

**VERSION 8** SEP 10, 2023

ONA extraction (BOMB) V.8

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**ABSTRACT** 

DNA extraction (BOMB)





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**Protocol status: Working** We use this protocol and it's working

**Created:** Sep 09, 2023

**MATERIALS** 

Last Modified: Sep 10,

2023

1. Lysis master mix (870 uL/sample)

**PROTOCOL** integer ID:

87598

| A                | В      |
|------------------|--------|
| TE buffer        | 225 uL |
| Lysis buffer     | 375 uL |
| Ammonium acetate | 270 uL |

### 2. TE buffer

| A                 | В    |
|-------------------|------|
| Tris HCI<br>pH8.0 | 10mM |
| EDTA              | 1mM  |

### 3. Lysis buffer

| A                 | В    |
|-------------------|------|
| GITC              | 4M   |
| Tris HCl<br>pH8.0 | 50mM |
| SDS               | 0.5g |
| EDTA              | 20mM |

# **Sample Collection**

3m



2 Add  $\underline{A}$  200  $\mu L$  of 1 mm beads to 2mL screw tube



3 Add  $\underline{A}$  870  $\mu L$  Lysis master mix to 2mL screw tube. The final look:

30s

30s



#### Note

In 11F, 4°C fridge

Lysis master mix: 225  $\mu$ L of TE buffer + 375  $\mu$ L of lysis buffer + 270  $\mu$ L of 10M ammonium acetate

4 Collect 4 20-50 mg of **sample** to 2mL screw tube

### Note

You can collect up to 100 mg of sample if you can until you bump into the low DNA quality or PCR success rate; by then it means too many inhibitors in the sample and you have to lower the input.

## **Sample crush**

4m

5 Put the 2mL screw tube in mixmill for sample crush, at 3200 rpm 00:04:00

#### Note

Remember to balance if you have odd number of samples

## Centrifugation

3m

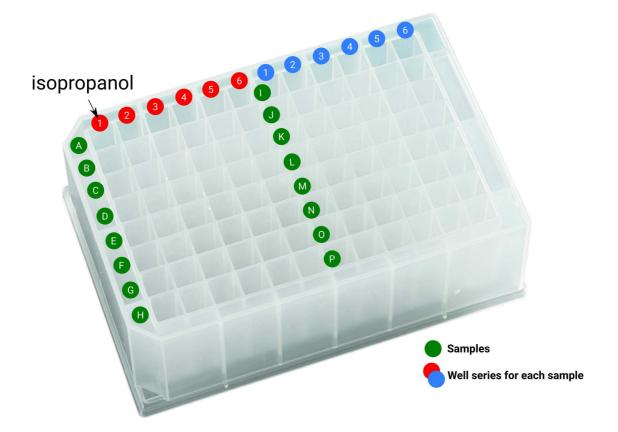
6 Put 2mL screw tube in centrifuge for centrifugation, at this condition: (3) 10 x g, 25°C, 00:03:00



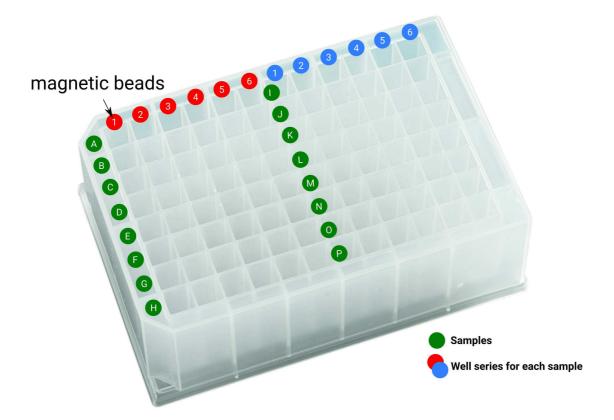
### **DNA** purification

37m 30s

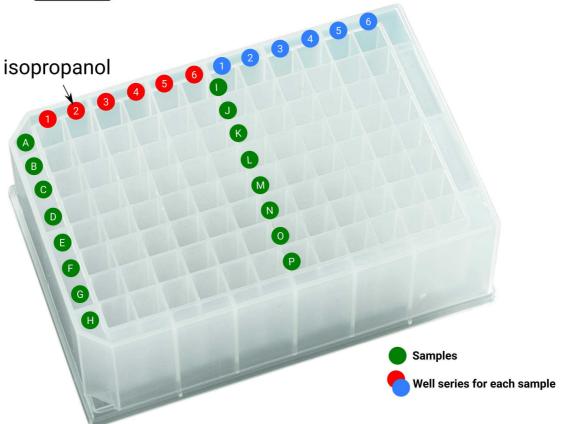
Add  $\pm$  350  $\mu L$  of **isopropanol** to the 1st well of 96 well plate 7



