

6



Version 2 ▼

Simon Blanchoud¹

¹University of Fribourg

Works for me

dx.doi.org/10.17504/protocols.io.brwxm7fn

Haematoxylin-Eosin stain for cryosections V.2

Jan 29, 2021

Blanchoud lab, UNIFR



ABSTRACT

This protocol has been successfully used to stain our samples of colonial tunicates sectioned at 12 µm.

Adapted from Anna Jazwinska and Marguerite Kaczorowski.

dx.doi.org/10.17504/protocols.io.brwxm7fn

PROTOCOL CITATION

Simon Blanchoud 2021. Haematoxylin-Eosin stain for cryosections. protocols.io https://dx.doi.org/10.17504/protocols.io.brwxm7fn

Version created by Marta Wawrzyniak

KEYWORDS

colonial tunicates, Haematoxylin-Eosin, cryosections, histology

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Jan 29, 2021

LAST MODIFIED

Jan 29, 2021

PROTOCOL INTEGER ID

46775

GUIDELINES

Note that the solutions need to be filtered after dilution, which takes typically 30 min.

mprotocols.io

01/29/2021

MATERIALS TEXT

- Ammonium iron(III) sulfate [Iron alum] (CAS 7783-83-7)
- Sulfuric acid (CAS 7664-93-9)
- Hematoxylin (CAS 517-28-2)
- Eosin Y (CAS 17372-87-1) or Erythrosine B (CAS 16423-68-0; for a redder coloring).
- Glacial acetic acid
- Ethanol
- HCl
- H20
- Histo-Clear II from National Diagnostics (HS-202) or Xylol
- Omnimount from National Diagnostics (HS-110).
- Coverslips
- Cellulose fliters

SAFETY WARNINGS

Note that Eosin solutions have to be discarded in the **Halogenated waste**.

Use Histo-Clear II which is a safer alternative to Xylol.

Groat's haematoxylin

- 1 Mix:
 - 1 g of Ammonium iron(III) sulfate
 - **⊒50 mL** of dH₂O
 - **Q.8** g of sulfuric acid
 - 1.1 Mix separately:
 - **Q0.5 g** of hematoxylin dissolved in **Q50 mL** of EtOH (96%)
 - 1.2 Pour the water-based mix into the EtOH one, filter if necessary.

0.1% HCl in 70% EtOH

- 2 Mix:
 - **250 μl** of HCl ([M] **37 % (V/V)**)
 - **□100 mL** of EtOH ([M]**70 % (v/v)**)

Solution can be stored and used to differentiate ~30 slides.

Eosin Y 0.1%

- 3 Mix:
 - **□0.1 g** of Eosin Y
 - \blacksquare 100 mL of dH₂0
 - **□0.5 mL** of glacial acetic acid

Filter if necessary.

Histo-Clear II (HC)

4 Use Histo-Clear II for the final washes.

 Thaw for **© 00:20:00** or until no more signs of condensation.

20m

Rehydrate the cryosection in tap water © 00:05:00

5m

5.2 Stain in Haematoxylin © 00:12:00 12m

- Haematoxylin solution can be reused many times and just needs to be filtered once in a while.
- 5.3 Differentiate each slide by shaking it vigorously in HCl-EtOH © 00:00:20

20s

5.4 Place in a coplin jar with tap water, place the jar in the sink and let tap water **run over it** for © 00:10:00

10m

- 5.5 Rinse in distilled water.
- 5.6 Stain in Eosin Y for \bigcirc **00:05:00**.

5m

- Rinse in EtOH [M]80 % (V/V) (slides could be checked for their Eosin staining).
- 3m
- 5.8 Start the wash "baths": EtOH (80%, 100%, 100 %) & HC (I, II, III) **© 00:03:00** each.

5.9 Mount under a coverslip using Omnimount.