

Wine Quality Regression Problem

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Abstract We are using a dataset related to red vinho verde wine samples, from the north of Portugal. The goal is to model wine quality based on physicochemical tests. The classes are ordered and not balanced (e.g. there are much more normal wines than excellent or poor ones). Outlier detection algorithms could be used to detect the few excellent or poor wines. Also, we are not sure if all input variables are relevant. So it could be interesting to test feature selection methods. The method chosen to solve the problem is Linear Regression.

Introduction

Once viewed as a luxury good, nowadays wine is increasingly enjoyed by all consumers. Portugal is a top ten wine exporting country with 3.17% of the market share in 2005 ([FAOSTAT](#)). Exports of its vinho verde wine (from the northwest region) have increased by yearly. To support its growth, the wine industry is investing in new technologies for both wine making and selling processes. Wine certification and quality assessment are key elements within this context. Certification prevents the illegal adulteration of wines (to safeguard human health) and assures quality for the wine market. Quality evaluation is often part of the certification process and can be used to improve wine making (by identifying the most influential factors) and to stratify wines such as premium brands (useful for setting prices). Wine certification is generally assessed by physicochemical and sensory tests ([Teranishi et al., 1999](#)). Physicochemical laboratory tests routinely used to characterize wine include determination of density, alcohol or pH values, while sensory tests rely mainly on human experts. It should be stressed that taste is the least understood of the human senses, thus wine classification is a difficult task. Moreover, the relationships between the physicochemical and sensory analysis are complex and still not fully understood ([Legin et al., 2003](#)).

Background

The two datasets are related to red and white variants of the Portuguese “Vinho Verde” wine. For more details, consult ([Cortez et al., 1998](#)). Due to privacy and logistic issues, only physicochemical (inputs) and sensory (the output) variables are available (e.g. there is no data about grape types, wine brand, wine selling price, etc.).

These datasets can be viewed as classification or regression tasks. The classes are ordered and not balanced (e.g. there are much more normal wines than excellent or poor ones). Outlier detection algorithms could be used to detect the few excellent or poor wines. Also, we are not sure if all input variables are relevant.

Due to specific purpose of this lab assignment, we are looking at Linear Regression problem only using red wine dataset. Full library of the wine datasets and their description are located here: ([UCI Wine Data Set](#)).

Objective

The objective of this article is to provide a reliable and feasible recommendation algorithm to predict wine quality based on physicochemical tests.

- Put all relevant variables in the model
- Leave the irrelevant variables out
- Check linearity
- Regression assumptions

** Residuals have a mean of zero ** Normality of errors ** Residuals are not autocorrelated ** Linearity variables ** Need more data than ** No excessive collinearity

Data understanding

The dataset ([UCI Wine Data Set](#)) has 8 attributes and 12960 instances. The following concept structure:

For more information, read [Cortez et al., 2009].
Input variables (based on physicochemical tests):

```

1 - fixed acidity
2 - volatile acidity
3 - citric acid
4 - residual sugar
5 - chlorides
6 - free sulfur dioxide
7 - total sulfur dioxide
8 - density
9 - pH
10 - sulphates
11 - alcohol
Output variable (based on sensory data):
12 - quality (score between 0 and 10)

```

Preparation

To perform the analysis, certain R libraries were used. The code below was used to load and initialize the libraries. The first line invoking seed function was applied to enforce the repeatability of the calculation results.

```

set.seed(42)
library(ggplot2)
library(rpart)
library(rpart.plot)
library(rattle)
library(caret)

```

Reading red wines dataset

```

library(readr)
wines_red_data <-
  read.csv(
    "http://archive.ics.uci.edu/ml/machine-learning-databases/wine-quality/winequality-red.csv",
    sep = ";",
    header = TRUE,
    col.names = c("FA", "VA", "CA", "RS", "CH", "FSD", "TSD", "DEN", "pH", "SUL", "ALC", "QLT"))

str(wines_red_data)

#> 'data.frame':   1599 obs. of  12 variables:
#> $ FA : num  7.4 7.8 7.8 11.2 7.4 7.4 7.9 7.3 7.8 7.5 ...
#> $ VA : num  0.7 0.88 0.76 0.28 0.7 0.66 0.6 0.65 0.58 0.5 ...
#> $ CA : num  0 0 0.04 0.56 0 0 0.06 0 0.02 0.36 ...
#> $ RS : num  1.9 2.6 2.3 1.9 1.9 1.8 1.6 1.2 2 6.1 ...
#> $ CH : num  0.076 0.098 0.092 0.075 0.076 0.075 0.069 0.065 0.073 0.071 ...
#> $ FSD: num  11 25 15 17 11 13 15 15 9 17 ...
#> $ TSD: num  34 67 54 60 34 40 59 21 18 102 ...
#> $ DEN: num  0.998 0.997 0.997 0.998 0.998 ...
#> $ pH : num  3.51 3.2 3.26 3.16 3.51 3.51 3.3 3.39 3.36 3.35 ...
#> $ SUL: num  0.56 0.68 0.65 0.58 0.56 0.56 0.46 0.47 0.57 0.8 ...
#> $ ALC: num  9.4 9.8 9.8 9.8 9.4 9.4 9.4 10 9.5 10.5 ...
#> $ QLT: int  5 5 5 6 5 5 5 7 7 5 ...

```

Distribution of target value in the dataset

Figure 1:

```

h <- hist(wines_red_data$QLT, freq=FALSE, xlab = "Wine Quality (QLT)", main="",
          col = colors()[626])

```

	FA	VA	CA	RS	CH	FSD	TSD	DEN	pH	SUL	ALC	QLT
1	7.40	0.70	0.00	1.90	0.08	11.00	34.00	1.00	3.51	0.56	9.40	5
2	7.80	0.88	0.00	2.60	0.10	25.00	67.00	1.00	3.20	0.68	9.80	5
3	7.80	0.76	0.04	2.30	0.09	15.00	54.00	1.00	3.26	0.65	9.80	5
4	11.20	0.28	0.56	1.90	0.07	17.00	60.00	1.00	3.16	0.58	9.80	6
5	7.40	0.70	0.00	1.90	0.08	11.00	34.00	1.00	3.51	0.56	9.40	5
6	7.40	0.66	0.00	1.80	0.07	13.00	40.00	1.00	3.51	0.56	9.40	5
7	7.90	0.60	0.06	1.60	0.07	15.00	59.00	1.00	3.30	0.46	9.40	5
8	7.30	0.65	0.00	1.20	0.06	15.00	21.00	0.99	3.39	0.47	10.00	7
9	7.80	0.58	0.02	2.00	0.07	9.00	18.00	1.00	3.36	0.57	9.50	7
10	7.50	0.50	0.36	6.10	0.07	17.00	102.00	1.00	3.35	0.80	10.50	5
11	6.70	0.58	0.08	1.80	0.10	15.00	65.00	1.00	3.28	0.54	9.20	5
12	7.50	0.50	0.36	6.10	0.07	17.00	102.00	1.00	3.35	0.80	10.50	5
13	5.60	0.61	0.00	1.60	0.09	16.00	59.00	0.99	3.58	0.52	9.90	5
14	7.80	0.61	0.29	1.60	0.11	9.00	29.00	1.00	3.26	1.56	9.10	5
15	8.90	0.62	0.18	3.80	0.18	52.00	145.00	1.00	3.16	0.88	9.20	5
16	8.90	0.62	0.19	3.90	0.17	51.00	148.00	1.00	3.17	0.93	9.20	5
17	8.50	0.28	0.56	1.80	0.09	35.00	103.00	1.00	3.30	0.75	10.50	7
18	8.10	0.56	0.28	1.70	0.37	16.00	56.00	1.00	3.11	1.28	9.30	5
19	7.40	0.59	0.08	4.40	0.09	6.00	29.00	1.00	3.38	0.50	9.00	4
20	7.90	0.32	0.51	1.80	0.34	17.00	56.00	1.00	3.04	1.08	9.20	6

Table 1: Red Wines Quality Dataset - first 20 rows

OLS Modeling

Default Linear Regression fit

The following code calculates the default OLS model using all the independent variables. Results of the calculations presented in Table 2.

