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## FindingNemo Extraction 3: NEB Monarch Kit

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1 Works for me

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### ABSTRACT

This is a sub-protocol designed to extract/isolate ultra-high molecular weight (UHMW) DNA to obtain ultra-long (UL) reads on Nanopore sequencers using **New England Biolabs (NEB) Monarch HMW DNA Extraction Kit for Cells & Blood**.

A DNA extraction protocol that yields clean and homogeneous UHMW DNA is important for a good UL sequencing output. The choice of protocol should be based on achieving these parameters.

The **New England Biolabs (NEB) Monarch HMW DNA Extraction Kit for Cells & Blood** is a quick, tweakable extraction protocol fitting for one-day library prep.

We tested this sub-protocol in **human cell line**, with input cells varied between 1-6 millions. As a rule of thumb, a million cells will suffice for one load on a MinION.

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### KEYWORDS

ultra-long sequencing, cohex, glass bead, nanopore, MinION, UHMW DNA, Monarch, Circulomics, phenol, SDS, CTAB, GM12878, Whatman, PromethION, Nanobind

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## GUIDELINES

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Please follow on Twitter for latest updates and results:

@NininUoN

@mattloose

## MATERIALS TEXT

### Chemicals/Compounds

[Tris-HCl pH 8.0 Thermo](#)

- [Scientific Catalog #J22638-AE](#)

[Ethanol](#)

- [Absolute Honeywell Catalog #32221-2.5L](#)

[Isopropanol Absolute Fisher](#)

- [Scientific Catalog #P/7500/15](#)

[1X Phosphate Buffer Saline Fisher](#)

- [Scientific Catalog #15453819](#)

[Nuclease-free](#)

- [Water Thermofisher Catalog #AM9920](#)

### Kits

[Monarch HMW DNA Extraction Kit for Cells & Blood New England](#)

- [Biolabs Catalog #T3050S](#) Step 1

### Disposables

[DNA LoBind Tubes, 1.5](#)

- [mL Eppendorf Catalog #0030108051](#)
- Wide-bore (or cut off) P1000 and P200 tips

#### SAFETY WARNINGS

When handling phenol always wear PPE, keep a solution of 50% (w/v) PEG-400 nearby to treat the burn in the case of accidental splashes.

#### BEFORE STARTING

##### Things to observe at all times:

- Excessive and vigorous pipetting and vortexing should be avoided as these may shear the DNA.
- Make up buffers with nuclease-free water to avoid introducing nucleases to solutions.
- Avoid unnecessary heating and freezing; isolated DNA should be stable for storage in the fridge for months.

#### UHMW DNA Extraction

- 1 This protocol is using **Monarch® HMW DNA Extraction Kit for Cells & Blood**.

[☒ Monarch HMW DNA Extraction Kit for Cells & Blood New England](#)

**Biolabs Catalog #T3050S**

- 2 We have obtained optimal extractions using the NEB Monarch kit. This combines speed with high quality UHMW DNA. Follow the manufacturer's instructions as described [here](#), BUT incorporate the following changes as described below.

Our most homogeneous extracted DNA samples were obtained by lysis at 600-700 rpm speed using the Monarch kit. This is one area that can be optimised depending on the input sample.

#### One-day Protocol

4h

- 3 To complete DNA extraction from cell to library prep and sequencing in one day, start early!

One million human cells are sufficient for a single library load on the MinION. At least two million human cells are required for a single load on the PromethION. Other samples can be scaled according to total amount of DNA recovered. So for a 1 gigabase genome you would require 3 million cells etc.

- 4 Dilute the eluted DNA with 150 µl of NEB Elution Buffer II.

- This is following the NEB UHMW Monarch kit protocol where DNA is first eluted with 100 µl buffer. After this step, sample volume will be 250 µl total.
- Quantification of a very viscous UHMW DNA is problematic and will not produce accurate results, hence the dilution.
- Gradual dilution is recommended to achieve homogeneous concentration of 50-100 ng/µl.

5 Incubate the eluted DNA at 37°C for about 2-3 hours with regular pipette mixing.

3h

 **37 °C**  **03:00:00 max (2-3 hours)**

During mixing, observe by eye that the viscous DNA 'blob' has been more or less dissolved to the different parts of the tube (*i.e.*, less heterogeneous). This is usually observed after 2 hours of incubation. Otherwise, continue the incubation to 3 hours and proceed to the next step.

6 Quantify DNA as per "**UHMW DNA QC**" and check homogeneity by calculating %CV values. If the DNA is not sufficiently homogeneous, incubate the DNA for longer.

7 Store at 4°C or continue to **UL Library Preparation** as per "**Modified ULK001**".  
If only SQK-RAD004 is available, follow library preparation as in "**Modified RAD004**" or "**KrazyStarFish (KSF)**".

 **4 °C for storage**