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Single-molecule Immunofluorescence Tissue Staining Protocol for Oligomer Imaging V.2

Rebecca Sonia
Andrews¹, Joanne Lachica², Steven F. Lee¹, Ghandi²

¹University of Cambridge; ²UCL

ASAP Collaborative Research Network



Rebecca Andrews
Univeristy of Cambridge

ABSTRACT

This protocol details background fluorescence quenching and immunofluorescence staining of human brain tissue for oligomer imaging.

ATTACHMENTS

kb3ib25np.pdf

GUIDELINES

- Use only clean bottles, flasks, magnetic stirrers, tweezers, weighing spatulas, measuring cylinders – everything should be cleaned, dried and covered if left on the side before next use.
- Everything should be handled with clean tweezers gloves should not touch the samples, solutions and ideally anything placed into the solutions where the slides are.

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Protocol status: Working We use this protocol and it's

working

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PROTOCOL integer ID:

75550

Keywords:

immunofluorescences, oligomer imaging

MATERIALS

Materials and Reagents

- Microtome
- Glass slides
- Xylene solution
- 100% alcohol
- Methanol
- Hydrogen peroxide (H₂0₂) solution
- Citrate buffer pH6
- Milli Q water
- Pressure cooker
- PBS
- Goat Serum 10%
- AlexaFluor antibody
- 0.1% Sudan black solution
- Vectashield
- Overslip

Immunofluorescences staining protocol for oligomer imaging 10m

- 1 Cut -1 + 8 µm tissue sections on a microtome and load onto glass slides.
- 2 Dry slides Overnight at 37 °C cover over the top.

