

# Package ‘fuzzySim’

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**Description** Functions to compute fuzzy versions of species occurrence patterns based on presence-absence data (including inverse distance interpolation, trend surface analysis, and prevalence-independent favourability obtained from probability of presence), as well as pair-wise fuzzy similarity (based on fuzzy logic versions of commonly used similarity indices) among those occurrence patterns. Includes also functions for model consensus and comparison (overlap and fuzzy similarity, loss or gain), and for data preparation, such as obtaining unique abbreviations of species names, gridding (thinning) point occurrence data onto raster maps, converting species lists (long format) to presence-absence tables (wide format), transposing part of a data frame, selecting relevant variables for models, assessing the false discovery rate, or analysing and dealing with multicollinearity. Initially described in Barbosa (2015) <[doi:10.1111/2041-210X.12372](https://doi.org/10.1111/2041-210X.12372)>.

**License** GPL-3

**URL** fuzzysim.r-forge.r-project.org,

<https://CRAN.R-project.org/package=fuzzySim>

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fuzzySim-package

*Fuzzy Similarity in Species Distributions***Description**

Functions to compute fuzzy versions of species occurrence patterns based on presence-absence data (including inverse distance interpolation, trend surface analysis, and prevalence-independent favourability obtained from probability of presence), as well as pair-wise fuzzy similarity (based on fuzzy logic versions of commonly used similarity indices) among those occurrence patterns. Includes also functions for model consensus and comparison (overlap and fuzzy similarity, loss or gain), and for data preparation, such as obtaining unique abbreviations of species names, grid-ding (thinning) point occurrence data onto raster maps, converting species lists (long format) to

presence-absence tables (wide format), transposing part of a data frame, selecting relevant variables for models, assessing the false discovery rate, or analysing and dealing with multicollinearity. Includes also sample datasets for providing practical examples. A step-by-step illustrated tutorial is available from the package homepage (<http://fuzzysim.r-forge.r-project.org>).

## Details

Package: fuzzySim  
Type: Package  
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License: GPL-3

## Author(s)

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## References

Barbosa A.M. (2015) fuzzySim: applying fuzzy logic to binary similarity indices in ecology. *Methods in Ecology and Evolution*, 6: 853-858.

## Examples

```
data(rotifers)

head(rotifers)

# add column with species name abbreviations:

rotifers$spcode <- spCodes(rotifers$species, sep.species = "_",
nchar.gen = 1, nchar.sp = 5, nchar.ssp = 0)

head(rotifers)

# convert species list (long format) to presence-absence table
# (wide format):

rotifers.presabs <- splist2presabs(rotifers, sites.col = "TDWG4",
sp.col = "spcode", keep.n = FALSE)

head(rotifers.presabs)
```

```
# get 3rd-degree spatial trend surface for some species distributions:

data(rotif.env)

names(rotif.env)

rotifers.tsa <- multTSA(rotif.env, sp.cols = 18:20,
coord.cols = c("Longitude", "Latitude"), id.col = 1)

head(rotifers.tsa)

# get inverse squared distance to presence for each species:

rotifers.isqd <- distPres(rotif.env, sp.cols = 18:20,
coord.cols = c("Longitude", "Latitude"), id.col = 1, p = 2, inv = TRUE)

head(rotifers.isqd)

# get prevalence-independent environmental favourability models
# for each species:

data(rotif.env)

names(rotif.env)

rotifers.fav <- multGLM(data = rotif.env, sp.cols = 18:20,
var.cols = 5:17, id.col = 1, step = FALSE, trim = TRUE,
Favourability = TRUE)

# get matrix of fuzzy similarity between species distributions:

# either based on inverse squared distance to presence:
rot.fuz.sim.mat <- simMat(rotifers.isqd[ , -1], method = "Jaccard")

# or on environmental favourability for presence:
rot.fuz.sim.mat <- simMat(rotifers.fav$predictions[ , 5:7],
method = "Jaccard")

head(rot.fuz.sim.mat)

# transpose fuzzy rotifer distribution data to compare
# regional species composition rather than species' distributions:

names(rotifers.isqd)

rot.fuz.reg <- transpose(rotifers.fav$predictions, sp.cols = 5:7,
reg.names = 1)

head(rot.fuz.reg)
```

```
# get matrix of fuzzy similarity between (some) regions'
# species compositions:

reg.fuz.sim.mat <- simMat(rot.fuz.reg[, 1:10], method = "Jaccard")

head(reg.fuz.sim.mat)
```

---

bioThreat	<i>Biotic threat of a stronger over a weaker species based on their favourability values</i>
-----------	--

---

## Description

This function takes two vectors of [Favourability](#) values at different localities for, respectively, a stronger and a weaker species (e.g., a superior vs. an inferior competitor, or an invasive predator vs. an unadapted native prey), and calculates the level of threat that the former may potentially pose to the latter in each locality.

## Usage

```
bioThreat(strong_F, weak_F, character = FALSE, ...)
```

## Arguments

strong_F	a numeric vector of favourability values (obtained, e.g., with functions <a href="#">Fav</a> or <a href="#">multGLM</a> ) for the stronger species.
weak_F	a numeric vector of favourability values for the weaker species. Must be of the same length and in the same order as 'strong_F'.
character	logical value indicating whether the result should be returned in character rather than numeric form. Defaults to FALSE.
...	additional arguments to pass to <a href="#">favClass</a> , namely the breaks for separating favourability values into low, intermediate and high (see Details).

## Details

Based on the notion of "favorableness" by Richerson & Lum (1980), according to which competing species may or may not be able to coexist depending on their relative environmental fitnesses, Acevedo et al. (2010, 2012) and some subsequent studies (e.g. Romero et al. 2014, Munoz et al. 2015, Chamorro et al. 2019) proposed possible biotic interaction outcomes of different combinations of favourability values for two species. Favourability has the advantage, in contrast with other types of potential distribution metrics, of being directly comparable among different species, independently of their relative occurrence frequencies (see [Fav](#)). This function builds on those proposals by including additional possible combinations of higher, intermediate or low favourability values (following Munoz & Real 2006), producing the following classification of biotic threat across a set of analysed localities:

0 ('grey'): areas where favourability is low for at least one of the species (abiotic exclusion), so biotic threat does not apply.

1 ('green'): areas where favourability is high for both species, so they should both be able to thrive and therefore co-occur (sympatric coexistence), hence biotic threat is low.

2 ('yellow'): areas where favourability is high for the weaker species and intermediate for the stronger species, so the level of threat is moderate.

3 ('orange'): areas where favourability is intermediate for both species, so the stronger one potentially prevails and the level of threat is high.

4 ('red'): areas where favourability is high for the stronger species and intermediate for the weaker species, in which case the level of threat is very high (biotic exclusion).

### Value

This function returns either an integer or a character vector (following the 'character' argument, which is set to FALSE by default) of the same length as 'strong\_F' and 'weak\_F', classifying each locality with the level of biotic threat posed by the former on the latter (see Details).

### Author(s)

A. Marcia Barbosa

### References

- Acevedo P., Ward A.I., Real R. & Smith G.C. (2010) Assessing biogeographical relationships of ecologically related species using favourability functions: a case study on British deer. *Diversity and Distributions*, 16: 515-528
- Acevedo P., Jimenez-Valverde A., Melo-Ferreira J., Real R. & Alves, P.C. (2012) Parapatric species and the implications for climate change studies: a case study on hares in Europe. *Global Change Biology*, 18: 1509-1519
- Chamorro D., Munoz A.R., Martinez-Freiria F. & Real R. (2019) Using the fuzzy logic in the distribution modelling of competitive interactions. Poster, IBS Malaga 2019 - 9th Biennial Conference of the International Biogeography Society
- Munoz A.R. & Real R. (2006) Assessing the potential range expansion of the exotic monk parakeet in Spain. *Diversity and Distributions*, 12: 656-665
- Munoz A.R., Real R. & Marquez A.L. (2015) Interacciones a escala nacional entre rapaces rupícolas en base a modelos de distribución espacial. Los casos del buitre leonado, alimoche y águila perdicera. Informe técnico, Universidad de Málaga & Fundación EDP
- Richerson P.J. & Lum K. (1980) Patterns of plant species diversity in California: relation to weather and topography. *American Naturalist*, 116:504-536
- Romero D., Baez J.C., Ferri-Yanez F., Bellido J. & Real R. (2014) Modelling favourability for invasive species encroachment to identify areas of native species vulnerability. *The Scientific World Journal*, 2014: 519710

### See Also

[sharedFav](#), [Fav](#), [favClass](#)

## Examples

```
data(rotif.env)
mods <- multGLM(rotif.env, sp.cols = 19:20, var.cols = 5:17)
head(mods$predictions)
favs <- mods$predictions[, 3:4]
threat <- bioThreat(strong_F = favs[,1], weak_F = favs[,2])
threat_chr <- bioThreat(strong_F = favs[,1], weak_F = favs[,2], char = TRUE)
data.frame(favs, threat = threat, threat_col = threat_chr)
```

---

corSelect

---

*Select among correlated variables based on a given criterion*


---

## Description

This function calculates pairwise correlations among the variables in a dataset and, among each pair of variables correlated above a given threshold, excludes the variable with either the highest variance inflation factor (VIF), or the least significant or least informative bivariate (individual) relationship with the response variable (if supplied), according to a specified criterion.

## Usage

```
corSelect(data, sp.cols = NULL, var.cols, cor.thresh = 0.8,
select = "p.value", family = "auto", use = "pairwise.complete.obs", method = "pearson")
```

## Arguments

data	a data frame containing the response and predictor variables.
sp.cols	name or index number of the column of 'data' that contains the response (e.g. species) variable. Currently, only one 'sp.cols' can be used at a time, so an error message is returned if <code>length(sp.cols) &gt; 1</code> . If <code>sp.cols = NULL</code> (the default), the function returns only the pairs of variables that are correlated over the given threshold, without selecting those that are more relevant for a target species.
var.cols	names or index numbers of the columns of 'data' that contain the predictor variables.
cor.thresh	threshold value of correlation coefficient above which (or below which, for negative correlations) predictor variables should be excluded. The default is 0.8.
select	character value indicating the criterion for excluding variables among those that are correlated. Can be "p.value" (the default), "AIC", "BIC", or "VIF" (see Details).
family	If 'sp.col' is not NULL, the error distribution and (optionally) the link function to use for assessing significant / informative variables (see <a href="#">glm</a> or <a href="#">family</a> for details). The default "auto" automatically uses "binomial" family for response variables containing only values of 0 and 1; "poisson" for positive integer responses (i.e. count data); "Gamma" for positive non-integer; and "gaussian" (i.e., linear models) otherwise.

use	argument to pass to <code>cor</code> indicating what to do when there are missing values. Can be "pairwise.complete.obs" (the default here), "everything", "all.obs", "complete.obs", "na.or.complete".
method	argument to pass to <code>cor</code> specifying the correlation coefficient to use. Can be "pearson" (the default), "kendall" or "spearman".

## Details

Correlations among variables are problematic in multivariate models, as they inflate the variance of coefficients and thus may bias the interpretation of the effects of those variables on the response (Legendre & Legendre 2012). One of the strategies to circumvent this problem is to eliminate one from each pair of correlated variables, but it is not always straightforward to choose the right variable to exclude a priori.

This function selects among correlated variables, based either on their variance inflation factor (VIF: Marquardt 1970; Mansfield & Helms 1982) within the variables dataset (obtained with the `multicol` function and recalculated iteratively after each variable exclusion); or on their relationship with the response, by building a bivariate model of each individual variable against the response and excluding, among each of two correlated variables, the one with the largest (worst) p-value, AIC (Akaike's Information Criterion: Akaike, 1973) or BIC (Bayesian Information Criterion, also known as Schwarz criterion, SBC or SBIC: Schwarz, 1978), which it calculates with the `FDR` function.

If 'sp.cols' is left NULL and the 'select' criterion is other than "VIF", the function returns only the pairs of variables that are correlated above the given threshold. If the 'select' criterion requires assessing bivariate relationships and 'sp.cols' is provided, the function uses only the rows of the dataset where this column (used as the response variable) contains finite values against which the predictor variables can be modelled; rows with NA or NaN in 'sp.cols' are thus excluded from the calculation of correlations among predictor variables.

## Value

This function returns a list of 7 elements, unless 'sp.cols = NULL', in which case it returns only the first of these elements:

high.correlations	data frame showing the pairs of input variables that are correlated beyond the given threshold, and their correlation coefficient.
bivariate.significance	data frame with the individual p-value, AIC and BIC (if one of these was the 'select' criterion) of each of the highly correlated variables against the response variable.
excluded.vars	character vector containing the names of the variables to be excluded (i.e., from each highly correlated pair, the variable with the worse 'select' score).
selected.vars	character vector containing the names of the variables to be selected (i.e., the non-correlated variables and, from each correlated pair, the variable with the better 'select' score).
selected.var.cols	integer vector containing the column indices of the selected variables in 'data'.



`strongest.remaining.corr`  
 numerical value indicating the strongest correlation coefficient among the selected variables.

`remaining.multicollinearity`  
 data frame showing the [multicollinearity](#) among the selected variables.

### Author(s)

A. Marcia Barbosa

### References

Akaike, H. (1973) Information theory and an extension of the maximum likelihood principle. In: Petrov B.N. & Csaki F., 2nd International Symposium on Information Theory, Tsahkadsor, Armenia, USSR, September 2-8, 1971, Budapest: Akademiai Kiado, p. 267-281.

Legendre P. & Legendre L. (2012) Numerical ecology (3rd edition). Elsevier, Amsterdam: 990 pp.

Marquardt D.W. (1970) Generalized inverses, ridge regression, biased linear estimation, and non-linear estimation. *Technometrics* 12: 591-612.

Mansfield E.R. & Helms B.P. (1982) Detecting multicollinearity. *The American Statistician* 36: 158-160.

Schwarz, G.E. (1978) Estimating the dimension of a model. *Annals of Statistics*, 6 (2): 461-464.

### See Also

[multicol](#), [FDR](#), [cor](#)

### Examples

```
data(rotif.env)

corSelect(rotif.env, var.cols = 5:17)

corSelect(rotif.env, sp.cols = 46, var.cols = 5:17)

corSelect(rotif.env, sp.cols = 46, var.cols = 5:17, cor.thresh = 0.7)

corSelect(rotif.env, sp.cols = 46, var.cols = 5:17, method = "spearman")
```

---

distPres

*(Inverse) distance to the nearest presence*

---

### Description

This function takes a matrix or data frame containing species presence (1) and absence (0) data and their spatial coordinates (optionally also a pre-calculated distance matrix between all localities), and calculates the (inverse) distance from each locality to the nearest presence locality for each species.

**Usage**

```
distPres(data, sp.cols, coord.cols = NULL, id.col = NULL,
dist.mat = NULL, method = "euclidian", suffix = "_D", p = 1,
inv = TRUE)
```

**Arguments**

<code>data</code>	a matrix or data frame containing, at least, two columns with spatial coordinates, and one column per species containing their presence (1) and absence (0) data, with localities in rows.
<code>sp.cols</code>	names or index numbers of the columns containing the species presences and absences in data. It must contain only zeros (0) for absences and ones (1) for presences.
<code>coord.cols</code>	names or index numbers of the columns containing the spatial coordinates in data (in this order, x and y, or longitude and latitude).
<code>id.col</code>	optionally, the name or index number of a column (to be included in the output) containing locality identifiers in data.
<code>dist.mat</code>	optionally, if you do not want distances calculated with any of the methods available in <a href="#">dist</a> , you may provide a distance matrix calculated elsewhere for the localities in data.
<code>method</code>	the method with which to calculate distances between localities. Available options are those of <a href="#">dist</a> . The default is "euclidian".
<code>suffix</code>	character indicating the suffix to add to the distance columns in the resulting data frame. The default is "_D".
<code>p</code>	the power to which distance should be raised. The default is 1; use 2 or higher if you want more conservative distances.
<code>inv</code>	logical value indicating whether distance should be inverted, i.e. standardized to vary between 0 and 1 and then subtracted from 1, so that it varies between 0 and 1 and higher values mean closer to presence. The default is TRUE, which is adequate as a fuzzy version of presence-absence (for using e.g. with <a href="#">fuzSim</a> and <a href="#">simMat</a> ). In this case, presences maintain the value 1, and inverse distance to presence is calculated only for absence localities.

**Details**

This function can be used to calculate a simple spatial interpolation model of a species' distribution (e.g. Barbosa 2015, Areias-Guerreiro et al. 2016).

**Value**

This function returns a matrix or data frame containing the identifier column (if provided in 'id.col') and one column per species containing the distance (inverse by default) from each locality to the nearest presence of that species.

