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🌐 Preparation of primary rat cortical neuron and astrocyte co-culture

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1 Works for me

 Sharedx.doi.org/10.17504/protocols.io.n92ldzq1xv5b/v1 Minee-Liane Choi
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ABSTRACT

This protocol describes how to prepare primary rat cortical neuron and astrocyte co-culture.

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MATERIALS TEXT


- Dissecting buffer: Hanks' Balanced Salt solution (HBSS) supplemented with [M]**10 millimolar (mM)** HEPES and [M]**20 % volume** Fetal bovine serum (FBS)
- Washing buffer: HBSS supplemented with 10mM HEPES
- Disgesting buffer: [M]**0.5 Mass / % volume** EDTA-trypsin supplemented with DNase
- Neurobasal completed medium: Neurobasal A medium supplemented with B27, [M]**2 millimolar (mM)** Glutamax, Pen/Strep

1 1-3 days postpartum Sprague Dawley rats (University College London breeding colony) are used.

Experimental procedures are performed according to the United Kingdom Animal (Scientific Procedures) Act of 1986.

2 Rat cortices are placed in an ice-cold Dissecting buffer (described in Materials).

3 Wash five times with Washing buffer (described in Materials).

4 Tissues are digested with a Disgesting buffer (described in Materials) for  **00:15:00**

15m

5 Digested tissues are neutralized with a dissecting buffer.

6 Washing twice with a Washing buffer,

7 Dissociate with a Washing buffer supplemented with DNase.

8 Dissociated pellets are collected in Neurobasal completed medium.

- 9 Approximately 600,000 cells are plated on ~~4~~**25 mm** Poly-D-Lysin (PDL) coated coverslips and 200,000 cells for 8-well ibidi chambers (PDL coated).
- 10 The cultures are maintained at ~~8~~**37 °C** (~~1M~~**5 % volume** CO₂), and the media are changed every 4-5 days.
- 11 Cells can be used at 12-16 days.