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	21.9652	21.1946	1.04	0.3002
FA	0.0250	0.0259	0.96	0.3357
VA	-1.0836	0.1211	-8.95	0.0000
CA	-0.1826	0.1472	-1.24	0.2150
RS	0.0163	0.0150	1.09	0.2765
CH	-1.8742	0.4193	-4.47	0.0000
FSD	0.0044	0.0022	2.01	0.0447
TSD	-0.0033	0.0007	-4.48	0.0000
DEN	-17.8812	21.6331	-0.83	0.4086
pH	-0.4137	0.1916	-2.16	0.0310
SUL	0.9163	0.1143	8.01	0.0000
ALC	0.2762	0.0265	10.43	0.0000

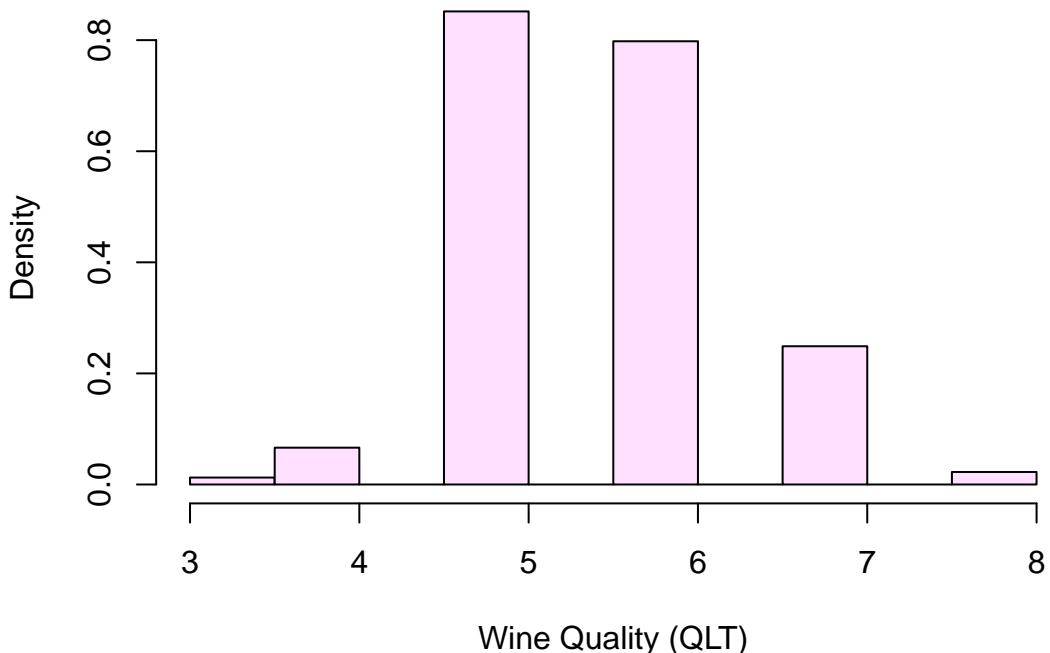
Table 2: Default OLS model - all variables included

More detailed summaries

```
hist(residuals(wines_red_data.fit), xlab = "Residuals", main = "")
```

Adjust the fit removing attributes with p > 0.05

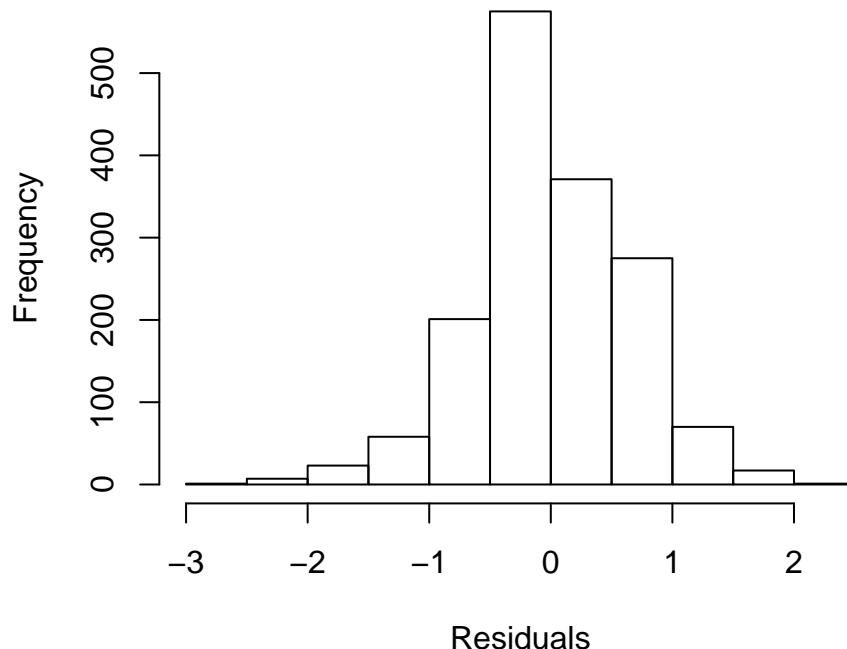
```
wines_red_data.fit1 <- lm (QLT ~ VA + CH + FSD + TSD + pH + SUL + ALC,
                           data=wines_red_data)
summary(wines_red_data.fit1)
```

**Figure 1:** Histogram of White Wine QUality

```

#>
#> Call:
#> lm(formula = QLT ~ VA + CH + FSD + TSD + pH + SUL + ALC, data = wines_red_data)
#>
#> Residuals:
#>   Min     1Q   Median     3Q    Max 
#> -2.68918 -0.36757 -0.04653  0.46081  2.02954
#>
#> Coefficients:
#>             Estimate Std. Error t value Pr(>|t|)    
#> (Intercept) 4.4300987  0.4029168 10.995 < 2e-16 ***
#> VA          -1.0127527  0.1008429 -10.043 < 2e-16 ***
#> CH          -2.0178138  0.3975417 -5.076 4.31e-07 ***
#> FSD          0.0050774  0.0021255  2.389  0.017 *  
#> TSD          -0.0034822  0.0006868 -5.070 4.43e-07 ***
#> pH           -0.4826614  0.1175581 -4.106 4.23e-05 ***
#> SUL          0.8826651  0.1099084  8.031 1.86e-15 ***
#> ALC          0.2893028  0.0167958 17.225 < 2e-16 ***
#> ---
#> Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
#>
#> Residual standard error: 0.6477 on 1591 degrees of freedom
#> Multiple R-squared:  0.3595, Adjusted R-squared:  0.3567 
#> F-statistic: 127.6 on 7 and 1591 DF,  p-value: < 2.2e-16

