



© RNA-Stable Isotope Probing V.3

Roey Angel¹, Eva Petrova¹, Ana Lara¹

¹Soil and Water Research Infrastructure

1 Works for me

dx.doi.org/10.17504/protocols.io.bpwympfw

Nov 23, 2020

SoWa RI Anaerobic and Molecular Microbiology (public) Tech. support email: eva.petrova@bc.cas.cz



Soil and Water Research Infrastructure

ABSTRACT

The following protocol describes how to perform an RNA-Stable Isotope Probing experiment. The scope of this protocol only covers the parts involving separating labelled RNA from unlabelled RNA using ultracentrifugation in a caesium trifluoroacetate density gradient and downstream quantification to evaluate whether the labelling and separation of the RNA were successful. Total RNA should be extracted from an environmental sample or an enrichment culture that was incubated with an isotopically-labelled substrate. Labelling can be of the carbon, oxygen or nitrogen in the RNA (or any combination of the 3). For environmental samples, we recommend extracting RNA using our protocol Total Nucleic Acids Extraction from Soil and purifying it using the Purification of RNA from Crude NA Extract protocol. This protocol is based on the following papers: Whiteley et al. (2007); Dumont et al. (2011); Angel and Conrad (2013). For a comprehensive discussion on how to design a SIP experiment and how to analyse the resulting data, we recommend referring to the recent book on the subject: Stable Isotope Probing: Methods and Protocols, especially chapters: 1-3 and 9-18.

Whiteley AS, Thomson B, Lueders T, Manefield M (2007). RNA stable-isotope probing. Nature protocols.

http://10.1038/nprot.2007.115

Angel R, Conrad R (2013). Elucidating the microbial resuscitation cascade in biological soil crusts following a simulated rain event.. Environmental microbiology.

https://doi.org/10.1111/1462-2920.12140

Dumont MG, Pommerenke B, Casper P, Conrad R (2011). DNA-, rRNA- and mRNA-based stable isotope probing of aerobic methanotrophs in lake sediment.. Environmental microbiology. https://doi.org/10.1111/j.1462-2920.2010.02415.x

THIS PROTOCOL ACCOMPANIES THE FOLLOWING PUBLICATION

Angel, R., and Conrad, R. (2013). Elucidating the microbial resuscitation cascade in biological soil crusts following a simulated rain event. Environ Microbiol 15, 2799–2815. doi:10.1111/1462-2920.12140.

DOI

dx.doi.org/10.17504/protocols.io.bpwympfw

mprotocols.io

11/23/2020

Citation: Roey Angel, Eva Petrova, Ana Lara (11/23/2020). RNA-Stable Isotope Probing. https://dx.doi.org/10.17504/protocols.io.bpwympfw

Roey Angel, Eva Petrova, Ana Lara 2020. RNA-Stable Isotope Probing. **protocols.io** https://dx.doi.org/10.17504/protocols.io.bpwympfw
Version created by Roey Angel

MANUSCRIPT CITATION please remember to cite the following publication along with this protocol

Angel, R., and Conrad, R. (2013). Elucidating the microbial resuscitation cascade in biological soil crusts following a simulated rain event. Environ Microbiol 15, 2799–2815. doi:10.1111/1462-2920.12140.

A

KEYWORDS

Stable isotope probing, SIP, RNA, Ultracentrifugation, Density gradient

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Nov 20, 2020

LAST MODIFIED

Nov 23, 2020

PROTOCOL INTEGER ID

44728

GUIDELINES

- Design of SIP experiments. SIP experiments are usually relatively complex, laborious, and time-consuming, and can, therefore, fail because of various reasons and at different stages. Therefore, the design of a SIP experiment should be carefully considered in advance and cover all aspects and phases, including preliminary knowledge of the environment and the targeted process, the nature and duration of the incubation, how many and what types of controls to include, how many fractions to collect and how deep to sequence. These considerations extend beyond the scope of this protocol. Comprehensive discussions and tips on how to best design a SIP experiment can be found at Angel (2019) and Sieradzki et al. (2020).
- RNA handling. Since RNA is very sensitive to both chemical and enzymatic degradation, some precautionary
 measures should be taken. The RNA molecules are protected from degradation while in the CsTFA gradient but
 are sensitive to degradation during the precipitation and washing steps and downstream applications. For
 more info see Total Nucleic Acids Extraction from Soil.
- Reducing the volume required for the refractometer. The typical handheld-refractometer such as the Reichert AR200 has a large lens size requiring 50-100 μl of liquid to cover its surface adequately. To minimise the volume of wasted sample, it is possible to cover the lens with a piece of strong dark adhesive tape, to which a hole was made using a perforator.
- **Timing.** The timings for each step listed SIP protocol assume that only two gradients are being processed simultaneously. We recommend processing more than 4-8 gradients at a time, but not more.
- Data analysis. Several statistical frameworks have been developed in recent years to analyse SIP datasets such as qSIP (<u>Hungate et al., 2015</u>), HR-SIP (<u>Youngblut et al., 2018</u>) and HR-RNA-SIP (<u>Angel et al., 2018</u>).

