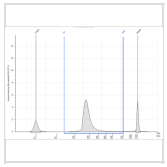

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Amplification and Pooling

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1 Works for me dx.doi.org/10.17504/protocols.io.beqkjduw

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ABSTRACT

This protocol describes the amplification procedure of dual-indexed double-stranded DNA libraries, for shotgun Illumina sequencing. It is typically used for libraries indexed using the following protocol: (https://dx.doi.org/10.17504/protocols.io.bakticwn)

DOI

dx.doi.org/10.17504/protocols.io.beqkjduw

PROTOCOL CITATION

Franziska Aron, Guido Brandt 2020. Amplification and Pooling . **protocols.io** https://dx.doi.org/10.17504/protocols.io.beqkjduw

KEYWORDS

DNA library, NGS, dual-index, ancient DNA, sequencing, nonUDG, double-stranded, DNA, genomic DNA, genomics, palaeogenetics, archaeogenetics, paleogenetics, archeogenetics, aDNA, Illumina, library preparation, nucleic acids, Amplification, PCR, Index Amplification

IMAGE ATTRIBUTION

Franziska Aron

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Apr 07, 2020

LAST MODIFIED

Oct 30, 2020

PROTOCOL INTEGER ID

35308

GUIDELINES

Working in an Molecular Biology Laboratory

This protocol can place in a typical DNA-based molecular biology lab.
Please keep in mind the safety guidelines of your specific country and institution.

Recommendations include wearing of:
- lab coats
- closed shoes and trousers
- safety glasses
- nitril or latex gloves

MATERIALS TEXT

MATERIALS

0.2 ml PCR Tube strips **Eppendorf Catalog #0030124359**

DNA LoBind Tube 1.5ml **Eppendorf Catalog #022431021**

2 ml LoBind Tubes **Eppendorf Catalog #0030108078**

Eppendorf Tubes® 5.0 mL with snap cap **Eppendorf Catalog #30119460**

dNTP Mix (25 mM each) **Thermo Fisher Scientific Catalog #R1121**

Sodium Acetate buffer solution 3M pH 52 for molecular biology **Sigma Aldrich Catalog #S7899-500ML**

Tween 20 **Sigma Aldrich Catalog #P9416-50ML**

Water HPLC Plus **Merck Millipore Sigma Catalog #34877-2.5L-M**

D1000 Ladder **Agilent Technologies Catalog #5067-5586**

D1000 ScreenTape **Agilent Technologies Catalog #5067-5582**

D1000 Reagents **Agilent Technologies Catalog #5067-5583**

Herculase II Fusion DNA Polymerase **Agilent Technologies Catalog #600679**

High Sensitivity D1000 Ladder **Agilent Technologies Catalog #5067-5587**

High Sensitivity D1000 Reagents **Agilent Technologies Catalog #5067-5585**

High Sensitivity D1000 ScreenTape **Agilent Technologies Catalog #5067-5584**

MinElute PCR Purification Kit **Qiagen Catalog #28004**

Primers

Oligo_ID	Sequence (5'-3')	Cocentration
IS5	AATGATACGGCGACCACCGA	10 µM
IS6	CAAGCAGAAGACGGCATACGA	10 µM

Lab equipment

PCR Thermocycler (e.g. Eppendorf Thermomaster Nexus)
Centrifuge 1.5/2.0 ml (e.g. Eppendorf 5424)
Rotor 1.5/2.0ml (e.g. Eppendorf F-45-24-11)
Mini table centrifuge

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TapeStation (e.g. Agilent Technologies, 4200 Tapestation System, SKU: G2991AA)
Vortex mixer (e.g. Scientific Industries Vortex-Genie® 2)

SAFETY WARNINGS

Reagents

Sodium Acetate

- H139: Causes serious eye irritation

Ethanol

- H225 Highly flammable liquid and vapour.
- H319 Causes serious eye irritation.

Guanidinium hydrochloride (GuHCl) (in PB buffer of Qiagen MinElute kit)

- H302 Harmful if swallowed.
- H332 Harmful if inhaled.
- H315 Causes skin irritation.
- H319 Causes serious eye irritation.

Kits

Check manufacturer's safety information for the TapeStation Kits used in this protocol.
Check manufacturer's safety information for the MinElute PCR Purification kit used in this protocol.
- Note that PBI must be stored at room temperature in the dark. PBI is light sensitive.

ABSTRACT

This protocol describes the amplification procedure of dual-indexed double-stranded DNA libraries, for shotgun Illumina sequencing. It is typically used for libraries indexed using the following protocol: (https://dx.doi.org/10.17504/protocols.io.bakticwn)

BEFORE STARTING

Planning

This protocol takes 1 day.

Check all waste disposal guidance for all reagents in this protocol against your corresponding laboratory regulations.

Preparation of buffers (Qiagen MinElute kit):

- Add ethanol to PE wash buffer acccording to manufacturer's instructions.
- Add **200 µl** pH-Indicator and **300 µl** Sodium Acetate to **48.5 mL** of PB binding buffer. This solution is referred to as **PBI** throughout the protocol. Must be stored at room temperature in the dark. PBI is light sensitive.
- Add Tween-20 to EB elution buffer to a final concentration of 0.05% Tween-20 in EB. This solution is referred to as **EBT** throughout the protocol.

Equipment

Make sure all necessary equipment is available (see Materials).

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Abbreviations
EBT = modified EB-Buffer (MinElute Kit), see Preparation of buffers
HPLC = High Performance Liquid Chromatography (-Grade Water)
PBI = modified PB-Buffer (MinElute Kit), see Preparation of buffers
PE = PE-Buffer from Qiagen MinElute Kit

Samples
This protocol is designed for the amplification of indexed libraries as prepared by the protocol described in (https://dx.doi.org/10.17504/protocols.io.bakticwn). The indexing protocol generates **50 µl** of indexed library, of which **20 µl** will be used for this protocol. Ensure sufficient indexed library is avaliable before starting this protocol.