```

**Figure 2:** Histogram of residuals

Stepwise Regression

Find the best model automatically

```
# Stepwise Regression
library(MASS)
fit <- lm(QLT ~ ., data=wines_red_data)
step <- stepAIC(fit, direction="both", trace = FALSE)
step$anova # display results

#> Stepwise Model Path
#> Analysis of Deviance Table
#>
#> Initial Model:
#> QLT ~ FA + VA + CA + RS + CH + FSD + TSD + DEN + pH + SUL + ALC
#>
#> Final Model:
#> QLT ~ VA + CH + FSD + TSD + pH + SUL + ALC
#>
#>
#>      Step Df  Deviance Resid. Df Resid. Dev      AIC
#> 1              1587   666.4107 -1375.489
#> 2 - DEN  1  0.2868924  1588   666.6976 -1376.801
#> 3 - FA   1  0.1079824  1589   666.8056 -1378.542
#> 4 - RS   1  0.2566805  1590   667.0623 -1379.926
#> 5 - CA   1  0.4748034  1591   667.5371 -1380.789
```

The model identical to the one found the the previous section

Plot pairwise scatter plots

Pairwise scatter plots 3 to inspect the result for relationships between the independent variable and the numerical dependent variables.

```
attach(wines_red_data)

#> The following object is masked from package:MASS:
#>
#>      VA

panel.points<-function(x,y){points(x,y,cex=.1)}
pairs(~QLT + VA + CH + FSD + TSD + pH + SUL + ALC,
      upper.panel=panel.points,lower.panel=panel.points)
```

Checking correlation matrix

	FA	VA	CA	RS	CH	FSD	TSD	DEN	pH	SUL	ALC	QLT
FA	1.00	-0.26	0.67	0.11	0.09	-0.15	-0.11	0.67	-0.68	0.18	-0.06	0.12
VA	-0.26	1.00	-0.55	0.00	0.06	-0.01	0.08	0.02	0.23	-0.26	-0.20	-0.39
CA	0.67	-0.55	1.00	0.14	0.20	-0.06	0.04	0.36	-0.54	0.31	0.11	0.23
RS	0.11	0.00	0.14	1.00	0.06	0.19	0.20	0.36	-0.09	0.01	0.04	0.01
CH	0.09	0.06	0.20	0.06	1.00	0.01	0.05	0.20	-0.27	0.37	-0.22	-0.13
FSD	-0.15	-0.01	-0.06	0.19	0.01	1.00	0.67	-0.02	0.07	0.05	-0.07	-0.05
TSD	-0.11	0.08	0.04	0.20	0.05	0.67	1.00	0.07	-0.07	0.04	-0.21	-0.19
DEN	0.67	0.02	0.36	0.36	0.20	-0.02	0.07	1.00	-0.34	0.15	-0.50	-0.17
pH	-0.68	0.23	-0.54	-0.09	-0.27	0.07	-0.07	-0.34	1.00	-0.20	0.21	-0.06
SUL	0.18	-0.26	0.31	0.01	0.37	0.05	0.04	0.15	-0.20	1.00	0.09	0.25
ALC	-0.06	-0.20	0.11	0.04	-0.22	-0.07	-0.21	-0.50	0.21	0.09	1.00	0.48
QLT	0.12	-0.39	0.23	0.01	-0.13	-0.05	-0.19	-0.17	-0.06	0.25	0.48	1.00

Table 3: Red Wines Quality Dataset Correlation Matrix

Evaluate Nonlinearity component + residual plot

```
library(car)

#> Loading required package: carData

crPlots(wines_red_data.fit1, layout = c(4,3), main = "")
```

Assessing Outliers

```
outlierTest(wines_red_data.fit) # Bonferroni p-value for most extreme obs

#>      rstudent unadjusted p-value Bonferroni p
#> 833 -4.194088      2.8912e-05     0.046231
```

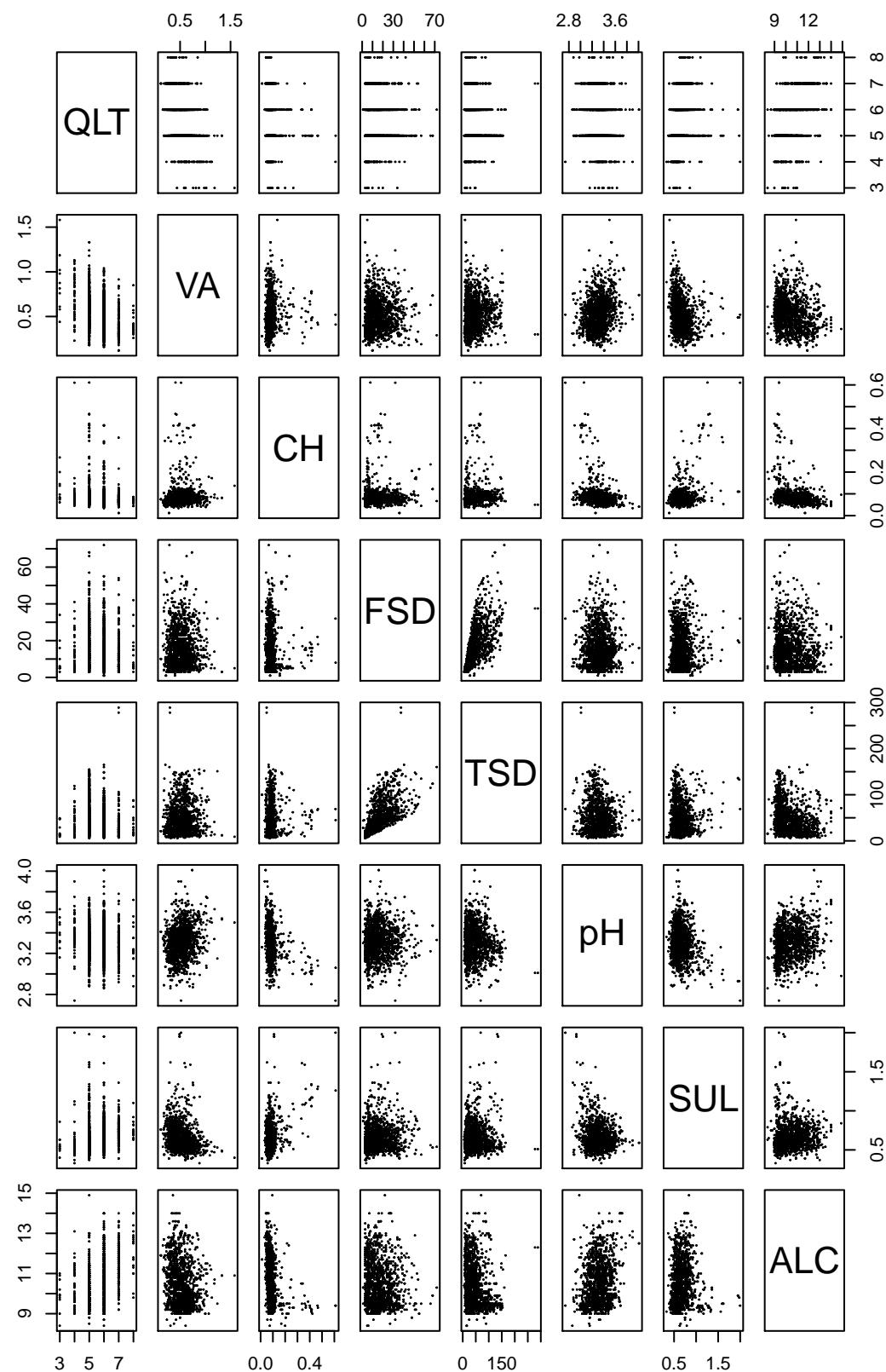
Both p-value and Bonferroni-corrected p-value are smaller than 0.05, so the model is acceptable.

```
qqPlot(wines_red_data.fit, main="", ylab="Studentized Residuals" ) #qq plot for studentized resid

#> [1] 653 833
```

