

EDDA - Assignment 1 - Group 77

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Exercise 1

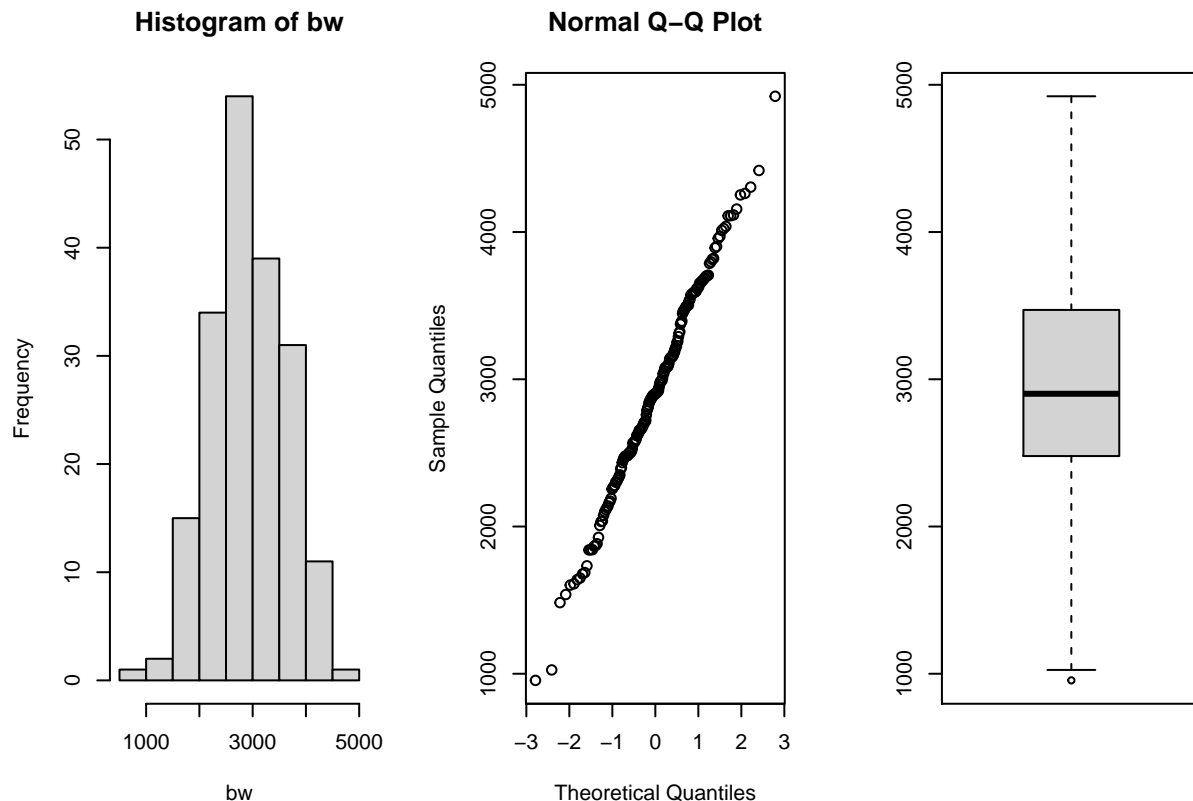
The data set birthweight.txt contains the birthweights of 188 newborn babies. We are interested in finding the underlying (population) mean μ of birthweights.

a) Check normality of the data. Compute a point estimate for μ . Derive, assuming normality (irrespective of your conclusion about normality of the data), a bounded 90% confidence interval for μ .

To check normality for the data we use a qqplot, histogram, box plot and shapiro-wilks test.

```
par(mfrow=c(1,3))
data=read.table(file="data/birthweight.txt",header=TRUE)

bw = data$birthweight
hist(bw)
qqnorm(bw)
boxplot(bw)
```



```
shapiro.test(bw)
```

```
##  
## Shapiro-Wilk normality test  
##  
## data:  bw  
## W = 0.99595, p-value = 0.8995
```

The graphical methods show that the data is normal. The Shapiro-Wilk test reinforces this assumption as it shows a p-value of 0.8995, meaning that the H_0 is not rejected and therefore the data is normal. Furthermore, a point estimate for μ is conducted along side a 90% confidence interval.

```
m = mean(bw)  
sd = sd(bw)  
n = length(bw)  
error = qnorm(0.95)*sd/sqrt(n)  
ci = c(m-error, m+error)  
m
```

```
## [1] 2913.293
```

```
ci
```

```
## [1] 2829.618 2996.967
```

b) An expert claims that the mean birthweight is bigger than 2800, verify this claim by using a t-test. What is the outcome of the test if you take $\alpha = 0.1$? And other values of α ?

