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Schistosoma mansoni miracidia

(Infection of Biomphalaria glabrata snails with

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Schistosoma mansoni



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ABSTRACT

To infect *Biomphalaria glabrata* snails with miracidia hatched from *Schistosoma mansoni* eggs

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Protocol status: Working We use this protocol and it's working

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MATERIALS

MilliQ water Contributed by users

- 24-well Clear TC-treated Multiple Well Plates Individually Wrapped Sterile Costar Catalog #3524
- **☒** Blunt featherweight forceps wide tip **BioQuip Catalog #4750**
- **⋈** 1x DPBS **Gibco Thermo Fischer Catalog #14190144**
- Signal Glass Pasteur pipettes with rubber bulbs Contributed by users

1x Aquarium Water (diluted from 10X; see recipes)

1-2L flask

Aluminum foil

B. glabrata snails 5-8mm

S. mansoni-infected livers in PBS

28°C room or incubator

Incubator at 37°C and 5% CO2

Bucket of ice

| Equipment | |
|---------------------------------------------------------|-------|
| Zoom stereomicroscope with diascopic illumination stand | NAME |
| stereomicroscope | TYPE |
| Nikon | BRAND |
| SMZ800N | SKU |

10X AQUARIUM WATER

5.56g CaCl2

12.28g MgSO4-7H2O

0.43g K2SO4

4.2g NaHCO3

480µl FeCl3-6H2O (0.5g/100ml water)

Fill to 10L and store in 1L bottles

Dilute to 1x: 9L MQ water + 1L 10x

Lung preparation

Collect lungs from patent mice into 50ml Falcon tubes containing pre-warmed 37°C 1x DPBS

1

- 2 Remove lungs from PBS and place in large mortar or laboratory blender
- **3** Gently homogenise lung tissue
- 4 Place homogenised lung slurry into a flask (size of flask depends on how many lungs you have) and fill with diH₂O or aquarium water

Hatching miracidia

- 5 Cover flask in aluminum foil and shine light horizontally across the opening of the flask for 1-2 hrs
- **6** Take a small aliquot of water from the top of the flask using a glass Pasteur pipette and place into a petri dish. Using a stereomicroscope, check for swimming miracidia

Exposing snails to miracidia for regular life cycle maintenan.

- 7 Under a microscope, collect 15 miracidia with a glass Pasteur pipette per well
 - Alternatively (but less preferred) estimate the number of miracidia by counting 12-5µl aliquots in a petri dish. After collecting (shaking the tube in between) and placing the aliquots in the petri dish add 5µl of Lugol to each drop (this kills and stains the miracidia)
- Place individual 5-8mm snails into 24-well culture plates and cover snails completely with 1x aquarium water

| 9 | Expose snails to miracidia for at least 3 hours (up to overnight) |
|----|--------------------------------------------------------------------------------------------------------------------------------------|
| 10 | After exposure, remove the snails carefully using featherweight forceps and put them in a new tank with food |
| | Exposing snails to miracidia for monomiracidium infections |
| 11 | Dilute miracidia so that one miracidium can easily be collected in ~3-5µl of water |
| 12 | Collected single miracidium using fresh 10µl pipette each collection and place in 24-well plate |
| 13 | After a plate is filled, check each well under a microscope to verify there is a single miracidium in each well |
| 14 | Place individual 5-8mm snails in the wells containing confirmed single miracidium and cover snails completely with 1x aquarium water |
| 15 | Leave the plate overnight (inside incubator or room at 28°C) |
| 16 | The following day, transfer the snails to a new tank with food |