phyloflows: Practical example

Xiaoyue Xi and Oliver Ratmann

2021-06-30

This vignette describes the analysis on real-world HIV data from Uganda. The analysis was explained in Inferring the sources of HIV infection in Africa from deepsequence data with semi-parametric Bayesian Poisson flow models (link to appear).

Getting started

1. RCCS_metadata includes individual-level meta-data on sequenced individuals. This adds ages and locations to the data.

```
library(data.table)
df <- read.csv('~/phyloscanner/phyloflows/data/RCCS_metadata.csv')
df <- data.table(df)
df[,X:=NULL]
head(df)</pre>
```

- *ID*: individual IDs
- SEX: genders
- AGE_AT_MID: ages at the mid of study
- COMM_TYPE: locations (f fishing sites, i inland)
- $INMIG_2YRS_LOC$: in migration status in 2 years
- 2. RCCS_pairs include reconstructed transmission pairs from Phyloscanner.R.utilities.

```
rtr <- read.csv('~/phyloscanner/phyloflows/data/RCCS_pairs.csv')
rtr <- data.table(rtr)
rtr[,X:=NULL]
head(rtr)</pre>
```

- TR RID: source IDs
- REC_RID: recipient IDs

Pre-processing

Meta data and reconstructed pairs

We process the pair data and meta data, in order to groups individuals into sub-populations and count transmissions between sub-populations. In particular, the subpopulation is defined by one-year age bands, genders and locations. Also, we considered the locations of sources to be the origin of inmigrantion, if the reconstructed sources migrated within 2 years.

```
library(data.table)
setnames(df,colnames(df),paste0('TR_',colnames(df)))
rtr <- merge(rtr, df, by='TR_ID')
setnames(df,colnames(df),gsub('TR_','REC_',colnames(df)))
rtr <- merge(rtr, df, by='REC_ID')</pre>
```

```
# set unknown source location to fishing
rtr[is.na(TR_COMM_TYPE),TR_COMM_TYPE:='f']
# set unknown recipient location to inland
rtr[is.na(REC COMM TYPE),REC COMM TYPE:='i']
# impute age
tmp <- which(is.na(rtr$TR_AGE_AT_MID))</pre>
set(rtr, tmp, 'TR AGE AT MID', mean(rtr$TR AGE AT MID[which(!is.na(rtr$TR AGE AT MID))]) )
tmp <- which(is.na(rtr$REC_AGE_AT_MID))</pre>
set(rtr, tmp, 'REC_AGE_AT_MID', mean(rtr$REC_AGE_AT_MID[which(!is.na(rtr$REC_AGE_AT_MID))]) )
# set unknown migration to residents
rtr[is.na(TR_INMIG_2YRS_LOC), TR_INMIG_2YRS_LOC:='resident']
rtr[TR_INMIG_2YRS_LOC=='resident',TR_COMM_TYPE_MIG:=TR_COMM_TYPE]
tmp <- rtr[, which(TR_INMIG_2YRS_LOC=='unknown')]</pre>
# set unknown origin to fishing
set(rtr, tmp, 'TR_COMM_TYPE_MIG', 'f')
# set known migration
rtr[TR_INMIG_2YRS_LOC=='inland',TR_COMM_TYPE_MIG:='i']
rtr[TR_INMIG_2YRS_LOC=='fish',TR_COMM_TYPE_MIG:='f']
rtr[TR_INMIG_2YRS_LOC=='external',TR_COMM_TYPE_MIG:='e']
# stratify age
rtr[, TR_AGE_AT_MID_C:= as.character(cut(TR_AGE_AT_MID, breaks=c(15,16:49,50), labels=paste0(15:49,'-',
rtr[, REC_AGE_AT_MID_C:= as.character(cut(REC_AGE_AT_MID, breaks=c(15,16:49,50), labels=paste0(15:49,'-
rtr <- subset(rtr, !is.na(REC_AGE_AT_MID_C))</pre>
   build category to match with sampling data tables
