

Feb 14, 2022

# Prokaryotes 16S-V4V5 rRNA Metabarcoding PCR protocol for NGS Illumina sequencing

Sarah Romac<sup>1</sup><sup>1</sup>UMR7144-Roscoff Marine Station

1

[dx.doi.org/10.17504/protocols.io.bzwwp7fe](https://dx.doi.org/10.17504/protocols.io.bzwwp7fe)

Ecology of Marine Plankton (ECOMAP) team - Roscoff AD2M

Sarah Romac  
UMR7144-Roscoff Marine Station

Nowadays metabarcoding approaches allow to explore the diversity of different communities using next-generation sequencing (NGS).

Here we describe the 16S-V4V5 DNA amplification method applied for prokaryote metabarcoding analyses using Illumina Miseq technology. This protocol has been used in many projects studying prokaryotic diversity (TARA-OCEANS 2009-2013, TARA-PACIFIC 2016-2018), and prokaryote monitoring projects (ROSCOFF ASTAN 2009-ongoing).

We developed the flowchart for 2 different sequencing platforms : Fasteris-Gene Support SA (Plan-Les-Ouates, Swiss) and GeT-PlaGE (Toulouse, France).

DOI

[dx.doi.org/10.17504/protocols.io.bzwwp7fe](https://dx.doi.org/10.17504/protocols.io.bzwwp7fe)

Sarah Romac 2022. Prokaryotes 16S-V4V5 rRNA Metabarcoding PCR protocol for NGS Illumina sequencing. **protocols.io**  
<https://dx.doi.org/10.17504/protocols.io.bzwwp7fe>



PCR, NGS, metabarcoding, prokaryotes, 16S-V4V5

protocol ,

Nov 09, 2021

Feb 14, 2022

54966

As metabarcoding is very sensitive to contaminations by exogen DNA, please respect some conditions :

- always wear a labcoat, and clean nitrile gloves;
- separate the work area for prePCR and postPCR manipulations.
- do your PCR under a PCR hood.

Specific Equipment (more details in the concerned steps) :

- PCR hood equipped with UV light and HEPA filter ;
- Thermocycler.
- Qubit 4 Fluorometer (Invitrogen) ;
- Fluorometer Plate reader ;
- Gel Tray Caster and Imager .

Optional Equipment :

- 2100 Bioanalyzer Instrument (Agilent)

Supplies :

Sterile microtubes 1,5mL

Semi-skirted PCR plates 0.2mL, 96 wells (like AB-0900) and thermoresistant seals.

Filter tips

Reagents and kis are mentioned in the protocol in the concerned steps.