A t-test is performed to verify the claim that the mean birthweight is bigger than 2800. The t-test shows a p-value of 0.014. This means that this claim is significant for an α of 0.1. The claim is significant for all α 's above 0.014 and insignificant for α 's below 0.014.

```
t.test(bw, mu=2800, alternative = "greater", conf.level = 0.95)
```

```
##  
## One Sample t-test  
##  
## data:  bw  
## t = 2.2271, df = 187, p-value = 0.01357  
## alternative hypothesis: true mean is greater than 2800  
## 95 percent confidence interval:  
##  2829.202      Inf  
## sample estimates:  
## mean of x  
##  2913.293
```

c) In the R-output of the test from b), also a confidence interval is given, but why is it different from the confidence interval found in a) and why is it one-sided?

The confidence interval is different because the one-sample t-test returns a 95% confidence interval while a 90% confidence interval is conducted in 1b). The confidence interval is one-sided because the critical area of the weight distribution is compared to a mean where it is greater than 2800, but not both greater and less than 2800.

Exercise 2

We study the power function of the two-sample t-test (see Section 1.9 of Assignment 0). For $n=m=30$, $\mu=180$, $\nu=175$ and $sd=5$, generate 1000 samples $x=rnorm(n,\mu,sd)$ and $y=rnorm(m,\nu,sd)$, and record the 1000 p-values for testing $H_0: \mu=\nu$. You can evaluate the power (at point $\nu=175$) of this t-test as fraction of p-values that are smaller than 0.05.

a) Set $n=m=30$, $\mu=180$ and $sd=5$. Calculate now the power of the t-test for every value of ν in the grid `seq(175,185,by=0.25)`. Plot the power as a function of ν .

```
n <- m <- 30
mu <- 180
nu <- 175
sd <- 5
grid <- seq(175,185, by=0.25)

power_function<-function(grid,n,m,mu,sd) {
  B <- 1000
  p <- numeric(B)
  G <- length(grid)
  fractions <- numeric(G)
  for (grid_nu in 1:G){
    p <- numeric(B)
    for (b in 1:B){
      x <- rnorm(n,mu,sd)
      y <- rnorm(m,grid[grid_nu],sd)
      p[b] <- t.test(x,y, var.equal = TRUE)[[3]]
    }
    fractions[grid_nu] <- mean(p<0.05)
  }
  return(fractions)
}

fractions_A <- power_function(grid,n,m,mu,sd)
```

b) Set $n=m=100$, $\mu=180$ and $sd=5$. Repeat the preceding exercise. Add the plot to the preceding plot.