A residual plot is a graph that shows the residuals on the vertical axis and the independent variable on the horizontal axis. If the points in a residual plot are randomly dispersed around the horizontal axis, a linear regression model is appropriate for the data; otherwise, a non-linear model is more appropriate. Our model is reasonably dispersed around 0,0 for each of the independent variables

```
# leverage plots
leveragePlots(wines_red_data.fit, layout = c(4,3), main = "")
```

**Figure 3:** Red Wines - relationships between variables

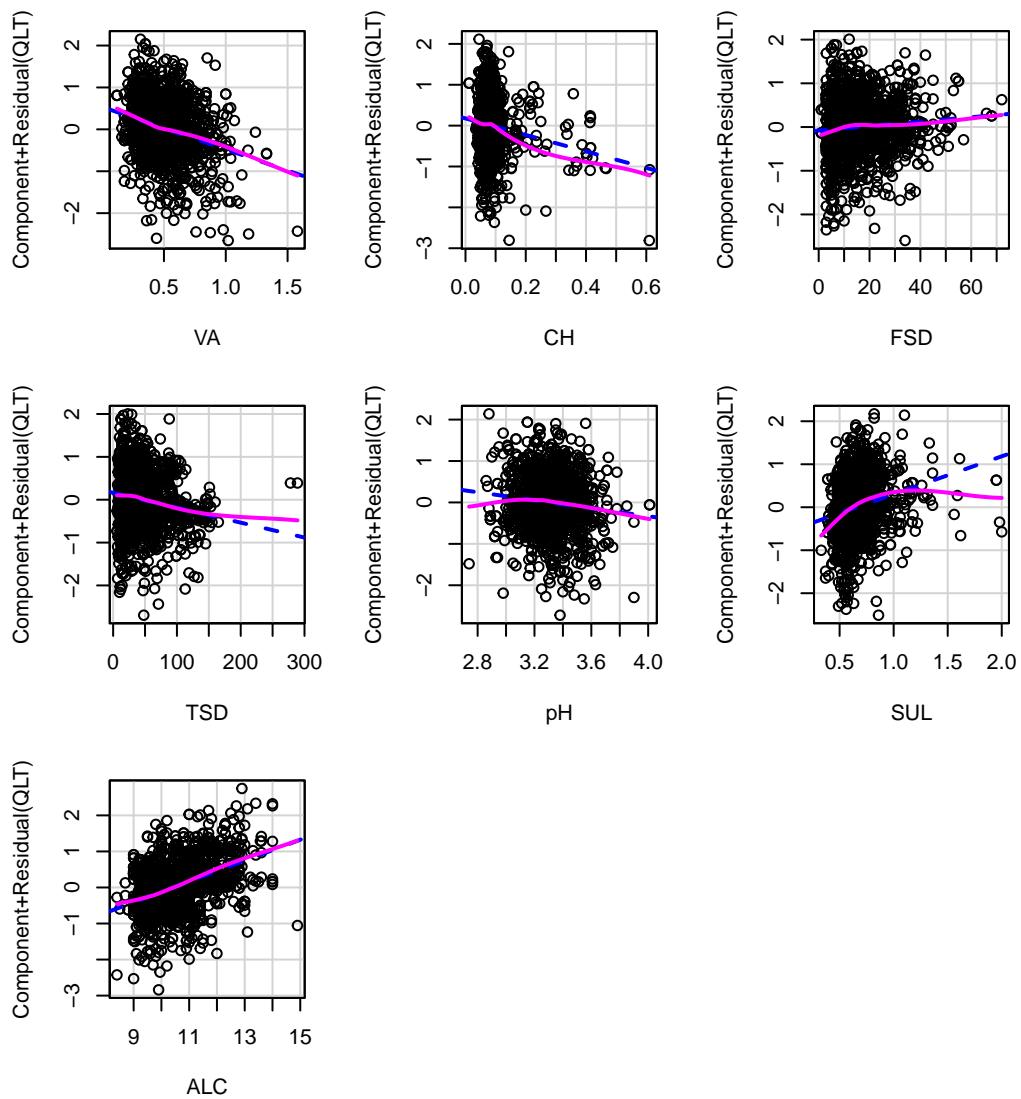


Figure 4: Red Wines - Component + Resudual Plots

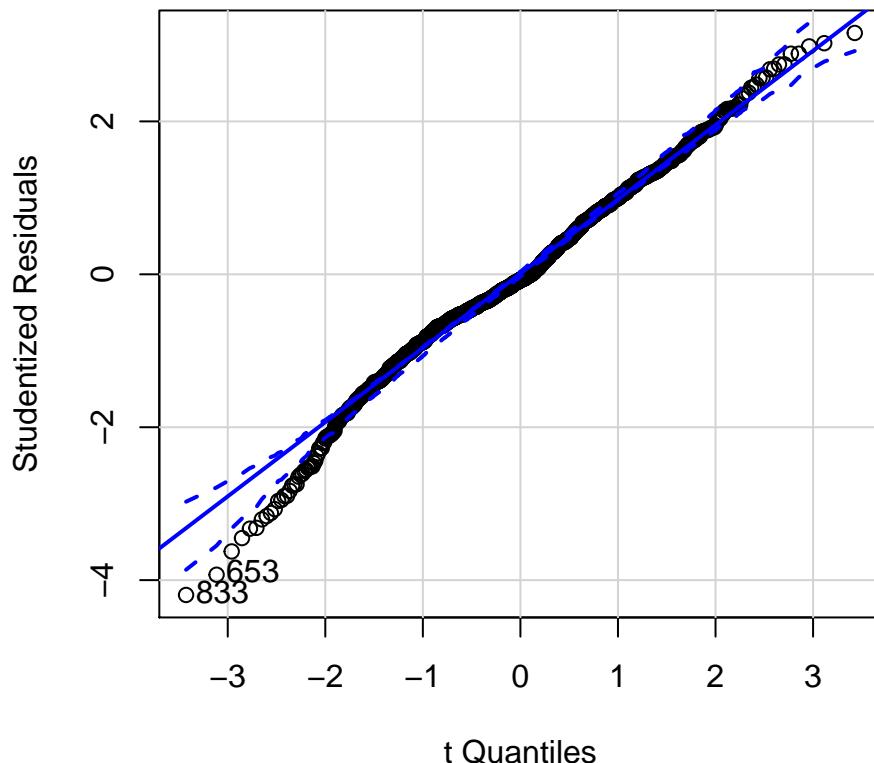


Figure 5: QQ Plot for studentized residuals

```
# Test for Autocorrelated Errors
durbinWatsonTest(wines_red_data.fit)

#> lag Autocorrelation D-W Statistic p-value
#> 1 0.121429 1.75714 0
#> Alternative hypothesis: rho != 0

# Global test of model assumptions
library(gvlma)
gvmmodel <- gvlma(wines_red_data.fit1)
summary(gvmmodel)

#>
#> Call:
#> lm(formula = QLT ~ VA + CH + FSD + TSD + pH + SUL + ALC, data = wines_red_data)
#>
#> Residuals:
#>    Min      1Q  Median      3Q     Max 
#> -2.68918 -0.36757 -0.04653  0.46081  2.02954
#>
#> Coefficients:
#>             Estimate Std. Error t value Pr(>|t|)    
#> (Intercept) 4.4300987  0.4029168 10.995 < 2e-16 ***
#> VA          -1.0127527  0.1008429 -10.043 < 2e-16 ***
#> CH          -2.0178138  0.3975417 -5.076 4.31e-07 ***
#> FSD          0.0050774  0.0021255  2.389  0.017 *  
#> TSD          -0.0034822  0.0006868 -5.070 4.43e-07 ***
#> pH          -0.4826614  0.1175581 -4.106 4.23e-05 ***
```

```

#> SUL          0.8826651  0.1099084   8.031 1.86e-15 ***
#> ALC          0.2893028  0.0167958  17.225 < 2e-16 ***
#> ---
#> Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
#>
#> Residual standard error: 0.6477 on 1591 degrees of freedom
#> Multiple R-squared:  0.3595, Adjusted R-squared:  0.3567
#> F-statistic: 127.6 on 7 and 1591 DF, p-value: < 2.2e-16
#>
#>
#> ASSESSMENT OF THE LINEAR MODEL ASSUMPTIONS
#> USING THE GLOBAL TEST ON 4 DEGREES-OF-FREEDOM:
#> Level of Significance = 0.05
#>
#> Call:
#> gvlma(x = wines_red_data.fit1)
#>
#>           Value    p-value            Decision
#> Global Stat      37.99895 1.121e-07 Assumptions NOT satisfied!
#> Skewness          6.45922 1.104e-02 Assumptions NOT satisfied!
#> Kurtosis          28.78544 8.086e-08 Assumptions NOT satisfied!
#> Link Function     2.70103 1.003e-01 Assumptions acceptable.
#> Heteroscedasticity 0.05326 8.175e-01 Assumptions acceptable.

```

Addressing Skeweness using log transformation

```

library(car)
summary(wines_red_data.fit2 <- lm (
  bcPower(QLT,1.25) ~ VA + CH + FSD + TSD + pH + SUL + ALC,
  data=wines_red_data))

#>
#> Call:
#> lm(formula = bcPower(QLT, 1.25) ~ VA + CH + FSD + TSD + pH +
#>       SUL + ALC, data = wines_red_data)
#>
#> Residuals:
#>   Min     1Q   Median     3Q    Max