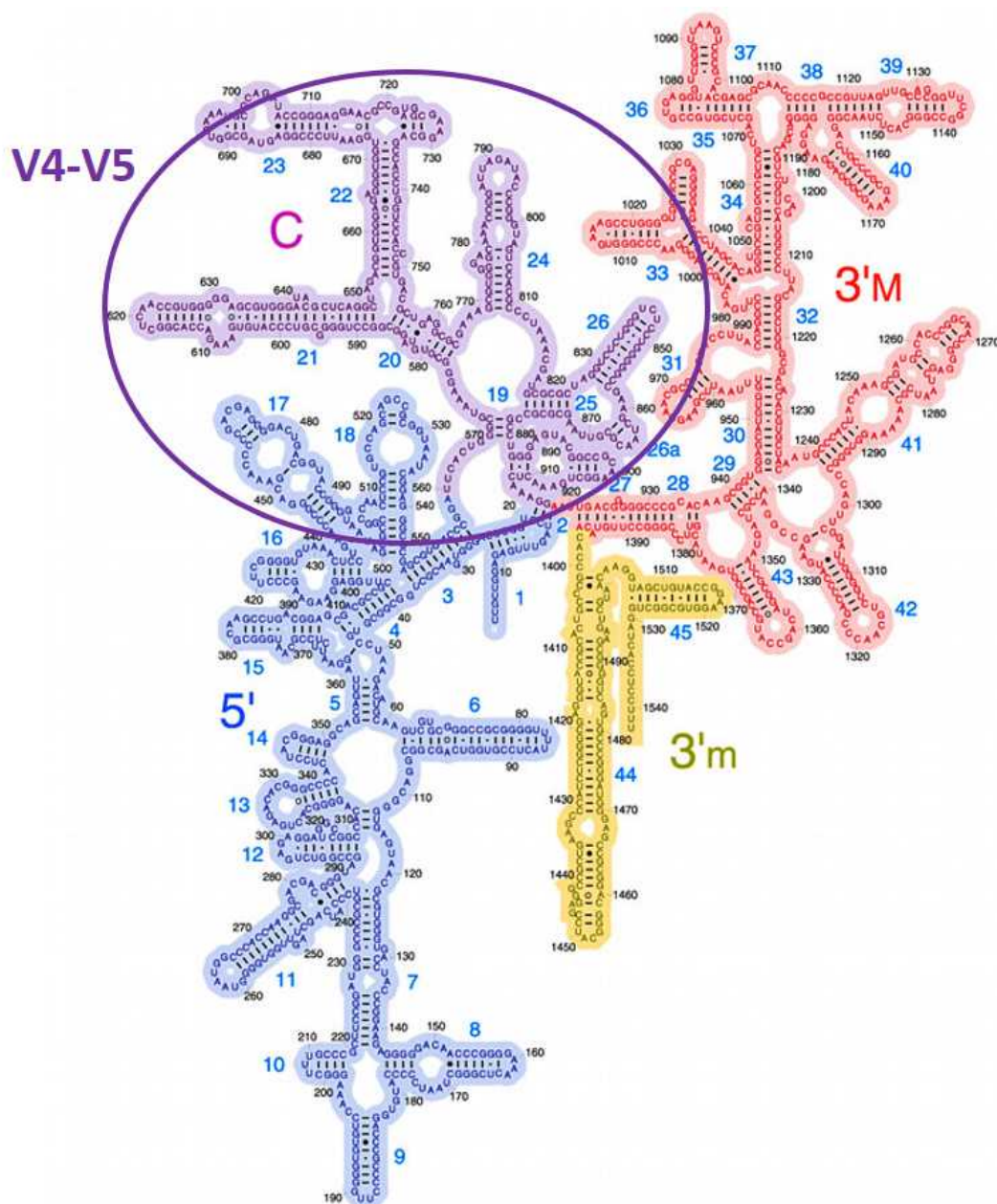
Do aliquots of 1 mL of sterile milliQ water.

Before starting, place all the supplies and sterile milliQ Water needed for the PCR under the PCR hood and switch on the UV light for at least 20 min.

## Tagged Primer Design and preparation

- 1 We use the prokaryotic 16SV4V5 primer set 515fY- 926r from Parada et al. 2016.

Primer	Target	Sense	Sequence 5'-3'	Length bp	Tm °C	Amplicon length bp	Source
515f-Y	16S_Prokaryotes	Forward	GTGYCAGCMGCCGCGGTAA	19	63	411	Parada et al. 2016
926r	16S_Prokaryotes	Reverse	CCGYCAATTYMTTTRAGTTT	20	63		



Location of the V4V5 part on the 16S structure.

2

Step 2 includes a Step case.

**Tagged-Primer Design for Fasteris platform**

**Tagged-Primer Design for GeT-PLaGE platform**

step case

### Tagged-Primer Design for Fasteris platform

Amplicons from each DNA sample are all pooled in a single microtube. Each pool of



amplicons will be considered as a "library sample " or called **"Pooled Amplicons"** and loaded on a Miseq 2x250 run.

To allow the latter separation of each sample in the Pooled Amplicon sample, each DNA is amplified using forward tagged primer built with a structure 5'-NNNN-MID-forwardprimer'. The reverse primer is not modified.

The tag, or Multiplex Identifier (MID) is a unique short sequence of 7 or 8 bases compatible with the forward primer V4f and generated using the matrix oligoTag program (Coissac et al. 2012). We also added 4 N at the 5' extremity of the forward primer to help MID sequence conservation during the cluster synthesis step on the Flowcell.

