**Abstract**

**Introduction**

For sedentary organisms such as plants, seed dispersal is essential to the movement of populations because it is the only stage in the life cycle that movement can occur. While plant propagules are often dispersed by abiotic vectors such as wind, water, and gravity, they are often dispersed biotically by organisms such as insects, birds, and mammals. For example, the seeds of the mahaleb cherry *Prunus mahaleb* are ingested, moved, and defecated by a variety of mammals such as foxes and badgers as well as birds such as crows, thrushes, warblers, and robins (Herrera and Jordano 1981; Guitián *et al*. 1992; Jordano *et al*. 2007). The acorns of various oak trees (*Quercus* sp.) are often dispersed and cached by a variety of rodents such as mice and squirrels (Jensen and Nielsen 1986; Vander Wall 2001; Gómez *et al*. 2008). Humans are even involved in the process of dispersing plant propagules, as numerous plants can be dispersed when propagules attach to clothing (Wichmann *et al*. 2009; Ansong and Pickering 2014), are caught in air currents generated by passing vehicles (Von Der Lippe *et al*. 2013) or on the vehicles themselves (Veldman and Putz 2010; Taylor *et al*. 2012), as a contaminant of horticultural stock (Hodkinson and Thompson 1997), and as impurities in agricultural produce such as grains (Shimono and Konuma 2008; Michael *et al*. 2010; Wilson *et al*. 2016). The prevalence in ecological literature of studies regarding these sorts of dispersal indicate a growing interest in identifying and quantifying dispersal by organisms such as insects, animals, and humans.

Such dispersal of seeds by biotic vectors is often part of a larger network of parallel and serial dispersal events for any given species. In the *Prunus mahaleb* example listed earlier, while seeds are often consumed directly from the tree by birds before primary dispersal via gravity, they may also be consumed off the ground and undergo secondary dispersal by birds and mammals after primary dispersal via gravity (SOURCE). Integration of these primary and secondary pathways and the vectors responsible into a total dispersal kernel, or probability distribution of how far seeds are dispersed when accounting for all possible dispersal vectors, can help us quantify how plant populations expand at a variety of different spatial scales (Nathan 2007, SOURCE). However significant challenges in identifying these biotic dispersers and quantifying how they disperse seeds make it challenging to fully understand their role in seed dispersal, especially for smaller seeds that cannot be easily tracked through empirical means (Rogers *et al*. 2019).

For these smaller seeds, ants and other insects are often an overlooked source of localised secondary dispersal and are responsible for moving seeds after they have been dispersed by wind, water, gravity, or another biotic dispersal vector (Vander Wall and Longland 2004). In particular, ant-mediated seed dispersal, or myrmecochory, has been shown to be a common occurrence in many plant species. In more than 80 plant families across the world, ant-dispersed plant species contain seeds that bear an elaiosome, a small structure on the seed achene that facilitates dispersal by ants (Edwards *et al*. 2006). The elaiosome has a high lipid content and likely serves as a reward for the ant dispersing the seed (Brew *et al*. 1989; Hughes and Westoby 1990, 1992), as ants will typically take the seeds back to their nest and consume the elaiosome, then store the seed achene in a midden (Berg 1975; Culver and Beattie 1978). This dispersal can be advantageous for a variety of different reasons, including moving propagules to areas with better germination and growing conditions as well as less competition from other plants (Handel and Beattie 1990).

Investigations of such ant-mediated secondary dispersal, as well as the primary dispersal that precedes it, can be possibly complicated by climate change, as climate change may affect various aspects and characteristics of these dispersal processes. For example, elevated temperatures, earlier flowering seasons, and increased CO2 levels can lead to changes in the number of seeds that plant can produce (SOURCES). The dynamics of seed release can also be affected by increased growing temperatures (Teller *et al*. 2016). Seed shape and size can also be affected (SOURCES), which can have implications for both abiotic and biotic dispersal distances. Climate change may cause shifts in the ranges or movement patterns of species that disperse a certain species’ seeds and may thus affect its dispersal capabilities (SOURCE). While there is amount of research addressing the effects of climate change on dispersal, much of it is focused on seed production and abiotic vectors of dispersal, with less research on how climate change may affect the dispersal of seeds by insects, animals, and humans.

