

Assesment_R_Programming_Basics

Igor Ruiz de los Mozos

Assessment: biomed_data.csv Exploration & Analysis

Instructions:

- Create a new R script or R Markdown document named `STUDENT_NAME_biomed_analysis.Rmd`. This could be a Word transformed to PDF but I encourage you to learn markdown.
 - For each question below, include (a) the R code you used, (b) any plots or summary tables, and (c) a brief written interpretation (2–4 sentences).
 - Knit your R Markdown to HTML or PDF and submit the rendered document.
-

1. Linear Modeling & Interpretation

0. Prepare enviroment and load dataset

```
# Set your working directory once per session  
getwd()  
  
[1] "/home/rstudio"  
  
setwd("/home/rstudio")  
  
# Install required libraries once:  
install.packages(c("tidyverse", "viridis", "GGally"))
```

```
Installing packages into '/usr/local/lib/R/site-library'  
(as 'lib' is unspecified)
```

```
# Load & packages
library(readr)
library(dplyr)
```

Attaching package: 'dplyr'

The following objects are masked from 'package:stats':

filter, lag

The following objects are masked from 'package:base':

intersect, setdiff, setequal, union

```
library(tidyr)
library(ggplot2)
library(GGally)
```

Registered S3 method overwritten by 'GGally':

method from
+.gg ggplot2

```
# Load & Inspect
biomed <- read_csv("/home/rstudio/biomed_data.csv", show_col_types = FALSE)
glimpse(biomed)      # columns & types
```

Rows: 200

Columns: 14

```
$ patient_id <chr> "P1", "P2", "P3", "P4", "P5", "P6", "P7", "P8", "P9", "P10"~
$ sex          <chr> "Female", "Female", "Female", "Female", "Male", "Female", "~
$ age          <dbl> 64, 27, 32, 43, 28, 44, 43, 31, 56, 41, 49, 59, 26, 61, 71,~
$ group         <chr> "Treated", "Treated", "Treated", "Control", "Control", "Con~
$ region        <chr> "North", "North", "North", "South", "South", "North", "Sout~
$ dose          <chr> "Low", "High", "High", "Low", "Medium", "High", "High", "Me~
$ marker_A      <dbl> 5.64, 3.61, 3.87, 3.86, 4.31, 5.17, 5.59, 4.18, 2.14, 5.95,~
$ marker_B      <dbl> 14.07, 11.44, 11.98, 13.23, 13.29, 10.17, 14.07, 8.11, 7.78~
$ marker_C      <dbl> 49.52, 46.27, 51.05, 45.76, 53.96, 50.15, 53.02, 45.03, 48.~
$ marker_D      <dbl> 1.23, 1.81, 1.91, 1.12, 0.99, 1.24, 1.54, 1.31, 0.79, 1.04,~
```

```

$ marker_E    <dbl> 11.38, 11.62, 10.65, 10.73, 9.75, 8.69, 10.27, 10.29, 9.00, ~
$ expression <dbl> 8.66, 12.54, 11.99, 11.27, 8.67, 10.80, 8.90, 8.97, 6.92, 1~
$ heart_rate <dbl> 69, 88, 84, 64, 91, 100, 64, 85, 92, 91, 99, 99, 94, 94, 63~
$ RBC_count   <dbl> 4.47, 4.38, 3.99, 4.02, 5.32, 4.45, 4.73, 4.74, 4.95, 4.41, ~

summary(biomed)      # summary stats & NAs

  patient_id          sex           age        group
Length:200      Length:200      Min.   :25.00  Length:200
Class :character Class :character 1st Qu.:38.00  Class :character
Mode  :character Mode  :character Median :49.00  Mode  :character
                           Mean   :50.09
                           3rd Qu.:61.25
                           Max.   :75.00

  region            dose         marker_A     marker_B
Length:200      Length:200      Min.   :2.140  Min.   : 6.72
Class :character Class :character 1st Qu.:4.300  1st Qu.:11.10
Mode  :character Mode  :character Median :4.900  Median :12.58
                           Mean   :4.896  Mean   :12.43
                           3rd Qu.:5.525  3rd Qu.:13.89
                           Max.   :7.350  Max.   :18.09
                           NA's   :5

  marker_C          marker_D     marker_E     expression
Min.   :35.35  Min.   :0.1400  Min.   : 5.850  Min.   : 4.000
1st Qu.:46.63  1st Qu.:0.8875  1st Qu.: 9.265  1st Qu.: 7.280
Median :49.93  Median :1.1200  Median :10.325  Median : 8.680
Mean   :49.89  Mean   :1.1838  Mean   :10.183  Mean   : 8.876
3rd Qu.:52.98  3rd Qu.:1.3550  3rd Qu.:11.110  3rd Qu.:10.340
Max.   :62.26  Max.   :2.9400  Max.   :15.040  Max.   :16.640
NA's   :5                  NA's   :5

  heart_rate       RBC_count
Min.   : 57.00  Min.   :3.550
1st Qu.: 67.00  1st Qu.:4.265
Median : 77.00  Median :4.550
Mean   : 78.89  Mean   :4.554
3rd Qu.: 91.00  3rd Qu.:4.860
Max.   :103.00  Max.   :5.490
NA's   :5