Primer	Sense	Tagged primer construction	Sequence 5'-3'	Amplicon Length (pb)
515f-Y-Fasteris	Forward	5' -NNNN-MID-primer - 3'	NNNNMID###GTGYCAGCMGCCGCGGTAA	423
926r-Fasteris	Reverse	5' - primer - 3'	CCGYCAATTYMTTTRAGTTT	

Figure 2 : Fasteris tagged-primer description.

Primer Name	MIDnumber	MIDsequence	Primer Name	MIDnumber	MIDsequence	Primer Name	MIDnumber	MIDsequence
515f_M8R0001	M8R0001	AACAACAA	515f_M8R0051	M8R0051	TATCCTCA	515f_M8R0095	M8R0095	GCGCTATG
515f_M8R0004	M8R0004	CCAGGTGA	515f_M8R0052	M8R0052	CACITCAA	515f_M8R0096	M8R0096	GCGCTGAA
515f_M8R0005	M8R0005	AGCATGCG	515f_M8R0053	M8R0053	AATTAAGT	515f_M8R0097	M8R0097	AATGGTTA
515f_M8R0006	M8R0006	AATGGAGG	515f_M8R0055	M8R0055	TGATGATT	515f_M8R0098	M8R0098	TTGCTTAA
515f_M8R0007	M8R0007	TTCTCTCG	515f_M8R0056	M8R0056	TCGATCGG	515f_M8R0099	M8R0099	AACCATTA
515f_M8R0008	M8R0008	CTTCTTCA	515f_M8R0057	M8R0057	GGCTGATG	515f_M8R0100	M8R0100	CTTGTGTA
515f_M8R0009	M8R0009	CGCAACAG	515f_M8R0059	M8R0059	CTATCGCG	515f_M8R0101	M8R0101	TCGCCTGT
515f_M8R0010	M8R0010	AACAATGG	515f_M8R0061	M8R0061	TATCGATA	515f_M8R0102	M8R0102	CTATTCAT
515f_M8R0011	M8R0011	GGCGTTAT	515f_M8R0064	M8R0064	GGCTGTCT	515f_M8R0103	M8R0103	TGTTGTTA
515f_M8R0012	M8R0012	GAGTACTA	515f_M8R0065	M8R0065	AACATATG	515f_M8R0105	M8R0105	AACCGAGT
515f_M8R0015	M8R0015	TATCACAT	515f_M8R0066	M8R0066	ATGTGCCG	515f_M8R0106	M8R0106	AGACTAAT
515f_M8R0016	M8R0016	GTCGCTGT	515f_M8R0067	M8R0067	TTGCGCCT	515f_M8R0107	M8R0107	CAGAGAAG
515f_M8R0017	M8R0017	CCGAGATT	515f_M8R0069	M8R0069	AATAGTAG	515f_M8R0108	M8R0108	TGCAATCG
515f_M8R0018	M8R0018	GAGACGAG	515f_M8R0070	M8R0070	ACGTACAG	515f_M8R0109	M8R0109	ACGTGTTA
515f_M8R0019	M8R0019	CGCAAGCA	515f_M8R0071	M8R0071	AGCCTCTT	515f_M8R0110	M8R0110	TATGAATG
515f_M8R0020	M8R0020	CCAGTCAG	515f_M8R0072	M8R0072	CGGTCACA	515f_M8R0111	M8R0111	ATAGAACT
515f_M8R0024	M8R0024	CTTGACAG	515f_M8R0073	M8R0073	TGATGTCG	515f_M8R0112	M8R0112	TTAACCAG
515f_M8R0025	M8R0025	AACACCGT	515f_M8R0074	M8R0074	CTATGACA	515f_M8R0113	M8R0113	TAATTGTA
515f_M8R0026	M8R0026	ATGTATAA	515f_M8R0075	M8R0075	TATCGGCT	515f_M8R0114	M8R0114	CGCGATGA
515f_M8R0028	M8R0028	TATCAGGA	515f_M8R0076	M8R0076	AGCCTTAA	515f_M8R0115	M8R0115	ATTAATAT
515f_M8R0029	M8R0029	GCGACAAT	515f_M8R0079	M8R0079	GTCTATGA	515f_M8R0116	M8R0116	CAGAGCCT
515f_M8R0030	M8R0030	GTCGGTCA	515f_M8R0080	M8R0080	AACATTAT	515f_M8R0118	M8R0118	GCGGCTTA
515f_M8R0031	M8R0031	AACGAGTG	515f_M8R0082	M8R0082	GGCTTGAA	515f_M8R0119	M8R0119	TATGTACA
515f_M8R0032	M8R0032	AACACGTA	515f_M8R0083	M8R0083	ATACGTCA	515f_M8R0123	M8R0123	TATGATAA
515f_M8R0034	M8R0034	GTGAATTA	515f_M8R0084	M8R0084	AACCAACG	515f_M8R0126	M8R0126	GAACATGA
515f_M8R0035	M8R0035	TGATCCTA	515f_M8R0085	M8R0085	TAATGCGT	515f_M8R0127	M8R0127	AACCTCAG
515f_M8R0038	M8R0038	ATACATGG	515f_M8R0086	M8R0086	TCGCAGTA	515f_M8R0128	M8R0128	TATGCAAT
515f_M8R0041	M8R0041	TATCCAGG	515f_M8R0087	M8R0087	ACAGCATA	515f_M8R0129	M8R0129	TGCACTGA
515f_M8R0042	M8R0042	TTGCCACG	515f_M8R0088	M8R0088	GGTAACCT	515f_M8R0130	M8R0130	ACAACCGA
515f_M8R0043	M8R0043	AGCCATGT	515f_M8R0090	M8R0090	TGATTGAT	515f_M8R0131	M8R0131	TTGGCAGT
515f_M8R0046	M8R0046	TTCTCAT	515f_M8R0091	M8R0091	AACCAGAT	515f_M8R0133	M8R0133	CTATACTG
515f_M8R0047	M8R0047	AGCCGATA	515f_M8R0093	M8R0093	GTCTCGCA	515f_M8R0134	M8R0134	CGCTAAGG
515f_M8R0049	M8R0049	CATGGCCA	515f_M8R0094	M8R0094	ATGTCTCA	515f_M8R0135	M8R0135	ACTAAGAG
515f_M8R0050	M8R0050	AACAGGAG						