Given the role that ants and other insects play in seed dispersal, a better understanding of how they move seeds and how far they do it can help us better understand how plant species dispersed by these insects spread. Such an understanding starts with better understanding seed removal, as this step is the beginning of the dispersal process by ants and other insects. Can it also affect seed nutrient contents and change how attractive they are to insect dispersers? Something about how we can use cafeteria experiments to address this.

Here, we seek to develop a better understanding of how seeds are moved by secondary dispersal vectors, namely ants, with a focus on investigating the first step of this secondary dispersal process: seed removal. We also seek to explore the role of the elaiosome, as well as whether increased growing temperatures on maternal plants, in seed removal rates and overall attractiveness to insect dispersers. We thus investigate three questions. First, what are the approximate rates of seed removal via insect after a seed has experienced primary dispersal? Second, does the seed elaiosome affect the rate of removal? And third, are seeds from maternal plants that experienced increased growing temperatures any more or less likely to be removed by insects than their unwarmed counterparts?

**Materials and Methods**

***Study Species***

*Carduus nutans* L. (“musk thistle” or “nodding thistle”) and *Carduus acanthoides* L. (“plumeless thistle”) are two closely-related invasive thistles in the Asteraceae family. Both species germinate in autumn or spring and bolt in the early summer (Zhang *et al*. 2012), reproduce exclusively by seed, and have monocarpic perennial life cycles that have been demonstrated to shift from biennial towards annual under warming conditions (Keller and Shea, *in press*). While sharing similar life histories, the two species display significant morphological differences in dispersal-related characteristics such as number of flower heads, flower head size, number of seeds produced per flower head, and distribution of flower heads across an individual (Desrochers *et al*. 1988). These invasive thistles have high reproductive potential and are a considerable agricultural pest since they thrive in pastures, are unpalatable to most grazers, and decrease pasture productivity (Trumble and Kok 1982). In addition to pastures, these thistles occur in other highly disturbed areas such as drainages and roadsides. Both species can be found across the U.S. and often co-occur (Allen and Shea 2006), and are listed as noxious weeds in several states (Skinner *et al.* 2000).

Wind serves as the primary dispersal vector in both *C. nutans* and *C. acanthoides*, as seeds of both species display a prominent pappus that increases hang time and makes it possible for them to be carried at long distances. However, dispersal of these seeds also extends beyond wind, with paths of secondary dispersal possible after seeds have hit the ground. Seeds from both species contain elaiosomes that are thought to play a role in ant-mediated dispersal (Pemberton and Irving 1990), and have been documented to be moved by insects and small mammals, with insects such as ants, crickets, and grasshoppers likely playing a significant role in the movement of seeds (Jongejans et al. 2015).

***Seed collection and preparation***

*C. nutans* and *C. acanthoides* from which seeds were harvested were grown under control and ambient warming treatments as part of an experiment in Drees and Shea (*in prep.*) to estimate the effects of warming on the distribution of flower heights; this publication discusses the methods and experimental setup in greater detail. In this experiment, individuals of each species were grown in a grid-like arrangement, and a subset of individuals within each species were randomly assigned a fibreglass open-top chamber. These chambers, built to specifications of the International Tundra Experiment Manual (Molau and Mølgaard 1996), have been demonstrated to increase the ambient temperature by approximately 0.6 °C (Zhang *et al*. 2011) without significantly affecting other possible growth factors such as soil moisture and snow depth. When flowers set seed, mesh pollen bags were wrapped around the flowers endure that seeds from these invasive thistles did not escape and contaminate other parts of the study area, as seeds can stay dormant for years and could pose problems for future experiments once germination occurs.

