



DEC 02, 2022

WORKS FOR ME

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## Expression and purification protocol of the Human (Homo sapiens) LC3B Ubiquitin-like modifier

Forked from [Expression and purification protocol of the Human \(Homo sapiens\) mCherry-LC3B Ubiquitin-like modifier](#)

DOI

[dx.doi.org/10.17504/protocols.io.j8nlkw82dl5r/v1](https://dx.doi.org/10.17504/protocols.io.j8nlkw82dl5r/v1)[Dorotea Fracchiolla](#)<sup>1</sup>, [Liv Jensen](#)<sup>2</sup><sup>1</sup>Team Hurley; <sup>2</sup>University of California, Berkeley

Liv Jensen

COMMENTS 0

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### ABSTRACT

This protocol describes expression and purification procedures for obtaining mCherry-tagged human recombinant Ubiquitin-like modifier LC3B (MAP1LC3B, Microtubule-associated proteins 1A/1B light chain 3B) lacking five C-terminal amino acids to allow in vitro protein conjugation to target PE, Phosphatidyl Ethanolamine.

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### PROTOCOL CITATION

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### FORK NOTE

#### FORK FROM

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#### KEYWORDS

expression, purification, recombinant, Microtubule-associated proteins 1A/1B light chain 3B, LC3B, mCherry-tag, ASAPCRN

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#### IMAGE ATTRIBUTION

© Dorotea Fracchiolla

#### CREATED

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#### LAST MODIFIED

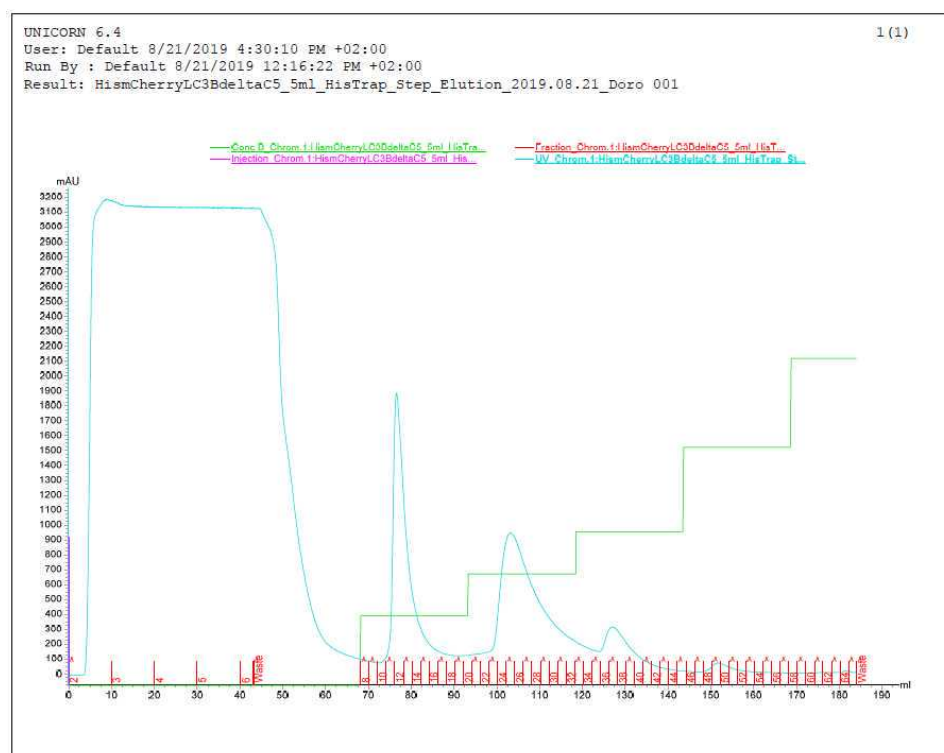
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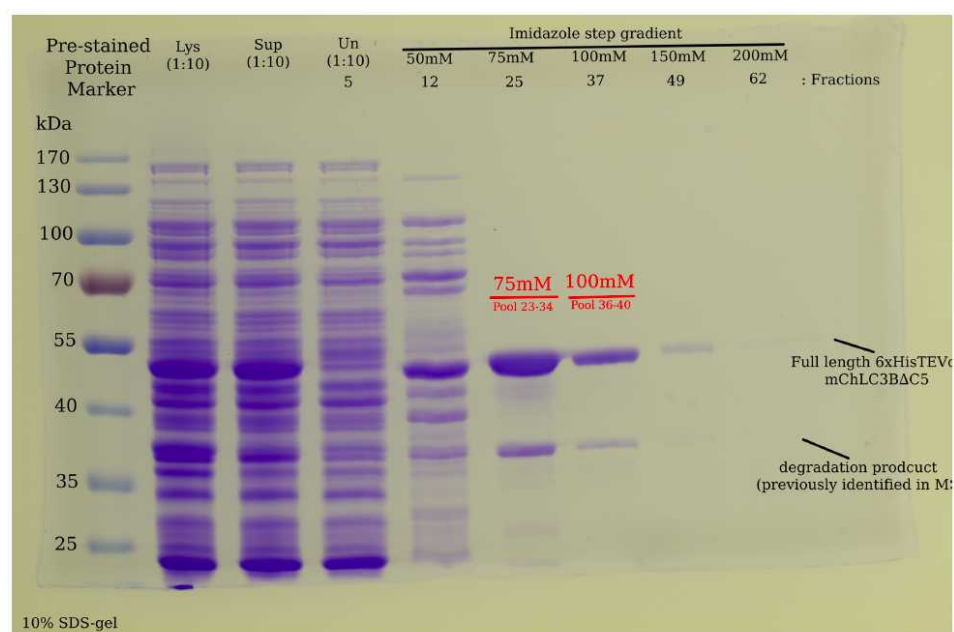
73497

