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Protein extraction form *Aurantiochytrium limacinum* (ATCC MYA-1381)

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1 Works for me dx.doi.org/10.17504/protocols.io.bc7gizjw

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ABSTRACT

Protein extraction protocol for *Aurantiochytrium limacinum* (ATCC MYA-1381; Stramenopile/ Heterokont, Thraustochytrid).

SAFETY WARNINGS

The lysis buffer contains detergents, and therefore it should be handled with care. Always wear gloves to minimize keratin contamination.

BEFORE STARTING

Get a bucket full of ice, and always keep the lysed culture or protein suspension on ice.

Lysis buffer preparation

10m

1 Start by making stocks of the following,

10m

- [M]1 Molarity (M) KCl
- [M]25 Milimolar (mM) MgCl₂
- [M]1 Molarity (M) Tris

Prepare lysis buffer (LyB) by adding the following chemicals (for 10ml),

Chemical	Volume (uL)
KCl (1M)	500
MgCl ₂ (25mM)	1000
Tris (1M)	500
NP40 (100%)	45
Tween (100%)	45
dH ₂ O	8310

Cell harvesting 10m

- 2
 - Pipette out ~3 ml of cell suspension from the test tube or conical flask.
 - Centrifuge the cells at **3000 rpm, 4°C 00:10:00**
 - Discard the supernatant and keep the cell pellet on ice.

10m

Lysis and soluble fraction extraction 20m

- 3
 - Add 1 ml of lysis buffer (LyB) to the pellet.
 - Vortex the mixture for 20 mins on high speed.
 - Centrifuge the suspension at **13500 rpm, 4°C 00:15:00**
 - Collect the supernatant in a separate tube and place it on ice. This is the soluble fraction.

20m

Insoluble fraction extraction 40m

- 4
 - Resuspend the pellet from the above step in [4x LDS sample buffer](#)
 - Add required amount of dH₂O
 - Add 1:10(v:v) 25% β-Mercaptoethanol

40m



Always handle β-Mercaptoethanol in fume hood!

- Heat the mixture at **70 °C** for **00:10:00**
- Centrifuge the mixture at **13500 rpm, 4°C 00:20:00**
- Carefully aspirate the supernatant (insoluble fraction) into a fresh tube and keep it on ice.



Alternatively, one could also use custom buffers (with high concentration of detergent) for insoluble fraction extraction.

Quantification and visualization

- 5 Quantify the protein extract using an appropriate method and the proteins can be visualized on a SDS-PAGE gel stained with Coomassie blue.



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