



Sep 08, 2020

# FLASH amp

eesha.sharma.phd<sup>1</sup><sup>1</sup>Quantification by Design, Stanford University

1

Works for me

dx.doi.org/10.17504/protocols.io.bk2hkyb6

eesha.sharma.phd

## ABSTRACT

FLASH amp is a method that allows for room temperature detection and amplification of a viral RNA sequence of interest submitted as part of the Xprize competition. The main components of the reaction are not included here and are given generic names such as enzyme X as they are currently not protected under IP. The steps outlined with tubes labeled with the generic names would allow any lab to reproduce the method.

Version 1 of this method outlines the protocol for a fluorescence-based readout that can be used with a qPCR machine or TECAN style plate reader.

Version 2 (to be released shortly) will be a colorimetric readout that can be assessed by eye.

The outlined protocol assumes the user performing this in a laboratory setting will setup reactions in a clean room and analyze results in a separate post-amplification room. In addition, since our amplification is rapid and at room temperature, care but be taken to add ligand to the master mix quickly and the reactions be sealed.

## DOI

dx.doi.org/10.17504/protocols.io.bk2hkyb6

## PROTOCOL CITATION

eesha.sharma.phd 2020. FLASH amp. **protocols.io**  
<https://dx.doi.org/10.17504/protocols.io.bk2hkyb6>

## KEYWORDS

Xprize

## LICENSE

————— This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

## CREATED

Sep 08, 2020

## LAST MODIFIED

Sep 08, 2020

## PROTOCOL INTEGER ID

41769

## STEPS MATERIALS

NAME	CATALOG #	VENDOR
Pyrophosphatase, Inorganic (E.coli) - 50 units	M0361L	New England Biolabs
Corning® Low Volume 384-well Black Flat Bottom Polystyrene	3821BC	

## EQUIPMENT

NAME	CATALOG #	VENDOR
SPARK	SPARK	



DISCLAIMER:

This method is currently used for research and development, not for diagnostic purposes.

#### Sample collection

- 1 Sample should be collected 1 hour after any food is consumed.  
10 minutes before collection rinse mouth well with water.  
Collect passive drool or pooled saliva below tongue into collection tube.

#### Set-up

- 2 Resuspend freeze dried master mix 1 with  **7.04 µl** of water
- 3 Resuspend freeze dried master mix 2 in  **1 µl** water and add to above

- 4 Add  **0.3 µl**



Pyrophosphatase, Inorganic (E.coli) -  
50 units  
by New England Biolabs  
Catalog #: M0361L

- 5 Add  **0.5 µl**

Undisclosed enzyme Z

- 6 Add  **0.5 µl**

Undisclosed Enzyme Y


- 7 Add  **0.13 µl**

Undisclosed Enzyme X

- 8 Put mix in



Corning® Low Volume 384-well Black  
Flat Bottom Polystyrene  
Catalog #: 3821BC

Add  **0.5 µl** of sample and seal plate.



A large master mix of the above components can be mixed and aliquoted into wells.

#### Data collection

- 9 Place plate in TECAN plate reader.



SPARK  
Microwell plate reader

TECAN

SPARK



Set gain to 120, excitation 480 and emission 550.

A qPCR machine can also be used.

Collect data at T= 30 min.

Data can also be collected every 1s for 30 min to monitor kinetics of reaction in the particular plate reader being used if not TECAN SPARK reader.

Fluorescence values of >4000 are positive and <4000 are negative.