#> -3.8872 -0.5845 -0.0839  0.7095  3.2585
#>
#> Coefficients:
#>             Estimate Std. Error t value Pr(>|t|)
#> (Intercept) 4.258050  0.620654  6.861 9.79e-12 ***
#> VA          -1.538767  0.155339 -9.906 < 2e-16 ***
#> CH          -3.112317  0.612374 -5.082 4.17e-07 ***
#> FSD          0.007722  0.003274  2.358  0.0185 *
#> TSD          -0.005436  0.001058 -5.139 3.11e-07 ***
#> pH           -0.751939  0.181087 -4.152 3.46e-05 ***
#> SUL          1.372027  0.169303  8.104 1.05e-15 ***
#> ALC          0.452131  0.025872 17.475 < 2e-16 ***
#> ---
#> Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
#>
#> Residual standard error: 0.9978 on 1591 degrees of freedom
#> Multiple R-squared:  0.3625, Adjusted R-squared:  0.3597
#> F-statistic: 129.2 on 7 and 1591 DF, p-value: < 2.2e-16

gvmodel <- gvlma(wines_red_data.fit2)
summary(gvmodel)

#>
#> Call:
#> lm(formula = bcPower(QLT, 1.25) ~ VA + CH + FSD + TSD + pH +
#>       SUL + ALC, data = wines_red_data)

```

```

#>
#> Residuals:
#>   Min     1Q Median     3Q    Max
#> -3.8872 -0.5845 -0.0839  0.7095  3.2585
#>
#> Coefficients:
#>             Estimate Std. Error t value Pr(>|t|)
#> (Intercept) 4.258050  0.620654  6.861 9.79e-12 ***
#> VA          -1.538767  0.155339 -9.906 < 2e-16 ***
#> CH          -3.112317  0.612374 -5.082 4.17e-07 ***
#> FSD         0.007722  0.003274  2.358  0.0185 *
#> TSD         -0.005436  0.001058 -5.139 3.11e-07 ***
#> pH          -0.751939  0.181087 -4.152 3.46e-05 ***
#> SUL         1.372027  0.169303  8.104 1.05e-15 ***
#> ALC         0.452131  0.025872 17.475 < 2e-16 ***
#> ---
#> Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
#>
#> Residual standard error: 0.9978 on 1591 degrees of freedom
#> Multiple R-squared:  0.3625, Adjusted R-squared:  0.3597
#> F-statistic: 129.2 on 7 and 1591 DF, p-value: < 2.2e-16
#>
#>
#> ASSESSMENT OF THE LINEAR MODEL ASSUMPTIONS
#> USING THE GLOBAL TEST ON 4 DEGREES-OF-FREEDOM:
#> Level of Significance = 0.05
#>
#> Call:
#> gvlma(x = wines_red_data.fit2)
#>
#>           Value p-value          Decision
#> Global Stat      24.80148 5.515e-05 Assumptions NOT satisfied!
#> Skewness          0.50847 4.758e-01 Assumptions acceptable.
#> Kurtosis          20.50362 5.952e-06 Assumptions NOT satisfied!
#> Link Function     3.77416 5.205e-02 Assumptions acceptable.
#> Heteroscedasticity 0.01524 9.018e-01 Assumptions acceptable.

hist(residuals(wines_red_data.fit2), xlab = "Residuals", main = "")

```

Tree-based regression methods

Tree-based methods, while simple and useful for interpretation, are typically not as competitive with the best supervised learning approaches such as polynomial regression. However, tree-based methods such as regression tree and random forests make up for this shortfall. By combining a large number of trees instead of one, the model usually results in dramatic improvements in terms of prediction accuracy. This improvement in accuracy comes at the expense of loss in interpretation.