Figure 3 : MID list adapted for 16S-V4V5 primer set.

- 3 Lyophilized Tagged-primers are obtained at Eurogentec, using the RP-Cartridge Gold purification.

Work always under the PCR hood.

- 3.1 Elute dried primers at 100  $\mu$ M with TE 1X sterile buffer under a PCR hood.
- 3.2 Primer dissolution is done for 15 min at room temperature under the hood. Short vortex and spin.
- 3.3 Primer working solutions are then prepared at 10  $\mu$ M with sterile milliQ water molecular grade.  
  
For each MID-primer, add 10  $\mu$ L of 100  $\mu$ M of Stock primer to 90  $\mu$ L in a 1.5mL sterile microtube correctly labeled (MID###, concentration, date, operator).
- 3.4 Stock primers and primer working solutions are stored at -20°C.

## PCR

- 4 PCR reactions are performed using the Taq polymerase **GoTaq G2 Flexi polymerase (Promega, Cat No M7806)**.

Keep the same annealing temperature as the one is used with the usual 16S-V4V5 primer set.

First test each of your tagged-primer sets on a positive and a negative control following steps 5, 6, 7 and 8.

Then you can perform DNA sample amplification following next steps.

- 5 **PCR plate's plan :**

Each DNA sample (DNA1, DNA2, DNA3...) will be amplified with its own tagged-MID-primer (515fY-M8R001, 515fY-M8R004, 515fY-M8R005...). **So there will be as many PCR mix preparations as there are DNA samples to amplify.**

In order to get enough material (50 ng), triplicate the PCR reactions on each DNA sample (you will pool them after).

One positive control and one negative control will be added for each PCR mix preparation.

PCR reactions are prepared on a semi-skirted 96-wells PCR plate (like AB-0900 PCR plate).

