




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# SOP for Spatial N-glycomics

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## ABSTRACT

This protocol describes the procedure to obtain high quality MALDI mass spectrometry images of N-linked glycans from formalin-fixed paraffin embedded tissue.

## PROTOCOL CITATION

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## OWNERSHIP HISTORY

Sep 28, 2022  bglarsonSep 29, 2022  Chris Anderton

## PROTOCOL INTEGER ID

70629

## Scope

- 1 This protocol describes the procedure to obtain high quality MALDI mass spectrometry images of N-linked glycans from formalin-fixed paraffin embedded tissue.

## Health and Safety

- 2 Wear nitrile gloves and safety glasses. Follow standard laboratory safety procedures.

## Equipment

- 3 Equipment Required:

- 3.1 8 Coplin jars

- 3.2



5 slides mailer with side opening

- 3.3 Vegetable steamer (antigen retrieval device): we use AROMA 8-Cup Cool-Touch Rice Cooker
- 3.4 Flatbed color scanner (we used EPSON PERFECTION V500 PHOTO)
- 3.5 Zeiss 710 laser scanning confocal (LSC) microscope equipped with a 20X EC Plan NEOFLUAR objective
- 3.6 Syringe pump capable of 25  $\mu$ l/min

3.7 Pump capable of 100  $\mu$ l/min

3.8 TM-Sprayer (HTXimaging)

3.9 Home-designed chamber for incubation:  
Note: This consists of a rubber gasket sealed glass container (the jar with lid, KORKEN, IKEA of Sweden. Diameter 11 cm; high 10.5 cm, volume 0.5 L) in which a 50 ml glass beaker and a set of weights is placed. The weights are required to keep the glass beaker from floating.

## Chemicals & Enzymes

### 4 Chemicals & Enzymes:

4.1 Xylenes

4.2 200 proof ethanol

4.3 Water

4.4 Citraconic anhydrous buffer

4.5 1 M HCl

4.6 PNGase F enzyme (PRIME, 50 U/ $\mu$ g (lyophilized))

## Procedure

- 5 Formalin fixed tissue should be sectioned at 7  $\mu\text{m}$  and mounted on indium-tin-oxide (ITO) glass slides (25 mm x 75 mm).

## Dewax Slides

- 5.1 Heat slides at 60°C for one hour.
- 5.2 Remove and cool to room temperature, usually 5 minutes.

## Dewax Washes

- 5.3 Xylenes 3 minutes, repeating a total of two times.
- 5.4 100% ethanol 1 minute, repeating a total of two times.
- 5.5 95% ethanol 1 minute
- 5.6 70% ethanol 1 minute
- 5.7 Distilled water 3 minutes, repeating a total of two times.
- 5.8 Dry slides in desiccator 5 minutes
- 5.9 Scan each slide, minus the surrounding sample holder, at a minimum of 1200 ppi resolution using flatbed scanner. This will be needed for image registration

in FlexImaging during imaging acquisition. Samples for higher resolution will require a higher resolution scanned image. For example, images acquired with a  $\leq 50 \mu\text{m}$  step size require a 2400 dpi scanned image.

#### Obtaining Autofluorescence (AF) and high-resolution brightfield images

- 5.10 Turn on main switch, laser, X-cite elimination unit and components.
- 5.11 Mount the slide in the slide holder and place it in the LSC microscope
- 5.12 Run the Zeiss Zen lite software
- 5.13 In "Locate" Tool Area find your tissue and focus 20X objective on it.
- 5.14 Click on "Live" button and fine adjust focusing
- 5.15 Open previously acquired image of KPMP sample, go in "acquisition" Tool Area and select "reuse" scanning parameters: 405nm and nm excitation laser, tile scan (size of tile:  $303.6 \mu\text{m} \times 303.6 \mu\text{m}$ ), pinhole size: 4.63 airy units.
- 5.16 Use enough tiles to capture entire tissue, and use minimum resolution of 512x512 to acquire image
- 5.17 "Start scanning" and save image after scanning is done.

#### Antigen Retrieval Prep

- 6 Prepare Citraconic buffer:

25 ml distilled water or HPLC grade water into a 50 ml falcon tube

6.1

6.2 Add 25 µl of citraconic buffer to the water

6.3 Add 24 µl of 1 M HCl

6.4 Agitate tube after capping.

6.5 Add water to a total of 50 ml

6.6 Agitate tube to mix

6.7 Check that pH is around  $3.0 \pm 0.5$  by spotting 2 µl of the prepared buffer onto a pH strip

7 Heat slides in vegetable steamer:

7.1 Preheat the vegetable steamer to generate steam by pressing the “cook” switch prior to retrieval procedure (example, preheating takes ~15 min)

7.2 Add ~ 10 ml of the buffer to a 5-slide mailer with side opening

7.3 Place no more than 3 slides per 5-slide mailer with side opening. Slides should be placed with tissue facing outward to the solution in positions 1 and 5, NOT facing the slide mailer walls. Position 3 may face either way

- 7.4 Completely fill the slide mailer the rest of the way with buffer
- 7.5 If the mailer has no holes punched in the lid, only snap close one corner of the mailer
- 7.6 Place the mailer in the corner of the vegetable steamer
- 7.7 Maintain “cook” option for 30 minutes

## 8 Cool the slides after antigen retrieval:

- 8.1 Remove mailer and place in a tub with cool water from the faucet. Water should not go over the top of the mailer
- 8.2 Allow to cool for 5 minutes
- 8.3 Remove half the buffer from the mailer and replace with distilled water.
- 8.4 Allow to cool 5 minutes on countertop
- 8.5 Repeat removal of half the buffer two more times, each time with 5 minutes of cooling
- 8.6 Complete by rinsing in 100% distilled water

