

The Tale of a Bacteria Battle

*A study on Staphylococcus Aureus, its prevalence, clinical possibilities and
our fighting tools*

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Version 2.13.23

Acknowledgements

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I would also like to acknowledge the magnificent work that the L^AT_EX community does to give the scientific community an incredible piece of software with which to write, as well as to Mark Olson for making his KOMA-Script T_EX template open source, allowing anyone and everyone to use it and modify it for free. On that note, I'd also like to thank my friend Gabriela, for helping me fix a weird issue with the bibliography system. But most importantly, I'd like to thank Núria Feliu, Imma Garcia and the rest of the science department for their monumental help which without this project could not have existed; as well as the subjects who volunteered to get their samples taken for this research.

Ethical considerations (position TBD)

This study requires taking samples from live human subjects. This is a one-off sampling process: they are required only once. The results are then communicated to the subjects via e-mail or by being delivered a physical piece of paper. They are informed previously on the process they are going to go through, as well as the purpose of the experiment. Each subject must read and agree to two documents: an informed consent which explains everything about the experiment¹ and a GDPR notice which documents the use of their data as well as an expected timeline for data anonymisation and destruction². All participants were screened to be over the age of 16, in order to ease the process and require no previous authorisation by parental figures on data collection. The experimentation followed has no effect on the subjects, and they were monitored during the process in order for them not to feel any kind of stress.

Since bacteria were used, some aspects of the experiment must be clarified and discussed. Previously to starting the experiment, I read profusely the WHO's Laboratory Biosafety Manual and Associated Monographs (4th Edition) as to mitigate any possible risk. During the experimentation there were 0 accidents or incidents. All plates were accounted for and controlled closely. No person other than me was allowed to come in contact with a plate that had been cultivated or with the used cotton swabs that were in the process of being disinfected. The cultivated plates were considered Biosecurity Level 2. All possibly infected material was disposed of taking into account the risks that the bacteria in question posed, using bleach.

Before starting the experimentation, I had an interview with my coordinator in order to solidify the fact that there was no alternative to taking cutaneous samples from human beings, as well as a discussion on bacteria and the risks that this experiment implies.

¹Can be found at <https://biblio.peiphy.xyz/TDR-IC.pdf>

²Can be found at <https://biblio.peiphy.xyz/GDPR-notice.pdf>

Abstract

Insert abstract here

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1

Introduction

§1.1 Bacteria and bacterial infections

Bacteria are prokaryote organisms, generally single-celled, which are part of the Monera animal kingdom. Their sizes range from between 30 µm and 100 µm and are ubiquitous¹ organisms. This form of life is believed to be the first one to have ever appeared on Earth, as well as the one responsible for the oxygen-rich atmosphere the Earth currently has. Some species are hard to culture in a laboratory environment, but generally, those that can be cultured in a controlled environment are grown in agar plates[1].

Agar is used as a place to grow bacteria due to the fact that it is indigestible for the majority of bacteria, yet keeps them humid. Together with growth mediums, such as Lysogeny Broth, bacteria thrive in this environment, allowing them to proliferate and create colonies, which can be observed without the need of optic magnifying equipment. Sometimes, together with the growth medium, additives such as mannitol or salt are added. These are used to improve or impede bacterial growth, modify their conditions so they develop differently or as an identification tool. For example, *Staphylococcus Aureus* ferments the mannitol, producing acid, which in turn generates an acid and decolorates the plate's integrated pH indicator from red to yellow, whilst the salt prevents the growth of bacteria that are not of interest to the study[2].

Pathogenic bacteria are bacteria that have the ability to cause disease². These are not the most common type of bacteria, as the majority of them are either harmless or beneficial to the human body through symbiosis, such as the bacteria that help with digestion in the stomach[1].

¹Ubiquitous: found everywhere

²A disease is a particular abnormal condition that negatively affects the structure or function of all or part of an organism, and that is not immediately due to any external injury[3].

