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**Protocol status:** Working We use this protocol and it's working

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# jer ID:

**Staining** 

1 Antigen Retrieval

Remove PBS from wells. Add 500ul 0.3% citrate buffer into each well.

## ( IHC\_cFOS+Parvalbumin

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#### **ABSTRACT**

Immunohistochemistry protocol for c-FOS and Parvalbumin co-staining of mouse brain tissue slices (30 microns thick).

#### **GUIDELINES**

All shaker incubations are done at 200 rpm speed.

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- 1.1 Incubate at RT on shaker for 30 min Incubate at \$\ 65 \circ\$ for 45 min Cool down on the bench for 20 min
- 2 3X PBS Washes
- 2.1 Remove citrate buffer from wells and replace with 500ul 1X PBS (or use a dropper pipette and fill the well halfway)
- 2.2 Incubate at RT on shaker for 15 min

**≡** remove PBS and replace with new 1X PBS, repeat two more times

- Block Non-Specific Signal Make enough: 10% NGS (Vector Labs, #S-1000-20) in 0.3% triton-X PBS for all samples (500ul per well)
- **3.1** Remove PBS and replace with blocking reagent 500ul per well.
- 3.2 Incubate on shaker RT for 60min.
- 4 Primary Antibody
  Make enough: mouse anti-parv (1:1000, Sigma-Aldrich, #P3088) and rabbi anti-cfos (1:1000, cell Signaling, #2250S) in 1% NGS (Vector Labs, #S-1000-20) in 0.3% triton-X PBS

4.1 Replace the blocking reagent with primary antibody mix, 500 ul per well, and incubate at



- 5 DAY 2 3X PBS Washes
- **5.1** Remove primary Ab mix from wells and replace with 500ul 1X PBS (or use a dropper pipette and fill the well halfway)
- 5.2 Incubate at RT on shaker for 15 min => repeat two more times
- 6 Secondary Antibody
  Make enough: goat anti-mouse AF-555 (1:500, Invitrogen, #A-21422) and biotin goat anti-rabbit (1:1000, Vector Labs, #PK-6101) in 1% NGS (Vector Labs, #S-1000-20) in 0.3% triton-X PBS
- **6.1** Remove PBS from wells and replace with 500ul Secondary antibody mix. Cover with aluminum foil. Note: secondary antibodies are light sensitive, so from this point on the plate MUST be covered from light as much as possible.

Incubate at RT on shaker for 120 min.

- 3X PBS washes as described in Step 5 and 5.1 with the plate covered
- Tertiary Antibody (for cFOS only)
  make enough: AF-488 streptavidin conjugated (1:1000, Thermo Fisher, #S11223) in 1% NGS
  (Vector Labs, #S-1000-20) in 0.3% triton-X PBS

- **8.1** Replace PBS in wells with 500uL of tertiary antibody solution, cover the plate with aluminum foil, and incubate at RT on shaker for 120 min.
- 3X PBS washes as described in step 5 and 5.1 with the plated covered

## **Mounting**

Mounting with medium with and without DAPI Step 10 includes a Step case.

If the mounting medium has DAPI included
If the mounting medium doesn't have DAPI included