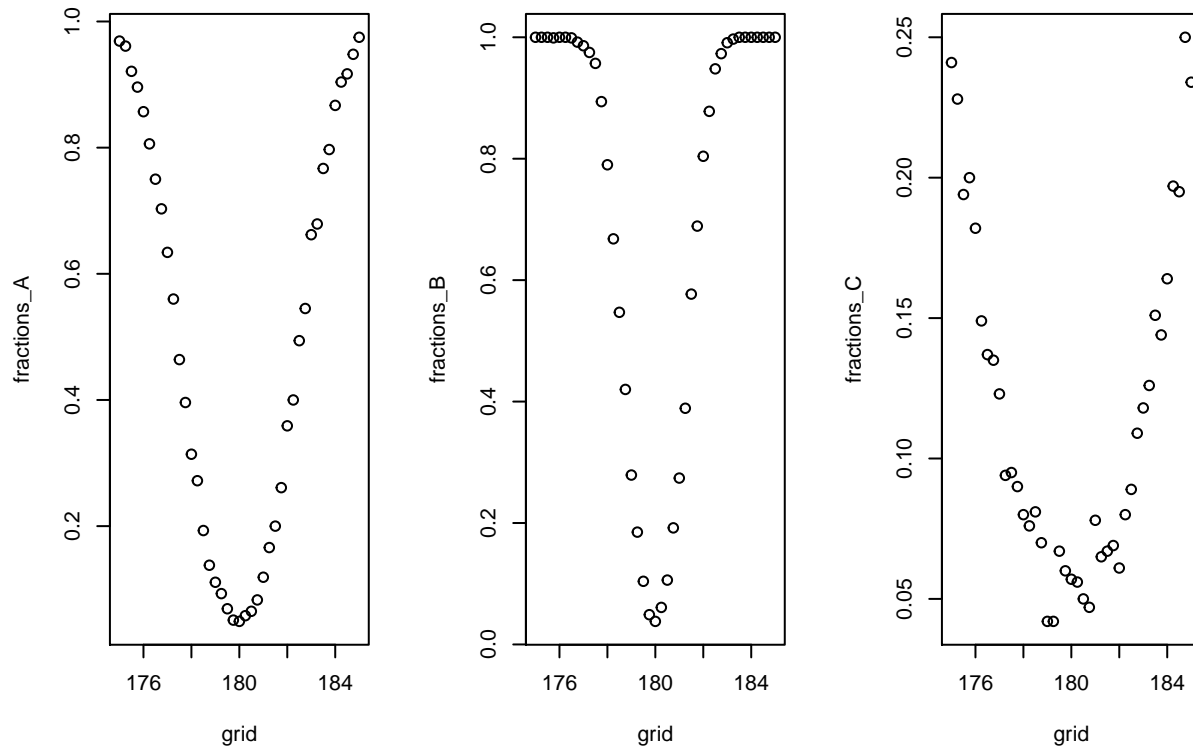
```
n <- m <- 100
mu <- 180
sd <- 5

fractions_B <- power_function(grid,n,m,mu,sd)
```

c) Set $n=m=30$, $\mu=180$ and $sd=15$. Repeat the preceding exercise.

```
n <- m <- 30
mu <- 180
sd <- 15

fractions_C <- power_function(grid,n,m,mu,sd)
par(mfrow=c(1,3))
plot(grid,fractions_A)
plot(grid,fractions_B)
plot(grid,fractions_C)
```



d) Explain your findings.

More datapoints seems to have an influence on the narrowness of the plot. Furthermore, a bigger standard deviations gives a more wider distribution of fractions as presented in the plot of C with lower fractions. This can be explained by the fact that a higher standard deviations gives a higher uncertainty which results in a lower amount of fractions with a p-value below 0.05.

Exercise 3

A telecommunication company has entered the market for mobile phones in a new country. The company's marketing manager conducts a survey of 200 new subscribers for mobile phones. The results of the survey are in the data set `telephone.txt`, which contains the first month bills X_1, \dots, X_{200} , in euros.