#### GUIDELINES

**Affinity purification**

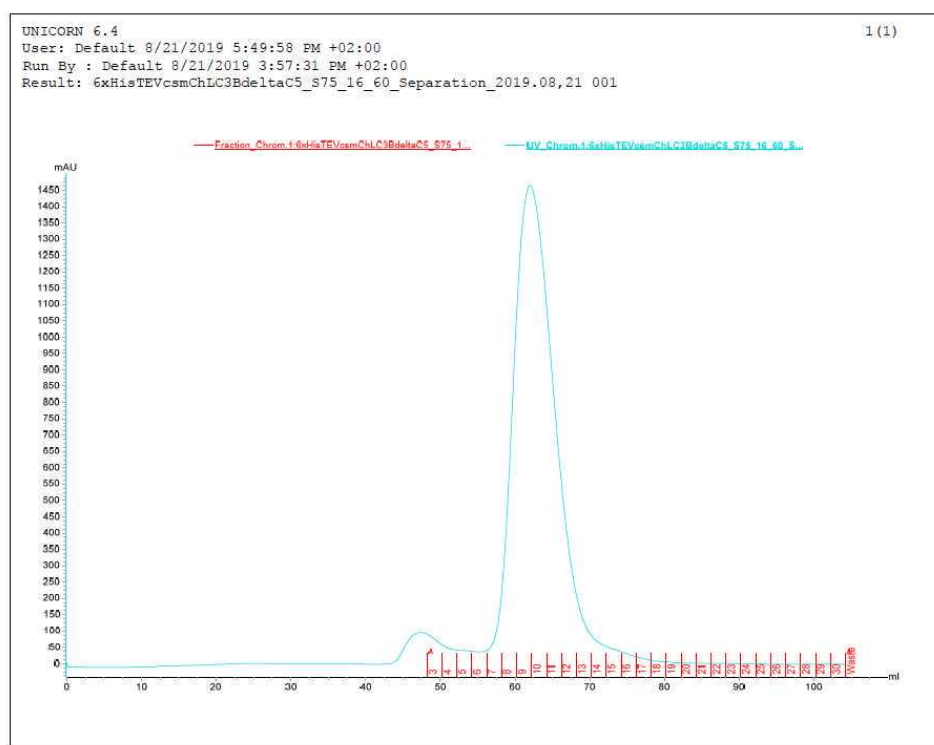


Chromatogram of His-tag affinity purification for mCherry-hLC3B.

