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NF-κB Controls Cell Growth and Differentiation through Transcriptional Regulation of Cyclin D1

DENIS C. GUTTRIDGE,¹ CHRIS ALBANESE,² JULIE Y. REUTHER,³ RICHARD G. PESTELL,²
AND ALBERT S. BALDWIN, JR.^{1,4*}

Lineberger Comprehensive Cancer Center,¹ Curriculum in Genetics and Molecular Biology,³ and Department of Biology,⁴
University of North Carolina, Chapel Hill, North Carolina 27599-7295, and Albert Einstein College of Medicine
Cancer Center, Department of Developmental and Molecular Biology and Department of Medicine,
Albert Einstein College of Medicine, Bronx, New York 10461²

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Accumulating evidence implicates the transcription factor NF-κB as a positive mediator of cell growth, but the molecular mechanism(s) involved in this process remains largely unknown. Here we use both a skeletal muscle differentiation model and normal diploid fibroblasts to gain insight into how NF-κB regulates cell growth and differentiation. Results obtained with the C2C12 myoblast cell line demonstrate that NF-κB functions as an inhibitor of myogenic differentiation. Myoblasts generated to lack NF-κB activity displayed defects in cellular proliferation and cell cycle exit upon differentiation. An analysis of cell cycle markers revealed that NF-κB activates cyclin D1 expression, and the results showed that this regulatory pathway is one mechanism by which NF-κB inhibits myogenesis. NF-κB regulation of cyclin D1 occurs at the transcriptional level and is mediated by direct binding of NF-κB to multiple sites in the cyclin D1 promoter. Using diploid fibroblasts, we demonstrate that NF-κB is required to induce cyclin D1 expression and pRb hyperphosphorylation and promote G₁-to-S progression. Consistent with results obtained with the C2C12 differentiation model, we show that NF-κB also promotes cell growth in embryonic fibroblasts, correlating with its regulation of cyclin D1. These data therefore identify cyclin D1 as an important transcriptional target of NF-κB and reveal a mechanism to explain how NF-κB is involved in the early phases of the cell cycle to regulate cell growth and differentiation.

NF-κB belongs to the Rel family of transcription factors which regulate genes involved in immune and inflammatory responses (3, 5, 70). In mammals, the Rel family is composed of RelA/p65, c-Rel, RelB, p50 (NF-κB1), and p52 (NF-κB2), which have sequence similarity over approximately 300 amino acids in the amino-terminal half of the protein. NF-κB subunits are able to homo- or heterodimerize to form transcription factor complexes with a range of DNA-binding and activation potentials. Although all Rel members bind DNA, only RelA/p65 (hereafter referred to as p65), c-Rel, and RelB have extended carboxy termini harboring transactivation function (70). The most widely studied form of NF-κB is a heterodimer of the p50 and p65 subunits and is a potent activator of gene transcription (56).

In most cells, NF-κB is found sequestered in the cytoplasm bound in an inactive complex with its natural biological inhibitor IκB (3, 70). The IκB family members include IκBα, IκBβ, p105/IκBγ (precursor of p50), p100 (precursor of p52), and IκBε (41, 74). Each has in common a series of ankyrin repeats which interact with the DNA-binding domain and the nuclear localization signal of NF-κB, thus maintaining the transcription factor as an inactive complex. Activation of NF-κB is induced by a variety of diverse stimuli including inflammatory cytokines, phorbol esters, bacterial toxins (such as lipopolysaccharide) viruses, UV light, and a variety of mitogens (4, 5). Treatment of cells with these stimuli activate the recently discovered IκB kinase complex, leading to the phosphorylation of

serines 32 and 36 of IκBα or serines 19 and 23 of IκBβ (19, 46, 52, 77). This phosphorylation event targets IκB for ubiquitin-dependent degradation through the 26S proteasome complex, resulting in the release and nuclear translocation of NF-κB (22, 68).

In addition to its well-established role in activating the transcription of genes involved in immunological responses, studies indicate that NF-κB also functions in promoting cell growth. For instance, lymphocytes from mice lacking p50, p65, or c-Rel are defective in mitogenic responses (20, 38, 58, 65), and p50/p52 double-knockout animals fail to generate mature osteoclasts and B cells (25, 35). Recent reports also demonstrate the expression of NF-κB/Rel proteins in the proliferative zone of the developing avian limb bud and the requirement of NF-κB for the proper growth of this tissue (15, 36). In addition, deregulated NF-κB activity has been associated with oncogenesis, since reports show elevated NF-κB/Rel levels in primary breast cancers (18, 66). NF-κB is activated by oncogenic Ras and is required by Ras to induce foci in NIH 3T3 cells (23). Similarly, the chimeric oncoprotein Bcr-Abl, implicated in acute lymphoblastic and chronic myelogenous leukemias, also requires NF-κB to induce cellular transformation (54). Consistent with this latter study, Hodgkin's lymphoma cells depleted of NF-κB activity revealed strongly impaired tumor growth in mice (7). The ability of NF-κB to protect cells against chemotherapeutic drugs or TNF-mediated apoptosis function (9, 69, 72, 75), suggests that NF-κB-regulated growth control may be related to its cell survival properties. In fact, inhibition of NF-κB led to apoptosis in cells expressing oncogenic forms of Ras (45). Finally, recent demonstrations that cellular proliferation defects, attributed to the absence of NF-κB, are associated with a delay in cell cycle progression in G₁

* Corresponding author. Mailing address: Lineberger Comprehensive Cancer Center, Campus Box 7295, Mason Farm Rd., UNC School of Medicine, Chapel Hill, NC 27599-7295. Phone: (919) 966-3652. Fax: (919) 966-0444. E-mail: jhall@med.unc.edu.

(7, 28), in addition to the previously described physical association with NF- κ B and CBP/p300 (26, 50), establishes a link between NF- κ B and regulators of the cell cycle. Although the above cited reports strongly suggest a role for NF- κ B in cell growth control, the molecular mechanism(s) underlying this regulation remains unclear.

To gain insight into the role of NF- κ B in regulating cell growth, we first used a well-established skeletal myogenesis model, which is characterized by the maturation of precursor myoblasts into differentiated contractile myotubes. This cellular process is dependent on the activation or induction of the myogenic basic helix-loop-helix (bHLH) and MEF2 families of transcription factors that stimulate tissue-specific gene expression, as well as changes in cell cycle regulators that cause myoblasts to undergo irreversible growth arrest (39, 48, 71). The latter process is regulated by a balance in activities of cyclin–cyclin-dependent kinase (cdk) complexes and their respective known kinase inhibitors. The signal to induce differentiation, achieved most commonly in tissue culture by removing growth factor-rich medium, causes the downregulated expression of cyclins A and D1 and kinases cdk2 and cdc2, with an induced synthesis of the cdk inhibitors p18 and p21 and stabilization of the p27 protein (71). This regulated switch in activities leads to the dephosphorylation of the product of the retinoblastoma susceptibility gene (pRb), which maintains cells in a G₁-arrested state by inhibiting the E2F-DP1 transcription factor complex. Exit from the cell cycle, therefore, is critical for myogenic transcription and completion of the differentiation program.

Using the murine C2C12 skeletal muscle cell line, we demonstrate that NF- κ B functions in proliferating myoblasts to inhibit their differentiation process. This was determined by showing that C2C12 myoblasts contain NF- κ B in their nuclei and that NF- κ B DNA-binding activity and transactivation function are reduced during myogenesis. In addition, myoblasts generated to lack NF- κ B activity are greatly accelerated in their differentiation program. Transfections in 10T1/2 cells showed that NF- κ B strongly blocks the ability of the myogenic transcription factor, MyoD, to induce myogenesis. Furthermore, this latter regulation is specific to the transactivation-competent p65 subunit of NF- κ B, arguing that NF- κ B inhibits myogenic differentiation through its activation of gene expression. The observation that C2C12 cells lacking NF- κ B display a reduction in their proliferation rate and exit the cell cycle faster than do control cells suggests that inhibition of myogenesis by NF- κ B is in part related to its growth-promoting activity. Importantly, these cells also exhibit a marked reduction in cyclin D1 protein and mRNA levels. The results of experiments performed with 10T1/2 fibroblasts indicate that NF- κ B regulation of cyclin D1 is one mechanism by which this transcription factor inhibits myogenic differentiation. From this differentiation model, we expanded our study to identify the level at which NF- κ B regulated cyclin D1. The results show that this regulation occurs at the transcriptional level and is mediated by several authentic NF- κ B DNA-binding sites in the cyclin D1 promoter. Furthermore, by using diploid fibroblasts, we addressed the potential relevance of NF- κ B regulation of cyclin D1 with respect to the cell cycle. Our data show that in cells stimulated to reenter the cell cycle, NF- κ B activity is required for cyclin D1 transcriptional initiation and hyperphosphorylation of pRb, leading to progression into S phase. Similar to what was observed in C2C12 cells, embryonic fibroblasts lacking NF- κ B activity also exhibit a reduction in proliferation, in conjunction with lower levels of cyclin D1. Taken together, these data establish that the ability of NF- κ B to control cellular

proliferation and differentiation are processes tightly coupled to its ability to transcriptionally regulate cyclin D1.

MATERIALS AND METHODS

Cell culture. Murine C2C12 myoblast cells obtained from the American Type Culture Collection and primary murine myoblasts (a generous gift from J. Samulski) were cultured at 37°C in Dulbecco's modified Eagle's medium with high glucose (DMEM-H), supplemented with 20% fetal bovine serum (FBS) and antibiotics (Life Technologies). The cells were grown at subconfluence and passaged every 2 to 3 days. To induce differentiation, the cells were grown overnight to 60 to 70% confluence in growth medium (GM), washed once with phosphate-buffered saline (PBS), and then switched to DMEM-H supplemented with 2% horse serum and 10 μ g of insulin per ml plus antibiotics (DM). C3H10T1/2 clone 8 mouse embryo fibroblasts (10T1/2), also obtained from American Type Culture Collection, were cultured in DMEM-H containing 15% FBS plus antibiotics and passaged every 2 to 3 days. HeLa cells were cultured in DMEM-H with 5% FBS and 5% calf serum, NIH 3T3 cells were grown in DMEM-H with 10% Colorado calf serum, and for mouse embryo fibroblasts (MEFs), cells were grown in DMEM-H plus 10% FBS.

Plasmids. For reporter plasmids, 3 κ B-Luc or 3 κ Bmut-Luc contain three tandem repeats of the wild-type or mutated κ B site, from the major histocompatibility complex (MHC) class I enhancer, respectively, fused to the luciferase reporter gene (obtained from B. Sugden, University of Wisconsin, Madison, Wis.). TnI-Luc contains a muscle-specific enhancer in the troponin I gene fused to luciferase, and 4RTK-Luc contains four E boxes from the muscle creatine kinase enhancer (gifts of S. Konieczny, Purdue University). Cyclin D1 promoter reporter constructs were described earlier (1). For expression plasmids, p50 and p65 subunits of NF- κ B were expressed from a cytomegalovirus (CMV)-driven promoter as previously described (10). The mutant $\text{I}\kappa\text{B}\alpha$ plasmid, designated $\text{I}\kappa\text{B}\alpha\text{SR}$, was a gift of D. Ballard (Vanderbilt University). A cyclin D1 expression plasmid was generated by removing a 1,300-bp *Eco*RI fragment containing the mouse cyclin D1 cDNA from the pBSSK plasmid and inserting it into the *Eco*RI site of pCMV5. pCDNA3-D3 plasmid expressing human cyclin D3 was a gift of C. Sherr (St. Jude Children's Research Hospital). pEMC11s plasmid expressing MyoD was obtained from the H. Weintraub laboratory (University of Washington). Oncogenic *ras* was expressed from the H-*ras* (V-12) plasmid as previously described (45).

EMSA. Nuclear extracts for electrophoretic mobility shift assays (EMSA) were prepared as previously described (16), except that 0.25% Nonidet P-40 was used to extract nuclei. A 5- μ g portion of extract were preincubated with 1 mM phenylmethylsulfonyl fluoride and 1 μ g of poly(dI-dC)-poly(dI-dC) in a volume of 12 μ l for 10 min. This mixture was subsequently incubated in a total volume of 20 μ l at room temperature for 20 min with 2×10^4 cpm of a ³²P-labeled oligonucleotide probe containing a κ B site (underlined) from the class I MHC promoter (5'-CAG GGC TGG GGA TTC CCC ATC TCC ACA GTT TCA CTT C-3'). The buffer consisted of 10 mM Tris-HCl (pH 7.7), 50 mM NaCl, 0.5 mM EDTA, 1 mM dithiothreitol, and 10% glycerol. Complexes were resolved on a 5% polyacrylamide gel in Tris-glycine buffer (25 mM Tris, 190 mM glycine, 1 mM EDTA) at 25 mA for 2 to 3 h at room temperature. The gels were dried and exposed on film for approximately 1 to 3 days. For supershift EMSAs, antibodies against specific NF- κ B were added to the nuclear extract and incubated for 10 min prior to the addition of phenylmethylsulfonyl fluoride and poly(dI-dC)-poly(dI-dC). The antibodies used for this portion of the study were p65 (Rockland), p50 (NLS; Santa Cruz Biotechnology), c-Rel (C; Santa Cruz Biotechnology), and RelB (C-19; Santa Cruz Biotechnology). NF- κ B-binding sites in the cyclin D1 promoter were determined by generating a series of oligonucleotides corresponding to both wild-type and mutant (M) putative NF- κ B sites within the human cyclin D1 promoter (47). The oligonucleotides have the following sequences (NF- κ B wild-type and mutated sites are underlined): -858, 5'-GTG CAG TTG GGG ACC CCC GCA AGG ACC GAC TGG TCA A-3'; -858(M), 5'-GTG CAG TTC CCG ACC CCC GCA AGG ACC GAC TGG TCA A-3'; -749, 5'-ACC ATC TTG GGC TGC TGC TGG AAT TTT CGG GCA TTT A-3'; -749(M), 5'-ACC ATC TTG GGC TGC TGC TCC CCT TTT CGG GCA TTT A-3'; -39, 5'-GGA CTA CAG GGG AGT TTT GTT GAA GTT GCA AAG TCC T-3'; and -39(M), 5'-GGA CTA CAC CCC AGT TTT GTT GAA GTT GCA AAG TCC T-3'.

