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Chapter 1

Installation

ASTEC (acronym of "adaptive segmentation and tracking of embryonic cells" [Gui15]) has been designed for unix-like systems (e.g. Linux, or MacOS). It has been developed with python2.7 and was not tested for python3.0. It is a set of python scripts, built over a set of C commands.

There are two distributions. The first one can be retrieved from github.com/astec-segmentation/astec-2019-published and includes both the python and the C codes. The installation procedure is dedicated to this distribution.

The second one is devoluted to more advanced users that may want to benefit from future developements of the ASTEC distribution:

- python scripts can be retrieved from github.com/astec-segmentation/astec
- C code can be retrieved from gitlab.inria.fr/morpheme/vt
- optional third-party librairies can be retrieved from gitlab.inria.fr/morpheme/vt-third-party

Both github.com/astec-segmentation/astec-2019-published and github.com/astec-segmentation/astec contains the following 4 sub-directories

| 2 | stec[-2019-published/] |
|---|--------------------------|
| | documentation/ |
| | parameter-file-examples, |
| | src/ |
| | tutorial/ |

- documentation/ contains this documentation.
- parameter-file-examples/ contains templates of parameter files for the python scripts. See chapter 3 for further details.
- src/ contains the python scripts and files (as well as the C codes for the astec-2019-published distribution).
- tutorial/ contains a toy data set and the associated parameter files. See chapter 2.

1.1 github.com/astec-segmentation/astec

1.1.1 Linux system

This section describes the required command to install the ASTEC distribution on a Linux system (was tested on a Ubuntu system (18.04.2, 64 bits) installed on a virtual machine¹) so the tutorial (chapter 2) can be run.

¹ virtualbox.org

1. Get the distribution. It is recommended (but not necessary) to use git, so keeping up to date with the distribution will be easier. git can be installed with

```
$ sudo apt install git
```

Then, choose the directory where to install the ASTEC distribution, and download it

- \$ cd /wherever/one/wants/
- \$ git clone https://github.com/astec-segmentation/astec-2019-published.git

It creates the directory /wherever/one/wants/astec-2019-published/ that will be denoted /path/to/astec/ from now on.

2. Prepare the compilation of the C code. Compilation is done within the cmake² framework. The standard Ubuntu distribution comes with a C compiler but not with a C++ one. Last a development version of the zlib is required. The next few lines allow to install the required components.

```
$ sudo apt install cmake
$ sudo apt install cmake-curses-gui
$ sudo apt install g++
$ sudo apt install zlib1g-dev
```

- 3. Compile the C code.
 - \$ cd /path/to/astec/
 - \$ cd src/ASTEC/CommunFunctions/cpp/vt/
 - \$ mkdir build
 - \$ cd build
 - \$ cmake ../
 - \$ make

4. Install the required python libraries. As mentioned, ASTEC has been developed with python2.7. pip is here used for the installation of the python libraries. Required libraries are numpy, scipy, libtiff, and h5py.

```
$ sudo apt install python2.7
$ sudo apt install python-pip
$ sudo pip install numpy
$ sudo pip install scipy
$ sudo pip install libtiff
$ sudo pip install h5py
```

5. Make the ASTEC scripts/commands available as on-line commands. It can be done in a terminal (but will be valid only for this terminal)

```
$ export PATH=$PATH:/path/to/astec/src
```

or by adding the above line in the right setup file (e.g. .bashrc, .profile, \dots).

 $^{^2 {\}tt cmake.org}$

Chapter 2

Tutorial

Before starting

It is advised to add to your PATH environment variable the paths to both the python and the C executable commands (the latter is important in case of non-standard installation). So, Astec commands can be launched without specifying the complete path to the command.

It can be done in a terminal (and will be valid only for this terminal)

```
$ export PATH=$PATH:/path/to/astec/src
```

\$ export PATH=\$PATH:/path/to/astec/src/ASTEC/CommunFunctions/cpp/vt/build/bin

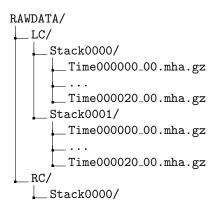
or by modifying a setup file (e.g. bashrc, .profile, ...).

2.1 Tutorial data

The directory /path/to/astec/tutorial/tuto-astec1/, also denoted by path/to/experiment/ or <EXPERIMENT>, contains the RAWDATA/ and parameters/ sub-directories and a README file

```
path/to/tuto-astec1/
RAWDATA/
README
parameters/
```

The RAWDATA/ contains 21 time points (indexed from 0 to 20) of subsampled (for file size consideration) raw data from a 3D+t movie acquired by a MuViSPIM microscope.



```
1 PATH_EMBRYO = '.'
2
3 EN = '2019-Tutorial100'
4
5 begin = 0
6 end = 20
7
8 acquisition_orientation = 'right'
9 acquisition_mirrors = False
10 acquisition_resolution = (1., 1., 1.)
11
12 target_resolution = 1.0
```

Figure 2.1: Tutorial parameter file for the fusion step (lines are numbered).

```
Time000000_00.mha.gz
...
Time0000020_00.mha.gz
Stack0001/
Time0000000_00.mha.gz
...
Time0000020_00.mha.gz
```

where LC/ and LC/ stand respectively for the left and the right cameras.

2.2 Fusion

We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Running the fusion is done with

```
$ 1-fuse.py -p parameters/1-fuse-tutorial-parameters.py
```

1-fuse-tutorial-parameters.py being the dedicated parameter file (figure 2.1).

- The variable PATH_EMBRYO is the path to the directory where the directory RAWDATA/ is located. It can be either relative (as in the above example) or global (it could have been /path/to/astec/tutorial/tuto-astec1/).
- The variable EN is the prefix after which the result fusion images will be named.
- The variables begin and end set respectively the first and the last index of the input time points to be processed.
- The variables acquisition_orientation and acquisition_mirrors are parameters describing the acquisition geometry.
- The variable acquisition_resolution is the voxel size (along the 3 dimensions X, Y and Z).
- The variable target_resolution is the desired isotropic (the same along the 3 dimensions) voxel size for the result fusion images.

After processing, a FUSE/ directory has been created

```
path/to/tuto-astec1/
__FUSE/
```

```
1 PATH_EMBRYO = '.'
2
3 EN = '2019-Tutorial100'
4
5 begin = 0
6 end = 20
```

Figure 2.2: Tutorial parameter file for the sequence intra-registration step.

```
RAWDATA/
README
parameters/

The FUSE/ directory contains
FUSE/
FUSE_RELEASE/
2019-Tutorial100_fuse_t000.inr
2019-Tutorial100_fuse_t020.inr
LOGS/
```

The fused images are named after <EN>_fuse<XXX>.inr (where <XXX> denotes the value of the variable XXX) and indexed from <begin> to <end> (as the input data).

The directory LOGS/ contains a copy of the parameter file (stamped with date and hour) as well as a log file (also stamped with date and hour) reporting information about the processing.

2.3 Sequence intra-registration (or drift compensation) [1]

We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Running the sequence intra-registration is done with

- \$ 1.5-intraregistration.py -p parameters/1.5-intraregistration-tutorial-parameters-fuse.py
- 1.5-intraregistration-tutorial-parameters-fuse.py being the dedicated parameter file (figure 2.2).
 - The variable PATH_EMBRYO is the path to the directory where the directory FUSE/ is located. It can be either relative (as in the above example) or global (it could have been /path/to/astec/tutorial/tuto-astec1/).
 - The variable EN is the prefix after which the images are named.
 - The variables begin and end set respectively the first and the last index of the input time points to be processed.

After processing, a INTRAREG/ directory has been created

```
path/to/tuto-astec1/
FUSE/
INTRAREG/
RAWDATA/
README
parameters/
```

INTRAREG/ __ INTRAREG_RELEASE/ _CO-TRSFS/ 2019-Tutorial100_intrareg_flo000_ref001.trsf _2019-Tutorial100_intrareg_flo019_ref020.trsf FUSE/ __ FUSE_RELEASE/ __2019-Tutorial100_intrareg_fuse_t000.inr __2019-Tutorial100_intrareg_fuse_t020.inr LOGS/ _MOVIES/ __FUSE/ __ FUSE_RELEASE/ ___2019-Tutorial100_intrareg_fuse_t000-020_xy0205.inr TRSFS_t0-20/ 2019-Tutorial100_intrareg_t000.trsf 2019-Tutorial100_intrareg_t020.trsf $_{ t template_t0-20.inr}$

- The directory CO-TRSF/ contains the co-registration transformations.
- The directory FUSE/FUSE_RELEASE/ contains the resampled fused images in the same geometry (images have the same dimensions along X, Y and Z), with drift compensation (the eventual motion of the sample under the microscope has been compensated).
- The directory MOVIES/FUSE/FUSE_RELEASE/ contains a 3D (which is a 2D+t) image, here 2019-Tutorial100_intrar which the #205 XY-section of the resampled fused images for all the time points.
- The directory TRSFS/ contains the transformation of every fused image towards the reference one as well as the template image (an image large enough to including each fused images after resampling).

The template image template_t0-20.inr is of size $422 \times 365 \times 410$ with a voxel size of 0.6 (the voxel size can be set by the variable intra_registration_resolution)

2.4 Segmentation of the first time point

We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Segmenting the first time point is done with

\$ 2-mars.py -p parameters/2-mars-tutorial-parameters.py

2-mars-tutorial-parameters.py being the dedicated parameter file (figure 2.3).

- The variable PATH_EMBRYO is the path to the directory where the directory FUSE/ is located. It can be either relative (as in the above example) or global (it could have been /path/to/astec/tutorial/tuto-astec1/).
- The variable EN is the prefix after which the images are named.
- The variable begin sets the index of the first input time point (to be processed).

