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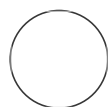
**Protocol status:** Working  
 We use this protocol and it's working

## 🌐 Immunohistochemistry Protocol for Free-floating Fixed Tissue with Tyramine Signal Amplification (TSA)

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### ABSTRACT

Immunohistochemistry protocol for staining free-floating fixed tissue with Tyramine signal amplification (TSA) in the Kordower Laboratory. TSA allows a much lower concentration of primary antibody to be used (typically 1:10K or 1:15K) at the expense of an extra day of staining.

### ATTACHMENTS

[nv3bbiaqf.docx](#)

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## GUIDELINES

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**PROTOCOL integer ID:**  
89625

**Keywords:**  
Immunohistochemistry

### HISTO- NOTES:

- Primate tissue staining dishes use  100 mL solution per dish
- Rodent tissue staining dishes  50 mL solution per dish
- If staining a large number of primate cases, incubate 1' & 2' Ab in individual cups to conserve volume of Ab used.
- Prepare bleach neutralizing solution prior to Step 12.
- Be conscious of tissue saturation while washing and incubating. i.e. Check that tissue is fully submerged in solution & not clumping. This will ensure proper penetration of antibodies & other reagents.
- Always include Positive & Negative Controls.
- Positive: Use relevant control tissue to confirm specific antibody detection. (i.e. pS129; control tissue should consist of nigral sections previously successfully stained for pS129).
- Negative: Ideally, use tissue that you know does not contain the targeted antigen. If not available, use a section of tissue not incubated in the 1' Ab (primary delete).
- When incubating 1' Ab overnight, leave on shaker in refrigerator.
- Can incubate in fridge on a shaker, covered in parafilm, over the weekend or up to 3 days.
- Select a secondary antibody directed against the species in which the primary antibody was raised (i.e. if a primary antibody raised in rabbit is used, an anti-rabbit secondary antibody raised in a species other than rabbit must be used).


## MATERIALS


- Dilution Media (DM) ([M] 0.2 Molarity (M) TBS plus [M] 0.05 % volume Triton X-100)
- [M] 0.2 Molarity (m) Tris-buffered saline (TBS)
- Sodium meta-periodate
- Normal Serum (species matching the host of the secondary antibody, e.g. horse, goat)
- Bovine Serum Albumin (BSA)
- Triton X-100
- Vectastain Elite ABC-HRP Kit (PK-6100)
- Imidazole
- Sodium Acetate
- Sodium tetraborate decahydrate
- Boric Acid
- Biotin tyramide
- 3,3-Diaminobenzidine Tetrahydrochloride (DAB)
- Nickel(II) sulfate hexahydrate
- [M] 30 % (v/v) hydrogen peroxide
- [M] 0.2 Molarity (m) Phosphate-buffered saline (PBS)
- Household Bleach
- Primary antibody against the target antigen
- Secondary antibody directed against the species in which the primary antibody was raised (i.e. if a primary antibody raised in rabbit is used, an anti-rabbit secondary antibody raised in a species other than rabbit must be used).

### DAY 1 (3.5hrs)

1 Wash sections (6 x  00:05:00) in Dilution Media (DM) ([M] 0.2 Molarity (M) TBS plus [M] 0.05 % volume Triton X-100). 5m



1.1 Wash sections for  00:05:00 in DM (1/6). 5m

1.2 Wash sections for  00:05:00 in DM (2/6). 5m

1.3 Wash sections for  00:05:00 in DM (3/6).

5m

1.4 Wash sections for  00:05:00 in DM (4/6).

5m

1.5 Wash sections for  00:05:00 in DM (5/6).




5m

1.6 Wash sections for  00:05:00 in DM (6/6).

5m

2 Endogenous peroxidase inhibition (  00:20:00 ).  0.1 Molarity (M) Sodium meta-periodate in TBS.

20m

-  100 mL  0.2 Molarity (m) Tris-buffered saline (TBS)
-  2.13 g sodium meta-periodate

3 Wash (2 x  00:00:00 ) in DM.



3.1 Wash for  00:10:00 in DM (1/2).

10m

3.2 Wash for  00:10:00 in DM (2/2).

10m

#### 4 Serum blocking step (🕒 01:00:00 incubation):

1h



- 🧪 100 mL DM
- 🧪 3 mL Normal Serum (species matching the host of the secondary antibody, e.g. horse, goat)
- 🧪 2 g Bovine Serum Albumin (BSA)

#### 5 Incubation in primary antibody (🕒 18:00:00 - 🕒 72:00:00 ). See antibody catalog for concentration of primary antibody (typically 1:10K or 1:15K with TSA).

3d 18h



- 🧪 100 mL DM
- 🧪 1 mL Normal Serum (species matching the host of the secondary antibody, e.g. horse, goat)
- 🧪 1 g BSA
- 🧪 0.5 mL Triton X-100

##### Note

**\*\*Optionally, refrigerate 🌡️ 4 °C to keep antibody stable\*\***

### DAY 2 (4hrs):

#### 6 Wash (6 x 🕒 00:05:00 ) in DM.

5m



##### 6.1 Wash for 🕒 00:05:00 in DM (1/6).

5m

##### 6.2 Wash for 🕒 00:05:00 in DM (2/6).

5m

##### 6.3 Wash for 🕒 00:05:00 in DM (3/6).

5m

6.4 Wash for  00:05:00 in DM (4/6).

5m

6.5 Wash for  00:05:00 in DM (5/6).




5m

6.6 Wash for  00:05:00 in DM (6/6).

5m

7 Incubation in secondary antibody ( 01:00:00 ). Concentration of secondary antibody is always 1:500 in solvent.

1h

-  100 mL DM
-  1 mL Normal Serum (species matching the host of the secondary antibody, e.g. horse, goat)
-  1 g BSA