Angel R (2019). Experimental Setup and Data Analysis
Considerations for DNA- and RNA-SIP Experiments in the Omics Era.
Methods in molecular biology (Clifton, N.J.).
https://doi.org/10.1007/978-1-4939-9721-3_1

Sieradzki ET, Koch BJ, Greenlon A, Sachdeva R, Malmstrom RR, Mau RL, Blazewicz SJ, Firestone MK, Hofmockel KS, Schwartz E, Hungate BA, Pett-Ridge J (2020). Measurement Error and Resolution in Quantitative Stable Isotope Probing: Implications for Experimental Design. mSystems.

https://doi.org/pii:e00151-20.10.1128/mSystems.00151-20

Youngblut ND, Barnett SE, Buckley DH (2018). HTSSIP: An R package for analysis of high throughput sequencing data from nucleic acid stable isotope probing (SIP) experiments. PloS one. https://doi.org/10.1371/journal.pone.0189616

Hungate BA, Mau RL, Schwartz E, Caporaso JG, Dijkstra P, van Gestel N, Koch BJ, Liu CM, McHugh TA, Marks JC, Morrissey EM, Price LB (2015). Quantitative microbial ecology through stable isotope probing. Applied and environmental microbiology. https://doi.org/10.1128/AEM.02280-15

Angel R, Panhölzl C, Gabriel R, Herbold C, Wanek W, Richter A, Eichorst SA, Woebken D (2018). Application of stable-isotope labelling techniques for the detection of active diazotrophs. Environmental microbiology.

https://doi.org/10.1111/1462-2920.13954

MATERIALS STEP MATERIALS Sigma Catalog #T5941 Step 1.1 Step 3 Step 3 Step 3 Step 3 Step 5 Step 5 Step 5 Step 5 Step 25



Scientific Catalog # AM9740 Step 25 Scientific Catalog #AM7000 Step 32 Scientific Catalog #N8080127 Step 33 ⊠ Bovine Serum Albumin (BSA) Thermo Fisher Scientific Catalog #B14 Step 35 Scientific Catalog #R0191 Step 35 **℧**USB Dithiothreitol (DTT) 0.1M Solution **Thermo Fisher** Scientific Catalog #707265ML Step 35 SuperScript™ IV First-Strand Synthesis System Thermo Fisher Scientific Catalog #18091050 Step 35 **⊠** RNaseOUT™ Recombinant Ribonuclease Inhibitor **Thermo Fisher** Scientific Catalog #10777019 Step 35

Apparatus

- 1) For gradient preparation
- Working bench in a climatised room at 20 °C
- Icebox
- 50 ml tube (for up to 8 gradients)
- Ultracentrifuge (capable of achieving 177,000 g) and a vertical rotor (e.g. Sorvall WX Ultra 100 Ultracentrifuge, TV-1665 rotor). A fixed-angle
- Ultracentrifugation tubes (e.g. Ultracrimp, PA centrifugation tubes 6 ml)
- Ultracentrifugation tube caps
- Refractometer
- Purified RNA samples (DNA-free) with a concentration >20 ng μl⁻¹
- Micropipettes and tips
- Lab-scale

2) For fractionation

- Working bench in a climatised room at 20°C
- Refractometer
- Low-binding tubes (one per fraction: 1.5 ml)
- Test tube utility clamp mounted on a stand
- Automatic syringe pump (e.g. NewEra's NE-300 Syringe Pump)
- 20 ml syringe
- Precision pump peroxide-cured silicone tube (or similar), 1/16", about 0.5-1 m long
- Luer fittings (1/16"), male and female, to fit the tube on a syringe on one end and a disposable needle on the other end
- Disposable syringe needles: 23G and 26G
- Stopwatch
- Micropipettes and tips

SAFETY WARNINGS



CSTFA is considered hazardous by the OSHA Hazard Communication Standard (29 CFR 1910.1200). Causes respiratory tract, eye and skin irritation. May be harmful if swallowed. Do not ingest. Avoid breathing vapour or mist. Use only with adequate ventilation. Avoid contact with

eyes, skin and clothing. Keep container tightly closed. Wash thoroughly after handling.

