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QGP-1 cell line maintenance protocol

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ABSTRACT

QGP-1 cell line maintenance protocol; thawing and passaging QGP-1 cells

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Thawing protocol

1 Warm media

2 Swirl cells in 37 degree water bath until vial is thawed

3 Add 7 mL of culture media to a T25 flask and gently pipette thawed cells into flask

4 Let cells adhere for 24 hours and replace half of the culture media with fresh media

5 Replace the media in the flask with fresh media after 48 hours

6 Split cells once cells reach 70-80% confluency: usually takes 3-4 days from thawing and plating in a T25 until they are ready for passage. (they won't ever become completely confluent, they will just start to grow on top of each other)

7 Lift cells using trypsin and plate all cells in a T75

8 They are usually ready to passage 3 days later.

passaging protocol

9

9.1 Warm media and trypsin in water bath

9.2 Clean hood with EtOH

9.3 Remove media

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9.4 Wash with HBSS

9.5 Discard HBSS, add Trypsin, and place flasks in incubator for 5 minutes

9.6 Bring flasks back to hood, obtain lifted cells and spin at 500 rcf for 5 minutes

9.7 Discard supernatant

9.8 Resuspend in 10 mL

9.9 Place 250 uL of cells into a T75 flask and add 13 ml of media

9.10 Place flask back in incubator and split next week

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