§1.2 The enemy: *Staphylococcus aureus*

Staphylococcus aureus (also known as Staph) is a GRAM-positive bacteria, the most virulent and studied of its genus³. Some of its distinctive characteristics include having a very thick glycopeptide wall, which allows it to withstand extreme temperatures and osmotic pressures, therefore rendering most classic methods of food conservation (such as cooking, smoking, freezing or salting⁴) completely useless against said bacteria; a protein A capsid, which binds to many eukaryote organism. It's an extremely resistant (and thus ubiquitous) bacteria. It can be found in human skin and mucous surfaces (such as the mouth or the nose), as well as in certain foods such as ham (cooked or cured), eggs, raw and cooked dough, as well as in poultry.

Staphylococcus Aureus has three main parts to its virulence: its cell wall, its membrane-bound factors and its secreted factors. Staph's **cell wall** is made up of three parts, going from inside to the outside of the cell: a plasma membrane, a peptidoglycan layer and a slime (sometimes also called capsule) layer.

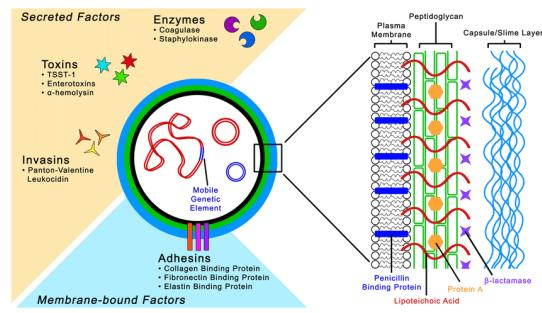


Figure 1.1: Parts of *Staphylococcus aureus*.
Source: PLoS Pathogens

³citation needed, got to check the proper terminology

⁴citation needed

The plasma membrane consists of a lipid bilayer that is semipermeable⁵, which regulates the transport of materials entering and exiting the cell. Integrated inside them are a type of integral protein called penicillin-binding protein (PBP), amongst other proteins such as protein channels. We will only talk about PBPs because they are the Achilles's Heel of bacteria, as long as you know how to exploit it. Whilst the name implies PBPs are only sensible to penicillin, the name actually comes because that's how they were discovered, and in fact could be resistant to it but sensible to other antibiotic agents. Variations in this protein may lead in some cases to antibiotic resistance, such as MRSA (*Methicillin-Resistant Staphylococcus Aureus*), a variation of Staph that is the result of a variation in this protein called PBP2A. The different variations of *Staphylococcus Aureus* will be discussed in more detail in a following section.

Staphylococcus aureus, like all other members of the *Staphylococcus* family, have very thick peptidoglycan layers. This grants them protection from extreme temperatures and high osmotic pressures, which means these bacteria can colonise cooked food and food that has been salted. The most notable example is ham, either cooked, smoked or cured.

§1.3 The enemy's attacks

Staphylococcus Aureus is a species that can cause a handful of different diseases, ranging from, most frequently, skin and respiratory tract infections to infective endocarditis, toxic shock syndrome or osteomyelitis. Several variations of this pathogen exist, with increasing levels of antibiotic resistance: MSSA (*Methicillin-Sensitive Staphylococcus Aureus*), having no resistance; MRSA (*Methicillin-Resistant Staphylococcus Aureus*); and VRSA (*Vancomycin-Resistant Staphylococcus Aureus*), the latter the latter for which no antibiotic concoction that can eradicate the infection is known, and the patients have to use experimental treatments. VISA (*Vancomycin-intermediate Staphylococcus Aureus*) is a variation that has medium resistance to vancomycin, being an intermediate step between MRSA and VRSA. VISA and VRSA are what we would call a superbug,

⁵Semipermeable: it lets water and ions through, but not other molecules. This transport will always be in favour of the pressure gradient, which means that it cannot insert any kind of substance into an environment that has a higher pressure than the other side

a microbe that has developed resistance to more quantity of antibiotic than is safe to consume. Studies have discovered that this genetic factor has been developed by different lineages separately, indicating that there is not a common ancestor of MRSA strains. This case is the bacteria equivalent of carcinisation, the discovery that several species have coevolved into crabs; as well as the bacteria equivalent of tree leaves, which were developed independently by several species at the same time, in completely different parts of the world.

