JUN 13, 2023

OPEN ACCESS

DOI:
dx.doi.org/10.17504/protocols.io.x54v9pezzg3e/v1

Protocol Citation: Tsu-Chun Hung, Yin-Tse Huang 2023. Size selection (Purification). **protocols.io** https://dx.doi.org/10.17504/protocols.io.x54v9pezzg3e/v1

Protocol status: Working
We use this protocol and it's working

Created: Jun 13, 2023

Last Modified: Jun 13, 2023

PROTOCOL integer ID:
83294

Size selection (Purification)

Forked from Size selection (Purification)

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ABSTRACT

Size selection (Purification)

1 Calculate the amount of magnetic beads you need and **DISTRIBUTE in a new 1.5 mL eppendorf tube (**Mix the bottle **VIGOROUSLY** to make sure it's homogeneous**).**

For example, if you have 5 samples to clean up, distribute 49.5 µl in a new 1.5 mL eppendorf

tube.

	A	B	C
	Sample	Magnetic beads need	Magnetic beads need *1.1
	1	9 µl	9.9 µl
	2	18 µl	19.8 µl
	3	27 µl	29.7 µl
	4	36 µl	39.6 µl
	5	45 µl	49.5 µl
	6	54 µl	59.4 µl
	7	63 µl	69.3 µl
	8	72 µl	79.2 µl

Note

The ratio is **20µl of sample** use **9µl of magnetic beads** to clean up (0.45X to keep long DNA fragments).

Note

Magnetic bead: BeaverBeads™ DNA Select Isolation

2 In each 20 µL sample, add 9 µL magnetic beads in.

Note

Pipetting thoroughly every time before adding to make sure all magnetic beads mixed well.

3 Mix sample and beads by **gently flicking** the tube for 5mins 00:05:00 5m

Note

Be gentle, mix too aggressive will make DNA fragmented

4 Transfer the tube to the 8-strip magnetic rack.
After all the magnetic beads are arrested, remove the supernatant.

5 Add 200 µL 80% ethanol, flip the magnetic rack around to clean the pellet.
Wait until all the magnetic beads are arrested, remove the supernatant.

Note

FRESHLY make 80% ethanol

6 Repeat the **step 5**.

7 Flash spin the tube, and put on the magnetic rack.
Remove **ANY trace of liquid residuals** using 10 µl pipette.

Note

Caution: DO NOT let the beads crack

8 Add 10 µL elution buffer.
Mix gently by flicking the tube and make sure all magnetic beads are dissolved in buffer, then flash spin down the sample.

9 Incubate the tube in PCR thermal cycler using "**37_incubation**" program for 10min 00:10:00 10m
.

10 Transfer the tube to the magnetic rack.
After all the magnetic beads are arrested, collect the 10 µL supernatant to a 200 µl PCR tube or a 8-strip PCR tube.

11 The clean-up sample is now ready for 2' PCR.