# Package 'SpaTalk'

May 26, 2022

```
Title Infer Cell-Cell Communication for Spatial Transcriptomics
Version 1.0
Depends R (>= 4.0.0), ggalluvial, doParallel
Description This package can infer cell-cell communication for spatial transcriptomics.
License GPL (>= 3)
Encoding UTF-8
LazyData true
Roxygen list(markdown = TRUE)
RoxygenNote 7.1.2
Imports Seurat (>= 3.0.0),
      methods,
      reshape2,
      Matrix,
      NNLM,
      stringr,
      progress,
      stats,
      ggplot2,
     ggExtra,
     crayon,
      ggpubr,
      grDevices,
      scales,
      scatterpie,
      pheatmap,
     corrplot,
      circlize,
      ggraph,
      ggrepel,
      igraph,
      foreach,
      parallel,
      iterators
Suggests rmarkdown,
      knitr,
      prettydoc
```

VignetteBuilder knitr

2 createSpaTalk

# R topics documented:

createSpaTalk	. 2
dec_cci	. 3
dec_cci_all	. 4
dec_celltype	. 5
demo_dec_result	. 6
demo_geneinfo	. 7
demo_lrpairs	. 7
demo_pathways	. 8
demo_sc_data	
demo_st_data	. 9
demo_st_meta	. 9
demo_st_sc_data	. 10
demo_st_sc_meta	. 10
find_lr_path	. 11
geneinfo	. 11
generate_spot	. 12
get_lr_path	. 12
lrpairs	. 13
pathways	. 13
plot_ccdist	. 14
plot_cci_lrpairs	. 15
plot_lrpair	. 15
plot_lrpair_vln	. 16
plot_lr_path	. 17
plot_path2gene	. 18
plot_st_celltype	. 19
plot_st_celltype_all	. 19
plot_st_celltype_density	. 20
plot_st_celltype_percent	. 21
plot_st_cor_heatmap	
plot_st_gene	. 22
plot_st_pie	. 23
plot_st_pie_generate	. 24
rev_gene	. 24
set_expected_cell	. 25
SpaTalk	. 25
	26

createSpaTalk

SpaTalk object

# Description

create SpaTalk object using spatial transcriptomics data.

# Usage

Index

```
createSpaTalk(st\_data,\ st\_meta,\ species,\ if\_st\_is\_sc,\ spot\_max\_cell)
```

dec\_cci 3

#### **Arguments**

st_data	A data frame or matrix or dgCMatrix containing counts of spatial transcriptomics, each column representing a spot or a cell, each row representing a gene.
st_meta	A data frame containing coordinate of spatial transcriptomics with three columns, namely 'spot', 'x', 'y' for spot-based spatial transcriptomics data or 'cell', 'x', 'y' for single-cell spatial transcriptomics data.
species	$Acharactermeaningspeciesofthespatialtranscriptomicsdata.{}^{\prime}Human{}^{\prime}or{}^{\prime}Mouse{}^{\prime}.$
if_st_is_sc	A logical meaning if it is single-cell spatial transcriptomics data. TRUE is FALSE.
spot_max_cell	A integer meaning max cell number for each plot to predict. If if_st_sc is FALSE, please determine the spot_max_cell. For 10X (55um), we recommend 30. For Slide-seq, we recommend 1.

#### Value

SpaTalk object

dec_cci	Decomposing cell-cell communications for spatial transciptomics data	
---------	--	--

# Description

Identify the cell-cell communications for single-cell or spot-based spatial transciptomics data with proximal ligand-receptor-target interactions.