One of Staph's most notorious abilities is using the body's own proteins to disguise itself and thus avoid detection and phagocytosis by the host's immune system. It accomplishes this task by using enzymes called coagulases, which enable the transformation of fibrinogen⁶ to fibrin⁷ [source: microbiology medical]. Only 11 other *Staphylococcus* family members are coagulase-positive. To test for this enzyme in the laboratory there are two main methods which are usually combined: culture of the sample on a Baird-Parker agar medium, a selective and differential medium which contains lithium chloride and tellurite as to inhibit the growth of other microbes. It also includes pyruvate and glycine, which promote the growth of *Staphylococci* colonies, showing in colour black and with an opaque zone around the colony. This opaque zone represents the effect of the coagulase. Another way to test for coagulase is to perform a coagulase test. This test generally requires a small quantity (generally 2 mL) of sheep blood serum, which will gelatinise if coagulase is present.

Staphylococcus Aureus contains an important quantity of **toxins**, which grants it most of its pathogenicity. Many of its virulence factors can be described as such. Toxins are usually defined as poisonous substances, which, in our case, means that they have the capacity to mess with the host body directly, without need of a mediating entity. This category doesn't include, for example, those molecules intended to combat the host's defence mechanisms or scavenge reactive oxygen. We'll also exclude those situated on its membrane for the purpose of cell binding. Staph has several kinds of toxin

⁶A glycoproteic complex produced in the liver and present in the blood of all vertebrates.

⁷fibrinogen after being stimulated by either thrombin or *staphylothrombin*, the result of a molecular pathway stimulated by coagulase. It helps in clotting the blood in the event of vascular or tissue injury

in its arsenal: membrane-damaging toxins (which can be receptor-mediated or not), receptor-interfering toxins (not membrane-damaging), enzymes, and pathway blockers.

- Membrane-damaging toxins. Several of *Staphylococcus aureus*' toxins target the cytoplasmic membrane of the host's cells. These lead to pore formation in it, which provokes the outflux of vital molecules of the cell which, in turn, leads to cytosis⁸.
 - Receptor-mediated. Many of the cytolytic toxins of *Staphylococcus aureus* have been shown to require receptor interaction for their lytic activity. The best-known toxin of this kind is Alpha-toxin, also known as Alpha-hemolysin, which is its major cytotoxic agent, and is lytic to red blood cells and certain leukocytes, but not to neutrophils. Whilst at low concentrations it has been shown to be dependent on the interaction with cells' ADAM10 receptors, in higher concentrations of this toxin, this interaction is no longer necessary. Other toxins of this type include PVL (Panton-Valentine Leucocidin) and Gamma-toxin.
 - Non-receptor-mediated. In 2007, a toxin family that includes the Delta-toxin called the Phenol-Soluble Modulins (PSMs). PSMs trigger an inflammatory response by interacting with the FPR2 receptor, however they can carry cytolytic activity independently from FPR2 interaction. Delta-toxin has been linked to allergic skin sease and atopic dermatitis by degrading mast cells. This kind of toxin contributes to neutrophil lysis after phagocytosis, which might partly explain why the development of *Staphylococcus Aureus* vaccines that work by enhancing a type of phagocytosis have failed so far.
- Receptor-function-interfering toxins. The toxins that fall into this category are enterotoxins⁹. These typically cause vomit and diarrhoea. *S. aureus* strains can produce a wide array (around 20) of entero and entero-like toxins. The most famous *Aureus* superantigen¹⁰, the 22-kD toxic shock syndrome toxin (TSST), belongs to this group. TSS is a very severe and potentially fatal disease. *Staphylococcus aureus* also secretes a series of proteins that interfere with leukocyte receptors to evade recognition and

⁸Cell bursting due to osmotic pressure imbalance between the inside and the outside of it

⁹Enterotoxins are those toxins that target the intestines.

¹⁰Superantigen: type of antigens that results in excessive activation by the immune system

thus activation of the immune system. CHIPS (Chemotaxis Inhibitory Protein of *Staphylococcus aureus*), which binds to the C5aR and FPR receptors, impairing the recognition of bacterial formylated peptides by the FPR receiver and blocking the activation of leukocytes via C5aR. *S. aureus* also has other proteins that work similarly to these, such as FLIPr.