	1	2	3	4	5	6	...
A	DNA1-M8R001	DNA1-M8R001	DNA1-M8R001	Control+_M8R001	Tneg_M8R001	...	...
B	DNA2-M8R004	DNA2-M8R004	DNA2-M8R004	Control+_M8R004	Tneg_M8R004	...	...
C	DNA3-M8R005	DNA3-M8R005	DNA3-M8R005	Control+_M8R005	Tneg_M8R005	...	...
D	DNA4-M8R006	DNA4-M8R006	DNA4-M8R006	Control+_M8R006	Tneg_M8R006	...	...
E	...	...	...	...	...	...	...
F	...	...	...	...	...	...	...
G	...	...	...	...	...	...	...
H	...	...	...	...	...	...	...

## 6 PCR Mix preparation :

### 6.1 Prepare the Master Mix :

Designation	Final concentration	Volume (µL)
Buffer Go Taq 5x	1x	6,12
MgCl2 25 mM	2 mM	2,45
dNTPs mix 10 mM	0,1 mM	0,3
Primer F 10 µM	0,2 µM	0,62
Primer R 10 µM	0,2 µM	0,62
Go Taq Flexi 5u/µL	1,5 u	0,3
H2O milliQ sterile		17,59
ADN 5ng/µL	10 ng	2
Vtotal µL		30

### 6.2 If you work on environmental sample :

Dispense 28 µL of each tagged-MID-primer PCR mix preparations per well under the PCR hood, and then add 2 µL of the DNA template on the bench.

### If you work on strain sample :

Dispense 26  $\mu\text{L}$  of each tagged-MID-primer PCR mix preparations per well under the PCR hood (so reduce the volume of H<sub>2</sub>O milliQ in the Master Mix), and then add 4  $\mu\text{L}$  of the DNA template on the bench to get 5-10ng in the MasterMix.

## 7 PCR Programm :

Initial Denaturation	10min @ 95°C	
Denaturation	30sec @ 95°C	x 30 cycles
Annealing	1min @ 50°C	
Elongation	1min @ 72°C	
Final Elongation	10min @ 72°C	
Storage	∞ @ 4°C	

In order to reduce the artificial building of chimeras during the PCR process, the # of cycles must be reduced to a minimum : 25, max 30 (chimeras formation mainly during the plateau-phase of the PCR reaction).

## 8 Check the quality of all the PCR products on an 1.2 % agarose gel.

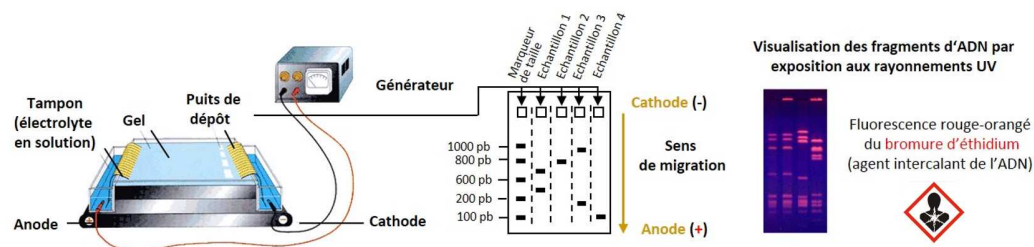


Figure 4 : Principe of DNA electropheris step (in French).



**Be very cautious with Ethidium Bromide manipulation!**

[FDS Ethidium Bromide.pdf](#)



## 8.1 Prepare 1,2 % agarose gel in TAE 0.5x buffer :

- In a Becher, put 1.2g agarose (Interchim, ref 31272L ) in 100mL TAE 0.5x buffer (TAE prediluted in milliQ water from TAE 10x-Thermofisher Scientific, ref 15558042).
- Heat under total dissolution of the agarose powder (you can use a microwave or a stirrer plate).
- Add one drop of Ethidium Bromide (Eurobio, ref GEPBET02AF).
- Prepare casting tray with combs according to your number of PCR products to check. (Biorad, ref 1704484)

 [Biorad\\_Gel Caster.pdf](#)

- Poor gel in the casting tray and check there are no bubbles.
- Let solidify for 20 min.

## 8.2 Sample loading and electrophoresis conditions :

Buffer Go Taq 5x (colored in green) already contains loading buffer so PCR products don't need to be mixed with a loading buffer before electrophoresis.