rtr[, REC_SAMPLING_CATEGORY:= paste0(REC_COMM_TYPE, ':', REC_SEX, ':', REC_AGE_AT_MID_C)]
rtr[, TR_SAMPLING_CATEGORY:= paste0(TR_COMM_TYPE, ':', TR_SEX, ':', TR_AGE_AT_MID_C)]
# build transmission flow category
rtr[, REC_TRM_CATEGORY:= paste0(REC_COMM_TYPE,':',REC_SEX,':',REC_AGE_AT_MID_C)]
rtr[, TR_TRM_CATEGORY:= paste0(TR_COMM_TYPE_MIG,':',TR_SEX,':',TR_AGE_AT_MID_C)]
# make all combinations of variables
dac <- expand.grid(COMM_TYPE_F= c('i','f'), SEX= c('F','M'), AGE_AT_MID_C= paste0(15:49,'-',16:50))
dac <- as.data.table(dac)</pre>
dac[, CATEGORY:= paste0(COMM_TYPE_F, ':', SEX, ':', AGE_AT_MID_C)]
dac <- as.data.table(expand.grid(TR_SAMPLING_CATEGORY= dac$CATEGORY, REC_SAMPLING_CATEGORY= dac$CATEGOR
# ignore Male-Male and Female-Female combinations
dac <- subset(dac, !(grep1('F',TR_SAMPLING_CATEGORY)&grep1('F',REC_SAMPLING_CATEGORY)) & !(grep1('M',TR
# add transmission categories
dac[, REC_TRM_CATEGORY:= REC_SAMPLING_CATEGORY]
dac[, TR_TRM_CATEGORY:= TR_SAMPLING_CATEGORY]
# add inmigrants from external communities
tmp <- copy(dac)</pre>
```

```
set(tmp, NULL, 'TR_TRM_CATEGORY', tmp[,gsub('^[f|i]','e',TR_SAMPLING_CATEGORY)])
dac <- rbind(dac, tmp)</pre>
# add inmigrants sampled in inland and migrated from fishing
tmp <- dac[grepl('^i',TR_SAMPLING_CATEGORY)]</pre>
set(tmp, NULL, 'TR_TRM_CATEGORY', tmp[,gsub('^i','f',TR_SAMPLING_CATEGORY)])
dac <- rbind(dac, tmp)</pre>
# add inmigrants sampled in fishing and migrated from inland
tmp <- dac[grepl('^f',TR_SAMPLING_CATEGORY)]</pre>
set(tmp, NULL, 'TR_TRM_CATEGORY', tmp[,gsub('^f','i',TR_SAMPLING_CATEGORY)])
dac <- rbind(dac, tmp)</pre>
# remove duplicated rows
# TR_SAMPLING_CATEGORY f: F: 15-24: 1 and i: F: 15-24: 1 are all set to e: F: 15-24: 1
dac <- unique(dac)</pre>
    calculate observed number of transmissions
        <- rtr[, list( TRM_OBS=length(unique(TR_ID))), by=c('TR_TRM_CATEGORY','REC_TRM_CATEGORY','TR_SA</pre>
dac[, DUMMY:= 1]
dobs <- merge(dac, dobs, by=c('TR_TRM_CATEGORY', 'REC_TRM_CATEGORY', 'TR_SAMPLING_CATEGORY', 'REC_SAMPLI
stopifnot( dobs[, !any(is.na(DUMMY))] )
set(dobs, NULL, 'DUMMY', NULL)
set(dobs, dobs[, which(is.na(TRM_OBS))], 'TRM_OBS', OL)
    make SMOOTH CATEGORY
dobs[,TR_SMOOTH_CATEGORY:= as.numeric(substr(TR_TRM_CATEGORY,5,6))+0.5]
dobs[,REC_SMOOTH_CATEGORY:= as.numeric(substr(REC_TRM_CATEGORY,5,6))+0.5]
dobs[,OUTPUT:=pasteO(substr(TR_TRM_CATEGORY,1,1),':',
                      substr(TR_TRM_CATEGORY,3,3),':',
                      substr(TR_SAMPLING_CATEGORY,1,1),':',
                      substr(REC_TRM_CATEGORY,1,1),':',
                      substr(REC_TRM_CATEGORY,3,3))]
tmp <- subset(dobs,select = 'OUTPUT')</pre>
tmp <- unique(tmp)</pre>
setkey(tmp, OUTPUT)
tmp[, OUTPUT_ID:= seq_len(nrow(tmp))]
dobs <- merge(dobs, tmp, by='OUTPUT')</pre>
setkey(dobs,OUTPUT_ID,TR_SMOOTH_CATEGORY,REC_SMOOTH_CATEGORY)
# make PAIR_ID
dobs[, TRM_CAT_PAIR_ID:= seq_len(nrow(dobs))]
setkey(dobs, TRM_CAT_PAIR_ID)
```

