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A Bioinformatics Analysis workflow for 16S rRNA Amplicon Sequencing data

In 3 collections

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Works for me

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ABSTRACT

Deal with the 16S rRNA amplicon sequencing reads, which sequenced by Ion PGM™ Sequencer with Ion 318™ Chip v2 with a read length of 400bp, and takes the OTU table(the measured abundance profile of detected operational taxonomic units (OTUs)).

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COLLECTIONS ⓘ

**The Female Urinary Microbiota Protocols Collection****Protocols for the female urinary microbiota in relation to the reproductive tract microbiota****Protocols for "The female urinary microbiota in relation to the reproductive tract microbiota."**

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[The Female Urinary Microbiota Protocols Collection](#)[Protocols for the female urinary microbiota in relation to the reproductive tract microbiota](#)[Protocols for "The female urinary microbiota in relation to the reproductive tract microbiota."](#)

ABSTRACT

Deal with the 16S rRNA amplicon sequencing reads, which sequenced by Ion PGM™ Sequencer with Ion

318TMChip v2 with a read length of 400bp, and takes the OTU table(the measured abundance profile of detected operational taxonomic units (OTUs)).

1 ## Convert the bam files to fastq files

```
../bam2fastq-1.1.0/bam2fastq -o vagina/01.trim.data/16_s1.fq ../rawdata/RFEMpxjMAADGAAPGM-47.bam
```

2 ## Delete the primer

```
perl ../iontorrent/re-primers.pl ../v4-v5-primers.fa 2 vagina/01.trim.data/16_s1.fq
```

3 ## Reverse fastq to fasta and quality

```
perl ../fq2qual.pl vagina/01.trim.data/16_s1.fq vagina/01.trim.data/16_s1
```

4 ## Trim the poly-A/T, multi-N and low-quality sequences.

```
mothur "#trim.seqs(fasta= vagina/01.trim.data/16_s1.fa, qfile= vagina/01.trim.data/16_s1.qual,  
minlength=200,maxambig=1,maxhomop=10,qwindowaverage=25,qwindowsize=50,processors=8)"
```

```
mothur "#reverse.seqs(fasta= vagina/01.trim.data/16_s1.trim.fasta, qfile=vagina/01.trim.data/16_s1.trim.qual)"
```

```
perl ../creat.group.pl vagina/01.trim.data/16_s1.trim.rc.fasta 16_s1 >> /vagina/02.processing.data/vagina.groups
```

```
cat vagina/01.trim.data/16_s1.trim.rc.fasta >> vagina/02.processing.data/vagina.trim.fasta
```

```
cat vagina/01.trim.data/16_s1.trim.rc.qual >> vagina/02.processing.data/vagina.trim.qual
```

5 ## Pick the representative sequences.

```
mothur "#unique.seqs(fasta=vagina/02.processing.data/vagina.trim.fasta)"
```

6 ## Assign to the SAILVA

```
mothur "#align.seqs(fasta=vagina/02.processing.data/vagina.trim.unique.fasta,  
reference=../bin/16s/database/silva.bacteria.fasta, processors=8)"
```

```
mothur "#summary.seqs(fasta=vagina/02.processing.data/vagina.trim.unique.align,  
name=vagina/02.processing.data/vagina.trim.names)" >  
vagina/02.processing.data/vagina.trim.unique.align.summary.log
```

7 ## Pick the high-scoring sequences

```
mothur "#screen.seqs(fasta=vagina/02.processing.data/vagina.trim.unique.align,  
name=vagina/02.processing.data/vagina.trim.names,group=vagina/02.processing.data/vagina.groups,  
optimize=start-end, criteria=90,processors=8)"
```

```
mothur "#filter.seqs(fasta=vagina/02.processing.data/vagina.trim.unique.good.align,  
vertical=T,trump=.,processors=8)"
```

```
mothur "#unique.seqs(fasta=vagina/02.processing.data/vagina.trim.unique.good.filter.fasta,  
name=vagina/02.processing.data/vagina.trim.good.names)"
```

```
rm vagina/02.processing.data/vagina.trim.unique.good.align vagina/02.processing.data/vagina.trim.unique.align
```

```
mothur "#summary.seqs(fasta=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.fasta,  
name=vagina/02.processing.data/vagina.trim.unique.good.filter.names)" >
```

vagina/02.processing.data/vagina.trim.unique.good.filter.unique.fasta.summary.log

8 ##Check chimera and delete chimeric sequences

```
mothur "#pre.cluster(fasta=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.fasta,  
name=vagina/02.processing.data/vagina.trim.unique.good.filter.names,group=vagina/02.processing.data/vagina.go  
od.groups,diffs=2)"
```

```
mothur "#chimera.uchime(fasta=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.fasta,  
name=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.names,group=vagina/02.processi  
ng.data/vagina.good.groups,processors=8)"
```

```
mothur "#remove.seqs(  
accnos=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.devono.uchime.accnos,  
fasta=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.fasta,  
name=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.names,group=vagina/02.processi  
ng.data/vagina.good.groups)"
```

```
mothur  
"#summary.seqs(fasta=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.fasta,  
name=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.names)" >  
vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.fasta.summary.log
```

9 ## Classify according to the Greengene reference sequences (gg_13_8_otus)

```
ln -s ../database/greengenes/gg_13_5_99.fasta vagina/02.processing.data/gg_13_5_99.fasta  
ln -s ../database/greengenes/gg_13_5_99.gg.tax vagina/02.processing.data/gg_13_5_99.gg.tax
```

```
mothur "#classify.seqs(fasta=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.fasta,  
template=vagina/02.processing.data/gg_13_5_99.fasta,taxonomy=vagina/02.processing.data/gg_13_5_99.gg.tax,cu  
toff=50,processors=8)"
```

10 ## Delete reads of Mitochondria and Chloroplast

```
mothur "#remove.lineage(  
fasta=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.fasta,  
name=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.names,group=vagina/02.proc  
essing.data/vagina.good.pick.groups,taxonomy=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.pr  
ecluster.pick.gg.wang.taxonomy,taxon=Mitochondria-Cyanobacteria-Chloroplast-unknown)"
```

11 ## Make the otu table

```
python ../bin/pick_otus.py -i vagina/03.otu.data/vagina.final.uclu.fasta -o vagina/03.otu.data
```

```
perl ../bin/OTU_format_trans.pl vagina/03.otu.data/vagina.final.uclu_otus.txt  
vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.pick.names 0.03 >  
vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.pick.an.list
```

```
perl ../bin/tags_taxonomy.stat.v2.pl  
vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.pick.names  
vagina/02.processing.data/vagina.good.pick.pick.groups  
vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.gg.wang.pick.taxonomy.pick  
vagina/03.otu.data/vagina.raw.data.info
```

```
mothur "#make.shared(  
list=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.pick.an.list,  
group=vagina/02.processing.data/vagina.good.pick.pick.groups,label=0.03)"
```

mothur

```
"#classify.otu(list=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.pick.an.list,  
name=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.pick.names,taxonomy=vagina  
/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.gg.wang.pick.taxonomy.pick,label=0.03)"
```

mothur "#summary.single(

```
shared=vagina/02.processing.data/vagina.trim.unique.good.filter.unique.precluster.pick.pick.an.shared,groupmode=  
t)"
```