After processing, a SEG/ directory has been created

path/to/tuto-astec1/

```
1 PATH_EMBRYO = '.'
2
3 EN = '2019-Tutorial100'
4
5 begin = 0
```

Figure 2.3: Tutorial parameter file for the segmentation of the first time point.

```
1 PATH_EMBRYO = '.'
2
3 EN = '2019-Tutorial100'
4
5 begin = 0
6
7 mancor_mapping_file='parameters/3-manualcorrection-tutorial.txt'
```

Figure 2.4: Tutorial parameter file for the segmentation correction of the first time point. See figure 2.5 for the <mancor_mapping_file> file.

```
FUSE/
INTRAREG/
RAWDATA/
README
SEG/
parameters/

The SEG/ directory contains

SEG/
SEG_RELEASE/
LOGS/
RECONSTRUCTION/
```

2019-Tutorial100_mars_t000.inr is the segmented first time point of the sequence.

2.5 Correction of the first time point segmentation

We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Correcting the first time point segmentation is done with

```
$ 3-manualcorrection.py -p parameters/3-manualcorrection-tutorial-parameters.py
```

3-manualcorrection-tutorial-parameters.py being the dedicated parameter file (figure 2.4).

• The variable PATH_EMBRYO is the path to the directory where the directory SEG/ is located. It can be either relative (as in the above example) or global (it could have been /path/to/astec/tutorial/tuto-astec1/).

```
10 6
20 13
9 4
26 11
21 11
27 15
32 23
39 31
35 20
38 43
46 45
52 42
58 62
63 60
78 67
74 66
68 66
83 75
82 77
```

Figure 2.5: The segmentation correction file 3-manualcorrection-tutorial.txt for the first time point. The first number is the line index (lines are numbered).

- The variable EN is the prefix after which the images are named.
- The variable begin set the index of the first input time point (to be processed).
- The variable mancor_mapping_file gives the file name containing the correction to be applied.

After processing, the SEG/ directory contains

```
SEG/
SEG_RELEASE/
2019-Tutorial100_mars_t000.inr
2019-Tutorial100_seg_t000.inr
LOGS/
RECONSTRUCTION/
```

2019-Tutorial100_seg_t000.inr is the corrected version of the segmentation obtained at the previous step.

2.6 Segmentation propagation

We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Segmenting the first time point is done with

```
$ 4-astec.py -p parameters/4-astec-tutorial-parameters.py
```

4-astec-tutorial-parameters.py being the dedicated parameter file (figure 2.6).

After processing, the SEG/ directory contains

```
1 PATH_EMBRYO = '.'
 3 EN = '2019-Tutorial100'
 5 \text{ begin} = 0
 6 \text{ end} = 20
 7 \text{ delta} = 1
 8 \text{ raw\_delay} = 0
10 ## General parameters for segmentation propagation
12 astec_sigma1 = 0.6
13 \operatorname{astec\_sigma2} = 0.15
14 \operatorname{astec\_h\_min\_min} = 4
15 \text{ astec\_h\_min\_max} = 18
16
17 ## Glace Parameters (if astec_membrane_reconstruction_method is set to 1 or 2):
18 ## membrane_renforcement
19
20 astec_sigma_membrane = 0.9
21 astec_sensitivity = 0.99
22 astec_manual = False
23 astec_manual_sigma = 15
24 astec_hard_thresholding = False
25 astec_hard_threshold = 1.0
27 ## Tensor voting framework
28
29 astec_sigma_TV = 3.6
30 astec_sigma_LF = 0.9
31 astec_sample = 0.2
32
33 ## Default parameters (for classical use, default values should not be changed)
35 astec_RadiusOpening = 20
36 \text{ astec\_Thau} = 25
37 astec_MinVolume = 1000
38 astec_VolumeRatioBigger = 0.5
39 astec_VolumeRatioSmaller = 0.1
40 astec_MorphosnakeIterations = 10
41 astec_NIterations = 200
42 astec_DeltaVoxels = 10**3
43 \text{ astec\_nb\_proc} = 10
```

Figure 2.6: Tutorial parameter file for the segmentation propagation.

```
SEG/

SEG_RELEASE/

2019-Tutorial100_mars_t000.inr

2019-Tutorial100_seg_lineage.test

2019-Tutorial100_seg_t000.inr

2019-Tutorial100_seg_t001.inr

2019-Tutorial100_seg_t020.inr

4-astec-tutorial-parameters.py

LOGS/

RECONSTRUCTION/
```

while a 2019-Tutorial100-seg_lineage.pkl file has been created in the path/to/tuto-astec1/ directory.

```
path/to/tuto-astec1/

2019-Tutorial100_seg_lineage.pkl

FUSE/
INTRAREG/
RAWDATA/
README
SEG/
parameters/
```

2019-Tutorial100-seg_lineage.pkl is a pickle python file containing a dictionary (in the python sense). It can be read by

```
$ python
...
>>> import cPickle as pkl
>>> f=open('2019-Tutorial100_seg_lineage.pkl')
>>> d=pkl.load(f)
>>> f.close()
>>> d.keys()
['h_mins_information', 'lin_tree', 'volumes_information',
'sigmas_information']
```

In this pickle file, cells have an unique identifier i * 1000 + c, which is made of both the image index i and the cell identifier c within a segmentation image (recall that, within an image, cells are numbered from 2, 1 being the background label).

2.7 Sequence intra-registration (or drift compensation) [2]

We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Running the sequence intra-registration is done with

- \$ 1.5-intraregistration.py -p parameters/1.5-intraregistration-tutorial-parameters-seg.py
- 1.5-intraregistration-tutorial-parameters-seg.py being the dedicated parameter file (figure 2.7).
 - The variable PATH_EMBRYO is the path to the directory where the directory FUSE/ is located. It can be either relative (as in the above example) or global (it could have been /path/to/astec/tutorial/tuto-astec1/).
 - The variable EN is the prefix after which the images are named.
 - The variables begin and end set respectively the first and the last index of the input time points to be processed.

```
1 PATH_EMBRYO = '.'
2
3 EN = '2019-Tutorial100'
4
5 begin = 0
6 end = 20
7
8 EXP_INTRAREG = 'SEG'
9
10 intra_registration_template_type = "SEGMENTATION"
11 intra_registration_template_threshold = 2
12 intra_registration_margin = 20
13
14 intra_registration_resample_segmentation_images = True
15 intra_registration_movie_segmentation_images = True
```

Figure 2.7: Tutorial parameter file for the sequence intra-registration step, segmentation images being used to build the template.

- the variable EXP_INTRAREG set the suffix of the sub-directory of the INTRAREG/ directory to be created.
- the variable intra_registration_template_type set the images to be used to build the template. Here, since it is equal to "SEGMENTATION'', they are the segmentation images obtained at the previous step.

The variable intra_registration_template_threshold set a threshold to be applied to the template images to define the information to be kept: we want all the points with a value equal or greater than 2 to be contained in the template after resampling. Since cells are labeled from 2 and above, the template is designed to contain all labeled cells after resampling, so it is built as small as possible.

The variable $intra_registration_margin$ allows to add margins (in the 3 dimensions) to the built template.

- The variable intra_registration_resample_segmentation_images indicates whether the segmentation images are to be resampled in the template geometry.
- The variable intra_registration_movie_segmentation_images indicates whether 2D+t movies have to be built from the resampled segmentation images.

After processing, a ${\tt INTRAREG_SEG/directory}$ has been created and the ${\tt INTRAREG/directory}$ now contains

```
2019-Tutorial100_intrareg_fuse_t020.inr
LOGS/
_MOVIES/
   FUSE/
     _FUSE_RELEASE/
      ___2019-Tutorial100_intrareg_fuse_t0-20_xy174.inr
   SEG/
   ___SEG_RELEASE/
      ___2019-Tutorial100_intrareg_seg_t0-20_xy174.inr
SEG/
___SEG_RELEASE/
      2019-Tutorial100_intrareg_seg_t000.inr
     _2019-Tutorial100_intrareg_seg_t020.inr
TRSFS_t0-20/
   2019-Tutorial100_intrareg_t000.trsf
  _2019-Tutorial100_intrareg_t020.trsf
  _template_t0-20.inr
```

In addition to directories already described in section 2.3, the INTRAREG_SEG/ directory contains

- The directory SEG/SEG_RELEASE contains the resampled segmentation images in the same geometry (images have the same dimensions along X, Y and Z), with drift compensation (the eventual motion of the sample under the microscope has been compensated).
- In addition to a 2D+t movie made from the resampled fusion images, the directory MOVIES/ contains a 2D+t movie made from the resampled segmentation images in the sub-directory SEG/SEG_RELEASE.
- The template image template_t0-20.inr in the directory TRSFS/ is now of size $323 \times 265 \times 348$ with a voxel size of 0.6, which is smaller than the one computed in section 2.3, even with the added margins. Note that all resampled images (in both the FUSE/FUSE_RELEASE and the SEG/SEG_RELEASE directories have the same geometry than the template image.

2.8 Sequence properties computation [1]

We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Computing cell properties as well as lineage assumes that segmentation or post-corrected segmentation (see section 2.11) images have been co-registered (see sections 2.7 and 2.10). Extracting the sequence properties from the co-registered segmentation images is done with

\$ X-embryoproperties.py -p parameters/X-embryoproperties-tutorial-parameters-seg.py X-embryoproperties-tutorial-parameters-seg.py being the dedicated parameter file (figure 2.8).

- The variable PATH_EMBRYO is the path to the directory where the directory FUSE/ is located. It can be either relative (as in the above example) or global (it could have been /path/to/astec/tutorial/tuto-astec1/).
- $\bullet\,$ The variable EN is the prefix after which the images are named.
- The variables begin and end set respectively the first and the last index of the input time points to be processed.
- the variable EXP_INTRAREG set the suffix of the sub-directory of the INTRAREG/ directory where to search post-corrected segmentation or segmentation images.
 - Since the directory INTRAREG/INTRAREG_SEG/ only contains the co-registered segmentation images (in the SEG/SEG_RELEASE/ sub-directory), properties will be computed from these images.

```
1 PATH_EMBRYO = '.'
2
3 EN = '2019-Tutorial100'
4
5 begin = 0
6 end = 20
7
8 EXP_INTRAREG = 'SEG'
```

Figure 2.8: Tutorial parameter file for the sequence properties from the co-registered segmentation images.