Splitting the dataset into train and test

The dataset has been split in such a way that train and test sets would have the same distribution of the 'QLT' attribute. The reason for this stratification strategy is to focus on the priority on the target value. We used 60:34 split ratio.

```

library(caret)
train.rows<- createDataPartition(y= wines_red_data$QLT, p=0.6, list = FALSE)
train.data<- wines_red_data[train.rows,]
prop.table((table(train.data$QLT)))

#>
#>      3       4       5       6       7       8
#> 0.005202914 0.029136316 0.430801249 0.398543184 0.124869927 0.011446410

test.data<- wines_red_data[-train.rows,]
prop.table((table(test.data$QLT)))

```

```
#>
#>      3      4      5      6      7      8
#> 0.007836991 0.039184953 0.418495298 0.399686520 0.123824451 0.010971787
```

Regression tree fit

In a regression tree, the tree arranges or segments observations into regions of a predictor space. For example, using the “Hitters” data set, which contains various statistics on baseball players, a tree might look something like in Figure 8 generated by the code below.

```
library(rpart)
library(rpart.plot)
library(rattle)
library(caret)

reg.tree <- rpart(QLT ~ ., method="anova", data = train.data)
fancyRpartPlot(reg.tree, main="", sub="")

# reg.tree$variable.importance
```

Regression Tree model evaluation

This Decision Tree favours the following attributes in order of their importance for the prediction of the target attribute: health, has_nurs, parents. It does not consider the rest of the attributes as important. Let’s apply the model to the test set and evaluate accuracy of the predictions.

```
dtPrediction <- predict(reg.tree, test.data)
cor(dtPrediction,test.data$QLT)

#> [1] 0.5480833

plot(jitter(test.data$QLT), dtPrediction,
      pch=20, col=rgb(0.1, 0.2, 0.8, 0.3),
      ylab="Prediction", xlab="Test Values", bty="n" )

qqPlot(dtPrediction, main="" )

#> 127 128
#> 44 45
```

Random Forest model fit

```
library(randomForest)
fitRF1 <- randomForest(
  QLT ~ ., method="anova",
  data=train.data, importance=TRUE, ntree=2000)

varImpPlot(fitRF1, main="")
```

Random Forest model prediction and evaluation

```
PredictionRF1 <- predict(fitRF1, test.data)
cor(PredictionRF1,test.data$QLT)

#> [1] 0.6748129

plot(jitter(test.data$QLT), PredictionRF1 ,
     pch=20, col=rgb(0.1, 0.2, 0.8, 0.3),
     ylab="Prediction", xlab="Test Values", bty="n" )

qqPlot(PredictionRF1, main="")

#> 199 128
#> 75 45
```

Conclusion

Bibliography

- P. Cortez, A. Cerdeira, F. Almeida, T. Matos, and J. Reis. Modeling wine preferences by data mining from physicochemical properties. *Decision Support Systems*, 47(4):547–553, 1998. [p1]
- FAOSTAT. Faostat. URL <http://faostat.fao.org/site/535/DesktopDefault.aspx?PageID=535>. [p1]
- A. Legin, A. Rudnitskaya, L. Lvova, Y. Vlasov, C. Di Natale, and A. D'Amico. Evaluation of Italian wine by the electronic tongue: recognition, quantitative analysis and correlation with human sensory perception. *Analytica Chimica Acta*, 484(1):33–44, May 2003. ISSN 00032670. doi: 10.1016/S0003-2670(03)00301-5. URL <http://linkinghub.elsevier.com/retrieve/pii/S0003267003003015>. [p1]
- R. Teranishi, E. L. Wick, and I. Hornstein, editors. *Flavor chemistry: thirty years of progress*. Kluwer Academic/Plenum Publishers, New York, 1999. ISBN 9780306461996. [p1]
- UCI Wine Data Set. Uci machine learning repository: wine quality data set. URL <http://archive.ics.uci.edu/ml/datasets/Wine+Quality>. [p1]

Note from the Authors

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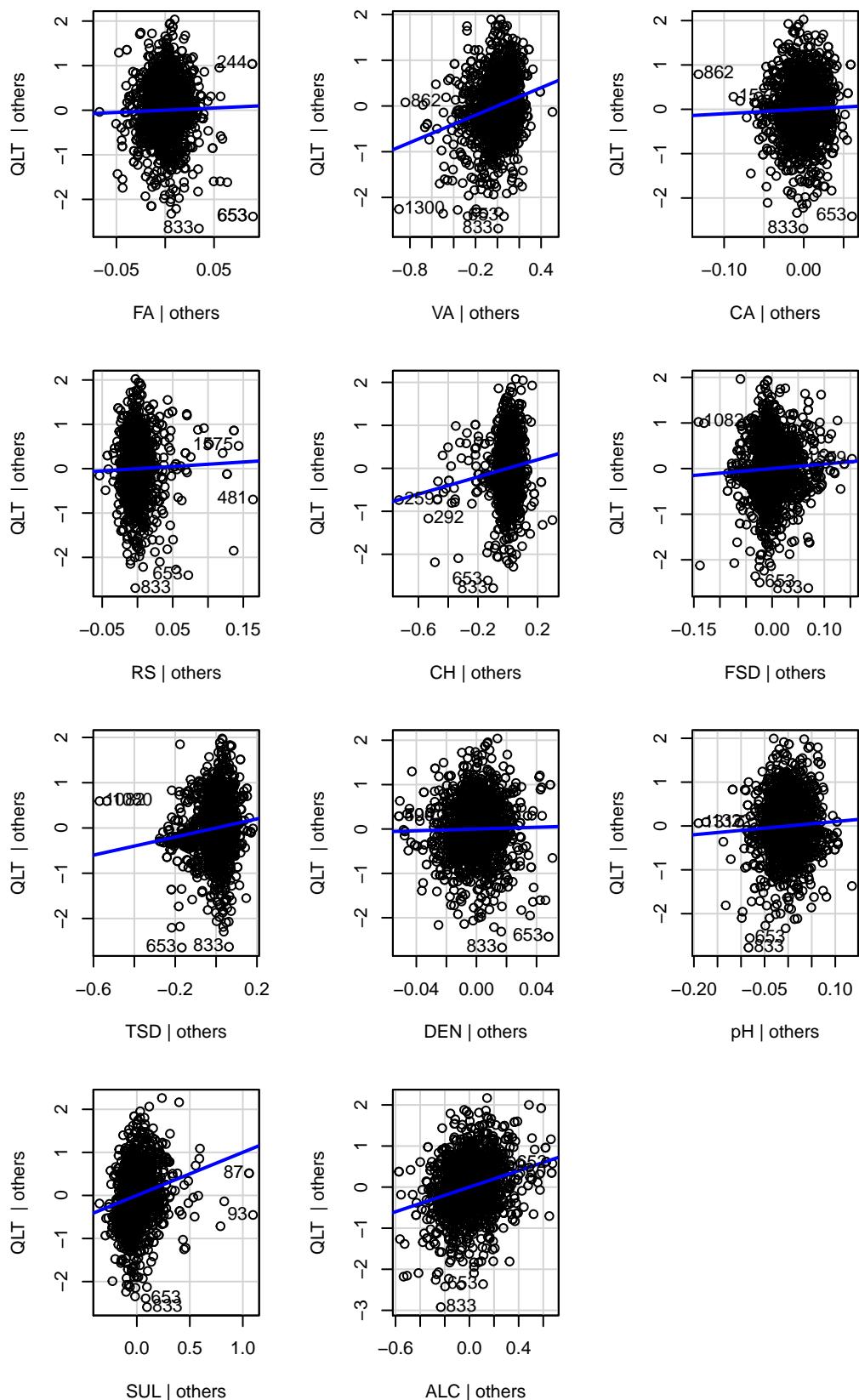


