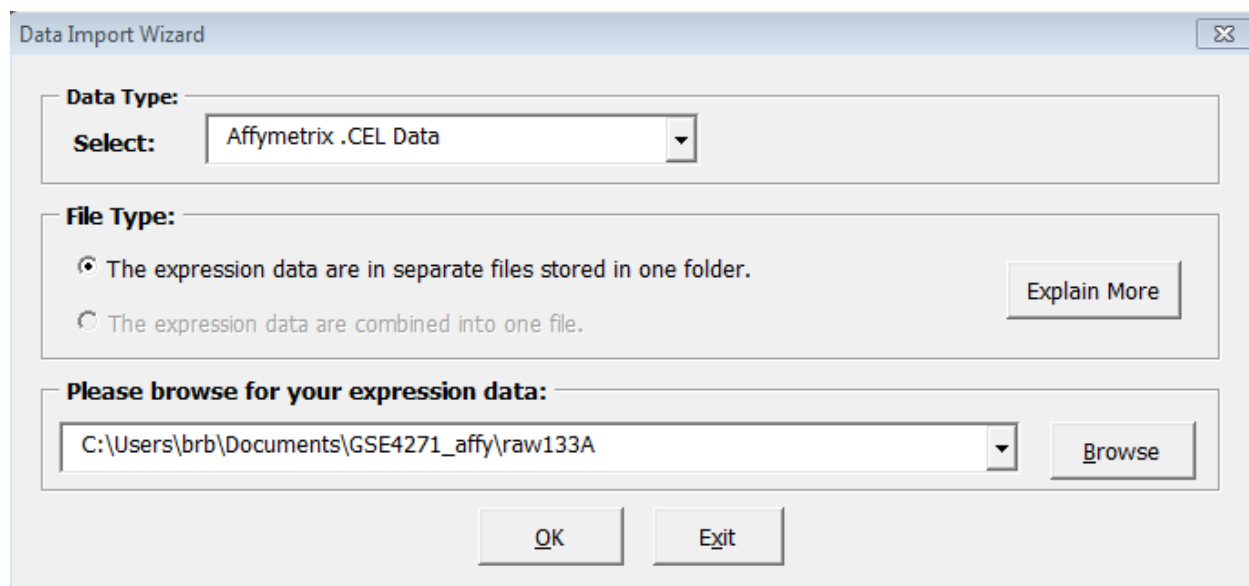


## Affy Cel file importer

[Case 1: Use standard affymetrix probe set id](#)

[Case 2: Use Affymetrix and import own identifier file](#)

[Case 3: Use custom cdf](#)



The 'Data Import Wizard' dialog box is shown. It has a title bar with a close button. The main area contains three sections. The first section, 'Data Type:', has a 'Select:' label and a dropdown menu currently showing 'Affymetrix .CEL Data'. The second section, 'File Type:', has two radio buttons. The first is selected and labeled 'The expression data are in separate files stored in one folder.', with an 'Explain More' button to its right. The second is labeled 'The expression data are combined into one file.'. The third section, 'Please browse for your expression data:', has a text box containing the path 'C:\Users\brb\Documents\GSE4271\_affy\raw133A' and a 'Browse' button to its right. At the bottom are 'OK' and 'Exit' buttons.