Place solidified gel in the proper orientation (electrophoresis occurs from cathode to anode).

Load :

- PCR products : 5 $\mu$ L ;
- Smartladder 200 to 10 000 bp : 3 $\mu$ L ; (Eurogentec, ref MW-1700-10).

Close the caster and connect it to the generator ( Bio-Rad, ref 1645050).

Let run the electrophoresis at 110 V for 45 min.

## 8.3 Amplification result observation :

After migration, observe amplification results under UV light using an Imager (for instance : ImageQuant LAZ4000, GE Healthcare).

 [LAS 4000 User manual.pdf](#)

**Amplifications worked very well if :**

- Negative control has not amplified;



- Positive control has amplified;
- Amplifications have band at the good size (423 pb), no smear.
- It is possible to observe a second band 200 bp longer than the target. This is due to the presence of eukaryotes in the sample.

Store the amplicons at -20°C until PCR purification.

#### PCR product purification

- 9 PCR products are purified using the purification kit: **NucleoSpin® PCR Clean-Up (Macherey-Nagel, cat. nb 740609.50 or 740609.250 )**.

Store the kit at room temperature.

 [Instruction-NucleoSpin-Gel-and-PCR-Clean-up.pdf](#)

- 10 Prepare Purification Run Table and pool the triplicate PCR into a single microtube with appropriate labelling (sample, target and tag-MID nb, PCR date).

Sample Code	Primer MID	PCR plate and wells	V pool amplicons (µL)	V NT buffer µL	V column µL	V NE elution µL
DNA1-M8R001	515fY_M8R0001	PCR_###_wells A1 to A3	60	120	180	22
DNA2-M8R004	515fY_M8R0004	PCR_###_wells B1 to B3	60	120	180	22
DNA3-M8R005	515fY_M8R0005	PCR_###_wells C1 to C3	60	120	180	22
DNA4-M8R006	515fY_M8R0006	PCR_###_wells D1 to D3	60	120	180	22
...	...	...	...	...	...	...

- 11 Mix 1 vol of sample with 2 volumes of buffer NT.  
Follow the instructions of manufacturer (mentioned in the Step 9), except for the elution step.

#### Elution Step :

- Place the NucleoSpin PCR Clean-Up Column into a clean 1.5mL microtube correctly labeled (sample, target tagged-MID, date).
- Add 22 µL buffer NE preheated at 65°C directly onto the column and incubate 5 min @ 65°C.
- Centrifuge 1 min @ 11 000 g.

Store the purified PCR products at -20°C or directly do the quantification.

#### PCR products quantification

- 12 PCR products are quantified using the quantification kit: Quant-iT™ PicoGreen® dsDNA reagent \*2000 assays\* (Invitrogen, cat nb P7581) and a Fluorometer Plate

readerFluorometer Plate reader, following the manufacturer's protocol.

 [Quant-it PicoGreen dsDNA kit.pdf](#)

### Final Pooled Amplicons preparation

- 13 After quantification of the PCR products, amplicons will be pooled before shipment to the NGS Fasteris sequencing platform.

One final tube (called "**Pooled Amplicons**") will contain all pooled amplicons at equimolar concentration, ready for the library preparation. The amounts and volume required by Fasteris are : **1 µg of equimolar amplicon pool in 30 µL (so Pool Amplicons concentration should be >35 ng/µL) .**

The volume of each amplicon that will be added in the tube "Pooled Amplicons" is calculated based on their average concentration as follows :

### Calculation table for Pooled Amplicons preparation :

Sample name	Primer_MID	nb lect att.	Average Concentration ng/µL	Amount in the final tube ng	Volume in the final tube µL
DNA1-M8R001	515fy_M8R0001	150000	c1	m	$= (m/c1)$
DNA2-M8R004	515fy_M8R0004	150000	c2	m	$= (m/c2)$
DNA3-M8R005	515fy_M8R0005	150000	c3	m	$= (m/c3)$
DNA4-M8R006	515fy_M8R0006	150000	c4	m	$= (m/c4)$
...	...	...	...	m	$= (m/c...)$
...	...	...	...	m	$= (m/c...)$
...	...	...	...	m	$= (m/c...)$
...	...	...	...	m	$= (m/c...)$
Total number of reads for Miseq		15-25 000 000		= Total amount ng (>1µg)	= Total volume µL (<30 µL)
			Pool Amplicons ng/µL		35 ng/µL