Figure 6: Red Wines - Leverage Plots

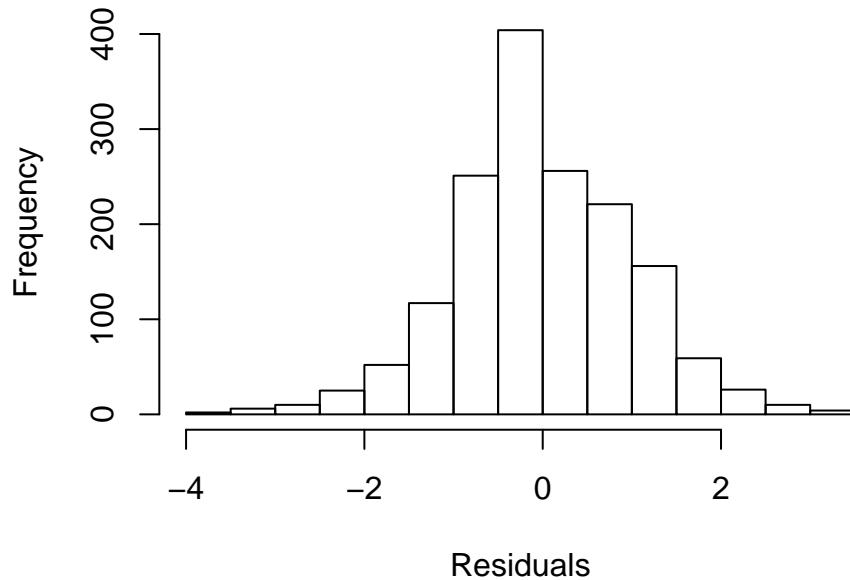


Figure 7: Histogram of residuals after correcting the Skewness

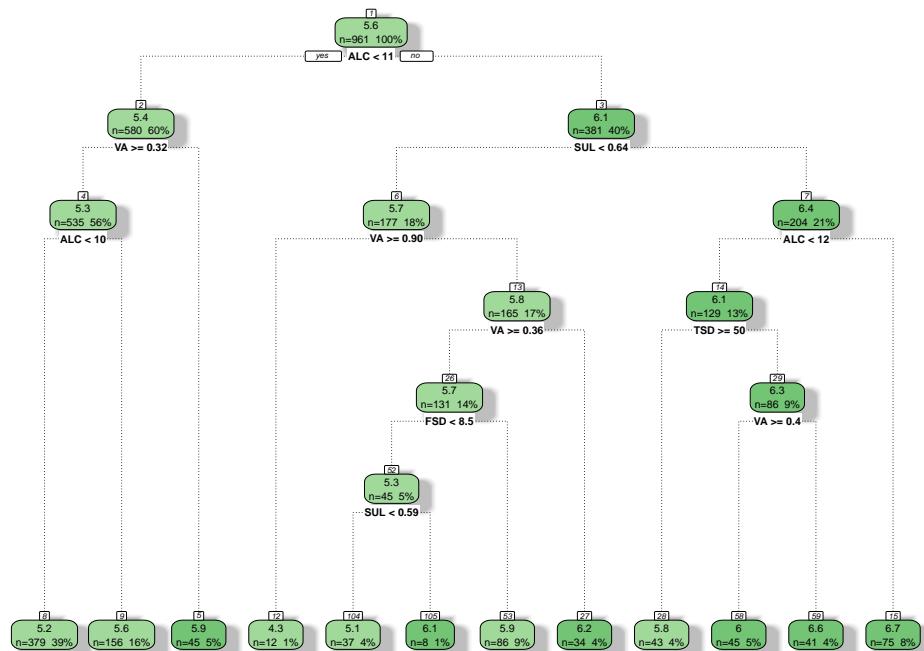


Figure 8: Regression Tree Diagram

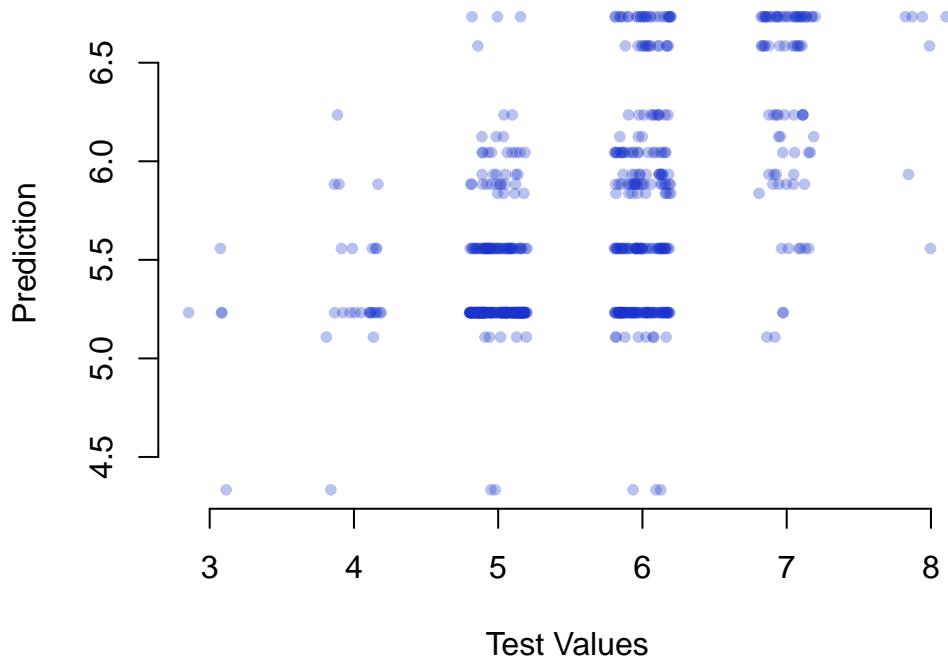


Figure 9: Regression Tree Prediction

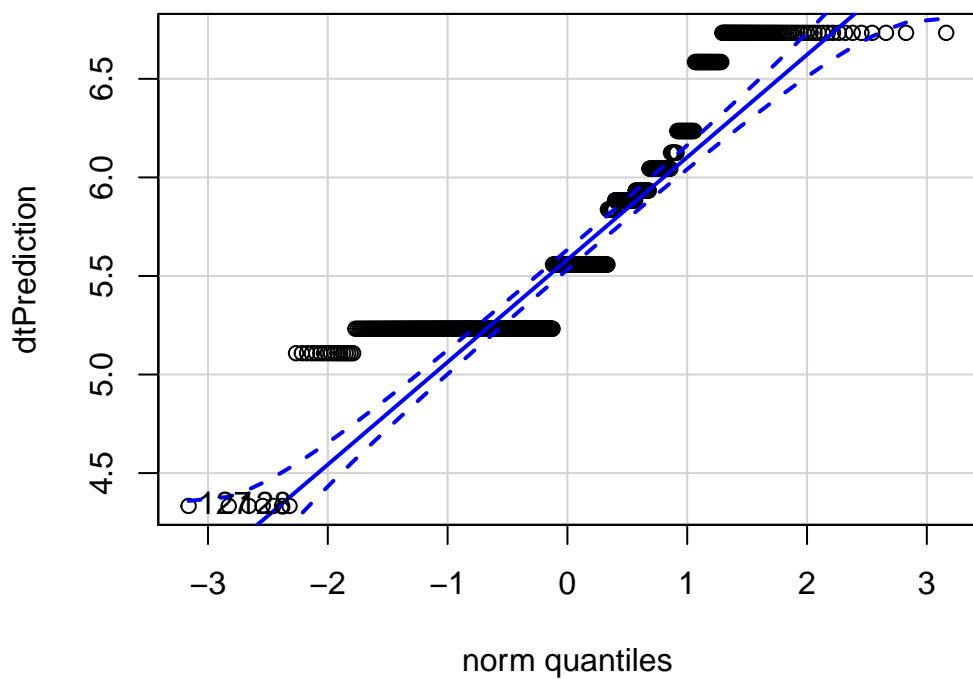
