13-Apul-sRNAseq-ShortStack

Sam White

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Use ShortStack (Axtell 2013; Shahid and Axtell 2014; Johnson et al. 2016)to perform alignment of sRNAseq data and annotation of sRNA-producing genes.

The A.millepora genome will be used as the reference genome for A.pulchra, as A.pulchra does not currently have a sequenced genome and A.millepora had highest alignment rates for standard RNAseq data compared to other published genomes tested.

Inputs:

- Requires trimmed sRNAseq files generated by 08-Apul-sRNAseq-trimming.Rmd
 - Filenames formatted: *flexbar_trim.25bp*.gz
- A.millepora genome FastA. See 12-Apul-sRNAseq-MirMachine.Rmd for download info if needed.

Outputs:

• See ShortStack outputs documentation for full list and detailed descriptions.

Software requirements:

• Utilizes a ShortStack Conda/Mamba environment, per the installation instructions.

Replace with name of your ShortStack environment and the path to the corresponding conda installation (find this after you've activated the environment).

E.g.

```
# Activate environment
conda activate ShortStack4_env
# Find conda path
which conda
```

1 Set R variables

```
shortstack_conda_env_name <- c("ShortStack4_env")
shortstack_cond_path <- c("/home/sam/programs/mambaforge/condabin/conda")
```

2 Create a Bash variables file

This allows usage of Bash variables across R Markdown chunks.

```
{
echo "#### Assign Variables ###"
echo ""

echo "# Trimmed FastQ naming pattern"
echo "export trimmed_fastqs_pattern='*flexbar_trim.25bp*.fastq.gz'"

echo "# Data directories"
echo 'export deep_dive_dir=/home/shared/8TB_HDD_01/sam/gitrepos/deep-dive'
echo 'export deep_dive_data_dir="${deep_dive_dir}/data"'
echo 'export output_dir_top=${deep_dive_dir}/D-Apul/output/13-Apul-sRNAseq-ShortStack'
echo 'export trimmed_fastqs_dir="${deep_dive_dir}/D-Apul/output/08-Apul-sRNAseq-trimming/trimmed-reads"
echo "# Input/Output files"
echo 'export genome_fasta_dir=${deep_dive_dir}/D-Apul/data/Amil/ncbi_dataset/data/GCF_013753865.1'
echo 'export genome_fasta_name="GCF_013753865.1_Amil_v2.1_genomic.fa"'
echo 'export shortstack_genome_fasta_name="GCF_013753865.1_Amil_v2.1_genomic.fa"'
```

```
echo 'export mirbase_mature_fasta=mature.fa'
echo 'export mirbase_mature_fasta_version=mirbase-mature-v22.1.fa'
echo 'export genome_fasta="${genome_fasta_dir}/${shortstack_genome_fasta_name}"'
echo ""
echo "# External data URLs"
echo 'export mirbase_fasta_url="https://mirbase.org/download_version_files/22.1/"'
echo ""
echo "# Set number of CPUs to use"
echo 'export threads=46'
echo ""
echo "# Initialize arrays"
echo 'export trimmed_fastqs_array=()'
} > .bashvars
cat .bashvars
#### Assign Variables ####
# Trimmed FastQ naming pattern
export trimmed_fastqs_pattern='*flexbar_trim.25bp*.fastq.gz'
# Data directories
export deep_dive_dir=/home/shared/8TB_HDD_01/sam/gitrepos/deep-dive
export deep_dive_data_dir="${deep_dive_dir}/data"
export output_dir_top=${deep_dive_dir}/D-Apul/output/13-Apul-sRNAseq-ShortStack
export trimmed_fastqs_dir="${deep_dive_dir}/D-Apul/output/08-Apul-sRNAseq-trimming/trimmed-reads"
# Input/Output files
export genome_fasta_dir=${deep_dive_dir}/D-Apul/data/Amil/ncbi_dataset/data/GCF_013753865.1
export genome_fasta_name="GCF_013753865.1_Amil_v2.1_genomic.fna"
export shortstack_genome_fasta_name="GCF_013753865.1_Amil_v2.1_genomic.fa"
export mirbase_mature_fasta=mature.fa
export mirbase mature fasta version=mirbase-mature-v22.1.fa
export genome_fasta="${genome_fasta_dir}/${shortstack_genome_fasta_name}"
# External data URLs
export mirbase_fasta_url="https://mirbase.org/download_version_files/22.1/"
# Set number of CPUs to use
export threads=46
# Initialize arrays
export trimmed_fastqs_array=()
```

