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General Taq PCR Master Mix -- CHEM 384/584 V.2

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SapphireAmp Fast PCR Master Mix contains a hot start PCR enzyme, optimized buffer, dNTP mixture, gel loading dye (blue), and a density reagent as a 2X premix. SapphireAmp Fast PCR Master Mix is optimized for fast PCR and offers a rapid extension rate (10 sec. per kb). The inclusion of blue dye and a density reagent allows direct loading of PCR products on an agarose gel for electrophoresis. The master mix format simplifies workflows and sample handling; simply add primers, template, and water and then begin PCR. SapphireAmp Fast PCR Master Mix is ideal for fast colony PCR screening. Fast colony PCR amplification of a 5 kb insert can be completed in approximately 1 hr 15 min. Furthermore, it is possible to amplify fragments up to 6 kb from genomic DNA templates.

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protocols.io<https://dx.doi.org/10.17504/protocols.io.b3gjqqjun>

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protocol

https://www.takarabio.com/assets/documents/User%20Manual/RR350A_DS.v1902Da.pdf

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Storage: -20°C for long-term storage. 4°C for short-term storage (up to 3 months).
(Note) If used frequently, store at 4°C ; the activity of the Master Mix may decrease with repeated freezing and thawing. Gently mix well before use and centrifuge briefly.

Application:

- DNA amplification by PCR
- Colony PCR

PCR Products: Since most PCR products amplified with SapphireAmp Fast PCR Master Mix have an A overhang added at 3'-termini, the obtained PCR product can be used directly for cloning into a T-vector. Additionally, it is possible to clone the product in a blunt-end vector after blunting and phosphorylation of the end.

Dye Migration During Electrophoresis: When 5 µl of the PCR sample is loaded on a 1% gel made with Agarose L03 [TAKARA] (Cat. #5003) and subjected to electrophoresis, the blue dye fronts are detected at positions corresponding to 1 kb and 3 - 5 kb. The absorption maxima for the dyes are ~ 260 nm and 620 nm, respectively. The dyes may be removed by isolating and purifying the DNA fragment from the gel or extracting DNA with NucleoSpin Gel and PCR Clean-Up (Cat. #740609.50/.250), if necessary

Setup Reaction

- 1 To a **25 µL** aliquot of a 2X Taq PCR Master Mix (e.g. TaqDog, or Sapphire Amp), add template (10-20 µl cleared lysate for colony PCR or **20-50 ng** of purified DNA for typical PCR), forward and reverse primers to a final concentration of **200 nanomolar (nM)** .
Adjust final volume to **50 µL** with nuclease free water or autoclaved water.

A	B
2X Master Mix	25 ul (pre- aliquoted and stored in the freezer)
Template	10-20 ul of bacterial lysate or 20-50 ng DNA
Forward Primer	1 ul of 10 uM primer dilution
Reverse Primer	1 ul of 10 uM primer dilution
ddH2O	to a final volume of 50 ul

Run Reaction

2 followed by 30 cycles of 98°C, 5 sec; 55°C, 5 sec; and 72°C, 40 sec.

A	B	C
Initial denature	98C	1 minute
Denature	94C	10 seconds
Anneal	55C	30 seconds
Extension	72C	1 min/kb
Repeat steps 2-4		30-40x
Final extension	72	1 minute
Cool	4C	Until cancelled

A typical thermocycling program for a PCR for amplicons less than 1 kb is 1 minute. For longer amplicons, adjust the program to 1 minute/kb seconds for the extension and final extension times.

