-fixed method: stress measurements, habituation dynamics, locomotion, and motor-skill learning in mice. Sci Rep 10, 12245 (2020). https://doi.org/10.1038/s41598-020-69132-6