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Axenic Diatoms cultures protocol^{v.4}

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Works for me

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ABSTRACT

Axenic cultures protocol

Axenicity of cultures is obtained by multi-antibiotic treatment: by adding antibiotics directly to the medium in which the strain will be inoculated. After a variable amount of days we refresh the cultures with new medium.

antibiotics concentration

- 1 The antibiotics used are ampicillin, streptomycin and penicillin;
We used the following final concentration:
 - Ampicillin 0,5 mg/ml
 - Streptomycin 0,05 mg/ml
 - Penicillin 0,03 mg/ml
- 2 Add antibiotics in F/2 medium.
Start from 20/50.000 cells in 25 ml of F/2 medium with antibiotics.
Refresh 3 times the cultures every 3 days with new medium with antibiotics.
- 3 Transfer the cultures into a variable higher volume flask containing F/2 medium and antibiotics.
Arrive at the desired final volume.

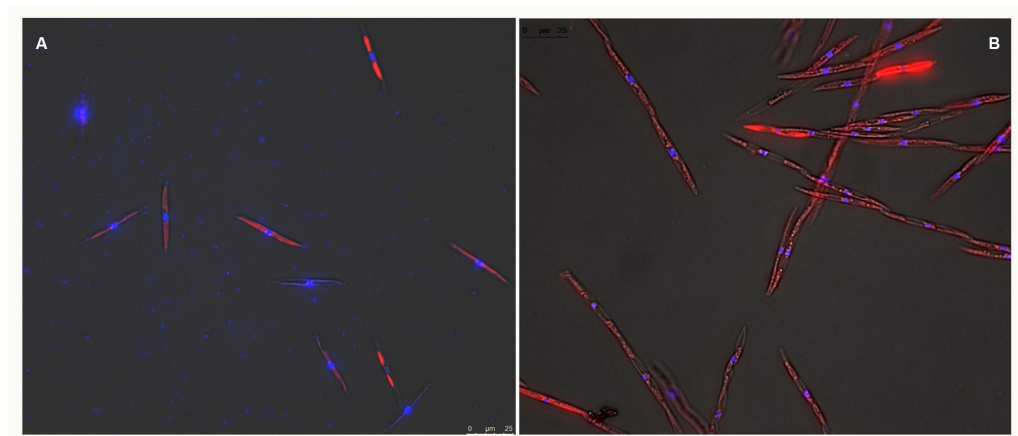
contamination test

- 4 Assess axenicity of the strains by fluorescence microscopy using DAPI staining (4',6-diamidino-2-phenylindole-a DNA stain).

(DAPI stains DNA, if bacteria are present in the culture bacterial nucleoids can be visualised as fluorescent spots).

To fix cells use neutralized formaldehyde 1.6%

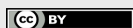
DAPI final concentration 1:1000



Pseudo-nitzschia multistriata fluorescence microscopy images using DAPI staining.

A. Normal culture with bacteria

B. Axenic culture without bacteria.



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