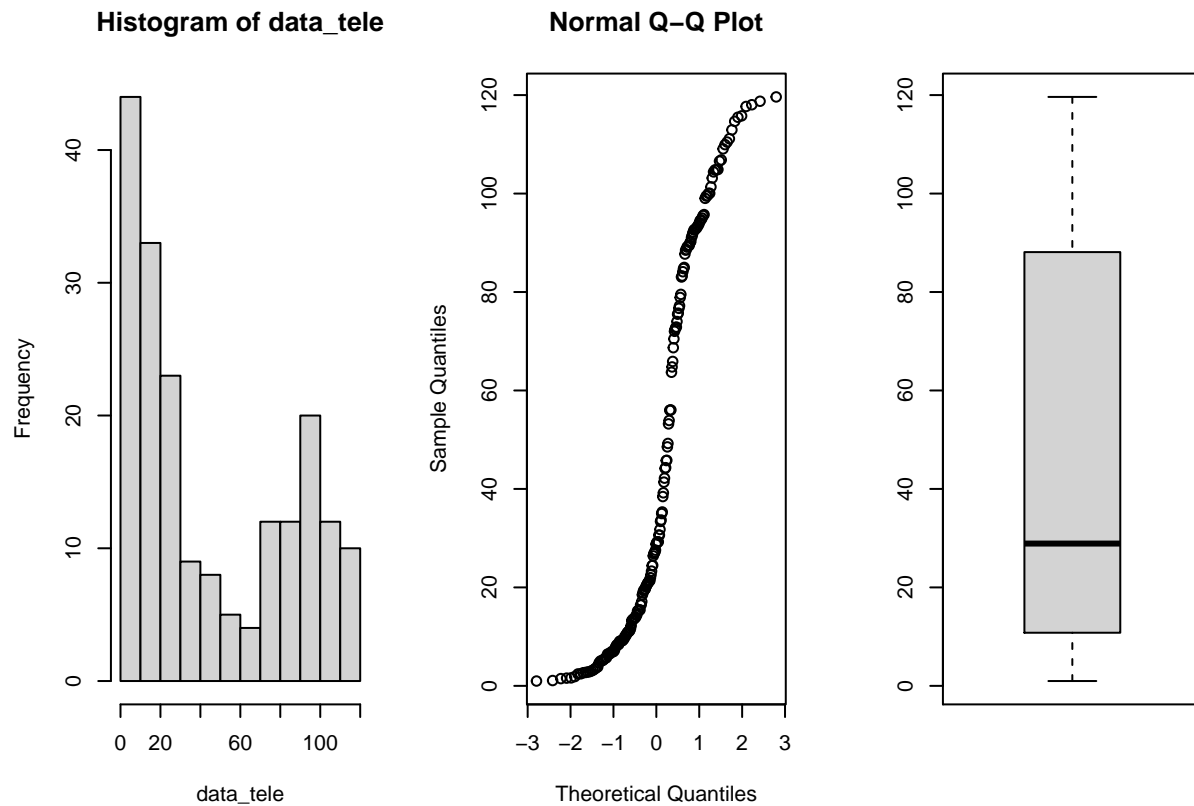
a) Make an appropriate plot of this data set. What marketing advice(s) would you give to the marketing manager? Are there any inconsistencies in the data? If so, try to fix these.

```
data<-read.table(file="data/telephone.txt",header=TRUE)

# remove zeros
data <- data %>%
  filter(Bills > 0)

data_tele <- data$Bills
par(mfrow=c(1,3))
hist(data_tele)
```

```
qqnorm(data_tele)
boxplot(data_tele)
```



From the survey it seems that there are two distinct peaks, therefore it would be good to run two separate marketing campaigns: a “premium” service campaign for customers who are willing to spend more and a campaign aimed at savers, usually people who use pre-paid services, - establishing a separate “cheaper” brand would be a good strategy here.

The survey data also encompassed people who did not have any spendings on the phone bills, therefore they were removed from the analysis.

b) By using a bootstrap test with the test statistic $T = \text{median}(X_1, \dots, X_{200})$, test whether the data `telephone.txt` stems from the exponential distribution $\text{Exp}(\lambda)$ with some λ from $[0.01, 0.1]$.