**Author(s)**

A. Marcia Barbosa

## References

Areias-Guerreiro J., Mira A. & Barbosa A.M. (2016) How well can models predict changes in species distributions? A 13-year-old otter model revisited. *Hystrix - Italian Journal of Mammalogy*, in press. DOI: <http://dx.doi.org/10.4404/hystrix-27.1-11867>

Barbosa A.M. (2015) fuzzySim: applying fuzzy logic to binary similarity indices in ecology. *Methods in Ecology and Evolution*, 6: 853-858

## See Also

[dist](#)

## Examples

```
data(rotif.env)

head(rotif.env)

names(rotif.env)

# calculate plain distance to presence:

rotifers.dist <- distPres(rotif.env, sp.cols = 18:47,
  coord.cols = c("Longitude", "Latitude"), id.col = 1, p = 1,
  inv = FALSE, suffix = "_D")

head(rotifers.dist)

# calculate inverse squared distance to presence:

rotifers.invd2 <- distPres(rotif.env, sp.cols = 18:47,
  coord.cols = c("Longitude", "Latitude"), id.col = 1, p = 2,
  inv = TRUE, suffix = "_iDsqr")

head(rotifers.invd2)
```

---

entropy	(Fuzzy) entropy
---------	-----------------

---

## Description

This function computes fuzzy entropy (Kosko 1986, Estrada & Real 2021), or optionally Shannon's (1948) entropy.

## Usage

```
entropy(data, sp.cols = 1:ncol(data), method = "fuzzy", base = exp(1),
  plot = TRUE, na.rm = TRUE, ...)
```

### Arguments

<code>data</code>	a vector, matrix or data frame containing the data to analyse.
<code>sp.cols</code>	names or index numbers of the columns of 'data' that contain the values for which to compute entropy (if 'data' is not a vector). The default is to use all columns.
<code>method</code>	character value indicating the method to use. Can be "fuzzy" (the default) or "Shannon". The former requires the input to be a fuzzy system (e.g. <a href="#">Favourability</a> values), while the latter requires probabilities. If <code>method="Shannon"</code> and the values for a vector or column do not sum up to 1, they are divided by their sum so that this additional requirement is met (Estrada & Real 2021).
<code>base</code>	base for computing the logarithm if <code>method="Shannon"</code> . The default is the natural logarithm.
<code>plot</code>	logical value indicating whether to produce a barplot with the results (if 'data' has more than one column). The default is TRUE.
<code>na.rm</code>	logical value indicating whether NA values should be removed before computations. The default is TRUE.
<code>...</code>	additional arguments to be passed to the <a href="#">barplot</a> function.

### Details

Fuzzy entropy (Kosko 1986) applies to fuzzy systems (such as [Favourability](#)) and it can take values between zero and one. Fuzzy entropy equals one when the distribution of the values is uniform, i.e. 0.5 in all localities. The smaller the entropy, the more orderly the distribution of the values, i.e. the closer they are to 0 or 1, distinguishing (potential) presences and absences more clearly. Fuzzy entropy can reflect the overall degree of uncertainty in a species' distribution model predictions, and it is directly comparable across species and study areas (Estrada & Real 2021).

Shannon's entropy requires that the input values are probabilities and sum up to 1 (Shannon 1948). This makes sense when analysing the probability that a unique event occurs in a finite universe. However, if a species has more than one presence, the sum of probabilities in all localities equals the number of presences. To satisfy the condition that the inputs sum up to 1, this function divides each value by the sum of values when this is not the case (if `method="Shannon"`). Notice that this has a mathematical justification but not a biogeographical sense, and (unlike fuzzy entropy) the results are comparable only between models based on the same number of presences + absences, e.g. in a context of selection of variables for a model (Estrada & Real 2021).

### Value

This function returns a numeric value of entropy for 'data' (if it is a numeric vector) or for each of 'sp.cols' in 'data' (if it is a matrix or data frame). Optionally (and by default), a barplot is also produced with these values (if there is more than one column) for visual comparison.

### Author(s)

A. Marcia Barbosa

## References

- Estrada A. & Real R. (2021) A stepwise assessment of parsimony and fuzzy entropy in species distribution modelling. *Entropy*, 23: 1014
- Kosko B. (1986) Fuzzy entropy and conditioning. *Information Sciences*, 40: 165-174
- Shannon C.E. (1948) A mathematical theory of communication. *Bell System Technical Journal*, 27: 379-423

## Examples

```
data(rotif.env)

pred <- multGLM(rotif.env, sp.cols = 18:20, var.cols = 5:17)$predictions

head(pred)

entropy(pred, sp.cols = c("Abrigh_F", "Afissa_F", "Apriod_F"))

entropy(pred, sp.cols = c("Abrigh_P", "Afissa_P", "Apriod_P"), method = "Shannon")
```

---

Fav

*Favourability (probability without the effect of sample prevalence)*


---

## Description

Computes prevalence-independent favourability for a species' presence, based on a presence/(pseudo)absence model object, or on a vector of predicted probability values plus either the modelled binary response variable, the total number of modelled ones and zeros, or the prevalence (proportion of ones) in the modelled binary response (i.e., in the training data).

## Usage

```
Fav(model = NULL, obs = NULL, pred = NULL, n1n0 = NULL, sample.preval = NULL,
method = "RBV", true.preval = NULL, verbosity = 2)
```

## Arguments

- |       |   |
|-------|---|
| model | a binary-response, presence/(pseudo)absence probability-producing model object of class "glm", "gam", "gbm", "randomForest" or "bart" (computed with <code>keeptrees=TRUE</code> ).   |
| obs   | alternatively to 'model', a vector of the 1 and 0 values of the binary response variable (e.g. presence-absence of a species) in the model training data. This argument is ignored if 'model' is provided.  |
| pred  | alternatively to 'model', a numeric vector, RasterLayer or SpatRaster of predicted presence probability values, produced by a presence/(pseudo)absence modelling method yielding presence probability. This argument is ignored if 'model' is provided. |

<code>n1n0</code>	alternatively to 'obs' or 'sample.preval', an integer vector of length 2 providing the total numbers of modelled ones and zeros (in this order) of the binary response variable in the model training data. Ignored if 'obs' or 'model' is provided.
<code>sample.preval</code>	alternatively to 'obs' or 'n1n0', the prevalence (proportion of ones) of the binary response variable in the model training data. Ignored if 'model' is provided.
<code>method</code>	either "RBV" for the original Real, Barbosa & Vargas (2006) procedure, or "AT" if you want to try out the modification proposed by Albert & Thuiller (2008) (but see Details!).
<code>true.preval</code>	the true prevalence (as opposed to sample prevalence), necessary if you want to try the "AT" method (but see Details!).
<code>verbosity</code>	numeric value indicating the amount of messages to display; currently meaningful values are 0, 1, and 2 (the default).

## Details

Methods such as Generalized Linear Models, Generalized Additive Models, Random Forests, Boosted Regression Trees / Generalized Boosted Models, Bayesian Additive Regression Trees and several others, are widely used for modelling species' potential distributions using presence/absence data and a set of predictor variables. These models predict presence probability, which incorporates the prevalence (proportion of presences) of the species in the modelled sample. So, predictions for restricted species are always generally low, while predictions for widespread species are always generally higher, regardless of the actual environmental quality. Barbosa (2006) and Real, Barbosa & Vargas (2006) proposed an environmental favourability function which is based on presence probability and cancels out uneven proportions of presences and absences in the modelled data. Favourability thus assesses the extent to which the environmental conditions change the probability of occurrence of a species with respect to its overall prevalence in the study area. Model predictions become, therefore, directly comparable among species with different prevalences. The favourability function is implemented in the **fuzzySim** package and is also in the SAM (Spatial Analysis in Macroecology) software (Rangel et al. 2010).

Using simulated data, Albert & Thuiller (2008) proposed a modification to the favourability function, but it requires knowing the true prevalence of the species (not just the prevalence in the modelled sample), which is rarely possible in real-world modelling. Besides, this suggestion was based on the misunderstanding that the favourability function was a way to obtain the probability of occurrence when prevalence differs from 50%, which is incorrect (see Acevedo & Real 2012).

To get environmental favourability with either the Real, Barbosa & Vargas ("RBV") or the Albert & Thuiller ("AT") method, you just need to get model predictions of presence probability from your data, together with the proportions of presences and absences in the modelled sample, and then use the 'Fav' function. Input data for this function are either a model object resulting from the `glm` function, or the vector of presences-absences (1-0) of your species and the corresponding presence probability values, obtained e.g. with `predict(mymodel, mydata, type = "response")`. Alternatively to the presences-absences, you can provide either the sample prevalence or the numbers of presences and absences in the dataset that was used to generate the presence probabilities. In case you want to use the "AT" method, you also need to provide the true (absolute) prevalence of your species.

**Value**

If 'model' is provided or if 'pred' is a numeric vector, the function returns a numeric vector of the favourability values. If 'model' is not provided (which would override other arguments) and 'pred' is a RasterLayer or a SpatRaster, the function returns an object of the same class, containing the favourability values.

**Author(s)**

A. Marcia Barbosa

**References**

Acevedo P. & Real R. (2012) Favourability: concept, distinctive characteristics and potential usefulness. *Naturwissenschaften* 99: 515-522

Albert C.H. & Thuiller W. (2008) Favourability functions versus probability of presence: advantages and misuses. *Ecography* 31: 417-422.

Barbosa A.M.E. (2006) Modelacion de relaciones biogeograficas entre predadores, presas y parásitos: implicaciones para la conservacion de mamiferos en la Peninsula Iberica. PhD Thesis, University of Malaga (Spain).

Rangel T.F.L.V.B, Diniz-Filho J.A.F & Bini L.M. (2010) SAM: a comprehensive application for Spatial Analysis in Macroecology. *Ecography* 33: 46-50.

Real R., Barbosa A.M. & Vargas J.M. (2006) Obtaining environmental favourability functions from logistic regression. *Environmental and Ecological Statistics* 13: 237-245.

**See Also**

[multGLM](#)

**Examples**

```
# obtain a probability model and its predictions:

data(rotif.env)

names(rotif.env)

mod <- with(rotif.env, glm(Abrigh ~ Area + Altitude +
  AltitudeRange + HabitatDiversity + HumanPopulation,
  family = binomial))

prob <- predict(mod, data = rotif.env, type = "response")

# obtain predicted favourability in different ways:

Fav(model = mod)

Fav(obs = rotif.env$Abrigh, pred = prob)
```

```
Fav(pred = mod$fitted.values, n1n0 = c(112, 179))

Fav(pred = mod$fitted.values, sample.preval = 0.38)
```

---

favClass

*Classify favourability into 3 categories (low, intermediate, high)*


---

## Description

This function takes a vector of [Favourability](#) values and reclassifies them into 3 increasing categories: low, intermediate or high. By default, the breaks between these classes are 0.2 and 0.8 (see [Details](#)), although these can be changed by the user.

## Usage

```
favClass(fav, breaks = c(0.2, 0.8), character = FALSE)
```

## Arguments

fav	a numeric vector of favourability values (obtained, e.g., with functions <a href="#">Fav</a> or <a href="#">multGLM</a> ).
breaks	a numeric vector of length 2 containing the two values which will divide fav into the 3 classes. Defaults to c(0.2, 0.8) following the literature (see <a href="#">Details</a> ).
character	logical value indicating whether the result should be returned in character rather than numeric form. Defaults to FALSE.

## Details

Some applications of species distribution models imply setting a threshold to separate areas with high and low probability or favourability for occurrence (see, e.g., [bioThreat](#)). However, it makes little sense to establish as markedly different areas with, for example, 0.49 and 0.51 favourability values (Hosmer & Lemeshow, 1989). It may thus be wiser to open a gap between values considered as clearly favourable and clearly unfavourable. When this option is taken in the literature, commonly used breaks are 0.8 as a threshold to classify highly favourable values, as the odds are more than 4:1 favourable to the species; 0.2 as a threshold below which to consider highly unfavourable values, as odds are less than 1:4; and classifying the remaining values as intermediate favourability (e.g., Munoz & Real 2006, Olivero et al. 2016).

## Value

This function returns either an integer or a character vector (following the 'character' argument, which is set to FALSE by default), of the same length as fav, reclassifying it into 3 categories: 1 ('low'), 2 ('intermediate'), or 3 ('high').

## Author(s)

A. Marcia Barbosa



## References

- Hosmer D.W. Jr & Lemeshow S. (1989) Applied logistic regression. John Wiley & Sons, New York
- Munoz A.R. & Real R. (2006) Assessing the potential range expansion of the exotic monk parakeet in Spain. *Diversity and Distributions*, 12: 656-665
- Olivero J., Fa J.E., Real R., Farfan M.A., Marquez A.L., Vargas J.M., Gonzalez J.P., Cunningham A.A. & Nasi R. (2017) Mammalian biogeography and the Ebola virus in Africa. *Mammal Review*, 47: 24-37

## See Also

[Fav](#), [multGLM](#)

## Examples

```
data(rotif.env)
mods <- multGLM(rotif.env, sp.cols = 20, var.cols = 5:17)
fav <- mods$predictions[, 2]
data.frame(fav = fav, favcl_num = favClass(fav),
  favcl_chr = favClass(fav, character = TRUE))
```

---

FDR

*False Discovery Rate*

---

## Description

Calculate the false discovery rate (type I error) under repeated testing and determine which variables to select and to exclude from multivariate analysis.

## Usage

```
FDR(data = NULL, sp.cols = NULL, var.cols = NULL, pvalues = NULL,
  model.type = NULL, family = "auto", correction = "fdr", q = 0.05,
  verbose = TRUE, simplif = FALSE)
```

## Arguments

<code>data</code>	a data frame containing the response and predictor variables (one in each column).
<code>sp.cols</code>	name or index number of the column containing the response variable (currently implemented for only one response variable at a time).
<code>var.cols</code>	names or index numbers of the columns containing the predictor variables.
<code>pvalues</code>	optionally, instead of 'data', 'sp.cols' and 'var.cols', a data frame with the names of the predictor variables in the first column and their bivariate p-values (obtained elsewhere) in the second column. Example: <code>pvalues &lt;- data.frame(var = letters[1:5], pval = c(0.02, 0.004, 0.07, 0.03, 0.05))</code> .

<code>model.type</code>	this argument (previously a character value, either "LM" or "GLM") is now deprecated and ignored with a warning if provided. This information is now included in argument 'family' – e.g., if you want linear models (LM), you can set 'family = "gaussian"".
<code>family</code>	The error distribution and (optionally) the link function to use (see <a href="#">glm</a> or <a href="#">family</a> for details). The default "auto" automatically uses "binomial" family for response variables containing only values of 0 and 1; "poisson" for positive integer responses (i.e. count data); "Gamma" for positive non-integer; and "gaussian" (i.e., linear models) otherwise.
<code>correction</code>	the correction procedure to apply to the p-values; see <a href="#">p.adjust.methods</a> for available options and <a href="#">p.adjust</a> for more information. The default is "fdr".
<code>q</code>	the threshold value of FDR-corrected significance above which to reject variables. Defaults to 0.05.
<code>verbose</code>	logical value indicating whether to display messages.
<code>simplif</code>	logical value indicating if simplified results should be provided (see Value).

## Details

It is common in ecology to search for statistical relationships between species' occurrence and a set of predictor variables. However, when a large number of variables is analysed (compared to the number of observations), false findings may arise due to repeated testing. Garcia (2003) recommended controlling the false discovery rate (FDR; Benjamini & Hochberg 1995) in ecological studies. The [p.adjust](#) R function performs this and other corrections to the significance (p) values of variables under repeated testing. The 'FDR' function performs repeated regressions (either linear or logistic) or uses already-obtained p values for a set of variables; calculates the FDR with 'p.adjust'; and shows which variables should be retained for or excluded from further multivariate analysis according to their corrected p values (see, for example, Barbosa, Real & Vargas 2009).

The FDR function uses the Benjamini & Hochberg ("BH", alias "fdr") correction by default, but check the [p.adjust](#) documentation for other available methods, namely "BY", which allows for non-independent data. Input data may be the response variable (for example, the presence-absence or abundance of a species) and the predictors (a table with one independent variable in each column, with the same number of rows and in the same order as the response); there should be no missing values in the data. Alternatively, you may already have performed the univariate regressions and have a set of variables and corresponding p values which you want to correct with FDR; in this case, get a table with your variables' names in the first column and their p values in the second column, and supply it as the 'pvalues' argument (no need to provide response or predictors in this case).

