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© Protocol for Dephosphorylation of 5' ends of DNA using Quick CIP (NEB #M0525)

New England Biolabs¹

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1 Works for me

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ABSTRACT

Quick CIP is a heat-labile version of calf intestinal alkaline phosphatase (CIP) purified from a recombinant source.

- Rapid and irreversible heat inactivation eliminates unwanted activity
- Improved storage stability versus native enzyme
- Faster reaction setup (no supplemental additives like zinc required) and shorter incubation time
- Flexible reaction conditions (active in any restriction enzyme buffer, no clean-up required)
- Less enzyme required (high specific activity), resulting in a lower cost per reaction
- No need for multiple phosphatases (Quick CIP removes 5'- and 3'- phosphates from DNA, RNA and dNTPs)
- Active on unincorporated dNTPs in PCR products improves DNA sequencing and SNP analysis
- Recombinant for purity, consistency and value

EXTERNAL LINK

https://neb.com/protocols/2019/06/04/protocol-for-dephosphorylation-of-5-ends-of-dna-using-quick-cip-neb-m0525

MATERIALS

NAME	CATALOG #	VENDOR
Quick CIP	M0525	New England Biolabs
CutSmart® Buffer	B7204S	New England Biolabs

SAFETY WARNINGS

Please see SDS (Safety Data Sheet) for hazards and safety warnings.

BEFORE STARTING

Dephosphorylation of DNA 5'-ends using Quick CIP in a Restriction Enzyme Reaction

- The phosphatase can be added directly into the digestion reaction during or after DNA digestion
- Add $\Box 1 \mu l$ of Quick CIP for every 1 pmol of DNA ends (about $\Box 1 \mu g$ of a 3 kb plasmid) and incubate at

§ 37 °C for 10 minutes

- Quick CIP is active in all NEB restriction enzyme buffers
- The restriction enzyme should be heat inactivated at the same time as the phosphatase after digest and dephosphorylation
- If restriction enzyme(s) cannot be heat inactivated, DNA purification is required before ligation



1 Prepare a **□20 µI** reaction as follows:

DNA	1 pmol of DNA ends*
CutSmart® Buffer (10X)	2 μΙ
Quick CIP	1 μΙ
H20, purified	to 20 μl**

^{*} Note: 1 pmol of DNA ends is about 1 μg of a 3 kb plasmid.

2 Incubate at § 37 °C for © 00:10:00.

3 Stop reaction by heat-inactivation at 880 °C for © 00:02:00.

^{**} Scale larger reaction volumes proportionally.