# Usage

```
dec_cci(
  object,
  celltype_sender,
  celltype_receiver,
  n_neighbor = 10,
  min_pairs = 5,
  min_pairs_ratio = 0,
  per_num = 1000,
  pvalue = 0.05,
  co_exp_ratio = 0.1,
  if_doParallel = T,
  use_n_cores = NULL
)
```

# Arguments

```
object SpaTalk object after find_lr_path.

celltype_sender

Name of celltype_sender.

celltype_receiver

Name of celltype_receiver.

n_neighbor Number of neighbor cells to select as the proximal cell-cell pair. Default is 10.
```

dec\_cci\_all

min_pairs	Min proximal cell-cell pairs between for sending and receiving cell types. Default is 5.	
min_pairs_ratio		
	Min proximal cell-cell pairs ratio between for sending and receiving cell types. Default is 0.	
per_num	Number of repeat times for permutation test. Default is 1000.	
pvalue	Include the significantly proximal LR pairs with this cutoff of p value from permutation test. Default is $0.05$ .	
co_exp_ratio	Min cell ratio in receiving cells with co-expressed source and target genes for predicting the downstream pathway activity.	
if_doParallel	Use doParallel. Default is TRUE.	
use_n_cores	Number of CPU cores to use. Default is all cores - 2.	

#### Value

SpaTalk object containing the inferred LR pairs and pathways.

-	Decomposing cell-cell ata	communications for	spatial transciptomics
---	------------------------------	--------------------	------------------------

#### **Description**

Identify the all cell-cell communications for single-cell or spot-based spatial transciptomics data with proximal ligand-receptor-target interactions.

### Usage

```
dec_cci_all(
  object,
  n_neighbor = 10,
  min_pairs = 5,
  min_pairs_ratio = 0,
  per_num = 1000,
  pvalue = 0.05,
  co_exp_ratio = 0.1,
  if_doParallel = T,
  use_n_cores = NULL
)
```

# Arguments

 $object \hspace{30pt} SpaTalk \hspace{30pt} object \hspace{30pt} after \hspace{30pt} find\_lr\_path.$ 

n\_neighbor Number of neighbor cells to select as the proximal cell-cell pair. Default is 10.

min\_pairs Min proximal cell-cell pairs between for sending and receiving cell types. De-

fault is 5.

min\_pairs\_ratio

Min proximal cell-cell pairs ratio between for sending and receiving cell types. Default is  $\emptyset$ .

dec\_celltype 5

per_num	Number of repeat times for permutation test. Default is 1000.
pvalue	Include the significantly proximal LR pairs with this cutoff of p value from permutation test. Default is $0.05$ .
co_exp_ratio	Min cell ratio in receiving cells with co-expressed source and target genes for predicting the downstream pathway activity.
if_doParallel	Use doParallel. Default is TRUE.
use_n_cores	Number of CPU cores to use. Default is all cores - 2.

#### Value

SpaTalk object containing the inferred LR pairs and pathways.

## Description

Identify the cellular composition for single-cell or spot-based spatial transcriptomics data with non-negative regression.

#### Usage

```
dec_celltype(
  object,
  sc_data,
  sc_celltype,
  min_percent = 0.5,
  min_nFeatures = 10,
  if_use_normalize_data = T,
  if_use_hvg = F,
  if_doParallel = T,
  use_n_cores = NULL,
  iter_num = 1000,
  method = 1,
  env = "base",
  anaconda_path = "~/anaconda3",
  dec_result = NULL
)
```

### **Arguments**

object	SpaTalk object generated from createSpaTalk.
sc_data	A A data frame or matrix or dgCMatrix containing counts of single-cell RNA-seq data as the reference, each column representing a cell, each row representing a gene.
sc_celltype	A character containing the cell type of the reference single-cell RNA-seq data.
min_percent	Min percent to predict new cell type for single-cell st_data or predict new cell for spot-based st_data. Default is 0.5.