## Value

If `simplif = TRUE`, this function returns a data frame with the variables' names as row names and 4 columns containing, respectively, their individual (bivariate) coefficients against the response, their individual AIC (Akaike's Information Criterion; Akaike, 1973), BIC (Bayesian Information Criterion, also known as Schwarz criterion, SBC, SBIC; Schwarz, 1978), p-value and adjusted p-value according to the applied 'correction'. If `simplif = FALSE` (the default), the result is a list of two such data frames:

<code>exclude</code>	with the variables to exclude.
<code>select</code>	with the variables to select (under the given 'q' value).

**Author(s)**

A. Marcia Barbosa

**References**

Akaike, H. (1973) Information theory and an extension of the maximum likelihood principle. In: Petrov B.N. & Csaki F., 2nd International Symposium on Information Theory, Tsahkadsor, Armenia, USSR, September 2-8, 1971, Budapest: Akademiai Kiado, p. 267-281.

Barbosa A.M., Real R. & Vargas J.M (2009) Transferability of environmental favourability models in geographic space: The case of the Iberian desman (*Galemys pyrenaicus*) in Portugal and Spain. *Ecological Modelling* 220: 747-754

Benjamini Y. & Hochberg Y. (1995) Controlling the false discovery rate: a practical and powerful approach to multiple testing. *Journal of the Royal Statistical Society, Series B* 57: 289-300

Garcia L.V. (2003) Controlling the false discovery rate in ecological research. *Trends in Ecology and Evolution* 18: 553-554

Schwarz, G.E. (1978) Estimating the dimension of a model. *Annals of Statistics*, 6 (2): 461-464.

**See Also**

[p.adjust](#)

**Examples**

```
data(rotif.env)

names(rotif.env)

FDR(data = rotif.env, sp.cols = 18, var.cols = 5:17)

FDR(data = rotif.env, sp.cols = 18, var.cols = 5:17, simplif = TRUE)

my_pvalues <- data.frame(var = letters[1:5], pval = c(0.02, 0.004, 0.07, 0.03, 0.05))
FDR(pvalues = my_pvalues)
```

---

fuzSim

*Fuzzy similarity*


---

**Description**

This function calculates fuzzy similarity, based on a fuzzy version of the binary similarity index specified in method, between two binary (0 or 1) or fuzzy (between 0 and 1) variables.

**Usage**

```
fuzSim(x, y, method, na.rm = TRUE)
```

## Arguments

<code>x</code>	a vector of (optionally fuzzy) presence-absence data, with 1 meaning presence, 0 meaning absence, and values in between meaning fuzzy presence (or the degree to which each locality belongs to the set of species presences, or to which each species belongs to the locality; Zadeh, 1965). Fuzzy presence-absence can be obtained, for example, with functions <code>multGLM</code> , <code>multTSA</code> or <code>distPres</code> in this package.
<code>y</code>	a vector similar to 'x', of the same length and in the same order.
<code>method</code>	the similarity index to use. Currently available options are "Jaccard", "Sorensen", "Simpson" and "Baroni" (see Details).
<code>na.rm</code>	logical value indicating whether NA values should be ignored. The default is TRUE.

## Details

Similarity between ecological communities, beta diversity patterns, biotic regions, and distributional relationships among species are commonly determined based on pair-wise (dis)similarities in species' occurrence patterns. Some of the most commonly employed similarity indices are those of Jaccard (1901), Sorensen (1948), Simpson (1960) and Baroni-Urbani & Buser (1976), which are here implemented in their fuzzy versions (Barbosa, 2015), able to deal with both binary and fuzzy data. Jaccard's and Baroni's indices have associated tables of significant values (Baroni-Urbani & Buser 1976, Real & Vargas 1996, Real 1999).

## Value

The function returns a value between 0 and 1 representing the fuzzy similarity between the provided 'x' and 'y' vectors. Note, for example, that Jaccard similarity can be converted to dissimilarity (or Jaccard distance) if subtracted from 1, while 1-Sorensen is not a proper distance metric as it lacks the property of triangle inequality (see [https://en.wikipedia.org/wiki/S%C3%B8rensen%E2%80%9393Dice\\_coefficient](https://en.wikipedia.org/wiki/S%C3%B8rensen%E2%80%9393Dice_coefficient)).

## Note

The formulas used in this function may look slightly different from some of their published versions (e.g. Baroni-Urbani & Buser 1976), not only because the letters are switched, but because here the A and B are the numbers of attributes present in each element, whether or not they are also present in the other one. Thus, our 'A+B' is equivalent to 'A+B+C' in formulas where A and B are the numbers of attributes present in one but not the other element, and our A+B-C is equivalent to their A+B+C. The formulas used here (adapted from Olivero et al. 1998) are faster to calculate, visibly for large datasets.

## Author(s)

A. Marcia Barbosa

## References

- Barbosa A.M. (2015) fuzzySim: applying fuzzy logic to binary similarity indices in ecology. *Methods in Ecology and Evolution*, 6: 853-858.
- Baroni-Urbani C. & Buser M.W. (1976) Similarity of Binary Data. *Systematic Zoology*, 25: 251-259
- Jaccard P. (1901) Etude comparative de la distribution florale dans une portion des Alpes et des Jura. *Memoires de la Societe Vaudoise des Sciences Naturelles*, 37: 547-579
- Olivero J., Real R. & Vargas J.M. (1998) Distribution of breeding, wintering and resident waterbirds in Europe: biotic regions and the macroclimate. *Ornis Fennica*, 75: 153-175
- Real R. (1999) Tables of significant values of Jaccard's index of similarity. *Miscellanea Zoologica* 22: 29:40
- Real R. & Vargas J.M (1996) The probabilistic basis of Jaccard's index of similarity. *Systematic Biology* 45: 380-385
- Simpson, G.G. (1960) Notes on the measurement of faunal resemblance. *Amer. J. Sci.* 258A, 300-311
- Sorensen T. (1948) A method of establishing groups of equal amplitude in plant sociology based on similarity of species and its application to analyses of the vegetation on Danish commons. *Kongelige Danske Videnskabernes Selskab*, 5(4): 1-34
- Zadeh L.A. (1965) Fuzzy sets. *Information and Control*, 8: 338-353

## See Also

[simMat](#); [modOverlap](#)

## Examples

```
data(rotif.env)

names(rotif.env)

# you can calculate similarity between binary species occurrence patterns:

fuzSim(rotif.env[, "Abrigh"], rotif.env[, "Afissa"], method = "Jaccard")
fuzSim(rotif.env[, "Abrigh"], rotif.env[, "Afissa"], method = "Sorensen")
fuzSim(rotif.env[, "Abrigh"], rotif.env[, "Afissa"], method = "Simpson")
fuzSim(rotif.env[, "Abrigh"], rotif.env[, "Afissa"], method = "Baroni")

# or you can model environmental favourability for these species
# and calculate fuzzy similarity between their environmental predictions
# which goes beyond the strict coincidence of their occurrence records:

fav <- multGLM(rotif.env, sp.cols = 18:19, var.cols = 5:17, step = TRUE,
FDR = TRUE, trim = TRUE, P = FALSE, Fav = TRUE) $ predictions

fuzSim(fav[, "Abrigh_F"], fav[, "Afissa_F"], method = "Jaccard")
fuzSim(fav[, "Abrigh_F"], fav[, "Afissa_F"], method = "Sorensen")
```

```
fuzSim(fav[, "Abrigh_F"], fav[, "Afissa_F"], method = "Simpson")
fuzSim(fav[, "Abrigh_F"], fav[, "Afissa_F"], method = "Baroni")
```

fuzzyConsensus

*Fuzzy consensus among model predictions*

## Description

This function takes a data frame or a (multilayer) `SpatRaster` map of favourability predictions (i.e., directly comparable predictions obtained from presence probability; see [Fav](#)) and it computes the consensus favourability, i.e., a row-wise weighted mean in which larger weights are assigned to models with higher loadings in the first axis of a principal components analysis (Baquero et al. 2021).

## Usage

```
fuzzyConsensus(data, weights = "PCA1", simplif = TRUE, plot = TRUE,
  biplot = TRUE, verbosity = 2, do.par = TRUE)
```

## Arguments

<code>data</code>	matrix, data frame or (multilayer) 'SpatRaster' map containing the favourability values to combine.
<code>weights</code>	method for computing the weights for the weighted average of favourability values. Currently only "PCA1" is implemented.
<code>simplif</code>	logical value. If TRUE (the default), the output includes only the numeric vector of weighted mean favourability. If set to FALSE, the output will include also the complete PCA result (if weights="PCA1").
<code>plot</code>	logical value indicating whether to produce a barplot of the PCA axis loadings. The default is TRUE.
<code>biplot</code>	logical value indicating whether to produce a <a href="#">biplot</a> of the PCA. The default is TRUE.
<code>verbosity</code>	integer value indicating the amount of messages to display in the console. The default is to emit all messages available.
<code>do.par</code>	logical value indicating whether to override the current plotting parameters (restoring them on exit). The default is TRUE.

## Details

Species distribution models are often computed using different modelling methods and/or climate scenarios. One way to summarize or combine them is to do a principal components analysis (PCA) of the different model predictions: The first axis of this PCA captures consistent spatial patterns in the predicted values across the different models (Araujo, Pearson, et al. 2005; Araujo, Whittaker, et al. 2005; Marmion et al. 2009; Thuiller 2004). However, the units of the PCA axes are difficult to interpret. Baquero et al. (2021) solved this by computing a weighted average of the favourability values (which are commensurable and therefore directly comparable across species and study areas;

Real et al. 2006, Acevedo & Real 2012), using the loadings of the first PCA axis as weights. The result is therefore in the same scale as favourability, and it incorporates the degree of consensus among models, which dictates how much weight each model has in the prediction, thus avoiding disparate predictions to be blindly mixed and averaged out (Baquero et al. 2021).

### Value

If `simplif=TRUE` (the default), the function returns a numeric vector with length equal to the number of rows in 'data' (if 'data' is a matrix or data frame), or a 'SpatRaster' layer (if 'data' is a 'SpatRaster' object), with the consensus among the input favourabilities. If `simplif=FALSE`, the function returns a list containing, additionally, the output of `prcomp`.

### Author(s)

A. Marcia Barbosa

### References

- Acevedo P. & Real R. (2012) Favourability: Concept, distinctive characteristics and potential usefulness. *Naturwissenschaften*, 99: 515-522
- Araujo M.B., Pearson R.G., Thuiller W. & Erhard M. (2005) Validation of species-climate impact models under climate change. *Global Change Biology*, 11: 1504-1513
- Araujo M.B., Whittaker R.J., Ladle R.J. & Erhard M. (2005) Reducing uncertainty in projections of extinction risk from climate change. *Global Ecology and Biogeography*, 14: 529-538
- Baquero R.A., Barbosa A.M., Ayllon D., Guerra C., Sanchez E., Araujo M.B. & Nicola G.G. (2021) Potential distributions of invasive vertebrates in the Iberian Peninsula under projected changes in climate extreme events. *Diversity and Distributions*, 27(11): 2262-2276
- Marmion M., Parviainen M., Luoto M., Heikkinen R.K. & Thuiller W. (2009) Evaluation of consensus methods in predictive species distribution modelling. *Diversity and Distributions* 15: 59-69
- Real R., Barbosa A.M. & Vargas J.M. (2006) Obtaining environmental favourability functions from logistic regression. *Environmental and Ecological Statistics* 13: 237-245
- Thuiller W. (2004) Patterns and uncertainties of species' range shifts under climate change. *Global Change Biology*, 10: 2020-2027

### See Also

[weighted.mean](#)

### Examples

```
## Not run:
# this example requires having the 'gam' package installed

data(rotif.env)

library(gam)
```

```
# get two different model predictions for one of the species in this dataset:

names(rotif.env)
vars <- names(rotif.env)[5:17]

form_glm <- as.formula(paste("Ttetra ~", paste(vars, collapse = "+")))
mod_glm <- glm(form_glm, family = binomial, data = rotif.env)
pred_glm <- predict(mod_glm, rotif.env, type = "response")

form_gam <- as.formula(paste("Ttetra ~", paste("s(", vars, ")", collapse = "+")))
mod_gam <- gam(form_gam, family = binomial, data = rotif.env)
pred_gam <- predict(mod_gam, rotif.env, type = "response")

# convert probability predictions to favourability:

fav_glm <- Fav(pred = pred_glm, sample.preval = prevalence(model = mod_glm))
fav_gam <- Fav(pred = pred_gam, sample.preval = prevalence(model = mod_gam))

# compute the consensus favourability of these two models:

fav_consensus <- fuzzyConsensus(cbind(fav_glm, fav_gam))

cor(cbind(fav_glm, fav_gam, fav_consensus))

## End(Not run)
```

---

fuzzyOverlay

*Row-wise overlay operations based on fuzzy logic*


---

## Description

Logical and set operations are useful for comparative distribution modelling, to assess consensus or mismatches between the predictions of different models, and to quantify differences between models obtained for different time periods. Fuzzy set theory (Zadeh 1965, Barbosa & Real 2012) allows performing such operations without converting model predictions from continuous to binary, thus avoiding the application of arbitrary thresholds and the distortion or over-simplification of those predictions. The result is a continuous numerical value quantifying the intersection, union, sum, or other operation among model predictions, whether binary or continuous.

## Usage

```
fuzzyOverlay(data, overlay.cols = 1:ncol(data), op = "intersection",
na.rm = FALSE, round.digits = 2)
```

## Arguments

**data**                      matrix or data frame containing the model predictions to compare.



overlay.cols	vector of the names or index numbers of the columns to compare. The default is all columns in data.
op	character value indicating the operation to perform between the prediction columns in 'data'. Can be "consensus" for the arithmetic mean of predictions (or the fuzzy equivalent of the proportion of models that agree that the species occurs at each site), "fuzzy_and" or "intersection" for fuzzy intersection; "fuzzy_or" or "union" for fuzzy union; "prob_and" or "prob_or" for probabilistic and/or, respectively (see Details); "maintenance" for the values where all predictions for the same row (rounded to the number of digits specified in the next argument) are the same. If 'data' has only two columns to compare, you can also calculate "xor" for exclusive 'or', "AnotB" for the occurrence of the species in column 1 in detriment of that in column 2, "expansion" for the prediction increase in rows where column 2 has higher values than column 1, "contraction" for the prediction decrease in rows where column 2 has lower values than column 1, or "change" for a mix of the latter two, with positive values where there has been an increase and negative values where there was decrease in favourability from columns 1 to 2. For expansion, contraction and maintenance, rows where the values do not satisfy the condition (i.e. second column larger, smaller, or roughly equal to the first column) get a value of zero.
na.rm	logical value indicating if NA values should be ignored. The default is FALSE, so rows with NA in any of the prediction columns get NA as a result.
round.digits	integer value indicating the number of decimal places to be used if op = "maintenance". The default is 2.

### Details

If your predictions are probabilities, "prob\_and" (probabilistic 'and') gives the probability of all species in 'data' occurring simultaneously by multiplying all probabilities; and "prob\_or" (probabilistic 'or') gives the probability of any of them occurring at each site. These can be quite restrictive, though; probabilistic "and" can give particularly unrealistically small values.

If you have (or convert your probabilities to) favourability predictions, which can be used directly with fuzzy logic (Real et al. 2006; see [Fav](#) function), you can use "fuzzy\_and" or "intersection" to get the favourability for all species co-occurring at each site, and "fuzzy\_or" or "union" to get favourability for any of them to occur at each site (Barbosa & Real 2012).

### Value

This function returns a vector, with length equal to the number of rows in data, containing the row-wise result of the operation performed.

### Author(s)

A. Marcia Barbosa

### References

Barbosa A.M. & Real R. (2012) Applying fuzzy logic to comparative distribution modelling: a case study with two sympatric amphibians. The Scientific World Journal, 2012, Article ID 428206

Real R., Barbosa A.M. & Vargas J.M. (2006) Obtaining environmental favourability functions from logistic regression. *Environmental and Ecological Statistics* 13: 237-245.

Zadeh, L.A. (1965) Fuzzy sets. *Information and Control*, 8: 338-353

### See Also

[fuzSim](#), [modOverlap](#) and [fuzzyRangeChange](#) for overall (not row-wise) comparisons among model predictions.