Transfections and viral infections. To generate C2C12 cells with stably integrated luciferase reporter plasmids, cells were seeded at a density of 2×10^5 cells in 6-cm dishes 24 h prior to transfection. Cotransfections with 4 μ g of reporter plasmid 3 κ B-Luc or 3 κ Bmut-Luc and 1 μ g of pcDNA3 (Invitrogen) containing the neomycin resistance marker were performed with Superfect reagent as recommended by the manufacturer (Qiagen). At 48 h posttransfection, the cells were trypsinized and cultured at 1/30 their density in 1 mg of Geneticin (G418; Life Technologies) per ml. Mixed populations containing either wild-type or mutant versions of the κ B sites were allowed to expand under selection, and from the wild-type population an individual clone expressing similar basal promoter activity was selected. Cell extracts were prepared and luciferase assays were performed as previously described (16).

C2C12 cells stably expressing $\text{I}\kappa\text{B}\alpha\text{SR}$ or empty vector were generated by retroviral infections as previously described (54). The cells were seeded under identical conditions to those stated above. Helper-free virus infection was per-

formed in 1 ml of culture in the presence of 2 μ g of Polybrene for 3 h. Culture medium was aspirated, and fresh medium was added for 48 h. The cells were then trypsinized and replated in G418-containing medium at 1 mg/ml at a ratio of 0.7 cell/well in a 96-well plate. The selection medium was replaced every 4 to 5 days, and individual clones were expanded for further study. To produce MEFs stably expressing I κ B α SR, retrovirus infections were performed as described above. The cells were then placed under a 3-day selection of G418 at 400 μ g/ml and subsequently expanded as a mixed population for further study.

Transient transfections into 10T1/2 fibroblasts were performed by seeding 5 \times 10⁵ cells in a 6-cm dish and growing the cells overnight in complete medium. The following day, a total of 2.5 μ g of plasmid DNA was incubated with Superfect as recommended by the manufacturer (Qiagen). This mixture was subsequently added to the cells with 1 ml of complete medium for approximately 3 h. The cells were rinsed with PBS and then refed with 4 ml of complete medium overnight. At this point, the cells were again rinsed once with PBS and then transferred to DM for a 48-h period. Cell extracts were prepared and luciferase activity was monitored as previously described (16).

For adenovirus infections in cycling cells, HeLa cells and MEFs were plated overnight in 10-cm culture dishes. The following day, replication-defective adenovirus (Ad5) expressing the I κ B α SR or empty vector (CMV) were diluted in 2.5 ml of complete medium and placed on the cells for 1 h. The volume was then raised to 10 ml, and infections were allowed to proceed for an additional 48 h, at which time the cells were harvested and total RNA was prepared. For infections performed in quiescent cells, MEFs were plated overnight in 10-cm culture plates at 50 to 60% confluence and then switched for 48 h to medium containing 0.2% FBS. Infections were performed as explained above, except that the cells were maintained quiescent for 24 h before being stimulated back into the cell cycle by the addition of complete medium.

Western blot analysis. Cells were harvested in PBS, and whole-cell lysates were prepared by resuspending cell pellets in ice-cold RIPA buffer (10 mM Tris-HCl [pH 8.0], 1 mM EDTA, 1% sodium dodecyl sulfate [SDS], 1% Nonidet P-40), and incubating the suspension on ice for 20 to 30 min. For pRb Western blots, lysis buffer included phosphatase inhibitors (50 mM sodium fluoride, 1 mM orthovanadate, 50 mM β -glycerophosphate, 80 μ M cantharidin). Supernatant lysates were collected following high-speed centrifugation for 20 min at 4°C. Equal amounts of extract were subjected to SDS-polyacrylamide gel electrophoresis and transferred to nitrocellulose membranes (Schleicher & Schuell). Blocking was performed in 5% nonfat dry milk-1× TBST (25 mM Tris-HCl [pH 8.0], 125 mM NaCl, 0.1% Tween 20). Primary and secondary antibodies were diluted in 0.5% nonfat dry milk-1× TBST, and the incubations proceeded for 30 min at room temperature, with the exception of pRb blots, where the primary antibody was incubated for 1 h. Washes were performed in 1× TBST for 5 to 10 min and repeated five times. Specific proteins were visualized by enhanced chemiluminescence (Amersham Life Science). Antibodies to I κ B α (C-21), myogenin (M-225), cyclin D1 (R-124), cyclin A (C-19), MyoD (M-318), and p21 (C-19) were obtained from Santa Cruz Biotechnology, pRb antibody was obtained from Pharmingen, cyclin D3, cdk4, and p27 antibodies were generous gifts of Y. Xiong (University of North Carolina).

Immunofluorescence. All immunofluorescence experiments were performed directly in 12-well plates. Myoblasts were grown to subconfluence in GM and then switched to DM for 48 h. For 10T1/2 fibroblasts, cells were grown overnight in complete medium following transfections then switched to DM for 72 h. All steps were performed at room temperature. The cells were washed in PBS and fixed in a 2% formaldehyde-1× PBS solution for 30 min. They were permeabilized with 0.5% Nonidet P-40-1× PBS for 5 min and then blocked with horse serum (1:100) in PBS for 30 min. They were washed with PBS, incubated for 1 h with anti-skeletal myosin heavy chain (MY-32; Sigma) diluted 1:500 in 3% bovine serum albumin-1× PBS, washed three times with PBS, and incubated for 1 h in the dark with fluorescein isothiocyanate (FITC)-conjugated anti-mouse immunoglobulin G (IgG; Sigma) diluted at 1:250. The cells were washed in PBS and photographed with an Olympus inverted microscope equipped with phase-contrast and UV illumination through an FITC filter. For bromodeoxyuridine (BrdU) staining, MEFs grown on glass coverslips were washed once in PBS, fixed for 5 min at room temperature with 100% cold methanol, washed again in PBS, and permeabilized with 0.25% Triton X-100 in PBS for 5 min at room temperature. Following a PBS wash, the cells were treated for 10 min with 1.5 M HCl, washed four times with PBS, and incubated for 30 min with an FITC-conjugated anti-BrdU antibody (1:10 dilution in 1% bovine serum albumin-PBS [Becton Dickinson]). After several washes in PBS, the cells were counterstained for 3 min with a 1- μ g/ml solution of Hoechst dye 33258 (Sigma) in PBS. Following two additional washes in PBS, the coverslips were mounted on glass slides and examined on a Zeiss microscope.

Northern blot analysis. Total RNA was isolated with Trizol reagent as recommended by the manufacturer (Life Technologies). RNA samples were fractionated on an agarose gel and transferred overnight onto a nylon filter. The next day, RNA was cross-linked with a UV cross-linker (Stratagene). For detection of I κ B α , Id-1, Hes-1, cyclin D3, and glyceraldehyde-3-phosphate dehydrogenase (GAPDH), mRNAs blots were hybridized in QuickHyb buffer supplemented with 100 μ g of salmon sperm DNA as recommended by the manufacturer (Stratagene). For detection of cyclin D1 mRNA, blots were hybridized overnight at 42°C in 50% formamide-5× SSC (1× SSC is 0.15 M NaCl plus 0.015 M sodium citrate)-1× PE (50 mM Tris-HCl [pH 7.5], 0.1% sodium pyrophosphate,

1% SDS, 0.25% polyvinylpyrrolidone, 0.25% Ficoll, 5 mM EDTA)-150 μ g of salmon sperm DNA. All the probes were generated with a random-primed labeling kit (Life Technologies) in the presence of [α -³²P]dCTP (NEN-Dupont). The DNA products were purified over micro-G-50 Sephadex columns (Life Technologies), boiled, and added to the hybridization mixture. Washes were performed twice in 2× SSC-0.1% SDS for 10 min at room temperature and then twice in 0.1× SSC-0.1% SDS for 20 min at 42°C for cyclin D1 detection and 65°C for detection of all other mRNAs.

RESULTS

Loss of NF- κ B activity correlates with myogenic differentiation. To analyze the role of NF- κ B in cell growth control, we initiated our study by using the C2C12 myogenic differentiation model, since in this cell culture system the gene products involved in regulating terminal cell cycle arrest are largely known (39, 48, 71). First, we analyzed the relative levels and subunit composition of NF- κ B/Rel complexes found in undifferentiated and differentiated C2C12 cell cultures. EMSAs performed with nuclear extracts prepared from C2C12 cells identified four potential NF- κ B-containing complexes in proliferating myoblasts (Fig. 1A, lane 1). The levels of complexes I, II, and III decreased as cells became terminally differentiated (Fig. 1A, lanes 1 to 4). The level of complex IV initially decreased but was elevated 72 h following initiation of the differentiation process. EMSA supershifts performed to identify which Rel proteins were contained in these complexes revealed the presence of the p65 subunit in complex III and p50 in complex IV in both undifferentiated and differentiated cells (Fig. 1B, lanes 2 and 3 and lanes 7 and 8). Although the antibody against p50 was not able to supershift complex III, the migration pattern of this complex nevertheless resembled that of the classical p50-p65 heterodimer. None of the antisera specific for NF- κ B/Rel proteins caused a supershift in complexes I and II, suggesting that NF- κ B is most probably not a component of these higher-molecular-weight complexes. To address whether these effects were specific to C2C12 cells, EMSA and supershift analyses were repeated with nuclear extracts prepared from primary myoblasts. As seen with the C2C12 cells, loss of p50 and p65 NF- κ B DNA-binding activity was observed in primary myoblasts undergoing differentiation (Fig. 1C), indicating that a similar reduction of NF- κ B may occur in naturally developing skeletal muscle.

Earlier reports established that upregulated expression and increased DNA-binding activity of the NF- κ B subunit c-Rel were associated with B-cell development (27). By EMSA, we were unable to detect c-Rel binding in skeletal muscle cells (Fig. 1B and C), suggesting that the role of NF- κ B subunits, with respect to differentiation, may be tissue specific. To ensure that the oligonucleotide probe used in our EMASAs did not favor p65 binding, we tested it with extract prepared from the mature B-cell line WEHI-231, known to contain high levels of c-Rel-binding activity (42). Supershift analysis showed that the probe was readily bound by c-Rel in these cells (Fig. 1D), supporting the notion that, unlike what is found in lymphocytes, c-Rel binding is not induced during myogenic differentiation.

To determine whether loss of NF- κ B binding activity correlated with its transactivation potential, a population of C2C12 cells were generated that stably integrated a luciferase reporter gene fused to three tandem repeats of the κ B site from the MHC class I enhancer (3x κ B-Luc). Upon differentiation of these cells, NF- κ B transcriptional activity was reduced nearly 50% (Fig. 2A), a level which is likely to underrepresent the total loss of κ B-dependent transcriptional activity, since not all C2C12 cells reach terminal differentiation under these culture conditions. Similar results were obtained when κ B-dependent

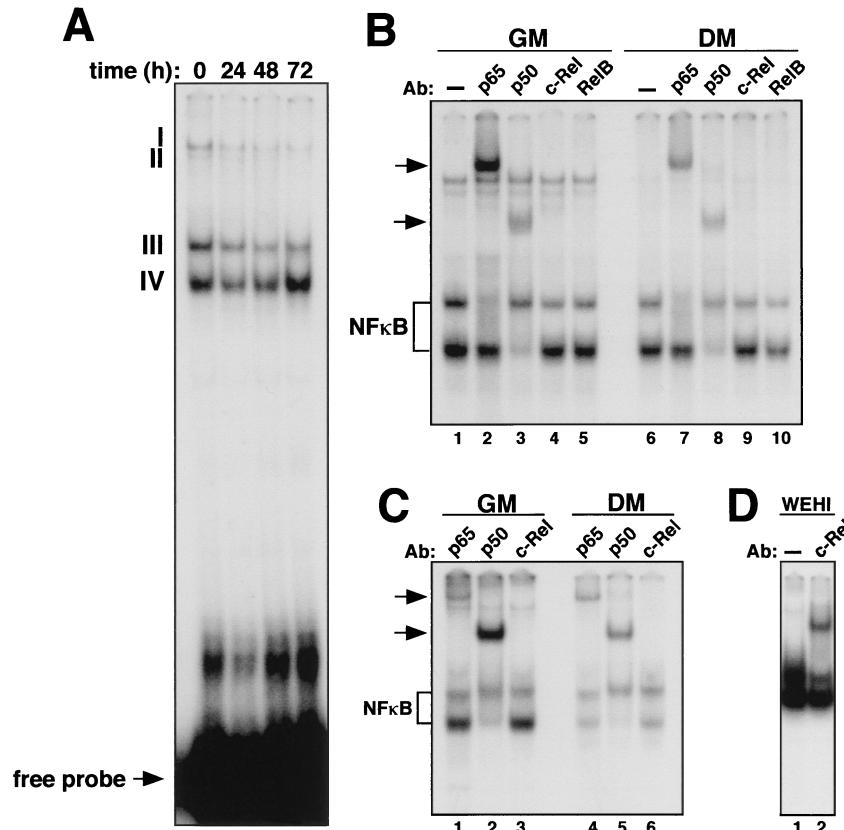


FIG. 1. Loss of NF- κ B binding activity during myogenic differentiation. (A) Proliferating C2C12 myoblasts (GM) were induced to differentiate (DM), and at the indicated times nuclear extracts were prepared and EMSA was performed with a radiolabeled oligonucleotide containing an NF- κ B-binding site. (B) C2C12 cells were maintained in GM or switched to DM for 72 h. Supershift EMSA was performed with nuclear extracts preincubated with either no antibody (lanes 1 and 6) or antisera specific for p65 (lanes 2 and 7), p50 (lanes 3 and 8), c-Rel (lanes 4 and 9), or RelB (lanes 5 and 10). NF- κ B complexes containing p50 and p65 subunits are shown. (C) EMSA and supershift EMSA were performed as described above with nuclear extracts prepared from primary myoblasts undergoing differentiation. (D) Supershift EMSA was performed with nuclear extract prepared from WEHI-231 cells, preincubated either with no antibody or with an antiserum specific to the c-Rel subunit of NF- κ B.

reporter activity was tested in an isolated clone undergoing differentiation (Fig. 2A). The functionality of the κ B-dependent reporter was demonstrated by showing that the promoter was responsive to the NF- κ B-activating cytokine tumor necrosis factor alpha (TNF- α) whereas a similar promoter containing a mutated version of the κ B sites ($3\chi\kappa$ Bmut-Luc) lacked basal activity and was unresponsive to cytokine treatment (Fig. 2A). To further investigate NF- κ B transcriptional activity in myogenesis, Northern blot analysis was performed to probe for I κ B α , whose transcription is known to be regulated by NF- κ B (5). The results showed that a significant reduction in the level of I κ B α mRNA was associated with differentiating C2C12 cells (Fig. 2B). Thus, the combined data from EMSAs, reporter assays, and Northern blotting confirmed that NF- κ B activity, most probably represented by the classical, transcriptionally active p50-p65 heterodimer, is reduced in differentiating skeletal muscle cells.