Once individuals completed their life cycle or collapsed under their own weight, they were cut down, and bagged seed heads were collected from all individuals that produced five or more viable flowers. During this process, seed heads from all individuals of the same species were mixed together to randomise possible differences in seed size, shape, nutrient content, and any other factors that vary between individuals and may affect seed attractiveness to insects. The mesh pollen bags were then removed from the decapitated seed heads, and the heads were placed in a large paper bag and allowed to desiccate in a dry, low-light environment at approximately 24 °C. After approximately a month of desiccation, the bags were sealed and shaken vigorously to separate the seeds from the seed heads and pappi from the seeds; separated seeds were then sifted to remove spines, pappi, and other plant debris, and were then stored in an airtight container.

After seeds were successfully extracted, they were irradiated to ensure they could be freely dispersed by ants and other insects without spreading these invasive thistles and contaminating study site. Seeds from *C. nutans* and *C. acanthoides* were irradiated using a 1000 KR dosage of gamma radiation; this dosage was demonstrated by Jongejans *et al*. (2014) to prevent any germination without affecting how attractive seeds from either of these species are to insects.

Once seeds were irradiated, a subset of seeds from each species and warming/ambient treatment were assigned an elaoisome removal treatment. Elaiosomes were removed my holding the seed with a pair of forceps, aligning the flattest side of the seed parallel to the workbench, and then using a teasing needle to press the elaiosome against the workbench until it was severed from the seed achene. In instances where the elaiosome did not cleanly separate, the teasing needle was used to gently dig out the remainder of the elaiosome from the achene. Seeds not receiving the elaiosome removal treatment were handled with forceps in the same manner mentioned above, for approximately the same duration as the seeds whose elaiosomes were removed; this was done so that possible differences in experimental outcomes could not be attributed to differences in the way the seeds were handled. After seeds had their elaiosomes removed and the control seeds were handled appropriately, seeds were again placed in airtight containers based on treatment type.

Seed depots were then constructed to serve as points where insects could easily access and remove seeds. All depots were constructed in a similar manner to those in Jongejans *et al*. (2014), using 95 mm diameter petri dishes with a 15 cm tall edge and placing black sandpaper at the bottom to mimic the appearance of soil. Sandpaper was adhered to the bottom of the petri dish in order to negate any warping or movement of the paper due to wind, moisture, sunlight exposure, or insect activity. Note that unlike in Jongejans *et al*. (2014), we do not seek to exclude specific types of insects as a treatment, so Tanglefoot was not used when constructing these seed depots. Once the depots were constructed, each one was loaded with 25 seeds placed near the centre of the petri dish, though spread out enough so that seeds were not piled on top of each other. All depots were prepared in a controlled environment and were transported to the field site, rather than being constructed *in situ*.

***Experimental setup***

The seed removal experiment was conducted at the Russell E. Larson Agricultural Research Farm in Rock Springs, Pennsylvania, which is the same study site at which the experiment from which the seeds were derived was conducted. All seed depots were arranged over an 8 x 10 grid in a small field approximately 50 m from the location of the warming experiment, with individual seed depots spaced 1 m apart. Seeds within a particular seed depot represented one of eight unique combinations of species (*C. nutans* vs *C. acanthoides*), warming treatment (warmed vs unwarmed), and elaiosome treatment (present vs absent), and each row of eight depots contained a randomised arrangement of these eight unique treatment combinations. Before placing each depot, the small patch of vegetation below was trimmed and lightly compacted so that the petri dish could be snugly fit in the grass canopy, reducing the chance of depots being disturbed by wind and ensuring that insects could access them.

Once all seed depots were placed, each seed depot was photographed from directly above by an observer in 30-minute intervals for the first 12 hours, from 09:00 to 21:00 on 07 September 2020; after 12 hours had elapsed, depots were then photographed once at 24 hours, again at 36 hours, and then once more at 48 hours. In instances where the sandpaper bottom of the dish was disturbed, it was fixed to the bottom of the petri dish again, and the remaining seeds gently placed back around the centre of the seed depot; such instances were extremely uncommon, though. No rain was observed over the duration of the experiment, and temperatures ranged between approximately 7°C and 27°C.