HiDi-formamide may damage fertility or the unborn child if swallowed. Suspected of causing cancer if swallowed. May cause damage to organs through prolonged or repeated exposure. Do not breathe fumes or spray. Wear protective gloves/protective clothing/eye protection/face

protocols.io

11/23/2020

Citation: Roey Angel, Eva Petrova, Ana Lara (11/23/2020). RNA-Stable Isotope Probing. https://dx.doi.org/10.17504/protocols.io.bpwympfw

protection.

- Storage and waste
- Store below eye level to prevent injuries in case of a spill.
- Dispose of CsTFA and HiDi-formamide in a sealed container as hazardous waste.

ABSTRACT

The following protocol describes how to perform an RNA-Stable Isotope Probing experiment. The scope of this protocol only covers the parts involving separating labelled RNA from unlabelled RNA using ultracentrifugation in a caesium trifluoroacetate density gradient and downstream quantification to evaluate whether the labelling and separation of the RNA were successful. Total RNA should be extracted from an environmental sample or an enrichment culture that was incubated with an isotopically-labelled substrate. Labelling can be of the carbon, oxygen or nitrogen in the RNA (or any combination of the 3). For environmental samples, we recommend extracting RNA using our protocol Total Nucleic Acids Extraction from Soil and purifying it using the Purification of RNA from Crude NA Extract protocol. This protocol is based on the following papers: Whiteley et al. (2007); Dumont et al. (2011); Angel and Conrad (2013). For a comprehensive discussion on how to design a SIP experiment and how to analyse the resulting data, we recommend referring to the recent book on the subject: Stable Isotope Probing: Methods and Protocols, especially chapters: 1-3 and 9-18.

Whiteley AS, Thomson B, Lueders T, Manefield M (2007). RNA stable-isotope probing. Nature protocols.

http://10.1038/nprot.2007.115

Angel R, Conrad R (2013). Elucidating the microbial resuscitation cascade in biological soil crusts following a simulated rain event.. Environmental microbiology.

https://doi.org/10.1111/1462-2920.12140

Dumont MG, Pommerenke B, Casper P, Conrad R (2011). DNA-, rRNA- and mRNA-based stable isotope probing of aerobic methanotrophs in lake sediment.. Environmental microbiology. https://doi.org/10.1111/j.1462-2920.2010.02415.x

BEFORE STARTING

- 1. Prepare all buffers and solutions in advance (see $\underline{\text{Step 1}}$).
- 2. Wipe all surfaces and apparatus with an RNase eliminating solution (e.g. RNAse Away).
- 3. Equilibrate CSTFA to room temperature (about 30-60 min).
- 4. Prepare one 50 ml tube (for up to 8 gradients; depending on the size of the centrifugation tubes) and one ultracentrifugation tube for each gradient.

⊠RNase AWAY™ Surface Decontaminant **Thermo Fisher**

Scientific Catalog #7002PK

Solutions for SIP 1h

1 Prepare the following solutions:

All glassware and plasticware must be clean and free of RNA and RNAse. Glassware can be baked at 8 180 °C for **© 04:00:00** 1.1 Gradient buffer (0.1 M Tris-HCl, 0.1 M KCl, 1 mM EDTA) pH8.0 : ■15.76 g Tris-HCl **■7.455** g KCl **■**0.37224 g EDTA Dissolve the salts in RNase-free water and fill up to $\boxed{1000}$ ml . Filter sterilise (0.1-0.2 μ m). Sigma Catalog #T5941 Aldrich Catalog #P9333 ⊠ Ethylenediaminetetraacetic acid disodium salt dihydrate BioUltra 98.5-101.5% Sigma Aldrich Catalog #E1644-100G Store at § Room temperature 1.2 Molecular-grade ethanol solution ([M]75 % (V/V): **■75** ml Absolute ethanol **■25 ml** RNase-free water Store at 8 -20 °C **Gradient preparation** 1h ■30 µl RNAse-free water Following calibration, the device should read 1.3330 ± 0.0002 nD-TC AR200 Automatic Digital Refractometer Digital Refractometer **(** Reichert 13950000

⋈ protocols.io 6 11/23/2020

For every two gradients, mix the following in a 50 ml tube (assuming 6 ml Ultracrimp, PA centrifugation tubes):

■9.696 ml CsFTA

■2.166 ml Gradient Buffer

& Room temperature

Adjust the volumes if using different-sized ultracentrifugation tubes.