After processing, some files appears in the INTRAREG/INTRAREG_SEG/SEG_RELEASE/ sub-directory

```
INTRAREG/
  _INTRAREG_RELEASE/
   __ . . .
   INTRAREG_SEG/
    _ CO-TRSFS/
      __...
     _FUSE/
      _ ...
     LOGS/
     MOVIES/
     ∟...
     SEG/
      __ SEG_RELEASE/
         __2019-Tutorial100_intrareg_seg_lineage.pkl
          __2019-Tutorial100_intrareg_seg_lineage.txt
           2019-Tutorial100_intrareg_seg_lineage.xml
          __2019-Tutorial100_intrareg_seg_t000.inr
          _ . . .
          _2019-Tutorial100_intrareg_seg_t020.inr
     TRSFS_t0-20/
```

2019-Tutorial100_intrareg_seg_lineage.pkl is a pickle python file containing a dictionary (in the python sense). It can be read by

```
$ python
...
>>> import cPickle as pkl
>>> f=open('2019-Tutorial100_intrareg_seg_lineage.pkl')
>>> d=pkl.load(f)
>>> f.close()
>>> d.keys()
['all_cells', 'cell_barycenter', 'cell_contact_surface',
'cell_principal_vectors', 'cell_principal_values', 'cell_volume',
'cell_compactness', 'cell_surface', 'cell_lineage']
```

In this pickle file (as in the one computed at section 2.6), cells have an unique identifier i * 1000 + c, which

```
<data>
  <cell_volume>
  </cell_volume>
  <cell_surface>
  </cell_surface>
  <cell_compactness>
  </cell_compactness>
  <cell_barycenter>
  </cell_barycenter>
  <cell_principal_values>
  </cell_principal_values>
  <cell_principal_vectors>
  </cell_principal_vectors>
  <cell_contact_surface>
  </cell_contact_surface>
  <all_cells>[2, 3, 4, 5, 6, 7, 8, 11, 12, 13,
     200097, 200099, 200100, 200101, 200102]</all_cells>
  <cell_lineage>
  </cell_lineage>
</data>
```

Figure 2.9: XML output properties file from the co-registered segmentation image.

is made of both the image index i and the cell identifier c within a segmentation image (recall that cells are numbered from 2, 1 being the background label).

2019-Tutorial100_intrareg_seg_lineage.xml contains the same information than the pickle file, but in xml format (see figure 2.9).

2019-Tutorial100_intrareg_seg_lineage.tst contains some diagnosis information (smallest and largest cells, weird lineages, etc.).

2.9 Segmentation post-correction

We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Segmentation post-correction is done with

```
$ 5-postcorrection.py -p parameters/5-postcorrection-tutorial-parameters.py
```

5-postcorrection-tutorial-parameters.py being the dedicated parameter file (figure 2.10). After processing, a POST/ directory has been created

```
1 PATH_EMBRYO = '.'
2
3 EN = '2019-Tutorial100'
4
5 begin = 0
6 end = 20
7 delta = 1
8 raw_delay = 0
9
10 postcor_Volume_Threshold=10000
11 postcor_Soon=True
```

Figure 2.10: Tutorial parameter file for the segmentation post-correction.

```
path/to/tuto-astec1/
   2019-Tutorial100_seg_lineage.pkl
  FUSE/
  INTRAREG/
  POST/
  RAWDATA/
  README
   SEG/
  _{	extstyle }parameters/
The POST/ directory contains
POST/
__ POST_RELEASE/
    _2019-Tutorial100_post_lineage.pkl
     _2019-Tutorial100_post_lineage.test
    _2019-Tutorial100_post_t000.inr
     _2019-Tutorial100_post_t020.inr
     _5-postcorrection-tutorial-parameters.py
     _5-postcorrection.log
```

2.10 Sequence intra-registration (or drift compensation) [3]

We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Running the sequence intra-registration is done with

- \$ 1.5-intraregistration.py -p parameters/1.5-intraregistration-tutorial-parameters-post.py
- 1.5-intraregistration-tutorial-parameters-post.py being the dedicated parameter file (figure 2.11).
 - The variable PATH_EMBRYO is the path to the directory where the directory FUSE/ is located. It can be either relative (as in the above example) or global (it could have been /path/to/astec/tutorial/tuto-astec1/).
 - The variable EN is the prefix after which the images are named.

```
1 PATH_EMBRYO = '.'
2
3 EN = '2019-Tutorial100'
4
5 begin = 0
6 end = 20
7
8 EXP_INTRAREG = 'POST'
9
10 intra_registration_template_type = "POST-SEGMENTATION"
11 intra_registration_template_threshold = 2
12 intra_registration_margin = 20
13
14 intra_registration_resample_post_segmentation_images = True
15 intra_registration_resample_segmentation_images = True
16 intra_registration_movie_post_segmentation_images = True
17 intra_registration_movie_segmentation_images = True
```

Figure 2.11: Tutorial parameter file for the sequence intra-registration step, post-segmentation images being used to build the template.

- The variables begin and end set respectively the first and the last index of the input time points to be processed.
- the variable EXP_INTRAREG set the suffix of the sub-directory of the INTRAREG/ directory to be created.
- the variable intra_registration_template_type set the images to be used to build the template. Here, since it is equal to "POST-SEGMENTATION'', they are the post-corrected segmentation images obtained at the previous step.

The variable intra_registration_template_threshold set a threshold to be applied to the template images to define the information to be kept: we want all the points with a value equal or greater than 2 to be contained in the template after resampling. Since cells are labeled from 2 and above, the template is designed to contain all labeled cells after resampling, so it is built as small as possible.

The variable intra_registration_margin allows to add margins (in the 3 dimensions) to the built template.

- The variable intra_registration_resample_post_segmentation_images indicates whether the post-corrected segmentation images are to be resampled in the template geometry.
- The variable intra_registration_resample_segmentation_images indicates whether the segmentation images are to be resampled in the template geometry.
- The variable intra_registration_movie_post_segmentation_images indicates whether 2D+t movies have to be built from the resampled post-corrected segmentation images.
- The variable intra_registration_movie_segmentation_images indicates whether 2D+t movies have to be built from the resampled segmentation images.

After processing, a $INTRAREG/INTRAREG_POST/$ directory has been created and the INTRAREG/ directory now contains

```
INTRAREG_POST/
L_CO-TRSFS/
```

```
2019-Tutorial100_intrareg_flo000_ref001.trsf
     _2019-Tutorial100_intrareg_flo019_ref020.trsf
  FUSE/
    __ FUSE_RELEASE/
        2019-Tutorial100_intrareg_fuse_t000.inr
       _2019-Tutorial100_intrareg_fuse_t020.inr
  LOGS/
  MOVIES/
    _{-} FUSE/
      ___ FUSE_RELEASE/
        __2019-Tutorial100_intrareg_fuse_t0-20_xy174.inr
     POST/
       _ POST_RELEASE/
        ___2019-Tutorial100_intrareg_post_t0-20_xy174.inr
     SEG/
     ___ SEG_RELEASE/
        ___2019-Tutorial100_intrareg_seg_t0-20_xy174.inr
  POST/
    __POST_RELEASE/
        2019-Tutorial100_intrareg_post_t000.inr
       _2019-Tutorial100_intrareg_post_t020.inr
  SEG/
  __ SEG_RELEASE/
      __2019-Tutorial100_intrareg_seg_t000.inr
       _2019-Tutorial100_intrareg_seg_t020.inr
  TRSFS_t0-20/
    _2019-Tutorial100_intrareg_t000.trsf
     2019-Tutorial100_intrareg_t020.trsf
    _template_t0-20.inr
INTRAREG_RELEASE/
 . . .
INTRAREG_SEG/
```

In addition to directories already described in section 2.3, the INTRAREG_POST/ directory contains

- The directory POST/POST_RELEASE/ contains the resampled post-corrected segmentation images in the same geometry (images have the same dimensions along X, Y and Z), with drift compensation (the eventual motion of the sample under the microscope has been compensated).
- In addition to a 2D+t movie made from the resampled fusion and the segmentation images, the directory MOVIES/ contains a 2D+t movie made from the resampled post-corrected segmentation images in the sub-directory POST/POST_RELEASE/.
- The template image template_to-20.inr in the directory TRSFS/ is now of size 323 × 265 × 348 with a voxel size of 0.6, has the same size than the one computed in section 2.7, which is expected since the post-correction does not change the background. Note that all resampled images (in the FUSE/FUSE_RELEASE/, the POST/POST_RELEASE/, and the SEG/SEG_RELEASE/ directories have the same geometry than the template image.

```
1 PATH_EMBRYO = '.'
2
3 EN = '2019-Tutorial100'
4
5 begin = 0
6 end = 20
7
8 EXP_INTRAREG = 'POST'
```

Figure 2.12: Tutorial parameter file for the sequence properties from the co-registered post-corrected segmentation images.

2.11 Sequence properties computation [2]

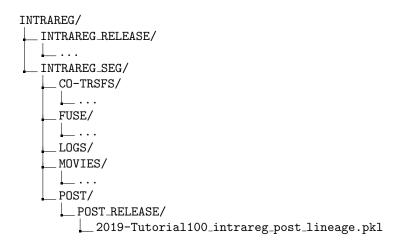
We assume that we are located in the directory /path/to/astec/tutorial/tuto-astec1/. Computing cell properties as well as lineage assumes that segmentation or post-corrected segmentation (see section 2.11) images have been co-registered (see sections 2.7 and 2.10). Extracting the sequence properties from the co-registered segmentation images is done with

\$ X-embryoproperties.py -p parameters/X-embryoproperties-tutorial-parameters-post.py X-embryoproperties-tutorial-parameters-post.py being the dedicated parameter file (figure 2.12).

- The variable PATH_EMBRYO is the path to the directory where the directory FUSE/ is located. It can be either relative (as in the above example) or global (it could have been /path/to/astec/tutorial/tuto-astec1/).
- The variable EN is the prefix after which the images are named.
- The variables begin and end set respectively the first and the last index of the input time points to be processed.
- the variable EXP_INTRAREG set the suffix of the sub-directory of the INTRAREG/ directory where to search post-corrected segmentation or segmentation images.