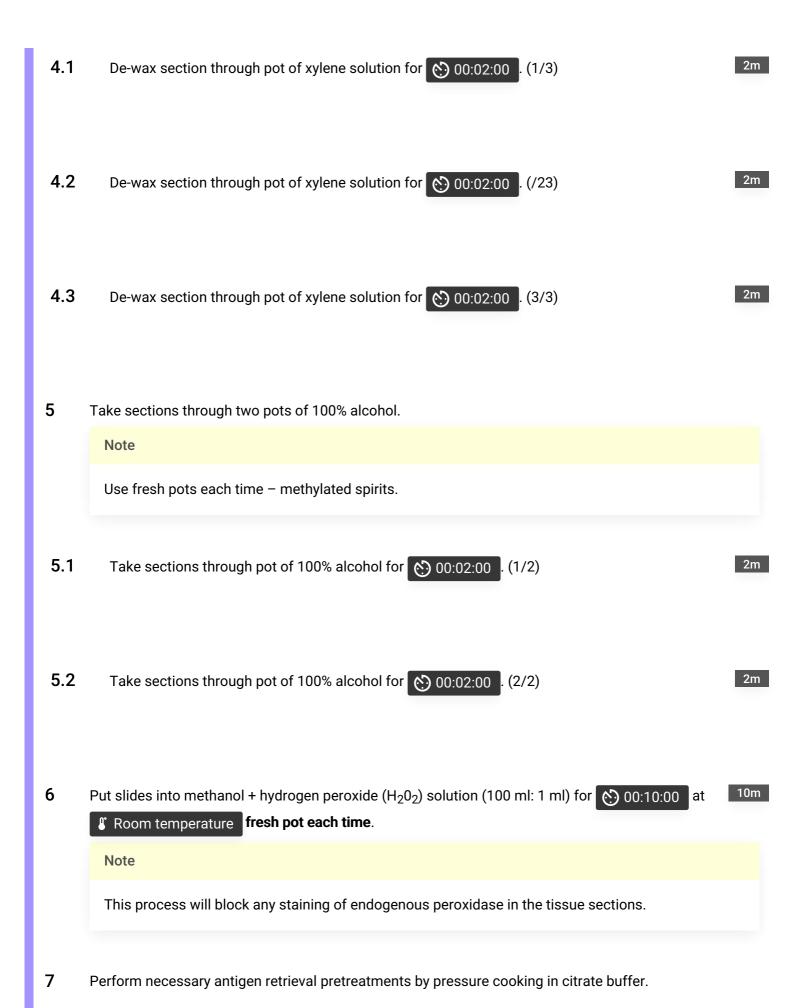
10m

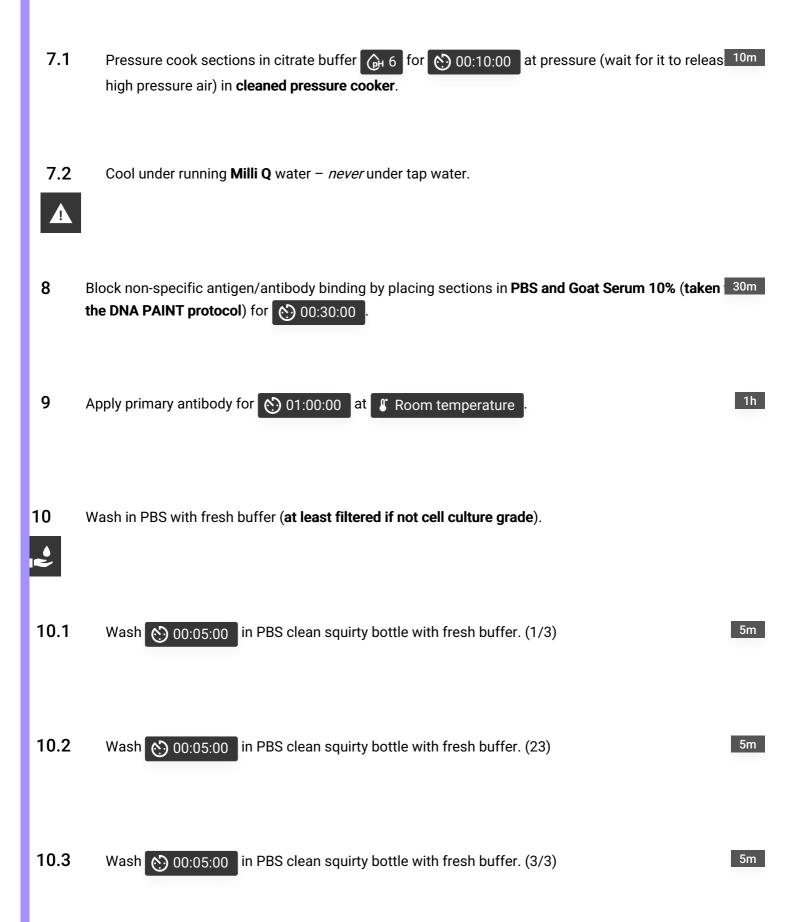
- 3
- Before staining commences keep slides for a few hours but ideally Overnight

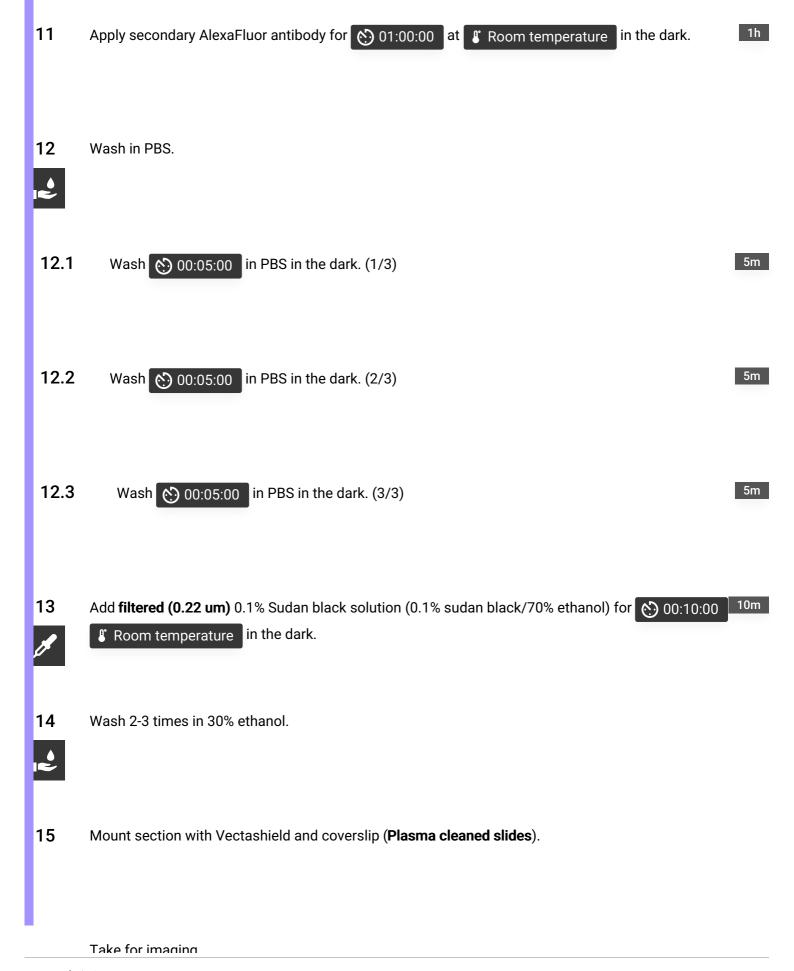




- 4
- De-wax sections through three pots of xylene solution. Use each fresh pots of xylene each time.







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