



APR 08, 2024

Inexpensive DNA Extraction Protocol

Valerie Warhol¹

¹Carnegie Museum of Natural History



Valerie Warhol
Carnegie Museum of Natural History

ABSTRACT

This is a protocol for doing extraction of DNA using inexpensive reagents rather than enzymes or kits. It can be used for plant, animal, or fungus samples.

MATERIALS

- Forceps (for handling sample)
- Pestle (for grinding sample in microtube)
- Hinged 1.5 μ L microtubes (3 per sample)
- Microcentrifuge
- Water bath
- Adjustable micropipette 1-10 μ L
- Adjustable micropipette 10-100 μ L
- Adjustable micropipette 100-1000 μ L
- Pipette tips
- Molecular grade water (100 μ L per sample)
- Silica resin (silica dioxide 50% w/v in water) (3 μ L per sample)
- Wash buffer (diluted 1:1 in 95% ethanol) (1 mL per sample)
- Guanidine Hydrochloride 6M (250 μ L per sample)

PROTOCOL MATERIALS

-  Guanidine Hydrochloride 6M **Carolina Biological Supply Catalog #C33427** Step 5
-  Silica Resin **Carolina Biological Supply Catalog #C33426** Step 12
-  Wash Buffer **Carolina Biological Supply Catalog #C33428** In 3 steps

OPEN  ACCESS



Protocol Citation: Valerie Warhol 2024. Inexpensive DNA Extraction Protocol. **protocols.io** <https://protocols.io/view/inexpensive-dna-extraction-protocol-dbtt2nnn>

License: This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited





Protocol status: Working
We use this protocol and it's working

Created: Apr 07, 2024

Last Modified: Apr 08, 2024




PROTOCOL integer ID: 97875

Prepare sample and equipment


- 1 Make sure all instruments, such as forceps and pestle, are clean and sterile. Make sure  Wash Buffer **Carolina Biological Supply Catalog #C33428** has been diluted 1:1 in 95% ethanol.
- 2 Prepare water bath at  65 °C .
- 3 Dissect sample from specimen. (This will be a piece of tissue approximately 10–20 mg. For small arthropods such as beetles or spiders, 1 or 2 legs will typically suffice.) Return specimen to freezer.
- 4 If sample was stored in ethanol, let sample dry for 5–10 minutes.
- 5 Prepare a clean 1.5 mL hinged tube by writing sample ID on it and filling with  250 µL of  Guanidine Hydrochloride 6M **Carolina Biological Supply Catalog #C33427**

Lyse cells


11m

- 6 Put sample in tube. Grind sample with pestle until broken up into tiny pieces.
- 7 Incubate sample tube in  65 °C water bath for  00:10:00 .
- 8 Remove tube and lower temperature of water bath to  57 °C .

10m



9 Centrifuge tube for  00:01:00 at maximum speed to pellet debris. 1m



10 Remove  Silica Resin **Carolina Biological Supply Catalog #C33426** from refrigerator.


11 Label a clean 1.5 μ L tube with sample number. Transfer  150 μ L of the supernatant to the clean tube. Discard old tube containing debris.

Bind DNA

5m 30s

12 Add  3 μ L of  Silica Resin **Carolina Biological Supply Catalog #C33426** to tube. Mix well by pipetting up and down several times.

13 Close tube and incubate for  00:05:00 in  57 °C water bath. 5m

14 Centrifuge for  00:00:30 at maximum speed to pellet the resin. 30s

15 Use a pipette with a fresh tip to remove the supernatant, being careful not to disrupt the pellet.

Wash

1m

- 16 Remove molecular grade water from refrigerator and

⊗ Wash Buffer **Carolina Biological Supply Catalog #C33428**

 from freezer.

- 17 Add

🧪 500 µL

 of ice-cold

⊗ Wash Buffer **Carolina Biological Supply Catalog #C33428**

 to the pellet. Mix well by pipetting up and down several times to resuspend the silica resin.

- 18 Close the tube and centrifuge for

⌚ 00:00:30

 at maximum speed to pellet the resin.

30s

- 19 Use a pipette with a fresh tip to remove the supernatant, being careful not to disrupt the pellet.

- 20 Again, add

🧪 500 µL

 of ice-cold

⊗ Wash Buffer **Carolina Biological Supply Catalog #C33428**

 to the pellet. Mix well by pipetting up and down to resuspend the silica resin.

- 21 Close the tube and centrifuge for

⌚ 00:00:30

 at maximum speed to pellet the resin.





30s

- 22 Return wash buffer to freezer.

- 23 Use a pipette with a fresh tip to remove the supernatant, being careful not to disrupt the pellet. Spin the tube briefly to collect any remaining drops of supernatant, and then remove these with a pipette.

Elute DNA

30s

- 24 Add  100 μ L of molecule grade water to the silica resin and mix by pipetting up and down several times.
- 25 Incubate the mixture at  57 °C for 5 minutes.
- 26 Centrifuge for  00:00:30 at maximum speed to pellet the resin. 30s
- 27 Label a clean 1.5 μ L tube with sample number. Transfer  90 μ L of the supernatant to the clean tube, being careful not to disturb the pellet. Discard old tube containing the resin.
- 28 Store sample in freezer until ready to PCR. If going directly to PCR, put sample in refrigerator.