⊠ Cesium Trifluoroacetate (CsTFA) illustra™ Thermo Fisher

Scientific Catalog #45-000-147

Thermo Scientific TUBE PA ULTRACRIMP 6ML PK/50

Ultracentrifugation tubes

Thermo Fisher Scientific 03945 👄

- 4 Mix by inverting several times, pipette 30 μl and measure the density in a refractometer. Make sure the density is:
 1.3702 ± 0.0002 nD-TC. Otherwise, add either CsTFA or GB to correct.
- 5

Add [M] 3.56 % (V/V) HiDi (\blacksquare 422 μ l if the volume was not corrected).

⊠ Hi-Di Formamide **Thermo Fisher**

Scientific Catalog # 4311320

6 Measure the density. Make sure the density is: 1.3725 ± 0.0002 nD-TC.

Due to potential variability between batches, it is recommended to add a slightly lower volume of HiDi at first to avoid exceeding the recommended refractive index.

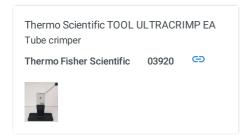
- 7 Transfer approx. **3.8 ml** of the mixture to each centrifugation tube using a micropipette. Make sure the volume reaches only the bottom of the neck.
- 8 Add the RNA sample. For downstream PCR purposes, ca. 200-350 ng is more than enough. Preferably, use a highly concentrated RNA solution to avoid diluting the gradient.

■4 µl RNA (1-8 µl)

[M]150 ng/µl RNA (75-600 ng)

The amount of RNA should not exceed 100 ng per 1 ml of gradient mixture.

- 9 Weigh each tube together with the caps and make sure every opposite pair of tubes weighs no more than 0.1 g apart from each other. Otherwise, adjust the weight by adding gradient mix solution.
- 10 Close the caps (using an appropriate crimper or by hand, depending on the type of tubes).



11 Place the tubes in the rotor, screw only the caps for the positions that contain tubes using a torque wrench up to about 120 in.-lb.

Ultracentrifugation

2d 17h

12



Centrifuge

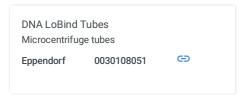
③130000 x g, 20°C, 65:00:00 , (37,900 rpm for the TV-1665 rotor)

Maximum acceleration and deceleration.

Because the density gradient will stabilise over time, centrifuging for a longer time period will make no difference but can be used for timing reasons. However, after the centrifugation has stopped the gradient will slowly diffuse back to its original state. Therefore, the gradients are best fractionated immediately.

Fractionation 1h

13 Prepare a rack filled with 2.0 ml low-binding collection tubes (one per fraction).



- 14 Fill a 30 ml syringe with RNase-free water. Remove any air bubbles.
- 15 Attach a female Luer fitting to one end of a precision pump tube (about 0.5 m long) and a male Luer fitting to the other

end. Attach the syringe to the precision pump tube on the female Luer fitting side. Attach a sterile **23G** needle to the other end of the tube on the male Luer fitting side. Lightly press the syringe piston to get water into the tube and mount the syringe on an automatic syringe pump.

NE-300 Just Infusion™ Syringe Pump Automatic syringe pump New Era Pump Systems, Inc. NE-300 ←

Masterflex L/S® Precision Pump Tubing, Peroxide-Cured Silicone, L/S 14; 25 ft Silicone tube

Masterflex 96400-14 👄

Masterflex Fitting, Polycarbonate, Straight, Female Luer to Low-Profile Semi-Rigid Barb Hose Adapter, 1/16" ID; 25/PK Luer fitting

Masterflex 45501-16

Masterflex Fitting, Polypropylene, Straight, Male Luer Lock to Hose Barb Adapter, 1/16" ID; 25/PK

Luer fitting

Masterflex 30800-16

Disposable needles Sterican® long bevel facet, 30 mm, 0.60 mm, Blue Disposable needles

Sterican X129.1

30s
Set the Rate to 1 ml min⁻¹ and collect fractions in **© 00:00:30** steps. If using a 6 ml tube, this will yield 12 fractions.
Volume should be set to "off" and diameter to "22 mm".

For collecting more or fewer fractions, adjust the speed or collection rate.