***Image processing***

All images were processed manually by counting the number of seeds for each seed depot at each recorded time; this was made easy by the contrast in colour between the seeds and the black sandpaper on the seed depots. Seeds were counted as removed only if they were completely removed from the seed depot; thus, even in instances where seeds or disturbed or scattered across the depot, they were still marked as present. For each image, the number of seeds was scored twice to ensure accuracy of the data.

***Statistical Analyses***

All statistical analyses were conducted in R (R Development Core Team 2009). To assess significance of the three treatment variables (species, warming, and elaiosome removal), we used the **glmer** function from the package **lme4** (Bates *et al*. 2012) to fit a generalised linear mixed-effects model to the data, using a logit link and treating the number of seeds as a binomial response. Each of the three treatment variables were encoded as a two-level factor and treated as a fixed effect; interactions between treatments were treated as fixed effects as well, and treatment block was treated as a random effect. To examine significance of treatment at different points in time, the model was fit separately at the 6, 12, 24, and 48 hour marks.

**References**

Allen, M. R., & Shea, K. (2006). Spatial segregation of congeneric invaders in central Pennsylvania, USA. Biological Invasions, 8(3), 509-521.

Ansong, M. and Pickering, C., 2014. Weed seeds on clothing: A global review. Journal of Environmental Management, 144, pp.203-211.

Bates, D., Maechler, M., Bolker, B., Walker, S., Christensen, R. H. B., Singmann, H., ... & Scheipl, F. (2012). Package ‘lme4’. CRAN. R Foundation for Statistical Computing, Vienna, Austria.

Berg, R.Y., 1975. Myrmecochorous plants in Australia and their dispersal by ants. Australian Journal of Botany, 23(3), pp.475-508.

Brew, C.R., O'Dowd, D.J. and Rae, I.D., 1989. Seed dispersal by ants: behaviour-releasing compounds in elaiosomes. Oecologia, 80(4), pp.490-497.

Culver, D.C. and Beattie, A.J., 1978. Myrmecochory in Viola: dynamics of seed-ant interactions in some West Virginia species. The journal of Ecology, pp.53-72.

Desrochers, AM, Bain, JF, & Warwick, SI (1988). The Biology of Canadian Weeds.: 89. Carduus nutans L. and Carduus acanthoides L. Canadian Journal of Plant Science , 68 (4), 1053-1068.

Edwards, W., Dunlop, M. and Rodgerson, L., 2006. The evolution of rewards: seed dispersal, seed size and elaiosome size. Journal of Ecology, 94(3), pp.687-694.

Gómez, J.M., Puerta-Piñero, C. and Schupp, E.W., 2008. Effectiveness of rodents as local seed dispersers of Holm oaks. Oecologia, 155(3), pp.529-537.

Guitián, J., Fuentes, M., Bermejo, T. and López, B., 1992. Spatial variation in the interactions between Prunus mahaleb and frugivorous birds. Oikos, pp.125-130.

Handel, S.N. and Beattie, A.J., 1990. Seed dispersal by ants. Scientific American, 263(2), pp.76-83B.

Herrera, C.M. and Jordano, P., 1981. Prunus mahaleb and birds: the high‐efficiency seed dispersal system of a temperate fruiting tree. Ecological monographs, 51(2), pp.203-218.

Hodkinson, D.J. and Thompson, K., 1997. Plant dispersal: the role of man. Journal of Applied Ecology, pp.1484-1496.

Hughes, L. and Westoby, M., 1990. Removal rates of seeds adapted for dispersal by ants. Ecology, 71(1), pp.138-148.

Hughes, L. and Westoby, M., 1992. Effect of diaspore characteristics on removal of seeds adapted for dispersal by ants. Ecology, 73(4), pp.1300-1312.

Jensen, T.S. and Nielsen, O.F., 1986. Rodents as seed dispersers in a heath—oak wood succession. Oecologia, 70(2), pp.214-221.

Jongejans, E., Silverman, E. J., Skarpaas, O., & Shea, K. (2015). Post-dispersal seed removal of Carduus nutans and C. acanthoides by insects and small mammals. Ecological research, 30(1), 173-180.

