



Version 3 ▼

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CTAB genomic DNA extraction from Arabidopsis leaf material V.3

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1 Works for me

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ABSTRACT

CTAB-based extraction of genomic DNA from Arabidopsis leaf tissue.

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KEYWORDS

genomic DNA, Arabidopsis

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GUIDELINES

Gives reasonable quality and yield of gDNA. Ideal for DNA in PCRs applications, Dye-terminator Sanger sequencing reactions, and cloning. Can be used for next-generation sequencing applications, however, ensure you perform additional cleaning steps and longer centrifuge times. Run on a 1% agarose gel to ensure you have good quality and clean DNA preparation (and Nanodrop), often these preps yield a substantial portion of sheared nucleotides (likely RNA, so make sure you add RNase A during extraction).

MATERIALS TEXT

MATERIALS

[RNase](#)

A Qiagen Catalog #19101

[EDTA](#) **Contributed by users**

[1.5 mL Eppendorf tubes](#) **Contributed by users**

[water](#) **Contributed by users**

[Ethanol](#) **Contributed by users**

[NaCl](#) **Sigma**

Aldrich Catalog #53014

[Hexadecyltrimethylammonium bromide](#) **Sigma**

Aldrich Catalog #H6269

[Tris-HCl \(Tris-Hydrochloride\),](#)

100gm Promega Catalog #H5121

[2-Propanol \(IsoPropanol\)](#) **Bio Basic**

Inc. Catalog #PC8601.SIZE.4L

[Tris-EDTA, pH](#)

8.0 Ambion Catalog #AM9849

[Chloroform](#) **Sigma Catalog #366919-1L**

[Centrifuge](#) **Contributed by users**

[Water bath set to 65°C](#) **Contributed by users**

Tissue lyser

1/8" steel ball bearings

Vortex

Centrifuge

RNase A (e.g. Promega #A7973 or Sigma #R6148)

SAFETY WARNINGS

Perform chloroform steps in fume hood.

BEFORE STARTING

Ensure you grind your leaf tissue into a fine powder using mortar and pestle or Qiagen tissue lyser (place 1/8" steel ball bearing into tube with tissue sample).

Make sure leaf tissue remains frozen until the addition of CTAB buffer.

Cell lysis 15m

- 1 Prepare 2% CTAB buffer. 15m

A	B
Reagent	[Cf]
hexadecyltrimethylammonium bromide	2% (w/v)
NaCl	1.4 M
EDTA (pH 8)	20 mM
Tris-Cl (pH 8)	100 mM

2% CTAB buffer recipe

- 2 Aliquot required volume of CTAB buffer and heat in water bath at 60 °C for 5-10 minutes immediately before use. 10m

- 3 Add 300 µL / 100 mg leaf tissue of CTAB buffer. 2m

- 4  2m


Add RNase A solution to a concentration 50-100 µg/mL.

- 5 Mix well with a vortex. Invert samples by hand to ensure that all ground tissue is in solution. 5m

- 6 Incubate in water bath at 60 °C for 30 - 60 minutes. Mix tubes periodically by inversion. 1h

- 7 Cool samples to room temperature . 10m

Phase separation 15m

- 8 Add 300 μ L chloroform and mix thoroughly with a vortex or vigorous shaking for 15 seconds. ^{5m}
- 9 Centrifuge samples for 10 minutes at 20,000 rcf. 10m
- 10 Transfer upper aqueous (approx. 200 μ L) phase to clean tube. 5m
- 11  20m
- Repeat steps 9-10 for a cleaner extract.

Precipitation 15m

- 12 Add equal volume of ice-cold 2-propanol and mix by inversion. 2m
- 13 Incubate for 30-60 min @ -20 °C 1h
- 14 Centrifuge for 15 min @ 20,000 rcf. 15m
- 15 Discard supernatant using pipette. 2m

Resuspend DNA 17m

- 16 Wash pellet with 1 mL of 70 % ethanol (mix by inversion). 2m
- 17 Centrifuge samples @ 9,200 rcf for 5 min. 5m

- 18 Remove as much ethanol as possible using a pipette, then allow pellet to air dry for 5 minutes.^{10m}
- 19 Resuspend gDNA in nuclease-free H₂O or low EDTA TE buffer (10 mM Tris-Cl pH 8, 0.1 mM EDTA).
- 20 Test yield and purity of samples using a Nanodrop and running samples on a 1 % agarose gel.