**Data Import Wizard**

**Data Type:**

**Select:** Affymetrix .CEL Data

**File Type:**

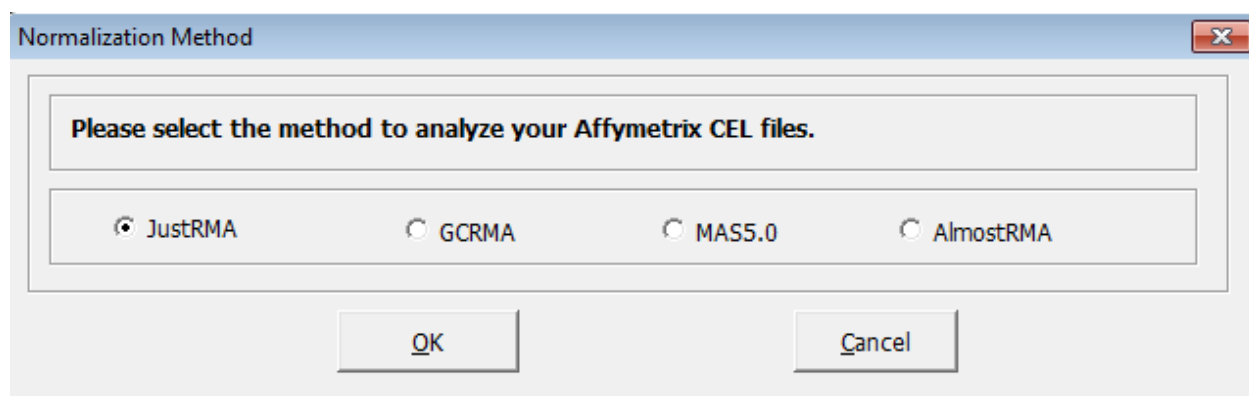
☒ The expression data are in separate files stored in one folder. [Explain More](#)

☐ The expression data are combined into one file.

**Please browse for your expression data:**

C:\Users\brb\Documents\GSE4271\_affy\raw133A [Browse](#)

[OK](#) [Exit](#)



The 'Normalization Method' dialog box is shown. It has a title bar with a close button. The main area contains a text box with the instruction 'Please select the method to analyze your Affymetrix CEL files.' Below this are four radio buttons: 'JustRMA' (selected), 'GCRMA', 'MAS5.0', and 'AlmostRMA'. At the bottom are 'OK' and 'Cancel' buttons.

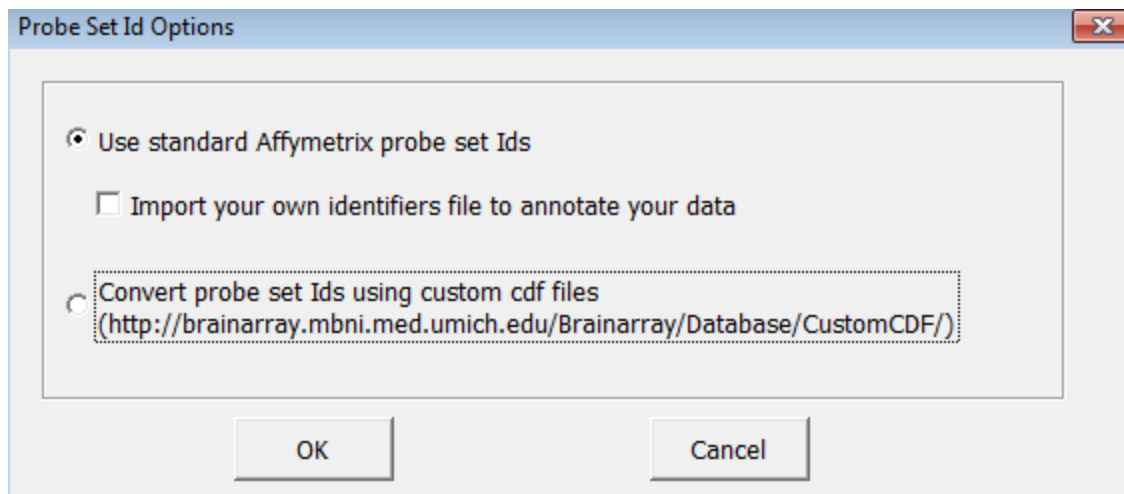
**Normalization Method**

Please select the method to analyze your Affymetrix CEL files.

