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# Expression and purification protocol of *Homo sapiens* E2-like enzyme ATG3

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Liv Jensen

This protocol describes expression and purification procedures for obtaining human recombinant autophagy E2-like enzyme ATG3 (ATG, AuTophagy-related protein) of the ATG8 ubiquitin-like conjugation system.

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protocol

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ATG3, expression, purification, recombinant protein, *Homo sapiens*, ASAPCRN

protocol ,

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## General information

Insert: *Homo sapiens* ATG3, NP\_071933.2; Expression system: *E. Coli* Rosetta pLyss; plasmid origin: Sascha Martens Lab, Addgene 169079, lab internal construct database number SMC861; backbone: pET-Duet1; plasmid resistance: Ampicillin; tags & cleavage sites: N-term 6xHis, followed by Tobacco Etch Virus (TEV) cleavage site, ATG3 ORF. Ext coeff: 45840 M<sup>-1</sup> cm<sup>-1</sup>, MW 35.8 kDa.

## Materials and Reagents

- *Escherichia coli* Rosetta pLyss cells
- Luria Bertani (LB) medium with antibiotics (final conc. 50µg/ml Ampicillin, 34µg/ml Chloramphenicol)
- IPTG (isopropyl-β-D-thiogalactopyranoside)
- 37°C shaker incubator
- sterile flasks/sterile pipettes
- tip sonicator
- **Lysis Buffer:** 50mM Hepes pH=7.5; 300mM NaCl, 10mM Imidazole, 2mM MgCl<sub>2</sub>, 2mM β-mercaptoethanol, 1mM Pefablock, Complete Protease Inhibitors (EDTA-free CIP tablet, Roche), DNase (Sigma).
- **Buffer A:** 50mM Hepes pH=7.5, 300mM NaCl, 10mM Imidazole (filtered and degassed) + 1mM β-mercaptoethanol
- **Buffer B:** 50mM Hepes pH=7.5, 300mM NaCl, 300mM Imidazole (filtered and degassed) + 1mM β-mercaptoethanol
- **Size Exclusion Chromatography (SEC) Buffer:** 25mM Hepes pH=7.5, 150mM NaCl (filtered and degassed) + 1mM DTT (Dithiothreitol)
- Note: all purification buffers are filtered and degassed. Reducing agents (β-mercaptoethanol and Dithiothreitol) are added after degassing step.

**Columns:** - NiNTA resin  
- S75\_16/60 (GE Healthcare)

**Gels:** 10% SDS-PAGE

For hazard information and safety warnings, please refer to the SDS (Safety Data Sheet).

## Protein Expression

1



Transform plasmid DNA (Addgene 169079, SMC861) into *E. Coli* Rosetta pLyss cells and plate on Ampicillin/Chloramphenicol LB agar plate for ☀ **Overnight** growth at 🌡 **37 °C**.

2 

The following day, inoculate a **5 mL LB + Amp/Cam pre-culture** with 1-2 colonies and grow **Overnight** at **37 °C** shaking.

3 The following day, use **5 mL pre-culture** to inoculate **1 L LB medium + Amp/Cam** at **37 °C** until an OD<sub>600</sub> (Optical Density at 600nm) of 0.4 is reached.

4 Cool down the culture to **18 °C** and grow until OD<sub>600</sub> = 0.8.

5 Induce protein expression with **200 micromolar (μM) IPTG** and keep shaking for a further **16:00:00** at **18 °C**. 16h

6 

15m

Pellet cells at **4000 rpm, 4°C, 00:15:00** in a Sorvall RC6+ centrifuge (Thermo Scientific), discard supernatant and resuspend pellets in ice cold lysis buffer (25 ml/1 lt culture).

7 

Flash freeze resuspended pellets in liquid nitrogen and store at **-80 °C** until purification.

Protein Purification 30m 30s

8 Perform His-Trap affinity purification followed by Size Exclusion Chromatography.

9 Cells are lysed via sonication: thaw pellet corresponding to **1 L** culture. All following steps are to be executed at **4 °C** or on ice.

10 Lyse cells by sonicating them using an immersion tip Sonicator (**2x 00:00:30**). Note: 30s

adjust times and intensity according to the available instrument.

11



30m

Clear lysate by spinning it down in a Beckman centrifuge at

 **40000 x g, 4 °C, 00:30:00 , Ti45 Rotor .**

12

Load supernatant onto 2ml Ni-NTA column at  **4 °C** pre-equilibrated in Buffer A.

13




Wash column with  **50 mL of Buffer A** to remove unspecific bound proteins.

14

Elute protein of interest with 300mM Imidazole concentration. Collect fractions.

15





Check fractions of each step on a SDS-PAGE. Pool and concentrate those containing the protein of interest by spinning at  **4 °C** down in a 10kDa cut-off Amicon Filter to

 **2 mL final volume .**


16

Cleave His tag overnight with TEV protease in dialysis against buffer containing 0mM Imidazole.

17

Inject  **2 mL protein** onto a S75\_16/600 column operating at  **4 °C** and pre-equilibrated in buffer containing SEC Buffer (see profile below).

18

Check fractions on a 10% SDS-PAGE. Pool and concentrate those containing the protein of interest down at  **4 °C** in a 10kDa cut-off Amicon Filter.

19



Measure protein Absorbance  $A_{280}$  using a Spectrophotometer blanking against SEC buffer (MW = 35,864 kDa ; Extinction coefficient =  $45840\text{M}^{-1}\text{cm}^{-1}$ ).

20

Resuspend protein in Glycerol to a final concentration of **30 % (v/v) for glycerol** . Store the protein batch at **-20 °C** . Estimated protein yield: **10 mg per 1 lt culture** . Protein activity is kept for 18 months.