



DEC 17, 2022

WORKS FOR ME

1

Nucleoside analysis with high performance liquid chromatography (HPLC)

In 1 collection

DOI

dx.doi.org/10.17504/protocols.io.5jyl8jn39g2w/v1Atanas Radkov¹¹Arcadia Science

Arcadia Science



Arcadia Science

COMMENTS 0

ABSTRACT

This protocol details the detection of modified nucleosides using HPLC.

DOI

dx.doi.org/10.17504/protocols.io.5jyl8jn39g2w/v1

PROTOCOL CITATION

Atanas Radkov 2022. Nucleoside analysis with high performance liquid chromatography (HPLC) . **protocols.io** <https://dx.doi.org/10.17504/protocols.io.5jyl8jn39g2w/v1>

COLLECTIONS ⓘ

[Protocol collection: Phage DNA isolation and chemical analysis](#)

LICENSE

————— This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Sep 19, 2022

LAST MODIFIED

Dec 17, 2022

OWNERSHIP HISTORY

Sep 19, 2022  Megan Hochstrasser Arcadia Science

Sep 20, 2022  Arcadia Science

Sep 20, 2022  Arcadia Science

PROTOCOL INTEGER ID

70252

PARENT PROTOCOLS

Part of collection

[Protocol collection: Phage DNA isolation and chemical analysis](#)

Prepare samples and solutions

- 1 Prepare the following **standards** at 100 μ M by dissolving each in a small volume of HPLC-grade methanol and bringing up the volume as necessary with MQ water. Filter each solution through a 0.22 μ m syringe filter (we used SLGSV255F - Millipore-Sigma), and keep solutions at 4 °C prior to HPLC analysis.

Note

- 2 Prior to HPLC analysis, adjust the total **experimental nucleoside sample** volume to 100 μ L by diluting with MQ water and then centrifuge each sample at 21000 \times g for 15 min to remove any debris.

Note

Step 2 includes a Step case.

Long binary Short isocratic

step case

Long binary

Using a 30 minute-long binary gradient to resolve nucleoside peaks.

- 3 Prepare solutions A and B. Filter solution A through a 0.22 μm filter (we used 10040-440 VWR) and then adjust pH with 1 M HCl.

Note

Run samples

- 4 Equilibrate the column with solution A. Once fully equilibrated, inject 10 μL of one of the **standards**.

Note

- 5 Begin the gradient at 100% solution A, then ramp up to 25% solution B over 16 min, then ramp down to 0% solution B in 1 min, and finally stay at 0% solution B for 13 min (30 min total method time). Flow rate for the entire run should be 0.5 mL per min. Detect elution of compounds at 260 nm.
- 6 Run in triplicate to obtain an average retention time and a standard deviation. Repeat for each **standard**.
- 7 Make sure the HPLC column is re-equilibrated in solution A, then inject 10 μL of your **experimental sample**. Run the same program used for the standards. Perform in triplicate to obtain an average retention time and a standard deviation.

Analysis

- 8 Extract peak elution times from the HPLC software report. Common calculations can include calculating average elution time and standard deviation, if you have ran replicates. We also often plot the A_{260} signal to look at the shape of individual peaks.