6 demo\_dec\_result

 $\verb|min_nFeatures| & Min number of expressed features/genes for each spot/cell in \verb|st_data|. Default| \\$ 

1s 10.

if\_use\_normalize\_data

Whether to use normalized  $st_data$  and  $sc_data$  with Seurat normalization. Default is TRUE. set it FALSE when the  $st_data$  and  $sc_data$  are already normal-

ized matrix with other methods.

if\_use\_hvg Whether to use highly variable genes for non-negative regression. Default is

FALSE.

if\_doParallel Use doParallel. Default is TRUE.

use\_n\_cores Number of CPU cores to use. Default is all cores - 2.

iter\_num Number of iteration to generate the single-cell data for spot-based data. Default

is 1000.

method 1 means using the SpaTalk deconvolution method, 2 means using RCTD, 3

means using Seurat, 4 means using SPOTlight, 5 means using deconvSeq, 6

means using stereoscope, 7 means using cell2location

env When method set to 6, namely use stereoscope python package to deconvolute,

please define the python environment of installed stereoscope. Default is the

'base' environment. Anaconda is recommended.

anaconda\_path When use python package, please define the path to anaconda, default is ~/ana-

conda3

dec\_result A matrix of deconvolution result from other upcoming methods, row repre-

sents spots or cells, column represents cell types of scRNA-seq reference. See

demo\_dec\_result

#### Value

SpaTalk object containing the decomposing results.

demo\_dec\_result

Demo data of dec\_result

#### **Description**

Demo data of dec\_result

#### Usage

demo\_dec\_result()

#### **Details**

dec\_result used in dec\_celltype must be a matrix object, each row representing a spot, each column representing a cell type.

#### Value

A matrix.

```
dec_result_demo <- demo_dec_result()</pre>
```

demo\_geneinfo 7

 ${\tt demo\_geneinfo}$ 

Demo data of geneinfo

# Description

Demo data of geneinfo

# Usage

```
demo_geneinfo()
```

#### **Details**

geneinfo used in dec\_celltype must be a data.frame object with three columns, namely 'symbol',
'synonyms', 'species'.

# **Examples**

```
geneinfo_demo <- demo_geneinfo()</pre>
```

demo\_lrpairs

Demo data of lrpairs

# Description

Demo data of Irpairs

### Usage

```
demo_lrpairs()
```

#### **Details**

```
lrpairs used in dec_cci must be a data.frame object with three columns, namely 'ligand',
'receptor', 'species'.
```

#### Value

A data.frame.

```
lrpairs_demo <- demo_lrpairs()</pre>
```

8 demo\_sc\_data

demo\_pathways

Demo data of pathways

### Description

Demo data of pathways

#### Usage

```
demo_pathways()
```

#### **Details**

```
pathways used in dec_cci must be a data.frame object with seven columns, namely 'src', 'dest', 'pathway', 'source', 'type', 'src_tf', 'dest_tf', 'species'.
```

#### Value

A data.frame.

#### **Examples**

```
pathways_demo <- demo_pathways()</pre>
```

demo\_sc\_data

Demo data of sc\_data

# Description

Demo data of sc\_data.

#### Usage

```
demo_sc_data()
```

### **Details**

sc\_data used in dec\_celltype must be a matrix object, each column representing a cell, each row representing a gene.

#### Value

A matrix.

```
sc_data_demo <- demo_sc_data()</pre>
```

demo\_st\_data 9

demo\_st\_data

Demo data of st\_data

### Description

Demo data of st\_data.

### Usage

```
demo_st_data()
```

#### **Details**

st\_data used in dec\_celltype must be a matrix object, each column representing a spot, each row representing a gene.

#### Value

A matrix.

#### **Examples**

```
st_data_demo <- demo_st_data()</pre>
```

demo\_st\_meta

Demo data of st\_meta

#### **Description**

Demo data of st\_meta

#### Usage

```
demo_st_meta()
```

### **Details**

st\_meta used in dec\_celltype must be a data.frame object with three columns, namely 'spot', 'x', 'y' for spot-based spatial transcriptomics data.

#### Value

A data.frame.

```
st_meta_demo <- demo_st_meta()</pre>
```

10 demo\_st\_sc\_meta

demo\_st\_sc\_data

Demo data of single-cell st\_data

### Description

Demo data of single-cell st\_data.