```

1. Fit a multiple linear regression

```
lm_fit <- lm(expression ~ marker_B + dose + group, data = biomed)
summary(lm_fit)
```

Call:

```
lm(formula = expression ~ marker_B + dose + group, data = biomed)
```

Residuals:

Min	1Q	Median	3Q	Max
-4.7909	-1.3004	0.0164	1.4084	6.0033

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)
(Intercept)	8.94424	0.87530	10.218	< 2e-16 ***
marker_B	0.04227	0.06624	0.638	0.524086
doseLow	-1.50361	0.35446	-4.242	3.46e-05 ***
doseMedium	-2.10358	0.35634	-5.903	1.61e-08 ***
groupTreated	1.15258	0.29319	3.931	0.000118 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 2.039 on 190 degrees of freedom

(5 observations deleted due to missingness)

Multiple R-squared: 0.2251, Adjusted R-squared: 0.2088

F-statistic: 13.8 on 4 and 190 DF, p-value: 6.719e-10

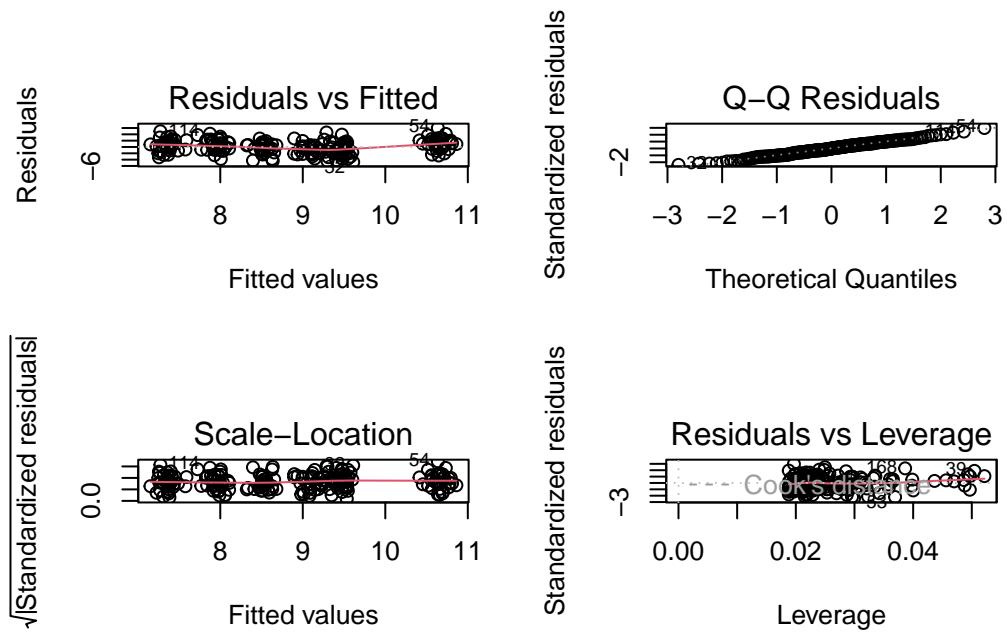
Report the estimated coefficient for marker_B, doseMedium, doseHigh, and groupTreated.

Interpret each coefficient in the context of how expression changes per unit increase in marker_B and between dose/group categories (holding other variables constant).

Assess model assumptions

Produce the 4 base R diagnostic plots:

```
par(mfrow = c(2,2))
plot(lm_fit)
```



Identify any potential outliers or non-constant variance. Which plot indicates this, and what would you do next?