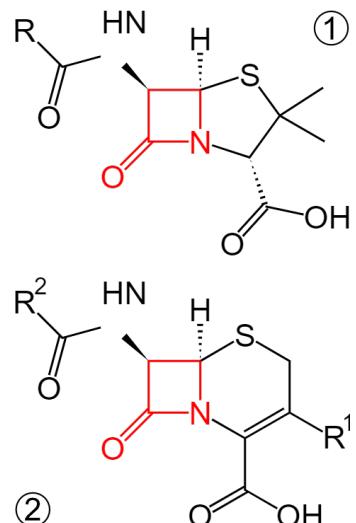
- Enzymes. Many enzymes secreted by *Staphylococcus Aureus* either degrade host molecules or interfere with its metabolic or signalling cascades. A few of them are proteases, which some non-specific ones have the ability to degrade host proteins in a broad proteins, leading to tissue destruction and necrosis, but may also have some more specific effects, for example the destruction of insulin B. Its two coagulases (staphylocoagulase and Willebrand factor) fall into this category.

§1.4 Our weapons

The tools we have at our disposal to fight off this infection fall into two main categories: chemical factors and biological factors.

The chemical factors are drugs, and they depend both in quantity and type on the variation a particular case falls in. It is **extremely important** to find out the level of antibiotic resistance that specific infection has before administering any antibiotic, as this treatment course will cause side effects such as killing gut bacteria, diminishing defense system capabilities, and increasing the possibility to develop yet more resistant infections. Generally, a large-spectrum antibiotic has an adequate risk-to-benefits ratio of causing the latter, so they may be used before switching to a more specific treatment.

Starting with the treatment to the least resistant strains of *Staphylococcus Aureus*, a -lactam antibiotic (such as methicillin, oxacillin, cloxacillin and penicillin) is the weapon of choice to fight against an MSSA infection. This is because this specific chemical part (just a -lactam ring does nothing by itself) has the ability to inhibit cell wall biosynthesis on the bacterial intruder's body. But once the -lactam ring is cut by an enzyme secreted by the bacteria itself, this type of antibiotic suddenly loses effect against them.



That's where Vancomycin comes in. It is a type of glycopeptide antibiotic, just like -lactam and works by blocking the construction of a cell wall, as all of its type do. This treatment is extremely invasive and only indicated for the treatment of extremely serious life-threatening infections by Gram-positive bacteria that have shown to be unresponsive to other antibiotics.

Figure 1.2: Organic chemistry structure of penicillin (top) and cephalexin (bottom). -lactam ring in red.
Source: WikiMedia

It can be taken as a pill or as an injectable fluid, which proves it to be much more effective. This treatment is incompatible with aminoglycosides, a type of antibiotic that inhibit protein synthesis, as it can lead to nephrotoxicity¹¹ and ototoxicity¹². Vancomycin can induce platelet-reactive antibodies in the patient, leading to severe thrombocytopenia and bleeding with petechial hemorrhages on the tongue and bruises. Unfortunately, even with use of Vancomycin, *Staphylococcus Aureus* can develop resistance. In this case, no other option than using a biological factor is left. There has been one study in 2020 that discovered that by modifying the bacteria with a cationic oligopeptide¹³, vancomycin resistance could be bypassed. This could be a good solution temporarily as we wait for phage therapy to get approved, if it was a sufficiently studied option,

¹¹Nephrotoxicity: damage to the kidneys

¹²Ototoxicity: damage to hearing

¹³Cationic oligopeptide: sequence of two or more aminoacids that is positively charged

which is not. This was discovered in 2020, while bacteriophage trials have been ongoing since the mid 2000s.