☒ JustRMA ☐ GCRMA ☐ MAS5.0 ☐ AlmostRMA

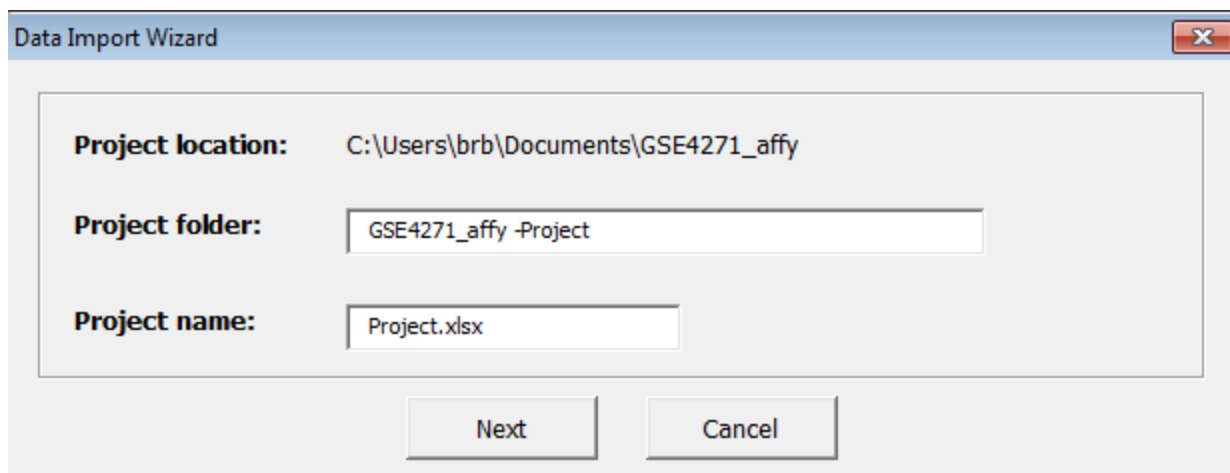
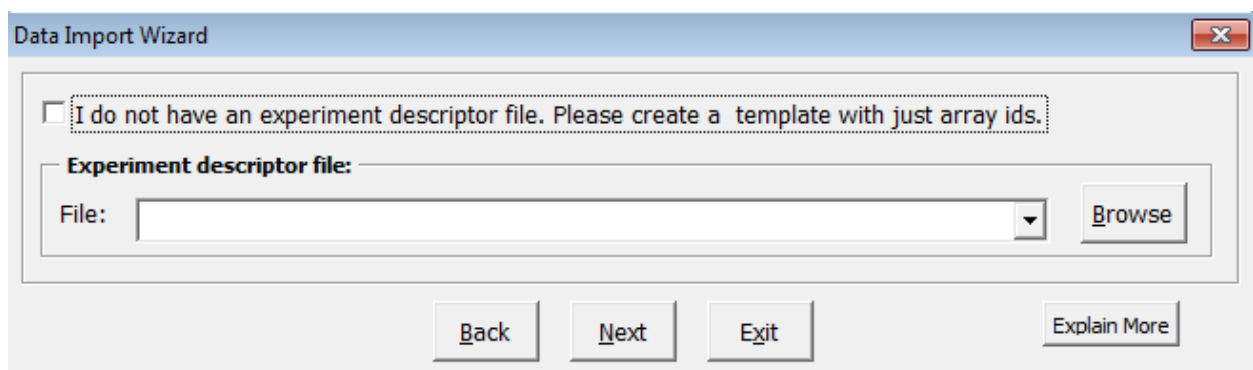
[OK](#) [Cancel](#)

## Case 1: Use standard affymetrix probe set id



It will install hgu133acdf (only one) package.

For gcrma option, it will download gcrma package.



Refilter, normalize and subset the data

1. Spot filters   2. Normalization   3. Gene filters

Background adjustment is performed before the intensity filtering and the averaging of replicate spots is done on filtered data.

☐ Apply background adjustment.

☐ Intensity Filter:
 

☐ EXCLUDE the spot if the intensity is below the minimum.

☒ THRESHOLD the intensity at the minimum value if the intensity is below the minimum.