NF- κ B functions as a negative regulator of myogenesis in the C2C12 model. The results described above suggested a functional role for NF- κ B in precursor myoblasts. To test this hypothesis, C2C12 cells were generated to express a mutant form of the I κ B α inhibitor for which serines at positions 32 and 36 had been changed to alanines. The resulting protein (referred to as I κ B α superrepressor, or I κ B α SR) is no longer subject to phosphorylation and subsequent proteasome degradation following an NF- κ B-activating stimulus and therefore

functions as a potent and specific inhibitor of NF- κ B activity (13). The absence of endogenous I κ B α protein in I κ B α SR-containing myoblasts (Fig. 3A), as well as the previously described sensitivity of I κ B α SR-expressing cells or p65 $^{-/-}$ cells to TNF-induced killing (9, 69, 72) (Fig. 3B), demonstrated that I κ B α SR was functioning properly to block NF- κ B activity. The inhibitory activity of the I κ B α SR was confirmed by EMSA, which showed that only I κ B α SR-expressing myoblasts were blocked in their ability to activate NF- κ B in response to TNF treatment (data not shown). Parental, vector control, or I κ B α SR-expressing myoblasts were examined for the rate at which they underwent differentiation. At 48 h following serum withdrawal, less than 10% of the parental or vector control cells had undergone differentiation, as determined by their myotube phenotype and by their ability to express the late differentiation marker, myosin heavy chain (Fig. 3C). In sharp contrast, nearly all the I κ B α SR-expressing cells had become fully differentiated by this time (Fig. 3C). Similar enhanced rates of differentiation were observed in pooled clones of I κ B α SR-containing cells, demonstrating that these effects were not due to clonal variations (Fig. 3C). An examination of a second temporally regulated myogenic differentiation marker, myogenin, demonstrated greatly accelerated expression of this myogenic transcription factor in C2C12-I κ B α SR cells compared to vector control cells (Fig. 3D). These data suggested

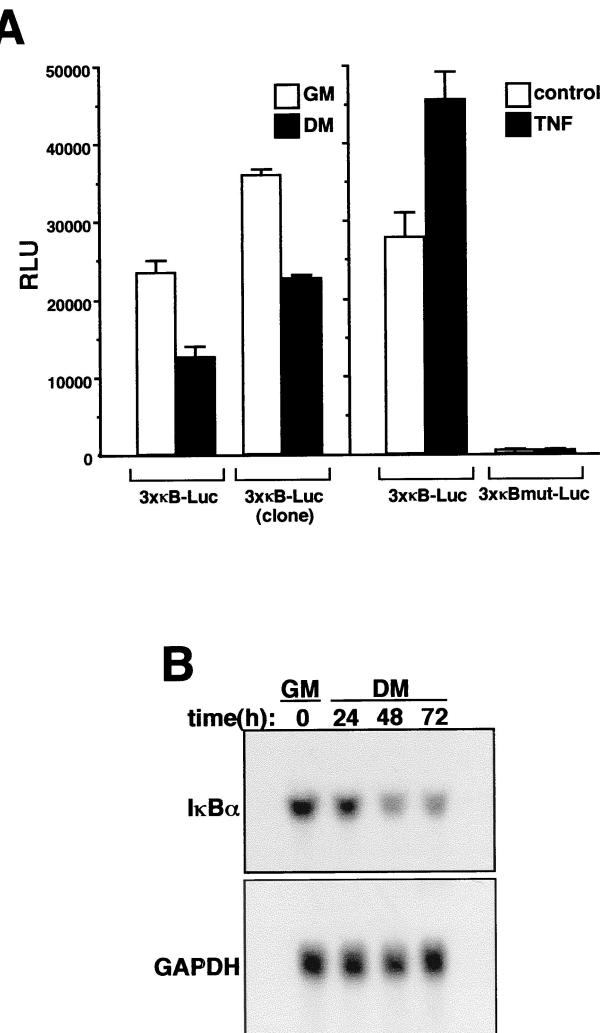


FIG. 2. Myogenic differentiation correlates with loss of NF- κ B transactivation function. (A) C2C12 myoblasts stably containing a 3xkB-Luc reporter plasmid were propagated as either a mixed population or a clonal isolate. Cells were plated in triplicate overnight in 6-cm culture dishes, and on the following day they were maintained in GM or switched to DM for 48 h. At that time, cell extracts were prepared, and relative luciferase units were determined by normalizing to total protein (RLU). Promoter activities were also determined for 3xkB-Luc and 3xkBmut-Luc populations that were treated or not treated with 10 ng of TNF- α per ml for 24 h. (B) C2C12 cells were maintained in GM or differentiated in DM for up to 72 h. At the indicated times, total RNA was prepared and 10 μ g of sample was used for Northern blot analysis. The blot was hybridized with an I κ B α -specific probe, and RNA loading was normalized by stripping the blot and reprobing for GAPDH mRNA.

that NF- κ B activity was required in proliferating myoblasts to block myogenic differentiation.

To verify that overexpression of I κ B α SR in C2C12 cells did not introduce nonspecific effects leading to false interpretation of NF- κ B function, we tested the ability of NF- κ B to regulate the expression of a muscle-specific gene. One of the hallmark features of MyoD, as well as other members of the myogenic bHLH family, myogenin, myf5, and MRF4, is their ability to induce myogenic differentiation in nonmuscle cells (39). We therefore performed cotransfection experiments with murine embryonic C3H10T1/2 (10T1/2) fibroblasts and a reporter plasmid containing the troponin I enhancer and promoter (TnI-Luc), along with expression plasmids for MyoD, NF- κ B,

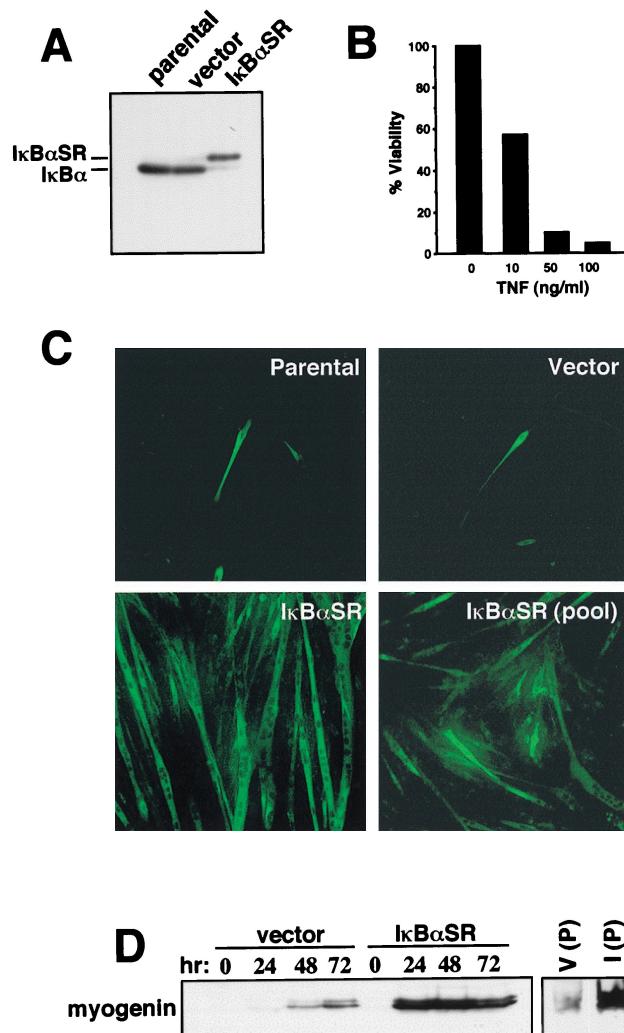


FIG. 3. C2C12 cells lacking NF- κ B activity have an accelerated rate of differentiation. (A) Whole-cell lysates were prepared from C2C12 parental, vector control, or I κ B α SR proliferating myoblasts, and 50 μ g of sample was used for Western blot analysis. I κ B α and I κ B α SR proteins were detected with an I κ B α polyclonal antibody (C-19; Santa Cruz Biotechnology) at a 1:1,500 dilution. The I κ B α SR protein is FLAG tagged and therefore migrates at a slightly higher mobility compared to the endogenous protein. (B) I κ B α SR-expressing myoblasts were seeded in triplicate overnight in 12-well plates, and the following day cells were treated with increasing concentrations of TNF- α for 48 h. Cell viability was scored by trypsinization and the trypan blue exclusion method. Cells not treated with TNF- α were designated 100% viable. (C) C2C12 parental cells, vector control, an I κ B α SR clone, or five pooled I κ B α SR clones were differentiated in DM for 48 h, at which time the cells were prepared for immunofluorescence to detect for the myosin heavy chain. (D) C2C12 vector control or I κ B α SR cells were induced to differentiate in DM for up to 72 h. At the indicated times, lysates were prepared and Western blot analysis was performed probing for myogenin expression. V (P) and I (P) denote myogenin expression from five pooled vector control or I κ B α SR clones, respectively, that were differentiated for 48 h.

and/or I κ B α SR. As expected, MyoD strongly activated the troponin I reporter when cells were placed in differentiation conditions (Fig. 4A). However, coexpression of the p50 and p65 NF- κ B subunits strongly repressed the activation by MyoD, at levels comparable to those for oncogenic H-Ras, a known potent negative regulator of myogenesis (37, 40, 49). Viability assays determined that the reduction in MyoD transactivation by NF- κ B was not due to cell death resulting from transfection conditions (data not shown). Similar inhibition

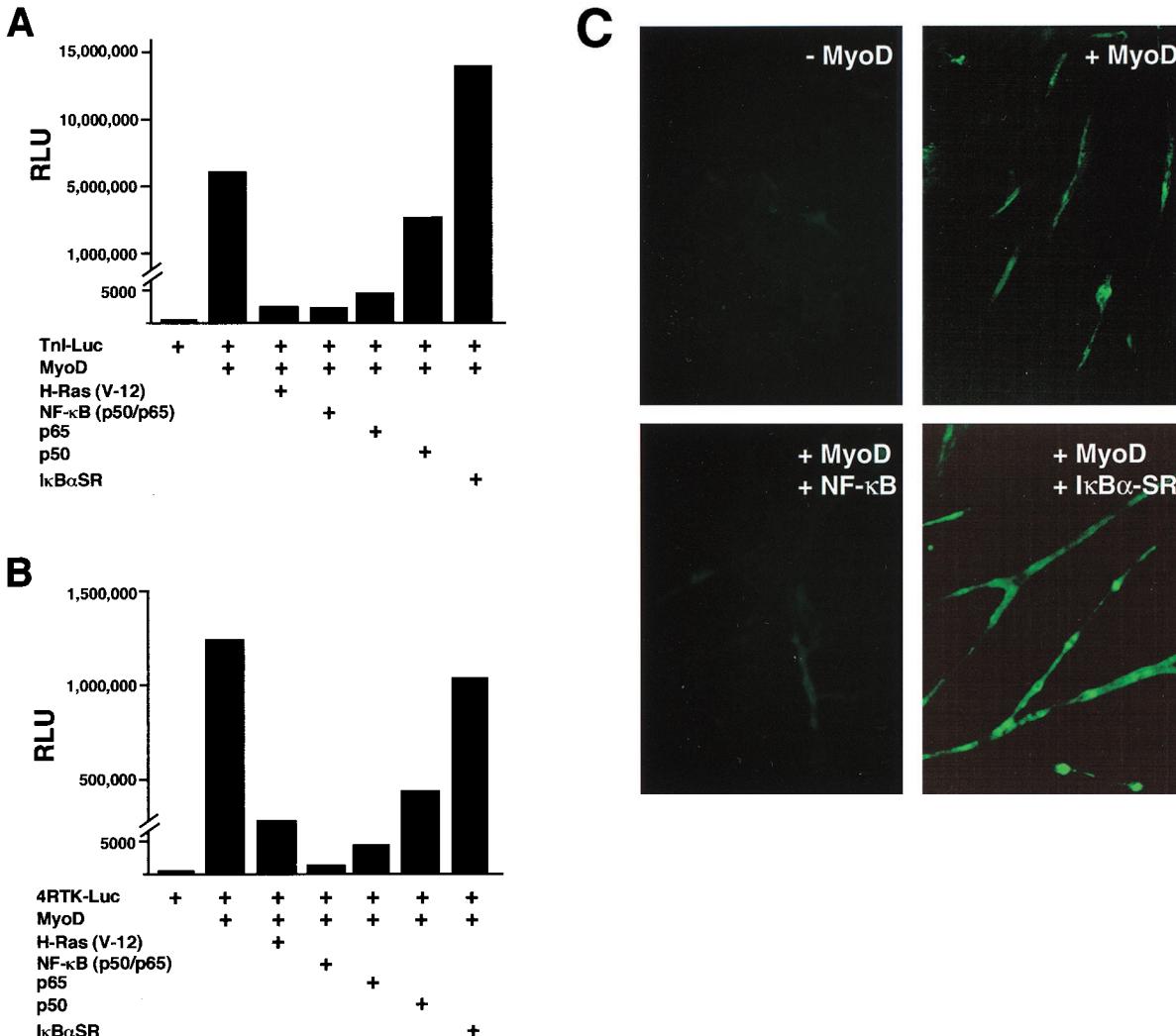


FIG. 4. NF-κB inhibits MyoD-induced myogenesis in 10T1/2 fibroblasts. Cells were maintained in growth medium containing 15% FBS and differentiated in DM. The cells were seeded in triplicate overnight in 6-cm dishes, and the following day, cotransfections were performed with Superfect (Qiagen). DNA consisted of 1 µg of a troponin-I-Luc reporter plasmid (TnI-Luc) (A) or 1 µg of the 4RTK-Luc plasmid (B), along with 0.25 µg of an expression plasmid for MyoD alone or in combination with 0.5 µg of expression plasmids for either the activated form of oncogenic ras (H-ras V-12), p65, p50, or 1 µg of IκBαSR. DNA was standardized to 2.5 µg by the addition of Bluescript plasmid (Stratagene). Cells were maintained in growth medium for 24 h following transfections and then switched to DM for 48 h, at which time cell extracts were prepared and relative light units (RLU) were determined, by normalizing values to total protein. (C) For immunofluorescence analysis, 10T1/2 cells were seeded overnight and the next day similar transfections were performed as described above, except that one-third of the amount of DNA was used. At 24 h following transfections, the cells were switched to DM for 72 h, at which time the cells were fixed and probed for the myosin heavy chain. To score for the number of myotubes formed, cells expressing myosin were counted and averaged from a minimum of 10 randomly selected fields.

