

MAR 14, 2024

OPEN ACCESS



DOI:

dx.doi.org/10.17504/protocols.io. dm6gp34p1vzp/v1

Protocol Citation: Ramhari Kumbhar, Hanseok Ko, Valina L. Dawson, Ted Dawson, Xiaobo Mao 2024. Stereotaxic α-Syn PFF injection into mouse striatum. protocols.io https://dx.doi.org/10.17504/protoc ols.io.dm6gp34p1vzp/v1

License: This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working We use this protocol and it's working

Stereotaxic α-Syn PFF injection into mouse striatum

Forked from Stereotaxic α-Syn PFF injection into mouse striatum

Ramhari Kumbhar^{1,2,3}. Hanseok Ko^{1,2,3}. Valina L. Dawson^{4,5,3,6,7,8}. Ted Dawson^{4,5,3,6,7,8,9}. Xiaobo Mao^{4,5,3,6,7,8,9}

¹Neuroregeneration and Stem Cell Programs, Institute for Cell Engineering, The Johns Hopkins University School of Medicine, Baltimore, Maryland, USA;

²Department of Neurology, The Johns Hopkins University School of Medicine, Baltimore, Maryland, USA;

³Aligning Science Across Parkinson's (ASAP) Collaborative Research Network, Chevy Chase, MD 20815, USA;

⁴Neuroregeneration and Stem Cell Programs, Institute for Cell Engineering, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA;

⁵Department of Neurology, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA;

⁶Adrienne Helis Malvin Medical Research Foundation, New Orleans, LA 70130-2685,

⁷Department of Physiology, Johns Hopkins University School of Medicine, Baltimore, MD 21205 USA;

⁸Solomon H. Snyder Department of Neuroscience, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA;

⁹Institute for NanoBioTechnology, Johns Hopkins University, Baltimore, MD, USA

ASAP Collaborative Research Network

Kaplitt Protocols



Eileen Ruth Torres Weill Cornell Medicine

ABSTRACT

For stereotaxic injection of α -Syn PFF into the mouse striatum using 2-3 month old mice. This forked protocol has slight variations in anesthesia solution and injection coordinations.

protocols.io

Created: Mar 14, 2024

Last Modified: Mar 14, 2024

PROTOCOL integer ID: 96713

Keywords: ASAPCRN

Funders Acknowledgement:

Aligning Science for Parkinson's

Grant ID: 020608

MATERIALS

- Anesthetic- isoflurane OR pentobarbital (45 mg/kg) diluted 1:1 with 0.9% NaCl OR ketamine cocktail (ketamine 100mg/kg; xylazine 20mg/kg; acepromazine 3mg/kg i.p. or s.c.)
- Small electric shaver
- scalpel
- 26 gauge needle and Hamilton syringe
- minisyringe pump (Model BS-800, Braintree Scientific)
- sterile wound healing clips (7 mm stainless steel wound clip; Reflex 7 skin closure system) or sutures
- bupivacaine (0.4-0.8 ml/kg of a 0.25% solution) OR 0.5% xylocaine 1.4 ml/kg, injected sc, injected) OR 0.5%
 xylocaine

BEFORE START INSTRUCTIONS

Personnel should use aseptic techniques and conduct the surgery using sterile gloves in a sterile environment. Surgical instruments should be sterilized before surgeries using an autoclave and during the procedure using glass bead sterilizer.

- 1 Anesthetize mice with a ketamine cocktail (ketamine 100mg/kg; xylazine 10mg/kg; i.p. or s.c.).
- 2 Shave the animals' skulls.
- 3 Create a 1.5 cm incision in the scalp after cleaning the scalp with alcohol or betadine swabs.
- 4 Retract the underlying fascia will and drill a hole in the skull.

protocols.io

- 5 Insert a 26 gauge needle attached to a 2uL Hamilton syringe containing recombinant α-syn PFF (5 μg/2 μL) or an equivalent volume of PBS into the striatum unilaterally (right hemisphere) at coordinates mediolateral 2.0 mm from bregma, anteroposterior -0.2 mm, and dorsoventral 2.8mm. at a rate of 0.4 ul per minute using a minisyringe pump (Model BS-800, Braintree Scientific).
- **6**Leave the needle in place for 5 minutes before withdrawing the needle.
- 7
 Close the incision using sterile wound healing clips (7 mm stainless steel wound clip; Reflex 7 skin closure system) or sutures.
- 8 Immediately after the stereotaxic procedure, treat the stapled wound site with bupivacaine (0.4-0.8 ml/kg of a 0.25% solution, injected sc) or 0.5% xylocaine 1.4 ml/kg, injected sc, injected) or 0.5% xylocaine when necessary or as determined by animal use policy.