#### Usage

```
demo_st_sc_data()
```

#### **Details**

st\_data used in dec\_celltype must be a matrix object, each column representing a cell, each row representing a gene.

#### Value

A matrix.

#### **Examples**

```
st_data_demo <- demo_st_sc_data()</pre>
```

demo\_st\_sc\_meta

Demo data of st\_sc\_meta

#### **Description**

Demo data of st\_sc\_meta

#### Usage

```
demo_st_sc_meta()
```

### **Details**

st\_sc\_meta used in dec\_celltype must be a data.frame object with three columns, namely 'cell', 'x', 'y' for single-cell spatial transcriptomics data.

#### Value

A data.frame.

```
st_sc_meta_demo <- demo_st_sc_meta()</pre>
```

find\_lr\_path

find_lr_path Find lrpairs and pathways
--

# Description

Find 1rpairs and pathways with receptors having downstream targets and transcriptional factors.

#### Usage

```
find_lr_path(
  object,
  lrpairs,
  pathways,
  max_hop = NULL,
  if_doParallel = T,
  use_n_cores = NULL
)
```

### Arguments

object	SpaTalk object generated from dec_celltype.
lrpairs	A data.frame of the system data containing ligand-receptor pairs of 'Human' and 'Mouse' from CellTalkDB.
pathways	A data.frame of the system data containing gene-gene interactions and pathways from KEGG and Reactome as well as the information of transcriptional factors.
max_hop	Max hop from the receptor to the downstream target transcriptional factor to find for receiving cells. Default is 3 for human and 4 for mouse.
if_doParallel	Use doParallel. Default is TRUE.
use_n_cores	Number of CPU cores to use. Default is all cores - 2.

### Value

SpaTalk object containing the filtered lrpairs and pathways.

|--|--|

# Description

Gene symbols of 'Human' and 'Mouse' updated on June 30, 2021 for revising count matrix.

### Usage

geneinfo

#### **Format**

An object of class data. frame with 250934 rows and 3 columns.

12 get\_lr\_path

#### **Source**

https://www.ncbi.nlm.nih.gov/gene

generate\_spot Generate pseudo spot st\_data

### Description

Generate pseudo spot st\_data with single-cell st\_data

#### Usage

```
generate_spot(st_data, st_meta, x_min, x_res, x_max, y_min, y_res, y_max)
```

#### **Arguments**

st_data	A data frame or matrix or dgCMatrix containing counts of spatial transcriptomics, each column representing a cell, each row representing a gene.
st_meta	A data.frame containing coordinate of spatial transcriptomics with three columns, 'cell', 'x', 'y', and celltype.
x_min	Min value of x axis.
x_res	Resolution of x coordinate.
x_max	Max value of x axis.
y_min	Min value of y axis.
y_res	Resolution of y coordinate.
y_max	Max value of y axis.

#### Value

A list of spot st\_data and st\_meta

get\_lr\_path Get LR and downstream pathways

# Description

Get LR and downstream pathways and get p value of receptor-related pathways with LR-target genes by the Fisher-exact test.

# Usage

```
get_lr_path(
  object,
  celltype_sender,
  celltype_receiver,
  ligand,
  receptor,
  min_gene_num = 5
)
```

13

#### **Arguments**

object SpaTalk object generated from dec\_cci.

celltype\_sender

Name of celltype\_sender.

celltype\_receiver

Name of celltype\_receiver.

ligand Name of ligand from celltype\_sender.

receptor Name of receptor from celltype\_receiver.

min\_gene\_num Min genes number for each pathway.

#### Value

A list containing two data.frame. One is LR and downstream pathways, another is the p value of receptor-related pathways with LR-target genes.

**lrpairs** 

lrpairs

## **Description**

Ligand-receptor pairs of 'Human' and 'Mouse' containing 3398 human and 2033 mouse pairs.

### Usage

lrpairs

# **Format**

An object of class data.frame with 5427 rows and 3 columns.

