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## Parallel rapid expression and purification of proteins for crystallography (PREPX): large scale 1 L cultures

 Forked from [Parallel rapid expression and purification of proteins for crystallography \(PREPX\): large scale 1 L cultures](#)

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ASAP Discovery



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### ABSTRACT

This protocol details the parallel rapid expression and purification of proteins for crystallography (PREPX) at a 1 L culture scale. Recombinant proteins are expressed in *Escherichia coli* using the autoinduction method and then purified in parallel using a IMAC, desalt, tag cleavage, reverse IMAC and gel filtration work flow.

### ATTACHMENTS

[nyikb5id7.docx](#)

### GUIDELINES

#### Method overview

Standard workflow is expression via autoinduction followed by purification using IMAC/PD-10/revIMAC and serial gel filtration

**Protocol status:** Working  
We use this protocol and it's working

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**Keywords:** parallel protein purification, Recombinant protein, Escherichia coli

## MATERIALS

- His Gravitrap columns (Cytiva) supplier item 11003399 - for capture and purification of histidine tagged proteins, maximum binding capacity of 40 mg of tagged protein per column  
[His GraviTrap | Cytiva \(cytivalifesciences.com\)](#)
- PD-10 buffer reservoir (Cytiva) supplier item 18321603 - increases the His Gravitrap and PD-10 desalting column load volumes to 40mL  
[Buffer Reservoir | Cytiva \(cytivalifesciences.com\)](#)
- PD-10 column spin adapters (Cytiva) supplier item 28923245 - used to prevent wobbling of columns in nalgene racks  
[Spin Adapters for Gravity Columns | Cytiva \(cytivalifesciences.com\)](#)
- PD-10 desalting columns (Cytiva) supplier item 17085101 - used for rapid buffer exchange  
[PD-10 desalting columns packed with Sephadex G-25 resin | Cytiva \(cytivalifesciences.com\)](#)

-  Nalgene® Unwire® Test Tube Racks: Resmer® Manufacturing Technology, for 30mm tubes, white **Thermo Fisher Catalog #5970-0030**
-  AIM – Terrific Broth Base including Trace elements **Formedium Catalog #AIMTB0210**
-  Ultra Yield 2.5L Flask, Sterile **Generon Catalog #931136-B**

## Optional but useful

-  BENCHMIXER™ XL MULTI-TUBE VORTEXER **Benchmark Scientific Catalog #BV1010**

## Materials (1 L cultures) for Expression:

- Plates with LB-agar+antibiotics
-  1 L of autoclaved autoinduction TB + 20 g/L glycerol + antibiotics
-  1 mL of 10 % Antifoam 204 (Sigma) in ethanol
-  2.5 L Ultra Yield flasks (fitted with loose foil cover\*\*)

## Materials (1 L cultures) for Purification:

- **1L of Base Buffer**

A	B
HEPES	10 mM
Glycerol	5%
NaCl	500 mM
TCEP, pH 7.5	0.5 mM

- **100 mL** of **[M] 3 Molarity (M)** imidazole pH 7.5.
- **100 mL** of 10 % Triton X-100 in water.
- **50 undetermined** Lysozyme solution (100 x).
- **1 undetermined** homemade benzonase (1000x). Maybe substituted for 10 mg/mL of commercial DNase I
- 2 x His GraviTrap column per litre of culture to be purified (Cytiva) fitted with LabMate extender (Cytiva) and PD-10 spin adapter (Cytiva) in 24 place Nalgene rack.
- 2 x PD-10 desalting column per litre of culture to be purified fitted with LabMate extender and PD-10 spin adapter in 24 place Nalgene rack.
- 2 x 50 mL centrifuge tubes per litre of culture to be purified in a 24 place Nalgene rack.

## SAFETY WARNINGS

! Triton x-100 is currently restricted for use in the EU and cannot be used without an exemption certificate REACH Annex XIV (Jan 2021). It can be readily substituted with IGEPAL CA-630 (which is likely to be subject to the same restrictions in the near future). Alternatives that also maybe used are Tergitol 15-S-9 or Tween-20 or octyl glucoside.

## Expression

12h 20m

- 1 Either transform *BL21 (DE3)* with the appropriate plasmid OR streak from glycerol stock onto agar plate and **incubate**  **Overnight**  **37 °C** \*.





## Note

\* Freshly transformed or re-streaked cells always give better yields than growing overnights directly from frozen glycerol stocks.

2

Grow 10 mL Overnight in 50 mL tube of each clone in superbroth + 1 % glucose + the appropriate antibiotics.

4h



3 Use 10 mL to inoculate 1 L AIM-TB (+ Antibiotics + Antifoam 204) in a baffled flask.

4

Grow 250 rpm, 37°C, 04:00:00 shaking using loose foil cover\*\*.

4h

**AERATION IS ESSENTIAL!**



## Note

\*\*An upturned 500 mL plastic beaker with a 2 mL microcentrifuge tube taped to the side of the flask to act as a spacer can also be used.

\*\*A breathable membrane such as an AirOtop enhanced flask seal may also be used.

