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Immunostaining- Fluorescent

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We use this protocol and it's working

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Abstract

Staining of mouse brain sections with immunofluroescent visualization.

1 Wash freshly sectioned slices of tissue 2-3 times with 1X PBS to remove OCT.

2 Block with 5% Normal donkey/Goat serum in 0.3% TRITON X-100 in 1X PBS for 01:00:00 at Room temperature . 1h

3 Wash sections 5 times in 1X PBS.

4 Following washes, transfer sections to a primary antibody solution containing a mix of primary antibody diluted in 0.3% TRITON X-100 in 1X PBS with 1% Normal Donkey/Goat serum Overnight at 4 °C . 1h

5 The following day, wash sections 5 times in 1X PBS.

6 Transfer sections to secondary solution containing the adequate secondary antibody diluted in 0.3%
TRITON X-100 in 1X PBS with 1% Normal Donkey/Goat serum for 04:00:00 at Room temperature . 4h

7 Wash sections 3 times in 1X PBS.

8 Mount sections on microscope slides with medium (Vectashield with 4',6-diamidino-2-phenylindole (DAPI), H-1800-10).