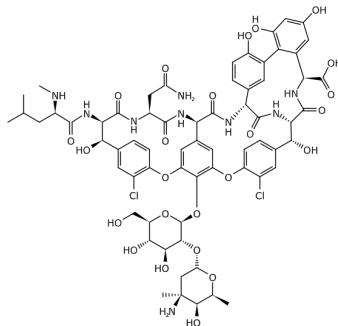


Figure 1.3: Organic chemistry structure of vancomycin. Source: WikiMedia

The biological factor is a bacteriophage, called P68. It comes from the *Caudovirales* order, which means that it is a bacteriophage with tail. This treatment is still in testing, but it appears to be effective and lead to low adverse results. If possible, it would be preferable to use bacteriophage therapy (shortened to phage therapy) instead of going for antibiotics, as can lead to less side effects than antibiotics, as it only attacks a specific bacteria. This means, unfortunately, that the infection has to be pinpointed with extreme accuracy. The use of this treatment also negates the risk of bacteria developing antibiotic resistance. It is, however, unclear whether the bacteriophage could mutate into a dangerous strain. This class of virii has been studied since the late 19th century after being discovered

by accident in water from a river in India. This research, however, was dropped due to penicillin being discovered and put into effect, as the main purpose of this research was to use it on war casualties as to reduce mortality due to infections.

Bacteriophages work in an interesting manner. They work by detecting one very specific bacteria, just like any other virus does with the type of cell they evolved for, then bind to it and inject their genetic material, which then in turn the bacteria considers as its own, inserts it into its own genetic sequence and starts producing the proteins the virus requires, but it doesn't eject them. Once the bacteria is full with phages, a special

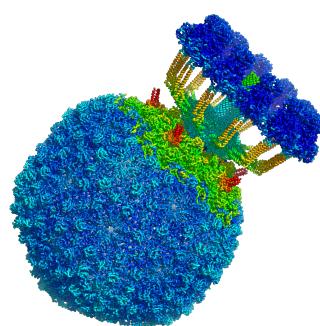


Figure 1.4: P68 structure.
Source: Own material

lytic compound is released which bursts the cell membrane in such a way that it resembles an explosion, but instead of heating up everything in a radius, spreads millions more of bacteriophages, which then bind to other bacteria and the cycle repeats until there's no more bacteria left. The fight from the bacteria point of view consists mostly on trying to outnumber and outreproduce the phages as to have a chance of survival, even if minimal. There is no known bacteria that shows resistance to phages. That is probably because unlike the chemical factors, phages can evolve and improve with each generation.

2 Physical experimentation

§2.1 Description

This experiment looks at finding out the prevalence of *Staphylococcus Aureus* in a sample of students from the school. The process used involves extracting a sample from underneath a subject's nalis by swabbing, cultivating that sample and then observing the results of said culture to determine the presence or not of *Staphylococcus Aureus* as part of the subject's resident bacterial flora. Each iteration of the process took less than two minutes to complete. However, all the safety measures and actions taken need more time to be taken care of properly.

§2.2 Protocol followed

The protocol followed was designed based on a similar protocol used in the many university laboratories^[4], modified to fit the needs of this research paper, and peer-reviewed by Olga Sánchez, and uploaded to the Protocols.io platform, to make it easier to follow the days of that the experiment took place in. This protocol underwent 9 different revisions^[5]. It can also be read below:

- 1) Prepare yourself for the experimentation: wash your hands, put on gloves, put on the lab coat, mask and goggles. Wash your hands again (gloves still on). Set up the work area; the Bunsen burner should be turned on in such a way that it can cover an acceptable surface to work. Turn it on and wash your hands again.
- 2) Divide each Petri dish between 2 parts. A guide should be used for this part. Get your subject to wash their hands. Observe their hands. If they are extremely short, it may be worth it to take the sample nasally.

Qty	Material/consumable	Qty	Reagent
x80	Sterile cotton swabs	~30mL	Bleach
x1	Kolle handle	~10mL	Methyl violet
x1	Optic microscope	~10mL	Iodine
x1	Binocular magnifier	~10mL	Alcoholic safranin
x1	Dissection tray	~10mL	Methanol
x1	Bunsen burner	<1mL	Ether
x1	Lab coat	x40	Agar MSA plates
x1	Lab goggles	>1L	Ringer solution (9% saline)
x8	Non-powdered gloves		
x10	Slides and slide covers		

- 3) Note down their information, crack open a sterile swab, dip it in Ringer solution and swab away at under their nails or nose. Then, populate the dish with this sample.
- 4) Incubate for 32-48h and then observe the results.
- 5) Observe the bacteria under a microscope after GRAM tinction.