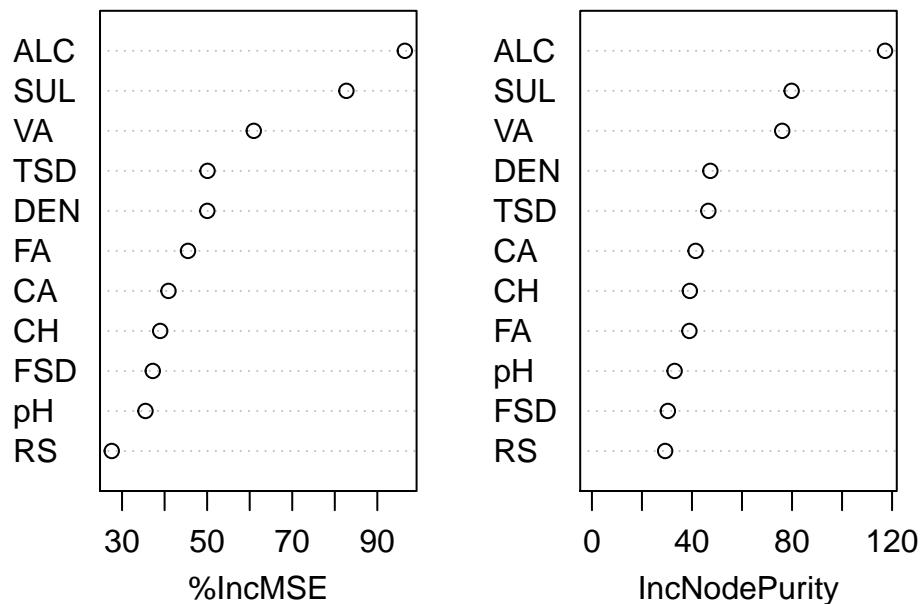
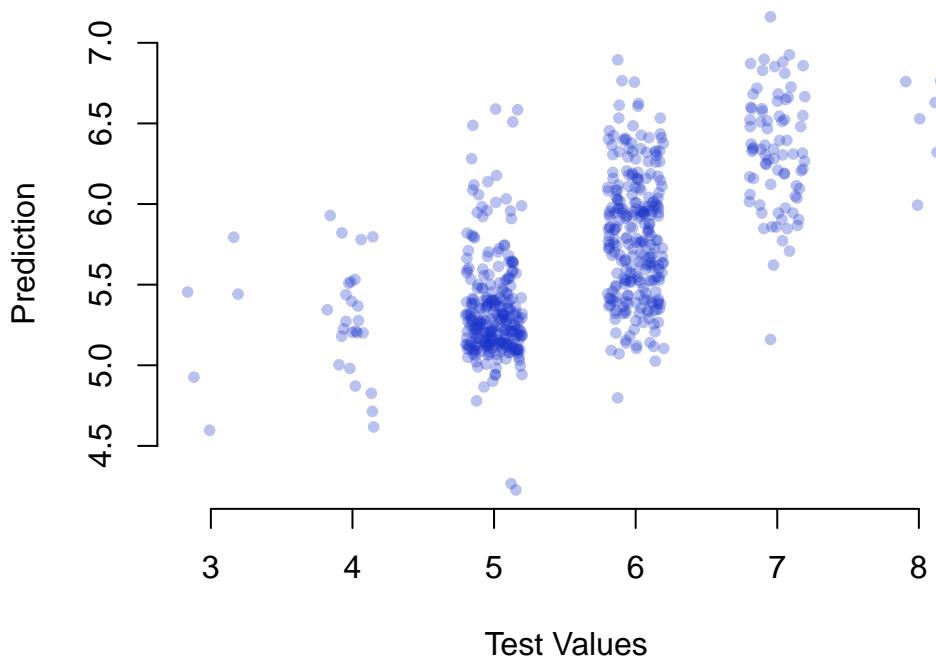


Figure 10: Regression Tree Prediction QQ Plot

**Figure 11:** Importance of the dataset attributes for the prediction of the 'class' attribute**Figure 12:** Random Forest Prediction

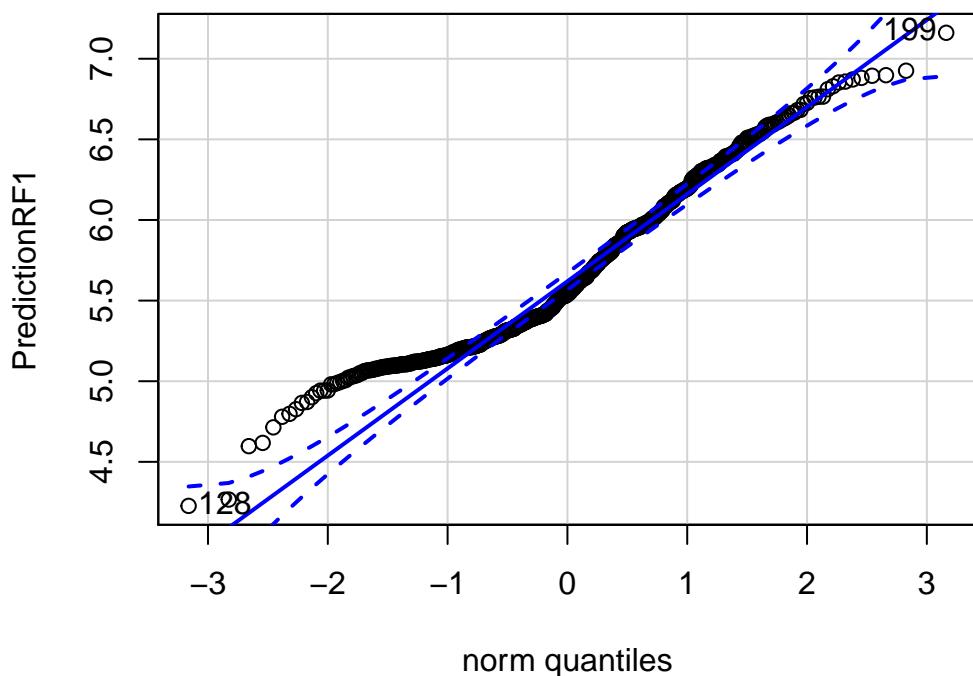


Figure 13: Random Forest Prediction QQ Plot