Calculations

1

Prepare amplification assay [**100 µl per reaction**]

Based on the quantification results of the indexed libaries (https://dx.doi.org/10.17504/protocols.io.bakticwn) calculate the number of PCR cycles (amplification factor) needed to reach 10^13 copies of DNA per indexed Library.

Formula in Excel to get the Cycles needed

=LOG((1*10^13/Copies per rxn),2)
(log base 2)

Example: The following calculation is for **5 µl** per reaction, with two indexed library samples (A and B) having different concentrations of DNA copies.
Optonial Changes: 1.If the Calculation shows up less then 3 Cycles, you also have the Option to add less then 5 µl.
2. Instead of 4 reactions of 5 µl each you can also split in 8 reactions of 2 µl each
Instead of 4 x 5 µl reactions you can also split in 8 x 2 µl reactions

Sample Name	Copies per µl	µl per rxn	Copies per rxn	Cycles needed	Real Cycles	Amplification Factor	Output per rxn [Copies]
A	7.32E+10	5	3.66E+11	4.772998	5	32	1.17E+13
B	5.79E+06	5	2.32E+07	18.72018251	19	524288	1.21E+13

Do not calculate the amount of cycles for a higher amount of copies than 1.4 *10^13 to avoid heteroduplexes.

Preparation

2 Prepare cleaned workspace with all necessary reagents and equipment.

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Label all **0.2 mL** PCR strips for the PCR reactions.

PCR

3

Set up four amplification reactions of **100 µl** each per library

Reagent	Stock concentration	Final concentration	1x Volume [µl]
Herculase II Reaction buffer	5x	1x	20
IS5 primer	10 µM	0.4 µM	4
IS6 primer	10 µM	0.4 µM	4
dNTP's	25 mM	0.25 mM	1
Herculase II Fusion	1 U	0.01 U	1
DNA			5
HPLC-Water			65
Total			**100**

3.1

Vortex master mix before adding the enzyme. After adding the enzyme, mix by pipetting or inverting the tube.

3.2

Pipette **95 µl** mastermix and **5 µl** indexed library into each tube (use **0.2 mL** PCR strips).

Keep the remaining library at **-20 °C** until further use.

4

Amplify in a thermocycler with the following program:

Tempreature	Time	
95°C	2 min	Inital denaturation
95°C	30 sec	Cycles (see Step 1)
60°C	30 sec	
72°C	30 sec	

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72°C	5 min	Final elongation
Finally hold the reactions at 10 °C.		

Adjust the number of cycles according to the amplification factor as calculated in step 1.

During this incubation take **MinElute columns** out of the fridge so they warm up to **room temperature** before use in the next step.

This is an ideal point to prepare downstream steps, including labelling of final elution tubes, MinElute Columns etc.

MinElute Purification

5 Purify with MinElute kit with the following modifications to the manufacturer's protocol:
Use one column for all four reactions *[=* **400 µl** *PCR product]* of a sample.

5.1

Add **2400 µl** PB or PBI* buffer to a **5 mL** tube for each sample (this is **600 µl** buffer for each PCR reaction). Add all 4 PCR reactions per sample to the same tube with PB buffer and vortex briefly.

After the PCR product is mixed with the PBI, the PBI should keep its yellow colour. If it turns purple the pH is too high and the efficency of the MinElute columns is not guaranteed.

5.2 Load **700 µl** of the mixture onto one MinElute column, incubate for **00:02:00** , spin **15800 x g, 00:01:00** , and discard flow-through.

Pour off the liquid into a waste tube, and pat the rim of the collection tube dry on a paper tissue or towel. Use just one spot on the paper tissue per sample. Be careful not to touch the rim of the tube on the waste container. After you are finished with all samples, discard the paper and wipe clean the surface underneath with water and soap.

5.3 Repeat loading until the complete mixture was run through the column. **go to step #5.2**

5.4 Add **700 µl** PE (wash) buffer, spin **15800 x g, 00:01:00** , and discard flow-through.

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5.5 Dry spin **15800 x g, 00:01:00** ,

5.6 Put column into new **1.5 mL** LoBind tubes.

5.7 Add **50 µl** EBT buffer to the center of the filter, incubate for **00:02:00** , and spin **15800 x g, 00:01:00** to elute the amplified indexed library.

Carefully pipette EBT directly onto the center of the membrane without touching the membrane.

Measurement and Dilution

6 Dilute amplified index library 1:10 with HPLC- water and check for fragment size, concentration, and heteroduplexes. (for example with the D1000 Kit's Tape, Reagent and Buffer - following the manufacturer's protocol on the TapeStation)

if you see heteroduplexes you need to perform a reconditoning PCR.

Reconditoning PCR: one cycle PCR using 100 ng library template in a 100 µl Herculase PCR reaction (same set up as in 3) and amplified with 1 cycles of 95°C for 2 min, 58°C for 2 min, and 72°C for 5 min. Purify with MinElute kit following the instructions from Step 5, but elute in 20µl EBT.

7 Dilute each amplified indexed library to **10 nM** with EBT buffer or HPLC-water for shotgun sequencing. Then pool the **10 nM** amplified indexed libraries in equimolar amounts (take the same volume for each sample).

The final concentration of a pool of several amplfied indexed libraries should be **10 nM** .

8 Check the **10 nM** library or the **10 nM** library pool for the correct concentration, (for example with the HighSensitivity D1000 Kit's Tape, Reagent and Buffer following the manufacturer's protocol on the TapeStation.)

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