

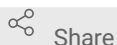


Jul 18, 2022

# DNA extraction (BOMB\_Soil)

Forked from [DNA extraction \(BOMB\)](#)Tsu-Chun Hung<sup>1</sup>, Yin-Tse Huang<sup>1</sup>, Hsin-Mao Wu<sup>2</sup><sup>1</sup>KMU; <sup>2</sup>Kaohsiung Medical College

In Development



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[dx.doi.org/10.17504/protocols.io.q26g74e8kgwz/v1](https://dx.doi.org/10.17504/protocols.io.q26g74e8kgwz/v1) Hsin-Mao Wu

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## ABSTRACT

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## DOI

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## PROTOCOL CITATION

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## FORK NOTE

## FORK FROM

Forked from [DNA extraction \(BOMB\)](#), Yin-Tse Huang

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#### CREATED

Jul 18, 2022

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Jul 18, 2022

#### PROTOCOL INTEGER ID

66907

#### MATERIALS TEXT

1. P1000 pipette
2. 96-deep well
3. MagBeads
4. TE buffer
5. Lysis buffer
6. Isopropanal
7. 80 % Ethanol
8. DEPC treated water
9. Zixpress 32 System
10. 0.5mm zirconia beads
11. 1.0mm zirconia beads
12. 0.133M Ammonium acetate
13. 0.06M Aluminium sulfat

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Sample Collection

3m

1 Measuring **250 mg** for a soil sample

2 Add **200 µL** of **1mm beads** to 2ml enppendorf tube 30s

3 Add **200 µL** of **0.5mm beads** to 2ml enppendorf tube 30s

4 Add **225 µL** of **TE buffer** to 2ml enppendorf tube 30s

TE buffer is in 4°C fridge

5 Add **375 µL** of **lysis buffer** to 2ml enppendorf tube 30s

Lysis buffer is in 4°C fridge

6 Take 2ml enppendorf tube out of the laminar flow and transfer soil samples to the 2ml enppendorf tube 1m










Sample crush 4m

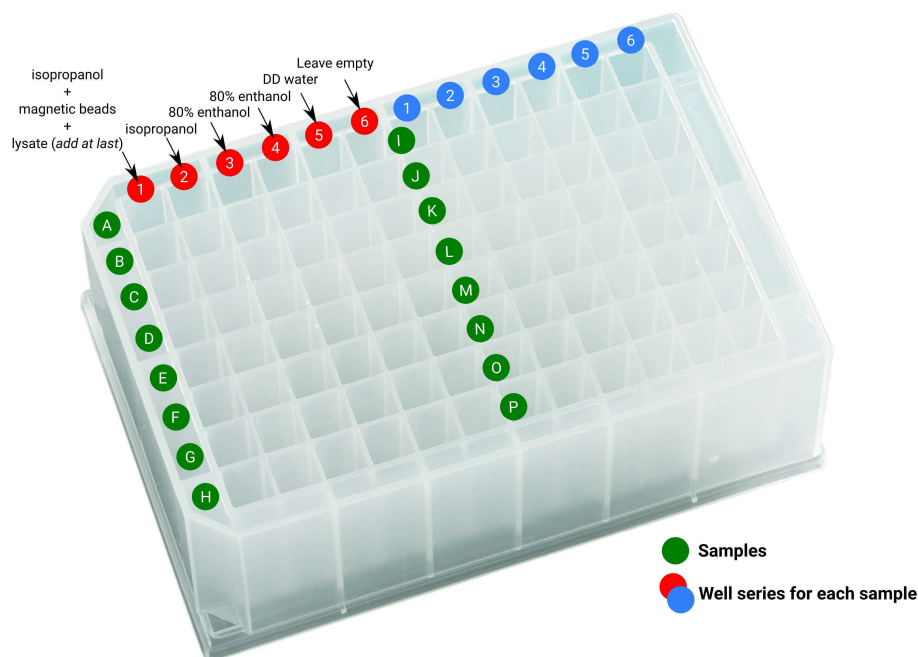
7 Put 2ml enppendorf tube in mixmill for sample crush, at this condition: 30 rpm/s, for 4mins 4m  
🕒 **00:04:00**


Centrifugation 3m

8 Put 2ml enppendorf tube in centrifuge for centrifugation, at this condition: 3m  
⚙️ **17.0 x g, 25°C, 00:03:00**


Remove proteins and humic acid

- 9 Add  **250 µL** 0.133M Ammonium acetate in a new 2ml enppendorf tube
- 10 Transfer supernatant  **300 µL** of step 8 into 2ml enppendorf tube in step 9
- 11 Incubate on ice for  **00:10:00** 10m
- 12 Put 2ml eppendorf tube in centrifuge for centrifugation, at this condition:  
 **17.0 x g, 25°C, 00:03:00** 3m
- 13 Add  **200 µL** 0.06M Aluminium sulfate in another 2ml enppendorf tube
- 14 Transfer step 12 supernatant  **500 µL** in step 13's 2ml enppendorf tube
- 15 Incubate on ice for  **00:10:00** 10m
- 16 Put 2ml eppendorf tube in centrifuge for centrifugation, at this condition:  
 **17.0 x g, 25°C, 00:10:00** 10m
- DNA purification 37m 30s
- 17 Add  **350 µL** of **isopropanol** to the 1st well of 96 well plate 30s







17.1 Add  **125  $\mu$ L** of **magnetic beads** (10 mg/ml) to the 1st well of 96 deep well plate 30s

Shake the bottle and pipetting before using magnetic beads

17.2 Add  **200  $\mu$ L** of the **sample (lysate)** from the 2ml centrifuged tube to the 1st well of 96 deep well plate 30s

**USUALLY ADD at LAST**

18 Add  **400  $\mu$ L** of **isopropanol** to the 2nd well of 96 deep well plate 30s

- 19 Add  **300 µL** of **80% ethanol** to the 3rd well of 96 deep well plate 30s
- 20 Add  **300 µL** of **80% ethanol** to the 4th well of 96 deep well plate 30s
- 21 Add  **100 µL** of **DD water** to the 5th well of 96 deep well plate 30s
- 22 Put the prepared 96 deep well plate in the automated DNA extraction machine 34m
- 23 After the extraction is done, collect  **100 µL** of the **eluted sample** as the DNA template for downstream experiments