



Jan 14, 2021

Scintillation Count of radiolabeled whole cell

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Works for me

This protocol is published without a DOI.



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ABSTRACT

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PROTOCOL CITATION

Elizabeth Fozo 2021. Scintillation Count of radiolabeled whole cell. **protocols.io**
<https://protocols.io/view/scintillation-count-of-radiolabeled-whole-cell-brghm3t6>



KEYWORDS

Scintillation Count of radiolabeled whole cell, Scintillation Count, radiolabeled whole cell

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CREATED

Jan 14, 2021

LAST MODIFIED

Jan 14, 2021

PROTOCOL INTEGER ID

46313

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ABSTRACT

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Steps

- 1 Grow OG1RF to mid log phase in BHI.
- 2 Harvest 5mL of cells.

- 3 Wash twice in 1X davis salts without glucose.
- 4 Re-suspend in 5mL 1X davis salt without glucose with 10mg/mL BSA.
- 5 Incubate at 37°C for 30 minutes as starvation treatment.
- 6 Move to hot room.
- 7 Place cells in 37°C heat block for assay or in 95°C heat block for certain death.
 - Prior to assay I make a stock of hot/cold OA.
- 8 Add the hot/cold mix to 5mL of cells in the davis/BSA buffer and start the timer.
- 9 Immediately take the 200ul of cells out for T0 - there are placed on the filter paper which is connected to a vacuum - these are washed with 2 mL of wash buffer (1X davis salts with 10mg/ml BSA).
- 10 Stager the time points every 5 minutes for about 20-25 minutes. Then , repeat this for the heat killed cells. Note that there isn't any difference between the heat killed cells and the 37°C cells. We are still working on control for this part of the assay.
- 11 At completion of the assay filters are added to vials with scintillation fluid and counted by the machine.