Intensity minimum:

☐ Detection Call
 

EXCLUDE the probeset if the Detection Call contains:

Any one of the following values (comma-separated): A,M,P,No Cal

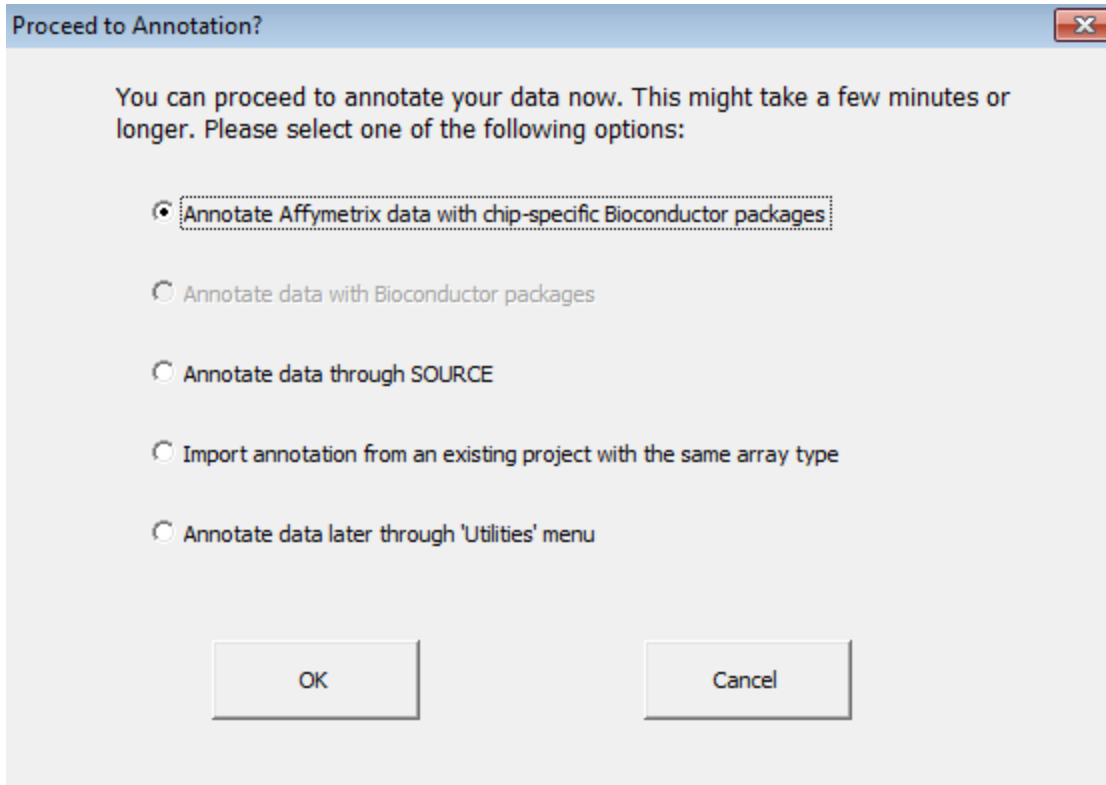
☐ Average the replicate spots within an array.

☐ Use a common reference design.

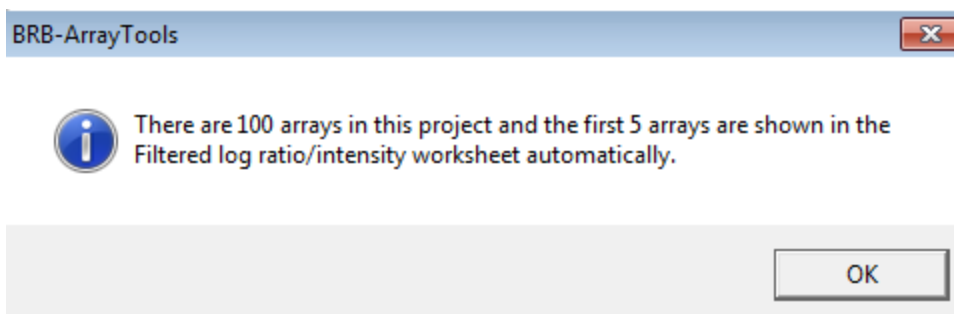
Note

There are 7200 genes which pass the filtering and subsetting criteria.

Matching genelists



It will install hgu133a.db package.



