



MultiQuas (Multiple reference quasispecies reconstruction protocol) V.1

Marco Cacciabue¹

¹Instituto de Agrobiotecnología y Biología Molecular (IABIMO, INTA-CONICET)

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In Development

Share

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FMDV_ARG_Lab

Marco Cacciabue

Instituto de Agrobiotecnología y Biología Molecular (IABIMO,...)

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ABSTRACT

The following protocol summarizes the major steps to run the MultiQuas pipeline to evaluate viral variability and reconstruct the viral quasispecies from NGS data (particularly Miseq reads). It is based on the assumption that 1 or more known references are available. These references could be obtained using other haplotype reconstruction softwares. Nonetheless, it is recommended that only a few trusted references are used.

PROTOCOL CITATION

Marco Cacciabue 2021. MultiQuas (Multiple reference quasispecies reconstruction protocol). [protocols.io](https://protocols.io/view/multiquas-multiple-reference-quasispecies-reconstr-bxahpib6)
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52265

MATERIALS TEXT

QuRe [↗](#)

[source](#) by Mattia C. F. Prospero

FastQC 0.11.9 [↗](#)

by Simon Andrews

Align_to_references.sh

#!/bin/bash

start=`date +%s`

bbduk.sh in1=\$2 out1=reads_1.fq in2=\$3 out2=reads_2.fq ref=[path/to/bbmap/instalation]/bbmap/resources/adapters.fa ktrim=r k=23 mink=11 hdist=1 tpe tbo qtrim=rl trimq=20 minlen=50 maq=20

bowtie2-build \$1 VFaref

bowtie2 --no-discordant --no-mixed -p \$4 -x VFaref -1 reads_1.fq -2 reads_2.fq | samtools view -@ 4 -bT \$1 - > SAMPLE.bam

samtools sort -@ 4 -m 2G SAMPLE.bam > SAMPLE_sorted.bam

samtools view -@ 4 -h -F 4 -b SAMPLE_sorted.bam > SAMPLE_map.bam

samtools index -@ 4 SAMPLE_map.bam SAMPLE_map.bai

samtools depth -d10000000 SAMPLE_map.bam > coverage.txt

lofreq viterbi -f \$1 -o SAMPLE_map_viterbi.bam SAMPLE_map.bam

samtools sort -@ 4 -m 2G SAMPLE_map_viterbi.bam > SAMPLE_map_viterbi_sorted.bam

samtools index -@ 4 SAMPLE_map_viterbi_sorted.bam SAMPLE_map_viterbi_sorted.bai

lofreq indelqual --dindel -f \$1 -o SAMPLE_map_viterbi_sorted_indels.bam SAMPLE_map_viterbi_sorted.bam

samtools index -@ 4 SAMPLE_map_viterbi_sorted_indels.bam SAMPLE_map_viterbi_sorted_indels.bai

lofreq call-parallel --pp-threads \$4 --call-indels --use-orphan -f \$1 SAMPLE_map_viterbi_sorted_indels.bam -o variants.vcf

end=`date +%s`

echo Execution time was `expr \$end - \$start` seconds.

bbduk [↗](#)

[source](#) by Brian Bushnell

samtools 1.12 [↗](#)

[source](#)

bcftools 1.12 [↗](#)

[source](#)

Bowtie2 2.4.4 [↗](#)

[source](#)

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Brief pipeline description

1

Reads are trimmed and filtered using

bbduk [↗](#)

[source](#) by Brian Bushnell

- 2 Filtered and trimmed reads are aligned to a set of user-defined references (multifasta format) with

Bowtie2 2.4.4 [↗](#)

[source](#)

- 3 Reads are then split into different classes (one for each reference and one for the unmapped reads) using

SAMtools 1.8 [↗](#)

Linux

[source](#) by Wellcome Trust Sanger Institute

- 4 For each class, reads are merged using

PEAR - Paired-End reAd mergeR

[source](#) by Alexandros Stamatakis

and then haplotypes are reconstructed with

QuRe [↗](#)

[source](#) by Mattia C. F. Prospero

Important note: If the time limit is reached this step is repeated with a subset of the reads in order to reduce computation time and resources required by QuRe. Steps include 0.99, 0.9, 0.8, 0.7, 0.6 and 0.5 proportions of the total reads for each class in that order.

- 5 Next, the proportions of each haplotype class (predicted by QuRe) are adjusted to reflect the number of reads of the corresponding class.
- 6 All reconstructed haplotypes are aligned to the first reference in the multifasta file using

mafft 7.487 [↗](#)

[source](#) by Kazutaka Katoh

- 7 Additionally, reads are then aligned to the first reference in the multifasta file. Single Nucleotide Variants (SNVs) are called using

Lofreq 2 [↗](#)

[source](#) by Andreas Wilm

- 8 Finally, concordance between the SNVs (expected minor allele frequency) from the predicted quasispecies and the Lofreq variants using an in-house R script. Higher value of R-squared indicates a better quasispecies reconstruction. Important: this step does not mean validation of the obtained results, but it allows the user to choose between different haplotypes reconstructions.

Installing docker

- 9 In order to run the pipeline, a wrapper file is available (bash) which automatically performs all the above numbered steps. A docker image is available that includes all the necessary dependencies. If you do not yet have docker installed, do so at this time, and ensure that is in your PATH. For more information please visit <https://www.docker.com/get-started>
- 10 The docker image ("rm") is available at Docker hub
To pull the image, use the command below:

```
docker pull
```

```
docker pull cacciabue/rm:latest
```

This will download and install the corresponding docker image. Only has to be run the first time (it may take several minutes depending on your internet connection)

- 10.1 Alternatively, If you don't want to use Docker, you can install all dependencies by yourself (only for linux users). The dependencies are:

- BCFtools v1.8 (or later version) <http://www.htslib.org/download/>
- Samtools v1.8 (or later version) <http://www.htslib.org/download/>
- Bowtie2 v2.2.4 <http://bowtie-bio.sourceforge.net/bowtie2/index.shtml>
- PEAR <https://cme.h-its.org/exelixis/web/software/pear/doc.html>
- seqtk <https://github.com/lh3/seqtk>
- bbmap <https://jgi.doe.gov/data-and-tools/bbtools/bb-tools-user-guide/bbduk-guide/>
- Lofreq v2 <https://csb5.github.io/lofreq/>
- mafft <https://mafft.cbrc.jp/alignment/software/>
- R v4.1 (or later version) <https://www.r-project.org/>
- R package seqinr <https://cran.r-project.org/web/packages/seqinr/index.html>
- R package ape <https://cran.r-project.org/web/packages/ape/>
- R package VariantAnnotation <https://bioconductor.org/packages/release/bioc/html/VariantAnnotation.html>

- R package Biostrings <https://bioconductor.org/packages/release/bioc/html/Biostrings.html>
 - R package ggplot2 <https://ggplot2.tidyverse.org/>
 - R package ggrepel <https://cran.r-project.org/web/packages/ggrepel/vignettes/ggrepel.html>
- and download the following file (bash script)

AGREGAR ARCHIVO O QUITAR!!!!!!

and the two following R script should also be downloaded

11 Depending on your operating system follow these steps

11.1 On windows: open a windows terminal (WIN + R), type "cmd" and press enter. A Windows terminal should be up and running.

11.2 Navigate to the folder containing the fastq files and the reference file.