5

Grow 40-48 h 250 rpm, 18°C shaking.



6

Harvest at 4000 x g, 4°C, 00:20:00 .

20m



7

Scrape out pellet and place in plastic polygrip bag and freeze -80 °C .

**Note**

Final wet cell weight is typically 50 g/L of culture

## Cell lysis

3h 30m

**8** Place polygrip bag on flat surface and smash cell pellet into small pieces and pour into 500 mL beaker.

**9** Add **3 mL** Base Buffer/g cell pellet ( **[M] 10 millimolar (mM)** HEPES, **[M] 500 millimolar (mM)** NaCl, 5% Glycerol, **[M] 0.5 millimolar (mM)** TCEP, **pH 7.5** ) + **0.5 undetermined** Lysozyme, **1 undetermined** Benzonase or **10 undetermined** DNase I, 1 % Triton X-100\*\*\*, **[M] 20 millimolar (mM)** imidazole.

**Note**

\*\*\*Triton x-100 is currently restricted for use in the EU and cannot be used without an exemption certificate REACH Annex XIV (Jan 2021). It can be readily substituted with IGEPAL CA-630 (which is likely to be subject to the same restrictions in the near future). Alternatives that also maybe used are Tergitol 15-S-9 or Tween-20 or octyl glucoside.

**10** Use stripette to dissolve pellet and put up to **45 mL** in a 50 mL tube (4 tubes in total).

**11** Leave **00:30:00** **Room temperature**.

30m

**12** Freeze **-80 °C** 1-2 h or overnight if preferred.



13 Thaw in Room temperature water bath 01:00:00 and mix.

1h



14 Centrifuge 4000 x g, 4°C, 01:00:00.

1h



#### Note

Higher speed centrifugation can also be performed if desired, e.g. 20,000 g but transfer to suitable centrifuge tubes will be necessary.

## Purification

3h 30m

15 Apply SN from 2 x 50 mL tubes to 1 mL His GraviTrap column (Cytiva) fitted with LabMate extender.

#### Note

Imidazole concentration can be increase to 40 mM in most cases, but may affect yield.

16 Wash 10 mL Base Buffer + 20 millimolar (mM) Imidazole\*\*.



#### Note

\*\*10 mL of a 40 mM or 70 mM imidazole wash can also be done, but this is very target dependent and may lead to significant reduction in final yield BUT can also increase purity substantially, worth trying if your purity is poor.

17 Slot His GraviTrap column into PD10 column (Cytiva) fitted with LabMate extender (pre-equilibrated in Base Buffer + 20 millimolar (mM) Imidazole).

18 Elute protein with 2.5 mL of Base Buffer + 500 millimolar (mM) Imidazole directly onto PD10 column.

19 Remove His GraviTrap column.

20 Place PD10 into 50 mL falcoln tube and add  3.5 mL Base Buffer +  20 millimolar (mM) Imidazole and collect.

21 Measure A280.

22 Add protease 1 OD unit TEV for every 10 OD units target and incubate  Overnight  4 °C . 

 Note

Some targets exhibit significant affinity for IMAC columns even after TEV cleavage try increasing the imidazole concentration to 40 or 70 mM or use an MBP-TEV construct so that the protease can be removed using an amylose column rather than reverse IMAC.

23 Run back over His GraviTrap column equilibrated in Base Buffer +  20 millimolar (mM) Imidazole.

24 Wash column 2.5 mL  20 millimolar (mM) Imidazole.



25 Check purity of  6 mL pool.

- 26 Concentrate to  1 mL ish.
- 27 Transfer to 1.6 mL glass autosampler vial ensure at least 1.1 mL in vial!.
- 28 Run through serial gel filtration system injecting  1 mL .
- 29 Take peak fraction(s) only (1-2 mL) and concentrated to 10-20 mg/mL if possible.

### Column regeneration: PD-10

- 30 Wash PD-10 columns with  50 mL  100 mL of Milli-Q water.  

- 31 Store all columns in water at  4 °C . For long term storage use 20 % Ethanol

### Column regeneration: His GraviTrap

- 32 Wash IMAC columns  40 mL Milli-Q.



33 Wash IMAC columns 10 mL 20 % Ethanol + 0.1 Molarity (M) EDTA\*.



\*PUT NICKEL WASTE IN APPROPRIATE CONTAINER FOR DISPOSAL!

34 Wash IMAC columns 40 mL Milli-Q.



35 Wash IMAC columns 10 mL 1 Molarity (M) NaOH.



36 Wash IMAC columns 40 mL Milli-Q.



37 Wash IMAC columns 10 mL 1 Molarity (M) Acetic Acid + 1 % Triton X-100.



38 Wash IMAC columns 40 mL Milli-Q.



39 Wash IMAC columns 0.5 mL 100 millimolar (mM) Nickel Sulfate + 20 millimolar (mM) Tris.HCl pH 8\*.

Note

\*PUT NICKEL WASTE IN APPROPRIATE CONTAINER FOR DISPOSAL!

40

Wash IMAC colums

40 mL

Milli-Q.



41

Store all columns in water at

4 °C

. For long term storage use 20 % Ethanol