Sampling data

We load samples drawn from the distribution of sampling fractions, as described in the basic example. Additional standard models to **phyloflows** were used to produce the samples, and details can be found in the paper. You may build other types of models for sampling.

```
dprior <- read.csv('~/phyloscanner/phyloflows/data/RCCS_sampling.csv')
dprior <- data.table(dprior)
dprior[,X:=NULL]
head(dprior)</pre>
```

Flow inference

Pre-processing gives the data needed for flow inference, including **dobs** and **dprior**. Now we use **phyloflows** to estimate transmission flows between subpopulations by MCMC.

Independent models

The first model considers flows to be independent between sub-populations, and estimate flows from the basic command in **phyloflow** through MCMC.

Correlated flows by ages

The second model treats flows correlated among the neighbouring age groups. We estimate flows by Stan through HMC. Note than Stan does not support samples drawn from distributions as inputs. Instead, we fit Beta distributions to samples in **dprior**. This model was facilated by reduced-rank Gaussian process approximation implemented in the file.

```
# fit beta to dprior
library(MASS)
dprior.fit <- dprior[,{tmp <- fitdistr(P, "beta", start=list(shape1=1,shape2=1/mean(P)),lower=c(0,0))</pre>
  list(SHAPE1=tmp$estimate[1],SHAPE2=tmp$estimate[2])
  },by=c('SAMPLING_CATEGORY','WHO')]
# index sampling groups
dprior.id <- subset(dprior.fit,select = c('SAMPLING_CATEGORY','WHO'))</pre>
setkey(dprior.id, SAMPLING_CATEGORY,WHO)
dprior.id[,ID:= seq_len(nrow(dprior.id))]
tmp <- subset(dobs, select = c('TR_SAMPLING_CATEGORY',</pre>
                                'REC_SAMPLING_CATEGORY',
                                'TRM_CAT_PAIR_ID'))
setnames(dprior.id,colnames(dprior.id),paste0('TR_',colnames(dprior.id)))
tmp <- merge(tmp,subset(dprior.id[TR_WHO=='TR_SAMPLING_CATEGORY',],select=c('TR_ID','TR_SAMPLING_CATEGORY')</pre>
setnames(dprior.id,colnames(dprior.id),gsub('TR_','REC_',colnames(dprior.id)))
tmp <- merge(tmp,subset(dprior.id[REC_WHO=='REC_SAMPLING_CATEGORY',],select=c('REC_ID','REC_SAMPLING_CATEGORY',]</pre>
setkey(tmp, TRM_CAT_PAIR_ID)
xi_id <- cbind(tmp$TR_ID, tmp$REC_ID)</pre>
# GP approximation
M < -30
D <- 2
indices <- matrix(NA, M^D, D)</pre>
mm=0:
for (m1 in 1:M){
 for (m2 in 1:M){
    mm = mm + 1
    indices[mm,] = c(m1, m2)
```

```
}
}
L \leftarrow matrix(rep(c(49.5, 49.5) * 5/4, each=nrow(indices)), nrow=nrow(indices))
sevalue <- pi * indices / (2 * L)
efunc <- do.call( rbind, lapply(1:1225, function(k){as.vector(apply(sqrt(1/L) * sin(sevalue *(matrix(r
