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Separation of Human Neutrophils (PMN) from Whole Blood

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1 Works for me This protocol is published without a DOI.



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ABSTRACT

Separation of Human Neutrophils (PMN) from Buffy Coat: list of published papers using this protocol

- Boydum A. Isolation of mononuclear cells and granulocytes from human blood. Scand. J. Clin. Lab. Invest. 21 (Suppl. 97): 77-89, 1968

- Alex Mabou Tagne, Franca Marino, Massimiliano Legnaro, Alessandra Luini, Barbara Pacchetti and Marco Cosentino. A Novel Standardized Cannabis sativa L. Extract and Its Constituent Cannabidiol Inhibit Human Polymorphonuclear Leukocyte Functions. Int J Mol Sci 2019 Apr; 20(8): 1833. Published online 2019 Apr 13. doi: 10.3390/ijms20081833.

- Angela Scanzano, Laura Schembri, Emanuela Rasini, Alessandra Luini, Jessica Dallatorre, Massimiliano Legnaro, Raffaella Bombelli, Terenzio Congiu, Marco Cosentino, Franca Marino. Adrenergic Modulation of Migration, CD11b and CD18 Expression, ROS and interleukin-8 Production by Human Polymorphonuclear Leukocytes. Inflamm Res. 2015 Feb; 64(2): 127-35. doi: 10.1007/s00011-014-0791-8. Epub 2015 Jan 6.

PROTOCOL CITATION

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<https://protocols.io/view/separation-of-human-neutrophils-pmn-from-whole-blo-bhatj2en>

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37939

MATERIALS

NAME	CATALOG #	VENDOR
Ficoll Paque PLUS	17144003-500 ml	Ge Healthcare
Fetal Bovine Serum (FBS)	ECS0180L-500 ml	EuroClone
RPMI 1640	ECM 0495L- 500 ml	EuroClone
NaCl	S9625	Sigma Aldrich



NAME	CATALOG #	VENDOR
NH4Cl	1.01145.1000	Merck Serono GmbH
KHCO3	1.04854.500	Merck Serono GmbH
EDTA	ED2SS	Sigma Aldrich
Acetic Acid 100%	A6283	Sigma Aldrich
Gentian violet 1%	not available	Marco Viti


EQUIPMENT

NAME	CATALOG #	VENDOR
Allegra AVANTI 30	Beckman Italy	Beckman Coulter
Cellometer Auto T4	EuroClone	
BD FACS Celesta	Milan Italy BD	




BEFORE STARTING


All reagents used in this protocol must be at room temperature

- 1 Place  **5 mL** of whole blood into a 10 ml volume centrifuge tube.
- 2 Add  **2 mL** of **dextran solution** and mix well by drawing in and out of a pipette.




Dextran
by Elisa Storelli,
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- 3 Incubate in the **dark** for  **00:30:00** at  **37 °C**
- 4 Place  **3 mL** of **Fycoll-HyPaque** media solution into a 10 ml volume centrifuge tube.
- 5 **Slowly** and **carefully** layer the supernatant from blood/dextran mixture onto the Fycoll-HyPaque media solution.



Important: when layering the sample, do not mix the Fycoll-HyPaque media solution and supernatant.

- 6 Centrifuge at  **400 x g, 20°C 00:30:00** with **no brake**.




Allegra AVANTI 30
Centrifuge
Beckman Coulter Beckman Italy

7 Draw off the mononuclear cell layer at the Ficoll/plasma interface along with plasma and Ficoll media, leaving the white cell layer of granulocytes above the red blood cell layer undisturbed.

8 Resuspend the remaining cell layer in  **5 mL** of **NaCl 0.15 M** and centrifuge at  **400 x g, 20°C 00:05:00**



Allegra AVANTI 30
Centrifuge
Beckman Coulter Beckman Italy




NaCl
by Elisa Storelli,
Center for Research in Medical Pharmacology, University of Insubria

9 Aspirate the supernatant with a plastic pipette Pasteur.

10 Lyse remaining red blood cells in  **5 mL** of [hypotonic lysis buffer](#) for  **00:05:00**


11 Centrifuge at  **400 x g, 20°C 00:05:00**



Allegra AVANTI 30
Centrifuge
Beckman Coulter Beckman Italy

12 Aspirate the supernatant with a plastic Pasteur pipette.

13 Resuspend the pellet in  **5 mL** **NaCl 0.15 M**.



NaCl
by Elisa Storelli,
Center for Research in Medical Pharmacology, University of Insubria

14 Centrifuge at  **400 x g, 20°C 00:05:00**





Allegra AVANTI 30
Centrifuge
Beckman Coulter Beckman Italy

15 Aspirate the supernatant with a plastic Pasteur pipette.

16 Resuspend the cell pellet in  **5 mL NaCl 0.15 M** for cell counting.




NaCl
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Center for Research in Medical Pharmacology, University of Insubria

17 Mix  **10 µl** of cell suspension with an equal amount of **Türk solution** (dilution factor=2) allow mixture
 **00:03:00** at Room temperature (RT).



Türk solution and Trypan Blue
by Elisa Storelli,
Center for Research in Medical Pharmacology, University of Insubria

Take  **10 µl** of the mixture and place it inside a **Bürker chamber** and view under an optical microscope using 40x magnification

Count cell in each square found in the four corners and in the central square (figure 1 below), including those that lie on the bottom and left-hand perimeters, but not those that lie on the top and right hand perimeters (see figure 2 below).

