




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# Support Protocol 2: Metagenotyping with a Custom Collection of Genomes

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1 Works for me

 Share[dx.doi.org/10.17504/protocols.io.dm6gpbp65lzp/v1](https://dx.doi.org/10.17504/protocols.io.dm6gpbp65lzp/v1) miriam.goldman

## ABSTRACT

This protocol describes how to build a MIDASDB from a custom collection of genomes and perform SNV metagenotyping with it. Other MIDAS2 commands can also be run with the new database. Single-sample SNV metagenotyping is shown as an example. There are three steps: construct a custom rep-genome database for a collection of representative genomes of interest, build a bowtie2 index, and execute run\_snps command with the prebuilt bowtie2 index.

## DOI

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## PROTOCOL CITATION

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## COLLECTIONS

 **MIDAS 2 Protocol**

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PARENT PROTOCOLS

Part of collection

[MIDAS 2 Protocol](#)

- 1 Download the example genome collection folder from Zenodo to the work directory (midas2\_protocol)

```
wget https://zenodo.org/record/6774976/files/midasdb_custom.zip  
unzip midasdb_custom.zip
```

We have prepared two genomes for two species from the 21 NCBI genomes in the HMP mock community: *Staphylococcus aureus* (GCF\_000013425.1) and *Staphylococcus epidermidis* (GCF\_006094375.1) for this protocol.

- genomes.tsv: the table-of-contents file specifying the assignment of representative genomes to species. We randomly assign each species a six-digit species\_id.
- cleaned\_imports/: the FASTA file of each representative genome, saved in the directory <species>/<genome>/<genome>.fna.
- metadata.tsv: taxonomic assignment of the randomly assigned species\_id.

MIDAS2 reserves the --midasdb\_name newdb for building any new MIDASDB, and the custom MIDASDB will be built at --midasdb\_dir midasdb\_custom

- 2 Construct rep-genome component of MIDASDB  
Annotate all the genomes and build the files needed for the rep-genome database. These commands should be executed in the work directory midas2\_protocol.

```
midas2 annotate_genome --species all \  
--midasdb_name newdb --midasdb_dir midasdb_custom \  
--debug --force
```

```
midas2 build_midasdb --generate_gene_feature \  
--genomes all \  
--midasdb_name newdb --midasdb_dir midasdb_custom \  
--debug --force
```

There are two command-line parameters that users need to pass:

--debug: keep the local file after successfully build the database

--force: re-build the database even if one already exists locally

### 3 Build one bowtie2 index with the representative genomes.

```
midas2 build_bowtie2db \  
--midasdb_name newdb --midasdb_dir midasdb_custom \  
--species_list 100001,100002 \  
--bt2_indexes_name repgenomes \  
--bt2_indexes_dir bt2_index_custom \  
--num_cores 8
```

We build the rep-genome bowtie2 index for the two species specified via --species\_list to the local directory bt2\_index\_custom/. Note we need to provide the custom MIDASDB midasdb\_custom/ to --midasdb\_dir.

Users can also specify --bt2\_indexes\_name pangenesomes to build the bowtie2 index for pangenesomes.

### 4 Execute run\_snps with the rep-genome database. For each sample, this code performs single-sample SNV calling for all the species in the bowtie2 database without any species filters (--select\_threshold=-1). The number of CPUs used is specified via --num\_cores 8.

```

for sample_name in SRR172902 SRR172903
do
    midas2 run_snps \
    --sample_name ${sample_name} \
    -1 reads/${sample_name}.fastq.gz \
    --midasdb_name newdb --midasdb_dir midasdb_custom \
    --prebuilt_bowtie2_indexes bt2_index_custom/repgenomes \
    --prebuilt_bowtie2_species bt2_index_custom/repgenomes.species \
    --select_threshold=-1 \
    --num_cores 8 midas2_output_custom
done

```

- 5 Confirm midas2 run\_snps has finished successfully.  
Once the single-sample SNV analysis is complete without any reported error, check for the output files (see Basic Protocol 3).