#### Source

http://tcm.zju.edu.cn/celltalkdb/

pathways

pathways

#### **Description**

KEGG pathways and Reactomes of 'Human' and 'Mouse' for intra-cellular genes and transcription factors.

#### Usage

pathways

#### **Format**

An object of class data.frame with 669197 rows and 8 columns.

14 plot\_ccdist

#### Source

```
https://www.genome.jp/kegg/pathway.html
https://reactome.org/
http://bioinfo.life.hust.edu.cn/AnimalTFDB/#!/
```

plot\_ccdist

Plot cell-cell distribution

#### **Description**

Point plot with spatial distribution of celltype\_sender and celltype\_receiver

#### Usage

```
plot_ccdist(
  object,
  celltype_sender,
  celltype_receiver,
  color = NULL,
  size = 1,
  if_plot_others = T,
  if_plot_density = T,
  if_plot_edge = T,
  if_show_arrow = T,
  arrow_length = 0.05,
  plot_cells = NULL
)
```

#### **Arguments**

```
object
                  SpaTalk object generated from dec_celltype.
celltype_sender
                  Name of celltype_sender.
celltype_receiver
                  Name of celltype_receiver.
                  Color for celltype_sender, celltype_receiver, and others. Three values.
color
size
                  Point size. Default is 1.
if_plot_others Whether to plot others. Default is TRUE.
if_plot_density
                  Whether to plot marginal density plots. Default is TRUE.
if_plot_edge
                  Whether to plot edge between neighbors. Default is TRUE.
                  Whether to show the arrow of the plotted edge. Default is TRUE.
if_show_arrow
arrow_length
                  Arrow length.
plot_cells
                  Which cells to plot. Default is all cells. Input a character vector of cell names to
                  plot.
```

plot\_cci\_lrpairs 15

### **Description**

Heatmap with LR pairs of celltype\_sender and celltype\_receiver

### Usage

```
plot_cci_lrpairs(
  object,
  celltype_sender,
  celltype_receiver,
  top_lrpairs = 20,
  color = NULL,
  border_color = "black",
  type = NULL,
  fontsize_number = 1,
  number_color = "black")
```

#### **Arguments**

object SpaTalk object generated from dec\_cci.

celltype\_sender

Name of celltype\_sender.

celltype\_receiver

Name of celltype\_receiver.

top\_lrpairs Number of top lrpairs for plotting. Default is 20.

color Color for the cells in heatmap.

border\_color color of cell borders on heatmap, use NA if no border should be drawn.

type Set 'sig' to plot significant LR pairs or set 'number' to plot the number of spatial

LR interactions.

fontsize\_number

fontsize of the numbers displayed in cells.

number\_color color of the text.

plot\_lrpair Plot LR pair

## Description

Point plot with LR pair from celltype\_sender to celltype\_receiver

16 plot\_lrpair\_vln

#### Usage

```
plot_lrpair(
  object,
  celltype_sender,
  celltype_receiver,
  ligand,
  receptor,
  color = NULL,
  size = 1,
  if_plot_density = T,
  if_plot_edge = T,
  if_show_arrow = T,
  arrow_length = 0.05,
  plot_cells = NULL
)
```

#### **Arguments**

object SpaTalk object generated from dec\_celltype.

celltype\_sender

Name of celltype\_sender.

celltype\_receiver

Name of celltype\_receiver.

ligand Name of ligand from celltype\_sender.

color Color for ligand, receptor, and others. Three values.

size Point size. Default is 1.

if\_plot\_density

receptor

Whether to plot marginal density plots. Default is TRUE.

if\_plot\_edge Whether to plot edge between neighbors. Default is TRUE.

Name of receptor from celltype\_receiver.

if\_show\_arrow Whether to show the arrow of the plotted edge. Default is TRUE.

arrow\_length Arrow length.

plot\_cells Which cells to plot. Default is all cells. Input a character vector of cell names to

plot.