G

Citation: Roey Angel, Eva Petrova, Ana Lara (11/23/2020). RNA-Stable Isotope Probing. https://dx.doi.org/10.17504/protocols.io.bpwympfw

Using a different syringe (other than 30 ml) will require adjusting the inner diameter setting on the pump

- 17 Switch the pump on to test the system and also to get rid of air trapped inside the needle and any air bubbles in the tube. Switch the pump back off.
- 18 Stop the ultracentrifuge. Remove the rotor and open the screw-caps. Take the first tube out of the rotor and carefully mount it on a stand with a clamp holder just above the collection tubes.

Make sure the tube stays upright during handling.

19 /

Pierce the ultracentrifugation tube, just below the neck, using the needle attached to to the precision pump tube.



Be careful not to pierce through the other end of the tube!

If the other end of the tube was accidentally pierced, a small amount of petroleum gel can be used to seal the hole.

Take a new, sterile **26G** needle, carefully puncture a hole at the bottom of the ultracentrifugation tube and remove the needle. The tube should not leak at this stage.

Disposable needles Sterican® long bevel facet, 25 mm, 0.45 mm, Brown Disposable needles

Sterican

c718.1

⊕

Open all the collection tubes in the rack and make sure the first tube is positioned just below the bottom hole of the ultracentrifugation tube.

Your set-up should look like this:



The SIP fraction collection set-up ready to start

- 22 Start the pump, as soon as the first drop falls off the ultracentrifugation tube start the stopwatch
- After © 00:00:30 (or your chosen time interval), shift the rack so that the drops will fall into the second collection tube. Continue in a similar fashion until all tubes have been filled. Close the tubes to avoid contamination and label them

24 &

Measure the density of each fraction using the refractometer. Start from the last (the lightest) fraction.

■30 µl of each fraction

The density of the fractions should increase at a linear rate as you progress from the lighter to the heavier fraction. The conversion between refractive index (n) to density (ρ) is (empirically):

$$\rho = 31.495n - 41.439$$

And can be easily determined in the lab by weighing a known volume of several fractions and establishing a calibration curve.

Citation: Roey Angel, Eva Petrova, Ana Lara (11/23/2020). RNA-Stable Isotope Probing. https://dx.doi.org/10.17504/protocols.io.bpwympfw

The gradient should be in the range of 1.77-1.80 g ml⁻¹, assuming a vertical rotor was used (a fixed-angle rotor will yield a steeper gradient, meaning a wider range of densities).

Typically the first and last fractions are discarded because they contain little to no nucleic acids.

RNA precipitation

25



To each tube add $\square 2 \mu I$ of GlycoBlue, [M]10 % (v/v) Na-Acetate ([M]3 Molarity (M)), and [M]250 % (v/v) of absolute ethanol. Assuming □500 μl fractions were collected and □30 μl were spent for determining the density, add $\blacksquare 47~\mu l$ Na-acetate and $\blacksquare 1175~\mu l$ ethanol (absolute) .

⊠GlycoBlue[™] coprecipitant **Thermo Fisher**

Scientific Catalog # AM9515

⊠3M Na-Acetate pH 5.5 **Thermo Fisher**

Scientific Catalog # AM9740

 $GlycoBlue\ is\ particularly\ advantageous\ here\ because\ otherwise, the\ pellet\ is\ completely\ invisible.$

26

Incubate at & -80 °C for (>00:30:00).

27

Centrifuge at (3)14000 rpm, 4°C, 00:30:00.

28



Decant the supernatant, wash once with 175% ethanol, ice-cold, invert the tube several times.

The pellet should be stable at this point and not detach from the tube's wall.

29



Centrifuge at **314000 rpm, 4°C, 00:10:00**.

mprotocols.io 11/23/2020

Remove as much as possible from the supernatant first using a 1 ml tip, spin down the remaining drops in the tube, and 30 remove them with a 100 µl tip. The pellet is unstable at this point. Be careful not to pipette the pellet with the liquid! Leave the tubes open at room temperature for around 5 min (preferably under a flame or in a laminar-flow hood) in order to evaporate the remaining ethanol. Alternatively, the pellets can be dried under a filtered stream of air. © 00:05:00 maximum time for drying The pellets might not be completely dry at this point, but the remaining liquid should be pure water. Resuspend the pellets in **10 μl** RNase-free water or the RNA Storage solution. Scientific Catalog #AM7000 cDNA synthesis 2h 33 For each fraction, prepare the following mixture in a PCR tube: 1. **□10** μl template RNA 2.
3 μl random hexamers ([M]50 Micromolar (μM) diluted 20x in RNase-free water: [M]2.5 Micromolar (µM)) Scientific Catalog #N8080127 34 Incubate the mixture at & 65 °C for © 00:05:00 in a thermocycler and chill at & 4 °C for at least © 00:01:00 . Prepare the following mixture (times the number of fractions) and add $\Box 7 \mu I$ into each tube: 1. **34 μl 5x Reaction buffer** 2. **1 μl 10 mM dNTP mix** 3. **1 μl 0.1 M DTT (optional)** 4.