### Examples

```
data(rotif.env)

names(rotif.env)

# get model predictions for 3 of the species in rotif.env:

mods <- multGLM(rotif.env, sp.cols = 18:20, var.cols = 5:17, id.col = 1,
step = TRUE, FDR = TRUE, trim = TRUE)

preds <- mods$predictions[ , c("Abrigh_F", "Afissa_F", "Apriod_F")]

# calculate intersection and union among those predictions:

preds$intersect <- fuzzyOverlay(preds, op = "intersection")

preds$union <- fuzzyOverlay(preds, op = "union")

head(preds)

# imagine you have a model prediction for species 'Abrigh' in a future time
# (here we will create one by randomly jittering the current predictions)

preds$Abrigh_imag <- jitter(preds[ , "Abrigh_F"], amount = 0.2)
preds$Abrigh_imag[preds$Abrigh_imag < 0] <- 0
preds$Abrigh_imag[preds$Abrigh_imag > 1] <- 1

# you can calculate row-wise prediction changes from Abrigh to Abrigh_imag:

preds$Abrigh_exp <- fuzzyOverlay(preds, overlay.cols = c("Abrigh_F",
"Abrigh_imag"), op = "expansion")

preds$Abrigh_contr <- fuzzyOverlay(preds, overlay.cols = c("Abrigh_F",
"Abrigh_imag"), op = "contraction")

preds$Abrigh_chg <- fuzzyOverlay(preds, overlay.cols = c("Abrigh_F",
"Abrigh_imag"), op = "change")
```

```

preds$Abrigh_maint <- fuzzyOverlay(preds, overlay.cols = c("Abrigh_F",
  "Abrigh_imag"), op = "maintenance")

head(preds)

```

fuzzyRangeChange

*Range change based on continuous (fuzzy) values*

## Description

This function quantifies overall range change (expansion, contraction, maintenance and balance) based on either presence-absence data or the continuous predictions of two models.

## Usage

```

fuzzyRangeChange(pred1, pred2, number = TRUE, prop = TRUE,
  na.rm = TRUE, round.digits = 2, measures = c("Gain", "Loss",
  "Stable presence", "Stable absence", "Balance"), plot = TRUE, x.lab = TRUE, ...)

```

## Arguments

pred1	numeric vector containing the predictions (between 0 and 1) of the model that will serve as reference.
pred2	numeric vector containing the predictions (between 0 and 1) of the model whose change will be calculated. Must be of the same length and in the same order as 'pred1'.
number	logical value indicating if results should include the fuzzy number of cases. The default is TRUE.
prop	logical value indicating if results should include the proportion of the total number of cases. The default is TRUE.
na.rm	logical value indicating whether NA values should be ignored. The default is TRUE.
round.digits	argument to pass to <a href="#">fuzzyOverlay</a> , indicating the number of decimal places to which to round 'pred' for calculating 'maintenance' or 'stability'. The default is 2.
measures	character vector listing the range change measures to calculate. The default is all available measures.
plot	logical value indicating whether to make a barplot with the results. The default is TRUE.
x.lab	logical value indicating whether to add the x axis labels to the plot (i.e., the names below each bar). The default is TRUE, but users may set it to FALSE and then add labels differently.
...	additional arguments to be passed to the <a href="#">barplot</a> function (if plot = TRUE).

**Value**

This function returns a data frame with the following values in different rows (among those that are included in 'measures'):

Gain	sum of the predicted values that have increased from 'pred1' to 'pred2' (fuzzy equivalent of the number of gained presences)
Loss	sum of the predicted values that have decreased from 'pred1' to 'pred2' (fuzzy equivalent of the number of lost presences)
Stable_presence	fuzzy equivalent of the number of predicted presences that have remained as such (when rounded to 'round.digits') between 'pred1' and 'pred2'
Stable_absence	fuzzy equivalent of the number of predicted absences that have remained as such (when rounded to 'round.digits') between 'pred1' and 'pred2'
Balance	sum of the change in predicted values from 'pred1' to 'pred2' (fuzzy equivalent of the balance of gained and lost presences)

If prop = TRUE (the default), there is an additional column named "Proportion" in which these values are divided by the total number of reference values (i.e., the fuzzy range or non-range size). If plot = TRUE (the default), a barplot is also produced representing the last column of the result data frame.

**Author(s)**

A. Marcia Barbosa

**See Also**

[fuzSim](#), [modOverlap](#) for other ways to compare models; [fuzzyOverlay](#) for row-wise model comparisons

**Examples**

```
# get an environmental favourability model for a rotifer species:

data(rotif.env)

names(rotif.env)

fav_current <- multGLM(rotif.env, sp.cols = 18, var.cols = 5:17,
  step = TRUE, FDR = TRUE, trim = TRUE, P = FALSE, Fav = TRUE) $
  predictions

# imagine you have a model prediction for this species in a future time
# (here we will create one by randomly jittering the current predictions)

fav_imag <- jitter(fav_current, amount = 0.2)
fav_imag[fav_imag < 0] <- 0
fav_imag[fav_imag > 1] <- 1
```

```
# calculate range change given by current and imaginary future predictions:

fuzzyRangeChange(fav_current, fav_imag)

fuzzyRangeChange(fav_current, fav_imag, number = FALSE)

fuzzyRangeChange(fav_current, fav_imag, ylim = c(-1, 1),
ylab = "Proportional change")
```

getPreds

*Get model predictions*

## Description

This function allows getting the predictions of multiple models when applied to a given dataset. It can be useful if you have a list of model objects (e.g. resulting from [multGLM](#)) and want to apply them to a new data set containing the same variables for another region or time period. There are options to include the logit link ('Y') and/or 'Favourability' (see [Fav](#)).

## Usage

```
getPreds(data, models, id.col = NULL, Y = FALSE, P = TRUE,
Favourability = TRUE, incl.input = FALSE, verbosity = 2)
```

## Arguments

data	an object of class either 'data.frame' or 'RasterStack' to which to apply the 'models' (below) to get their predictions; must contain all variables (with the same names, case-sensitive) included in any of the 'models'.
models	an object of class 'list' containing one or more model objects, obtained e.g. with function <a href="#">glm</a> or <a href="#">multGLM</a> .
id.col	optionally, the index number of a column of 'data' containing row identifiers, to be included in the result. Ignored if incl.input = TRUE, or if 'data' is a RasterStack rather than a data frame.
Y	logical, whether to include the logit link (y) value in the predictions.
P	logical, whether to include the probability value in the predictions.
Favourability	logical, whether to include Favourability in the predictions (see <a href="#">Fav</a> ).
incl.input	logical, whether to include input columns in the output data frame (if the 'data' input is a data frame too). The default is FALSE.
verbosity	numeric value indicating the amount of messages to display; currently meaningful values are 0, 1, and 2 (the default).

## Value

This function returns the model predictions in an object of the same class as the input 'data', i.e. either a data frame or a RasterStack.

**Author(s)**

A. Marcia Barbosa

**See Also**

[multGLM](#), [predict](#)

**Examples**

```
data(rotif.env)

names(rotif.env)

# identify rotifer data in the Eastern and Western hemispheres:

unique(rotif.env$CONTINENT)

rotif.env$HEMISPHERE <- "Eastern"

rotif.env$HEMISPHERE[rotif.env$CONTINENT %in%
c("NORTHERN_AMERICA", "SOUTHERN_AMERICA")] <- "Western"

head(rotif.env)

# separate the rotifer data into hemispheres

east.hem <- rotif.env[rotif.env$HEMISPHERE == "Eastern", ]
west.hem <- rotif.env[rotif.env$HEMISPHERE == "Western", ]

# make models for 3 of the species in rotif.env based on their distribution
# in the Eastern hemisphere:

mods <- multGLM(east.hem, sp.cols = 18:20, var.cols = 5:17,
id.col = 1, step = FALSE, FDR = FALSE, trim = FALSE)

# get the models' predictions for the Western hemisphere dataset:

preds <- getPreds(west.hem, models = mods$models, P = TRUE,
Favourability = TRUE)

head(preds)
```

---

gridRecords	<i>Grid (or thin) point occurrence records to the resolution of a raster map</i>
-------------	--

---

## Description

This function takes a Raster\* or SpatRaster object and a set of spatial coordinates of a species' presence (and optionally absence) records, and returns a data frame with the presences and absences as well as the corresponding values of the rasters in the grid of pixels (cells). This is analogous to removing duplicates and thinning points (both presences and absences) with a distance equal to the pixel size of the raster map(s) on which analysis will be based.

## Usage

```
gridRecords(rst, pres.coords, abs.coords = NULL, absences = TRUE,
  species = NULL, na.rm = TRUE)
```

## Arguments

rst	a Raster* or SpatRaster object (the latter is processed faster) with the desired spatial resolution and extent for the species presence-(pseudo)absence data, and the layer(s) whose values to extract for those data.
pres.coords	a matrix or data frame with two columns containing, respectively, the x and y, or longitude and latitude coordinates (in this order, and in the same coordinate reference system as 'rst') of the points where species presence was detected.
abs.coords	(optional) same as 'pres.coords' but for points where the species was not detected. If NULL and if absences=TRUE (the default), all pixels that are not intersected by 'pres.coords' will be returned as absence cells.
absences	logical value indicating whether pixels without presence records should be returned as absences. The default is TRUE.
species	(optional) character vector, of the same length as 'nrow(pres.coords)', indicating the species to which each pair of coordinates corresponds. Useful for gridding records of more than one species at a time. If specified, 'abs.coords' cannot be used.
na.rm	Logical value indicating whether pixels without values in any of the 'rst' layers should be removed from the resulting data frame. The default is TRUE.

## Details

See e.g. Baez et al. (2020), where this function was used to get unique presences and absences from point occurrence data at the spatial resolution of marine raster variables.

**Value**

This function returns a data frame with the following columns:

'presence'	integer, 1 for the cells (pixels) with at least one presence point, and 0 for the cells without any presence point (if <code>absences = TRUE</code> ) or with at least one absence point (if <code>'abs.coords'</code> are provided) AND with no presence points. If the <code>'species'</code> argument is provided, instead of <code>'presence'</code> you get one column named as each species.
'x', 'y'	centroid coordinates of each cell (pixel).
'cell'	the pixel identifier in <code>'rst'</code> .
one column for each layer in <code>'rst'</code>	value of each pixel for each layer.

**Note**

This function requires either the **raster** or the **terra** package, depending on the class of `'rst'`.

**Author(s)**

A. Marcia Barbosa

**References**

Baez J.C., Barbosa A.M., Pascual P., Ramos M.L. & Abascal F. (2020) Ensemble modelling of the potential distribution of the whale shark in the Atlantic Ocean. *Ecology and Evolution*, 10: 175-184

**See Also**

`'extract'` in package **raster** or in package **terra**

**Examples**

```
## Not run:

# you can run these examples if you have the 'terra' package installed
require(terra)

# import a raster map and aggregate it to a coarser resolution:
r <- rast(system.file("ex/elev.tif", package = "terra"))
r <- aggregate(r, 6)
plot(r)

# generate some random presence and absence points:
set.seed(123)
presences <- terra::spatSample(as.polygons(r), 100)
set.seed(456)
absences <- terra::spatSample(as.polygons(r), 70)
points(presences, pch = 20, cex = 0.3, col = "black")
points(absences, pch = 20, cex = 0.3, col = "white")
```



```

# use 'gridRecords' on these random points:
gridded_pts <- gridRecords(rst = r, pres.coords = terra::crds(presences),
  abs.coords = terra::crds(absences))
head(gridded_pts)

# map the gridded points (presences black, absences white):
points(gridded_pts[ , c("x", "y")], col = gridded_pts$presence)

# you can also do it with only presence (no absence) records
# in this case, by default (with 'absences = TRUE'),
# all pixels without presence points are returned as absences:

gridded_pres <- gridRecords(rst = r, pres.coords = terra::crds(presences))
head(gridded_pres)
plot(r)
points(presences, pch = 20, cex = 0.2, col = "black")
points(gridded_pres[ , c("x", "y")], col = gridded_pres$presence)

# with only presence (no absence) records, as in this latter case,
# you can grid records for multiple species at a time
# by adding a 'species' argument

presences$species <- rep(c("species1", "species2", "species3"), each = 33)
data.frame(presences)
plot(r)
plot(presences, col = as.factor(presences$species), add = TRUE)
gridded_pres_mult <- gridRecords(rst = r, pres.coords = terra::crds(presences),
  species = presences$species)
head(gridded_pres_mult)

## End(Not run)

```

---

integerCols

---

*Classify integer columns*


---

## Description

This function detects which numeric columns in a data frame contain only whole numbers, and converts those columns to integer class, so that they take up less space.

## Usage

```
integerCols(data)
```

## Arguments

**data** a data frame containing possibly integer columns classified as "numeric".

**Value**

The function returns a data frame with the same columns as 'data', but with those that are numeric and contain only whole numbers (possibly including NA) now classified as "integer".

**Author(s)**

A. Marcia Barbosa

**See Also**

[is.integer](#), [as.integer](#), [multConvert](#)

**Examples**

```
dat <- data.frame(
  var1 = 1:10,
  var2 = as.numeric(1:10),
  var3 = as.numeric(c(1:4, NA, 6:10)),
  var4 = as.numeric(c(1:3, NaN, 5, Inf, 7, -Inf, 9:10)),
  var5 = as.character(1:10),
  var6 = seq(0.1, 1, by = 0.1),
  var7 = letters[1:10]
) # creates a sample data frame

dat

str(dat)
# var2 classified as "numeric" but contains only whole numbers
# var3 same as var2 but containing also NA values
# var4 same as var2 but containing also NaN and infinite values
# var5 contains only whole numbers but initially classified as factor

dat <- integerCols(dat)

str(dat)
# var2 and var3 now classified as "integer"
# var4 remains as numeric because contains infinite and NaN
# (not integer) values
# var5 remains as factor
```

---

modelTrim

*Trim off non-significant variables from a model*

---

**Description**

This function performs a stepwise removal of non-significant variables from a model, following Crawley (2005, 2007). See [stepwise](#) for a newer, more complete function that can be used instead.

**Usage**

```
modelTrim(model, method = "summary", alpha = 0.05)
```

**Arguments**

model	a model object.
method	the method for getting the individual p-values. Can be either "summary" for the p-values of the coefficient estimates, or "anova" for the p-values of the variables themselves (see Details).
alpha	the p-value above which a variable is removed.

**Details**

Stepwise variable selection is a common procedure for simplifying models. It maximizes predictive efficiency in an objective and reproducible way, and is useful when the individual importance of the predictors is not known a priori (Hosmer & Lemeshow, 2000). The [step](#) R function performs such procedure using an information criterion (AIC) to select the variables, but it often leaves variables that are not significant in the model. Such variables can be subsequently removed with a manual stepwise procedure (e.g. Crawley 2005, p. 208; Crawley 2007, p. 442 and 601; Barbosa & Real 2010, 2012; Estrada & Arroyo 2012). The 'modelTrim' function performs such removal automatically until all remaining variables are significant. It can also be applied to a full model (i.e., without previous use of the 'step' function), as it serves as a backward stepwise selection procedure based on the significance of the coefficients (if method = "summary", the default) or on the significance of the variables (if method = "anova", better when there are categorical variables in the model). See [stepwise](#) for a more complete stepwise selection procedure.

**Value**

The input model object after removal of non-significant variables.

**Author(s)**

A. Marcia Barbosa

**References**

- Barbosa A.M. & Real R. (2010) Favourable areas for expansion and reintroduction of Iberian lynx accounting for distribution trends and genetic diversity of the European rabbit. *Wildlife Biology in Practice* 6: 34-47
- Barbosa A.M. & Real R. (2012) Applying fuzzy logic to comparative distribution modelling: a case study with two sympatric amphibians. *The Scientific World Journal*, Article ID 428206
- Crawley, M.J. (2005) *Statistics: An introduction using R*. John Wiley & Sons, Ltd.
- Crawley, M.J. (2007) *The R Book*. John Wiley & Sons, Ltd.
- Estrada A. & Arroyo B. (2012) Occurrence vs abundance models: Differences between species with varying aggregation patterns. *Biological Conservation*, 152: 37-45
- Hosmer D. W. & Lemeshow S. (2000) *Applied Logistic Regression* (2nd ed). John Wiley and Sons, New York

**See Also**[step](#)**Examples**

```
# load sample data:

data(rotif.env)

names(rotif.env)

# build a stepwise model of a species' occurrence based on
# some of the variables:

mod <- with(rotif.env, step(glm(Abrigh ~ Area + Altitude + AltitudeRange +
HabitatDiversity + HumanPopulation, family = binomial)))

# examine the model:

summary(mod) # contains non-significant variables

# use modelTrim to get rid of non-significan effects:

mod <- modelTrim(mod)

summary(mod) # only significant variables now
```

---

modOverlap

---

*Overall overlap between model predictions*


---

**Description**

This function calculates the degree of overlap between the predictions of two models, using niche comparison metrics such as Schoener's D, Hellinger distance and Warren's I.

**Usage**

```
modOverlap(pred1, pred2, na.rm = TRUE)
```

**Arguments**

pred1	numeric vector of the predictions of a (values between 0 and 1).
pred2	numeric vector of the predictions of another model; must be of the same length and in the same order as 'pred1'.
na.rm	logical value indicating whether NA values should be removed prior to calculation. The default is TRUE.