Project - Microsoft Excel

	A	B	C	D	E	F	G	H	I	J	K
	ProbeSet (Double-click)	Name	Accessi	UGClus	Symbol	Entrezl	Chromo	Cytoba	GO	Filter	
1											
2	1007 s at		U48705						Biological	TRUE	
3	1053 at	replication	M87338	Hs.647062	RFC2	5982	7	7q11.23	Biological	TRUE	
12	1438 at	EPH recep	X75208	Hs.2913	EPHB3	2049	3	3q27.1	Biological	TRUE	
19	1773 at		L00635						Molecular	TRUE	
21	1861 at	BCL2-asso	U66879	Hs.370254	BAD	572	11	11q13.1	Biological	TRUE	
22	200000 s	pre-mRNA	NM_00644	Hs.181368	PRPF8	10594	17	17p13.3	Biological	TRUE	
23	200001 at	calpain, sr	NM_00174	Hs.515371	CAPNS1	826	19	19q13.12	Molecular	TRUE	
24	200008 s	GDP disc	D13988	Hs.299055	GDI2	2665	10	10p15	Molecular	TRUE	

## Case 2: Use Affymetrix and import own identifier file

Probe Set Id Options

☐ Use standard Affymetrix probe set Ids  
☒ Import your own identifiers file to annotate your data  
☐ Convert probe set Ids using custom cdf files  
[\(http://brainarray.mbni.med.umich.edu/Brainarray/Database/CustomCDF/\)](http://brainarray.mbni.med.umich.edu/Brainarray/Database/CustomCDF/)

OK Cancel

**Data Import Wizard**

**Please specify the location of your gene identifiers:**

☐ The identifiers are stored alongside the expression data. ☒ The identifiers are stored in a separate file.

**Please select your Gene identifiers file:**

File:   Header line #:

**Please select the available gene identifiers:**

Unique ID (Well or Spot ID, etc.):	<input type="text" value="Col 1: ProbeSet"/>	EntrezId:	<input type="text" value="Col 4: EntrezID"/>
Clone ID (IMAGE or ATCC ID, etc.):	<input type="text"/>	Gene Name ,Title or Description:	<input type="text" value="Col 3: Name"/>
UniGene Cluster ID:	<input type="text" value="Col 6: UGCluster"/>	GenBank Accession:	<input type="text" value="Col 5: Accession"/>
Gene Symbol:	<input type="text" value="Col 2: Symbol"/>	Map Location:	<input type="text"/>
microRNA ID:	<input type="text"/>		

☒ **Annotate the project with these gene ids, instead of using the data from SOURCE database.** **Organisms:**

**Data Import Wizard**

☒ I do not have an experiment descriptor file. Please create a template with just array ids.

**Experiment descriptor file:**

File:

**Data Import Wizard**

**Project location:** C:\Users\brb\Documents\GSE4271\_affy

**Project folder:**

**Project name:**

Refilter, normalize and subset the data

1. Spot filters 2. Normalization 3. Gene filters

Background adjustment is performed before the intensity filtering and the averaging of replicate spots is done on filtered data.

☐ Apply background adjustment.

☐ Intensity Filter:

- ☐ EXCLUDE the spot if the intensity is below the minimum.
- ☒ THRESHOLD the intensity at the minimum value if the intensity is below the minimum.

Intensity minimum:

☐ Detection Call

- EXCLUDE the probeset if the Detection Call contains:

Any one of the following values (comma-separated): A,M,P,No Cal

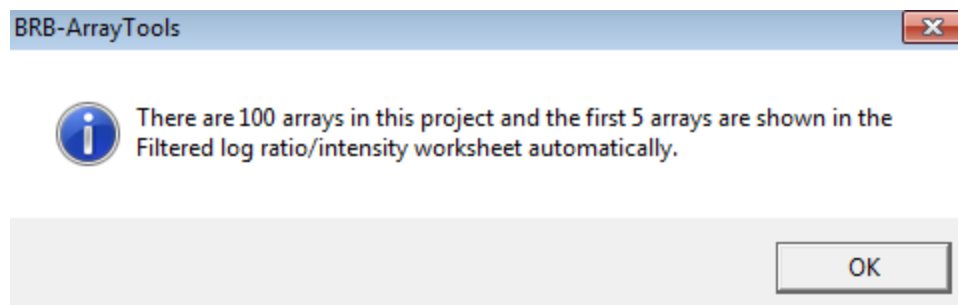
☐ Average the replicate spots within an array.

