

VERSION 2

OCT 24, 2023

OPEN ACCESS



DOI:

dx.doi.org/10.17504/protocols.io.6qpvr385ovmk/v2

Protocol Citation: Verena Dederer, Deep Chatterjee, Sebastian Mathea, Stefan Knapp 2023. In vitro LRRK2 kinase activity assay using mass-spectrometry as readout. **protocols.io** <https://dx.doi.org/10.17504/protocols.io.6qpvr385ovmk/v2> Version created by Verena Dederer

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

🌐 In vitro LRRK2 kinase activity assay using mass-spectrometry as readout V.2

Verena

Dederer^{1,2},

Stefan

Knapp^{1,2}

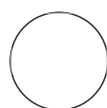
Deep

Chatterjee^{1,2},

Sebastian Mathea^{1,2},

¹Institute of Pharmaceutical Chemistry, Goethe University Frankfurt, Max-von-Laue-Straße 9, Frankfurt 60438, Germany;

²Structural Genomics Consortium, Buchman Institute for Molecular Life Science (BMLS), Max-von-Laue-Straße 15, Frankfurt 60438, Germany



Verena Dederer

ABSTRACT

This protocol can be used in this form or with small adjustments regarding concentration and reaction time to determine in vitro substrate phosphorylation for any purified kinase substrate pair.

It can be used to determine inhibition curves by addition of concentration series of kinase inhibitors.

Reaction is carried out in 50 μ L reaction mix containing final concentration of 50 nM kinase and 5 μ M substrate.

Protocol status: Working
We use this protocol and it's working

Created: Sep 07, 2023

Last Modified: Oct 24, 2023

PROTOCOL integer ID:
87485

Funders
Acknowledgement:
Aligning Science Across
Parkinson's
Grant ID: ASAP-000519

MATERIALS

Purified proteins

kinase: LRRK2^{RCKW}

protein sequence:

KKAVPYNRMKLMIVGNTGSGKTTLLQQLMKTKKSDLGMQSATVGIDVKDWPIQIRDKRK
RDLVLNVWDFAGREEFYSTHPHFM TQRALYLAVYDLSKGQAEVDAMKPWLFNIKARASS
SPVILVGTHLDVSDEKQRKACMSKITKELLNKRGFPAIRDYHFVNATEESDALAKLRKTIIN
ESLNFKIRDQLVVGQLIPDCYVELEKIILSERKNVPIEFVIDRKRLQLVRENQLQLDENELP
HAVHFLNESGVLLHFQDPALQLSDLYFVEPKWLCKIMAQILTVKVEGCPKHPKGIISRRDV
EKFLSKKRKFPKNYMSQYFKLLEKFQIALPIGEEYLLVPSSLS DHRPVIELPHCENSEIIIRLY
EMPYFPMGFWSRLINRLLLEISPYMLSGRERLRPNRM YWRQGIYLNWSPEAYCLVGSEVL
DNHPESFLKITVPSCRKGCILLGQVVDHIDSLMEEWFPGLLEIDICGEGETLLKKWALYSFN
DGEEHQKILLDDL MKKAEEGDLLVNPDPRLTIPISQIAPDLILADLP RNIMLNNDLEFEQ
APEFLLGDGSFGSVYRAAYEGEEVAVKIFNKHTSLRLLRQELVVLCHLHHP SLISLLAAGIR
PRMLVMELASKGSLDRLLQQDKASLTRLQHRIALHVADGLRYLHSAMIIYRDLKPHNVLL
FTLYPNAIIAKIADYGIAQYCCRMGIKTSEGT PGFRAPEVARGNVIYNQQADV SFGLLLY
DILTTGGRIVEGLKFPNEFDELEIQGKLDPVKEYGCAPWPMVEKLIKQCLKENPQERPTSA
QVFDILNSAELVCLTRILLPKNVIVECMVATHHNSRNASIWLGC GHTDRGQLSFLDLNTE
GYTSEEVADSRILCLALVHLPVEKESWIVSGTQSGTLLVINTEDGKKRHTLEKMTDSVTCL
YCNSFSKQSKQKNFLLVGTADGKLAIFEDKTVKLKGAAPL KILNIGNVSTPLMCLSESTNS
TERNVMWGGCGTKIFSFSNDFTIQKLIETRTSQLFSYAAFSDSNIITVVVD TALYIAKQNSP
VVEVWDKKTEKLCGLDCVHFLREVMVKENKESKHKMSYSGRVKTLCLQKNTALWIGTG
GGHILLDLSTRRLIRVIYNFCNSVRVMMTAQLGSLKNVMLVLGYNRKNT EG TQKQKEIQS
CLTVWDINLPHEVQNLEKHIEVRKELAEKMRRTSVE

substrate: Rab8A

protein sequence:

GHMDYLFKLLLLIGDSGVGKTCVLF RFSEDAFNSTFISTIGIDFKIRTIELDGKRIKLQIWD TAG
QERFRTITTAYYRGAMGIMLVYDITNEKSFDNIRNWIRNIEEHASADVEKMILGNKCDVND
KRQVSKERGEKLALDYGIKFMETSAKANINVENAFFTLARDIKAKMDKK

Buffers and Reagents

reaction buffer: 20 mM Hepes pH 7.4, 150 mM NaCl, 5% glycerol, 0.5 mM TCEP, 20
μM GDP, 2.5 mM MgCl₂

ATP 100 mM stock in dH₂O

mass spec buffer: dH₂O + 0.1% formic acid

Step-by-Step Protocol

- 1 Prepare 50 μL reaction mix I: 50 nM purified LRRK2^{RCKW} and 5 μM Rab8 substrate in buffer containing 20 mM Hepes pH 7.4, 150 mM NaCl, 5% glycerol, 0.5 mM TCEP, 20 μM GDP, 2.5 mM MgCl_2 .
Aliquot 25 μL per tube.

Note

If you want to run multiple reactions prepare a master mix and aliquot 25 μL per tube.

- 2 Prepare 25 μL reaction mix II: 2 mM ATP + 2.5 mM MgCl_2 in buffer containing 20 mM Hepes pH 7.4, 150 mM NaCl, 5% glycerol, 0.5 mM TCEP, 20 μM GDP, 2.5 mM MgCl_2 .

Note

Multiply if you want to run more than one reaction.

- 3 Prepare 25 μL reaction mix III: same as reaction mix II but no ATP as negative control.

- 4 Start reaction by adding 25 μL reaction II to reaction mix I.
Note: for negative control add 25 μL reaction mix III to second aliquot of reaction mix I.

- 5 Mix by vortexing and brief centrifugation for 30 sec with 500xg.

- 6 Incubate reaction for 3 h at room temperature.
Note: depending on kinase this may be shorter or longer.

- 7 Stopp reaction by adding 50 µL mass spec buffer (dH₂O+0.1% formic acid).
- 8 Store at -80°C or proceed directly with mass spectrometry analysis.

Note

For determination of inhibition curves, prepare a concentration series of inhibitor in DMSO. Prepare a master mix of kinase and substrate pair and aliquot 25 µL per tube. Add the desired amount of inhibitor to each of the tube and incubate all reactions at room temperature.

Note

For analysis: substrate phosphorylation/turnover can be determined as ratio between phosphorylated and unphosphorylated peak intensity for each sample analyzed.