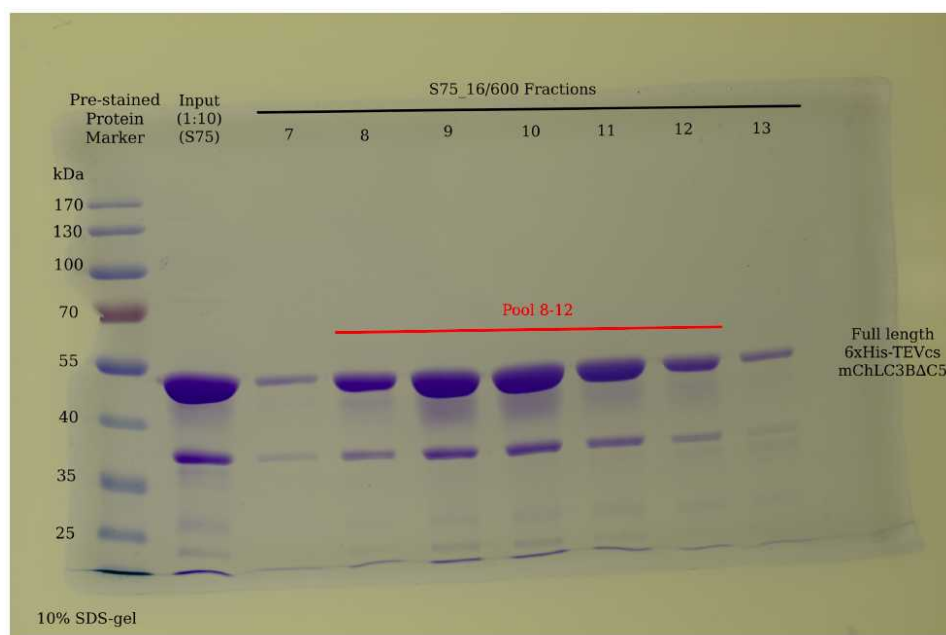


Coomassie BB stained gel of His-tag affinity purification for mCherry-hLC3B.

## Size Exclusion Chromatography



Chromatogram of Size Exclusion purification for mCherry-hLC3B.



Coomassie BB stained gel of Size Exclusion Chromatography purification for mCherry-hLC3B.

## MATERIALS TEXT

### Materials and Reagents

- Escherichia coli Rosetta pLyss cells
  - Luria Bertani (LB) medium with antibiotics (final conc. 50µg/ml Ampicillin, 34µg/ml Chloramphenicol)
  - IPTG (isopropyl-b-d-thiogalactopyranoside)
  - 37°C shaker incubator
  - sterile flasks/sterile pipettes
  - tip sonicator
- 
- **Lysis Buffer:** 50mM Hepes pH=7.5, 300mM NaCl, 10mM Imidazole, 2mM MgCl<sub>2</sub>, 2mM β-mercaptoethanol, 1mM Pefablock, Complete Protease Inhibitors (EDTA-free CIP tablet, Roche), DNase (Sigma).
  - **Buffer A:** 50mM Hepes pH=7.5, 300mM NaCl, 10mM Imidazole (filtered and degassed) + 1mM β-mercaptoethanol;
  - **Buffer B:** 50mM Hepes pH=7.5, 300mM NaCl, 300mM Imidazole (filtered and degassed) + 1mM β-mercaptoethanol;
  - **Size Exclusion Chromatography (SEC) Buffer:** 25mM Hepes pH=7.5, 150mM NaCl, 1mM DTT (Dithiothreitol).