☐ Use a common reference design.

Note

There are 7200 genes which pass the filtering and subsetting criteria.

Matching genelist



Project - Microsoft Excel

Home Insert Page Layout Formulas Data Review View Add-Ins Team

ArrayTools  
CGHTools

Menu Commands

A1 fx ProbeSet

	A	B	C	D	E	F	G	H	I
	ProbeSet	Name	Accession	UGCluster	Symbol	EntrezID	Defined_Genelists	Filter	
1	1007_s_at		U48705					TRUE	
2	1053_at	replication factor C (activator)	M87338	Hs.647062	RFC2	5982	DNA replication, Mismatch re	TRUE	
12	1438_at	EPH receptor B3	X75208	Hs.2913	EPHB3	2049	Axon guidance	TRUE	
19	1773_at		L00635					TRUE	
21	1861_at	BCL2-associated agonist of ce	U66879	Hs.370254	BAD	572	AKT Signaling Pathway, Apop	TRUE	
22	200000_s_at	pre-mRNA processing factor	NM_00644	Hs.181368	PRPF8	10594	Spliceosome	TRUE	
23	200001_at	calpain, small subunit 1	NM_00174	Hs.515371	CAPNS1	826	Deregulation of CDK5 in Alzh	TRUE	
30	200008_s_at	GDP dissociation inhibitor 2	D13988	Hs.299055	GDI2	2665		TRUE	
33	200011_s_at	ADP-ribosylation factor 3	NM_00165	Hs.119177	ARF3	377		TRUE	
42	200020_at	TAR DNA binding protein	NM_00737	Hs.300624	TARDBP	23435		TRUE	
44	200022_at	ribosomal protein L18	NM_00097	Hs.515517	RPL18	6141	Ribosome	TRUE	
46	200024_at	ribosomal protein S5	NM_00100	Hs.378103	RPS5	6193	Ribosome	TRUE	
58	200036_s_at	ribosomal protein L10a	NM_00710	Hs.148340	RPL10A	4736	Ribosome	TRUE	
59	200037_s_at	chromobox homolog 3	NM_01658	Hs.381189	CBX3	11335		TRUE	
69	200047_s_at	YY1 transcription factor	NM_00340	Hs.388927	YY1	7528	The PRC2 Complex Sets Long	TRUE	
72	200050_at	zinc finger protein 146	NM_00714	Hs.643436	ZNF146	7705		TRUE	
73	200051_at	squamous cell carcinoma ant	NM_00514	Hs.502883	SART1	9092	Spliceosome	TRUE	

### Case 3: Use custom cdf

Select custom cdf file

Select chip name:

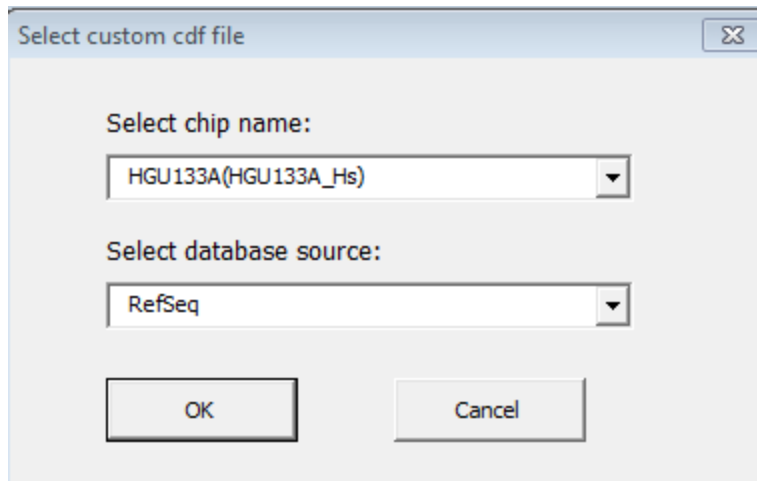
HGU133A(HGU133A\_Hs)