levels were produced when the p65 subunit, but not p50, was cotransfected along with MyoD, suggesting that the transcriptional activation function of NF-κB is required to mediate this regulation. It was possible that expression of NF-κB subunits sequestered a cofactor required for MyoD transcriptional function. Therefore, we asked whether inhibition of NF-κB function would enhance MyoD transcriptional activity. Expression of IκBαSR enhanced MyoD transcriptional activation of troponin I over that of MyoD alone (Fig. 4A), reaffirming the notion that NF-κB functions as an inhibitor of differentiation. We also observed that expression of IκBαSR partially overcame the ability of Ras to inhibit MyoD transcriptional activity (data not shown), confirming that NF-κB is a Ras-responsive transcription factor which also can mediate antidiifferentiation. Similar results to the ones described above were obtained when transfactions were repeated with a reporter plasmid con-

taining four E-box sites from the muscle creatine kinase enhancer (4RTK-Luc) (Fig. 4B), which is also known to be strongly regulated by myogenic bHLH proteins (39). These results argue that negative regulation on myogenesis by NF-κB is not specific to the troponin gene. As an additional approach to assay the regulatory potential of NF-κB on differentiation, cotransfections with MyoD and NF-κB subunits were performed with 10T1/2 fibroblasts and analyzed for their effects on myotube formation and myosin heavy-chain expression. As can be seen in Fig. 4C, the expression of MyoD in 10T1/2 cells caused the formation of small myotubes 72 h following serum withdrawal but addition of NF-κB was sufficient to completely abolish this myogenic event. In contrast, the expression of IκBαSR with MyoD enhanced both the overall number (quantitatively approximated to be threefold over that of MyoD alone) and size of myotubes formed in the culture wells (Fig.

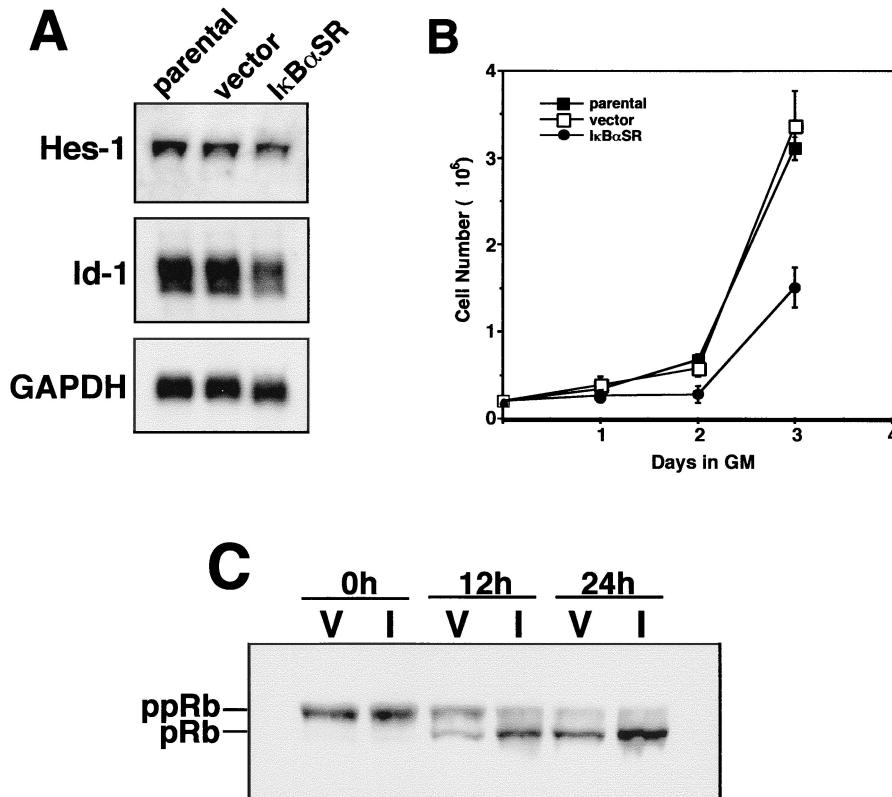


FIG. 5. C2C12 cells lacking NF- κ B exhibit a growth defect and changes in pRb phosphorylation. (A) Total RNA was prepared from C2C12 parental, vector control, or I κ B α SR proliferating myoblasts, and Northern analysis was performed to detect the expression of Hes-1 or Id-1 genes. (B) C2C12 parental, vector control, or I κ B α SR myoblasts were plated in triplicate in 10-cm plates and maintained in GM for 3 days. Every 24 h, the total cell number was determined by trypsinization and trypan blue exclusion. (C) C2C12 vector control (V) or I κ B α SR cells (I) were induced to differentiate for up to 24 h. At the indicated times, cell lysates were prepared with lysis buffer containing phosphatase inhibitors and Western blotting was performed to probe for the hypo- and hyperphosphorylated forms of Rb with a monoclonal antibody (14001A; Pharmigen).

4C). These findings correlated strongly with the reporter assay data and confirmed that NF- κ B functions as a negative regulator of myogenic differentiation in vitro.

NF- κ B inhibits C2C12 myogenesis through its growth-promoting activity and regulation of cyclin D1. To determine the mechanism by which NF- κ B regulated myogenesis in vitro, we first examined whether NF- κ B had any effect on the expression of Id-1 or Hes-1, which are known inhibitors of differentiation (12, 55). Northern analysis performed with mRNA isolated from proliferating C2C12 parental, vector control, or I κ B α SR-expressing myoblasts revealed that the steady-state levels of Hes-1 was only slightly altered whereas Id-1 was more strongly altered in cells lacking NF- κ B activity (Fig. 5A). This result may indicate that these genes could be involved in NF- κ B-mediated inhibition of myogenic differentiation. Relevant to the regulatory mechanism of NF- κ B was that C2C12-I κ B α SR cells maintained in growth medium exhibited a substantial increase in cell doubling time compared to parental or vector control cells (Fig. 5B). Since successful progression of myogenic differentiation is highly dependent on the ability of cells to irreversibly exit the cell cycle in the G₁ state (71), we asked whether the effect on cell growth as a result of the absence of NF- κ B activity translated to changes in cell cycle progression. Exit from cell cycle was assessed by immunoblot analysis probing for the phosphorylation status of pRb. The results showed that while no obvious differences in pRb states were observed in either vector control or I κ B α SR cells maintained in fresh growth medium ($t = 0$ h), once cells were induced to differ-

entiate ($t = 12$ and 24 h) pRb hypophosphorylation occurred much faster in I κ B α SR cells than in control cells (Fig. 5C). These data imply that in immortalized C2C12 cells, inhibition of NF- κ B accelerates the rate at which cells exit the cell cycle.

The above finding prompted us to search for other cell cycle regulatory factors that function upstream of pRb and that could potentially be regulated by NF- κ B. Immunoblotting results showed that under proliferating conditions, I κ B α SR-expressing myoblasts exhibited a striking reduction in cyclin D1 levels compared to vector control cells (Fig. 6A). In addition, cyclin D1 levels decreased more rapidly in differentiating I κ B α SR cells than in control cells. Potential regulation by NF- κ B for other cell cycle regulators, cyclin D3, cyclin A, and p21, was observed but was less dramatic than the effect on cyclin D1 and may therefore be due to secondary effects of changes on cyclin D1. Although basal levels of p21 were reduced in C2C12 cells lacking NF- κ B (Fig. 6A), inhibition of NF- κ B did not block the accumulation of p21, which is associated with myogenic differentiation (29, 30). The lack of any significant regulation of cdk4, as well as for the cdk inhibitor protein p27, also indicated that effects of NF- κ B on cyclin D1 expression were specific. Consistent with immunoblotting results, steady-state levels of cyclin D1 mRNA were also significantly lower in proliferating I κ B α SR-expressing myoblasts than in either parental or vector control cells (Fig. 6B). In comparison, we detected only a slight effect on cyclin D3 mRNA expression in the NF- κ B-inhibited cells, consistent with observations showing that cyclin D3 levels increase during skel-

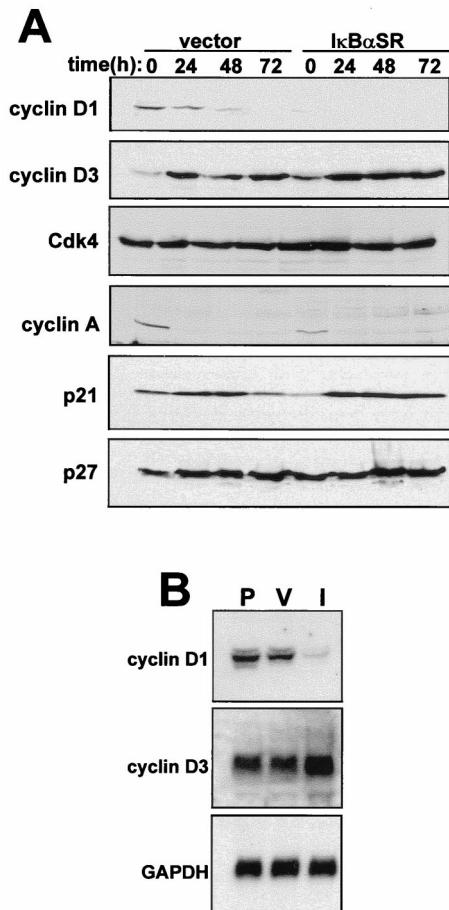


FIG. 6. C2C12 cells lacking NF- κ B are downregulated for cyclin D1 protein and mRNA. (A) C2C12 vector control or I κ B α SR cells were differentiated in DM for up to 72 h. At the indicated times, whole-cell lysates were prepared and 50 μ g was used in Western blot analyses to probe for various cell cycle proteins. (B) Total RNA was prepared from proliferating parental (P), vector control (V), or I κ B α SR (I) myoblasts, and 10 μ g of sample was used in Northern blotting to probe for cyclin D1 or cyclin D3 mRNA.

etal muscle differentiation (51). These results therefore demonstrate that NF- κ B is a specific regulator of cyclin D1.

As shown in Fig. 6A and consistent with previous reports (51, 64), downregulated expression of cyclin D1 correlates with myogenic differentiation. One mechanism by which cyclin D1 is thought to function as a negative regulator of this cellular process is by blocking the transactivation function of MyoD (51, 64). Indeed, in agreement with these previous reports, cotransfections in 10T1/2 fibroblasts showed that while cyclin D1 strongly blocked the ability of MyoD to activate the troponin I gene, cyclin D3 was only partially inhibitory toward MyoD (Fig. 7A). To examine whether NF- κ B inhibits myogenesis through its regulation of cyclin D1, NF- κ B activity was inhibited in 10T1/2 cells through expression of I κ B α SR and cyclin D1 was reexpressed to determine if the block on MyoD transcriptional function could be restored. The inhibitory potential of the I κ B α SR plasmid in these cells was confirmed on an NF- κ B-responsive promoter (data not shown). The results showed that increasing the amounts of the cyclin D1 expression plasmid could restore the inhibition of MyoD activity in cells where NF- κ B was inhibited (Fig. 7A). In comparison, similar cotransfections with the cyclin D3 expression vector were significantly less effective at inhibiting MyoD function. Impor-

tantly, similar results were obtained when myogenesis was assessed by scoring for myotube formation (Fig. 7B), demonstrating that these effects were not specific to the troponin I gene.

To further explore whether cyclin D1 is involved in NF- κ B regulation of myogenesis, NF- κ B was activated by TNF- α while C2C12 myoblasts were induced to undergo differentiation. The results showed that TNF treatment almost completely repressed the ability of C2C12 cells to form myotubes, again supporting the claim that NF- κ B functions as a negative regulator of myogenesis (Fig. 8). In addition, while the levels of cyclin D1 were diminished in untreated differentiating cells, cyclin D1 was stabilized in differentiating C2C12 cells activated for NF- κ B. Although it is possible that these effects of TNF occur independently of NF- κ B activation, this result, taken together with previous data, strongly suggests that one mechanism by which NF- κ B inhibits skeletal muscle differentiation is through the regulation of cyclin D1 expression.

NF- κ B is a direct transcriptional activator of cyclin D1. The use of the C2C12 myogenesis model allowed us to demonstrate that NF- κ B regulates cyclin D1 expression. Next, we were interested in determining whether this regulation was tissue specific and at what level it was controlled. To address the first of these points, an adenoviral delivery system was used to transiently overexpress I κ B α SR in HeLa cells and MEFs. Northern analysis showed that, similar to what was seen in C2C12 cells stably expressing I κ B α SR, transient inhibition of NF- κ B activity led to a decrease in steady-state levels of cyclin D1 mRNA (Fig. 9A). This result demonstrated that this regulation was therefore not unique to skeletal muscle cells and, perhaps equally important, did not result from a clonally selectable process. Conversely, transfections in fibroblasts with a plasmid expressing the p65 subunit of NF- κ B led to increased levels of endogenous cyclin D1 (Fig. 9B), thus underscoring the specificity of the I κ B α SR protein and demonstrating that expression of p65 is sufficient to induce cyclin D1 expression.

