



Aug 04, 2020

Low-input DNA extractions in 96-well plates

Forked from [Honeybee DNA extractions in 96-well plates](#)

Tom Harrop¹, Reuben McKay Vercoe¹, Ciarán Cuddy¹

¹University of Otago

1 *Works for me* dx.doi.org/10.17504/protocols.io.birqkd5w

Tom Harrop

DOI

dx.doi.org/10.17504/protocols.io.birqkd5w

PROTOCOL CITATION

Tom Harrop, Reuben McKay Vercoe, Ciarán Cuddy 2020. Low-input DNA extractions in 96-well plates.
protocols.io
dx.doi.org/10.17504/protocols.io.birqkd5w

FORK FROM

Forked from [Honeybee DNA extractions in 96-well plates](#), Tom Harrop

LICENSE

This is an open access protocol distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

CREATED

Jul 19, 2020

LAST MODIFIED

Aug 04, 2020

PROTOCOL INTEGER ID

39440

MATERIALS

NAME	CATALOG #	VENDOR
3.2 mm stainless steel beads, RNase free	NEXSSB32-RNA	
96 Well 0.8mL Plate (Bulk)	AB0859	Thermo Fisher
ZymoBIOMICS Lysis Solution	D4300-1-150	Zymo Research
Quick-DNA Magbead Plus Kit	D4082	Zymo Research

MATERIALS TEXT

RNase A solution in 10 mM Tris-HCl, pH 7.5, 15 mM NaCl at a concentration of 10 mg/mL:

250 mg RNase A
250 µL 1 M Tris-HCl pH 7.5
75 µL 5 M NaCl
Make up to 25 mL

Proteinase K solution in 50 mM Tris, pH 8, 3 mM CaCl₂, 50% Glycerol at a concentration of 20 mg/mL:

100 mg Proteinase K
250 µL 1 M Tris-HCl pH 8
6 µL 2.5 M CaCl₂
Make up to 5 mL

Prepare lysate for extraction

1 Prepare lysate for extraction

1.1 Prepare 100 µL lysis buffer per sample.

Lysis solution	99 µL
RNase solution (10 mg/mL)	1 µL
Total	100 µL

The Zymo lysis solution can be bought separately.

This protocol also works with the Zymo Solid Tissue Buffer II that is supplied with the Quick-DNA Magbead Plus kit. Solid Tissue Buffer II comes as a 2x concentrate and has to be diluted with nuclease-free water.

1.2 Dissect e.g. 1 *M. hyperodae* pupa for a single sample.

Place up to 88 individuals in a 1000 µL, round well deepwell plate

Add 2 3.2 mm stainless steel ball bearings and 100 µL of lysis buffer with RNase to each well.

1.3 Homogenise the tissue for **90 seconds at 1200 RPM** using a plate-compatible tissue homogenizer, e.g. SPEX SamplePrep 2010 Geno/Grinder.

🕒 **00:01:30**

1.4 Seal the plate and mix on the plate shaker. Make sure the paste is resuspended.

Incubate at 37°C for 30 minutes.

🕒 **00:30:00**

🌡 **37 °C**

1.5 Add 5 µL Proteinase K solution (20 mg/mL) and mix on the plate shaker.

Incubate at 55°C for 120 minutes, shaking the plate for one minute every 30 minutes.

🕒 **02:00:00**

🌡 **55 °C**

1.6 Spin the plate for 10 minutes at 1,000 RPM.

🌀 **1000 rpm, 15°C 00:10:00**

1.7 Transfer 100 µL of lysate to a 0.8 mL deepwell plate.

DNA extraction

2 Set up the reagents from the ZYMO *Quick*-DNA Magbead Plus Kit

2.1 Add the following reagents to the 7-position ReservoirRack

Reagent	Reservoir volume (mL)	8	16	24	32	40	48	56	64	72	80	88
1: 300 µL MBB : 5 µl beads	30	2645	5085	7525	9965	12405	14845	17285	19725	22165	24605	27045
2: DNA pre-wash buffer	100	3340	5340	7340	9340	11340	13340	15340	17340	19340	21340	23340
3: g-DNA wash buffer	100	5340	9340	13340	17340	21340	25340	29340	33340	37340	41340	45340
4: Tris-HCl	10	702	902	1102	1302	1502	1702	1902	2102	2302	2502	2702

3 Run the epMotion protocol 88x Quick-DNA Magbead Plus Low Volume 40_6.

3.1

Run the protocol with level sensing enabled.