



Jan 26, 2022

Making LB NGM Plates

Making LB NGM Plates

Priota Islam¹

¹Imperial College London

1



dx.doi.org/10.17504/protocols.io.b4c2qsye

Behavioural Genomics

e.warren

C. elegans is maintained in the laboratory on Nematode Growth Medium (NGM) agar which has been aseptically poured into petri plates. Sometimes they can also be grown on LB-NGM, where the LB in the media will promote better growth of the bacteria the worms feed on.

The LB-NGM agar medium can be poured into petri plates easily and aseptically using a peristaltic pump. This pump can be adjusted so that a constant amount of LB-NGM agar is dispensed into each petri plate. A constant amount of agar in the plates reduces the need for refocusing the microscope when you switch from one plate to another.

DOI

dx.doi.org/10.17504/protocols.io.b4c2qsye

Priota Islam 2022. Making LB NGM Plates. **protocols.io**
<https://dx.doi.org/10.17504/protocols.io.b4c2qsye>



Making LB NGM Plates, Priota Islam

protocol ,

Jan 26, 2022

Jan 26, 2022

57466

Reagents:

For 1000ml

A) Pre-Autoclave

Sodium Chloride- 3g, Sigma- Aldrich-71376-1KG

Bio Agar- 17g, Biogene- 400-050

LB Broth Powder- 2.5g, Fisher BioReagents- BP- 1426-2

Cholesterol (5mg/ml in EtOH)- 1ml, Sigma- C1145-250MG [Cholesterol should be stored at 4C away from light]

Sterile water- 975ml

B) Post-Autoclave

1M CaCl₂- 1ml, Sigma- C3881-1KG

1M MgSO₄- 1ml, Fisher- M/1050/53

1M KPO₄ (pH 6.0)- 25ml,

Pre-Autoclave:

- 1 Book the autoclave (notebook on top of the machine).
- 2 Take clean flasks from the glass kitchen (Only the ones with autoclave tape on are sterile)
- 3 Measure all the pre-autoclave reagents and add to the flask (Use a new weighing boat and spatula for each reagent. Also, the cholesterol is kept in the fridge.)
- 4 Once water is added mix thoroughly and label with autoclave tape ('LB NGM Rm 5020').
Make sure the bottle is not screwed completely when placing it inside the autoclave machine.

Using the Autoclave:

- 5 Turn ON the autoclave
- 6 Make sure that the autoclave's probe bottle is the same size as the largest bottle you use and fill it with water.
- 7 Place the temperature probe in it.

- 8 Fill up the autoclave with water until it reaches the grill.
- 9 Place the bottles in the autoclave and make sure that the cap is not screwed completely.
- 10 Check the waste flask is not too full
- 11 Use 'media' program.
- 12 Press START.
- 13 It will take about 2.5 hours for 1L or larger bottles.

Post-Autoclave:

- 14 When autoclave is complete, remove the probe flask
- 15 Make sure to wear gloves as the flask will be hot
- 16 Let the agar to cool to around 55°C, ie the bottle is cool enough to hold for a second with a gloved hand.
- 17 Add the post autoclave reagents.

- 18 Mix it well and start pouring onto desired sized plates (See Protocol for plate pouring)
- 19 Try not to shake the bottle too much while mixing to avoid air bubbles.
- 20 The agar needs to be warm to be poured without blocking the tubings, so try to pour as quickly as possible and if not poured immediately put the bottle on a waterbath set to 60C until being used.