3 Load ShortStack conda environment

If this is successful, the first line of output should show that the Python being used is the one in your [ShortStack](https://github.com/MikeAxtell/ShortStack conda environment path.

```
E.g.
```

python: /home/sam/programs/mambaforge/envs/mirmachine_env/bin/python

```
use_condaenv(condaenv = shortstack_conda_env_name, conda = shortstack_cond_path)
# Check successful env loading
py_config()
```

python: /home/sam/programs/mambaforge/envs/ShortStack4_env/bin/python

libpython: /home/sam/programs/mambaforge/envs/ShortStack4_env/lib/libpython3.10.so

pythonhome: /home/sam/programs/mambaforge/envs/ShortStack4_env:/home/sam/programs/mambaforge/envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack4_envs/ShortStack

version: 3.10.13 | packaged by conda-forge | (main, Oct 26 2023, 18:07:37) [GCC 12.3.0]

numpy: /home/sam/programs/mambaforge/envs/ShortStack4_env/lib/python3.10/site-packages/numpy

numpy_version: 1.26.0

NOTE: Python version was forced by use_python() function

4 Download miRBase mature miRNA FastA

```
# Load bash variables into memory
source .bashvars
wget \
--directory-prefix ${deep_dive_data_dir} \
--recursive \
--no-check-certificate \
--continue \
--no-host-directories \
--no-directories \
--no-parent \
--quiet \
--execute robots=off \
${mirbase_fasta_url}/${mirbase_mature_fasta}
# Rename to indicate miRBase FastA version
mv ${deep_dive_data_dir}/${mirbase_mature_fasta} ${deep_dive_data_dir}/${mirbase_mature_fasta_version}
ls -lh "${deep_dive_data_dir}"
total 3.7M
```

-rw-r--r- 1 sam sam 3.7M Nov 6 12:40 mirbase-mature-v22.1.fa

5 Run ShortStack

5.1 Modify genome filename for ShortStack compatability

```
# Load bash variables into memory
source .bashvars

# Copy genome FastA to ShortStack-compatible filename (ending with .fa)
cp ${genome_fasta_dir}/${genome_fasta_name} ${genome_fasta_dir}/${shortstack_genome_fasta_name}

# Confirm
ls -lh ${genome_fasta_dir}/${shortstack_genome_fasta_name}
```

-rw-r--r- 1 sam sam 460M Nov 6 12:40 /home/shared/8TB_HDD_01/sam/gitrepos/deep-dive/D-Apul/data/Amil/

5.2 Excecute ShortStack command

Uses the --dn_mirna option to identify miRNAs in the genome, without relying on the --known_miRNAs. This part of the code redirects the output of time to the end of shortstack.log file.

• ; } \ 2>> \${output_dir_top}/shortstack.log

```
# Load bash variables into memory
source .bashvars
# Create array of trimmed FastQs
trimmed_fastqs_array=(${trimmed_fastqs_dir}/${trimmed_fastqs_pattern})
# Pass array contents to new variable as space-delimited list
trimmed_fastqs_list=$(echo "${trimmed_fastqs_array[*]}")
###### Run ShortStack ######
{ time \
ShortStack \
--genomefile "${genome_fasta}" \
--readfile ${trimmed_fastqs_list} \
--known_miRNAs ${deep_dive_data_dir}/${mirbase_mature_fasta_version} \
--dn_mirna \
--threads ${threads} \
--outdir ${output_dir_top}/ShortStack_out \
&> ${output_dir_top}/shortstack.log ; } \
2>> ${output_dir_top}/shortstack.log
```

5.3 Check runtime

```
# Load bash variables into memory
source .bashvars

tail -n 3 ${output_dir_top}/shortstack.log \
| grep "real" \
| awk '{print "ShortStack runtime:" "\t" $2}'
```

ShortStack runtime: 142m36.973s

6 Results

6.1 ShortStack synopsis

```
# Load bash variables into memory
source .bashvars
tail -n 20 ${output_dir_top}/shortstack.log
Screening of possible de novo microRNAs
No microRNA loci were found!
Writing final files
Non-MIRNA loci by DicerCall:
N 18676
22 45
23 36
21 10
24 5
Mon 06 Nov 2023 11:50:36 -0800 PST
Run Completed!
        142m36.973s
real
        2955m32.601s
sys 1100m59.754s
```