Note: all purification buffers are filtered and degassed. Reducing agents (β-mercaptoethanol and Dithiothreitol) are added after degassing step.

**Columns:-** HT 5ml column (GE Healthcare)  
- S75\_16/60 (GE Healthcare)

**Gels:** 10% SDS-PAGE

## SAFETY WARNINGS

Please refer to the Safety Data Sheets (SDS) for health and environmental hazards.

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## BEFORE STARTING

### General information



Insert: Homo sapiens LC3B, [NP\\_073729.1](https://doi.org/10.17504/protocols.io.j8nlkw82dl5r/v1); Expression system: E.Coli Rosetta pLyss; plasmid origin: Sascha Martens Lab, Addgene 169168, lab internal construct database number SMC948; backbone: pET-Duet1; plasmid resistance: Ampicillin; tags & cleavage sites: N-term 6xHis, followed by Tobacco Etch Virus (TEV) cleavage site, mCherry tag, LC3B ORF. Ext coeff: 41830 M<sup>-1</sup> cm<sup>-1</sup>, MW 47,89 kDa.

**Citation:** Dorotea Fracchiolla, Liv Jensen Expression and purification protocol of the Human (Homo sapiens) LC3B Ubiquitin-like modifier <https://dx.doi.org/10.17504/protocols.io.j8nlkw82dl5r/v1>

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16h 15m

## Protein Expression




- 1 Transform plasmid DNA (Addgene\_190237) into E.Coli Rosetta pLyss cells and plate on Ampicillin LB agar plate for  Overnight at  37 °C .






- 2 The following day, inoculate a  5 mL LB + Amp/Cam pre-culture with 1-2 colonies and grow




 Overnight at  37 °C shaking.

- 3 The following day, use  5 mL pre-culture to inoculate  1 L LB medium + Amp/Cam at  37 °C until an OD<sub>600</sub> (Optical Density) of 0.6 is reached.


- 4 Induce protein expression with  400 micromolar (μM) IPTG and grow for a further  16:00:00 at  18 °C shaking.

16h

- 5 Pellet cells at  4000 rpm, 4°C, 00:15:00 in a Sorvall RC6+ centrifuge (Thermo Scientific), discard supernatant and resuspend pellets in ice cold lysis buffer (25 ml/1 lt culture).



15m

- 6 Flash freeze resuspended pellets in liquid nitrogen and store at  -80 °C until purification.






## Protein Purification


- 7 Perform His-Trap affinity purification followed by Size Exclusion Chromatography.

30m 30s


## Purification of LC3B Ubiquitin-like modifier

- 8 Cells are lysed via freeze/thaw cycles and sonication: thaw pellet corresponding to  1 L culture by


freeze/thawing in  Room temperature water bath. All following steps are to be executed at  4 °C or  On ice .

- 9 Lyse cells by sonicating them with an immersion Tip Sonicator (2x  00:00:30 ). Note: adjust times and intensity according to the available instrument.

30s

- 10 Clear lysate by spinning in a Beckman centrifuge, at  40000 x g, 4°C, 00:30:00 , Ti45 Rotor .

30m

- 11 Inject soluble fraction onto a 5ml HT column operating at  4 °C pre-equilibrated in Buffer A at 1 ml/min flow rate.


- 12 Wash column for  5 undetermined with Buffer A at 2 ml/min flow rate to remove unspecific bound proteins.

- 13 Elute protein at 300mM Imidazole concentration. Perform elution at 1ml/min flow rate.

- 14 Perform TEV cleavage with 1mg/ml TEV protease overnight at 4°C while dialyzing against elution buffer containing no imidazole.

- 15 Apply cleavage reaction to equilibrated HT column, and collect the flow-through.

- 16 Measure protein absorbance at A<sub>280</sub> with a Spectrophotometer against dialysis buffer.

- 17 Aliquot protein, flash freeze in liquid Nitrogen and store at  -80 °C .