## Details

See Warren et al. (2008).

## Value

This function returns a list of 3 metrics:

SchoenerD	Schoener's (1968) D statistic for niche overlap, varying between 0 (no overlap) and 1 (identical niches).
WarrenI	the I index of Warren et al. (2008), based on Hellinger distance (below) but re-formulated to also vary between 0 (no overlap) and 1 (identical niches).
HellingerDist	Hellinger distance (as in van der Vaart 1998, p. 211) between probability distributions, varying between 0 and 2.

## Author(s)

A. Marcia Barbosa

## References

- Schoener T.W. (1968) Anolis lizards of Bimini: resource partitioning in a complex fauna. *Ecology* 49: 704-726
- van der Vaart A.W. (1998) Asymptotic statistics. Cambridge Univ. Press, Cambridge (UK)
- Warren D.L., Glor R.E. & Turelli M. (2008) Environmental niche equivalency versus conservatism: quantitative approaches to niche evolution. *Evolution*, 62: 2868-83 (and further ERRATUM)

## See Also

[fuzSim](#); [fuzzyOverlay](#); `niche.overlap` in package **phyloclim**; `ecospat.niche.overlap` in package **ecospat**

## Examples

```
# get an environmental favourability model for a rotifer species:

data(rotif.env)

names(rotif.env)

fav_current <- multGLM(rotif.env, sp.cols = 18, var.cols = 5:17,
step = TRUE, FDR = TRUE, trim = TRUE, P = FALSE, Fav = TRUE) $
predictions

# imagine you have a model prediction for this species in a future time
# (here we will create one by randomly jittering the current predictions)

fav_imag <- jitter(fav_current, amount = 0.2)
fav_imag[fav_imag < 0] <- 0
```

```

fav_imag[fav_imag > 1] <- 1

# calculate niche overlap between current and imaginary future predictions:

modOverlap(fav_current, fav_imag)

```

---

multConvert

*Multiple conversion*


---

### Description

This function can simultaneously convert multiple columns of a matrix or data frame.

### Usage

```
multConvert(data, conversion, cols = 1:ncol(data))
```

### Arguments

data	A matrix or data frame containing columns that need to be converted
conversion	the conversion to apply, e.g. <a href="#">as.factor</a> or a custom-made function
cols	the columns of 'data' to convert

### Details

Sometimes we need to change the data type (class, mode) of a variable in R. There are various possible conversions, performed by functions like [as.integer](#), [as.factor](#) or [as.character](#). If we need to perform the same conversion on a number of variables (columns) in a data frame, we can convert them all simultaneously using this function. By default it converts all columns in 'data', but you can specify just some of those. 'multConvert' can also be used to apply other kinds of transformations – for example, if you need to divide some of your columns by 100, just write a function to do this and then use 'multConvert' to apply this function to any group of columns.

### Value

The input data with the specified columns converted as specified in 'conversion'.

### Author(s)

A. Marcia Barbosa

## Examples

```
data(rotif.env)

str(rotif.env)

# convert the first 4 columns to character:
converted.rotif.env <- multConvert(data = rotif.env,
  conversion = as.character, cols = 1:4)

str(converted.rotif.env)

names(rotif.env)

# divide some columns by 100:

div100 <- function(x) x / 100

rotif.env.cent <- multConvert(data = rotif.env,
  conversion = div100, cols = c(6:10, 12:17))

head(rotif.env.cent)
```

---

multGLM

*GLMs with variable selection for multiple species*


---

## Description

This function performs selection of variables and calculates generalized linear models for a set of presence/absence records in a data frame, with a range of options for data partition, variable selection, and output form.

## Usage

```
multGLM(data, sp.cols, var.cols, id.col = NULL, family = "binomial",
  test.sample = 0, FDR = FALSE, correction = "fdr", corSelect = FALSE,
  cor.thresh = 0.8, cor.method = "pearson", step = TRUE, trace = 0,
  start = "null.model", direction = "both", select = "AIC", trim = TRUE,
  trim.fun = "modelTrim", Y.prediction = FALSE, P.prediction = TRUE,
  Favourability = TRUE, group.preds = TRUE, TSA = FALSE, coord.cols = NULL,
  degree = 3, verbosity = 2, ...)
```

## Arguments

data	a data frame in wide format (see <a href="#">splist2presabs</a> ) containing, in separate columns, your species' binary (0/1) occurrence data and the predictor variables.
sp.cols	names or index numbers of the columns containing the species data to be modelled.

<code>var.cols</code>	names or index numbers of the columns containing the predictor variables to be used for modelling.
<code>id.col</code>	(optional) name or index number of column containing the row identifiers (if defined, it will be included in the output 'predictions' data frame).
<code>family</code>	argument to be passed to the <a href="#">glm</a> function; currently, only 'binomial' is implemented here.
<code>test.sample</code>	a subset of data to set aside for subsequent model testing. Can be a value between 0 and 1 for a proportion of the data to choose randomly (e.g. 0.2 for 20%); or an integer number for a particular number of cases to choose randomly among the records in 'data'; or a vector of integers for the index numbers of the particular rows to set aside; or "Huberty" for his rule of thumb based on the number of variables (Huberty 1994, Fielding & Bell 1997).
<code>FDR</code>	logical value indicating whether to do a preliminary exclusion of variables based on the false discovery rate (see <a href="#">FDR</a> ). The default is FALSE.
<code>correction</code>	argument to pass to the <a href="#">FDR</a> function if FDR = TRUE. The default is "fdr", but see <a href="#">p.adjust</a> for other options.
<code>corSelect</code>	logical value indicating whether to do a preliminary exclusion of highly correlated variables (see <a href="#">corSelect</a> ). The default is FALSE.
<code>cor.thresh</code>	numerical value indicating the correlation threshold to pass to <a href="#">corSelect</a> (used only if <code>corSelect</code> = TRUE).
<code>cor.method</code>	character value to pass to <a href="#">corSelect</a> (used only if <code>corSelect</code> = TRUE) specifying the correlation coefficient to use. Can be "pearson" (the default), "kendall" or "spearman".
<code>step</code>	logical, whether to use the <a href="#">step</a> function to perform a stepwise variable selection (based on AIC or BIC).
<code>trace</code>	if positive, information is printed during the running of <a href="#">step</a> . Larger values may give more detailed information.
<code>start</code>	character string specifying whether to start with the 'null.model' (so that variable selection starts forward) or with the 'full.model' (so selection starts backward). Used only if <code>step</code> = TRUE.
<code>direction</code>	argument to be passed to <a href="#">step</a> specifying the direction of variable selection ('forward', 'backward' or 'both'). Used only if <code>step</code> = TRUE.
<code>select</code>	character string specifying the criterion for stepwise selection of variables. Options are "AIC" (Akaike's Information Criterion; Akaike, 1973), the default; or BIC (Bayesian Information Criterion, also known as Schwarz criterion, SBC or SBIC; Schwarz, 1978). Used only if <code>step</code> = TRUE.
<code>trim</code>	logical value indicating whether to trim off non-significant variables from the model(s), using either <a href="#">modelTrim</a> or backward <a href="#">stepwise</a> selection (depending on the specified 'trim.fun' below). This argument is TRUE by default, and it can be used whether or not <code>step=TRUE</code> – e.g. Crawley (2005, p. 208) and Crawley (2007, p. 442 and 601) recommend that <a href="#">step</a> be followed by significance-based backward elimination).
<code>trim.fun</code>	character value indicating the name of the function to use for removing non-significant variables if <code>trim=TRUE</code> . Can be " <a href="#">modelTrim</a> " (the default) or " <a href="#">stepwise</a> " (which will be performed backward).



Y.prediction	logical, whether to include output predictions in the scale of the predictor variables (type = "link" in <code>predict.glm</code> ).
P.prediction	logical, whether to include output predictions in the scale of the response variable, i.e. probability (type = "response" in <code>predict.glm</code> ).
Favourability	logical, whether to apply the <code>Favourability</code> function to remove the effect of prevalence on predicted probability (Real et al. 2006) and include its results in the output.
group.preds	logical, whether to group together predictions of similar type ('Y', 'P' or 'F') in the output 'predictions' table (e.g. if FALSE: sp1_Y, sp1_P, sp1_F, sp2_Y, sp2_P, sp2_F; if TRUE: sp1_Y, , sp2_Y, sp1_P, sp2_P, sp1_F, sp2_F).
TSA	logical, whether to add a trend surface analysis (calculated individually for each species) as a spatial variable in each model. See <code>multTSA</code> for more details. The default is FALSE.
coord.cols	argument to pass to <code>multTSA</code> (if TSA = TRUE).
degree	argument to pass to <code>multTSA</code> (if TSA = TRUE).
verbosity	numeric value indicating the amount of messages to display; currently meaningful values are 0, 1, and 2 (the default).
...	additional arguments for variable selection criteria to pass to the function specified in 'trim.fun'.

## Details

This function automatically calculates binomial GLMs for one or more species (or other binary variables) in a data frame. The function can optionally perform `stepwise` variable selection (and it does so by default) instead of forcing all variables into the models, starting from either the null model (the default, so selection starts forward) or from the full model (so selection starts backward) and using Akaike's information criterion (AIC) as a variable selection criterion. Instead or subsequently, it can also perform stepwise removal of non-significant variables from the models using the `modelTrim` function.

There is also an optional preliminary selection of non-correlated variables, and/or of variables with a significant bivariate relationship with the response, based on the false discovery rate (FDR). Note, however, that some variables can be significant in a multivariate model even if they would not have been selected by FDR.

`Favourability` is also calculated, removing the effect of species prevalence from occurrence probability and thus allowing direct comparisons between models (Real et al. 2006).

By default, all data are used in model training, but you can define an optional 'test.sample' to be reserved for model testing afterwards. You may also want to do a previous check for multicollinearity among variables, e.g. the variance inflation factor (VIF), using `multicol`.

The 'multGLM' function will create a list of the resulting models (each with the name of the corresponding species column) and a data frame with their predictions ('Y', 'P' and/or 'F', all of which are optional). If you plan on representing these predictions in a GIS based on .dbf tables, remember that dbf only allows up to 10 characters in column names; 'multGLM' predictions will add 2 characters (\_Y, \_P and/or \_F) to each of your species column names, so use species names/codes with up to 8 characters in the data set that you are modelling. You can create (sub)species name abbreviations with the `spCodes` function.

**Value**

This function returns a list with the following components:

predictions	a data frame with the model predictions (if either of Y.prediction, P.prediction or Favourability are TRUE).
models	a list of the resulting model objects.
variables	a list of character vectors naming the variables finally included in each model according to the specified selection criteria.

**Note**

With step=TRUE (the default), an error may occur if there are missing values in some of the variables that are selected (see "Warning" in [step](#)). If this happens, you can use something like `data=na.omit(data[, var.cols])`.

**Author(s)**

A. Marcia Barbosa

**References**

- Akaike, H. (1973) Information theory and an extension of the maximum likelihood principle. In: Petrov B.N. & Csaki F., 2nd International Symposium on Information Theory, Tsahkadsor, Armenia, USSR, September 2-8, 1971, Budapest: Akademiai Kiado, p. 267-281.
- Crawley, M.J. (2005) Statistics: An introduction using R. John Wiley & Sons, Ltd.
- Crawley, M.J. (2007) The R Book. John Wiley & Sons, Ltd.
- Fielding A.H. & Bell J.F. (1997) A review of methods for the assessment of prediction errors in conservation presence/absence models. *Environmental Conservation* 24: 38-49
- Huberty C.J. (1994) Applied Discriminant Analysis. Wiley, New York, 466 pp.
- Schaafsma W. & van Vark G.N. (1979) Classification and discrimination problems with applications. Part IIa. *Statistica Neerlandica* 33: 91-126
- Real R., Barbosa A.M. & Vargas J.M. (2006) Obtaining environmental favourability functions from logistic regression. *Environmental and Ecological Statistics* 13: 237-245.
- Schwarz, G.E. (1978) Estimating the dimension of a model. *Annals of Statistics*, 6 (2): 461-464.

**See Also**

[glm](#), [step](#), [stepwise](#)

**Examples**

```
data(rotif.env)

names(rotif.env)

# make models for 2 of the species in rotif.env:
```

```

mods <- multGLM(rotif.env, sp.cols = 46:47, var.cols = 5:17, id.col = 1,
step = TRUE, FDR = TRUE, trim = TRUE)

names(mods)
head(mods$predictions)
names(mods$models)
mods$models[[1]]
mods$models[["Ttetra"]]

# include each species' spatial trend in the models:

mods <- multGLM(rotif.env, sp.cols = 46:47, var.cols = 5:17, id.col = 1,
step = TRUE, FDR = TRUE, trim = TRUE, TSA = TRUE, coord.cols = c(11, 10))

mods$models[[1]]
mods$models[["Ttetra"]]

mods$variables
# you can then use these selected variables elsewhere

```

---

multicol

---

*Analyse multicollinearity in a dataset, including VIF*


---

## Description

This function analyses multicollinearity in a set of variables or in a model, including the R-squared, tolerance and variance inflation factor (VIF).

## Usage

```
multicol(vars = NULL, model = NULL, reorder = TRUE)
```

## Arguments

vars	A matrix or data frame containing the numeric variables for which to calculate multicollinearity. Only the 'independent' (predictor, explanatory, right hand side) variables should be entered, as the result obtained for each variable depends on all the other variables present in the analysed data set.
model	Alternatively to 'vars', a model object of class "glm" to calculate 'multicol' among the included variables.
reorder	logical, whether variables should be output in decreasing order or VIF value rather than in their input order. The default is TRUE.

## Details

Testing collinearity among covariates is a recommended step of data exploration before applying a statistical model (Zuur et al. 2010). However, you can also calculate multicollinearity among the variables already included in a model.

The multicol function calculates the degree of multicollinearity in a set of numeric variables, using three closely related measures: R squared (the coefficient of determination of a linear regression of each predictor variable on all other predictor variables, i.e., the amount of variation in each variable that is accounted for by other variables in the dataset); tolerance ( $1 - R^2$ ), i.e. the amount of variation in each variable that is not included in the remaining variables; and the variance inflation factor:  $VIF = 1 / (1 - R^2)$ , which, in a linear model with these variables as predictors, reflects the degree to which the variance of an estimated regression coefficient is increased due only to the correlations among covariates (Marquardt 1970; Mansfield & Helms 1982).

## Value

The function returns a matrix with one row per analysed variable, the names of the variables as row names, and 3 columns: R-squared, Tolerance and VIF.

## Author(s)

A. Marcia Barbosa

## References

- Marquardt D.W. (1970) Generalized inverses, ridge regression, biased linear estimation, and non-linear estimation. *Technometrics* 12: 591-612.
- Mansfield E.R. & Helms B.P. (1982) Detecting multicollinearity. *The American Statistician* 36: 158-160.
- Zuur A.F., Ieno E.N. & Elphick C.S. (2010) A protocol for data exploration to avoid common statistical problems. *Methods in Ecology and Evolution* 1: 3-14.

## See Also

vif in package **HH**, vif in package **usdm**

## Examples

```
data(rotif.env)
names(rotif.env)

# calculate multicollinearity among the predictor variables:
multicol(rotif.env[ , 5:17], reorder = FALSE)
multicol(rotif.env[ , 5:17])

# you can also calculate multicol among the variables included in a model:
mod <- step(glm(Abrigh ~ Area + Altitude + AltitudeRange +
HabitatDiversity + HumanPopulation + Latitude + Longitude +
Precipitation + PrecipitationSeasonality + TemperatureAnnualRange
+ Temperature + TemperatureSeasonality + UrbanArea,
```

```

data = rotif.env))
multicol(model = mod)

# more examples using R datasets:
multicol(trees)

# you'll get a warning and some NA results if any of the variables
# is not numeric:
multicol(OrchardSprays)

# so define the subset of numeric 'vars' to calculate 'multicol' for:
multicol(OrchardSprays[ , 1:3])

```

multTSA

*Trend Surface Analysis for multiple species*

## Description

This function performs trend surface analysis for one or more species at a time. It converts categorical presence-absence (1-0) data into continuous surfaces denoting the spatial trend in species' occurrence patterns.

## Usage

```

multTSA(data, sp.cols, coord.cols, id.col = NULL, degree = 3,
step = TRUE, criterion = "AIC", type = "P", Favourability = FALSE,
suffix = "_TS", save.models = FALSE, verbosity = 2, ...)