Since the directory INTRAREG/INTRAREG_POST/ contains the co-registered post-corrected segmentation images (in the POST/POST_RELEASE/ sub-directory), properties will be computed from these images preferably to the co-registered segmentation images (in the SEG/SEG_RELEASE/ sub-directory).

After processing, some files appears in the INTRAREG/INTRAREG_POST/POST/ sub-directory



```
2019-Tutorial100_intrareg_post_lineage.txt
2019-Tutorial100_intrareg_post_lineage.xml
2019-Tutorial100_intrareg_post_t000.inr
2019-Tutorial100_intrareg_post_t020.inr
SEG/
TRSFS_t0-20/
```

Those files have the same content than the ones already presented in section 2.8.

Chapter 3

User guide: command line interfaces

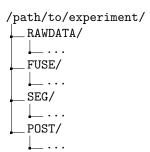
The Astec distribution contains 4 sub-directories

```
path/to/astec/
    documentation/
    parameter-file-examples/
    src/
    tutorial/
```

- documentation/ contains this documentation
- parameter-file-examples/ contains examples of parameter files for the command line interfaces (CLIs) that are introduced below. These files are named after the CLI name.
- src/ contains the command line interfaces (CLIs) and the code.
- tutorial/ contains a tutorial (see chap. 2) along with a toy example.

Data organization

It is assumed that there will be one directory per experiment. This directory contains the acquired data, but will also contain the result data as depicted below.



RAWDATA/ is assumed to contain the raw data (ie acquired images from the MuViSPIM microscope), while the other subdirectories will contain processing results.

3.1 1-fuse.py

The fusion is made of the following steps.

- 1. Optionally, a slit line correction. Some Y lines may appear brighter in the acquisition and causes artifacts in the reconstructed (i.e. fused) image. By default, it is not done.
- 2. A change of resolution in the X and Y directions only (Z remains unchanged). It allows to decrease the data volume (and then the computational cost) if the new pixel size (set by target_resolution) is larger than the acquisition one.
- 3. Optionally, a crop of the resampled acquisitions. It allows to decrease the volume of data, hence the computational cost. The crop is based on the analysis of a MIP view (in the Z direction) of the volume, and thus is sensitive to hyper-intensities if any. By default, it is done.
- 4. Optionally, a mirroring of the 'right' image. It depends on the value of raw_mirrors variable.
- 5. Co-registration of the 3 last images onto the first one (the acquisition from the left camera for stack #0) considered as a reference. The reference image is resampled again, to get an isotropic voxel (whose size is given by target_resolution), i.e. the voxel size is the same along the 3 directions: X, Y, Z. There are two alternative methods.
 - (a) The direct fusion method. Each of the 3 last images is *linearly* co-registered onto the reference image.
 - (b) The hierarchical method. Each stack is first reconstructed (with the acquisition couple of both left and right cameras), then stack #1 is non-linearly co-registered onto stack #0. From this last registration, non-linear co-registrations are deduced for the stack #1 acquisitions, while linear co-registration is still considered for the right camera acquisition of stack #0.
- 6. Linear combination of images, weighted by an ad-hoc function.
- 7. Optionally, a crop of the fused image, still based on the analysis of a MIP view (in the Z direction). By default, it is done.

3.1.1 1-fuse.py options

The following options are available:

- -h prints a help message
- -p <u>file</u> set the parameter file to be parsed
- -e path set the path to the directory where the RAWDATA/ directory is located
- -k allows to keep the temporary files
- -f forces execution, even if (temporary) result files are already existing
- -v increases verboseness (both at console and in the log file)
- -nv no verboseness
- -d increases debug information (in the log file)
- -nd no debug information

3.1.2 Important parameters in the parameter file

A simple parameter file for fusion is described in the tutorial section 2.2. A more comprehensive parameter file example is provided in the parameter-file-examples/ directory.

Indicating the right values of the acquisition parameters is crucial; these parameters are

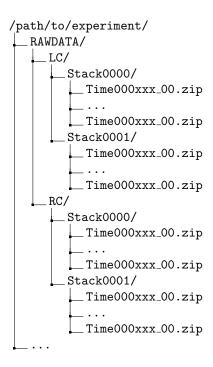
- raw_ori is a parameter describing the acquisition orientation of the acquisition of the second pair of images. Its value can be either 'left' (orientation of 270°) or 'right' (orientation of 90°).
- raw_mirrors is a parameter indicating whether the right camera images have to be mirrored or not. Its value is either False or True.
- raw_resolution is the voxel size (along the 3 dimensions X, Y and Z) of the acquired images.
- target_resolution is the desired isotropic (the same along the 3 dimensions) voxel size for the result fusion images.
- begin gives the index of the first time point to be processed.
- end gives the index of the last time point to be processed.

When one may not be sure of the raw_ori and raw_mirrors right values, it is advised to perform the fusion on only one time point (by indicating the same index for both begin and end), with the four possibilities for the variable couple (raw_ori, raw_mirrors), i.e. ('left', False), ('left', True), ('right', False), and ('right', True). It comes to write four parameter files that differ only for the parameters raw_ori, raw_mirrors, and EXP_FUSE (to store the fusion result in different directories, see section 3.1.4). For these first experiments, it is also advised to set target_resolution to a large value, in order to speed up the calculations.

3.1.3 Input data

Input data (acquired images from the MuViSPIM microscope) are assumed to be organized in a separate RAWDATA/ directory in the /path/to/experiment/ directory as depicted below.

- RAWDATA/LC/Stack000 contains the images acquired at the first angulation by the left camera.
- RAWDATA/LC/Stack001 contains the images acquired at the second angulation by the left camera.
- RAWDATA/RC/Stack000 contains the images acquired at the first angulation by the right camera.
- RAWDATA/RC/Stack001 contains the images acquired at the second angulation by the right camera.



where xxx denotes a three digit number (e.g. 000, 001, ...) denoting the time point of each acquisition. The range of time points to be fused are given by the variables begin and end, while the path/path/to/experiment/ has to be assigned to the variable PATH_EMBRYO

Hence a parameter file containing

```
PATH_EMBRYO = /path/to/experiment/
begin = 0
end = 10
```

indicates that time points in [0,10] of the RAWDATA/ subdirectory of /path/to/experiment/ have to be fused.

3.1.3.1 Input data directory names

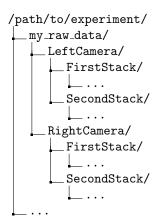
However, directories may be named differently. The variables DIR_RAWDATA, DIR_LEFTCAM_STACKZERO, DIR_RIGHTCAM_STACKONE DIR_LEFTCAM_STACKONE, and DIR_RIGHTCAM_STACKONE allow a finer control of the directory names. The images acquired at the first angulation by the left and the right cameras are searched in the directories

```
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_LEFTCAM_STACKZERO>
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_RIGHTCAM_STACKZERO>
```

while the images acquired at the second angulation by the left and the right cameras are searched in the directories

```
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_LEFTCAM_STACKONE>
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_RIGHTCAM_STACKONE>
```

where <XXX> denotes the value of the variable XXX. Then, to parse the following data architecture



one has to add the following lines in the parameter file

```
DIR_RAWDATA = 'my_raw_data'
DIR_LEFTCAM_STACKZERO = 'LeftCamera/FirstStack'
DIR_RIGHTCAM_STACKZERO = 'RightCamera/FirstStack'
DIR_LEFTCAM_STACKONE = 'LeftCamera/SecondStack'
DIR_RIGHTCAM_STACKONE = 'RightCamera/SecondStack'
```

It has to be noted that, when the stacks of a given time point are in different directories, image file names are tried to be guessed from the directories parsing. It has to be pointed out that indexes have to be encoded with a 3-digit integer with 0 padding (i.e. 000, 001, ...) and that has to be the only variation in the file names (within each directory).

3.1.3.2 Input data image file names

Images acquired from the left and the right cameras may be stored in the same directory, but obviously with different names as in

```
/path/to/experiment/

__RAWDATA/
__stack_0_channel_0
__Cam_Left_00xxx.zip
__...
__Cam_Right_00xxx.zip
```

```
_____stack_1_channel_0
____Cam_Left_00xxx.zip
_____.
___Cam_Right_00xxx.zip
```

The parameter file has then to contain the following lines to indicate the directory names.

```
DIR_LEFTCAM_STACKZERO = 'stack_0_channel_0'
DIR_RIGHTCAM_STACKZERO = 'stack_0_channel_0'
DIR_LEFTCAM_STACKONE = 'stack_1_channel_0'
DIR_RIGHTCAM_STACKONE = 'stack_1_channel_0'
```

In addition, to distinguish the images acquired by the left camera to those acquired by the right one, one has to give the image name prefixes, i.e. the common part of the image file names before the 3-digit number that indicates the time point. This is the purpose of the variables acquisition_leftcam_image_prefix and acquisition_rightcam_image_prefix. The parameter file has then to contain the following lines not only to indicate the directory names but also the image file name prefixes.

```
DIR_LEFTCAM_STACKZER0 = 'stack_0_channel_0'
DIR_RIGHTCAM_STACKZER0 = 'stack_0_channel_0'
DIR_LEFTCAM_STACKONE = 'stack_1_channel_0'
DIR_RIGHTCAM_STACKONE = 'stack_1_channel_0'
acquisition_leftcam_image_prefix = 'Cam_Left_00'
acquisition_rightcam_image_prefix = 'Cam_Right_00'
```

3.1.3.3 Multichannel acquisition

In case of multichannel acquisition, the fusion is computed for the first channel, and the computed parameters (e.g. transformations, etc.) are also used for the other channels.

