



Version 1

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# Cleaning colonial ascidians V.1

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Works for me

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MATERIALS TEXT

## Material required

- 2 containers with system water
- 2 plastic slide rack
- 1 paintbrush
- 1 single-edge razor blade (used)
- 1 curved-edge scalpel blade

## Optional

- 1 container with filtered artificial seawater
- 1 glass Petri dish with a slide holder
- 1 stereoscope
- 1 pair of plastic tweezers

1. Select the slides to be cleaned.

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1.1 Place them in the first container in a slide rack.

2 Take one slide.

3 1. Using the single-edge razor blade, wipe clean as much surface as possible.

3.1 Make sure to avoid the colonies and the vascular system.

3.2 Do not forget to clean the sides of the slides too.

4 1. Using the wet paintbrush, wipe gently the systems and the tunic of the colonies.

4.1 Try brushing from the center of the colony towards its rim, to avoid detaching it.

4.2 Brush out all the weird-looking tissue.

5 1. Using the curved-edge scalpel blade, ablate the detached or decaying tissue.

5.1 Use a rolling movement to avoid detaching more tissue: press the tip of the blade on the glass, do not slide it, roll the blade to press the edge through the tissue.

6 Place the clean slide into the second container inside another slide rack.

7 If the colony has grown too much or outside of the center of the slide, subclone it (see protocol).

1. **Optional:** If slide needs to be super extra clean:

- 8.1 Place the cleaned slide under the stereoscope.
- 8.2 Inspect the slide.
- 8.3 Use the wet paintbrush to clean the remaining dirt.
- 8.4 Use the plastic tweezers to pluck recalcitrant pieces of foreign tissue.
- 8.5 Let the slide rest in FASW (filtered artificial sea water) for 10 min

- 9 Once all slides are clean, put them back in the system.