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Human fibroblast culturing

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Fibroblasts are cultured in Dulbecco's modified eagle media (DMEM) 4500 (mg/L) growth medium supplemented with Glutamax (Gibco), 10% foetal bovine serum (FBS), non-essential amino acids (NEAA: 0.1 mM of: glycine, L-alanine, L-asparagine, L-aspartic acid, L-glutamic acid, L-proline and L-serine) and penicillin/streptomycin antibiotic cocktail (50ng/ml) at 37°C and 5% CO₂.

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SAFETY:-**Biological Hazards:**

- The culturing of human cells is a class II activity and all appropriate safety issues need to be addressed.
- If using GMOs read the Laboratory Codes of practice for working with GMOs

Characteristics**1 Characteristics**

- Obtained /developed from human adult dermal biopsies

- Proliferative until p20
- Doubling time ~ 24h
- Maintain between 60 – 90% confluent

Complete growth medium

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 - DMEM (LT # 61965-059; 4500 mg/L and no pyruvate) Glutamax medium
 - Fetal bovine serum (FBS, 10% final) – 1 / 10 dilution
 - Pen / Strep (LT # 15140-122, 50 U/ml and 50 ng/ml final) – 1 / 200 dilution
 - MEM / NEAA (LT # 11140-035) – 1 / 100 dilution
 - As required – Sodium pyruvate (Sigma # S8636, 1 mM final) – 1 / 100 dilution
 - As required – Uridine (Sigma # U3003, stock of 25 mg/ml in H₂O filter-sterilised; 50 ug/ml final) – 1 / 500 dilution
 - As required, as permitted by DSO – Fungizone (LT # 15290-026; original of 250 ug/ml) – 1 / 100 dilution

Subculturing by trypsinisation, and cell counting

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 1. Wash cells grown onto 10 cm Ø plate with PBS.
 2. Add 1.5 ml diluted trypsin (4 ml of 2.5%, LT # 15090-046, into 100 ml Versene LT # 15040-033) and leave at 37°C for several mins.
 3. Tap the plates and add 2 ml complete medium to stop trypsination.
 4. To a 50 ul sample, add equal vol of 0.4% Trypan blue (Sigma # T8154 or LT # 15250-061); feed it to a hemacytometer.
 5. Optional – to pellet cells and resuspend into fresh medium.
 6. Replate the cells accordingly; typically 10 ml medium is used for 10 cm Ø plate.
 7. Subcultivation ratio of 1:4 is recommended; medium renewal every 2-3d.

Freezing and thawing

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 1. After centrifugation to pellet cells, resuspend in Freezing medium (90% FBS / 10% DMSO filter sterilised) and aliquot into freezing vials; typically ¼ of a 50% confluent 10 cm Ø plate of 125K cells in 0.5ml. Freeze cells in a stepwise fashion from -80°C to liq N.
 2. Thaw cells at 37°C.