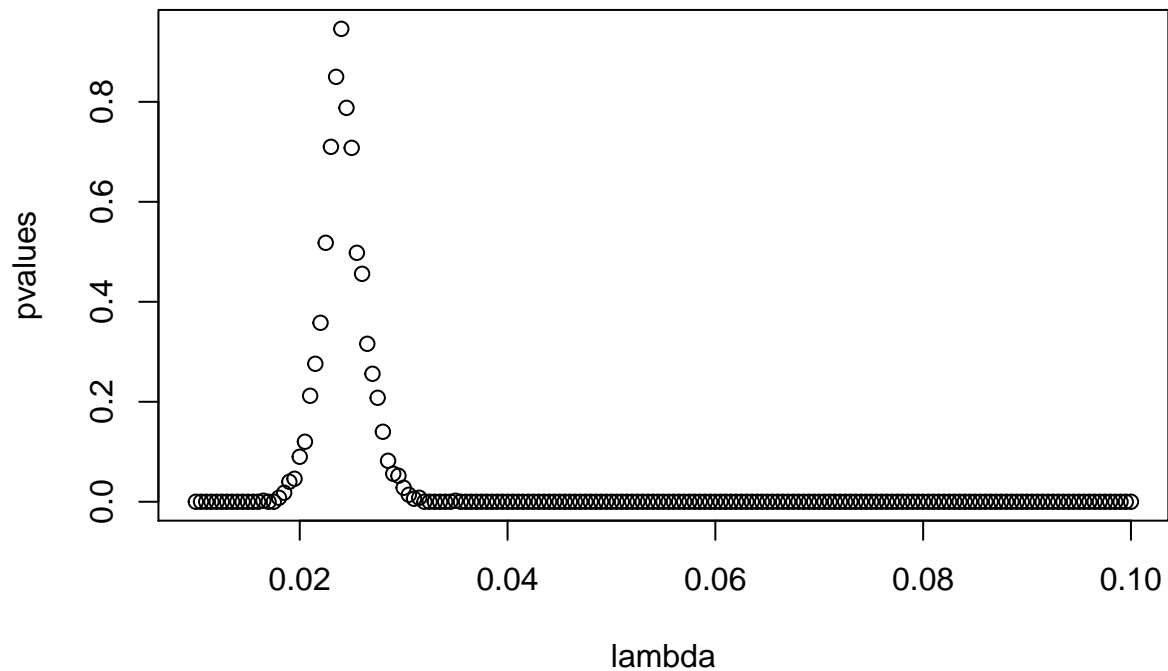
```
lambda <- seq(0.01, 0.1, 0.0005)
pvalues <- c()
t <- median(data_tele)
for (x in lambda){
  B <- 1000
  tstar <- numeric(B)
  n <- length(data_tele)

  for (i in 1:B){
    xstar <- rexp(n,x)
    tstar[i] <- median(xstar)
  }
}
```

```

pl<-sum(tstar<t)/B
pr<-sum(tstar>t)/B
p<-2*min(pl,pr)
pl;pr;p
pvalues <- c(pvalues,p)
}
plot(lambda, pvalues)

```



```

best_p <- which(pvalues > 0.05)
lambda[best_p]

```

```

## [1] 0.0200 0.0205 0.0210 0.0215 0.0220 0.0225 0.0230 0.0235 0.0240 0.0245
## [11] 0.0250 0.0255 0.0260 0.0265 0.0270 0.0275 0.0280 0.0285 0.0290 0.0295

```

The figure above shows the p-values for different lambda values. There can be concluded that the data stems from an exponential distribution for the lambda values 0.02 till 0.029.

c) Construct a 95% bootstrap confidence interval for the population median of the sample.

```

B <- 1000
T1 <- median(data_tele)
Tstar <- numeric(B)
for (i in 1:B){
  Xstar <- sample(data_tele,replace=TRUE)
  Tstar[i] <- median(Xstar)
}

```

```

}
Tstar25 <- quantile(Tstar,0.025)
Tstar975 <- quantile(Tstar, 0.975)

T1

```

```
## [1] 28.905
```

```
c(2*T1-Tstar975, 2*T1-Tstar25)
```

```
##      97.5%      2.5%
## 17.51500 36.72062
```

d) Assuming $X_1, \dots, X_n \sim \text{Exp}(\lambda)$ and using the central limit theorem for the sample mean, estimate λ and construct again a 95% confidence interval for the population median. Comment on your findings.

```

max_index <- which.max(pvalues)

opt_Lambda <- lambda[max_index]

```

The variable `opt_Lambda` is the optimal λ value with the highest P-value.

e) Using an appropriate test, test the null hypothesis that the median bill is bigger or equal to 40 euro against the alternative that the median bill is smaller than 40 euro. Next, design and perform a test to check whether the fraction of the bills less than 10 euro is less than 25%.

```

bill_bigeq40 <- sum(data_tele>=40)
bill_smal40 <- sum(data_tele<40)

binom.test(bill_bigeq40, length(data_tele),p=0.5)

```

```

##
## Exact binomial test
##
## data: bill_bigeq40 and length(data_tele)
## number of successes = 83, number of trials = 192, p-value = 0.07092
## alternative hypothesis: true probability of success is not equal to 0.5
## 95 percent confidence interval:
##  0.3611512 0.5055558
## sample estimates:
## probability of success
##                0.4322917

```

```
binom.test(bill_smal40, length(data_tele),p=0.5)
```

```

##
## Exact binomial test
##
## data: bill_smal40 and length(data_tele)
## number of successes = 109, number of trials = 192, p-value = 0.07092