# **Description**

Violin plot spatial distance of LR pair between expressed senders and receivers and between expressed cell-cell pairs.

plot\_lr\_path 17

#### Usage

```
plot_lrpair_vln(
  object,
  celltype_sender,
  celltype_receiver,
  ligand,
  receptor,
  vln_color = NULL,
  if_plot_boxplot = T,
  box_width = 0.2
)
```

#### **Arguments**

```
object
                  SpaTalk object generated from dec_celltype.
celltype_sender
                  Name of celltype_sender.
celltype_receiver
                  Name of celltype_receiver.
ligand
                  Name of ligand from celltype_sender.
receptor
                  Name of receptor from celltype_receiver.
vln_color
                  Color for violins. Two values.
if_plot_boxplot
                  Whether to plot boxplot. Default is TRUE.
box_width
                  Box width. Default is 0.2.
```

plot\_lr\_path

Plot LR and downstream pathways

#### **Description**

Plot network with LR and downstream pathways

#### Usage

```
plot_lr_path(
  object,
  celltype_sender,
  celltype_receiver,
  ligand,
  receptor,
  color = NULL,
  size = 5,
  arrow_length = 0.1
```

plot\_path2gene

#### **Arguments**

object SpaTalk object generated from dec\_cci.
celltype\_sender
Name of celltype\_sender.
celltype\_receiver
Name of celltype\_receiver.
ligand Name of ligand from celltype\_sender.
receptor Name of receptor from celltype\_receiver.
color Color for points Two values.
size Size of points.

Arrow length.

plot\_path2gene

arrow\_length

River plot of significantly activated pathways and related downstream genes of receptors.

#### **Description**

River plot of significantly activated pathways and related downstream genes of receptors.

#### Usage

```
plot_path2gene(
  object,
  celltype_sender,
  celltype_receiver,
  ligand,
  receptor,
  min_gene_num = 5,
  pvalue = 0.5,
  color = NULL,
  color_flow = "blue"
)
```

#### **Arguments**

object SpaTalk object generated from dec\_cci.

celltype\_sender

Name of celltype\_sender.

celltype\_receiver

Name of celltype\_receiver.

ligand Name of ligand from celltype\_sender.
receptor Name of receptor from celltype\_receiver.
min\_gene\_num Min genes number for each pathway.

pvalue P value of the Fisher-exact test.

color Color of pathways and genes. Two values.

color\_flow Color of the flow.

plot\_st\_celltype 19

## **Description**

Ponit plot with spatial distribution of a single predicted cell type for transcriptomics data

### Usage

```
plot_st_celltype(
  object,
  celltype,
  size = 1,
  color_celltype = "blue",
  color_others = "grey"
)
```

### Arguments

```
object SpaTalk object generated from dec_celltype.

celltype Name of cell type in the sc_celltype.

size Point size. Default is 1.

color_celltype Color for the celltype of interest.

color_others Color for the others.
```

### Description

Plot spatial distribution of all predicted cell types for transcriptomics data

# Usage

```
plot_st_celltype_all(object, size = 1, color = NULL)
```

## Arguments

```
object SpaTalk object generated from dec_celltype.
size Point size. Default is 1.
```

color Color for all predicted cell types.

```
plot_st_celltype_density

Plot spatial density of a single cell type
```

#### **Description**

Plot spatial density of a single predicted cell type for transcriptomics data

#### Usage

```
plot_st_celltype_density(
  object,
  celltype,
  type,
  if_plot_point = T,
  point_color = NULL,
  point_size = 1,
  color_low = "grey",
  color_mid = NULL,
  color_high = "blue",
  color_midpoint = NULL,
  size = 1
)
```