Second, to investigate the mechanism of this regulation, cotransfections were performed with reporter plasmids containing different lengths of the human cyclin D1 promoter and an expression plasmid for p65. These experiments showed that p65 strongly activated cyclin D1 gene expression and that this regulation mapped to regions from -963 to -630 and -66 to -22 within the promoter (Fig. 10A). Transcriptional activation of the cyclin D1 promoter was also observed when c-Rel was expressed in these cells (data not shown), indicating that this regulation is not specific to the p65 subunit of NF- κ B. Examination of the sequence from the human cyclin D1 promoter (47) identified potential NF- κ B-binding sites at positions -858, -749, and -39 that matched the NF- κ B consensus binding sequence, GGG(G/A)NNYYCC, and that mapped to the regions we had identified to be regulated by p65. Oligonucleotides to these sites were generated and tested by EMSA to determine whether NF- κ B binding occurred. With nuclear extracts prepared from both NIH 3T3 and HeLa cells, complexes were formed at all three sites (Fig. 10B). TNF treatment, which activates nuclear translocation of NF- κ B, displayed increased complex formation, providing greater evidence that NF- κ B was the component bound to these sites. Supershift analysis determined that NF- κ B complexes were predominantly represented by the p65 subunit and that the -39 site contained both the p50 homodimer and p50-p65 heterodimer complexes, since at this site both antibodies either supershifted or blocked complex formations (Fig. 10C). Furthermore, oligonucleotide competition analysis established that the binding of these complexes was specific to κ B sites (Fig. 10C). Finally, site-directed changes made in the NF- κ B site located at position -39, within

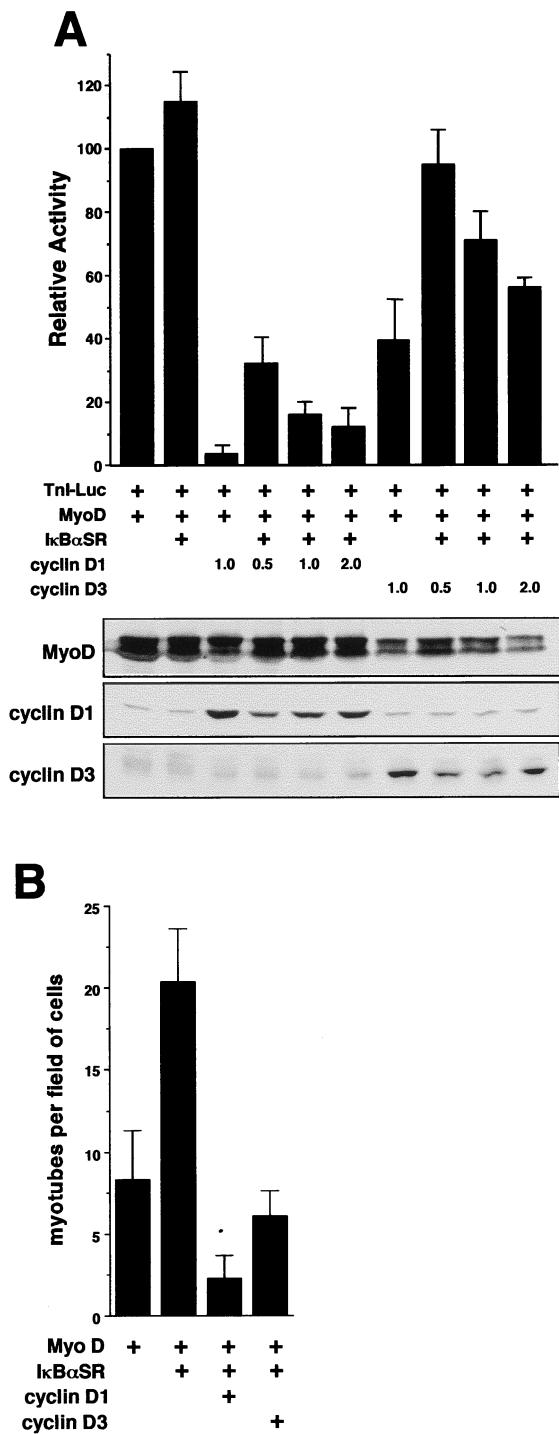


FIG. 7. NF-κB inhibits MyoD transactivation function through the regulation of cyclin D1. (A) 10T1/2 fibroblasts were seeded in triplicate overnight in 6-cm culture dishes, and the next day cotransfection was performed with DNA consisting of 1 μg of the troponin-I reporter plasmid (TnI-Luc) and 0.25 μg of a MyoD expression plasmid, along with either 0.25 μg of an IκBαSR expression plasmid or the indicated amounts of cyclin D1 or cyclin D3 expression plasmid. DNA was normalized by the addition of Bluescript plasmid (Stratagene). Transfected cells were maintained in growth medium overnight and on the following day were switched to DM for 48 h, at which time extracts were prepared and a luciferase assay was performed. The level of troponin-I activation by MyoD alone was set to a value of 100. (Below) To verify the expression of proteins, parallel transfections were performed and immunoblot analysis was performed to probe for MyoD, cyclin D1, and cyclin D3. (B) Similar transfections were performed in 10T1/2 fibroblasts with the following amounts of expression plasmids: 0.25 μg of MyoD, 1 μg of IκBαSR, and 2 μg of cyclin D1 or cyclin D3. The cells were differentiated as described above for 72 h, at which point myogenesis was quantitated by counting myotubes from a minimum of 10 fields of cells.

the −66CD1-Luc reporter plasmid, demonstrated the requirement of this regulatory site for cyclin D1 transcriptional activation relative to expression of the NF-κB p65 subunit (Fig. 10D). Taken together, these data conclusively demonstrate that NF-κB regulation of cyclin D1 occurs at the transcriptional level mediated by direct binding of NF-κB to potentially multiple regions within the cyclin D1 promoter.

NF-κB is required in early G₁ for cyclin D1 activation and progression into S phase. Having established that cyclin D1 is a novel transcriptional target of NF-κB and provided evidence to show how this regulation is important for controlling the differentiation of skeletal muscle cells, we focused the last part of our study on examining the relevance of this regulatory mechanism to cell cycle progression and cell growth. Cyclin D1 was originally identified as a suppressor of G₁ arrest in yeast cells and as a gene that is inducible in the G₁ phase of the cell cycle (44, 76). The expression of cyclin D1 in G₁ is important for cell cycle progression (60, 61), and results demonstrating that acceleration into S phase by cyclin D1 is greater for synchronized cultures emerging from quiescence than for asynchronous cycling cells suggest a specialized role for this D-type cyclin in the G₀/G₁ transition (53). Interestingly, earlier results from our laboratory showed that NF-κB was strongly activated when growth-arrested fibroblasts were stimulated by serum to re-enter cell cycle (6). This finding prompted us to investigate whether NF-κB was required for cyclin D1 induction in cells reinitiating cell cycle.

Since differentiated C2C12 cells are not capable of reinitiating a cell cycle, we instead turned to the use of diploid fibroblasts. Cotransfections were performed in quiescent fibroblasts with a reporter plasmid containing the cyclin D1 promoter and the IκBαSR expression plasmid, and cells were subsequently stimulated back into the cell cycle with the addition of serum. The results showed that in the absence of the IκBαSR plasmid, cyclin D1 promoter activity was induced approximately fourfold in response to mitogen addition (Fig. 11A). In contrast, inhibition of NF-κB by IκBαSR expression blocked this induction in a dose-dependent manner, indicating that NF-κB is required for cyclin D1 transcriptional activation in early G₁. To examine whether NF-κB regulation of cyclin D1 was also important for the G₁-to-S progression, quiescent MEFs were infected with adenovirus containing either empty vector (CMV) or IκBαSR and cells were allowed to reinitiate the cell cycle. G₁-to-S progression was first assessed by examining the phosphorylation status of pRb. The result of this experiment showed that cells lacking NF-κB contained a substantially lower level of hyperphosphorylated pRb than did CMV control cells (Fig. 11B). Importantly, the decrease in pRb hyperphosphorylation in these cells correlated with a marked reduction of cyclin D1, supporting the notion that the Rb protein is a direct target of the cyclin D1-cdk4 complex. By using similar infection conditions, MEFs lacking NF-κB activity were also shown to be significantly impaired for entry into S phase, as assessed by BrdU incorporation (Fig. 11C). Importantly, the defect in S-phase entry in these cells could be restored by addition of cyclin D1, arguing that NF-κB activation of cyclin D1 is important for G₁-to-S progression.

Finally, based on the above findings, we asked whether NF-κB regulation of cyclin D1 was a relevant mechanism involved in the proliferation of primary fibroblasts. To investigate this, we used a retrovirus delivery system to stably generate a mixed population of MEFs expressing either empty vector or the IκBαSR protein. Again, immunoblot analysis showing that endogenous levels of IκBα were downregulated in IκBαSR-expressing fibroblasts (as shown in Fig. 3A) confirmed that NF-κB activity was effectively blocked (data not shown). When growth rates were monitored, the results showed that over an extended number of passage doublings,

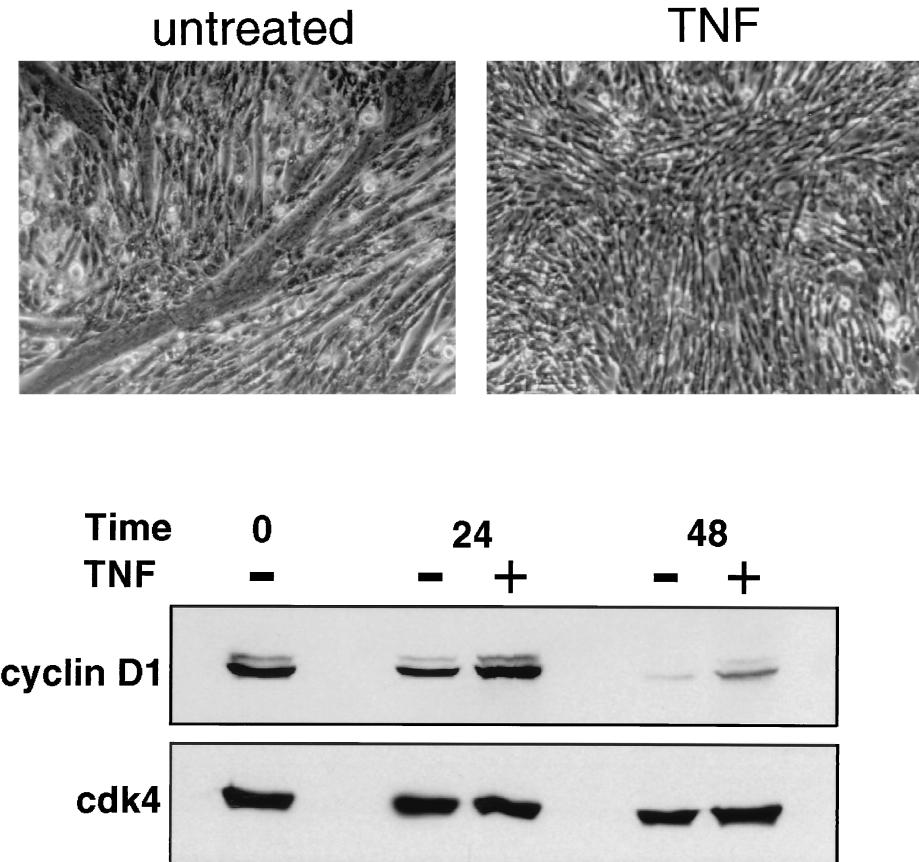


FIG. 8. TNF- α inhibits C2C12 myogenesis and stabilizes cyclin D1. C2C12 cells were induced to differentiate in the absence or presence of 20 ng of TNF- α per ml. Cytokine addition was repeated at 6 h and every additional 12 h after the induction of differentiation. At 72 h, the cells were washed with PBS, fixed for 10 min at room temperature with 4% paraformaldehyde, and photographed by phase-contrast microscopy. In parallel treatment cultures, cells were harvested at 24 and 48 h and whole-cell lysates were prepared for Western blot analysis. A 50- μ g portion of total protein was fractionated by SDS-polyacrylamide gel electrophoresis, and immunoblotting was performed to probe for cyclin D1. The blot was subsequently stripped and reprobed for cdk4, used as an internal control.

fibroblasts devoid of NF- κ B activity exhibited a significant defect in cellular proliferation (Fig. 12). Importantly, NF- κ B inhibition in these cells again led to a persistent decrease in the levels of cyclin D1 (Fig. 12, inset), suggestive that NF- κ B growth-promoting activity is tightly coupled to its transcriptional regulation of cyclin D1.

DISCUSSION

This study was undertaken to gain an insight into how NF- κ B participates in regulating cell growth and differentiation. To examine this, we first used a skeletal differentiation model, since many of the genes that regulate this cellular process have been elucidated. We discovered that in this system NF- κ B activity is reduced as cells undergo differentiation. In myoblasts generated to lack NF- κ B activity, the differentiation was accelerated, suggesting that NF- κ B plays a role in proliferating myoblasts to inhibit their differentiation. This claim was supported in transfection experiments performed with 10T1/2 fibroblasts, where we demonstrated that the inhibitory action of NF- κ B was as potent as oncogenic *ras* and inhibition of myogenesis was specific to the p65 subunit of NF- κ B. Our observation that myoblasts lacking NF- κ B activity increased their cell-doubling time and, upon receiving a differentiation signal, appeared to exit the cell cycle faster than control cells did, led

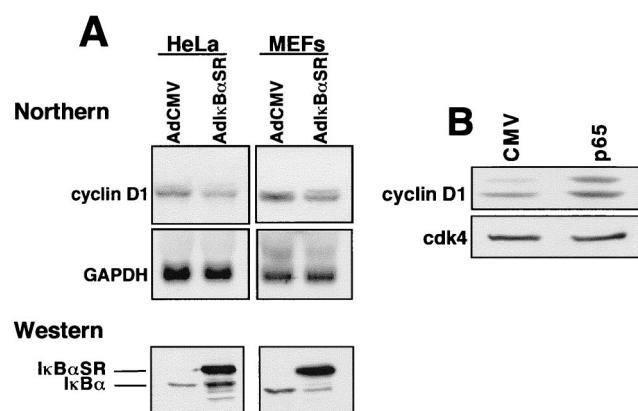


FIG. 9. Regulation of cyclin D1 is specific to NF- κ B occurring in multiple cell types. (A) HeLa cells or MEFs were infected with adenovirus containing either the I κ B α SR or empty vector (AdCMV) at a multiplicity of infection of 50 or 200, respectively. Total RNA was prepared 48 h postinfection, and Northern analysis was performed to detect cyclin D1 mRNA expression. Western blot analysis with an I κ B α -specific antibody is shown to demonstrate the expression of the I κ B α SR in HeLa cells and MEFs by using the adenovirus delivery system. (B) Fibroblasts were transfected with either empty vector or an expression plasmid expressing the p65 subunit of NF- κ B. The following day, cells were switched to serum-deprived conditions for a 48-h period. Subsequently, whole-cell extracts were prepared and 50 μ g was used in immunoblot analysis probing for cyclin D1. The loading efficiency was normalized by reprobing the blot for cdk4 expression.