§2.3 Bill of materials

The materials used, as well as the quantities used can be found in the following table. On the left, laboratory equipment and, on the right, regents and consumables used:

§2.4 Risk assessment and prevention

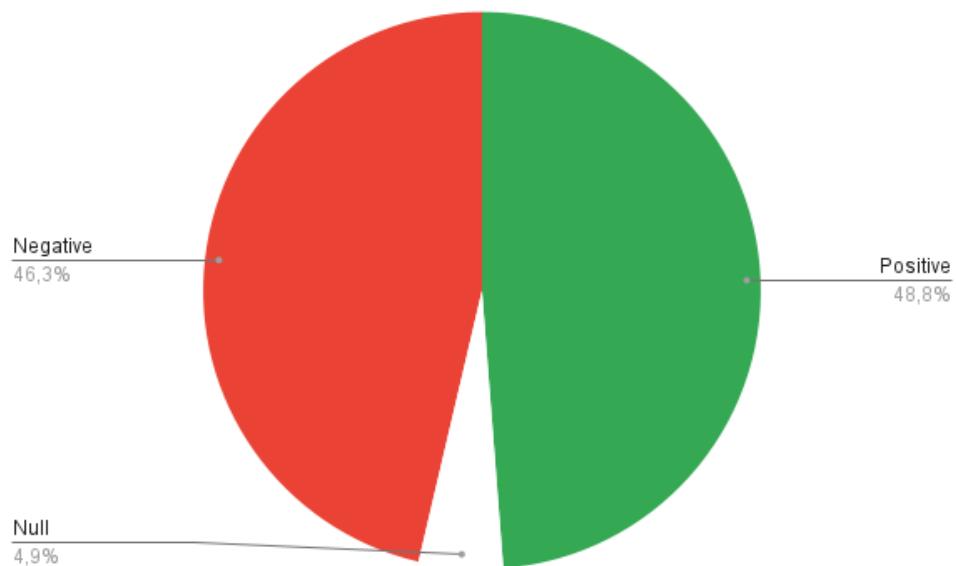
Staph is considered a Biosecurity Level (BSL) 2 pathogenic bacteria. This means that it is associated with a human disease that can pose a moderate human health hazard.

In a laboratory where BSL-2 pathogens are handled, regular lab rules should be followed (mechanical pipetting only, hand washing, prohibiting the consumption of food and drinks in the lab, proper PPE use...), as well as avoiding splashes or aerosols, biohazard warning signs present on all material used, as well as proper surface and material disinfection via the use of autoclave or alternative decontamination method.[source: book] The risks associated with this bacteria were assessed following the protocol designated by the World Health Organisation [cite], and proper security measures were followed at all times when handling biohazardous material. No incidents occurred during the research part of this project, and the protocol defined previous to the start was followed to a T. While the laboratory used may not be the most ideal type of laboratory for this type of research, it is certainly adequate enough to perform a research project like this one

§2.5 Results and analysis

The results obtained can be found as Annex II in the form of a raw data table with personal identificative data removed following GDPR regulations. The data was then recounted and graphed into the following pie chart:

. As we can see, almost 50% of the samples taken tested positive for Staphylococcus Aureus, compared to the expected 30%[[StaphylococcusAureusHealthcare2020](#)]. We can, however, see in the UK's Public Health bactaeremia data that Staph infections have been on the rise lately, so it may not be a case of wrong data[[englandMSSABacteraemiaAnnual2021](#)]. On top of that, my advisers, both Olga and Margarita, have also found themselves getting more prevalence than usual of this bacteria, and are finding cases that were once negative but turned positive in the last few years. There may be several reasons for the infection rate and thus natural prevalence to be increasing. One of them could be that since antibiotic abuse is growing with each passing year, the typical resident microbiota is getting killed, leaving more resources for Staph to thrive in that environment. The other could be climate change. An increase of ambient temperatures could mean a better breeding ground for this specific species and thus leading to a higher-than-usual prevalence.