```

## Arguments

data	a matrix or data frame containing, at least, two columns with spatial coordinates, and one column per species containing their presence (1) and absence (0) data, with localities in rows.
sp.cols	names or index numbers of the columns containing the species presences and absences in data. Must contain only zeros (0) for absences and ones (1) for presences.
coord.cols	names or index numbers of the columns containing the spatial coordinates in data (x and y, or longitude and latitude, in this order!).
id.col	optionally, the name or index number of a column (to be included in the output) containing locality identifiers in data.
degree	the degree of the spatial polynomial to use (see Details). The default is 3.
step	logical value indicating whether the regression of presence-absence on the spatial polynomial should do a stepwise inclusion of the polynomial terms (using the <a href="#">step</a> function with default settings, namely backward AIC selection), rather than forcing all terms into the equation. The default is TRUE.

criterion	character value indicating whether the backward stepwise selection of variables (if <code>step = TRUE</code> ) should be made according to "AIC" (the default, using the <code>step</code> function) or to "significance" (using the <code>modelTrim</code> function).
type	the type of trend surface to obtain. Can be either "Y" for the raw polynomial equation (i.e. in the scale of the predictors, e.g. if you want to use the spatial trend as a predictor variable in a model), "P" for the logit-transformed probability (e.g. if you want to use the output as a prediction of presence probability based on spatial trend alone), or "F" for spatial favourability, i.e., prevalence-independent probability (see <code>Fav</code> ).
Favourability	deprecated argument; <code>linktype</code> should now be used instead, although (at least for the timebeing) this argument will still be accepted (with <code>Favourability=TRUE</code> internally resulting in <code>type="F"</code> ) for back-compatibility.
suffix	character indicating the suffix to add to the trend surface columns in the resulting data frame. The default is "_TS".
save.models	logical value indicating whether the models obtained from the regressions should be saved and included in the output. The default is <code>FALSE</code> .
verbosity	integer value indicating the amount of messages to display; currently meaningful values are 0, 1, and 2 (the default).
...	additional arguments to be passed to <code>modelTrim</code> (if <code>step = TRUE</code> and <code>criterion = "significance"</code> ).

## Details

Trend Surface Analysis is a way to model the spatial structure in species' distributions by regressing occurrence data on the spatial coordinates  $x$  and  $y$ , for a linear trend, or on polynomial terms of these coordinates ( $x^2$ ,  $y^2$ ,  $x*y$ , etc.), for curvilinear trends (Legendre & Legendre, 1998; Borcard et al., 2011). Second- and third-degree polynomials are often used. 'multTSA' allows specifying the degree of the spatial polynomial to use. By default, it uses a 3rd-degree polynomial and performs stepwise AIC selection of the polynomial terms to include.

## Value

This function returns a matrix or data frame containing the identifier column (if provided in 'id.col') and one column per species containing the value predicted by the trend surface analysis. If `save.models = TRUE`, the output is a list containing this dataframe plus a list of the model objects.

## Author(s)

A. Marcia Barbosa

## References

Borcard D., Gillet F. & Legendre P. (2011) Numerical Ecology with R. Springer, New York.  
 Legendre P. & Legendre L. (1998) Numerical Ecology. Elsevier, Amsterdam.

## See Also

`distPres`, `poly`, `multGLM`

## Examples

```
data(rotif.env)

head(rotif.env)

names(rotif.env)

tsa <- multTSA(rotif.env, sp.cols = 18:20,
coord.cols = c("Longitude", "Latitude"), id.col = 1)

head(tsa)
```

---

pairwiseRangemaps	<i>Pairwise intersection (and union) of range maps</i>
-------------------	--

---

## Description

This function takes a set of rangemaps and returns a matrix containing the areas of their pairwise intersections; optionally, also their individual areas and/or their areas of pairwise unions.

## Usage

```
pairwiseRangemaps(rangemaps, projection, diag = TRUE, unions = TRUE,
verbosity = 2, Ncpu = 1, nchunks = 1, subchunks = NULL,
filename = "rangemap_matrix.csv")
```

## Arguments

rangemaps	a character vector of rangemap filenames, including folder paths if not in the working directory. ESRI shapefile (.shp) is currently the only accepted format. Specifying the extension is optional.
projection	argument to be passed to function 'importShapefile' of package <b>PBSmapping</b>
diag	logical, whether to fill the diagonal of the resulting matrix with the area of each rangemap. The default is TRUE, and it is also automatically set to TRUE (as it is necessary) if unions = TRUE.
unions	logical, whether to fill the upper triangle of the resulting matrix with the area of union of each pair of rangemaps. The default is TRUE. It is not as computationally intensive as the intersection, as it is calculated not with spatial but with algebraic operations within the matrix (union = area1 + area2 - intersection).
verbosity	integer number indicating the amount of progress messages to display.
Ncpu	integer indicating the number of CPUs (central processing units) to employ if parallel computing is to be used. The default is 1 CPU, which implies no parallel computing, but you may want to increase this if you have many and/or large rangemaps and your machine has more cores that can be used simultaneously. You can find out the total number of cores in your machine with the <a href="#">detectCores</a> function of the <a href="#">parallel</a> package; a usually wise option is to use all cores except one (i.e., Ncpu = parallel::detectCores()-1).

nchunks	either an integer indicating the number of chunks of rows in which to divide the results matrix for calculations, or character "decreasing" to indicate that the matrix should be divided into chunks of decreasing number of rows (as intersections are calculated in the lower triangle, rows further down the matrix have an increasing number of intersections to compute). Note, however, that rangemap size, not rangemap number, is the main determinant of computation time. The default is 1 (no division of the matrix) but, if you have many rangemaps, the process can get clogged. With chunks, each set of rows of the matrix is calculated and saved to disk, and the memory is cleaned before the next chunk begins.
subchunks	optional integer vector specifying which chunks to actually calculate. This is useful if a previous, time-consuming run of pairwiseRangemaps was interrupted (e.g. by a power outage) and you want to calculate only the remaining chunks.
filename	optional character vector indicating the name of the file to save the resulting matrix to.

### Details

This calculation can be very intensive and slow, especially if you have many and/or large rangemaps, due to the time needed for spatial operations between maps. You can set `nchunks = "decreasing"` for the matrix to be calculated in parts and the memory cleaned between one part and the next; and, if your computer has more than one core that you can use, you can increase `'Ncpu'` to get parallel computing.

### Value

This function returns a square matrix containing, in the lower triangle, the area of the pair-wise intersections among the input 'rangemaps'; in the diagonal (if `diag = TRUE` or `union = TRUE`), the area of each rangemap; and in the upper triangle (if `union = TRUE`), the area of the pair-wise unions among the rangemaps.

### Note

This function uses the **PBSmapping** package to import and intersect the polygon rangemaps and to calculate areas. Remember to use projected rangemaps, preferably with an equal-area reference system, so that calculations are correct.

### Author(s)

A. Marcia Barbosa

### References

Barbosa A.M. & Estrada A. (2016) Calcular corotipos sin dividir el territorio en OGU: una adaptacion de los indices de similitud para su utilizacion directa sobre areas de distribucion. In: Gomez Zotano J., Arias Garcia J., Olmedo Cobo J.A. & Serrano Montes J.L. (eds.), *Avances en Biogeografia. Areas de Distribucion: Entre Puentes y Barreras*, pp. 157-163. Editorial Universidad de Granada & Tundra Ediciones, Granada (Spain)



**See Also**[rangemapSim](#)

---

percentTestData	<i>Percent test data</i>
-----------------	--------------------------

---

**Description**

Based on the work of Schaafsma & van Vark (1979), Huberty (1994) provided a heuristic ("rule of thumb") for determining an adequate proportion of data to set aside for testing species presence/absence models, based on the number of predictor variables that are used (Fielding & Bell 1997). The 'percentTestData' function calculates this proportion as a percentage.

**Usage**

```
percentTestData(nvar)
```

**Arguments**

nvar                      the number of variables in the model.

**Value**

A numeric value of the percentage of data to leave out of the model for further model testing.

**Author(s)**

A. Marcia Barbosa

**References**

Huberty C.J. (1994) Applied Discriminant Analysis. Wiley, New York, 466 pp.  
Schaafsma W. & van Vark G.N. (1979) Classification and discrimination problems with applications. Part IIa. Statistica Neerlandica 33: 91-126  
Fielding A.H. & Bell J.F. (1997) A review of methods for the assessment of prediction errors in conservation presence/absence models. Environmental Conservation 24: 38-49

**See Also**[multGLM](#)**Examples**

```
# say you're building a model with 15 variables:

percentTestData(15)

# the result tells you that 21% is an appropriate percentage of data
# to set aside for testing your model, so train it with 79% of the data
```

prevalence

*Prevalence***Description**

Prevalence is the proportion of presences of a species in a dataset, which is required (together with presence probability) for computing [Favourability](#).

**Usage**

```
prevalence(obs, model = NULL, event = 1, na.rm = TRUE)
```

**Arguments**

obs	a vector or a factor of binary observations (e.g. 1 vs. 0, male vs. female, disease vs. no disease, etc.). This argument is ignored if 'model' is provided.
model	alternatively to 'obs', a binary-response model object of class "glm", "gam", "gbm", "randomForest" or "bart". If this argument is provided, 'obs' will be extracted with <a href="#">mod2obspred</a> .
event	the value whose prevalence we want to calculate (e.g. 1, "present", etc.). This argument is ignored if 'model' is provided.
na.rm	logical, whether NA values should be excluded from the calculation. The default is TRUE.

**Value**

Numeric value of the prevalence of event in the obs vector.

**Author(s)**

A. Marcia Barbosa

**Examples**

```
# calculate prevalence from binary vectors:

(x <- rep(c(0, 1), each = 5))

(y <- c(rep(0, 3), rep(1, 7)))

(z <- c(rep(0, 7), rep(1, 3)))

prevalence(x)

prevalence(y)

prevalence(z)
```

```

(w <- c(rep("yes", 3), rep("nope", 7)))

prevalence(w, event = "yes")

# calculate prevalence from a model object:

data(rotif.env)

mod <- glm(Abrigh ~ HabitatDiversity + HumanPopulation, family = binomial, data = rotif.env)

prevalence(model = mod)

# same as:
prevalence(obs = rotif.env$Abrigh)

```

---

rangemapSim	<i>Pairwise similarity between rangemaps</i>
-------------	--

---

## Description

Calculate pairwise similarity among rangemaps from a matrix of their areas of intersection and union

## Usage

```

rangemapSim(rangemap.matrix, total.area,
method = c("Jaccard", "Sorensen", "Simpson", "Baroni"),
diag = FALSE, upper = FALSE, verbosity = 2)

```

## Arguments

rangemap.matrix	a matrix like the one produced by function <a href="#">pairwiseRangemaps</a> , containing the areas of pairwise intersection among rangemaps in the lower triangle, individual rangemap areas in the diagonal, and pairwise union areas in the upper diagonal.
total.area	numeric value indicating the total size of the study area, in the same units as the areas in the rangemap.matrix. Used only if 'method' uses shared absences (as is the case of "Baroni")
method	character value indicating the similarity index to use. Currently implemented indices are "Jaccard", "Sorensen", "Simpson" and "Baroni". The default is the first one.
diag	logical value indicating if the diagonal of the resulting matrix should be filled
upper	logical value indicating if the upper triangle of the resulting matrix should be filled (symmetrical to the lower triangle)
verbosity	integer number indicating the amount of messages to display.

### Details

Distributional relationships among species are commonly determined based on pair-wise (dis)similarities in species' occurrence patterns. Some of the most commonly employed similarity indices are those of Jaccard (1901), Sorensen (1948), Simpson (1960) and Baroni-Urbani & Buser (1976), which are here implemented for comparing rangemaps based on their areas of intersection and union (Barbosa & Estrada, in press).

### Value

This function returns a square matrix of pairwise similarities between the rangemaps in 'rangemap.matrix', calculated with the (first) similarity index specified in 'method'.

### Author(s)

A. Marcia Barbosa

### References

- Barbosa A.M. & Estrada A. (in press) Calcular corotipos sin dividir el territorio en OGUs: una adaptacion de los indices de similitud para su utilizacion directa sobre areas de distribucion. In: Areas de distribucion: entre puentes y barreras. Universidad de Granada, Spain.
- Baroni-Urbani C. & Buser M.W. (1976) Similarity of Binary Data. *Systematic Zoology*, 25: 251-259
- Jaccard P. (1901) Etude comparative de la distribution florale dans une portion des Alpes et des Jura. *Memoires de la Societe Vaudoise des Sciences Naturelles*, 37: 547-579
- Simpson G.G. (1960) Notes on the measurement of faunal resemblance. *Amer. J. Sci.* 258A, 300-311
- Sorensen T. (1948) A method of establishing groups of equal amplitude in plant sociology based on similarity of species and its application to analyses of the vegetation on Danish commons. *Kongelige Danske Videnskabernes Selskab*, 5(4): 1-34

### See Also

[pairwiseRangemaps](#); [simFromSetOps](#); [simMat](#)

---

rarity

(Fuzzy) rarity

---

### Description

This function computes the index of species rarity of Real et al. (2006), using either crisp (presence/absence, i.e. ones and zeros) or fuzzy values (e.g. [Favourability](#), between zero and one), for a single species or for several species across a study area. Rarity is like a (potential) richness index in which rarer species have higher weight.

**Usage**

```
rarity(data, sp.cols = 1:ncol(data), na.rm = TRUE)
```

**Arguments**

data	a numeric vector, matrix or data frame containing the presence/absence or the <a href="#">Favourability</a> (fuzzy presence) values for the target species.
sp.cols	names or index numbers of the columns of 'data' that contain the species values for which to compute rarity. The default is to use all columns.
na.rm	logical value indicating whether NA values should be removed before the computation.

**Details**

If the input data include only one species (i.e. a numeric vector or a one-column table, with one value for each locality), rarity is 1 divided by the sum of its values. If the input includes more than one species or column, rarity is the sum of the product of each (fuzzy) presence value by the rarity of the corresponding species, so that rarer species have higher weight in the resulting sum (Real et al. 2006). See also Estrada et al. (2011) for a more complex version of fuzzy rarity.

**Value**

If 'data' is a vector or a one-column table, or if 'sp.cols' is of length 1, this function returns a single value of rarity for the underlying species, which is simply 1 divided by the sum of its values. If 'data' and 'sp.cols' refer to more than 1 column, the function returns the total combined rarity value of all corresponding species for each row in 'data' (see Examples).

**Author(s)**

A. Marcia Barbosa

**References**

Real R., Estrada A., Barbosa A.M. & Vargas J.M. (2006) Aplicacion de la logica difusa al concepto de rareza para su uso en Gap Analysis: el caso de los mamíferos terrestres en Andalucía. Serie Geografica 13: 99-116

Estrada A., Real R. & Vargas J.M. (2011) Assessing coincidence between priority conservation areas for vertebrate groups in a Mediterranean hotspot. Biological Conservation, 144: 1120-1129

**See Also**

[vulnerability](#)

**Examples**

```
data(rotif.env)
```

```
head(rotif.env)
```

```

rarity(rotif.env[, 18])

rarity(rotif.env, sp.cols = "Abrigh")

rarity(rotif.env, sp.cols = 18:47)
# yields one value of combined rarity for each row in 'data'

# fuzzy rarity (from favourability values):

pred <- multGLM(rotif.env, sp.cols = 18:20, var.cols = 5:17)$predictions

head(pred)

rarity(pred, sp.cols = "Abrigh_F")

rarity(pred, sp.cols = c("Abrigh_F", "Afissa_F", "Apriod_F"))
# yields one value of combined rarity for each row in 'data'

```

---

rotif.env	<i>Rotifers and environmental variables on TDWG level 4 regions of the world</i>
-----------	--

---

## Description

These data were extracted from a database of monogonont rotifer species presence records on the geographical units used by the Biodiversity Information Standards (formerly Taxonomic Database Working Group, TDWG: <https://www.tdwg.org>) and a few environmental (including human and spatial) variables on the same spatial units. The original data were compiled and published by Fontaneto et al. (2012) in long (narrow, stacked) format. Here they are presented in wide or unstacked format (presence-absence table, obtained with the `splist2presabs` function), reduced to the species recorded in at least 100 (roughly one third) different TDWG level 4 units, and with abbreviations of the species' names (obtained with the `spCodes` function). Mind that this is not a complete picture of these species' distributions, due to insufficient sampling in many regions.

## Usage

```
data(rotif.env)
```

## Format

A data frame with 291 observations on the following 47 variables.