*DNA1 has a different concentration than DNA2, DNA3...*

*But DNA1 needs to have the same mass than DNA2, DNA3... in the « Pool Amplicon »*

*Volume calculated of DNA1 to mix in the same microtube with DNA2, DNA3... in the « Pool Amplicon »*  
DNA1, DNA2, DNA3... will be each added with a different volume

*Volume, amount and so concentration required by Fasteris*

If the final volume of Pooled Amplicons is higher than 30 µL, (so concentration inferior to 35ng/µL), an additional concentration step will be necessary. For this we used the purification kit: **NucleoSpin® PCR Clean-Up (Macherey-Nagel, cat. nb 740609.50 or 740609.250) .**

Step 13 includes a Step case.

### Pooled Amplicons Concentration

### Pooled Amplicons Quantification and Quality Checking

step case

### Pooled Amplicons Concentration

This step is performed only if the "Pooled Amplicons" has the following parameters  $>30\mu\text{L}$  and  $<35\text{ ng}/\mu\text{L}$ .

We use the kit **NucleoSpin® PCR Clean-Up (Macherey-Nagel, cat. No 740609.50)**.

The **Final Concentration is fixed at  $50\text{ ng}/\mu\text{L}$** , to be sure to be in excess.

Prepare the Concentration File :

Sample name	Initial Conc. Ci ng/ $\mu\text{L}$	Vi $\mu\text{L}$	V NT buffer $\mu\text{L}$	Vcolumn $\mu\text{L}$	Velution $\mu\text{L}$	Final Conc. Cf ng/ $\mu\text{L}$
PoolAmplicon 1	Ci	Vi	$= 2 * Vi$	$= Vi + VNT$	$= (Ci * Vi) / Cf$	50

The elution volume is calculated depending on the Initial Concentration and Initial Volume of the Pooled Amplicons.

2. Add  $2 * Vi$   $\mu\text{L}$  of NT buffer and follow the recommendations of the manufacturer as mentioned in **Step 9**, except for the Elution Step.

#### Elution Step :

- Place the NucleoSpin PCR Clean-Up Column into a clean 1.5mL microtube correctly labeled (Pool Amplicon name, Quotation nb given by Fasteris, date).
- Add **Vf of buffer NE (calculated in your table above)** preheated at  $65^{\circ}\text{C}$  directly onto the column and incubate 1 min @ room temperature.
- Centrifuge 1 min @ 11 000 g.

Store the concentrated Pooled Amplicons at  $-20^{\circ}\text{C}$ .

- 14 After the preparation of the Pooled Amplicons (and concentration if needed), the final concentration is checked by quantification using **Qubit 4 Fluorometer** (Invitrogen) with the kit **Qubit 1x dsDNA HS Assay** (Invitrogen, Thermofisher Scientific cat. No Q33230).

 [MAN0019617\\_Qubit\\_1X\\_dsDNA\\_BR\\_Assay\\_UG.pdf](#)

- 15 If possible, check the final Pooled Amplicons quality on a Bioanalyzer using the kit **Agilent DNA 1000** (Agilent Technologies, Cat. No 5067-1504).

 [G2938-90014\\_DNA1000Assay\\_KG.pdf](#)

- 16 Store the Pooled Amplicons at  $-80^{\circ}\text{C}$  until the shipment to Fasteris.

## Shipment Conditions to Fasteris

- 17 Pooled Amplicons must be shipped via Dry Ice to the following address :
- FASTERIS SA  
NGS Services  
Chemin du Pont-du-Centenaire 109  
CH-1228 Plan-les-Ouates  
Switzerland

Additonnal documents to include in the package (that must also be sent by e-mail to Fasteris NGS services (ngs@fasteris.com) :

- Quotation number Q##### ;
- Order form ;
- Purchase oder edited by your company.

Don't forget to add a Pro Forma Invoice to your package.