Total number of cell per ml = mean number of cell x dilution factor x 10⁴ (hemacytometer volume)

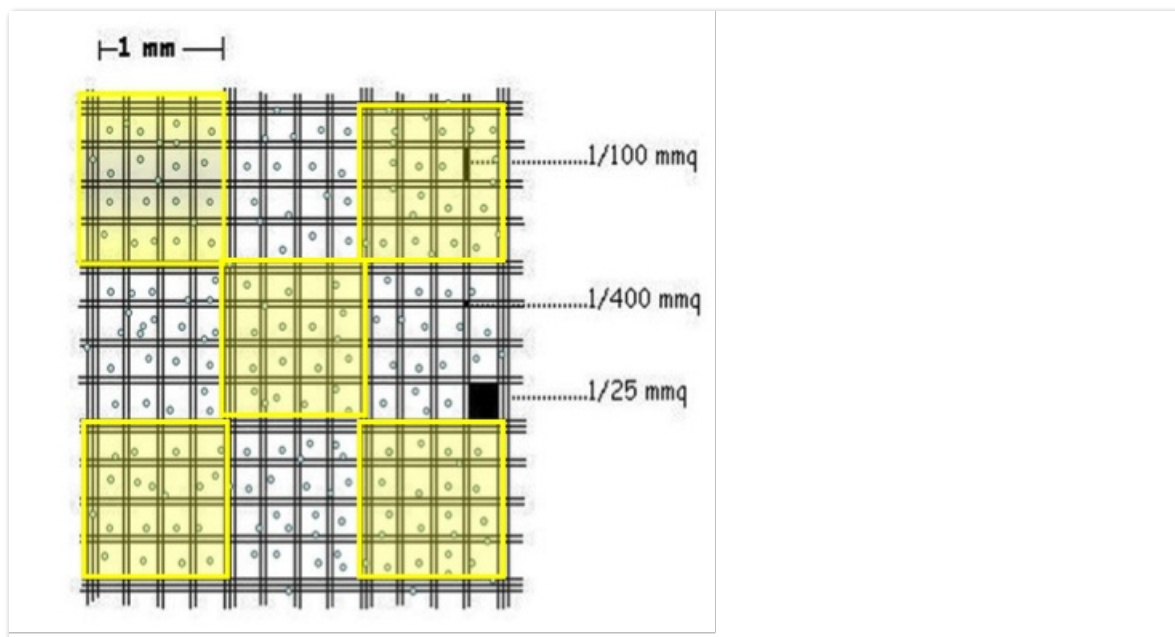


Figure 1

The gridded area of the chamber consists of nine 1 mmq squares. These squares are subdivided in three directions; 0.0625 mmq, 0.05 mmq and 0.04 mmq. The central square here in Figure 1 is further subdivided into 0.0025 mmq = 1/25 mmq squares. Count cells in 5 squares as shown.

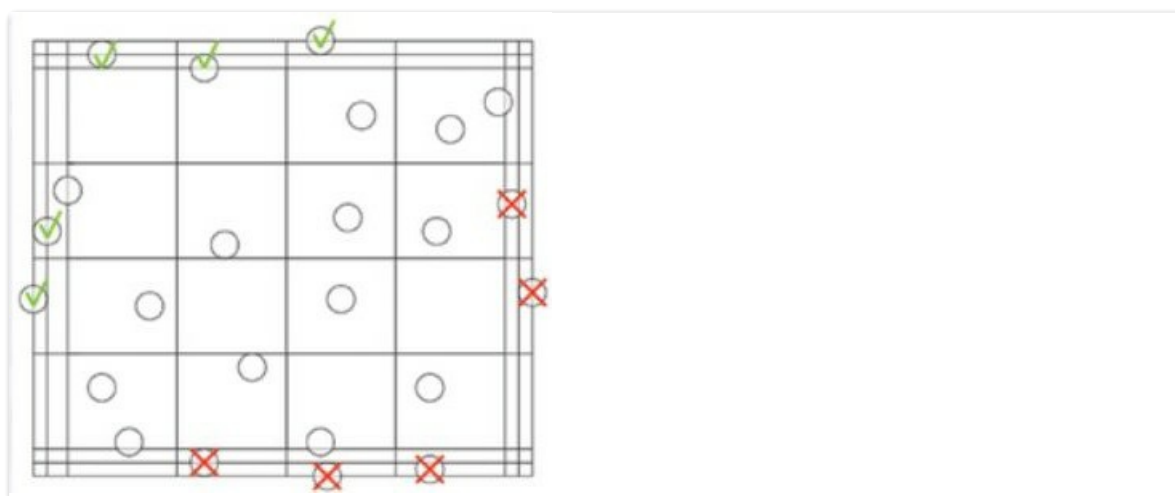


Figure 2


Concerning those cells that lay on the perimeter of the square, count following this scheme.

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OPTIONAL STEP

For automatic cell count with Cellometer machines use **Trypan Blue**. The machine will calculate the number of cells /ml and the % of viability.

Take **10 μ l** of cell suspension and add an equal amount of Trypan Blue. Use all the volume to place it in a counting chamber. Place the chamber inside Cellometer and count.



Cellometer Auto T4
Automated cell counter
Nexcelom Bioscience EuroClone




Türk solution and Trypan Blue
by **Elisa Storelli**,
Center for Research in Medical Pharmacology, University of Insubria

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OPTIONAL STEP

If needed, check the purity of PMN suspension by using morphological parameters of the flow cytometer.

For this test $0,5 \times 10^6$ PMN in **500 μ l** of PBS are enough.




BD FACS Celesta
Flow Cytometer
Becton Dickinson Milan Italy BD



PBS 1X
by **Elisa Storelli**,
Center for Research in Medical Pharmacology, University of Insubria

20 EXPECTED RESULTS



VIABILITY: the expected viability by Trypan Blue should be $\geq 90\%$

CELL YIELD: $\pm 1 \times 10^6$ cells starting from 1 mL of Whole Blood