O.2 μl RNase OUT (40 U/μl; optional) 5. **Q0.2** μl BSA (20 μg/μl) 6. Q0.1 μl SuperScript IV RT (200 U/μl) 7. **■0.5** µl RNase-free water SuperScript™ IV First-Strand Synthesis System Thermo Fisher Scientific Catalog #18091050 **⊠** RNaseOUT™ Recombinant Ribonuclease Inhibitor **Thermo Fisher** Scientific Catalog #10777019

⊠ Bovine Serum Albumin (BSA) Thermo Fisher

Scientific Catalog #B14

⊠dNTP Mix (10 mM each) Thermo Fisher

Scientific Catalog #R0191

⊠USB Dithiothreitol (DTT) 0.1M Solution **Thermo Fisher**

Scientific Catalog #707265ML

- 36 Incubate the mixture in a thermocycler for \bigcirc 00:10:00 at & 23 °C followed by \bigcirc 01:00:00 at & 50 °C and then \bigcirc 00:10:00 at & 80 °C . Chill at & 4 °C .
- 37 Dilute 11 μl cDNA in 14 μl RNase-free water for use as qPCR template. No dilution is required for use as a PCR template.

This dilution step here is required to not exceed the range of detection of the qPCR assay. Higher or lower dilutions might be required depending on the amount of RNA that was loaded on the gradient and the recovery efficiency.

Evaluate the level of enrichment

2h 30m

38



Evaluate the level of isotopic enrichment using a qPCR assay. We recommend

qPCR: Bacterial SSU rRNA 338F-516P-805R
by Roey Angel,
Soil and Water Research Infrastructure

38.1

Name	Туре	Sequence	Target region ¹
BAC338F	Forward	ACT CCT ACG GGA GGC AG	338-354
BAC516P ²	Probe	TGC CAG CAG CCG CGG TAA TA	516-536
BAC805R	Reverse	GAC TAC CAG GGT ATC TAA TC	785-805

- 1. Relative to *E. coli* SSU rRNA gene
- 2. The probe must be dual-labelled either with 5'-6-FAM, 3'-BHQ1 or any other valid combination

38.2 Reagent Final concentration 1 tube (20 μl) plate (20 μl x 100)

PCR H ₂ O		4.6	460
iQ TM Supermix	1x	10	1000
MgCl ₂ (25 mM)	4.0 mM	0.81	80
BSA (20 μg μl ⁻¹)	0.2 μg μl ⁻¹	0.2	20
338F (10 μM)	0.5 μΜ	1.0	100
805R (10 μM)	0.5 μΜ	1.0	100
516P (10 μM)	0.2 μΜ	0.4	40
Template		2	2 x 100

¹ Buffer contains MgCl₂ at final conc. of 3.0 mM

38.3

```
1. § 95 °C for © 00:05:00
2. x 40 {
2.1 § 95 °C for © 00:00:30
2.2 § 62 °C for © 00:00:30 take snapshot
```

39



Plot the cDNA copy numbers against the density of each fraction. It is common to normalise the qPCR results to the highest copy number in the gradient or to the total copy numbers of all the fractions in the gradient.



Expect a peak of unlabelled RNA at around 1.78 g ml^{-1} and a peak of labelled RNA at around 1.82 g ml^{-1}

If the amount of labelled RNA is too small it might not be visible through qPCR. However, it might still be detectable through qSIP or HT-SIP analysis (see e.g. <u>Youngblut et al., 2018, Angel, 2019</u>)

Youngblut ND, Barnett SE, Buckley DH (2018). HTSSIP: An R package for analysis of high throughput sequencing data from nucleic acid stable isotope probing (SIP) experiments.. PloS one. https://doi.org/10.1371/journal.pone.0189616

Angel R (2019). Experimental Setup and Data Analysis
Considerations for DNA- and RNA-SIP Experiments in the Omics
Era.. Methods in molecular biology (Clifton, N.J.).
https://doi.org/10.1007/978-1-4939-9721-3_1