



Oct 27, 2022

O Determining MLST allele sequences in novel STs

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dx.doi.org/10.17504/protocols.io.36wgqj7kovk5/v1



ABSTRACT

Steps required to determine Novel allele sequence of MLST from the assembly files using MLSTaR R package

DOI

dx.doi.org/10.17504/protocols.io.36wgqj7kovk5/v1

DOCUMENT CITATION

Varun Shamanna 2022. Determining MLST allele sequences in novel STs. **protocols.io**

https://dx.doi.org/10.17504/protocols.io.36wgqj7kovk5/v1

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CREATED

Oct 27, 2022

LAST MODIFIED

Oct 27, 2022

DOCUMENT INTEGER ID

71866

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GHRU Determining MLST allele sequences in novel STs

There are many methods both manual and programmatic for achieving this. Here is just one method using an existing software tool - MLSTar (https://github.com/iferres/MLSTar)

- 1. First install the blast dependency. It is recommended to perform this using a conda package conda install -c bioconda blast
- 2. MLSTar is an R package so install R if not already installed (RStudio recommended). Then within the console in R/RStudio
- 3. Install the devtools package install.packages("devtools")
- 4. Install the MLSTar package devtools::install_github('iferres/MLSTar')

5.

6. This package will only work "out of the box" with MLST schemes on the PubMLST database/website, so for other schemes (e.g Klebsiella on the Pasteur website) you need to download the profiles and allele sequences.

For klebsiella the profiles can be found

https://bigsdb.web.pasteur.fr/cgi-bin/bigsdb/bigsdb.pl?db=pubmlst_klebsiella_segdef allele sequences can be found here

https://bigsdb.web.pasteur.fr/cgi-bin/bigsdb/bigsdb.pl?

db=pubmlst_klebsiella_seqdef&page=downloadAlleles

I have pre-downloaded them and zipped them up here

Please note of providing your own profiles and sequences check that the header names in the profiles TSV match the names of the sequence file prefixes and that there are no trailing tabs in either the header or profile rows.

```
e.g
sed -i 's/[[:space:]]*$//' kpn_profiles.txt
on Mac OSX
```

sed -i " 's/[[:space:]]*\$//' kpn_profiles.txt

- 1. Then to find the profiles and extract the sequences we run the following commands
- 2. Find supported organisms listPubmlst_orgs()
- 3. Find schemes for a supported organism

listPubmlst_schemes(org = "escherichia")

There are many schemes including some very long ones which are the cgMLST schemes, however you will see 2 familiar ones for E.coli

\$scheme_1

```
[1] "adk" "fumC" "gyrB" "icd" "mdh" "purA" "recA"
attr(,"Desc")
[1] "MLST (Achtman)"
```

\$scheme_2

[1] "dinB" "icdA" "pabB" "polB" "putP" "trpA" "trpB" "uidA"



```
attr(,"Desc")
  [1] "MLST (Pasteur)"
  1. To call MLST and write alleles to a file
     results <- doMLST(
     c("G18000002.fasta", "G18000051.fasta"),
     org = "escherichia",
     scheme = 1,
     write = "all")
     results$result
       adk fumC gyrB icd mdh purA recA ST
  G18000002 92 4 87 96 70 58 2 648
  G18000051 53 40 47 13 36 28 29 131
  1. The allele sequences are found in a directory as shown below. In this case there are no new alleles
     and so there is no need to examine these on the linux terminal (not R console)
     Is -l alleles_escherichia_1/
  total 28
                                                                            ()
  -rw-rw-r-- 1 biouser biouser 1196 Nov 2 11:07 adk.fasta
  -rw-rw-r-- 1 biouser biouser 1061 Nov 2 11:07 fumC.fasta
  -rw-rw-r-- 1 biouser biouser 1044 Nov 2 11:07 gyrB.fasta
  -rw-rw-r-- 1 biouser biouser 1160 Nov 2 11:07 icd.fasta
  -rw-rw-r-- 1 biouser biouser 1026 Nov 2 11:07 mdh.fasta
  -rw-rw-r-- 1 biouser biouser 1081 Nov 2 11:07 pur A. fasta
  -rw-rw-r-- 1 biouser biouser 1145 Nov 2 11:07 recA.fasta
  1. For other schemes the paths to the profiles and allele sequences need to be provided (in this case
     they are in a directory called mlst_scheme) and you'll need to provide a dummy organism e.g test
     since "klebsiella" is not an officially supported organism
     results <- doMLST(
     c("G18583057.fasta", "G18583075.fasta"),
     org = "test",
     scheme = 1,
     schemeFastas = c(
  "mlst_scheme/gapA.fas",
  "mlst_scheme/infB.fas",
   "mlst_scheme/mdh.fas",
  "mlst_scheme/pgi.fas",
  "mlst_scheme/phoE.fas",
   "mlst_scheme/rpoB.fas",
  "mlst_scheme/tonB.fas"),
  schemeProfile = "mlst_scheme/kpn_profiles.txt",
  write = "all")
  When looking at the result this time you will see that the profiles have an unknown allele 'u1'
  results$result
       gapA infB mdh pgi phoE rpoB tonB ST
  G18583057 2 5 u1 1 4 1 4 NA
  G18583075 2 5 u1 1 4 1 4 NA
protocols.io
```

Citation: Varun Shamanna Determining MLST allele sequences in novel STs https://dx.doi.org/10.17504/protocols.io.36wqqi7kovk5/v1

Or

If you have novel profile instead of a unknown allele 'u1' the result will be a profile with all allele numbers assigned but ST still NA since the alleles present represent a new combination.

gapA infB mdh pgi phoE rpoB tonB ST G18250048 3 3 1 1 1 1 4 NA

1. The novel allele sequences can be found in the output file cat alleles_test_1/mdh.fasta >mdh_u1;G18583057;NODE_2_length_734790_cov_17.426302 catcgacaaggtcgccgacccgccgccgctttcgcttccacgacttcggtaccggcgtt ctgaatacgtttagtcaggtcggcaatttcctgatcgctaaagctgacgccggggatctg cgacagtaaaggcagaatggtgaccccggagtgaccaccaatgaccgggacttccacctc ggttgccgatttacctttcagctccgccacaaaggtattggaacggatgatgtcaagcgt ggtaacgccgaacagtttgtttttatcgtacacgccggcttttttcagtacttcggcggc gatagccacggtggtattcaccgggttggtgataatgccgatgcaggcctgcgggcaggt tttggcaatctgctgcacgaggttcttcacgatacccgcattcacattaaacaggtcgga acgatccatgccgggcttacgcgccacgcccgcggagatcagcactacatccgcg >mdh_u1;G18583075;NODE_2_length_734148_cov_16.523686 cgcggatgtagtgctgatctccgcgggcgtggcgcgtaagcccggcatggatcgttccga cctgtttaatgtgaatgcgggtatcgtgaagaacctcgtgcagcagattgccaaaacctg cccgcaggcctgcatcggcattatcaccaacccggtgaataccaccgtggctatcgccgc tgacatcatccgttccaatacctttgtggcggagctgaaaggtaaatcggcaaccgaggt ggaagtcccggtcattggtggtcactccggggtcaccattctgcctttactgtcgcagat ccccggcgtcagctttagcgatcaggaaattgccgacctgactaaacgtattcagaacgccggtaccgaagtcgtggaagcgaaagcgggcggcgggtcggcgaccttgtcgatg