2. Two-Way ANOVA & Interaction

Fit a two-way ANOVA

```
aov_fit <- aov(expression ~ group * dose, data = biomed)
summary(aov_fit)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)						
group	1	68.2	68.20	19.68	1.55e-05 ***						
dose	2	159.5	79.74	23.01	1.14e-09 ***						
group:dose	2	136.1	68.07	19.64	1.78e-08 ***						
Residuals	189	655.1	3.47								

Signif. codes:	0	'***'	0.001	'**'	0.01	'*'	0.05	'.'	0.1	' '	1
5 observations deleted due to missingness											

Report the F-value and p-value for the group:dose interaction term.

Interpret whether there is a statistically significant interaction between treatment group and dose on expression.

Post-hoc comparisons (optional, bonus)

Use TukeyHSD(aov_fit) to examine pairwise differences. Which combinations of group × dose differ significantly?

3. Missing-Value Imputation & Re-visualisation

Impute marker_C with its median

```
med_C <- median(biomed$marker_C, na.rm = TRUE)
biomed_imputed <- biomed %>%
  mutate(marker_C = ifelse(is.na(marker_C), med_C, marker_C))
```

Re-plot the histogram of marker_C before and after imputation side by side (use gridExtra::grid.arrange or patchwork).

Comment on how the distribution changed.

4. New Factor & Comparative Boxplots

Create an age_group factor

```
biomed2 <- biomed %>%
  mutate(age_group = cut(age,
    breaks = c(-Inf, 35, 60, Inf),
    labels = c("<35", "35-60", ">60")))
```

Boxplot heart_rate by age_group with points overlaid (geom_jitter).

Describe any trends you observe (e.g., does heart rate vary by age group?).

5. Tidying & Faceted Violin Plots (Challenge)

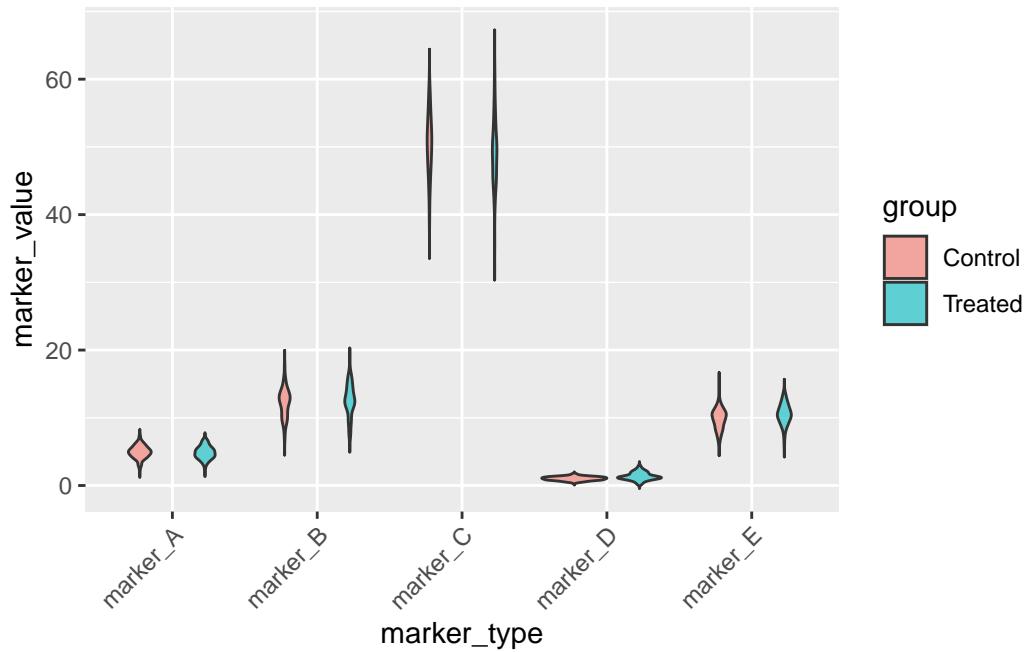
Pivot all marker_ columns longer

```
biomed_long <- biomed %>%
  pivot_longer(cols = starts_with("marker_"),
    names_to = "marker_type",
    values_to = "marker_value")
```

Create violin plot of marker_value by marker_type, colored by group.

