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We use this protocol and it's working

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SOP88v1_TGD_Lentivirus transduction in EndoC-bH1

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ABSTRACT

For lentivirus transduction in EndoC-bH1 cells

1 After generating lentivirus (Virus core facility), keep them at -80C.

2 Seed 4x10^6 cells in T25 flask or 7x10^6 cells in T75 flask (EndoC-bH1 cells).

3 The next day, calculate MOI and left the appropriate amount of lentivirus (MOI must be tested to ensure appropriate before transduction) on the cells for 8 days.

4 Check whether GFP is detected in the cells using fluorescence microscope if GFP is included in the construct (In my experience, the signals became stronger over time, so I checked it everyday, and Day 8 was the best. I think it is related we split EndoC-bH1 cells once a week (cell doubling time)).

5 8 days later, split them into separate plates.

6 **CAUTION:**
1. There is no significant difference depending on whether polybrene is present or not.
2. When puromycin was used, the cells did not grow well, thus it was not possible to make cell stocks or any experiments. Additionally, since GFP is present in almost all cells after viral (containing GFP) transduction, selection by puromycin is unlikely to be necessary.