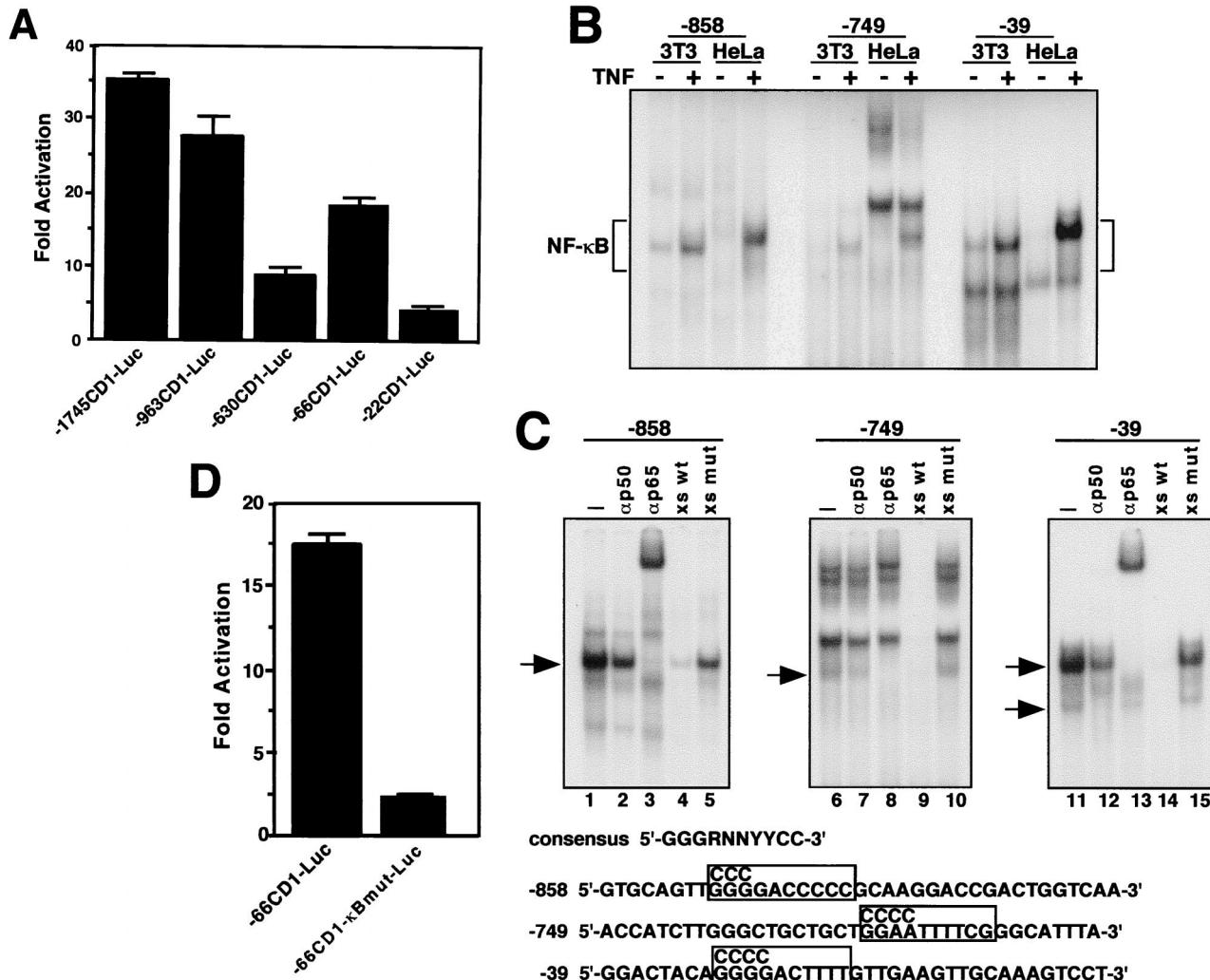


FIG. 10. NF- κ B regulation of cyclin D1 occurs at the transcriptional level. (A) NIH 3T3 cells were plated in triplicate in 12-well plates. Transfections were performed on the following day with Lipofectamine reagent (Life Technologies) mixed with DNA consisting of 0.2 μ g of reporter plasmids containing various 5' deletions of the human cyclin D1 promoter, along with 0.15 μ g of a p65 expression plasmid. DNA remained on the cells for 3 h in serum-free medium, and the cells were then switched for 48 h to complete medium containing 10% calf serum, at which time the luciferase activity was determined. Values were normalized to basal levels of promoter activity obtained by transfecting cyclin D1 promoter constructs in the absence of p65. (B) EMSAs were performed with nuclear extracts prepared from either NIH 3T3 cells or HeLa cells treated (+) or not treated (-) with TNF- α for 30 min. Putative NF- κ B-binding sites, located at positions -858, -749, and -39 in the human cyclin D1 promoter, are indicated within the oligonucleotide sequences used for EMSAs. (C) Supershift EMSAs were performed with nuclear extracts prepared from HeLa cells treated with TNF, which were preincubated either with no addition (lanes 1, 6, and 11) or with addition of antisera specific for p50 (lanes 2, 7, and 12), or the p65 subunit (lanes 3, 8, and 13). Arrows denote p65-containing complexes. For competition EMSAs, extracts were preincubated with either a 100-fold molar excess of unlabeled oligonucleotides containing wild-type (xs wt) NF- κ B-binding sites (lanes 4, 9, and 14), or a 100-fold molar excess of unlabeled oligonucleotides containing mutations in the NF- κ B sites (xs mut) (lanes 5, 10, and 15). The mutations are shown in the boxed regions above the NF- κ B-binding sites. (D) The same mutation at position -39 was made in the NF- κ B-binding site within the -66CD1-Luc reporter plasmid. Both the wild-type and mutant (-66CD1- κ Bmut-Luc) reporter plasmids were separately cotransfected along with a p65 expression plasmid in NIH 3T3 cells under the same transfection conditions as described for panel A.

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us to conclude that NF- κ B regulates cyclin D1 expression. Regulation of this cyclin was shown to be one mechanism by which NF- κ B acted as a negative regulator of myogenesis. We extended our study to examine at what level cyclin D1 regulation by NF- κ B occurred. The results showed that the p65 subunit of NF- κ B was a potent transcriptional activator of the cyclin D1 gene. In addition, we found that NF- κ B bound the cyclin D1 promoter at multiple sites, and for at least the -39 site, we showed by site-directed mutagenesis that NF- κ B sequences are required for transcriptional activation of the cyclin D1 promoter. Furthermore, we demonstrated that NF- κ B activity is required for both cyclin D1 transcriptional activation

and S-phase entry, suggesting an important role for NF- κ B in early G₁. Finally, we showed that similar to immortalized C2C12 cells, primary fibroblasts also lacking NF- κ B activity exhibited reduced proliferation rates in conjunction with lower cyclin D1 levels, arguing that the regulation of this cyclin by NF- κ B is important for proper cell growth control. While our manuscript was in preparation, Hinz et al. reported similar findings that NF- κ B transcriptional regulation of cyclin D1 is necessary in G₁-to-S progression (33). Below, we discuss in greater detail the implications of our findings with respect to the function of NF- κ B in cellular differentiation, the cell cycle, and oncogenesis.

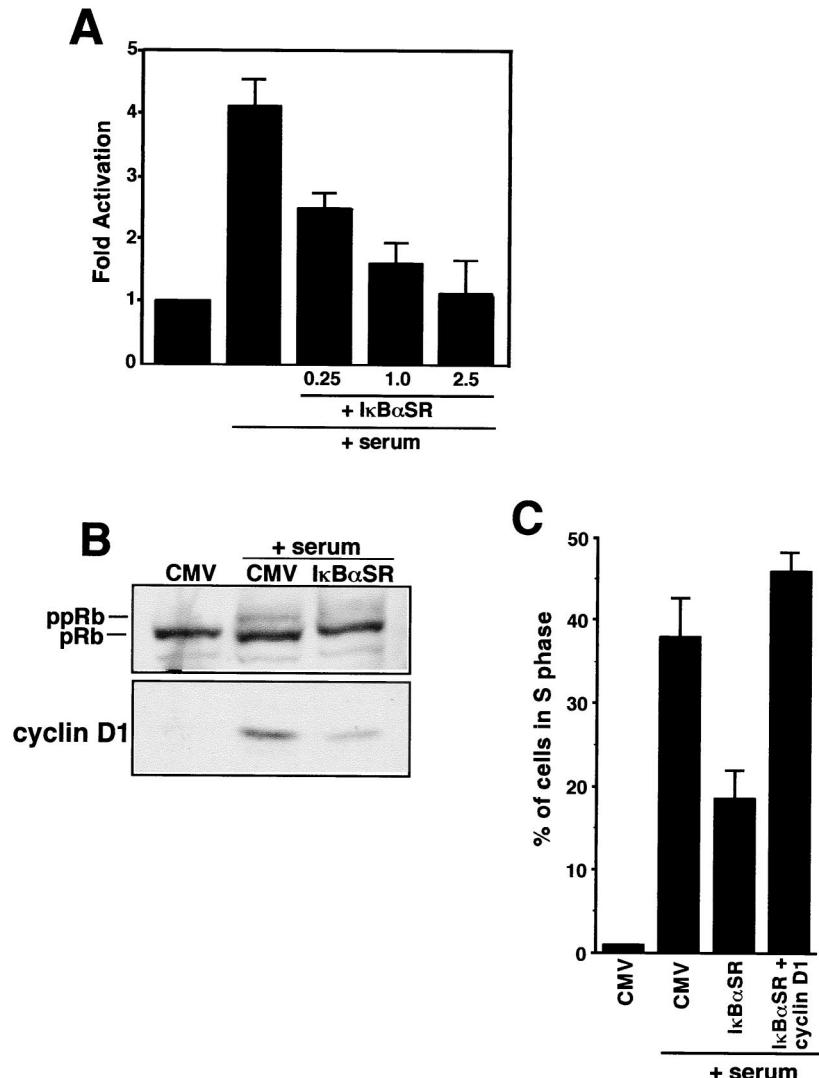


FIG. 11. Requirement for NF- κ B activity in the early G₁ phase of the cell cycle. (A) NIH 3T3 cells were made quiescent by being switched for 48 h from complete medium containing 10% serum to medium containing 0.25% calf serum. Serum-deprived cells were cotransfected either with a cyclin D1 promoter reporter plasmid alone ($-963CD1-Luc$) (2.5 μ g) or in combination with indicated amounts of I_KB_aSR plasmid. The cells were maintained in quiescence or switched to complete medium to induce cyclin D1 transcription and reentry into the cell cycle. Extracts were prepared, and relative luciferase activity was determined by standardizing to total cellular protein. (B) MEFs were made quiescent by culturing cells in 0.1% FBS for 48 h. The cells were then infected under serum-deprived conditions with adenovirus containing either empty vector (CMV) or I_KB_aSR at an MOI of 200. At 24 h postinfections, MEFs were either maintained under serum-deprived conditions or induced to reenter the cell cycle by the addition of 10% FBS. Whole-cell lysates were prepared 24 h later, and immunoblot analysis was performed to probe for both pRb and cyclin D1 (under these culture conditions, we determined that pRb hyperphosphorylation in MEFs is maximally detectable between 20 and 24 h following serum stimulation). (C) Subconfluent MEFs were grown on coverslips overnight and were rendered quiescent on the following day. Cells were infected with either control adenovirus (CMV), I_KB_aSR, or a combination of I_KB_aSR and cyclin D1 viruses (both at a multiplicity of infection of 200). At 24 h following infections, the cells were maintained quiescent or switched for 12 h to growth medium containing 10% serum, at which point the medium was supplemented with 100 μ M BrdU (Sigma) for an additional 10 h. The cells were fixed and prepared for immunofluorescence analysis as described in Materials and Methods. The percentage of cells in the S phase was calculated by determining the number of BrdU-positive cells with respect to the total number of cells in a given field.

