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## Expression and purification of PI3KC3-C1 complex

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**Protocol status:** Working We use this protocol and it's working

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## **ABSTRACT**

Expression and purification of PI3KC3-C1 complex

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## **PROTOCOL** integer ID:

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	Expression	2d 0h 40m
1	Transfect HEK GNTi cells at concentration of 2 × 10 <sup>6</sup> cells/ml	
2	Dilute PEI with Warm Hybridoma-SFM(1X)	
3	In a separate tube, dilute DNA with Hybridoma-SFM(1X)	
4	Add PEI to DNA dilution. Incubate mixture for 00:30:00 at 37 °C	30m
5	Add mixture to cells. Let cells grow for 48:00:00	2d
6	Harvest Cells 500 rpm , 4 °C , 00:10:00	10m

7 Wash pellet with cold PBS. Store pellet at \$\circ\$ -80 °C until purification.

## **Purification**

2h 50m

- 8 Pellets were homogenized 20 times by Pyrex douncer (Corning) in lysis buffer (25 mM HEPES pH 7.5, 200 mM NaCl, 2 mM MgCl2, 10% Glycerol) with 25mM TCEP/proteinase inhibitors (Thermo Scientific), and then add 10% Triton X-100 stock to final 1% concentration.
- 9 Rocking at **4** °C for **(\*)** 01:00:00

1h

10 Clarify lysate for ( 17000 rpm for ( 00:40:00 at \$ 4 °C

40m

- 11 Wash strep-tactin resin (IBA Lifesciences, Germany) into lysis buffer (without Triton). Load clarified lysate onto resin
- 12 Rock supernatant with equilibrated Strep-Tactin Sepharose resin for 🔊 Overnight



- 13 Wash with 5CV lysis buffer (25 mM HEPES pH 7.5, 200 mM NaCl, 2 mM MgCl2, 25 mM TCEP, 10% Glycerol)
- 14 Elute with lysis buffer plus 4 mM desthiobiotin for STREP resin

Concentrate elution and inject onto pre-equilibrated Superose 6 Increase 10/300 GL column (Cytiva)

(25 mM HEPES pH 7.5, 300 mM NaCl, 1 mM MgCl2, 1 mM TCEP)

Pool peak fractions, concentrate, snap freeze, and store at F-80 °C