Passage cells

Adita Ayu Permanasari[1]

[1]Institute of Tropical Disease, Airlangga University

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Adita Ayu Permanasari

ABSTRACT

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1 Observe cells under microscope

2 Wipe outside surface of cell culture dish with ethanol cotton and bring it inside Biosafety Cabinet (BSC)

3 Bring medium, PBS, Trypsin-EDTA 2x to BSC

4 Bring autopipetter, disposable pipet (10ml, 2ml), one 15ml tube to BSC

5 Discard old medium

6 Rinse the bottom of dish with 10ml PBS

7 Add 1ml Trypsin-EDTA 2x to dish

8 Incubate the dish in CO2 Incubator 37oC 4min

9 Check under microscope (cell shape become rounded)

10 Neutralize trypsin-EDTA 2x by add 8ml medium

11 Suspend detached cells and transfer to 15ml tube

12 Centrifuge 1200 rpm, 4 min

13 Bring to BSC, discard medium

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14 Add 20 ml DMEM and divided each 10 ml to dish (total 2 dish with @50% cells)

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