TDWG4 a factor with 291 levels indicating the abbreviation code of each TDWG4 region

LEVEL\_NAME a factor with 291 levels indicating the name of each TDWG4 region

REGION\_NAME a factor with 47 levels indicating the name of the main geographical region to which each TDWG4 level belongs

CONTINENT a factor with 9 levels indicating the continent to which each TDWG4 level belongs

Area a numeric vector  
Altitude a numeric vector  
AltitudeRange a numeric vector  
HabitatDiversity a numeric vector  
HumanPopulation a numeric vector  
Latitude a numeric vector  
Longitude a numeric vector  
Precipitation a numeric vector  
PrecipitationSeasonality a numeric vector  
TemperatureAnnualRange a numeric vector  
Temperature a numeric vector  
TemperatureSeasonality a numeric vector  
UrbanArea a numeric vector  
Abrigh a numeric vector  
Afissa a numeric vector  
Apriod a numeric vector  
Bangul a numeric vector  
Bcalyc a numeric vector  
Bplica a numeric vector  
Bquadr a numeric vector  
Burceo a numeric vector  
Cgibba a numeric vector  
Edilat a numeric vector  
Flongi a numeric vector  
Kcochl a numeric vector  
Kquadr a numeric vector  
Ktropi a numeric vector  
Lbulla a numeric vector  
Lclost a numeric vector  
Lhamat a numeric vector  
Lluna a numeric vector  
Llunar a numeric vector  
Lovali a numeric vector  
Lpatel a numeric vector  
Lquadr a numeric vector  
Mventr a numeric vector  
Ppatul a numeric vector

Pquadr a numeric vector  
 Pvulga a numeric vector  
 Specti a numeric vector  
 Tpatin a numeric vector  
 Tsimil a numeric vector  
 Ttetra a numeric vector

### Source

Fontaneto D., Barbosa A.M., Segers H. & Pautasso M. (2012) The 'rotiferologist' effect and other global correlates of species richness in monogonont rotifers. *Ecography*, 35: 174-182.

### Examples

```
data(rotif.env)

head(rotif.env)
```

---

rotifers

*Rotifer species on TDWG level 4 regions of the world*

---

### Description

These data were extracted from a database of monogonont rotifer species records on the geographical units used by the Biodiversity Information Standards (formerly Taxonomic Database Working Group, TDWG: <https://www.tdwg.org>). The original data were compiled and published by Fontaneto et al. (2012) for all TDWG levels. Here they are reduced to the TDWG - level 4 units and to the species recorded in at least 100 (roughly one third) of these units. Mind that this is not a complete picture of these species' distributions, due to insufficient sampling in many regions.

### Usage

```
data("rotifers")
```

### Format

A data frame with 3865 observations on the following 2 variables.

TDWG4 a factor with 274 levels corresponding to the code names of the TDWG level 4 regions in which the records were taken

species a factor with 30 levels corresponding to the names of the (sub)species recorded in at least 100 different TDWG level 4 regions

### Source

Fontaneto D., Barbosa A.M., Segers H. & Pautasso M. (2012) The 'rotiferologist' effect and other global correlates of species richness in monogonont rotifers. *Ecography*, 35: 174-182.



**Examples**

```
data(rotifers)

head(rotifers, 10)
```

---

sharedFav	<i>Shared favourability for two competing species</i>
-----------	---

---

**Description**

This function implements the graphical analyses of Acevedo et al. (2010, 2012) on biogeographical interactions. It takes two vectors of favourability values at different localities for, respectively, a stronger and a weaker competing species (or two equally strong competitors), and plots their favourableness or shared favourability to assess potential competitive interactions.

**Usage**

```
sharedFav(strong_F, weak_F, conf = 0.95, bin_interval = "0.1", ...)
```

**Arguments**

strong_F	a numeric vector of favourability values (obtained, e.g., with functions <a href="#">Fav</a> or <a href="#">multGLM</a> ) for the stronger species.
weak_F	a numeric vector of favourability values for the weaker species. Must be of the same length and in the same order as strong_F.
conf	confidence level for the confidence intervals in the plot. The default is 0.95. Set it to NA for no confidence intervals (see "Note" below).
bin_interval	character value specifying the method for grouping the favourability values into bins, to plot and compare mean favourability for each species. Currently implemented options are "0.1" (the default, dividing the values at 0.1 intervals as per Acevedo et al. 2010, 2012) and "quantiles" (as the former method may generate an error if there are overly small bins to calculate confidence intervals). See "Note" below.
...	some additional arguments can be passed to <a href="#">barplot</a> , such as "main" (for the plot title) or "las" (for the orientation of the axis labels).

**Details**

This function implements the biogeographic analyses of Acevedo et al. (2010, 2012), assessing the trends of environmental favourability across a range of favourability intersection values between two interacting species. It first calculates the fuzzy intersection (minimum value) between the two species' favourability values at each locality (i.e., favourability for the occurrence of both species simultaneously); it groups these values into 10 bins; and calculates the mean favourability (and its confidence interval) for each of the two species within each interval.

According to the notion of "favorableness" by Richerson & Lum (1980), competing species may or may not be able to coexist depending on their relative environmental fitnesses; competition between

species increases and competitive exclusion decreases as their favourability intersection increases (Acevedo et al. 2010, 2012). The shaded area in the shared favourability plot, where at least one of the species is at intermediate favourability, is the area where competitive interactions may limit species occurrence. Outside this shaded area, where favourability is either very low for at least one of the species (left) or very high for both species (right side of the plot), competition is not limiting (see also [bioThreat](#) for details).

### Value

This function returns the numeric value of the fuzzy overlap index (FOvI; Dubois & Prade 1980, Acevedo et al. 2010, 2012), a data frame with the bin values and the shared favourability plot, with circles and a continuous line representing favourability for the stronger species, and squares and a dashed line representing favourability for the weaker species. The height of the bars at the bottom represents the proportional sample size of each bin.

### Note

This function may generate an error if one or more bins don't have enough values for the confidence interval to be computed. If this occurs, you can try a different 'bin\_interval' (e.g. "quantiles") or set the 'conf' argument to NA (in which case confidence intervals will not be computed). Either will affect only the plot, not the overall fuzzy overlap value.

### Author(s)

A. Marcia Barbosa

### References

- Acevedo P., Ward A.I., Real R. & Smith G.C. (2010) Assessing biogeographical relationships of ecologically related species using favourability functions: a case study on British deer. *Diversity and Distributions*, 16: 515-528
- Acevedo P., Jimenez-Valverde A., Melo-Ferreira J., Real R. & Alves, P.C. (2012) Parapatric species and the implications for climate change studies: a case study on hares in Europe. *Global Change Biology*, 18: 1509-1519
- Dubois D. & Prade H. (1980) *Fuzzy sets and systems: theory and applications*. Academic Press, New York
- Richerson P.J. & Lum K. (1980) Patterns of plant species and diversity in California: relation to weather and topography. *American Naturalist*, 116: 504-536

### See Also

[bioThreat](#), [Fav](#)

### Examples

```
# get favourability model predictions for two species:
data(rotif.env)
mods <- multGLM(rotif.env, sp.cols = 19:20, var.cols = 5:17)
head(mods$predictions)
```

```

favs <- mods$predictions[ , 3:4]

# get shared favourability:
sharedFav(strong_F = favs[,1], weak_F = favs[,2], main = "Shared favourability")

sharedFav(strong_F = favs[,1], weak_F = favs[,2], bin_interval = "quantiles",
main = "Shared favourability", las = 2)

```

---

simFromSetOps

---

*Calculate similarity from set operations*


---

## Description

This function calculates pair-wise similarity based on the results of set operations (intersection, union) among the subjects.

## Usage

```

simFromSetOps(size1, size2, intersection, union, total.size = NULL,
method = c("Jaccard", "Sorensen", "Simpson", "Baroni"),
verbosity = 1)

```

## Arguments

size1	size of subject 1 (e.g., area of the distribution range of a species, or its number of presences within a grid). Not needed if method = "Jaccard".
size2	the same for subject 2.
intersection	size of the intersection among subjects 1 and 2 (area of the intersection among their distribution ranges, or number of grid cells in which they co-occur).
union	size of the union of subjects 1 and 2.
total.size	total size of the study area. Needed only when calculating a similarity index that takes shared absences into account (i.e., method = "Baroni").
method	the similarity index to use. Currently implemented options are "Jaccard", "Sorensen", "Simpson" or "Baroni".
verbosity	integer indicating whether to display messages.

## Details

Similarities among ecological communities, beta diversity patterns, biotic regions, and distributional relationships among species are commonly determined based on pair-wise (dis)similarities in species' occurrence patterns. This function implements some of the most commonly employed similarity indices, namely those of Jaccard (1901), Sorensen (1948), Simpson (1960) and Baroni-Urbani & Buser (1976), based on the amount of occupied and overlap area between two species.

## Value

The numeric value of similarity among subjects 1 and 2.

**Author(s)**

A. Marcia Barbosa

**References**

Baroni-Urbani C. & Buser M.W. (1976) Similarity of Binary Data. *Systematic Zoology*, 25: 251-259

Jaccard P. (1901) Etude comparative de la distribution florale dans une portion des Alpes et des Jura. *Memoires de la Societe Vaudoise des Sciences Naturelles*, 37: 547-579

Simpson, G.G. (1960) Notes on the measurement of faunal resemblance. *Amer. J. Sci.* 258A, 300-311

Sorensen T. (1948) A method of establishing groups of equal amplitude in plant sociology based on similarity of species and its application to analyses of the vegetation on Danish commons. *Kongelige Danske Videnskabernes Selskab*, 5(4): 1-34

**See Also**

[fuzSim](#), [simMat](#)

**Examples**

```
# take two species which occur in 22 and 35 area units, respectively
# and which overlap in 8 of those units:
```

```
sp1 <- 22
sp2 <- 35
int <- 8
uni <- sp1 + sp2 - int
```

```
# calculate similarity between their distributions based on
# different indices:
```

```
simFromSetOps(intersection = int, union = uni, method = "Jaccard")
```

```
simFromSetOps(sp1, sp2, int, uni, method = "Sorensen")
```

```
simFromSetOps(sp1, sp2, int, uni, method = "Simpson")
```

```
# if you want Baroni-Urbani & Buser's index
# you need to provide also the total size of your study area:
```

```
simFromSetOps(sp1, sp2, int, uni, total = 100, method = "Baroni")
```

---

simMat	<i>Pair-wise (fuzzy) similarity matrix</i>
--------	--

---

## Description

simMat takes a matrix or data frame containing species occurrence data or regional species composition, either categorical (0 or 1) or fuzzy (between 0 and 1), and uses the [fuzSim](#) function to calculate a square matrix of pair-wise similarities between them, using a fuzzy logic version (Barbosa, 2015) of the specified similarity index.

## Usage

```
simMat(data, method, diag = TRUE, upper = TRUE, verbosity = 2)
```

## Arguments

data	a matrix or data frame containing (optionally fuzzy) species presence-absence data (in wide format, i.e. one column per species), with 1 meaning presence, 0 meaning absence, and values in between for fuzzy presence (or the degree to which each locality belongs to the set of species presences; see Zadeh, 1965). Fuzzy presence-absence can be obtained, for example, with <a href="#">multGLM</a> , <a href="#">distPres</a> or <a href="#">multTSA</a> . These data can also be <a href="#">transposed</a> for comparing regional species compositions.
method	the similarity index whose fuzzy version to use. See <a href="#">fuzSim</a> for available options.
diag	logical value indicating whether the diagonal of the matrix should be filled (with ones). Defaults to TRUE.
upper	logical value indicating whether the upper triangle of the matrix (symmetric to the lower triangle) should be filled. Defaults to TRUE.
verbosity	integer value indicating the amount of messages to display; currently meaningful values are 0, 1, and 2 (the default).

## Details

The fuzzy versions of species occurrence data and of binary similarity indices introduce tolerance for small spatial differences in species' occurrence localities, allow for uncertainty about species occurrence, and may compensate for under-sampling and geo-referencing errors (Barbosa, 2015).

## Value

This function returns a square matrix of pair-wise similarities among the species distributions (columns) in data. Similarity is calculated with the fuzzy version of the index specified in method, which yields traditional binary similarity if the data are binary (0 or 1), or fuzzy similarity if the data are fuzzy (between 0 and 1) (Barbosa, 2015).

**Author(s)**

A. Marcia Barbosa

**References**

Barbosa A.M. (2015) fuzzySim: applying fuzzy logic to binary similarity indices in ecology. *Methods in Ecology and Evolution*, 6: 853-858.

**See Also**

[fuzSim](#)

**Examples**

```
# load and look at the rotif.env presence-absence data:

data(rotif.env)

head(rotif.env)

names(rotif.env)

# build a matrix of similarity among these binary data
# using e.g. Jaccard's index:

bin.sim.mat <- simMat(rotif.env[, 18:47], method = "Jaccard")

head(bin.sim.mat)

# calculate a fuzzy version of the presence-absence data
# based on inverse distance to presences:

rotifers.invd <- distPres(rotif.env, sp.cols = 18:47,
  coord.cols = c("Longitude", "Latitude"), id.col = 1, suffix = ".d",
  p = 1, inv = TRUE)

head(rotifers.invd)

# build a matrix of fuzzy similarity among these fuzzy
# distribution data, using the fuzzy version of Jaccard's index:

fuz.sim.mat <- simMat(rotifers.invd[, -1], method = "Jaccard")

head(fuz.sim.mat)

# plot the similarity matrices as colours:
```

```

image(x = 1:ncol(bin.sim.mat), y = 1:nrow(bin.sim.mat),
      z = bin.sim.mat, col = rev(heat.colors(256)), xlab = "", ylab = "",
      axes = FALSE, main = "Binary similarity")
axis(side = 1, at = 1:ncol(bin.sim.mat), tick = FALSE,
      labels = colnames(bin.sim.mat), las = 2)
axis(side = 2, at = 1:nrow(bin.sim.mat), tick = FALSE,
      labels = rownames(bin.sim.mat), las = 2)

image(x = 1:ncol(fuz.sim.mat), y = 1:nrow(fuz.sim.mat),
      z = fuz.sim.mat, col = rev(heat.colors(256)), xlab = "", ylab = "",
      axes = FALSE, main = "Fuzzy similarity")
axis(side = 1, at = 1:ncol(fuz.sim.mat), tick = FALSE,
      labels = colnames(fuz.sim.mat), las = 2, cex = 0.5)
axis(side = 2, at = 1:nrow(fuz.sim.mat), tick = FALSE,
      labels = rownames(fuz.sim.mat), las = 2)

# plot a UPGMA dendrogram from each similarity matrix:

plot(hclust(as.dist(1 - bin.sim.mat), method = "average"),
     main = "Binary cluster dendrogram")

plot(hclust(as.dist(1 - fuz.sim.mat), method = "average"),
     main = "Fuzzy cluster dendrogram")

# you can get fuzzy chorotypes from these similarity matrices
# (or fuzzy biotic regions if you transpose 'data'),
# so that localities are in columns and species in rows)
# using the RMACOQUI package (Olivero et al. 2011)

```

---

spCodes

---

*Obtain unique abbreviations of species names*


---

## Description

This function takes a vector of species names and converts them to abbreviated species codes containing the specified numbers of characters from the genus, the specific and optionally also the subspecific name. Separators can be specified by the user. The function checks that the resulting codes are unique.

## Usage

```

spCodes(species, nchar.gen = 3, nchar.sp = 3, nchar.ssp = 0,
        sep.species = " ", sep.spcode = "", verbosity = 2)

```

## Arguments

**species**                      a character vector containig the species names to be abbreviated.

nchar.gen	the number of characters from the genus name to be included in the resulting species code.
nchar.sp	the number of characters from the specific name to be included in the resulting species code.
nchar.ssp	optionally, the number of characters from the subspecific name to be included in the resulting species code. Set it to 0 if you have subspecific names in 'species' but do not want them included in the resulting species codes.
sep.species	the character that separates genus, specific and subspecific names in 'species'. The default is a white space.
sep.spcode	the character you want separating genus and species abbreviations in the resulting species codes. The default is an empty character (no separator).
verbosity	integer value indicating the amount of messages to display. Defaults to 2, for showing all messages.

### Value

This function returns a character vector containing the species codes resulting from the abbreviation. If the numbers of characters specified do not make for unique codes, an error message is displayed showing which 'species' names caused it, so that you can try again with different 'nchar.gen', 'nchar.sp' and/or 'nchar.ssp'.