For a second channel, the images acquired at the first angulation by the left and the right cameras are searched in the directories

```
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_LEFTCAM_STACKZERO_CHANNEL_2>
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_RIGHTCAM_STACKZERO_CHANNEL_2>
```

while the images acquired at the second angulation by the left and the right cameras are searched in the directories

```
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_LEFTCAM_STACKONE_CHANNEL_2>
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_RIGHTCAM_STACKONE_CHANNEL_2>
```

For a third channel, the images acquired at the first angulation by the left and the right cameras are searched in the directories

```
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_LEFTCAM_STACKZERO_CHANNEL_3>
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_RIGHTCAM_STACKZERO_CHANNEL_3>
```

while the images acquired at the second angulation by the left and the right cameras are searched in the directories

```
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_LEFTCAM_STACKONE_CHANNEL_3>
<PATH_EMBRYO>/<DIR_RAWDATA>/<DIR_RIGHTCAM_STACKONE_CHANNEL_3>
```

3.1.4 Output data

The variable target_resolution allows to set the desired isotropic (the same along the 3 dimensions) voxel size for the result fusion images.

3.1.4.1 Output data directory names

The resulting fused images are stored in sub-directory FUSE/FUSE_<EXP_FUSE> under the /path/to/experiment/directory

```
/path/to/experiment/
| RAWDATA/
| L ...
| FUSE/
| FUSE_<EXP_FUSE>/
| L ...
```

where <EXP_FUSE> is the value of the variable EXP_FUSE (its default value is 'RELEASE'). Hence, the line

```
EXP_FUSE = 'TEST'
```

in the parameter file will create the directory FUSE/FUSE_TEST/ in which the fused images are stored. For instance, when testing for the values of the variable couple (raw_ori, raw_mirrors), a first parameter file may contain

```
raw_ori = 'left'
raw_mirrors = False
begin = 1
end = 1
EXP_FUSE=TEST-LEFT-FALSE
a second parameter file may contain
raw_ori = 'left'
raw_mirrors = True
begin = 1
end = 1
```

EXP_FUSE=TEST-LEFT-TRUE

etc. The resulting fused images will then be in different directories

```
/path/to/experiment/
__RAWDATA/
____...
__FUSE/
____FUSE_TEST-LEFT-FALSE/
____...
__FUSE_TEST-LEFT-TRUE/
____...
```

This will ease their visual inspection to decide which values of the variable couple (raw_ori, raw_mirrors) to use for the fusion.

3.1.4.2 Output data file names

Fused image files are named after the variable EN: <EN>_fuse_t<xxx>.inr where <xxx> is the time point index encoded by a 3-digit integer (with 0 padding).

3.1.4.3 Multichannel acquisition

Variables EXP_FUSE_CHANNEL_2 and EXP_FUSE_CHANNEL_3 allows to set the directory names for the resulting fused images of the other channels.

3.1.5 Fusion parameters

3.1.5.1 Step 3: raw data cropping

For computational cost purposes, raw data (images acquired by the MuViSPIM microscope) are cropped (only in X and Y dimensions) before co-registration. A threshold is computed with Otsu's method [Ots79] on the maximum intensity projection (MIP) image. The cropping parameters are computed to keep the above-threshold points in the MIP image, plus some extra margins. Hyper-intense areas may biased the threshold computation, hence the cropping.

To desactivate this cropping, the line

raw_crop = False

has to be added in the parameter file.

3.1.5.2 Step 5: image co-registration

To fuse the images, they are co-registered onto a reference one. Co-registration are conducted only on the first channel (in case of multiple channel acquisitions), and the computed transformations are also applied onto the other channels. The reference image is chosen as being the acquisition from the left camera for the first stack (also denoted stack #0). The co-registration strategy is given by the variable fusion_strategy.

• The line

fusion_strategy = 'direct-fusion'

will set the co-registration strategy to the one described in [Gui15, ?]: each acquisition image is linearly co-registered with the reference one, i.e. the one from the left camera and for the first stack.

• The line

fusion_strategy = 'hierarchical-fusion'

defines a hierarchical co-registration strategy. First, stacks are reconstructed with the two acquisition from the left and the right cameras: here the left and right acquisitions are linearly co-registered. Then, the second stack (stack #1) is non-linearly co-registered onto the first one (stack #0). Transformation compositions yield the final transformations that co-registered each acquisition onto the reference one. Note that the acquisition from the right camera of the first stack is only linearly co-registered with the reference acquisition, while both acquisitions for the second stack are non-linearly co-registered.

3.1.5.2.1 Acquisitions linear co-registration The linear co-registrations are either used to co-registered each acquisition onto the reference one in the 'direct-fusion' strategy, or to build stacks from the left and right cameras in the 'hierarchical-fusion' strategy. Variables that controls the linear co-registrations are either prefixed by fusion_preregistration_ or by fusion_registration_.

To decrease the computational cost, images are normalized and cast on one byte before registration. While it generally does not degrade the registration quality, it may induce troubles when hyper-intensities areas are present in the image. In such a case, the useful information may then be summarized in only a few intensity values.

Intensity normalization in registration can be deactivated by adding the following line in the parameter file

fusion_registration_normalization = False

To verify whether a good quality registration can be conducted, the searched transformation type can be changed for a simpler one than affine. Adding the following line in the parameter file.

fusion_registration_transformation_type = translation

will search for a translation which is supposed to be sufficient, according that only translations relates the 4 acquisitions of the MuViSPIM microscope (in a perfect setting). If the search for an affine transformation (the default behavior) failed (the fusion looks poor) while the search for a translation is successful (the fusion looks good), a two-steps registration may help to refine the found translation by a subsequent affine transformation as explained below.

Hyper-intensities areas may also bias the threshold calculation used for the automatic crop (step 3 of fusion). In such cases, the iterative registration method may find a local minimum that is not the desired one, because the relative positions of the two images to be co-registered are too far apart. To circumvent such a behavior, a two-steps registration can be done. It consists on a first pre-registration with a transformation with fewer degrees of freedom (i.e. a 3D translation).

This pre-registration can be activated by adding the following line in the parameter file.

```
fusion_preregistration_compute_registration = True
```

It may be also preferable to deactivate the image normalization for both registration steps with

```
fusion_preregistration_normalization = False
fusion_registration_normalization = False
```

3.1.5.2.2 Stacks non-linear co-registration Variables that controls the non-linear co-registrations are either prefixed by fusion_stack_preregistration_ or by fusion_stack_registration_.

3.1.5.3 Step 6: linear combination of co-registered stacks

The resampled co-registered stacks are fused together by the means of a weighted linear combination. The choice of the weighting function is controlled by the variable fusion_weighting, eventually suffixed by _channel_[1,2,3].

• The line

```
fusion_weighting = 'guignard-weighting'
```

will set the weighting function to the one described in [Gui15]: more weight are given to sections close to the camera and it also takes into account the traversed material. Such a weighting depends on the stacks to be fused.

• The line

```
fusion_weighting = 'uniform'
```

comes to the average of the resampled co-registered stacks. Such a weighting does not depend on the stacks to be fused.

When using the variable fusion_weighting, the same weights (computed on the first channel to be processed) are used for all fusion. However, different weighting functions can be used for the channels to be fused by using the variables fusion_weighting_channel_[1,2,3], eg

```
fusion_weighting_channel_1 = 'guignard-weighting'
fusion_weighting_channel_2 = 'uniform'
```

3.1.5.4 Step 7: fused data cropping

To save disk storage, fused images are cropped at the end of the fusion stage. To desactivate this cropping, the line

```
fusion_crop = False
```

has to be added in the parameter file.

3.1.6 Troubleshooting

- The fused images are obviously wrong.
 - 1. Are the values of the variable couple (raw_ori, raw_mirrors) the right ones? Conduct experiments as suggested in section 3.1.2 (see also section 3.1.4) to get the right values.
 - 2. The registration may have failed.
 - (a) Deactivate the 1-byte normalization (see section 3.1.5.2).
 - (b) Try to register with a simpler transformation type (i.e. translation) and/or with a two-steps registration (see section 3.1.5.2).
- The imaged sample is cropped by the image border in the fused image.
 - 1. Check whether the imaged sample was not already cropped in the raw data.
 - 2. The automated cropping may have failed. It is more likely to happen when cropping the raw data, so deactivate it (see section 3.1.5.1). If it still happens, try to deactivate also the fused image cropping (see section 3.1.5.4).

3.1.7 Parameter list

Please also refer to the file parameter-file-examples/1-fuse-parameters.py

- DIR_LEFTCAM_STACKONE see section 3.1.3
- DIR_LEFTCAM_STACKONE_CHANNEL_2 see section 3.1.3
- DIR_LEFTCAM_STACKONE_CHANNEL_3 see section 3.1.3
- \bullet DIR_LEFTCAM_STACKZERO see section 3.1.3
- DIR_LEFTCAM_STACKZERO_CHANNEL_2 see section 3.1.3
- DIR_LEFTCAM_STACKZERO_CHANNEL_3 see section 3.1.3
- DIR_RAWDATA see section 3.1.3
- DIR_RAWDATA_CHANNEL_2 see section 3.1.3
- DIR_RAWDATA_CHANNEL_3 see section 3.1.3
- \bullet DIR_RIGHTCAM_STACKONE see section 3.1.3
- DIR_RIGHTCAM_STACKONE_CHANNEL_2 see section 3.1.3
- DIR_RIGHTCAM_STACKONE_CHANNEL_3 see section 3.1.3
- DIR_RIGHTCAM_STACKZERO see section 3.1.3
- DIR_RIGHTCAM_STACKZERO_CHANNEL_2 see section 3.1.3
- DIR_RIGHTCAM_STACKZERO_CHANNEL_3 see section 3.1.3
- EN see section 3.1.4