Select database source:

RefSeq

OK Cancel

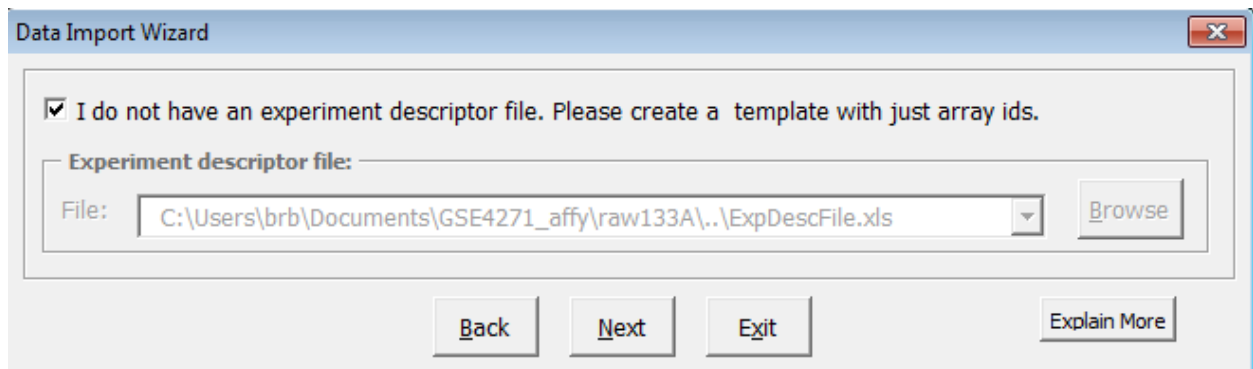
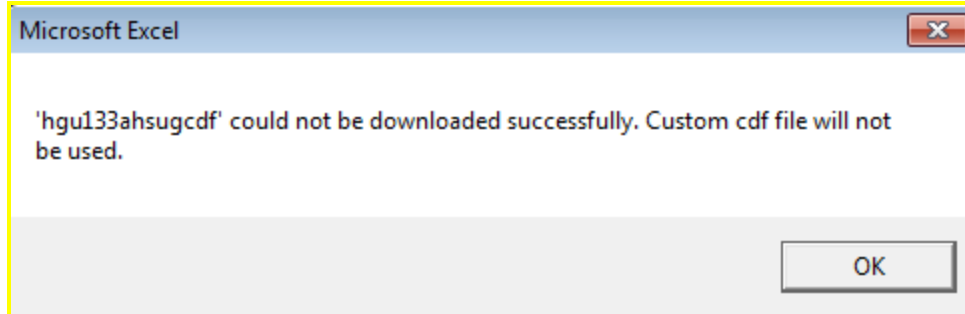




database source: EntrezGene, RefSeq, UniGene

It will download hgu133ahsentrezgprobe, hgu133ahsentrezg.db, hgu133ahsentrezgcdf together if we choose RefSeq.

it will download hgu133ahsugcdf if we choose UniGene



**Data Import Wizard**

**Project location:** C:\Users\brb\Documents\GSE4271\_affy

**Project folder:** GSE4271\_affy -Project

**Project name:** Project.xlsx

Next Cancel

**Refilter, normalize and subset the data**

1. Spot filters 2. Normalization 3. Gene filters

Background adjustment is performed before the intensity filtering and the averaging of replicate spots is done on filtered data.

☐ Apply background adjustment.

☐ **Intensity Filter:**

☐ EXCLUDE the spot if the intensity is below the minimum.

☒ THRESHOLD the intensity at the minimum value if the intensity is below the minimum.

Intensity minimum: 10

☐ **Detection Call**

EXCLUDE the probeset if the Detection Call contains:

Any one of the following values (comma-separated): A,M,P,No Cal

☐ Average the replicate spots within an array.

☐ Use a common reference design. Change current setting

OK Cancel Reset Help