6.2 Inspect Results.txt

ShortStack didn't identify any miRNAs.

```
End Length Reads UniqueReads FracTop Strand MajorRNA
Locus
       Name
               Chrom
                      Start
                                                                                        MajorRNARea
                                      NC_058066.1 161118 161784 667 1363
NC_058066.1:161118-161784
                         Cluster_1
                                                                            392 0.6573734409391049
NC 058066.1:171557-171958
                          Cluster 2
                                      NC 058066.1 171557 171958 402 366 108 0.5683060109289617
NC_058066.1:204734-205143
                          Cluster_3
                                      NC_058066.1 204734 205143 410 525 180 0.6342857142857142
NC_058066.1:205754-206966
                          Cluster 4
                                      NC_058066.1 205754 206966 1213
                                                                        3040
                                                                                509 0.3769736842105
NC 058066.1:210858-211343
                         Cluster 5
                                      NC 058066.1 210858 211343
                                                                 486 1422
                                                                            317 0.2883263009845288
                          Cluster 6
                                      NC 058066.1 243461 243885
                                                                 425 446 46 0.8497757847533632 +
NC 058066.1:243461-243885
                          Cluster 7
NC_058066.1:349656-351296
                                      NC 058066.1 349656 351296 1641
                                                                         5821
                                                                                1435
                                                                                        0.515718948
NC 058066.1:351494-353435
                          Cluster 8
                                      NC_058066.1 351494 353435
                                                                 1942
                                                                        17924
                                                                                2140
                                                                                        0.571356839
NC_058066.1:776275-776775
                                      NC_058066.1 776275 776775 501 2260
                          Cluster_9
                                                                            216 0.8433628318584071
```

.....

Nummber of potential loci: 18772

Column 20 of the Results.txt file identifies if a cluster is a miRNA or not (Y or N).

```
# Load bash variables into memory
source .bashvars

echo "Number of loci characterized as miRNA:"
awk '$20=="Y" {print $0}' ${output_dir_top}/ShortStack_out/Results.txt \
| wc -1
echo ""
echo "------"

echo "Number of loci _not_ characterized as miRNA:"
awk '$20=="N" {print $0}' ${output_dir_top}/ShortStack_out/Results.txt \
| wc -1
```

Number of loci characterized as miRNA:

Number of loci _not_ characterized as miRNA: 18772

Column 21 of the Results.txt file identifies if a cluster aligned to a known miRNA (miRBase) or not (Y or NA).

Since there are no miRNAs, the following code will not print any output.

The echo command after the awk command is simply there to prove that the chunk executed.

```
# Load bash variables into memory
source .bashvars

echo "Number of loci matching miRBase miRNAs:"
awk '$21!="NA" {print $0}' ${output_dir_top}/ShortStack_out/Results.txt \
| wc -1
```

```
echo ""

echo ""

echo ""

echo "Number of loci _not_ matching miRBase miRNAs:"

awk '$21=="NA" {print $0}' ${output_dir_top}/ShortStack_out/Results.txt \
| wc -1

Number of loci matching miRBase miRNAs:

46

Number of loci _not_ matching miRBase miRNAs:

18727
```

Although there are loci with matches to miRBase miRNAs, ShortStack did *not* annotated these clusters as miRNAs likely because they do not *also* match secondary structure criteria.

6.2.1 Directory tree of all ShortStack outputs

Many of these are large (by GitHub standards) BAM files, so will not be added to the repo.