- EXP_FUSE see section 3.1.4
- EXP_FUSE_CHANNEL_2 see section 3.1.4
- EXP_FUSE_CHANNEL_3 see section 3.1.4
- PATH_EMBRYO see section 3.1.3
- RESULT_IMAGE_SUFFIX_FUSE
- acquisition_leftcam_image_prefix see section 3.1.3
- acquisition_mirrors same as raw_mirrors
- acquisition_orientation same as raw_ori
- \bullet acquisition_resolution same as raw_resolution
- acquisition_rightcam_image_prefix see section 3.1.3
- acquisition_slit_line_correction
- begin see section 3.1.2
- default_image_suffix
- delta
- end see section 3.1.2
- fusion_crop see section 3.1.5.4
- fusion_margin_x_0
- fusion_margin_x_1
- fusion_margin_y_0
- fusion_margin_y_1
- fusion_strategy see section 3.1.5.2
- fusion_preregistration_compute_registration see section 3.1.5.2
- fusion_preregistration_lts_fraction
- fusion_preregistration_normalization see section 3.1.5.2
- fusion_preregistration_pyramid_highest_level
- fusion_preregistration_pyramid_lowest_level
- fusion_preregistration_transformation_estimation_type
- fusion_preregistration_transformation_type
- fusion_registration_compute_registration
- fusion_registration_lts_fraction
- fusion_registration_normalization see section 3.1.5.2
- fusion_registration_pyramid_highest_level
- fusion_registration_pyramid_lowest_level
- fusion_registration_transformation_estimation_type
- fusion_registration_transformation_type see section 3.1.5.2
- fusion_stack_preregistration_compute_registration
- fusion_stack_preregistration_lts_fraction
- fusion_stack_preregistration_normalization
- fusion_stack_preregistration_pyramid_highest_level
- fusion_stack_preregistration_pyramid_lowest_level
- fusion_stack_preregistration_transformation_estimation_type
- fusion_stack_preregistration_transformation_type
- fusion_stack_registration_compute_registration
- fusion_stack_registration_lts_fraction
- fusion_stack_registration_normalization
- fusion_stack_registration_pyramid_highest_level
- fusion_stack_registration_pyramid_lowest_level
- $\bullet \ \, fusion_stack_registration_transformation_estimation_type$
- fusion_stack_registration_transformation_type
- fusion_weighting
- fusion_weighting_channel_1
- fusion_weighting_channel_2
- fusion_weighting_channel_3

- raw_crop see section 3.1.5.1
- raw_delay
- raw_margin_x_0
- raw_margin_x_1
- raw_margin_y_0
- raw_margin_y_1
- raw_mirrors see section 3.1.2
- raw_ori see section 3.1.2
- raw_resolution see section 3.1.2
- result_image_suffix
- target_resolution see section 3.1.4

3.2 1.5-intraregistration.py

The sequence intra-registration procedure can be done either after the fusion step, or after the (post-)segmentation step. It aims at

- compensating for the eventual motion of the imaged sample with respect to the microscope
- resampling the fusion and/or the segmentation images into a common frame/geometry, so they can better be compared, and
- building 2D+t images made of 2D sections from either the fusion and/or the segmentation images, so that the quality of the fusion and/of the tracking step can be visually assessed.

The intra-registration procedure is made of the following steps:

- 1. Co-registration of pairs of successive fused images (section 3.2.3.1). This yields the transformations $T_{t+1\leftarrow t}$. Fused images are located in <EMBRYO>/FUSE/FUSE_<EXP_FUSE>: the parameter EXP_FUSE is either set in the parameter file or is set at RELEASE. This step may be long.
- 2. Composition of transformations issued from the co-registration step. This step computes the transformations $T_{ref\leftarrow t}$ towards a reference image ref given by the parameter intra_registration_reference_index.
- 3. Computation of the *template* image (section 3.2.3.2). This *template* image dimension are computed so that the useful information of all resampled images fits into it. Useful information can be issued from either the fused sequence, the segmentation sequence or the post-segmentation sequence. It is indicated by the intra_registration_template_type which value can be either 'FUSION', 'SEGMENTATION', or 'POST-SEGMENTATION'. This step may be long.
- 4. Resampling of either the fused or the segmentation images (section 3.2.3.3). Note that changing the parameters for this step will not require to re-compute the first steps.
- 5. Extraction of 2D+t images from the resampled sequences (section 3.2.3.4). Note that changing the parameters for this step (i.e. requiring extra movies) will not require to re-compute the first steps, with an eventual exception for the resampling step.

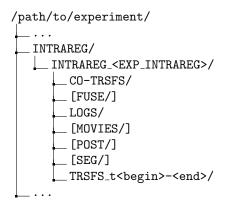
3.2.1 1.5-intraregistration.py options

The following options are available:

- -h prints a help message
- -p <u>file</u> set the parameter file to be parsed
- -e path set the path to the directory where the RAWDATA/ directory is located
- -t file set the resampling transformation file for the reference image (see section 3.2.3.2)
- -a string set the resampling transformation angles for the reference image (see section 3.2.3.2)
- -k allows to keep the temporary files
- -f forces execution, even if (temporary) result files are already existing
- -v increases verboseness (both at console and in the log file)
- -nv no verboseness
- -d increases debug information (in the log file)
- -nd no debug information

3.2.2 Output data

The results are stored in sub-directories INTRAREG/INTRAREG-<EXP_INTRAREG> under the /path/to/experiment/directory where <EXP_INTRAREG> is the value of the variable EXP_INTRAREG (its default value is 'RELEASE').



Output data are of two kinds: image series (fused images, segmentation images, post-corrected segmentation images) can be resampled in the same common geometry (also known as the *template*), see section 3.2.3.3, and 3D (ie 2D+t) images of the evolution (with respect to time) of one section (XY, XZ, or YZ) of the images of the series can be built, see section 3.2.3.4.

3.2.3 Intra-registration parameters

3.2.3.1 Step 1: co-registration

Default registration parameters for the co-registration are set by:

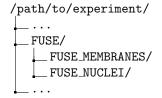
```
# intra_registration_compute_registration = True
# intra_registration_transformation_type = 'rigid'
# intra_registration_transformation_estimation_type = 'wlts'
# intra_registration_lts_fraction = 0.55
# intra_registration_pyramid_highest_level = 6
# intra_registration_pyramid_lowest_level = 3
# intra_registration_normalization = True
```

Computed transformations are stored in INTRAREG/INTRAREG_<EXP_INTRAREG>/CO-TRSFS. It may be advised to set the pyramid lowest level value to some higher value to speed up the co-registrations (recall that all pairs of successive images will be co-registered, i.e.

```
intra_registration_pyramid_lowest_level = 4
```

Co-registration are computed using the fused images of /path/to/experiment/FUSE/FUSE_<EXP_FUSE>. If EXP_FUSE is a list of strings (ie indicates a list a directories) rather than a single string, the fused image from the first directory are used for the co-registration computation.

Typically, if there are several fused series (eg, in case of multi-channel acquisition) as in



Specifying

```
EXP_FUSE = ['MEMBRANES', 'NUCLEI']
```

in the parameter file implies that co-registrations will be done on the fused images from FUSE/FUSE_MEMBRANES/.

3.2.3.2 Step 3: template building

```
# intra_registration_reference_index = None
# intra_registration_reference_resampling_transformation_file = None
# intra_registration_reference_resampling_transformation_angles = None
# intra_registration_template_type = "FUSION"
# intra_registration_template_threshold = None
# intra_registration_margin = None
# intra_registration_resolution = 0.6
# intra_registration_resolution = False
```

- The intra_registration_reference_index allows to choose the reference image (the one which remains still, i.e. up to a translation), by default it is the first image image of the series (associated to begin). However, it may happen that this image has to be reoriented to fit the user's expectation. The resampling transformation¹, that re-orient the reference image, can then be given and will be applied to the whole series.
 - intra_registration_reference_resampling_transformation_file can be given a resampling transformation file name.
 - intra_registration_reference_resampling_transformation_angles can be given a string describing the successive rotations (with respect to the frame axis) to be applied. E.g. the string "X 30 Y 50" defines a resampling transformation equal to $R_X(30) \circ R_Y(50)$ where $R_X(30)$ is a rotation of 30 degrees around the X axis and $R_Y(50)$ is a rotation of 50 degrees around the Y axis.
- Depending on intra_registration_template_type ('FUSION', 'SEGMENTATION' or 'POST-SEGMENTATION'), the two latter assume obviously that the segmentation has been done), the template image can be built either after the fusion or the segmentation images. If no threshold is given by intra_registration_template_threshold template will be large enough to include all the transformed fields of view (in this case, the template is the same whatever intra_registration_template_type is).

If intra_registration_template_type='FUSION' (respectively 'SEGMENTATION' and 'POST-SEGMENTATION'), the template is built from the images of the first directory indicated by EXP_FUSE (respectively EXP_SEG and EXP_POST) in case of EXP_FUSE contains a list of strings.

If a threshold is given, the built template will be large enough to include all the transformed points above the threshold. E.g., the background is labeled with either '1' or '0' in segmentation images, then a threshold of '2' ensures that all the embryo cells will not be cut by the resampling stage. In this case, adding an additional margin (with intra_registration_margin) to the template could be a good idea for visualization purpose.

• Specifying using a different resolution for the drift-compensated series than the target_resolution (the resolution of the fused images) allows to decrease the resampled images volume. This can be achieved by setting intra_registration_resolution to the desired value (default is 0.6).

¹The resampling transformation is the one that goes from the destination image towards the input image.

• Last, co-registrations may have been computed during a first computation, fused images being used to compute the template. However, if a subsequent segmentation has been conducted, a smaller template is likely to be computed (with the segmentation images to build the template), without recomputing the co-registration. This is the purpose of the variable intra_registration_rebuild_template. If set to True, it forces to recompute the template as well as the transformations from the co-registrations (that are not re-computed). Obviously, resampling as well as 2D+t movies are also re-generated.

As an example, building a template image after the segmentation images can be done with

```
# intra_registration_reference_index = None
intra_registration_template_type = "SEGMENTATION"
intra_registration_template_threshold = 2
# intra_registration_resolution = 0.6
intra_registration_margin = 10
```

Computed transformations from the *template* image as well as the *template* image itself are stored in INTRAREG/INTRAREG</TRSFS_t<F>-<L>/ where <F> and L are the first and the last index of the series (specified by begin and end from the parameter file).

3.2.3.3 Step 4: resampling fusion/segmentation images

The resampling of the fused and/or segmentation images are done depending on the value of the following variables (here commented). Resampling is done either if the following parameters are set to True or if movies are requested to be computed (section 3.2.3.4).

```
# intra_registration_resample_fusion_images = True
# intra_registration_resample_segmentation_images = False
# intra_registration_resample_post_segmentation_images = False
```

This default behavior implies that the fusion images will be resampled while the segmentation and the post-corrected segmentation images are not.

