



Nov 23, 2020

## Cytochrome C Assay

## Elizabeth Fozo<sup>1</sup>

<sup>1</sup>In-house protocol

1 Works for me

This protocol is published without a DOI.

Eadewunm

## PROTOCOL CITATION

Elizabeth Fozo 2020. Cytochrome C Assay. **protocols.io** https://protocols.io/view/cytochrome-c-assay-bpz6mp9e

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Nov 23, 2020

LAST MODIFIED

Nov 23, 2020

PROTOCOL INTEGER ID

44830

DISCLAIMER:

DISCLAIMER: THIS IS A WORK IN PROGRESS. IT IS FOR INFORMATIONAL PURPOSES ONLY; USE AT YOUR OWN RISK

The protocol content here is for informational purposes only and does not constitute legal, medical, clinical, or safety advice, or otherwise; content added to <a href="protocols.io">protocols.io</a> is not peer-reviewed and may not have undergone a formal approval of any kind. Information presented in this protocol should not substitute for independent professional judgment, advice, diagnosis, or treatment. Any action you take or refrain from taking using or relying upon the information presented here is strictly at your own risk. You agree that neither the Company nor any of the authors, contributors, administrators, or anyone else associated with <a href="protocols.io">protocols.io</a>, can be held responsible for your use of the information contained in or linked to this protocol or any of our Sites/Apps and Services.

## Steps

- 1 Start ON cultures of a strain of interest.
- 2 The next day, measure 100 mL BHI into each flask to be used.
- 3 Measure  $OD_{600}$  of each overnight and calculate how much of your overnight culture you need for  $OD_{600} \sim 0.01$  in 100 mL.

TKO - 20 min - WT - 10 min - DKO

4	Add supplement if doing Long Term exposure or place BHI + 0.01 cells directly into 37*C for a later spike-in
5	Harvest cells at 0.3 if doing Long Term or 0.225-0.25 if doing Spike
	Check OD after 2.5 hours.
6	Obtain $OD_{600}$ values for each of your cultures and then calculate how much of your culture is needed to reach an $OD_{600}$ of 7 in 3 mL
	<ol> <li>Formula: (7-OD<sub>600</sub>)(3mL) = (measured log phase-OD<sub>600</sub>)(X mL)</li> <li>Will need around 60-80mL to achieve this concentration</li> </ol>
7	Pipet half the total volume needed into a 50mL conical and centrifuge for~10 minutes at 3500 RPM. Pour off supernatant. Add the other half of the cell volume. Centrifuge again.
8	Wash the cells with 10 mL of 20mM MOPS buffer*(stock is 1M) by centrifugation for 5 min at 3500 RPM. 20mM MOPS buffer is made in $\rm H_2O$
	C1V1=C2V2 formula to figure out how much stock to make 20mM.
	*MOPS-3-(N-Morpholino)propane sulfonic acid resuspend in H2O- autoclave.
9	Repeat the wash
0	Resuspend the pellet in 3 mL of 20mM MOPS buffer
1	Add 1mg/mL of Cytochrome C (stock is 10mg/mL in H2O)-vortex
	C1V1 = C2V2
2	Incubate for 10 minutes

Centrifuge for 10 minutes at 3500 RPM

13

- 14 Add 2 mL of the supernatant to 4mL plastic cuvettes
- 15 While cytochrome cells are spinning, make standards. To 3 mL of 20mM MOPS buffer add:
  - 0.1 = 30uL of cytochrome
  - 0.2 = 60uL of cytochrome
  - 0.3 = 90uL
  - 0.4 = 120uL
- 16 Use the Cytochrome program ("CYTOCHROME") on the Fozo Lab Spec =  $OD_{530}$

Standards listed above cover a range of predetermined mg/mL results, if your results fall outside of this range, you may need to adjust your standards accordingly.