- 8.7 Dry the slides 5 minutes in the desiccator
- 8.8 Check to ensure scanning of the slides has been performed
- 8.9 For scanning, scan one slide each at 1200 dpi

#### Application of PNGase F Solution

### 9 Prepare PNGase F solution:

- 9.1 Prepare 0.1 µg/µl PNGase F in water: resuspend lyophilized enzyme in 1 mL water
- 9.2 Ensure that enough solution is prepared, e.g. three full slides takes approximately 1 ml of solution, spraying at 25 µl/min

### 10 Spray the PNGase F solution:

- 10.1 Using the syringe dedicated to PNGase F enzyme solution, rinse the syringe with water by screwing in the needle tip, filling with 3 or more ml of water, and aspirating into waste
- 10.2 Fill with PNGase F solution ensuring that there are no bubbles in syringe. Tip: After loading all the PNGase F solution required, pull a small volume of air into the syringe. Gently dispense the syringe until the large air bubble is gone.
- 10.3 Remove the needle tip and fasten the syringe to the TM-Sprayer line used for PNGaseF. Place the syringe onto the red syringe pump. Check that the syringe head is snug against the dispense head of the syringe pump. Ensure that the



diameter is set appropriate to the syringe and the rate is set at 25 µl/min. Do not start the pump at this time.

- 10.4 Place the samples in the TM-Sprayer tray, fastening them with tape.
- 10.5 Set up TM-Sprayer, referring to the guide for the TM-Sprayer. Temperature should be set to 45°C with 15 passes, velocity of 1200, and 3 mm offset.
- 10.6 Pressure reading on the front of the TM-Sprayer should be 10 psi.
- 10.7 Start the syringe pump.
- 10.8 Use a dummy slide to check the TM-Sprayer nozzle for spraying of solution. It generally takes about 1-3 minutes (100 µL) to start spraying.
- 10.9 Once moisture is detected on the dummy slide, press Start on the TM-Sprayer. PNGase F solution will be applied in a thin layer onto target tissue.

## 11 Incubation PNGase F digest:

*To prevent liquid from evaporating too fast and the enzyme from becoming inactive, a wet atmosphere is maintained by placing the ITO slide into a sealed incubation chamber filled with 150 ml saturated  $K_2SO_4$  solution and pre-incubated at 40 °C.*

- 11.1 After application of PNGase F onto the slide, place it on top of a 50 ml glass beaker in the incubation chamber.
- 11.2 Incubate 2 hours
- 11.3 After incubation, remove the slide from the incubation chamber and let dry in the desiccator (15 min).

- 11.4 Store the slide in a 5-slide mailer to protect the released glycans. If matrix cannot be sprayed the same day, store at -20°C. It is preferred to immediately spray matrix onto the slide.

#### Application of Matrix

### 12 Prepare the CHCA Matrix:

- 12.1 Prepare CHCA matrix at 7 mg/ml in 50% acetonitrile/0.1% TFA. Add 0.042 g CHCA to 6 ml 50% acetonitrile/0.1% TFA. Prepare fresh each time in a 50 ml falcon tube.
- 12.2 Vortex briefly and sonicate 5 minutes.
- 12.3 Small chunks may remain in the bottom of the falcon tube. Make sure that there are not loaded into the TM-Sprayer loop as they will clog components of the TM-Sprayer.
- 12.4 Filter CHCA solution using Millex (Millipore) 0.2 µm syringe filter.

### 13 Spray the CHCA Matrix:

- 13.1 Fill the glass-5ml syringe with CHCA solution ensuring that there are no bubbles in syringe. Tip: After loading all the solution required, pull a small volume of air into the syringe. Gently dispense the syringe until the large air bubble is gone.
- 13.2 Remove the needle tip and fasten the syringe to the TM-Sprayer line going to the 6-port valve.
- 13.3 Move the switch to "LOAD" and steadily depress the syringe until all the sample is loaded. Note: Do not load air bubbles or undissolved matrix.

- 13.4 Make sure that the pump is flowing at 0.1 ml/minute. Pump pressure should be 30-40 psi when flowing at 0.1 ml/min.
- 13.5 Place the sample in the TM-Sprayer tray, fasten them with tape.
- 13.6 Set up TM-Sprayer referring to the guide for the TM-Sprayer. Temperature should be set to 80°C with 10 passes, velocity of 1300, and 2.5 mm offset.
- 13.7 Pressure reading on the front of the TM-Sprayer should be 10 psi.
- 13.8 Move the 6-port valve switch to "Spray".
- 13.9 Use a dummy slide to check the TM-Sprayer nozzle for spraying of solution. It generally takes about one minute to start spraying matrix.
- 13.10 Once matrix is detected as an opaque solution on the dummy slide, press Start on the TM-Sprayer. CHCA solution will be applied in a thin layer onto target tissue.
- 13.11 When finished, matrix coated slides may be imaged immediately or stored in a desiccator.

#### MALDI Imaging MS Acquisition

- 14 Put the slide in the MALDI holder and load it in the 15 T FTCIR instrument
- 15 Load the method for N-glycan analysis Parameters of the method: m/z range 800-4,000; R=512k, laser power=28-32%, laser focus=minimum, laser shots=200, frequency=2,000
- 16 Teach plate

17 Select measurement region in FlexImaging

18 Run acquisition through FlexImaging

#### MALDI Imaging Analysis

19

20 Load data to SCILS lab

21 Convert data to imZML

22 Upload imZML files to METASPACE

22.1 Under Annotation settings select: "NGlycDB" for Metabolite database

22.2 Under Annotation settings select: "+Na" as Adducts