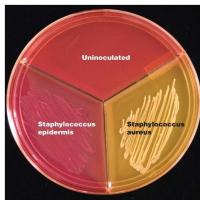
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Appendix

Annex 1 - Protocol as published in protocols.io



Staphylococcus Aureus Sampling V.10

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Olga Sánchez: Peer review;

Version 10 ▾

Sep 19, 2022

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ABSTRACT

This protocol is intended to study the affection of *Staphylococcus Aureus*, including the MRSA, VISA and VRSA variants, even if it makes the test more difficult to perform. It outlines the basic protocol for a multi-subject study, while using basic and minimal resources found in almost every biology lab.

DOI

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PROTOCOL CITATION

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Version created by [Mar Roca Cugat](#)

KEYWORDS

Microbiology, sampling, swab, staphilococcus aureus

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CREATED

Jan 12, 2022

LAST MODIFIED

Sep 19, 2022

PROTOCOL INTEGER ID

70244

GUIDELINES

This protocol is intended to study the affection of *Staphylococcus Aureus*, which is a Biosecurity Level 2 bacterial agent. As such, the laboratory should be adequately to those standards or take measures in order to prevent infection, cross-contamination, or leaks.

MATERIALS TEXT

PPE

- Face shield or protective goggles
- FFP2/KN95 or higher-rated mask
- Latex, non-powdered gloves
- Lab coat

Sampling material

- Clean and sterile cotton swabs ($n+n/10$ being n the number of tests required)
- MSA Agar Petri dishes ($n/(2 \text{ to } 4)$ being n the number of dishes required)
- Sterile Ringer solution
- Permanent marker

Support material

- Bunsen burner
- Incubator
- Ethanol: >80%
- Bleach solution at 50%v in water.
- Reagents to make LB
- Inactivation buffer
- Photospectrometer

SAFETY WARNINGS

This protocol requires interaction with people, possibly infected with a pathogen (especially in 2022, when this protocol was written and put into practice, as COVID-19 was still going strong). As such, there is a risk of infection which can be minimized with proper PPE use.

Proper ventilation is recommended at all times, even when the pandemic situation is over. However, the sterile field must be preserved at all costs, so try to direct the airflow in order for it not to affect the results.

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Preparation

1h

10m



Wash your hands with soap. Put on your lab coat, your mask, and your goggles or face shield. Make sure your mask is airtight and air cannot escape through the sides.

- 1.1 Prepare the area where you are going to work. Disinfect the surfaces with the bleach solution.

The subjects should not be able to walk behind you, only to the side or to the front. Make sure to leave enough distance between the sampler and the subject, but not enough distance as for the sampling to be uncomfortable. The environment should be comfortable, within the following range of temperatures: **20 °C – 35 °C**

You should have a plastic, sealable box to your side or on the table to store the sampled Petri dishes.

The Bunsen burner should be to the front of you, within a hand of distance. The fresh swabs and Petri dishes should never be accessible by the subjects.

- 1.2 Using a permanent marker, divide each plate into two to four equal parts. You should help yourself by using a guide, such as a ruler.

- 2 Observe the subject's hands. If their nails are longer than 1-4 mm (the white part of the nail that can overgrow).
Bitten-down nails could lead to invalid results. Too long nails could lead to cross-contamination.
Ask the subject for their identificative and contact information if this has not been done previously.
- 3 Place a Petri dish on the side of the Bunsen burner. The burner will be the center of our sterile field.



Watch out so as not to break the sterile field

Step 3 includes a Step case.

Nail sampling

Nose sampling

step case

Nail sampling

For cases where the nails are 3-6mm long

- 4 Ask the subject to wash their hands thoroughly and below the nails with soap and lukewarm water. Note their subject ID on the bottom (agar side) of the Petri dish.
 - 4.1 Bring the subject's hands below the sterile area generated by the Bunsen burner. Open the Ringer solution and soak the swab. Proceed by swabbing below every nail in both hands. Once done, the subject can be dismissed.
 - 4.2 "Paint" one half of the Petri dish with the swab, softly so as not to break the agar but firmly as to get the sample to transfer to the plate.

5



1d

⌚ Repeat n times

Once the plate has 3 samples, place it in the full plates box.