A role for NF- κ B as a negative regulator of differentiation. NF- κ B regulation of cyclin D1 gene expression was revealed by examining the role of NF- κ B in a myogenic differentiation model. Results obtained from analyses which included EMSAs, reporter assays, generation of C2C12 cells lacking NF- κ B activity, and transfections in 10T1/2 fibroblasts (Fig. 1 to 4), confirmed that NF- κ B functions, at least in vitro, as a negative regulator of skeletal muscle differentiation. Our results also show that this regulation requires the transcriptional activation function provided by the p65 component of NF- κ B. One point, which will be addressed in future experiments, is

whether the p65 subunit also functions as a negative regulator of myogenesis in vivo. Such an experiment may be difficult to perform since mice with deletions of the p65 subunit die in utero (11) and since functional redundancy is most likely to exist among NF- κ B family members. In addition, genes such as pRb and p21^{WAF1/CIP1}, considered important in regulating skeletal myogenesis in vitro (29, 30), are not required for normal skeletal development in the animal (71). It is also noteworthy that defects in skeletal muscle development were not indicated in cyclin D1 knockout animals (62), underscoring the complexity of this cellular process. Other reports support a

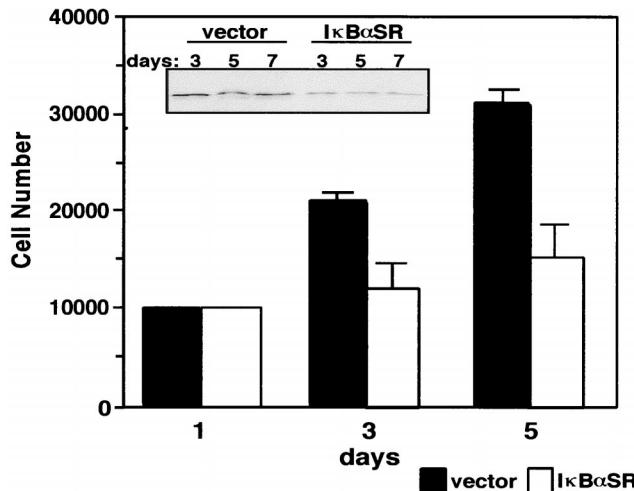


FIG. 12. NF- κ B is required for proper cellular proliferation in primary fibroblasts. Early-passaged MEFs were infected by retrovirus either harboring empty vector or the I κ B α SR. At 2 days following infection, the cells were placed on a 4-day drug selection with 400 μ g of G418 per ml. A mixed population of selected cells was then seeded in triplicate in either 12-well plates or 10-cm dishes. Cells seeded in 12-well plates were trypsinized every 24 to 48 h, and cell numbers were determined by the trypan blue exclusion method. Cells seeded in 10-cm dishes were collected on days 3, 5, and 7, and lysates were prepared for immunoblot analysis detecting for cyclin D1 (inset).

role for NF- κ B in regulating cellular differentiation. For instance, the induction of c-Rel gene expression and DNA-binding activity in mature B cells implicates this NF- κ B subunit in the development of hematopoietic cells (27). Data has also shown that mice with deletions of both p50 and p52 subunits exhibited defects in osteoclast development (25, 35). More recently, the inhibition of NF- κ B activity in keratinocytes, using a similar I κ B α transdominant mutant to that used in the present study, prevented the maturation process of these cells (57). In addition, the inhibition of NF- κ B activity in the proliferative zone of the developing avian limb bud led to impaired growth of this tissue (15, 36). These data, in conjunction with our results, suggest that NF- κ B functions in multiple tissues to regulate their differentiation. We speculate that whether NF- κ B promotes or represses cellular differentiation most probably depends on which specific homo- or heterodimer forms of this transcription factor are represented during the development of a particular tissue and on cell type differences which control distinct transcriptional responses.

Mechanisms of NF- κ B inhibition of myogenesis. C2C12 cells containing the I κ B α SR, and therefore devoid of NF- κ B activity, were observed to be accelerated in their differentiation program (Fig. 3). These cells also exhibited significant reductions in their proliferation rates and, upon receiving a differentiation signal, appeared to show an acceleration in the rate at which they exited the cell cycle (Fig. 5). The defect in the growth of C2C12 cells lacking NF- κ B activity most probably contributes to their ability to rapidly exit cell cycle. Since terminal cell cycle arrest is known to be coupled to myogenic transcription (39), we reasoned that the ability of C2C12-expressing I κ B α SR cells to undergo rapid differentiation is due to the fact that these cells are capable of early G₁ arrest. These results therefore indicate that one mechanism by which NF- κ B inhibits differentiation is through its growth-promoting activity.

Our analysis of cell cycle markers identified the downregulated expression of cyclin D1 in C2C12 cells lacking NF- κ B activity. Since cyclin D1 is an important regulator of cell cycle

progression in many cell types, our results suggest that the premature cell cycle arrest observed in differentiating C2C12-expressing I κ B α SR cells is derived from its lower levels of cyclin D1. These results further imply that the ability of NF- κ B to inhibit myogenic differentiation through its growth-promoting activity is derived directly from its regulation of cyclin D1 expression. This hypothesis is supported by previous findings that ectopic expression of cyclin D1, in association with its catalytic partners cdk4 and cdk6, inhibits myogenesis (64). It should be noted, however, that in these studies the inhibition of myogenesis was assessed by the ability of cyclin D1 to block MyoD transactivation function of muscle-specific genes. In addition, evidence has been presented showing that cyclin D1 inhibition of the transactivation function of MyoD is independent of the activity of this cyclin to phosphorylate pRb (63). These data therefore suggest that the ability of cyclin D1-cdk complexes to inhibit skeletal muscle differentiation is a process that may be uncoupled from the ability of these kinase complexes to promote cell cycle progression. Based on this assumption, it is possible that NF- κ B inhibits myogenesis by at least two distinct mechanisms: (i) by promotion of growth and cell cycle progression independent of cyclin D1 regulation, and (ii) by positive regulation of cyclin D1, which functions to block the activities of myogenic transcription factors (as demonstrated in Fig. 7).

Elucidation of a novel transcriptional target of NF- κ B. Part of our analysis in this study was to identify whether NF- κ B directly regulated cyclin D1 gene expression. The results of cyclin D1 promoter-reporter assays and EMSAs confirmed that NF- κ B regulation of cyclin D1 was mediated at the transcriptional level by authentic NF- κ B binding in at least three sites, -858, -749, and -39 (Fig. 10). Mutational analysis demonstrated that NF- κ B binding to the -39 site was important for cyclin D1 transcriptional regulation. These results therefore indicate that NF- κ B transcriptional regulation of cyclin D1 occurs directly via binding to multiple sites within the promoter region. To further explore this mechanism, we have attempted to address whether NF- κ B, on its own, can induce cyclin D1 mRNA expression when quiescent fibroblasts are stimulated back into the cell cycle. However, Northern analysis indicated that in the absence of ongoing protein synthesis, the cyclin D1 mRNA is highly labile (data not shown), making our interpretation with respect to NF- κ B technically limiting. Therefore, although our results indicate quite clearly that NF- κ B transcriptionally activates cyclin D1, we cannot exclude the possibility that other transcription factors are required that function in a synergistic fashion with NF- κ B to obtain full transcriptional activation of the cyclin D1 gene. Support for this latter hypothesis comes from earlier studies demonstrating that cyclin D1 is transcriptionally regulated separately by the AP-1 and Ets transcription factor complexes (1, 32), both of which are known to physically associate with NF- κ B (8, 67). It is clear that the transcriptional regulatory mechanism of the cyclin D1 gene is complex and requires further detailed experimentation.

Establishing a role for NF- κ B in early G₁ relative to cell growth. Several reports have described an association with NF- κ B activation and the early G₁ phase of the cell cycle. Previously, we found that NF- κ B was strongly activated when quiescent fibroblasts were stimulated by serum addition to reenter the cell cycle (6). Others have described a rapid NF- κ B DNA-binding activity following partial hepatectomy, when hepatocytes progress in the cell cycle from G₀ to G₁ (17, 24). Absence of NF- κ B activity has also been correlated with defects in early G₁, which contributed to a failure of resting B cells to proliferate in response to an activating stimulus (28).

The results in this report showing that NF- κ B is a transcriptional regulator of cyclin D1 now provide a molecular mechanism with which we can better understand the role of this transcription factor in the early G₁ phase of the cell cycle. Based on our findings that cell cycle progression was not affected in cycling immortalized C2C12 cells lacking NF- κ B (Fig. 5 and data not shown), we believe that the relevance of cyclin D1 regulation by NF- κ B may be more meaningful in cells either reentering or exiting the cell cycle. This hypothesis is supported by results obtained with C2C12 cells, where a defect in cell cycle exit was associated with cells lacking NF- κ B activity. Support also comes from our analysis of fibroblasts, where we demonstrated that upon progression from G₀ to G₁ to S, NF- κ B activity is required for cyclin D1 transcriptional activation, pRb hyperphosphorylation, and proper S-phase entry (Fig. 11). These latter findings are consistent with a recent report showing the requirement of NF- κ B activity during the G₁-to-S phase transition, in conjunction with its regulation on cyclin D1 expression (33). The complementary data of both studies, including our additional results showing that embryonic fibroblasts lacking NF- κ B activity also exhibit a severe growth defect correlating with reduced levels of cyclin D1 (Fig. 12), implicates the regulation of cyclin D1 as a mechanism to explain how NF- κ B promotes cell growth. This hypothesis is consistent with the role of cyclin D1 as a regulator of cell growth, since mice lacking cyclin D1 display reduced body size and exhibit a dramatic reduction in cell number in specialized tissues (62). Although cyclin D1^{-/-} fibroblasts were reported to maintain normal growth characteristics when cultured under standard conditions (21), recent evidence suggests that these effects are cell density dependent, since substantial proliferation defects do occur in fibroblasts lacking cyclin D1 when seeded at lower cell density (14).

Implications in oncogenesis. Finally, the results presented in this study may also have broader implications for understanding how NF- κ B participates in oncogenesis. Cyclin D1 levels are deregulated in many human cancers as a result of gene amplification or translocations or of aberrant overexpression (31, 34, 59). Studies with transgenic mice also show that targeted overexpression of cyclin D1 leads to the development of mammary carcinomas (73). In some instances, overexpression of cyclin D1 in transformed cells is regulated by activated *ras* genes (2, 43), which is consistent with findings that *ras* transcriptionally regulates this cyclin (1). Interestingly, our laboratory has found that oncogenic *ras* stimulates the transactivation function of NF- κ B, and that NF- κ B is required by *ras* to induce cellular transformation (23). Therefore, NF- κ B may be considered a component of oncogene-induced signaling pathways leading to the activation of cyclin D1, potentially contributing to the onset and/or progression of oncogenesis.

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REFERENCES

- Albanese, C., J. Johnson, G. Watanabe, N. Eklund, D. Vu, A. Arnold, and R. G. Pestell. 1995. Transforming p21ras mutants and c-Ets-2 activate the cyclin D1 promoter through distinguishable regions. *J. Biol. Chem.* **270**: 23589–23597.
- Arber, N., T. Sutter, M. Miyake, S. M. Kahn, V. S. Venkatraj, A. Sobrino, D. Warburton, P. R. Holt, and I. B. Weinstein. 1996. Increased expression of cyclin D1 and the Rb tumor suppressor gene in c-K-ras transformed rat enterocytes. *Oncogene* **12**:1903–1908.
- Baeuerle, P. A., and D. Baltimore. 1996. NF-kappa B: ten years after. *Cell* **87**:13–20.
- Baeuerle, P. A., and T. Henkel. 1994. Function and activation of NF-kappa B in the immune system. *Annu. Rev. Immunol.* **12**:141–179.
- Baldwin, A. S., Jr. 1996. The NF-kappa B and I kappa B proteins: new discoveries and insights. *Annu. Rev. Immunol.* **14**:649–683.
- Baldwin, A. S., Jr., J. C. Azizkhan, D. E. Jensen, A. A. Beg, and L. R. Coodly. 1991. Induction of NF-kappa B DNA-binding activity during the G₀-to-G₁ transition in mouse fibroblasts. *Mol. Cell. Biol.* **11**:4943–4951.
- Bargou, R. C., F. Emmerich, D. Krappmann, K. Bommert, M. Y. Mapara, W. Arnold, H. D. Royer, E. Grinstein, A. Greiner, C. Scheidereit, and B. Dorken. 1997. Constitutive nuclear factor-kappaB-RelA activation is required for proliferation and survival of Hodgkin's disease tumor cells. *J. Clin. Investig.* **100**:2961–2969.
- Basuk, A. G., R. T. Anandappa, and J. M. Leiden. 1997. Physical interactions between Ets and NF-kappaB/NFAT proteins play an important role in their cooperative activation of the human immunodeficiency virus enhancer in T cells. *J. Virol.* **71**:3563–3573.
- Beg, A. A., and D. Baltimore. 1996. An essential role for NF- κ B in preventing TNF- α -induced cell death. *Science* **274**:782–784.
- Beg, A. A., S. M. Ruben, R. J. Scheinman, S. Haskill, C. A. Rosen, and A. S. Baldwin, Jr. 1992. I kappa B interacts with the nuclear localization sequences of the subunits of NF-kappa B: a mechanism for cytoplasmic retention. *Genes Dev.* **6**:1899–1913.
- Beg, A. A., W. C. Sha, R. T. Bronson, S. Ghosh, and D. Baltimore. 1995. Embryonic lethality and liver degeneration in mice lacking the RelA component of NF-kappa B. *Nature* **376**:167–170.
- Beneza, R., R. L. Davis, D. Lockshon, D. L. Turner, and H. Weintraub. 1990. The protein Id: a negative regulator of helix-loop-helix DNA binding proteins. *Cell* **61**:49–59.
- Brockman, J. A., D. C. Scherer, T. A. McKinsey, S. M. Hall, X. Qi, W. Y. Lee, and D. W. Ballard. 1995. Coupling of a signal response domain in I κ B α to multiple pathways for NF- κ B activation. *Mol. Cell. Biol.* **15**:2809–2818.
- Brown, J. R., E. Nigh, R. J. Lee, H. Ye, M. A. Thompson, F. Saudou, R. G. Pestell, and M. E. Greenberg. 1998. Fos family members induce cell cycle entry by activating cyclin D1. *Mol. Cell. Biol.* **18**:5609–5619.
- Bushdid, P. B., D. M. Brantley, F. E. Yull, G. L. Blaauw, L. H. Hoffman, L. Niwander, and L. D. Kerr. 1998. Inhibition of NF- κ B activity results in disruption of the apical ectodermal ridge and aberrant limb morphogenesis. *Nature* **392**:615–618.
- Cheshire, J. L., and A. S. Baldwin, Jr. 1997. Synergistic activation of NF- κ B by tumor necrosis factor alpha and gamma interferon via enhanced I κ B α degradation and de novo I κ B β degradation. *Mol. Cell. Biol.* **17**:6746–6754.
- Cressman, D. E., L. E. Greenbaum, B. A. Haber, and R. Taub. 1994. Rapid activation of post-hepatectomy factor/nuclear factor kappa B in hepatocytes, a primary response in the regenerating liver. *J. Biol. Chem.* **269**:30429–30435.
- Dejardin, E., G. Bonizzi, A. Bellahcene, V. Castronovo, M. P. Merville, and V. Bours. 1995. Highly-expressed p100/p52 (NFKB2) sequesters other NF-kappa B-related proteins in the cytoplasm of human breast cancer cells. *Oncogene* **11**:1835–1841.
- DiDonato, J. A., M. Hayakawa, D. M. Rothwarf, E. Zandi, and M. Karin. 1997. A cytokine-responsive I κ kappaB kinase that activates the transcription factor NF- κ p κ B. *Nature* **388**:548–554.
- Doi, T. S., T. Takahashi, O. Taguchi, T. Azuma, and Y. Obata. 1997. NF- κ B RelA-deficient lymphocytes: normal development of T cells and B cells, impaired production of IgA and IgG1 and reduced proliferative responses. *J. Exp. Med.* **185**:953–961.
- Fantl, V., G. Stamp, A. Andrews, I. Rosewell, and C. Dickson. 1995. Mice lacking cyclin D1 are small and show defects in eye and mammary gland development. *Genes Dev.* **9**:2364–2372.
- Finco, T. S., and A. S. Baldwin. 1995. Mechanistic aspects of NF- κ B regulation: the emerging role of phosphorylation and proteolysis. *Immunity* **3**:263–272.
- Finco, T. S., J. K. Westwick, J. L. Norris, A. A. Beg, C. J. Der, and A. S. Baldwin, Jr. 1997. Oncogenic Ha-Ras-induced signaling activates NF- κ p κ B transcriptional activity, which is required for cellular transformation. *J. Biol. Chem.* **272**:24113–24116.
- Fitzgerald, M. J., E. M. Webber, J. R. Donovan, and N. Fausto. 1995. Rapid DNA binding by nuclear factor kappa B in hepatocytes at the start of liver