8 Add  $\perp$  50  $\mu$ L of magnetic beads (10mg/ml) to the 1st well of 96 deep well plate



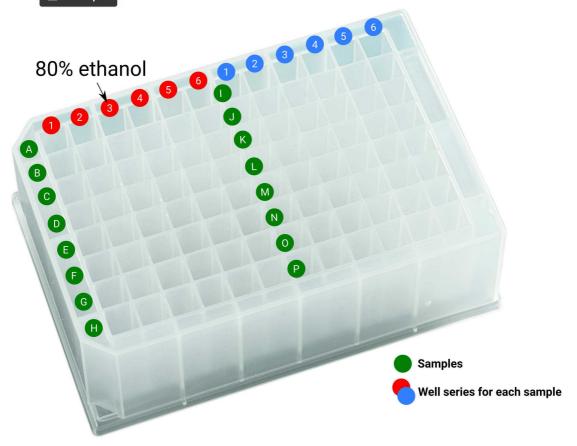




30s

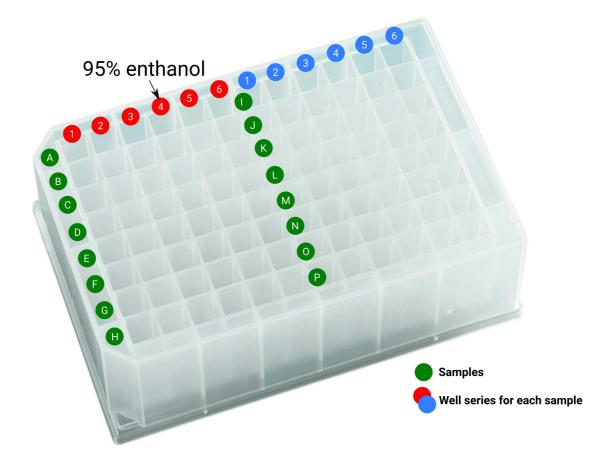
Add Δ 300 μL of **80% ethanol** to the 3rd well of 96 deep well plate



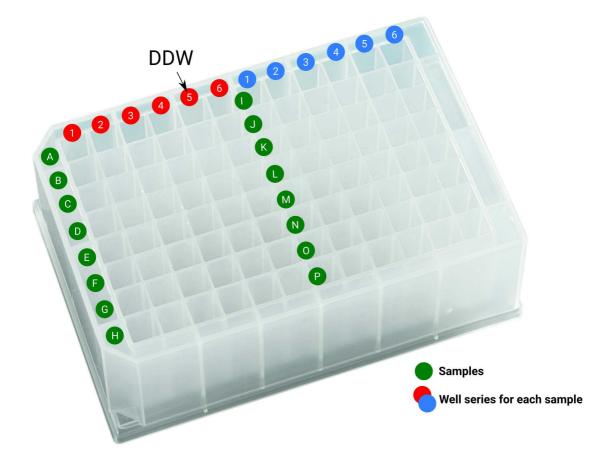


Add  $\pm$  300  $\mu$ L of **95% ethanol** to the 4th well of 96 deep well plate

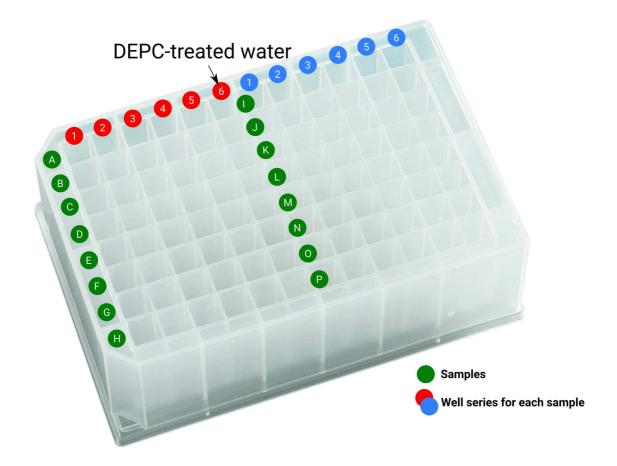
30s



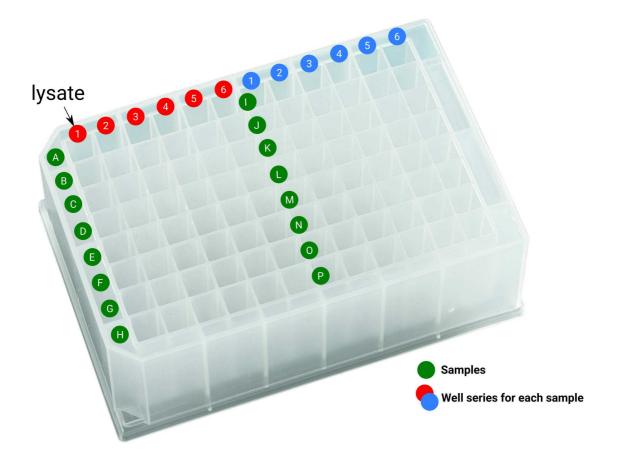
12 Add  $\coprod$  300  $\mu$ L of **DDW** to the 5th well of 96 deep well plate



Add  $\triangle$  100  $\mu$ L of **DEPC-treated water** to the 6th well of 96 deep well plate



Add  $\pm$  300-500  $\mu$ L of the **sample (lysate)** from the 1.5mL centrifuged tube to the 1st well of 96 deep well plate



### Note

Pipetting **as many lysate as you can**, as long as it's free of any cell debris (no solids in your tip)

Put the prepared 96 deep well plate in the automated DNA extraction machine and select the BOMB protocol

34m

16 After the extraction is done, put on the 96 magnetic plate to pellet the magnetic bead residues.



