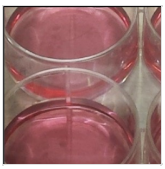

Sep 10, 2020

Feeding T75, T150, and 6wp Backup

In 1 collection

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1 Works for me dx.doi.org/10.17504/protocols.io.8gkhtuw

Neurodegeneration Method Development Community
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ABSTRACT

Protocol includes feeding of T75 flask, T150 flask, or the back up 6 well plate.

DOI

dx.doi.org/10.17504/protocols.io.8gkhtuw

PROTOCOL CITATION

Andrea Argouarch 2020. Feeding T75, T150, and 6wp Backup. **protocols.io** https://dx.doi.org/10.17504/protocols.io.8gkhtuw

COLLECTIONS

Dural Cell Isolation and Culturing - Collection

CREATED

Oct 19, 2019

LAST MODIFIED

Sep 10, 2020

PROTOCOL INTEGER ID

28908

PARENT PROTOCOLS

Part of collection
Dural Cell Isolation and Culturing - Collection

STEPS MATERIALS

NAME	CATALOG #	VENDOR
Fetal Bovine Serum	97068-091	Vwr
DMEM, high glucose, pyruvate	11995073	Thermo Fisher
Penicillin-Streptomycin	15140122	Gibco - Thermo Fisher

Observations

1 Observe cell morphology and outgrowth

2 Feed every 2-3 days until 90-100% confluent

Preparation

3 Turn off UV lights and clean hood with 70% ethanol

4 Clean items with 70% ethanol and bring into hood
a. Sterile Filtered Media with PenStrep

DMEM, high glucose, pyruvate
by Thermo Fisher
Catalog #: 11995073

Penicillin-Streptomycin
by Gibco - Thermo Fisher
Catalog #: 15140122

Fetal Bovine Serum
by Vwr
Catalog #: 97068-091

5 Write down observations on confluency

Culturing

6 Aspirate old media

7 Add 20 mls **20 mL** of media with penstrep to T75 flask

8 Add 30 mls **30 mL** of media with penstrep to T150 flask

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9 Add 2 mls **2 mL** of media with penstrep to 6wp backup

10 Continue to feed every 2-3 days until 90-100% confluent

Clean Up

11 Throw away biohazard materials properly

12 Clean and sterilize hood with 70% ethanol and turn on UV

13 Update cell culture notes in lab notebook

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