8 Wash (6 x  00:05:00 ) in DM.

5m



#### Note

**\*\*(incubate ABC in solvent during these washes)\*\***

8.1 Wash for  00:05:00 in DM (1/6).

5m

8.2 Wash for  00:05:00 in DM (2/6).

5m


8.3 Wash for  00:05:00 in DM (3/6).

5m

8.4 Wash for  00:05:00 in DM (4/6). 5m




8.5 Wash for  00:05:00 in DM (5/6). 5m

8.6 Wash for  00:05:00 in DM (6/6). 5m

9 Actin Biotin Complex Step ( 01:00:00 ) - Vectastain Elite ABC-HRP Kit (PK-6100). 1h

#### Note

**\*\*Re-use for step 15. Can be stored overnight in refrigerator  4 °C \*\***

-  100 mL DM
-  1 mL Normal Serum (species matching the host of the secondary antibody, e.g. horse, goat)
-  1 g BSA

9.1 Add ABC Reagent A and B to 1/10th of total desired volume of solvent.



9.2 Incubate for  00:30:00 at  Room temperature . 30m



#### Note

Then dilute 1:10 using the same solvent. This is your working solution. See chart below for example volumes.

A	B	C	D
Working Solution	A (drops)	B (drops)	1/10th Working solution
25 mL	1	1	2.5mL
50 mL	2	2	5mL
100mL	4	4	10mL

10 Wash for 00:10:00 in DM.

10m



11 Wash 00:10:00 in TBS.

10m



12 Wash (2 x 00:10:00 ) in 0.05 Molarity (M) Borate buffer pH 8.5 .

10m



- 1000 mL dH<sub>2</sub>O
- 4.77 g Sodium tetraborate decahydrate
- 2.32 g Boric Acid

12.1 Wash for 00:10:00 in Borate buffer (1/2).

10m

12.2 Wash for 00:10:00 in Borate buffer (2/2).

10m

13 Incubate sections with biotin tyramide solution ( 00:30:00 ).

30m



#### Note

**DO NOT USE IF >6 MONTHS OLD.**

- 100 mL Borate buffer



- 2  $\mu\text{L}$  of 50 mg/mL biotin tyramide stock
- 10  $\mu\text{L}$  30 % (v/v) Hydrogen Peroxide ( $\text{H}_2\text{O}_2$ )

14 Wash (3 x 00:10:00) in TBS (Antigen is now labeled with biotin).

10m



14.1 Wash for 00:10:00 in TBS (1/3).

10m

14.2 Wash for 00:10:00 in TBS (2/3).

10m

14.3 Wash for 00:10:00 in TBS (3/3).

10m

## DAY 3 (4 hrs):

40m

15 Repeat Actin Biotin Complex Step 9 then wash for 00:10:00 with TBS.

10m



16 Wash (3 x 00:10:00) in 0.2 Molarity (M) Imidazole/ 1.0 Molarity (M) Sodium Acetate buffer, pH 7.2 to pH 7.4.


10m




- 1000 mL  $\text{dH}_2\text{O}$
- 0.68 g Imidazole
- 6.8 g Sodium Acetate
- Retain 100 mL of non-pH'd buffer for DAB preparation

16.1 Wash for 00:10:00 in Imidazole/Sodium Acetate buffer (1/3).

10m

16.2 Wash for  00:10:00 in Imidazole/Sodium Acetate buffer (2/3).




10m

16.3 Wash for  00:10:00 in Imidazole/Sodium Acetate buffer (3/3).

10m

17 DAB step (Neutralize DAB with bleach when done)

17.1 Make DAB solution




-  100 mL non-pH'd imidazole acetate buffer from above
-  50 mg 3,3-Diaminobenzidine Tetrahydrochloride (DAB)
-  2 g Nickel(II) sulfate hexahydrate **\*\***(Only used with certain primary antibodies, chromagen enhancer that changes brown DAB precipitate to blue-purple)**\*\***

17.2 Make  1 % (v/v) Hydrogen Peroxide (H<sub>2</sub>O<sub>2</sub>)

-  3 mL of dH<sub>2</sub>O
-  100 µL of  30 % (v/v) hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>)


17.3 Start reaction -- add  500 µL of  1 % (v/v) hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) to the above DAB mixture just prior to use.



OR add  16.7 µL of  30 % (v/v) hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), per  100 mL .

17.4 Place tissue in DAB solution.

5m

- Develop tissue for approximately  00:05:00 .
- Timing is critical, ensure all tissue spends the same amount of time in DAB solution.


17.5 To monitor signal, move all tissue to imidazole buffer, remove one section and mount on an UNSUBBED slide and view under microscope. Place all tissue back in DAB solution to





increase signal intensity, if needed.

18

Wash developed tissue in imidazole acetate buffer (3 x  00:10:00 ).


10m



Note


**\*\*Neutralize DAB with BLEACH!!!\*\***

18.1

Wash developed tissue in imidazole acetate buffer for  00:10:00 (1/3).


10m

18.2

Wash developed tissue in imidazole acetate buffer for  00:10:00 (2/3).



10m

18.3

Wash developed tissue in imidazole acetate buffer for  00:10:00 (3/3).

10m

19

Store tissue in  0.2 Molarity (m) Phosphate-Buffered Saline (PBS) in refrigerator  4 °C until mounted on slides.