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RNAlater recipe

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[1]KMU

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ABSTRACT

RNAlater recipe

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Before you start 1d

1 Submerge all your equipment and glassware in **1% bleach to get rid of nuclease**

Reagents 1d

2 **DEPC treated water** 1d

1. Add **1 mL** DEPC in **1 L** ddH2O (**0.1 % (v/v)**); **MIX it well.**
2. Let it sit for **24:00:00** at **25 °C**
3. Autoclave it to deactivate DEPC
4. Store the DEPC treated water at **4 °C**

3 **0.5 M Disodium dihydrate EDTA**:
1. Add **18.61 g** disodium dihydrate EDTA in **100 mL** ddH2O
2. Adjust pH to 8.0 with NaOH while stiring

4 **1 M tri-Sodium Citrate Dihydrate**:
Add **29.4 g** tri-Sodium Citrate Dihydrate in **100 mL** ddH2O, stir to dissolve

5 **1 M sulfuric acid**:
Add **5.33 mL** sulfuric acid in **100 mL** ddH2O, stir to dissolve

Start to make RNAlater

6 Make a mixture of:
40 mL of **0.5 M EDTA** + **25 mL** **1 M tri-Sodium Citrate** + **700 g** **Ammonium Sulfate** + **935 mL** **ddH2O**; stir on a hot plate with low heat until ammonium sulfate dissolved

7 Allow it to cool; adjust the pH of the solution to **pH 5.2** using **1 M sulfuric acid**

> To avoid contamination from the immersed pH meter, adjust pH by pouring 10% of the solution in a new beaker and add base or acid to hit the right pH; then scale it up to the needed amount for the original solution and blindly adjust its pH

8 The final concentration: 25 mM Sodium Citrate, 10 mM EDTA, 70 g ammonium sulfate in 100 ml ddH2O; pH 5.2.

9 Transfer to screw top, **nuclease-free** bottles; label the date of make; store at **4 °C**

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