



Jul 23, 2021

Flow Cytometry ICS Nuclear Antigens V.2

Michael Betts¹, Gregory Golden¹

¹University of Pennsylvania

1 Works for me



dx.doi.org/10.17504/protocols.io.bwuupeww

Human Islet Research Network



ABSTRACT

High-parameter flow cytometry enables identification and characterization of a wide range of cell populations within a biological sample. A combined analysis of extracellular epitope staining (ECS) and intra-cellular epitope staining (ICS) using a collection of fluorophore-labeled antibodies sufficiently identifies discrete cell populations and their respective phenotypes. Importantly, ECS/ICS can be applied to cryo-preserved cell suspensions recovered in tissue culture media, enabling samples to be conveniently analyzed after collection and proper cryopreservation. However, consistent cryopreserved sample recovery and ECS procedures are critical to data comparison across multiple experiments. Herein, we describe a standardized protocol for cryopreserved sample recovery and ECS/ICS procedures for cell-surface and intra-cellular epitope labeling.

DO

dx.doi.org/10.17504/protocols.io.bwuupeww

PROTOCOL CITATION

Michael Betts, Gregory Golden 2021. Flow Cytometry ICS Nuclear Antigens. **protocols.io** https://dx.doi.org/10.17504/protocols.io.bwuupeww
Version created by Lili Liang

KEYWORDS

null, extracellular epitope staining (ECS), intra-cellular epitope staining (ICS), HIRN

LICENSE

This is an open access protocol distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Jul 23, 2021

LAST MODIFIED

Jul 23, 2021

PROTOCOL INTEGER ID

51828

Materials Required

- **⊠**1x phosphate saline buffer
- 1. (PBS) Corning Catalog #21-031-CM
 - ⊠ Bovine serum albumin (BSA) Gemini
- 2. Bioproducts Catalog #700-101P
 - Sodium Azide Fisher
- 3. Scientific Catalog #S2271-500
- 4. FACS buffer (1xPBS, 10g/L BSA, 1 g/L sodium azide)
- 5. RPMI Corning Catalog #10-040-CM
- 6. Bioproducts Catalog #900-108
 - ⊠ Penicillin/streptomycin 10000 U/mL penicillin 10000 μg/mL
- 7. streptomycin Lonza Catalog #17-602E

Lonza (RRID:SCR_000377)

8. "R10" medium (RPMI, 10% FCS, 1% penicillin/streptomycin)

⊠ DNAse

9. Roche Catalog #04716728001

⊠ Live/Dead fixable Aqua Dead Cell Stain kit Invitrogen - Thermo

10. Fisher Catalog #L34966

diluted 1:60 in 1x PBS (prepare fresh each day from DMSO stock)

11.

🛭 🛱 e-Bioscience Foxp3 / Transcription Factor Staining Buffer Set Invitrogen - Thermo

Fisher Catalog # 00-5523-00

12. Sciences Catalog #15712-S

diluted to 1% in PBS

- 13. Fluorophore-labelled antibodies of choice
- 14. Hemocytometer

Procedure

- | Thawing and Resting
 - a. Pre-warm R10 media in a § 37 °C water bath.

- **b.** Thaw samples in-vial using a § 37 °C water bath.
- c. Add thawed cells to 114 mL of R10, then spin cells at 500 xg for 5 min.
- **d.** Resuspend cell pellet in **3 mL** of R10 and count cells.
- e. Rest cells at least 3 hours (up to overnight) at $2x10^6$ cells/mL in R10 medium + 1 μ L/mL DNAse I at 8 37 °C , 5% CO₂

Note: during resting, prepare antibody cocktail master mix. Adjust volume of ECS for ■50 µl per test with FACS buffer

- f. After resting, add PBS up to □15 mL or □50 mL (whichever is closer, rounding up) to cells and transfer to □15 mL or □50 mL conical tube.
- g. Spin cells at 500 xg for 5 minutes at room temperature (RT).
- **h.** Resuspend cells in PBS to $1x10^7$ cells/mL and count. If cells are too dilute, re-spin cells and resuspend at $1x10^7$ cells/mL. Transfer 200 mL of cells $2x10^6$ cells) into each well of a V-bottom 96 well plate.

7 Viability and extracellular staining (ECS)

- a. Spin plate at 500 xg for 5 minutes at RT
- **b.** Using a multichannel pipette, carefully remove the supernatant.
- **c.** Add **5** μl of 1:60 Aqua viability dye directly to cell pellet and resuspend cells.
- d. Incubate for 10 minutes at RT in the dark.
- **e.** Add **50** μl of ECS antibody cocktail to cells and incubate for 20 minutes at RT in the dark (prepare 1% PFA fixation buffer in the meantime).
- f. Add 100 µl of FACS buffer to each well and spin plate at 500 xg for 5 minutes. Remove supernatant.

3 Fixation, Permeabilization, and ICS Staining

a. Fix cells with $\frac{100 \, \mu l}{100 \, \mu l}$ of 1xFixation/permeabilization buffer (1 part concentrate + 3 parts diluent) for 30 minutes at RT in the dark.

Note: Make ICS abs here

- **b.** Add **100** μl of 1x perm/wash buffer and centrifuge 800 xg for 5 min.
- c. Remove supernatant and add 1100 μl ICS cocktail (made in perm/wash buffer, made with diH₂0) to the cells.
- d. Incubate for 1h at RT in the dark.
- e. Add 100 µl perm/wash buffer. Centrifuge at 800 xg for 5 min.

 f. Discard supernatant and resuspend pellet in ■200 µl 1% PFA and transfer to FACS tubes.

 ${\bf g.}$ Wrap in aluminum foil and store at $\ \ {\bf \$} \ {\bf 4\ ^{\circ}C}$ until flow cytometry.