### Author(s)

A. Marcia Barbosa

### See Also

[substr](#), [strsplit](#)

### Examples

```
data(rotifers)

head(rotifers)

## add a column to 'rotifers' with shorter versions of the species names:

## Not run:
rotifers$spcode <- spCodes(rotifers$species, sep.species = "_",
nchar.gen = 1, nchar.sp = 4, nchar.ssp = 0, sep.spcode = ".")

# this produces an error due to resulting species codes not being unique

## End(Not run)

rotifers$spcode <- spCodes(rotifers$species, sep.species = "_",
nchar.gen = 1, nchar.sp = 5, nchar.ssp = 0, sep.spcode = ".")

# with a larger number of characters from the specific name,
```



```
# resulting codes are now unique

## check out the result:
head(rotifers)
```

---

`splist2presabs`*Convert a species list to a presence-absence table*

---

## Description

This function takes a locality+species dataset in long (stacked) format, i.e., a matrix or data frame containing localities in one column and their recorded species in another column, and converts them to a presence-absence table (wide format) suitable for mapping and for computing distributional similarities (see e.g. [simMat](#)). Try out the Examples below for an illustration).

## Usage

```
splist2presabs(data, sites.col, sp.col, keep.n = FALSE)
```

## Arguments

<code>data</code>	a matrix or data frame with localities in one column and species in another column. Type or paste <code>'data(rotifers); head(rotifers)'</code> (without the quote marks) in the R console for an example.
<code>sites.col</code>	the name or index number of the column containing the localities in <code>'data'</code> .
<code>sp.col</code>	the name or index number of the column containing the species names or codes in <code>'data'</code> .
<code>keep.n</code>	logical value indicating whether to get in the resulting table the number of times each species appears in each locality; if <code>FALSE</code> (the default), only presence (1) or absence (0) is recorded.

## Value

A data frame containing the localities in the first column and then one column per species indicating their presence or absence (or their number of records if `keep.n = TRUE`). Type `'data(rotif.env); head(rotif.env[,18:47])'` (without the quote marks) in the R console for an example.

## Author(s)

A. Marcia Barbosa

## See Also

[table](#)

## Examples

```
data(rotifers)

head(rotifers)

rotifers.presabs <- splist2presabs(rotifers, sites.col = "TDWG4",
sp.col = "species", keep.n = FALSE)

head(rotifers.presabs)
```

---

stepByStep

---

*Analyse and compare stepwise model predictions*


---

## Description

This function builds a generalized linear model with forward stepwise inclusion of variables, using AIC as the selection criterion, and provides the values predicted at each step, as well as their correlation with the final model predictions.

## Usage

```
stepByStep(data, sp.col, var.cols, family = binomial(link = "logit"),
Favourability = FALSE, trace = 0, direction = "forward", k = 2,
cor.method = "pearson")
```

## Arguments

data	a data frame containing your target and predictor variables.
sp.col	name or index number of the column of 'data' that contains the response variable.
var.cols	names or index numbers of the columns of 'data' that contain the predictor variables.
family	argument to be passed to the <a href="#">glm</a> function indicating the family (and error distribution) to use in modelling. The default is binomial distribution with logit link (for binary response variables).
Favourability	logical, whether to apply the <a href="#">Favourability</a> function to remove the effect of prevalence from predicted probability (Real et al. 2006). Applicable only to binomial GLMs. Defaults to FALSE.
trace	argument to pass to the <a href="#">step</a> function. If positive, information is printed during the stepwise procedure. Larger values may give more detailed information. The default is 0 (silent).
direction	argument to pass to the <a href="#">step</a> function. Implementation contributed by Alba Estrada. Can be "forward" (the default, for back-compatibility with former versions of 'stepByStep'), "backward", or "both".

<code>k</code>	argument to pass to the <a href="#">step</a> function indicating the multiple of the number of degrees of freedom used for the penalty. The default is 2, which yields the original AIC. You can use larger values for a more stringent selection– e.g., for a critical p-value of 0.05, use <code>k = qchisq(0.05, 1, lower.tail = F)</code> .
<code>cor.method</code>	character string to pass to <a href="#">cor</a> indicating which coefficient to use for correlating predictions at each step with those of the final model. Can be "pearson" (the default), "kendall", or "spearman".

## Details

Stepwise variable selection often includes more variables than would a model selected after examining all possible combinations of the variables (e.g. with package **MuMIn** or **glmulti**). The 'stepByStep' function can be useful to assess if a stepwise model with just the first few variables could already provide predictions very close to the final ones (see e.g. Fig. 3 in Munoz et al., 2005). It can also be useful to see which variables determine the more general trends in the model predictions, and which variables just provide additional (local) nuances.

## Value

This function returns a list of the following components:

<code>predictions</code>	a data frame with the model's fitted values at each step of the variable selection.
<code>correlations</code>	a numeric vector of the correlation between the predictions at each step and those of the final model.
<code>variables</code>	a character vector of the variables in the final model, named with the step at which each was included.
<code>model</code>	the resulting model object.

## Author(s)

A. Marcia Barbosa

## References

Munoz, A.R., Real R., BARBOSA A.M. & Vargas J.M. (2005) Modelling the distribution of Bonelli's Eagle in Spain: Implications for conservation planning. *Diversity and Distributions* 11: 477-486

Real R., Barbosa A.M. & Vargas J.M. (2006) Obtaining environmental favourability functions from logistic regression. *Environmental and Ecological Statistics* 13: 237-245.

## See Also

[step](#), [glm](#), [modelTrim](#)

Examples

```
data(rotif.env)

stepByStep(data = rotif.env, sp.col = 18, var.cols = 5:17,
cor.method = "spearman")

stepByStep(data = rotif.env, sp.col = 18, var.cols = 5:17,
cor.method = "spearman", Favourability = TRUE)

stepByStep(data = rotif.env, sp.col = 9, var.cols = c(5:8, 10:17),
family = poisson)
```

---

stepwise	<i>Stepwise regression</i>
----------	----------------------------

---

Description

This function runs a stepwise regression, selecting and/or excluding variables based on the significance (p-value) of the statistical tests implemented in the [add1](#) and [drop1](#) functions of R.

Usage

```
stepwise(data, sp.col, var.cols, id.col = NULL, family = binomial(link="logit"),
direction = "both", test.in = "Rao", test.out = "LRT", p.in = 0.05, p.out = 0.1,
trace = 1, simplif = TRUE, preds = FALSE, Favourability = FALSE, Wald = FALSE)
```

Arguments

data	a data frame (or an object that can be coerced with 'as.data.frame') containing your target and predictor variables.
sp.col	name or index number of the column of 'data' that contains the response variable.
var.cols	names or index numbers of the columns of 'data' that contain the predictor variables.
id.col	(optional) name or index number of column containing the row identifiers (if defined, it will be included in the output 'predictions' data frame).
family	argument to be passed to <a href="#">glm</a> indicating the error distribution (and optionally the link function) to be used in the model. The default is binomial distribution with logit link (i.e. logistic regression, for binary response variables), and it is the only one that has been tested so far. If you try other options, please carefully check your results and let me know if you find a bug.
direction	the mode of stepwise search. Can be either "forward", "backward", or "both" (the default).
test.in	argument to pass to <a href="#">add1</a> specifying the statistical test whose 'p.in' a variable must pass to enter the model. Can be "Rao" (the default), "LRT", "Chisq" or "F".

<code>test.out</code>	argument to pass to <a href="#">drop1</a> specifying the statistical test whose 'p.out' a variable must fail to be expelled from the model. Can be "LRT" (the default), "Rao", "Chisq" or "F".
<code>p.in</code>	threshold p-value for a variable to enter the model. Defaults to 0.05.
<code>p.out</code>	threshold p-value for a variable to leave the model. Defaults to 0.1.
<code>trace</code>	if positive, information is printed to the console at each step. The default is 1, for naming each variable that was added or removed. With <code>trace=2</code> , the summary of the model at each step is also printed.
<code>simplif</code>	logical, whether to return a simpler output containing only the model object (the default), or a list with, additionally, a data frame with the variable included or excluded at each step.
<code>preds</code>	logical, whether to return also the predictions given by the model at each step. This argument is ignored if <code>simplif=TRUE</code> .
<code>Favourability</code>	logical, whether to convert the predictions (if <code>preds=TRUE</code> ) with the <a href="#">Fav</a> function. This argument is ignored if <code>simplif=TRUE</code> .
<code>Wald</code>	logical, whether to report the Wald test statistics (rather than the z test normally returned by <a href="#">summary</a> ). Used only if <code>trace &gt; 1</code> . Requires the <b>aod</b> package.

## Details

Stepwise variable selection is a way of selecting a subset of significant variables to get a simple and easily interpretable model (e.g. Munoz et al., 2005). It is more computationally efficient than best subset selection. This function uses [add1](#) for selecting and [drop1](#) for excluding variables. The default parameters mimic the "Forward Selection (Conditional)" stepwise procedure implemented in the IBM SPSS software. This is a widely used, but also widely criticized method for variable selection (e.g. Harrell 2001; Flom & Cassell, 2007; Smith 2018), though its AIC-based counterpart (implemented in the [step](#) function) is also not without flaws (Real et al., in prep.).

## Value

If `simplif=TRUE` (the default), this function returns the model object obtained after the variable selection procedure. If `simplif=FALSE`, it returns a list with the following components:

<code>model</code>	the model object obtained after the variable selection procedure.
<code>steps</code>	a data frame where each row shows the variable included or excluded at each step.
<code>predictions</code>	(if <code>preds=TRUE</code> ) a data frame where each column contains the predictions of the model at each step. These predictions are probabilities by default, or favourabilities if <code>Favourability=TRUE</code> .

## Author(s)

A. Marcia Barbosa

## References

- Flom P.L. & Cassell D.L. (2007) Stopping stepwise: Why stepwise and similar selection methods are bad, and what you should use. NESUG 2007
- Harrell F.E. (2001) Regression modeling strategies: With applications to linear models, logistic regression, and survival analysis. Springer-Verlag, New York
- Munoz, A.R., Real R., Barbosa A.M. & Vargas J.M. (2005) Modelling the distribution of Bonelli's Eagle in Spain: Implications for conservation planning. Diversity and Distributions 11: 477-486
- Smith G. (2018) Step away from stepwise. Journal of Big Data 32 (<https://doi.org/10.1186/s40537-018-0143-6>)

## See Also

[step](#), [stepByStep](#), [modelTrim](#)

## Examples

```
data(rotif.env)

stepwise(data = rotif.env, sp.col = 18, var.cols = 5:17)
```

---

summaryWald

---

*Model summary with Wald (instead of z) test statistics*


---

## Description

This function produces a summary of a generalized linear model, with the Wald test (instead of the z test) and associated statistics.

## Usage

```
summaryWald(model, interceptLast = TRUE)
```

## Arguments

model	a model object of class "glm".
interceptLast	logical, whether to place the intercept in the last (rather than the first) row of the output. Defaults to TRUE.

## Details

This function requires the **aod** package, whose `wald.test` function is used for computing the Wald test.

## Value

This function returns a data frame with the model summary statistics.

**Author(s)**

A. Marcia Barbosa

**See Also**

[summary](#)

**Examples**

```
# load sample data:

data(rotif.env)
names(rotif.env)

# build a model of a species' occurrence based on
# some of the variables:

model <- glm(Abrigh ~ Area + Altitude + AltitudeRange + HabitatDiversity +
HumanPopulation, family = binomial, data = rotif.env)

# get the Wald-based model summary:

summaryWald(model)
```

---

timer

*Timer*

---

**Description**

Reporting of time elapsed since a given start time. This function is used internally by other functions in the package.

**Usage**

```
timer(start.time)
```

**Arguments**

start.time      A date-time object of class [POSIXct](#), e.g. as given by [Sys.time](#).

**Value**

The function returns a message informing of the time elapsed since the input 'start.time'.

**Author(s)**

A. Marcia Barbosa

See Also

[Sys.time](#), [proc.time](#), [difftime](#)

Examples

```
# get starting time:
start <- Sys.time()

# do some random analysis:
sapply(rnorm(50000), function(x) x*5)

# see how long it took:
timer(start)
```

---

transpose	<i>Transpose (part of) a matrix or dataframe</i>
-----------	--

---

Description

This function transposes (a specified part of) a matrix or data frame, optionally using one of its columns as column names for the transposed result. It can be useful for turning a species presence-absence table into a regional species composition table.

Usage

```
transpose(data, sp.cols = 1:ncol(data), reg.names = NULL)
```

Arguments

- `data` a matrix or data frame containing the species occurrence data to transpose.
- `sp.cols` names or index numbers of the columns containing the species occurrences in 'data' which are meant to be transposed.
- `reg.names` name or index number of the column in 'data' containing the region names, to be used as column names in the transposed result.

Value

This function returns the transposed 'sp.cols' of 'data', with the column specified in 'reg.names' as column names.

Author(s)

A. Marcia Barbosa

See Also

[t](#)



**Examples**

```
data(rotif.env)

head(rotif.env)

names(rotif.env)

rotif.reg <- transpose(rotif.env, sp.cols = 18:47, reg.names = 1)

head(rotif.reg)
```

---

triMatInd	<i>Triangular matrix indices</i>
-----------	----------------------------------

---

**Description**

This function outputs the indices of one triangle (the lower one by default) of an input square matrix. It is used by `simMat` and, for large matrices, makes it faster than e.g. with `lower.tri` or `upper.tri`.

**Usage**

```
triMatInd(mat, lower = TRUE, list = FALSE)
```

**Arguments**

<code>mat</code>	a square matrix.
<code>lower</code>	logical indicating whether the indices should correspond to the lower triangle. The default is TRUE; FALSE produces the upper triangle indices.
<code>list</code>	logical indicating whether the results should be output as a list instead of a matrix. The default is FALSE.

**Value**

The indices (row, column) of the elements of the matrix that belong to the requested triangle.

**Author(s)**

A. Marcia Barbosa

**References**

<http://stackoverflow.com/questions/20898684/how-to-efficiently-generate-lower-triangle-indices-of-a-symmetric-matrix>

**See Also**

`lower.tri`, `upper.tri`

Examples

```
mat <- matrix(nrow = 4, ncol = 4)
mat
triMatInd(mat)
triMatInd(mat, list = TRUE)
```

---

vulnerability	(Fuzzy) vulnerability
---------------	-----------------------

---

Description

This function computes the index of species vulnerability of Estrada et al. (2011), using either crisp (presence/absence, i.e. ones and zeros) or fuzzy (Favourability, between zero and one) values, taking into account the conservation status of each species. Vulnerability is like a (potential) richness index in which more vulnerable species (i.e., those with a more threatened conservation status) have higher weight.

Usage

```
vulnerability(data, sp.cols = 1:ncol(data), categories, na.rm = TRUE)
```

Arguments

data	a numeric vector, matrix or data frame containing the presence/absence (ones and zeros) or the Favourability (fuzzy presence, between zero and one) values for the target species.
sp.cols	names or index numbers of the columns of 'data' that contain the species values for which to compute vulnerability. The default is to use all columns.
categories	numeric vector of the same length as 'sp.cols' (or of length 1 if 'data' is a vector) indicating the IUCN Red List category of each species. This vector should be provided in the same order as the columns in data[ , sp.cols]. See Details.
na.rm	logical value indicating whether NA values should be removed before the computation.

Details

The numeric values for the 'categories' argument are suggested by Estrada et al. (2011) to be as follows for each species, according to its IUCN Red List category (available at <https://www.iucnredlist.org>):

- Critically endangered (CR): 16
- Endangered (EN): 8
- Vulnerable (VU): 4
- Near Threatened (NT): 2
- Least Concern (LC): 1
- Data Deficient (DD): 1

Not evaluated (NE): 0

These values follow an exponential scale, because a critically endangered species is generally considered more important than two endangered species, an endangered species more important than two vulnerable species, and so on (Estrada et al. 2011).

### Value

This function returns a numeric vulnerability value for each value or each row in 'data'.

### Author(s)

A. Marcia Barbosa

### References

Estrada A., Real R. & Vargas J.M. (2011) Assessing coincidence between priority conservation areas for vertebrate groups in a Mediterranean hotspot. *Biological Conservation*, 144: 1120-1129

### See Also

[rarity](#)

### Examples

```
data(rotif.env)

# note the 'categories' below are made up, as rotifers are not on yet redlisted
# see Details above for how to get actual values for your species

vulnerability(rotif.env[, 18], categories = 8)

vulnerability(rotif.env, sp.cols = "Abrigh", categories = 8)

vulnerability(rotif.env, sp.cols = c("Apriod", "Burceo", "Kcochl"), categories = c(8, 16, 2))

# fuzzy vulnerability (from favourability values):

pred <- multGLM(rotif.env, sp.cols = c("Apriod", "Burceo", "Kcochl"), var.cols = 5:17)$predictions

head(pred)

vulnerability(pred, sp.cols = "Apriod_F", categories = 8)

vulnerability(pred, sp.cols = c("Apriod_F", "Burceo_F", "Kcochl_F"), categories = c(8, 16, 2))
```

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