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Binary HPLC method for determination of 4-hydroxybenzoic acid (4-HBA), chlorogenic acid (CGA), quercetin (QUE), and salicylic acid (SAL)

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ABSTRACT

We cut 1 g of each sample in such way that it contained the cuticle from one side of the sample. We homogenized it with 35 ml of the methanol 100%, and left them in ultrasonic 6 l bath during 30 min at room temperature. Methanol extracts obtained were filtered, placed in amber bottles to avoid degradation by light, and stored at – 20°C until processed. We used Waters 717 liquid chromatograph with autosampler, Waters 2487 HPLC Absorbance UV-Vis Detector, Waters 1525 Binary HPLC Pump, Waters control module with SAT/IN Bus (Waters, Milford, MA, USA), Symmetry HPLC C18 column (particle size 5 µm, length 250 mm, internal Ø = 4.6 cm; Waters, Milford, MA, USA). We filtered the extracts with 0.45 µm pore size nylon-membrane filter. To obtain the calibration curves we used standards for salicylic acid (SAL), 4-hydroxybenzoic acid (4-HBA), chlorogenic acid (CGA), and quercetin (QUE), all supplied by Sigma-Aldrich. The mobile phase consisted of 0.1% v/v acetic acid (A) together with 100% acetonitrile (B). For the mobile phase A, we dissolved 1 ml of glacial acetic acid with HPLC water, until 1 L. For the mobile phase B we used 100% acetonitrile. We filtered both mobile phases with 0.45 µm nylon membrane. We degasified them with ultrasonic bath during 30 min. We put the column temperature at 25°C, used the 254 nm UV detector, established the flow of the mobile phase at 0.2–0.8 mL/min, injection volume at 8 µL, and run time at 35 min. To wash the piston seals we used MeOH : H₂O (60 : 50). To quantify the four compounds, we used the calibration curves prepared out of 10 mg stock solution in 5 mL of MeOH. We took known concentrations from these solutions to obtain the relationship between the area corresponding to absorbance and concentration of each metabolite.

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KEYWORDS

4-hydroxybenzoic acid, 4-HBA, chlorogenic acid, CGA, quercetin, QUE

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Extraction of secondary metabolites

- 1 We cut 1 g of each sample in such way that it contained the cuticle from one side of the sample. We homogenized it with 35 ml of the methanol 100%, and left them in ultrasonic 6 l bath during 30 min at room temperature.

 **35 mL methanol**

[M] **100 % volume methanol**

 **00:30:00**

Mobile phase composition

- 2 The mobile phase consisted of 0.1% v/v acetic acid (A) together with 100% acetonitrile (B). For the mobile phase A, we dissolved 1 ml of glacial acetic acid with HPLC water, until 1 L. For the mobile phase B we used 100% acetonitrile.

Filtration of mobile phases

- 3 We used 0.45 µm nylon membrane.

Degasification of mobile phases

- 4 We degasified them with ultrasonic bath during 30 min.

Column temperature

5 We put the column temperature at 25°C

UV detection light frequency

6 We used the 254 nm UV detector

Mobile phase flow rate

7	Time	Flow rate (ml/min)	%A (acetic acid)	%B (acetonitrile)
	0.00	0.30	80.0	20.0
	5.30	0.30	70.0	30.0
	11.00	0.20	65.0	35.0
	13.00	0.20	50.0	50.0
	13.30	0.20	40.0	60.0
	15.00	0.20	30.0	70.0
	16.00	0.30	40.0	60.0
	17.00	0.70	50.0	50.0
	19.00	0.70	65.0	35.0
	22.0	0.80	70.0	30.0
	30.0	0.80	80.0	20.0
	35.0	0.80	80.0	20.0

Calibration curves

8 To quantify the four compounds, we used the calibration curves prepared out of 10 mg stock solution in 5 mL of MeOH. We took known concentrations from these solutions to obtain the relationship between the area corresponding to absorbance and concentration of each metabolite. We obtained the following calibration curves:

$$y_1 = 1109.4x_1 + 481.67,$$

$$y_1 = 296.01x_1 + 133.74,$$

$$y_1 = 551.41x_1 + 263.64, \text{ and}$$

$$y_1 = 919.96x_1 + 201.64,$$

where y_1 – area below the absorbance curve, x_i – concentration of the secondary metabolite, and $i = 1, 2, 3$ or 4 for 4-hydroxybenzoic acid (4-HBA), chlorogenic acid (CGA), quercetin (QUE), or salicylic acid (SAL), respectively.

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