Jordano, P., García, C., Godoy, J.A. and García-Castaño, J.L., 2007. Differential contribution of frugivores to complex seed dispersal patterns. Proceedings of the National Academy of Sciences, 104(9), pp.3278-3282.

Keller and Shea. *In press*.

Michael, P.J., Owen, M.J. and Powles, S.B., 2010. Herbicide-resistant weed seeds contaminate grain sown in the Western Australian grainbelt. Weed Science, 58(4), pp.466-472.

Molau U, P. Mølgaard P (1996). International Tundra Experiment Manual. Danish Polar Centre, Copenhagen.

Nathan, R., 2007. Total dispersal kernels and the evaluation of diversity and similarity in complex dispersal systems. Seed dispersal: theory and its application in a changing world, pp.252-276.

Pemberton, R. W., & Irving, D. W. (1990). Elaiosomes on weed seeds and the potential for myrmecochory in naturalized plants. Weed Science, 615-619.

R Development Core Team (2009) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.

Rogers, H.S., Beckman, N.G., Hartig, F., Johnson, J.S., Pufal, G., Shea, K., Zurell, D., Bullock, J.M., Cantrell, R.S., Loiselle, B. and Pejchar, L., 2019. The total dispersal kernel: a review and future directions. AoB Plants, 11(5), p.plz042.

Shimono, Y. and Konuma, A., 2008. Effects of human‐mediated processes on weed species composition in internationally traded grain commodities. Weed Research, 48(1), pp.10-18.

Skinner, K., Smith, L., & Rice, P. (2000). Using noxious weed lists to prioritize targets for developing weed management strategies. Weed Science, 48(5), 640-644.

Teller, B.J., Zhang, R. and Shea, K., 2016. Seed release in a changing climate: initiation of movement increases spread of an invasive species under simulated climate warming. Diversity and Distributions, 22(6), pp.708-716.

Taylor, K., Brummer, T., Taper, M.L., Wing, A. and Rew, L.J., 2012. Human‐mediated long‐distance dispersal: an empirical evaluation of seed dispersal by vehicles. Diversity and Distributions, 18(9), pp.942-951.

Trumble, J.T. and Kok, L.T., 1982. Integrated pest management techniques in thistle suppression in pastures of North America. Weed Research, 22(6), pp.345-359.

Vander Wall, S.B., 2001. The evolutionary ecology of nut dispersal. The Botanical Review, 67(1), pp.74-117.

Vander Wall, S.B. and Longland, W.S., 2004. Diplochory: are two seed dispersers better than one?. Trends in ecology & evolution, 19(3), pp.155-161.

Veldman, J.W. and Putz, F.E., 2010. Long‐distance dispersal of invasive grasses by logging vehicles in a tropical dry forest. Biotropica, 42(6), pp.697-703.

Von Der Lippe, M., Bullock, J.M., Kowarik, I., Knopp, T. and Wichmann, M., 2013. Human-mediated dispersal of seeds by the airflow of vehicles. PloS one, 8(1), p.e52733.

Wichmann, M.C., Alexander, M.J., Soons, M.B., Galsworthy, S., Dunne, L., Gould, R., Fairfax, C., Niggemann, M., Hails, R.S. and Bullock, J.M., 2009. Human-mediated dispersal of seeds over long distances. Proceedings of the Royal Society B: Biological Sciences, 276(1656), pp.523-532.

Wilson, C.E., Castro, K.L., Thurston, G.B. and Sissons, A., 2016. Pathway risk analysis of weed seeds in imported grain: A Canadian perspective. NeoBiota, 30, p.49.

Zhang, R., Jongejans, E., & Shea, K. (2011). Warming increases the spread of an invasive thistle. PLoS One, 6(6), e21725.

Zhang, R., Post, E., & Shea, K. (2012). Warming leads to divergent responses but similarly improved performance of two invasive thistles. Population ecology, 54(4), 583-589.

**Figure 1.** Ant removing a *C. acanthoides* seed with an elaiosome (11:30).



**Figure 2.** Cricket removing a *C. acanthoides* seed without an elaiosome (20:30).

