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Preparing 10x PCR buffer

In 1 collection

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Low-cost, high-quality ...

Nadine Mowoh

ABSTRACT

10X ThermoPol Reaction Buffer is optimized for use with Vent® and Deep Vent® DNA Polymerases. This buffer also provides superior reaction conditions for other thermophilic DNA polymerases, including *Taq*DNA Polymerase, OpenVent as well as various other DNA and RNA modifying enzymes.

Here we describe the protocol used in making a 10x PCR buffer recipe that provides optimum reaction condition for OpenVent polymerase enzyme.

10x PCR buffer comprise of:

100 mM KCL 100 mM (NH)2SO4 200 mM Tris-HCL pH8.8 @ 25c 20 mM MgSO₄

PROTOCOL CITATION

1 % Triton x100

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https://protocols.io/view/preparing-10x-pcr-buffer-cc79szr6

COLLECTIONS (i)

Beneficial Bio Products

KEYWORDS

10x PCR buffer recipe, Composition of 10x PCR buffer



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GUIDELINES

This protocol describes the steps in preparing a low cost 10x PCR buffer.

MATERIALS TEXT

Chemicals

⊠Tris **Roche Diagnostics**

⋈ Hydrochloric acid Thermo Scientific

Ammonium sulphate Sigma Aldrich

 Magnesium sulphate Sigma Aldrich

⊠ Distilled Water **Contributed by users**

Materials and Equipment

Sterile Pipette tips (P-10, P200, P-1000) Micropipettes Sterile falcon tubes Sterile Eppendorf tubes

SAFETY WARNINGS



- HCL is very corrosive and should be handled with care.
- Wear protective clothing (Lab coat), face masks and gloves.

BEFORE STARTING

Make sure all chemicals are available in their right concentrations needed to make the 10x PCR buffer.

Preparing reagent stocks

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Before preparing the 10x PCR buffer, prepare 1M and initial stocks of the different chemicals that would be used.

- We adopt 3 M stock of HCL to use in preparing Tris-HCL
- Prepare 10 mL of 3 M HCL by diluting 3.129 mL of concentrated or 37% HCL in 6.871 mL of distilled water.



When you mix acid with water, it's extremely important to add the **acid to the water** rather than the other way around. This is because acid and water react in a vigorous exothermic reaction, releasing heat, sometimes boiling the liquid.

Prepare 10 mL of 1 M concentration of each of the chemical stocks by weighing the salt powders as indicated in the table 1 below:

- Weigh out the salts as indicated in the table and add to a 100 mL glass beaker.
- Measure out 10 mL of distilled water and add to the glass beaker.
- Add a magnetic flea and place on a magnetic stirring plate to mix the solution as shown in figure 1
- To sterilize, autoclave the solutions by using a steam autoclave at 121° C for at least 30 minutes under at least 15 psi. (All the solutions should be transferred into Duran bottles that can withstand heating).

Α	В
Chemical	Amount
	needed in (g)
	for 1 M stock
Tris	1.211
KCL	0.74
NH(SO4)2	1.30
MgSO4	1.20

Table 1

To make 1M Tris HCL pH 8.8

- 1. Weigh out **1.211** g Tris and add to a 50 mL glass beaker.
- 2. Measure out 8 mL of distilled water and add to the glass beaker.
- 3. Add a magnetic flea and place on a magnetic stirring plate to mix the solution as shown in figure 1.
- 4. Add a pH meter into the solution to observe the pH.
- 5. Slowly add the 3 M hydrochloric acid (HCl) solution using a Pasteur pipette to reduce the pH to p+8.8. Be careful not to add too much at a time, since the pH will change rapidly.
- 6. Once the desired pH has been reached, top up the solution to 10 mL using distilled water.



Figure 1

Pipetting to make 10x PCR buffer - 1mL

2



Pipette the following reagent stocks into a clean 1.5 mL Eppendorf tube as described in the table below to compose 1 mL of 10x PCR buffer.

1M KCL

1M (NH)2SO4



1M Tris-HCL pH8.8 @ 25c 1M MgSO $_4$

Triton x100

Α	В
Chemical	Amount
stock	required in uL
	to make a
	1000 mL of
	10x PCR
	Buffer
1M KCL	100
1M (NH)2S04	100
1M Tris-HCL	200
pH8.8 @ 25c	
1M MgSO4	20
Triton x100	10

Table 2

- Cork the tube and invert several times to mix the solutions
- Label the tube appropriately and store at -20c.
 - Before using the buffer, carryout a functionality and Nuclease test to show that the buffer formulation is able to provide a suitable PCR condition for DNA amplification and also free from contaminating nucleases.
 - We typically use our internal quality control protocol to check this.