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# Granulate formulation protocol

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## ABSTRACT

**Bioformulation** is used in agriculture for various reasons like soil fertility, plant growth promotion, and suppression of phytopathogens. The bacterial inoculants are applied as a formulated product like powder, spray, or pellet. This is a green strategy that is being developed as a less harmful method to protect crops other than pesticides. In the paper published by *Alvarez et al, 2016*, they developed a talc-based powder formulation based on *Bacillus* B25 spores and evaluated some of its characteristics, like shelf life and efficacy against the pathogenic fungus *Fusarium verticillioides*.



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S. A. El-Hassan and S. R. Gowen (2006). Formulation and Delivery of the Bacterial Antagonist *Bacillus subtilis* for Management of Lentil Vascular Wilt Caused by *Fusarium oxysporum* f. sp. *lentis*. *Journal of Phytopathology*, Volume 154, Issue 3.  
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**Fatty acid analysis** is an important means to characterize fats and oils and to determine the total fat content. The result can be used to identify the *Bacillus mycoides* strain in the soil and determine its survival with the granulate formulation.



Friedrich von Wintzingerode, Frederick A. Rainey, Reiner M. Kroppenstedt, Erko Stackebrandt (1997). Identification of environmental strains of *Bacillus mycoides* by fatty acid analysis and species-specific 16S rDNA oligonucleotide probe. *FEMS Microbiology Ecology*, Volume 24, Issue 3.  
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#### Bacterium Inoculum

- 1 Grow a single colony of bacteria in an assay tube with **5 mL** of Luria Broth (LB) medium
- 2 Incubate it in an orbital shaker at 200 rev.min<sup>-1</sup> at **30 °C** for **18:00:00**.
- 3 After bacterial growth, take a **500 mL** Erlenmeyer flask and add **100 mL** of LB medium in it.
- 4 Add **1 mL** of the culture (**1 % (v/v)**) in the flask and incubate at **30 °C** and 200 rev.min<sup>-1</sup> for **24:00:00**, until an optical density of close to 1 is obtained.

#### Spore Production

- 5 Add **100 mL** of Difco Sporulation Medium (DSM; 5g l<sup>-1</sup> peptone, 3g l<sup>-1</sup> yeast extract, 1g l<sup>-1</sup> KCl and 0.12 g l<sup>-1</sup> MgSO<sub>4</sub>·7H<sub>2</sub>O) in a **500 mL** Erlenmeyer flask.
- 6 Sterilize this DSM medium at **121 °C** and 1.5 psi for **00:15:00**.
- 7 Add **1 mL** each of: **1 Molarity (M)** Ca(NO<sub>3</sub>)<sub>2</sub>, **10 Milimolar (mM)** MnCl<sub>2</sub>·4H<sub>2</sub>O and **1 Milimolar (mM)** FeSO<sub>4</sub>
- 8 Inoculate with 1X10<sup>6</sup> c.f.u ml<sup>-1</sup> of the bacterial strain. Keep the culture conditions at **30 °C** and 200 rev.min<sup>-1</sup> for **72:00:00**.

#### Powder Formulation

- 9 Mix talc (which will be the carrier) with, carboxy-methyl-cellulose (CMC; 1% w/w), CaCO<sub>3</sub> (15% w/w) and glucose (0.25% w/w) in powder form.
- 10 Autoclave the mixture at **121 °C** and 15 psi for **00:15:00**.
- 11 Mix the same material with the bacterial spore suspension and dry at **55 °C** for **36:00:00**.
- 12 Using sterile porcelain mortar and pestle pulverize the formulation.
- 13 Pack it in plastic bags and store it at room temperature.

#### CFU determination

- 14 Colony Forming Units (CFU) can be determined by estimating the OD of spore suspension using a tube-reading spectrophotometer adjusted at 1.978 [corresponding to  $8.5 \cdot 10^{10}$  CFU/ml] at 600nm absorbance wavelength
- 15 The formulation will be placed on sterile aluminum foil in pans and air-dried for **24:00:00** with occasional stirring in a laminar airflow cabinet.
- 16 Dried formulations (35% moisture content) of *B. mycoides* will be passed through a 250µm mesh sieve to attain the desired particle size.
- 17 Pack in sterilized polypropylene bags, seal and store at **Room temperature** prior to use.
- 18 Count CFUs to estimate the number of viable propagules of *B. mycoides* using the standard dilution plating method.

#### STD dilution method

- 19 Take three **1 g** aliquots of the dried powder and place in **99 mL** sterile PBST solution (this will include PBS + **0.05 % (v/v)** Tween 20). Stir magnetically at high speed for **00:15:00**. Now dilute this suspension with approximately and take **0.2 mL** of this suspension and plate on Nutrient Agar (NA) media.

#### Fatty acid analysis

- 20 By performing saponification, methylation, and then extracting we can obtain fatty acid methyl esters from wet biomass.

- 21 Next, separate the fatty acid methylester mixtures by using a microbial identification system. Peaks can be automatically integrated, and the Microbial ID will calculate the fatty acid names and percentages.