# stan input
data.fit <- list(N=nrow(dobs),</pre>
                N_group = max(dobs$OUTPUT_ID),
                N_{per_group} = 1225,
                y=matrix(dobs$TRM_OBS,nrow=1225,ncol=max(dobs$OUTPUT_ID)),
                D=2
                x=cbind(dobs$TR_SMOOTH_CATEGORY[1:1225],dobs$REC_SMOOTH_CATEGORY[1:1225]),
                M_nD=900,
                Xgp=efunc,
                slambda=sevalue,
                N_xi = nrow(dprior.fit),
                shape = cbind(dprior.fit$SHAPE1,dprior.fit$SHAPE2),
                xi_id_src = matrix(xi_id[,1],nrow=1225,ncol=max(dobs$0UTPUT_ID)),
                xi_id_rec = matrix(xi_id[,2],nrow=1225,ncol=max(dobs$OUTPUT_ID)),
                id_mf = dobs[grep1(':M:',TR_TRM_CATEGORY),unique(OUTPUT_ID)],
                id_fm = dobs[grep1(':F:',TR_TRM_CATEGORY),unique(OUTPUT_ID)],
                id eh = dobs[grep1('e:',TR TRM CATEGORY) & grep1('f:',REC TRM CATEGORY),unique(OUTPUT I
                id_el = dobs[grepl('e:',TR_TRM_CATEGORY) & grepl('i:',REC_TRM_CATEGORY),unique(OUTPUT_I
                id hh = dobs[grep1('f:',TR TRM CATEGORY) & grep1('f:',REC TRM CATEGORY),unique(OUTPUT I
                id_hl = dobs[grep1('f:',TR_TRM_CATEGORY) & grep1('i:',REC_TRM_CATEGORY),unique(OUTPUT_I
                id_lh = dobs[grep1('i:',TR_TRM_CATEGORY) & grep1('f:',REC_TRM_CATEGORY),unique(OUTPUT_I
                id_mf_h = dobs[grep1(':M:',TR_TRM_CATEGORY) & grep1('f:',REC_TRM_CATEGORY),unique(OUTP')
                id_mf_l = dobs[grepl(':M:',TR_TRM_CATEGORY) & grepl('i:',REC_TRM_CATEGORY),unique(OUTP')
                id_fm_h = dobs[grep1(':F:',TR_TRM_CATEGORY) & grep1('f:',REC_TRM_CATEGORY),unique(OUTP'
                id_fm_l = dobs[grepl(':F:',TR_TRM_CATEGORY) & grepl('i:',REC_TRM_CATEGORY),unique(OUTP'
                id_sh = dobs[grep1('f:',TR_TRM_CATEGORY),unique(OUTPUT_ID)],
                id_sl = dobs[grepl('i:',TR_TRM_CATEGORY),unique(OUTPUT_ID)],
                id_se = dobs[grep1('e:',TR_TRM_CATEGORY),unique(OUTPUT_ID)])
# run stan
library(rstan)
fit <- stan(file = file.path('gpa4.stan'),</pre>
            data = data.fit,
            iter = 3000, warmup = 500, chains=1, thin=1, seed = 42,
            algorithm = "NUTS", verbose = FALSE,
            control = list(adapt_delta = 0.999,max_treedepth=15))
```

Going forward

So far, we have covered how to reproduced the main analysis in the paper. Next, please use your R wizadry or other functions in **phyloflows** to process the output further and answer practical questions in disease surveillance.