```

```
## alternative hypothesis: true probability of success is not equal to 0.5
## 95 percent confidence interval:
##  0.4944442 0.6388488
## sample estimates:
## probability of success
##           0.5677083
```

```
bill_less10 <- sum(data_tele < 10)
bill_less10/length(data_tele)
```

```
## [1] 0.2291667
```

Exercise 4

To study the effect of energy drink a sample of 24 high school pupils were randomized to drinking either a softdrink or an energy drink after running for 60 meters. After half an hour they were asked to run again. For both sprints they were asked to sprint as fast they could, and the sprinting time was measured. The data is given in the file run.txt. [Courtesy class 5E, Stedelijk Gymnasium Leiden, 2010.]

a) Disregarding the type of drink, test whether the run times before drink and after are correlated.

```
data <- read.table(file="data/run.txt",header=TRUE)
cor(data$before, data$after)
```

```
## [1] 0.638803
```

Run times before and after the drink seem to be positively correlated.

b) Test separately, for both the softdrink and the energy drink conditions, whether there is a difference in speed in the two running tasks.

```
# calculate differences
data <- data %>%
  mutate(diff = before - after)

# filter for lemo
lemo <- data %>%
  filter(drink == "lemon")

t.test(lemo$before, lemo$after, paired = TRUE)
```

```
##
## Paired t-test
##
## data: lemo$before and lemo$after
## t = -0.80596, df = 11, p-value = 0.4373
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -0.5409781 0.2509781
## sample estimates:
## mean of the differences
##           -0.145
```



```
# filter for energy

energy <- data %>%
  filter(drink == "energy")

t.test(energy$before, energy$after, paired = TRUE)

##
## Paired t-test
##
## data: energy$before and energy$after
## t = 1.6538, df = 11, p-value = 0.1264
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -0.05101059 0.35934392
## sample estimates:
## mean of the differences
## 0.1541667
```

For both energy and soft-drink groups there does not seem to be a significant difference in running times.

c) For each pupil compute the time difference between the two running tasks. Test whether these time differences are effected by the type of drink.

```
# perform t-test

t.test(lemo$diff, energy$diff)

##
## Welch Two Sample t-test
##
## data: lemo$diff and energy$diff
## t = -1.4764, df = 16.509, p-value = 0.1586
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -0.7276409 0.1293076
## sample estimates:
## mean of x mean of y
## -0.1450000 0.1541667
```

The p-value is > 0.05 therefore the means of the two populations are not significantly different.

d) Can you think of a plausible objection to the design of the experiment in b) if the main aim was to test whether drinking the energy drink speeds up the running? Is there a similar objection to the design of the experiment in c)? Comment on all your findings in this exercise.

In both experiments the participants were asked to run on the same day. This could strongly influence the outcomes in data. Therefore, the setup was certainly not ideal to check the influence of both drinks.

