

Aug 23, 2024



High-Capacity cDNA Reverse Transcription

DOI

dx.doi.org/10.17504/protocols.io.6qpvr8wq2lmk/v1

Hector Martell Martinez¹

¹University of Minnesota

ASAP Collaborative Rese...

Team Lee



Jane Balster

ASAP - Team Lee





DOI: dx.doi.org/10.17504/protocols.io.6qpvr8wq2lmk/v1

Protocol Citation: Hector Martell Martinez 2024. High-Capacity cDNA Reverse Transcription. protocols.io

https://dx.doi.org/10.17504/protocols.io.6qpvr8wq2lmk/v1

License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits

unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working We use this protocol and it's

working

Created: August 21, 2024

Last Modified: August 23, 2024

Protocol Integer ID: 106358

Keywords: ASAPCRN

Funders Acknowledgement:

ASAP

Grant ID: 000592



Abstract

This protocol details the high-capacity cDNA reverse transcription.

Materials

Applied BiosystemsTM High-Capacity cDNA Reverse Transcription Kit Applied Biosystems (ThermoFisher Scientific) Catalog #4368814

Master Mix:

A	В
Component	Volume
1) Nuclease-free Water	3.2 µL
2) 10X RT Buffer	2.0 μL
3) 10X Random Primers	2.0 μL
4) RNase Inhibitor	1.0 μL
5) 25X dNTP Mix	0.8 μL
6) MultiScribe Reverse Trans criptase	1.0 µL ***add last!!
TOTAL per reaction	10 μL



Nanodrop

- Nanodrop each isolated sample of RNA.
- 1.1 A good concentration of RNA is between 🚨 200 undetermined - 🚨 2000 undetermined .
 - a final concentration below \(\brace 2000 \) undetermined \(.
- 1.2 A good 260/280 value is \sim 2.0.
- 1.3 A good 260/230 value is $\sim 2.0-2.2$.

cDNA Calculations

2 cDNA for all brain regions is made with 🚨 2000 ng of RNA. cDNA for isolated cells is made with the highest amount of RNA that can be made from the least concentrated sample.

TV per reaction = Δ 20 μ L (Δ 10 μ L of RNA/Water + Δ 10 μ L of Master Mix)

■ Calculate the RNA amount needed to make

2000 ng of RN for each sample

Ex. For a RNA concentration of <u>A</u> 1250.0 undetermined .

 $2000/1250 = 4.6 \,\mu L$ of RNA

3 Calculate the amount of water to be added to the RNA for a TV = 4 10 µL

Ex. For 4 1.6 µL of RNA

 \perp 10 μ L - \perp 1.6 μ L of RNA = \perp 8.4 μ L of Water



Sample Number	RNA (ng/ul	260/280	260/230	Sample #	cDNA (2000 ng)	Water
1	849.1	2.11	2.34	1	2.36	7.64
2	1038.6	2.1	2.24	2	1.93	8.07
3	604.8	2.07	2.21	3	3.31	6.69
4	985.7	2.1	2.32	4	2.03	7.97
5	948.8	2.09	2.32	5	2.11	7.89
6	736.2	2.08	2.26	6	2.72	7.28
7	1185.5	2.1	2.33	7	1.69	8.31
8	450.5	2.12	2.19	8	4.44	5.56
9	1070.6	2.1	2.32	9	1.87	8.13
10	1000.9	2.11	2.33	10	2.00	8.00

Making cDNA

- 4 Thaw the isolated RNA and the following components of the High-Capacity cDNA reverse transcription kit 3 On ice.

- 10X RT Buffer
- 25X dNTP Mix ([M] 100 millimolar (mM))
- 10X Random Primers Can thaw at

 Room temperature
- RNase Inhibitor

Note

- DO NOT thaw MultiScribe Reverse Transcriptase it does not freeze at 2 -20 °C and is prone to denaturing at higher temperatures. Keep at 20 °C until creating your master mix in **■**5 below.
- 5 While the above components thaw

 On ice pipette the calculated amount of water (from of cDNA calculations) to PCR tubes. This step can be done at 8 Room temperature.
- 6 Place the PCR tubes with water | On ice | and then add the calculated amount of RNA (from of cDNA calculations) to its respective PCR tube.



- 7 Create the following master mix On ice . Make enough master mix for each sample plus a little extra (if you have 10 samples, make enough master mix for 11).
- 7.1 • Add reagents to a 1.5 mL tube in the following order.

Α		В	
С	omponent	Volume	
1	Nuclease-free Water	3.2 µL	
2) 10X RT Buffer	2.0 μL	
3	10X Random Primers	2.0 µL	
4) RNase Inhibitor	1.0 µL	
5	25X dNTP Mix	0.8 μL	
6	MultiScribe Reverse Transcriptase	1.0 µL ***add last!!	
Т	OTAL per reaction	10 μL	

7.2 Mix gently.



8 Add \perp 10 μ L of master mix to each PCR tube \parallel On ice .



9 Mix PCR tubes gently then spin down briefly.



10 Keep On ice until performing the reverse transcription.



Perform Reverse Transcription

11 Place PCR tubes into the thermal cycler.



- 12
- 13 Set the following conditions:





Settings	Step 1	Step 2	Step 3	Step 4	
Temp.	25°C	37°C	85°C	4°C	
Time	10 minutes	120 minutes	5 minutes	Hold	

Start the thermal cycler run. 14



15 When the samples reach take the PCR tubes out and store at 4 °C for short term use and at 🖁 -20 °C for long term use.



Protocol references

Refer to the applied biosystems "High Capacity cDNA Reverse Transcription Kit User Guide" for reference.