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# Fitting enzyme kinetics data with Solver

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## ABSTRACT

Reaction kinetics are a fundamental component of the biochemical characterization of a biomolecule. The  $V_{\max}$  and the  $K_M$  values derived from experiments describe how fast a biomolecule catalyzes the reaction and the approximate affinity between biomolecule and substrate. For almost a century, these properties have been used to describe the mechanism and functions of enzyme and other biomolecules. Fitting data to calculate kinetic values like the  $V_{\max}$  and the  $K_M$  values is a fundamental skill in biochemistry.

This protocol describes fitting data determined at several substrate concentrations to calculate  $V_{\max}$  and  $K_M$  values using Excel/Google Sheets and the macro with these programs Solver. This protocol is based on the work of Kemmer and Keller.

This protocol is written for students in Biochemistry at James Madison University, however the approach is general enough for any enzyme kinetics data.

## THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Kemmer, G., and Keller, S. (2010) Nonlinear least-squares data fitting in Excel spreadsheets. Nat. Protoc. 5, 267–281.

## MATERIALS TEXT

Microsoft Excel or Google Sheets

Kinetics data from at least 5 concentrations of substrate

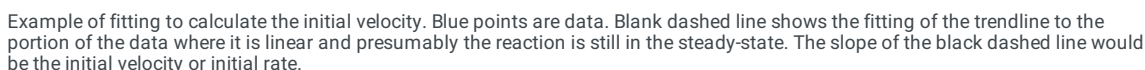
## Calculating initial rates

- 1 Calculate the initial rate from the slope of the product formed vs. time plot.



It is important to calculate the rate when the plot of product formed vs. time is linear with time. This ensures that the enzyme is in the **steady-state**. Typically this is in the early part of the reaction before more than 10% of the substrate has been converted to product (see figure). *The rate is the slope of the line.*

In the steady-state, the concentration of enzyme bound complexes is not changing with time and the enzyme is catalyzing the reaction at the fastest possible rate at that concentration of substrate.



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1.3 Record the rate at each concentration of substrate in the table below. Be sure to modify the units to match the units from the experiment. Add rows to the table as needed.

[illegible]

## Plotting data to estimate kinetic values

- Plot rate (y-axis) vs Substrate concentration (x-axis) and estimate the  $V_{\max}$  and  $K_M$  values using the plot. Plot the data as a scatter plot (no lines)



The  $V_{\max}$  estimate is the y-value where the graph levels out. The  $K_M$  is the x-value at  $1/2 V_{\max}$ .

- Record your estimates with units of  $V_{\max}$  and  $K_M$  in the table below.

$V_{\max}$		
$K_M$		

## Fitting data to calculate $V_{\max}$ and $K_M$

- To draw a trendline, you'll need to calculate theoretical values for rates using the estimated  $V_{\max}$  and  $K_M$  values from 2.1 and the Michaelis-Menten Equation. This is your  $V$  Calculated ( $V_{\text{calc}}$ ). This will also help you calculate the correct  $V_{\max}$  and  $K_M$  values later on.
- In Microsoft Excel or Google Sheets, set up a table with headers as shown below:

Concentration of substrate	Observed rate	Theoretical rate ( $V_{\text{Calc}}$ )	Squared residual		Variables	Estimates	Units
					$V_{\max}$		
					$K_M$		
					SSR		

- Input your data table from step 1.3 into columns A and B.

- Input your estimates of  $V_{\max}$  and  $K_M$  from step 2.1 into the appropriate box in column G

- In column C, insert the following code to use the Michaelis-Menten equation to calculate the theoretical rate given the estimates for  $V_{\max}$  and  $K_M$ .

```
=($G$2 * A2) / ($G$3 + A2)
```



This code uses the Michaelis-Menten equation  $v = \frac{V_{\max} \times [S]}{K_M + [S]}$  to calculate the rate based on the

estimates. The \$ locks those values in so that the equation can be "dragged" down for all concentrations of substrate without needing to re-write the equation.

- 5.1 "Drag" the equation down until the theoretical rate has been calculated for all the concentrations of substrate. After this step the table will appear similar to below:

Conc	Obs	V Calc
10	05628495	0.05
20	09715311	0.08571429
50	15269997	0.15
100	17799929	0.2
200	26219102	0.24

- 5.2 In Column D, insert the following code to calculate the squared residual value.

```
=(B2 - C2)^2
```



The squared residual will be used to calculate error and refine the estimates of  $V_{\max}$  and  $K_M$ . It is the squared difference between the real rate and the calculated rate. For a perfect match the squared residual should be zero.

- 5.3 "Drag" the equation down until the squared residual has been calculated for all the concentrations of substrate.

- 5.4 In cell G4, insert the following code to calculate the sum of squared residuals. Replace DXX with D followed by the number of the last substrate concentration (e.g. D10 if there are 9 substrate concentrations).

The resulting number is the sum of squared residuals or SSR.

```
=sum(D2 : DXX)
```

- 6 Use SOLVER to fit the data by changing the  $V_{\max}$  and  $K_M$  estimates to minimize the sum of squared residuals.



- 6.1 If Solver has not been used before, it may not have been installed as described below. If it has been used before skip to step 6.2.