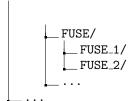
Resampled images will be stored in the INTRAREG/INTRAREG-<EXP_INTRAREG/> directory, with the same hierarchy than under /path/to/experiment. E.g.

```
/path/to/experiment/
_______FUSE/
_______FUSE_1/
_______FUSE_2/
_________Specifying

EXP_FUSE = ['1', '2']
```

in the parameter file causes the resampling of both fused image series (FUSE/FUSE_1/ and FUSE/FUSE_2/)

```
/path/to/experiment/
...
FUSE/
FUSE_1/
FUSE_2/
INTRAREG/
INTRAREG/
```



The same behavior stands for ${\tt EXP_SEG}$ and ${\tt EXP_POST}.$

3.2.3.4 Step 5: 2D+t movies

For either visual assessment or illustration purposes, 2D+t (i.e. 3D) images can be built from 2D sections extracted from the resampled temporal series. This is controlled by the following parameters:

```
# intra_registration_movie_fusion_images = True
# intra_registration_movie_segmentation_images = False
# intra_registration_movie_post_segmentation_images = False
# intra_registration_xy_movie_fusion_images = [];
# intra_registration_xz_movie_fusion_images = [];
# intra_registration_yz_movie_fusion_images = [];
# intra_registration_xy_movie_segmentation_images = [];
# intra_registration_xz_movie_segmentation_images = [];
# intra_registration_yz_movie_segmentation_images = [];
# intra_registration_xy_movie_post_segmentation_images = [];
# intra_registration_xz_movie_post_segmentation_images = [];
# intra_registration_yz_movie_post_segmentation_images = [];
```

If intra_registration_movie_fusion_images is set to True, a movie is made with the XY-section located at the middle of each resampled fusion image (recall that, after resampling, all images have the same geometry). Additional XY-movies can be done by specifying the wanted Z values in intra_registration_xy_movie_fusion E.g.

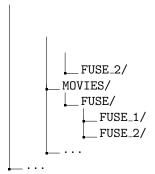
```
intra_registration_xy_movie_fusion_images = [100, 200];
```

will build two movies with XY-sections located respectively at Z values of 100 and 200. The same stands for the other orientation and for the resampled segmentation images.

Movies will be stored in the INTRAREG/INTRAREG-<EXP_INTRAREG>/MOVIES/ directory, with the same hierarchy than under /path/to/experiment. E.g.,

```
EXP_FUSE = ['1', '2']
```

in the parameter file results in



The same behavior stands for EXP_SEG and EXP_POST.

3.2.4 Parameter list

Please also refer to the file parameter-file-examples/1.5-intraregistration-parameters.py

- EN
- EXP_FUSE

String (str type) or list (list type) of strings. It indicates what are the fused images directories, of the form /path/to/experiment/FUSE/FUSE_<EXP_FUSE>.

```
EXP_FUSE = 'exp1'
EXP_FUSE = ['exp1', 'exp2']
```

are both valid. Recall that the default value of EXP_FUSE is 'RELEASE'.

- EXP_INTRAREG
- EXP_POST
- EXP_SEG
- PATH_EMBRYO
- begin
- default_image_suffix
- delta
- end
- intra_registration_compute_registration
- intra_registration_lts_fraction
- intra_registration_margin
- intra_registration_movie_fusion_images
- intra_registration_movie_post_segmentation_images
- intra_registration_movie_segmentation_images
- intra_registration_normalization
- intra_registration_pyramid_highest_level
- intra_registration_pyramid_lowest_level
- intra_registration_rebuild_template

If set to True, force to recompute the template as well as the transformations from existing coregistrations (that are not re-computed). It is useful when a first intra-registration has been done with only the fusion images: a second intra-registration with the segmentation images as template can be done without recomputing the co-registrations.

- intra_registration_reference_index
- intra_registration_reference_resampling_transformation_file
- intra_registration_reference_resampling_transformation_angles
- intra_registration_resample_fusion_images
- intra_registration_resample_post_segmentation_images
- intra_registration_resample_segmentation_images
- intra_registration_resolution
- intra_registration_sigma_segmentation_images

- intra_registration_template_threshold
- intra_registration_template_type
- intra_registration_transformation_estimation_type
- intra_registration_transformation_type
- intra_registration_xy_movie_fusion_images
- intra_registration_xy_movie_post_segmentation_images
- intra_registration_xy_movie_segmentation_images
- intra_registration_xz_movie_fusion_images
- intra_registration_xz_movie_post_segmentation_images
- intra_registration_xz_movie_segmentation_images
- intra_registration_yz_movie_fusion_images
- intra_registration_yz_movie_post_segmentation_images
- intra_registration_yz_movie_segmentation_images
- result_image_suffix

3.3 2-mars.py

The name mars comes from $[FDM^+10]$ where MARS is the acronym of multiangle image acquisition, 3D reconstruction and cell segmentation.

This method aims at producing a segmentation of a membrane cell image (e.g. a fused image) into a segmention image. This segmentation image is a integer-valued image where each integer labeled an unique cell in the image. By convention, '1' is the background label, while cells have labels greater than 2. It is is made of the following steps:

- 1. Optionally, a transformation of the input image.
- 2. A seeded watershed.

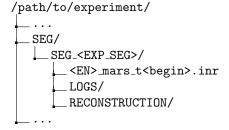
3.3.1 2-mars.py options

The following options are available:

- -h prints a help message
- -p <u>file</u> set the parameter file to be parsed
- -e path set the path to the directory where the RAWDATA/ directory is located
- -k allows to keep the temporary files
- -f forces execution, even if (temporary) result files are already existing
- -v increases verboseness (both at console and in the log file)
- -nv no verboseness
- -d increases debug information (in the log file)
- -nd no debug information

3.3.2 Output data

The results are stored in sub-directories SEG/SEG_<EXP_SEG> under the /path/to/experiment/ directory where where <EXP_SEG> is the value of the variable EXP_SEG (its default value is 'RELEASE').



3.3.3 Segmentation parameters

3.3.3.1 Input image for watershed computation

Before the watershed segmentation, the input image may be pre-processed. This pre-processing is controlled by the two variables.

- mars_intensity_transformation whose values are to be chosen in None, 'Identity', or 'Normalization_to_u8'.

 Default is 'Identity'.
- mars_intensity_enhancement whose values are to be chosen in None or GACE. Default is None.

Each of these variables, if not None, induce a transformation of the input (i.e. fused) image. If both values are not known, the input image for the watershed is the result of the maximum operator over the two images.

- mars_intensity_transformation = 'Identity': the input image is not transformed.
- mars_intensity_transformation = 'Normalization_to_u8': input images are usually encoded on 2 bytes. The choice transformed the input image in an 1-byte image by linearly mapping the input image values from $[I_{min}, I_{max}]$ to [0, 255]. I_{min} and I_{max} correspond respectively to the 1% and to the 99% percentiles of the input image cumulative histogram. Values below I_{min} are set to 0 while values above I_{max} are set to 255.
- mars_intensity_enhancement = 'GACE': GACE stands for Global Automated Cell Extractor. This is the method described in [MGFM14, Mic16]. It consists in
 - 1. extracting a centerplane image of the membranes,
 - 2. thresholding this centerplane image, and
 - 3. reconstruct the membranes through a tensor voting method.

If the input image is transformed before segmented, the transformed image is named <EN>_fuse_t<begin>_membrane.in and stored in the directory SEG/SEG_<EXP_SEG>/RECONSTRUCTION/ if the value of the variable mars_keep_reconstruction is set to True.

3.3.3.2 Seed extraction

The seed extraction is made of the following steps:

- 1. Gaussian smoothing of the input image, the gaussian standard deviation being given by the variable watershed_seed_sigma.
- 2. Extraction of the h-minima of the previous image, h being given by the variable watershed_seed_hmin.
- 3. Hysteresis thresholding (and labeling) of the h-minima image, with a high threshold equal to watershed_seed_high_t (default is h) and and a low threshold equal to 1. It then only selects the h-minima that have an actual depth of h.

Given the seeds, the watershed is performed on the smoothed input image (gaussian standard deviation being given by the variable watershed_membrane_sigma).

3.3.3.3 Seed correction

Several rounds of correction of the computed seeds can be done. At each round, different seeds can be assigned the same label (and this will fuse the further reconstructed cells) or new seeds (each new seed is a single voxel) can be added. See the 'seed_edition_files' variable for details.

When correcting seeds, it is advised to launch 2-mars.py with the '-k' option. Indeed, temporary files, as the seed image, are kept in a temporary directory located in the SEG/SEG_'EXP_SEG'/ directory and then re-used, and not recomputed at each 2-mars.py use.

3.3.3.4 Seeded watershed

The watershed is performed with the previously computed seeds on a smooth input image.

3.3.4 Parameter list

Please also refer to the file parameter-file-examples/2-mars-parameters.py

- EN
- EXP_FUSE
- EXP_SEG
- PATH_EMBRYO
- begin
- default_image_suffix
- delta
- mars_begin
- mars_end
- mars_hard_threshold
- mars_intensity_enhancement
- mars_intensity_transformation
- mars_keep_reconstruction
- mars_manual
- mars_manual_sigma
- mars_sample: this parameter sets the fraction of the binary centerplanes that will be used for tensor voting (step 3). Points being randomly drawn, results are not strictly reproducible if the code is re-run with the same sets of parameters. Using a larger value (smaller than or equal to 1.0) increases the reproductibility but induces a larger computational cost.
- mars_sensitivity: this parameter sets the sensitivity for the centerplanes thresholding of step 2. It is set to 0.99 by default. Using larger value (smaller than or equal to 1.0, say 0.9999) allows to extract less-contrasted membranes (for instance cell/background membranes).
- mars_sigma_TV
- mars_sigma_membrane
- result_image_suffix
- seed_edition_dir:
- seed_edition_files: it is a list of lists of 2 elements, each element being a file name. E.g.

These files are assumed to be located in the seed_edition_dir directory.