- regeneration. *Cell Growth Differ.* **6**:417–427.
25. Franzoso, G., L. Carlson, L. Xing, L. Poljak, E. W. Shores, K. D. Brown, A. Leonardi, T. Tran, B. F. Boyce, and U. Siebenlist. 1997. Requirement for NF-κappaB in osteoclast and B-cell development. *Genes Dev.* **11**:3482–3496.
 26. Gerritsen, M. E., A. J. Williams, A. S. Neish, S. Moore, Y. Shi, and T. Collins. 1997. CREB-binding protein/p300 are transcriptional coactivators of p65. *Proc. Natl. Acad. Sci. USA* **94**:2927–2932.
 27. Grumont, R. J., and S. Gerondakis. 1994. The subunit composition of NF-κappa B complexes changes during B-cell development. *Cell Growth Differ.* **5**:1321–1331.
 28. Grumont, R. J., I. J. Rourke, L. A. O'Reilly, A. Strasser, K. Miyake, W. Sha, and S. Gerondakis. 1998. B lymphocytes differentially use the rel and nuclear factor κB1(NF-κB1) transcription factors to regulate cell cycle progression and apoptosis in quiescent and mitogen-activated cells. *J. Exp. Med.* **187**:663–674.
 29. Guo, K., J. Wang, V. Andres, R. C. Smith, and K. Walsh. 1995. MyoD-induced expression of p21 inhibits cyclin-dependent kinase activity upon myocyte terminal differentiation. *Mol. Cell. Biol.* **15**:3823–3829.
 30. Halevy, O., B. G. Novitch, D. B. Spicer, S. X. Skapek, J. Rhee, G. J. Hannon, D. Beach, and A. B. Lassar. 1995. Correlation of terminal cell cycle arrest of skeletal muscle with induction of p21 by MyoD. *Science* **267**:1018–1021.
 31. Hall, M., and G. Peters. 1996. Genetic alterations of cyclins, cyclin-dependent kinases, and Cdk inhibitors in human cancer. *Adv. Cancer Res.* **68**:67–108.
 32. Herber, B., M. Truss, M. Beato, and R. Muller. 1994. Inducible regulatory elements in the human cyclin D1 promoter. *Oncogene* **9**:1295–1304.
 33. Hinz, M., D. Krappmann, A. Eichten, A. Heder, C. Scheidereit, and M. Strauss. 1999. NF-κB function in growth control: regulation of cyclin D1 expression and G₀/G₁-to-S-phase transition. *Mol. Cell. Biol.* **19**:2690–2698.
 34. Hunter, T., and J. Pines. 1994. Cyclins and cancer. II: cyclin D and CDK inhibitors come of age. *Cell* **79**:573–582.
 35. Iotsova, V., J. Caamano, J. Loy, Y. Yang, A. Lewin, and R. Bravo. 1997. Osteopetrosis in mice lacking NF-κappaB1 and NF-κappaB2. *Nat. Med.* **3**:1285–1289.
 36. Kanegae, Y., A. T. Tavares, J. C. I. Belmonte, and I. M. Verma. 1998. Role of Rel/NF-κB transcription factors during the outgrowth of the vertebrate limb. *Nature* **392**:611–614.
 37. Konieczny, S. F., B. L. Drobis, S. L. Menke, and E. J. Taparowsky. 1989. Inhibition of myogenic differentiation by the H-ras oncogene is associated with the down regulation of the MyoD1 gene. *Oncogene* **4**:473–481.
 38. Kontgen, F., R. J. Grumont, A. Strasser, D. Metcalf, R. Li, D. Tarlinton, and S. Gerondakis. 1995. Mice lacking the c-rel proto-oncogene exhibit defects in lymphocyte proliferation, humoral immunity, and interleukin-2 expression. *Genes Dev.* **9**:1965–1977.
 39. Lassar, A. B., S. X. Skapek, and B. Novitch. 1994. Regulatory mechanisms that coordinate skeletal muscle differentiation and cell cycle withdrawal. *Curr. Opin. Cell Biol.* **6**:788–794.
 40. Lassar, A. B., M. J. Thayer, R. W. Overell, and H. Weintraub. 1989. Transformation by activated ras or fos prevents myogenesis by inhibiting expression of MyoD1. *Cell* **58**:659–667.
 41. Li, Z., and G. J. Nabel. 1997. A new member of the I kappaB protein family, I kappaB epsilon, inhibits RelA (p65)-mediated NF-κappaB transcription. *Mol. Cell. Biol.* **17**:6184–6190.
 42. Liou, H. C., W. C. Sha, M. L. Scott, and D. Baltimore. 1994. Sequential induction of NF-κappa B/Rel family proteins during B-cell terminal differentiation. *Mol. Cell. Biol.* **14**:5349–5359.
 43. Liu, J. J., J. R. Chao, M. C. Jiang, S. Y. Ng, J. J. Yen, and H. F. Yang-Yen. 1995. Ras transformation results in an elevated level of cyclin D1 and acceleration of G1 progression in NIH 3T3 cells. *Mol. Cell. Biol.* **15**:3654–3663.
 44. Matsushima, H., M. F. Roussel, R. A. Ashmun, and C. J. Sherr. 1991. Colony-stimulating factor 1 regulates novel cyclins during the G1 phase of the cell cycle. *Cell* **65**:701–713.
 45. Mayo, M. W., C.-Y. Wang, P. C. Cogswell, K. S. Rogers-Graham, S. W. Lowe, C. J. Der, and A. S. J. Baldwin. 1997. Requirement of NF-κB activation to suppress p53-independent apoptosis induced by oncogenic ras. *Science* **278**:1812–1815.
 46. Mercurio, F., H. Zhu, B. W. Murray, A. Shevchenko, B. L. Bennett, J. Li, D. B. Young, M. Barbosa, M. Mann, A. Manning, and A. Rao. 1997. IKK-1 and IKK-2: cytokine-activated IkappaB kinases essential for NF-κappaB activation. *Science* **278**:860–866.
 47. Motokura, T., and A. Arnold. 1993. PRAD1/cyclin D1 proto-oncogene: genomic organization, 5' DNA sequence, and sequence of a tumor-specific rearrangement breakpoint. *Genes Chromosomes Cancer* **7**:89–95.
 48. Olson, E. N. 1992. Interplay between proliferation and differentiation within the myogenic lineage. *Dev. Biol.* **154**:261–272.
 49. Olson, E. N., G. Spizz, and M. A. Tainsky. 1987. The oncogenic forms of N-ras or H-ras prevent skeletal myoblast differentiation. *Mol. Cell. Biol.* **7**:2104–2111.
 50. Perkins, N. D., L. K. Felzen, J. C. Betts, K. Leung, D. H. Beach, and G. J. Nabel. 1997. Regulation of NF-κappaB by cyclin-dependent kinases associated with the p300 coactivator. *Science* **275**:523–527.
 51. Rao, S. S., C. Chu, and D. S. Kohtz. 1994. Ectopic expression of cyclin D1 prevents activation of gene transcription by myogenic basic helix-loop-helix regulators. *Mol. Cell. Biol.* **14**:5259–5267.
 52. Regnier, C. H., H. Y. Song, X. Gao, D. V. Goeddel, Z. Cao, and M. Rothe. 1997. Identification and characterization of an IkappaB kinase. *Cell* **90**:373–383.
 53. Resnitzky, D., M. Gossen, H. Bujard, and S. I. Reed. 1994. Acceleration of the G1/S phase transition by expression of cyclins D1 and E with an inducible system. *Mol. Cell. Biol.* **14**:1669–1679.
 54. Reuther, J. Y., G. W. Reuther, D. Cortez, A. M. Pendergast, and A. S. Baldwin, Jr. 1997. A requirement for NF-κB activation in Bcr-Abl-mediated transformation. *Genes Dev.* **12**:968–981.
 55. Sasaki, Y., R. Kageyama, Y. Tagawa, R. Shigemoto, and S. Nakanishi. 1992. Two mammalian helix-loop-helix factors structurally related to *Drosophila* hairy and Enhancer of split. *Genes Dev.* **6**:2620–2634.
 56. Schmitz, M. L., and P. A. Baeuerle. 1991. The p65 subunit is responsible for the strong transcription activating potential of NF-κappa B. *EMBO J.* **10**:3805–3817.
 57. Seitz, C. S., Q. Lin, H. Deng, and P. A. Khavari. 1998. Alterations in NF-κB function in transgenic epithelial tissue demonstrate a growth inhibitory role for NF-κB. *Proc. Natl. Acad. Sci. USA* **95**:2307–2312.
 58. Sha, W. C., H. C. Liou, E. I. Tuomanen, and D. Baltimore. 1995. Targeted disruption of the p50 subunit of NF-κappa B leads to multifocal defects in immune responses. *Cell* **80**:321–330.
 59. Sherr, C. J. 1996. Cancer cell cycles. *Science* **274**:1672–1677.
 60. Sherr, C. J. 1995. D-type cyclins. *Trends Biochem. Sci.* **20**:187–190.
 61. Sherr, C. J. 1994. G1 phase progression: cycling on cue. *Cell* **79**:551–555.
 62. Sicinski, P., J. L. Donaher, S. B. Parker, T. Li, A. Fazeli, H. Gardner, S. Z. Haslam, R. T. Bronson, S. J. Elledge, and R. A. Weinberg. 1995. Cyclin D1 provides a link between development and oncogenesis in the retina and breast. *Cell* **82**:621–630.
 63. Skapek, S. X., J. Rhee, P. S. Kim, B. G. Novitch, and A. B. Lassar. 1996. Cyclin-mediated inhibition of muscle gene expression via a mechanism that is independent of pRB hyperphosphorylation. *Mol. Cell. Biol.* **16**:7043–7053.
 64. Skapek, S. X., J. Rhee, D. B. Spicer, and A. B. Lassar. 1995. Inhibition of myogenic differentiation in proliferating myoblasts by cyclin D1-dependent kinase. *Science* **267**:1022–1024.
 65. Snapper, C. M., P. Zelazowski, F. R. Rosas, M. R. Kehry, M. Tian, D. Baltimore, and W. C. Sha. 1996. B cells from p50/NF-κappa B knockout mice have selective defects in proliferation, differentiation, germ-line CH transcription, and Ig class switching. *J. Immunol.* **156**:183–191.
 66. Sovak, M. A., R. E. Bellas, D. W. Kim, G. J. Zanieski, A. E. Rogers, A. M. Traish, and G. E. Sonenschein. 1997. Aberrant nuclear factor-κappaB/Rel expression and the pathogenesis of breast cancer. *J. Clin. Investig.* **100**:2952–2960.
 67. Stein, B., A. S. Baldwin, Jr., D. W. Ballard, W. C. Greene, P. Angel, and P. Herrlich. 1993. Cross-coupling of the NF-κappa B p65 and Fos/Jun transcription factors produces potentiated biological function. *EMBO J.* **12**:3879–3891.
 68. Thanos, D., and T. Maniatis. 1995. NF-κappa B: a lesson in family values. *Cell* **80**:529–532.
 69. Van Antwerp, D. J., S. J. Martin, K. Tal, D. R. Green, and I. M. Verma. 1996. Suppression of TNF-α-induced apoptosis by NF-κB. *Science* **274**:787–789.
 70. Verma, I. M., J. K. Stevenson, E. M. Schwartz, D. Van Antwerp, and S. Miyamoto. 1995. Rel/NF-κB/IκB family: intimate tales of association and dissociation. *Genes Dev.* **9**:2723–2735.
 71. Walsh, K., and H. Perlman. 1997. Cell cycle exit upon myogenic differentiation. *Curr. Opin. Genet. Dev.* **7**:597–602.
 72. Wang, C.-Y., M. W. Mayo, and A. S. J. Baldwin. 1996. TNF- and cancer therapy-induced apoptosis: potentiation by inhibition of NF-κB. *Science* **274**:784–787.
 73. Wang, T. C., R. D. Cardiff, L. Zukerberg, E. Lees, A. Arnold, and E. V. Schmidt. 1994. Mammary hyperplasia and carcinoma in MMTV-cyclin D1 transgenic mice. *Nature* **369**:669–671.
 74. Whiteside, S. T., J. C. Epinat, N. R. Rice, and A. Israel. 1997. I kappa B epsilon, a novel member of the I kappa B family, controls RelA and cRel NF-κappa B activity. *EMBO J.* **16**:1413–1426.
 75. Wu, M., H. Lee, R. E. Bellas, S. L. Schauer, M. Arsura, D. Katz, M. J. Fitzgerald, T. L. Rothstein, D. H. Sherr, and G. E. Sonenschein. 1996. Inhibition of NF-κappaB/Rel induces apoptosis of murine B cells. *EMBO J.* **15**:4682–4690.
 76. Xiong, Y., T. Connolly, B. Futcher, and D. Beach. 1991. Human D-type cyclin. *Cell* **65**:691–699.
 77. Zandi, E., D. M. Rothwarf, M. Delhase, M. Hayakawa, and M. Karin. 1997. The IkappaB kinase complex (IKK) contains two kinase subunits, IKKα and IKKβ, necessary for IkappaB phosphorylation and NF-κappaB activation. *Cell* **91**:243–252.