Additionally, it's unlikely we'll utilize most of the other files (bigwig) generated by ShortStack.

```
# Load bash variables into memory
source .bashvars

tree -h ${output_dir_top}/
```

```
/home/shared/8TB_HDD_01/sam/gitrepos/deep-dive/D-Apul/output/13-Apul-sRNAseq-ShortStack/
  [ 28K] shortstack.log
  [ 36K] ShortStack_out
      [ 47K]
              alignment_details.tsv
      [1.4M] Counts.txt
      [ 87K]
              known_miRNAs.gff3
      [1.8M]
              known_miRNAs_unaligned.fasta
      [9.8M]
              merged_alignments_21_m.bw
      [ 10M]
              merged_alignments_21_p.bw
      [9.5M]
              merged_alignments_22_m.bw
      [9.9M]
              merged_alignments_22_p.bw
      [ 19M]
              merged alignments 23-24 m.bw
      [ 20M]
              merged_alignments_23-24_p.bw
      [2.7G]
              merged_alignments.bam
      [227K]
              merged_alignments.bam.csi
      [123M]
              merged_alignments_other_m.bw
      [126M]
              merged alignments other p.bw
              merged_alignments_sRNA-ACR-140-S1-TP2.flexbar_trim.25bp_1.bw
      [48M]
      [48M]
              merged_alignments_sRNA-ACR-140-S1-TP2.flexbar_trim.25bp_2.bw
      [52M]
              merged_alignments_sRNA-ACR-145-S1-TP2.flexbar_trim.25bp_1.bw
      [ 52M]
              merged_alignments_sRNA-ACR-145-S1-TP2.flexbar_trim.25bp_2.bw
```

```
[ 50M]
       merged alignments sRNA-ACR-150-S1-TP2.flexbar trim.25bp 1.bw
[49M]
       merged_alignments_sRNA-ACR-150-S1-TP2.flexbar_trim.25bp_2.bw
[43M]
       merged alignments sRNA-ACR-173-S1-TP2.flexbar trim.25bp 1.bw
[43M]
       merged_alignments_sRNA-ACR-173-S1-TP2.flexbar_trim.25bp_2.bw
[44M]
       merged_alignments_sRNA-ACR-178-S1-TP2.flexbar_trim.25bp_1.bw
[43M]
       merged alignments sRNA-ACR-178-S1-TP2.flexbar trim.25bp 2.bw
[1.9M]
       Results.gff3
[2.8M]
       Results.txt
[246M]
       sRNA-ACR-140-S1-TP2.flexbar_trim.25bp_1.bam
[224K]
       sRNA-ACR-140-S1-TP2.flexbar_trim.25bp_1.bam.csi
[266M]
       sRNA-ACR-140-S1-TP2.flexbar_trim.25bp_2.bam
[229K]
       sRNA-ACR-140-S1-TP2.flexbar_trim.25bp_2.bam.csi
[279M]
       sRNA-ACR-145-S1-TP2.flexbar_trim.25bp_1.bam
[228K]
       sRNA-ACR-145-S1-TP2.flexbar_trim.25bp_1.bam.csi
[298M]
       sRNA-ACR-145-S1-TP2.flexbar_trim.25bp_2.bam
[230K]
       sRNA-ACR-145-S1-TP2.flexbar_trim.25bp_2.bam.csi
[297M]
       sRNA-ACR-150-S1-TP2.flexbar_trim.25bp_1.bam
[228K]
       sRNA-ACR-150-S1-TP2.flexbar trim.25bp 1.bam.csi
[316M]
       sRNA-ACR-150-S1-TP2.flexbar_trim.25bp_2.bam
[229K]
       sRNA-ACR-150-S1-TP2.flexbar trim.25bp 2.bam.csi
[255M]
       sRNA-ACR-173-S1-TP2.flexbar_trim.25bp_1.bam
[229K]
       sRNA-ACR-173-S1-TP2.flexbar trim.25bp 1.bam.csi
[275M]
       sRNA-ACR-173-S1-TP2.flexbar_trim.25bp_2.bam
[230K]
       sRNA-ACR-173-S1-TP2.flexbar trim.25bp 2.bam.csi
[234M]
       sRNA-ACR-178-S1-TP2.flexbar trim.25bp 1.bam
[229K]
       sRNA-ACR-178-S1-TP2.flexbar trim.25bp 1.bam.csi
[248M]
       sRNA-ACR-178-S1-TP2.flexbar_trim.25bp_2.bam
[230K]
       sRNA-ACR-178-S1-TP2.flexbar_trim.25bp_2.bam.csi
```

1 directory, 47 files

Citations

Axtell, Michael J. 2013. "ShortStack: Comprehensive Annotation and Quantification of Small RNA Genes." *RNA* 19 (6): 740–51. https://doi.org/10.1261/rna.035279.112.

Johnson, Nathan R, Jonathan M Yeoh, Ceyda Coruh, and Michael J Axtell. 2016. "Improved Placement of Multi-Mapping Small RNAs." *G3 Genes/Genomes/Genetics* 6 (7): 2103–11. https://doi.org/10.1534/g3. 116.030452.

Shahid, Saima, and Michael J. Axtell. 2014. "Identification and Annotation of Small RNA Genes Using ShortStack." Methods 67 (1): 20–27. https://doi.org/10.1016/j.ymeth.2013.10.004.