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# Protocol for Exo-CIP™ Rapid PCR Cleanup (#E1050)

New England Biolabs<sup>1</sup><sup>1</sup>New England Biolabs

1 Works for me

[dx.doi.org/10.17504/protocols.io.8yahxse](https://dx.doi.org/10.17504/protocols.io.8yahxse)

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## ABSTRACT

**Exo-CIP™ Rapid PCR Cleanup Kit**

- Rapidly degrade residual PCR primers and dephosphorylate excess dNTPs after amplification
- Reaction complete in 4 minutes
- Thermolabile formulation can be heat inactivated in 1 minute at 80°C
- PCR product can be used directly in downstream applications
- Compatible with commonly-used reaction buffers

## EXTERNAL LINK

<https://neb.com/protocols/2019/01/16/protocol-for-exo-ciprapid-pcr-cleanup-e1050>

## MATERIALS

NAME	CATALOG #	VENDOR
Exo-CIP Rapid PCR Cleanup Kit - 100 rxns	E1050S	New England Biolabs

## STEPS MATERIALS

NAME	CATALOG #	VENDOR
BigDye™ Terminator v3.1 Cycle Sequencing Kit	4337455	Thermo Fisher Scientific

## SAFETY WARNINGS

Please see SDS (Safety Data Sheet) for hazards and safety warnings.



- 1 Transfer **5 µl** of PCR product to a new PCR tube and add **1 µl** of Exo-CIP A and Exo-CIP B respectively. The final volume is **7 µl**.



- 2 Mix thoroughly and briefly centrifuge at **1000 x g**.



- 3 Incubate the reaction tube for **00:04:00** at **37 °C** followed by **00:01:00** at **80 °C**.

- 4 Submit **3 µl** or less (in a range of 15-200 fmol)\* of treated PCR product for sequencing using BigDye™ Terminator v3.1 Cycle Sequencing Kit or store the treated samples at **-20 °C** for longer term storage.



**BigDye™ Terminator v3.1 Cycle Sequencing Kit**  
by Thermo Fisher Scientific  
Catalog #: 4337455



\* A simple way to determine the amount of your amplicon is to load **3 µl** on an agarose gel along with a known amount of a control DNA for comparison. Alternatively, direct measurement using fluorescent dye based kit (e.g., Qubit™) will ensure the proper amount of DNA is submitted.

Size of PCR amplicon	ng of DNA (in 3 µ sample)
100 bp	1 - 12
500 bp	5 - 60
1000 bp	10 - 120
3000 bp	30 - 360
5000 bp	50 - 600