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Assessing IL-15 bioavailability ("the bioassay")

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1 Works for me



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ABSTRACT

Assessing the capacity of IL-15 analogs (e.g. within TriKE molecules) to stimulate proliferation of NK-92 cells. NK-92 are deprived of cytokines overnight, then cultured with IL-15 analogs for two days. After this time, the expansion of viable cells is quantitatively assessed using a redox-sensitive dye that changes from blue/non-fluorescent in media to pink/fluorescent upon reduction in viable cells.

It should be noted that NK-92 lack CD16, so delivery of IL-15 to these cells should not be enhanced by the anti-CD16 component of the TriKE as it would be for CD16+ NK-92 or CD16+ pNK cells.

This protocol is adapted from the following assays:

<https://linkinghub.elsevier.com/retrieve/pii/S0022175994903964>

<http://doi.wiley.com/10.1046/j.1440-1711.1998.00733.x>

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MATERIALS TEXT

MATERIALS

[96 well black assay plate with clear](#)

[bottom Corning Catalog #3603](#)

[Resazurin R&D](#)

[Systems Catalog #AR002](#) Step 6

[NK-](#)

[92 ATCC Catalog #CRL-247](#) Step 1

NK-92 culture

- 1 Defrost NK-92 (malignant non-Hodgkin's lymphoma; see *Cell Line Information*) and culture for at least one week prior to initiation of the assay.

[NK-](#)

[92 ATCC Catalog #CRL-247](#)

1.1 NK-92 media

- Alpha Minimum Essential Medium plus ribonucleosides and deoxyribonucleosides (Gibco Cat. No. 12571)
- 0.1 mM 2-mercaptoethanol (Sigma Aldrich, Cat. No. M7522)
- 12.5% horse serum (Fisher Scientific Cat. No. 26050088)
- 12.5% fetal bovine serum (FBS; Gibco Cat. No. 26140079)
- 100 U/mL penicillin streptomycin (Gibco Cat. No. 15140122)

For normal culture, supplement with 500 U/mL recombinant human IL-2 (Prometheus, Cat. No. NDC 65483-116-07).

[2-Mercaptoethanol Sigma](#)

[Aldrich Catalog #M7522](#)

[Horse Serum, heat inactivated, New Zealand origin Thermo](#)

[Fisher Catalog #26050088](#)

[Fetal Bovine Serum, qualified, United States Thermo](#)

[Fisher Catalog #26140079](#)

[Penicillin-Streptomycin Gibco - Thermo](#)

[Fisher Catalog #15140122](#)

[Proleukin \(recombinant IL-2\) Contributed by](#)

[users Catalog #National Drug Code 65483-116-07](#)

1.2 NK-92 culture conditions:

Humidified incubator at 37.0°C, 5% CO₂

Culture conditions: Replace medium every 2 - 3 days

Passage interval: Passage 2 - 3 times a week

Sub-cultivation ratio: 1:2 - 1:3

Seeding conditions: 2×10^5 - 3×10^5 cells/mL

1.3 NK-92 freezing protocol:

- Freeze medium: 50% FBS; 40% NK92 media; 10% DMSO
- Freeze no more than 5×10^6 cells/mL
- Storage temperature: liquid nitrogen vapor phase

Day - 1

2 Deprive NK-92 of IL-2 at least 16 h prior to plating cells.

2.1 Count NK cells (at least 4.8 million will be required for a full plate).

2.2 Spin NK-92 at 1600 RPM for 5 min and remove the supernatant.

2.3 Resuspend cells in media without IL-2.

2.4 Spin NK-92 at 1600 RPM for 5 min and remove supernatant.

2.5 Resuspend cells at 7.5×10^5 cells/mL in media without IL-2 and return to the incubator overnight.

Day 0


3 Dilute all drugs in media (without IL-2) to 5x the highest concentration to be tested (e.g. 150 nM). This will be used to make a serial dilution of the drug (3x, 5x or 10x depending on range required).

4 Plate NK-92 in triplicate into a 96 well black assay plate at 5×10^4 cells/well in 80 μ L of media without IL-2. Avoid using the wells at the edge of the plate as these undergo the greatest evaporation.

5 Add 20 μ L of diluted drug to each well containing NK-92. Include a no drug condition as a negative control.

Day 2

6 After 24 h, add 10 μ L resazurin to each well, mix and incubate for 1 h at 37°C and 5% CO₂.

 **Resazurin R&D**

Systems Catalog #AR002

- 7 Ensuring there are no air bubbles in the media, read the fluorescence at 530-570 nm, with a correction at 590-620 nm using a plate reader (Tecan).
- 8 Replace the plate in the incubator and wait 1 h before repeating step 7. Repeat this process to obtain four readings in total. Select the most stable readings for analysis (usually 3 h).
- 9 Analyze the adjusted readings using Graphpad Prism. Log transform the data and calculate a non-linear fit using "log(agonist) vs. normalized response -- Variable slope - Least squares fit" in order to obtain an EC 50.

Prism 8.0 [↗](#)

by GraphPad