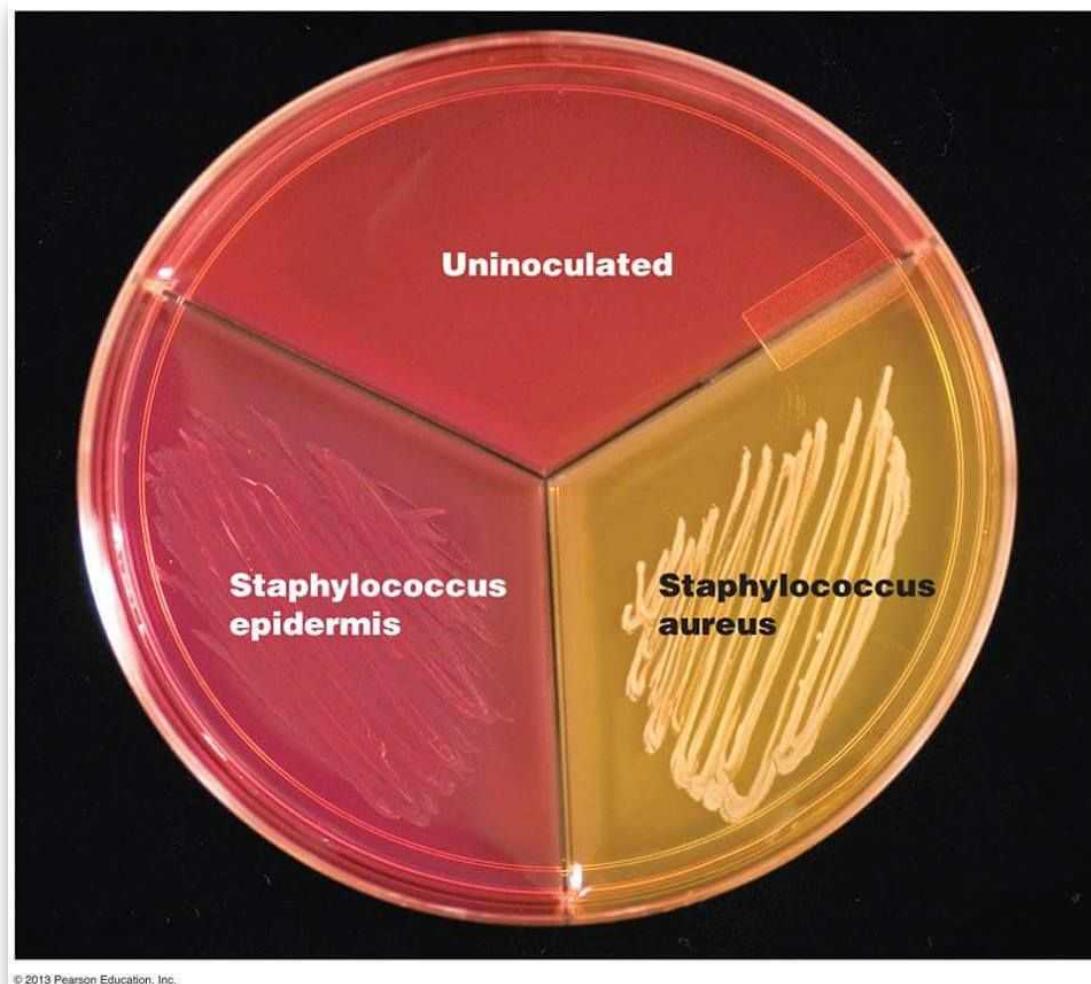
Once there are 20 plates in the box, group them together with tape, write an identificative group number and place it in the incubator.

The incubator should be set to **⌚ 37 °C** and left to incubate for **⌚ 24:00:00**.

Study

1d 2h

6



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It is expected to one of these results. A bright red colour means the sample was uninoculated. A pinkish colour with translucent streaks means there is *Staphylococcus epidermidis* present. A faint yellow colour or a bright yellow colour means there is *Staphylococcus aureus* present

The samples should be taken out of the incubator, the results introduced into the database and communicated to the subject.

7



Using one of the extra MSA Agar plates, culture and purify a sample of *Staphilococcus Aureus*. This can then be treated with GRAM tinture in order to observe it under an optical microscope (ideally at x1000-x1200).



Watch out for impurities/contaminations

Annex 2 - Raw data table (data anonimised)

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