#### **Arguments**

```
object
                  SpaTalk object generated from dec_celltype.
                  Name of cell type in the sc_celltype.
celltype
                  Select 'contour' or 'raster'.
type
                  Whether to plot points when type is 'contour'.
if_plot_point
                  Point color.
point_color
point_size
                  Point size. Default is 1.
color_low
                  Color for the lowest value.
color_mid
                  Color for the middle value for using scale_color_gradient2. Default is NULL.
color_high
                  Color for the highest value.
color_midpoint Value for the middle scale. Default is NULL.
                  Line size when type is 'contour'. Default is 1.
size
```

```
plot_st_celltype_percent
```

Plot spatial distribution of a single cell type percent

#### **Description**

Plot spatial distribution of a single predicted cell type percent for transcriptomics data

#### Usage

```
plot_st_celltype_percent(
  object,
  celltype,
  size = 1,
  color_low = NULL,
  color_mid = NULL,
  color_high = NULL,
  color_midpoint = NULL)
```

#### **Arguments**

```
object SpaTalk object generated from dec_celltype.

celltype Name of cell type in the sc_celltype.

size Point size. Default is 1.

color_low Color for the lowest value.

color_mid Color for the middle value for using scale_color_gradient2. Default is NULL.

color_high Color for the highest value.

color_midpoint Value for the middle scale. Default is NULL.
```

plot\_st\_cor\_heatmap

Plot heatpmap of correlation between marker genes and cell types

## Description

Plot heatpmap of correlation between the expression of marker genes and the predicted score of cell types among all spatial cells or spots.

#### Usage

```
plot_st_cor_heatmap(
  object,
  marker_genes,
  celltypes,
  color_low = NULL,
  color_mid = NULL,
  color_high = NULL,
```

plot\_st\_gene

```
scale = "none",
if_show_top = T,
top_direction = "row",
border_color = NA
)
```

### Arguments

object	SpaTalk object generated from dec_celltype.
marker_genes	A character containing the known marker genes to plot, provide at least two marker genes of interest.
celltypes	A character containing name of cell type in the sc_celltype. Default is to plot all cell types.
color_low	Color for the lowest value.
color_mid	$Color for the \ middle \ value \ for \ using \ \ scale\_color\_gradient 2. \ Default \ is \ \ NULL.$
color_high	Color for the highest value.
scale	Character indicating if the values should be centered and scaled in either the row direction or the column direction, or none. Corresponding values are 'row', 'column' and 'none'.
if_show_top	Whether to plot a symbol to the highest value across rows or columns. Default is TRUE.
top_direction	Direction to identify the highest value, select 'row' or 'column'.
border_color	Color of the cell border. Default is 'NA'.

plot\_st\_gene Plot spatial distribution of gene

# Description

Point plot with spatial distribution of a gene for transcriptomics data

# Usage

```
plot_st_gene(
  object,
  gene,
  size = 1,
  color_low = "grey",
  color_mid = NULL,
  color_high = "blue",
  color_midpoint = NULL,
  if_use_newmeta = T,
  celltype = NULL,
  if_plot_others = T
)
```

plot\_st\_pie 23

#### **Arguments**

object SpaTalk object generated from dec\_celltype.

gene Symbol of gene, e.g., 'AKT1'.

size Point size. Default is 1.

color\_low Color for the lowest value.

color\_mid Color for the middle value for using scale\_color\_gradient2. Default is NULL.

color\_high Color for the highest value.

color\_midpoint Value for the middle scale. Default is NULL.

if\_use\_newmeta Whether to use newmeta o plot the spatial distribution of gene after dec\_celltype for spot-based data. Default is TRUE.

celltype gene in which celltype to plot. Default is NULL. Set Nif\_use\_newmeta TRUE

when using this parameter.

if\_plot\_others Whether to plot other cells when to use defined celltype.