Exercise 5

a) Test whether the distributions of the chicken weights for meatmeal and sunflower groups are different by performing three tests: the two samples t-test (argue whether the data are paired or not), the Mann-Whitney test and the Kolmogorov-Smirnov test. Comment on your findings.

```

# filter for meatmeal

meatmeal <- chickwts %>%
  filter(feed == "meatmeal") %>%
  select(weight)

# filter for sunflower

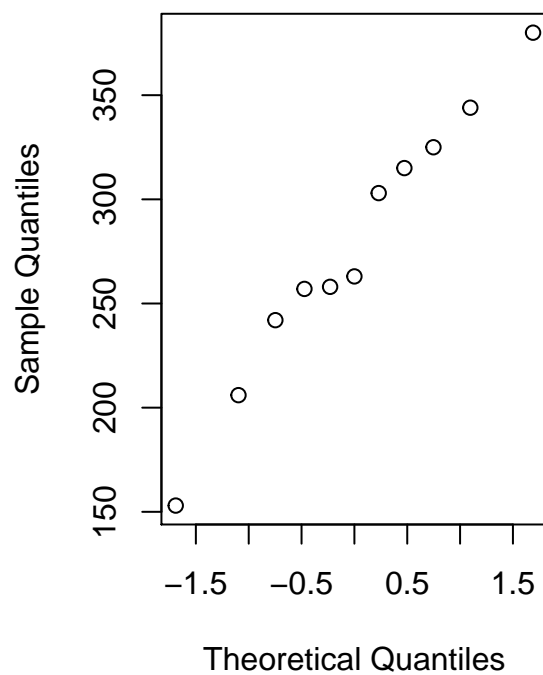
sunflower <- chickwts %>%
  filter(feed == "sunflower") %>%
  select(weight)

# check for data normality

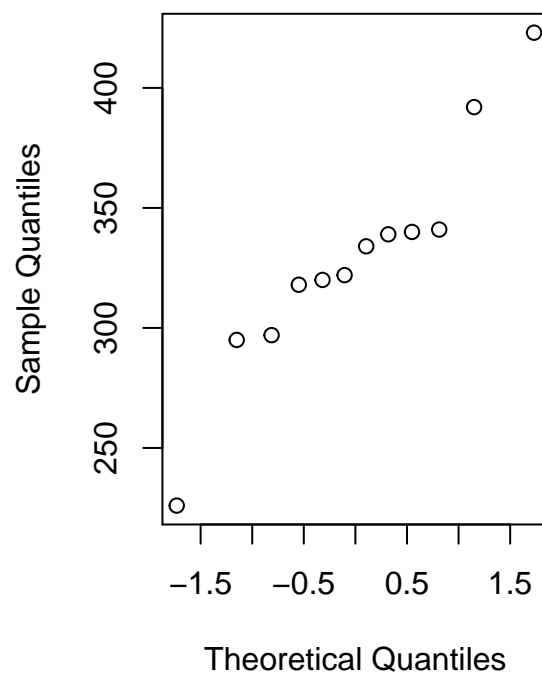
par(mfrow=c(1,2))
qqnorm(meatmeal$weight)
qqnorm(sunflower$weight)

```

Normal Q-Q Plot



Normal Q-Q Plot



```

# perform t-test, the data is not paired

t.test(meatmeal, sunflower)

```

```

##
##  Welch Two Sample t-test
##

```

```
## data: meatmeal and sunflower
## t = -2.1564, df = 18.535, p-value = 0.04441
## alternative hypothesis: true difference in means is not equal to 0
## 95 percent confidence interval:
## -102.572435 -1.442716
## sample estimates:
## mean of x mean of y
## 276.9091 328.9167
```

```
# Mann-Whitney test
```

```
wilcox.test(meatmeal$weight, sunflower$weight)
```

```
##
## Wilcoxon rank sum exact test
##
## data: meatmeal$weight and sunflower$weight
## W = 36, p-value = 0.06882
## alternative hypothesis: true location shift is not equal to 0
```

```
# Kolmogorov-Smirnov test
```

```
ks.test(meatmeal$weight, sunflower$weight)
```

```
##
## Two-sample Kolmogorov-Smirnov test
##
## data: meatmeal$weight and sunflower$weight
## D = 0.47727, p-value = 0.1085
## alternative hypothesis: two-sided
```

Data in chickwts is not paired as the “treatment” of different feed was applied to different newly-hatched chicks not the same chick. From t-test we can see that the p-values < 0.05 , therefore the means between the two groups are significantly different. From Mann-Whitney test we can see that p-value is > 0.05 therefore we can not conclude that the medians of the two datasets are different. From Kolmogorov-Smirnov test we can not conclude that the means are different.

b) Conduct a one-way ANOVA to determine whether the type of feed supplement has an effect on the weight of the chicks. Give the estimated chick weights for each of the six feed supplements. What is the best feed supplement?

```
chickaov <- lm(weight~feed, data = chickwts)
# performing one-way ANOVA
anova(chickaov)
```

```
## Analysis of Variance Table
##
## Response: weight
##          Df Sum Sq Mean Sq F value    Pr(>F)
## feed         5 231129   46226  15.365 5.936e-10 ***
## Residuals  65 195556     3009
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