**Note:** For some users of Excel, add-ins are disabled, therefore Google Sheets has to be used.

**FOR EXCEL USERS:**

If Solver has not been installed, it can be added to Excel by going to File -> Options -> Add-ins -> Manage Excel Add-ins -> Solver Add-in

#### FOR GOOGLE SHEETS USERS:

If Solver has not been installed, it can be added to Sheets by going to Add-ons -> Get Add-ons, and searching for Solver.



6.2 Load the Solver window. In Excel, Solver is found in the Data tab. In Sheets, it is found in the Add-ons menu.

6.3 The Solver window (example from Excel shown below) has 4 parameters to set:

- **Set Objective:** \$G\$4
- **To:** Min
- **By Changing Variable Cells:** \$G\$2, \$G\$3
- **Select a Solving Method:** GRG Nonlinear

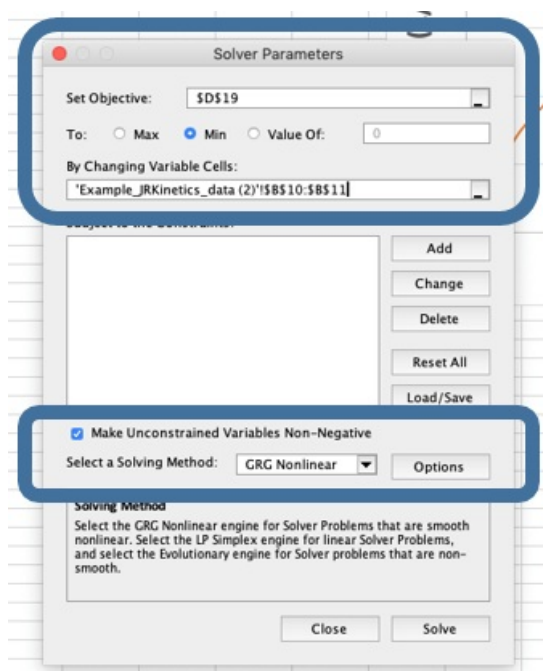
The Objective and Variable cells can also be selected using the button at the right of the window and using the cursor to click on the appropriate cells.

The image is a screenshot of the Excel Solver window. The title bar says "Solver" with a close button. The window contains the following fields and buttons:

- Set Objective:** A text box with a selection icon on the right.
- To:** Radio buttons for "Max", "Min", and "Value Of:", followed by a text box.
- By Changing:** A text box with a selection icon on the right.
- Subject To:** A large empty text box.
- Buttons:** "Add", "Change", and "Delete" buttons below the "Subject To" box.
- Solving Method:** A dropdown menu showing "Standard LSGRG Nonlinear".
- Buttons:** "Reset All" and "Insert Example" buttons below the "Solving Method" dropdown.
- Buttons:** "Solve" and "Options" buttons at the bottom.

At the bottom of the window, it says "Copyright © 2019 Frontline Systems, Inc." and has a logo with a lightbulb and the word "Solver".

Solver in Google Sheets



Solver in Excel

- 6.4 Click Solve and wait for the fitting to finish (<10 seconds).



Solver will substitute in various values for  $V_{\max}$  and  $K_M$  to reduce the SSR value as much as possible which means that the theoretical values match the experimental values closely. At the end, the values will be changed automatically in the spreadsheet

- 6.5 **In Sheets:** Solver will run and change the values.

**In Excel:** A window will pop-up asking to accept the values and display if there were errors. Click OK and the estimated  $V_{\max}$  and  $K_M$  will be changed to the optimized values.

- 6.6 Record the values  $V_{\max}$ ,  $K_M$ , and SSR below. These  $V_{\max}$  and  $K_M$  values are accurate compared to your original estimates.

Vmax	
Km	
SSR	1.99566E-07

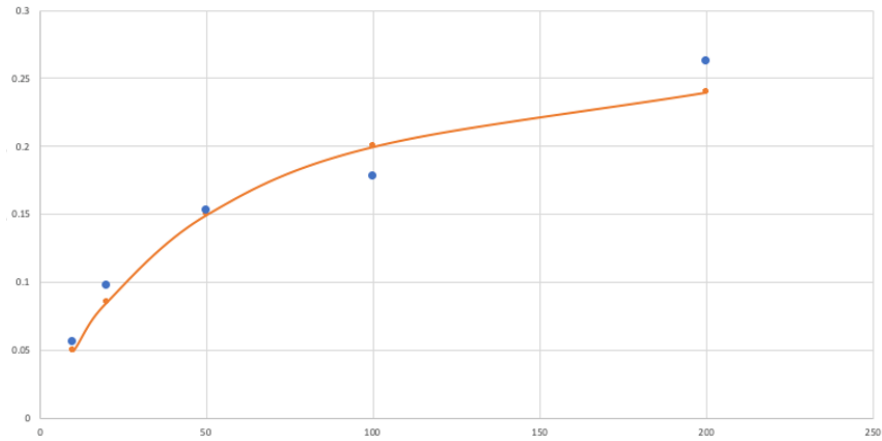
#### Visualizing the fit to the data

- 7 Add a second set of data to the original plot of observed rate vs. concentration of substrate.

7.1 Plot the theoretical (V Calc) rates vs. concentration of substrate.

7.2

Click on the newly plotted V Calc data and edit the data set to show only the line. The plot should look similar to that shown below:



7.3 Label the plot axes with appropriate labels and units.

7.4 Save your plot as a picture and upload the image.