#### **Details**

Please set if\_use\_newmeta as FALSE to plot the spatial distribution of gene before dec\_celltype for spot-based data.

|--|

#### **Description**

Plot scatterpie for spatial transcriptomics data

#### Usage

```
plot_st_pie(object, pie_scale = 1, xy_ratio = 1, color = NULL)
```

### **Arguments**

object SpaTalk object generated from dec\_celltype.

pie\_scale Scale of each pie to plot. Default is 1.

xy\_ratio Ratio of y and x coordinates. Default is 1.

color Filled of colors for pie plot, length of color must be equal to the number of

unique cell types in sc\_celltype.

rev\_gene

# Description

Plot scatterpie for spot-based ST data

### Usage

```
plot_st_pie_generate(st_meta, pie_scale = 1, xy_ratio = 1, color = NULL)
```

### **Arguments**

st_meta	st_meta generated from generate_spot
pie_scale	Scale of each pie to plot. Default is 1.
xy_ratio	Ratio of y and x coordinates. Default is 1.
color	Filled of colors for pie plot, length of color must be equal to the number of unique cell types in sc_celltype.

# Description

Revise genes according to NCBI Gene symbols updated in June 30, 2021 for count matrix, user-custom lrpairs data.frame, and user-custom pathways data.frame.

#### Usage

```
rev_gene(data = NULL, data_type = NULL, species = NULL, geneinfo = NULL)
```

### Arguments

data	A matrix containing count data each column representing a spot or a cell, each row representing a gene; Or a data.frame containing ligand-receptor pairs; Or a data.frame containing gene-gene interactions and pathways from KEGG and Reactome as well as the information of transcriptional factors.
data_type	A character to define the type of data, select 'count' for the data matrix, 'lrpairs' for the data.frame containing lrpairs, 'pathways' for the data.frame containing pathways.
species	Species of the data. 'Human' or 'Mouse'.
geneinfo	A data frame of the system data containing gene symbols of 'Human' and 'Mouse' updated on June 30, 2021 for revising count matrix.

# Value

A new matrix or data.frame.

set\_expected\_cell 25

#### **Description**

Set the expected cell in SpaTalk object

#### Usage

```
set_expected_cell(object, value)
```

#### **Arguments**

object SpaTalk object

value Th number of expected cell for each spot, must be equal to the spot number.

#### Value

SpaTalk object

SpaTalk Definition of 'SpaTalk' class

# Description

An S4 class containing the data, meta, and results of inferred cell type compositions, LR pairs, and pathways.

#### Slots

data A list containing the raw and normalized data.

meta A list containing the raw and new meta data.

para A list containing the parameters.

coef A matrix containing the results of deconvolution.

cellpair A list containing the cell-cell pairs based on the spatial distance.

dist A matrix containing the Euclidean distance among cells.

1rpair A data frame containing the inferred LR pairs.

tf A data frame containing the TFs of receptors.

lr\_path A list containing the lrpairs and pathways.

# **Index**

rev\_gene, 24

```
* datasets
                                                   set_expected_cell, 25
    geneinfo, 11
                                                   SpaTalk, 25
    1rpairs, 13
                                                   SpaTalk-class (SpaTalk), 25
    pathways, 13
createSpaTalk, 2, 5
dec_cci, 3, 7, 8, 13, 15, 18
dec_cci_all, 4
dec_celltype, 5, 6-11, 14, 16, 17, 19-23
demo\_dec\_result, 6, 6
demo_geneinfo, 7
demo_lrpairs, 7
demo_pathways, 8
{\sf demo\_sc\_data}, \textcolor{red}{8}
demo_st_data, 9
demo_st_meta, 9
demo_st_sc_data, 10
demo_st_sc_meta, 10
find_lr_path, 3, 4, 11
geneinfo, 11
generate_spot, 12, 24
get_lr_path, 12
1rpairs, 13
pathways, 13
plot_ccdist, 14
plot_cci_lrpairs, 15
plot_lr_path, 17
plot_lrpair, 15
plot_lrpair_vln, 16
plot_path2gene, 18
plot_st_celltype, 19
plot_st_celltype_all, 19
plot_st_celltype_density, 20
plot_st_celltype_percent, 21
plot_st_cor_heatmap, 21
plot_st_gene, 22
plot_st_pie, 23
plot_st_pie_generate, 24
```