```
#extracting more information
```

```
summary_table <- summary(chickaov)
knitr::kable(data.frame(summary_table$coefficients), "latex")
```

	Estimate	Std..Error	t.value	Pr...t..
(Intercept)	323.583333	15.83391	20.4360920	0.0000000
feedhorsebean	-163.383333	23.48549	-6.9567776	0.0000000
feedlinseed	-104.833333	22.39254	-4.6816194	0.0000149
feedmeatmeal	-46.674242	22.89580	-2.0385502	0.0455667
feedsoybean	-77.154762	21.57799	-3.5756235	0.0006654
feedsunflower	5.333333	22.39254	0.2381746	0.8124949

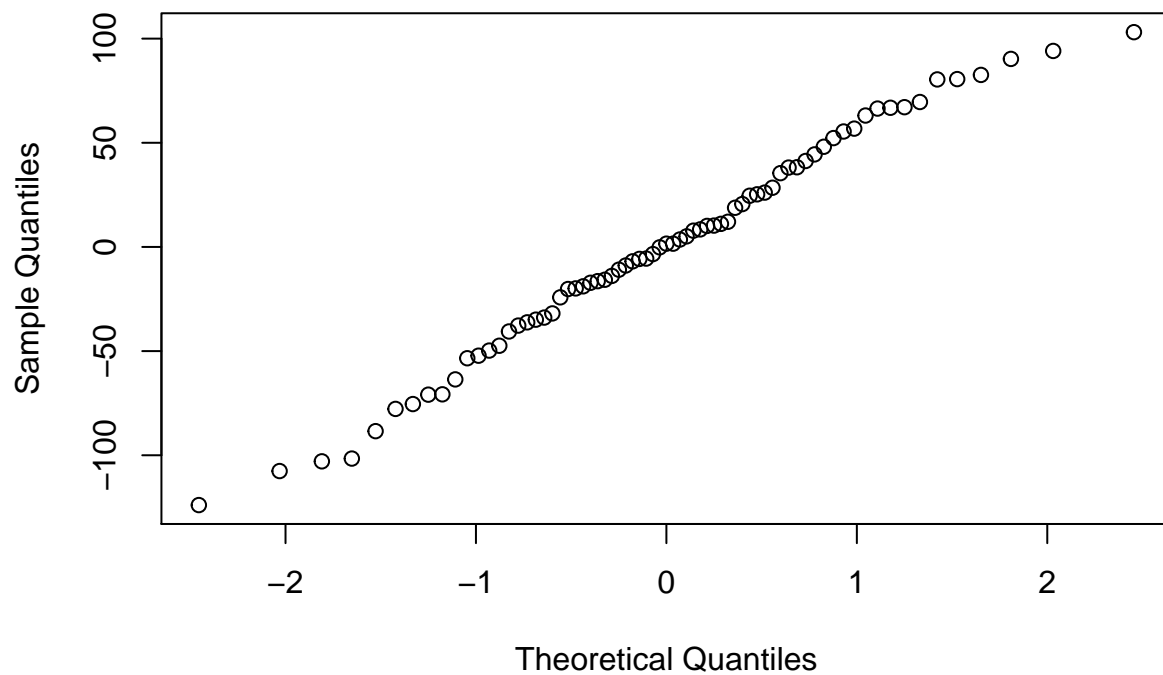
From the results of one-way ANOVA we can see that the p-values is < 0.05 therefore we can conclude that the means between all of the feed varieties are significantly different. From summary statistics it seems that “sunflower” feed is the feed resulting in highest weight. therefore it is the best.

c) Check the ANOVA model assumptions by using relevant diagnostic tools.

```
# check for normality
```

```
qqnorm(chickaov$residuals)
```

Normal Q-Q Plot



```
# check if the variances are equal
```

```
chickwts %>%
  group_by(feed) %>%
  summarise(variance = var(weight))
```

```
## # A tibble: 6 x 2
##   feed      variance
## * <fct>      <dbl>
## 1 casein      4152.
## 2 horsebean   1492.
## 3 linseed     2729.
## 4 meatmeal    4212.
## 5 soybean     2930.
## 6 sunflower   2385.
```

From qqplot assumption of normality holds. However the assumption of equal variances does not hold.

d) Does the Kruskal-Wallis test arrive at the same conclusion about the effect of feed supplement as the test in b)? Explain possible differences between conclusions of the Kruskal-Wallis and ANOVA tests.

```
kruskal.test(weight~feed, data = chickwts)
```

```
##
##  Kruskal-Wallis rank sum test
##
## data:  weight by feed
## Kruskal-Wallis chi-squared = 37.343, df = 5, p-value = 5.113e-07
```

With Kruskal-Wallis test we arrive to the same conclusion as with ANOVA.