```
ggplot(biomed_long, aes(marker_type, marker_value, fill = group)) +
  geom_violin(alpha = .6, trim = FALSE) +
  theme(axis.text.x = element_text(angle = 45, hjust = 1))
```

Warning: Removed 10 rows containing non-finite outside the scale range
(`stat_ydensity()`).



Explain which markers show the largest differences between Control and Treated.

6. Extended Exploratory Questions for biomed_data.csv

Use these prompts to guide deeper, self-driven investigations of the mock cohort. Pick a handful for your project or discussion. *Choose at least five of these prompts to pursue in your final report. For each, provide code, output (tables/plots), and a brief biological or statistical interpretation.*

1. Multivariate Correlations

- Which pairs of biomarkers (A–E) show the strongest positive or negative correlations? How does this change when stratified by treatment group or dose?

2. Predicting Expression

- Build a multiple regression (or regularized model) predicting `expression` from all five `marker_*` variables plus `age` and `sex`. Which predictors are most important?

3. Dose–Response Curves

- For each `group` (Control vs Treated), plot the mean \pm standard error of `marker_D` across `dose` levels. Do dose–response curves diverge between groups?

4. Age Effects and Interactions

- Create an `age_band` (< 35 , $35\text{--}60$, > 60) and test whether the effect of `dose` on `expression` differs across age bands (three-way interaction: `expression ~ group * dose * age_band`).

5. Regional Variation

- Compare biomarker distributions among the four `regions` using box-plots and one-way ANOVA. Which region shows the highest mean `marker_C`, and is it statistically different?

6. Missing-Data Sensitivity

- Impute missing values in `marker_A` and `marker_C` using median, mean, and k-nearest neighbors. Recompute a key summary (e.g., mean of each marker by `group`) and describe how results vary by imputation method.

7. Patient Clustering

- Perform hierarchical clustering or k-means on the numeric biomarkers. How many clusters “make sense”? Do clusters align with `group`, `dose`, or `region`?

8. Principal Component Analysis (PCA)

- Run PCA on the five `marker_*` columns. Plot PC1 vs PC2, coloured by `group` and sized by `dose`. What biological patterns emerge on the principal component axes?

9. Outlier Investigation

- Identify patients whose `expression` lies > 3 SDs from the mean. Examine their full profiles—do they share any common characteristics (age, group, dose, region)?

10. Time-to-Event Simulation (*Bonus*)

- Assume `expression` is a surrogate for time to clinical response. Simulate a binary outcome (`response = expression > threshold`) and fit a logistic regression. Interpret odds ratios for `marker_B` and `dose`.

11. Model Diagnostics

- For your `lm(expression ~ marker_B + dose + group)` model, generate residual vs fitted, QQ, and Cook’s distance plots. Which observations drive the model fit?

12. Interaction Visualization

- Use `ggplot2::geom_interaction()` or custom line plots to visualize any significant three-way interactions (e.g., `marker_E ~ group * dose * sex`).

13. Heatmap of Biomarker Z-Scores

- Standardize each `marker_*` to Z-scores and plot a patient-by-marker heatmap (rows = patients, columns = markers). Cluster both dimensions to reveal co-expression modules.

14. Effect of Sex

- Test whether the `group:dose` interaction on `expression` differs between `sex` by fitting separate models or including a three-way interaction: `expression ~ group * dose * sex`. Summarize the findings.

15. Reproducible Reporting

- Package your entire analysis in an R Markdown report with clear section headers, narrative interpretations, and embedded code: set `code-copy: true` in the YAML and ensure every plot is accompanied by a concise caption.

16. Correlation heatmap

- Compute the correlation matrix for all numeric columns (`marker_A`-`marker_E`, `expression`, `heart_rate`, `RBC_count`). Plot it as a heatmap (`geom_tile`) with a diverging color scale. Identify the strongest positive and negative correlations and speculate on their biological meaning.

17. Group-wise summary table

- Produce a summary table showing, for each region, the mean and standard deviation of `expression` and `heart_rate`. Display it in R as a nicely formatted tibble, and export to CSV.

18. Outlier detection

- For each numeric variable, compute how many observations lie more than 3 standard deviations from the mean. Discuss whether these outliers should be retained or investigated further.

Submission Checklist

R Markdown file (`biomed_analysis.Rmd`) with answers, code, and narrative.

Rendered HTML/PDF output with plots and tables.

Any export files (e.g., `region_summary.csv`) if requested.