Each line of a seeds_to_be_fused_00x.txt file contains the labels to be fused, e.g. "10 4 2 24". A same label can be found in several lines, meaning that all the labels of these lines will be fused.

Each line of a seeds_to_be_created_00x.txt contains the coordinates of a seed to be added

- watershed_membrane_sigma: gaussian standard deviation σ (in real unit) to smooth the input image (i.e. the reconstructed image, see section 3.3.3.1) before the watershed segmentation.
- watershed_seed_high_threshold: threshold value to segment the h-minima. has to be choose in [1, h].
- watershed_seed_hmin: h-value for the regional minima extraction.
- watershed_seed_sigma: gaussian standard deviation (in real unit) to smooth the input image (i.e. the reconstructed image, see section 3.3.3.2) before the seed extraction.

3.4 3-manualcorrection.py

The seeded watershed is likely to produce segmentation errors, even with a careful choice of parameters. It is advised to set the parameters to favour over-segmentations insted of under-segmentations since the former are much more easier to correct, which is the purpose of 3-manualcorrection.py.

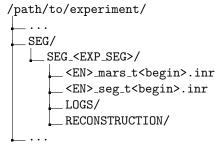
3.4.1 3-manualcorrection.py options

The following options are available:

- -h prints a help message
- $-p \ \underline{file}$ indicates the parameter file to be parsed
- -e path indicates the path to the directory where the RAWDATA/ directory is located
- -k allows to keep the temporary files
- -f forces execution, even if (temporary) result files are already existing
- -v increases verboseness (both at console and in the log file)
- -nv no verboseness
- -d increases debug information (in the log file)
- -nd no debug information
- -i input_image set the input_image file to be corrected. Allows to skip the automated naming of files.
- -o output_image set the resulting ouput_image file to be saved. Allows to skip the automated naming of files.
- -m mapping_file set the mapping_file to be used for the correction.
- -nsc <u>smallest_cells</u> set the number of the smallest cells to be displayed after correction. The smallest cells are the most likely to be issued from an over-segmentation.
- -nlc <u>largest_cells</u> set the number of the largest cells to be displayed after correction. The largest cells are the most likely to be issued from an under-segmentation.

3.4.2 Output data

The results are stored in sub-directories SEG/SEG_<EXP_SEG> under the /path/to/experiment/ directory where <EXP_SEG> is the value of the variable EXP_SEG (its default value is 'RELEASE'). <EN>_seg_t
begin>.inr is the correction of the segmentation image <EN>_mars_t
begin>.inr.



3.4.3 Segmentation correction parameters

3-manualcorrection.py parses a correction file whose name is given by the variable mancor_mapping_file. The syntax of this file is very simple. Lines beginning with # are ignored (and can be used to insert comments in the files). Non-empty lines should contain two numbers separated by a space, and 3-manualcorrection.py will replace the first number by the second in the segmentation file.

E.g. a cell c is recognized to be over-segmented, and then is represented by two labels, says 9 and 10. Thus the line

```
10 9
```

will replace all 10's by 9's in the segmentation image, thus c will only be represented by 9's after correction. See also the tutorial section 2.5 for an other example.

3.4.4 Parameter list

Please also refer to the file parameter-file-examples/3-manualcorrection-parameters.py

- EN
- EXP_SEG
- PATH_EMBRYO
- begin
- default_image_suffix
- delta
- mancor_input_seg_file
- mancor_mapping_file
- mancor_output_seg_file
- mars_begin
- mars_end
- result_image_suffix

3.5 4-astec.py

The name astec comes from the Phd work of L. Guignard [Gui15] where ASTEC is the acronym of adaptive segmentation and tracking of embryonic cells.

3.6 5-postcorrection.py

3.7 X-embryoproperties.py

X-embryoproperties.py can be used either to extract cell properties as well as cell lineage from a coregistered image sequence or to handle a property file (pkl or xml).

3.7.1 X-embryoproperties.py options

The following options are available:

- -h prints a help message
- -p <u>file</u> set the parameter file to be parsed
- -e path set the path to the directory where the RAWDATA/ directory is located
- -i <u>files</u> ... input files (pkl or xml) to be read
- -o <u>files ...</u> output files (pkl or xml) to be read
- ${\tt -c}$ ${\tt \underline{files}}$... files (pkl or xml) to be compared to those given by ${\tt -i}$
- -feature <u>features</u> ... features to be extracted from the input files, that are to be written in the output files. Features have to be chosen in 'lineage', 'h_min', 'volume', 'surface', 'sigma', 'label_in_time', 'barycenter', 'fate', 'fate2', 'fate3', 'fate4', 'all-cells', 'principal-value', 'name', 'contact', 'history', 'principal-vector', 'name-score', 'cell-compactness'
- -property <u>features</u> ... same as -feature
- --diagnosis performs some test on the read properties
- $\hbox{\tt --diagnosis-minimal-volume} \ \, \underline{\hbox{\tt DIAGNOSIS_MINIMAL_VOLUME}} \ \, \mathrm{displays} \ \, \mathrm{all} \ \, \mathrm{cells} \ \, \mathrm{with} \ \, \mathrm{volume} \ \, \mathrm{smaller} \ \, \mathrm{than} \ \, \underline{\hbox{\tt DIAGNOSIS_MINIMAL_VOLUME}}$
- --diagnosis-items <u>DIAGNOSIS_ITEMS</u> minimal number of items to be displayed
- --print-content print the keys of the input file(s) (read as python dictionary)
- --print-keys same as --print-content
- --print-types print types of read features (for debug purpose)
- -k allows to keep the temporary files
- -f forces execution, even if (temporary) result files are already existing

- -v increases verboseness (both at console and in the log file)
- -nv no verboseness
- -d increases debug information (in the log file)
- -nd no debug information

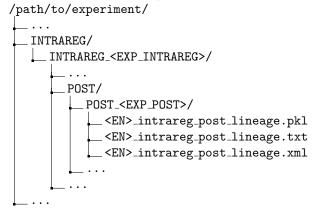
3.7.2 Extracting properties from a co-registered image sequence

When a parameter file is passed after the -p option, X-embryoproperties.py will compute image sequence properties. Computing cell related informations as well as the lineage tree requires that the (post-corrected) segmentation images have already been co-registered (with 1.5-intraregistration.py see section 3.2). X-embryoproperties.py will parse the INTRAREG/INTRAREG<EXP_INTRAREG>/ directory, and will compute the properties from the images in the POST/POST_<EXP_POST>/ sub-directory, if existing, else of from the SEG/SEG_<EXP_SEG>/ sub-directory.

3.7.2.1 Output data

The results are stored in the POST/POST_<EXP_POST>/ or SEG/SEG_<EXP_SEG>/ sub-directory under the INTRAREG/INTRAREG-<EXP_INTRAREG> where <EXP_INTRAREG> is the value of the variable EXP_INTRAREG (its default value is 'RELEASE'). The resulting properties will be stored in the same directory than the images they are issued. It will be stored as a pickle python file, and also as a XML file. Both files contain exactly the same information.

According that the POST/POST_<EXP_POST>/ sub-directory exists (that post-corrected segmentation images have been co-registered), 3 files will be created, named after <EN>



The computed information are

- all_cells All the cell identifiers. Each cell (in a segmentation image) has a given label (ranging from 2 and above, 1 being used for the background) in each image. To uniquely identify a cell in the sequence, it has been given an unique identifier computed by i*1000 + c, i and c denoting respectively the image index (ranging in [<begin>, <end>]) and the cell label.
- cell_barycenter Cell center of mass (in voxel coordinates)
- cell_contact_surface For each cell, give for each neighboring cell the contact surface. The sum of these contact surfaces is the cell surface.
- cell_principal_vectors The cell principal vectors are issued from the diagonalization of the cell covariance matrix (in voxel unit).
- cell_principal_values The cell principal value are issued from the diagonalization of the cell covariance matrix (in voxel unit).
- cell_volume (in voxel unit)
- cell_compactness The cell compactness is defined by $C = \frac{\sqrt[3]{\mathcal{V}}}{\sqrt[3]{\mathcal{S}}}$ where \mathcal{V} is the volume of the cell and \mathcal{S} is its surface.

cell_surface Cell surface (in pixel unit). For this computation, is mandatory that the co-registered images are isotropic (the same voxel size along the 3 dimensions X, Y, and Z).

cell_lineage

The text file <EN>_intrareg_post_lineage.txt contains diagnosis information about the sequence. It lists

- the cell with the smallest sizes as well as the ones with the largest sizes
- the cell with a weird lineage: cells without a mother cell, or cells without daughter cells or having more than 2 daughter cells
- cells having a small intersection with its mother cell with respect to either the mother cell volume or the cell volume.

3.7.2.2 Parameter list

Please also refer to the file parameter-file-examples/X-embryoproperties-parameters.py

- EN
- EXP_INTRAREG
- EXP_POST
- EXP_SEG
- PATH_EMBRYO
- begin
- end
- properties_nb_proc the property computation supports parallelism. However, it appears that the opening of several files at the same time may cause the computation to fail. Thus, the default behavior is a sequential processing. To enable parallelism, this parameter can be set to either any negative value (causing a default parallel behavior) or to a positive value indicating the number of threads to be created.

3.7.3 Handling properties files

X-embryoproperties.py can also help managing property files.

- Converting from xml to pkl and the other way around.
 - \$ X-embryoproperties.py -i file.pkl -o file.xml

convert the pickle file file.pkl into the xml file file.xml

- Converting the lineage information from either an xml or an pkl file to a tlp² file for lineage visualization
 - \$ X-embryoproperties.py -i file.pkl -o file.tlp

convert the pickle file file.pkl into the tlp file file.tlp

- Merging files.
 - \$ X-embryoproperties.py -i file1.pkl file2.xml ...filen.pkl -o
 merge.xml merge.pkl

will merge the files file1.pkl, file2.xml, ..., filen.pkl (note that they can be either xml or pkl) and write the result both in xml and pkl formats.

- Extracting properties.

will extract the cell volume and surface information from the pickle file file.pkl and write them into the xml file file.xml

²Tulip is a Data Visualization Software, see tulip.labri.fr.

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