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Gene knockout

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1 Works for me dx.doi.org/10.17504/protocols.io.bnygmftw

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ABSTRACT

We use this protocol to knock out *yhaK* in *E.coli* BW25113

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MATERIALS TEXT

E. coli BW25113
pCas9 plasmid
LB medium
Sterilized 10% glycerol
1 mM IPTG
Kana
Str

SAFETY WARNINGS

We use this protocol to knock out *yhaK* in *E.coli* BW25113.

ABSTRACT

We use this protocol to knock out *yhaK* in *E.coli* BW25113

BEFORE STARTING

Please read the protocol carefully before doing the experiment.

1 Transfer the pCas9 plasmid into the recipient *E. coli* BW25113 (Kana resistance, temperature sensitive).

2 Preparation of pCas9 competent cells:

2.1 Add 50 µL of bacterial solution to 5 mL of LB medium, incubate at 30°C, 200 rpm for about 12 h, Add 2% to 100 mL LB medium, add 0.5% of arabinose at a final concentration after 0.5 hours of culture.

2.2 Incubate at 30°C for about 2~2.5 h, until the OD600 is about 0.55~0.6, then take it out and let it stand on ice for 30 min.

2.3 Collect cells at 4200 rpm for 10 min in a 4°C refrigerated centrifuge.

2.4 Resuspend the cells gentlely with 10 ml of pre-cooled and sterilized 10% glycerol on ice.

2.5 Repeat steps 2.3 and 2.4 three times, using 10 mL of pre-cooled and sterilized 10% glycerol for the first two washes. The last time it was resuspended with 0.5 mL of glycerol.

2.6 Dispense in 100 µL/tube into a pre-cooled centrifuge tube, it can be used for electroporation or immediately placed in a -80°C refrigerator. The bacteria can be stored at -80°C for half a year.

3 Prepare target segment: Clone upstream and downstream sequences of *yhaK* by PCR to about 500 bp each, then connect upstream and downstream homology arms together by overlap.

4 pTarget-yhaK vector construction (Str resistance): Download the yhaK gene sequence, Find the appropriate gRNA affinity site and replace 1961 to 1980 bases of the pTarget plasmid by digestion and ligation or overlap.

5 Electroporation:

5.1 Add >200ng target fragment and >40ng pTarget-yhaK plasmid to 100µL competent cells on ice for 10~30 min, the plasmid and the ligation product should not exceed 5µL;

5.2 UV-sterilize the washed and dried 2 mm electroporation cuvette for 20 min and pre-cooled on ice, then quickly transfer the competent cells to the bottom of the electroporation cuvette;

5.3 Dry the outer wall of the cup. Immediately after the electric shock, add 900~1000µL of 37 °C preheated

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LB medium, mix gently, transfer to a 1.5 mL centrifuge tube, shake at 30 °C, 150 rpm, 45 ~ 60 min, then apply an appropriate amount of bacterial solution to the plate containing Kana and Str.

6 Knockout verification: Primers were designed at 700~1000 bp upstream and downstream of the *yhaK* in the genome to perform colony PCR validation on clones on the plates.

7 pTarget-yhaK elimination: Take the correct clone, add 1 mM IPTG at 30℃ for 12 h, streak to isolate the monoclone. The clones were tested for the presence of Str resistance, clones with pTarget eliminated is not resistant to str. The pCas9 plasmid can be eliminated by incubation at 37℃.

8 Strains that retain the pCas9 plasmid can be used to knock out other genes.

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