

Sep 13, 2024

Preparation of B cells for scRNAseq

DOI

dx.doi.org/10.17504/protocols.io.n92ld8e1nv5b/v1

Nicholas Pease¹

¹University of Pittsburgh



Nicholas Pease

University of Pittsburgh

OPEN  ACCESS



DOI: **dx.doi.org/10.17504/protocols.io.n92ld8e1nv5b/v1**

Protocol Citation: Nicholas Pease 2024. Preparation of B cells for scRNAseq. **protocols.io**

<https://dx.doi.org/10.17504/protocols.io.n92ld8e1nv5b/v1>

License: This is an open access protocol distributed under the terms of the **[Creative Commons Attribution License](#)**, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

Protocol status: Working

We use this protocol and it's working

Created: September 13, 2024

Last Modified: September 13, 2024

Protocol Integer ID: 107559

Abstract

This protocol details activated B cell preparation and sample hashtagging for Chromium Single Cell Gene Expression (10x Genomics).



Materials

- EasySep™ Dead Cell Removal (Annexin V) Kit (STEMCell Technologies, Cat. # 17899)
- TotalSeq™-C0262 anti-human Hashtag 12 Antibody (Biolegend, Cat. # 394683)
- TotalSeq™-C0263 anti-human Hashtag 13 Antibody (Biolegend, Cat. # 394683)
- TotalSeq™-C0264 anti-human Hashtag 14 Antibody (Biolegend, Cat. # 394683)
- TotalSeq™-C0265 anti-human Hashtag 15 Antibody (Biolegend, Cat. # 394683)



Dead cell removal

- 1 Pellet cells at 300xg for 10min
- 2 Resuspend in 100uL Dead Cell Removal Buffer and filter through 40uM cell strainer
- 3 Add 5uL of Annexin V Cocktail to sample
- 4 Add 5uL of Biotin Selection Cocktail to sample, mix and incubate at RT for 3min
- 5 Vortex RapidSpheres for 30 sec and add 10uL to each sample, flick to mix
- 6 Add 2.4mL of Dead Cell Removal buffer and mix again
- 7 Place tubes on magnet and incubate at RT for 10 min
- 8 Transfer supernatant to new 15mL tube
- 9 Aliquot

Sample hashtagging

- 10 Resuspend samples in 100uL of wash buffer (PBS + 10% FBS)
- 11 Add 1uL of HTO antibodies
- 12 Incubate for 30 min on ice



- 13 Add 3.5mL of wash buffer to labeled cells, mix and pellet at 200xg for 10min at 4C
- 14 Remove supernatant with pipette, resuspend pellet with 3.5mL of wash buffer, and transfer to new tube
- 15 Pellet at 200xg for 10min at 4C
- 16 Remove supernatant with pipette and resuspend pellet with 3.5mL of wash buffer
- 17 Pellet at 200xg for 10min at 4C
- 18 Assuming 50% cell loss, resuspend pellet in wash buffer to obtain ~1,000 cells/uL
- 18.1 Count cells and adjust concentration, if needed

10x loading

- 19 Mix hashtag samples at desired ratio and load 52,500 cells per well